

Six-probe scintillator dosimeter for treatment verification in HDR-brachytherapy

Mathieu Gonod, Miguel Angel Suarez, Carlos Chacon Avila, Vage Karakhanyan, Clément Eustache, Samir Laskri, Julien Crouzilles, Jean-François Vinchant, Léone Aubignac, Thierry Grosjean

▶ To cite this version:

Mathieu Gonod, Miguel Angel Suarez, Carlos Chacon Avila, Vage Karakhanyan, Clément Eustache, et al.. Six-probe scintillator dosimeter for treatment verification in HDR-brachytherapy. Medical Physics, 2023, 50 (11), pp.7192 - 7202. 10.1002/mp.16745 . hal-04801764

HAL Id: hal-04801764 https://hal.science/hal-04801764v1

Submitted on 25 Nov 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

¹ Six-probe scintillator dosimeter for treatment verification in ² HDR-brachytherapy

 Mathieu Gonod¹, Miguel Angel Suarez², Carlos Chacon Avila², Vage Karakhanyan², Clément Eustache², Samir Laskri³, Julien Crouzilles³, Jean-François Vinchant³, Léone Aubignac¹ and Thierry Grosjean²
 ¹Centre Georges François Leclerc (CGFL) - Dijon, France
 ²University of Franche-Comté, CNRS, FEMTO-ST Institute, UMR 6174, Besançon, France
 ³SEDI-ATI Fibres Optiques, 8 Rue Jean Mermoz, 91080 Évry-Courcouronnes, France
 Version typeset November 15, 2024

Author to whom correspondence should be addressed. email: thierry.grosjean@univfcomte.fr

12

13

21

22

23

24

25 26

Abstract

Background: In vivo dosimetry (IVD) is gaining interest for treatment delivery verification in HDR-brachytherapy. Time resolved methods, including source tracking, have the ability both to detect treatment errors in real time and to minimize experimental uncertainties. Multiprobe IVD architectures holds promise for simultaneous dose determinations at the targeted tumor and surrounding healthy tissues while enhancing measurement accuracy. However, most of the multiprobe dosimeters developed so far either suffer from compactness issues or rely on complex data post-treatment.

Purpose: We introduce a novel concept of a compact multiprobe scintillator detector and demonstrate its applicability in HDR-brachytherapy. Our fabricated seven-fiber probing system is sufficiently narrow to be inserted in a brachytherapy needle or in a catheter.

Methods: Our multiprobe detection system results from the parallel implementation 27 of six miniaturized inorganic Gd_2O_2S : Tb scintillator detectors at the end of a bundle 28 of seven fibers, one fiber is kept bare to assess the stem effect. The resulting system, 29 which is narrower than 320 microns, is tested with a MicroSelectron 9.14 Ci Ir-192 30 HDR afterloader, in a water phantom. The detection signals from all six probes are 31 simultaneously read with a sCMOS camera (at a rate of 0.06 s). The camera is coupled 32 to a chromatic filter to cancel Cerenkov signal induced within the fibers upon exposure. 33 By implementing an aperiodic array of six scintillating cells along the bundle axis, we 34 first determine the range of inter-probe spacings leading to optimal source tracking 35 accuracy (first tracking method). Then, three different source tracking algorithms in-36 volving all the scintillating probes are tested and compared. In each of these four 37 methods, dwell positions are assessed from dose measurements and compared to the 38

³⁹ treatment plan. Dwell time is also determined and compared to the treatment plan.

Results: The optimum inter-probe spacing for an accurate source tracking ranges from 15 mm to 35 mm. The optimum detection algorithm consists of adding the readout signals from all detector probes. In that case, the error to the planned dwell positions is of 0.01 ± 0.14 mm and 0.02 ± 0.29 mm at spacings between the source and detector axes of 5.5 and 40 mm, respectively. Using this approach, the average deviations to the expected dwell time are of -0.006 ± 0.009 s and -0.008 ± 0.058 s, at spacings between source and probe axes of 5.5 mm and 20 mm, respectively.

Conclusions: Our six-probe Gd₂O₂S:Tb dosimeter coupled to a sCMOS camera can
 perform time-resolved treatment verification in HDR brachytherapy. This detection
 system of high spatial and temporal resolutions (0.25 mm and 0.06s, respectively)
 provides a precise information on the treatment delivery via a dwell time and position
 verification of unmatched accuracy.

54

40

55

57

56 This is a sample note.

58 Contents

59	١.	Introduction	1
60	н.	Material and Methods	2
61		II.A. Multiprobe system	2
62		II.B. Optical readout	3
63		II.C. Brachytherapy system	5
64		II.D. Phantom	5
65		II.E. Detector calibration	5
66		II.F. Dwell position and dwell time verification	6
67		II.F.1. Measurements	6
68		II.F.2. Source position monitoring	6
69		II.F.3. Dwell time verification	7
70		. Results	8
71		III.A. Detector specification and calibration	8

72	III.B. Source position monitoring	8
73	III.B.1. Optimal probe-to-probe spacing	8
74	$III.B.2.$ Source tracking: two-probe versus all-probe detection strategies $\ .$.	11
75	III.C. Dwell time verification	13
76	IV. Discussion	14
77	IV.A. Detector specification and calibration	14
78	IV.B. Dwell position verification	15
79	$IV.B.1.$ Optimal probe-to-probe spacing \hdots	15
80	$IV.B.2.$ Source tracking: two-probe versus all-probe detection strategies $\ .$.	16
81	IV.C. Dwell time verification	17
82	IV.D. Clinical use	18
83	V. Conclusion	19
84	References	20

85 I. Introduction

High dose rate brachytherapy (HDR-BT) is a standard modality in cancer treatment which
offers advantages of highly localized dose distributions and minimum number of treatment
fractions.^{1,2,3,4}. To ensure that the planned dose is properly delivered, time-resolved in vivo
dosimetry (IVD) has been proposed for monitoring treatments and detecting errors ^{5,6,7,8,9}.
Among time-resolved IVD approaches, optical fibers coupled to scintillators have shown
promise in time-resolved verification of the dose rate ^{10,11,12,13,14,15,16,17}, as well as dwell position and dwell time monitoring of a stepping radioactive source ^{7,15,18,19,20}.

IVD in multiprobe architectures has recently attracted attention for its ability to in-93 crease the spatial extent of treatment monitoring to volumes including the targeted tumor 94 and surrounding healthy tissues. By use of individual detectors in various parallel catheters, 95 Wang et al.²¹ and Guiral et al.²² performed extended source tracking along the source 96 catheter. Cartwright et al²³ implemented a source tracking with an array of 16 plastic 97 scintillator dosimeters embedded in a 20 mm-diameter rectal applicator. However, the ac-98 quisition rate of the detector was limited to 1 s, short dwell times of the source could not 99 be assessed. Moreover, the diameter of the resulting multiprobe dosimeter in the centimeter 100 range limits its field of application in BT. Therriault-Proulx et al.²⁴ developed a three-probe 101 plastic scintillator detector sufficiently narrow to be inserted within a BT needle or catheter, 102 thereby making multiprobe tracking applicable in a broader range of BT. Because they in-103 volve one-millimeter outer diameter fibers, the three scintillator probes were engineered on 104 the same fiber to be insertable into a BT needle or catheter. In this specific setup, where 105 the luminescence from all three scintillators travels through the same physical path within 106 the fiber, the optical signals are entangled at the fiber output. By analyzing the optical 107 signal with a spectrometer, dosimetry has been successfully demonstrated with a detection 108 rate as large as 3 seconds. A recent study has introduced a hyper-spectral optical detec-109 tion system enabling time-resolved dose rate monitoring as well as dwell position and dwell 110 time verification with such a fiber detector 19,25 . To determine the dose received by each 111 individual scintillator from the detected optical signals, a calibration process based on the 112 AAPM TG-43 dose parameters²⁶ has been implemented²⁵. The detected optical signals were 113 combined in a 4×4 linear equation system to obtain a detector overall response that is free 114 from the stem effect 25,27 . With this system, triangulation approaches for source tracking 115

¹¹⁶ were demonstrated.

