

# Using a common morphometric-based method to sex a migratory bird along its entire flyway despite geographical and temporal variations in body size and sexual size dimorphism

Thomas Pagnon, Léa Etchart, Maria Teixeira, François-Xavier Dechaume-Moncharmont, Gunnar Hallgrimsson, Jannik Hansen, Johannes Lang, Jérôme Moreau, Jeroen Reneerkens, Niels Schmidt, et al.

#### ▶ To cite this version:

Thomas Pagnon, Léa Etchart, Maria Teixeira, François-Xavier Dechaume-Moncharmont, Gunnar Hallgrimsson, et al.. Using a common morphometric-based method to sex a migratory bird along its entire flyway despite geographical and temporal variations in body size and sexual size dimorphism. Journal of Ornithology, 2024, 165 (4), pp.923-934. 10.1007/s10336-024-02178-9. hal-04771784

# HAL Id: hal-04771784 https://hal.science/hal-04771784v1

Submitted on 7 Nov 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

- 1 Using a common morphometric-based method to sex a migratory
- bird along its entire flyway despite geographical and temporal
- variations in body size and sexual size dimorphism
- 5 Thomas Pagnon<sup>1,2</sup> (0009-0003-1204-5230), Léa Etchart<sup>1</sup> (0009-0004-0095-1196), Maria
- 6 Teixeira<sup>3</sup>, François-Xavier Dechaume-Moncharmont<sup>4</sup> (0000-0001-7607-8224), Gunnar T.
- 7 Hallgrimsson<sup>5</sup> (0000-0002-3697-9148), Jannik Hansen<sup>6</sup> (0000-0002-2716-9928), Johannes
- 8  $Lang^{2,7}$  (0000-0002-7387-795X), Jérôme Moreau<sup>2,3,8</sup> (0000-0001-9984-0998), Jeroen
- 9 Reneerkens<sup>9,10</sup> (0000-0003-0674-8143), Niels M. Schmidt<sup>6</sup> (0000-0002-4166-6218),
- 10 Mikhail Soloviev<sup>11</sup> (0000-0003-4040-1477), Job ten Horn<sup>10</sup> (0000-0002-8767-5929), Pavel
- 11 Tomkovich<sup>11</sup>, Andrew G. Wood<sup>12</sup>, Glenn Yannic<sup>2,13</sup> (0000-0002-6477-2312), Loïc
- 12 Bollache<sup>1,2</sup> (0000-0003-0316-6746), Olivier Gilg<sup>1,2</sup> (0000-0002-9083-4492)
- <sup>1</sup> UMR 6249 Chrono-environnement, CNRS, Université de Franche-Comté, 25030 Besançon,
- 15 France

- <sup>2</sup> Groupe de Recherche en Ecologie Arctique, 21440 Francheville, France
- 17 <sup>3</sup> UMR 6282 Biogéosciences, CNRS, Université de Bourgogne, 21000 Dijon, France
- <sup>4</sup> Univ. Lyon, Université Claude Bernard Lyon 1, CNRS, ENTPE, UMR 5023 LEHNA, F-
- 19 69622 Villeurbanne, France
- <sup>5</sup> Faculty of Life and Environmental Sciences, University of Iceland, 102 Reykjavík, Iceland
- <sup>6</sup> Department of Ecoscience and Arctic Research Centre, Aarhus University, 4000 Roskilde,
- 22 Denmark
- 23 <sup>7</sup> Working Group for Wildlife Research at the Clinic for Birds, Reptiles, Amphibians and
- Fish, Justus Liebig University Giessen, 35392 Giessen, Germany

- 25 8 UMR 7372, Centre d'Études Biologiques de Chizé, CNRS, Université de La Rochelle,
- 26 79360 Villiers en Bois, France
- 27 <sup>9</sup> Groningen Institute for Evolutionary Life Sciences (GELIFES), University of Groningen,
- 28 9747 AG Groningen, The Netherlands
- 29 10 Department of Coastal Systems, NIOZ Royal Netherlands Institute for Sea Research, PO
- box 59, 1790 AB Den Burg, Texel, The Netherlands
- 31 <sup>11</sup> Zoological Museum, Lomonosov Moscow State University, 125009 Moscow, Russia
- 32 <sup>12</sup> British Antarctic Survey, Natural Environment Research Council, Cambridge CB3 0ET,
- 33 United Kingdom
- 34 <sup>13</sup> Univ. Grenoble Alpes, Univ. Savoie Mont Blanc, CNRS, LECA, Laboratoire
- d'Ecologie Alpine, 38000 Grenoble, France
- 36 Corresponding author: Thomas Pagnon, thomas.pagnon@gmail.com

# 38 Abstract

- 39 Sexing bird species with monomorphic plumage is generally challenging, and sexual size
- 40 dimorphism (SSD) is often used to develop morphometric-based sexing tools, e.g., using
- 41 discriminant functions. Within species, local selection pressures, age-related and season-
- 42 related growth, may however induce geographical and temporal variations in body size and
- 43 SSD. Such variations may complicate the development of reliable morphometric-based sexing
- 44 methods at a broad scale.
- We first investigated body size variations in a migratory shorebird species with wide breeding
- and wintering ranges, the Sanderling Calidris alba, within the two breeding populations
- 47 (Greenland and Russia) and three staging/wintering populations (United Kingdom, Iceland
- and Mauritania), which belong to the same flyway. Then, for samples from each region, we
- 49 tested whether site-specific (i.e., "regional") functions performed better than functions

50 developed for birds from the other sites (i.e., "foreign" functions) or than an overall

51 ("flyway") function that combined all sampled individuals.

52 We found minor variations in SSD between regions, but significant differences in body size

53 between sexes and regions. Females were larger than males and, for instance, breeders had

longer wings than staging and wintering birds. Regional functions had similar sexing

efficiency as any other functions applied to sample from each region, except for Western

Africa where the regional function performed slightly better than some of the other functions.

Furthermore, the flyway function developed after merging all subsamples had a similar

58 efficiency than the regional functions, i.e., from 75.4% to 90% of correct sex assignment

59 depending on the region.

60 Given the small or lack of benefit in using regional functions, we conclude that the flyway

61 function can be used reliably to sex Sanderlings measured at different sites, years or seasons

within the East Atlantic flyway. Our results may help to develop global sexing function for

63 other bird species.

65 Keywords

66 Discriminant function, Molecular sexing, East Atlantic flyway, Shorebird, Sanderling.

68

67

54

55

56

57

62

64

# Introduction

70

72

69

71 Sex-related variations are commonly reported in ecological and evolutionary bird studies

(e.g., Remisiewicz and Wennerberg 2006; Saino et al. 2010; Carvalho Provinciato et al.

73 2018). In species which exhibit phenotypic differences between males and females,

determining the sex is an easy task. Conversely, in species with no or limited sexual 74 dimorphism and/or dichromatism, distinguishing between males and females can be 75 challenging. To solve this problem, molecular sexing is often the best and most accurate 76 option, but it requires the collection and preservation of biological samples, and comes with 77 economical and time constraints (Lessells and Mateman 1998; Morinha et al. 2012). 78 In many species, males and females differ slightly in some morphometric traits (Fairbairn et 79 al. 2007). In birds, such sexual size dimorphism (SSD) mainly concerns wing, bill, tarsus, and 80 head lengths (globally referred to as "body size"). Thus, based on morphological 81 measurements, sexing tools can be developed. Discriminant functions (DF), allowing sex 82 determination from a selection of measured morphometric data, are the most widely used 83 statistical tools (Witkowska and Meissner 2020; Almeida et al. 2020). These methods have 84 become popular among biologists because, after the initial effort in designing DFs, their 85 further applications are cheap, easy-to-use, minimally invasive and time effective (Dechaume-86 Moncharmont et al. 2011; Yannic et al. 2016). 87 The accuracy of a sexing DF depends on the degree of SSD in the studied species (Kocijan et 88 al. 2011). In species with low to moderate SSD, i.e., with large overlap in morphometrics 89 between sexes, the risk of sex assignment errors is higher than in species with high SSD 90 (Baker et al. 1999; Remisiewicz and Wennerberg 2006). In addition, in species with a wide 91 92 geographical range, different populations may be exposed to local selection pressures resulting in regional variations in body size and/or SSD (Badyaev et al. 2000; Helfenstein et 93 al. 2004; Santiago-Alarcon and Parker 2007; de Abreu 2018). In such cases, morphometric-94 based sexing methods might be region-specific (Granadeiro 1993; Weidinger and van 95 Francker 1998; Ellrich et al. 2010). For example, a sexing DF generated for Black Terns 96 (Chlidonias niger) in Wisconsin (USA) performed equally well on neighbouring populations, 97 whereas a sexing DF from the Oregon State performed poorly on the Wisconsin population 98

