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The Monogenic Landscape of Human Infectious Diseases

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Abstract

The spectrum of known monogenic inborn errors of immunity is growing, with certain disorder underlying a specific and narrow range of infectious diseases. These disorders reveal the core mechanisms by which these infections occur in various settings, including inherited and acquired immunodeficiencies, thereby delineating the essential mechanisms of protective immunity to the corresponding pathogens. These findings also have medical implications, facilitating diagnosis and improving the management of individuals at risk of disease.

Keywords: Mendelian and monogenic infections, Inborn errors of immunity, infections, protection against common pathogens

Abbreviations:

IEI: inborn errors of immunity GWAS: genome-wide association studies MSMD: Mendelian susceptibility to mycobacterial disease EM: environmental mycobacteria AR: autosomal recessive XL: X-linked recessive **TB:** Tuberculosis Mtb: Mycobacterium tuberculosis ROS: Reactive oxygen species CMC: Chronic mucocutaneous candidiasis AD: Autosomal dominant GOF: gain-of-function IFD: Invasive fungal diseases auto-Abs: autoantibodies MAC: membrane attack complex TLR: Toll-like receptor TIR: Toll-like and IL-1 receptor LTA: lipoteichoic acid CRP: C reactive protein EV: Epidermodysplasia verruciformis HPV: Human papillomavirus NMSC: non-melanoma skin cancer APC: antigen-presenting cell DC: dendritic cells **RW: Recalcitrant warts** HSV-1: Herpes simplex virus 1 HSE: Herpes simplex encephalitis h- and i- PSC: human and induced pluripotent stem cell CNS: central nervous system EBV: Epstein Barr virus WD: Whipple's disease DARC: Duffy antigen receptor for chemokines HBGA: histo-blood group antigens

Introduction

Inborn errors of immunity (IEI) were initially thought to affect all leukocyte subsets and to predispose individuals to a wide range of pathogens ^[1, 2]. IEI have been classified into combined immunodeficiency groups (see Tables 1 and 2 of the classification of IEI) ^[3]. However, with the increasing number of IEIs being discovered, some exhibit a very narrow spectrum of susceptibility, often limited to a single pathogen [4-8]. In this review, susceptibility is defined as the unusual recurrence or severity of infection with a given pathogen that necessitates medical intervention or hospitalization. These patients are often cared for by specialized centers and undergo ancillary investigations (e.g., immunophenotyping, biopsy, etc.). For monogenic disorders, susceptibility arises from defects having a strong individual effect in increasing the risk of developing a specific infectious disease. This risk is classically measured by odds ratio (OR) or relative risk, with monogenic defects associated with high ORs (e.g. >10)^[9]. Although these disorders are more common than previously thought and can manifest with a full spectrum of clinical manifestations, ranging from localized symptoms to severe and potentially lethal conditions in otherwise healthy individuals, they highlight the specific involvement of and impact on distinct immunological pathways, and therefore underscore redundant pathways for other pathogens. However, we define a syndromic disorder as a clinical phenotype characterized by the association of a primary infectious disease with additional clinical manifestations, whether infectious or not. The advent of next-generation sequencing has significantly improved our ability to provide a genetic diagnosis for an ever-growing number of these conditions, allowing for more precise and tailored medical interventions ^[10]. In this review, we will attempt to highlight the most important IEIs conferring a predisposition to narrow infectious diseases, focusing particularly on susceptibility to mycobacterial diseases (caused by both weakly virulent non-tuberculous and highly virulent tuberculous species), and certain bacterial, fungal, and viral diseases, and highlighting the genes recently implicated in IEI and the cellular and molecular functions affected in patients. Conversely, we also highlight genetic variants important for resistance to infection. We will not discuss genetic investigations based on genome-wide association studies (GWAS), which are more commonly used to identify common variants for complex traits through large-scale comparisons of patients and controls, scanning the entire genome, and detecting several loci with possible small independent effects on the disease. These approaches have been previously reviewed [11-13]. Instead, we focus here on the genetic investigation of patients (and their relatives) to identify pathogenic causal variants with a strong effect (Table 1). We will deal, in particular, with monogenic predispositions, in which the causal relationship between the genotype and phenotype has been demonstrated.

Inborn errors of immunity underlying mycobacterial diseases

Mendelian susceptibility to mycobacterial disease (MSMD)

MSMD is a rare group of IEI characterized by a selective predisposition to clinical disease caused by weakly virulent mycobacterial species, such as Mycobacterium bovis-BCG vaccines and environmental mycobacteria (EM), in otherwise healthy individuals normally resistant to other microbes [14, 15]. MSMD occurs in ~ 1/50,000 individuals of diverse ancestries. About half the patients are also particularly susceptible to Salmonella and a significant proportion of MSMD patients also suffer from mucocutaneous infection with Candida species ^[16]. The genetic dissection of MSMD has identified deleterious variants in 22 genes underlying 46 different genetic etiologies^[14, 17-23], considering factors such as mode of inheritance, whether the defect is complete or partial, whether the mutant protein being expressed, and the specific function affected (e.g. phosphorylation)(Figure 1). They are all physiologically related and impair the activity of interferon-gamma (IFN-y), the "macrophage-activating factor"^[24, 25]. ZNFX1 is the only gene implicated in MSMD for which the underlying mechanism has yet to be determined [26]. The essential role of IFN-y in protective immunity to mycobacteria was confirmed with the identification of the first patients with complete IFN-y deficiency in 2020 ^[27], showing that IFN-y is the only agonist of its receptor. The last four years have revealed four new molecular players in MSMD: TBX21, MCTS1, IRF1, and CCR2. Patients with autosomal recessive (AR) T-bet/TBX21 deficiency display impaired development of the NK, iNKT, MAIT, Vδ2+ γδ T and TH1 lymphoid cell subsets and impaired IFN-y production by innate and innate-like lymphocytes ^[19]. Xlinked recessive (XL)-MCTS1 deficiency impairs JAK2 synthesis, leading to impaired IL-23-dependent IFNy production ^[18]. AR IRF1 deficiency highlights the essential role of this protein in IFN-y dependent macrophagic immunity to mycobacteria and its probable redundancy in type I IFN-mediated immunity in humans [22]. Finally, AR CCR2 deficiency may underlie MSMD in some patients due to migration of too few peripheral blood monocytes to tissues ^[17]. The identification of new genetic disorders of MSMD should extend the list of molecules known to be involved in the control of human IFN-y immunity.

Tuberculosis (TB)

Tuberculosis, caused by the highly virulent Mycobacterium tuberculosis (Mtb), remains a devastating disease ^[28, 29]. However, building on the discoveries made for MSMD, rare patients suffering solely from TB and homozygous for variants causing deficiencies of IL-12R^β1 and TYK2, for example, have been identified ^[5, 8, 15, 16, 30, 31], providing the first proof-of-principle that TB can be a monogenic disorder, upon infection with *Mtb* that acts as a trigger ^[9, 30] (Figure 1). New genetic etiologies, such as AR ITK and PD1 deficiencies, have been shown to underlie TB [32-34] through the impairment of IFN-y-mediated immunity [35]. The later has been corroborated by the susceptibility to mycobacterial disease observed in patients treated with anti-PD1/PDL1 antibodies ^[36, 37]. Impaired IFN-y production was also observed in the first two PDL1-deficient patients ^[38]. In addition, five forms of AR TYK2 deficiency were characterized, based on expression, pathway affected and minor allele frequency of the deleterious variants [39, 40], all affecting IL-23-dependent IFN-y-mediated immunity and underlying susceptibility to mycobacterial diseases, highlighting the role of IL-23 in anti-TB immunity ^[39, 40]. Moreover, homozygosity for a common missense TYK2 variant (P1104A) was identified as the first common monogenic cause of tuberculosis in two different patient cohorts^[41, 42]. P1104A has been subject to negative selection over the last 4,000 years in Europe, probably due to the selective pressure exerted by TB^[43, 44]. Homozygosity for this variant was probably responsible for 10 million deaths (1%) from TB over the last 2,000 years ^[42]. Homozygosity for P1104A specifically impairs (but does not abolish) IL-23-mediated IFN-y-dependent immunity, which is exerted mostly by MAIT and V $\delta 2^+ \gamma \delta$ Tcells ^[23], challenging the conventional doctrine and demonstrating that IL-23 is predominantly an antimycobacterial cytokine. Two patients with complete AR TNF deficiency and recurrent TB were recently identified (Figure 1). Reactive oxygen species (ROS) production is impaired in the patients' alveolar-like macrophages but not in other myeloid cells, explaining the narrow clinical phenotype of the patients ^[45] and revealing the redundant nature of TNF for other infectious diseases. The discovery of additional IEI underlying TB is anticipated, with different genetic architectures depending on ethnicity and inheritance.

· Inborn errors of immunity underlying fungal diseases

Non-invasive fungal diseases

Chronic mucocutaneous candidiasis (CMC) manifests as persistent or recurrent infections of the skin, mucosa, and nails with Candida spp. Severe CMC disease is associated with weight loss, failure to thrive, and complications, such as esophageal stricture, squamous cell carcinoma, and cerebral aneurysms in some cases [46, 47]. The first IEI of the IL-17A/F signaling pathway were identified in 2011 [48, 49]. Autosomal dominant (AD) IL-17F, and AR IL-17RA, IL-17RC, and ACT1 deficiencies were described in patients with CMC who were otherwise healthy apart from mucocutaneous diseases caused by Staphylococcus aureus in some patients (Figure 2). In parallel, IEI affecting the production alone or both the production of and response to IL-17A/F were progressively described and deciphered, mostly in patients with syndromic CMC (ie: CMC and additional symptoms, infectious or not). AD JNK1 or IL6ST deficiency, AR IL-23R, RORy/RORyT, ZNF341, or cRel deficiency, and AD STAT1 gain-of-function (GOF) have all been described in patients with mild or severe CMC and other infectious and/or autoimmune features. These IEI were shown to impair the production of IL-17A/F, probably underlying the CMC observed in the patients ^[48-51]. CMC was recently reported in a third of the patients from a cohort of individuals with complete AR IL-23R deficiency and abolished responses to IL-23, a cytokine reported to be essential for the maintenance of Th17 cells in mice ^[23]. These findings suggest that human IL-23 plays a crucial role in IL-17A/F-mediated protection against mucocutaneous Candida disease in a subset of affected individuals, whereas it appears to be redundant in most individuals ^[23]. None of the patients with IEI of IL-17-mediated immunity reported (except for a few of the patients with AD STAT1 GOF and most patients with AR CARD9 deficiency) have been documented to have invasive candidiasis, suggesting that IL-17A/F play a redundant role in protection against invasive C. albicans disease.

Invasive fungal diseases (IFD)

The second emblematic example of an IEI underlying fungal diseases is AR CARD9 deficiency ^[52, 53]. Over 90 patients have now been reported, all suffering from mucocutaneous or invasive diseases caused exclusively by fungal pathogens, with a disease onset ranging from early childhood to late adulthood. CARD9 is expressed primarily by myeloid cells and serves as an adaptor downstream from C-type lectin receptors (e.g. dectin-1), promoting the production of proinflammatory cytokines and chemokines upon fungal infection ^[53]. The fungal diseases concerned include superficial and deep dermatophytosis,

superficial and invasive Candida infections, subcutaneous and invasive phaeohyphomycosis caused by fungi such as Exophiala spp., Corynespora cassiicola, and Phialophora verrucosa. Rare cases of mucormycosis or aspergillosis have been reported [49]. By contrast, no cases of cryptococcosis "reported in some patients with neutralizing autoantibodies (auto-Abs) against GM-CSF", Pneumocystis jirovecii pneumonia "frequently reported in patients with IEI of the IKAROS zinc finger transcription factor family", or cases following dimorphic fungal infections have yet been reported in patients with AR CARD9 deficiency. Impaired CARD9-dependent induction of Th17 cytokines and neutrophil-recruiting chemokines (CXCL1, CXCL2 or CXCL5 ^[54]) has been identified as a potentially important mechanism underlying superficial and invasive fungal diseases, respectively, in patients with CARD9 deficiency (Figure 2). Biallelic deleterious variants of CLEC7A, encoding dectin-1 (a β-glucan-binding receptor), which signals through CARD9, have recently been reported in patients with severe C. cassiicola phaeohyphomycosis [55]. However, human dectin-1 deficiency is a frequent condition, with the p.Y238* allele present at a frequency of about 7% in European populations and up to 40% in the San population of South Africa, suggesting that it cannot be considered per se as an IEI underlying rare C. cassiicola phaeohyphomycosis, but should instead be seen as a risk factor. Further studies of the molecular and cellular bases of isolated CMC and IFD in humans with inherited CARD9 deficiency should shed light on the mechanisms of antifungal immunity in humans.

Inborn errors of immunity underlying pyogenic bacterial diseases

Neisseria disease

Diseases due to *Neisseria* result in an estimated 600,000 deaths worldwide^[56]. Complement deficiencies, first identified in the early 1970s, confer a predisposition to *Neisseria* infections^[57]. The complement system — discovered by Bordet in 1896 and shown to have antibacterial properties — is a proteolytic cascade of multiple proteins ^[58, 59] defining three activation pathways: the classical, mannose, and alternative complement pathways. All these pathways can cleave C3, the central component, and activate the terminal pathway^[58, 59]. Genetic studies of patients with a particularly susceptibility only to *Neisseria* infections ^[60-62]. These terminal components form the membrane attack complex (MAC) on the surface of the bacterium, creating a pore that kills it by osmolysis ^[63]. In 2023, two studies suggested a role for C5 in human immunity

to fungi^[64, 65], but these studies were unable to explain the absence of fungal infections in hundreds of previously reported C5-deficient patients. Inherited deficiencies of early alternative pathway components (XLR properdin, AR factor D, AR factor B deficiencies) ^[60, 62] and classical pathway components (AR C1q, AR C1r, AR C1s, AR C2, AR C3, AR C4, AR, AR factor H, AR factor I deficiencies) increase susceptibility to *Neisseria*, but also to pneumococcal diseases, and such deficiencies may also be associated with autoimmune conditions ^[60, 62]. Investigations of complement are therefore essential in child and adult patients who have suffered from invasive infections due to encapsulated bacteria, even after a single episode ^[66, 67]. Deficiencies of components of the mannose pathway do not cause Mendelian disorders ^[68]. Interestingly, alleles underlying deficiencies of C2, C6, and C9 are present in the European, African and Asian populations, respectively, each with an minor allele frequency close to 1%^[69]. The high frequencies of these deleterious alleles, at least in certain populations, are intriguing and warrant further investigation through population and evolutionary genetics.

