



HAL
open science

**Biased sex ratio in the European eel (*Anguilla anguilla*)
swimbladder parasite *Anguillicola crassus*,
experimentally induced by 11-ketosterone**

G. Fazio, H. Moné, G. Mouahid, P. Sasal

► **To cite this version:**

G. Fazio, H. Moné, G. Mouahid, P. Sasal. Biased sex ratio in the European eel (*Anguilla anguilla*) swimbladder parasite *Anguillicola crassus*, experimentally induced by 11-ketosterone. *Journal of Parasitology*, 2008, 94, pp.956-958. 10.1645/GE-1486.1 . halsde-00344534

HAL Id: halsde-00344534

<https://hal.science/halsde-00344534v1>

Submitted on 5 Dec 2008

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 RH: RESEARCH NOTE

2
3 **BIASED SEX RATIO IN THE EUROPEAN EEL (*ANGUILLA ANGUILLA*)**
4 **SWIMBLADDER PARASITE *ANGUILLICOLA CRASSUS*, EXPERIMENTALLY**
5 **INDUCED BY 11-KETOTESTOSTERONE**

6 **G. Fazio, H. Moné, G. Mouahid, and P. Sasal**, UMR 5244 CNRS-EPHE-UPVD,
7 Biologie et Ecologie Tropicale et Méditerranéenne, Université Via Domitia, 52 Avenue
8 Paul Alduy, 66860 Perpignan Cedex, France. *e-mail: fazio@univ-perp.fr*

9 **ABSTRACT:** Parasites are intimately connected to the host in which they live, and some
10 may be affected by the polluted environment of their host. The present study describes the
11 effect of a steroid hormone (11-ketotestosterone) on the sex ratio of the invasive
12 hematophagous nematode *Anguillicola crassus* Kuwahara, Niimi & Itagaki, 1974, when
13 experimentally injected to European eels, *Anguilla anguilla*, Our results showed that this
14 steroid induced a significant male-biased ratio in the nematode *A. crassus* infrapopulations,
15 suggesting that the presence of endocrine disruptors in the environment may lead to
16 skewed sex ratios among parasites.

17 The biological effects of environmental pollutants on aquatic organisms have recently
18 become a major concern. Pollutants are being discharged continuously into water
19 throughout the world, resulting in substantive changes in aquatic habitats, mainly in
20 freshwater and coastal areas such as estuaries and lagoons. The toxicity of these chemical
21 and biological substances has mostly been evaluated (both in laboratory and field
22 conditions) in fishes, revealing disturbances in feeding (Eddy, 2005), growth (Robinet and
23 Feunteun, 2002), immunity (Dunier and Siwicki, 1993), and reproduction (Jones and
24 Reynolds, 1997; Sumpter, 1997). A great interest was taken in the effects of toxic pollution

1 on parasites, for 2 major reasons. First, parasites, in particular metazoans, may be reliable
2 biological indicators of water contamination (Poulin, 1992; MacKenzie et al., 1995;
3 Marcogliese 2005). Second, the impacts of parasites on their hosts can be positively or
4 negatively affected by pollution (Sures, 2006). However, the former considerations
5 demonstrated the need to understand how pollutants might directly or indirectly influence
6 the prevalence, intensity, and pathogenicity of a parasite. General (non-exclusive) trends
7 have emerged from different studies. Thus, on the one hand, pollutants may increase
8 parasitism by impairing the host immune response or by favoring the survival and
9 reproduction of the intermediate host (Khan and Thulin, 1991; Poulin, 1992; Sures, 2006).
10 On the other hand, pollutants may decrease parasitism by being more toxic to parasites
11 than to their (final) hosts (for example high toxicity to free-living stages), negatively
12 affecting intermediate hosts, or altering the host physiology (Khan and Thulin, 1991;
13 Poulin, 1992; Sures, 2006). In any case, studies are required for each new
14 pollutant/parasite/host system.

