



**HAL**  
open science

## Mesostructural changes in cellulose within wood cell wall upon hydrothermal treatment at 200 °C

Tomoko Kuribayashi, Yu Ogawa, I. Morfin, Yuji Matsumoto, Yoshiharu Nishiyama

► **To cite this version:**

Tomoko Kuribayashi, Yu Ogawa, I. Morfin, Yuji Matsumoto, Yoshiharu Nishiyama. Mesostructural changes in cellulose within wood cell wall upon hydrothermal treatment at 200 °C. *Cellulose*, 2023, 30 (13), pp.8405-8413. 10.1007/s10570-023-05388-1 . hal-04876854

**HAL Id: hal-04876854**

**<https://hal.science/hal-04876854v1>**

Submitted on 9 Jan 2025

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

1 **Mesostructural changes in cellulose within wood cell wall upon hydrothermal**  
2 **treatment at 200 °C**

3 Tomoko Kuribayashi <sup>a, b, c, \*</sup>, Yu Ogawa <sup>d</sup>, Isabelle Morfin <sup>e</sup>, Yuji Matsumoto <sup>a</sup>,  
4 Yoshiharu Nishiyama <sup>d, \*</sup>

5 <sup>a</sup> Graduate School of Agricultural and Life Sciences, The University of Tokyo, Yayoi 1-  
6 1-1, Bunkyo-Ku, 113-8657 Tokyo, Japan

7 <sup>b</sup> Department of Bioproducts and Biosystems, Aalto university, Vuorimiehentie 1, 02150  
8 Espoo, Finland

9 <sup>c</sup> Department of Food and Nutrition, University of Helsinki, P.O. Box 66, 00014  
10 Helsinki, Finland

11 <sup>d</sup> Univ. Grenoble Alpes, CNRS, CERMAV, 38000 Grenoble, France

12 <sup>e</sup> Univ. Grenoble Alpes, CNRS, LiPhy, 38000 Grenoble, France

13 \*Corresponding authors:

14 T. Kuribayashi, [tomoko.kuribayashi@aalto.fi](mailto:tomoko.kuribayashi@aalto.fi)

15 Y. Nishiyama, [yoshiharu.nishiyama@cermav.cnrs.fr](mailto:yoshiharu.nishiyama@cermav.cnrs.fr)

16

17 **Abstract**

18 Hydrothermal treatment between 150 °C and 230 °C is widely used in wood processing,  
19 from the steam treatment of timber for better dimensional stability and durability to the  
20 pretreatment for enzymatic saccharification and chemical pulping. Understanding the  
21 ultrastructural changes of wood cell walls through hydrothermal treatments is crucial for  
22 controlling and optimizing these hydrothermal treatment-based processes. Here, we  
23 studied the ultrastructure of wood cell walls of 24 hardwood species using simultaneous  
24 small- and wide-angle X-ray scattering measurements before and after the hydrothermal

25 treatment at 200 °C. Most hardwoods show similar equatorial scattering features,  
26 representing the structure in the cross-section of the cell walls. In a water-saturated  
27 native state, there is a **prominent** correlation peak between 0.1 and 0.2 Å<sup>-1</sup> and a second  
28 peak between 0.2 and 0.4 Å<sup>-1</sup>. The hydrothermal treatment above 160 °C drastically  
29 altered the structure at this nanometric scale: the two native correlation peaks  
30 disappeared, coincident with a buildup of a correlation peak in the 0.03-0.04 Å<sup>-1</sup> range.  
31 The hydrothermal treatment likely removed the cell wall matrix component between the  
32 microfibrils through autohydrolysis and phase separation, leading to the collapse of  
33 microfibrils with each other in the normal wood. In cellulose-rich cell walls, **such as the**  
34 **G-layer in tension wood**, cellulose microfibrils are already **collated** in the native state.

35

36 **Keywords:** Wood; Cellulose microfibril; Mesostructure; X-ray scattering;  
37 Hydrothermal treatment; Co-crystallization

38

### 39 **Introduction**

40 Thermal treatment between 150°C and 230°C with moisture is commonly used in  
41 timber processing. Heat steam treatment decreases **wood's equilibrium moisture content**,  
42 **improving** dimensional stability and durability (Kocaeffe et al. 2015; Sandberg and  
43 Kutnar 2016). Hydrothermal treatment with excess water causes partial hydrolysis of  
44 the hemicelluloses that generates water-soluble saccharides and increases the specific  
45 surface area of the wood (Garrote et al. 1999). Such effects are **helpful in the**  
46 pretreatment of woody biomass for saccharification and chemical pulping (Garrote et al.  
47 1999; Nitsos et al. 2013, 2016; Romani et al. 2010; Wojtasz-Mucha et al. 2017). The  
48 ultrastructural changes of wood cell walls by hydrothermal treatments are thus an

49 important parameter.

50 X-ray scattering can probe the structural features over wide length scales, typically  
51 from angstroms to a few hundred nanometers, without special sample preparation,  
52 which makes it a suitable analytical tool for structural investigations of plant cell walls  
53 (Jakob et al. 1994, 1996; Fernandes et al. 2011; Penttilä et al. 2019). Small- (SAXS) and  
54 wide-angle X-ray scattering (WAXS) have been used to study the structural evolutions  
55 of fully hydrated aspen (Nishiyama et al. 2014) and bagasse (Driemeier et al. 2015)  
56 during hydrothermal treatments. These studies found several common structural  
57 changes, such as co-crystallization of cellulose and phase separation of the cell wall  
58 matrix component (Nishiyama et al. 2014; Kuribayashi et al. 2016). We extended this  
59 study to broader spectrum wood samples, including 24 hardwood species before and  
60 after hydrothermal treatment at 200 °C for 2 hours using simultaneous S/WAXS  
61 measurements.

62

## 63 **Experimental**

### 64 **Materials**

65 Green beech (*Fagus crenata*) was harvested from The University of Tokyo Chichibu  
66 Forest (Saitama, Japan). Blocks (dimensions: 13 mm×13 mm×60 mm in the direction of  
67 tangential (T), radial (R), and longitudinal (L), respectively) were sawn out from the  
68 mature sapwood and stored in water with sodium azide at 4°C.

