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# New records of South American Cerambycidae (Coleoptera), with notes on the authorship of *Homogenes* and corrections in the species-group names of some species

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## Keywords:

Coleoptera;  
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*Atrypanius*;  
*Nyssodrysternum*;  
*Oxathres*;  
taxonomy;  
Neotropical region;  
South America.

**Abstract.** – The authorship of the genus *Homogenes* (Cerambycinae, Dichophyiini) is corrected and a new Brazilian state record is provided for *Homogenes albolineatus* (Buquet, 1844). Two new country records for Peru are provided as well as eight new regional records. A host plant is recorded for the first time for *Atrypanius unpanus* Juárez-Noé & González-Coronado, 2021 (Lamiinae, Acanthocinini). Additionally, *Nyssodrysternum instabile* Monné, 1992 is recorded from the Brazilian state of Minas Gerais and *Oxathres guyanensis* Monné & Tavakilian, 2011 is recorded for the first time from Brazil. Twenty-six species-group names are corrected in Cerambycinae and Lamiinae.

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## Introduction

New records have been quite frequently reported recently in Cerambycidae, demonstrating that the fauna of many regions is still not well known. Even in regions that have been extensively collected in the past, it is relatively common to find species that had not yet been collected in those places.

Here, we are recording some species for Brazil (new state records and a new country record) and several for regions of Peru. One new record for Peru also included the host plant of the species. Additionally, twenty-six corrections in species-group names of Cerambycidae are provided.

## Materials and Methods

Photographs were taken by the first author using a Canon EOS TD Mark II camera equipped with a Canon MP-E 65 mm f/2.8 1-5X macro lens. The images were processed with Zerene Stacker AutoMontage software; except the photographs of *Atrypanius unpanus* Juárez-Noé & González-Coronado, 2021 and the fruits of its host plant, taken by the third author. In “Material examined,” only

the specimens used to establish the new records are listed. The references on each species are restricted to the original description. For full references, see Monné (2024a, b) and/or Tavakilian & Chevillotte (2024).

The collection acronyms used in the text are as follows:

- **LGBC:** Larry G. Bezark Collection, Sacramento, California, USA.
- **MNRJ:** Museu Nacional, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil.
- **MZSP:** Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil.
- **SENASA:** Servicio Nacional de Sanidad Agraria, Colección Entomológica Referencial, Lima, Peru.

## New records and notes

**CERAMBYCIDAE** Latreille, 1802

**ACHRYSONINI** Lacordaire, 1868

*Achryson* Audinet-Serville, 1834

*Achryson pictum* Bates, 1870

*Achryson pictum* Bates, 1870a: 248.

**Remarks.** – This species was described based on syntypes from Brazil (Pará) and French Guiana. Joly (2000) listed it from “Rio Santiago,”

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which is in Condorcanqui province (Peru), in the Amazonas Region. Currently, it is known from Colombia, Bolivia (Santa Cruz), Venezuela (Bolívar and Amazonas), Ecuador, Peru (Amazonas), French Guiana, Suriname, Brazil (Amapá, Amazonas, Pará, Roraima, and Maranhão) (Bezark, 2024; Monné, 2024a; Tavakilian & Chevillotte, 2024).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata NP, Bello Horizonte, 1 specimen, November 2023 (LGBC).

**EBURIINI** Blanchard, 1845

*Pantomallus* Lacordaire, 1868

*Pantomallus tristis* (Blanchard, 1847)

*Coccoderus tristis* Blanchard, 1847: 208.

**Remarks.** – This species was described based on syntypes from Bolivia. Currently, it is known from Peru (Junín), Bolivia (Pando, Santa Cruz, and La Paz), Brazil (Pará, Rondônia, Mato Grosso, Maranhão, and Espírito Santo), and Paraguay (Bezark, 2024; Monné, 2024a; Tavakilian & Chevillotte, 2024).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Sudadero, 5 specimens, January, 2023, local collector (LGBC); 9 specimens, February 2023, local collector (LGBC); 2 specimens, September 2023, local collector (LGBC); 4 specimens, October 2023, local collector (LGBC); Tambopata National Park, Bello Horizonte, 14 specimens, November 2023, local collector (LGBC); Tambopata National Park, Planchon, 1 specimen, March 2023, local collector (LGBC); 1 specimen, September 2023, local collector (LGBC).

*Susuacanga* Martins, 1997

*Susuacanga octoguttata* (Germar, 1821)

*Cerambyx octoguttatus* Germar, 1821: 168.

