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# Rheological and mechanical behavior of soy protein-polysaccharide composite paste for extrusion-based 3D food printing: Effects of type and concentration of polysaccharides

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2 **Rheological and mechanical behavior of soy protein-polysaccharide**  
3 **composite paste for extrusion-based 3D food printing: effects of type**  
4 **and concentration of polysaccharides**

5

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21

22 **Abstract**

23 Soy protein isolate (SPI)-based pastes are widely employed for 3D printing of food  
24 materials. However, their rheological properties often do not meet the 3D printing  
25 requirements. To enhance the 3D printing characteristics of SPI pastes, this study  
26 combined SPI with carrageenan and sodium alginate to form composite materials. The  
27 effect of polysaccharide type and concentration on the rheological and textural  
28 properties of the ink was investigated to elucidate the mechanism of interaction between  
29 proteins and polysaccharides. The results demonstrated that the addition of carrageenan  
30 (1.5%, wt %) and sodium alginate (1.5%, wt %) effectively improved the mechanical  
31 properties of the paste, such as hardness and adhesiveness, enhanced thus the quality of  
32 3D printed products. With the increase of polysaccharide proportion, the apparent  
33 viscosity and storage modulus ( $G'$ ) of the composite paste increased significantly.  
34 Fourier-transform infrared spectroscopy (FT-IR), protein solubility measurement, X-  
35 ray diffraction (XRD), and differential scanning calorimetry (DSC) were employed and  
36 the results revealed notable interactions among SPI and two kinds of polysaccharides,  
37 with electrostatic interactions and hydrophobic interactions being the primary driving  
38 forces. The addition of polysaccharides induced changes in the paste crystalline state  
39 and improved its thermal stability. Scanning electron microscopy (SEM) confirmed a  
40 uniform and compact microstructure due to the incorporation of polysaccharides.  
41 Consequently, the SPI-polysaccharide paste can be considered as novel material for 3D  
42 printing food materials, and the results of this study provides a theoretical foundation  
43 for its application in the field of plant-based food products.

44 **Keywords:** 3D printing; SPI-polysaccharide composite paste; Printability; Rheological  
45 properties

46

## 47 **1. Introduction**

48 3D printing is an efficient and convenient processing technology. Because of its  
49 bottom-up stacking approach, can print and manufacture complex and complicated  
50 structures that traditional processing methods cannot construct (Dankar et al., 2018). It  
51 has broad applications in various fields such as textiles, architecture, healthcare, and  
52 food (Keerthana et al., 2020). 3D food printing is revolutionizing the food processing  
53 industry by creating novel foods with complex shapes, distinctive textures, and  
54 enhanced nutritional value. This allows for personalized nutrition and customized food  
55 design, catering to the specific needs of aging populations, children, and sick patients.  
56 (Phuhongsung et al., 2020). Typically, extrusion, binder jetting, and inkjet printing  
57 technologies have been explored for various 3D food printing applications (Cheng et  
58 al., 2022). Among them, extrusion-based printing technique can be effectively used for  
59 semi-solid pastes such as grain dough, meat puree, and chocolate. The principle of this  
60 technique is to extrude material layer by layer through a nozzle to form a complex 3D  
61 structure (Rowat et al., 2021). Extrusion technology is currently the most widely studied  
62 method for 3D food printing. The emergence of 3D food printing technology helps to  
63 accelerate the development of new food products and become a potential technological  
64 mean to rebuild the conventional food supply chain (Chen et al., 2019).

65 Proteins and polysaccharides are two essential nutrients in the human food system  
66 that can alter their structural and functional properties, such as the ability to form gels  
67 or emulsions, through electrostatic forces and hydrogen bonds (Zhang et al., 2022). Soy  
68 protein isolate (SPI) as a food component has been widely used in 3D printing (Chen  
69 et al., 2019; Phuhongsung et al., 2020). However, SPI exhibits limitations in terms of  
70 printing accuracy and stability, as it is prone to breakage under high shearing forces. To  
71 address this issue, the addition of hydrocolloids or salt ions to SPI has been identified  
72 as a viable solution (Xu et al., 2023). Previous research has demonstrated the  
73 effectiveness of incorporating apricot polysaccharide into SPI, resulting in improved  
74 gelatinization performance and viscoelasticity of the SPI gel (Xu et al., 2023).  
75 Polysaccharide is a kind of natural macromolecular carbohydrate with complex and  
76 huge molecular structure obtained from multiple monosaccharides after dehydration

77 and condensation. Although polysaccharides are widely distributed in nature, not all  
78 polysaccharides are called food polysaccharides. Food polysaccharide refers to the  
79 polysaccharide that is allowed as a food additive in the food processing process to  
80 control the sensory quality, physical properties and improve the nutritional value of  
81 food. Kappa-Carrageenan ( $\kappa$ C), a widely used polysaccharide in the food industry,  
82 exhibits rapid thermal-reversible behavior and can form robust and brittle gel structures  
83 suitable for 3D printing (Liu et al., 2019; Yu et al., 2022). Additionally, alginates are  
84 widely used as 3D printing materials because of their good gelling properties due to  
85 their many carboxyl and hydroxyl groups (Chen et al., 2019). Dick et al. (2020)  
86 investigated the effects of different ink polysaccharides on the rheology and texture of  
87 pig meat and found that the addition of ink polysaccharides improved the printability  
88 and texture of the meat. Studies have reported that SPI mixed with sodium alginate and  
89 gelatin as a 3D printing material improved the hardness and chewiness of 3D printed  
90 geometries, and SPI paste will be a promising material for 3D food printing (Chen et  
91 al., 2019).

92 Currently, 3D printing of food is still in its early stages. Warner et al. (2019)  
93 investigated the 3D printability of a mixture of gelatin and  $\kappa$ C and found that the  
94 addition of  $\kappa$ C resulted in a more stable structure than pure gelatin. Commercially, the  
95 3D printed plant-based beef product “Alt Steak” produced by Israeli startup Redefine  
96 Meat has been sold in over 120 restaurants in Israel, the Netherlands, Germany, and the  
97 United Kingdom. Japanese company “Open Meals” uses 3D printer to print edible  
98 pixelated sushi. Each sushi is composed of edible gel with specific color and taste.  
99 However, to date, there has been limited exploration of 3D printing of SPI-  
100 polysaccharide composite pastes. At the same time, carrageenan and sodium alginate  
101 are extracted from plants, and we consider maintaining a plant-based property in the  
102 material properties of the ink, which can be used as a raw material to support the  
103 production of plant meat in the future. Therefore, this study developed a 3D printable  
104 ink by mixing SPI with  $\kappa$ C and alginate polysaccharides. The structural and functional  
105 properties of the ink were evaluated using rheology, texture analysis, Fourier transform  
106 infrared spectroscopy, and scanning electron microscopy. The study investigated the

107 interactions between protein and polysaccharides and their effects on the printability of  
108 the ink.

## 109 **2. Materials and methods**

### 110 **2.1. Materials**

111 SPI (protein content > 90%) was purchased from Shanghai yuanye Bio-  
112 Technology Co., Ltd (Shanghai, China). The ionic content of SPI was determined by  
113 atomic absorption spectrometry, and its composition was  $0.02\pm 0.01$  wt% of  $\text{Ca}^{2+}$ ,  
114  $0.04\pm 0.00$  wt% of  $\text{K}^+$  and  $0.03\pm 0.01$  wt% of  $\text{Na}^+$ . Kappa-carrageenan ( $\kappa\text{C}$ ) and  $\beta$ -  
115 mercaptoethanol were acquired from Shanghai Aladdin Ltd (Shanghai, China). The  
116 ionic content of  $\kappa\text{C}$  was determined by the same procedure, the contents were  $3.5\pm 0.05$   
117 wt% of  $\text{K}^+$ ,  $1.25\pm 0.01$  wt% of  $\text{Na}^+$ ,  $0.47\pm 0.04$  wt% of  $\text{Ca}^{2+}$ ,  $2.1\pm 0.02$  wt% of  $\text{Cu}^{2+}$ , and  
118  $3.2\pm 0.03$  wt% of  $\text{Zn}^{2+}$ . Sodium alginate (SA), tris (hydroxymethyl)methyl  
119 aminomethane (Tris), glycine, urea, sodium dodecyl sulfate (SDS), and disodium  
120 ethylenediaminetetraacetic acid ( $\text{Na}_2\text{EDTA}$ ) were supplied by Shanghai Macklin  
121 Biochemical Technology Co., Ltd (Shanghai, China). The ionic content of SA was  
122 determined by the same procedure, the contents were  $2.31\pm 0.01$  wt% of  $\text{K}^+$ ,  $11.23\pm 0.02$   
123 wt% of  $\text{Na}^+$ ,  $0.03\pm 0.01$  wt% of  $\text{Fe}^{2+}$ . Bovine serum albumin (BSA) was obtained from  
124 Solarbio Science & Technology Co., Ltd (Beijing, China). All other reagents and  
125 chemicals were of analytical grade.

### 126 **2.2. Food 3D Printing Ink Preparation**

127 The mixtures of SPI with sodium alginate and  $\kappa\text{C}$  were prepared as followed. SA  
128 and  $\kappa\text{C}$  solutions were dissolved in deionized water at three concentrations (1, 1.5, and  
129 2% w/w). Subsequently, mixed the two solutions and added SPI powder to the mixed  
130 solution to achieve a concentration of 15% (w/w) of SPI. Table 1 presents eight  
131 representative ink formulations with a gradient cross rise in two kinds of  
132 polysaccharides, the original ratio of SPI and polysaccharide was chosen according to  
133 Hu et al., (2017). The resulting mixed solution was homogenized using a homogenizer  
134 (HFJ-25, Hengao, Tianjin, China) at 500 rpm for 5 min until a stable suspension was  
135 formed. All samples had pH values within  $6.8\pm 0.3$ , which were above the isoelectric  
136 point of soy proteins (Lu et al., 2023). The resulting paste-like mixture was kept in a

137 water bath at 80 °C for 30 min and cooled at room temperature for 15 min. The samples  
138 were stored overnight in the refrigerator and heated to 25 °C before 3D printing.

### 139 **2.3. 3D printing process**

140 The samples were printed using an extrusion-based food 3D printer (FOODBOT-  
141 S2, Hangzhou Shiyin Technology Co., Ltd., Hangzhou, China). The prepared ink filled  
142 into the 3D printing nozzle syringe, and the printer extruded the material by controlling  
143 the movement of the plunger in accordance with the preset 3D mesh model (18.6 mm  
144 × 18.6 mm × 8 mm). The main printing parameters were set as follows: nozzle diameter  
145 of 0.84 mm, nozzle printing rate of 25 mm/s, layer height of 0.8 mm, printing  
146 temperature of 25 °C, extruded rate of 24 mm<sup>3</sup>/s and infill density of 100% printing. To  
147 ensure maximum aseptic operation, all containers, nozzle syringes, and nozzle tips were  
148 sterilized at 121 °C in an autoclave for 15 min. The printing environment was  
149 disinfected with 75% ethanol and UV radiation, and all operations were conducted in a  
150 laminar flow hood.

### 151 **2.4. Rheological Properties of Printing Inks**

152 The rheological properties of the printing inks were measured using a hybrid  
153 rheometer (Discovery HR-2, DHR, TA Instruments, New Castle, DE, USA). A parallel  
154 plate with a diameter of 40 mm was used, with a measurement gap of 1 mm. When  
155 loading the sample, any excess material on the edges was removed with a scraper, and  
156 a thin layer of silicone oil was applied to the edges to prevent moisture evaporation.  
157 The configuration was set as follow: soak time was 180 s, 7 points were detected per  
158 decade, the interval between the points was 10 s, and the steady state sensing in DHR  
159 rheometer was applied to make sure the equilibrium time was 30 s for each point.

#### 160 2.4.1. Flow sweep test

161 The flow sweep test was measured at shear rates ranging from 0.01 to 100 s<sup>-1</sup>, with  
162 a temperature set at 25 °C, to obtain the viscosity  $\eta$  as a function of shear rate and  
163 investigate the shear-thinning behavior of the sample.

#### 164 2.4.2. Amplitude sweep test

165 The linear viscoelastic region (LVR) was determined through an amplitude sweep  
166 test. The test was conducted within a strain range of 0.01-100% at a constant frequency

167 of 10 rad/s, and the experiment was performed at a temperature of 25 °C.

#### 168 2.4.3. Frequency sweep test

169 The dynamic viscoelastic properties of the sample were evaluated through a  
170 frequency sweep test. The oscillation was performed within a frequency range of 0.1-  
171 100 rad/s, with a constant strain of 0.5% (within the LVR), and the experiment was  
172 conducted at a temperature of 25 °C. The storage modulus ( $G'$ ) loss modulus ( $G''$ ) and  
173 loss tangent ( $\tan \delta = G''/G'$ ) were recorded. All tests were repeated in triplicates, and  
174 experimental curves were plotted accordingly.

### 175 **2.5. Texture analysis**

176 Texture characteristics of the 3D printed samples were measured using a texture  
177 analyzer (TA. XT Plus, Stable Micro Systems, UK) equipped with a P36/R probe. The  
178 samples were tested as  $18.6 \times 18.6 \times 8$  mm grid-like architectures printed products for  
179 two compression cycles. The testing parameters were set as follows: the pre-test, test,  
180 and post-test speeds were all set at 1 mm/s, the trigger force was 10 g, the compression  
181 ratio was 50%, the time interval between the two compressions was 5 s, and the  
182 measurement temperature was 25 °C. Each sample was tested three times for  
183 repeatability.

### 184 **2.6. Field-emission Scanning electron microscopy (FE-SEM) analysis**

185 The microstructure of the paste was observed using a scanning electron  
186 microscope. The paste samples were freeze-dried, and the surface of the samples was  
187 sputter-coated with gold under vacuum conditions. The samples were then observed  
188 under the field-emission scanning electron microscope (S-4700, HITACHI, Japan) at  
189 different magnifications (1000 $\times$ , 2500 $\times$  and 5000 $\times$ ) with an accelerating voltage of 15  
190 kV.

### 191 **2.7. Molecular interaction analysis**

192 To elucidate the paste cross-linking mechanism resulting from SPI-polysaccharide  
193 interactions, the protein solubility of the composite paste in five different solvents was  
194 measured to assess the intermolecular forces within the paste according to the method  
195 described by Wang et al. (2018). The paste sample (protein concentration 2 mg/mL)  
196 was dissolved in different solvents. The solvent S1 was distilled water, the solvent S2  
197 consisted of 0.086 M Tris-0.09 M Glycine-4 mM- $\text{Na}_2\text{EDTA}$  (pH = 8.0), the solvent S3



198 was 0.5% SDS, the solvent S4 was 8 M Urea, and solvent S5 was 2%  $\beta$ -  
199 mercaptoethanol. The mixture samples ( $2.00 \pm 0.02$  g) were mixed with 10 mL buffer,  
200 the resulting solution was incubated in a water bath at 25 °C for 20 min, and then  
201 centrifuged at 5000 rpm for 10 min. The protein supernatant was determined using  
202 bovine serum albumin as standard with ultraviolet spectrophotometry at 595 nm  
203 (GENESYS 150, Thermo Fisher Scientific Inc, Waltham, MA, USA). The protein  
204 solubility is expressed as the percentage of protein content of the supernatant in the  
205 total protein content.

206 Tris-glycine buffer disrupts electrostatic interactions, SDS disrupts hydrophobic  
207 interactions by disrupting the medium between the hydrophobic and hydrophilic  
208 regions, urea disrupts hydrogen bonding in the protein by dehydrating the protein  
209 molecules' side chains and backbone, and  $\beta$ -mercaptoethanol reduces protein disulfide  
210 bonds by competing with thiol groups (Qin et al., 2017; Wang et al., 2021). According  
211 to this method, the differences in protein solubility in S2 (Tris-Gly buffer)-S1 (distilled  
212 water), S3 (0.5% SDS)-S2, S4 (urea)-S3, and S5 ( $\beta$ -mercaptoethanol)-S4 were used as  
213 indications of the electrostatic interactions, hydrophobic interactions, hydrogen  
214 bonding interactions, and disulfide bonding interactions, respectively.

## 215 **2.8. Fourier transform infrared spectroscopy (FTIR)**

216 The samples were mixed with dry KBr at a ratio of 1:100 (w/w). Then the mixed  
217 powder was ground in the quartz mortar and pressed into a tablet. FTIR spectra (Nicolet  
218 6700, Thermo Fisher Scientific Inc, USA) was performed at 25 °C in the range of 4000  
219 to 500  $\text{cm}^{-1}$ . The resolution remains at 1  $\text{cm}^{-1}$ .

## 220 **2.9. X-ray diffraction (XRD)**

221 The X-ray diffraction (XRD) patterns of Cu  $K\alpha$  radiation ( $\lambda = 0.1541$  nm) were  
222 recorded by powder diffractometer (Empyrean, Malvern PANalytical, NL). The  
223 working voltage was 40 kV and the working current was 40 mA. The patterns were  
224 collected with a  $2\theta$  range from 5° to 60° at a step of 0.0262°.

