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# Interaction Between Plants Through Litter Input in Mangrove Succession in the Red River Delta

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Keywords: allelopathy | Avicennia marina | Kandelia obovata | litter decomposition | mangrove | plant-plant interaction | soil metabolomics | specialized metabolites | succession

### ABSTRACT

**Aims:** Dominant or pioneer species litter decomposition was found to be involved in the recruitment of following species during succession in terrestrial plant communities, while knowledge remains scarce in wetland forests such as mangroves. Our study evaluated the influence of pioneer species leaf litter decomposition on (i) the settlement success of species following them or not in the natural succession and (ii) on soil characteristics.

Location: Red River Delta, Vietnam.

**Methods:** We set up an in situ experiment in mesocosms that followed the survival and growth of three species (*Kandelia obovata*, *Aegiceras corniculatum*, and *Rhizophora stylosa*) planted in soils of *Avicennia marina* or *Sonneratia caseolaris*, with or without leaf litter addition for 203 days. Soil nutrient contents were determined at the beginning and end of the experiment, and their metabolic fingerprints were compared using an untargeted metabolomic approach to highlight potential allelochemicals when a phytotoxic effect was observed.

**Results:** Seedling response to litter addition was species-specific, which can be linked to their successional status. *K. obovata* was most affected by litter addition, showing better growth with both litter types. Meanwhile, *A. marina* litter substantially decreased survival rates by 50% for *K. obovata* and 33% for *R. stylosa*, suggesting a strategy to delay its replacement by these species through the release of phytotoxic decomposition by-products. Metabolic fingerprints of *A. marina* soils planted with *K. obovata* showed no shift in global chemical composition, but 12 features were found significantly more abundant in the presence of litter. Putative annotations revealed sulfur-containing metabolites, which are probable products of organic matter sulfurization, a frequently occurring process in mangrove sediments. These unusual compounds may act as allelochemicals and should be further assessed.

**Conclusions:** Our findings suggest that allelopathic processes through litter decomposition may be at play in mangrove community assembly and could be a tool for restoration purposes.

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# 1 | Introduction

The diversity and composition of plant communities is the result of a complex network of interactions with their environment. Competition for resources, facilitation, and allelopathy are among these structuring interactions (Callaway and Walker 1997; Reigosa et al. 2006). Allelopathy can act as a process in forest renewal and species composition (Mallik 2003; Hashoum et al. 2017), particularly during succession, when plant species are replaced (Gallet and Pellissier 2002; Reigosa et al. 2006; Fernandez et al. 2006), with effects that can either interfere or facilitate the establishment of species from the next successional stage (Fernandez et al. 2013; Meiners 2014).

Litter decomposition is a key element in ecosystem productivity, but its by-products also showed various allelopathic effects on neighboring plants according to litter chemistry and stage of decomposition (Sarker et al. 2020; Bonanomi et al. 2021). Through leaching and microbial degradation, this fundamental process causes the progressive release of essential mineral nutrients (Vitousek and Sanford 1986) as well as low and high molecular weight soluble organic compounds in the soil, which can be beneficial for plant growth (Chomel et al. 2016). Meanwhile, through physicochemical and biological transformations, these chemicals can either increase or decrease their allelopathic effects, ultimately altering neighboring plant growth and functioning, directly or indirectly (An, Pratley, and Haig 2000; Bonanomi et al. 2006; Delory et al. 2021).

In the context of plant succession, litter accumulation and decomposition from pioneer or dominant species is one of the leading mechanisms at play in the recruitment of following species, along with light and nutrient availability, or soil characteristics (Facelli and Pickett 1991; Janišová et al. 2007; Koorem, Price, and Moora 2011). Previous studies addressed the implication of allelopathic processes in this phenomenon mainly in terrestrial ecosystems (Kaligarič et al. 2011; Mudrák and Frouz 2012; Meiners 2014), while knowledge remains scarce in wetland forests such as mangroves (Lang et al. 2021). A wide variety of mangrove communities exist as a result of various geomorphological, ecological, and climatic conditions (Feller et al. 2009). Avicennia, Aegiceras, and Sonneratia are among the main pioneer genera that colonize newly formed mudbanks in the Indo-West Pacific region of mangroves (Duke, Ball, and Ellison 1998; Tomlinson 2016).

In the Red River Delta (RRD) in North Vietnam, Avicennia marina, Rhizophora stylosa, Bruguiera gymnorhiza, Aegiceras corniculatum, Kandelia obovata, and Sonneratia caseolaris are naturally occurring true mangrove species. Two main pioneer stages co-exist, mainly according to distance to the shore and water salinity. A. marina colonizes newly formed mudbanks in more saline waters whereas S. caseolaris establishes more inland where waters are brackish (Hong and San 1993). This allows two different succession dynamics to occur: in the mixed stage, A. corniculatum, R. stylosa, B. gymnorhiza, and K. obovata establish under A. marina canopy while A. corniculatum and three semi-mangrove species (Acanthus ilicifolius, Acrostichum aureum, Cyperus malaccensis) establish under S. caseolaris canopy (Hong and San 1993).

Few studies addressed the implications of mangrove allelopathic effects in their successional dynamics. Chen et al. (2009) monitored the development of Kandelia obovata (previously identified as K. candel (Sheue, Liu, and Yong 2003)) watered with aqueous extracts from leaves of A. marina, A. corniculatum, B. gymnorhiza, and R. stylosa and showed contrasted speciesspecific responses. Extracts from A. corniculatum were the least limiting to K. obovata development, which may partly explain the frequent colonization of A. corniculatum stands by K. obovata in the Guangdong province in southern China from which the plants originated. Recent attention was given to the effects of allelochemicals from litterfall in mangrove natural regeneration in the same province, where Lang et al. (2021) showed that leaf litter leachates from K. obovata significantly inhibited the germination and growth of its associated species A. corniculatum in a microcosm experiment. Their results highlighted condensed tannins as the main allelochemical compounds, but also suggested that other bioactive materials from leaf litter could be at play. To our knowledge, Chapman and Feller (2011) is the only study that addressed the direct effect of mangrove leaf litter addition on mangrove community composition, showing an influence of litter identity on seedling growth of dominant species in the Atlantic East Pacific. No study evaluated the influence of leaf litter decomposition in the settlement success of species from later successional stages in mangroves in the Indo West Pacific.

Here, we evaluated the influence of two pioneer species leaf litter decomposition on (i) the settlement success of species following them or not in the natural succession to test if pioneer species facilitate their following species specifically and (ii) on soil chemical composition. For this purpose, we set-up an in situ experiment in mesocosms that followed survival and growth of three target species seedlings of the RRD (K. obovata, A. corniculatum, and R. stylosa) planted in soils derived from monospecific communities of dominant A. marina or S. caseolaris pioneer species, with or without leaf litter addition. We analyzed the chemical composition of tested soils, namely their nutrient contents for all tested interactions. For the most marked target response, we compared the global metabolic fingerprints of soils with or without litter by an untargeted metabolomic approach using liquid chromatography coupled with mass spectrometry, to highlight potential allelochemicals that could explain the effect. We hypothesize a species-specific response where pioneer species litter will (i) specifically facilitate the settlement of species following them in the mixed stage through the release of allelochemicals in soils during decomposition, and therefore (ii) induce a shift in soil chemical fingerprints.