69 Air-dried 24 hardwood specimens are summarized in Table 1. These were harvested  
70 in Japan and kindly provided by the Research Institute for Sustainable Humanosphere  
71 (RISH), Kyoto University, Japan.

72

73 **Table 1.** A list of wood specimens and their moisture contents

No.	Scientific name	Common name	Mc [%] <sup>(1)</sup>		
			Air Dry	Wet	
			Untreated	Untreated	Thermal treated
1	<i>Acer</i> sp.	Maple	7	121	123
2	<i>Aesculus turbinata</i>	Japanese horse-chestnut	8	123	128
3	<i>Aphananthe aspera</i>	Muku tree	8	82	87
4	<i>Castanea crenata</i>	Japanese chestnut	8	117	110
5	<i>Castanopsis</i> sp.	Chinquapin	8	116	105
6	<i>Celtis sinensis</i>	Chinese hackberry <sup>(2)</sup>	7	142	130
7	<i>Cercidiphyllum japonicum</i>	Katsura	7	139	139
8	<i>Cinnamomum camphora</i>	Camphor tree	7	108	128
9	<i>Cleyera japonica</i>	Japanese cleyra	6	187	183
10	<i>Diospyros kaki</i>	Japanese persimmon	6	94	97
11	<i>Distylium racemosum</i>	Isu tree	10	98	97
12	<i>Firmiana simplex</i>	Chinese parasol tree	7	90	86
13	<i>Fraxinus mandshurica</i>	Manchurian ash	6	133	149
14	<i>Ilex integra</i>	Mochi tree	6	82	88
15	<i>Kalopanax septemlobus</i>	Castor aralia <sup>(2)</sup>	6	135	143
16	<i>Liriodendron tulipifera</i>	Yellow poplar	7	141	154
17	<i>Magnolia obovata</i>	Japanese bigleaf magnolia	9	114	148
18	<i>Morus</i> sp.	Mulberry	7	135	130
19	<i>Quercus</i> sp.	Oak	9	110	83
20	<i>Quercus acutissima</i>	Sawtooth oak	9	85	83
21	<i>Quercus crispula</i>	Mizunara	7	100	97
22	<i>Styphnolobium japonicum</i>	Japanese pagoda tree	6	96	98
23	<i>Ulmus davidiana</i>	David elm	8	193	209
24	<i>Zelkova serrata</i>	Japanese elm	9	120	116

(1) Mc [%] = Dry basis moisture content. (2) Tension wood.

74

75 **Preparation of water-swollen wood samples**

76 Cylindrical samples with about 2 mm diameter and 10 mm length in fiber direction  
77 were cut out from the air-dried wood blocks. Specimens were cropped from the part  
78 without visible large vessel pores and medullary rays. Deionized water was soaked in  
79 the samples by vacuum pressure impregnation until the sample weight did not increase.  
80 The dry-basis moisture content of water-saturated samples was estimated as follows. A  
81 part of air-dried samples was dried overnight in an oven at 105 °C to obtain the dry  
82 weight. We calculated the equilibrium moisture contents of water-saturated samples in  
83 the ambient condition and considered it to be the same for each wood species. Moisture

84 contents in the water saturated samples were estimated taking into account the  
85 equilibrium moisture contents and weight gain by water impregnation. The calculated  
86 moisture contents in air-dried and water-saturated states were summarized in Table 1.  
87 Immediately after weighing the water-saturated sample, each wet sample was sealed  
88 into a glass tube (3 mm outer diameter, 200  $\mu\text{m}$  wall thickness).

### 89 **Hydrothermal treatment**

90 The glass tubes containing a wood sample and a drop of water for pressure regulation  
91 were placed in a 30 ml pressure vessel and heat-treated in an oil bath at 200 °C for 2  
92 hours. The partial water vapor pressure during hydrothermal treatment is 1.6 MPa  
93 according to the Tetens approximation formula. After the treatment, the vessel was  
94 quenched with ice.

### 95 **Simultaneous SAXS and WAXS measurement**

96 X-ray scattering experiments were performed at the D2AM beamline of the  
97 European Synchrotron Radiation Facility (ESRF). The tubes containing samples were  
98 probed with a monochromatic X-ray of 18 keV ( $\lambda = 0.6889 \text{ \AA}$ ) before and after the  
99 hydrothermal treatment. The incident beam was orthogonal to the sample tubes. The  
100 scattering intensity was measured using two-dimensional pixel detection (XPAD-WOS  
101 and D5). Two sets of camera distances, D5 at 2.2 m and WOS at 32 cm or D5 at 1 m  
102 and WOS at 12 cm were used to cover a scattering vector range of  $q (2\pi/d) = 0.005$  to 6  
103  $\text{\AA}^{-1}$ .

### 104 **In-situ X-ray scattering measurement over the thermal treatment**

105 We used an aluminum block with 3 mm perforations to fit sample tubes and holes  
106 for incoming X-rays and scattered X-rays at D2AM beamline (ESRF). The temperature  
107 was regulated by using a heater regulated by a Eurotherm temperature controller.

108 Sample tubes containing green wood and wood with moisture content of 0-30 % were  
109 sealed by flame and placed in the sample holder. The samples were first heated from  
110 25 °C to 80 °C, and the temperature was elevated stepwise by 10 °C with 10 minutes of  
111 holding time. After reaching 200 °C, the temperature was kept for 2 hours, then cooled  
112 to 23 °C by heat dissipation. The X-ray energy was 16 keV ( $\lambda = 0.775 \text{ \AA}$ ). The scattered  
113 X-rays were recorded on a fiber optic coupled CCD camera. The experiment was  
114 repeated twice with two detector distances, 1.6 m and 16 cm, to cover a q range from  
115 0.03 to  $2 \text{ \AA}^{-1}$ .