**Remarks.** – This species was described, apparently, based on a single specimen from Brazil (no further details). Currently, it is known from Colombia, Venezuela, Peru (Loreto, Junín, and Huánuco), Brazil (Amazonas, Pará, Acre, Rondônia, Maranhão, Mato Grosso, Mato Grosso do Sul, Goiás, Paraíba, Ceará, Bahia, Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina, and Rio Grande do Sul), Bolivia (Beni, Pando, and Santa Cruz), Paraguay, and Argentina (Misiones, Corrientes, Entre Ríos, Tucumán, and Salta) (Bezark, 2024; Monné 2024a; Tavakilian & Chevillotte, 2024).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Sudadero 1 specimen, January 2023, local collector (LGBC); 1 specimen, February 2023, local collector (LGBC); 1 specimen, October 2023, local collector (LGBC); Tambopata National Park, Planchon, 1 specimen, September 2023, local collector (LGBC).

**DICHOPHYIINI** Gistel, 1848

*Homogenes* Chevrolat, 1863

**Remarks.** – According to Chevrolat (1863: 747) (translated): “Mr. Lucien Buquet, *IC. du Règne animal de Cuvier*, volume II, p. 226, described, under the names of *Er. albolineatus* and *Leprieuri*, two species of which I make a division in the genus *Mallosoma*,” and on the page 757: “I have no new species to make known; only the *Eriphus Leprieuri* and *albolineatus* of M. Buquet establish for me a division (Genus *Homogenes* Th. Mss. [Thomson manuscripts]) in the genus *Mallosoma*. Although the name *Homogenes* was apparently based on unpublished manuscripts by James Thomson, the genus cannot be attributed to that author. According to Lacordaire (1869: 124) on *Mallosoma* (translated): “Mr. Chevrolat (Ann. d. l. Soc. entom. 1862) adds to these species [“*M. fuligeneum*, Newm. The Entomol. p. 7 (*thoracicum*, A. White, Longie. of the Brit. Mus. p. 111); Brésil. – *scutellare*, A. White, *ibid.* p. 110; meme pays”], by forming a separate section, a new species (*albolineatum*, Brazil) and *Eriphus Leprieuri* of Mr. Buquet (in Guérin-Méneville. Icon.; Ins. p. 226), from Cayenne, more, with doubt, *M. transversale*

of Dejean (loc. cit.), which is from Brazil. These three insects are very probably foreign to the genus.” Although the text by Lacordaire (1869) does not make it clear, *Mallosoma transversale* was doubtfully included in this genus, but not in the subgenus *Homogenes*. Furthermore, *Homogenes albolineatus* was described by Buquet (1844) and not by Chevrolat (1863). Lacordaire (1869) was probably mistaken because Chevrolat (1863) did not provide a bibliographic list for this species, while all the other species included by him in *Mallosoma* and *Homogenes* present references.

Although *Homogenes* has been attributed to Thomson in Chevrolat (e.g. Gemminger 1872; Monné & Giesbert 1992; Napp & Santos 1996; Monné 2024a; Tavakilian & Chevillotte 2024), the genus was erected by Chevrolat, as correctly reported by Aurivillius (1912), who considered it as a junior synonym of *Mallosoma* Audinet-Serville, 1834. Monné (2024a) listed: “*Homogenes* Thomson in Chevrolat, 1862d: 751.” However, the correct year is 1863 (Desmarest 1863; see also Monné *et al.* 2023).

*Homogenes* is both masculine and feminine gender. The determination of the gender of the genus cannot be attributed to Chevrolat (1863). Although he listed “*albolineatus*,” he did so in combination with *Eriphus*, the original combination of the species, and not with *Homogenes*. In Chevrolat’s text (1863) appears “*Mallosoma albolineatum*.” However, “*albolineatum*” is neuter gender. We consider that the determination of masculine gender was correctly adopted by Monné & Giesbert (1992) (ICZN 1999: 30.1.4.2).

*Homogenes albolineatus* (Buquet, 1844)

(Fig. 1)

*Eriphus albolineatus* Buquet, 1844: 226.

**Remarks.** – This species was described based on a single specimen from Brazil, without further details. According to Monné (2024a) it was described based on syntypes. However, there is no evidence that Jean Baptiste Lucien Buquet had more than one specimen when he described the species. He provided the measurements of a single specimen and did not provide information in the description indicating that he examined more than one specimen. According to Tavakilian & Chevillotte (2024), the holotype is a female. Currently, it is known from Brazil (Bahia, Espírito Santo, Rio de Janeiro, São Paulo, and Santa Catarina) (Bezark, 2024; Monné, 2024a; Tavakilian & Chevillotte, 2024).