### 2 | Material and Methods

### 2.1 | Study Site and Mangrove Species

To evaluate the effect of the pioneer species leaf litter decomposition (*Avicennia marina* and *Sonneratia caseolaris*) on the following species in the succession (i.e., the target species, *Kandelia obovata, Aegiceras corniculatum*, and *Rhizophora stylosa*), we performed an in situ mesocosm experiment at the

Mangrove Ecosystem Research Station (MERS, Appendix S1) in a mangrove plantation in the Giao Lac Commune, Giao Thuy District, Nam Dinh Province (20°14'27.1"N 106°30'51.6" E). The major soil type in this area is fluvisol (Nguyen Van et al. 2016). With an average humidity of 84.5% throughout the year, the region has a tropical wet climate with two monsoon seasons. The rainy season between June and October receives 85% of the annual precipitation (1660mm in 2006) with July being the rainiest month (Minh et al. 2014). The mean seasonal temperature during summertime (May to September) varies between 27°C and 29°C, the highest temperature recorded exceeds 40°C. The Ba Lat estuary is subject to a diurnal, neap tide-spring tide cycle of 14 days, with an average tidal amplitude of 1.92 m during the 1972-1990 period. The tidal waves travel from South to North along the shorelines, and salinity intrusion can be observed up to 20 km inland from the Ba Lat mouth (Minh et al. 2014). However, during the flood season of the Red River (June-October), due to the high fresh-water discharge, the tidal variation and its influences are restricted.

# 2.2 | Soil and Plant Collection for Mesocosms Preparation

Soils from *A. marina* and *S. caseolaris* (40 cm depth), and their green leaves (5 individuals) were collected from monospecific mangroves stands in the Giao Lac commune (Appendix S1) 1 day before the experiment. Initial soil characteristics are shown in Table 1. After soil homogenization, macroscopic plant parts were manually removed to focus on the effect of leaf litter addition. Homogenized initial soil subsamples, as well as mixed leaves subsamples were kept at  $-18^{\circ}$ C for nutrient content (soils only) and metabolomic analysis.

All the target seedlings were collected from multiple parent trees in the Giao Lac site and kept in separated soil bags in nursery outdoor (MERS) for 6 months before the beginning of the experiment. Healthy seedlings (approximately 5 cm in height and 2 leaves for *A. corniculatum* and 25 cm in height and usually 4–6 leaves for *K. obovata* and *R. stylosa*) were selected for this study and were kept under the same conditions. All mangrove species were identified according to the plant morphology description (Spalding 2011; Tomlinson 2016).

## 2.3 | Experimental Setup

A graphical representation of the experimental design is shown in Figure 1. In January 2020, 1 day after soils and leaves collection, 60 plastic pots  $(40 \text{ cm} \times 40 \text{ cm} \times 45 \text{ cm})$  were filled with 17 kg of soil for mesocosm settlement. To study the effect of pioneer species litter on growth and survival of nursed target species, 30 mesocosms were amended with 170g of leaves (1% of total FW) of one of the two pioneer species that were mixed with the soil. Thirty mesocosms for each pioneer species did not receive leaves and thus constitute control treatment (no litter). This experimental design leads to the following four treatments: A. marina soil with no litter (A-NL), A. marina soil with A. marina litter (A-L), S. caseolaris soil with no litter (S-NL), S. caseolaris soil with S. caseolaris litter (S-L). For each treatment, the five replicate mesocosms were planted with either four seedling of K. obovata and R. stylosa or five seedlings for A. corniculatum. In the field, the five mesocosms of each treatment were disposed randomly in five independent blocks, leading to a total of sixty mesocosms (two sources species (A. marina and S. caseolaris)×2 modalities (with or without leaf litter addition)×3 target species (A. corniculatum, K. obovata, and R. stylosa)). Each block was protected by nets set up underneath and around at 1.5m high to protect seedlings from predators. Mesocosms were submerged once a day by tide during our assay.

### 2.4 | Plant Growth and Survival

Regular follow-up measurements were carried monthly from the beginning  $(t_0)$  of the experiment 20th of January 2020, until the end of the experiment on the 10th of August 2020  $(t_f)$  where the height and the number of leaves were recorded on each of the seedlings of each of the mesocosms in all five plots (six measurements in total). For survival rates, the numbers of living and dead seedlings were recorded seven times in total.

The seedlings survival rates were determined in each of the mesocosms by considering the number of seedlings in the current and initial time steps:

Survival rate at 
$$t_i$$
 (%) = 100 ×  $\left(\frac{\text{Nb of seedlings at } t_i}{\text{Initial Nb of seedlings at } t_0}\right)$ 

**TABLE 1** | Pioneer species initial soil characteristics  $(t_0)$ . Mean  $\pm$  SE.

Pioneer species	Avicennia marina	Sonneratia caseolaris		
Location	20°13'17.0"N 106°29'40.7"E	20°14'34.8"N 106°34'15.7"E		
TOC (%)	$0.39 \pm 0.04$	$0.84 \pm 0.05$		
N (%)	$0.08 \pm 0.00$	$0.11\pm0.01$		
CN ratio	$5.19 \pm 0.09$	$8.05 \pm 0.19$		
Available N (mg/100g)	$1.96 \pm 0.09$	$2.91\pm0.19$		
Available $P_2O_5$ (mg/100g)	$0.46 \pm 1.15$	$0.51 \pm 1.21$		
Cl (%)	$0.46 \pm 0.01$	$0.51 \pm 0.01$		



**FIGURE 1** | Schematic representation of the experimental design showing the two main steps: (i) assessing three mangrove species growth and survival responses to litter addition in soils collected in *S. caseolaris* or *A. marina* communities (left) and (ii) evaluating the effect of leaf litter addition on the soil chemical fingerprints, and individual metabolite variation after 203 days of experiment, with a focus on *A. marina* soils planted with *K. obovata* (right). At  $t_0$ , soils were collected from two pioneer species monospecific communities and transferred in mesocosms with (L) or without (NL) leaf litter from respective pioneer species and planted with each of the three target species.

with Nb of seedlings at  $t_i$  the number of seedlings alive at  $t_i$ .

### 2.5 | Chemical Analysis

#### 2.5.1 | Soil Nutrients

Nutrient content analysis was carried out on initial soil subsamples  $(t_0)$  (see section Soil and plant collection for mesocosms preparation) and soils at the end of the experiment ( $t_{\rm f}$  = 203 days). At  $t_{p}$ , soils from each mesocosms (five replicates per treatment) were collected and stored at -18°C prior to their analysis. The National Institute of Agricultural Planning and Projection (NIAPP, Hanoï) performed the quantification of the following parameters: chloride concentrations were determined with Mohr's method using silver nitrate titration with chromate indicator (Rajković, Sredović, and Miloradović 2010); total organic carbon (TOC) was determined using the Walkley Black Method (Meersmans, van Wesemael, and van Molle 2009); total nitrogen content (N) was determined with a modified Kjeldahl method using TiO<sub>2</sub> catalyst (Singh, Singh, and Sharma 2021); available nitrogen was determined with a nitrogen distillation and titration method using sodium hydroxide solution (Singh, Singh, and Sharma 2021); available phosphorus was determined using molybdenum blue spectrophotometry, with soil samples extracted by Olsen method (Lin et al. 2021).

# 2.5.2 | Metabolomic Analysis, Data Pre-Processing and Annotation of Metabolites

For chemical profile determination, we chose to focus on *A. marina* soils planted with *K. obovata*, as this target species showed the most marked decrease in survival in response to litter addition at the end of the experiment. *A. marina* initial leaves added to the mesocosms at the beginning of the assay (A-leaves-t0)

were analyzed to serve as analytical references for the dereplication of litter decomposition by-products, which could be allelochemicals. Our objectives were (i) to evaluate the differences between *A. marina* initial soils (A-t0) and soils planted with *K. obovata* with (AL-tf) and without (ANL-tf) *A. marina* leaf litter 203 days after their addition and (ii) to assess the similarities between AL-tf and A-leaves-t0.

**2.5.2.1** | **Initial Leaves Extraction.** After freeze-drying (Lyovac GT2) and powdering (MM400, Retsch GmbH.D, Hann Germany), a dry mass of 10 mg of each sample was weighed in 1.5 mL Eppendorf tubes and suspended in 400  $\mu$ L of methanol (LC–MS grade, Carlo Erba, France) before ultra-sonication at room temperature for 5 min for metabolite extraction. Subsequent sample preparation is shown in Appendix S2.