99 (Shealer and Cleary 2007). Moreover, individual measurements may change over time. Wing 100 length, like other plumage morphometric, can fluctuate during the year with moult and wear (Low 2006; Fernández and Lank 2007), reducing the reliability of morphometrics-based 101 sexing tools developed and used during different seasons. 102 To assess the potential effects of geographical and temporal variations of body size and SSD 103 on morphometrics-based sexing, we compared measurements from different populations 104 using the same flyway and during different seasons in a migratory shorebird species with a 105 wide geographical range, the Sanderling (Calidris alba). In breeding plumage, males of 106 Sanderling often (but not always) have brown-spotted cinnamon feathers on the sides of the 107 108 neck and upper breast, and tend to be slightly more colourful than females, while winter plumage is identical in both sexes (Pienkowski and Green 1976). As plumage cannot be 109 reliably used for sexing, several sexing DF have already been proposed for this species along 110 111 the East Atlantic flyway, from one breeding site (Russia; Soloviev and Tomkovich 1995), one staging-wintering site (United Kingdom; Wood 1987) and one wintering region (Western 112 Africa; Mauritania and Ghana combined; Lourenço et al. 2016). 113 In the present study, our aim was to determine whether the efficiency of a morphometric-114 based sexing tool is region-specific and time-specific, or if the development of a single 115 116 global-scale DF could be relevant. First, we compared body size and SSD of birds sampled in different regions using morphometrics from the three published studies mentioned above as 117 well as from unpublished data collected in Greenland and Iceland, respectively additional 118 breeding and staging sites belonging to the same flyway. Second, we developed new sexing 119 DFs for (1) Greenland and Iceland and for (2) the East Atlantic flyway using all available 120 data. Finally, we cross-compared the respective efficiency of each of these six sexing DFs to 121 assess the extent to which geographical and temporal variations in body size and SSD affected 122 the predictive power of each sexing DF. 123

125

126

#### Materials and methods

127

128

#### **Study species**

The Sanderling is a high Arctic shorebird species split into two subspecies. C. alba alba 129 130 breeds in Greenland and Russia and winters along the Atlantic coasts of Europe and Africa (East Atlantic flyway), the east coast of Africa (East and South African flyway) and Western 131 Asia (Southwest Asian flyway). C. alba rubidus, slightly larger than C. a. alba, breeds in 132 Canada and Alaska and winters along the coasts of the Americas (Reneerkens et al. 2009). 133 Along East Atlantic flyway, most Sanderlings wintering in Western Europe and Western 134 Africa appear to belong to the Greenland breeding population and to stop in Iceland during 135 their northbound migration (Gudmundsson and Lindström 1992; Reneerkens et al. 2009, 136 137 2020). 138 To date, no DF has ever been published to sex birds breeding in Greenland and migrating

141

142

139

140

unknown.

### Data collection and morphometric measurements

We collected morphometric data and genetic samples from 422 breeding adults (from June to
August during 2007-2021) at two study sites along the North-East coast of Greenland:
Hochstetter Forland (75.16666°N, 19.75000°W; 74 individuals) and Zackenberg
(74.46665°N, 20.56684°W; 348 individuals). Incubating adults were caught using a 40 cm
wide clap net (model BE40 from "Moudry traps", Czech Republic, www.moudry.cz) placed
over the nest and automatically released by birds returning on nests when sitting on their

through Iceland. The reliability of available DFs from other locations to sex these birds is also

clutch. The same data and samples were also collected in Sandgerði, Iceland (64.04261°N, 149 22.71404°W; 560 adults and immatures birds trapped with canon nets), between 2007 and 150 2013, in May and early June (i.e., just before Greenland birds return to their breeding sites; 151 Reneerkens et al. 2020). 152 In addition, we made use of the morphometric and sexing data available from the three 153 previous studies mentioned above, (i) 71 breeding adults trapped from June to August in the 154 Knipovich Bay, Russia (76.08333°N, 98.53333°E), in 1990-1992 (Soloviev and Tomkovich 155 1995), (ii) 45 adults birds staging or wintering from September to May around the estuary of 156 the Tees river, United Kingdom (54.64162°N, 1.15293°W) in 1983-1984 (Wood 1987), and 157 (iii) 928 birds (all age classes) wintering in Iwik, Mauritania (19.87754°N, 16.30356°W), 158 trapped in November-December between 2002 and 2011 (ca. 75% of birds used to develop 159 Western Africa DF; Lourenço et al. 2016). 160 161 For morphometric traits, the multivariate approach recommended by Engelmoer et al. (1987) was used in all sites and the following variables were measured with a single method: (1) bill 162 length (B), from the anterior edge of feathering to tip of culmen, (2) total head (TH), from the 163 back of the head to tip of culmen, (3) tarsus length (T), from the tarsal joint to the distal end 164 of the tarso-metatarsus (except in United Kingdom and Iceland), and (4) wing length (W), 165 166 straightened and flattened as described by Evans (1986). Bill length, total head and tarsus length were measured to the nearest 0.1 mm with a calliper; wing length was measured with a 167 stop ruler to the nearest 0.5 mm in all sites but United Kingdom (to the nearest millimetre). 168

169

170

171

172

173

#### Molecular and anatomical sexing

In birds from Greenland, Iceland and Mauritania, sexes were determined molecularly with a similar method. Three plucked pectoral feathers or one blood sample (with a capillary tube after puncturing the brachial vein with a 25-gauge needle) were collected from each bird.

Blood from capillary tubes was transferred and stored in microtubes filled with absolute ethanol. Cellular DNA was extracted following the protocol of Richardson et al. (2001) or according to manufacturer's instructions with 96-Well Plate Genomic DNA Miniprep Kit (Bio Basic). Sex of each bird was determined by polymerase chain reaction (PCR) analysis of the extracted DNA using the primers 2602F (5' CAGATGGTGAGGATGCTGGAC 3') and 2669R (5' CCCTTTTATTGATCCATCAAGYCTCTRAAGAG 3') designed by van der Velde et al. (2017). PCR reactions were carried out on a C1000 Touch TM thermal cycler (Bio-Rad) in 10 µL reaction mixture containing 0.1 U of Taq polymerase (Quantabio or Roche), 1X buffer, 0.2 mM dNTPs, 0.2 µM of each primer and 2 µL of DNA template. The thermal profile consisted of an initial denaturation for 1.5 min at 94°C, followed by 34 cycles of 30 sec at 94°C, 30 sec at 53°C or 62°C, 30 sec at 65°C, and a final extension at 65°C or 72°C during 10 min. 4 µL of PCR products were separated on a 2% agarose gel and visualized by UV-transillumination using GelDoc system (Bio-Rad) after ethidium bromide staining. Molecular sexing relied on the presence or absence of a sex-dependent DNA fragment, i.e., W chromosome fragment only found in females birds. The absence of this sex-dependent fragment is indistinguishable from a failure to amplify this fragment due to technical issues (Griffiths 2000). As in most comparable studies, we used sexes determined by this molecular method and assumed that they are correct (Baker et al. 1999), although we are aware that sexing errors could have happened (Roberston and Gemmell 2006). Birds measured in England were sexed by gonadal examination after dissection. Birds from Russia were sexed either by gonadal examination, cloaca measurements, or by mating behaviour of colour-marked birds.