## 116 **Data analysis**

117 The beam center and the detector positions were refined using PyFAI (Ashiotis et al.  
118 2015) software using silver behenate and  $\text{Cr}_2\text{O}_3$  diluted with microcrystalline cellulose  
119 powder in the same sample tube as the calibrant. The data were scaled by transmission  
120 intensity and remapped into polar coordinates. The intensity of the overlapping region  
121 was used to scale the two data sets measured at different detector distances and merged  
122 into one figure. The isotropic scattering level was estimated for each scattering angle by  
123 averaging 10 % of pixels of the lowest intensities. For the in-situ experiment data, the  
124 azimuthal intensity distributions were fitted with a constant and a Gaussian function for  
125 each scattering angle to separate the isotropic and the equatorial (anisotropic)  
126 contributions (Nishiyama et al. 2014).

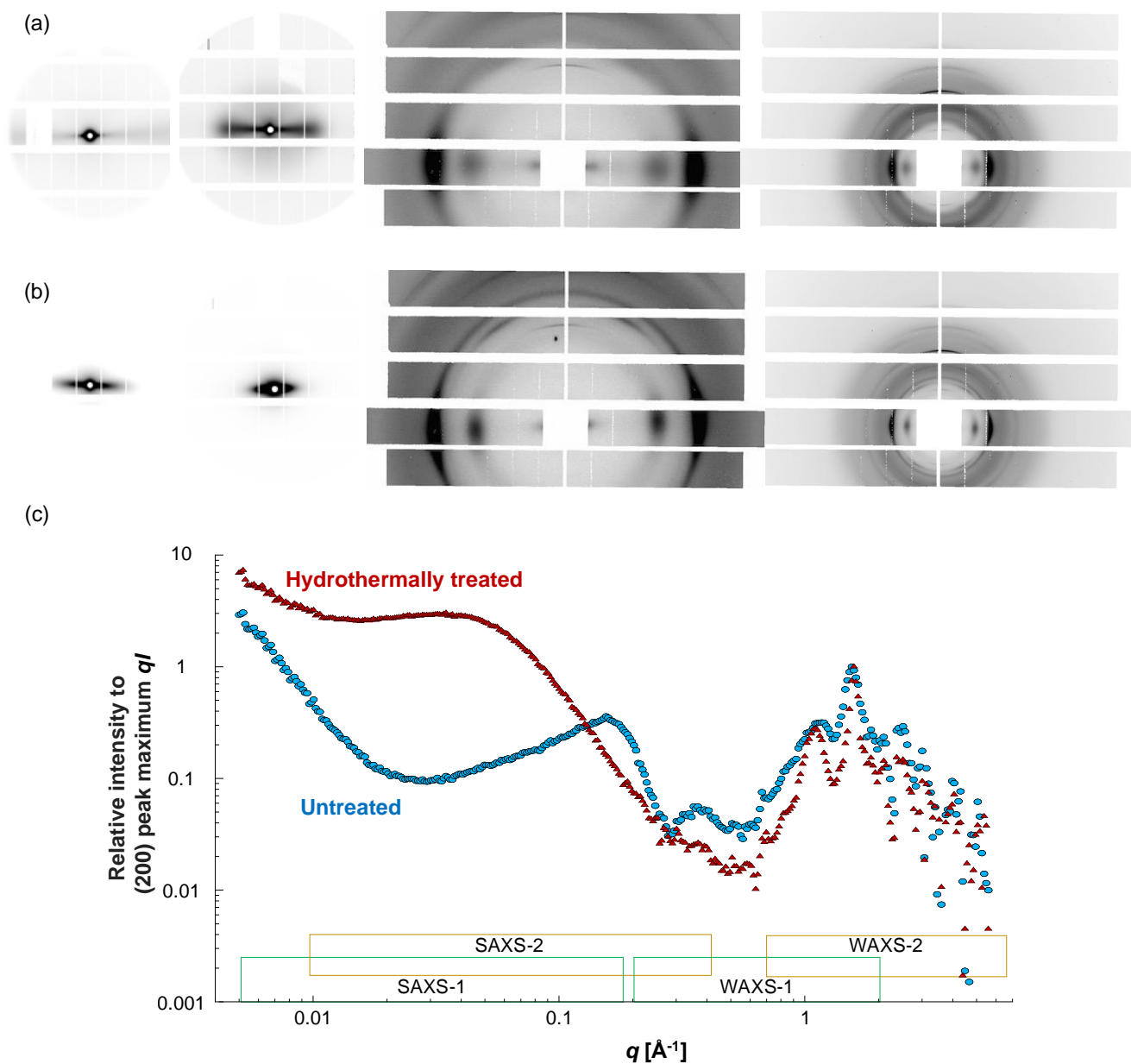
## 127 **Results and discussion**

### 128 **Changes in the scattering profile of anisotropic components**

129 Examples of scattering patterns obtained from *Quercus* sp. are shown in Fig. 1. The  
130 untreated and treated wood show anisotropic scattering features over the whole q range:  
131 SAXS-1 and 2 show a streak in the equatorial direction (Fig. 1(a)), while WAXS-1 and

132 2 show fiber diffraction patterns where cellulose is oriented along the long axis of wood  
133 (Fig. 1(b)). The scattering profile of equatorial components of *Quercus* sp. is shown in  
134 Fig. 1(c), where  $q \times I$  is plotted as a function of  $q$ , where  $I$  is an intensity count at a given  
135  $q$ . **Multiplying by  $q$  is equivalent to making an abstraction of the misalignment of**  
136 **cylindrically symmetric structure and extracting the power spectrum of the electron**  
137 **density of the cross-section.**





