

Addressing health equity for breastfeeding women: primaquine for Plasmodium vivax radical cure

Nada Abla, Anne Claire Marrast, Elodie Jambert, Naomi Richardson, Stephan Duparc, Lisa Almond, Karen Rowland Yeo, Xian Pan, Joel Tarning, Ping Zhao, et al.

To cite this version:

Nada Abla, Anne Claire Marrast, Elodie Jambert, Naomi Richardson, Stephan Duparc, et al.. Addressing health equity for breastfeeding women: primaquine for Plasmodium vivax radical cure. Malaria Journal, 2024, 23 (1), pp.287. $10.1186 \text{/s}12936-024-05112-9$. hal-04840606

HAL Id: hal-04840606 <https://hal.science/hal-04840606v1>

Submitted on 16 Dec 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

PERSPECTIVE

Addressing health equity for breastfeeding women: primaquine for *Plasmodium vivax* radical cure

Nada Abla^{1*}, Anne Claire Marrast¹, Elodie Jambert¹, Naomi Richardson², Stephan Duparc¹, Lisa Almond³, Karen Rowland Yeo³, Xian Pan³, Joel Tarning^{4,5}, Ping Zhao⁶, Janice Culpepper⁶, Catriona Waitt^{7,8}, Charlotte Koldeweij⁹, Susan Cole¹⁰, Andrew S. Butler¹⁰, Sonia Khier¹¹, Jörg J. Möhrle¹ and Myriam El Gaaloul¹

Abstract

Plasmodium vivax malaria remains a global health challenge, with approximately 6.9 million estimated cases in 2022. The parasite has a dormant liver stage, the hypnozoite, which reactivates to cause repeated relapses over weeks, months, or years. These relapses erode patient health, contribute to the burden of malaria, and promote transmission. Radical cure to prevent relapses requires administration of an 8-aminoquinoline, either primaquine or tafenoquine. However, malaria treatment guidelines updated by the World Health Organization (WHO) in October 2023 restrict primaquine use for women breastfeeding children<6 months of age, or women breastfeeding older children if their child is G6PD defcient or if the child's G6PD status is unknown. Primaquine restrictions assume that 8-aminoquinoline exposures in breast milk would be sufficient to cause haemolysis in the nursing infant should they be G6PD defcient. WHO recommendations for tafenoquine are awaited. Notably, the WHO recommends that infants are breastfed for the frst 2 years of life, and exclusively until 6 months old. Repeated pregnancies, followed by extended breastfeeding leaves women in *P. vivax* endemic regions potentially vulnerable to relapses for many years. This puts women's health at risk, increases the malaria burden, and perpetuates transmission, hindering malaria control and elimination. The benefts of lifting restrictions on primaquine administration to breastfeeding women are signifcant, avoiding the adverse consequences of repeated episodes of acute malaria, such as severe anaemia. Recent data challenge the restriction of primaquine in breastfeeding women. Clinical pharmacokinetic data in breastfeeding infants≥28 days old show that the exposure to primaquine is very low and less than 1% of the maternal exposure, indicating negligible risk to infants, irrespective of their G6PD status. Physiologically-based pharmacokinetic modelling complements the clinical data, predicting minimal primaquine exposure to infants and neonates via breast milk from early post-partum. This article summarizes the clinical and modelling evidence for a favourable beneft:risk evaluation of *P. vivax* radical cure with primaquine for breastfeeding women without the need for infant G6PD testing, supporting a change in policy. This adjustment to current treatment guidelines would support health equity in regard to efective interventions to protect women and their children, enhance malaria control strategies, and advance *P. vivax* elimination.

Keywords *Plasmodium vivax*, Radical cure, Primaquine, Physiologically-based pharmacokinetic modelling, Breastfeeding, Pregnancy, Malaria

*Correspondence: Nada Abla ablan@mmv.org Full list of author information is available at the end of the article

© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modifed the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit<http://creativecommons.org/licenses/by-nc-nd/4.0/>.

Background

Globally, there were an estimated 6.9 million cases of *Plasmodium vivax* malaria in 2022 [1]. *Plasmodium vivax* is dominant in Latin America and South-East Asia, causing 75% and 51% of all malaria cases, respectively [1]. *P. vivax* is also a key pathogen in the Eastern Mediterranean (42% of cases), Western Pacifc (27% of cases) and the horn of Africa [1]. Chloroquine is the frst-line treatment for acute uncomplicated *P. vivax* malaria in most countries, though artemisinin-based combination therapy (ACT) is also used [2].

The *P. vivax* lifecycle is complicated by the ability of the parasite to form hypnozoites. This arrested or dormant stage shelters in the liver, is not susceptible to treatment with chloroquine or ACT, and cannot be detected symptomatically. Although serological testing to assess prior exposure to *P. vivax* has been suggested as a presumptive test for hypnozoites to support test and treat strategies for malaria elimination, there is no direct diagnostic available for the presence of hypnozoites [3].

Hypnozoites become reactivated within weeks or months of the initial infection causing repeated malaria episodes (relapses) and onward transmission (Fig. 1).

Multiple relapses are damaging to patient health, causing chronic anaemia with associated morbidity and an increased mortality risk [4–6]. Short-latency tropical strains of *P. vivax* have relapse intervals of approximately 3 weeks [7]. The contribution of relapses to the burden of *P. vivax* malaria varies but often represents more than half of cases and can be over 80% of cases in endemic countries $[8-10]$. Thus, it is essential to break the cycle of repeated *P. vivax* relapses by not only treating the acute infection but also eliminating hypnozoites, termed radical cure.

Plasmodium vivax radical cure requires the administration of a blood schizonticide (chloroquine or artemisininbased combination therapy) to eliminate the blood-stage parasites and an 8-aminoquinoline to eliminate hypnozoites. Two 8-aminoquinolines are approved for radical cure of *P. vivax*—primaquine and tafenoquine—and both can cause haemolysis in individuals with glucose-6-phosphate dehydrogenase (G6PD) deficiency $[11-15]$. Haemolysis may be severe, causing acute haemolytic anaemia requiring hospitalisation, blood transfusion and potentially renal dialysis $[16–20]$. The haemolytic risk depends on the dose and duration of 8-aminoquinoline

Fig. 1 Breastfeeding women remain at risk of multiple *P. vivax* relapses without access to primaquine. Following a single infectious bite from a mosquito during pregnancy or when breastfeeding, without access to efective radical cure with primaquine breastfeeding women are at high risk of repeated *P. vivax* malaria relapses. Relapses are damaging to a patient's health, causing chronic anaemia with associated morbidity and an increased mortality risk. Although chloroquine prophylaxis during pregnancy and while breastfeeding is recommended, it is not deployed in endemic regions so most women will be trapped in this cycle of repeated relapses until they complete breastfeeding or until their child is ≥6 months old and proven not to be G6PD defcient

administration and the level of G6PD enzyme activity [11–13, 16, 21].

G6PD deficiency is the most common X-linked enzymopathy worldwide, and prevalence is highest in populations that are exposed currently or historically to malaria, with the allelic frequency averaging around 8%, but as high as 33% in some populations $[22]$. The high prevalence of G6PD deficiency and the potential adverse consequences have hampered *P. vivax* radical cure using 8-aminoquinolines. To address the challenge of G6PD deficiency diagnosis, point-of-care quantitative G6PD tests have recently become available, with the potential to direct appropriate therapy and expand access to *P. vivax* radical cure [23–26]. This permits single-dose tafenoquine treatment for patients≥2 years old with normal G6PD activity, a daily primaquine regimen (7 or 14 days) for patients≥6 months of age who have G6PD normal or intermediate activity, and a weekly primaquine dose (0.75 mg/kg/week) for 8 weeks for G6PD-defcient patients [2]. Potentially, the extended weekly regimen can also be used where G6PD testing is not available.