In this paper, we use the miniaturized scintillator detector (MSD) approach 28,29,30 to 117 demonstrate a seven-channel multiprobe detector that is narrow enough to perform time-118 resolved treatment monitoring within a single BT needle or catheter. Our 320-micron outer 119 diameter fiber detector consists of 6 scintillating probes and a bare test-fiber, engineered 120 at the end of a narrow seven-fiber bundle. The parallel measurement of the seven optical 121 signals at the bundle output with a sCMOS camera avoids inter-probe cross-talk, i.e., signal 122 entanglement at the bundle output. As a result, the calibration of our multiprobe detector 123 does not require the use of the AAPM TG-43 algorithm. Moreover, Cerenkov signal is simply 124 removed by positioning a bandpass filter in front of the camera. Each MSD of the detection 125 system ensures minimum volume averaging within the steep dose gradients of BT sources, 126 leading to unmatched source tracking performances in space and time. 127

¹²⁸ II. Material and Methods

¹²⁹ II.A. Multiprobe system

The multiprobe detector (MPD) shown in Fig. 1(b) involves a 10-meter-long bundle of seven 130 biocompatible fibers arranged in a hexagonal lattice (cf. Fig. 1(a); fabricated by SEDI-ATI). 131 Each fiber is of 80-micron outer diameter (50-micron core diameter) and is covered with a 5-132 micron-thick polyimide protective coating. The total width of the bundle is of 270 microns. 133 Each fiber tip is tapered in the form of a leaky-wave nano-optical antenna^{28,31} aimed at 134 improving the transfer of the X-ray excited luminescence from the scintillators to the fiber. 135 Scintillating powder (Gd_2O_2S :Tb) is locally attached to the tapered tip of six of the seven 136 fibers to form the probes P1-P4, P6 and P7 (see Fig. 1(b)). Gd₂O₂S:Tb is chosen as the 137 scintillating material owing to its high scintillation efficiency, stability, linearity and fast 138 temporal response^{32,33,34} and with very low sensitivity to temperature (in the range of 15°-139 $(40^{\circ})^{35,36}$. This inorganic scintillator shows an energy dependence³⁰ that need to be corrected 140 if a direct dose rate measurement, rather than a source tracking, is targeted. The last bare 141 fiber, labelled as P5, is used to evaluate the level of spurious Cerenkov signal generated 142 within fibers upon irradiation. The overall fabrication process is detailed in Refs.^{29,30}. The 143 MPD is locally enlarged to diameters around 320 microns by the presence of the scintillating 144

cells (see Fig. 1(c)). The scintillation cells forming the six parallel probes are shown in 145 Fig. 1(d). The scintillation volume varies from 0.008 mm^3 (P1) to 0.009 mm^3 (P3), due 146 to slight imperfections in our fabrication process, in terms of reproducibility. Four different 147 inter-probe spacings along the bundle axis are defined by adapting the length of each optical 148 fiber of the bundle. P1 and P2, P2 and P3, and P4 and P6 are spaced by Δ , 2Δ , and 4Δ , 149 respectively, where $\Delta = 5$ mm, whereas P3 and P4 as well as P6 and P7 are both spaced 150 by 1.7 Δ . Interprobe spacings are defined with an accuracy of 0.3 mm. Measurements where 151 realized by positioning the MPD onto a calibrated motorized stage coupled to a binocular 152 equipped with a crosshair. The fiber bundle is positioned within a black 0.9-mm hytrel 153 cladding to minimize collection of the background light from the test room. This opaque 154 shield stops about 10 cm before the first probe P1 so that all six probes plus the bare fiber 155 are directly in contact to the water phantom. To avoid contribution from external spurious 156 light, the experiments are carried out in the dark. In addition, an opaque cover is used to 157 protect the water phantom from residual room light. 158

159 II.B. Optical readout

The optical signals at the end of the MPD are simultaneously recorded with a sCMOS camera 160 (Andor Technology, Zyla 4.2 model) whose maximum detection yield spectrally matches 161 the emission of the Gd_2O_2S :Tb material. A 35 mm camera objective (Fujinon HF35SA) 162 is positioned in front of the camera to image the bare output face of the fiber bundle at 163 a rate of 0.06 s. Prior to acquisitions, we define seven regions of interest (ROI) tightly 164 enclosing the seven light spots that are observable in the image (one spot per probe, see 165 the green circles in Fig. 1(e) delimiting the ROIs). Our code, developed under Labview 166 environment, automatically defines the ROIs as 16-pixel diameter disks centered with respect 167 to the maximum signal. The image pixels located within each ROI are integrated to obtain 168 seven detection signals sampled at 0.06 s. A chromatic filter (544/24 nm band pass filter 169 from Semrock) is positioned in front of the camera to filter out the spurious Cerenkov signal 170 (stem effect) generated in the fibers upon exposure.²⁹. 171



Figure 1: (a) Schematic of the bundle of 7 fibers arranged in an hexagonal lattice. (b) Photograph of the multiprobe detector. Green laser light is coupled to the free bare facet of the fiber bundle to identify in the image the six scintillation cells (six green spots are observed due to light scattering of the fiber modes by the scintillators). (c) Schematic of a cross-section of the MPD at the location of probe P4 (cf. (b)). The green and gray disks represent the cross-sections of the scintillating cell and fibers, respectively. (d) Magnified optical images of the six scintillation cells at the end of the fiber bundle. (e) Image of the bare face of the fiber bundle by the sCMOS camera when white light is projected onto the seven probes. (f) Schematics of the experimental setup involving a water tank, the multiprobe detector positioned onto a 2D motorized stage and a photometer based on a sCMOS camera coupled to an objective (Obj.) and a band-pass filter (BP). (g) Photograph of the experimental setup.

¹⁷² II.C. Brachytherapy system

A MicroSelectron afterloader with a 9.14 Ci Ir-192 HDR source (Air kerma strength of 37309
U) is used for irradiation.

