

Transgenic corn syrup enhances in vitro yeast ethanol production: Hidden cause of the US MASH epidemic?

Sibiri Sissoko, Armstrong Nicholas, Tidjani Alou, Amael Fadlane, Thera Ma,

René Gerolami, Didier Raoult, Stéphane Ranque

▶ To cite this version:

Sibiri Sissoko, Armstrong Nicholas, Tidjani Alou, Amael Fadlane, Thera Ma, et al.. Transgenic corn syrup enhances in vitro yeast ethanol production: Hidden cause of the US MASH epidemic?. 2025. hal-04930585

HAL Id: hal-04930585 https://hal.science/hal-04930585v1

Preprint submitted on 5 Feb 2025

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Transgenic corn syrup enhances in vitro yeast ethanol production:
2	Hidden cause of the US MASH epidemic?
3	
4	Short title: Transgenic corn syrup enhances in vitro yeast ethanol production
5	
6 7	Sissoko S ^{1,2,3} , Armstrong N ⁴ , Tidjani Alou M ^{1,3} , Fadlane A ^{1,4} , Thera MA ² , Gerolami R ^{1,3,5} , Ranque S ^{1,4,6} , Raoult D ^{1,3} .
8	
9 10 11 12 13 14 15	 Institut Hospitalo-Universitaire Méditerranée Infection (IHU), 13005 Marseille, France. Malaria Research and Training Center (MRTC), FMOS-FAPH, Mali-NIAID-ICER, Université des Sciences, des Techniques et des Technologies de Bamako, BP 423, Mali. Aix-Marseille University, MEPHI, 13005 Marseille. France Assistance Publique - Hôpitaux de Marseille (AP-HM), Marseille, France Unité Hépatologie, Hôpital de la Timone, APHM, 13005 Marseille, France Aix-Marseille University, SSA, UMR D 257, RITMES, 13005 Marseille. France
16 17	For correspondence: stephane.ranque@univ-amu.fr
18	
19	

- 20 Abstract
- 21

The incidence of metabolic dysfunction-associated liver steatohepatitis (MASH) has risen over 22 the last decade, making it a major health problem and the second leading cause of liver disease in 23 24 the US. Recent studies suggest that MASH may be linked to the endogenous production of 25 alcohol in the digestive tract after sugar ingestion, with bacteria such as *Klebsiella* and *Lactobacillus* playing a key role in this fermentation. More recently, the role of the gut 26 27 mycobiota has also been suggested. In particular, MASH has been recently associated with the 28 presence of yeast strains that produce high concentrations of alcohol in the gut microbiota. As 29 the source of sugar varies by region - transgenic maize in the USA, sugar cane in Africa and beet 30 in France - we hypothesised that consumption of sweetened beverages, in combination with the 31 gut mycobiota, may be a contributing factor to MASH. 32 Analysis of American and French Coca-Cola and World-Cola from Mali revealed significant differences in carbohydrate composition, with the presence of complex oligosaccharides in 33 American Coca-Cola. A significantly higher ethanol production was observed when a *Candida* 34

albicans yeast strain was cultured in American Coca-Cola compared to the other two colas. The results suggest that consumption of transgenic maize sweeteners may increase the risk of autobrowery syndrome and underline the need for epidemiological studies to assess the association of these carbohydrates with the MASH epidemic in the US.

39

- 42
- 43

Keywords: Soda; coke; Coca-Cola, carbohydrates; transgenic maize, corn syrup, alcohol, gut
 mycobiota; *Candida albicans*, in vitro.

