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Killing or Cheating: two strategies to colonize an oyster

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High prevalence of the Vibrio Harveyi clade during a Pacific Oyster **Mortality Syndrome episode in the Thau Lagoon**

Since 2008, the worldwide production of *Crassostreae gigas* oysters has been dramatically affected by the Pacific Oyster Mortality Syndrome (**POMS**)¹. This polymicrobial disease is caused by a Ostreid herpesvirus, **OsHV-1** μ **Var**, and opportunistic bacteria, among which members of the Vibrionaceae family are the best characterized².

Oysters susceptible to POMS present an overall lower basal expression of immune related genes and an inefficient transcriptomic response against OsHV-1 μ Var infection. Viral replication in susceptible oysters leads to the host entering an immune-compromised state, causing a dysbiosis that evolves towards bacteremia³.

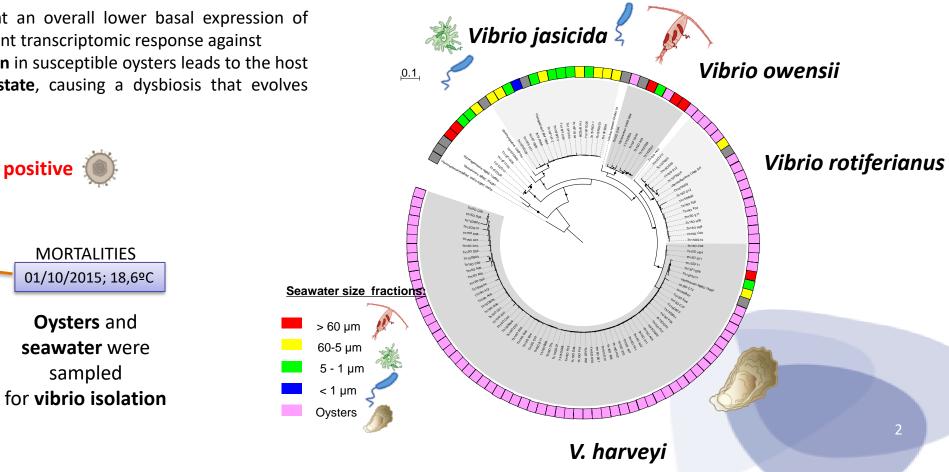
Mortality outbreak with oysters positive (

for OsHV-1 µvar-1

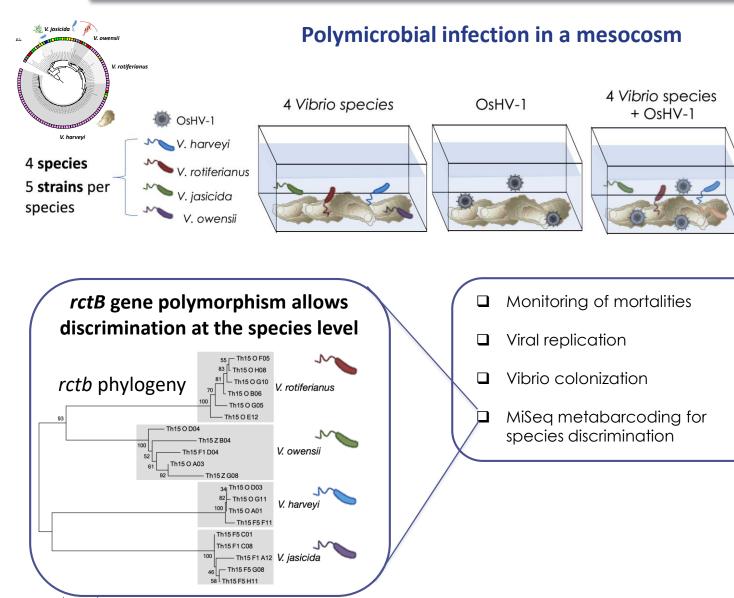
Thau Lagoon, France

2015

Here, we investigated the Vibrio community associated with the disease in the Thau lagoon (south of France), were we found a strong prevalence of the Harveyi clade in diseased oysters.



Can we reproduce the association of *V. harveyi* and *V. rotiferianus* found in the field?



Tanks with 2L of seawater and 10 oysters were prepared for **sampling** at different time points and tanks with 3L of seawater with 15 oysters to **track the daily mortalities** on each condition.

Seawater containing **OsHV-1** μ **Var** virions was produced trough **donors oyster. Donors** were injected with the **OsHV-1** virus and released virions into the seawater trough a **24hr** period. This **contaminated seawater** was used to fill the tanks for the different experimental conditions.