175 II.D. Phantom

The probe characterization is conducted in a $40 \times 30 \times 30$ cm³ water tank. The source catheter 176 crosses the tank widthwise (Figs. 1(f) and (g)). During experiments, temperature in the 177 water phantom varies from 17° to 19° (temperature assessment with a thermometer before 178 and after the experiments). The MPD is fixed to a solid-water holder that is attached to a 179 2D translation stage via a plastic adaptor (Figs. 1(f) and (g)). The fiber probe is set parallel 180 to the source catheter. A coordinate frame of the set-up is defined so that the origin of the 181 frame coincides with the scintillators of the proximal probe P1. The source-probe spacing 182 along the (0x) and (0z) axes is determined with the motorized stage and the afterloader, 183 respectively. 184

¹⁸⁵ II.E. Detector calibration

The MPD is calibrated along seven lines parallel to the (0z)-axis. These lines are spaced 186 by 5.5, 8, 10, 15, 20, 30 and 40 mm (along (0x)) from the axis of the source catheter. 187 First, the MPD is positioned at the desired source-probe inter-catheter spacing x with the 188 motorized stage. Then, the source is displaced along the fixed source catheter by 2.5-mm 189 steps. The calibration curves are obtained by integrating 165 images per source position. 190 An interpolation (performed with the "interp" function of Matlab software) is applied to 191 the measured profiles to obtain a 0.1-mm sampling rate. During calibration, we verify with 192 the scintillator-free fiber probe P5 that no optical signal (stem effect) is detected with the 193 chromatic filter positioned in front of the camera. 194

The signal-to-noise ratio (SNR) of the six scintillating probes forming the detector is assessed at the dwell positions corresponding to the maxima of all the above-mentioned calibration curves, at the seven source-probe spacings x ranging from 5.5 mm to 40 mm. The SNR is the average amplitude of the signal divided by its standard deviation.

5

¹⁹⁹ II.F. Dwell position and dwell time verification

200 II.F.1. Measurements

To test the MPD, an irradiation protocol consisting of 40 dwell positions is applied for 201 each value of source-probe spacing along (0x). The dwell positions are spaced by 2.5 mm 202 and the dwell time is fixed to 10 s. Note that 2.5-mm is the minimum displacement step 203 allowed by our after loader. With a source tracking accuracy of $0.023 \pm 0.077 \text{ mm}^{30}$, our sub-204 millimeter scintillating probes are suitable to accurately track shorter sources steps (e.g., 1 205 mm steps). The dwell time value has been fixed to 10s for a direct comparison of our source-206 tracking results with those of Linares et al.¹⁹. Note that our scintillating probes have already 207 demonstrated their effectiveness in monitoring dwell times ranging from 0.2s to 11s.³⁰. 208

²⁰⁹ II.F.2. Source position monitoring

The instant position of the source at each acquisition time is retrospectively determined from the output signals of the MPD and the source activity using the calibration curves presented in section II.E.. To ensure that the transitory phases between two successive dwell positions are not taken into account in the source tracking, the first and the last signal points for each dwell position are ignored. These transitory phases, which last a few tens of milliseconds³⁷, correspond to the displacement of the radioactive source between two dwell positions.

During a treatment delivery, each probe j (1 < j < 7) of the MPD delivers a temporal signal $S_j(t)$. At each instant $t = k\tau$, where $k \in \mathbb{N}$ and τ is the acquisition time of the camera, the instant source position is deduced from the readout signal S_j as follows.

First, function f_j^k is defined for each probe j as:

220

$$f_j^k(z) = \left| C_j(z) - S_j^k \right|,\tag{1}$$

where $C_j(z)$ and S_j^k are the calibration curve and the readout signal of the j^{th} probe at the k^{th} time step, respectively. z corresponds to the spatial coordinate along the axis of the source catheter. Calibration curves being symmetric regarding the z-coordinate, each probe provides two likely instant source positions located on both sides of function f_j^k . Therefore, at minimum two probes are necessary to unambiguously determine the position of a stepping 226 BT-source.

235

2

The instant source position Z^k is determined using four different methods involving 227 various manipulations of functions f_i^k . To find the inter-probe spacing which optimizes source 228 tracking accuracy (for detector design purpose), source position verification is realized from 229 a "two-probe" dosimetry using Eq. 2 (m = 1). $j_1 \in [1, 6] \setminus 5$ and $j_2 \in [2, 7] \setminus 5$ are the indices 230 of the probes forming the 15 probe pairs (j_1, j_2) allowed by our detector. The inter-probe 231 spacing ranges from 5 mm to 52 mm. Probe P5, which is bare to assess in-fiber Cerenkov 232 effect, is not involved in the source position monitoring. Source tracking is systematically 233 analyzed from each of the 15 probe pairs of the detector. 234

$$Z_m^k(z) = \min\left[f_{j_1}^k(z) + f_{j_2}^k(z)\right],$$
(2)

We also tested and compared three different source tracking algorithms. In each case, 236 all the six scintillating probes are involved in the source position monitoring during the 237 treatment delivery. The two first values of the instant source position $(Z_m^k, m = 2 \text{ and } 3)$ 238 are calculated from the readout signals of probe pairs dynamically chosen among the six 239 available probes of the MSD. In both of these two-probe measurements, the source position 240 is defined from Eq. 2 with indices j_1 and j_2 which vary with the source position along (0z). 241 Probe pairs are dynamically chosen to provide the higher readout signals (m=2) or on the 242 basis of a maximum gradient of their calibration curves at the source position (m=3). The 243 last z-coordinate of the source Z_4^k is determined by adding functions f_j^k of all seven probes. 244 We have: 245

46
$$Z_4^k(z) = \min\left[\sum_{i=1}^7 f_j^k(z)\right],$$
 (3)

²⁴⁷ II.F.3. Dwell time verification

The monitoring of an HDR-BT treatment is known to produce a staircase temporal signal^{14,15,29}. The dwell times of the stepping source, which correspond to the duration of the plateaus in between two successive signal edges, can be simply determined from an edge detection within all readout signals of our MPD system. Our edge detection approach involves function F^k defined as:

$$F^{k} = \sum_{j=1}^{6} \left(S_{j}^{k+1} - S_{j}^{k} \right), \tag{4}$$

By adding the signal derivatives from all probes, our algorithm is expected to reduce undesired fluctuations in the edge detection function (due to readout noise), as compared to that of a single probe detector.

²⁵⁷ III. Results

²⁵⁸ III.A. Detector specification and calibration

The specification of the scintillating probes forming the MPD can be found in Ref.³⁰. The 259 linearity coefficient of the probes exceeds 0.999 regardless of the source-probe spacings x. 260 The deviation to repeatability remains below 2% for all values of x. The energy dependence 261 of the fiber probes forming the MPD, referred to as the single-probe MSD, has already 262 been characterized in a past study³⁰. Figure 2(b) shows the calibration curves of the MPD 263 acquired at source-probe spacings x of 10, 20 and 30 mm. Six gaussian-like profiles are shown 264 per source-probe inter-catheter spacing (one profile per probe), whose maxima coincide with 265 the probe positions along (0z), as depicted in Fig. 2(a). The SNR of the probes forming the 266 detector varies from 110-140 down to 20-25 at source-probe spacings x of 5.5 mm and 40 267 mm, respectively. 268

²⁶⁹ III.B. Source position monitoring

²⁷⁰ III.B.1. Optimal probe-to-probe spacing

The aperiodic scintillator array of our MPD enables 15 probe pairs whose inter-probe spacings vary from 5 mm to 52 mm. To identify optimal inter-probe spacings for the future MPD designs, we analyzed the accuracy of the source position verification over these 15 probe pairs, versus the inter-probe distance δz along the detector axis (0z) (see Fig. 3). The instant source position is determined from Z_1^k function, cf. Eq. 2 (m = 1). The displacement range of the BT source along (0z) varies with the inter-probe distance δz as $\delta z + 2 * 0.75$ cm (see inset of Fig. 3(b)).