Pneumococcal disease

Pneumococcal diseases cause an estimated 800,000 deaths annually worldwide^[59]. In 1953, Bruton reported that immunoglobulins play an important role in controlling *S. pneumoniae* infection ^[1]. Studies of patients with agammaglobulinemia led to the discovery of the most common genetic etiology underlying this condition: X-recessive BTK deficiency ^[70, 71]. It is now well established that IEI affecting the development, differentiation or maturation of B cells confer a predisposition to *S. pneumoniae* ^[72]. More than 50 genetic etiologies with an impact on the quantity or quality of immunoglobulins have now been identified and recently reviewed ^[73]. Complement studies revealed that the classical complement pathway and antibodies binding to the polysaccharide on the surface of *S. pneumoniae* cooperate to initiate opsonization of the bacteria (see above). Patients with AR deficiencies of C2, C3 and C4 may therefore develop severe pneumococcal disease, but they may also present autoimmunity. From 2001 onwards, studies of a series of patients with Toll-like receptor (TLR)-IL-1R (TIR)-NF-κB pathway deficiencies (XLR-IKBKG, AD NFKBIA, AR HOIL-1 and AR HOIP deficiencies)^[74-79], in addition to the identification of IRAK-4^[80] and MyD88^[81] deficiencies, highlighted the crucial role of TLR/IL-1R pathway in the myeloid lineage for controlling pneumococcal infections in humans and defining narrow non-redundant antibacterial immunity.

However, the effector mechanism remains unknown. These three key non-redundant mechanismscomplement and myeloid cells (both innate immunity) and anti-polysaccharide antibodies (humoral immunity)-are required to protect humans against pneumococcal disease. The absence of a genetic diagnosis for most people suffering from pneumococcal disease ^[66] despite active vaccination, suggests the presence of other immune mechanisms capable of controlling this disease.

Staphylococcal disease

Staphylococcal diseases lead to an estimated 1,000,000 deaths annually worldwide [56]. Human phagocytes play a key role in defenses against S. aureus infection^[82]. Indeed, IEI such as congenital neutropenia^[83] and phagocytosis defects^[84, 85] confer a predisposition to S. aureus infections. Disorders of the TLR/IL-1R -NF-kB pathway (XLR-IKBKG, AD NFKBIA, AR IRAK4 and AR MyD88 deficiencies)[74-81], confer a predisposition to severe S. aureus and a few other infections. The description of inherited TIRAP deficiency, which is an adaptor protein interacting with TLR2 and TLR4, and acts upstream of MyD88 and IRAK-4, has narrowed the search for the main player in selective predisposition to S. aureus infection to TLR1/2 and TLR2/6^[86]. The incomplete penetrance of staphylococcal infection among individuals with AR TIRAP deficiency in this kindred was explained by the production of anti-lipoteichoic acid antibody in all relatives tested but not in the patient, which rescue TLR2-dependent immunity to staphylococcal LTA. Studies of patients with human OTULIN haploinsufficiency have recently shed light on the essential role of toxin receptor expression in immunity. OTULIN haploinsufficiency in fibroblasts dysregulates CAVEOLIN/ADAM10 expression on the cell surface, increasing the cells' sensitivity to a-toxin. It has indicated that other, non-hematopoietic immune cells also play crucial roles in controlling pyogenic infections through intrinsic mechanisms^[87]. Incomplete penetrance for staphylococcal infection in individuals with OTULIN haploinsufficiency has been shown to be due to the presence of anti- α -toxin antibody [87]. Finally, further investigations into patients suffering from staphylococcal diseases and hyper-IgE syndrome ultimately led to the discovery of the first patients with AR IL-6 receptor deficiency [88-90]. The studies of these patients have revealed the importance of IL-6 in T-cell differentiation (circulating follicular helper cells and Th17 T cells) confirming the observation described in patients with AD STAT3, or AD or AR GP130 deficiency (encoded by IL6ST), who are characterized by a deficit of STAT3-dependent IL-6

response ^[91-93]. Since 2008, a few patients with recurrent subcutaneous staphylococcal abscesses or other severe pyogenic infections have been reported to have neutralizing auto-Abs against IL-6 ^[94-96]. Similarly, severe bacterial infections have been reported in patients treated with Tocilizumab ^[97]. Auto-Abs against IL-6 should therefore be considered in patients, particularly adults, with unexplained bacterial infections, especially when there is no adequate increase in CRP levels during the infection. Together, these discoveries demonstrate the key role of IL-6 in immunity to staphylococci and highlight the contribution of adaptive immunity.

• Inborn errors of immunity underlying viral diseases of the skin

Epidermodysplasia verruciformis (EV)

Human papillomaviruses (HPVs) are small DNA viruses, each with a specific tropism to mucosal (α -) or cutaneous (α -, β -, γ -, μ - and ν -) tissues. In the general population, the α -, γ -, μ - and ν -HPVs cause benign cutaneous common and plantar warts^[98]. The α -HPVs also cause anogenital and oropharyngeal cancers, whereas β -HPVs generally cause asymptomatic infections. Some β -HPVs are responsible for flat wart-like lesions and non-melanoma skin cancer (NMSC) in rare patients with epidermodysplasia verruciformis (EV)^[99, 100]. In 1946, EV was the first Mendelian IEI described, characterized by a narrow range or "isolated" form of infections^[99-101]. Three IEI have been identified in patients with isolated EV: EVER1, EVER2 and CIB1 deficiencies^[101-103] (Figure 3). CIB1, EVER1, and EVER2 form a complex that binds E5 and E8, governing keratinocyte-intrinsic immunity to β -HPVs and potentially acting as a viral restriction factor^[104]. Rare patients with EV and other phenotypes, mostly infectious, referred to as atypical, or "syndromic" forms of EV^[101] present various inherited disorders of T cells, including RhoH, CORO1A, DOCK8, LCK, TPP2, STK4, and ITK deficiencies^[32, 105-110]. Complete AR FLT3L deficiency was recently reported in a multiplex family with syndromic EV or recalcitrant warts. FLT3L deficiency induces a severe deficit of antigenpresenting cells (APCs) of all kinds, including dendritic cells. FLT3L-deficient individuals have a similar skin composition to healthy donor, with the presence of Langerhans cells and macrophages, but with much lower levels of dermal DCs. These findings strongly suggest a non-redundant role of dermal DCs in the control of cutaneous HPVs [111]. Together, these data demonstrate that both isolated and syndromic EV display genetic heterogeneity but physiological homogeneity. Both cellular and intrinsic immune responses

are therefore involved in cutaneous immunity to EV-HPVs. Disorders of T-cell responses underlie syndromic EV and other HPV-related skin infections, whereas disorders of keratinocytes underlie isolated EV.

Recalcitrant warts (RW)

In rare cases, common warts, mostly due to α -HPVs, can spread and persist, in which case they are described as recalcitrant warts (RW)^[112]. Unlike EV, RW often present as a syndromic form, with multiple HPV types or manifestations (cutaneous, mucosal). The best characterized of these genetic defects are those in patients with monoallelic mutations of CXCR4 [113-115] and GATA2 [116-119], and biallelic mutations of DOCK8^[120-122], who suffer from RW in 80, 50 and 40%, respectively, of cases^[105, 123]. These IEI with a high penetrance for HPV diseases may selectively affect APCs counts, as recently reported for FLT3L deficiency^[111], or both APCs and T lymphocytes counts, migration or function (Figure 3). Other IEI affecting mostly T lymphocytes have also been reported in patients with severe HPV infection^[105, 123]. Many of these IEI impair αβ T-cell counts, as reported for TRAC and IL-7 deficiencies^[124-126], or primarily CD4⁺ T-cell counts, as reported for STK4 deficiency or the recently described SASH3 deficiency^[127-132]. Other IEI conferring a predisposition to RW are associated with normal or near-normal T-cell counts and differentiation. CD28 deficiency has been reported in patients with isolated RW, including one patient with tree man syndrome ^[133]. Interestingly multiple IEI conferring predisposition to HPV underlie a CD28 signaling defect. Deficiencies of CARMIL2, a cytoplasmic signaling molecule, impair NF-KB activation downstream from CD28^[134-136]. MAGT1 deficiency results in a glycosylation defect, the effects of which include an impairment of CD28 expression on T lymphocytes^[137]. AD CARD11 dominant-negative variants impair NF-κB activation upon CD3/CD28 costimulation ^[138], and CARD11 is a direct partner of CARMIL2^{[139,} ^{140]}. Together, these findings suggest a major role of the NF- κ B pathway downstream from CD28 in the control of skin papillomavirus infection by T cells. Furthermore, deficiencies of CD4 ^[141], another T cellcostimulating molecule, have recently been shown to underlie RW^[142] by an as yet unclear mechanism, possibly involving CD28 and LCK^[143]. Thus, studies of IEI have shown that impairments of the numbers or functions of APCs and T cells can confer a predisposition to severe RW.

• Inborn errors of immunity underlying viral encephalitis

Herpes simplex virus 1 (HSV-1) encephalitis (HSE) is the most common sporadic viral encephalitis in Western countries and, perhaps, worldwide. The causal role of HSV in HSE was established in 1941 [144], and HSV-1 was distinguished from HSV-2 in 1977 ^[145]. About 1-2/10,000 infected individuals develop lifethreatening HSE. In the early 2000s, it was hypothesized that childhood HSE might result from single-gene IEI to HSV-1 in the central nervous system (CNS) [146-148]. The evidence supporting genetic susceptibility to HSE came from two very rare cases of "syndromic" HSE observed in combination with mycobacterial diseases, in a child with AR complete STAT1 deficiency ^[149], and a child with a NEMO mutation ^[150-152]. From 2006 onwards, 12 IEI underlying the typical form of isolated childhood HSE were progressively discovered. Most of the HSE-causing IEI identified to date affect type I IFN-mediated antiviral immunity through mutations of genes encoding proteins involved in type I IFN production (TLR3, UNC93B1, TRIF, TRAF3, TBK1, IRF3, NEMO, GTF3A) or responsiveness (IFNAR1, STAT1, STAT2, TYK2)^[6, 149, 152-162] pathways (Figure 4). HSE-causing IEI disrupting novel type I IFN-independent antiviral mechanisms have also recently been shown to underlie HSE in other HSE patients, the genes encode DBR1 ^[163], snoRNA31 ^[164], RIPK3 ^[165] and TMEFF1 ^[166]. Unlike deficits of the type I IFN circuit that impair type I IFN-mediated first line antiviral responses in a broad range of tissues and cells, hence underlying various severe viral infections in humans including those resulted from live attenuated virus vaccination [40, 167, 168], the four HSEcausing IEI impairing type I IFN-independent antiviral immunity have been found only in patients with isolated HSE so far. Regardless of the molecular mechanisms, human pluripotent stem cell (hPSC)-derived forebrain neurons harboring HSE-causing IEIs display defective cell-intrinsic immunity to HSV-1, resulting in unrestricted HSV-1 replication in neurons. Such IEI therefore presumably underlie HSE pathogenesis in vivo, with incomplete clinical penetrance, accounting for up to 10% of the cases studied. The last two decades of human genetic and immunological studies have, therefore, progressively established a new paradigm that childhood HSE can result from inborn errors of CNS-specific, cell-intrinsic immunity to HSV-1, expanding the concept of host defense *in natura* from the immune system to the whole organism.

Inborn errors of immunity underlying viral pneumonia

The first IEI identified to cause life-threatening viral pneumonia was IRF7-deficiency in a patient with influenza^[169]. IRF7-deficient plasmacytoid DCs (pDCs) and induced pluripotent stem cell (iPSC)–derived

pulmonary epithelial cells (PECs) from this patient produced abnormally small amounts of type I and III IFNs in response to influenza A viruses (IAVs), promoting IAV proliferation^[169]. AD deficiencies of TLR3, and AR deficiencies of IRF9, STAT1, TYK2 and STAT2- have been identified in patients with life-threatening influenza pneumonia, further demonstrating that the type I IFN response in PECs is essential to control influenza infection^[40, 167, 168, 170-172] (Figure 4). Studies of critical COVID-19 pneumonia led to the discovery of a new IEI, XR TLR7 deficiency, accounting for more than 1% of cases of critical COVID-19 in men, especially those under 60 years of age^[173]. The blood B-cell lines and myeloid cell subsets of these patients did not respond to TLR7 stimulation and their pDCs produced low levels of type I IFNs in response to SARS-CoV-2 ^[173]. TLR7 is a MyD88/IRAK-4-dependent endosomal receptor for single-stranded RNA in pDCs. Patients with AR MyD88 or IRAK-4 deficiency have been shown to be at high risk of life-threatening COVID-19 due to impaired TLR7-dependent type I IFN production by pDCs^[174]. The discoveries of IE of TLR3- and IRF7-dependent type I IFN immunity, including AR deficiencies of IRF7, IRF9, IFNAR1, IFNAR2, STAT2, TYK2 and TBK1-, and AD deficiencies of TLR3 in patients with critical COVID-19 pneumonia suggested that influenza and COVID-19 pneumonia are allelic on five loci (IRF7, IRF9, STAT2, TYK2, and TLR3)^{[175-} ^{180]}, where variants on the same loci underlie both diseases. AR deficiencies of IFIH1, a RIG-I-like receptor involved in the sensing of viral RNA leading to the induction of type I interferons, have been reported in children suffering of severe respiratory illness due to respiratory syncytial virus, human rhinovirus or influenza ^[181, 182]. Intriguingly, the penetrance of all these inborn errors of type I IFN immunity is far from complete, even for the most virulent pathogen, SARS-CoV-2.