196

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

### **Data analysis**

197

198

otherwise stated, the level of significance was set at P < 0.05. 199 All the assumptions required to run analysis were met, including normal distributions of the 200 data, homogeneity of covariances (Box's M test) and homogeneity of variance (Levene's test), 201 both for males and females. 202 203 We first tested variations in morphometrics according to site, sex and their interaction with multivariate and univariate analysis of variance (MANOVA/ANOVA). Then, we compared 204

All statistical tests were performed using R software version 4.2 (R Core Team 2022). Unless

body size between sexes using unpaired Student's t-test and Cohen's d with its confidence 205 206

interval 95% (effect size calculation; see Nagawaka and Cuthill 2007).

Morphometric means were given with standard deviation (SD) and coefficient of variation 207 (CV). The degree of sexual size dimorphism (SSD) for a given trait was determined by the 208 209 following equation (Almeida et al. 2020, modified from Lovich and Gibbons 1992):

210

$$SSD = \left[ \left( \frac{average \ of \ the \ larger \ sex}{average \ of \ the \ smaller \ sex} \right) - 1 \right] X100$$

211

Next, we compared body size and SSD between regions. The overall differences in 212 morphometric characters by sex between sites were determined with a MANOVA. Then, we 213 used the same method as the comparison between sexes to compare morphometric 214 215 measurements between sites. Tarsus length was not used in both multivariate analyses and in comparisons between sites, as this measurement is missing in UK and Iceland. 216 Second, we performed three linear discriminant analysis with the 'lda' function from MASS 217 package (Ripley et al. 2013) to determine functions which could distinguish sexes by the most 218 relevant morphometric characteristics for Sanderlings breeding in Greenland (Greenland DF), 219 staging in Iceland (Iceland DF), and using the East Atlantic flyway (flyway DF), including 220

datasets from all birds from all locations pooled. We used only data from birds having all measurements. Tarsus length data was only integrated in the process of discriminant analyses for Greenland. Potential outliers were initially detected by the Mahalanobis distances, using the chisq.plot and aq.plot functions from the mvoutlier package (Yannic et al. 2016). Detected outliers (n=5 from Greenland, n=0 from Iceland, n=14 from the flyway) were removed from the dataset due to uncorrectable mistakes in measurement or transcription. The final sample sizes and sex-ratios used to perform the discriminant analyses are indicated in table 1. We estimated the discriminant rate of all combinations of morphometric variables using the leaveone-out cross-validation (LOOCV) method to determine the best model. This procedure predicts the sex of an individual after this individual has been removed from the data, instead of automated stepwise techniques which can capitalize on sampling error and lead to nonreplicable results (Dechaume-Moncharmont et al. 2011). Note that DFs previously developed in the United Kingdom and in Russia followed similar discriminant analyses as ours, whereas the DF developed for Western Africa opted for a generalized linear model (GLM) with binomial errors and a logit-link function to examine the relationship between the molecular sex, each morphometrics and their interactions (with sex as the binary response variable). To test the ability of regional DFs (i.e., from Greenland, Iceland, Russia, United Kingdom and Western Africa) and the flyway DF to correctly assign sexes for birds measured in a given region, we compared result outputs between DFs using McNemar's test (Western Africa DF was not tested on UK, Iceland and all pooled samples because they do not include tarsus length measurements). To enable comparisons, success rates of DFs with their related samples were determined by resubstitution, i.e., the sex of each individual was predicted using the function obtained from the complete dataset and then compared with the sex identified using molecular or anatomical sexing.

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

247

248

#### Results

- 250 Size variability between breeding, staging and wintering sites
- 251 Multivariate analysis of variance indicated that body size varied significantly between sexes
- 252 (F<sub>3,1976</sub> = 397.254, P < 0.0001) and among regions (F<sub>12,5925</sub> = 24.523, P < 0.0001). Sex and
- region, but not their interaction, had also significant effects on each morphometrical variable
- 254 (Table 2).
- 255 Females were larger than males and bill length was the most dimorphic trait across sites (from
- 5.9% to 8.1%). Sexual size dimorphism of each set of morphometrics was similar between all
- sites, although the mean SSD was slightly higher in Russian birds (ca. 4.2% with very large
- effect sizes) and lower in Icelandic birds (ca. 4.03% with smaller effect sizes) than in others
- 259 (Table 3).
- Males differed in size among regions (MANOVA:  $F_{12,2796} = 10.348$ , P < 0.0001). Adult males
- 261 measured in Greenland and in United Kingdom had shorter bills than males measured in
- 262 Iceland (GL:  $t_{282.47} = -2.424$ , P = 0.0159, Cohen's  $d = 0.27 \pm 0.21$ ; UK: ns), in Russia (GL:
- 263  $t_{57.76} = -4.324$ , P < 0.0001, Cohen's  $d = 0.61 \pm 0.37$ ; UK:  $t_{33.40} = 2.525$ , P = 0.0164, Cohen's  $d = 0.61 \pm 0.37$ ; UK:  $t_{33.40} = 0.61$
- 264 0.75±0.57), and in Mauritania (GL:  $t_{342.71}$  = -6.010, P < 0.0001, Cohen's  $d = 0.51\pm0.17$ ; UK:
- 265  $t_{22.45} = 2.340$ , P = 0.0285, Cohen's  $d = 0.55 \pm 0.43$ ). Males from Mauritania had shorter wings
- 266 than males from Greenland ( $t_{442.21} = 2.620$ , P = 0.009, Cohen's  $d = 0.20 \pm 0.16$ ) and from
- 267 Russia ( $t_{38.88} = -2.694$ , P = 0.0103, Cohen's  $d = 0.38 \pm 0.35$ ).
- Females also differed in size among regions (MANOVA:  $F_{12,3114} = 13.826$ , P < 0.0001).
- Females from Greenland had longer wings than females from Iceland ( $t_{434.61} = 4.370$ , P <
- 270 0.0001, Cohen's  $d = 0.37 \pm 0.16$ ) and from Mauritania ( $t_{507.08} = 3.852$ , P = 0.0001, Cohen's d =

- 271 0.32±0.17), and shorter bills than females from Russia ( $t_{54.11} = -3.677$ , P = 0.0005, Cohen's d
- 272 = 0.60±0.35) and from Mauritania ( $t_{510.65}$  = -6.959, P < 0.0001, Cohen's  $d = 0.57\pm0.17$ ).
- Females measured in Mauritania had shorter wings than females measured in Russia ( $t_{51.18} = -$
- 274 4.038, P = 0.0002, Cohen's  $d = 0.53\pm0.34$ ) and in the United Kingdom ( $t_{24.63} = -2.583$ , P =
- 275 0.0161, Cohen's  $d = 0.57 \pm 0.42$ ).
- 276 In all morphometrics and for each sex, we observed large intra-regional variability and large
- overlap between regions (Table 3, Figure 1).

279

## Discriminant functions for Greenland, Iceland and the East Atlantic flyway

- 280 The selection of variables that best predicted sex led to the combination of wing length [W],
- bill length [B] and total head [TH] for the three discriminant functions (Table 4).
- For Greenland, the discriminant function correctly assigned 84% of the males and 84.4% of
- 283 the females, with overall accuracy of 84.2%, by cross-validation (see Table 1).
- The Icelandic discriminant function, correctly classified the sex of 80.6% of the birds, with
- 92.4% of the females and 48.7% of the males by cross-validation.
- 286 The East Atlantic flyway discriminant function yielded correct assignment for 80% of the
- males and 81.3% of the females, with 80.7% of overall accuracy, by cross-validation.
- 288 Misclassified birds had all D values ranged [-4 to 4.90] for Greenland, [-0.56 to 2.21] for
- 289 Russia, [-6.86 to 7.08] for Iceland, [-2.09 to 3.85] for UK, and [-9.36 to 6.57] for birds from
- 290 Mauritania (see Figure 2).