Figure 2: (a) Schematic of the experimental set-up in the water tank. The coordinate frame defining the axis convention is shown in the top-left corner. The source and MPD are represented in red and green, respectively. All seven positions of the MPD defining source-probe spacings x ranging from 5.5 to 40 mm are represented with dashed lines. The source and MPD move along the (0z) and (0x) axes, respectively. (b) Calibration curves of the MPD used for source tracking at x equal to 10, 20 and 30 mm.



Figure 3: Optimal inter-probe spacing for source tracking. (a) Offset between the measured and planned dwell positions along (0z) (\bar{z}_{exp} and z_{TPS} , respectively) as a function of the inter-probe distance δz . The measured dwell position \bar{z}_{exp} corresponds to the average of the instant source position Z_1^k (see Eq. 2) over a dwell time. Each error bar shows for one source-probe inter-catheter spacing x (see inset of (a)) the offset analysis to the planned dwell positions (mean and standard deviation). This analysis is performed over z-coordinates spanning over $\delta z + 2 * 0.75$ cm (see inset of (b)). (b) SD of the experimentally determined source position Z_1^k versus the inter-probe distance δz . Each error bar shows the distribution (mean and standard deviation) of the SD for dwell positions within $\delta z + 2 * 0.75$ cm (see inset of (b)), at a given source-probe inter-catheter spacing x (see inset of (a)).

We see from Fig. 3(a) that the minimum deviation to the planned dwell positions 278 occurs at δz values in-between 15 mm and 35 mm, regardless of the source-probe inter-279 catheter spacing x. In that δz range, the offset distribution to the planned dwell positions 280 does not exceed 0.05 ± 0.15 mm at x=30 mm $(0.15 \pm 0.41$ mm at x=40 mm). The SD of the 281 measured instant source position Z_1^k reaches minimum values at δz in between 0.87x and x282 (see Fig. 3(b)). This property, which is observed for all values of x ranging from 5.5 mm to 283 40 mm, is imputed to the broadening of the calibration curves along (0z) as the source-probe 284 inter-catheter spacing x increases (cf. Fig 2). The tighter distributions of SD are of 0.2 \pm 285 $0.022 \text{ mm}, 0.82 \pm 0.12 \text{ mm}$ and $1.78 \pm 0.12 \text{ mm}$ at x = 10, 20 and 30 mm, respectively. 286

²⁸⁷ III.B.2. Source tracking: two-probe versus all-probe detection strategies

A six-probe detector enables numerous detection strategies for source tracking. In Fig. 4, we compare three algorithms which involve dosimetry either from probe pairs dynamically chosen during the treatment delivery (cf. Z_2^k and Z_3^k of Eq. 2) or from all the probes of the detector (cf. Z_4^k of Eq 3).

At source-probe spacings x below 15 mm, all three source-tracking algorithms show 292 similar accuracy. The mean and standard deviation of the offset to the planned dwell po-293 sitions do not vary by more than 0.008 and 0.014 mm, respectively, from one method to 294 another (Fig. 4(a)). With the two higher signal method (cf. Z_2^k of Eq. 2), the deviation 295 to the planned dwell positions, which is of 0.007 ± 0.138 mm at x=20 mm, increases up to 296 0.20 ± 1.12 mm at x = 40 mm. As a comparison, the dwell position verification from the 297 two signals of steeper gradients (calculation of Z_3^k) leads to a mismatch to the treatment 298 plan of 0.027 ± 0.115 mm and -0.11 ± 0.68 mm at x equal to 20 and 40 mm, respectively. 299 Source tracking from all detected signals (calculation of Z_4^k) is much less impacted by the 300 enhancement of the source-probe inter-catheter spacing x. The offset to the planned dwell 301 positions is of 0.029 ± 0.078 mm at x=20 mm and 0.02 ± 0.19 mm at x=40 mm, respectively. 302

The SD of the instant source position determined from the three above-mentioned methods is reported in Fig. 4(b). For source-probe distances below 15 mm, all three methods determine the instant source positions with almost the same accuracy. When x exceeds 20 mm, the dwell position verification from the two higher detected intensities (calculation of Z_2^k) is the less accurate. A detection from all probes (cf. Z_4^k) minimizes signal fluctuations.

Fig. 5 displays a detailed representation of the source position determined from all 308 scintillating probes, which correspond to the analysis shown in Fig. 4 (cf. black error bars). 309 Fig. 5(a) reports the error to the planned dwell positions, which corresponds to the difference 310 between the planned and measured dwell positions (z_{TPS} and \bar{z}_{exp} , respectively). Fig. 5(b) 311 reports the SD of the instant source position Z_4^k , i.e., the SD of the distribution of source 312 positions measured at a rate of 0.06s during a dwell time. Noticeable enhancement of the 313 SD is observed when the source is positioned out of the region where the probes are located, 314 i.e., in between the detector and probe P_1 ($\bar{z}_{exp} < 0$) or beyond probe P_7 ($\bar{z}_{exp} > 5.2$ cm 315 cf. Fig. 1(b) and Fig. 2). This local fluctuation enhancement is maximum when x=5.5316



Figure 4: Source tracking: two-probe versus all-probe algorithms. (a) Offset between the measured and planned dwell positions along (0z) (\bar{z}_{exp} and z_{TPS} , respectively) as a function of the source-probe inter-catheter spacing x. The measured dwell position \bar{z}_{exp} corresponds to the average of the instant source position over a dwell time. The instant source position corresponds to Z_2^k , Z_3^k or Z_4^k (cf. Eqs. 3 and 2; see inset of (b)). Each error bar shows for one algorithm (see inset of (b)) the analysis of the offset to the planned dwell positions (mean and SD). This analysis is performed over a range of source positions along (0z) of 10 cm. (b) SD of the experimentally determined instant source positions Z_2^k , Z_3^k and Z_4^k . Each error bar shows for one algorithm (see inset of (b)) the analysis (mean and SD) of the distribution of SD for dwell positions spanning over 10 cm (along (0z)).



Figure 5: Analysis of an "all-probe" source tracking. (a) Offset between the planned and experimentally determined dwell positions along (0z) (z_{TPS} and \bar{z}_{exp} , respectively). \bar{z}_{exp} is the average of the measured instant source position Z_4^k over a dwell time. The mismatch to the planned dwell positions is shown for seven values of the source-probe inter-catheter spacing x (see legend of (b)). (b) SD of the measured instant source position Z_4^k along (0z) as a function of the z-coordinate. Here again, seven source-probe spacings x are considered (see legend). Insets of (a) and (b), schematic of the multiprobe detector which identifies the z-coordinates of the scintillators on the graphs (with dashed lines).

mm and lessens as the source-probe spacing x increases, to finally vanish at x=20 mm. On the contrary, the offset to the planned dwell positions remains at the same level over the entire displacement range of the stepping source (i.e., 10 cm), regardless of the source-probe spacing x (cf. Fig 5(a)).