## 225 **2.10. Differential scanning calorimetry (DSC)**

226 Differential scanning calorimeter (DSC 8000, PE Instruments Inc., USA) was used  
227 to evaluate the thermal properties of the composite pastes. The thermal analysis was  
228 conducted according to the method described by Wang et al. (2018) with slight

229 modifications. Briefly, the sample mixture (10 mg) was accurately weighed and loaded  
230 into a DSC pan. Prior to analysis, the DSC pan was sealed and maintained at the initial  
231 scanning temperature of 20 °C. The sample was analyzed from 20 °C to 180 °C at a  
232 scanning rate of 10 °C/min, and the temperature of each endothermic or exothermic  
233 peak was determined. A sealed empty pan was used as a blank reference.

#### 234 **2.11. Statistical Analysis**

235 The results are shown as the mean  $\pm$  standard deviation (SD) of triplicate  
236 experiments. SPSS software (SPSS26; IBM SPSS Statistics, Chicago, IL, USA) was  
237 used for one-way ANOVA and Duncan tests at 95% confidence level for data analysis.  
238 All graphs in the experiment were drawn using Origin software (2020b, OriginLab, MA,  
239 USA).

### 240 **3. Results and discussion**

#### 241 **3.1. Printability of composite inks**

242 Soy protein isolate (SPI), as the main material for printing, cannot be directly  
243 printed due to its high flowability and relatively weak cross-linking in water (Shi et al.,  
244 2023). Sodium alginate (SA) is a commonly used hydrophilic colloid with good binding  
245 capacity for proteins (Chen et al., 2022). Kappa-carrageenan ( $\kappa$ C) is a soluble  
246 polysaccharide with high solubility and gelation properties (Bi et al., 2018). The  
247 addition of SA and  $\kappa$ C to the SPI paste could improve the stability and printability of  
248 food ink. In order to screen ink formulations with optimal printing performance, this  
249 study prepared inks with different polysaccharide concentration ratios and examined  
250 their printability. The 3D printing results of different ink formulations are shown in  
251 Figure 1. The successful implementation of food 3D printing requires two conditions:  
252 the material in the extrusion process has the right viscosity to flow through the nozzle  
253 and adhere layer by layer, and the paste structure after extrusion has sufficient  
254 mechanical strength to ensure its formability (Osi et al., 2021). For preliminary tests of  
255 printability, the ink should be continuously extruded from the nozzle as a uniform  
256 filament, and the printed product should show the effect of perfect detail, smooth  
257 surface, and structural stability, rather than being unable to form or causing blockages  
258 and tangled (Figure 1).

259 In the blue irregular region, SPI paste collapsed immediately after continuous and  
260 uniform extrusion from the nozzle, showing the characteristics of some liquid-like  
261 fluids, and the 3D printing molding effect was poor, unable to form and support the  
262 designed three-dimensional grid structure. This could be due to the insufficient gelation  
263 capacity and the low viscosity, and the ink forms irregular printing filaments at the  
264 nozzle, resulting in the inability to form a homogeneous structure (Jiang et al., 2019).  
265 In the orange region, the ink formulation aggregates significantly at the nozzle when  
266 subjected to stress, which may be due to the inappropriate surface tension of the ink,  
267 leading to a tendency for rapid recovery and aggregation of the system after extrusion  
268 (Fu et al., 2021). In the gray region, as the polysaccharide content in the ink gradually  
269 increased, the viscosity of the sample increased, resulting in poor flowability of the ink  
270 material and the nozzle clogging (Fu et al., 2021). In the green printable region, ink  
271 components were able to be continuously and smoothly extruded. The ink can be  
272 printed when the concentration of SA changed from 1 wt% to 2.5 wt% with a  $\kappa$ C  
273 concentration of 1 wt%, or when the concentration of SA changed from 1 wt% to 2.5  
274 wt% with a  $\kappa$ C concentration of 1.5 wt%. However, some ink formulations, such as  
275 S15A1K1.5, S15A1K1, and S15A1.5K1.5, print three-dimensional mesh structure  
276 filaments that are uneven and have broken structures. In contrast, the 3D mesh  
277 structures printed by the SPI15A1.5K1 and S15A2K1.5 formulations had good self-  
278 supporting properties, smooth surfaces and showed uniform layer thickness. Therefore,  
279 to investigate the reasons for the differences in printability among the different  
280 formulations and the effects of different polysaccharide concentrations on ink  
281 printability, five representative ink formulations (irregular: SPI 15; tangled: S15A2K2;  
282 clogged: S15A1.5K2; printable: S15A1.5K1.5 and S15A2K1.5) were selected for in-  
283 depth study.

### 284 **3.2. Rheological properties**

285 The rheological properties of the inks directly influence their printability for 3D  
286 printing based on extrusion samples, with viscosity being a key indicator of extrusion  
287 capability. As the polysaccharide concentration increased, the viscosity of the paste also  
288 increased. As shown in Figure 2a, all samples exhibited a shear rate-dependent

289 reduction in viscosity, demonstrating typical shear-thinning behavior, which is  
290 generally favorable for extrusion-based 3D printing (Dankar et al., 2018). This may be  
291 attributed to the disruption of polymer entanglement during the shear process, causing  
292 polymer chains to stretch and align parallel to the shear direction (Hou et al., 2022;  
293 Zhang et al., 2022). The stress and shear rate applied to the fluid during extrusion  
294 depends on the fluid and extrusion conditions, and these conditions can vary greatly in  
295 different applications (Lopez Hernandez et al., 2021). The apparent shear rate of a  
296 steady-state pipe can be derived by extruding the corresponding flow rate and tube  
297 radius (Xie et al., 2020).

$$298 \quad \dot{\gamma} = \frac{3.34Q}{\pi R^3}$$

299 Where Q is the volumetric flow rate and R is the tube inner radius. According to  
300 the volumetric flow (24 mm<sup>3</sup>/s) and inner radius (0.42 mm), the calculated shear rate is  
301 about 344.4 s<sup>-1</sup>. This result is fitted with the report of (Lopez Hernandez et al., 2021).  
302 The viscosity of the SPI 15 ink at the high shear rate (100 s<sup>-1</sup>) is much lower than the  
303 viscosity at the low shear rate of 0.1 s<sup>-1</sup>. This is because at very low shear rates, the  
304 stable aggregates formed by the glycine subunits of SPI are stabilized by disulfide  
305 bonds, and the side chains are entangled with adjacent chains, resulting in high viscosity.  
306 As the shear rate increases, the unraveling of the side chains and the arrangement of the  
307 molecules provide less shear resistance, thereby reducing the viscosity (Dorishetty et  
308 al., 2019). Some studies have added *Lycium barbarum* polysaccharide to soy protein  
309 isolate, and the results show that the apparent viscosity increases significantly with the  
310 addition of polysaccharide in a concentration-dependent manner (Xing et al., 2024). In  
311 our experiment, the slope of the viscosity versus shear rate was not smooth, which is  
312 also observed in other studies (Lu et al., 2023, Liu et al., 2022, Yu et al., 2022 and Chen  
313 et al., 2019). This may be because the sample has not yet reached equilibrium,  
314 especially at a high shear rate. In the high shear rate region (i.e. from 1 to 100·s<sup>-1</sup>),  
315 molecules or particles within the fluid may not have enough time to rearrange and adjust  
316 their positions. Besides, it is speculated that the turbulent flow happened or the sample  
317 slipped out at the higher shear rate. At the shear rate of 1 s<sup>-1</sup>, the viscosity of S15A1.5K2,

318 S15A1.5K1.5, and SPI 15 food inks was  $1.22 \times 10^3$  Pa s,  $9.81 \times 10^2$  Pa s and  $9.06 \times 10^2$   
319 Pa s, respectively. The viscosity range of S15A1.5K1.5 ink is similar to the results of  
320 (Dick et al., 2020). Among them, the viscosity of more than 1200 Pa s will lead to  
321 entanglement of ink molecules, insufficient pressure in the cylinder, resulting in nozzle  
322 clogging. If the viscosity was less than 900 Pa s, although it is easy to squeeze, the  
323 fluidity was too large to provide sufficient viscoelasticity and stability. Therefore, an  
324 ideal viscosity for the ink should be known during the extrusion process, which allows  
325 the ink to pass through the narrow diameter of the printing nozzle while maintaining its  
326 deposition structure and shape.

327 The nonlinear rheological behavior of samples can be monitored by large  
328 amplitude oscillatory shear test. The linear viscoelastic range (LVER) and critical strain  
329 of ink formulations with different compositions are shown in Figure 2b. LVER is a  
330 range where the storage modulus  $G'$  and loss modulus  $G''$  remain relatively constant as  
331 shear intensity increases. The  $G'$  is a measure of the elastic solid-like behavior, that is,  
332 the resistance to elastic deformation of the sample, while the  $G''$  is the viscous response,  
333 which is the ratio of stress to strain under vibration conditions (Yang et al., 2018). When  
334  $G'$  is greater than  $G''$ , the material behaves primarily as a solid, and vice versa for a  
335 viscous fluid, exhibited a slight frequency dependence (Pan et al., 2022). Within this  
336 range, the material undergoes elastic deformation and can recover to its original state  
337 after the stress is removed. As shown in Figure 2b, when the applied strain is less than  
338 1%, different inks have similar LVERs, maintaining stable values. As the strain  
339 increased, the value of  $G'$  started to decrease, indicating the critical point of the elastic  
340 properties of the samples about 1-2%. The weak gelation ability of SPI at room  
341 temperature results in disordered polymer chains and poor stability, leading to a liquid-  
342 like state after extrusion (Fu et al., 2021). There is a clear positive correlation between  
343  $G'$  and the polysaccharide concentration in the ink, where  $G'$  increases sequentially for  
344 inks S15A1.5K1.5 and S15A2K1.5, indicating higher structural strength, which aligns  
345 with the results of viscosity tests. Among them, ink S15A1.5K1.5 showed the lowest  
346  $G'$  value in all the ink formulations with added polysaccharides. This may be due to a  
347 low polysaccharide concentration, which prevents the ink system from forming a stable

348 paste structure.

349 Frequency sweep tests can provide information about the internal structure of the  
350 printing inks (Hou et al., 2022). As shown in Figure 2c and, 2d, the  $G'$  and  $G''$  of all  
351 pastes exhibit a steady upward trend, with  $G'$  being superior to  $G''$ , indicating that the  
352 ink primarily possesses elastic and solid characteristics. Higher polysaccharide content  
353 in the paste could lead to higher cross-linking density in the SPI-polysaccharide system,  
354 resulting in stronger viscoelasticity and mechanical strength of the paste, as well as  
355 good self-supporting properties and resistance to shape deformation (Pan et al., 2022).  
356 This might be attributed to the formation of a denser and more robust intermolecular  
357 cross-linking structure through molecular interactions between polysaccharides and SPI,  
358 exhibiting stronger solid characteristics. The loss tangent,  $\tan \delta = G''/G'$ , is used as a  
359 characteristic parameter to account for the different viscoelastic behaviors. A  $\tan \delta$  value  
360 less than 1 indicates a predominantly elastic property, while a  $\tan \delta$  value greater than  
361 1 indicates a predominantly viscous property (Yang et al., 2018). The  $\tan \delta$  values of all  
362 samples were less than 1, which indicated that the addition of polysaccharide enhanced  
363 the gel properties of the mixture and made it form a rigid gel structure. The addition of  
364  $\kappa$ C can induce electrostatic interactions with proteins, leading to the predominant solid-  
365 like behavior of these fluids (Lu et al., 2022). Among them, with a constant  $\kappa$ C  
366 concentration of 1.5 wt% and angular frequency of 10 rad/s, when the SA concentration  
367 was increased from 1.5 wt% to 2 wt%,  $G'$  and  $G''$  increase from 24970.3 Pa and 4042.66  
368 Pa to 30697.3 Pa and 4845.1 Pa, respectively.  $G'$  increased by 22.94% and  $G''$  increased  
369 by 19.85%. However, with a constant SA concentration of 2 wt% and angular frequency  
370 of 10 rad/s, when the  $\kappa$ C concentration was increased from 1.5 wt% to 2 wt%,  $G'$  and  
371  $G''$  increased from 30697.3 Pa and 4845.1 Pa to 32279.5 Pa and 5002.04 Pa,  
372 respectively.  $G'$  increased by 51.54% and  $G''$  increased by 3.24%. These results suggest  
373 that polysaccharides participate in the formation of the paste structure, and different  
374 types of polysaccharides lead to different structures. It has been reported that with the  
375 increase of  $\kappa$ C concentration, the interaction between  $\kappa$ C-proteins may gradually  
376 decrease and be replaced by the  $\kappa$ C- $\kappa$ C interaction (Lu et al., 2022). Some studies have  
377 shown that the gel strength of calcium alginate gels is relatively weak (Fernández Farrés

378 et al., 2013). Compared to SA,  $\kappa$ C has a greater impact on the viscoelasticity of the  
379 paste, and at higher concentrations, the paste structure becomes more rigid. However,  
380 excessively high viscoelasticity of the paste may have adverse effects on 3D printing,  
381 as the generated high shear forces can hinder ink extrusion or cause fracture after  
382 extrusion. On the other hand, the higher the concentration of SA, the smoother the  
383 extrusion process, the flatter the filaments, making it more suitable for printing, which  
384 will be confirmed by the results of texture analysis.

385 In general, the printability of soybean protein-polysaccharide composite paste is  
386 aimed at printing out self-supporting, flat sample and smooth surface. To ensure  
387 printability, the ink should exhibit contrasting properties. Initially, it must be fluid  
388 enough to be pumped and extruded, yet it should also stiffen rapidly after deposition to  
389 enable construction. Therefore, the amount of different polysaccharides added is  
390 particularly important, and the range of proper mechanical properties should be  
391 between 900 Pa s and 1200 Pa s.

### 392 **3.3. Texture profile analysis (TPA)**

393 Texture profile analysis can detect several texture parameters, including hardness,  
394 springiness, adhesiveness, and others (Yuan et al., 2016). As shown in Table 2, paste  
395 samples without polysaccharides cannot determine the TPA parameters due to their  
396 extremely weak gel properties, highlighting the importance of incorporating  
397 polysaccharides in the ink for the production of self-supporting 3D printed food  
398 products. With an increase in polysaccharide concentration, the springiness,  
399 cohesiveness, gumminess, and resilience of the printed samples all significantly  
400 decreased (Table 2). This could be attributed to the changes in the repulsive forces  
401 between polysaccharide charges that affect molecular aggregation. Both carrageenan  
402 and sodium alginate are anionic polysaccharides that can form complexes with the  
403 amino group ( $-\text{NH}_3^+$ ) of the protein through electrostatic interactions, thereby altering  
404 the printing characteristics of the paste (Huang et al., 2021). Sulfated polysaccharides  
405 such as carrageenan have the ability to interact with proteins above their isoelectric  
406 point.  $\kappa$ C contains a sulfate group ( $-\text{OSO}_3^-$ ) that can be bonded to the amino group ( $-\text{NH}_3^+$ )  
407 of the protein by electrostatic interaction. The electrostatic interaction between

408 sulfate group ( $-OSO_3^-$ ) and protein is stronger than that between carboxyl group ( $-$   
409  $COO^-$ ) and protein (Huang et al., 2021). Furthermore, SPI possesses a globular  
410 structure, and the folded SPI structure can stretch and expose internal functional group  
411 structures upon heating, facilitating cross-linking with polysaccharide ions.