Access to *P. vivax* radical cure is restricted in pregnant and breastfeeding women. Updated in October 2023, the World Health Organization (WHO) guidelines prohibit primaquine during pregnancy or to women breastfeeding infants aged<6 months or women breastfeeding infants aged≥6 months if their child is G6PD defcient or has unknown G6PD status [2]. As a relatively new treatment option, the inclusion of tafenoquine in the WHO malaria guidelines is currently under review. However, the product insert states that breastfeeding women should not take tafenoquine if they have a child who is known to have G6PD defciency or who has not been tested for G6PD defciency and women should not breastfeed for three months following the last dose of tafenoquine [14].

Historically, restrictions on the administration of 8-aminoquinolines during breastfeeding have been based on infant safety. However, primaquine has been in clinical use for around 70 years, and yet there has been no systematic effort to assess the primaquine risk: benefit during breastfeeding. Although a weekly primaquine regimen is recommended for G6PD-defcient adults, this regimen appears to be contraindicated in women breastfeeding children of any age [2].

Lactating women are underserved with limited options and resources to adequately meet their needs for malaria prevention and treatment [27]. Recent studies supporting the administration of primaquine to breastfeeding women for *P. vivax* radical cure have prompted calls for policy change [28–32]. Here, the totality of the available evidence is presented, emphasising the urgent need and supporting equitable access to primaquine to better serve the health of breastfeeding women.

Limitations for prevention of *P. vivax* **relapses in breastfeeding women**

A study at the Thailand–Myanmar border found that a history of *P. vivax* infection during pregnancy dramatically increased the risk of subsequent *P. vivax* malaria by 1398% [33]. Tis 14-fold increase in risk for *P. vivax* malaria is likely because pregnant women who experience an acute *P. vivax* malaria episode subsequently experience repeated *P. vivax* relapses. Figure 1 illustrates the impact of restricting primaquine in pregnant and breastfeeding women on the prevention of *P. vivax* relapses. Extending the availability of *P. vivax* radical cure to breastfeeding women would likely greatly decrease the burden of malaria in this population.

To prevent *P. vivax* infections in women who are pregnant or breastfeeding and not eligible for primaquine, the WHO recommends weekly chemoprophylaxis with chloroquine (300 mg base adult dose) [2, 8, 34–36]. However, in low transmission areas, where the risk of a new infection is lower than the beneft of weekly chloroquine chemoprophylaxis, the intervention could be used following an acute *P. vivax* malaria episode in women who are at risk of relapse $[8, 34-36]$. The presence of therapeutic concentrations of chloroquine in the blood should prevent both newly acquired blood-stage infections and suppress most acute *P. vivax* malaria episodes caused by relapses [8, 34]. However, hypnozoites will persist and the probability of a relapse increases if adherence is sub-optimal.

The operational feasibility and cost-effectiveness of chloroquine chemoprophylaxis for relapse suppression have not been investigated. There are no data on the adherence of breastfeeding women to the regimen in operational settings or its long-term safety. Information on its deployment and accessibility in *P. vivax* endemic regions is also lacking. The effectiveness of this regimen in suppressing relapse in areas with chloroquine-resistant *P. vivax* is unknown, though artemisinin-based combination therapy has been suggested $[2, 37]$. Thus, once infected with *P. vivax,* it is likely that most women will remain vulnerable to multiple *P. vivax* relapses while breastfeeding (Figs. 1, 2A).

Breastfeeding versus protection from *P. vivax* **relapse**

To promote infant and child health and development, the WHO recommends exclusive breastfeeding for the frst 6 months of life with continued breastfeeding until at least 2 years of age [38]. Although WHO guidance does not advocate that women should pause breastfeeding in order to receiving primaquine, clinicians may advise suspending breastfeeding, with harmful efects on breastfeeding outcomes [39]. Women must either Treatment of blood-stage infection: chloroquine or ACT as needed for acute P. vivax malaria

Chloroquine (CQ) weekly chemoprophylaxis (vertical bars indicate doses) $\overline{\mathbf{u}}$

Radical cure: chloroquine or ACT plus primaguine as needed for the treatment of acute P. vivax malaria and prevention of relapse

Fig. 2 *Plasmodium vivax* radical cure with primaquine during pregnancy and breastfeeding. Diferent possible scenarios for employing radical cure (green bars) for the management of *P. vivax* malaria and prevention of relapse for a single pregnancy in terms of the timing of restrictions to drugs during pregnancy and breastfeeding for 24 months. (A) and (B) are based on the WHO recommendations updated in October 2023, (C) is based on clinical pharmacokinetic data and (D) on PBPK modelling. The aim is to maximize the time during which *P. vivax* radical cure could be administered (green bars) and negate the need for infant G6PD testing. Thus, scenario D is preferred, followed by C, whereas scenarios B and A restrict the availability of radical cure to breastfeeding women unless their child is ≥6 months old and shown not to be G6PD defcient. (A) The most common scenario where chloroquine or an artemisinin-based combination therapy (ACT) is available as needed for the treatment of uncomplicated acute *P. vivax* malaria, and chloroquine chemoprophylaxis is not available. Radical cure, which is chloroquine or an ACT plus primaquine to treat uncomplicated *P. vivax* malaria and prevent relapse, can only be given to women breastfeeding infants≥6 months old if the child is known to not be G6PD defcient. (B) Chloroquine chemoprophylaxis is not usually available and is unlikely to be cost-efective in low transmission zones unless the mother has had acute *P. vivax* malaria [2]. However, the October 2023 recommendations are for weekly chloroquine for *P. vivax* malaria prevention for at least 15 months (~65 chloroquine doses) while pregnant and while breastfeeding if radical cure is contraindicated. Once the child is ≥6 months old, radical cure, which is chloroquine or an ACT plus primaquine to treat uncomplicated *P. vivax* malaria and prevent relapse, can be given to women only if their child is known to not be G6PD defcient. (C) Based on clinical pharmacokinetic data of primaquine exposures in breast milk and plasma in infants≥28 days old, radical cure with chloroquine or ACT plus primaquine could be given from 28 days post-partum to breastfeeding women, regardless of the G6PD status of the child (normal, deficient or unknown). If available, chloroquine chemoprophylaxis could be given while pregnant and for the frst month of breastfeeding. If chloroquine chemoprophylaxis is not available, treatment with chloroquine could be given during pregnancy and during the frst month of breastfeeding. (D) PBPK modelling of infant exposure via breast milk in children<28 days old supports radical cure with chloroquine or ACT plus primaquine for breastfeeding women soon after birth, regardless of the G6PD status of the child (normal, defcient or unknown). If available, chloroquine chemoprophylaxis could be given while pregnant. If chloroquine chemoprophylaxis is not available, treatment with chloroquine could be given during pregnancy

face the uncertainty of repeated *P. vivax* relapses and the attendant health risks or pause breastfeeding and risk the health of their child (Fig. 1).

Assuming chloroquine chemoprophylaxis is available, to simultaneously follow the WHO requirements for breastfeeding, as well as the recommendations for *P. vivax* prevention throughout pregnancy until breastfeeding is completed would require 15 to 33 months of weekly chloroquine chemoprophylaxis per pregnancy. The regimen would potentially be shorter if started following an acute episode of *P. vivax* malaria, which may occur later in the pregnancy (Fig. 2B).