321 III.C. Dwell time verification

In Fig. 6, we report the analysis of the offset ΔT between the measured and planned dwell times (T_{exp} and T_{TPS} , respectively) as a function of the source probe inter-catheter spacing x. The error bars show the mean and SD of the offset to the planned dwell times (calculated over 40 dwell positions). The average of ΔT is estimated to be -0.006 \pm 0.009 s.



Figure 6: Mismatch ΔT between the measured and planned dwell times (T_{exp} and T_{TPS} , respectively) as a function of the source-probe spacing x.

326

IV. Discussion

³²⁷ IV.A. Detector specification and calibration

Our MPD shows an acceptable SNR for an accurate dwell time and position verification 328 (larger than 20). At a source probe spacing x of 20 mm, Linares et al. reported a SNR 329 ranging from 13 to 23 with their multipoint detector system involving three photomultiplier 330 tubes (PMT) as photometers²⁵. At the same source-probe spacing, we measure a SNR 331 spanning from 33 to 39 with a sCMOS camera, a detection rate of 0.06 s and sensitive 332 volumes that are 250 to 650 times smaller than those of Linares's multipoint detector²⁵. 333 SNR could even be enhanced either by slightly broadening the scintillation cell and the fiber 334 core (cf. Ref.³⁰) or by replacing the sCMOS camera by six PMTs which provide higher light 335 detection sensitivity and an acceptable detection speed. However, six (or seven) PMTs may 336 be more expensive than a sCMOS camera. Moreover, the performance of these ultrasensitive 337 photometers can be seriously reduced after misuse (such as, e.g., exposing the PMT to room 338 light). Note that the range of SNR observed across the probe array for each source-probe 339 spacing x is due to small discrepancies in scintillation volume and in-fiber luminescence 340 coupling efficiency of the probes. We did not observe noticeable signal variation of the 341 MPD with the slight temperature drift in the water phantom during our experiments, thus 342 confirming the very low sensitivity of the Gd_2O_2S :Tb to changes in temperature^{35,36}. 343

344

Beyond the opportunity of inserting a seven-probe detector within a brachytherapy

needle, the advantage of the miniaturization of a water nonequivalent fiber dosimeter resides
in a smaller volume averaging effect in the strong dose gradients of the radioactive source,
as well as a minimum electron fluence perturbation.

Miniaturization does however not resolve the energy dependence of inorganic scintil-348 lator detectors. In contrast to the multipoint detector approach based on organic materi-349 als^{19,24,25,38}, our water non-equivalent multiprobe system requires a space-dependent correc-350 tion factor for a direct accurate determination of the dose rate at a measured dwell position³⁰. 351 To overcome this issue, our strategy is to define a dose rate value from the dwell position and 352 source activity by use of the AAPM TG-43 dose parameters²⁶. This indirect dosimetry can 353 be seen to bring the required correction factor 30 . Note that the space-dependent calibration 354 of the organic multipoint detector also relies on the AAPM TG-43 formalism²⁵. 355

³⁵⁶ IV.B. Dwell position verification

357 IV.B.1. Optimal probe-to-probe spacing

At an interprobe spacing δz of 36.8 mm and at a source-probe spacing x=5.5 mm, our MPD 358 used as a two-probe detector ensures a deviation to the planned dwell positions of 0.03 \pm 359 0.15 mm. We followed the measurement process depicted in Fig. 3(b), with source positions 360 expanding over 51.8 mm along (0z) (i.e., 36.8 + 2 * 7.5 mm). As a comparison, the deviation 361 to the planned dwell positions is estimated to be 0.45 ± 0.3 mm (1SD) with the three-362 probe fiber detector of Linares et al¹⁹, at a source-probe spacing x = 5 mm and a source 363 activity of 10.73 Ci. Linares's detection system consists of three 10-mm spaced scintillation 364 cells integrated at the end of an individual fiber detection line. Two reasons may explain 365 the higher source-tracking accuracy of our two probe device, despite detection efficiencies 366 of equivalent SNR. First, the sensitive volume of our probes is two orders of magnitude 367 smaller, thereby limiting the averaging effect in the steep dose gradients near an HDR-BT 368 source. Second, the source tracking accuracy of Linares's system may be constrained due to 369 the utilization of non-exclusively experimental calibration, which incorporates the AAPM 370 TG-43 formalism 25 . 371

With a 21-mm spaced four-probe detection system, Guiral et al demonstrated a mismatch to the planned dwell positions of 0.11 ± 0.7 mm at a 0.1 s detection rate and along a $_{374}$ 60 mm portion of source catheter. At a similar source probe spacing estimated to be of 20 mm, we find a deviation to the planned dwell positions of 0.03 ± 0.14 mm over dwell positions expanding over 50 mm. Our sensitive volume being 45 times smaller than Guiral et al.'s²², volume averaging effect is reduced, thereby improving dwell position verification.

³⁷⁸ IV.B.2. Source tracking: two-probe versus all-probe detection strategies

Cascading scintillating probes along the source catheter allows for extending optimum source 379 tracking capabilities over longer source paths. The source tracking accuracy reported above 380 can be improved when the probe pair is dynamically chosen among the six available probes of 381 the detector to follow the source during the treatment delivery (all six probes are sequentially 382 involved in the treatment monitoring). The detection accuracy for the "two best gradients" 383 surpass that of the "two best intensities" algorithm. At a 10 s dwell time, source tracking 384 accuracy from these two algorithms is only slightly better than for the fixed two-probe 385 system but the gap should be noticeably enhanced for shorter dwell times, especially the 386 sub-second regime. The two investigated algorithms could also show improved performances 387 if a constant interprobe spacing was considered. 388

Source tracking accuracy is noticeably enhanced when the source position monitoring 389 is done via the accumulation of all the six readout signals of the detection system. In 390 that case, the deviation to the planned dwell positions is reduced to -0.017 ± 0.063 mm and 391 0.029 ± 0.078 mm at the source-probe spacings x of 5.5 and 20 mm, respectively. At x = 5.5392 mm, we observe a maximum offset to the treatment plan of 0.12 mm. As a comparison, 393 Linares et al reported a maximum deviation of 1.8 mm at x = 5 mm and at similar source 394 displacement range (10 cm) and dwell time (10s)¹⁹. At x = 20 mm, Guiral et al found a 395 discrepancy to the treatment plan of 0.11 ± 0.7 mm with their four-probe detection system 396 (at a dwell time of 5s)²². The accuracy of the MPD in dwell position verification is fully 397 compatible with the requirements of HDR brachytherapy in terms of medical treatment and 398 quality assurance 1,7,39,40 . 399

Over the entire displacement range of the stepping source (i.e., 10 cm) and at a dwell time of 10 s, the "all-probe" detection approach provides a dwell position verification that is not affected by the fluctuations of the instant source position \bar{z}_{exp} , whatever the source-probe spacing x. Although the SD of \bar{z}_{exp} varies by one order of magnitude during the treatment at x = 5.5 mm (Fig. 5(b)), the dwell position measurement remains at the same accuracy level (Fig. 5(a)). The offset to the planned dwell position shows a narrow distribution of 0.002 $\pm 0.049 \text{ mm}$ and a maximum value which does not exceed 0.12 mm. Such performances exceed those of competing multipoint scintillator detectors¹⁹. Note that all these results are obtained at a noticeably long dwell time. Measurements within a range of shorter dwell times will be studied in the future. A measurement accuracy of 0.023 \pm 0.077 mm has already been shown with a single MSD for dwell times ranging from 0.1 to 11 s³⁰.