**2.5.2.2** | **Soil** Extraction. For soil extraction, the microwave-assisted extraction (MAE, CEM MARS 230/60 Microwave Accelerated Reaction System, CEM Corporation, USA) modified from Montesdeoca-Esponda et al. (2013) was used. After freeze-drying (Lyovac GT2) and powdering (MM400, Retsch GmbH.D, Hann Germany), 5g of dry soil samples were transferred to microwave tubes and suspended in 10 mL of MeOH/Water (70:30 v:v). Subsequent sample preparation is shown in Appendix S2.

**2.5.2.3** | **UHPLC-ESI-qToF Analysis.** The samples were analyzed by Ultra-High Performance Liquid Chromatography instrument, including UHPLC system (Dionex Ultimate 3000, Thermo Electron, France) coupled to a qToF mass spectrometer (Impact II, Bruker Daltonics, Germany). Separation was conducted on a Kinetex XB-C18 column (150 mm  $\times$  2.1 mm, 1.7  $\mu$ m, Phenomenex, France). More details are shown in Appendix S2.

The raw data obtained from UHPLC-MS/MS were calibrated and converted into Computable Document Format (.cdf) files

using Compass DataAnalysis (Bruker Daltonics version 5.0). These files were then preprocessed (via chromatographic peak detection, integration, alignment, and grouping) using the open-source XCMS package (version 3.6.2 (Smith et al. 2006)) in R 3.6.0 version. To ensure data quality and remove redundant signals, three successive filtering steps were applied as described in Favre et al. (2017). After statistical analysis, the biomarkers specific to *tf*-Am-L soils were selected for putative annotation. The process of metabolite annotations were based on a dereplication strategy as described in Dhaou et al. (2022) with an additional public database interrogation using MetFrag (Ruttkies et al. 2016).

#### 2.6 | Data Analysis

### 2.6.1 | Growth and Survival Responses of Target Species

Growth and survival responses according to leaf litter addition (NL, L), planted target species (*A. corniculatum, K. obovata*, and *R. stylosa*) and their interaction were tested for each pioneer species (*A. marina* and *S. caseolaris*). To take into account repeated measures of seedlings over the days of measurements and over experimental plots (time and spatial dependencies), we used linear mixed-effect models (LMMs) and generalized linear mixed-effect models (GLMMs) (R package *lme4*) that include both fixed (litter addition and target species) and random factors (time and plots). GLMMs with binomial distribution were used for survival rates (proportion data), LMMs for height (continuous data) and GLMMs with Poisson distribution for the number of leaves (count data). When a significant effect was observed, we used a post hoc Tukey test for pairwise comparison.

#### 2.6.2 | Soil Nutrient Contents

The soil nutrient contents according to litter addition, planted target species and their interaction were also tested for each pioneer species. For this analysis, time and spatial dependencies were not considered since soil samples were collected only at *tf* with one sample per experimental plot for each treatment. We therefore used two-way ANOVAs followed by a post hoc Tukey test for pairwise comparison.

When necessary, data were transformed using the *bestNormalize* package to meet the assumptions of normality and homoscedasticity of variances (arcsinh transformation for *A. marina* soils TOC and available N). All the tests were carried out using the R software (version 3.6.0, The R Foundation for Statistical Computing, Vienna, Austria) and plots were created using *gplot2* package. The map of the experimental site was created using the Free and Open Source QGIS.

#### 2.6.3 | Metabolomics Data

For metabolomic analysis, figures, multivariate (Principal Component Analysis (PCA)), univariate analyses (ANOVA) and boxplots plots were realized using MetaboAnalyst 6.0 online resource (https://www.metaboanalyst.ca/, accessed in April

2024). PERMANOVA permutation test (*RVAideMemoire* package on log-transformed data) was performed (10,000 permutations) to test global differences between leaf and soil extract groups and pairwise differences using R software.

#### 3 | Results

# 3.1 | Survival and Growth Response to Litter Decomposition

The three planted species responded differently to pioneer species leaf litter decomposition. *Aegiceras corniculatum* did not show any significant response to litter decomposition for all variables measured here, whereas *Kandelia obovata* and *Rhizophora stylosa*'s response depended on parameters and pioneer species (Figure 2, Table 2).

Considering seedling mortality, Sonneratia caseolaris litter addition and planted target species did not affect seedling survival (Table 2), while a significant interaction of these two factors was observed for Avicennia marina (Table 2), where K. obovata and R. stylosa survival significantly decreased by 46% and 33% in L soils compared with their NL control respectively, after 203 days of decomposition (Figure 2). K. obovata showed the most rapid and marked mortality in A. marina soils with litter (L) where approximately 31% more seedling died after 81 days of decomposition compared to soils without litter (no litter: NL), when this mortality level was reached by R. stylosa seedlings in L soils compared to NL after 203 days of decomposition (Figure 2). Considering seedling growth, litter addition, and target species interacted in their effects for height response for both pioneer species (Table 2), while this interaction was found significant for the number of leaves response for S. caseolaris only (Table 2). R. stylosa height significantly decreased in A. marina L soils compared to NL (-8.3% at the end of the experiment, Figure 2). However, K. obovata height was found substantially higher in A. marina and S. caseolaris L soils compared to NL with a 36% and 24% increase after 203 days of decomposition respectively, while its number of leaves only increased in S. caseolaris L soils compared to NL (+50% at the end of the experiment, Figure 2).

# 3.2 | Soil Nutrient Content Response to Litter Decomposition

At the end of the experiment (203 days), soil nutrient compositions were not equally affected by litter addition or target species, depending on the considered pioneer species (Table 3). Globally, no effect of these two factors or their interaction was observed in *S. caseolaris* soils for all assessed parameters: TOC, N, available N, available P contents, or CN ratios (Appendix S3). For *A. marina* however, no interaction effects were found for all parameters, but we observed significant effects of litter addition on TOC (F=7.71, p <0.05), N (F=8.09, p <0.01), available N (F=6.97, p <0.05) contents and CN ratios (F=5.16, p <0.05), while no effect of target species was highlighted (F=0.29, p >0.05 for TOC; F=0.22, p >0.05 for N, F=0.07, p >0.05 for available N, and F=0.45, p >0.05 for CN). For all target species combined, litter decomposition



**FIGURE 2** | Survival and growth response of planted target species (*A. corniculatum*, *K. obovata* and *R. stylosa*) to leaf litter addition from two pioneer species communities (*S. caseolaris*, left and *A. marina*, right; plain line in dark color: Litter addition (L); dashed line in light: No litter (NL)) during 203 days of decomposition. Mean  $\pm$  SE,  $\nearrow$ L and  $\searrow$ L indicate a significant increase or decrease when litter is added for each target species (post hoc pairwise comparison *p* < 0.05).

**TABLE 2** Image: Results of linear (LMMs) and generalized linear mixed models (GLMMs) testing the effects of litter addition, planted target species and their interaction on seedling survival, height and number of leaves for each tested pioneer species during the experiment.

	Survival		Height		Number of leaves				
	$\chi^2$	df	р	$\chi^2$	df	р	$\chi^2$	df	р
Avicennia marina									
Litter addition	0.045	1		0.3899	1		0.8748	1	
Planted species	8.469	2	*	324.0785	2	***	10.8047	2	**
L×T	13.2113	2	**	48.4623	2	***	4.6038	2	
Sonneratia caseolari	S								
Litter addition	0.1301	1		2.356	1		4.4684	1	*
Planted species	0.3367	2		406.5496	2	***	7.4497	2	*
L×T	3.4478	2		9.6864	2	**	22.9507	2	***

*Note:* Significant *p* values are shaded in gray.

substantially increased *A. marina* L soil contents in TOC (1.8-fold), N (1.5-fold), available N (1.8-fold), and CN ratios (1.13-fold) compared to NL (Table 3). Only available P contents were not affected by the tested factors or their interaction in the case of *A. marina* soils and remained similar between NL and L soils (Table 3).