291

292

#### Compared accuracy of the six discriminant functions

- 293 When comparing overall (i.e., combining males and females) sex assignment success rates
- obtained with each regional DF (using equations from Table 1), we found that regional DFs
- showed similar success rates than foreign DFs. The only exception was for the Western Africa

DF which performed significantly better with Mauritanian birds than some of foreign DFs did. At the sex level, some discrepancies in success rate appeared for both sexes for birds from Greenland, Icelandic and Mauritania when using foreign DFs (Table 5).

DFs showed similar performances in sexing each foreign samples as their own sample at an overall level, except Russia DF which performed better for Russian birds than for birds from Greenland ( $\chi^2_1 = 4.05$ , P = 0.0439), United Kingdom ( $\chi^2_1 = 5.02$ , P = 0.0249), Iceland ( $\chi^2_1$ 

302 =10.38, P = 0012) and Mauritania ( $\chi^2_1 = 5.25, P = 0.0219$ ).

Overall, the flyway DF provided similar sex assignment success rates to the regional DFs for each regional sample. There were exceptions for birds from Iceland for which the flyway DF performed better than the Iceland DF (McNemar's test:  $\chi^2_1 = 4.82$ , P = 0.0279), and for each sex when applied to Mauritanian birds (McNemar's test: males  $\chi^2_1 = 23.51$ , P < 0.0001, females  $\chi^2_1 = 8.04$ , P = 0.0045; Table 5, last column). The flyway DF was better to sex mixed-origins birds than regional DFs (Table 5, last row).

Globally, Icelandic sample got the lowest sexing success rates (≤75%) and Russian sample got the highest ones (ca. 90%) with any DF, while for other samples the success rates were similar (ca. 80%).

**Discussion** 

In the Scolopacidae family (sandpipers and related species), most species present some SSD with females larger than males (Jehl and Murray 1986), which allows the development of DFs (e.g., Niemc et al. 2018; Meissner et al. 2018; Almeida et al. 2020; Witkowska and Meissner 2020; Pohlen et al. 2021). In this study, we investigated whether variation in SSD and body size within a flyway could limit the efficiency of DFs to sex Sanderling on a large scale. We

found that, despite quite similar SSD between the regions, the body size may differ. Still, each regional DF gave comparable results for any regional sample. The use of a sole flyway-scale DF appears to be relevant and could facilitate future studies.

324

325

321

322

323

#### Geographical and temporal size variations

Regional variations occurred in body size. Birds from Russia were larger than any other birds, 326 327 and birds on wintering/staging grounds had shorter wing lengths than birds on breeding grounds. 328 Sanderlings breeding in Greenland and in Russia belong to the same subspecies C. alba alba 329 (Engelmoer and Roselaar 1998) and show only a low genetic differentiation (Conklin et al. 330 2016), while they are separated by approximately 2000 km. Bill size discrepancies between 331 these two breeding populations could thus suggest a phenotypic or developmental plasticity in 332 333 Sanderling, and reflect differences in local environments experienced by adults or young birds (Lafuente and Beldade 2019). The two regions have relatively similar climatic conditions 334 (Hijmans et al. 2005) but host different insectivorous avian communities, which might result 335 in different food competition pressures shaping divergent bill morphology (Kelly et al. 2024). 336 Interestingly, birds sampled in Mauritania and Iceland had on average longer bills than those 337 338 measured in Greenland. This could indicate that populations wintering in Western Africa are composed not only by birds from North-East Greenland, but also include some birds from 339 Russia, while populations staging in Iceland also include larger birds from the North 340 341 American subspecies breeding in the Eastern Canadian Arctic (Reneerkens et al. 2008). We also noted that wings were on average shorter in Mauritania and Iceland, where adults and 342 immatures were measured, compared to breeding sites in Greenland where only adults were 343 measured. Wing length varies seasonally due to wear and moult, as well as weather and 344 feeding conditions during the growth of the primaries (Hall and Fransson 2000; Milá et al. 345

2008). Also, immature Sanderlings wear their primaries faster than adults (Pienkowski and Minton 1973), and replace them during their first summer at the earliest (Demongin 2016), whereas adult Sanderlings moult their primaries on wintering grounds (Loonstra et al. 2016). Immatures included in datasets could induce shorter wing lengths (for seabirds see e.g., Bugoni and Furness 2009). In Mauritania, for example, the proportion of immatures among wintering birds is relatively high (Reneerkens et al. 2020).

However, unlike skeletal elements, for which standard measurements are reliably repeatable (Rising and Somers 1989), wing length is difficult to calibrate and has limited accuracy (measured to the nearest 0.5 mm at most). Most changes in wing length between seasons and with age could be small relative to measurement errors and hence difficult to quantify (Francis and Wood 1989). Furthermore, body size can "drift" over time (Anderson et al. 2019; Zimova et al. 2023). Data used in this study were collected between the early 1980s for the oldest and the early 2020s for the latest. Some morphometric changes may have occurred among populations sampled 30 or 40 years ago, potentially adding a cohort effect.

# Morphometric-based sexing over large scale, any limits?

Morphometrics are usually effective in sexing birds from the *Calidrinae* subfamily (e.g., Meissner and Pilacka 2008; Almeida et al. 2020; Pohlen et al. 2021). Bill length is often the most discriminant morphometric amongst *Calidris* species, but appears seldom efficient alone for sexing because of substantial overlapping ranges between sexes (e.g., Niemc et al. 2018; Witkowska and Meissner 2020). Sexing requires the use of several other morphometrics in most of the cases, as for Sanderling.

Despite a moderate sexual dimorphism in Sanderling (ca. 4.6%), the DFs developed for this species allowed for 67% to 100% of correct sex assignments, depending on the sex and origin of the birds (Table 5). Such success rates might not be high enough for certain scientific

applications, e.g., sex-specific behaviour studies, which thus would require molecular 371 372 analysis. All DFs performed better for males than for females. This difference is most likely related to 373 sex-specific differences in size distribution, with many small females assigned as males and 374 only a few large males assigned as females. This may weakly affect breeding population 375 studies if both members of a pair are captured, but should be considered for staging and 376 377 wintering population studies (Koloski et al. 2016). Overall, foreign DFs performed equally well as regional DFs to assign sex for most regions. 378 In Western Africa only, the regional DF performed slightly better than some of the foreign 379 380 DFs. This may be due to the methodological difference (i.e., GLM compared to DFA) used to develop the Western Africa DF (Hallgrimsson et al. 2016). 381 To summarize, although we found significant geographical differences in body size (Table 1, 382 383 Fig. 1) and specific models were developed for each regional dataset, using regional DFs provided almost no benefit to sex Sanderlings as compared with the use of foreign DFs. 384 Correct assignment rates varied between sexes and regions, reflecting small differences in 385 SSD between regions, but did not vary according to the functions used (all but one DFs being 386 equally reliable to sex regional or foreign individuals), likely as a result of large variability in 387 388 the measured populations (see above). As expected, the flyway DF did not improve sexing efficiency at a regional scale, except for Icelandic birds, but performed equally well as 389 regional DFs, and could hence provide a more universal and practical tool than regional DFs. 390 391 However, as body size is predicted to change with ongoing climate change (Sheridan and Bickford 2011; Gardner et al. 2014; Youngflesh et al. 2022), current discriminant functions 392 may require updates in the future. 393 Our results have important practical implications for the study of Sanderling populations 394 using the same flyway, regardless of the location and the season. In particular, they can be 395

useful to design or reframe scientific programs in the field, helping to target a sex or to balance the sex-ratio in sampling for instance, without the need for molecular tools. Moreover, similar methodological considerations might be valuable in sexing other bird species with large geographical range.