Inborn errors of immunity underlying EBV diseases

Epstein Barr virus (EBV) belongs to the γ-herpes family ^[183]. Most EBV infections are benign and symptomfree, with about 90% of adults worldwide experiencing infection at some time in their lives, but this virus has been implicated in about 1% of all cancer cases ^[184]. EBV primarily infects epithelial cells and B lymphocytes, establishing latency within B cells. Uncontrolled EBV infections can manifest as infectious mononucleosis (IM), hemophagocytic lymphohistiocytosis (HLH), and various B-cell proliferative disorders. CD8⁺ T-cell responses play a key role in immunity to EBV ^[183, 185, 186]. Two types of IEI corresponding to different lymphoproliferative disorders have been identified. The first group, characterized by variants of

PRF1, *RAB27A*, *UNC13D*, *SYT11*, *LYST* and *SH2D1A*, impairs the granule-dependent cytotoxicity function of NK and CD8⁺ T cells, leading to EBV-induced HLH. The second group comprises IEI in which EBV infection predominantly drives B-cell lymphoproliferative disorders, including B-cell lymphomas. Patients with IEI of this type present an impaired activation and expansion of EBV-specific T cells due to deficiencies of RASGRP1, ITK, MAGT1, DEF6, CTPS1, TNFRSF9, CD27 or its ligand CD70 ^[187, 188]. Patients with ITK deficiency are also susceptible to TB ^[32]. For this group of IEI, a recent report identified patients homozygous for loss-of-function variants of *IL27RA* displaying selective vulnerability to EBV^[189]. The phosphorylation of STAT1 and STAT3 by IL-27 was abolished in the T cells of these patients. This deficiency impairs the expansion of potent anti-EBV effector cytotoxic CD8⁺ T cells, probably due to the lack of a synergistic effect on T-cell receptor-dependent T-cell proliferation. In addition, a specific allele of *IL27RA* (rs201107107) for which enrichment has been detected in the Finish population is associated with a higher risk of developing IM when present in the homozygous state. Finally, neutralizing auto-Abs against IL-27 have also been reported in patients with sporadic IM and chronic EBV infection. These findings demonstrate the crucial role of *IL27RA* in immunity to EBV ^[189].

Autoimmune phenocopies of inborn errors of immunity

Some autoimmune disorders can phenocopy IEIs, resulting in an almost undistinguishable infectious phenotype^[190]. Auto-Abs against a component of the complement system (C3 convertase) has been shown to enhance its activity (thus being an "enhancing" auto-Ab), lowering complement levels and underlying severe bacterial infections such as meningitis due to *Neisseria*^[191], pneumonia due to *Streptococcus pneumoniae*^[192] or *Haemophilus influenza*^[193]. Autoimmune phenocopies of at least four cytokines have also been described^[72, 194, 195]: IL-6 and staphylococcal disease ^[94], IFN- γ and MSMD^[194, 196-199], IL-17A/F and autoimmune polyendocrinopathy syndrome type I (APS-1 or APECED syndrome /CMC^[200, 201] and GM-CSF and idiopathic pulmonary alveolar proteinosis and cerebral/pulmonary nocardiosis^[202-204] or cryptococcosis^[205-216]. More recently, auto-Abs neutralizing IFN- α and/or ω were detected in about 15% of cases of life-threatening COVID-19 pneumonia ^[217], 20% of the fatal COVID-19 pneumonia despite an appropriate Ab response to mRNA vaccine ^[221, 222]. The risk of critical disease increases with the

number and concentration of type I IFNs neutralized. These auto-Abs were also found to underlie about one third of the rare cases of severe adverse reactions to yellow fever YFV-17D live-attenuated viral vaccine $^{[223]}$, 5% of cases of critical influenza pneumonia $^{[176]}$, 20% of cases of critical Middle East respiratory syndrome (MERS) pneumonia $^{[224]}$, 40% of cases of West Nile virus encephalitis $^{[225]}$, and various herpes viral infections $^{[226-231]}$. They can be detected in 0.3-1% of individuals under the age of 65 years (including 0.2-2% of children), after which their prevalence increases sharply, reaching 4-8% in individuals over 70 years of age. Auto-Abs neutralizing type I IFNs are present before infection and are causal for severe viral diseases. Patients with neutralizing anti-IL-27 auto-Abs have recently been reported to be prone to Epstein– Barr virus diseases, like patients with AR IL-27RA deficiency^[189]. It remains unknown why they are produced, but their production can be genetically driven, as in patients with anti-IFN- γ auto-Abs and specific HLA class II haplotypes, or in a few patients with IEI of T-cell thymic selection and auto-Abs against type I IFNs ^[232, 233]. The clinical consequences may be much broader than initially thought, such as high titers of neutralizing auto-Abs against IL-10 recently reported in children with mild to severe colitis, phenotypically mimicking inborn errors of the IL-10 pathway ^[234].

Other infections studied from a monogenic angle

Other infectious diseases have also been investigated to identify the human genetic component involved. Whipple's disease (WD), caused by the bacterium *Trophyrema whipplei*, is associated with chronic or acute infections ^[235, 236]. Two genetic etiologies have been identified: AD IRF4 deficiency due to haploinsufficiency ^[237] and CD4 deficiency in patients with WD among other symptoms ^[141]. However, the pathophysiology of this disease remains unknown. Two mycobacterial diseases, Buruli ulcer, caused by *M. ulcerans* and recently shown to be transmitted by mosquitoes in Australia ^[238] and leprosy, caused by *M. leprae*, have been studied. A monogenic contribution to Buruli ulcer was recently reported, with the identification of a microdeletion on chromosome 8 encompassing a long non-coding (Inc) RNA gene expressed in the skin and located close to a cluster of defensin genes ^[239, 240], in a familial form of severe Buruli ulcer ^[241]. Recent studies of twins with early-onset leprosy led to identification of three rare variants of two genes (*LRRK2* and *NOD2*), with deleterious and additive effects on disease development ^[242]. Kaposi sarcoma is caused by infection with HHV8, a γ -herpes virus with oncogenic properties, the seroprevalence of which varies

considerably worldwide, from 20 to 40 % ^[243]. Three AR genetic etiologies have been linked to susceptibility to KS (IFNGR1, STIM1 and OX40), and three XL genetic etiologies have been linked to cutaneous KS (WAS, MAGT1 and CTLA4), highlighting the crucial role of T cells, and of OX40-dependent T-cell immunity in particular, for protection against HHV8 ^[244-246]. In addition, fulminant viral hepatitis, caused by hepatitis A virus (HAV) is a rare life-threatening disease, for which two genetic etiologies have been discovered: IL10RB ^[247] and IL18BP deficiencies ^[248]. These findings suggest that the molecular basis of FVH in these patients may involve excessive IFN-γ activity during HAV infections of the liver ^[249]. Finally, NOS2 deficiency was found to be associated with fatal CMV disease in an otherwise healthy adult ^[250]. Altogether, these studies suggest that human genetic determinants play an important role in the course of infectious diseases.

Mendelian resistance to infection

Mendelian susceptibility to infectious disease is indissociable from Mendelian resistance to infection. The role of Duffy antigen receptor for chemokines (DARC)-expressing erythrocytes in protection against Plasmodium vivax infection was first reported in the 1970s [251]. In Plasmodium vivax malaria, the parasite makes use of the interaction between its Duffy binding protein and DARC to invade reticulocytes. AR DARC deficiency, due to a single-nucleotide variant, inhibits DARC transcription in erythroid cells [252]. Natural selection, in areas in which Pv is endemic, led to an increase in the prevalence of resistance associated with an absence of the Duffy binding protein [253]. Similarly, FUT2-expressing intestinal epithelial cells play a key role in resistance to norovirus and rotavirus infections, which cause diarrheal diseases [254]. FUT2 controls the secretion of ABO histo-blood group antigens (HBGAs) at the surface of the gut, where they act as receptors for nororovirus and rotavirus. AR loss-of-function FUT2 variants confer a non-secretor phenotype, preventing the expression of HBGAs on the surface of the gut, making it more difficult for the virus to bind ^[255, 256]. However, the idea that HBGAs function as receptors is controversial and some authors have suggested that these antigens may simply enhance viral replication and disease development without being essential for infection per se [257]. FUT2 non-secretor status has also been linked to susceptibility to various diseases. Consistently, a complex pattern of natural selection has been described for FUT2, with most variants displaying a long history of balancing selection in Eurasian and African populations ^[258]. These advances in our understanding of monogenic resistance have highlighted the importance of non-

professional immune cells in protective immunity. Human genotypes at the DARC and FUT2 loci blocking the entry of parasites and viruses into non-professional immune cells have demonstrated how genetic variants in erythrocytes and the intestinal epithelium can have a life-saving effect. Finally, AR CCR5 deficiency, which has been found to confer resistance to HIV-1, illustrates the role of complete deficiencies of pathogen entry receptors in mechanisms of Mendelian resistance ^[259-261]. The most common loss-of-function variant of CCR5 is a 32-base pair deletion with a minor allele frequency of 10% in the European population. The genetic mechanisms of protection against severe infectious outcomes and resistance to infection are subject to positive selection, providing a survival advantage.

Future directions

The human genetic basis of infectious diseases remains elusive in most affected individuals, driving ongoing efforts to unravel the genetic determinants underlying diseases, including those occurring sporadically. In large cohorts of patients with narrow infectious phenotypes, the identification of causal genetic etiologies depends largely on the disease itself and its prevalence (ranging from 50% for MSMD to 1% for tuberculosis)^[8]. Other genetic hypotheses (e.g. weak clinical penetrance, digenic or oligenic models) have not yet been thoroughly tested and might explain many infectious phenotypes. Recent discoveries of common monogenic alleles in the general population — such P1104A TYK2 and the role of this allele in the homozvgous state in TB^[41] through a selective biochemical effect on IL-23-mediated IFN-y immunity, or D31N PTCRA ^[262], which increase the risk of autoimmunity — indicate promising future directions for IEI research. Digenic or oligogenic models with strong effects will probably shape the future of this field. Nevertheless, many patients still have no known genetic etiology, particular for the more frequent diseases. Widespread access to WES has already resolved many genetic cases, and the possibility of performing WGS combined with long-read technology may soon make it possible to determine not only the role of noncoding variants but also that of more complex variants involved in chromosome architecture, such as short and long repeats, CNVs, inversions, or translocations. The computing challenges posed by the large amounts of data generated are gradually being resolved. The bottleneck is not currently the identification of variants, but substantiating the deleterious nature of the variants identified and their causal role in the disease studied. Diagnosis based on the identification of a variant without functional validation persists and

poses inherent risks^[173, 262-264]. The ongoing elucidation of interindividual variability in responses to specific pathogens is improving our genetic understanding and providing deeper insight into the phenomenon of incomplete penetrance (Table 1). Furthermore, the identification of novel genetic causes of specific infectious diseases is enriching our knowledge of cellular, immunological, and pathophysiological processes in an unbiased manner. This should help to define the cell subsets and pathways crucial for the control of particular infections *in vivo* and will serve as a basis for the development of preventive and therapeutic strategies based on physiological mechanisms. These discoveries will also facilitate the development of genetic counseling for affected families.

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References

1. Bruton OC. **Agammaglobulinemia (congenital absence of gamma globulin); report of a case**. *Med Ann Dist Columbia* 1953; 22(12):648-650; passim.

2. Bonthron DT, Markham AF, Ginsburg D, Orkin SH. Identification of a point mutation in the adenosine deaminase gene responsible for immunodeficiency. *J Clin Invest* 1985; 76(2):894-897.

3. Tangye SG, Al-Herz W, Bousfiha A, Cunningham-Rundles C, Franco JL, Holland SM, et al. Human Inborn Errors of Immunity: 2022 Update on the Classification from the International Union of Immunological Societies Expert Committee. *J Clin Immunol* 2022; 42(7):1473-1507.

4. Casanova JL, Abel L. Human genetics of infectious diseases: Unique insights into immunological redundancy. *Semin Immunol* 2018; 36:1-12.

5. Casanova JL, Abel L. The human genetic determinism of life-threatening infectious diseases: genetic heterogeneity and physiological homogeneity? *Hum Genet* 2020.

6. Casanova JL, Abel L. Mechanisms of viral inflammation and disease in humans. *Science* 2021; 374(6571):1080-1086.

7. Casanova JL, Abel L. Lethal Infectious Diseases as Inborn Errors of Immunity: Toward a Synthesis of the Germ and Genetic Theories. *Annu Rev Pathol* 2021; 16:23-50.

8. Casanova JL, Abel L. From rare disorders of immunity to common determinants of infection: Following the mechanistic thread. *Cell* 2022; 185(17):3086-3103.

9. Casanova JL, Abel L. The Microbe, the Infection Enigma, and the Host. Annu Rev Microbiol 2024.

10. Carrasco-Ramiro F, Peiró-Pastor R, Aguado B. **Human genomics projects and precision medicine**. *Gene Therapy* 2017; 24(9):551-561.

11. Gibbs KD, Schott BH, Ko DC. **The Awesome Power of Human Genetics of Infectious Disease**. *Annu Rev Genet* 2022; 56:41-62.

12. Thorball CW, Fellay J, Borghesi A. **Immunological lessons from genome-wide association studies** of infections. *Curr Opin Immunol* 2021; 72:87-93.

13. Cobat A, Zhang Q, Effort CHG, Abel L, Casanova JL, Fellay J. Human Genomics of COVID-19 **Pneumonia: Contributions of Rare and Common Variants**. *Annu Rev Biomed Data Sci* 2023; 6:465-486.

14. Bustamante J, Boisson-Dupuis S, Abel L, Casanova JL. **Mendelian susceptibility to mycobacterial disease: genetic, immunological, and clinical features of inborn errors of IFN-gamma immunity**. *Seminars in immunology* 2014; 26(6):454-470.

15. Bustamante J. **Mendelian susceptibility to mycobacterial disease: recent discoveries**. *Hum Genet* 2020; 139(6-7):993-1000.

16. Casanova JL, Abel L. Genetic dissection of immunity to mycobacteria: the human model. *Annu Rev Immunol* 2002; 20:581-620.