15 The present study focused on steroid hormones, which are considered as emerging
16 pollutants (Lopez de Alda et al., 2003). A recent investigation conducted in North America,
17 steroid compounds such as 17α and β -estradiol, estriol, estrone, progesterone, and
18 testosterone, which are generally considered as important reproductive hormones in
19 vertebrate animals, were the ones most frequently found as pollutants in aquatic habitats,
20 and occurred in the highest concentrations (Kolpin et al., 2002). These hormones should
21 disrupt important endocrine-mediated processes and would thus have an effect on various
22 reproductive functions, potentially leading to masculinization or feminization of
23 organisms, as well as biased sex ratios in populations (Christiansen et al., 2002; Khanal et
24 al., 2006; Martinovi et al., 2007).

1 Specifically, 11-ketotestosterone (11-KT) belongs to the family of 11-oxygenated
2 androgens that are typically produced in fish testes (Borg, 1994). It is known that aquatic
3 environments are exposed to 11-oxygenated androgen contamination because of an
4 increase of intensive aquaculture practices (Foresti, 2000; Boyd et al., 2005). Moreover,
5 several xenobiotics that may disrupt endocrine functions are extensively discharged in the
6 environment due to agricultural and industrial practices (Kime, 1999; Zala and Penn,
7 2004). We, therefore, attempted to assess the biological effects of 11-KT on the invasive
8 parasite species, *Anguillicola crassus* Kuwahara, Niimi & Itagaki, 1974, a parasite of the
9 swim bladder of European eels (*Anguilla anguilla* Linnaeus, 1758). In addition to being an
10 invasive species, *A. crassus* is an hematophagus species, bringing it into direct contact with
11 the host blood, in which potential pollutants that may be circulating. Because the parasite is
12 also gonochoric, the aim of the study was to investigate a potential effect of the 11-KT on
13 the parasites' sex ratio.

14 Eels were caught in July 2005 at the Palavasian Lagoons (43.54°N 3.92°E, Hérault,
15 France) by a professional fisherman. Epidemiological surveys carried out in July 2004 on
16 20 eels and 119 more in June and July 2005 revealed surprisingly low prevalences of *A.*
17 *crassus*, and no more than 2 parasites per host (unpubl. obs.). During the July 2005
18 collection, the eels were sorted in order to select only males. Catch effort was focused on
19 eels about to silver (to metamorphose before their oceanic migration to the Sargasso Sea)
20 in order to limit individual variability. The selection was based on 4 criteria, i.e., the total
21 body length (L_T), the ocular index (I_O), the skin coloration, and the differentiation of the
22 lateral line. Eels that were approximately 40 cm and that exhibited ocular hypertrophy, a
23 differentiated lateral line, and a contrasting color (Acou et al., 2005; Durif et al., 2005),
24 were brought to the laboratory in oxygenated lagoon water and transferred to 10, 100-L

1 tanks filled with artificial sea water ($S = 37 \text{ g/L}$). They all received a mebendazole (Sigma;
2 St. Louis, Missouri) treatment (1 mg/L for 24 hr) for monogeneans (Buchmann 1993).
3 Freshwater copepods (*Cyclops* spp.) were collected at the Villeneuve-de-la-Raho Lake
4 ($42.63^\circ \text{ N } 2.90^\circ \text{ E}$, Pyrénées-Orientales, France) with a plankton net. They were fed with
5 second stage (L2) larvae of *A. crassus* recovered from swim bladders of naturally infected
6 eels (around 10 L2 larvae per copepod). They were then maintained at 24 C in oxygenated
7 water and fed once a day with *Paramecium* sp. Third stage larvae developed by 11 days
8 post-infection. The L3 stage was confirmed microscopically by the presence of a brace-
9 shaped sclerified structure at the anterior end of the larvae, called the “buccal
10 ornamentation” (Blanc et al., 1992). At this point, L3 larvae were harvested from the
11 copepods with a pestle in physiological serum (8.5‰) and counted using a binocular
12 microscope.