For women breastfeeding for the 2 years recommended by the WHO, assuming they have three children (based on fertility rates in Indonesia and Pakistan of 2.4–3.4), this represents 2.25 years of pregnancy and 6 years of breastfeeding $[30]$. Thus, a woman could be excluded from primaquine treatment and potentially vulnerable to *P. vivax* relapse for more than 8 years if her children were G6PD-defcient or if their G6PD status was unknown [30]. A study conducted on the Thailand–Myanmar border reported that one woman was excluded from radical cure of *P. vivax* malaria for >10 years due to consecutive cycles of pregnancy and breastfeeding [28].

Even if the child has been tested and is not G6PD defcient, the restriction on primaquine for breastfeeding mothers of infants<6 months old would still exclude the mother from radical cure for a total of 3.75 years over three pregnancies (Fig. 2A, B). However, this would be reduced to as little as 2.25 years in total if primaquine was only contraindicated during pregnancy and could be given post-partum regardless of the G6PD status of the child (Fig. 2C, D).

Substantial risks to women from *P. vivax* **relapse**

The greatest risk to post-partum women with *P*. *vivax* malaria is anaemia, which can be severe or lifethreatening [40, 41]. Maternal anaemia is common in many malaria endemic countries, but pregnant women with symptomatic *P. vivax* infection are over five times more likely to be anaemic compared with non-infected pregnant women [42]. Even asymptomatic *P. vivax* malaria during pregnancy can increase the risk of maternal anaemia [40]. Consequently, during post-partum, women who are at risk of *P. vivax* relapse are likely to be already anaemic. Repeated *P. vivax* relapses compound their anaemia, causing morbidity and leaving the individual susceptible to other diseases [18, 27]. Severe *P. vivax* malaria is also a risk, particularly in breastfeeding women who may be malnourished or have additional co-morbidities [4, 27, 41, 43]. It is, therefore, essential to provide radical cure for new mothers who have had *P. vivax* malaria during pregnancy as soon as possible following birth to prevent relapse, allowing haematological recovery and preventing severe anaemia [27].

It is also necessary to ensure that women who have had *P. vivax* during pregnancy or while breastfeeding receive radical cure before their next pregnancy. If there is no opportunity for administration of 8-aminoquinolines between children, women remain vulnerable to relapses during pregnancy, with potentially serious consequences for the mother, foetus, and newborn [40, 41, 44–47].

Malaria elimination requires access to *P. vivax* **radical cure**

To accelerate malaria elimination, the greatest proportion of the population possible must have access to *P. vivax* radical cure. The persistence of the hypnozoite reservoir signifcantly contributes to malaria morbidity, mortality and transmission [48, 49]. To deplete this reservoir requires efective radical cure with high coverage of the population at risk. Where dosing is inadequate, adherence sub-optimal, or coverage is insufficient, the hypnozoite reservoir will continue to fuel relapses and transmission [8, 9, 30, 50, 51].

There has been considerable progress in modelling *P*. *vivax* transmission dynamics to consider the impact of the hypnozoite reservoir on malaria control and elimination. In a recent model, increasing *P. vivax* radical cure coverage reduced disease prevalence, with high coverage (82% of *P. vivax* infections) driving the parasite to near elimination over a period of 10 years, even without the use of mass drug administration [52]. A diferent model based on data from the Republic of Korea showed a decline in *P. vivax* prevalence with increasing coverage of radical cure [53]. Any new strategies for *P. vivax* malaria elimination based on radical cure will need to achieve high coverage to be efective and must therefore include breastfeeding women [3, 54].

Around 13% of women are excluded from radical cure because they are pregnant or breastfeeding [30]. This is a substantial fraction of the population in need of treatment and will sustain the hypnozoite reservoir, potentially hindering *P. vivax* control and elimination. However, if breastfeeding women could receive primaquine, then only around 4% of women would be excluded from radical cure $[30]$. Thus, extending primaquine radical cure to breastfeeding women is crucial to successfully support *P. vivax* malaria control and elimination.

Primaquine exposure in breastfeeding infants

Restrictions on the use of primaquine in breastfeeding mothers assumes that primaquine concentrations in breast milk may be sufficient to provoke haemolysis in a G6PD-defcient infant. However, the available clinical data for primaquine provide strong evidence that this is not the case [28].

Conducted in the Thailand–Myanmar border region, primaquine pharmacokinetics and haematological safety parameters were evaluated in 21 healthy G6PD-normal women with previous *P. vivax* infection and their healthy,

breastfeeding, G6PD-normal infants aged between 28 days and 2 years [28]. Primaquine was administered to the mothers for 14 days at the standard dose of 0.5 mg/ kg/day. The nursing infants showed no differences in haematological fndings versus age-matched controls [28].

Drug concentrations of primaquine and its major metabolite carboxyprimaquine were measured in maternal plasma, breast milk and infant plasma [28]. Plasma pharmacokinetic parameters for primaquine and carboxyprimaquine in the breastfeeding women were similar to those reported for non-pregnant or nonbreastfeeding adults $[55-58]$. The median values for the maximum primaquine concentration (C_{max}) and the area under the concentration time curve 24 h after dosing (AUC_{0-t}) were around threefold lower in breast milk

than in maternal plasma, i.e., median (range) C_{max} was 44.0 (31.4–99.0) ng/mL versus 139 (66.1–215) ng/mL, and AUC_{0-t} was 420 (179–1150) ng⋅h/mL versus 1220 (499–2360) ng∙h/mL, respectively (Fig. 3) [28]. For carboxyprimaquine the difference in C_{max} and AUC between maternal plasma and breast milk was more pronounced, being at least 113-fold and 163-fold lower in breast milk, respectively (Fig. 3) [28].

In the clinical lactation study, infant plasma primaquine concentrations were all measured to be below the lower limit of quantifcation (1.14 ng/mL), except for one sample that was 2.59 ng/mL on day 7 $[28]$. The resulting infant plasma carboxyprimaquine concentrations were negligible (0–25.8 ng/mL) compared to observed maximum plasma concentrations in mothers

Fig. 3 Pharmacokinetic parameters for primaquine and carboxyprimaquine in maternal plasma and breast milk [28]. Primaquine dose in mother was 0.5 mg/kg/day for 14 days; data are presented as median±range. AUC_{0-t}, area under the concentration–time curve until the last sampling point (24 h after administration); C_{max}, maximum drug concentration. The lower limit of quantification was 1.14 ng/mL for primaquine and 4.88 ng/ mL for carboxyprimaquine

receiving the treatment (486–2250 ng/mL) [28]. To put this in context, the single 0.25 mg/kg dose of primaquine, which is recommended to block transmission in patients with falciparum malaria without the need for G6PD testing [2], had a median (range) plasma primaquine C_{max} of 103 (38–174) ng/mL and an AU $C_{0-\text{last}}$ of 730 (297–1251) ng∙h/mL in children with acute falciparum malaria aged > 6 months to 1 year [59].

In a follow-up analysis, the clinical pharmacokinetic plasma and breast milk data from the mothers were evaluated using a simultaneously ftted population pharmacokinetic model. The data did not allow for developing a model for the infant plasma data as almost all of the observed concentrations were below the lower limit of quantifcation. However, based on breast milk concentrations and a mechanistic breastfeeding model, primaquine and carboxyprimaquine concentrations in infant plasma were simulated $[31]$. These simulations showed an infant primaquine plasma concentration below 1 ng/mL and carboxyprimaquine plasma concentrations below 2 ng/mL, comparable to that of the observed infant data. The simulations also demonstrated that for all of the currently recommended and used primaquine dosing regimens for radical cure and transmission blocking (0.5 mg/kg/day for 14 days, 1 mg/kg/day for 7 days, 0.5 mg/kg/day for 7 days, 0.75 mg/kg/week and 0.25 mg/kg single dose), breastfeeding infants received less than 1% of the weightadjusted maternal dose [31].