**Material examined.** – BRAZIL, *Minas Gerais* (new state record): Parque Estadual do Rio Doce, 1 male (MZSP 61528), XI.2014, L. Migliore leg. (MZSP).

*Tobipuranga* Napp & Martins, 1996

*Tobipuranga ruficoxis* (Bates, 1870)

*Chrysoprasia ruficoxis* Bates, 1870b: 412.

**Remarks.** – This species was described based on syntypes from Brazil (Amazonas and Pará). Currently, it is known from Peru (Loreto), Ecuador, Brazil (Amazonas, Pará, Rondônia, and Mato Grosso), and Bolivia (La Paz and Santa Cruz) (Bezark, 2024; Monné, 2024a; Tavakilian & Chevillotte, 2024).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Sudadero, 2 specimens, September 2023, local collector (LGBC).

*Tobipuranga ybyra* Napp & Martins, 1996

*Tobipuranga ybyra* Napp & Martins, 1996: 205.

**Remarks.** – This species was described based on specimens from Brazil (Pará, Rondônia, Mato Grosso, Mato Grosso do Sul, Goiás, and São Paulo). It remained known only from those Brazilian states (Bezark, 2024; Monné, 2024a; Tavakilian & Chevillotte, 2023).

**Material examined.** – PERU (new country record), *Madre de Dios*: Tambopata National Park, Planchon, 2 specimens, September 2023, local