# 3.3 | Focus on *A. marina* Soils Planted With *K. obovata* for Chemical Investigation

To further investigate soil chemical compositions and highlight potential allelochemicals that could be released during litter decomposition through an untargeted metabolomic approach, we chose to focus on the most marked survival and growth response to litter addition of the assessed target species. Among the species that were significantly affected by litter addition, *K. obovata* seedlings showed the most rapid and important decrease in survival rates and the most important increase in height in *A. marina* L soils compared to NL (Figure 2). For these reasons, these soils were selected for analysis at the end of the experiment (AL-tf and ANL-tf for *A. marina* soils planted with *K. obovata*, with and without litter addition respectively, after 203 days of decomposition), along with *A. marina* initial soil (A-t0) and leaves that were added to the mesocosms at the beginning of the assay (A-leaves-t0) (Figure 1). Our objectives were (i) to evaluate the differences between A-t0, AL-tf, and ANL-tf extracts and (ii) to assess the similarities between AL-tf and A-leaves-t0.

# 3.4 | Soil Chemical Fingerprints

After soil extract analysis by an untargeted UHPLC-MSbased metabolomics approach, data preprocessing and filtering allowed to obtain 1232 m/z features dataset (766 features detected in negative mode combined to 466 features detected in positive mode). The resulting PCA score plot (Figure 3a) revealed significant differences between tested soil chemical fingerprints (PERMANOVA, p < 0.01). *A. marina* initial soils (A-t0) separated from A-NL-tf and A-L-tf soils fingerprints on the first component (46.9% of total variance), while the latter two clusters overlapped on components 1 and 2 (13.7% of total variance), indicating fingerprint similarity.

**TABLE 3** | Nutrient contents of soils collected from assessed mesocosms at the end of the experiment ( $t_r$ =203 days) for each pioneer species.

Pioneer species	Avicenni	a marina	Sonneratia caseolaris			
Litter addition	NL	L	NL	L		
TOC (%)	$0.40 \pm 0.04$ a	$0.72 \pm 0.12$ b	$0.68\pm0.08$	$0.8\pm0.11$		
N (%)	$0.06 \pm 0.00$ a	$0.09 \pm 0.01 \text{ b}$	$0.08\pm0.01$	$0.09\pm0.01~\mathrm{b}$		
Available N (mg/100g)	$1.12 \pm 0.08$ a	$2.02 \pm 0.33$ b	$2.05 \pm 0.27$	$2.12 \pm 0.26$		
Available $P_2O_5$ (mg/100g)	$67.72 \pm 1.04$	$66.43 \pm 0.69$	$71.41 \pm 0.76$	$69.88 \pm 0.55$		
CN ratio	$6.69 \pm 0.2$ a	$7.56 \pm 0.4$ b	$7.98 \pm 0.24$	$7.68 \pm 0.25$		

*Note:* NL and L indicate the absence or presence of leaf litter (added at  $t_0$ ), respectively. Mean ±SE of data pooled for all planted target species, different letters indicate statistical difference between NL and L with a < b for each tested pioneer species, significant differences are shaded in gray (post hoc pairwise comparison p < 0.05).



**FIGURE 3** | (a) Principal component analysis showing chemical fingerprints of extracts from *A. marina* initial soils (A-t0) and soils collected from mesocosms planted with *K. obovata* with (A-L-tf) or without (A-NL-tf) *A. marina* litter addition, 203 days after decomposition (*tf*); (b) Soil type variation in relative concentrations of 12 selected and putatively annotated biomarkers specific to A-L-tf condition (post hoc pairwise comparison p < 0.05). The dataset obtained by UHPLC-HRMS (1232 detected features in total) was log-transformed and mean-centered.

## 3.5 | *A. marina* Litter Decomposition Biomarkers Selection and Annotation

Univariate analysis of each detected m/z feature allowed to highlight 13 biomarkers significantly more abundant in A-L-tf than A-t0 and A-NL-tf (ANOVA, p < 0.01). We focused on these 13 features (all detected in negative mode) for putative annotations with MS-based dereplication. When observed, insource fragments of the same molecule were combined to avoid chemical redundancy and keep metabolites with their monoisotopic molecular ion [M-H]<sup>-</sup>, which resulted in 12 biomarkers (Figure 3b). Except for the feature NM147T485, all biomarkers molecular formulas could be deduced (Appendix S4). Isotopic patterns and the presence of a MS/MS fragment at m/z 79.956  $[O_3S]^{\circ-}$  indicated the presence of one or more sulfur atoms in all biomarkers. The only feature that could be putatively identified is NM211T380, suggested as 2-(2-thioxopropanoyloxy) ethanesulfonic acid, with seven MS/MS fragments out of eight matching with MetFrag database (final match score = 1.0), its MS/MS spectrum and fragmentation pattern is shown in Appendix S5. When available, MS/MS spectra for each biomarker are shown in Appendixes S5–S12.

### 4 | Discussion

In this study, we investigated whether leaf litter decomposition of pioneer mangrove species in the Red River Delta could (i) influence species recruitment during succession and (ii) modify the chemical composition of soils with decomposition byproducts potentially acting as allelochemicals.

Our mesocosm experiment revealed contrasted effects of pioneer mangrove litter decomposition on target seedling growth and survival, with a species-specific response. Indeed, in the case of Avicennia marina litter, Rhizophora stylosa and Kandelia obovata were the only species affected by its decomposition, with heights either slightly inhibited or substantially stimulated respectively, and most of all, an important mortality for both species after 203 days of decomposition. In the case of Sonneratia caseolaris, the only species affected by its litter decomposition was K. obovata, with a significant increase in height and number of leaves at the end of our experiment, but overall, seedling survival rates remained unchanged. This in accordance with the findings of Sarker et al. (2020), who observed facilitating and inhibitory effects of litter addition on seedling growth in relation to target species identity and litter type in tropical rainforests. Other studies showed litter beneficial effects linked to nutrient enrichment (Ganade and Brown 2002; Brearley, Press, and Scholes 2003) or inhibitory effects linked to allelopathic processes (Fernandez et al. 2006; Meiners 2014; de Heras et al. 2020; Bonanomi et al. 2021).

The more rapid growth of *K. obovata* seedlings in the presence of *A. marina* litter may have occurred in response to its important decrease in survival throughout the experiment, as previous studies showed that low density populations can support tree settlement success with less resource competition and concentration for specific pathogen (Chesson 2000; Liu et al. 2015). Moreover, the two-fold increase in available nitrogen in *A. marina* soils with litter addition compared to no litter addition at the end of our experiment may have favored K. obovata growth, as several field studies showed that nutrient enrichment, and especially nitrogen, favors mangrove tree growth (Feller et al. 2003; Lovelock et al. 2004; Krauss et al. 2008; Reef, Feller, and Lovelock 2010). In the presence of S. caseolaris litter however, seedling height and number of leaves were also found higher than seedlings grown in S. caseolaris soils without litter despite the absence of soil nutrient enrichment. This is in accordance with Chapman and Feller (2011) findings, who also observed a more rapid mangrove seedling growth in the presence of conspecific litter, irrespective of litter chemical quality and nitrogen release in a mesocosm study conducted in Florida. Still, these studies were conducted in the Atlantic East Pacific regions while knowledge remains very scarce about the specific response of species from the Indo West Pacific regions (Krauss et al. 2008), it is therefore difficult to explain the species-specific growth responses observed in our study through this framework exclusively, which only partially validates our first hypothesis of a better growth through nutrient enrichment.