13 Experimental infections of the eels were performed after at least 1 wk of
14 acclimatization in aquaria. The eels were anesthetized (0.1 ml/L of Eugenol) and measured
15 (total length, L_T). The horizontal (D_h) and vertical (D_v) eye diameters were measured to the
16 nearest 0.1 mm on the left side of exposed eels. The ocular index (I_o) (Pankhurst 1982)
17 was calculated as: $I_o = [((D_h + D_v)/4)^2 * \pi / L_T] * 100$. Batches of 50 L3 larvae were prepared in
18 physiological serum. The larval suspension was drawn into a syringe with a blunt cannula
19 and orally administered intubated into the eels' stomachs. The eels were infected with 50
20 L3 (1 syringe of 50 L3). Of the 36 exposed eels, 16 received an injection of 11-
21 ketotestosterone (11-KT, Sigma); $2 \mu\text{g}$ of 11-KT per 1 g of eel, homogenized in about 0.5
22 ml of physiological serum (6‰), were injected each wk into the body cavity (S. Dufour,
23 pers. comm.). This treatment began 1 wk after the experimental infections and continued
24 for 5 wk. Following the same timing protocol, the remaining 20 eels were injected just with

1 serum.

2 At time of initial exposure to the third-stage larvae, the biological characteristics
3 of the eels were $352 < L_T < 438$ mm and $6.0 < I_O < 8.7$ for (further) non-11-KT treated
4 eels (N=20) and $343 < L_T < 449$ mm and $5.5 < I_O < 8.9$ for (further) 11-KT treated eels
5 (N=16). These differences in treated and untreated eels were not significantly different
6 (Mann-Whitney *U*-tests, L_T : $U=130.5$, $P>0.05$ and I_O : $U=160.0$, $P>0.05$). After 6
7 injections, the eels were anesthetized (0.1 ml / l of Eugenol), weighed and measured
8 (total length) to the nearest 0.1 g and mm, respectively, then instantly killed by
9 beheading.

10 Five mo post-infection, swim bladders were removed and examined using a
11 binocular microscope to recover parasites. The developmental stages, i.e., L3 larvae, L4
12 larvae, and/or adults, as well as sex of the adults, were determined. The recovery
13 success was calculated as the number of recovered parasites divided by the number of
14 intubated L3 larvae. The male-ratio was calculated as the number of recovered males
15 divided by the number of recovered adults, for each eel. Both male and female
16 infrapopulations were singly weighed to the nearest mg. We calculated a mean
17 individual male (female) biomass per infected eel, considering all the males (females),
18 divided by the number of males (females) recovered in each fish. The mean individual
19 male and female biomass and infrapopulation biomass \pm Standard Deviation for each
20 sex were further calculated for 11-KT treated and untreated eels. Non-parametric
21 Mann-Whitney *U*-tests were performed comparing 11-KT treated and 11-KT untreated
22 eels for observed male-ratios, recoveries of male/female parasites, and male/female
23 biomass data.

24 When the 2 sexes are combined, the mean number of parasites recovered was

1 7.4±5.2 (min-max =1-19). The mean recovery success was 0.13±0.09 for 11-KT
2 untreated eels (Ø) and 0.19±0.12 for 11-KT treated (11-KT) eels, and are not
3 significantly different ($n_{\text{Ø}}=20$, $n_{11\text{-KT}}=16$, $U=116$, $P>0.05$). The mean number of males
4 was 2.7±2.2 (min-max=0-8) in untreated eels and 5.4±3.9 (min-max=0-14) in 11-KT
5 injected eels (Fig. 1). The mean number of females was 3.4±2.3 (min-max=0-7) in
6 untreated eels and 3.7±3.2 (min-max=0-12) in 11-KT injected eels (Fig. 1). Mann-
7 Whitney U -tests revealed that the mean number of recovered males was significantly
8 higher in 11-KT eels than in Ø eels ($n_{\text{Ø}}=20$, $n_{11\text{-KT}}=16$, $U=95$, $P=0.037$). However, the
9 numbers of recovered females in Ø and 11-KT eels were not different ($n_{\text{Ø}}=20$, $n_{11\text{-}}$
10 $n_{\text{KT}}=16$, $U=156$, $P>0.05$). The mean male-ratios were 0.42±0.27 and 0.59±0.27 for Ø
11 and 11-KT eels, respectively. A Chi square test did not indicate a significantly biased
12 sex ratio in untreated eels ($\chi^2=0.6$, $\text{ddl}=1$, $P=0.44$), while it was significantly male
13 biased in 11-KT treated eels ($\chi^2=34.1$, $\text{ddl}=1$, $P<0.001$). Figure 2 shows the male and
14 female *A. crassus* individual biomass in Ø and 11-KT eels. Mann-Whitney U -tests did
15 not revealed any significant differences ($n_{\text{Ø}}=17$ and 18, and $n_{11\text{-KT}}= 15$ and 14, for
16 either male and female mean biomass, respectively, $P>0.05$) between Ø and 11-KT
17 eels.