17. Neehus AL, Carey B, Landekic M, Panikulam P, Deutsch G, Ogishi M, et al. **Human inherited CCR2** deficiency underlies progressive polycystic lung disease. *Cell* 2024; 187(2):390-408 e323.

18. Bohlen J, Zhou Q, Philippot Q, Ogishi M, Rinchai D, Nieminen T, et al. Human MCTS1-dependent translation of JAK2 is essential for IFN-gamma immunity to mycobacteria. *Cell* 2023; 186(23):5114-5134 e5127.

19. Yang R, Mele F, Worley L, Langlais D, Rosain J, Benhsaien I, et al. **Human T-bet governs both innate and innate-like adaptive IFN-γ immunity against mycobacteria**. *Cell* 2020; 183(7):1826-1847.

20. Khavandegar A, Mahdaviani SA, Zaki-Dizaji M, Khalili-Moghaddam F, Ansari S, Alijani S, et al. **Genetic**, immunologic, and clinical features of 830 patients with Mendelian susceptibility to mycobacterial diseases (MSMD): A systematic review. *J Allergy Clin Immunol* 2024.

21. Ogishi M, Yang R, Rosain J, Bustamante J, Casanova JL, Boisson-Dupuis S. **Inborn errors of human transcription factors governing IFN-gamma antimycobacterial immunity**. *Curr Opin Immunol* 2023; 81:102296.

22. Rosain J, Neehus AL, Manry J, Yang R, Le Pen J, Daher W, et al. **Human IRF1 governs macrophagic IFN-gamma immunity to mycobacteria**. *Cell* 2023; 186(3):621-645 e633.

23. Philippot Q, Ogishi M, Bohlen J, Puchan J, Arias AA, Nguyen T, et al. **Human IL-23 is essential for IFN-gamma-dependent immunity to mycobacteria**. *Science immunology* 2023; 8(80):eabq5204.

24. Dupuis S, Doffinger R, Picard C, Fieschi C, Altare F, Jouanguy E, et al. **Human interferon-gamma-mediated immunity is a genetically controlled continuous trait that determines the outcome of mycobacterial invasion**. *Immunol Rev* 2000; 178:129-137.

25. Nathan CF, Murray HW, Wiebe ME, Rubin BY. Identification of interferon-gamma as the lymphokine that activates human macrophage oxidative metabolism and antimicrobial activity. *J Exp Med* 1983; 158(3):670-689.

26. Le Voyer T, Neehus AL, Yang R, Ogishi M, Rosain J, Alroqi F, et al. **Inherited deficiency of stress** granule **ZNFX1** in patients with monocytosis and mycobacterial disease. *Proc Natl Acad Sci U S A* 2021; 118(15).

27. Kerner G, Rosain J, Guerin A, Al-Khabaz A, Oleaga-Quintas C, Rapaport F, et al. **Inherited human IFN-gamma deficiency underlies mycobacterial disease**. *J Clin Invest* 2020; 130(6):3158-3171.

28. WHO. Global Tuberculosis Report 2022. World Health Organization, Geneva; 2022 2022.

29. Bloom BR. A half-century of research on tuberculosis: Successes and challenges. *J Exp Med* 2023; 220(9).

30. Boisson-Dupuis S. The monogenic basis of human tuberculosis. *Hum Genet* 2020; 139(6-7):1001-1009.

31. Casanova JL, Abel L, Quintana-Murci L. **Immunology taught by human genetics**. *Cold Spring Harb Symp Quant Biol* 2013; 78:157-172.

32. Ogishi M, Yang R, Rodriguez R, Golec DP, Martin E, Philippot Q, et al. **Inherited human ITK deficiency impairs IFN-gamma immunity and underlies tuberculosis**. *J Exp Med* 2023; 220(1).

33. Ogishi M, Yang R, Gruber C, Zhang P, Pelham SJ, Spaan AN, et al. **Multibatch Cytometry Data Integration for Optimal Immunophenotyping**. *J Immunol* 2021; 206(1):206-213.

34. Ogishi M, Yang R, Aytekin C, Langlais D, Bourgey M, Khan K, et al. **Inherited PD-1 deficiency underlies tuberculosis and autoimmunity in a child**. *Nature Medicine* 2021; 27(9):1646-1654.

35. Casanova JL, MacMicking JD, Nathan CF. Interferon-gamma and infectious diseases: Lessons and prospects. *Science* 2024; 384(6693):eadl2016.

36. Anand K, Sahu G, Burns E, Ensor A, Ensor J, Pingali SR, et al. **Mycobacterial infections due to PD-1 and PD-L1 checkpoint inhibitors**. *ESMO Open* 2020; 5(4).

37. Tsai CC, Chen JH, Wang YC, Chang FY. **Re-activation of pulmonary tuberculosis during anti- programmed death-1 (PD-1) treatment**. *QJM* 2019; 112(1):41-42.

38. Johnson MB, Ogishi M, Domingo-Vila C, De Franco E, Wakeling MN, Imane Z, et al. **Human inherited PD-L1 deficiency is clinically and immunologically less severe than PD-1 deficiency**. *J Exp Med* 2024; 221(6).

39. Kreins AY, Ciancanelli MJ, Okada S, Kong XF, Ramirez-Alejo N, Kilic SS, et al. **Human TYK2** deficiency: Mycobacterial and viral infections without hyper-IgE syndrome. *J Exp Med* 2015; 212(10):1641-1662.

40. Ogishi M, Arias AA, Yang R, Han JE, Zhang P, Rinchai D, et al. **Impaired IL-23-dependent induction** of **IFN-gamma underlies mycobacterial disease in patients with inherited TYK2 deficiency**. *J Exp Med* 2022; 219(10).

41. Boisson-Dupuis S, Ramirez-Alejo N, Li Z, Patin E, Rao G, Kerner G, et al. **Tuberculosis and impaired IL-23-dependent IFN-gamma immunity in humans homozygous for a common TYK2 missense variant**. *Science immunology* 2018; 3(30):eaau8714.

42. Kerner G, Ramirez-Alejo N, Seeleuthner Y, Yang R, Ogishi M, Cobat A, et al. **Homozygosity for TYK2 P1104A underlies tuberculosis in about 1% of patients in a cohort of European ancestry**. *Proc Natl Acad Sci U S A* 2019; 116(21):10430-10434.

43. Kerner G, Laval G, Patin E, Boisson-Dupuis S, Abel L, Casanova JL, et al. **Human ancient DNA** analyses reveal the high burden of tuberculosis in Europeans over the last 2,000 years. *American Journal of Human Genetics* 2021; 108:517-524.

44. Kerner G, Neehus AL, Philippot Q, Bohlen J, Rinchai D, Kerrouche N, et al. **Genetic adaptation to pathogens and increased risk of inflammatory disorders in post-Neolithic Europe**. *Cell Genom* 2023; 3(2):100248.

45. Arias AA, Neehus AL, Ogishi M, Meynier V, Krebs A, Lazarov T, et al. **Tuberculosis in otherwise** healthy adults with inherited TNF deficiency. *Nature* 2024; 633(8029):417-425.

46. Lilic D. New perspectives on the immunology of chronic mucocutaneous candidiasis. *Curr Opin Infect Dis* 2002; 15(2):143-147.

47. Puel A, Cypowyj S, Marodi L, Abel L, Picard C, Casanova JL. **Inborn errors of human IL-17 immunity underlie chronic mucocutaneous candidiasis**. *Curr Opin Allergy Clin Immunol* 2012; 12(6):616-622.

48. Tangye SG, Puel A. **The Th17/IL-17 Axis and Host Defense Against Fungal Infections**. *J Allergy Clin Immunol Pract* 2023; 11(6):1624-1634.

49. Puel A. Human inborn errors of immunity underlying superficial or invasive candidiasis. *Hum Genet* 2020; 139(6-7):1011-1022.

50. Toubiana J, Okada S, Hiller J, Oleastro M, Lagos Gomez M, Aldave Becerra JC, et al. **Heterozygous STAT1 gain-of-function mutations underlie an unexpectedly broad clinical phenotype**. *Blood* 2016; 127(25):3154-3164.

51. Okada S, Asano T, Moriya K, Boisson-Dupuis S, Kobayashi M, Casanova JL, et al. Human STAT1 Gain-of-Function Heterozygous Mutations: Chronic Mucocutaneous Candidiasis and Type I Interferonopathy. *J Clin Immunol* 2020; 40(8):1065-1081.

52. Corvilain E, Casanova JL, Puel A. Inherited CARD9 Deficiency: Invasive Disease Caused by Ascomycete Fungi in Previously Healthy Children and Adults. *J Clin Immunol* 2018; 38(6):656-693.

53. Drummond RA, Franco LM, Lionakis MS. Human CARD9: A Critical Molecule of Fungal Immune Surveillance. *Front Immunol* 2018; 9:1836.

54. Drummond RA, Collar AL, Swamydas M, Rodriguez CA, Lim JK, Mendez LM, et al. **CARD9-Dependent Neutrophil Recruitment Protects against Fungal Invasion of the Central Nervous System**. *PLOS Pathogens* 2015; 11(12):e1005293.

55. Drummond RA, Desai JV, Hsu AP, Oikonomou V, Vinh DC, Acklin JA, et al. **Human Dectin-1 deficiency impairs macrophage-mediated defense against phaeohyphomycosis**. *J Clin Invest* 2022; 132(22).

56. Collaborators GBDAR. Global mortality associated with 33 bacterial pathogens in 2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet* 2022; 400(10369):2221-2248.

57. Lim D, Gewurz A, Lint TF, Ghaze M, Sepheri B, Gewurz H. Absence of the sixth component of complement in a patient with repeated episodes of meningococcal meningitis. *J Pediatr* 1976; 89(1):42-47.

58. Merle NS, Church SE, Fremeaux-Bacchi V, Roumenina LT. **Complement System Part I - Molecular Mechanisms of Activation and Regulation**. *Front Immunol* 2015; 6:262.

59. Merle NS, Noe R, Halbwachs-Mecarelli L, Fremeaux-Bacchi V, Roumenina LT. **Complement System Part II: Role in Immunity**. *Front Immunol* 2015; 6:257.

60. Figueroa JE, Densen P. Infectious diseases associated with complement deficiencies. *Clin Microbiol Rev* 1991; 4(3):359-395.

61. Rosain J, Hong E, Fieschi C, Martins PV, El Sissy C, Deghmane AE, et al. **Strains Responsible for Invasive Meningococcal Disease in Patients With Terminal Complement Pathway Deficiencies**. *J Infect Dis* 2017; 215(8):1331-1338.

62. El Sissy C, Rosain J, Vieira-Martins P, Bordereau P, Gruber A, Devriese M, et al. **Clinical and Genetic Spectrum of a Large Cohort With Total and Sub-total Complement Deficiencies**. *Front Immunol* 2019; 10:1936.

63. Xie CB, Jane-Wit D, Pober JS. **Complement Membrane Attack Complex: New Roles, Mechanisms** of Action, and Therapeutic Targets. *The American Journal of Pathology* 2020; 190(6):1138-1150.

64. Lortholary O, El-Sissy C, Leporrier J, Sze Wah Wong S, Dannaoui E, Fremeaux-Bacchi V, et al. **Disseminated Cryptococcosis Following Eculizumab Therapy: Insight Into Pathogenesis**. *Open Forum Infect Dis* 2023; 10(4):ofad159.

65. Desai JV, Kumar D, Freiwald T, Chauss D, Johnson MD, Abers MS, et al. **C5a-licensed phagocytes** drive sterilizing immunity during systemic fungal infection. *Cell* 2023; 186(13):2802-2822 e2822.

66. Gaschignard J, Levy C, Chrabieh M, Boisson B, Bost-Bru C, Dauger S, et al. **Invasive pneumococcal** disease in children can reveal a primary immunodeficiency. *Clin Infect Dis* 2014; 59(2):244-251.

67. Sanges S, Wallet F, Blondiaux N, Theis D, Verin I, Vachee A, et al. **Diagnosis of primary antibody** and complement deficiencies in young adults after a first invasive bacterial infection. *Clin Microbiol Infect* 2017; 23(8):576 e571-576 e575.

68. Heitzeneder S, Seidel M, Forster-Waldl E, Heitger A. **Mannan-binding lectin deficiency - Good news**, **bad news**, **doesn't matter?** *Clin Immunol* 2012; 143(1):22-38.

69. Chen S, Francioli LC, Goodrich JK, Collins RL, Kanai M, Wang Q, et al. **A genomic mutational** constraint map using variation in **76,156** human genomes. *Nature* 2024; 625(7993):92-100.

70. Tsukada S, Saffran DC, Rawlings DJ, Parolini O, Allen RC, Klisak I, et al. **Deficient expression of a B** cell cytoplasmic tyrosine kinase in human X-linked agammaglobulinemia. *Cell* 1993; 72(2):279-290.

71. Vetrie D, Vořechovský I, Sideras P, Holland J, Davies A, Flinter F, et al. **The gene involved in X-linked** agammaglobulinaemia is a member of the src family of protein-tyrosine kinases. *Nature* 1993; 361(6409):226-233.

72. Tangye SG, Al-Herz W, Bousfiha A, Chatila T, Cunningham-Rundles C, Etzioni A, et al. **Human Inborn Errors of Immunity: 2019 Update on the Classification from the International Union of Immunological Societies Expert Committee**. *J Clin Immunol* 2020; 40(1):24-64.

73. Edwards ESJ, Bosco JJ, Ojaimi S, O'Hehir RE, van Zelm MC. **Beyond monogenetic rare variants:** tackling the low rate of genetic diagnoses in predominantly antibody deficiency. *Cellular & Molecular Immunology* 2021; 18(3):588-603.

74. Boisson B, Laplantine E, Dobbs K, Cobat A, Tarantino N, Hazen M, et al. **Human HOIP and LUBAC deficiency underlies autoinflammation, immunodeficiency, amylopectinosis, and lymphangiectasia**. *J Exp Med* 2015; 212(6):939-951.