Within our findings, two other observations further suggest that other factors are involved. First, litter addition effect was not found to be dependent on target species for soil nutrient contents, as opposed to growth responses. Secondly, only *A. marina* soils with litter addition showed higher nutrient contents compared to soils without litter, with almost doubled amounts of total carbon and available nitrogen, and amounts of total nitrogen increased by half, while no significant enrichment was highlighted for *S. caseolaris* soils with litter addition compared to soils without litter. Yet, *K. obovata* grew better in soils amended with litter from both pioneer species in our assay. These results reinforced our second hypothesis of allelopathic processes potentially at play here, further supported by the substantially higher mortality of *K. obovata* and *R. stylosa* that could be due to phytotoxic by products from *A. marina* leaf litter decomposition.

A strong allelopathic potential of A. marina leaves was previously shown on agricultural species (Chen et al. 2009; Dhaou et al. 2022). To our knowledge, Chen et al. (2009) study is the only assessment of A. marina's allelopathic potential on mangrove seedlings. Contrary to our findings, they found that leachates from green leaves of A. marina significantly inhibited shoot length of K. obovata (previously identified as K. candel) in a mesocosm experiment. The variable chemical composition between A. marina green leaves and its decomposed litter (Zhou et al. 2010, 2023) may explain our observations. Moreover, although seedling mortality was not assessed, a significant reduction in root length was observed in their study. We may therefore suggest that belowground parts may have been negatively affected by the end of our experiment. Overall, by showing the allelopathic effect of A. marina green leaves macerates, Chen et al. (2009) findings suggest a possible phytotoxicity of its litter through the liberation of its decomposition by-products in the soil, to which K. obovata could be sensitive. This could have led to the important mortality found in our study for this target species.

For these reasons, we investigated the assessed soil chemical compositions, with a focus on the most affected target seedling, namely *K. obovata* and the most impacting pioneer species, namely *A. marina*. Using an untargeted metabolomic

approach, we compared the chemical composition of initial A. marina soils (A-t0) with soils from mesocosms planted with K. obovata where A. marina leaf litter was initially added (A-L-tf) or not (A-NL-tf), after 203 days of decomposition (i.e., end of our experiment). Contrary to our third hypothesis, leaf litter decomposition did not modify A-L-tf soils' global chemical fingerprints compared to A-NL-tf, and no proximity was observed with our initial leaves analytical reference. Previous studies showed contrasted average times of decomposition of A. marina leaf litter according to mangrove types and regions, time to lose half of the initial mass  $(t_{50})$  ranged from 96 to 78 days according to dry and wet season respectively in urban dwarf mangroves and 171-48 days in a rural well-developed mangrove in South Mozambique (Fernando and Bandeira 2009). In South Vietnam, t<sub>50</sub> ranged from 36.5 days in the dry season to 12.2 days in the wet season (Vinh et al. 2020), while in South China, t<sub>50</sub> was reached after 19 days of decomposition (Zhou et al. 2010). Mangrove litter decomposition is characterized by an initial leaching of soluble organic (which can occur rapidly for A. marina within the first 14 days (Zhou et al. 2010)) and inorganic compounds, followed by microorganism colonization, which initiates physical and biological transformation (Hossain and Hoque 2008). With a maximum  $t_{50}$  reaching 171 days for A. marina leaf litter decomposition according to previous assessments, A. marina litter probably already underwent substantial biological and physical transformations after 203 days in the present study, which could explain the absence of chemical fingerprints proximity between A. marina initial litter and A-L-tf soils. This was confirmed by the MS-based annotation of biomarkers specific to A-L-tf soils that did not highlight any tannin or phenolic compound.

Instead, A-L-tf soil biomarkers putative annotations revealed 12 sulfur-containing organic compounds that suggest possible by-products of litter decomposition through a sulfur (S) reducing process. Mangrove sediments are frequently anoxic and rich in sulfates mainly of seawater origin (Balk, Keuskamp, and Laanbroek 2016), with generally high salinity, which favors organic matter (OM) sulfurization that can contribute to up to 50% of carbon burial (Chambers, Osborne, and Reddy 2013). The redox gradients around O2-releasing mangrove roots can facilitate the formation of several OM-S complexes by reacting with free polysulfides in porewaters (Raven et al. 2019). Nevertheless, sulfated ellagic acid derivates were recently described for the first time in roots and leaves of the mangrove genus Lumnitzera spp. (Manurung et al. 2021) and Sonneratia apetala (Dhaou et al. 2022) respectively, suggesting that some annotated biomarkers could also have been sulfurized prior to their physicochemical or biological transformation in the sediments here.

The leaves of *A. marina* contain abundant and diverse specialized metabolites (Liebezeit 2012; Thatoi, Samantaray, and Das 2016), among them, phenolics were suggested to be responsible for allelopathic effect of this species (Chen and Peng 2008; Chen et al. 2009). No allelopathic assessment of its leaf litter was investigated to our knowledge. Lang et al. (2021) is the only investigation of litter allelopathy conducted on mangrove species, namely *K. obovata* on *Aegiceras corniculatum*. Their results showed the allelopathic potential of its leaf litter leachates, with condensed tannins as the main responsible compounds for *A. corniculatum* germination and growth reduction, while suggesting that

other phytotoxic by-products of decomposition could be at play. Similarly, in other plant models, de Heras et al. (2020) also observed the predominance of phytotoxic effects over positive effects when assessing leaf litter from exotic and native riverine trees on the germination and growth of herbaceous species. They characterized the chemical composition of tested litter samples over time of decomposition and found that among nine species, only Ailanthus altissima litter inhibitory effect on growth could be linked to higher phenolic acids and flavonoids contents, suggesting that other compound classes acted as allelochemicals in their study. Quan et al. (2022) assessed the allelopathic potential of aqueous extracts of Salix caprea litter and soil layers. After highlighting a significant germination inhibition of three target species, a total of 26 allelochemicals were identified, with mainly organic acids in the soil and humus layers, and phenolic acids in the litter layer. The organic acid annotated as 2-(2-thioxopropanoyloxy)ethanesulfonic acid may therefore be involved in the phytotoxicity observed in A-L-tf soils. The isolated compounds' effects need to be further assessed and other types of specialized metabolites may be involved in the present study. As previously mentioned, the main bioactive compounds may have already been degraded after 203 days of decomposition (i.e., the time after which soils amended with A. marina leaf litter and planted with K. obovata were analyzed, A-L-tf), considering that the maximum reduction of K. obovata survival rates was reached after 116 days, and the relatively fast decomposition rate of A. marina leaf litter (Fernando and Bandeira 2009; Vinh et al. 2020).

The allelopathic effect suggested here may have occurred directly or indirectly (i) through chemical modifications of litter by microorganisms during their decay, allowing a phytotoxic effect to occur in the soil (Jilani et al. 2008), or (ii) through the modification of microbial communities exposed to allelochemicals or by-products that caused unfavorable conditions for plant growth or survival in the soil, by limiting nutrient absorption for example (Inderjit 2003; Scavo, Abbate, and Mauromicale 2019). Moreover, allelochemicals can influence not only their plant neighbors but also fungi and bacteria (Schandry and Becker 2020), which can ultimately affect competition for resources between these organisms (mainly saprophytic microbes) and plants in the context of litter decomposition. In the case of mangroves, Lang et al. (2024) showed that leaf litter of K. obovata, A. marina and S. apetala boosts sediment microbial activities and diversities. Their results revealed that the release of mangrove tannins during leaf litter decomposition is one of the key factors driving N cycling, and microbial activities and diversities in mangrove wetlands. Overall, other processes may be at play in combination with resources competition and allelopathy in the present work, this complexity makes the study of litter-plant interactions quite challenging in situ.