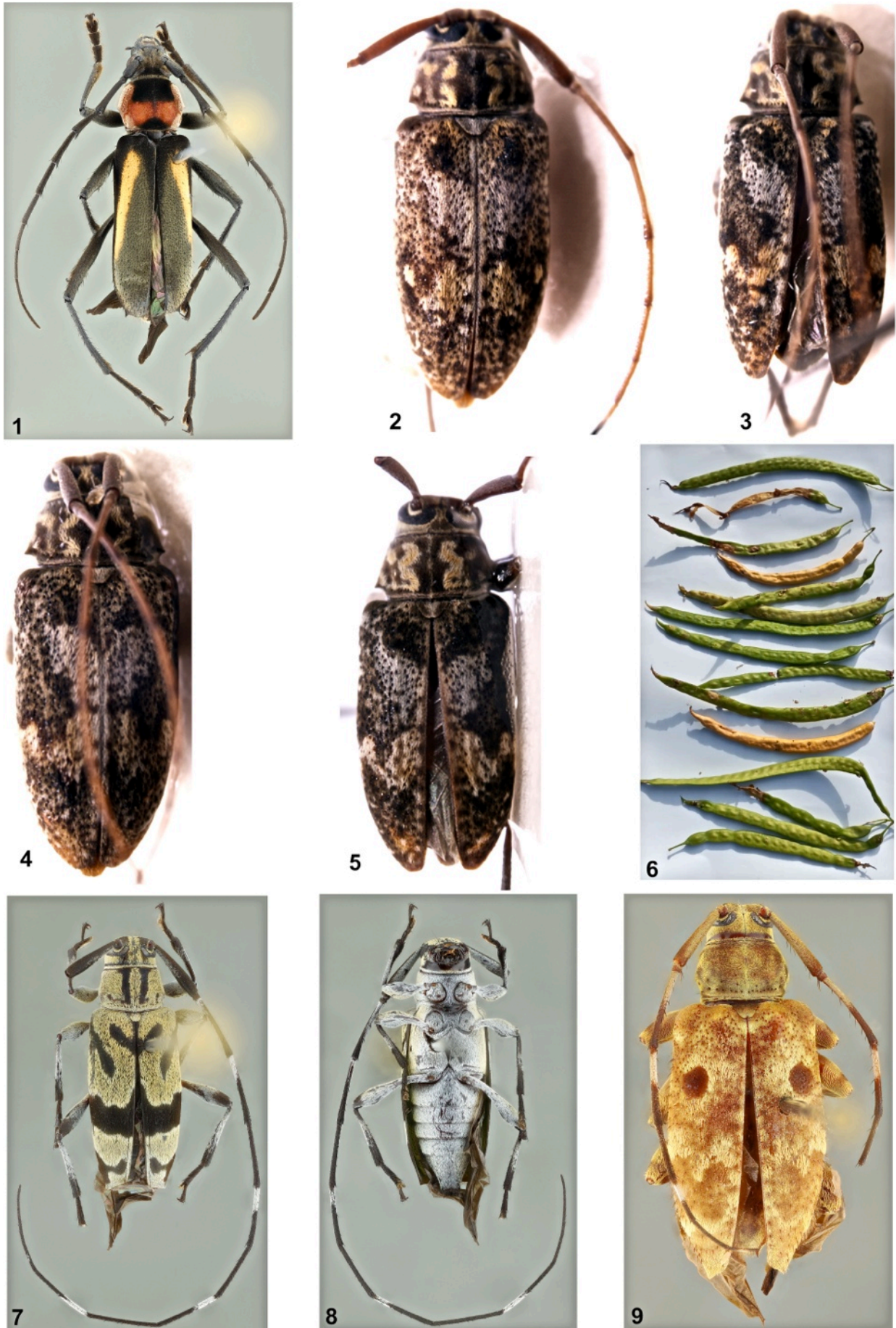


Fig. 1-9. Cerambycidae spp.

1. *Homogenes albolineatus* (Buquet, 1844), male (MZSP 61528), dorsal habitus. 2-5. *Atrypanius unpanus* Juárez-Noé & González-Coronado, 2021, dorsal habitus. 2. Specimen 1. 3. Specimen 2. 4. Specimen 3. 5. Specimen 4. 6. Fruits of *Prosopis pallida* (Humb. & Bonpl. ex Willd.) Kunth. 7-8. *Nyssodrysternum instabile* Monné, 1992, female (MZSP 61527). 6. Dorsal habitus. 7. Ventral habitus. 8. Ventral habitus. 9. *Oxathres guyanensis* Monné & Tavakilian, 2011, female (MZSP 61530), dorsal habitus.



collector (LGBC); 1 specimen, March 2023, local collector (LGBC); Tambopata National Park, Sudadero, 12 specimens, October 2023, local collector (LGBC); 16 specimens, September 2023, local collector (LGBC); Tambopata National Park, Bello Horizonte, 13 specimens, November 2023, local collector (LGBC); 1 specimen, January 2023, local collector (LGBC); Rio Huallaga, 1 specimen, January 1970, Peña leg. (LGBC).

#### LAMIINAE Latreille, 1825

#### ACANTHOCININI Blanchard, 1845

##### *Atrypanius* Bates, 1864

##### *Atrypanius unpanus* Juárez-Noé & González-Coronado, 2021

(Fig. 2-5)

*Atrypanius unpanus* Juárez-Noé & González-Coronado, 2021: 23.

**Remarks.** – This species was described based on one male and one female from Peru (Piura region). According to Juárez-Noé & González-Coronado (2021) it differs from *A. implexus* (Erichson, 1847) by “the continuous yellowish pronotal bands (forming four yellowish spots in *A. implexus*), the X-shaped grayish pubescent band not reaching the suture (reaching the suture in *A. implexus*) and the short yellowish subtransverse band on the apical third of elytra (without subtransverse [*sic*] band in *A. implexus*).” However, the innermost yellowish-brown pubescent maculae on the pronotum may be divided into two maculae (Fig. 2) or not divided (Figs 3–5). Furthermore, the large grayish-white pubescent area on the elytra never forms an X-shaped macula; in fact, it forms a V-shaped macula which reaches the suture and is sometimes projected forward adjacent to the suture.

**Host plant.** – The specimens were collected in fruits of carob, *Prosopis pallida* (Humb. & Bonpl. ex Willd.) Kunth (Fabaceae) (Fig. 6). The fruits were placed in recovery boxes where adults of *A. unpanus* were recovered.

**Material examined.** – PERU, *Tumbes* (new region record): “Bosque seco de algarrobo de la Asociación de Ganaderos el Charan,” La Cruz, 5 specimens, 4.IV.2024, 25.IV.2024, 2.V.2024, Javier Marchena Cruz & Marco Antonio Alemán Ramírez leg. (SENASA).

##### *Neopalame* Martins & Monné, 1972

##### *Neopalame albomaculata* Monné & Martins, 1976

*Neopalame albomaculata* Monné & Martins, 1976: 74.

**Remarks.** – This species was described based on males and females from Peru (Junín). Currently, it is known from Peru (Junín) and Brazil (Paraná) (Tavakilian & Chevillotte 2023).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Bello Horizonte, 2 specimens, November, 2023, local collector (LGBC).

##### *Nyssodrysternum* Gilmour, 1960

##### *Nyssodrysternum instabile* Monné, 1992

(Fig. 7-8)

*Nyssodrysternum instabile* Monné, 1992: 775.

**Remarks.** – This species was described based on males and females from Brazil (Espírito Santo, Rio de Janeiro, and São Paulo). Currently, it remains known only from those localities (Bezark 2024; Monné 2024b; Tavakilian & Chevillotte 2023). According to Monné (1992), there was a paratype female in the MZSP collection: “São Paulo: Peruíbe, 1 [symbol of female], 4.II.1943, Coll. H. Zelliger (MZSP).” Unfortunately, the specimen was never sent to the MZSP. Thus, the holotype and the 10 paratypes were destroyed during the fire that devastated the MNRJ.