# Acknowledgements

We are particularly grateful to all people who searched for the nests and helped to collect the field data. Yvonne Verkuil, Marco Van de Velde (University of Groningen) and Anneke Bol (NIOZ) molecularly sexed the birds from Mauritania, Iceland and Zackenberg (Greenland). Data provided by AW were collected by the Shorebird Research Group at Durham University. The Government of Greenland (Ministry of Domestic Affairs, Nature and Environment) in Nuuk granted access and research permits for field work in the National Park. We thank the two reviewers whose comments have greatly contributed to improve the manuscript.

# **Declarations**

#### 413 Conflict of interest

The authors declare no confict of interest. The authors have no competing interests to declare that are relevant to the content of this article.

#### **Ethical approval**

All data were collected by people who have licences for handling animals for scientifc purposes. All our fieldworks complied with the ethical standards and national laws on animal welfare.

421	
422	Funding
423	This research was supported by the French Polar Institute (IPEV, program 1036
424	'Interactions'), the Agence Nationale de la Recherche (ANR-21-CE02-0024 PACS to L.
425	Bollache), the Groupe de Recherche en Ecologie Arctique (GREA) and the University of
426	Bourgogne Franche-Comté (Chrono-environnement lab.).
427	
428	Authors contributions
429	TP and OG designed the study and wrote the first draft of the manuscript; TP and LE carried
430	out data analyses; MT helped with the genetic analysis; FXDM, GY and LB provided
431	methodological support and contributed to drafting the manuscript; GH, JH, JL, JM, JR, NS,
432	MS, JtH, PT, AW contributed to data collection and drafting the manuscript. All authors read
433	and approved the final manuscript.
434	
435	Data availability
436	Data are available on request from the concerned authors.
437	
438	
439	References
440	
441 442 443 444 445	Almeida JB, Lopes IF, Oring LW, Tibbitts TL, Pajot LM, Lanctot RB (2020) After-hatch and hatch year Buff-breasted Sandpipers <i>Calidris subruficollis</i> can be sexed accurately using morphometric measures. Wader Study 127(2):147–155. <a href="https://doi.org/10.18194/ws.00189">https://doi.org/10.18194/ws.00189</a> Anderson AM, Friis C, Gratto-Trevor CL, Morrison RG, Smith PA, Nol E (2019) Consistent declines in wing lengths of Calidridine sandpipers suggest a rapid morphometric response to

447 448

449 450 environmental

change.

https://doi.org/10.1371/journal.pone.0213930

**PLoS** 

dimorphism in the house finch. II. Population divergence in relation to local selection. Evolution 54(6):2134-2144. <a href="https://doi.org/10.1111/j.0014-3820.2000.tb01255.x">https://doi.org/10.1111/j.0014-3820.2000.tb01255.x</a>

Badyaev AV, Hill GE, Stoehr AM, Nolan PM, McGraw KJ (2000) The evolution of sexual size

One

18

14(4):e0213930.

- Baker AJ, Piersma T, Greenslade AD (1999) Molecular vs. phenotypic sexing in Red Knots. Condor 101(4):887–893. <a href="https://doi.org/10.2307/1370083">https://doi.org/10.2307/1370083</a>
- Bugoni L, Furness RW (2009) Age composition and sexual size dimorphism of albatrosses and petrels off Brazil. Mar Ornithol 37:253–260
- Carvalho Provinciato IC, Araújo MS, Jahn AE (2018) Drivers of wing shape in a widespread Neotropical bird: a dual role of sex-specific and migration-related functions. Evol Ecol 32:379–393. https://doi.org/10.1007/s10682-018-9945-4
- Conklin JR, Reneerkens J, Verkuil YI, Tomkovich PS, Palsbøll PJ, Piersma T (2016) Low genetic
   differentiation between Greenlandic and Siberian Sanderling populations implies a different
   phylogeographic history than found in Red Knots. J Ornithol 157(1):325–332.
   <a href="https://doi.org/10.1007/s10336-015-1284-4">https://doi.org/10.1007/s10336-015-1284-4</a>
  - de Abreu FHT, Schietti J, Anciães M (2018) Spatial and environmental correlates of intraspecific morphological variation in three species of passerine birds from the Purus–Madeira interfluvium, Central Amazonia. Evol Ecol 32:191–214. <a href="https://doi.org/10.1007/s10682-018-9929-4">https://doi.org/10.1007/s10682-018-9929-4</a>
- Dechaume-Moncharmont FX, Monceau K, Cezilly F (2011) Sexing birds using discriminant function analysis: a critical appraisal. The Auk: Ornithol Adv 128:78–86. https://doi.org/10.1525/auk.2011.10129
- Demongin L (2016) Identification guide to birds in the hands. Beauregard-Vendon

463

464

465

478 479

480

484

485

486 487

488

489

- Ellrich H, Salewski V, Fiedler W (2010) Morphological sexing of passerines: not valid over wider geographical scales. J Ornithol 151:449–458. <a href="https://doi.org/10.1007/s10336-009-0478-z">https://doi.org/10.1007/s10336-009-0478-z</a>
- Engelmoer M, Roselaar CS, Nieboer E, Boere GC (1987) Biometrics in waders. Wader Study Group Bull 51:44–47
- 474 Engelmoer M, Roselaar CS (1998) Geographic variation in waders. Kluwer, Dordrecht
- Evans PR (1986) Correct measurement of wing-length of waders. Wader Study Group Bull 48:11
- Fairbairn DJ, Blanckenhorn WU, Szekely T (2007) Sex, size and gender roles: evolutionary studies of sexual size dimorphism. Oxford university press, London
  - Fernández G, Lank DB (2007) Variation in the wing morphology of Western Sandpipers (*Calidris Mauri*) in relation to sex, age class, and annual cycle. The Auk: Ornithol Adv 124(3):1037–1046. https://doi.org/10.1093/auk/124.3.1037
- Francis CM, Wood DS (1989) Effects of age and wear on wing length of Wood-Warblers (Efecto de la edad y el desgaste en el largo del ala de Emberísidos (Emberizidae)). J Field Ornithol 60(4):495–503
  - Gardner JL, Amano T, Backwell PR, Ikin K, Sutherland WJ, Peters A (2014) Temporal patterns of avian body size reflect linear size responses to broadscale environmental change over the last 50 years. J Avian Biol 45(6):529–535. <a href="https://doi.org/10.1111/jav.00431">https://doi.org/10.1111/jav.00431</a>
  - Granadeiro JP (1993) Variation in measurements of Cory's Shearwater between populations and sexing by discriminant analysis. Ringing and Migration 14:103–112. https://doi.org/10.1080/03078698.1993.9674051
  - Griffiths R (2000) Sex identification using DNA markers. In: Baker AJ (ed) Molecular methods in ecology. Blackwell Science, Oxford, pp 295–321
- Gudmundsson GA, Lindström Å (1992) Spring migration of Sanderlings *Calidris alba* through SW
   Iceland: wherefrom and whereto? Ardea 80:315–326
- 494 Hall S, Fransson T (2000) Lesser Whitethroats under time-constraint moult more rapidly and grow 495 shorter wing feathers. J Avian Biol 31:583–587. <a href="https://doi.org/10.1034/j.1600-048X.2000.310419.x">https://doi.org/10.1034/j.1600-048X.2000.310419.x</a>
- Hallgrimsson GT, Helgason HH, Palsdottir ES, Palsson S (2016) Sexing adult and fledgling Lesser
   Black-backed Gulls from morphometrics. Ringing and Migration 31(1):68-73.
   https://doi.org/10.1080/03078698.2016.1190617
- Helfenstein F, Danchin E, Wagner RH (2004) Assortative mating and sexual size dimorphism in Black-legged Kittiwakes. Waterbirds 27(3):350–354. <a href="https://doi.org/10.1675/1524-4695(2004)027[0350:AMASSD]2.0.CO;2">https://doi.org/10.1675/1524-4695(2004)027[0350:AMASSD]2.0.CO;2</a>
- Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A (2005) Very high resolution interpolated climate surfaces for global land areas. Int J Climatol 25:1965–1978. https://doi.org/10.1002/joc.1276