Based on the clinical fndings for the 0.5 mg/kg/day primaquine dose for 14 days, the relative primaquine dose in infants versus mothers was estimated to be between 0.23 and 1.82% on day 0 and 0.21% to 1.64% on day 13 [28]. A relative infant dose surpassing 10% of the maternal dose is conventionally regarded as a threshold warranting concern [60, 61], but as a fxed threshold, this overlooks the potential toxicity of the drug $[61]$. However, it has been demonstrated that a single 0.25 mg/kg primaquine dose does not cause clinically signifcant haemolysis in G6PD-defcient adults [62]. In comparison, infants in this study received a total dose via breast milk of 0.042 mg/ kg spread over 14 days $[28]$. The very low calculated relative infant dose, along with the negligible primaquine and carboxyprimaquine concentrations in infant plasma, are highly unlikely to pose a haemolytic risk to a G6PD-defcient infant and should not prevent the administration of primaquine to breastfeeding women of nursing infants between 1 and 24 months of age [28].

These clinical data are compelling and support the use of primaquine in women with acute *P. vivax* malaria who are breastfeeding, without the requirement to determine the infant's G6PD status. This would reduce the period for which women are vulnerable to relapse to 10 months for any pregnancy and negate the risk of carrying that risk to subsequent pregnancies (Fig. 2C).

The role of physiologically‑based pharmacokinetic modelling

Although highly reassuring, the clinical fndings do not include all circumstances for breastfeeding mothers. For example, the results cannot be generalised to newborns (<28 days) because the composition of breast milk in the frst days after birth (colostrum) difers signifcantly from mature milk. To complement clinical studies, researchers are addressing the challenges of drug development in breastfeeding women by using PBPK modelling [29, 63–73]. PBPK modelling combines information on human physiology and known drug properties to understand and predict the pharmacokinetic properties of a drug in a target population. Drug factors within the model include physicochemical properties and metabolism, and intrinsic physiological factors include bodyweight, height, age, organ sizes and the expression of metabolising enzymes. Extrinsic factors such as food intake and co-medications can also be considered.

This approach allows exploration of various drug dosing regimens under diferent scenarios that are challenging to test clinically. Relevant demographic and physiological factors can be incorporated in PBPK models to simulate drug pharmacokinetics in diferent populations of interest. For example, virtual populations of women at diferent stages of breastfeeding or infants with diferent feeding patterns can be incorporated into the model and exposures estimated in neonates and infants via breast milk [29, 63–72].

PBPK modelling is used routinely in drug development and is well accepted by regulatory authorities, with clear guidance on the factors that must be considered, such as the uncertainties and bias [74–77]. For example, a credibility assessment framework has been developed with the aim of standardising regulatory evaluation of PBPK models across therapeutic products [78]. Similarly, a structured framework has been proposed using the Grading of Recommendations Assessment, Development, and Evaluation (GRADE) conceptual approach, for integrating model-derived evidence into health decision-making processes [79]. PBPK models can be verifed against existing clinical pharmacokinetic data for drugs, with integration of clinical and model-derived data for optimal decision-making.

Simulated primaquine concentrations in breast milk, neonates and infants

A recent paper used a previously verifed primaquine PBPK model to estimate the pharmacokinetics and variability of primaquine in breastfeeding women and in their infants via breast milk $[29]$. The model was optimized using the reported observed milk to plasma (M/P) ratio and verifed using the data from the clinical lactation study described above, with the virtual infant population defned based on the heights and weights of the clinical population [28, 29]. The simulated primaquine dose $(0.5 \text{ mg/kg/day}$ for 14 days) was the same as in the clinical lactation study [28, 29].

Model-predicted primaquine plasma concentrations in virtual infants were below 2 ng/mL across the duration of 14-day primaquine dosing in breastfeeding women [29]. These predicted concentrations were consistent with the reported clinical data for infants (\geq 28 days old) who had primaquine exposures below the lower limit of quantifcation [28] (Fig. 4A).

The model was also used to simulate primaquine concentrations in neonates (<28 days old), where no clinical data are available, given the difficulty in recruiting neonates into clinical trials. Primaquine is mainly metabolised by monoamine oxidase (MAO), and diferent scenarios were modelled considering varying degrees of maturation of MAO activity in the neonate. In all scenarios, the predicted exposures of primaquine in neonates were $<$ 1% of those reported in maternal plasma (Fig. 4A).

Using the published PBPK model, an additional analysis was performed for this paper investigating the 0.75 mg/ kg/week for 8 weeks primaquine dosing regimen, which is used clinically for G6PD-defcient patients or where G6PD testing is not available [2]. In this case, the simulated concentrations of primaquine in infants and neonates were below 3 ng/mL, which is less than 1% of the maternal plasma levels, throughout the 8-week dosing period (Fig. 4B).

Another factor to consider is the impact of changes in composition of human milk (from colostrum to mature milk) in the frst 2 months post-partum on the primaquine intake of infants via breast milk. Using the composition of mature milk, the predicted M/P ratio was 0.47, similar to the observed value of 0.34 [28, 29]. A sensitivity analysis was performed within the PBPK model, varying breast milk fat content (2–5%) and pH (7.2–7.6) to simulate the fuctuations in milk composition [29]. These indicated a range of M/P ratios of 0.2 to 0.48, with the predicted primaquine exposures via breast milk to infants receiving colostrum being lower than with mature milk [29].

There are limitations to the PBPK model, including limited understanding of drug absorption processes in young children and of the degree and efect of protein

binding within breast milk, as well as the sparse clinical data available for model verifcation in children, particularly in neonates $[67]$. There is also uncertainty regarding the sensitivity of neonates and infants to primaquine-induced oxidative toxicity. However, the model was conservative in its design. It incorporated the clinically reported M/P ratio in infants, assumed complete drug absorption from the gastrointestinal tract and a slow maturation of metabolising enzymes, representing a 'worst-case' scenario. Simulated variations in milk composition and the degree of metabolising enzyme maturation consistently indicated very low predicted exposures to primaquine in breastfeeding infants which were congruent with the available clinical data [28, 29].

The findings from the PBPK model support and supplement the clinical lactation study and population pharmacokinetic modelling. Taken together, the evidence of low primaquine exposures in breastfeeding neonates and infants suggests that primaquine could be safely given to breastfeeding women, regardless of the G6PD status of the child. This would allow administration of radical cure soon after birth, further reducing the period during which women are vulnerable to *P. vivax* relapse to just the duration of the pregnancy (Fig. 2D). It would also ensure that women received treatment with primaquine before being potentially 'lost to follow up' post-partum.

Conclusion: health equity for *P. vivax* **radical cure**

Addressing health equity for *P. vivax* radical cure is a public health priority, with signifcant societal and economic implications associated with inadequate access [5, 6, 18, 27, 41, 80, 81]. Women living in areas where *P. vivax* malaria is endemic may be excluded for many years from radical cure with primaquine while breastfeeding, threatening their health. Additionally, the indirect benefts to children when their mothers are treated appropriately versus the harmful efects of repeated relapses must be considered. There are also public health consequences, with healthcare systems having to choose between supporting adherence to many months of chloroquine chemoprophylaxis and managing the clinical consequences of repeated relapses, such as anaemia and potentially severe malaria [5, 6, 18, 27, 41]. Moreover, there are uncertainties around the prolonged use of chloroquine chemoprophylaxis, in terms of feasibility, adherence and potential safety risks to the mother and child. Additionally, the persistence of the hypnozoite transmission reservoir in the approximately 9% of women who are breastfeeding in endemic populations undermines malaria elimination and control activities [30].