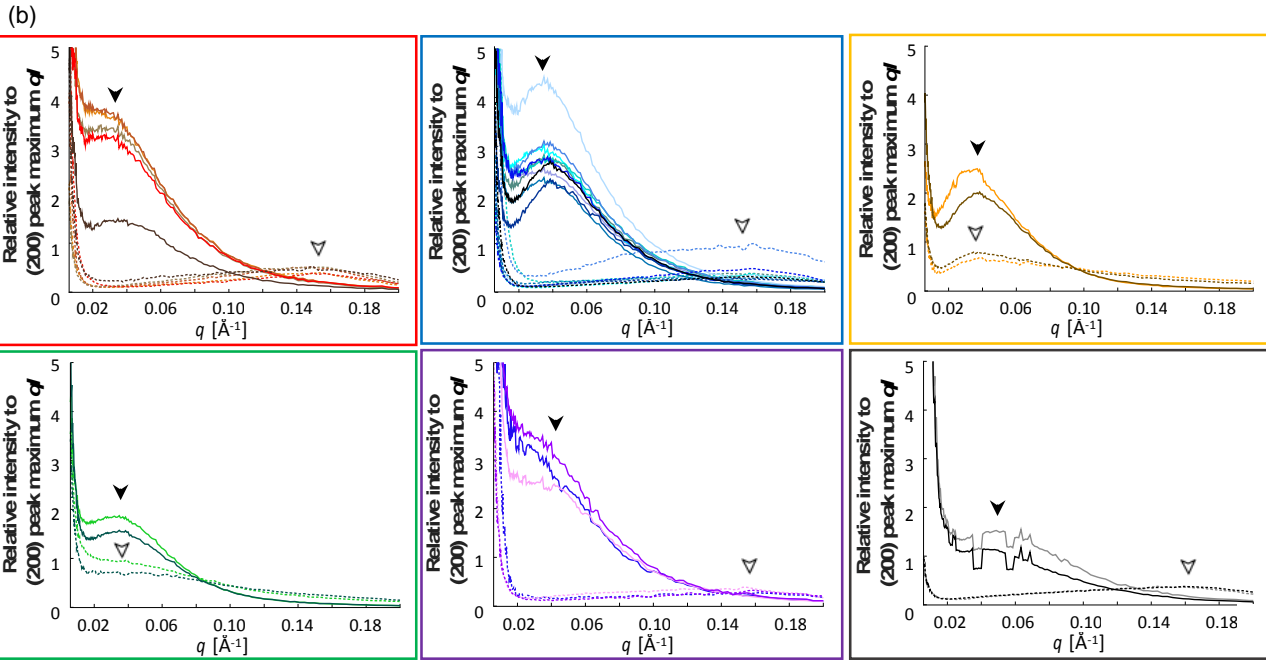
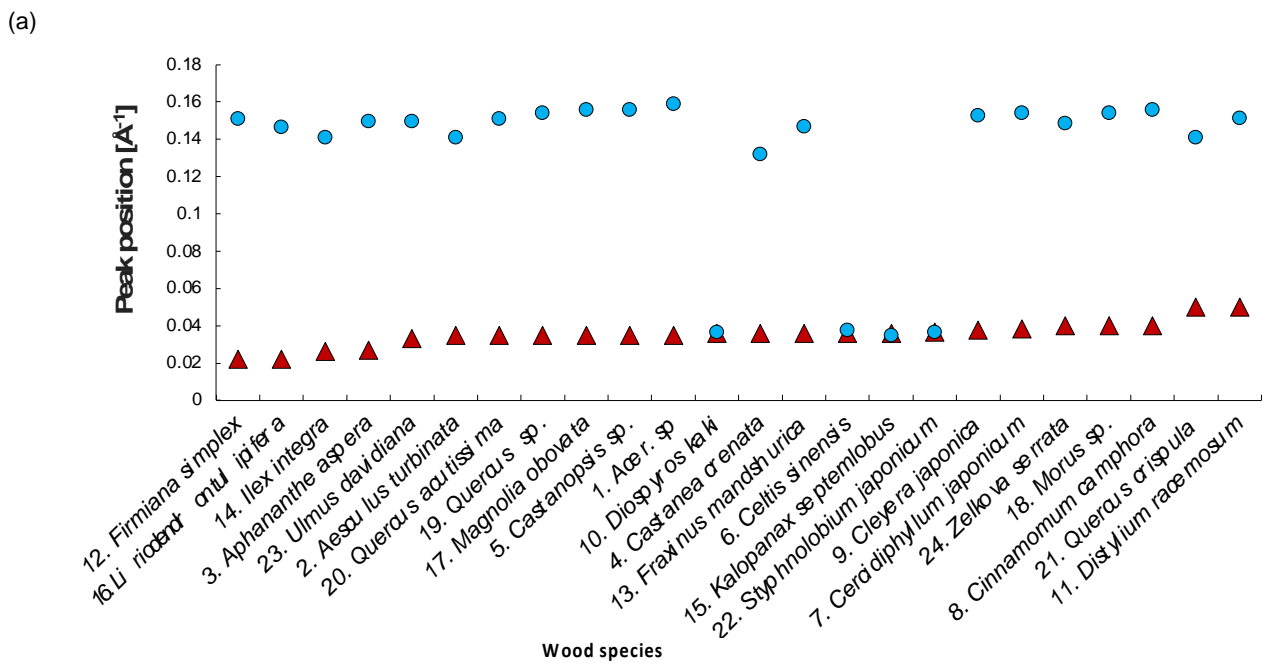
138 **Figure 1.** Two-dimensional scattering images (a, b) and profiles of equatorial  
 139 component (c) of the wet oak wood sample (*Quercus* sp.) before ((a) and blue profile in  
 140 (c)) and after hydrothermal treatment at 200 °C for 2 hours ((b) and red profile in (c)).  
 141 Scattering images in (a) and (b) are SAXS-1, SAXS-2, WAXS-1, and WAXS-2 from left  
 142 to right. The X-ray beam was irradiated orthogonal to the wood fiber direction (i.e., the  
 143 fiber direction was vertical in the diagrams).