**Material examined.** – BRAZIL, *Minas Gerais* (new state record): Itajubá, Reserva Biológica Municipal Serra dos Toledos, 1 female (MZSP 61527), 27.III–3.V.2018, Rosa *et al.* leg. (MZSP).

##### *Oxathres* Bates, 1864

##### *Oxathres guyanensis* Monné & Tavakilian, 2011

(Fig. 9)

*Oxathres guyanensis* Monné & Tavakilian, 2011: 23.

**Remarks.** – This species was described based on males and females from French Guiana and Brazil (Rondônia). Currently, it remains known only from those localities (Bezark 2024; Monné 2024b; Tavakilian & Chevillotte 2023).

**Material examined.** – BRAZIL, *Amazonas* (new state record): Manaus, AM10, km 16, 1 male (MZSP 61529), 16.IV.1988, R.A. Rocha leg. (MZSP). *Pará* (new state record): Santarenzinho (município de Itaituba), Rio Tapajós, 1 female (MZSP 61530), IV.1961 (MZSP, formerly Diringshofen collection).

#### ACANTHODERINI Thomson, 1860

##### *Acanthoderes* (*Acanthoderes*) Audinet-Serville, 1835

##### *Acanthoderes* (*Acanthoderes*) *thammi* Bates, 1880

*Acanthoderes thammi* Bates, 1880: 59.

**Remarks.** – This species was described based on syntypes from Peru (Junín). Currently, it is known from Peru (Junín), Brazil (Amazonas), and Bolivia (Beni and Santa Cruz) (Bezark 2024; Monné 2024b; Tavakilian & Chevillotte 2023).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Bello Horizonte, 2 specimens, November 2023, local collector (LGBC); Tambopata National Park, Planchon, 1 specimen, March 2023, local collector (LGBC); Tambopata National Park, Sudadero, 1 specimen, January 2023, local collector (LGBC); 1 specimen, February 2023, local collector (LGBC); 1 specimen, April 2023 (LGBC); 1 specimen, October 2023, local collector (LGBC).

##### *Aegomorphus* Haldeman, 1847

##### *Aegomorphus nigricans* (Lameere, 1884)

*Acanthoderes* (*Psapharochrus*) *nigricans* Lameere, 1884: 110.

**Remarks.** – This species was described based on syntypes from Venezuela. Currently, it is known from Guatemala, Honduras, Nicaragua, Costa Rica, Panama, Colombia, Peru, Venezuela, Ecuador, French Guiana, Brazil (Roraima, Rondônia, Mato Grosso, Rio Grande do Norte, Bahia, Minas Gerais, Rio de Janeiro, São Paulo, Paraná, Santa Catarina, and Rio Grande do Sul), Bolivia (La Paz and Santa Cruz), Paraguay, Argentina (Misiones, Corrientes, Entre Ríos, and Buenos Aires), and Uruguay (Tavakilian & Chevillotte 2023).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Sudadero, 1 specimen, January 2023, local collector (LGBC).

**Note.** – Carrasco (1978) reported this species from Peru. However, he did not state the region where the specimen (or specimens) was (or were) from. In the abstract, he only mentioned (translated): “This note reports the existence of 56 species of Cerambycidae (Coleoptera) recorded in different localities of the Departments of Cusco, Apurímac, Madre de Dios, and Arequipa, with representatives of three known subfamilies.” Therefore, we are considering the region of Madre de Dios as a new region record.

##### *Discopus* Thomson, 1864

##### *Discopus spectabilis* (Bates, 1861)

*Acanthoderes spectabilis* Bates, 1861: 264.

**Remarks.** – This species was described based on syntypes from Brazil (Amazonas). Bezark (2024) and Monné (2024b) reported it from Peru, without further details, and Brazil (Amazonas and Pará). Tavakilian & Chevillotte (2023) listed: Aurivillius (1909) / Peru:

Linguiyata; and Brèthes (1920) / “Pérou.” Chanchamayo. Linguiyata (the correct spelling of this place is Linquiata) is in the Region of Puno, and Chanchamayo in the Region of Junín. Therefore, currently it is known from Peru (Puno and Junín) and Brazil (Amazonas and Pará).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Sudadero, 1 specimen, January 2023, local collector (LGBC); 2 specimens, February 2023, local collector (LGBC); Tambopata National Park, Bello Horizonte, 3 specimens, November 2023, local collector (LGBC); Tambopata National Park, Planchon, 1 specimen, September 2023, local collector (LGBC).