412 Hardness refers to the force required to compress a sample and can mimic the  
413 chewing behavior of the oral cavity on solids or semi-solids, serving as an indicator of  
414 the structural stability of 3D printed food. Compared to pastes with the same  
415 concentration of SA, the addition of  $\kappa$ C significantly affects the properties, with an  
416 increase in sample hardness and a decrease in adhesiveness and springiness. When the  
417  $\kappa$ C concentration reached 2 wt%, high shear forces occur during the extrusion process,  
418 leading to filament breakage at the nozzle, resulting in poor shape fidelity and  
419 significant discrepancies between the printed samples and the intended model, making  
420 it also impossible to determine the TPA parameters. In contrast, the addition of SA, with  
421 the same concentration of  $\kappa$ C, yields opposite results. This may be attributed to the  
422 disruption of the molecular chain structure of the samples during preparation, where  
423 longer polysaccharide chains exhibit stronger gelation properties with proteins  
424 (Monteiro & Lopes-da-Silva, 2017). The springiness of the samples refers to the  
425 physical reboundness after deformation, which can resist to external forces. At higher  
426 concentrations, polysaccharides dominate the springiness of the mixture, leading to a  
427 weakening of samples structure, as also reflected in the aforementioned rheological  
428 analysis (Warnakulasuriya & Nickerson, 2018). This is contrary to the experimental  
429 conclusion of (Xu et al., 2023). This result may be due to the fact that the sample has a  
430 hollow structure, and when the extrusion degree is 50%, the sample is forced to extrude  
431 from all sides to widen, resulting in a decrease in data presentation. Cohesiveness refers  
432 to the adhesive forces within the sample particles, reflecting the magnitude of  
433 intermolecular binding forces that allow the sample to maintain its integrity even under  
434 pressure. The data indicates that there is only a minor difference in cohesiveness  
435 between the samples, and the low concentration of polysaccharide improved the  
436 structural properties of the paste. When the concentration of polysaccharide was too  
437 high ( $>1.5\%$ , wt %), the Cohesiveness and resilience of the sample decreased, which



438 was consistent with the results of (Xu et al.,2023). This may be attributed to changes in  
439 the repulsive forces between the negative charges in the polysaccharide, which affect  
440 molecular aggregation and thus alter the texture properties of the sample (Xu et  
441 al.,2023). The above results suggested a positive correlation between paste textural  
442 properties and polysaccharide concentration. Considering that plant-based foods are  
443 extruded with current inks as the main material, the protein paste having the  
444 formulations S15A1.5K1.5 exhibits optimal texture properties and is most suitable for  
445 3D printing. In fact, the texture of plant-based foods largely depends on their structure,  
446 mechanical properties, and surface characteristics (Shi et al., 2023).

#### 447 **3.4. Microstructure**

448 Figure 3 illustrates the microstructures of different ink formulations at various  
449 magnifications (1000×, 2500×, and 5000×). All food inks exhibit a heterogeneous  
450 microstructure composed of protein-polysaccharide particles forming aggregates. The  
451 structure of soybean isolate protein (SPI) has a rough surface because it is a paste at  
452 room temperature. However, the structural support provided by these concentrations is  
453 still insufficient to meet the required criteria, which may be the reason for the inability  
454 to achieve proper printability (Figure 1). Therefore, an appropriate polysaccharide  
455 concentration needs to be added to achieve gelation. Different ink formulations have  
456 different densification and orientation of the cavity, which is especially obvious after  
457 the addition of polysaccharides. Sodium alginate (SA) and kappa carrageenan ( $\kappa$ C) bind  
458 strongly to protein particles, forming small fragmented block formations. SA,  $\kappa$ C, and  
459 SPI particles form non-covalent bonds due to their hydrophilic nature and abundance  
460 of carboxyl and hydroxyl groups, which promote hydrophobic interactions and  
461 hydrogen bonding (Huang et al., 2021; Wei et al., 2023). As the  $\kappa$ C content increased,  
462 the cross-linking structure with SPI-polysaccharide particles became rougher, resulting  
463 in a more disordered and chaotic ink system, forming large aggregates. This could be  
464 an increased interaction between  $\kappa$ C and  $\kappa$ C (Lu et al., 2022). This may explain why  
465 ink S15A1.5K1.5 was able to be extruded from the nozzle while ink S15A1.5K2  
466 became clogged and ink S15A2K2 agglomerated (Figure 1). SA more ink formula  
467 surface is more compact and smoother, which is consistent with (Chen et al., 2022).

468 Overall, ink S15A1.5K1.5 has the best network density and can resist structural changes  
469 under high shear stress during the extrusion process.

### 470 **3.5. Molecular interaction force analysis**

471 As shown in Figure 4a, the solubility of the composite paste was significantly  
472 enhanced, indicating the presence of hydrophobic interactions, electrostatic interactions,  
473 disulfide bonds, and hydrogen bonds within the protein paste structure. The interaction  
474 forces between protein molecules and polysaccharides followed the order of  
475 hydrophobic interactions > electrostatic interactions > disulfide bond interactions >  
476 hydrogen bond interactions. This observation was consistent with the previously  
477 reported results (Lu et al., 2023). A slight decrease in electrostatic interactions was  
478 observed in the composite paste. This result can be attributed to the influence of  
479 negatively charged groups on the protein and polysaccharide surfaces, which are  
480 affected by the electrostatic binding under high polysaccharide concentrations, leading  
481 to conformational changes in the protein and exposing more hydrophobic groups (Wang  
482 et al., 2021). When electrostatic repulsion further increases, the distance between  
483 molecules becomes longer, making it difficult to detect the signal of electrostatic  
484 repulsion, resulting in decreased solubility. Compared to pure SPI paste, the addition of  
485 polysaccharides significantly increased hydrophobic interactions and hydrogen bond  
486 interactions in the paste ( $p < 0.05$ ). This finding indicates that the addition of  
487 polysaccharides greatly enhances hydrophobic interactions and hydrogen bond  
488 interactions, playing a crucial role in the formation and maintenance of the SPI-  
489 polysaccharide paste system. This phenomenon may be attributed to the exposure of  
490 chemical groups within the protein matrix after thermal treatment. SA and  $\kappa$ C are  
491 anionic acidic polysaccharides rich in carboxyl and hydroxyl groups. Although there is  
492 an intrinsic repulsion between them, but other forces can maintain the state without  
493 causing the system to separate, especially at high concentrations. Studies have shown  
494 that there is an electrostatic interaction between SPI and  $\kappa$ C, where the negatively  
495 charged sulfate groups on  $\kappa$ C can interact with the positively charged amino groups ( $-$   
496  $\text{OSO}_3^- \text{NH}_3^+$ ) on proteins, even at pH above the isoelectric point (Huang et al., 2021; Lu  
497 et al., 2023). Additionally, SPI and polysaccharides can absorb moisture, promoting

498 further aggregation of proteins and overcoming the biocompatibility and inherent  
499 repulsion between SPI and polysaccharides, ultimately forming a paste cross-linking  
500 structure (Cao et al., 2022; Min et al., 2022).

### 501 **3.6. Composite paste forming mechanism**

502 Based on the intermolecular interactions and rheological properties discussed  
503 above, this study presents a plausible hypothesis regarding the gelation mechanism of  
504 SPI-polysaccharide (Figure 5). The experimental results revealed that hydrophobic  
505 interactions and electrostatic interactions are the predominant forces in the composite  
506 paste. Heating facilitates the exposure of hydrophobic groups in the protein, while the  
507 polysaccharides SA and  $\kappa$ C contain numerous carboxyl and hydroxyl groups,  
508 enhancing hydrophobic interactions and hydrogen bonding with SPI particles (Huang  
509 et al., 2021; Wei et al., 2023). The presence of a substantial number of hydrophobic  
510 groups leads to protein crosslinking and aggregation.  $\kappa$ C, on average, contains an  
511 anionic sulfate group ( $-\text{OSO}_3^-$ ) per disaccharide repeating unit, which can engage in  
512 electrostatic interactions with cationic groups ( $-\text{NH}_3^+$ ) of the protein. The electrostatic  
513 interaction between sulfate group ( $-\text{OSO}_3^-$ ) and protein is stronger than that between  
514 carboxyl group ( $-\text{COO}^-$ ) and protein (Huang et al., 2021; Lu et al., 2023). In the SPI  
515 system, the addition of  $\kappa$ C forms protein-polysaccharide interactions, establishing a  
516 balance between protein-protein and protein-polysaccharide interactions. This is  
517 consistent with the fact that in all oscillation studies, the composite gel is visually  
518 homogeneous and there is no phase separation. When the polysaccharide concentration  
519 exceeds 1.5 wt%, the self-aggregation of polysaccharides hinders SPI cross-linking,  
520 reducing its interaction with SPI. As the  $\kappa$ C content increases, the  $\kappa$ C- $\kappa$ C interaction  
521 increases and the  $\kappa$ C-protein interaction decreases, resulting in a stronger gel that is  
522 unable to extrude from the nozzle. The molecular structure of SA is rich in hydroxyl  
523 and carboxylic groups, and it is easy to form intermolecular hydrogen bonds with the  
524 functional groups of other polymers (Wei et al., 2023). However, it has been reported  
525 that SA alone exhibits inadequate mechanical properties in soft tissue engineering  
526 applications, failing to provide sufficient mechanical support under stress conditions,  
527 which aligns with our experimental results (Wei et al., 2023). But when the addition

528 amount of SA exceeds a certain critical value (2.5 wt%), it forms a gel with soybean  
529 protein, yielding improved printing performance. Some studies have successfully  
530 prepared alginate/carrageenan composite hydrogels with different concentrations of  
531 carrageenan, and bio-printed the alginate/carrageenan scaffold, the results show that the  
532 gel is suitable for using bioprinting to manufacture 3D hydrogel scaffold (Kim et al.,  
533 2019).

### 534 **3.7. Fourier transform infrared (FTIR) analysis**

535 In order to investigate the changes in chemical bonds in the inks, a comparison of  
536 their FTIR spectra was conducted, as shown in Figure 4b. The FTIR spectra of the paste  
537 samples did not exhibit any new peaks upon the addition of polysaccharides, and there  
538 were no new functional groups observed with varying ratios of SA and  $\kappa$ C, indicating  
539 that the covalent interactions were minor. This is consistent with the research results of  
540 (Xing et al., 2024). All inks displayed a strong and broad absorption peak in the range  
541 of 3200-3600  $\text{cm}^{-1}$ , which is typically associated with the stretching of O-H bonds and  
542 partially formed hydrogen bonds (Yu et al., 2022). Compared to the SPI 15, the O-H  
543 stretching bands of the ink with added polysaccharides shifted to lower wavenumbers.  
544 For instance, in the inks S15A2K1.5 and S15A2K2, the band of the SPI 15 ink at 3273  
545  $\text{cm}^{-1}$  shifted to 3282  $\text{cm}^{-1}$  and 3286  $\text{cm}^{-1}$ , respectively, suggesting stronger hydrogen  
546 bonding and increased molecular interactions in the S15A2K2 ink (Min et al., 2022).  
547 In fact, hydrogen bonding can form connections between proteins and polysaccharides,  
548 enhancing cross-linking strength, which is crucial for controlling the printability of  
549 different biomacromolecules in food inks (Warnakulasuriya & Nickerson, 2018). In all  
550 mixed pastes, weak peaks were also observed in the range of 2998-3018  $\text{cm}^{-1}$ , which  
551 could be attributed to the stretching vibrations of C-H bonds (Min et al., 2022).  
552 Furthermore, all inks exhibited a peak at approximately 1633  $\text{cm}^{-1}$ , which is related to  
553 the asymmetric and symmetric stretching of carboxylate groups ( $\text{COO}^-$ ) and has been  
554 suggested to be caused by the bending vibrations of water molecules (1650  $\text{cm}^{-1}$ ) (Hu  
555 et al., 2017; Shi et al., 2023). It is noteworthy that when different polysaccharides were  
556 added to SPI, all inks showed absorption peaks in the range of 1515-1531  $\text{cm}^{-1}$ , with  
557 the bands shifting from higher to lower wavenumbers. These peaks are associated with

558 the amide I (1600–1700  $\text{cm}^{-1}$ ) and amide II (1500–1600  $\text{cm}^{-1}$ ) regions in the FT-IR  
559 spectra. The amide regions primarily represent the stretching vibrations of C=O, the  
560 bending vibrations of N-H, and the stretching vibrations of C-N, which are reliable  
561 indicators of protein secondary structure conformation (Carbonaro & Nucara, 2010).  
562 The shift of bands from higher to lower wavenumbers indicates a decrease in the  
563 content of  $\alpha$ -helical structures and an increase in the proportion of  $\beta$ -sheet structures in  
564 the paste (Liu et al., 2022). The results demonstrate that the combination of  
565 polysaccharides and SPI significantly alters the spatial structure of the paste.

### 566 **3.8. X-ray diffraction (XRD) analysis**

567 The X-ray diffraction patterns can be used to determine the amorphous or  
568 crystalline nature of molecules (Dong et al., 2021). As shown in Figure 4c, SPI exhibits  
569 two sharp peaks at diffraction angles of  $9^\circ$  and  $19^\circ$ , indicating its crystalline structure.  
570 Its primary structure is believed to consist of an  $\alpha$ -helical and  $\beta$ -sheet structure (Chen  
571 et al., 2013; Dong et al., 2021). In contrast, the addition of polysaccharides to the  
572 composite paste results in two relatively flat peaks around  $9^\circ$ , suggesting a significant  
573 reduction in the  $\alpha$ -helical structure and a presumed transition to the  $\beta$ -sheet structure.  
574 This also indicates the formation of amorphous complexes between SPI and  
575 polysaccharide molecules. Such amorphous nature is directly related to the  
576 hydrophilicity and hygroscopicity of the polymer (Wang et al., 2018). Composite pastes  
577 prepared with formulations S15A1.5K1.5 and S15A1.5K2 exhibit broad diffraction  
578 peaks at approximately  $19^\circ$ , while formulations S15A2K1.5 and S15A2K2 show  
579 increased peak intensities and sharpened peaks. These structural changes suggested a  
580 corresponding interplay between the  $\alpha$ -helical and  $\beta$ -sheet structures, with an increase  
581 in SA concentration potentially leading to the transformation of the  $\alpha$ -helical structure  
582 to the  $\beta$ -sheet structure in the paste samples (Pan et al., 2022). It has been found that  $\beta$ -  
583 sheet structures have greater potential energy compared to  $\alpha$ -helices, which can explain  
584 why the composite paste exhibits a more stable structure than the SPI paste (Ding et al.,  
585 2003). When the SA concentration is 1.5 wt%, the peak intensity of the composite paste  
586 significantly weakens. This indicates good compatibility between the two polymers (Y.  
587 Dong et al., 2021). At an SA concentration of 2 wt%, the peak intensity increases and

588 becomes sharper around 19°, indicating the formation of amorphous complexes and  
589 intermolecular interactions. These results demonstrated that the presence of κC and SA  
590 influences the crystallinity of the SPI paste system making it promising for 3D printing.

### 591 **3.9. Differential scanning calorimetry (DSC) analysis**

592 The properties and structure of most substances vary with temperature, and  
593 therefore, the stability of the sample structure can be reflected by the changes in  
594 endothermic and exothermic behavior in the DSC spectra. The enthalpy change and  
595 transition temperature can be determined based on the temperature and peak area  
596 corresponding to the largest peak in the graph (Hou et al., 2022). The influence of  
597 different polysaccharide concentrations on the thermal stability of the composite paste  
598 is shown in Figure 4d. The addition of the polysaccharides significantly affects the  
599 thermal transition temperature, resulting in an upward shift of the melting temperature  
600 from 89.6 °C to higher temperatures as the polysaccharide content increases. The  
601 melting temperatures for S15A1.5K1.5, S15A1.5K2, S15A2K1.5, and S15A2K2 are  
602 95.3 °C, 92.8 °C, 93.5 °C, and 95.2 °C, respectively. This can be attributed to the  
603 interaction between the amino and carboxyl groups of SPI and the hydroxyl groups of  
604 the polysaccharides through hydrogen bonding, consistent with the aforementioned  
605 intermolecular forces. As the polysaccharide quantity increases, the melting  
606 temperature of the composite paste gradually rises, indicating a higher amount of heat  
607 absorbed during the sample melting process and an enhancement in thermal stability.  
608 Within the melting temperature range, SA and κC in the composite paste may form a  
609 cross-linked structure, with some SPI potentially participating in the formation of the  
610 structure and being encapsulated within it. This promotes the stability of the dense  
611 cross-linking system, ultimately leading to changes in thermal stability. These findings  
612 suggest a certain degree of interaction between SPI and polysaccharides, resulting in a  
613 covalently cross-linked composite paste system, which could explain the paste's  
614 elasticity, viscosity, and strength.

### 615 **4. Conclusion**

616 In this study, a 3D printable ink was prepared using soy protein isolate,  
617 carrageenan, and sodium alginate. The addition of polysaccharides to soy protein at

618 appropriate concentrations facilitated the interaction between proteins and  
619 polysaccharides, resulting in protein-polysaccharide pastes with optimal 3D printing  
620 quality. This combination provides a viable method for the production innovative of  
621 plant-based foods. Rheological and textural analyses revealed that the addition of  
622 polysaccharides increased the viscosity and hardness of the printing ink. The  
623 S15A1.5K1.5 ink exhibited favorable viscosity and storage modulus ( $G'$ ), enabling  
624 stable printability. FT-IR analysis and molecular interaction within the dense cross-  
625 linking indicated that the introduction of polysaccharides led to the formation of  
626 hydrogen bonds through the -OH groups, transforming  $\alpha$ -helices into  $\beta$ -sheets.  
627 Hydrophobic interactions, electrostatic interactions, and disulfide bonding were  
628 identified as the main driving forces maintaining the paste structure. SEM images  
629 displayed the rough surface of the composite material, confirming the formation of a  
630 dense cross-linking structure between SPI and polysaccharides. XRD results indicated  
631 that the interactions between SPI and polysaccharides led to a transition from a  
632 crystalline to an amorphous structure. DSC analysis demonstrated that the addition of  
633 polysaccharides significantly enhanced the thermal stability of the composite paste.  
634 These findings contribute to the improvement of texture and quality in protein-  
635 polysaccharide pastes, as well as the practical application of 3D printing technology in  
636 plant-based food, establishing both theoretical and practical foundations. In our next  
637 studies, we will explore various ink formulations using other types of food ingredients,  
638 such as pea protein isolate, to meet specific nutritional requirements of individuals.