44 Introduction

45 There are several stages of liver damage in metabolic dysfunction-associated liver steatohepatitis (MASH). The first is steatosis, which in some cases can lead to fibrosis,¹ then to cirrhosis,^{2,3,4} 46 and finally to liver cancer^{5,6}. Currently, MASH is a major liver problem, being the second most 47 common cause of liver disease in patients awaiting liver transplantation in the United States^{7,8}. In 48 the last decade, the prevalence of MASH has increased by almost 100%⁹. Recently, several 49 studies have shown that MASH is probably related to the endogenous production of alcohol in 50 the digestive tract after the ingestion of sugar^{10,11,12}. Some authors have shown that *Klebsiella* 51 bacteria are responsible for this fermentation¹³; others have highlighted the role of *Lactobacillus* 52 bacteria^{14,15} and confirmed the presence of alcohol in the hepatic portal vein of MASH patients 53 sampled during bariatric surgery¹⁶. Another team also showed that alcohol in stool or blood 54 samples from MASH patients was associated with specific Klebsiella pneumoniae strains¹⁷. 55 56 Fungal colonisation of the human gastrointestinal tract is common, particularly with *Candida* spp., and half of the human population in the US is colonised with at least one *Candida* spp^{18} . 57 Our team has shown that the presence of alcohol in stool samples from MASH patients is 58 associated with *Candida* yeast colonisation¹⁸. Others have shown that alcohol in stool or blood 59 samples from MASH patients is associated with specific Klebsiella pneumoniae strains¹⁹ or 60 Saccharomyces cerevisiae yeast²⁰. It has long been known that MASH is associated with sugar 61 consumption¹². In addition, the use of antibiotics prior to the onset of MASH can undoubtedly 62 explain the change in the digestive ecosystem that favours the proliferation and spread of 63 yeasts²¹. Struck by the significance of the MASH epidemic in the United States, we hypothesised 64 that it might be related to the consumption of sweetened products, since the composition of sweet 65 soft drinks, especially colas, varies from country to country. In the USA, colas are sweetened 66 with sugar extract from transgenic maize ^{22,23} whereas in Africa the sugar comes from sugar 67

cane,^{24,25} and in France (Europe) from beetroot (Coca Cola European Partners).²⁶ The aim of this
study was to evaluate the in vitro ethanol production of yeasts incubated in colas produced in the
USA, Africa or France.

71

72 Materials and Methods

73 Cola samples

We analysed a total of three different sugared colas produced by the Coca-Cola Company in the USA and in France. In Mali, World-Cola is produced by the Castel Group, which is present in 14 countries in sub-Saharan Africa. Coca-Cola original taste and World-Cola was purchased in local stores in France or Mali. Classic American Coca-Cola was purchased from the My American Market online store (21 August 2023). It is worth noting that the source of the sugar used to sweeten colas varies depending on where they are produced. Transgenic maize is used in the USA, sugar beet in France and sugar cane in Mali.

81 *Analysis of the carbohydrate composition of colas*

The carbohydrate content of colas was studied by ultra-high performance liquid 82 chromatography-mass spectrometry (UHPLC-MS) according to an application note from Waters 83 Corporation (Shelton, USA), with modifications.²⁷ Mono-, di- and oligosaccharides were 84 85 separated by hydrophilic interaction liquid chromatography (HILIC) using a BEH amide column (2.1x50 mm, Waters) installed in an iClass UHPLC system (Waters), which in turn was 86 87 connected to a Vion IMS mass spectrometer (Waters). To study the composition of free carbohydrate structures, samples were diluted in water only, whereas to study the 88 89 monosaccharide composition of complex carbohydrates, samples were first evaporated and hydrolysed with trifluoroacetic acid (4 h, 100%); the acid was then evaporated and the resulting 90 samples dissolved in water. All samples were injected directly into the amide column and the 91 compounds were separated according to the following solvent composition gradient (A: 80:20% 92 93 acetonitrile/water with 0.1% NH4OH; B: 30:70% acetonitrile/water with 0.1% NH4OH): linear increase of B from 0 to 60% between 0 and 6 min. The carbohydrates were then ionised in 94 negative mode using an electrospray ion source and the resulting ions were monitored using an 95

accurate mass screening method including ion mobility separation and MSe data acquisition
(HDMSe). Data processing was performed using UNIFI software (version 8.1.4).