18 Our results revealed a significant male biased sex ratio in the nematode *A.*
19 *crassus* infrapopulation when eels received a 11-ketotestosterone treatment. Studies on
20 the sex ratio of the worm in the field are rare, but the results suggest a naturally
21 equilibrated sex ratio (Belpaire et al., 1989; data not shown). Moreover, we did not
22 observe a significantly biased sex ratio in untreated eels. The distortion observed in
23 injected eels is due to the development of a larger number of males since the overall

1 number of females did not decrease in 11-KT treated eels. Moreover, the bias of the
2 male-ratio was not linked to any change in collective mean male and female biomass.
3 This suggests that the injection of 11-KT into the hosts induced a distortion in the
4 blood sucking parasite sex ratio by promoting male development.

5 We suggest 2 (non-exclusive) hypotheses to explain the results obtained for the
6 host parasite model employed in the present study, and the experimental protocol
7 followed. First, the 11-KT treatment may have increased the eel susceptibility to the
8 nematode by affecting the immune system. Second, the 11-KT treatment may have
9 favored the development of male parasites.

10 The deleterious effect of dihydrotestosterone and testosterone on both innate
11 and acquired immunity has been well studied in mammals, i.e., sex hormone receptors
12 are known to be localized in immune cells such as lymphocytes, macrophages,
13 granulocytes, and mast cells (see Klein, 2004, for a review). However, very little is
14 known with respect to teleost fishes. However, recent studies failed to show an effect of
15 androgen treatments on some components of the immune system in tilapia
16 (*Oreochromis niloticus*) and common carp (*Cyprinus carpio*) (Law et al., 2001) and in
17 tench (*Tinca tinca*) (Vainikka et al., 2005). Furthermore, an immune effect should
18 rather have had consequences for infectivity of the nematode and thus have led to an
19 increase in both the male and female components of the parasite infrapopulation.
20 However, the number of females recovered was not different between treated and
21 untreated eels. Nonetheless, we cannot reject the immunity hypothesis, since other
22 pollutants have been shown to act as immunomodulators in the eel. For example, Sures
23 and Knopf (2004) demonstrated that European eels experimentally infected with L3
24 larvae of *A. crassus* were not able to produce specific antibodies when they were

1 simultaneously exposed to polychlorinated biphenyls (PCBs).

2 The second hypothesis that could explain the male biased sex ratio would be an
3 enhancement of the development of male parasites, by modifying the density-
4 dependent relationships between parasites at the infrapopulation level. Two density-
5 dependent mechanisms are known to constrain the infrapopulation size of *A. crassus*
6 within the eel (Ashworth and Kennedy, 1999). First, adults in the swim bladder of eels
7 were shown to inhibit the development of L3 larvae, in cases of heavy infections, i.e.
8 more than 20 parasites. Second, density-dependence limits the number of female
9 worms reaching maturity because of large size of already present adult females. This
10 latter “crowding effect”, was found by these authors at a mean number of adults
11 ranging between 3 and 9 and could occur in our studied infrapopulations where the
12 mean number of adults \pm standard deviation (min-max) was 7.4 ± 5.2 (1-19). This
13 hypothesis is supported by the blood diet of the nematode, which should account for
14 the transfer of 11-KT from the digestive system to the nematode, as well as the
15 presence of a large number of steroid receptors among various species of nematodes
16 (Höss and Weltje, 2007).