639

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798

799 **Table captions**

800 **Table 1.** Composition of different 3D printing ink formulations (wt%).

801 **Table 2.** Texture profile analysis (TPA) indicators for different 3D printed pastes with  
802 different formulations.

803

804

**Table 1.** Composition of different 3D printing ink formulations (wt%).

<b>Inks</b>	<b>Water/(ml)</b>	<b>SPI/(g)</b>	<b>SA/(g)</b>	<b><math>\kappa</math>C /(g)</b>
SPI 15	85	15	-	-
S15A1K1	85	15	1.0	1.0
S15A1K1.5	85	15	1.0	1.5
S15A1.5K1	85	15	1.5	1.0
S15A1.5K1.5	85	15	1.5	1.5
S15A1.5K2	85	15	1.5	2.0
S15A2K1.5	85	15	2.0	1.5
S15A2K2	85	15	2.0	2.0

805 In the codes listed for formulation column, S15AmKn food ink means that SPI, SA, and  $\kappa$ C

806 constitute 15 wt%, m wt% and n wt%, respectively.

807

808 **Table 2.** Texture profile analysis (TPA) indicators for different 3D printed pastes with  
 809 different formulations.

Sample	Hardness (g)	Adhesiveness	Springiness	Cohesiveness	Gumminess	Chewiness	Resilience (g•sec)
SPI 15	-	-	-	-	-	-	-
S15A1K1	529.27±14.13 <sup>a</sup>	-7.22±3.31 <sup>a</sup>	0.78±0.02 <sup>a</sup>	0.42±0.03 <sup>a</sup>	223.55±19.90 <sup>a</sup>	173.18±13.32 <sup>a</sup>	0.17±0.01 <sup>a</sup>
S15A1K1.5	552.91±20.28 <sup>a</sup>	-23.60±2.73 <sup>c</sup>	0.46±0.02 <sup>c</sup>	0.37±0.01 <sup>b</sup>	205.01±2.26 <sup>b</sup>	94.93±4.41 <sup>b</sup>	0.11±0.01 <sup>b</sup>
S15A1.5K1	480.10±8.76 <sup>b</sup>	-14.14±3.39 <sup>b</sup>	0.59±0.02 <sup>b</sup>	0.36±0.01 <sup>bc</sup>	172.20±4.56 <sup>c</sup>	101.65±5.83 <sup>b</sup>	0.11±0.01 <sup>b</sup>
S15A1.5K1.5	538.73±12.16 <sup>a</sup>	-56.24±1.93 <sup>e</sup>	0.45±0.02 <sup>c</sup>	0.33±0.01 <sup>c</sup>	180.06±7.39 <sup>c</sup>	81.88±2.62 <sup>c</sup>	0.08±0.00 <sup>d</sup>
S15A1.5K2	-	-	-	-	-	-	-
S15A2K1.5	429.84±7.18 <sup>c</sup>	-43.74±3.38 <sup>d</sup>	0.45±0.01 <sup>c</sup>	0.35±0.00 <sup>bc</sup>	150.22±2.08 <sup>d</sup>	67.47±0.47 <sup>d</sup>	0.10±0.00 <sup>c</sup>
S15A2K2	-	-	-	-	-	-	-

810 Values were expressed as the mean ± standard deviation (n = 3). Different letters indicated  
 811 significant differences ( $p < 0.05$ ) between values in the same column. "-" means not shaped.  
 812 S15AmKn food ink means that SPI, SA, and κC constitute 15 wt%, m wt% and n wt%,  
 813 respectively.  
 814

815 **Figure captions**

816 **Figure 1.** Analysis of 3D printing of different ink formulations and pictures of the  
817 printed products.

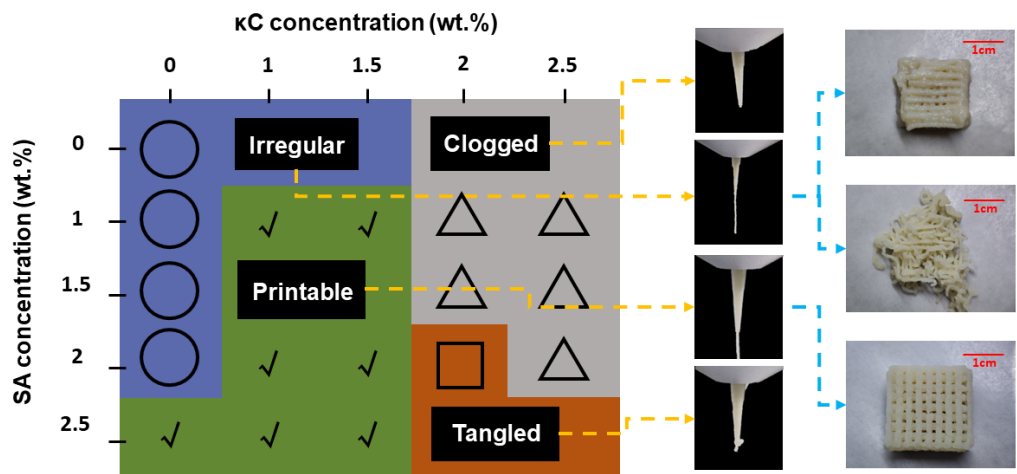
818 **Figure 2.** Rheological properties analysis of different ink formulations: (a): viscosity;  
819 (b): strain provide information which lines for  $G'$  and  $G''$ ; (c): storage modulus  $G'$ ; (d):  
820 loss modulus  $G''$ .

821 **Figure 3.** Microstructure of ink formulations with different magnifications of  $1000\times$ ,  
822  $2500\times$  and  $5000\times$ .

823 **Figure 4.** (a): Changes in molecular interaction force of the composite pastes in the  
824 presence of different polysaccharide concentrations. Mean values with different letters  
825 (a-d) differ significantly ( $p < 0.05$ ); (b): The Fourier transform infrared, (FTIR)  
826 spectrum characterization, (c): X-ray diffractograms and (d): DSC thermograms of  
827 composite pastes with different formulations.

828 **Figure 5.** The formation mechanism diagram of SPI-polysaccharide composite pastes.  
829

830 **Figure 1**

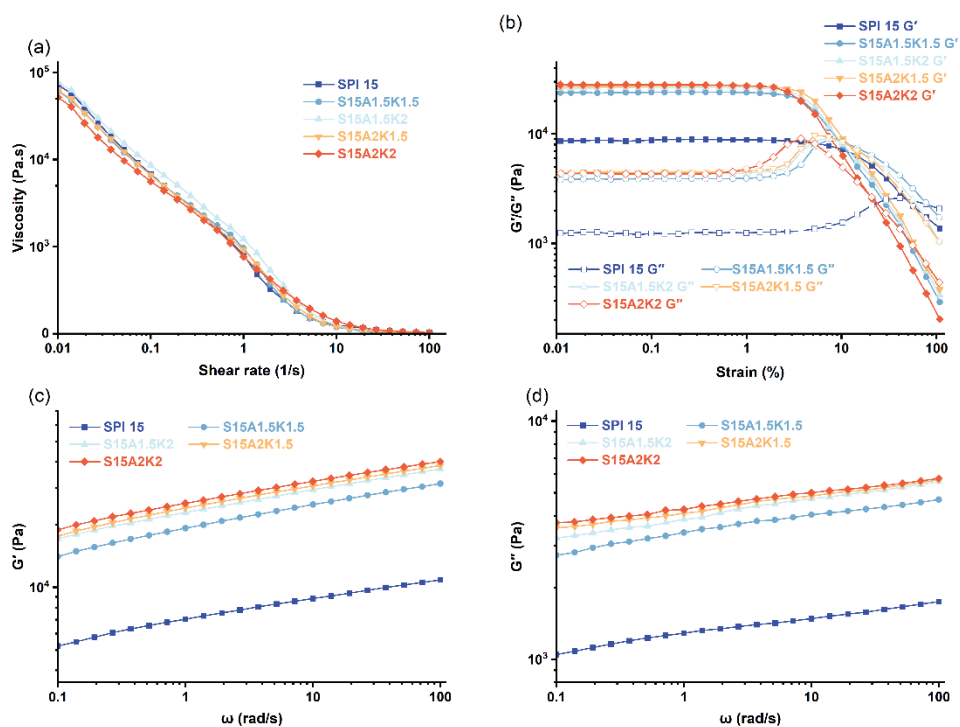


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832 Figure 1. Analysis of 3D printing of different ink formulations and pictures of the  
833 printed products.

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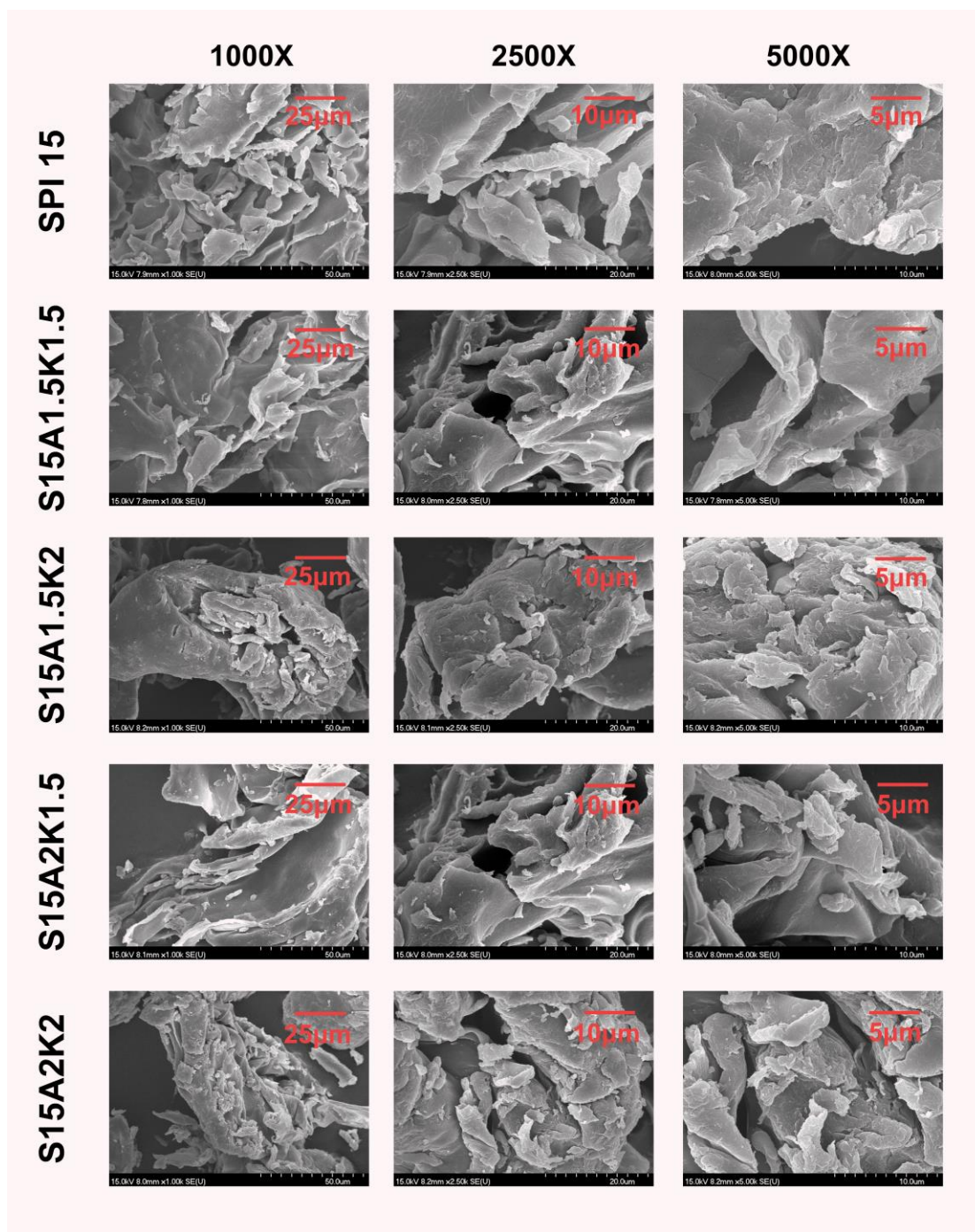
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837 Figure 2. Rheological properties analysis of different ink formulations: (a): viscosity;

838 (b): strain provide information which lines for G' and G''; (c): storage modulus G'; (d):

839 loss modulus G''.

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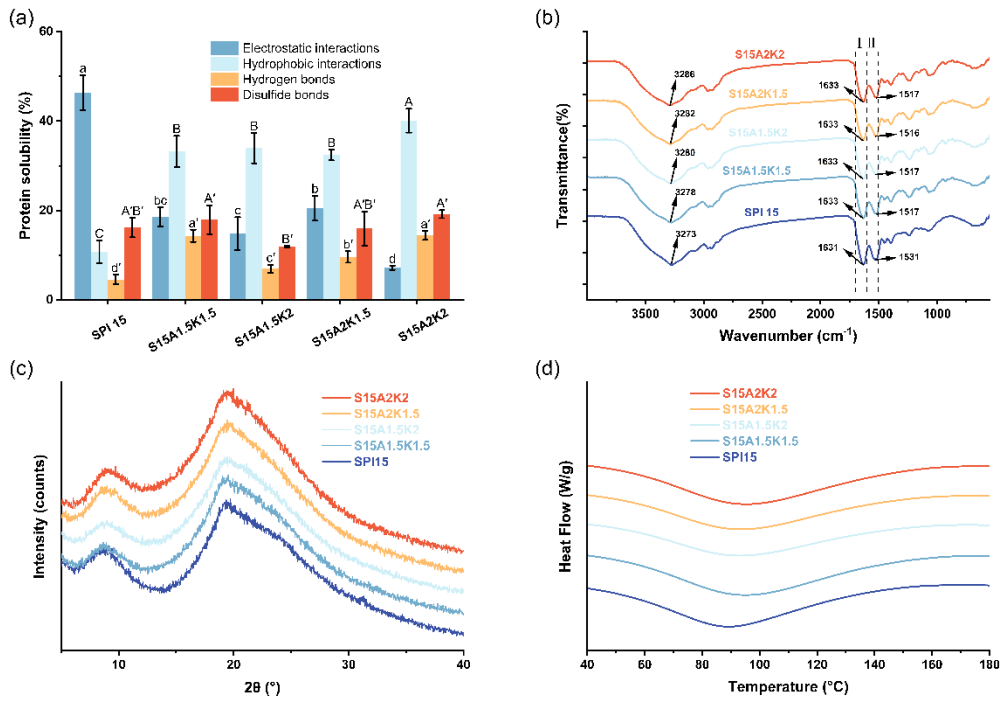


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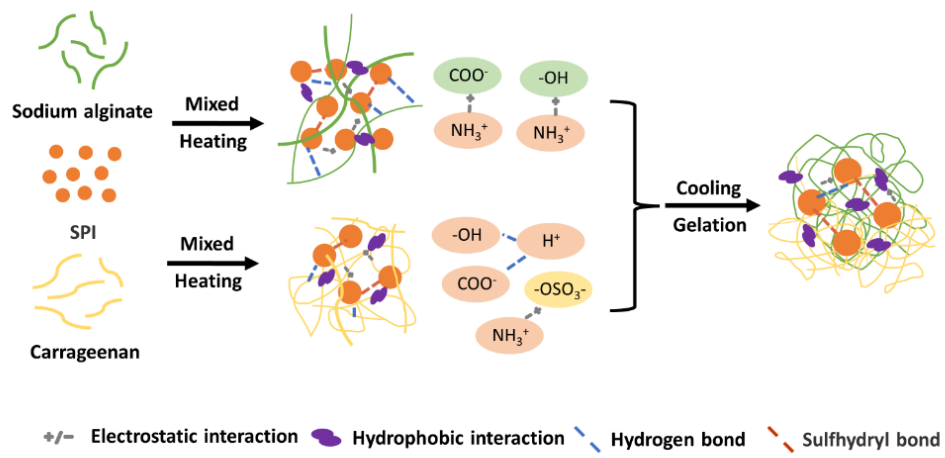
843 Figure 3. Microstructure of ink formulations with different magnifications of 1000 ×,

844 2500 × and 5000 ×.

845



847 Figure 4. (a): Changes in molecular interaction force of the composite pastes in the  
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 851 composite pastes with different formulations.  
 852



854 Figure 5. The formation mechanism diagram of SPI-polysaccharide composite pastes.

The authors declared that they have no conflicts of interest to this work. We declare that we do not have any commercial or associative interest that represents a conflict of interest in connection with the work submitted.

