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Simulation of data-driven multi-omic benchmark data for cellular deconvolution methods evaluation Hugo Barbot¹, David Causeur¹, Yuna Blum², Magali Richard³

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⇒ Cell deconvolution infers relative abundance of cell types using one or more -omic data [1].

Cell Deconvolution

Cellular heterogeneity in a bulk:

- \rightarrow refers to the variety of cell types within the bulk,
- \rightarrow reflects progression of **disease state**,
- \rightarrow is a complex mixture signal,
- \rightarrow is difficult to assess from bulk molecular profiles.

For now, we want **control on 4 hypotheses** of our deconvolution model based on Ordinary Least Squares optimisation:

A benchmark dataset generated in vitro is accessible (from COMETH project [2]) with:

$\overline{}$ 21104 gene expressions,

Bulks

- $\overline{}$ ∼ 800000 CpG probes methylation, \bullet $N = 30$ independent bulk,
- $\bullet K = 9$ cell types commonly found in PDAC.

Dependance with high dimensionnality

This allows us to see those uniform values as **quantile**. Then we only have to **define or** infer the marginal law of each feature to simulate any data types, thanks to the inversion of the Cumulative Distribution Function:

> $\left(X_{1},\ldots,X_{M}\right)=\left(F_{1}^{-1}\right)$ $F_1^{-1}(U_1),\ldots,F_M^{-1}(U_M)\big),$

where here X_i represents values for the **specific** feature *j* (gene/probe) and F_i is the marginal distribution function defined or inferred for the **specific** feature j (gene/probe) with its **specific** parameter which can change for each feature.

Natural way to deal with dependant data is by using a multivariate normal law. However, inferring a conditional correlation matrix with ∼ 20000 or ∼ 800000 features (gene/probe) is time consuming and quite inoperable for simulation with this approach.

Benchmark dataset

The true proportions of each cell type in each bulk are controlled and therefore can be assumed to be known.

> Since Ψ and B result from a decomposition of a correlation matrix, each line of $\varepsilon_{generated}$ is centered and scaled residuals.

Moreover, both omics have significative conditional two by two correlations between features:

We propose two different simulation methodologies for dependent data following specific marginals distribution functions:

 \rightarrow reproduce and make explicit hypothesis on different levels of complexity (dependencies, intrinsic nature of data, . . .),

 \rightarrow need at least one *in vivo* or *in vitro* dataset with known and controlled ground truth, however more datasets are needed to avoid overfitted simulation procedure.

-
- dependence between multiple variables.

Generation procedure

 \rightarrow Copulas generate dependant **uniform** random vectors (U_1, \ldots, U_M)

[1] Clémentine Decamps, Alexis Arnaud, Florent Petitprez, et al. DECONbench: a benchmarking platform dedicated to deconvolution methods for tumor heterogeneity quantification. BMC Bioinformatics, 22(1):473, October 2021.

[2] Yuna Blum, Jérôme Cros, Sergio Escalera et al. COMETH – COmputational METhods in Health.

[3] M Sklar. Fonctions de répartition à n dimensions et leurs marges. Annales de l'ISUP, 8(3):229-231, 1959.

[4] Chloé Friguet, Maela Kloareg, and David Causeur. A factor model approach to multiple testing under dependence. Journal of the American Statistical Association, 104(488):1406-1415, 2009.

loadings for each feature on each factor.

Generation procedure

$$
\varepsilon_{generated} = \mathcal{N}(0, \Psi) + B \times \mathcal{N}(0, I_q), \ \varepsilon_{generated} \in \mathcal{M}_{M,N}(\mathbb{R}).
$$

$$
\mathcal{E}_{generated} \leftarrow (\mathcal{E}_{generated} \times \hat{\sigma}_{feature}) + \hat{\mu}_{feature}
$$

The procedure mimics the behaviour of the data provided.

Results

Perspectives

Both methodologies:

 \rightarrow are computationally **fast**,

Ongoing works:

Here, Copulas and factor model methodology capture dependance structure empirically. We focus now on defining controlled parameters for each approach to simulate different scenarios.

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