⁴¹¹ IV.C. Dwell time verification

Across the source-probe spacings x ranging from 5.5 to 20 mm, the total average of the 412 differences ΔT between the measured and planned dwell times is of -0.006 ± 0.009 s, at a 413 detection rate of 0.06 s. At x = 20 mm, ΔT is found to be -0.008 ± 0.058 s. With their 414 four-probe detector, Guiral et al measured a difference ΔT of 0.05 ± 0.9 s at approximately 415 similar source-probe spacing, a detection rate of 0.1 s and for 5-s dwell times²². In their 416 study, the source positions expanded over 6 cm within the source catheter, rather than 10 cm 417 as in our case. At x = 5.5 mm, we find an offset ΔT of 0.0009 ± 0.0497 s. As a comparison, 418 Linares et al reported an offset to the planned dwell time of 0.33 ± 0.37 s at x = 5 mm and 419 a dwell times of 1s. 420

It is noteworthy that the measurement errors of the dwell time discussed here are all 421 obtained from an edge detection in a staircase detection signal. In all the proposed methods, 422 dwell times are defined as the elapsed time in between two successive signal edges, which 423 are identified from a signal derivative calculation. Since all the considered dwell times in the 424 above-cited studies are at least 50 fold longer than the integration time of the photometers 425 used, one can assert that the measurement accuracy will not significantly change as the dwell 426 time increases. We recently verified this property in a single probe detection³⁰. Therefore, 427 the results from Guiral's and Linares's multiprobe detectors obtained with 5 s and 1 s dwell 428 times, respectively, can be directly compared to our results measured at a dwell time of 10 429 s. As an example, the lower measurement accuracy of Linares's approach may be partly 430 explained by their shorter 1 mm inter-dwell spacing, leading to noticeably smaller temporal 431 edges (given the strong dose gradients involved), rather than the use of a shorter dwell time 432 of 1s. 433

Finally, note that the individual component of the MPD, referred to as the single-probe MSD, has already demonstrated its capability in monitoring prostate treatment sequences encompassing dwell times ranging from 0.2 to 11 seconds.³⁰. With our probe approach for a MPD, 94% of the 966 dwell positions were successfully identified, with an average deviation to the planned dwell times of 0.005 ± 0.060 and a 100 % detection rate for dwell times exceeding 0.5 s (17%, 86%, 91% and 95% of the dwell times of 0.2 s, 0.3 s, 0.4 s and 0.5 s were successfully identified, respectively).

⁴⁴¹ IV.D. Clinical use

⁴⁴² Our multiprobe detector is compatible with clinical applications. It indeed consists of bio-⁴⁴³ compatible elements and it is sufficiently narrow to be inserted in a BT needle or in a ⁴⁴⁴ catheter. As a preliminary step, we successfully positioned our detector in a one-millimeter ⁴⁴⁵ wide sealed encapsulation pipe made of PEEK material.

In vivo applications forbid the use of our motorized stage for probe positioning. In 446 gynecologic BT, the probe and the source would be inserted in two parallel catheters of 447 an applicator. The inter-catheter spacing would be precisely known, as in the case of the 448 present study. In prostate BT, the probe and the source are inserted in two independent 449 needles which are implanted manually in a patient. Therefore, x and z coordinates (cf. 450 Fig. 2(a) are usually coupled since the needles are rarely implanted perfectly parallel from 451 each other, due to operational uncertainties. x and z coordinates of the source relatively to 452 the probe could however be simultaneously determined by various triangulation approaches 453 that would be rendered possible by our six-probe detection system (see for instance Ref.¹⁹). 454 Source tracking via a triangulation process requires a refined 2D calibration plot of the 455 system, which would not represent a challenge here³⁰. Moreover, parallel IVD from a pair of 456 multi-probes detectors connected to the same camera and inserted in two different needles 457 would allow a 3D positioning of the source by triangulation³⁸ with minimum equipment. 458

⁴⁵⁹ Note that further experiments are required to assess the compatibility of our probe with ⁴⁶⁰ various clinical scenarios. Firstly, it is necessary to repeat measurements using sources of ⁴⁶¹ lower activities, specifically those nearing the end of their clinical lifespan (around 3 or 4 ⁴⁶² months). This will help evaluate the probe's performance under non-optimum conditions ⁴⁶³ leading to lower SNR. Secondly, it is important to test our fiber detector while it is positioned

V. CONCLUSION

within a plastic catheter or either a plastic or metallic needle, depending on the specific 464 treatment being targeted. This evaluation will provide insights into the probe's functionality 465 and reliability when used in conjunction with different types of delivery devices potentially 466 attenuating or spectrally modifying radiations at probe locations. Lastly, it is recommended 467 to consider treatment sequences that involve dwell times of fractions of seconds, various 468 source steps (reduced to 1 mm), and source-probe spacings larger than 4 cm. By examining 469 these parameters, we could gain a comprehensive understanding of the probe's performance 470 across a wider range of clinical scenarios. 471

As mentioned earlier, the individual component of the MPD, specifically the single probe 472 fiber detector, has already been successfully demonstrated in accurately monitoring dwell 473 times down to 0.2 seconds, with source-probe distances of up to 4.7 cm^{30} . In cases where 474 the source-probe distance exceeds 4 cm, an alternative multiprobe architecture is envisioned. 475 This architecture would involve multiple MPDs, each incorporating 2 to 4 probes equally 476 spaced apart. These MPDs, inserted within different catheters or needles and engineered 477 from the same fiber bundle, would be positioned parallel to each other, and separated by 478 approximately 3 cm within the treatment volume. This proposed configuration would provide 479 the capability to accurately monitor the source in three dimensions (3D) throughout the 480 larger treatment volumes typically encountered in brachytherapy. 481

482 V. Conclusion

We have demonstrated in a water phantom a monitoring device for HDR-BT based on a 483 six-probe scintillator dosimeter coupled to a sCMOS camera. Being engineered at the end of 484 a narrow 270- μ m diameter fiber bundle, our miniaturized probes combine high spatial reso-485 lution and high detection speed while ensuring a minimum perturbation of the therapeutic 486 process, even if water nonequivalent (inorganic) materials are used. Moreover, the overall 487 dosimeter is totally free from inter-probe cross-talk. The use of a sCMOS camera, rather 488 than seven photomultiplier tubes of higher sensitivity offers the possibility of a simultaneous 489 parallel readout of the six scintillating probe signals, plus the residual Cerenkov signal after 490 chromatic filtering (from the bare fiber), in a simple and low cost architecture that is suitable 491 for clinical use. The lower SNR of the camera is compensated by a higher probe detection 492 efficiency enabled by our concept of an IVD micro-pixel based on a nano-optical interface in 493

⁴⁹⁴ between scintillators and a fiber 28 .