### Author Contributions

Jian Wang: Conceptualization, Methodology, Writing - Review & Editing, Funding acquisition. Qianjin Jiang: Investigation, Data curation, Software, Writing - Original draft preparation. Zhenyu Huang: Visualization, Methodology, Investigation. Ahsan Hafiz Muhammad: Writing - Reviewing and Editing. Adem Gharsallaoui: Writing - Reviewing and Editing. Ming Cai: Supervision, Funding acquisition. Kai Yang: Supervision, Funding acquisition. Peilong Sun: Supervision, Project administration.

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**Rheological and mechanical behavior of soy protein-polysaccharide composite paste for extrusion-based 3D food printing: effects of type and concentration of polysaccharides**

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22 **Abstract**

23 Soy protein isolate (SPI)-based pastes are widely employed for 3D printing of food  
24 materials. However, their rheological properties often do not meet the 3D printing  
25 requirements. To enhance the 3D printing characteristics of SPI pastes, this study  
26 combined SPI with carrageenan and sodium alginate to form composite materials. The  
27 effect of polysaccharide type and concentration on the rheological and textural  
28 properties of the ink was investigated to elucidate the mechanism of interaction between  
29 proteins and polysaccharides. The results demonstrated that the addition of carrageenan  
30 (1.5%, wt %) and sodium alginate (1.5%, wt %) effectively improved the mechanical  
31 properties of the paste, such as hardness and adhesiveness, enhanced thus the quality of  
32 3D printed products. With the increase of polysaccharide proportion, the apparent  
33 viscosity and storage modulus ( $G'$ ) of the composite paste increased significantly.  
34 Fourier-transform infrared spectroscopy (FT-IR), protein solubility measurement, X-  
35 ray diffraction (XRD), and differential scanning calorimetry (DSC) were employed and  
36 the results revealed notable interactions among SPI and two kinds of polysaccharides,  
37 with electrostatic interactions and hydrophobic interactions being the primary driving  
38 forces. The addition of polysaccharides induced changes in the paste crystalline state  
39 and improved its thermal stability. Scanning electron microscopy (SEM) confirmed a  
40 uniform and compact microstructure due to the incorporation of polysaccharides.  
41 Consequently, the SPI-polysaccharide paste can be considered as novel material for 3D  
42 printing food materials, and the results of this study provides a theoretical foundation  
43 for its application in the field of plant-based food products.

44 **Keywords:** 3D printing; SPI-polysaccharide composite paste; Printability; Rheological  
45 properties

46



## 47 **1. Introduction**

48 3D printing is an efficient and convenient processing technology. Because of its  
49 bottom-up stacking approach, can print and manufacture complex and complicated  
50 structures that traditional processing methods cannot construct (Dankar et al., 2018). It  
51 has broad applications in various fields such as textiles, architecture, healthcare, and  
52 food (Keerthana et al., 2020). 3D food printing is revolutionizing the food processing  
53 industry by creating novel foods with complex shapes, distinctive textures, and  
54 enhanced nutritional value. This allows for personalized nutrition and customized food  
55 design, catering to the specific needs of aging populations, children, and sick patients.  
56 (Phuhongsung et al., 2020). Typically, extrusion, binder jetting, and inkjet printing  
57 technologies have been explored for various 3D food printing applications (Cheng et  
58 al., 2022). Among them, extrusion-based printing technique can be effectively used for  
59 semi-solid pastes such as grain dough, meat puree, and chocolate. The principle of this  
60 technique is to extrude material layer by layer through a nozzle to form a complex 3D  
61 structure (Rowat et al., 2021). Extrusion technology is currently the most widely studied  
62 method for 3D food printing. The emergence of 3D food printing technology helps to  
63 accelerate the development of new food products and become a potential technological  
64 mean to rebuild the conventional food supply chain (Chen et al., 2019).

65 Proteins and polysaccharides are two essential nutrients in the human food system  
66 that can alter their structural and functional properties, such as the ability to form gels  
67 or emulsions, through electrostatic forces and hydrogen bonds (Zhang et al., 2022). Soy  
68 protein isolate (SPI) as a food component has been widely used in 3D printing (Chen  
69 et al., 2019; Phuhongsung et al., 2020). However, SPI exhibits limitations in terms of  
70 printing accuracy and stability, as it is prone to breakage under high shearing forces. To  
71 address this issue, the addition of hydrocolloids or salt ions to SPI has been identified  
72 as a viable solution (Xu et al., 2023). Previous research has demonstrated the  
73 effectiveness of incorporating apricot polysaccharide into SPI, resulting in improved  
74 gelatinization performance and viscoelasticity of the SPI gel (Xu et al., 2023).  
75 Polysaccharide is a kind of natural macromolecular carbohydrate with complex and  
76 huge molecular structure obtained from multiple monosaccharides after dehydration

77 and condensation. Although polysaccharides are widely distributed in nature, not all  
78 polysaccharides are called food polysaccharides. Food polysaccharide refers to the  
79 polysaccharide that is allowed as a food additive in the food processing process to  
80 control the sensory quality, physical properties and improve the nutritional value of  
81 food. Kappa-Carrageenan ( $\kappa$ C), a widely used polysaccharide in the food industry,  
82 exhibits rapid thermal-reversible behavior and can form robust and brittle gel structures  
83 suitable for 3D printing (Liu et al., 2019; Yu et al., 2022). Additionally, alginates are  
84 widely used as 3D printing materials because of their good gelling properties due to  
85 their many carboxyl and hydroxyl groups (Chen et al., 2019). Dick et al. (2020)  
86 investigated the effects of different ink polysaccharides on the rheology and texture of  
87 pig meat and found that the addition of ink polysaccharides improved the printability  
88 and texture of the meat. Studies have reported that SPI mixed with sodium alginate and  
89 gelatin as a 3D printing material improved the hardness and chewiness of 3D printed  
90 geometries, and SPI paste will be a promising material for 3D food printing (Chen et  
91 al., 2019).

92 Currently, 3D printing of food is still in its early stages. Warner et al. (2019)  
93 investigated the 3D printability of a mixture of gelatin and  $\kappa$ C and found that the  
94 addition of  $\kappa$ C resulted in a more stable structure than pure gelatin. Commercially, the  
95 3D printed plant-based beef product “Alt Steak” produced by Israeli startup Redefine  
96 Meat has been sold in over 120 restaurants in Israel, the Netherlands, Germany, and the  
97 United Kingdom. Japanese company “Open Meals” uses 3D printer to print edible  
98 pixelated sushi. Each sushi is composed of edible gel with specific color and taste.  
99 However, to date, there has been limited exploration of 3D printing of SPI-  
100 polysaccharide composite pastes. At the same time, carrageenan and sodium alginate  
101 are extracted from plants, and we consider maintaining a plant-based property in the  
102 material properties of the ink, which can be used as a raw material to support the  
103 production of plant meat in the future. Therefore, this study developed a 3D printable  
104 ink by mixing SPI with  $\kappa$ C and alginate polysaccharides. The structural and functional  
105 properties of the ink were evaluated using rheology, texture analysis, Fourier transform  
106 infrared spectroscopy, and scanning electron microscopy. The study investigated the

107 interactions between protein and polysaccharides and their effects on the printability of  
108 the ink.

## 109 **2. Materials and methods**

### 110 **2.1. Materials**

111 SPI (protein content > 90%) was purchased from Shanghai yuanye Bio-  
112 Technology Co., Ltd (Shanghai, China). The ionic content of SPI was determined by  
113 atomic absorption spectrometry, and its composition was  $0.02\pm 0.01$  wt% of  $\text{Ca}^{2+}$ ,  
114  $0.04\pm 0.00$  wt% of  $\text{K}^+$  and  $0.03\pm 0.01$  wt% of  $\text{Na}^+$ . Kappa-carrageenan ( $\kappa\text{C}$ ) and  $\beta$ -  
115 mercaptoethanol were acquired from Shanghai Aladdin Ltd (Shanghai, China). The  
116 ionic content of  $\kappa\text{C}$  was determined by the same procedure, the contents were  $3.5\pm 0.05$   
117 wt% of  $\text{K}^+$ ,  $1.25\pm 0.01$  wt% of  $\text{Na}^+$ ,  $0.47\pm 0.04$  wt% of  $\text{Ca}^{2+}$ ,  $2.1\pm 0.02$  wt% of  $\text{Cu}^{2+}$ , and  
118  $3.2\pm 0.03$  wt% of  $\text{Zn}^{2+}$ . Sodium alginate (SA), tris (hydroxymethyl)methyl  
119 aminomethane (Tris), glycine, urea, sodium dodecyl sulfate (SDS), and disodium  
120 ethylenediaminetetraacetic acid ( $\text{Na}_2\text{EDTA}$ ) were supplied by Shanghai Macklin  
121 Biochemical Technology Co., Ltd (Shanghai, China). The ionic content of SA was  
122 determined by the same procedure, the contents were  $2.31\pm 0.01$  wt% of  $\text{K}^+$ ,  $11.23\pm 0.02$   
123 wt% of  $\text{Na}^+$ ,  $0.03\pm 0.01$  wt% of  $\text{Fe}^{2+}$ . Bovine serum albumin (BSA) was obtained from  
124 Solarbio Science & Technology Co., Ltd (Beijing, China). All other reagents and  
125 chemicals were of analytical grade.

### 126 **2.2. Food 3D Printing Ink Preparation**

127 The mixtures of SPI with sodium alginate and  $\kappa\text{C}$  were prepared as followed. SA  
128 and  $\kappa\text{C}$  solutions were dissolved in deionized water at three concentrations (1, 1.5, and  
129 2% w/w). Subsequently, mixed the two solutions and added SPI powder to the mixed  
130 solution to achieve a concentration of 15% (w/w) of SPI. Table 1 presents eight  
131 representative ink formulations with a gradient cross rise in two kinds of  
132 polysaccharides, the original ratio of SPI and polysaccharide was chosen according to  
133 Hu et al., (2017). The resulting mixed solution was homogenized using a homogenizer  
134 (HFJ-25, Hengao, Tianjin, China) at 500 rpm for 5 min until a stable suspension was  
135 formed. All samples had pH values within  $6.8\pm 0.3$ , which were above the isoelectric  
136 point of soy proteins (Lu et al., 2023). The resulting paste-like mixture was kept in a

137 water bath at 80 °C for 30 min and cooled at room temperature for 15 min. The samples  
138 were stored overnight in the refrigerator and heated to 25 °C before 3D printing.

### 139 **2.3. 3D printing process**

140 The samples were printed using an extrusion-based food 3D printer (FOODBOT-  
141 S2, Hangzhou Shiyin Technology Co., Ltd., Hangzhou, China). The prepared ink filled  
142 into the 3D printing nozzle syringe, and the printer extruded the material by controlling  
143 the movement of the plunger in accordance with the preset 3D mesh model (18.6 mm  
144 × 18.6 mm × 8 mm). The main printing parameters were set as follows: nozzle diameter  
145 of 0.84 mm, nozzle printing rate of 25 mm/s, layer height of 0.8 mm, printing  
146 temperature of 25 °C, extruded rate of 24 mm<sup>3</sup>/s and infill density of 100% printing. To  
147 ensure maximum aseptic operation, all containers, nozzle syringes, and nozzle tips were  
148 sterilized at 121 °C in an autoclave for 15 min. The printing environment was  
149 disinfected with 75% ethanol and UV radiation, and all operations were conducted in a  
150 laminar flow hood.

### 151 **2.4. Rheological Properties of Printing Inks**

152 The rheological properties of the printing inks were measured using a hybrid  
153 rheometer (Discovery HR-2, DHR, TA Instruments, New Castle, DE, USA). A parallel  
154 plate with a diameter of 40 mm was used, with a measurement gap of 1 mm. When  
155 loading the sample, any excess material on the edges was removed with a scraper, and  
156 a thin layer of silicone oil was applied to the edges to prevent moisture evaporation.

157 The configuration was set as follow: soak time was 180 s, 7 points were detected per  
158 decade, the interval between the points was 10 s, and the steady state sensing in DHR  
159 rheometer was applied to make sure the equilibrium time was 30 s for each point.

#### 160 2.4.1. Flow sweep test

161 The flow sweep test was measured at shear rates ranging from 0.01 to 100 s<sup>-1</sup>, with  
162 a temperature set at 25 °C, to obtain the viscosity  $\eta$  as a function of shear rate and  
163 investigate the shear-thinning behavior of the sample.

#### 164 2.4.2. Amplitude sweep test

165 The linear viscoelastic region (LVR) was determined through an amplitude sweep  
166 test. The test was conducted within a strain range of 0.01-100% at a constant frequency

167 of 10 rad/s, and the experiment was performed at a temperature of 25 °C.

#### 168 2.4.3. Frequency sweep test

169 The dynamic viscoelastic properties of the sample were evaluated through a  
170 frequency sweep test. The oscillation was performed within a frequency range of 0.1-  
171 100 rad/s, with a constant strain of 0.5% (within the LVR), and the experiment was  
172 conducted at a temperature of 25 °C. The storage modulus ( $G'$ ) loss modulus ( $G''$ ) and  
173 loss tangent ( $\tan \delta = G''/G'$ ) were recorded. All tests were repeated in triplicates, and  
174 experimental curves were plotted accordingly.

### 175 **2.5. Texture analysis**

176 Texture characteristics of the 3D printed samples were measured using a texture  
177 analyzer (TA. XT Plus, Stable Micro Systems, UK) equipped with a P36/R probe. The  
178 samples were tested as  $18.6 \times 18.6 \times 8$  mm grid-like architectures printed products for  
179 two compression cycles. The testing parameters were set as follows: the pre-test, test,  
180 and post-test speeds were all set at 1 mm/s, the trigger force was 10 g, the compression  
181 ratio was 50%, the time interval between the two compressions was 5 s, and the  
182 measurement temperature was 25 °C. Each sample was tested three times for  
183 repeatability.

### 184 **2.6. Field-emission Scanning electron microscopy (FE-SEM) analysis**

185 The microstructure of the paste was observed using a scanning electron  
186 microscope. The paste samples were freeze-dried, and the surface of the samples was  
187 sputter-coated with gold under vacuum conditions. The samples were then observed  
188 under the field-emission scanning electron microscope (S-4700, HITACHI, Japan) at  
189 different magnifications (1000 $\times$ , 2500 $\times$  and 5000 $\times$ ) with an accelerating voltage of 15  
190 kV.

### 191 **2.7. Molecular interaction analysis**

192 To elucidate the paste cross-linking mechanism resulting from SPI-polysaccharide  
193 interactions, the protein solubility of the composite paste in five different solvents was  
194 measured to assess the intermolecular forces within the paste according to the method  
195 described by Wang et al. (2018). The paste sample (protein concentration 2 mg/mL)  
196 was dissolved in different solvents. The solvent S1 was distilled water, the solvent S2  
197 consisted of 0.086 M Tris-0.09 M Glycine-4 mM- $\text{Na}_2\text{EDTA}$  (pH = 8.0), the solvent S3

198 was 0.5% SDS, the solvent S4 was 8 M Urea, and solvent S5 was 2%  $\beta$ -  
199 mercaptoethanol. The mixture samples ( $2.00 \pm 0.02$  g) were mixed with 10 mL buffer,  
200 the resulting solution was incubated in a water bath at 25 °C for 20 min, and then  
201 centrifuged at 5000 rpm for 10 min. The protein supernatant was determined using  
202 bovine serum albumin as standard with ultraviolet spectrophotometry at 595 nm  
203 (GENESYS 150, Thermo Fisher Scientific Inc, Waltham, MA, USA). The protein  
204 solubility is expressed as the percentage of protein content of the supernatant in the  
205 total protein content.

206 Tris-glycine buffer disrupts electrostatic interactions, SDS disrupts hydrophobic  
207 interactions by disrupting the medium between the hydrophobic and hydrophilic  
208 regions, urea disrupts hydrogen bonding in the protein by dehydrating the protein  
209 molecules' side chains and backbone, and  $\beta$ -mercaptoethanol reduces protein disulfide  
210 bonds by competing with thiol groups (Qin et al., 2017; Wang et al., 2021). According  
211 to this method, the differences in protein solubility in S2 (Tris-Gly buffer)-S1 (distilled  
212 water), S3 (0.5% SDS)-S2, S4 (urea)-S3, and S5 ( $\beta$ -mercaptoethanol)-S4 were used as  
213 indications of the electrostatic interactions, hydrophobic interactions, hydrogen  
214 bonding interactions, and disulfide bonding interactions, respectively.