144

145 In the WAXS range, a composite peak of 1 -1 0/1 1 0 reflections and the 2 0 0  
146 reflections of cellulose I $\beta$  are visible at 1.1 and 1.6  $\text{\AA}^{-1}$ , respectively. The pattern before  
147 the thermal treatment (blue profile) is typical of wood samples, and the positions and  
148 the intensity ratio roughly correspond to those obtained from highly crystalline cellulose  
149 I $\beta$ . After hydrothermal treatment at 200°C, the equatorial reflections became much  
150 sharper due to the co-crystallization of microfibrils as reported previously (Fig. 1(a) and  
151 (b)) (Kuribayashi et al. 2016, 2019).

152 In the lower q range recorded, two peaks are present at  $q = 0.1 - 0.2 \text{\AA}^{-1}$  and  $q = 0.3 -$   
153  $0.4 \text{\AA}^{-1}$  in the untreated sample (Fig. 1 (a)). Several studies have previously observed the  
154 former peak using X-ray (Jakob et al. 1996; Chen et al. 2021; Paajanen et al. 2022) or  
155 neutron (Fernandes et al. 2011; Nishiyama et al. 2014). The peak can be explained by  
156 the correlation between hard cylinders (i.e. cellulose microfibrils) randomly packed in  
157 the cell wall (Jakob et al. 1996) or the center-to-center distance of cylinders (Fernandes  
158 et al. 2011) or the convolution of the form factor of cylindrical microfibrils and their  
159 correlation (Chen et al. 2021). The latter peak has not been clearly explained, while  
160 Viljanen and co-workers (2020) reported a peak fitting of the peak observed in a SAXS  
161 profile of a tropical wood using a form factor of a cylinder. In Paajanen et al. (2022), the  
162 sample volume in the beam was probably too small compared to the Kapton window  
163 film whose scattering overlapped with this peak. In Jakob et al. (1996), the part of the  
164 peak can be recognized, but the limited q-range did not allow the appreciation of the  
165 whole feature. In our study, we used a sample thickness of about 2.5 mm, leading to  
166 sufficiently high scattering intensity with respect to the contribution of optical elements  
167 in the beam, revealing that this peak is another common scattering feature of many  
168 hardwood species.

169           The hydrothermal treatment drastically changed the scattering features: the  $0.3 \text{ \AA}^{-1}$   
170 and the  $0.15 \text{ \AA}^{-1}$  peaks completely disappeared, and a new peak appeared at a much  
171 lower  $q$  range ( $0.01\text{-}0.05 \text{ \AA}^{-1}$ ). The streak intensities increased in the  $q$  range of  $0.06\text{-}$   
172  $0.6 \text{ \AA}^{-1}$ . Below  $q = 0.01 \text{ \AA}^{-1}$ , the intensity decay followed a power law. We carried out  
173 the same hydrothermal treatment on the 24 species. As summarized in Fig. 2, the small  
174 angle peaks (white arrowheads in Fig. 2(b)) were visible between  $0.12\text{-}0.16 \text{ \AA}^{-1}$  in  
175 most wood species before the treatment except 4 species (*D. kaki*, *C. sinensis*, *K.*  
176 *septemlobus*, and *S. japonicum*). Similar to the case of *Quercus* sp., this peak  
177 disappeared after the thermal treatment, and the new broad peak appeared centered  $0.02$   
178  $\text{-}0.04 \text{ \AA}^{-1}$  (black arrowheads in Fig. 2(b)). This change indicates that the hydrothermal  
179 treatment commonly induced the structural coarsening of approximately one order of  
180 magnitude in length scale in the cell walls of these 20 species.



181 **Figure 2.** (a) Peak positions of prominent correlation peaks of 24 wood species before  
 182 (blue circle) and after the hydrothermal treatment (red triangle). (b) Corresponding X-  
 183 ray scattering profiles before (dotted lines) and after the hydrothermal treatment (solid  
 184 lines) in the double linear scale. The line colors in (b) correspond to those in (a).