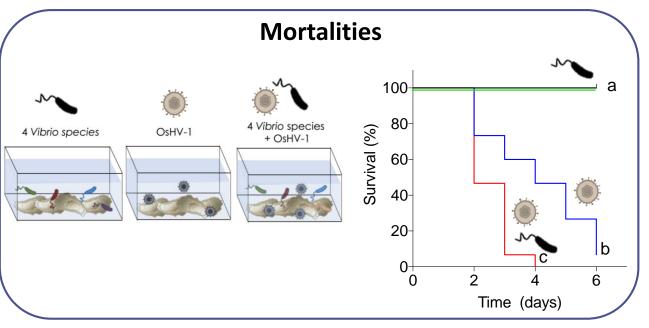
After placing the oysters in the tanks, bacterial suspensions in seawater adjusted to OD600 = 1 (10^9 CFU/mL) were added to each tank to reach a final concentration of 10^7 CFU/mL.

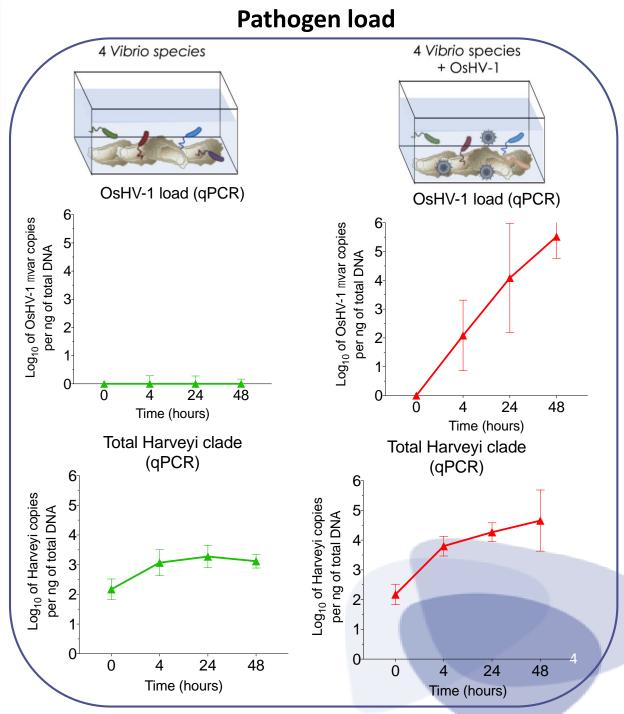
10 oysters at **each time** point and **for each condition** were sampled for analysis

OsHV-1 favors colonization by vibrios and synergizes with members of the Harveyi clade to kill oysters

No mortalities were observed when oysters were immersed in seawater containing **only the Harveyi community**. Consistently, a transient increase in total Harveyi load was observed from 4h to 24h followed by a decreased at 48h, revealing **a control of** *Vibrio* **colonization**

Mortalities were **significantly enhanced (100% at day 4)** when the Harveyi community and OsHV-1 μ var were **combined into the mesocosm** In this condition the total Harveyi load in tissues of live oysters was **1-2 log higher** than in other conditions at 2 day



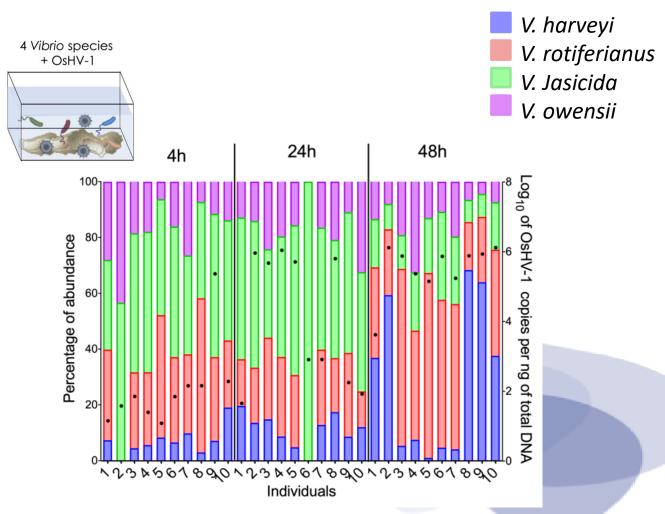


OsHV-1 µVar favors colonization by V. harveyi and V. rotiferianus only

Individuals collected during the time course of the infection were therefore subjected to illumina Miseq sequencing of the *rctB* **polymorphic region** which discriminated the four species introduced into the mesocosm. **The relative abundance of the 4 species remained stable in the oysters only exposed to the Harveyi** community during 48h, as revealed by *rctB* barcoding analysis

4 Vibrio species 4h 24h 48h 100 _og₁₀ 우 OsHV-1 80 Percentage of abundance 60 copies per ng 40 of total 20 DN へりらかららく もみのへりらかららく もみのへりらかららく もみの Individuals

On the contrary, **in the presence of OsHV-1** µvar, not only the total Harveyi load increased over time but there was also a **significant enrichment of the species** *V. harveyi* **and** *V. rotiferianus* at 48 h, after viral replication occurred (24h)