- Intergovernmental Panel on Climate Change (2022) Climate Change 2022: Impacts, Adaptation and Vulnerability. Contribution of working groups I, II and III to the fifth assessment report of the intergovernmental panel on climate change. Geneva
- Jehl JR Jr, Murray G Jr (1986) The evolution of normal and reverse sexual size dimorphism in shorebirds and other birds. Curr Ornithol 3:1–86. <a href="https://doi.org/10.1007/978-1-4615-6784-4">https://doi.org/10.1007/978-1-4615-6784-4</a> 1
- Kelly DJ, O'Connell DP, Ó Marcaigh F, Kelly SBA, Karya A, Analuddin K, Marples N (2024)
  Rolling with the punches—How competition shapes the morphology of small passerines on small islands. J Biogeogr 00:1–9. https://doi.org/10.1111/jbi.14838
- Kocijan I, Dolenec P, Sinko T, Nenadic DD, Pavokovic G, Dolenec Z (2011) Sex-typing bird species with little or no sexual dimorphism: an evaluation of molecular and morphological sexing. J Biol Res Thessaloniki 15:145–150
- 517 Koloski L, Coulson S, Nol E (2016) Sex determination in breeding Dunlin (*Calidris alpina hudsonia*). Waterbirds 39(1):27–33. https://doi.org/10.1675/063.039.0104
- Lafuente E, Beldade P (2019) Genomics of developmental plasticity in animals. Front Genet 10:720.
- Lessells CM, Mateman AC (1998) Sexing birds using random amplified polymorphic DNA (RAPD) markers. Mol Ecol 7(2):187–195. <a href="https://doi.org/10.1046/j.1365-294x.1998.00331.x">https://doi.org/10.1046/j.1365-294x.1998.00331.x</a>
- Loonstra AHJ, Piersma T, Reneerkens J (2016) Staging duration and passage population size of Sanderlings in the western Dutch Wadden Sea. Ardea 104:49–61. https://doi.org/10.5253/arde.v104i1.a4
- Low M (2006) Sex, age and season influence morphometrics in the New Zealand Stitchbird (or Hihi; *Notiomystis cincta*). Emu 106:297–304. https://doi.org/10.1071/MU06003
- Lourenço PM, Alves JA, Reneerkens J, Loonstra AJ, Potts PM, Granadeiro JP, Catry T (2016) Influence of age and sex on winter site fidelity of sanderlings *Calidris alba*. PeerJ 4:e2517. https://doi.org/10.7717/peerj.2517
- Lovich JE, Gibbons JW (1992) A review of techniques for quantifying sexual size dimorphism. Growth, Development and Aging 56:269–281
- 532 Meissner W, Pilacka L (2008) Sex identification of adult Dunlins *Calidris alpina alpina* migrating in autumn through Baltic region. Ornis Fennica 85:135–139

535

536

546

- Meissner W, Pinchuk P, Karlionova N, Fischer I, Pilacka L (2018) Sexual size dimorphism and sex determination by external measurements in the Redshank *Tringa totanus*. Turk J Zool 42:1–5. https://doi.org/10.3906/zoo-1705-52
- 537 Milá B, Wayne RK, Smith TB (2008) Ecomorphology of migratory and sedentary populations of the 538 yellow-rumped warbler (*Dendroica coronata*). The Condor: Ornithol appl 110:335–244. 539 https://doi.org/10.1525/cond.2008.8396
- Morinha F, Cabral JA, Bastos E (2012) Molecular sexing of birds: A comparative review of polymerase chain reaction (PCR)-based methods. Theriogenology 78:703–714. https://doi.org/10.1016/j.theriogenology.2012.04.015
- Nakagawa S, Cuthill IC (2007) Effect size, confidence interval and statistical significance: a practical guide for biologists. Biol Rev 82(4):591–605. <a href="https://doi.org/10.1111/j.1469-185X.2007.00027.x">https://doi.org/10.1111/j.1469-185X.2007.00027.x</a>
  - Niemc A, Remisiewicz M, Avni J, Underhill LG (2018) Sexual dimorphism in adult Little Stints (*Calidris minuta*) revealed by DNA sexing and discriminant analysis. PeerJ 6:e5367 https://doi.org/10.7717/peerj.5367
- Pienkowski MW, Minton CDT (1973) Wing length changes of the Knot with age and time since moult. Bird Study 20(1):63–68. <a href="https://doi.org/10.1080/00063657309476359">https://doi.org/10.1080/00063657309476359</a>
- Pienkowski MW, Green GH (1976) Breeding biology of Sanderlings in north-east Greenland. Br Birds 69:165–177
- Pohlen ZM, Decicco LH, Johnson JA, Buchanan JB, Tomkovich PS (2021) Sex determination of Red Knots *Calidris canutus roselaari* using morphometrics. Wader Study 128(2):183–188. https://doi.org/10.18194/ws.00241
- R Development Core Team (2022) R: A language and environment for statistical computing, reference index Version 4.2.1. R Foundation for Statistical Computing, Vienna
- Remisiewicz M, Wennerberg L (2006) Differential migration strategies of the Wood Sandpiper (*Tringa glareola*)—genetic analyses reveal sex differences in morphology and spring migration phenology. Ornis Fennica 83:1–10

- Reneerkens J, Morrison RIG, Coulomb G (2008) First re-sighting of an individually marked Sanderling *Calidris alba* from Ellesmere Island. Wader Study Group Bull 115:116–118
- Reneerkens J, Benhoussa A, Boland H, Collier M, Grond K, Günther K, Hallgrimsson GT, Hansen J,
   Meissner W, de Meulenaer B, Ntiamoa-Baidu Y, Piersma T, Poot M, van Roomen M, Summers
   RW, Tomkovich PS, Underhill LG (2009) Sanderlings using African–Eurasian flyways: a
   review of current knowledge. Wader Study Group Bull 116(1):2–20