17 Our results suggest an enhancement in the development of male parasites, but we do
18 not actually know on which developmental stage the 11-KT has an effect. Following
19 the findings of Ashworth and Kennedy (1999) on the development of L3, the 11-KT
20 may have had an effect on the relationship between L3 larvae and adults. Moreover,
21 their findings on the constraint of the number of gravid females, suggest the existence
22 of a competition between the adult stages. Both intraspecific interactions (larvae vs.
23 adults or adults vs. adults) involve independent mechanisms that could occur
24 simultaneously. However, more experimental work, especially a molecular protocol to

1 determine the sex of L3, could help in answering these questions.

2 The fact that 11-KT had no effect on the mean parasite biomass suggests that this
3 hormone does not affect morphological characteristics of the nematode. More work is
4 needed to test if other life history traits (for males in particular), i.e., acceleration of
5 growth and acquisition of puberty, or an increase of gonad size and fecundity, would be
6 affected by 11-KT.

7 In conclusion, these preliminary results should be confirmed in field studies by
8 comparing the parasites' sex ratios between polluted and unpolluted habitats.

9 The authors would like to thank Mr Molle for supplying with eels. This work
10 was financially supported by the French Ministère de l'écologie et du développement
11 durable, the CNRS and the University of Perpignan Via Domitia. G.F. was financially
12 supported by a grant from the French Ministère de l'enseignement supérieur et de la
13 recherche.

14 **LITERATURE CITED**

15 Acou, A., P. Boury, P. Laffaille, A.J. Crivelli, and E. Feunteun 2005. Towards a
16 standardized characterization of the potentially migrating silver European eel (*Anguilla*
17 *anguilla*, L.). Archives für Hydrobiologia **164**: 237-255.

18 Ashworth S.T., and C.R. Kennedy. 1999. Density-dependent effects on *Anguillicola*
19 *crassus* (Nematoda) within its European eel definitive host. Parasitology **118**: 289-296.

20 Belpaire, C., D. de Charleroy, K. Thomas, P. van Damme, and F. Ollevier. 1989. Effects
21 of eel restocking on the distribution of the swimbladder nematode *Anguillicola crassus* in
22 Flanders, Belgium. Journal of Applied Ichthyology **5**: 151-153.

23 Blanc, G., S. Bonneau, S. Bagianti, and A.J. Petter. 1992. Description of the larval

1 stages of *Anguillicola crassus* (Nematoda, Dracunculoidea) using light and scanning
2 electron microscopy. *Aquatic Living Resources* **5**: 307-318.

3 Borg, B. 1994. Androgens in teleost fishes. *Comparative Biochemistry and Physiology*
4 **C 109**: 219-245.

5 Boyd, C.E., A.A. McNevin, J. Clay, and H.M. Johnson. 2005. Certification issues for
6 some common aquaculture species. *Review in Fisheries Sciences* **13**: 231-279.

7 Buchmann, K. 1993. Epidemiology and control of *Pseudodactylogyrus* infections in
8 intensive eel culture systems: recent trends. *Bulletin Français de la Pêche et de la*
9 *Pisciculture* **328**: 66-73.

10 Christiansen, L.B., M. Winther-Nielsen, and C. Helwig. 2002. Feminisation of fish.
11 The effect of estrogenic compounds and their fate in sewage treatment plants and
12 nature. Environmental Project No. 729, Danish Environmental Protection Agency 184
13 p. Available at: [http://www.mst.dk/udgiv/publications/2002/87-7972-305-
14 5/html/default_eng.htm](http://www.mst.dk/udgiv/publications/2002/87-7972-305-5/html/default_eng.htm).

15 Dunier, M., and A.K. Siwicki. 1993. Effects of pesticides and other organic pollutants
16 in the aquatic environment on immunity of fish - a review. *Fish and Shellfish*
17 *Immunology* **3**: 423-438.

18 Durif, C., S. Dufour, and P. Elie. 2005. The silvering process of *Anguilla anguilla*: a
19 new classification from the yellow resident to the silver migrating stage. *Journal of Fish*
20 *Biology* **66**: 1025-1043.

21 Eddy, F.B. 2005. Ammonia in estuaries and effects on fish. *Journal of Fish Biology* **67**:
22 1495-1513.

23 Foresti, F. 2000. Biotechnology and fish culture. *Hydrobiologia* **420**: 45-47.

24 Harvey, S.C., and M.E. Viney. 2001. Sex determination in the parasitic nematode

1 *Strongyloides ratti*. *Genetics* **158**: 1527-1533.