98 *Quantification of ethanol after inoculation with the* C. albicans strain CSURL 0240

Inoculum and fermentation conditions: C. albicans strain CSURL 0240. This yeast was 99 inoculated in triplicate at a cell density of 3 McFarland into 1 mL of each cola sample in the 100 flask vial tube. The inoculated flasks were incubated at 28 °C for 48 hours. Ethanol was 101 quantified at 24, 36 and 48 hours of incubation by headspace gas chromatography-mass 102 spectrometry (HS-GC/MS) as previously described.²⁸ Briefly, the samples and ethanol standards 103 (20-1800 mM) were all spiked with isopropanol as an internal standard. They were then heated 104 to 60 °C to evaporate the alcohol molecules. The pressurised headspace gases were transferred to 105 the gas chromatography system consisting of a Turbomatrix HS110 sampler connected to a 106 107 Clarus 690 chromatograph and an SQ8 T single quadrupole mass spectrometer (Perkin Elmer, 108 Waltham, MA, USA). The analytes were separated on an Elite BAC-2 column (Perkin Elmer). 109 Ethanol and isopropanol were selectively detected and automated quantification was performed 110 using Turbomass 6.1 software (Perkin Elmer). The influence of both the origin of the colas and 111 the incubation time on the ethanol concentration was tested by two-way ANOVA using the statistical software SAS Stat (ver. 9.4). A P value less that 0.05 was considered statistically 112 significant. 113

114

115 **Results**

116 *The carbohydrate composition of colas*

As illustrated in **Figure 1**, UHPLC-MS detected a unique disaccharide, identified as sucrose, in both Malian World-Cola and French Coca-Cola. In contrast, sugars with a high degree of polymerisation were present in American Coca-Cola. The structures of the sugars from transgenic maize are drastically different from those of sugar cane or sugar beet, as the analysis revealed several oligosaccharides with up to 8 polymerised sugar branches (hexoses), and another disaccharide of unknown composition.

123

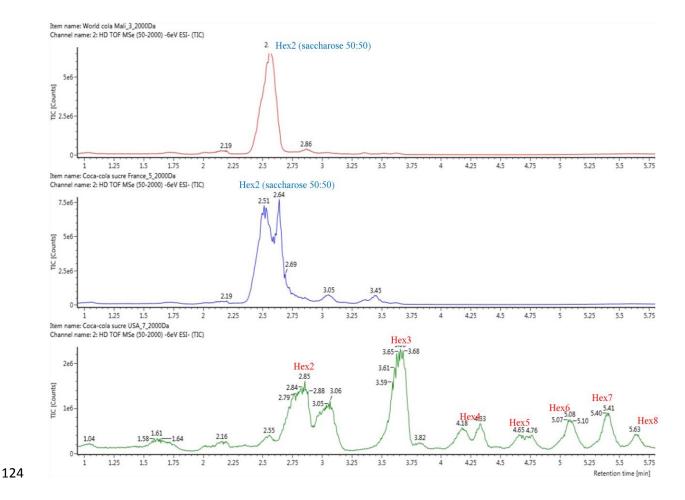
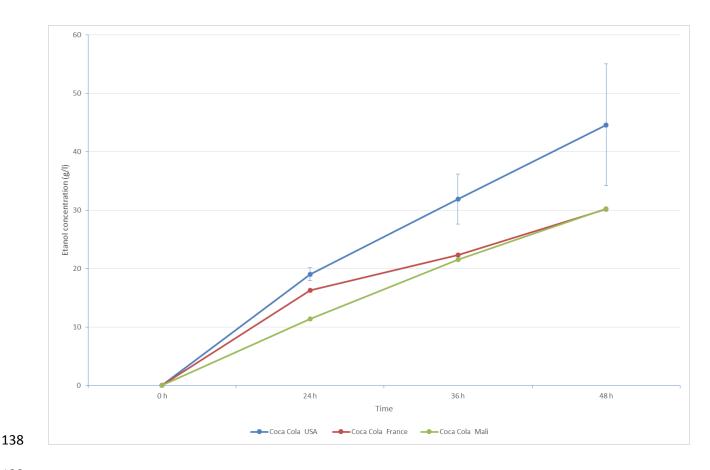


Figure 1. Ultra-high performance liquid chromatography-mass spectrometry carbohydrate analysis showing the high degree of polymerisation of free sugars present in American Coca-Cola, in contrast to the unique presence of saccharose in colas from France or Mali. A: Coca-Cola from Mali; B: Coca-Cola from France; C: Coca-Cola from the USA.