First, we found a range of probe-to-probe spacings which minimizes source tracking 495 uncertainties. This will be an important information for future MPD designs. We then 496 studied and compared three different source tracking algorithms from the large array of 497 options available within our six-probe system. The best detection approach was found by 498 adding the parallel readout signals from all the probes of the detector. Realizing a source 499 tracking based on this overall accumulated readout signal led to an offset to the planned 500 dwell position as small as 0.01 ± 0.14 mm and 0.02 ± 0.29 mm over a 10-cm long source 501 displacement in the source catheter and at spacings between source and probe catheters of 502 5.5 and 40 mm, respectively. Using this method, we also measured deviations to the planned 503 dwell time of -0.006 ± 0.009 s and -0.008 ± 0.058 s, at source-probe spacings x of 5.5 mm and 504 20 mm, respectively (detection rate of 0.6 s). All the studied configurations were found to 505 surpass current fiber-integrated multiprobes and multipoints detection systems. The next 506 steps will be to test our detection system with various treatment plans used for instance in 507 prostate brachytherapy. The detection performances demonstrated here need to be assessed 508 at shorter dwell times down to a fraction of a second. Triangulation approach will also be 509 realized to simultaneously define the dwell time and the 2D coordinates x and z of a stepping 510 HDR-BT source. 511

512 Acknowledgments

The authors thank Lionel Pazart, Thomas Lihoreau and Karine Charriere for helpful discussions. This study is funded by the SAYENS Agency, the French Agency of Research (contract ANR-18-CE42-0016), the Region "Bourgogne Franche-Comte" and the EIPHI Graduate School (contract ANR-17-EURE-0002). This work is supported by the French RENATECH network and its FEMTO-ST technological facility.

519 .

518 **References**

¹ H. D. Kubo, G. P. Glasgow, T. D. Pethel, B. R. Thomadsen, and J. F. Williamson, High
 dose-rate brachytherapy treatment delivery: report of the AAPM Radiation Therapy
 Committee Task Group No. 59, Med. Phys. 25, 375–403 (1998).

- J. Crook, M. Marbán, and D. Batchelar, HDR prostate brachytherapy, in *Semin. Radiat.* Oncol., volume 30, pages 49–60, Elsevier, 2020.
- ⁵²⁵ ³ C. Shah et al., The American Brachytherapy society consensus statement for skin ⁵²⁶ brachytherapy, Brachytherapy **19**, 415–426 (2020).
- A. N. Viswanathan, S. Beriwal, F. Jennifer, D. J. Demanes, D. Gaffney, J. Hansen,
 E. Jones, C. Kirisits, B. Thomadsen, and B. Erickson, American Brachytherapy Society
 consensus guidelines for locally advanced carcinoma of the cervix. Part II: high-dose-rate
 brachytherapy, Brachytherapy 11, 47–52 (2012).
- ⁵ L. A. DeWerd, G. S. Ibbott, A. S. Meigooni, M. G. Mitch, M. J. Rivard, K. E. Stump,
 ⁵³² B. R. Thomadsen, and J. L. Venselaar, A dosimetric uncertainty analysis for photon ⁵³³ emitting brachytherapy sources: Report of AAPM Task Group No. 138 and GEC ⁵³⁴ ESTRO, Med. Phys. 38, 782–801 (2011).
- ⁵³⁵ ⁶ J. Valentin et al., Prevention of high-dose-rate brachytherapy accidents. ICRP Publica-⁵³⁶ tion 97., Annals of the ICRP **35**, 1–51 (2005).
- ⁷ G. P. Fonseca, J. G. Johansen, R. L. Smith, L. Beaulieu, S. Beddar, G. Kertzscher,
 ⁵³⁸ F. Verhaegen, and K. Tanderup, In vivo dosimetry in brachytherapy: requirements and
 ⁵³⁹ future directions for research, development, and clinical practice, Phys. Imag. Radiat.
 ⁵⁴⁰ Oncol. 16, 1–11 (2020).
- ⁸ F. Verhaegen et al., Future directions of in vivo dosimetry for external beam radiotherapy
 ⁵⁴¹ and brachytherapy, Physics and imaging in radiation oncology 16, 18–19 (2020).
- ⁵⁴³ ⁹ K. Tanderup, S. Beddar, C. E. Andersen, G. Kertzscher, and J. E. Cygler, In vivo dosimetry in brachytherapy, Med. Phys. 40, 070902 (2013).
- J. Lambert, D. McKenzie, S. Law, J. Elsey, and N. Suchowerska, A plastic scintillation
 dosimeter for high dose rate brachytherapy, Phys. Med. Biol. 51, 5505 (2006).
- J. Lambert, T. Nakano, S. Law, J. Elsey, D. R. McKenzie, and N. Suchowerska, In vivo dosimeters for HDR brachytherapy: a comparison of a diamond detector, MOSFET,
 TLD, and scintillation detector, Med. Phys 34, 1759–1765 (2007).

- F. Therriault-Proulx, T. M. Briere, F. Mourtada, S. Aubin, S. Beddar, and L. Beaulieu,
 A phantom study of an in vivo dosimetry system using plastic scintillation detectors for
 real-time verification of 192Ir HDR brachytherapy, Med. Phys. 38, 2542–2551 (2011).
- ¹³ C. E. Andersen, S. K. Nielsen, S. Greilich, J. Helt-Hansen, J. C. Lindegaard, and
 ⁵⁵⁴ K. Tanderup, Characterization of a fiber-coupled luminescence dosimetry system for
 ⁵⁵⁵ online in vivo dose verification during brachytherapy, Med. Phys. **36**, 708–718 (2009).
- ¹⁴ M. D. Belley, O. Craciunescu, Z. Chang, B. W. Langloss, I. N. Stanton, T. T. Yoshizumi,
 M. J. Therien, and J. P. Chino, Real-time dose-rate monitoring with gynecologic
 ⁵⁵⁸ brachytherapy: Results of an initial clinical trial, Brachytherapy 17, 1023–1029 (2018).

J. Johansen, G. Kertzscher, E. Jørgensen, S. Rylander, L. Bentzen, S. Hokland, C. S.
 Søndergaard, A. With, S. Buus, and K. Tanderup, Dwell time verification in brachyther apy based on time resolved in vivo dosimetry, Phys. Med. 60, 156–161 (2019).

- ¹⁶ G. Kertzscher, C. E. Andersen, F.-A. Siebert, S. K. Nielsen, J. C. Lindegaard, and
 ⁵⁶³ K. Tanderup, Identifying afterloading PDR and HDR brachytherapy errors using real ⁵⁶⁴ time fiber-coupled Al2O3: C dosimetry and a novel statistical error decision criterion,
 ⁵⁶⁵ Radiother. Oncol. 100, 456–462 (2011).
- E. B. Jørgensen, J. G. Johansen, J. Overgaard, D. Piché-Meunier, D. Tho, H. M. Rosales,
 K. Tanderup, L. Beaulieu, G. Kertzscher, and S. Beddar, A high-Z inorganic scintillator–
 based detector for time-resolved in vivo dosimetry during brachytherapy, Med. Phys.
 48, 7382–7398 (2021).
- J. G. Johansen, S. Rylander, S. Buus, L. Bentzen, S. B. Hokland, C. S. Søndergaard,
 A. K. M. With, G. Kertzscher, and K. Tanderup, Time-resolved in vivo dosimetry for
 source tracking in brachytherapy, Brachytherapy 17, 122–132 (2018).
- ¹⁹ H. M. Linares Rosales, L. Archambault, S. Beddar, and L. Beaulieu, Dosimetric performance of a multipoint plastic scintillator dosimeter as a tool for real-time source tracking
 ⁵⁷⁵ in high dose rate Ir brachytherapy, Med. Phys. 47, 4477–4490 (2020).
- ⁵⁷⁶ ²⁰ E. B. Jørgensen, G. Kertzscher, S. Buus, L. Bentzen, S. B. Hokland, S. Rylander,
 ⁵⁷⁷ K. Tanderup, and J. G. Johansen, Accuracy of an in vivo dosimetry-based source