- Reneerkens J, Versluijs TS, Piersma T, Alves JA, Boorman M, Corse C, Gilg O, Hallgrimsson GT, Lang J, Loos B, Ntiamoa-Baidu Y, Nuoh AA, Potts PM, ten Horn J, Lok T (2020) Low fitness at low latitudes: Wintering in the tropics increases migratory delays and mortality rates in an Arctic breeding shorebird. J Anim Ecol 89(3):691–703. <a href="https://doi.org/10.1111/1365-2656.13118">https://doi.org/10.1111/1365-2656.13118</a>
- Richardson DS, Jury FL, Blaakmeer K, Komdeur J, Burke T (2001) Parentage assignment and extragroup paternity in a cooperative breeder: The Seychelles Warbler (*Acrocephalus sechellensis*). Mol Ecol10:2263–2273. https://doi.org/10.1046/j.0962-1083.2001.01355.x
- Ripley B, Venables B, Bates DM, Hornik K, Gebhardt A, Firth D, Ripley MB (2013) Package 'mass'. Cran r 538, pp 113–120
- Rising JD, Somers KM (1989) The measurement of overall body size in birds. The Auk: Ornithol Adv 106(4):666–674. https://doi.org/10.1093/auk/106.4.666
- Robertson BC, Gemmell NJ (2006) PCR-based sexing in conservation biology: wrong answers from an accurate methodology? Conserv Genet 7:267–271. <a href="https://doi.org/10.1007/s10592-005-9105-6">https://doi.org/10.1007/s10592-005-9105-6</a>
- Saino N, Rubolini D, Serra L, Caprioli M, Morganti M, Ambrosini R, Spina F (2010) Sex-related variation in migration phenology in relation to sexual dimorphism: a test of competing hypotheses for the evolution of protandry. J Evol Biol 23:2054–2065. https://doi.org/10.1111/j.1420-9101.2010.02068.x
- Santiago-Alarcon D, Parker PG (2007) Sexual size dimorphism and morphological evidence supporting the recognition of two subspecies in the Galápagos Dove. Condor 109(1):132–141. https://doi.org/10.1093/condor/109.1.132
- Shealer DA, Cleary CM (2007) Sex determination of adult Black Terns by DNA and morphometrics: Tests of sample size, temporal stability and geographic specificity in the classification accuracy of discriminant function models. Waterbirds 30(2):180–188. <a href="https://doi.org/10.1675/1524-4695(2007)30[180:SDOABT]2.0.CO;2">https://doi.org/10.1675/1524-4695(2007)30[180:SDOABT]2.0.CO;2</a>
- Sheridan JA, Bickford D (2011) Shrinking body size as an ecological response to climate change. Nat Clim Change 1:401–406. <a href="https://doi.org/10.1038/nclimate1259">https://doi.org/10.1038/nclimate1259</a>
- Soloviev MY, Tomkovich RS (1995) Biometrics of Sanderlings *Calidris alba* from the Taimyr. Ringing and Migration 16:91–99. <a href="https://doi.org/10.1080/03078698.1995.9674097">https://doi.org/10.1080/03078698.1995.9674097</a>
- van der Velde M, Haddrath O, Verkuil YI, Baker AJ, Piersma T (2017) New primers for molecular sex identification of waders. Wader Study 124(2):147–151
- van Gils JA, Lisovski S, Lok T, Meissner W, Ożarowska A, de Fouw J, Rakhimberdiev E, Soloviev MY, Piersma T, Klaassen M (2016) Body shrinkage due to Arctic warming reduces red knot fitness in tropical wintering range. Science 352:819–821. <a href="https://doi.org/10.1126/science.aad6351">https://doi.org/10.1126/science.aad6351</a>
- Weidinger K, van Francker JA (1998) Applicability of external measurements to sexing of the Cape petrel *Daption capense* at within-pair, within-population and between-population scales. J Zool 245:473–482. <a href="https://doi.org/10.1017/S0952836998008115">https://doi.org/10.1017/S0952836998008115</a>
- Witkowska M, Meissner W (2020) Sexual dimorphism in size and plumage in adult Curlew Sandpipers (*Calidris ferruginea*) migrating in autumn through the Baltic Sea region. Ornis Fennica 97:186–199
- Wood AG (1987) Discriminating the sex of Sanderling *Calidris alba*: some results and their implications. Bird Study 34(3):200–204. <a href="https://doi.org/10.1080/00063658709476962">https://doi.org/10.1080/00063658709476962</a>
- Yannic G, Broquet T, Strøm H, Aebischer A, Dufresnes C, Gavrilo MV, Gilchrist HG, Mallory ML,
  Morrison RIG, Sabard B, Sermier R, Gilg O (2016) Genetic and morphological sex
  identification methods reveal a male-biased sex ratio in the Ivory Gull *Pagophila eburnea*. J
  Ornithol 157(3):861–873. https://doi.org/10.1007/s10336-016-1328-4

Youngflesh C, Saracco JF, Siegel RB, Tingley MW (2022) Abiotic conditions shape spatial and temporal morphological variation in North American birds. Nat Ecol Evol 6:1860–1870. <a href="https://doi.org/10.1038/s41559-022-01893-x">https://doi.org/10.1038/s41559-022-01893-x</a>
Zimova M, Weeks BC, Willard DE, Giery ST, Jirinec V, Burner RC, Winger BM (2023) Body size predicts the rate of contemporary morphological change in birds. Proc Natl Acad Sci USA 120(20):e2206971120. <a href="https://doi.org/10.1073/pnas.2206971120">https://doi.org/10.1073/pnas.2206971120</a>

# **Tables and figures**

**Table 1** Discriminant functions developed for birds from Greenland (present study), Russia (Soloviev and Tomkovich 1995), Iceland (present study), United Kingdom (Wood 1987), Western Africa (Laurenço et al. 2016) and all sites combined (referred as East Atlantic flyway; present study)

Location	Sample size	Age class	Trapping period	Statistical method <sup>a</sup>	Discriminant Function <sup>b</sup>
Greenland	411 (200M/211F)	Adults	Summer	DFA	D = 110.28 - 0.397W - 0.919B - 0.748TH (male if D>0)
Russia	47 (18M/29F)	Adults	Summer	DFA	D = -166.059 + 0.932W + 0.645B + 0.625TH (male if D<0)
Iceland	557 (150M/407F)	Adults + Immatures	Spring	DFA	D = 98.38 - 0.359W - 0.764B - 0.67TH (male if D>0)
United Kingdom	42 (22M/20F)	Adults	Autumn + Winter + Spring	DFA	D = 0.375W + 1.13B - 75.5 (male if D<0)

Western Africa	990 (549M/441F)	Adults + Immatures	Winter	GLM	$\begin{array}{l} D=-0.5+e^{(\text{-}6486,316+276,9894B+128,3699TH+50,50511W-0,3635402T-5,455647BTH-2,148423BW-0,9945474THW+0,04216853BTHW)}/1+e^{(\text{-}6486,316+276,9894B+128,3699TH+50,50511W-0,3635402T-5,455647BTH-2,148423BW-0,9945474THW+0,04216853BTHW)} \\ \text{(male if D>0)} \end{array}$
East Atlantic flyway	1967 (929M/1038F)	Adults + Immatures	All seasons	DFA	D = 101.25 - 0.352W - 0.825B - 0.727TH (male if D>0)

<sup>628 &</sup>lt;sup>a</sup> DFA: discriminant function analysis; GLM: generalized linear model.

<sup>629 &</sup>lt;sup>b</sup> W: wing length; B: bill length; TH: total head; T: tarsus length.

**Table 2** Results of the univariate analysis of variance for factors contributing to variation in morphometric measures of Sanderlings in Greenland, Iceland, Russia, United Kingdom and Mauritania. Values correspond to *F*-ratios for each factors, with associated probabilities

Factors	df	Wing length	Bill length	Total head
Sex	1	561.27****	774.77***	926.45****
Region	3	12.63****	11.98****	10.62****
Sex * Region	4	1.87 (ns)	1.42 (ns)	1.48 (ns)

636 ns = not significant at P > 0.05

637 \*\*\*\* = *P* < 0.001

**Table 3** Descriptive statistics of morphometric measurements (in mm) of Sanderlings captured in Greenland (present study), Iceland (present study), Russia (Soloviev and Tomkovich 1995), United Kingdom (Wood 1987) and Mauritania (Laurenço et al. 2016). Standard deviation (SD), coefficient of variation (CV) and sample size (N) are given for each sex. Student *t*-test with degree of freedom (df) and *P*-value, Cohen's *d* and degree of sexual size dimorphism (SSD) between sexes are given for each morphometric

	Males				Females								
	Mean	Mean SD CV N		N	Mean SD CV N		t df		P	Cohen's <i>d</i> [CI95%]	SSD		
				Gree	enland								
Wing length	125.5	2.3	1.8	203	128.8	2.8	2.2	216	-13.50	410.8	< 0.0001	1.31 [1.10-1.52]	2.6
Bill length	23.3	1.2	5.1	205	25	1.1	4.4	217	-14.73	405.8	< 0.0001	1.44 [1.23-1.66]	7.3
Total head	49.1	2.1	4.3	204	51.3	1.3	2.5	216	-12.39	340.5	< 0.0001	1.22 [1.02-1.43]	4.5
Tarsus length	24.8	0.9	3.6	205	25.7	1.1	4.3	215	-9.22	409.9	< 0.0001	0.90 [0.69-1.10]	3.6
-				Rı	ıssia								
Wing length	126	2.2	1.7	33	129.6	2.3	1.8	37	-6.55	67.8	< 0.0001	1.56 [1.02-2.11]	2.8
Bill length	24.1	0.8	3.3	33	25.6	1	3.9	38	-7.39	69	< 0.0001	1.74 [1.18-2.29]	6.2
Total head	49.3	0.9	1.8	20	51.1	1.2	2.3	30	-6.01	47.2	< 0.0001	1.64 [0.97-2.31]	3.6
Tarsus length	24.8	0.7	2.8	33	25.8	1	3.9	38	-5.10	66.7	< 0.0001	1.19 [0.67-1.70]	4.0
				Ice	land								
Wing length	124.9	2.8	2.2	150	127.8	2.8	2.2	410	-10.91	264.1	< 0.0001	1.04 [0.85-1.24]	2.3
Bill length	23.7	1.5	6.3	150	25.1	1.2	4.8	410	-10.48	232.3	< 0.0001	1.08 [0.88-1.28]	5.9
Total head	49.2	1.7	3.4	150	51.1	1.4	2.7	407	-12.00	233.1	< 0.0001	1.23 [1.03-1.44]	3.9
Tarsus length	_	_	_	_	_	_			_	_	_	_	_