129

130 *Quantification of ethanol production by* C. albicans *strain CSURL 0240*

The concentrations of ethanol produced by *C. albicans* in colas of different origins are shown in *Figure 2*. Both incubation time and cola substrate ($P < 10^{-4}$ each) had a statistically significant effect on the measured ethanol concentration. As shown in **Figure 2**, yeast fermentation of carbohydrates produced a significantly higher ethanol concentration in American Coca-Cola than in the other colas after 36 hours of incubation. Ethanol production was similar in French and Malian colas (**Figure 2**).



139

Figure 2: Overtime in vitro ethanol production by *Candida albicans* CSURL 0240 in colas from the USA, France, or Mali. The yeast fermentation of carbohydrates produced a significantly higher ethanol concentration in American Coca-Cola than in the other colas (ANOVA, $P < 10^{-4}$).

143

144 **Discussion**

In this exploratory study, we highlighted the highly different carbohydrate composition between 145 the colas from the USA compared to those from France or Europe. The variety of 146 oligosaccharides with a degree of polymerization ranging from 3 to 8 in the American Coca-147 Cola contrasted with the presence of unique shorter disaccharide structure in the other colas. In a 148 previous work, we have shown a very high production of ethanol by the yeasts Pichia 149 kudriavzevii, C. glabrata and C. albicans¹⁸. Here, we point out the association of a high degree 150 of polymerisation of the carbohydrates with a significantly higher yeast alcohol production. 151 152 Indeed, the ethanol production of C. albicans CSURL 0240 in American Coca-Cola reached up

to 49.7g/L at 48 hours, which is equivalent to drinking 1 litre of beer at 6% alcohol. People who have yeast in their gut microbiota and drink soda or eat any food sweetened with corn syrup derived from transgenic maize are therefore likely to be at increased risk of autobrowery syndrome. Further epidemiological studies are warranted to assess the influence of consumption of carbohydrates derived from transgenic maize on MASH epidemics in the United States. We believe that if this hypothesis is confirmed, there is likely to be a major public health problem associated with the massive consumption of glucose and fructose oligosaccharide combinations.

160

161

162 **References**

- 163 (1) Anstee, Q. M.; McPherson, S.; Day, C. P. How Big a Problem Is Non-Alcoholic Fatty
 164 Liver Disease? *BMJ* 2011, *343*, d3897. https://doi.org/10.1136/bmj.d3897.
- 165 (2) Caldwell, S. H.; Oelsner, D. H.; Iezzoni, J. C.; Hespenheide, E. E.; Battle, E. H.; Driscoll,
 166 C. J. Cryptogenic Cirrhosis: Clinical Characterization and Risk Factors for Underlying
 167 Disease. *Hepatol. Baltim. Md* 1999, 29 (3), 664–669.
 168 https://doi.org/10.1002/hep.510290347.
- 169 (3) Edmison, J.; McCullough, A. J. Pathogenesis of Non-Alcoholic Steatohepatitis: Human
 170 Data. *Clin. Liver Dis.* 2007, *11* (1), 75–104, ix. https://doi.org/10.1016/j.cld.2007.02.011.
- 171 (4) Ertle, J.; Dechêne, A.; Sowa, J.-P.; Penndorf, V.; Herzer, K.; Kaiser, G.; Schlaak, J. F.;
 172 Gerken, G.; Syn, W.-K.; Canbay, A. Non-Alcoholic Fatty Liver Disease Progresses to
 173 Hepatocellular Carcinoma in the Absence of Apparent Cirrhosis. *Int. J. Cancer* 2011, *128*174 (10), 2436–2443. https://doi.org/10.1002/ijc.25797.
- Bugianesi, E.; Leone, N.; Vanni, E.; Marchesini, G.; Brunello, F.; Carucci, P.; Musso, A.;
 De Paolis, P.; Capussotti, L.; Salizzoni, M.; Rizzetto, M. Expanding the Natural History
 of Nonalcoholic Steatohepatitis: From Cryptogenic Cirrhosis to Hepatocellular
 Carcinoma. *Gastroenterology* 2002, *123* (1), 134–140.
 https://doi.org/10.1053/gast.2002.34168.
- (6) White, D. L.; Kanwal, F.; El-Serag, H. B. Non-Alcoholic Fatty Liver Disease and
 Hepatocellular Cancer: A Systematic Review. *Clin. Gastroenterol. Hepatol. Off. Clin. Pract. J. Am. Gastroenterol. Assoc.* 2012, *10* (12), 1342-1359.e2.
 https://doi.org/10.1016/j.cgh.2012.10.001.
- (7) Cholankeril, G.; Wong, R. J.; Hu, M.; Perumpail, R. B.; Yoo, E. R.; Puri, P.; Younossi,
 Z. M.; Harrison, S. A.; Ahmed, A. Liver Transplantation for Nonalcoholic Steatohepatitis
 in the US: Temporal Trends and Outcomes. *Dig. Dis. Sci.* 2017, *62* (10), 2915–2922.
 https://doi.org/10.1007/s10620-017-4684-x.