185

186 On the other hand, the above-mentioned 4 species had small-angle peaks at  $0.03 \text{ \AA}^{-1}$

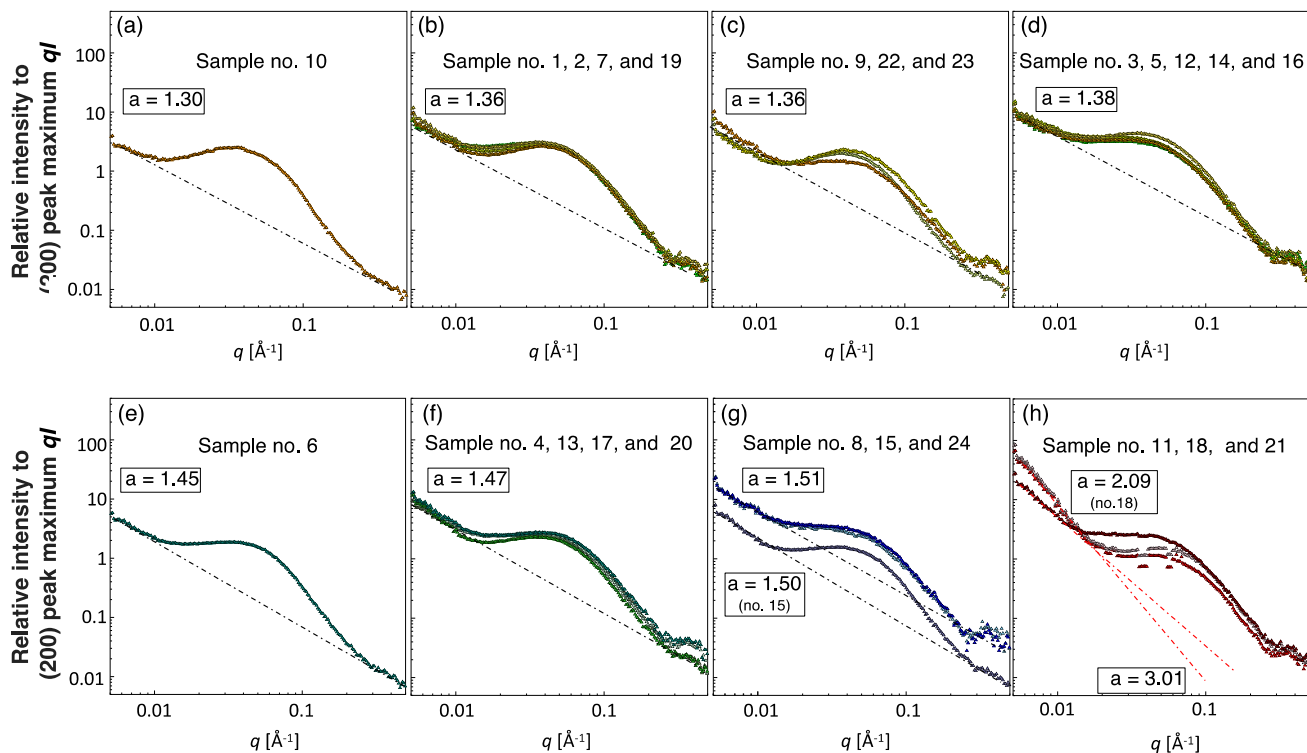
187 <sup>1</sup> before the hydrothermal treatment (Fig. 2(a)). While the peak intensities increased, the  
188 peak positions remained at the same positions after the treatment of these species (Fig.  
189 2(b)). Thus, the hydrothermal treatment did not strongly affect the structural features at  
190 this ten-nanometer length scale related to the organization of cellulose microfibrils in  
191 the cell walls. Optical micrographs of *C. sinensis* (6), *K. septemlobus* (15) before the  
192 treatment show thick G-layers in the fiber cells (Fig. S1). The G-layer is a non-lignified  
193 cell wall layer composed mostly of cellulose. The lack of the matrix components in the  
194 G-layer may cause the coalescence of cellulose microfibrils, similarly in the  
195 hydrothermally treated cell walls, resulting in a coarse cell wall structure and hence  
196 similar scattering features before and after the treatment. The other two species, *D. kaki*  
197 (10), *S. japonicum* (22), as well as the remaining 22 specimens, did not show G-layer in  
198 their micrographs, while the scattering patterns were similar to the aforementioned  
199 species with the tension wood feature (Fig. S1). The solid-state <sup>13</sup>C NMR analysis  
200 suggested that specimens of *C. sinensis* (6), *K. septemlobus* (15), *D. kaki* (10) contained  
201 a higher amount of crystalline cellulose compared to other specimens (Kuribayashi,  
202 2019).

203

#### 204 **Q range below 0.01 Å<sup>-1</sup>**

205 In the smaller angle region below 0.01 Å<sup>-1</sup>, power-law behavior can be seen after  
206 hydrothermal treatment, as shown in Fig. 3. The straight line extends to q = 0.4 Å<sup>-1</sup> in  
207 21 species, as seen in the double logarithmic scale. The slope constants were between -  
208 1.3 to -1.5. The remaining 3 species, *D. racemosum* (11), *Morus* sp. (18), *Q. crispula*  
209 (21), show higher exponents in the low q range below 0.01 Å<sup>-1</sup>. In particular, the  
210 constants for *D. racemosum* and *Q. crispula* are -3, corresponding to the presence of a

211 clear interface in the structure. This might be due to air bubbles in the specimens,



212 resulting in the air-solid interfaces.

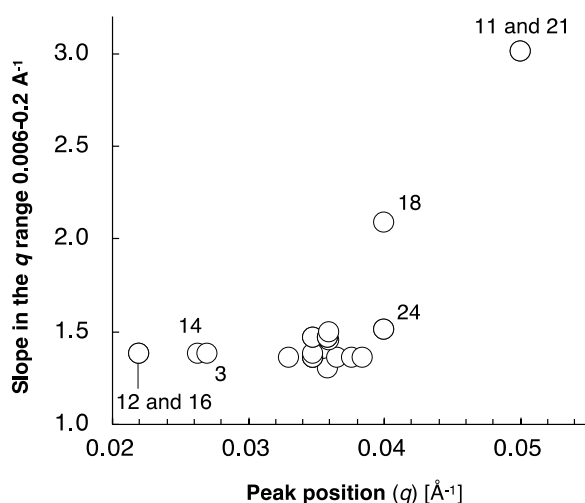
213 **Figure 3.** Scattering profiles of 24 hydrothermally treated wood species between  $q =$   
214  $0.006$  and  $0.3 \text{ \AA}^{-1}$  in double log scale with power-law slopes as dotted lines (a).

215 Numbers correspond to those in Table 1.

216

217 The relationship between the slope constants and the peak positions after the  
218 hydrothermal treatment is shown in Fig. 4. Among 24 species studied in this study, 17 of  
219 them show a notable similarity in these two scattering features in the small angle  
220 regions, with the power-law exponent of about -1.4 and the peak positions at  $0.032$  to  
221  $0.04 \text{ \AA}^{-1}$ . Despite the taxonomical and ultrastructural differences found before the  
222 treatment, the hydrothermal treatment at  $200 \text{ }^\circ\text{C}$  converges to a similar structure in the

223 cell wall at a nanometric length scale.