Despite the difficulty of describing the mechanisms behind the contrasted seedling responses here (i.e., distinguishing allelopathic effects from competition for nutrients (Weidenhamer, Hartnett, and Romeo 1989)), our results can be interpreted in the framework of mangrove species composition and successional dynamics. Seedling establishment success is critical for mangrove regeneration and community replacement, as vegetative reproduction is rarely employed by these species (Tomlinson 2016). During this process, stem elongation is one of the most important parameter, allowing better access to light and avoidance of tidal immersion. Here, *K. obovata* is the only

target species that showed more rapid growth in the presence of S. caseolaris and A. marina litter. This suggests a facilitation of K. obovata settlement by both pioneer species, which could participate to its dominant status in the studied area. Still, survival of K. obovata and R. stylosa seedlings was substantially reduced where A. marina litter decomposed. In the RRD, these two target species specifically follow A. marina in the mixed stage, where they gradually surpass the latter to become dominant, ultimately replacing A. marina in the dominant stage by forming dense canopies (Hong and San 1993). We hypothesize that the release of decomposition by-products of A. marina litter, specifically phytotoxic to K. obovata and R. stylosa, may allow the delay of its replacement during succession, as suggested in previous studies in other plant communities (Fernandez et al. 2006; Meiners 2014). On a longer term, the reduction of seedling survival may have indirect beneficial consequences for species settlement by reducing competition for resources (Liu et al. 2015), this could ultimately facilitate the progressive replacement of A. marina by K. obovata and R. stylosa. Overall, A. corniculatum did not show any response to the decomposition of both pioneers. Its position in successional dynamics of the RRD might be a reason for this specificity. Indeed, this shrub is the only assessed species that co-exists with both pioneers in the mixed stage as understory, without replacing them in the dominant stage (Hong and San 1993).

#### 5 | Conclusions

The present study shows a species-specific response of mangrove seedlings settlement success to pioneer species leaf litter decomposition, with contrasted beneficial effects on growth and detrimental effects on seedling survival. Kandelia obovata is the only species that grew faster with both types of litter, which could participate to its dominant status in the studied area. Meanwhile, contrary to our first hypothesis of pioneer species specifically facilitating the settlement of the species following them in succession, we found a substantial decrease in K. obovata and Rhizophora stylosa survival rates in the presence of Avicennia marina, indicating a potential strategy to delay the pioneer's replacement through the release of phytotoxic by-products of litter decomposition which could act as allelochemicals. Nevertheless, we may hypothesize an indirect beneficial effect for the settlement of these species in the long run through the reduction of intra-specific competition for resources. Finally, we focused on the most marked phytotoxic effect observed in our study to compare soil chemical fingerprints and highlight potential allelochemicals using an untargeted metabolomic approach, namely in A. marina soils planted with K. obovata amended or not with leaf litter. Contrary to our second hypothesis, we did not observe a shift in global chemical fingerprints in soils where A. marina litter decomposed for 203 days, yet our analysis revealed 12 sulfur-containing specific biomarkers, among which the organic acid 2-(2-thioxopropanoyloxy)ethanesulfonic acid. These annotated compounds are probable by-products of litter decomposition through a sulfur reducing process, suggesting that mangrove litter allelochemicals may exhibit previously undescribed original structures. Further studies are necessary to unravel the identity and role of mangrove allelochemicals. Plant succession is an important process during ecosystem development, especially in forests that are recovering from disturbances such as Vietnamese mangroves. Further studies are needed to better understand the successional process in mangroves and the importance of allelopathy to support restoration projects.

#### **Author Contributions**

D.D., C.F., T.D.V., and A.B.-M. conceived the research idea; D.D., A.B.-M., C.F., T.D.V., D.S., H.K.T., S.G., and C.L. collected data; D.D., C.F., D.S., S.G., H.K.T., and R.G. performed statistical analyses; D.D., D.S., H.K.T., and A.B.-M. wrote the paper; all authors discussed the results and commented on the manuscript; A.B.-M., and D.D. acquired the fundings.

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#### **Conflicts of Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### Data Availability Statement

Data are available from the Dryad Digital Repository: https://doi.org/ 10.5061/dryad.4qrfj6qjw.

#### References

An, M., J. E. Pratley, and T. Haig. 2000. "Phytotoxicity of *Vulpia* Residues: IV. Dynamics of Allelochemicals During Decomposition of Vulpia Residues and Their Corresponding Phytotoxicity." *Journal of Chemical Ecology* 26: 2603–2617. https://doi.org/10.1023/A:10055 45014843.

Balk, M., J. A. Keuskamp, and H. J. Laanbroek. 2016. "Potential for Sulfate Reduction in Mangrove Forest Soils: Comparison Between Two Dominant Species of the Americas." *Frontiers in Microbiology* 7: 1855. https://doi.org/10.3389/fmicb.2016.01855.

Bonanomi, G., M. G. Sicurezza, S. Caporaso, A. Esposito, and S. Mazzoleni. 2006. "Phytotoxicity Dynamics of Decaying Plant Materials." *New Phytologist* 169: 571–578. https://doi.org/10.1111/j. 1469-8137.2005.01611.x.

Bonanomi, G., M. Zotti, M. Idbella, S. Mazzoleni, and A. M. Abd-ElGawad. 2021. "Microbiota Modulation of Allelopathy Depends on Litter Chemistry: Mitigation or Exacerbation?" *Science of the Total Environment* 776: 145942. https://doi.org/10.1016/j.scitotenv.2021.145942.

Brearley, F. Q., M. C. Press, and J. D. Scholes. 2003. "Nutrients Obtained From Leaf Litter Can Improve the Growth of *Dipterocarp* Seedlings." *New Phytologist* 160: 101–110. https://doi.org/10.1046/j.1469-8137.2003. 00851.x.

Callaway, R. M., and L. R. Walker. 1997. "Competition and Facilitation: A Synthetic Approach to Interactions in Plant Communities." *Ecology* 78: 21.

Chambers, L. G., T. Z. Osborne, and K. R. Reddy. 2013. "Effect of Salinity-Altering Pulsing Events on Soil Organic Carbon Loss Along an Intertidal Wetland Gradient: A Laboratory Experiment." *Biogeochemistry* 115: 363–383. https://doi.org/10.1007/s10533-013-9841-5. Chapman, S. K., and I. C. Feller. 2011. "Away-Field Advantage: Mangrove Seedlings Grow Best in Litter From Other Mangrove Species." *Oikos* 120: 1880–1888. https://doi.org/10.1111/j.1600-0706.2011.19381.x.

Chen, L., and S. Peng. 2008. "Allelopathic Potential of Mangrove Plants (Avicennia marina, Aegiceras corniculata and Bruguiera gymnorrhiza)." Allelopathy Journal 22: 213–220.

Chen, L., S. L. Peng, B.-M. Chen, et al. 2009. "Effects of Aqueous Extracts of 5 Mangrove Spp. on Cabbage Germination and Hypocotyl Growth of *Kandelia candel*." *Allelopathy Journal* 23: 469–476.

Chesson, P. 2000. "Mechanisms of Maintenance of Species Diversity." *Annual Review of Ecology, Evolution, and Systematics* 31: 343–366.

Chomel, M., M. Guittonny-Larchevêque, C. Fernandez, et al. 2016. "Plant Secondary Metabolites: A Key Driver of Litter Decomposition and Soil Nutrient Cycling." *Journal of Ecology* 104: 1527–1541. https:// doi.org/10.1111/1365-2745.12644.

de las Heras, P., S. Medina-Villar, M. E. Pérez-Corona, and B. R. Vázquez-de-Aldana. 2020. "Leaf Litter Age Regulates the Effect of Native and Exotic Tree Species on Understory Herbaceous Vegetation of Riparian Forests." *Basic and Applied Ecology* 48: 11–25. https://doi.org/10.1016/j.baae.2020.07.005.