The individual and societal benefits of being able to administer radical cure among breastfeeding women are likely to be substantial. The health of breastfeeding women should not be jeopardised when clinical data and population and PBPK modelling provide consistent evidence of very low primaquine exposures in breastfeeding infants, suggesting negligible haemolytic risk, regardless of the child's G6PD status [28, 29, 31]. *P. vivax* radical cure in this population should be integrated with strategies to provide comprehensive care for maternal and infant health [27, 82].

Despite there being no evidence of risk, the treatment guidelines have restricted breastfeeding women from using primaquine presumably because of the perceived risk that exposures through breast milk could be suffcient to cause haemolysis should the infant be G6PD deficient. However, the health interests of breastfeeding women should not be secondary to an unproven assumption of risk to the infant. Even considering the complex regulatory environment for the approval of drugs for breastfeeding women [83], it is unacceptable that primaquine, which has been used for more than 70 years, has been withheld from breastfeeding women [84].

Although robust, the clinical pharmacokinetic data are limited to one study including 21 mother and infant pairs [28]. However, more than 5 years after publication, neither a change in guidelines nor additional clinical studies occurred. Plans for a clinical trial evaluating the impact of diferent breast milk composition on primaquine and tafenoquine concentrations in breast milk in the early post-partum period were recently withdrawn because of challenges in obtaining ethical approval (ClinicalTrials.gov Identifer: NCT04984759). Although the study is being redesigned (NCT06191458), it highlights the diffculty in progressing clinical studies in breastfeeding women and the knowledge gaps that need addressing. Support for studies investigating the safety and efficacy of drugs in lactating women needs to come from all levels of the research continuum, starting with drug development to clinical trials implementation, with sufficient funding and appropriate regulatory and ethical oversight [82, 85, 86]. Where clinical data are limited, the integration of clinical data and PBPK modelling can be used to strengthen the evidence base and inform treatment recommendations and policy change. Similar approaches should become best practice for studying underserved populations, such as breastfeeding women, in an efficient manner to achieve equitable access for other medicines [29, 63, 82, 84–86].

Breastfeeding women should have access to primaquine without the need for G6PD testing of their infant. The evidence supports primaquine 0.5 mg/kg day for 7 or 14 days in breastfeeding women from early post-partum, unless the woman is G6PD defcient or if G6PD testing is not available, in which case primaquine 0.75 mg/kg/week for 8 weeks can be administered.

Based on the totality of evidence, it is of paramount importance to re-examine the recommendations around primaquine for *P. vivax* radical cure in breastfeeding women, and thus provide health equity for this critically important intervention.

Abbreviations

-
- MAO Monoamine oxidase
MDA Mass drug administr
- MDA Mass drug administration
M/P Milk to plasma Milk to plasma
-
- PBPK Physiologically-based pharmacokinetic modelling WHO World Health Organization
-

Acknowledgements

The authors thank Tim Wells (MMV) for his review of the manuscript and helpful suggestions.

Author contributions

NA and PZ developed the concept for the paper. NR wrote the frst draft of the paper. All authors critically reviewed and contributed to the paper and take responsibility for its publication.

Funding

This article was funded by MMV Medicines for Malaria Venture. This work was supported, in whole or in part, by the Bill & Melinda Gates Foundation (INV-007155). Under the grant conditions of the Foundation, a Creative Commons Attribution 4·0 Generic License has already been assigned to the Author Accepted Manuscript version that might arise from this submission. JT receives funding from the Wellcome Trust (220211).

Availability of data and materials

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

NA, ACM, EJ, SD, JJM and MEG are employees of MMV Medicines for Malaria Venture. PZ and JC are employees of the Bill & Melinda Gates Foundation. LA, KRY and XP are employees of Certara Ltd and may hold shared in the company. NR is a paid consultant for MMV Medicines for Malaria Venture. SK is employee of Montpellier University. CW is funded by Wellcome Clinical Research Career Development Fellowship (222075_Z_20_Z). CK is funded by the Bill & Melinda Gates Foundation (INV-023795). AB and SC work for the Medicines and Healthcare products Regulatory Agency and are funded by the Bill & Melinda Gates Foundation (INV-009383). JT declares no competing interests.

Author details

¹MMV Medicines for Malaria Venture, 20 Route de Pré-Bois, 1215 Geneva 15, Switzerland. ²Magenta Communications Ltd, Abingdon, UK. ³Certara Predictive Technologies, Simcyp Division, Sheffield, UK. ⁴Mahidol Oxford Tropical Medicine Research Unit, Faculty of Tropical Medicine, Mahidol University, Bangkok, Thailand. ⁵Nuffield Department of Clinical Medicine, Centre for Tropical Medicine and Global Health, University of Oxford, Oxford, UK. Bill & Melinda Gates Foundation, Seattle, WA, USA. ⁷ Department of Pharmacology and Therapeutics, University of Liverpool, Liverpool, UK. ⁸Infectious Diseases Institute, Makerere University College of Health Sciences, Kampala, Uganda. ⁹ Division of Pharmacology Toxicology, Department of Pharmacy, Radboud University Medical Centre, Nijmegen, The Netherlands. ¹⁰Medicines and Healthcare products Regulatory Agency (MHRA), 10 South Colonnade,

London, UK.¹¹Pharmacokinetic and Modelling Department, School of Pharmacy, IMAG, CNRS, INRIA, UMR 5149, University of Montpellier, Montpellier, France.