*Symperasmus* Thomson, 1864

*Symperasmus thoracicus* (White, 1855)

*Acanthoderes thoracicus* White, 1855: 359.

**Remarks.** – This species was described based on a single specimen from Brazil (Pará). Currently, it is known from Peru (Loreto and Junín), French Guiana, Bolivia (La Paz), and Brazil (Amazonas, Pará, Rondônia, and Mato Grosso) (Bezark 2024; Monné 2024b; Tavakilian & Chevillotte 2023). Additionally, Aurivillius (1909) recorded this species from “Plabobamba” (Santa Cruz de Pablo Bamba), which is located in the Peruvian region of Puno.

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Sudadero, 6 specimens, September 2023, local collector (LGBC); 1 specimen, October 2023, local collector (LGBC); Tambopata National Park, Bello Horizonte, 3 specimens, November 2023, local collector (LGBC).

ANISOCERINI Thomson, 1860

*Cyclopeplus* Thomson, 1860

*Cyclopeplus peruvianus* Tippmann, 1939

*Cyclopeplus peruvianus* Tippmann, 1939: 77.

**Remarks.** – This species was described based on two specimens from Peru (Junín). Tippmann (1939) also described four aberrations of this species from the same place. He reported that the holotypes and paratypes were in his private collection. Monné (2024b) and Tavakilian & Chevillotte (2023) reported the existence of holotype for these aberrations. However, according to ICZN (1999: 45.6.2), aberrations are infrasubspecific. Therefore, they are not available and the holotypes have no value. Currently it is known from Peru (Junín, Ucayali, Huánuco, and Loreto).

**Material examined.** – PERU, *Madre de Dios* (new region record): Tambopata National Park, Planchon, 1 specimen, September 2023, local collector (LGBC). *Pasco* (new region record): Pozuzo, 1 specimen, November 2013, local collector (LGBC).

## Corrections in the species-group names

**Note:** In each species, the first species-group name is the one currently used; the second is the corrected one.

CERAMBYCIDAE

CERAMBYCINAE

CALLICHRMATINI Swainson, 1840

*Callichroma* Latreille, 1817

**Remarks.** – *Callichroma* is neuter gender because “χρόμα” (chroma, meaning “color”) is neuter. Therefore, one species-group name needs to be corrected to agree with the gender of the genus:

– *Callichroma minima* Podany, 1965 – *C. minimum* [“minima” is feminine gender].

*Mionochroma* Schmidt, 1924

**Remarks.** – *Mionochroma* is neuter gender (see remarks in *Callichroma*). Therefore, two species-group names need to be corrected to agree with the gender of the genus:

– *Mionochroma guyanensis* Juhel & Dalens, 2017 – *M. guyanense* [the suffix “-ensis”, meaning “of / from” is both masculine and feminine gender];

– *Mionochroma novella* (Bates, 1885) – *M. novellum* [“novella” is feminine gender].

*Monnechroma* Napp & Martins, 2005

**Remarks.** – *Monnechroma* is neuter gender (see remarks in *Callichroma*). Therefore, two species-group names need to be corrected to agree with the gender of the genus:

– *Monnechroma azurea* (Demets, 1976) – *M. azureum* (*azurea* is feminine gender);

– *Monnechroma tibialis* (Giesbert, 1987) – *M. tibiale* (“*tibialis*” is both masculine and feminine gender).

**Note.** – These two corrections already appear on Tavakilian & Chevillotte (2023).

*Plinthocoelium* Schmidt, 1924

**Remarks.** – *Plinthocoelium* is neuter gender because “*coelium*” is neuter. Therefore, two species-group names need to be corrected to agree with the gender of the genus:

– *Plinthocoelium chilensis* (Blanchard, 1851) – *P. chilense* [see remarks in *Mionochroma*];

– *Plinthocoelium domingoensis* (Fisher, 1922) – *P. domingoense* [see remarks in *Mionochroma*].

COMPSOCERINI Thomson, 1864

*Aglaoschema* Napp, 1994

**Remarks.** – *Aglaoschema* is neuter gender because “*schema*” (meaning “form”) is the transliteration of “σχῆμα” (ICZN 1985: Appendix B). Therefore, according to ICZN (1999: 30.1.2) it takes the gender given for that word in Greek dictionaries: neuter. Therefore, one species-group name needs to be corrected to agree with the gender of the genus:

– *Aglaoschema vinolenta* Dalens, Tavakilian & Touroult, 2010; – *A. vinolentum* [“*vinolenta*” is feminine gender].