- tracking method for afterloading brachytherapy—A phantom study, Med. Phys. 48,
 2614–2623 (2021).
- ²¹ R. Wang, J. Ribouton, P. Pittet, P. Guiral, P. Jalade, and G.-N. Lu, Implementation of
 ⁵⁸¹ GaN based real-time source position monitoring in HDR brachytherapy, Radiat. Meas.
 ⁵⁸² **71**, 293–296 (2014).
- P. Guiral, J. Ribouton, P. Jalade, R. Wang, J.-M. Galvan, G.-N. Lu, P. Pittet, A. Rivoire,
 and L. Gindraux, Design and testing of a phantom and instrumented gynecological
 applicator based on GaN dosimeter for use in high dose rate brachytherapy quality
 assurance, Med. Phys. 43, 5240–5251 (2016).
- L. Cartwright, N. Suchowerska, Y. Yin, J. Lambert, M. Haque, and D. McKenzie, Dose
 mapping of the rectal wall during brachytherapy with an array of scintillation dosimeters,
 Med. Phys. 37, 2247–2255 (2010).
- F. Therriault-Proulx, S. Beddar, and L. Beaulieu, On the use of a single-fiber multipoint
 plastic scintillation detector for 192Ir high-dose-rate brachytherapy, Med. Phys. 40,
 062101 (2013).
- ⁵⁹³²⁵ H. M. Linares Rosales, P. Duguay-Drouin, L. Archambault, S. Beddar, and L. Beaulieu,
 Optimization of a multipoint plastic scintillator dosimeter for high dose rate brachyther apy, Med. Phys. 46, 2412–2421 (2019).
- J. Perez-Calatayud, F. Ballester, R. K. Das, L. A. DeWerd, G. S. Ibbott, A. S. Meigooni,
 Z. Ouhib, M. J. Rivard, R. S. Sloboda, and J. F. Williamson, Dose calculation for photonemitting brachytherapy sources with average energy higher than 50 keV: report of the
 AAPM and ESTRO, Medical physics **39**, 2904–2929 (2012).
- L. Archambault, F. Therriault-Proulx, S. Beddar, and L. Beaulieu, A mathematical for malism for hyperspectral, multipoint plastic scintillation detectors, Physics in Medicine
 & Biology 57, 7133 (2012).
- ⁶⁰³²⁸ M. A. Suarez, T. Lim, L. Robillot, V. Maillot, T. Lihoreau, P. Bontemps, L. Pazart, and
 ⁶⁰⁴ T. Grosjean, Miniaturized fiber dosimeter of medical ionizing radiations on a narrow
 ⁶⁰⁵ optical fiber, Opt. Express 27, 35588–35599 (2019).

- ⁶⁰⁶ ²⁹ M. Gonod, C. C. Avila, M. A. Suarez, J. Crouzilles, S. Laskri, J.-F. Vinchant,
 ⁶⁰⁷ L. Aubignac, and T. Grosjean, Miniaturized scintillator dosimeter for small field ra⁶⁰⁸ diation therapy, Phys. Med. Biol. **66**, 115016 (2021).
- ³⁰ M. Gonod, M. A. Suarez, C. C. Avila, V. Karakhanyan, C. Eustache, J. Crouzilles,
 S. Laskri, J.-F. Vinchant, L. Aubignac, and T. Grosjean, Characterization of a miniatur⁶¹¹ ized scintillator detector for time-resolved treatment monitoring in HDR-brachytherapy,
 ⁶¹² Phys. Med. Biol. **67**, 245016 (2022).
- ³¹ J. D. Kraus, R. J. Marhefka, and A. S. Khan, Antennas and wave propagation, Tata
 McGraw-Hill Education, 2006.
- ³² Z.Qin, Y.Hu, Y. Ma, W. Zhao, W. Sun, D. Zhang, Z. Chen, and E. Lewis, Embedded
 structure fiber-optic radiation dosimeter for radiotherapy applications, Opt. Express 24,
 ⁵¹⁷ 5172–5185 (2016).
- ³³ Y. Hu, Z. Qin, Y. Ma, W. Zhao, W. Sun, D. Zhang, Z. Chen, B. Wang, H. Tian,
 ⁶¹⁹ and E. Lewis, Characterization of fiber radiation dosimeters with different embedded
 ⁶²⁰ scintillator materials for radiotherapy applications, Sensors and Actuators A: Physical
 ⁶²¹ 269, 188–195 (2018).
- ³⁴ M. Alharbi, S. Gillespie, P. Woulfe, P. McCavana, S. O'Keeffe, and M. Foley, Dosimetric
 ⁶²³ characterization of an inorganic optical fiber sensor for external beam radiation therapy,
 ⁶²⁴ IEEE Sensors Journal 19, 2140–2147 (2018).
- ³⁵ D. O'Reilly, K. Qayyum, M. Alharbi, and M. Foley, Temperature Dependence of Novel
 ⁶²⁵ Inorganic Scintillation Detectors, in 2020 IEEE Sensors, pages 1–4, IEEE, 2020.

³⁶ O. McLaughlin, M. Martyn, C. Kleefeld, and M. Foley, Investigation of temperature
 dependence of inorganic scintillators using the HYPERSCINT research platform, Rad.
 Meas. 164, 106936 (2023).

G. P. Fonseca, R. S. Viana, M. Podesta, R. A. Rubo, C. P. de Sales, B. Reniers,
H. Yoriyaz, and F. Verhaegen, HDR 192Ir source speed measurements using a high
speed video camera, Med. Phys. 42, 412–415 (2015).

- ³⁸ H. M. Linares Rosales, J. G. Johansen, G. Kertzscher, K. Tanderup, L. Beaulieu, and
 S. Beddar, 3D source tracking and error detection in HDR using two independent
 scintillator dosimetry systems, Med. Phys. 48, 2095–2107 (2021).
- ³⁹ M. J. Rivard, B. M. Coursey, L. A. DeWerd, W. F. Hanson, M. Saiful Huq, G. S. Ibbott,
 M. G. Mitch, R. Nath, and J. F. Williamson, Update of AAPM Task Group No. 43
 Report: A revised AAPM protocol for brachytherapy dose calculations, Med. Phys. 31,
 ⁶³⁹ 633–674 (2004).
- ⁴⁰ R. Nath, L. L. Anderson, J. A. Meli, A. J. Olch, J. A. Stitt, and J. F. Williamson,
 ⁶⁴¹ Code of practice for brachytherapy physics: report of the AAPM Radiation Therapy
 ⁶⁴² Committee Task Group No. 56, Med. Phys. 24, 1557–1598 (1997).