Received: 2 May 2024 Accepted: 13 September 2024

References

- WHO. World malaria report 2023. Geneva: World Health Organization; 2023. [https://www.who.int/teams/global-malaria-programme/reports/](https://www.who.int/teams/global-malaria-programme/reports/world-malaria-report-2023) [world-malaria-report-2023.](https://www.who.int/teams/global-malaria-programme/reports/world-malaria-report-2023) Accessed 8 Jan 2024.
- 2. WHO. Guidelines for malaria (version 7.1). Geneva: World Health Organization; 2023. [https://www.who.int/publications/i/item/guidelines-for-malar](https://www.who.int/publications/i/item/guidelines-for-malaria) [ia.](https://www.who.int/publications/i/item/guidelines-for-malaria) Accessed 8 Jan 2024.
- 3. Nekkab N, Obadia T, Monteiro WM, Lacerda MVG, White M, Mueller I. Accelerating towards *P. vivax* elimination with a novel serological testand-treat strategy: a modelling case study in Brazil. Lancet Reg Health Am. 2023;22: 100511.
- 4. Douglas NM, Pontororing GJ, Lampah DA, Yeo TW, Kenangalem E, Poespoprodjo JR, et al. Mortality attributable to *Plasmodium vivax* malaria: a clinical audit from Papua, Indonesia. BMC Med. 2014;12:217.
- 5. Lanca EF, Magalhaes BM, Vitor-Silva S, Siqueira AM, Benzecry SG, Alexandre MA, et al. Risk factors and characterization of *Plasmodium vivax*associated admissions to pediatric intensive care units in the Brazilian Amazon. PLoS ONE. 2012;7: e35406.
- 6. Poespoprodjo JR, Fobia W, Kenangalem E, Lampah DA, Hasanuddin A, Warikar N, et al. Vivax malaria: a major cause of morbidity in early infancy. Clin Infect Dis. 2009;48:1704–12.
- 7. White NJ. Determinants of relapse periodicity in *Plasmodium vivax* malaria. Malar J. 2011;10:297.
- 8. Corder RM, de Lima ACP, Khoury DS, Docken SS, Davenport MP, Ferreira MU. Quantifying and preventing *Plasmodium vivax* recurrences in primaquine-untreated pregnant women: an observational and modeling study in Brazil. PLoS Negl Trop Dis. 2020;14: e0008526.
- 9. Commons RJ, Rajasekhar M, Edler P, Abreha T, Awab GR, Baird JK, et al. Efect of primaquine dose on the risk of recurrence in patients with uncomplicated *Plasmodium vivax*: a systematic review and individual patient data meta-analysis. Lancet Infect Dis. 2023;24:172–83.
- 10. Soto AM, Gonzalez-Ceron L, Santillan-Valenzuela F, Parrales ME, Montoya A. Recurrent *Plasmodium vivax* cases of both short and long latency increased with transmission intensity and were distributed year-round in the most afected municipalities of the RACCN, Nicaragua, 2013–2018. Int J Environ Res Public Health. 2022;19:6195.
- 11. Bancone G, Chu CS. G6PD variants and haemolytic sensitivity to primaquine and other drugs. Front Pharmacol. 2021;12: 638885.
- 12. Rueangweerayut R, Bancone G, Harrell EJ, Beelen AP, Kongpatanakul S, Mohrle JJ, et al. Hemolytic potential of tafenoquine in female volunteers heterozygous for glucose-6-phosphate dehydrogenase (G6PD) defciency (G6PD Mahidol variant) versus G6PD-normal volunteers. Am J Trop Med Hyg. 2017;97:702–11.
- 13. Chu CS, Bancone G, Nosten F, White NJ, Luzzatto L. Primaquine-induced haemolysis in females heterozygous for G6PD defciency. Malar J. 2018;17:101.
- 14. GSK. KRINTAFEL prescribing information. Durham, GSK; 2023. [https://](https://gskpro.com/content/dam/global/hcpportal/en_US/Prescribing_Information/Krintafel/pdf/KRINTAFEL-PI-PIL.PDF) [gskpro.com/content/dam/global/hcpportal/en_US/Prescribing_Infor](https://gskpro.com/content/dam/global/hcpportal/en_US/Prescribing_Information/Krintafel/pdf/KRINTAFEL-PI-PIL.PDF) [mation/Krintafel/pdf/KRINTAFEL-PI-PIL.PDF.](https://gskpro.com/content/dam/global/hcpportal/en_US/Prescribing_Information/Krintafel/pdf/KRINTAFEL-PI-PIL.PDF) Accessed 13 Mar 2024.
- 15. Sanof-Aventis. Primaquine phosphate prescribing information. Bridgewater, Sanof-Aventis; 2017. [https://www.accessdata.fda.gov/drugsatfda_](https://www.accessdata.fda.gov/drugsatfda_docs/label/2017/008316s023lbl.pdf) [docs/label/2017/008316s023lbl.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/label/2017/008316s023lbl.pdf). Accessed 13 Mar 2024.
- 16. Rajasekhar M, Simpson JA, Ley B, Edler P, Chu CS, Abreha T, et al. Primaquine dose and the risk of haemolysis in patients with uncomplicated *Plasmodium vivax* malaria: a systematic review and individual patient data meta-analysis. Lancet Infect Dis. 2023;24:184–95.
- 17. Chu CS, Bancone G, Soe NL, Carrara VI, Gornsawun G, Nosten F. The impact of using primaquine without prior G6PD testing: a case series describing the obstacles to the medical management of haemolysis. Wellcome Open Res. 2019;4:25.
- 18. Commons RJ, Simpson JA, Thriemer K, Chu CS, Douglas NM, Abreha T, et al. The haematological consequences of *Plasmodium vivax* malaria after chloroquine treatment with and without primaquine: a WorldWide Antimalarial Resistance Network systematic review and individual patient data meta-analysis. BMC Med. 2019;17:151.
- 19. Yilma D, Groves ES, Brito-Sousa JD, Monteiro WM, Chu C, Thriemer K, et al. Severe hemolysis during primaquine radical cure of *Plasmodium vivax* malaria: two systematic reviews and individual patient data descriptive analyses. Am J Trop Med Hyg. 2023;109:761–9.
- 20. Brito-Sousa JD, Santos TC, Avalos S, Fontecha G, Melo GC, Val F, et al. Clinical spectrum of primaquine-induced hemolysis in glucose-6-phosphate dehydrogenase defciency: a 9-year hospitalization-based study from the Brazilian Amazon. Clin Infect Dis. 2019;69:1440–2.
- 21. Pfeffer DA, Satyagraha AW, Sadhewa A, Alam MS, Bancone G, Boum Y 2nd, et al. Genetic variants of glucose-6-phosphate dehydrogenase and their associated enzyme activity: a systematic review and meta-analysis. Pathogens. 2022;11:1045.
- 22. Howes RE, Piel FB, Patil AP, Nyangiri OA, Gething PW, Dewi M, et al. G6PD defciency prevalence and estimates of afected populations in malaria endemic countries: a geostatistical model-based map. PLoS Med. 2012;9: e1001339.
- 23. Gerth-Guyette E, Nguyen HT, Nowak S, Hoang NT, Mai DTT, Thi Sang V, et al. Assessing the operational feasibility of integrating point-of-care G6PD testing into *Plasmodium vivax* malaria management in Vietnam. Pathogens. 2023;12:689.
- 24. Adhikari B, Tripura R, Dysoley L, Peto TJ, Callery JJ, Heng C, et al. Glucose-6-phosphate dehydrogenase (G6PD) measurement using biosensors by community-based village malaria workers and hospital laboratory staf in Cambodia: a quantitative study. Pathogens. 2023;12:400.
- 25. Brito-Sousa JD, Peixoto HM, Devine A, Silva-Neto AV, Balieiro PCS, Sampaio VS, et al. Real-life quantitative G6PD screening in *Plasmodium vivax* patients in the Brazilian Amazon: a cost-efectiveness analysis. PLoS Negl Trop Dis. 2022;16: e0010325.
- 26. Brito-Sousa JD, Murta F, Vitor-Silva S, Sampaio V, Mendes M, Souza B, et al. Quantitative G6PD defciency screening in routine malaria diagnostic units in the Brazilian Amazon (SAFEPRIM): an operational mixed-methods study. Pathogens. 2022;11:1328.
- 27. Brummaier T, Gilder ME, Gornsawun G, Chu CS, Bancone G, Pimanpanarak M, et al. Vivax malaria in pregnancy and lactation: a long way to health equity. Malar J. 2020;19:40.
- 28. Gilder ME, Hanpithakphong W, Hoglund RM, Tarning J, Win HH, Hilda N, et al. Primaquine pharmacokinetics in lactating women and breastfed infant exposures. Clin Infect Dis. 2018;67:1000–7.
- 29. Pan X, Abduljalil K, Almond LM, Pansari A, Yeo KR. Supplementing clinical lactation studies with PBPK modeling to inform drug therapy in lactating mothers: prediction of primaquine exposure as a case example. CPT Pharmacomet Syst Pharmacol. 2024;13:386–95.
- 30. Watson J, Taylor WRJ, Bancone G, Chu CS, Jittamala P, White NJ. Implications of current therapeutic restrictions for primaquine and tafenoquine in the radical cure of vivax malaria. PLoS Negl Trop Dis. 2018;12: e0006440.
- 31. Wattanakul T, Gilder ME, McGready R, Hanpithakpong W, Day NP, White N, et al. Population pharmacokinetic modelling of primaquine exposures in lactating women and breastfed infants. Nat Commun. 2024;14:3851.
- 32. Price RN, Douglas NM. Expanding the use of primaquine for the radical cure of *Plasmodium vivax*. Clin Infect Dis. 2018;67:1008–9.
- 33. Boel ME, Rijken MJ, Leenstra T, Phyo AP, Pimanpanarak M, Keereecharoen NL, et al. Malaria in the post-partum period; a prospective cohort study. PLoS ONE. 2013;8: e57890.
- 34. Villegas L, McGready R, Htway M, Paw MK, Pimanpanarak M, Arunjerdja R, et al. Chloroquine prophylaxis against vivax malaria in pregnancy: a randomized, double-blind, placebo-controlled trial. Trop Med Int Health. 2007;12:209–18.
- 35. Wolfe MS, Cordero JF. Safety of chloroquine in chemosuppression of malaria during pregnancy. Br Med J (Clin Res Ed). 1985;290:1466–7.
- 36. Ogunbona FA, Onyeji CO, Bolaji OO, Torimiro SE. Excretion of chloroquine and desethylchloroquine in human milk. Br J Clin Pharmacol. 1987;23:473–6.
- 37. Ahmed R, Poespoprodjo JR, Syafruddin D, Khairallah C, Pace C, Lukito T, et al. Efficacy and safety of intermittent preventive treatment and intermittent screening and treatment versus single screening and

treatment with dihydroartemisinin–piperaquine for the control of malaria in pregnancy in Indonesia: a cluster-randomised, open-label, superiority trial. Lancet Infect Dis. 2019;19:973–87.