188 189 190 191	(8)	 Wong, R. J.; Aguilar, M.; Cheung, R.; Perumpail, R. B.; Harrison, S. A.; Younossi, Z. M.; Ahmed, A. Nonalcoholic Steatohepatitis Is the Second Leading Etiology of Liver Disease Among Adults Awaiting Liver Transplantation in the United States. <i>Gastroenterology</i> 2015, <i>148</i> (3), 547–555. https://doi.org/10.1053/j.gastro.2014.11.039.
192 193 194 195	(9)	Hamid, O.; Eltelbany, A.; Mohammed, A.; Alsabbagh Alchirazi, K.; Trakroo, S.; Asaad, I. The Epidemiology of Non-Alcoholic Steatohepatitis (NASH) in the United States between 2010-2020: A Population-Based Study. <i>Ann. Hepatol.</i> 2022 , <i>27</i> (5), 100727. https://doi.org/10.1016/j.aohep.2022.100727.
196 197 198 199	(10)	Kaji, H.; Asanuma, Y.; Ide, H.; Saito, N.; Hisamura, M.; Murao, M.; Yoshida, T.; Takahashi, K. The Auto-Brewery Syndromethe Repeated Attacks of Alcoholic Intoxication Due to the Overgrowth of Candida (Albicans) in the Gastrointestinal Tract. <i>Mater. Medica Pol. Pol. J. Med. Pharm.</i> 1976 , <i>8</i> (4), 429–435.
200 201 202	(11)	Logan, B. K.; Jones, A. W. Endogenous Ethanol "auto-Brewery Syndrome" as a Drunk- Driving Defence Challenge. <i>Med. Sci. Law</i> 2000 , <i>40</i> (3), 206–215. https://doi.org/10.1177/002580240004000304.
203 204 205 206	(12)	Tseng, TS.; Lin, WT.; Ting, PS.; Huang, CK.; Chen, PH.; Gonzalez, G. V.; Lin, HY. Sugar-Sweetened Beverages and Artificially Sweetened Beverages Consumption and the Risk of Nonalcoholic Fatty Liver (NAFLD) and Nonalcoholic Steatohepatitis (NASH). <i>Nutrients</i> 2023 , <i>15</i> (18), 3997. https://doi.org/10.3390/nu15183997.
207 208 209 210 211 212	(13)	Xue, G.; Feng, J.; Zhang, R.; Du, B.; Sun, Y.; Liu, S.; Yan, C.; Liu, X.; Du, S.; Feng, Y.; Cui, J.; Gan, L.; Zhao, H.; Fan, Z.; Cui, X.; Xu, Z.; Fu, T.; Li, C.; Huang, L.; Zhang, T.; Wang, J.; Yang, R.; Yuan, J. Three Klebsiella Species as Potential Pathobionts Generating Endogenous Ethanol in a Clinical Cohort of Patients with Auto-Brewery Syndrome: A Case Control Study. <i>eBioMedicine</i> 2023 , <i>91</i> , 104560. https://doi.org/10.1016/j.ebiom.2023.104560.
213 214 215 216 217	(14)	Tsai, MC.; Liu, YY.; Lin, CC.; Wang, CC.; Wu, YJ.; Yong, CC.; Chen, KD.; Chuah, SK.; Yao, CC.; Huang, PY.; Chen, CH.; Hu, TH.; Chen, CL. Gut Microbiota Dysbiosis in Patients with Biopsy-Proven Nonalcoholic Fatty Liver Disease: A Cross-Sectional Study in Taiwan. <i>Nutrients</i> 2020 , <i>12</i> (3), 820. https://doi.org/10.3390/nu12030820.
218 219 220 221 222	(15)	Vallianou, N.; Christodoulatos, G. S.; Karampela, I.; Tsilingiris, D.; Magkos, F.; Stratigou, T.; Kounatidis, D.; Dalamaga, M. Understanding the Role of the Gut Microbiome and Microbial Metabolites in Non-Alcoholic Fatty Liver Disease: Current Evidence and Perspectives. <i>Biomolecules</i> 2021 , <i>12</i> (1), 56. https://doi.org/10.3390/biom12010056.
223 224 225 226 227	(16)	Meijnikman, A. S.; Davids, M.; Herrema, H.; Aydin, O.; Tremaroli, V.; Rios-Morales, M.; Levels, H.; Bruin, S.; de Brauw, M.; Verheij, J.; Kemper, M.; Holleboom, A. G.; Tushuizen, M. E.; Schwartz, T. W.; Nielsen, J.; Brandjes, D.; Dirinck, E.; Weyler, J.; Verrijken, A.; De Block, C. E. M.; Vonghia, L.; Francque, S.; Beuers, U.; Gerdes, V. E. A.; Bäckhed, F.; Groen, A. K.; Nieuwdorp, M. Microbiome-Derived Ethanol in