75. Boisson B, Laplantine E, Prando C, Giliani S, Israelsson E, Xu Z, et al. **Immunodeficiency**, **autoinflammation and amylopectinosis in humans with inherited HOIL-1 and LUBAC deficiency**. *Nat Immunol* 2012; 13(12):1178-1186.

76. Courtois G, Smahi A, Reichenbach J, Doffinger R, Cancrini C, Bonnet M, et al. A hypermorphic lkappaBalpha mutation is associated with autosomal dominant anhidrotic ectodermal dysplasia and T cell immunodeficiency. *J Clin Invest* 2003; 112(7):1108-1115.

77. Doffinger R, Smahi A, Bessia C, Geissmann F, Feinberg J, Durandy A, et al. X-linked anhidrotic ectodermal dysplasia with immunodeficiency is caused by impaired NF-kappaB signaling. *Nat Genet* 2001; 27(3):277-285.

78. Picard C, Casanova JL, Puel A. Infectious diseases in patients with IRAK-4, MyD88, NEMO, or IkappaBalpha deficiency. *Clin Microbiol Rev* 2011; 24(3):490-497.

79. Boisson B. The genetic basis of pneumococcal and staphylococcal infections: inborn errors of human TLR and IL-1R immunity. *Hum Genet* 2020; 139(6-7):981-991.

80. Picard C, Puel A, Bonnet M, Ku CL, Bustamante J, Yang K, et al. **Pyogenic bacterial infections in humans with IRAK-4 deficiency**. *Science* 2003; 299(5615):2076-2079.

81. von Bernuth H, Picard C, Jin Z, Pankla R, Xiao H, Ku CL, et al. **Pyogenic bacterial infections in humans with MyD88 deficiency**. *Science* 2008; 321(5889):691-696.

82. Spaan AN, Henry T, van Rooijen WJM, Perret M, Badiou C, Aerts PC, et al. **The staphylococcal toxin Panton-Valentine Leukocidin targets human C5a receptors**. *Cell host & microbe* 2013; 13(5):584-594.

83. Ward AC, Dale DC. Genetic and molecular diagnosis of severe congenital neutropenia. *Curr Opin Hematol* 2009; 16(1):9-13.

84. Winkelstein JA, Marino MC, Johnston RB, Jr., Boyle J, Curnutte J, Gallin JI, et al. **Chronic** granulomatous disease. Report on a national registry of 368 patients. *Medicine (Baltimore)* 2000; 79(3):155-169.

85. Svensson L, Howarth K, McDowall A, Patzak I, Evans R, Ussar S, et al. Leukocyte adhesion deficiency-III is caused by mutations in KINDLIN3 affecting integrin activation. *Nat Med* 2009; 15(3):306-312.

86. Israel L, Wang Y, Bulek K, Della Mina E, Zhang Z, Pedergnana V, et al. **Human Adaptive Immunity Rescues an Inborn Error of Innate Immunity**. *Cell* 2017; 168(5):789-800 e710.

87. Spaan AN, Neehus AL, Laplantine E, Staels F, Ogishi M, Seeleuthner Y, et al. **Human OTULIN** haploinsufficiency impairs cell-intrinsic immunity to staphylococcal alpha-toxin. *Science* 2022; 376(6599):eabm6380.

88. Spencer S, Köstel Bal S, Egner W, Lango Allen H, Raza SI, Ma CA, et al. Loss of the interleukin-6 receptor causes immunodeficiency, atopy, and abnormal inflammatory responses. *J Exp Med* 2019; 216(9):1986-1998.

89. Nahum A, Sharfe N, Broides A, Dadi H, Naghdi Z, Mandola AB, et al. **Defining the biological responses of IL-6 by the study of a novel IL-6 receptor chain immunodeficiency**. *Journal of Allergy and Clinical Immunology* 2020; 145(3):1011-1015.e1016.

90. Barman P, Chawla S, Sharma J, Tyagi R, Karim A, Rawat A, et al. **Cold abscess and high IgE beyond Job's syndrome: Four cases of IL-6R deficiency**. *Pediatr Allergy Immunol* 2024; 35(9):e14244.

91. Wilson RP, Ives ML, Rao G, Lau A, Payne K, Kobayashi M, et al. **STAT3 is a critical cell-intrinsic regulator of human unconventional T cell numbers and function**. *Journal of Experimental Medicine* 2015; 212(6):855-864.

92. Beziat V, Tavernier SJ, Chen YH, Ma CS, Materna M, Laurence A, et al. **Dominant-negative mutations** in human IL6ST underlie hyper-IgE syndrome. *J Exp Med* 2020; 217(6).

93. Chen Y-H, Grigelioniene G, Newton PT, Gullander J, Elfving M, Hammarsjö A, et al. **Absence of GP130** cytokine receptor signaling causes extended Stüve-Wiedemann syndrome. *Journal of Experimental Medicine* 2020; 217(3).

94. Puel A, Picard C, Lorrot M, Pons C, Chrabieh M, Lorenzo L, et al. **Recurrent staphylococcal cellulitis** and subcutaneous abscesses in a child with autoantibodies against IL-6. *J Immunol* 2008; 180(1):647-654.

95. Nanki T, Onoue I, Nagasaka K, Takayasu A, Ebisawa M, Hosoya T, et al. **Suppression of elevations** in serum C reactive protein levels by anti-IL-6 autoantibodies in two patients with severe bacterial infections. *Ann Rheum Dis* 2013; 72(6):1100-1102.

96. Bloomfield M, Parackova Z, Cabelova T, Pospisilova I, Kabicek P, Houstkova H, et al. Anti-IL6 Autoantibodies in an Infant With CRP-Less Septic Shock. *Front Immunol* 2019; 10:2629.

97. Broca F, Souchaud-Debouverie O, Liuu E, Roblot P, Martin M. Severe Infections in Patients Treated with Tocilizumab for Systemic Diseases Other Than Rheumatoid Arthritis: A Retrospective Multicenter Observational Study. *Eur J Rheumatol* 2023; 10(1):18-22.

98. Doorbar J, Egawa N, Griffin H, Kranjec C, Murakami I. **Human papillomavirus molecular biology and disease association**. *Rev Med Virol* 2015; 25 Suppl 1(Suppl Suppl 1):2-23.

99. Orth G. Genetics of epidermodysplasia verruciformis: Insights into host defense against papillomaviruses. *Semin Immunol* 2006; 18(6):362-374.

100. Orth G. Host defenses against human papillomaviruses: lessons from epidermodysplasia verruciformis. *Curr Top Microbiol Immunol* 2008; 321:59-83.

101. de Jong SJ, Imahorn E, Itin PH, Uitto J, Orth G, Jouanguy E, et al. **Epidermodysplasia verruciformis:** inborn errors of immunity to human beta-papillomaviruses. *Front Microbiol* 2018; 9:1222.

102. Ramoz N, Rueda LA, Bouadjar B, Montoya LS, Orth G, Favre M. **Mutations in two adjacent novel** genes are associated with epidermodysplasia verruciformis. *Nat Genet* 2002; 32(4):579-581.

103. Ramoz N, Taieb A, Rueda LA, Montoya LS, Bouadjar B, Favre M, et al. **Evidence for a nonallelic** heterogeneity of epidermodysplasia verruciformis with two susceptibility loci mapped to chromosome regions 2p21-p24 and 17q25. *J Invest Dermatol* 2000; 114(6):1148-1153.

104. de Jong SJ, Crequer A, Matos I, Hum D, Gunasekharan V, Lorenzo L, et al. **The human CIB1-EVER1-EVER2 complex governs keratinocyte-intrinsic immunity to beta-papillomaviruses**. *J Exp Med* 2018; 215(9):2289-2310.

105. Beziat V, Casanova JL, Jouanguy E. Human genetic and immunological dissection of papillomavirus-driven diseases: new insights into their pathogenesis. *Curr Opin Virol* 2021; 51:9-15. 106. Crequer A, Troeger A, Patin E, Ma CS, Picard C, Pedergnana V, et al. Human RHOH deficiency causes T cell defects and susceptibility to EV-HPV infections. *J Clin Invest* 2012; 122(9):3239-3247.

107. Crequer A, Picard C, Patin E, D'Amico A, Abhyankar A, Munzer M, et al. **Inherited MST1 Deficiency Underlies Susceptibility to EV-HPV Infections**. *PLoS One* 2012; 7(8):e44010.

108. Stray-Pedersen A, Jouanguy E, Crequer A, Bertuch AA, Brown BS, Jhangiani SN, et al. **Compound** heterozygous CORO1A mutations in siblings with a mucocutaneous-immunodeficiency syndrome of epidermodysplasia verruciformis-HPV, molluscum contagiosum and granulomatous tuberculoid leprosy. *J Clin Immunol* 2014; 34(7):871-890.

109. Youssefian L, Saeidian AH, Tavasoli AR, Kalamati E, Naghipoor K, Hozhabrpour A, et al. **Recalcitrant Cutaneous Warts in a Family with Inherited ICOS Deficiency**. *J Invest Dermatol* 2022; 142(9):2435-2445.

110. Youssefian L, Vahidnezhad H, Yousefi M, Saeidian AH, Azizpour A, Touati A, et al. Inherited Interleukin 2-Inducible T-Cell (ITK) Kinase Deficiency in Siblings With Epidermodysplasia Verruciformis and Hodgkin Lymphoma. *Clin Infect Dis* 2019; 68(11):1938-1941.

111. Momenilandi M, Lévy R, Sobrino S, Li J, Lagresle-Peyrou C, Esmaeilzadeh H, et al. **FLT3L governs the development of partially overlapping hematopoietic lineages in humans and mice**. *Cell* 2024.

112. Leiding JW, Holland SM. **Warts and all: human papillomavirus in primary immunodeficiencies**. *J Allergy Clin Immunol* 2012; 130(5):1030-1048.

113. McDermott DH, Murphy PM. WHIM syndrome: Immunopathogenesis, treatment and cure strategies. *Immunol Rev* 2019; 287(1):91-102.

114. Kawai T, Malech HL. **WHIM syndrome: congenital immune deficiency disease**. *Curr Opin Hematol* 2009; 16(1):20-26.

115. Tarzi MD, Jenner M, Hattotuwa K, Faruqi AZ, Diaz GA, Longhurst HJ. **Sporadic case of warts, hypogammaglobulinemia, immunodeficiency, and myelokathexis syndrome**. *J Allergy Clin Immunol* 2005; 116(5):1101-1105.

116. West ES, Kingsbery MY, Mintz EM, Hsu AP, Holland SM, Rady PL, et al. **Generalized verrucosis in a patient with GATA2 deficiency**. *Br J Dermatol* 2014; 170(5):1182-1186.

117. Spinner MA, Sanchez LA, Hsu AP, Shaw PA, Zerbe CS, Calvo KR, et al. **GATA2 deficiency: a protean disorder of hematopoiesis, lymphatics, and immunity**. *Blood* 2014; 123(6):809-821.

118. Kuriyama Y, Hattori M, Mitsui T, Nakano H, Oikawa D, Tokunaga F, et al. **Generalized verrucosis** caused by various human papillomaviruses in a patient with GATA2 deficiency. *J Dermatol* 2018; 45(5):e108-e109.

119. Toboni MD, Bevis KS. Vulvar Cancer as a Result of GATA2 Deficiency, a Rare Genetic Immunodeficiency Syndrome. *Obstet Gynecol* 2018; 132(5):1112-1115.

120. Sanal O, Jing H, Ozgur T, Ayvaz D, Strauss-Albee DM, Ersoy-Evans S, et al. **Additional diverse findings expand the clinical presentation of DOCK8 deficiency**. *J Clin Immunol* 2012; 32(4):698-708.

121. Liu YQ, Zhang GL, Mo XH, Wang B, Wu F, Chen J, et al. A novel homozygous DOCK8 mutation associated with unusual coexistence of gross molluscum contagiosum and epidermodysplasia verruciformis in a DOCK8 deficiency patient. *J Eur Acad Dermatol Venereol* 2017; 31(11):e504-e505.

122. Aydin SE, Kilic SS, Aytekin C, Kumar A, Porras O, Kainulainen L, et al. **DOCK8 deficiency: clinical and immunological phenotype and treatment options - a review of 136 patients**. *J Clin Immunol* 2015; 35(2):189-198.

123. Beziat V. Human genetic dissection of papillomavirus-driven diseases: New insight into their pathogenesis. *Hum Genet* 2020; In press.

124. Horev L, Unger S, Molho-Pessach V, Meir T, Maly A, Stepensky P, et al. **Generalized verrucosis and HPV-3 susceptibility associated with CD4 T-cell lymphopenia caused by inherited human interleukin-7 deficiency**. *J Am Acad Dermatol* 2015; 72(6):1082-1084.

125. Kosumi H, Natsuga K, Takashima S, Miyauchi T, Huang YT, Nomura T, et al. **Two Cases of** Interleukin-7-Deficient Generalized Verrucosis. *Clin Infect Dis* 2020; 71(6):1561-1563.

126. Rawat A, Singh A, Dobbs K, Pala F, Delmonte OM, Vignesh P, et al. **Skewed TCR Alpha, but not Beta, Gene Rearrangements and Lymphoma Associated with a Pathogenic TRAC Variant**. *J Clin Immunol* 2021; 41(6):1395-1399. 127. Delmonte OM, Bergerson JRE, Kawai T, Kuehn HS, McDermott DH, Cortese I, et al. **SASH3 variants cause a novel form of X-linked combined immunodeficiency with immune dysregulation**. *Blood* 2021; 138(12):1019-1033.

128. Abdollahpour H, Appaswamy G, Kotlarz D, Diestelhorst J, Beier R, Schaffer AA, et al. **The phenotype of human STK4 deficiency**. *Blood* 2012; 119(15):3450-3457.

129. Abolnezhadian F, Iranparast S, Ahmadpour F. **Identical Twins with a Mutation in the STK4 Gene Showing Clinical Manifestations of the Mutation at Different Ages: A Case Report**. *Iran J Immunol* 2020; 17(4):333-340.

130. Jorgensen SE, Al-Mousawi A, Assing K, Hartling U, Grosen D, Fisker N, et al. **STK4 Deficiency Impairs Innate Immunity and Interferon Production Through Negative Regulation of TBK1-IRF3 Signaling**. *J Clin Immunol* 2021; 41(1):109-124.