- 38. World Health Organization. Breastfeeding recommendations. Geneva: WHO; 2023. [https://www.who.int/health-topics/breastfeeding#tab](https://www.who.int/health-topics/breastfeeding#tab=tab_2)= [tab_2.](https://www.who.int/health-topics/breastfeeding#tab=tab_2) Accessed 9 Jan 2024.
- 39. Akus M, Bartick M. Lactation safety recommendations and reliability compared in 10 medication resources. Ann Pharmacother. 2007;41:1352–60.
- 40. Pincelli A, Neves PAR, Lourenco BH, Corder RM, Malta MB, Sampaio-Silva J, et al. The hidden burden of *Plasmodium vivax* malaria in pregnancy in the amazon: an observational study in Northwestern Brazil. Am J Trop Med Hyg. 2018;99:73–83.
- 41. Phyo AP, Dahal P, Mayxay M, Ashley EA. Clinical impact of vivax malaria: a collection review. PLoS Med. 2022;19: e1003890.
- 42. Bardaji A, Martinez-Espinosa FE, Arevalo-Herrera M, Padilla N, Kochar S, Ome-Kaius M, et al. Burden and impact of *Plasmodium vivax* in pregnancy: a multi-centre prospective observational study. PLoS Negl Trop Dis. 2017;11: e0005606.
- 43. Boltena MT, El-Khatib Z, Sahlemichael Kebede A, Asamoah BO, Tadesse Boltena A, Yeshambaw M, et al. Comorbidity of geo-helminthes among malaria outpatients of the health facilities in Ethiopia: systematic review and meta-analysis. Int J Environ Res Public Health. 2021;18:862.
- 44. Cardona-Arias JA, Carmona-Fonseca J. Congenital malaria: frequency and epidemiology in Colombia, 2009–2020. PLoS ONE. 2022;17: e0263451.
- 45. Dombrowski JG, Barateiro A, Peixoto EPM, Barros A, Souza RM, Clark TG, et al. Adverse pregnancy outcomes are associated with *Plasmodium vivax* malaria in a prospective cohort of women from the Brazilian Amazon. PLoS Negl Trop Dis. 2021;15: e0009390.
- 46. Botto-Menezes C, Silva Dos Santos MC, Lopes Simplicio J, Menezes de Medeiros J, Barroso Gomes KC, de Carvalho Costa IC, et al. *Plasmodium vivax* malaria in pregnant women in the Brazilian amazon and the risk factors associated with prematurity and low birth weight: a descriptive study. PLoS ONE. 2015;10: e0144399.
- 47. Chandrashekhar VN, Punnath K, Dayanand KK, Kakkilaya SB, Jayadev P, Kumari SN, et al. Impact of oxidative stress in response to malarial infection during pregnancy: complications, histological changes, and pregnancy outcomes. Trop Parasitol. 2022;12:21–33.
- 48. Wells TN, Burrows JN, Baird JK. Targeting the hypnozoite reservoir of *Plasmodium vivax*: the hidden obstacle to malaria elimination. Trends Parasitol. 2010;26:145–51.
- 49. Ahmad SS, Rahi M, Sharma A. Relapses of *Plasmodium vivax* malaria threaten disease elimination: time to deploy tafenoquine in India? BMJ Glob Health. 2021;6: e004558.
- 50. John GK, Douglas NM, von Seidlein L, Nosten F, Baird JK, White NJ, et al. Primaquine radical cure of *Plasmodium vivax*: a critical review of the literature. Malar J. 2012;11:280.
- 51. Milligan R, Daher A, Villanueva G, Bergman H, Graves PM. Primaquine alternative dosing schedules for preventing malaria relapse in people with *Plasmodium vivax*. Cochrane Database Syst Rev. 2020;8:CD012656.
- 52. Walker CR, Hickson RI, Chang E, Ngor P, Sovannaroth S, Simpson JA, et al. A model for malaria treatment evaluation in the presence of multiple species. Epidemics. 2023;44: 100687.
- 53. Kim S, Byun JH, Park A, Jung IH. A mathematical model for assessing the efectiveness of controlling relapse in *Plasmodium vivax* malaria endemic in the Republic of Korea. PLoS ONE. 2020;15: e0227919.
- 54. Obadia T, Nekkab N, Robinson LJ, Drakeley C, Mueller I, White MT. Developing sero-diagnostic tests to facilitate *Plasmodium vivax* serological test-and-treat approaches: modeling the balance between public health impact and overtreatment. BMC Med. 2022;20:98.
- 55. Elmes NJ, Bennett SM, Abdalla H, Carthew TL, Edstein MD. Lack of sex efect on the pharmacokinetics of primaquine. Am J Trop Med Hyg. 2006;74:951–2.
- 56. Ward SA, Mihaly GW, Edwards G, Looareesuwan S, Phillips RE, Chanthavanich P, et al. Pharmacokinetics of primaquine in man. II. Comparison of acute vs chronic dosage in Thai subjects. Br J Clin Pharmacol. 1985;19:751–5.
- 57. Mihaly GW, Ward SA, Edwards G, Nicholl DD, Orme ML, Breckenridge AM. Pharmacokinetics of primaquine in man. I. Studies of the absolute bioavailability and efects of dose size. Br J Clin Pharmacol. 1985;19:745–50.
- 58. Mihaly GW, Ward SA, Edwards G, Orme ML, Breckenridge AM. Pharmacokinetics of primaquine in man: identifcation of the carboxylic

acid derivative as a major plasma metabolite. Br J Clin Pharmacol. 1984;17:441–6.