## 215 **2.8. Fourier transform infrared spectroscopy (FTIR)**

216 The samples were mixed with dry KBr at a ratio of 1:100 (w/w). Then the mixed  
217 powder was ground in the quartz mortar and pressed into a tablet. FTIR spectra (Nicolet  
218 6700, Thermo Fisher Scientific Inc, USA) was performed at 25 °C in the range of 4000  
219 to 500  $\text{cm}^{-1}$ . The resolution remains at 1  $\text{cm}^{-1}$ .

## 220 **2.9. X-ray diffraction (XRD)**

221 The X-ray diffraction (XRD) patterns of Cu  $K\alpha$  radiation ( $\lambda = 0.1541$  nm) were  
222 recorded by powder diffractometer (Empyrean, Malvern PANalytical, NL). The  
223 working voltage was 40 kV and the working current was 40 mA. The patterns were  
224 collected with a  $2\theta$  range from 5° to 60° at a step of 0.0262°.

## 225 **2.10. Differential scanning calorimetry (DSC)**

226 Differential scanning calorimeter (DSC 8000, PE Instruments Inc., USA) was used  
227 to evaluate the thermal properties of the composite pastes. The thermal analysis was  
228 conducted according to the method described by Wang et al. (2018) with slight

229 modifications. Briefly, the sample mixture (10 mg) was accurately weighed and loaded  
230 into a DSC pan. Prior to analysis, the DSC pan was sealed and maintained at the initial  
231 scanning temperature of 20 °C. The sample was analyzed from 20 °C to 180 °C at a  
232 scanning rate of 10 °C/min, and the temperature of each endothermic or exothermic  
233 peak was determined. A sealed empty pan was used as a blank reference.

#### 234 **2.11. Statistical Analysis**

235 The results are shown as the mean  $\pm$  standard deviation (SD) of triplicate  
236 experiments. SPSS software (SPSS26; IBM SPSS Statistics, Chicago, IL, USA) was  
237 used for one-way ANOVA and Duncan tests at 95% confidence level for data analysis.  
238 All graphs in the experiment were drawn using Origin software (2020b, OriginLab, MA,  
239 USA).

### 240 **3. Results and discussion**

#### 241 **3.1. Printability of composite inks**

242 Soy protein isolate (SPI), as the main material for printing, cannot be directly  
243 printed due to its high flowability and relatively weak cross-linking in water (Shi et al.,  
244 2023). Sodium alginate (SA) is a commonly used hydrophilic colloid with good binding  
245 capacity for proteins (Chen et al., 2022). Kappa-carrageenan ( $\kappa$ C) is a soluble  
246 polysaccharide with high solubility and gelation properties (Bi et al., 2018). The  
247 addition of SA and  $\kappa$ C to the SPI paste could improve the stability and printability of  
248 food ink. In order to screen ink formulations with optimal printing performance, this  
249 study prepared inks with different polysaccharide concentration ratios and examined  
250 their printability. The 3D printing results of different ink formulations are shown in  
251 Figure 1. The successful implementation of food 3D printing requires two conditions:  
252 the material in the extrusion process has the right viscosity to flow through the nozzle  
253 and adhere layer by layer, and the paste structure after extrusion has sufficient  
254 mechanical strength to ensure its formability (Osi et al., 2021). For preliminary tests of  
255 printability, the ink should be continuously extruded from the nozzle as a uniform  
256 filament, and the printed product should show the effect of perfect detail, smooth  
257 surface, and structural stability, rather than being unable to form or causing blockages  
258 and tangled (Figure 1).

259 In the blue irregular region, SPI paste collapsed immediately after continuous and  
260 uniform extrusion from the nozzle, showing the characteristics of some liquid-like  
261 fluids, and the 3D printing molding effect was poor, unable to form and support the  
262 designed three-dimensional grid structure. This could be due to the insufficient gelation  
263 capacity and the low viscosity, and the ink forms irregular printing filaments at the  
264 nozzle, resulting in the inability to form a homogeneous structure (Jiang et al., 2019).  
265 In the orange region, the ink formulation aggregates significantly at the nozzle when  
266 subjected to stress, which may be due to the inappropriate surface tension of the ink,  
267 leading to a tendency for rapid recovery and aggregation of the system after extrusion  
268 (Fu et al., 2021). In the gray region, as the polysaccharide content in the ink gradually  
269 increased, the viscosity of the sample increased, resulting in poor flowability of the ink  
270 material and the nozzle clogging (Fu et al., 2021). In the green printable region, ink  
271 components were able to be continuously and smoothly extruded. The ink can be  
272 printed when the concentration of SA changed from 1 wt% to 2.5 wt% with a  $\kappa$ C  
273 concentration of 1 wt%, or when the concentration of SA changed from 1 wt% to 2.5  
274 wt% with a  $\kappa$ C concentration of 1.5 wt%. However, some ink formulations, such as  
275 S15A1K1.5, S15A1K1, and S15A1.5K1.5, print three-dimensional mesh structure  
276 filaments that are uneven and have broken structures. In contrast, the 3D mesh  
277 structures printed by the SPI15A1.5K1 and S15A2K1.5 formulations had good self-  
278 supporting properties, smooth surfaces and showed uniform layer thickness. Therefore,  
279 to investigate the reasons for the differences in printability among the different  
280 formulations and the effects of different polysaccharide concentrations on ink  
281 printability, five representative ink formulations (irregular: SPI 15; tangled: S15A2K2;  
282 clogged: S15A1.5K2; printable: S15A1.5K1.5 and S15A2K1.5) were selected for in-  
283 depth study.

### 284 **3.2. Rheological properties**

285 The rheological properties of the inks directly influence their printability for 3D  
286 printing based on extrusion samples, with viscosity being a key indicator of extrusion  
287 capability. As the polysaccharide concentration increased, the viscosity of the paste also  
288 increased. As shown in Figure 2a, all samples exhibited a shear rate-dependent



289 reduction in viscosity, demonstrating typical shear-thinning behavior, which is  
290 generally favorable for extrusion-based 3D printing (Dankar et al., 2018). This may be  
291 attributed to the disruption of polymer entanglement during the shear process, causing  
292 polymer chains to stretch and align parallel to the shear direction (Hou et al., 2022;  
293 Zhang et al., 2022). The stress and shear rate applied to the fluid during extrusion  
294 depends on the fluid and extrusion conditions, and these conditions can vary greatly in  
295 different applications (Lopez Hernandez et al., 2021). The apparent shear rate of a  
296 steady-state pipe can be derived by extruding the corresponding flow rate and tube  
297 radius (Xie et al., 2020).

$$\dot{\gamma} = \frac{3.34Q}{\pi R^3}$$

299 Where Q is the volumetric flow rate and R is the tube inner radius. According to  
300 the volumetric flow (24 mm<sup>3</sup>/s) and inner radius (0.42 mm), the calculated shear rate is  
301 about 344.4 s<sup>-1</sup>. This result is fitted with the report of (Lopez Hernandez et al., 2021).  
302 The viscosity of the SPI 15 ink at the high shear rate (100 s<sup>-1</sup>) is much lower than the  
303 viscosity at the low shear rate of 0.1 s<sup>-1</sup>. This is because at very low shear rates, the  
304 stable aggregates formed by the glycine subunits of SPI are stabilized by disulfide  
305 bonds, and the side chains are entangled with adjacent chains, resulting in high viscosity.  
306 As the shear rate increases, the unraveling of the side chains and the arrangement of the  
307 molecules provide less shear resistance, thereby reducing the viscosity (Dorishetty et  
308 al., 2019). Some studies have added *Lycium barbarum* polysaccharide to soy protein  
309 isolate, and the results show that the apparent viscosity increases significantly with the  
310 addition of polysaccharide in a concentration-dependent manner (Xing et al., 2024). In  
311 our experiment, the slope of the viscosity versus shear rate was not smooth, which is  
312 also observed in other studies (Lu et al., 2023, Liu et al., 2022, Yu et al., 2022 and Chen  
313 et al., 2019). This may be because the sample has not yet reached equilibrium,  
314 especially at a high shear rate. In the high shear rate region (i.e. from 1 to 100·s<sup>-1</sup>),  
315 molecules or particles within the fluid may not have enough time to rearrange and adjust  
316 their positions. Besides, it is speculated that the turbulent flow happened or the sample  
317 slipped out at the higher shear rate. At the shear rate of 1 s<sup>-1</sup>, the viscosity of S15A1.5K2,

318 S15A1.5K1.5, and SPI 15 food inks was  $1.22 \times 10^3$  Pa s,  $9.81 \times 10^2$  Pa s and  $9.06 \times 10^2$   
319 Pa s, respectively. The viscosity range of S15A1.5K1.5 ink is similar to the results of  
320 (Dick et al., 2020). Among them, the viscosity of more than 1200 Pa s will lead to  
321 entanglement of ink molecules, insufficient pressure in the cylinder, resulting in nozzle  
322 clogging. If the viscosity was less than 900 Pa s, although it is easy to squeeze, the  
323 fluidity was too large to provide sufficient viscoelasticity and stability. Therefore, an  
324 ideal viscosity for the ink should be known during the extrusion process, which allows  
325 the ink to pass through the narrow diameter of the printing nozzle while maintaining its  
326 deposition structure and shape.

327 The nonlinear rheological behavior of samples can be monitored by large  
328 amplitude oscillatory shear test. The linear viscoelastic range (LVER) and critical strain  
329 of ink formulations with different compositions are shown in Figure 2b. LVER is a  
330 range where the storage modulus  $G'$  and loss modulus  $G''$  remain relatively constant as  
331 shear intensity increases. The  $G'$  is a measure of the elastic solid-like behavior, that is,  
332 the resistance to elastic deformation of the sample, while the  $G''$  is the viscous response,  
333 which is the ratio of stress to strain under vibration conditions (Yang et al., 2018). When  
334  $G'$  is greater than  $G''$ , the material behaves primarily as a solid, and vice versa for a  
335 viscous fluid, exhibited a slight frequency dependence (Pan et al., 2022). Within this  
336 range, the material undergoes elastic deformation and can recover to its original state  
337 after the stress is removed. As shown in Figure 2b, when the applied strain is less than  
338 1%, different inks have similar LVERs, maintaining stable values. As the strain  
339 increased, the value of  $G'$  started to decrease, indicating the critical point of the elastic  
340 properties of the samples about 1-2%. The weak gelation ability of SPI at room  
341 temperature results in disordered polymer chains and poor stability, leading to a liquid-  
342 like state after extrusion (Fu et al., 2021). There is a clear positive correlation between  
343  $G'$  and the polysaccharide concentration in the ink, where  $G'$  increases sequentially for  
344 inks S15A1.5K1.5 and S15A2K1.5, indicating higher structural strength, which aligns  
345 with the results of viscosity tests. Among them, ink S15A1.5K1.5 showed the lowest  
346  $G'$  value in all the ink formulations with added polysaccharides. This may be due to a  
347 low polysaccharide concentration, which prevents the ink system from forming a stable

348 paste structure.

349 Frequency sweep tests can provide information about the internal structure of the  
350 printing inks (Hou et al., 2022). As shown in Figure 2c and, 2d, the  $G'$  and  $G''$  of all  
351 pastes exhibit a steady upward trend, with  $G'$  being superior to  $G''$ , indicating that the  
352 ink primarily possesses elastic and solid characteristics. Higher polysaccharide content  
353 in the paste could lead to higher cross-linking density in the SPI-polysaccharide system,  
354 resulting in stronger viscoelasticity and mechanical strength of the paste, as well as  
355 good self-supporting properties and resistance to shape deformation (Pan et al., 2022).  
356 This might be attributed to the formation of a denser and more robust intermolecular  
357 cross-linking structure through molecular interactions between polysaccharides and SPI,  
358 exhibiting stronger solid characteristics. The loss tangent,  $\tan \delta = G''/G'$ , is used as a  
359 characteristic parameter to account for the different viscoelastic behaviors. A  $\tan \delta$  value  
360 less than 1 indicates a predominantly elastic property, while a  $\tan \delta$  value greater than  
361 1 indicates a predominantly viscous property (Yang et al., 2018). The  $\tan \delta$  values of all  
362 samples were less than 1, which indicated that the addition of polysaccharide enhanced  
363 the gel properties of the mixture and made it form a rigid gel structure. The addition of  
364  $\kappa$ C can induce electrostatic interactions with proteins, leading to the predominant solid-  
365 like behavior of these fluids (Lu et al., 2022). Among them, with a constant  $\kappa$ C  
366 concentration of 1.5 wt% and angular frequency of 10 rad/s, when the SA concentration  
367 was increased from 1.5 wt% to 2 wt%,  $G'$  and  $G''$  increase from 24970.3 Pa and 4042.66  
368 Pa to 30697.3 Pa and 4845.1 Pa, respectively.  $G'$  increased by 22.94% and  $G''$  increased  
369 by 19.85%. However, with a constant SA concentration of 2 wt% and angular frequency  
370 of 10 rad/s, when the  $\kappa$ C concentration was increased from 1.5 wt% to 2 wt%,  $G'$  and  
371  $G''$  increased from 30697.3 Pa and 4845.1 Pa to 32279.5 Pa and 5002.04 Pa,  
372 respectively.  $G'$  increased by 51.54% and  $G''$  increased by 3.24%. These results suggest  
373 that polysaccharides participate in the formation of the paste structure, and different  
374 types of polysaccharides lead to different structures. It has been reported that with the  
375 increase of  $\kappa$ C concentration, the interaction between  $\kappa$ C-proteins may gradually  
376 decrease and be replaced by the  $\kappa$ C- $\kappa$ C interaction (Lu et al., 2022). Some studies have  
377 shown that the gel strength of calcium alginate gels is relatively weak (Fernández Farrés

378 et al., 2013). Compared to SA,  $\kappa$ C has a greater impact on the viscoelasticity of the  
379 paste, and at higher concentrations, the paste structure becomes more rigid. However,  
380 excessively high viscoelasticity of the paste may have adverse effects on 3D printing,  
381 as the generated high shear forces can hinder ink extrusion or cause fracture after  
382 extrusion. On the other hand, the higher the concentration of SA, the smoother the  
383 extrusion process, the flatter the filaments, making it more suitable for printing, which  
384 will be confirmed by the results of texture analysis.

385 In general, the printability of soybean protein-polysaccharide composite paste is  
386 aimed at printing out self-supporting, flat sample and smooth surface. To ensure  
387 printability, the ink should exhibit contrasting properties. Initially, it must be fluid  
388 enough to be pumped and extruded, yet it should also stiffen rapidly after deposition to  
389 enable construction. Therefore, the amount of different polysaccharides added is  
390 particularly important, and the range of proper mechanical properties should be  
391 between 900 Pa s and 1200 Pa s.

### 392 **3.3. Texture profile analysis (TPA)**

393 Texture profile analysis can detect several texture parameters, including hardness,  
394 springiness, adhesiveness, and others (Yuan et al., 2016). As shown in Table 2, paste  
395 samples without polysaccharides cannot determine the TPA parameters due to their  
396 extremely weak gel properties, highlighting the importance of incorporating  
397 polysaccharides in the ink for the production of self-supporting 3D printed food  
398 products. With an increase in polysaccharide concentration, the springiness,  
399 cohesiveness, gumminess, and resilience of the printed samples all significantly  
400 decreased (Table 2). This could be attributed to the changes in the repulsive forces  
401 between polysaccharide charges that affect molecular aggregation. Both carrageenan  
402 and sodium alginate are anionic polysaccharides that can form complexes with the  
403 amino group ( $-\text{NH}_3^+$ ) of the protein through electrostatic interactions, thereby altering  
404 the printing characteristics of the paste (Huang et al., 2021). Sulfated polysaccharides  
405 such as carrageenan have the ability to interact with proteins above their isoelectric  
406 point.  $\kappa$ C contains a sulfate group ( $-\text{OSO}_3^-$ ) that can be bonded to the amino group ( $-\text{NH}_3^+$ )  
407 of the protein by electrostatic interaction. The electrostatic interaction between

408 sulfate group ( $-OSO_3^-$ ) and protein is stronger than that between carboxyl group ( $-$   
409  $COO^-$ ) and protein (Huang et al., 2021). Furthermore, SPI possesses a globular  
410 structure, and the folded SPI structure can stretch and expose internal functional group  
411 structures upon heating, facilitating cross-linking with polysaccharide ions.