2 Hodgkin, J. 1983. Two types of sex determination in a nematode. *Nature* **304**: 267-268.

3 Höss, S., and L. Weltje. 2007. Endocrine disruption in nematodes: effects and
4 mechanisms. *Ecotoxicology* **16**: 15-28.

5 Jones, J.C., and J.D. Reynolds. 1997. Effects of pollution on reproductive behaviour of
6 fishes. *Review in Fish Biology and Fisheries* **7**: 463-491.

7 Khan, R.A., and J. Thulin. 1991. Influence of pollution on parasites of aquatic animals.
8 *Advances in Parasitology* **30**: 201-238.

9 Khanal, S.K., B. Xie , M.L. Thompson, S.W. Sung, S.K. Ong, and J. Van Leeuwen.
10 2006. Fate, transport, and biodegradation of natural estrogens in the environment and
11 engineered systems. *Environmental Sciences and Technology* **40**: 6537-6546.

12 Kime, D.E. 1999. A strategy for assessing the effects of xenobiotics on fish
13 reproduction. *The Science of the Total Environment* **225**: 3-11.

14 Klein, S.L. 2004. Hormonal and immunological mechanisms mediating sex differences
15 in parasite infection. *Parasite Immunology* **26**: 247-264.

16 Kolpin, D.W., E.T. Furlong, M.T. Meyer, E.M. Thurman, S.D. Zaugg, L.B. Barber, and
17 H.T. Buxton. 2002. Pharmaceuticals, hormones, and other organic wastewater
18 contaminants in U.S. streams, 1999-2000: a national reconnaissance. *Environmental*
19 *Sciences and Technology* **36**: 1202-1211.

20 Law, W.Y., W.H. Chen, Y.L. Song, S. Dufour, and C.F. Chang. 2001. Differential in
21 vitro suppressive effects of steroids on leukocyte phagocytosis in two teleosts, tilapia
22 and common carp. *General and Comparative Endocrinology* **121**: 163-172.

23 Lopez de Alda, M.J., S. Diaz-Cruz, M. Petrovic, and D. Barcelo. 2003. Liquid
24 chromatography-(tandem) mass spectrometry of selected emerging pollutants (steroid

1 sex hormones, drugs and alkylphenolic surfactants) in the aquatic environment. Journal
2 of Chromatography A **1000**: 503-526.

3 MacKenzie, K., H.H. Williams, B. Williams, A.H. McVicar, and R. Siddall. 1995.
4 Parasites as indicators of water-quality and the potential use of helminth transmission
5 in marine pollution studies. *Advances in Parasitology* **35**: 85-144.

6 Marcogliese, D.J. 2005. Parasites of the superorganism: are they indicators of
7 ecosystem health? *International Journal for Parasitology* **35**: 705-716.

8 Martinovi, D., W.T. Hogarth, R.E. Jones, and P.W. Sorensen. 2007. Environmental
9 estrogens suppress hormones, behaviour, and reproductive fitness in male fathead
10 minnows. *Environmental Toxicology and Chemistry* **26**: 271-278.

11 Pankhurst, N.W. 1982. Relation of visual changes to the onset of sexual maturation in
12 the European eel *Anguilla anguilla* (L.). *Journal of Fish Biology* **21**: 127-140.

13 Poulin, R. 1992. Toxic pollution and parasitism in freshwater fish. *Parasitology Today*
14 **8**: 58-61.

15 Robinet T.T., and E.E. Feunteun. 2002. Sublethal effects of exposure to chemical
16 compounds: a cause for the decline in Atlantic eels? *Ecotoxicology* **11**: 265-277.

17 Sumpter, J.P. 1997. Environmental control of fish reproduction: a different perspective.
18 *Fish Physiology and Biochemistry* **17**: 25-31.

19 Sures, B. 2006. How parasitism and pollution affect the physiological homeostasis of
20 aquatic hosts. *Journal of Helminthology* **80**: 151-157.