131. Sharafian S, Ziaee V, Shahrooei M, Ahadi M, Parvaneh N. **A Novel STK4 Mutation Presenting with** Juvenile Idiopathic Arthritis and Epidermodysplasia Verruciformis. *J Clin Immunol* 2019; 39(1):11-14. 132. El Kettani A, Ailal F, Marnissi F, Hali F, El Bakkouri J, Benhsaien I, et al. Atypical Cutaneous Viral Infections Reveal an Inborn Error of Immunity in 8 Patients. *Microorganisms* 2023; 11(5).

133. Beziat V, Rapaport F, Hu J, Titeux M, Bonnet des Claustres M, Bourgey M, et al. **Humans with** inherited T cell CD28 deficiency are susceptible to skin papillomaviruses but are otherwise healthy. *Cell* 2021; 184(14):3812-3828 e3830.

134. Schober T, Magg T, Laschinger M, Rohlfs M, Linhares ND, Puchalka J, et al. **A human immunodeficiency syndrome caused by mutations in CARMIL2**. *Nat Commun* 2017; 8:14209.

135. Levy R, Gothe F, Momenilandi M, Magg T, Materna M, Peters P, et al. Human CARMIL2 deficiency underlies a broader immunological and clinical phenotype than CD28 deficiency. *J Exp Med* 2023; 220(2).

136. Wang Y, Ma CS, Ling Y, Bousfiha A, Camcioglu Y, Jacquot S, et al. **Dual T cell- and B cell-intrinsic deficiency in humans with biallelic RLTPR mutations**. *J Exp Med* 2016; 213(11):2413-2435.

137. Ravell JC, Chauvin SD, He T, Lenardo M. An Update on XMEN Disease. *J Clin Immunol* 2020; 40(5):671-681.

138. Dorjbal B, Stinson JR, Ma CA, Weinreich MA, Miraghazadeh B, Hartberger JM, et al. **Hypomorphic** caspase activation and recruitment domain 11 (CARD11) mutations associated with diverse immunologic phenotypes with or without atopic disease. *J Allergy Clin Immunol* 2019; 143(4):1482-1495.

139. Park J, Yang J, Wenzel AT, Ramachandran A, Lee WJ, Daniels JC, et al. **Genomic analysis of 220 CTCLs identifies a novel recurrent gain-of-function alteration in RLTPR (p.Q575E)**. *Blood* 2017; 130(12):1430-1440.

140. Roncagalli R, Cucchetti M, Jarmuzynski N, Gregoire C, Bergot E, Audebert S, et al. **The scaffolding function of the RLTPR protein explains its essential role for CD28 co-stimulation in mouse and human T cells**. *J Exp Med* 2016; 213(11):2437-2457.

141. Guerin A, Moncada-Velez M, Jackson K, Ogishi M, Rosain J, Mancini M, et al. **Helper T cell immunity** in humans with inherited CD4 deficiency. *J Exp Med* 2024; 221(5).

142. Fernandes RA, Perez-Andres M, Blanco E, Jara-Acevedo M, Criado I, Almeida J, et al. **Complete Multilineage CD4 Expression Defect Associated With Warts Due to an Inherited Homozygous CD4 Gene Mutation**. *Front Immunol* 2019; 10:2502.

143. Holdorf AD, Lee KH, Burack WR, Allen PM, Shaw AS. **Regulation of Lck activity by CD4 and CD28** in the immunological synapse. *Nat Immunol* 2002; 3(3):259-264.

144. Smith MG, Lennette EH, Reames HR. Isolation of the virus of herpes simplex and the demonstration of intranuclear inclusions in a case of acute encephalitis. *Am J Pathol* 1941; 17:55-68.

145. Wolman B, Longson M. Herpes encephalitis. Acta Paediatr Scand 1977; 66(2):243-246.

146. Casanova JL. Severe infectious diseases of childhood as monogenic inborn errors of immunity. *Proc Natl Acad Sci U S A* 2015; 112(51):E7128-7137.

147. Casanova JL. Human genetic basis of interindividual variability in the course of infection. *Proc Natl Acad Sci U S A* 2015; 112(51):E7118-7127.

148. Alcais A, Quintana-Murci L, Thaler DS, Schurr E, Abel L, Casanova JL. Life-threatening infectious diseases of childhood: single-gene inborn errors of immunity? *Ann N Y Acad Sci* 2010; 1214:18-33.

149. Dupuis S, Jouanguy E, Al-Hajjar S, Fieschi C, Al-Mohsen IZ, Al-Jumaah S, et al. **Impaired response to interferon-alpha/beta and lethal viral disease in human STAT1 deficiency**. *Nat Genet* 2003; 33(3):388-391.

150. Niehues T, Reichenbach J, Neubert J, Gudowius S, Puel A, Horneff G, et al. **A NEMO-deficient child with immunodeficiency yet without anhidrotic ectodermal dysplasia**. *J Allergy Clin Immunol* 2004; 114:1456-1462.

151. Puel A, Reichenbach J, Bustamante J, Ku CL, Feinberg J, Doffinger R, et al. **The NEMO Mutation Creating the Most-Upstream Premature Stop Codon Is Hypomorphic Because of a Reinitiation of Translation**. *Am J Hum Genet* 2006; 78(4):691-701.

152. Audry M, Ciancanelli M, Yang K, Cobat A, Chang HH, Sancho-Shimizu V, et al. **NEMO is a key component of NF-kappaB- and IRF-3-dependent TLR3-mediated immunity to herpes simplex virus**. *J Allergy Clin Immunol* 2011; 128(3):610-617 e611-614.

153. Andersen LL, Mork N, Reinert LS, Kofod-Olsen E, Narita R, Jorgensen SE, et al. **Functional IRF3** deficiency in a patient with herpes simplex encephalitis. *J Exp Med* 2015; 212(9):1371-1379.

154. Casrouge A, Zhang SY, Eidenschenk C, Jouanguy E, Puel A, Yang K, et al. Herpes simplex virus encephalitis in human UNC-93B deficiency. *Science* 2006; 314(5797):308-312.

155. Guo Y, Audry M, Ciancanelli M, Alsina L, Azevedo J, Herman M, et al. Herpes simplex virus encephalitis in a patient with complete TLR3 deficiency: TLR3 is otherwise redundant in protective immunity. *J Exp Med* 2011; 208(10):2083-2098.

156. Lim HK, Seppanen M, Hautala T, Ciancanelli MJ, Itan Y, Lafaille FG, et al. **TLR3 deficiency in herpes simplex encephalitis: high allelic heterogeneity and recurrence risk**. *Neurology* 2014; 83(21):1888-1897.

157. Perez de Diego R, Sancho-Shimizu V, Lorenzo L, Puel A, Plancoulaine S, Picard C, et al. Human TRAF3 adaptor molecule deficiency leads to impaired Toll-like receptor 3 response and susceptibility to herpes simplex encephalitis. *Immunity* 2010; 33(3):400-411.

158. Sancho-Shimizu V, Perez de Diego R, Lorenzo L, Halwani R, Alangari A, Israelsson E, et al. **Herpes** simplex encephalitis in children with autosomal recessive and dominant TRIF deficiency. *J Clin Invest* 2011; 121(12):4889-4902.

159. Zhang SY, Jouanguy E, Ugolini S, Smahi A, Elain G, Romero P, et al. **TLR3 deficiency in patients** with herpes simplex encephalitis. *Science* 2007; 317(5844):1522-1527.

160. Zhang SY, Casanova JL. **Inborn errors underlying herpes simplex encephalitis: From TLR3 to IRF3**. *J Exp Med* 2015; 212(9):1342-1343.

161. Herman M, Ciancanelli M, Ou YH, Lorenzo L, Klaudel-Dreszler M, Pauwels E, et al. **Heterozygous TBK1 mutations impair TLR3 immunity and underlie herpes simplex encephalitis of childhood**. *J Exp Med* 2012; 209(9):1567-1582.

162. Naesens L, Muppala S, Acharya D, Nemegeer J, Bogaert D, Lee JH, et al. **GTF3A mutations** predispose to herpes simplex encephalitis by disrupting biogenesis of the host-derived RIG-I ligand **RNA5SP141**. *Science immunology* 2022; 7(77):eabq4531.

163. Lafaille FG, Harschnitz O, Lee YS, Zhang P, Hasek ML, Kerner G, et al. Human SNORA31 variations impair cortical neuron-intrinsic immunity to HSV-1 and underlie herpes simplex encephalitis. *Nat Med* 2019; 25(12):1873-1884.

164. Zhang SY, Clark NE, Freije CA, Pauwels E, Taggart AJ, Okada S, et al. **Inborn Errors of RNA Lariat Metabolism in Humans with Brainstem Viral Infection**. *Cell* 2018; 172(5):952-965 e918.

165. Liu Z, Garcia Reino EJ, Harschnitz O, Guo H, Chan YH, Khobrekar NV, et al. **Encephalitis and poor neuronal death-mediated control of herpes simplex virus in human inherited RIPK3 deficiency**. *Science immunology* 2023; 8(82):eade2860.

166. Chan Y-H, Liu Z, Bastard P, Khobrekar N, Hutchison KM, Yamazaki Y, et al. **Human TMEFF1 is a** restriction factor for herpes simplex virus in the brain. *Nature* 2024; 632(8024):390-400.

167. Le Voyer T, Sakata S, Tsumura M, Khan T, Esteve-Sole A, Al-Saud BK, et al. **Genetic, Immunological, and Clinical Features of 32 Patients with Autosomal Recessive STAT1 Deficiency**. *J Immunol* 2021; 207(1):133-152.

168. Bucciol G, Moens L, Ogishi M, Rinchai D, Matuozzo D, Momenilandi M, et al. **Human inherited** complete STAT2 deficiency underlies inflammatory viral diseases. *J Clin Invest* 2023.

169. Ciancanelli MJ, Huang SX, Luthra P, Garner H, Itan Y, Volpi S, et al. **Infectious disease. Lifethreatening influenza and impaired interferon amplification in human IRF7 deficiency**. *Science* 2015; 348(6233):448-453. 170. Hernandez N, Melki I, Jing H, Habib T, Huang SSY, Danielson J, et al. Life-threatening influenza pneumonitis in a child with inherited IRF9 deficiency. *J Exp Med* 2018; 215(10):2567-2585.

171. Lim HK, Huang SXL, Chen J, Kerner G, Gilliaux O, Bastard P, et al. **Severe influenza pneumonitis** in children with inherited TLR3 deficiency. *J Exp Med* 2019; 216(9):2038-2056.

172. Freij BJ, Hanrath AT, Chen R, Hambleton S, Duncan CJA. Life-Threatening Influenza, Hemophagocytic Lymphohistiocytosis and Probable Vaccine-Strain Varicella in a Novel Case of Homozygous STAT2 Deficiency. *Front Immunol* 2020; 11:624415.

173. Asano T, Boisson B, Onodi F, Matuozzo D, Moncada-Velez M, Maglorius Renkilaraj MRL, et al. Xlinked recessive TLR7 deficiency in ~1% of men under 60 years old with life-threatening COVID-19. Science immunology 2021; 6(62):eabl4348.

174. Garcia-Garcia A, Perez de Diego R, Flores C, Rinchai D, Sole-Violan J, Deya-Martinez A, et al. Humans with inherited MyD88 and IRAK-4 deficiencies are predisposed to hypoxemic COVID-19 pneumonia. *J Exp Med* 2023; 220(5).

175. Zhang Q, Bastard P, Liu Z, Le Pen J, Moncada-Velez M, Chen J, et al. **Inborn errors of type I IFN immunity in patients with life-threatening COVID-19**. *Science* 2020; 370(6515).

176. Zhang Q, Bastard P, Effort CHG, Cobat A, Casanova JL. **Human genetic and immunological determinants of critical COVID-19 pneumonia**. *Nature* 2022.

177. Abolhassani H, Landegren N, Bastard P, Materna M, Modaresi M, Du L, et al. Inherited IFNAR1 Deficiency in a Child with Both Critical COVID-19 Pneumonia and Multisystem Inflammatory Syndrome. *J Clin Immunol* 2022; 42(3):471-483.

178. Khanmohammadi S, Rezaei N, Khazaei M, Shirkani A. A Case of Autosomal Recessive Interferon Alpha/Beta Receptor Alpha Chain (IFNAR1) Deficiency with Severe COVID-19. *J Clin Immunol* 2022; 42(1):19-24.

179. Schmidt A, Peters S, Knaus A, Sabir H, Hamsen F, Maj C, et al. **TBK1 and TNFRSF13B mutations** and an autoinflammatory disease in a child with lethal COVID-19. *NPJ Genom Med* 2021; 6(1):55.

180. Zhang Q, Matuozzo D, Le Pen J, Lee D, Moens L, Asano T, et al. **Recessive inborn errors of type I IFN immunity in children with COVID-19 pneumonia**. *J Exp Med* 2022; 219(8).

181. Asgari S, Schlapbach LJ, Anchisi S, Hammer C, Bartha I, Junier T, et al. **Severe viral respiratory infections in children with IFIH1 loss-of-function mutations**. *Proc Natl Acad Sci U S A* 2017; 114(31):8342-8347.

182. Lamborn IT, Jing H, Zhang Y, Drutman SB, Abbott JK, Munir S, et al. **Recurrent rhinovirus infections** in a child with inherited MDA5 deficiency. *J Exp Med* 2017; 214(7):1949-1972.

183. Fournier B, Latour S. **Immunity to EBV as revealed by immunedeficiencies**. *Curr Opin Immunol* 2021; 72:107-115.

184. Taylor GS, Long HM, Brooks JM, Rickinson AB, Hislop AD. **The immunology of Epstein-Barr virus**induced disease. *Annu Rev Immunol* 2015; 33:787-821.

185. Tangye SG. Genetic susceptibility to EBV infection: insights from inborn errors of immunity. *Hum Genet* 2020; 139(6-7):885-901.