224

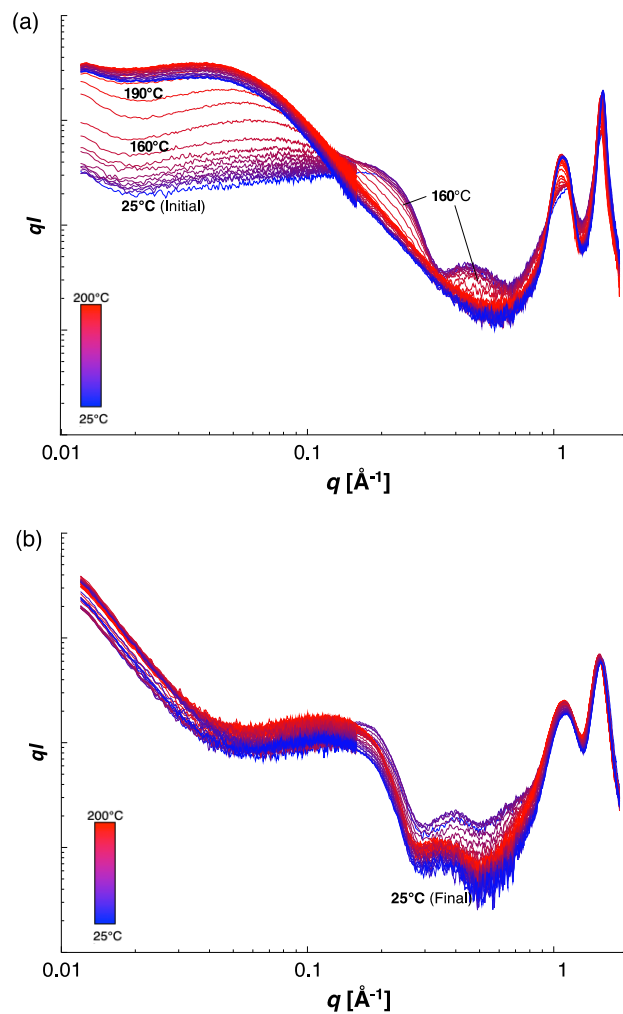
225 **Figure 4.** Relationship between power-law slope constants and peak positions of the  
226 main correlation peak of 24 wood species after hydrothermal treatment. Numbers  
227 correspond to those in Table 1.

228

### 229 **Structural changes during the in-situ experiment**

230 To follow the structural changes due to the hydrothermal treatment, we performed in  
231 situ S/WAXS measurements during the hydrothermal treatment of beech wood (*Fagus*  
232 *crenata*) at 200 °C, as shown in Fig. 5. This series of the equatorial scattering profiles  
233 indicates that these changes occurred only in the water-saturated sample but not in the  
234 air-dried sample (Fig. 5(a) and (b)). All the major structural changes happened during  
235 the heating process, and the cooling did not affect the structure. The increase in the  
236 streak intensity at  $q < 0.2 \text{ \AA}^{-1}$  started at 160 °C. Fig. S1(a)). The  $0.3\text{-}\text{\AA}^{-1}$  peak weakened  
237 and eventually disappeared during the heating from 130°C to 150°C. The  $0.15\text{-}\text{\AA}^{-1}$  peak  
238 also disappeared during the heating, where the intensity weakening started at around  
239 160 °C. This temperature range of around 160 °C is typically the onset of autohydrolysis

240 of hemicelluloses (Santucci et al. 2015). Thus, the loss of the  $0.15 \text{ \AA}^{-1}$  peak may  
241 originate from the collapse of cellulose microfibrils induced by removing  
242 hemicelluloses from the interfibrillar space. This possible mechanism also explains the  
243 less pronounced structural change in the air-dried sample, as the extensive  
244 autohydrolysis requires excess water in the system.



245  
246 **Figure 5.** In situ S/WAXS profiles of the equatorial component during hydrothermal  
247 treatment. Green (a) and air-dry (b) beech wood (*F. crenata*).

248  
249 **Conclusion**



250 Most hardwoods show similar equatorial scattering features that represent the  
251 structure in the cross-section of the cell wall, in the  $q$  range spanning 3 decades between  
252 0.004 and  $2 \text{ \AA}^{-1}$ . In a water-swollen native state, there is a prominent correlation peak  
253 between 0.1 and  $0.2 \text{ \AA}^{-1}$  and a second peak between 0.2 and  $0.4 \text{ \AA}^{-1}$ . These correlation  
254 peaks disappear upon hydrothermal treatment above  $160^\circ\text{C}$ , accompanied by a buildup  
255 of a correlation peak in the  $0.03 - 0.04 \text{ \AA}^{-1}$  range. Some exceptions were seen in  
256 cellulose-rich samples, where the correlation peaks between 0.1 and  $0.4 \text{ \AA}^{-1}$  are **absent**  
257 in the native state. The hydrothermal treatment is probably leaching out the matrix  
258 component between the microfibrils leading to the collapse of microfibrils with each  
259 other in the normal wood. **In contrast**, cellulose microfibrils are already collapsed to  
260 each other in the native state in cellulose-rich cell walls.

261

#### 262 **Competing interests**

263 The authors have no relevant financial or non-financial interests to disclose.

#### 264 **Consent for publication**

265 All authors have provided their consent for publication.

#### 266 **Availability of data**

267 All data generated or analyzed during this study are included in this article and the  
268 supporting information file.

#### 269 **Authors' contributions**

270 T.K., Y.O., and Y.N. conceptualized the study. T.K., Y.O., I.M., and Y.N. performed  
271 the experiments. T.K., Y.O., and Y.N. analyzed the data. T.K., Y.O., and Y.N. wrote the  
272 manuscript. Y.M. and Y.N. supervised the study. All authors read and approved the final  
273 manuscript.

#### 274 **Acknowledgements and funding.**

275 This study was supported by JSPS KAKENHI Grant Number JP17J05156 (T.K.).  
276 The dried wood specimens were obtained through collaborative research using wood  
277 collection databases with Research Institute for Sustainable Humanosphere (RISH)  
278 (Y.M. and T.K.). We thank Prof. Junji Sugiyama and Hajime Sorimachi (Kyoto  
279 University, Japan) for advising us on the selection of wood specimens. We thank the  
280 support of Prof. Yukie Saito (The University of Tokyo, Japan) and The University of  
281 Tokyo Chichibu Forest and Arboricultural Research Institute (UTCF) during the  
282 sampling of beech green wood specimens. We acknowledge Glyco@Alps (ANR-15-  
283 IDEX-02) for the financial support. The wide-angle X-ray detector (WOS) was funded  
284 by the French National Research Agency (ANR) under the ‘Investissement d’Avenir’  
285 program (Grant No. ANR-11-EQPX-0010). ESRF is acknowledged for the provision of  
286 beamtimes (experiment numbers A02-1-892 and A02-1-864, D2AM beamline).