Delory, B. M., H. Schempp, S. M. Spachmann, et al. 2021. "Soil Chemical Legacies Trigger Species-Specific and Context-Dependent Root Responses in Later Arriving Plants." *Plant, Cell & Environment* 44: 1215–1230. https://doi.org/10.1111/pce.13999.

Dhaou, D., V. Baldy, D. van Tan, et al. 2022. "Allelopathic Potential of Mangroves From the Red River Estuary Against the Rice Weed *Echinochloa crus-galli* and Variation in Their Leaf Metabolome." *Plants* 11: 2464. https://doi.org/10.3390/plants11192464.

Duke, N., M. Ball, and J. Ellison. 1998. "Factors Influencing Biodiversity and Distributional Gradients in Mangroves." *Global Ecology and Biogeography Letters* 7: 27–47. https://doi.org/10.1111/j.1466-8238.1998. 00269.x.

Facelli, J. M., and S. T. A. Pickett. 1991. "Plant Litter: Its Dynamics and Effects on Plant Community Structure." *Botanical Review* 57: 1–32.

Favre, L., A. Ortalo-Magné, S. Greff, et al. 2017. "Discrimination of Four Marine Biofilm-Forming Bacteria by LC–MS Metabolomics and Influence of Culture Parameters." *Journal of Proteome Research* 16: 1962–1975. https://doi.org/10.1021/acs.jproteome.6b01027.

Feller, I. C., C. E. Lovelock, U. Berger, K. L. McKee, S. B. Joye, and M. C. Ball. 2009. "Biocomplexity in Mangrove Ecosystems." *Annual Review of Marine Science* 2: 395–417. https://doi.org/10.1146/annurev.marine. 010908.163809.

Feller, I. C., D. F. Whigham, K. L. McKee, and C. E. Lovelock. 2003. "Nitrogen Limitation of Growth and Nutrient Dynamics in a Disturbed Mangrove Forest, Indian River Lagoon, Florida." *Oecologia* 134: 405– 414. https://doi.org/10.1007/s00442-002-1117-z.

Fernandez, C., B. Lelong, B. Vila, et al. 2006. "Potential Allelopathic Effect of *Pinus halepensis* in the Secondary Succession: An Experimental Approach." *Chemoecology* 16: 97–105. https://doi.org/10.1007/s0004 9-006-0334-z.

Fernandez, C., M. Santonja, R. Gros, et al. 2013. "Allelochemicals of *Pinus halepensis* as Drivers of Biodiversity in Mediterranean Open Mosaic Habitats During the Colonization Stage of Secondary Succession." *Journal of Chemical Ecology* 39: 298–311. https://doi.org/10.1007/s10886-013-0239-6.

Fernando, S. M. C., and S. O. Bandeira. 2009. "Litter Fall and Decomposition of Mangrove Species *Avicennia marina* and *Rhizophora mucronata* in Maputo Bay, Mozambique." *Western Indian Ocean Journal of Marine Science* 8. https://doi.org/10.4314/wiojms.v8i2.56975.

Gallet, C., and F. Pellissier. 2002. "Allelopathy in Forest." *Ecosystems*. https://doi.org/10.4267/2042/4944.

Ganade, G., and V. K. Brown. 2002. "Succession in Old Pastures of Central Amazonia: Role of Soil Fertility and Plant Litter." *Ecology* 83: 743–754. https://doi.org/10.1890/0012-9658(2002)083[0743:SIOPOC] 2.0.CO;2.

Hashoum, H., M. Santonja, T. Gauquelin, et al. 2017. "Biotic Interactions in a Mediterranean Oak Forest: Role of Allelopathy Along Phenological Development of Woody Species." *European Journal of Forest Research* 136: 699–710. https://doi.org/10.1007/s10342-017-1066-z.

Hong, P. N., and H. T. San. 1993. "Mangroves of Vietnam." IUCN Southeast Asia Regional Office, Bangkok: IUCN, Wetlands Conservation Programme.

Hossain, M., and A. K. F. Hoque. 2008. "Litter Production and Decomposition in Mangroves—A Review." *Indian Journal of Forestry* 31: 227–238. https://doi.org/10.54207/bsmps1000-2008-8TS8TD.

Inderjit, C. R. M. 2003. "Experimental Designs for the Study of Allelopathy." *Plant and Soil* 256: 1–11. https://doi.org/10.1023/A:10262 42418333.

Janišová, M., R. Hrivnák, D. Gömöry, et al. 2007. "Changes in Understorey Vegetation After Norway Spruce Colonization of an Abandoned Grassland." *Annales Botanici Fennici* 44: 256–266.

Jilani, G., S. Mahmood, A. N. Chaudhry, I. Hassan, and M. Akram. 2008. "Allelochemicals: Sources, Toxicity and Microbial Transformation in Soil—A Review." *Annales de Microbiologie* 58: 351–357. https://doi.org/ 10.1007/BF03175528.

Kaligarič, M., M. H. Meister, S. Škornik, N. Šajna, B. Kramberger, and H. R. Bolhar-Nordenkampf. 2011. "Grassland Succession Is Mediated by Umbelliferous Colonizers Showing Allelopathic Potential." *Plant Biosystems - An International Journal Dealing with all Aspects of Plant Biology* 145: 688–698. https://doi.org/10.1080/11263504.2011.601338.

Koorem, K., J. N. Price, and M. Moora. 2011. "Species-Specific Effects of Woody Litter on Seedling Emergence and Growth of Herbaceous Plants." *PLoS One* 6: e26505. https://doi.org/10.1371/journal.pone.0026505.

Krauss, K. W., C. E. Lovelock, K. L. McKee, L. López-Hoffman, S. M. Ewe, and W. P. Sousa. 2008. "Environmental Drivers in Mangrove Establishment and Early Development: A Review." *Aquatic Botany* 89: 105–127. https://doi.org/10.1016/j.aquabot.2007.12.014.

Lang, T., X. Ke, J. Wei, et al. 2024. "Dynamics of Tannin Variations in Mangrove Leaf Litter Decomposition and Their Effects on Environmental Nitrogen and Microbial Activity." *Science of the Total Environment* 908: 168150. https://doi.org/10.1016/j.scitotenv.2023.168150.

Lang, T., P. Wei, X. Chen, et al. 2021. "Microcosm Study on Allelopathic Effects of Leaf Litter Leachates and Purified Condensed Tannins From *Kandelia obovata* on Germination and Growth of *Aegiceras cornicula-tum*." *Forests* 12, no. 8: 1000. https://doi.org/10.20944/preprints202106. 0445.vl.

Liebezeit, G. 2012. "Natural Products Chemistry of the Mangrove Species *Avicennia* Spp. Review and New Data." *Natural Products: An Indian Journal* 11: 10–20.

Lin, X., J. Zhang, H. Chen, and L. Han. 2021. "Determination of Available Phosphorus in Alkaline Soil by Molybdenum Blue Spectrophotometry." *IOP Conference Series: Earth and Environmental Science* 781: 052003. https://doi.org/10.1088/1755-1315/781/5/052003.

Liu, Y., S. Fang, P. Chesson, and F. He. 2015. "The Effect of Soil-Borne Pathogens Depends on the Abundance of Host Tree Species." *Nature Communications* 6: 10017. https://doi.org/10.1038/ncomms10017.

Lovelock, C. E., I. C. Feller, K. L. Mckee, et al. 2004. "The Effect of Nutrient Enrichment on Growth, Photosynthesis and Hydraulic Conductance of Dwarf Mangroves in Panamá." *Functional Ecology* 18: 25–33. https://doi.org/10.1046/j.0269-8463.2004.00805.x.