PTEROPLATINI Thomson, 1861

*Deltosoma* Thomson, 1864

**Remarks.** – *Deltosoma* is neuter gender because “σῶμα” (soma, meaning “body”) is neuter. Therefore, two species-group names need to be corrected to agree with the gender of the genus:

– *Deltosoma humeralis* Dalens & Giuglaris, 2014 – *D. humerale* [“*humeralis*” is both masculine and feminine gender];

– *Deltosoma xerophila* Di Iorio, 1995 – *D. xerophilum* [“*xerophila*” is feminine gender].

TORNEUTINI Thomson, 1861

*Gigantotrichoderes* Tippmann, 1953

**Remarks.** – “*Deres*” is from the Greek (δέρη (*dére*), meaning neck, throat) and is feminine gender. Therefore, *Gigantotrichoderes* is feminine gender and one species-group name needs to be corrected to agree with the gender of the genus:

– *Gigantotrichoderes brunneus* Joly, 2017 – *G. brunnea* [“*brunneus*” is masculine gender].

**Note.** – This correction already appears on Tavakilian & Chevillotte (2023).



**TRACHYDERINI** Dupont, 1836**TRACHYDERINA** Dupont, 1836*Chlorotherion* Zajciw, 1962

**Remarks.** – “Therion” is from Greek (θηρίον (therion), meaning wild animal, beast) and is neuter gender. Therefore, *Chlorotherion* is neuter gender and one species-group name need to be corrected to agree with the gender of the genus:

– *Chlorotherion consimilis* Zajciw, 1962 – *C. consimile* [“*consimilis*” is both masculine and feminine gender].

**Note.** – This correction already appears on Tavakilian & Chevillotte (2023).

*Placoschema* Chemsak & Hovore, 2010

**Remarks.** – *Placoschema* is neuter gender (see remarks in *Aglaoschema*). Therefore, one species-group name needs to be corrected to agree with the gender of the genus:

– *Placoschema dimorpha* Chemsak & Hovore, 2010 – *P. dimorphum* [“*dimorpha*” is feminine gender].

**LAMIINAE****ACANTHOCININI** Blanchard, 1845*Alcidion* Sturm, 1843

**Remarks.** – “Idion” is a neuter diminutive suffix from Greek (-ιδιον). Therefore, *Alcidion* is neuter gender and three species-group names need to be corrected to agree with the gender of the genus:

– *Alcidion aestimabilis* (Melzer, 1934) – *A. aestimabile* [“*aestimabilis*” is both masculine and feminine gender];

– *Alcidion apicalis* (Bates, 1864) – *A. apicale* [“*apicalis*” is both masculine and feminine gender];

– *Alcidion albosparsus* (Melzer, 1934) – *A. albosparsum* [“*albosparsus*” is masculine gender].

**Note**<sup>1</sup>. – *Alcidion sulphurifer* (White, 1855) – this species group-name remains unchanged and does not need to agree with the neuter gender of *Alcidion* (ICZN 1999: 31.2.2). It is not possible to know whether this was an error by White (1855) or whether he used it as a noun in apposition.

**Note**<sup>2</sup>. – In Monné (2024b) appears *Alcidion humeralis* (Perty, 1832). However, the species-group name was already changed in Monné *et al.* (2020): *A. humerale*, as correctly registered by Sturm (1843), who transferred the species to *Alcidion*.

**AERENICINI** Lacordaire, 1872*Hydraschema* Thomson, 1864

**Remarks.** – *Hydraschema* is neuter gender (see remarks in *Aglaoschema*), as correctly indicated by Gemminger (1873). Therefore, four species-group names need to be corrected to agree with the gender of the genus:

– *Hydraschema cribripennis* Lane, 1966 – *H. cribripenne* [“*cribripennis*” is both masculine and feminine gender; Lane (1966) correctly used “*cribripenne*”];

– *Hydraschema fabulosa* Thomson, 1864 – *H. fabulosum* [“*fabulosa*” is feminine gender; Gemminger (1873) used “*fabulosum*”];

– *Hydraschema obliquevittata* (Lane, 1966) – *H. obliquevittatum* [“*obliquevittata*” is feminine gender];

– *Hydraschema petila* Martins & Galileo, 1998 – *H. petilum* [“*petila*” is feminine gender];

**Note.** – These species-group names already appear in Tavakilian & Chevillotte (2023) as neuter gender.

**APOMEYCYNINI** Thomson, 1860*Acrepidopterum* Fisher, 1926

**Remarks.** – “Pterum” is a neuter Greek adjective. Therefore, one species-group name needs to be corrected to agree with the gender of the genus, as was written by Vitali (2002) and ignored in subsequent catalogs and checklists:

– *Acrepidopterum jamaicensis* Fisher, 1942 – *A. jamaicense* [“*jamaicensis*” is both masculine and feminine gender].