Nonalcoholic Fatty Liver Disease. *Nat. Med.* 2022, 28 (10), 2100–2106.
 https://doi.org/10.1038/s41591-022-02016-6.

Yuan, J.; Chen, C.; Cui, J.; Lu, J.; Yan, C.; Wei, X.; Zhao, X.; Li, N.; Li, S.; Xue, G.;

(17)

230

Cheng, W.; Li, B.; Li, H.; Lin, W.; Tian, C.; Zhao, J.; Han, J.; An, D.; Zhang, Q.; Wei, 231 232 H.; Zheng, M.; Ma, X.; Li, W.; Chen, X.; Zhang, Z.; Zeng, H.; Ying, S.; Wu, J.; Yang, R.; Liu, D. Fatty Liver Disease Caused by High-Alcohol-Producing Klebsiella 233 Pneumoniae. Cell Metab. 2019, 30 (4), 675-688.e7. 234 https://doi.org/10.1016/j.cmet.2019.08.018. 235 236 (18)Mbaye, B.; Borentain, P.; Magdy Wasfy, R.; Alou, M. T.; Armstrong, N.; Mottola, G.; Meddeb, L.; Ranque, S.; Gérolami, R.; Million, M.; Raoult, D. Endogenous Ethanol and 237 238 Triglyceride Production by Gut Pichia Kudriavzevii, Candida Albicans and Candida Glabrata Yeasts in Non-Alcoholic Steatohepatitis. Cells 2022, 11 (21), 3390. 239 https://doi.org/10.3390/cells11213390. 240 Bernhardt, H. H. J. Winner Und Rosalinde Hurley, Candida Albicans. X u. 306 S., 45 241 (19)Abb., London 1964: J. & A. Churchill Sh 60.-.. Z. Für Allg. Mikrobiol. 1965, 5 (5), 410-242 410. https://doi.org/10.1002/jobm.19650050519. 243 Demir, M.; Lang, S.; Hartmann, P.; Duan, Y.; Martin, A.; Miyamoto, Y.; Bondareva, M.; (20)244 Zhang, X.; Wang, Y.; Kasper, P.; Bang, C.; Roderburg, C.; Tacke, F.; Steffen, H.-M.; 245 246 Goeser, T.; Kruglov, A.; Eckmann, L.; Stärkel, P.; Fouts, D. E.; Schnabl, B. The Fecal Mycobiome in Non-Alcoholic Fatty Liver Disease. J. Hepatol. 2022, 76 (4), 788–799. 247 https://doi.org/10.1016/j.jhep.2021.11.029. 248 d'Enfert, C.; Kaune, A.-K.; Alaban, L.-R.; Chakraborty, S.; Cole, N.; Delavy, M.; 249 (21)250 Kosmala, D.; Marsaux, B.; Fróis-Martins, R.; Morelli, M.; Rosati, D.; Valentine, M.; Xie, Z.; Emritloll, Y.; Warn, P. A.; Bequet, F.; Bougnoux, M.