186. Tangye SG, Latour S. **Primary immunodeficiencies reveal the molecular requirements for effective host defense against EBV infection**. *Blood* 2020; 135(9):644-655.

187. Fournier B, Tusseau M, Villard M, Malcus C, Chopin E, Martin E, et al. **DEF6 deficiency, a mendelian susceptibility to EBV infection, lymphoma, and autoimmunity**. *J Allergy Clin Immunol* 2021; 147(2):740-743 e749.

188. Fournier B, Hoshino A, Bruneau J, Bachelet C, Fusaro M, Klifa R, et al. **Inherited TNFSF9 deficiency** causes broad Epstein-Barr virus infection with EBV+ smooth muscle tumors. *J Exp Med* 2022; 219(7).

189. Martin E, Winter S, Garcin C, Tanita K, Hoshino A, Lenoir C, et al. **Role of IL-27 in Epstein-Barr virus** infection revealed by IL-27RA deficiency. *Nature* 2024.

190. Casanova JL, Peel J, Donadieu J, Neehus AL, Puel A, Bastard P. **The ouroboros of autoimmunity**. *Nat Immunol* 2024; 25(5):743-754.

191. Teisner B, Elling P, Svehag SE, Poulsen L, Lamm LU, Sjoholm A. **C3 nephritic factor in a patient** with recurrent Neisseria meningitidis infections. *Acta Pathol Microbiol Immunol Scand C* 1984; 92(6):341-349.

192. Edwards KM, Alford R, Gewurz H, Mold C. **Recurrent bacterial infections associated with C3 nephritic factor and hypocomplementemia**. *N Engl J Med* 1983; 308(19):1138-1141.

193. Thompson RA, Yap PL, Brettle RB, Dunmow RE, Chapel H. **Meningococcal meningitis associated** with persistent hypocomplementaemia due to circulating C3 nephritic factor. *Clin Exp Immunol* 1983; 52(1):153-156.

194. Ku CL, Chi CY, von Bernuth H, Doffinger R. Autoantibodies against cytokines: phenocopies of primary immunodeficiencies? *Hum Genet* 2020; 139(6-7):783-794.

195. Puel A, Bastard P, Bustamante J, Casanova JL. Human autoantibodies underlying infectious diseases. *J Exp Med* 2022; 219(4).

196. Shih HP, Ding JY, Yeh CF, Chi CY, Ku CL. Anti-interferon-gamma autoantibody-associated immunodeficiency. *Curr Opin Immunol* 2021; 72:206-214.

197. Peel JN, Yang R, Le Voyer T, Gervais A, Rosain J, Bastard P, et al. **Neutralizing IFN-gamma autoantibodies are rare and pathogenic in HLA-DRB1*15:02 or 16:02 individuals**. *J Clin Invest* 2024; 134(8).

198. Ku CL, Lin CH, Chang SW, Chu CC, Chan JF, Kong XF, et al. **Anti-IFN-gamma autoantibodies are** strongly associated with HLA-DR*15:02/16:02 and HLA-DQ*05:01/05:02 across Southeast Asia. *J Allergy Clin Immunol* 2016; 137(3):945-948 e948.

199. Chi CY, Chu CC, Liu JP, Lin CH, Ho MW, Lo WJ, et al. Anti-IFN-gamma autoantibodies in adults with disseminated nontuberculous mycobacterial infections are associated with HLA-DRB1*16:02 and HLA-DQB1*05:02 and the reactivation of latent varicella-zoster virus infection. *Blood* 2013; 121(8):1357-1366.

200. Kisand K, Boe Wolff AS, Podkrajsek KT, Tserel L, Link M, Kisand KV, et al. **Chronic mucocutaneous** candidiasis in **APECED** or thymoma patients correlates with autoimmunity to Th17-associated cytokines. *J Exp Med* 2010; 207(2):299-308.

201. Puel A, Doffinger R, Natividad A, Chrabieh M, Barcenas-Morales G, Picard C, et al. Autoantibodies against IL-17A, IL-17F, and IL-22 in patients with chronic mucocutaneous candidiasis and autoimmune polyendocrine syndrome type I. *J Exp Med* 2010; 207(2):291-297.

202. Rosen LB, Rocha Pereira N, Figueiredo C, Fiske LC, Ressner RA, Hong JC, et al. **Nocardia-induced** granulocyte macrophage colony-stimulating factor is neutralized by autoantibodies in disseminated/extrapulmonary nocardiosis. *Clin Infect Dis* 2015; 60(7):1017-1025.

203. Kitamura T, Tanaka N, Watanabe J, Uchida, Kanegasaki S, Yamada Y, et al. **Idiopathic pulmonary** alveolar proteinosis as an autoimmune disease with neutralizing antibody against granulocyte/macrophage colony-stimulating factor. *J Exp Med* 1999; 190(6):875-880.

204. Seymour JF, Presneill JJ. **Pulmonary alveolar proteinosis: progress in the first 44 years**. *Am J Respir Crit Care Med* 2002; 166(2):215-235.

205. Rosen LB, Freeman AF, Yang LM, Jutivorakool K, Olivier KN, Angkasekwinai N, et al. **Anti-GM-CSF autoantibodies in patients with cryptococcal meningitis**. *J Immunol* 2013; 190(8):3959-3966.

206. Browne SK, Burbelo PD, Chetchotisakd P, Suputtamongkol Y, Kiertiburanakul S, Shaw PA, et al. **Adult-onset immunodeficiency in Thailand and Taiwan**. *N Engl J Med* 2012; 367(8):725-734.

207. Perrineau S, Guery R, Monnier D, Puel A, Lanternier F. **Anti-GM-CSF Autoantibodies and Cryptococcus neoformans var. grubii CNS Vasculitis**. *J Clin Immunol* 2020; 40(5):767-769.

208. Kuo CY, Wang SY, Shih HP, Tu KH, Huang WC, Ding JY, et al. **Disseminated Cryptococcosis Due** to Anti-Granulocyte-Macrophage Colony-Stimulating Factor Autoantibodies in the Absence of Pulmonary Alveolar Proteinosis. *J Clin Immunol* 2017; 37(2):143-152.

209. Saijo T, Chen J, Chen SC, Rosen LB, Yi J, Sorrell TC, et al. **Anti-granulocyte-macrophage colony**stimulating factor autoantibodies are a risk factor for central nervous system infection by **Cryptococcus gattii in otherwise immunocompetent patients**. *mBio* 2014; 5(2):e00912-00914.

210. Huynh J, Saddi V, Cooper P, Cheng AT, Meyer W, Chen S, et al. **Unusual Presentation of Severe Endobronchial Obstruction Caused by Cryptococcus gattii in a Child**. *J Pediatric Infect Dis Soc* 2020; 9(1):67-70.

211. Applen Clancey S, Ciccone EJ, Coelho MA, Davis J, Ding L, Betancourt R, et al. **Cryptococcus** deuterogattii VGIIa Infection Associated with Travel to the Pacific Northwest Outbreak Region in an Anti-Granulocyte-Macrophage Colony-Stimulating Factor Autoantibody-Positive Patient in the United States. *mBio* 2019; 10(1).

212. Stevenson B, Bundell C, Mulrennan S, McLean-Tooke A, Murray R, Brusch A. **The significance of** anti-granulocyte-macrophage colony-stimulating factor antibodies in cryptococcal infection: case series and review of antibody testing. *Intern Med J* 2019; 49(11):1446-1450.

213. Viola GM, Malek AE, Rosen LB, DiNardo AR, Nishiguchi T, Okhuysen PC, et al. **Disseminated** cryptococcosis and anti-granulocyte-macrophage colony-stimulating factor autoantibodies: An underappreciated association. *Mycoses* 2021; 64(6):576-582.

214. Panackal AA, Rosen LB, Uzel G, Davis MJ, Hu G, Adeyemo A, et al. Susceptibility to Cryptococcal Meningoencephalitis Associated With Idiopathic CD4(+) Lymphopenia and Secondary Germline or Acquired Defects. Open Forum Infect Dis 2017; 4(2):ofx082.

215. Crum-Cianflone NF, Lam PV, Ross-Walker S, Rosen LB, Holland SM. Autoantibodies to Granulocyte-Macrophage Colony-Stimulating Factor Associated With Severe and Unusual Manifestations of Cryptococcus gattii Infections. *Open Forum Infect Dis* 2017; 4(4):ofx211.

216. Demir S, Chebib N, Thivolet-Bejui F, Cottin V. **Pulmonary alveolar proteinosis following** cryptococcal meningitis: a possible cause? *BMJ Case Rep* 2018; 2018.

217. Casanova JL, Anderson MS. Unlocking life-threatening COVID-19 through two types of inborn errors of type I IFNs. *J Clin Invest* 2023; 133(3):e166283.

218. Bastard P, Rosen LB, Zhang Q, Michailidis E, Hoffmann HH, Zhang Y, et al. **Autoantibodies against** type I IFNs in patients with life-threatening COVID-19. *Science* 2020; 370(6515).

219. Bastard P, Gervais A, Le Voyer T, Rosain J, Philippot Q, Manry J, et al. Autoantibodies neutralizing type I IFNs are present in ~4% of uninfected individuals over 70 years old and account for ~20% of COVID-19 deaths. *Science immunology* 2021; 6(62).

220. Manry J, Bastard P, Gervais A, Le Voyer T, Rosain J, Philippot Q, et al. **The risk of COVID-19 death is much greater and age dependent with type I IFN autoantibodies**. *Proc Natl Acad Sci U S A* 2022; 119(21):e2200413119.

221. Bastard P, Vazquez S, Liu J, Laurie MT, Wang CY, Gervais A, et al. Vaccine breakthrough hypoxemic COVID-19 pneumonia in patients with auto-Abs neutralizing type I IFNs. *Science immunology* 2022:eabp8966.

222. Sokal A, Bastard P, Chappert P, Barba-Spaeth G, Fourati S, Vanderberghe A, et al. **Human type I IFN** deficiency does not impair B cell response to SARS-CoV-2 mRNA vaccination. *J Exp Med* 2023; 220(1).

223. Bastard P, Michailidis E, Hoffmann HH, Chbihi M, Le Voyer T, Rosain J, et al. Auto-antibodies to type I IFNs can underlie adverse reactions to yellow fever live attenuated vaccine. *J Exp Med* 2021; 218(4). 224. Alotaibi F, Alharbi NK, Rosen LB, Asiri AY, Assiri AM, Balkhy HH, et al. Type I interferon autoantibodies in hospitalized patients with Middle East respiratory syndrome and association with outcomes and treatment effect of interferon beta-1b in MIRACLE clinical trial. *Influenza Other Respir Viruses* 2023; 17(3):e13116.

225. Gervais A, Rovida F, Avanzini MA, Croce S, Marchal A, Lin SC, et al. Autoantibodies neutralizing type I IFNs underlie West Nile virus encephalitis in approximately 40% of patients. *J Exp Med* 2023; 220(9).

226. Nagafuchi S, Umene K, Yamanaka F, Oohashi S, Shindo M, Kurisaki H, et al. **Recurrent herpes** simplex virus infection in a patient with autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy associated with L29P and IVS9-1G>C compound heterozygous autoimmune regulator gene mutations. *J Intern Med* 2007; 261(6):605-610.

227. Mathian A, Breillat P, Dorgham K, Bastard P, Charre C, Lhote R, et al. Lower disease activity but higher risk of severe COVID-19 and herpes zoster in patients with systemic lupus erythematosus with pre-existing autoantibodies neutralising IFN-alpha. *Ann Rheum Dis* 2022.

228. Hetemäki I, S. L, Välimaa H, livari K, Kekäläinen E, Petteri AT. **Patients with autoimmune** polyendocrine syndrome type 1 have an increased susceptibility to severe herpesvirus infections. *Clinical Immunology* 2021; 231:108851.

229. Busnadiego I, Abela IA, Frey PM, Hofmaenner DA, Scheier TC, Schuepbach RA, et al. **Critically ill COVID-19 patients with neutralizing autoantibodies against type I interferons have increased risk of herpesvirus disease**. *PLoS Biol* 2022; 20(7):e3001709.

230. Batra A, Davison S, Rajwal S, Hale A, Stringer MD, McClean P. **Varicella recurrence complicated by pneumonia after liver transplantation for APECED**. *J Pediatr Gastroenterol Nutr* 2007; 44(5):637-639.

231. Walter JE, Rosen LB, Csomos K, Rosenberg JM, Mathew D, Keszei M, et al. **Broad-spectrum antibodies against self-antigens and cytokines in RAG deficiency**. *J Clin Invest* 2015; 125(11):4135-4148.

232. Le Voyer T, Parent AV, Liu X, Cederholm A, Gervais A, Rosain J, et al. **Autoantibodies against type** I IFNs in humans with alternative NF-kappaB pathway deficiency. *Nature* 2023; 623(7988):803-813.

233. Bastard P, Gervais A, Le Voyer T, Philippot Q, Cobat A, Rosain J, et al. Human autoantibodies neutralizing type I IFNs: From 1981 to 2023. *Immunol Rev* 2024; 322(1):98-112.

234. Griffin H, Ceron-Gutierrez L, Gharahdaghi N, Ebrahimi S, Davies S, Loo PS, et al. **Neutralizing Autoantibodies against Interleukin-10 in Inflammatory Bowel Disease**. *N Engl J Med* 2024; 391(5):434-441.

235. Lagier JC, Raoult D. Whipple's disease and Tropheryma whipplei infections: when to suspect them and how to diagnose and treat them. *Curr Opin Infect Dis* 2018; 31(6):463-470.

236. Boumaza A, Ben Azzouz E, Arrindell J, Lepidi H, Mezouar S, Desnues B. Whipple's disease and Tropheryma whipplei infections: from bench to bedside. *Lancet Infect Dis* 2022; 22(10):e280-e291.

237. Guerin A, Kerner G, Marr N, Markle JG, Fenollar F, Wong N, et al. **IRF4 haploinsufficiency in a** family with Whipple's disease. *Elife* 2018; 7.

238. Mee PT, Buultjens AH, Oliver J, Brown K, Crowder JC, Porter JL, et al. **Mosquitoes provide a transmission route between possums and humans for Buruli ulcer in southeastern Australia**. *Nat Microbiol* 2024; 9(2):377-389.