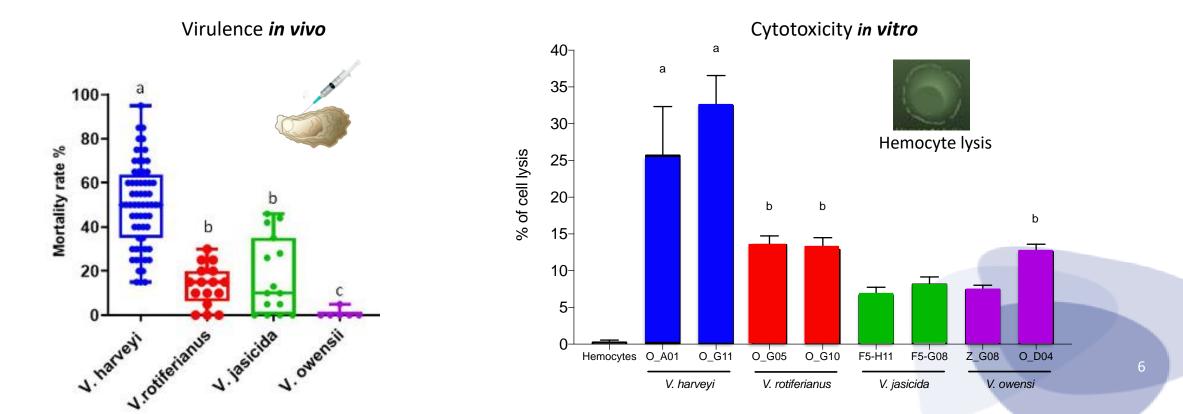


What makes V. harveyi and V. rotiferianus succesful colonizers?

V. harveyi, the strategy of a killer

In experimental infection by injection, *V. harveyi* isolates were by far the most virulent with \geq 50% of *V. harveyi* isolates inducing \geq 50% mortality after 1 day. The cytotoxic potential of *V. harveyi* was confirmed experimentally in vitro by comparing two strains representative of the four Harveyi species from our study

We found that *V. harveyi* strains exerted a potent cytotoxicity toward hemocytes, the oyster immune cells, 25-35% of which were lysed upon incubation with *V. harveyi* strains. *V. harveyi* harbored a type 3 secretion system homologous to the *V. parahaemolyticus* T3SS1 that could contribute to its virulence and cytotoxicity.

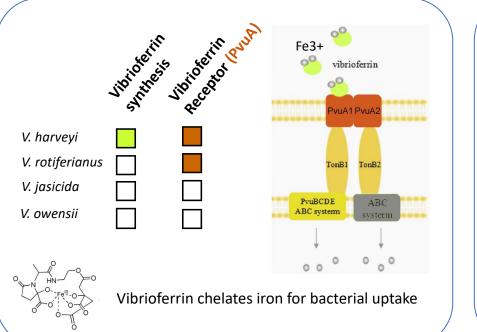


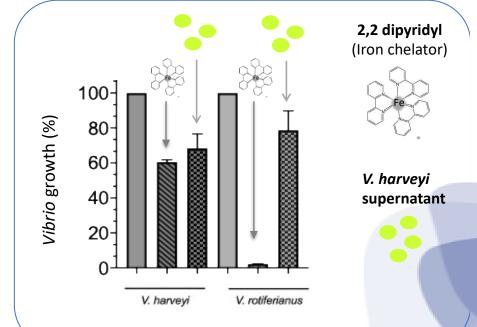
What makes V. harveyi and V. rotiferianus successful colonizers?

V. rotiferianus, the strategy of a cheater

Indeed, *V. harveyi* was found to harbor a conserved set of genes responsible for the synthesis of vibrioferrin, a tricarboxylic acid siderophore. In contrast, *V. rotiferianus* does not exhibit known *Vibrionaceae* siderophore biosynthetic proteins **but it carries the vibrioferrin receptor, present in both species, necessary for the siderophore uptake.** *V. harveyi* growth was not impaired in iron depleted media (EDDA used as iron chelator. On the contrary, iron depletion significantly affected the growth of *V. rotiferianus*. This growth defect was rescued by the addition of cell-free culture supernatant from *V. harveyi*. This demonstrates that *V. rotiferianus* is able to acquire important resources (most likely vibrioferrin) produced by *V. harveyi*, which are needed to grow in iron-poor environments

Comparative genomics identifies a **siderophore receptor** present in *V. harveyi* and *V. rotiferianus* only V. rotiferianus uses V. harveyi secretion products to grow in iron-poor conditions





7

Conclusions



) *V. harveyi* acts as a **killer**: it uses **virulence** and **cytotoxicity** to colonize its **immunocompromised** host

V. rotiferianus acts as a cheater: it benefits from the virulence/cytotoxicity and the siderophore production of V. harveyi

References

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