-E.; Bornes, S.; Gresnigt, M. S.; 251 Hube, B.; Jacobsen, I. D.; Legrand, M.; Leibundgut-Landmann, S.; Manichanh, C.; 252 253 Munro, C. A.; Netea, M. G.; Queiroz, K.; Roget, K.; Thomas, V.; Thoral, C.; Van den Abbeele, P.; Walker, A. W.; Brown, A. J. P. The Impact of the Fungus-Host-254 Microbiota Interplay upon Candida Albicans Infections: Current Knowledge and New 255 Perspectives. FEMS Microbiol. Rev. 2020, 45 (3), fuaa060. 256 https://doi.org/10.1093/femsre/fuaa060. 257 (22)Christophe Rondot. Qu'y a-t-il dans le Coca-Cola? ChimieParis 2012. 337, 12-15. 258 https://www.mediachimie.org/sites/default/files/CP 337 12.pdf 259 (23)Tom Philpott, T. The secret history of why soda companies switched from sugar to high-260 fructose corn syrup. Mother Jones. https://www.motherjones.com/food/2019/07/the-261 262 secret-history-of-why-soda-companies-switched-from-sugar-to-high-fructose-corn-syrup/ (accessed 2023-12-28). 263 (24)Rogier Mathieu, M. SOMDIAA-RAPPORT DEVELOPPEMENT DURABLE. 264 https://docplayer.fr/89462951-Rapport-developpement-durable.html (accessed 2024-01-265 02). 266

267 268 269	(25)	Oxfam-France, O. <i>Les géants de l'agroalimentaire et la fièvre du sucre</i> . Oxfam France. https://www.oxfamfrance.org/communiques-de-presse/les-geants-de-lagroalimentaire-et-la-fievre-du-sucre/ (accessed 2023-12-28).
270 271 272	(26)	<i>Coca-Cola une production au coeur des territoires français</i> . France. https://www.cocacolaep.com/fr/actualites/2019/coca-cola-une-production-au-coeur-des-territoires-francais/ (accessed 2023-09-19).
273 274 275	(27)	Fountain, K. J.; Hudalla, C. J.; McCabe, D. R.; Morrison, D. Analysis of Carbohydrates by UltraPerformance Liquid Chromatography and Mass Spectrometry. 2009 . 720003212en, Waters Corporation
276 277 278 279 280	(28)	Magdy Wasfy, R.; Mbaye, B.; Borentain, P.; Tidjani Alou, M.; Murillo Ruiz, M.L.; Caputo, A.; Andrieu, C.; Armstrong, N.; Million, M.; Gerolami, R. Ethanol-Producing Enterocloster bolteae Is Enriched in Chronic Hepatitis B-Associated Gut Dysbiosis: A Case–Control Culturomics Study. <i>Microorganisms</i> 2023 , <i>11</i> , 2437. https://doi.org/10.3390/microorganisms11102437

281