#### 287 **Ethics approval and consent to participate**

288 Not applicable.

289

#### 290 **References**

291 Ashiotis G, Deschildre A, Nawaz Z, et al (2015) The fast azimuthal integration Python  
292 library: pyFAI J Appl Crystallogr 48:510–519.  
293 <https://doi.org/10.1107/S1600576715004306>

294 Chen P, Li Y, Nishiyama Y, et al (2021) Small Angle Neutron Scattering Shows  
295 Nanoscale PMMA Distribution in Transparent Wood Biocomposites. Nano Lett  
296 21:2883–2890. <https://doi.org/10.1021/acs.nanolett.0c05038>

297 Driemeier C, Mendes FM, Santucci BS, Pimenta MTB (2015) Cellulose co-  
298 crystallization and related phenomena occurring in hydrothermal treatment of  
299 sugarcane bagasse. Cellulose 22:2183–2195. [https://doi.org/10.1007/s10570-015-](https://doi.org/10.1007/s10570-015-0638-7)  
300 [0638-7](https://doi.org/10.1007/s10570-015-0638-7)

301 Fernandes AN, Thomas LH, Altaner CM, et al (2011) Nanostructure of cellulose  
302 microfibrils in spruce wood. Proc Natl Acad Sci 108:E1195–E1203.  
303 <https://doi.org/10.1073/pnas.1108942108>

304 Garrote G, Domínguez H, Parajó JC (1999) Hydrothermal processing of lignocellulosic  
305 materials. Holz als Roh - und Werkst 57:191–202.  
306 <https://doi.org/10.1007/s001070050039>

307 Jakob HF, Fratzl P, Tschegg SE (1994) Size and Arrangement of Elementary Cellulose  
308 Fibrils in Wood Cells: A Small-Angle X-Ray Scattering Study of *Picea abies*. *J*  
309 *Struct Biol* 113:13–22. [https://doi.org/https://doi.org/10.1006/jsbi.1994.1028](https://doi.org/10.1006/jsbi.1994.1028)

310 Jakob HF, Tschegg SE, Fratzl P (1996) Hydration Dependence of the Wood-Cell Wall  
311 Structure in *Picea abies*. A Small-Angle X-ray Scattering Study. *Macromolecules*  
312 29:8435–8440. <https://doi.org/10.1021/ma9605661>

313 Kocafe D, Huang X, Kocafe Y (2015) Dimensional stabilization of wood. *Curr For*  
314 *Reports* 1:151–161. <https://doi.org/10.1007/s40725-015-0017-5>

315 Kuribayashi T, Ogawa Y, Rochas C, et al (2016) Hydrothermal Transformation of  
316 Wood Cellulose Crystals into Pseudo-Orthorhombic Structure by Cocrystallization.  
317 *ACS Macro Lett* 5:730–734. <https://doi.org/10.1021/acsmacrolett.6b00273>

318 Kuribayashi, T., Ogawa, Y., Matsumoto, et al (2019). Changes in Crystal Structure of  
319 Cellulose in Hardwood Cell Walls by Hydrothermal Treatment at 200 °C under 1.6  
320 Mpa. *Mokuzai Gakkaishi*, 65:212-217.

321 Nishiyama Y, Langan P, O'Neill H, et al (2014) Structural coarsening of aspen wood by  
322 hydrothermal pretreatment monitored by small- and wide-angle scattering of X-  
323 rays and neutrons on oriented specimens. *Cellulose* 21:1015–1024.  
324 <https://doi.org/10.1007/s10570-013-0069-2>

325 Nitsos CK, Matis KA, Triantafyllidis KS (2013) Optimization of hydrothermal  
326 pretreatment of lignocellulosic biomass in the bioethanol production process.  
327 ChemSusChem 6:110–122. <https://doi.org/10.1002/cssc.201200546>

328 Nitsos CK, Choli-Papadopoulou T, Matis KA, Triantafyllidis KS (2016) Optimization  
329 of hydrothermal pretreatment of hardwood and softwood lignocellulosic residues  
330 for selective hemicellulose recovery and improved cellulose enzymatic hydrolysis.  
331 ACS Sustain Chem Eng 4:4529–4544.  
332 <https://doi.org/10.1021/acssuschemeng.6b00535>

333 Paajanen A, Zitting A, Rautkari L, et al (2022) Nanoscale Mechanism of Moisture-  
334 Induced Swelling in Wood Microfibril Bundles. Nano Lett 22:5143–5150.  
335 <https://doi.org/10.1021/acs.nanolett.2c00822>

336 Penttilä PA, Rautkari L, Österberg M, Schweins R (2019) Small-angle scattering model  
337 for efficient characterization of wood nanostructure and moisture behaviour. J  
338 Appl Crystallogr 52:369–377. <https://doi.org/10.1107/S1600576719002012>

339 Romaní A, Garrote G, Alonso JL, Parajó JC (2010) Bioethanol production from  
340 hydrothermally pretreated Eucalyptus globulus wood. Bioresour Technol  
341 101:8706–8712. <https://doi.org/10.1016/j.biortech.2010.06.093>

342 Sandberg D, Kutnar A (2016) Thermally modified timber: Recent developments in  
343 Europe and North America Wood Fiber Sci 48:28–39

344 Santucci BS, Maziero P, Rabelo SC, et al (2015) Autohydrolysis of Hemicelluloses  
345 from Sugarcane Bagasse During Hydrothermal Pretreatment: a Kinetic Assessment.  
346 BioEnergy Res 8:1778–1787. <https://doi.org/10.1007/s12155-015-9632-z>

347 Viljanen M, Ahvenainen P, Penttilä P, Svedström K. (2020) Ultrastructural X-ray  
348 scattering studies of tropical and temperate hardwoods used as tonewoods. IAWA  
349 J., 41:301–319. <https://doi.org/10.1163/22941932-bja10010>

350 Wojtasz-Mucha J, Hasani M, Theliander H (2017) Hydrothermal pretreatment of wood  
351 by mild steam explosion and hot water extraction. Bioresour Technol 241:120–126.  
352 <https://doi.org/10.1016/j.biortech.2017.05.061>

353