Mallik, A. U. 2003. "Conifer Regeneration Problems in Boreal and Temperate Forests With Ericaceous Understory: Role of Disturbance, Seedbed Limitation, and Keytsone Species Change." *Critical Reviews in Plant Sciences* 22: 341–366. https://doi.org/10.1080/713610860.

Manurung, J., J. Kappen, J. Schnitzler, et al. 2021. "Analysis of Unusual Sulfated Constituents and Anti-Infective Properties of Two Indonesian Mangroves, *Lumnitzera littorea* and *Lumnitzera racemosa* (Combretaceae)." *Separations* 8: 82. https://doi.org/10.3390/separation s8060082.

Meersmans, J., B. van Wesemael, and M. van Molle. 2009. "Determining Soil Organic Carbon for Agricultural Soils: A Comparison Between the Walkley & Black and the Dry Combustion Methods (North Belgium)." *Soil Use and Management* 25: 346–353. https://doi.org/10.1111/j.1475-2743.2009.00242.x.

Meiners, S. J. 2014. "Functional Correlates of Allelopathic Potential in a Successional Plant Community." *Plant Ecology* 215: 661–672. https://doi.org/10.1007/s11258-014-0331-1.

Minh, L. T. N., D. Orange, T. H. Thai, J. Garnier, L. L. Anh, and T. A. Duc. 2014. "Hydrological Regime of a Tidal System in the Red River Delta, Northern Vietnam." *Hydrology in a Changing World: Environmental and Human Dimensions*: 451–456.

Montesdeoca-Esponda, S., Z. Sosa-Ferrera, and J.-J. Santana-Rodríguez, 2013. "Microwave-Assisted Extraction Combined With On-Line Solid Phase Extraction Followed by Ultra-High-Performance Liquid Chromatography With Tandem Mass Spectrometric Determination of Benzotriazole UV Stabilizers in Marine Sediments and Sewage Sludges." *Journal of Separation Science* 36: 781–788. https://doi.org/10. 1002/jssc.201200664.

Mudrák, O., and J. Frouz. 2012. "Allelopathic Effect of *Salix caprea* Litter on Late Successional Plants at Different Substrates of Post-Mining Sites: Pot Experiment Studies." *Botany* 90: 311–318. https://doi.org/10. 1139/b2012-005.

Nguyen Van, T., A. Ozaki, H. Nguyen Tho, et al. 2016. "Arsenic and Heavy Metal Contamination in Soils Under Different Land Use in an Estuary in Northern Vietnam." *International Journal of Environmental Research and Public Health* 13: 1091. https://doi.org/10.3390/ijerp h13111091.

Quan, W., A. Wang, C. Li, and L. Xie. 2022. "Allelopathic Potential and Allelochemical Composition in Different Soil Layers of *Rhododendron delavayi* Forest, Southwest China." *Frontiers in Ecology and Evolution* 10. https://doi.org/10.3389/fevo.2022.963116.

Rajković, M. B., I. D. Sredović, and Z. N. Miloradović. 2010. "Comparison of Different Methods for Determination of Sodium Chloride in Cheese." *Journal of Agricultural Sciences, Belgrade* 55: 65–77.

Raven, M. R., D. A. Fike, M. L. Gomes, and S. M. Webb. 2019. "Chemical and Isotopic Evidence for Organic Matter Sulfurization in Redox Gradients Around Mangrove Roots." *Frontiers in Earth Science* 7. https://doi.org/10.3389/feart.2019.00098.

Reef, R., I. C. Feller, and C. E. Lovelock. 2010. "Nutrition of Mangroves." *Tree Physiology* 30: 1148–1160. https://doi.org/10.1093/treephys/tpq048.

Reigosa, M. J., M. J. R. Roger, L. Gonzâalez, et al. 2006. *Allelopathy: A Physiological Process With Ecological Implications*. Springer Science & Business Media.

Ruttkies, C., E. L. Schymanski, S. Wolf, et al. 2016. "MetFrag Relaunched: Incorporating Strategies Beyond In Silico Fragmentation." *Journal of Cheminformatics* 8: 3. https://doi.org/10.1186/s13321-016-0115-9.

Sarker, T. C., G. Maisto, A. de Marco, et al. 2020. "Species-Specific Root Proliferation of Tree Seedlings in Tropical Litter: Do Nutrients Matter?" *Oikos* 129: 598–606. https://doi.org/10.1111/oik.06786.

Scavo, A., C. Abbate, and G. Mauromicale. 2019. "Plant Allelochemicals: Agronomic, Nutritional and Ecological Relevance in the Soil System." *Plant and Soil* 442: 23–48. https://doi.org/10.1007/s11104-019-04190-y.

Schandry, N., and C. Becker. 2020. "Allelopathic Plants: Models for Studying Plant–Interkingdom Interactions." *Trends in Plant Science* 25: 176–185. https://doi.org/10.1016/j.tplants.2019.11.004.

Sheue, C.-R., H.-Y. Liu, and J. W. H. Yong. 2003. "*Kandelia obovata* (Rhizophoraceae), a New Mangrove Species From Eastern Asia." *Taxon* 52: 287–294. https://doi.org/10.2307/3647398.

Singh, Y.-S., B. Singh, and S. Sharma. 2021. "Determination of Total, Available, Mineral, Non-exchangeable Ammonium and Potentially Mineralizable Nitrogen in Soil." Accessed January 2024, https://www. researchgate.net/publication/350123952\_Determination\_of\_total\_ available\_mineral\_non-exchangeable\_ammonium\_and\_potentially\_ mineralizable\_nitrogen\_in\_soil.

Smith, C. A., E. J. Want, G. O'Maille, et al. 2006. "XCMS: Processing Mass Spectrometry Data for Metabolite Profiling Using Nonlinear Peak Alignment, Matching, and Identification." *Analytical Chemistry* 78: 779–787. https://doi.org/10.1021/ac051437y.

Spalding, M. 2011. World Atlas of Mangroves. London: Routledge.

Thatoi, H., D. Samantaray, and S. K. Das. 2016. "The Genus Avicennia, a Pioneer Group of Dominant Mangrove Plant Species With Potential Medicinal Values: A Review." *Frontiers in Life Science* 9: 267–291. https://doi.org/10.1080/21553769.2016.1235619.

Tomlinson, P. B. 2016. *The Botany of Mangroves*. 2nd ed. Cambridge: Cambridge University Press.

Vinh, T. V., M. Allenbach, K. T. V. Linh, and C. Marchand. 2020. "Changes in Leaf Litter Quality During Its Decomposition in a Tropical Planted Mangrove Forest (Can Gio, Vietnam)." *Frontiers in Environmental Science* 8: 10. https://doi.org/10.3389/fenvs.2020.00010.

Vitousek, P. M., and R. L. Sanford. 1986. "Nutrient Cycling in Moist Tropical Forest." Annual Review of Ecology and Systematics 17: 137–167.

Weidenhamer, J. D., D. C. Hartnett, and J. T. Romeo. 1989. "Density-Dependent Phytotoxicity: Distinguishing Resource Competition and Allelopathic Interference in Plants." *Journal of Applied Ecology* 26: 613– 624. https://doi.org/10.2307/2404086.

Zhou, H.-C., H.-X. Kang, J. Wei, et al. 2023. "Microcosm Study on Fate and Dynamics of Mangrove Tannins During Leaf Litter Leaching." *Ecological Processes* 12: 37. https://doi.org/10.1186/s13717-023-00453-w.

Zhou, H.-C., S.-D. Wei, Q. Zeng, et al. 2010. "Nutrient and Caloric Dynamics in *Avicennia marina* Leaves at Different Developmental and Decay Stages in Zhangjiang River Estuary, China." *Estuarine, Coastal and Shelf Science* 87: 21–26. https://doi.org/10.1016/j.ecss.2009.12.005.

#### **Supporting Information**

Additional supporting information can be found online in the Supporting Information section.