239. Manry J, Vincent QB, Johnson C, Chrabieh M, Lorenzo L, Theodorou I, et al. **Genome-wide** association study of Buruli ulcer in rural Benin highlights role of two LncRNAs and the autophagy pathway. *Commun Biol* 2020; 3(1):177.

240. Manry J. Human genetics of Buruli ulcer. Hum Genet 2020; 139(6-7):847-853.

241. Vincent QB, Belkadi A, Fayard C, Marion E, Adeye A, Ardant MF, et al. **Microdeletion on** chromosome 8p23.1 in a familial form of severe Buruli ulcer. *PLoS Negl Trop Dis* 2018; 12(4):e0006429.

242. Dallmann-Sauer M, Xu YZ, da Costa ALF, Tao S, Gomes TA, Prata R, et al. Allele-dependent interaction of LRRK2 and NOD2 in leprosy. *PLoS Pathog* 2023; 19(3):e1011260.

243. Mesri EA, Cesarman E, Boshoff C. **Kaposi's sarcoma and its associated herpesvirus**. *Nat Rev Cancer* 2010; 10(10):707-719.

244. Beziat V, Jouanguy E. Human inborn errors of immunity to oncogenic viruses. *Curr Opin Immunol* 2021; 72:277-285.

245. Jouanguy E, Beziat V, Mogensen TH, Casanova JL, Tangye SG, Zhang SY. **Human inborn errors of immunity to herpes viruses**. *Curr Opin Immunol* 2020; 62:106-122.

246. Jackson CC, Dickson MA, Sadjadi M, Gessain A, Abel L, Jouanguy E, et al. **Kaposi Sarcoma of Childhood: Inborn or Acquired Immunodeficiency to Oncogenic HHV-8**. *Pediatr Blood Cancer* 2016; 63(3):392-397.

247. Korol CB, Belkaya S, Alsohime F, Lorenzo L, Boisson-Dupuis S, Brancale J, et al. **Fulminant Viral Hepatitis in Two Siblings with Inherited IL-10RB Deficiency**. *J Clin Immunol* 2023; 43(2):406-420.

248. Belkaya S, Michailidis E, Korol CB, Kabbani M, Cobat A, Bastard P, et al. **Inherited IL-18BP** deficiency in human fulminant viral hepatitis. *J Exp Med* 2019; 216(8):1777-1790.

249. Jouanguy E. Human genetic basis of fulminant viral hepatitis. *Hum Genet* 2020; 139(6-7):877-884.

250. Drutman SB, Mansouri D, Mahdaviani SA, Neehus AL, Hum D, Bryk R, et al. **Fatal Cytomegalovirus Infection in an Adult with Inherited NOS2 Deficiency**. *N Engl J Med* 2020; 382(5):437-445.

251. Miller LH, Mason SJ, Clyde DF, McGinniss MH. The resistance factor to Plasmodium vivax in blacks. The Duffy-blood-group genotype, FyFy. *N Engl J Med* 1976; 295(6):302-304.

252. Tournamille C, Colin Y, Cartron JP, Le Van Kim C. **Disruption of a GATA motif in the Duffy gene promoter abolishes erythroid gene expression in Duffy-negative individuals**. *Nat Genet* 1995; 10(2):224-228.

253. McManus KF, Taravella AM, Henn BM, Bustamante CD, Sikora M, Cornejo OE. **Population genetic** analysis of the DARC locus (Duffy) reveals adaptation from standing variation associated with malaria resistance in humans. *PLoS Genet* 2017; 13(3):e1006560.

254. Le Pendu J, Ruvoen-Clouet N, Kindberg E, Svensson L. **Mendelian resistance to human norovirus infections**. *Semin Immunol* 2006; 18(6):375-386.

255. Zhang SY, Jouanguy E, Zhang Q, Abel L, Puel A, Casanova JL. **Human inborn errors of immunity to infection affecting cells other than leukocytes: from the immune system to the whole organism**. *Curr Opin Immunol* 2019; 59:88-100.

256. Beziat V, Abel L, Casanova JL, Cobat A. **Human genetics of infectious diseases**. *Vogel and Motulsky's Human Genetics* 2024; in press.

257. Barbe L, Le Moullac-Vaidye B, Echasserieau K, Bernardeau K, Carton T, Bovin N, et al. **Histo-blood** group antigen-binding specificities of human rotaviruses are associated with gastroenteritis but not with in vitro infection. *Sci Rep* 2018; 8(1):12961.

258. Ferrer-Admetlla A, Sikora M, Laayouni H, Esteve A, Roubinet F, Blancher A, et al. **A natural history** of FUT2 polymorphism in humans. *Mol Biol Evol* 2009; 26(9):1993-2003.

259. Dean M, Carrington M, Winkler C, Huttley GA, Smith MW, Allikmets R, et al. **Genetic restriction of** HIV-1 infection and progression to AIDS by a deletion allele of the CKR5 structural gene. Hemophilia Growth and Development Study, Multicenter AIDS Cohort Study, Multicenter Hemophilia Cohort Study, San Francisco City Cohort, ALIVE Study. *Science* 1996; 273(5283):1856-1862.

260. Liu R, Paxton WA, Choe S, Ceradini D, Martin SR, Horuk R, et al. **Homozygous defect in HIV-1** coreceptor accounts for resistance of some multiply-exposed individuals to HIV-1 infection. *Cell* 1996; 86(3):367-377.

261. Samson M, Libert F, Doranz BJ, Rucker J, Liesnard C, Farber CM, et al. **Resistance to HIV-1 infection** in caucasian individuals bearing mutant alleles of the CCR-5 chemokine receptor gene. *Nature* 1996; 382(6593):722-725.

262. Materna M, Delmonte OM, Bosticardo M, Momenilandi M, Conrey PE, Charmeteau-De Muylder B, et al. **The immunopathological landscape of human pre-TCRalpha deficiency: From rare to common variants**. *Science* 2024; 383(6686):eadh4059.

263. Li J, Lei WT, Zhang P, Rapaport F, Seeleuthner Y, Lyu B, et al. **Biochemically deleterious human NFKB1 variants underlie an autosomal dominant form of common variable immunodeficiency**. *J Exp Med* 2021; 218(11).

264. Asano T, Khourieh J, Zhang P, Rapaport F, Spaan AN, Li J, et al. **Human STAT3 variants underlie** autosomal dominant hyper-IgE syndrome by negative dominance. *J Exp Med* 2021; 218(8).

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Figure 1: Schematic representation of the cooperation between phagocytic cells and T and NK cells during bacterial invasion.

Molecules for which the genes are found mutated in patients with MSMD are represented in pink. Molecules for which the genes are found mutated in patients with TB are represented with a red surrounding. Molecules in pink with a red surrounding indicate that patients were found to suffer from MSMD or TB or both. Autoantibodies to IFN-g in patients with MSMD are also represented.

Figure 2: Schematic representation of the signaling pathways activated during *C. albicans* infection.

Molecules for which the genes are mutated and responsible for syndromic chronic mucocutaneous candidiasis (CMC) are represented in pink. Molecules for which the genes are mutated and responsible for isolated CMC are represented with red surroundings. Autoantibodies neutralizing IL-17A and/or IL-17F in patients with syndromic CMC (APS-1/APECED patients) are also represented.

Figure 3: Schematic representation of the skin and the cooperation between target cells (keratinocytes), antigen-presenting cells (APCs) et T lymphocytes in the cutaneous anti-HPV immune response.

Mutated genes found mutated in patients with isolated EV are in red. Molecules for which the genes are found mutated in patients with syndromic EV or recalcitrant warts are represented in pink.

Figure 4: Schematic representation of the cooperation between Epithelial/neuron cells sand plasmacytoid dendritic cells during viral invasion.

Molecules for which the genes are found mutated in patients with viral encephalitis are represented in pink. Molecules for which the genes are found mutated in patients with viral pneumonia are represented with a red surrounding. Molecules in pink with a red surrounding indicate that patients were found to suffer from HSE or viral pneumonia or both. Autoantibodies to IFN- α and ω in patients with pneumonia are also represented.

	Infectious agent	Clinical phenotype	Immunological phenotype	Gene	Inheritance #	
Mycobacterial diseases	BCG vaccines and environmental mycobacteria	Mendelian Susceptibility to Mycobacterial Diseases (MSMD)	IFN-γ deficiency	IFNGR1, IFNGR2, IL12RB1, IL12B, NEMO, STAT1, CYBB, IRF8, ISG15, TYK2, RORC, IL12RB2, IL23R, SPPL2A, JAK1, IFNG, CCR2, IRF1, MCTS1, ZNFX1, TBX21, USP18	Mendelian or monogenic	AR, AD, XR
	Mycobacterium tuberculosis	Tuberculosis (TB)	IFN-γ or TNF deficiency	IL12RB1, TYK2, ITK, PDCD1, TNF	Monogenic	AR
	Mycobacterium ulcerans	Buruli ulcer	Defensin deficiency	Chr. 8 microdeletion	Monogenic	AR
	Mycobacterium leprosy	Leprosy	LRRK2 and NOD2 deficiency	LRRK2 and NOD2	Digenic	AR, AD
Fungal diseases	Candida sp. Dermatophytes	Chronic Mucocutaneous Candidiasis (CMC) Invasive fungal	IL-17 deficiency CARD9	IL17F, IL17RA, IL17RC, TRAP3IP2, STAT1, JNK1, IL6ST, CARD9, IL12B, IL12RB1, IL23R, STAT3, REL, ZNF341, RORC CARD9	Mendelian Mendelian	AR, AD AR
		diseases	deficiency			
Bacteria disease	Neisseria	Meningitis	Complement deficiency	C1QA, C1QB, C1QC, C1S, C2, C3, C4A, C4B, CFH, CFI, C5, C6, C7, C8A, C8B, C9, CFB, CFD, CEP	Monogenic	AR, XR
	Streptococcus pneumoniae (and other encapsulated bacteria)	Meningitis, pneumonia, ENT	Complement deficiency	C1QA, C1QB, C1QC, C1S, C2, C3, C4A, C4B, CFH, CFI, CFP	Monogenic	AR
	,		TIR response deficiency	IRAK4, MYD88, NEMO, HOIL1, HOIP	Monogenic	AR, XR
			Specific antibody defect/asplenia	RPSA	Monogenic	AD
	Staphylococcus aureus	Severe and recurrent pneumonia, skin infection (CMC, Abscesses)	TLR2 response deficiency or IL-6 deficiency	TIRAP, IL6RA, ZNF341, STAT3, IL6ST	Mendelian or monogenic	AR, AD
	Trophervma	Whipple's disease	Increase sensitivity to α -toxin in non- hematopoietic cells IRF4 or CD4	OTULIN IRF4. CD4	Monogenic	AD AD. AR
	whipplei		deficiency		Sienegonio	
Viral diseases of the skin	Human papillomavirus (HPV)	Epidermodysplasia verruciformis	EVER-CIB1 deficiency and APC and/or T cells deficiency	EVER1, EVER2, CIB1, RhoH, CORO1A, DOCK8, LCK, TPP2, STK4, ITK, FLT3L	Monogenic	AR

	Human herpes virus 8	Recurrent warts Kaposi sarcoma	APC and/or T cells deficiency T cells and OX40-	STK4, DOCK8, GATA2, CXCR4, FLT3L, CD4, ICOSLG, CARMIL2, IL7, CD28, CARD11, MAGT1, TRAC OX40, IFNGR1, CTLA4, MAGT1,	Monogenic	AR, AD, XLR AR, AD
			dependent T cell immunity	STIM1		
Viral encephalitis	Herpes simplex virus (HSV)	Encephalitis	TLR3-IFN-α/β deficiency, TFIIIA-RIG-I- IFN-α/β deficiency	TLR3, UNC93B1, TRIF, TRAF3, TBK1, IRF3, NEMO, GTF3A, IFNAR1, STAT1, STAT2, TYK2,	Monogenic	AR, AD
			independent deficiency	DBR1	Monogenic	AR, AD
Viral pneumonia	SARS-CoV-2	COVID-19 pneumonia	Type I and III IFN deficiency	IRF7, IRF9, STAT2, TYK2, TLR3, STAT1	Mendelian	AR, AD, XR
	Influenza virus	Severe influenza	Type I and III IFN deficiency	IRF7, IRF9, TLR3, TLR7, IFNAR1, IFNAR2, STAT2, TYK2, TBK1	Monogenic	AR, AD
	Rhinovirus, Respiratory syncitial virus (RSV)	Recurrent/severe infections	MDA5 deficiency	IFIH1	Monogenic	AR, AD
EBV diseases	Epstein–Barr virus (EBV)	X-linked lymphoproliferative disease; severe infection; B-cell lymphoma	Cytotoxic T/NK cell deficiency	PRF1, RAB27A, UNC13D, SYT11, LYST, SH2D1A, RASGRP1, ITK, MAGT1, DEF6, CTPS1, THFRSF9, CD27, CD70, IL27RA	Mendelian or monogenic	AR, XR
	Cytomegalovirus (CMV)	Lethal infection	NOS2 deficiency	NOS2	Mendelian	AR
Hepatitis	Hepatitis A virus	Fulminant hepatitis	Increased IFN-γ activity	IL18BP, IL10RB	Monogenic	AR
Resistance to infection	Human immunodeficiency virus-1	Resistance to infection	Lack of receptor for pathogen in CD4 ⁺ T cells	CCR5	Mendelian	AR
	Plasmodium vivax	Resistance to infection	Lack of receptor for pathogen in erythrocytes	DARC	Mendelian	AR
	Norovirus	Resistance to infection	Lack of receptor for pathogen in intestinal epithelium	FUT2	Mendelian	AR

Table 1 : Genetic etiologies of infectious diseases categorized by infectious agents, clinical, and immunological phenotypes.

Footnote: # Mendelian refers to complete concordance between the genotype and phenotype, with full penetrance of the trait. In contrast, monogenic diseases may exhibit incomplete penetrance.







Epithelial cells / neurons