21 _____, and K. Knopf. 2004. Individual and combined effects of cadmium and 3,3
22 ', 4,4 ', 5-pentachlorobiphenyl (PCB 126) on the humoral immune response in European
23 eel (*Anguilla anguilla*) experimentally infected with larvae of *Anguillicola crassus*
24 (Nematoda). *Parasitology* **128**: 445-454.

1 Underwood, A.P., and A.E. Bianco. 1999. Identification of a molecular marker for the
2 Y chromosome of *Brugia malayi*. *Molecular Biochemistry and Parasitology* **99**: 1-10.
3 Vainikka, A., E.I. Jokinen, R. Kortet, S. Paukku, J. Pirhonen, M.J. Rantala, and J.
4 Taskinen . 2005. Effects of testosterone and beta-glucan on immune functions in tench.
5 *Journal of Fish Biology* **66**: 348-361.
6 Zala, S.M., and D.J. Penn. 2004. Abnormal behaviours induced by chemical pollution:
7 a review of the evidence and new challenges. *Animal Behaviour* **68**: 649-664.
8
9
10
11
12

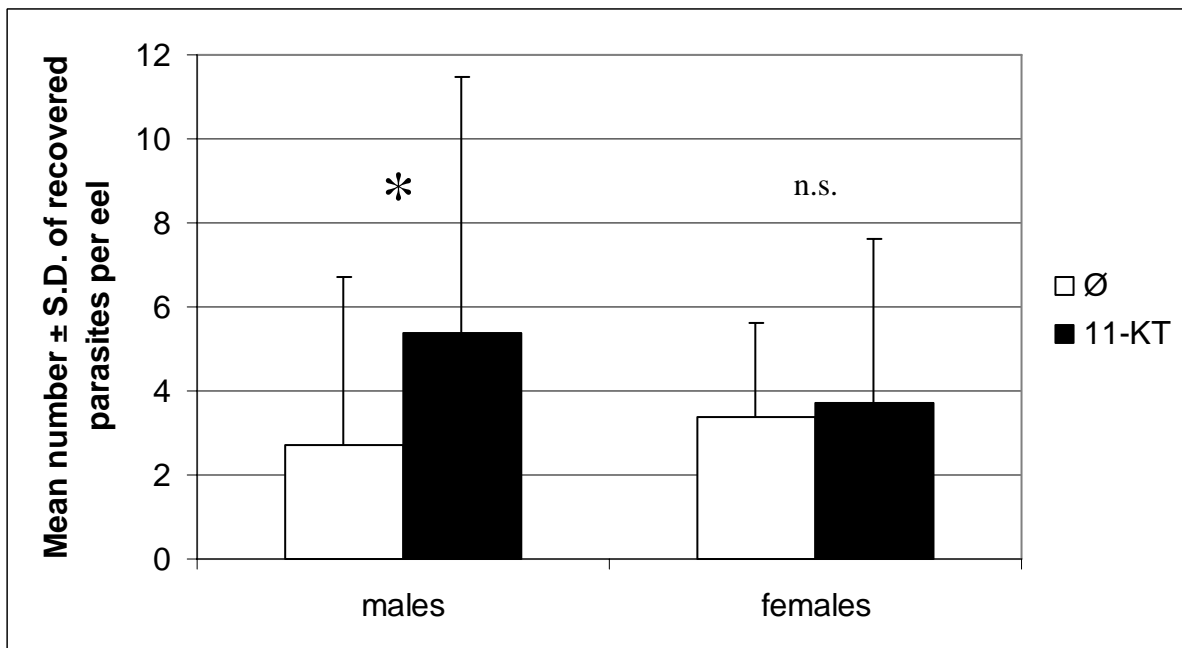


Fig. 1. Mean number (\pm SD) of male and female *A. crassus* recovered in experimentally infected eels, after a 6 weeks-long 11-ketotestosterone treatment (black bars, N = 16 eels) or no treatment (white bars, N = 20 eels). “*” $p < 0.05$ (Mann-Whitney test between treated and non-treated eels). S.D., standard deviation; n.s., not significant.