**CALLIINI** Thomson, 1864*Callisema* Martins & Galileo, 1990

**Remarks.** – “Sema” is from Greek “σημα,” meaning “mark,” and is neuter gender. Therefore, one species-group name needs to be corrected to agree with the gender of the genus:

– *Callisema elongata* Galileo & Martins, 1992 – *C. elongatum* [“*elongata*” is feminine gender].

**COMPOSOMATINI** Thomson, 1857*Composoma* Lacordaire, 1830

**Remarks.** – *Composoma* is neuter gender (see remarks in *Deltosoma*). Therefore, one species-group name needs to be corrected to agree with the gender of the genus:

– *Composoma oculata* Lingafelter & Wappes, 2014 – *C. oculatum* [“*oculata*” is feminine gender].

**DESMIPHORINI** Thomson, 1860*Paroetropsis* Cerda, 1953

**Remarks.** – The Greek suffix “-opsis” is feminine gender. Therefore, the species-group name of the only species included in the genus needs to be corrected to agree with the gender of the genus:

– *Paroetropsis decoratus* Cerda, 1953 – *P. decorata* [“*decoratus*” is masculine gender].

**Note.** – This species-group name already appears correctly in Tavakilian & Chevillotte (2023) as feminine gender.

**ONCIDERINI** Thomson, 1860*Strioderes* Giorgi, 2001

**Remarks.** – “Deres” is neuter gender (see remarks in *Gigantotrichoderes*). Therefore, the species-group name of the only species included in the genus needs to be corrected to agree with the gender of the genus:

– *Strioderes peruanus* Giorgi, 2001 – *S. peruana* [“*peruanus*” is masculine gender].

**Note.** – This species-group name already appears in Tavakilian & Chevillotte (2023) as feminine gender.

**TETRAOPINI** Thomson, 1860*Mecasoma* Chemsak & Linsley, 1974

**Remarks.** – *Mecasoma* is neuter gender (see remarks in *Deltosoma*). Therefore, the species-group name of the only species included in the genus needs to be corrected to agree with the gender of the genus:

– *Mecasoma validicornis* (Bates, 1885) – *D. validicorne* [“*cornis*” is both masculine and feminine gender].

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- Publisher Correspondence.* – LB  
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*Material study.* – SS, LB  
*Author of the figures.* – SS (Fig. 1, 7-9), TH (2-6) & LB (cover)

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## Résumé

Santos-Silva A., Bezark L. G. & Tejada Hurtado G., 2024. – Nouveaux enregistrements de Cerambycidae (Coleoptera) d'Amérique du Sud, avec des notes sur la paternité dans le genre *Homogenes* et des corrections sur les noms de groupes d'espèces. *Faunitaxys*, 12(53): 1–8.

La paternité du genre *Homogenes* (Cerambycinae, Dichophyiini) est corrigée et un nouveau signalement du Brésil est fourni pour *Homogenes albolineatus* (Buquet, 1844). Deux nouveaux signalements du Pérou sont fournis ainsi que huit nouveaux signalements régionaux. Une plante hôte est signalée pour la première fois pour *Atrypanius unpanus* Juárez-Noé & González-Coronado, 2021 (Lamiinae, Acanthocini). De plus, *Nyssodrysternum instabile* Monné, 1992 est signalé dans l'état brésilien du Minas Gerais et *Oxathres guyanensis* Monné & Tavakilian, 2011 est signalé pour la première fois du Brésil. Vingt-six noms de groupes d'espèces sont corrigés dans les Cerambycinae et Lamiinae.

Mots-clés. – Coleoptera, Cerambycidae, longicorne, *Homogenes*, *Atrypanius*, *Nyssodrysternum*, *Oxathres*, taxonomie, Région néotropicale, Amérique du Sud.

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Nouveaux enregistrements de Cerambycidae (Coleoptera) d'Amérique du Sud, avec des notes sur la paternité dans le genre *Homogenes* et des corrections sur les noms de groupes d'espèces.

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***Illustration de la couverture :***

*A creek in the Osa Peninsula of Costa Rica.*

Crédits:

**Antonio Santos-Silva:** Fig. 1, 7-9.

**Graciano Tejada Hurtado:** Fig. 2-6.

**Larry G. Bezark:** couverture

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