412 Hardness refers to the force required to compress a sample and can mimic the  
413 chewing behavior of the oral cavity on solids or semi-solids, serving as an indicator of  
414 the structural stability of 3D printed food. Compared to pastes with the same  
415 concentration of SA, the addition of  $\kappa$ C significantly affects the properties, with an  
416 increase in sample hardness and a decrease in adhesiveness and springiness. When the  
417  $\kappa$ C concentration reached 2 wt%, high shear forces occur during the extrusion process,  
418 leading to filament breakage at the nozzle, resulting in poor shape fidelity and  
419 significant discrepancies between the printed samples and the intended model, making  
420 it also impossible to determine the TPA parameters. In contrast, the addition of SA, with  
421 the same concentration of  $\kappa$ C, yields opposite results. This may be attributed to the  
422 disruption of the molecular chain structure of the samples during preparation, where  
423 longer polysaccharide chains exhibit stronger gelation properties with proteins  
424 (Monteiro & Lopes-da-Silva, 2017). The springiness of the samples refers to the  
425 physical reboundness after deformation, which can resist to external forces. At higher  
426 concentrations, polysaccharides dominate the springiness of the mixture, leading to a  
427 weakening of samples structure, as also reflected in the aforementioned rheological  
428 analysis (Warnakulasuriya & Nickerson, 2018). This is contrary to the experimental  
429 conclusion of (Xu et al., 2023). This result may be due to the fact that the sample has a  
430 hollow structure, and when the extrusion degree is 50%, the sample is forced to extrude  
431 from all sides to widen, resulting in a decrease in data presentation. Cohesiveness refers  
432 to the adhesive forces within the sample particles, reflecting the magnitude of  
433 intermolecular binding forces that allow the sample to maintain its integrity even under  
434 pressure. The data indicates that there is only a minor difference in cohesiveness  
435 between the samples, and the low concentration of polysaccharide improved the  
436 structural properties of the paste. When the concentration of polysaccharide was too  
437 high ( $>1.5\%$ , wt %), the Cohesiveness and resilience of the sample decreased, which

438 was consistent with the results of (Xu et al.,2023). This may be attributed to changes in  
439 the repulsive forces between the negative charges in the polysaccharide, which affect  
440 molecular aggregation and thus alter the texture properties of the sample (Xu et  
441 al.,2023). The above results suggested a positive correlation between paste textural  
442 properties and polysaccharide concentration. Considering that plant-based foods are  
443 extruded with current inks as the main material, the protein paste having the  
444 formulations S15A1.5K1.5 exhibits optimal texture properties and is most suitable for  
445 3D printing. In fact, the texture of plant-based foods largely depends on their structure,  
446 mechanical properties, and surface characteristics (Shi et al., 2023).

### 447 **3.4. Microstructure**

448 Figure 3 illustrates the microstructures of different ink formulations at various  
449 magnifications (1000×, 2500×, and 5000×). All food inks exhibit a heterogeneous  
450 microstructure composed of protein-polysaccharide particles forming aggregates. The  
451 structure of soybean isolate protein (SPI) has a rough surface because it is a paste at  
452 room temperature. However, the structural support provided by these concentrations is  
453 still insufficient to meet the required criteria, which may be the reason for the inability  
454 to achieve proper printability (Figure 1). Therefore, an appropriate polysaccharide  
455 concentration needs to be added to achieve gelation. Different ink formulations have  
456 different densification and orientation of the cavity, which is especially obvious after  
457 the addition of polysaccharides. Sodium alginate (SA) and kappa carrageenan ( $\kappa$ C) bind  
458 strongly to protein particles, forming small fragmented block formations. SA,  $\kappa$ C, and  
459 SPI particles form non-covalent bonds due to their hydrophilic nature and abundance  
460 of carboxyl and hydroxyl groups, which promote hydrophobic interactions and  
461 hydrogen bonding (Huang et al., 2021; Wei et al., 2023). As the  $\kappa$ C content increased,  
462 the cross-linking structure with SPI-polysaccharide particles became rougher, resulting  
463 in a more disordered and chaotic ink system, forming large aggregates. This could be  
464 an increased interaction between  $\kappa$ C and  $\kappa$ C (Lu et al., 2022). This may explain why  
465 ink S15A1.5K1.5 was able to be extruded from the nozzle while ink S15A1.5K2  
466 became clogged and ink S15A2K2 agglomerated (Figure 1). SA more ink formula  
467 surface is more compact and smoother, which is consistent with (Chen et al., 2022).

468 Overall, ink S15A1.5K1.5 has the best network density and can resist structural changes  
469 under high shear stress during the extrusion process.

### 470 **3.5. Molecular interaction force analysis**

471 As shown in Figure 4a, the solubility of the composite paste was significantly  
472 enhanced, indicating the presence of hydrophobic interactions, electrostatic interactions,  
473 disulfide bonds, and hydrogen bonds within the protein paste structure. The interaction  
474 forces between protein molecules and polysaccharides followed the order of  
475 hydrophobic interactions > electrostatic interactions > disulfide bond interactions >  
476 hydrogen bond interactions. This observation was consistent with the previously  
477 reported results (Lu et al., 2023). A slight decrease in electrostatic interactions was  
478 observed in the composite paste. This result can be attributed to the influence of  
479 negatively charged groups on the protein and polysaccharide surfaces, which are  
480 affected by the electrostatic binding under high polysaccharide concentrations, leading  
481 to conformational changes in the protein and exposing more hydrophobic groups (Wang  
482 et al., 2021). When electrostatic repulsion further increases, the distance between  
483 molecules becomes longer, making it difficult to detect the signal of electrostatic  
484 repulsion, resulting in decreased solubility. Compared to pure SPI paste, the addition of  
485 polysaccharides significantly increased hydrophobic interactions and hydrogen bond  
486 interactions in the paste ( $p < 0.05$ ). This finding indicates that the addition of  
487 polysaccharides greatly enhances hydrophobic interactions and hydrogen bond  
488 interactions, playing a crucial role in the formation and maintenance of the SPI-  
489 polysaccharide paste system. This phenomenon may be attributed to the exposure of  
490 chemical groups within the protein matrix after thermal treatment. SA and  $\kappa$ C are  
491 anionic acidic polysaccharides rich in carboxyl and hydroxyl groups. Although there is  
492 an intrinsic repulsion between them, but other forces can maintain the state without  
493 causing the system to separate, especially at high concentrations. Studies have shown  
494 that there is an electrostatic interaction between SPI and  $\kappa$ C, where the negatively  
495 charged sulfate groups on  $\kappa$ C can interact with the positively charged amino groups ( $-$   
496  $\text{OSO}_3^- \text{NH}_3^+$ ) on proteins, even at pH above the isoelectric point (Huang et al., 2021; Lu  
497 et al., 2023). Additionally, SPI and polysaccharides can absorb moisture, promoting

498 further aggregation of proteins and overcoming the biocompatibility and inherent  
499 repulsion between SPI and polysaccharides, ultimately forming a paste cross-linking  
500 structure (Cao et al., 2022; Min et al., 2022).

### 501 **3.6. Composite paste forming mechanism**

502 Based on the intermolecular interactions and rheological properties discussed  
503 above, this study presents a plausible hypothesis regarding the gelation mechanism of  
504 SPI-polysaccharide (Figure 5). The experimental results revealed that hydrophobic  
505 interactions and electrostatic interactions are the predominant forces in the composite  
506 paste. Heating facilitates the exposure of hydrophobic groups in the protein, while the  
507 polysaccharides SA and  $\kappa$ C contain numerous carboxyl and hydroxyl groups,  
508 enhancing hydrophobic interactions and hydrogen bonding with SPI particles (Huang  
509 et al., 2021; Wei et al., 2023). The presence of a substantial number of hydrophobic  
510 groups leads to protein crosslinking and aggregation.  $\kappa$ C, on average, contains an  
511 anionic sulfate group ( $-\text{OSO}_3^-$ ) per disaccharide repeating unit, which can engage in  
512 electrostatic interactions with cationic groups ( $-\text{NH}_3^+$ ) of the protein. The electrostatic  
513 interaction between sulfate group ( $-\text{OSO}_3^-$ ) and protein is stronger than that between  
514 carboxyl group ( $-\text{COO}^-$ ) and protein (Huang et al., 2021; Lu et al., 2023). In the SPI  
515 system, the addition of  $\kappa$ C forms protein-polysaccharide interactions, establishing a  
516 balance between protein-protein and protein-polysaccharide interactions. This is  
517 consistent with the fact that in all oscillation studies, the composite gel is visually  
518 homogeneous and there is no phase separation. When the polysaccharide concentration  
519 exceeds 1.5 wt%, the self-aggregation of polysaccharides hinders SPI cross-linking,  
520 reducing its interaction with SPI. As the  $\kappa$ C content increases, the  $\kappa$ C- $\kappa$ C interaction  
521 increases and the  $\kappa$ C-protein interaction decreases, resulting in a stronger gel that is  
522 unable to extrude from the nozzle. The molecular structure of SA is rich in hydroxyl  
523 and carboxylic groups, and it is easy to form intermolecular hydrogen bonds with the  
524 functional groups of other polymers (Wei et al., 2023). However, it has been reported  
525 that SA alone exhibits inadequate mechanical properties in soft tissue engineering  
526 applications, failing to provide sufficient mechanical support under stress conditions,  
527 which aligns with our experimental results (Wei et al., 2023). But when the addition



528 amount of SA exceeds a certain critical value (2.5 wt%), it forms a gel with soybean  
529 protein, yielding improved printing performance. Some studies have successfully  
530 prepared alginate/carrageenan composite hydrogels with different concentrations of  
531 carrageenan, and bio-printed the alginate/carrageenan scaffold, the results show that the  
532 gel is suitable for using bioprinting to manufacture 3D hydrogel scaffold (Kim et al.,  
533 2019).

### 534 **3.7. Fourier transform infrared (FTIR) analysis**

535 In order to investigate the changes in chemical bonds in the inks, a comparison of  
536 their FTIR spectra was conducted, as shown in Figure 4b. The FTIR spectra of the paste  
537 samples did not exhibit any new peaks upon the addition of polysaccharides, and there  
538 were no new functional groups observed with varying ratios of SA and  $\kappa$ C, indicating  
539 that the covalent interactions were minor. This is consistent with the research results of  
540 (Xing et al., 2024). All inks displayed a strong and broad absorption peak in the range  
541 of 3200-3600  $\text{cm}^{-1}$ , which is typically associated with the stretching of O-H bonds and  
542 partially formed hydrogen bonds (Yu et al., 2022). Compared to the SPI 15, the O-H  
543 stretching bands of the ink with added polysaccharides shifted to lower wavenumbers.  
544 For instance, in the inks S15A2K1.5 and S15A2K2, the band of the SPI 15 ink at 3273  
545  $\text{cm}^{-1}$  shifted to 3282  $\text{cm}^{-1}$  and 3286  $\text{cm}^{-1}$ , respectively, suggesting stronger hydrogen  
546 bonding and increased molecular interactions in the S15A2K2 ink (Min et al., 2022).  
547 In fact, hydrogen bonding can form connections between proteins and polysaccharides,  
548 enhancing cross-linking strength, which is crucial for controlling the printability of  
549 different biomacromolecules in food inks (Warnakulasuriya & Nickerson, 2018). In all  
550 mixed pastes, weak peaks were also observed in the range of 2998-3018  $\text{cm}^{-1}$ , which  
551 could be attributed to the stretching vibrations of C-H bonds (Min et al., 2022).  
552 Furthermore, all inks exhibited a peak at approximately 1633  $\text{cm}^{-1}$ , which is related to  
553 the asymmetric and symmetric stretching of carboxylate groups ( $\text{COO}^-$ ) and has been  
554 suggested to be caused by the bending vibrations of water molecules (1650  $\text{cm}^{-1}$ ) (Hu  
555 et al., 2017; Shi et al., 2023). It is noteworthy that when different polysaccharides were  
556 added to SPI, all inks showed absorption peaks in the range of 1515-1531  $\text{cm}^{-1}$ , with  
557 the bands shifting from higher to lower wavenumbers. These peaks are associated with

558 the amide I (1600–1700  $\text{cm}^{-1}$ ) and amide II (1500–1600  $\text{cm}^{-1}$ ) regions in the FT-IR  
559 spectra. The amide regions primarily represent the stretching vibrations of C=O, the  
560 bending vibrations of N-H, and the stretching vibrations of C-N, which are reliable  
561 indicators of protein secondary structure conformation (Carbonaro & Nucara, 2010).  
562 The shift of bands from higher to lower wavenumbers indicates a decrease in the  
563 content of  $\alpha$ -helical structures and an increase in the proportion of  $\beta$ -sheet structures in  
564 the paste (Liu et al., 2022). The results demonstrate that the combination of  
565 polysaccharides and SPI significantly alters the spatial structure of the paste.

### 566 **3.8. X-ray diffraction (XRD) analysis**

567 The X-ray diffraction patterns can be used to determine the amorphous or  
568 crystalline nature of molecules (Dong et al., 2021). As shown in Figure 4c, SPI exhibits  
569 two sharp peaks at diffraction angles of  $9^\circ$  and  $19^\circ$ , indicating its crystalline structure.  
570 Its primary structure is believed to consist of an  $\alpha$ -helical and  $\beta$ -sheet structure (Chen  
571 et al., 2013; Dong et al., 2021). In contrast, the addition of polysaccharides to the  
572 composite paste results in two relatively flat peaks around  $9^\circ$ , suggesting a significant  
573 reduction in the  $\alpha$ -helical structure and a presumed transition to the  $\beta$ -sheet structure.  
574 This also indicates the formation of amorphous complexes between SPI and  
575 polysaccharide molecules. Such amorphous nature is directly related to the  
576 hydrophilicity and hygroscopicity of the polymer (Wang et al., 2018). Composite pastes  
577 prepared with formulations S15A1.5K1.5 and S15A1.5K2 exhibit broad diffraction  
578 peaks at approximately  $19^\circ$ , while formulations S15A2K1.5 and S15A2K2 show  
579 increased peak intensities and sharpened peaks. These structural changes suggested a  
580 corresponding interplay between the  $\alpha$ -helical and  $\beta$ -sheet structures, with an increase  
581 in SA concentration potentially leading to the transformation of the  $\alpha$ -helical structure  
582 to the  $\beta$ -sheet structure in the paste samples (Pan et al., 2022). It has been found that  $\beta$ -  
583 sheet structures have greater potential energy compared to  $\alpha$ -helices, which can explain  
584 why the composite paste exhibits a more stable structure than the SPI paste (Ding et al.,  
585 2003). When the SA concentration is 1.5 wt%, the peak intensity of the composite paste  
586 significantly weakens. This indicates good compatibility between the two polymers (Y.  
587 Dong et al., 2021). At an SA concentration of 2 wt%, the peak intensity increases and

588 becomes sharper around 19°, indicating the formation of amorphous complexes and  
589 intermolecular interactions. These results demonstrated that the presence of κC and SA  
590 influences the crystallinity of the SPI paste system making it promising for 3D printing.

### 591 **3.9. Differential scanning calorimetry (DSC) analysis**

592 The properties and structure of most substances vary with temperature, and  
593 therefore, the stability of the sample structure can be reflected by the changes in  
594 endothermic and exothermic behavior in the DSC spectra. The enthalpy change and  
595 transition temperature can be determined based on the temperature and peak area  
596 corresponding to the largest peak in the graph (Hou et al., 2022). The influence of  
597 different polysaccharide concentrations on the thermal stability of the composite paste  
598 is shown in Figure 4d. The addition of the polysaccharides significantly affects the  
599 thermal transition temperature, resulting in an upward shift of the melting temperature  
600 from 89.6 °C to higher temperatures as the polysaccharide content increases. The  
601 melting temperatures for S15A1.5K1.5, S15A1.5K2, S15A2K1.5, and S15A2K2 are  
602 95.3 °C, 92.8 °C, 93.5 °C, and 95.2 °C, respectively. This can be attributed to the  
603 interaction between the amino and carboxyl groups of SPI and the hydroxyl groups of  
604 the polysaccharides through hydrogen bonding, consistent with the aforementioned  
605 intermolecular forces. As the polysaccharide quantity increases, the melting  
606 temperature of the composite paste gradually rises, indicating a higher amount of heat  
607 absorbed during the sample melting process and an enhancement in thermal stability.  
608 Within the melting temperature range, SA and κC in the composite paste may form a  
609 cross-linked structure, with some SPI potentially participating in the formation of the  
610 structure and being encapsulated within it. This promotes the stability of the dense  
611 cross-linking system, ultimately leading to changes in thermal stability. These findings  
612 suggest a certain degree of interaction between SPI and polysaccharides, resulting in a  
613 covalently cross-linked composite paste system, which could explain the paste's  
614 elasticity, viscosity, and strength.