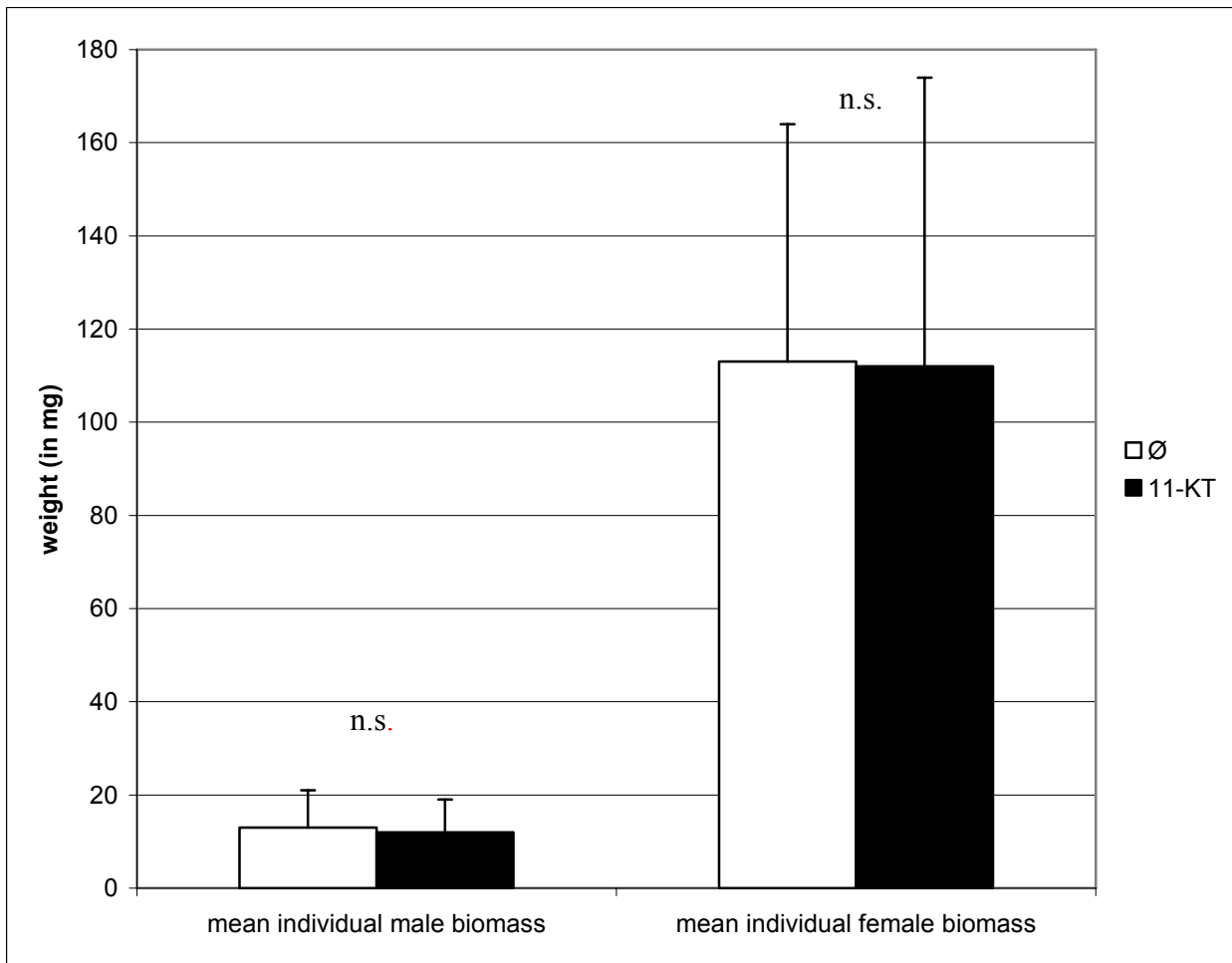


Fig. 2. Male and female *A. crassus* individual biomasses in experimentally infected eels, after a 6 weeks-long 11-ketotestosterone treatment (black bars, N = 15 and 14 eels for male and female data, respectively) or no treatment (white bars, N = 17 and 18 eels for male and female data, respectively). No significant difference (n.s.) was found between treated and non-treated eels (Mann-Whitney test).