				United 1	Kingdom								
Wing length	124.7	4.4	3.5	22	129.7	3.4	2.6	23	4.27	39.1	0.0001	1.28 [0.62-1.94]	4.0
Bill length	23.3	1.2	5.1	22	25.2	1.5	5.9	23	4.61	41.5	< 0.0001	1.37 [0.70-2.04]	8.1
Total head	48.8	1.5	3.1	22	50.9	1.7	3.3	23	4.50	42.8	< 0.0001	1.34 [0.67-2.00]	4.3
Tarsus length	_	_	_	_	_	_	_	_		_	_	_	_
Mauritania													
				Mau	ritania								
Wing length	124.9	2.9	2.3	Maur 545	ritania 127.9	3.3	2.6	372	-14.03	724	< 0.0001	0.97 [0.83-1.11]	2.4
Wing length Bill length	124.9 23.9	2.9 1.1	2.3 4.6			3.3 1.2	2.6 4.7	372 376	-14.03 -21.52	724 748.8	< 0.0001 < 0.0001	0.97 [0.83-1.11] 1.47 [1.32-1.61]	2.4 7.1
6 6				545	127.9								
Bill length	23.9	1.1	4.6	545 552	127.9 25.6	1.2	4.7	376	-21.52	748.8	< 0.0001	1.47 [1.32-1.61]	7.1

**Table 4** Statistical significances from the Sanderling morphometric variables selections in Greenland, Iceland and East Atlantic flyway (all available samples combined) to determine the best sexing discriminant functions. Values correspond to Wilks'  $\lambda$  with associated Fisher F and P-value

		Wilks' $\lambda$	F	P
Greenland	W	0.697	177.4	< 0.001
	В	0.632	237.8	< 0.001
	TH	0.618	252.4	< 0.001
Iceland	W	0.822	119.9	< 0.001
	В	0.813	126.9	< 0.001
	TH	0.768	167.2	< 0.001
East Atlantic flyway	W	0.765	602.3	< 0.001
	В	0.707	813.4	< 0.001
	TH	0.631	1145.7	< 0.001

W: wing length; B: bill length; TH: total head

**Table 5** Success rates (percentage of correct sex assignments) of the six discriminant functions developed for Sanderling from Greenland, Russia, Iceland, United Kingdom, Western Africa and East Atlantic flyway (all samples pooled). Numbers in brackets indicate the difference (percentage point) to the success rate obtained with regional function by resubstitution (i.e., region were birds were captured, marked in grey). Values with significant differences (P < 0.05; McNemar's test) in success rate between the regional function and foreign functions for each sample are in bold

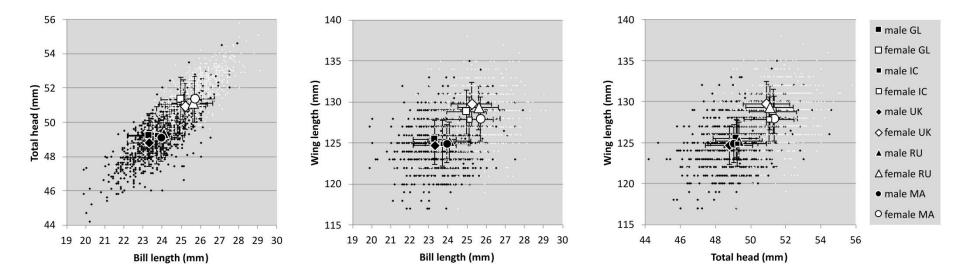
		n	Greenland function	Russia function	Iceland function	UK function	W Africa function <sup>a</sup>	Flyway function
Greenland sample	M	200	84.5%	88% (+3.5)	91.5% (+7.0)	90.5% (+6.0)	91% (+6.5)	84.5%
	F	211	82.5%	75.3% (-7.2)	73.4% (-9.1)	73% (-9.5)	72.5% (-10.0)	82% (-0.5)
	All	411	83.4%	81.5% (-1.9)	82.2% (-1.2)	81.5% (-1.9)	81.5% (-1.9)	83.2% (-0.2)
Russia sample	M	20	90% (-10.0)	100%	100%	90% (-10.0)	90% (-10.0)	90% (-10.0)
	F	30	93.3% (+3.3)	90%	80% (-10.0)	90%	90%	90%
	All	50	92% (-2.0)	94%	88% (-6.0)	90% (-4.0)	90% (-4.0)	90% (-4.0)
Iceland sample	M	150	78% (-7.3)	85.3%	85.3%	82% (-3.3)	_	78% (-7.3)
	F	407	74.9% (+6.4)	67.1% (-1.4)	68.5%	67.8% (-0.7)	_	74.4% (+5.9)
	All	557	75.7% (+2.6)	72% (-1.1)	73.1%	71.6% (-1.5)	_	75.4% (+2.3)

UK sample	M	22	81.8% (+4.5)	77.3%	86.3% (+9.0)	77.3%	_	81.8% (+4.5)
	F	23	78.3% (+4.4)	73.9%	69.5% (-4.4)	73.9%	_	78.3% (+4.4)
	All	45	80% (+4.5)	75.5%	77.8% (+2.3)	75.5%	_	80% (+4.5)
Mauritania sample	M	542	81.4% (-5.3)	87.6% (+1.1)	88% (+1.3)	83% (-3.7)	86.7%	81.7% (-5.0)
	F	372	81.4% (+3.7)	68.3% (-9.4)	75.8% (-1.9)	76.3% (-1.4)	77.7%	81.2% (+3.5)
	All	914	81.4% (-1.6)	79.7% (-3.3)	83%	80.3% (-2.7)	83%	81.5% (-1.5)
Flyway sample	M	929	82.2% (-0.2)	87.8% (+5.4)	89% (+6.6)	85.1% (+2.7)	-	82.4%
	F	1038	79.7% (+0.5)	70.2% (-9.0)	72.7% (-6.5)	72.8% (-6.4)	-	79.2%
	All	1967	80.9% (+0.2)	78.5% (-2.2)	80.4% (-0.3)	78.6% (-2.1)	_	80.7%

n denotes sample size for individuals genetically or anatomically sexed with all morphometrics measured

<sup>&</sup>lt;sup>a</sup> Western Africa function was not tested on UK, Iceland and flyway samples because they do not include tarsus length measurements

**Fig. 1** Relationships between wing length, bill length and total head in Greenland (GL), Russia (RU), Iceland (IC), United Kingdom (UK) and Mauritania (MA) Sanderlings males (black symbols) and females (white symbols). Small dots represent the individuals (symbols slightly larger for males to show overlapping colours). Mean values + SD are indicated with large symbols.



**Fig. 2** Frequency distribution (number) of males (light grey) and females (dark grey) Sanderlings measured for each regional sample and the pooled samples (flyway sample), as assigned with the flyway discriminant functions. The black parts of the bars present the range of overlapping scores. Vertical lines present the discrimination value