- 59. Mukaka M, Onyamboko MA, Olupot-Olupot P, Peerawaranun P, Suwannasin K, Pagornrat W, et al. Pharmacokinetics of single low dose primaquine in Ugandan and Congolese children with falciparum malaria. EBioMedicine. 2023;96: 104805.
- 60. Bennett P. Use of the monographs on drugs. In: Bennett P, editor. Drugs and human lactation: a comprehensive guide to the content and consequences of drugs, micronutrients, radiopharmaceuticals, and environmental and occupational chemicals in human milk. 2nd ed. Amsterdam: Elsevier; 1996. p. 67–74.
- 61. Ojara FW, Kawuma AN, Waitt C. A systematic review on maternal-toinfant transfer of drugs through breast milk during the treatment of malaria, tuberculosis, and neglected tropical diseases. PLoS Negl Trop Dis. 2023;17: e0011449.
- 62. Bancone G, Chowwiwat N, Somsakchaicharoen R, Poodpanya L, Moo PK, Gornsawun G, et al. Single low dose primaquine (0.25 mg/kg) does not cause clinically signifcant haemolysis in G6PD defcient subjects. PLoS ONE. 2016;11: e0151898.
- 63. Abduljalil K, Pansari A, Ning J, Jamei M. Prediction of drug concentrations in milk during breastfeeding, integrating predictive algorithms within a physiologically-based pharmacokinetic model. CPT Pharmacomet Syst Pharmacol. 2021;10:878–89.
- 64. Allegaert K, Abbasi MY, Annaert P, Olafuyi O. Current and future physiologically based pharmacokinetic (PBPK) modeling approaches to optimize pharmacotherapy in preterm neonates. Expert Opin Drug Metab Toxicol. 2022;18:301–12.
- 65. Nauwelaerts N, Macente J, Deferm N, Bonan RH, Huang MC, Van Neste M, et al. Generic workfow to predict medicine concentrations in human milk using physiologically-based pharmacokinetic (PBPK) modelling—a contribution from the ConcePTION project. Pharmaceutics. 2023;15:1469.
- 66. Pressly MA, Schmidt S, Guinn D, Liu Z, Ceresa C, Samuels S, et al. Informing a comprehensive risk assessment of infant drug exposure from human milk: application of a physiologically based pharmacokinetic lactation model for sotalol. J Clin Pharmacol. 2023;63(Suppl 1):S106–16.
- 67. Van Neste M, Bogaerts A, Nauwelaerts N, Macente J, Smits A, Annaert P, et al. Challenges related to acquisition of physiological data for physiologically based pharmacokinetic (PBPK) models in postpartum, lactating women and breastfed infants—a Contribution from the ConcePTION Project. Pharmaceutics. 2023;15:2618.
- 68. Damoiseaux D, Amant F, Beijnen JH, Barnett S, Veal GJ, Huitema ADR, et al. Physiologically-based pharmacokinetic model to predict doxorubicin and paclitaxel exposure in infants through breast milk. CPT Pharmacomet Syst Pharmacol. 2023;12:1931–44.
- 69. Yeung CHT, Autmizguine J, Dalvi P, Denoncourt A, Ito S, Katz P, et al. Maternal ezetimibe concentrations measured in breast milk and its use in breastfeeding infant exposure predictions. Clin Pharmacokinet. 2024;63:317–32.
- 70. Cardoso E, Guidi M, Nauwelaerts N, Nordeng H, Teil M, Allegaert K, et al. Safety of medicines during breastfeeding—from case report to modeling: a contribution from the ConcePTION project. Expert Opin Drug Metab Toxicol. 2023;19:269–83.
- 71. Guinn D, Pressly MA, Liu Z, Ceresa C, Samuels S, Wang YM, et al. Exploring the knowledge gaps in infant drug exposure from human milk: a clinical pharmacology perspective. J Clin Pharmacol. 2023;63:273–6.
- 72. Pan X, Rowland YK. Physiologically based pharmacokinetic modeling to determine the impact of CYP2B6 genotype on efavirenz exposure in children, mothers and breastfeeding infants. Clin Pharmacol Ther. 2023;114:182–91.
- 73. Center for Drug Evaluation and Research. Clinical lactation studies: considerations for study design. Rockville: United States Food and Drug Administration; 2019. [https://www.fda.gov/regulatory-information/](https://www.fda.gov/regulatory-information/search-fda-guidance-documents/clinical-lactation-studies-considerations-study-design) [search-fda-guidance-documents/clinical-lactation-studies-considerat](https://www.fda.gov/regulatory-information/search-fda-guidance-documents/clinical-lactation-studies-considerations-study-design) [ions-study-design](https://www.fda.gov/regulatory-information/search-fda-guidance-documents/clinical-lactation-studies-considerations-study-design). Accessed 8 Apr 2024.
- 74. Zhang X, Yang Y, Grimstein M, Fan J, Grillo JA, Huang SM, et al. Application of PBPK modeling and simulation for regulatory decision making and its impact on US prescribing information: an update on the 2018–2019 submissions to the US FDA's Office of Clinical Pharmacology. J Clin Pharmacol. 2020;60(Suppl 1):S160–78.
- 75. Johnson TN, Small BG, Rowland YK. Increasing application of pediatric physiologically based pharmacokinetic models across academic

and industry organizations. CPT Pharmacomet Syst Pharmacol. 2022;11:373–83.

- 76. Lin W, Chen Y, Unadkat JD, Zhang X, Wu D, Heimbach T. Applications, challenges, and outlook for PBPK modeling and simulation: a regulatory, industrial and academic perspective. Pharm Res. 2022;39:1701–31.
- 77. European Medicines Agency. Guideline on the reporting of physiologically based pharmacokinetic (PBPK) modelling and simulation. London, European Medicines Agency; 2018. [https://www.ema.europa.](https://www.ema.europa.eu/en/documents/scientific-guideline/guideline-reporting-physiologically-based-pharmacokinetic-pbpk-modelling-and-simulation_en.pdf) [eu/en/documents/scientifc-guideline/guideline-reporting-physiologi](https://www.ema.europa.eu/en/documents/scientific-guideline/guideline-reporting-physiologically-based-pharmacokinetic-pbpk-modelling-and-simulation_en.pdf) [cally-based-pharmacokinetic-pbpk-modelling-and-simulation_en.pdf.](https://www.ema.europa.eu/en/documents/scientific-guideline/guideline-reporting-physiologically-based-pharmacokinetic-pbpk-modelling-and-simulation_en.pdf) Accessed 14 Mar 2024.
- 78. Kuemmel C, Yang Y, Zhang X, Florian J, Zhu H, Tegenge M, et al. Consideration of a credibility assessment framework in model-informed drug development: potential application to physiologically-based pharmacokinetic modeling and simulation. CPT Pharmacomet Syst Pharmacol. 2020;9:21–8.
- 79. Brozek JL, Canelo-Aybar C, Akl EA, Bowen JM, Bucher J, Chiu WA, et al. GRADE guidelines 30: the GRADE approach to assessing the certainty of modeled evidence—an overview in the context of health decisionmaking. J Clin Epidemiol. 2021;129:138–50.
- 80. Devine A, Battle KE, Meagher N, Howes RE, Dini S, Gething PW, et al. Global economic costs due to vivax malaria and the potential impact of its radical cure: a modelling study. PLoS Med. 2021;18: e1003614.
- 81. White MT, Yeung S, Patouillard E, Cibulskis R. Costs and cost-efectiveness of *Plasmodium vivax* control. Am J Trop Med Hyg. 2016;95:52–61.
- 82. El Gaaloul M, Tornesi B, Lebus F, Reddy D, Kaszubska W. Re-orienting anti-malarial drug development to better serve pregnant women. Malar J. 2022;21:121.
- 83. Alexe A, Garg A, Kovacs B, Abramova N, Apara O, Eisele O, et al. Regulations governing medicines for maternal and neonatal health: a landscape assessment. Ther Innov Regul Sci. 2024;58:242–57.
- 84. Weld ED, Bailey TC, Waitt C. Ethical issues in therapeutic use and research in pregnant and breastfeeding women. Br J Clin Pharmacol. 2022;88:7–21.
- 85. Illamola SM, Bucci-Rechtweg C, Costantine MM, Tsilou E, Sherwin CM, Zajicek A. Inclusion of pregnant and breastfeeding women in research eforts and initiatives. Br J Clin Pharmacol. 2018;84:215–22.
- 86. Jorgensen SCJ, Miljanic S, Tabbara N, Somanader D, Leung F, De Castro C, et al. Inclusion of pregnant and breastfeeding women in nonobstetrical randomized controlled trials. Am J Obstet Gynecol MFM. 2022;4: 100700.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.