### 615 **4. Conclusion**

616 In this study, a 3D printable ink was prepared using soy protein isolate,  
617 carrageenan, and sodium alginate. The addition of polysaccharides to soy protein at

618 appropriate concentrations facilitated the interaction between proteins and  
619 polysaccharides, resulting in protein-polysaccharide pastes with optimal 3D printing  
620 quality. This combination provides a viable method for the production innovative of  
621 plant-based foods. Rheological and textural analyses revealed that the addition of  
622 polysaccharides increased the viscosity and hardness of the printing ink. The  
623 S15A1.5K1.5 ink exhibited favorable viscosity and storage modulus ( $G'$ ), enabling  
624 stable printability. FT-IR analysis and molecular interaction within the dense cross-  
625 linking indicated that the introduction of polysaccharides led to the formation of  
626 hydrogen bonds through the -OH groups, transforming  $\alpha$ -helices into  $\beta$ -sheets.  
627 Hydrophobic interactions, electrostatic interactions, and disulfide bonding were  
628 identified as the main driving forces maintaining the paste structure. SEM images  
629 displayed the rough surface of the composite material, confirming the formation of a  
630 dense cross-linking structure between SPI and polysaccharides. XRD results indicated  
631 that the interactions between SPI and polysaccharides led to a transition from a  
632 crystalline to an amorphous structure. DSC analysis demonstrated that the addition of  
633 polysaccharides significantly enhanced the thermal stability of the composite paste.  
634 These findings contribute to the improvement of texture and quality in protein-  
635 polysaccharide pastes, as well as the practical application of 3D printing technology in  
636 plant-based food, establishing both theoretical and practical foundations. In our next  
637 studies, we will explore various ink formulations using other types of food ingredients,  
638 such as pea protein isolate, to meet specific nutritional requirements of individuals.

639

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799 **Table captions**

800 **Table 1.** Composition of different 3D printing ink formulations (wt%).

801 **Table 2.** Texture profile analysis (TPA) indicators for different 3D printed pastes with  
802 different formulations.

803

804

**Table 1.** Composition of different 3D printing ink formulations (wt%).

<b>Inks</b>	<b>Water/(ml)</b>	<b>SPI/(g)</b>	<b>SA/(g)</b>	<b><math>\kappa</math>C /(g)</b>
SPI 15	85	15	-	-
S15A1K1	85	15	1.0	1.0
S15A1K1.5	85	15	1.0	1.5
S15A1.5K1	85	15	1.5	1.0
S15A1.5K1.5	85	15	1.5	1.5
S15A1.5K2	85	15	1.5	2.0
S15A2K1.5	85	15	2.0	1.5
S15A2K2	85	15	2.0	2.0

805 In the codes listed for formulation column, S15AmKn food ink means that SPI, SA, and  $\kappa$ C

806 constitute 15 wt%, m wt% and n wt%, respectively.

807

808 **Table 2.** Texture profile analysis (TPA) indicators for different 3D printed pastes with  
 809 different formulations.

Sample	Hardness (g)	Adhesiveness	Springiness	Cohesiveness	Gumminess	Chewiness	Resilience (g•sec)
SPI 15	-	-	-	-	-	-	-
S15A1K1	529.27±14.13 <sup>a</sup>	-7.22±3.31 <sup>a</sup>	0.78±0.02 <sup>a</sup>	0.42±0.03 <sup>a</sup>	223.55±19.90 <sup>a</sup>	173.18±13.32 <sup>a</sup>	0.17±0.01 <sup>a</sup>
S15A1K1.5	552.91±20.28 <sup>a</sup>	-23.60±2.73 <sup>c</sup>	0.46±0.02 <sup>c</sup>	0.37±0.01 <sup>b</sup>	205.01±2.26 <sup>b</sup>	94.93±4.41 <sup>b</sup>	0.11±0.01 <sup>b</sup>
S15A1.5K1	480.10±8.76 <sup>b</sup>	-14.14±3.39 <sup>b</sup>	0.59±0.02 <sup>b</sup>	0.36±0.01 <sup>bc</sup>	172.20±4.56 <sup>c</sup>	101.65±5.83 <sup>b</sup>	0.11±0.01 <sup>b</sup>
S15A1.5K1.5	538.73±12.16 <sup>a</sup>	-56.24±1.93 <sup>e</sup>	0.45±0.02 <sup>c</sup>	0.33±0.01 <sup>c</sup>	180.06±7.39 <sup>c</sup>	81.88±2.62 <sup>c</sup>	0.08±0.00 <sup>d</sup>
S15A1.5K2	-	-	-	-	-	-	-
S15A2K1.5	429.84±7.18 <sup>c</sup>	-43.74±3.38 <sup>d</sup>	0.45±0.01 <sup>c</sup>	0.35±0.00 <sup>bc</sup>	150.22±2.08 <sup>d</sup>	67.47±0.47 <sup>d</sup>	0.10±0.00 <sup>c</sup>
S15A2K2	-	-	-	-	-	-	-

810 Values were expressed as the mean ± standard deviation (n = 3). Different letters indicated  
 811 significant differences ( $p < 0.05$ ) between values in the same column. "-" means not shaped.  
 812 S15AmKn food ink means that SPI, SA, and κC constitute 15 wt%, m wt% and n wt%,  
 813 respectively.  
 814

815 **Figure captions**

816 **Figure 1.** Analysis of 3D printing of different ink formulations and pictures of the  
817 printed products.

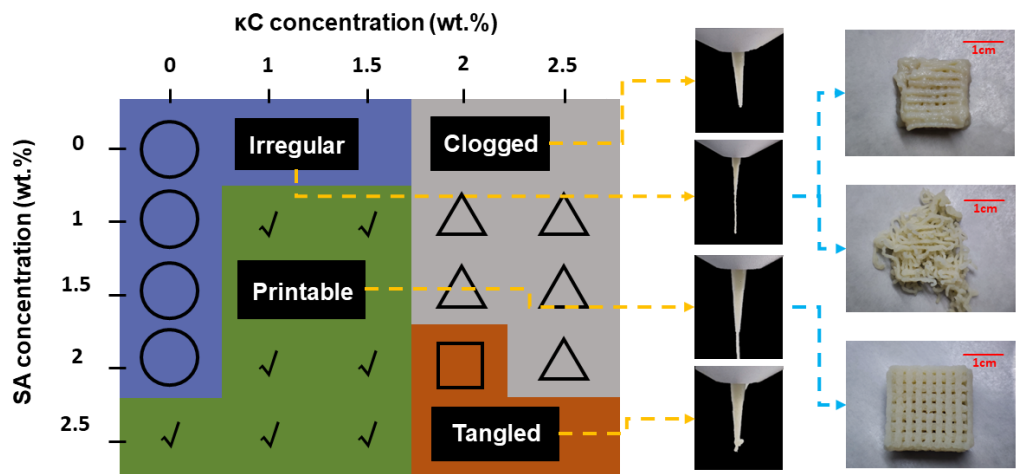
818 **Figure 2.** Rheological properties analysis of different ink formulations: (a): viscosity;  
819 (b): strain provide information which lines for  $G'$  and  $G''$ ; (c): storage modulus  $G'$ ; (d):  
820 loss modulus  $G''$ .

821 **Figure 3.** Microstructure of ink formulations with different magnifications of 1000  $\times$ ,  
822 2500  $\times$  and 5000  $\times$ .

823 **Figure 4.** (a): Changes in molecular interaction force of the composite pastes in the  
824 presence of different polysaccharide concentrations. Mean values with different letters  
825 (a-d) differ significantly ( $p < 0.05$ ); (b): The Fourier transform infrared, (FTIR)  
826 spectrum characterization, (c): X-ray diffractograms and (d): DSC thermograms of  
827 composite pastes with different formulations.

828 **Figure 5.** The formation mechanism diagram of SPI-polysaccharide composite pastes.  
829

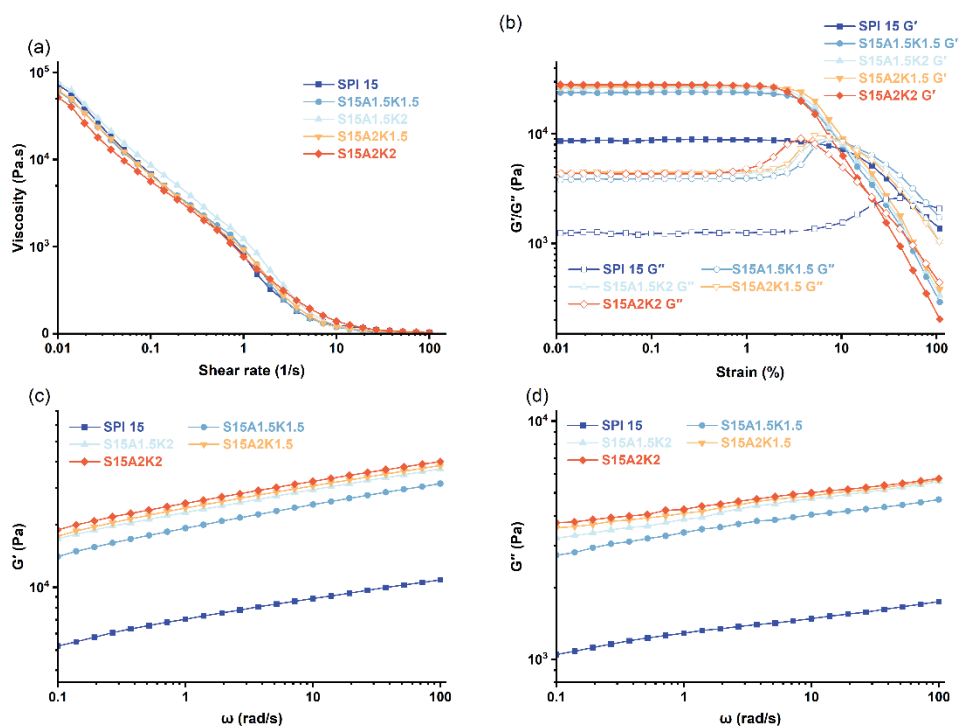
830 **Figure 1**



831

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833 printed products.

834



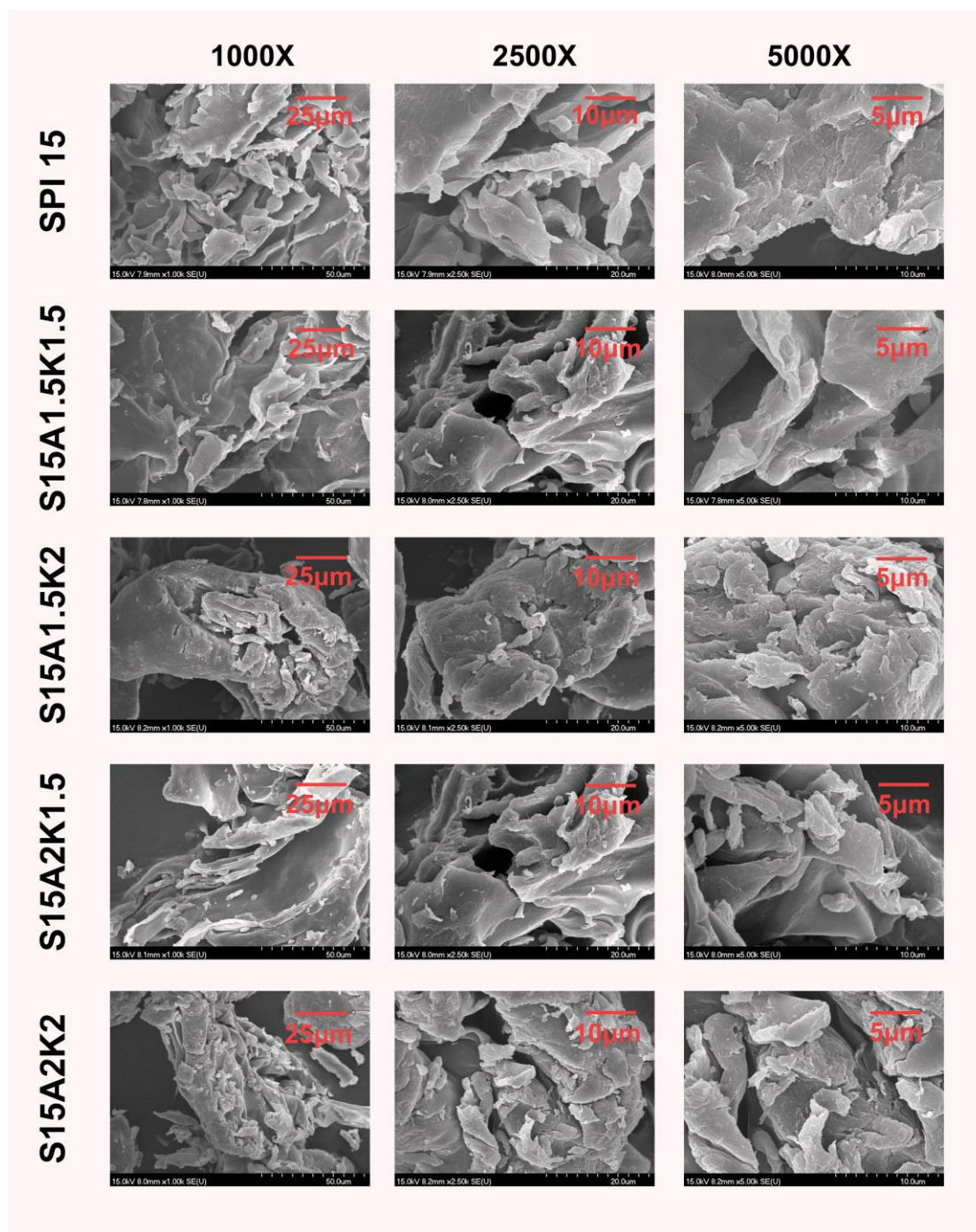
836

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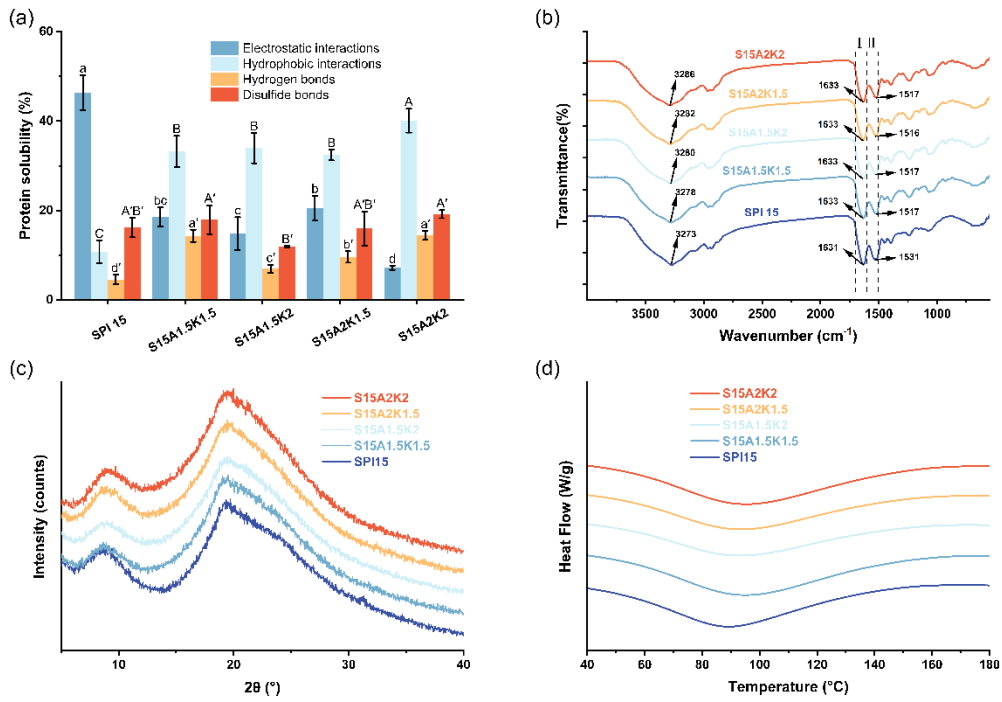


842

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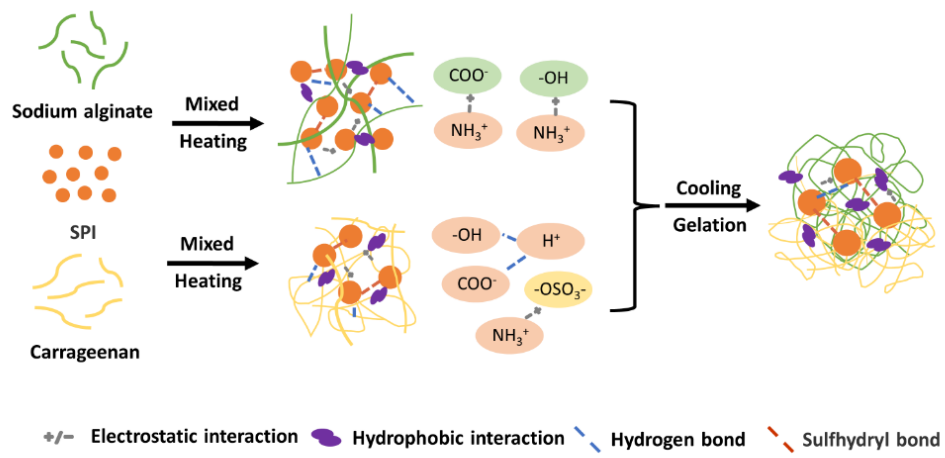
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845



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 851 composite pastes with different formulations.  
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854 Figure 5. The formation mechanism diagram of SPI-polysaccharide composite pastes.