1	Toll-like receptor 4 -
2	a multifunctional virus recognition receptor
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13 Abstract

Since the initial description of toll receptors in Drosophila and their mammalian counterparts 14 Toll-like receptors (TLRs), numerous fundamental and applied studies have explored their 15 crucial role as sensors of pathogen-associated molecular patterns (PAMPs). Among the ten 16 human TLRs, TLR4 is particularly well known for its ability to detect lipopolysaccharides (LPS). 17 a component of gram-negative bacterial cell wall. In addition to its archetypal functions, TLR4 18 is also a versatile virus sensor. This review provides a background on the discovery of TLR4 19 and how this knowledge laid a foundation for characterization of its diverse roles in antiviral 20 examined through genetic, biochemical, structural, and immunological 21 responses, approaches. These advances have led to a deeper understanding of the molecular functions 22 that enable TLR4 to orchestrate multi-nodal control by professional antigen-presenting cells 23 24 (APCs) to initiate appropriate and regulated antiviral immune responses.

25 **Keywords**: *TLR4*, viruses, antigen-presenting cells, immune response, polymorphisms

26 Highlights

- TLR4 is a single-pass transmembrane protein whose structure and function as a PRR are highly conserved in mammals
- TLR4 can be located on the cell surface and/or within cytoplasmic vesicles
- TLR4 participates in direct and indirect interactions with viruses and viral proteins to trigger antiviral responses
- In several instances, TLR4 polymorphisms can influence virulence
- TLR4 agonist are being used to improve vaccine efficacy
- Extracellular proteins, including antimicrobial peptides and danger associated molecular patterns (DAMPs) impact TLR-mediated antiviral responses

36 Milestones in toll-mediated immunity

Until the end of the 20th century, most immunologists focused on the adaptive arm of the 37 immune system. In 1989, Charles Janeway proposed that in addition to the interaction of 38 specific ligands with somatically rearranged antigen receptors, lymphocytes require another 39 signal from antigen-presenting cells (APCs) [1]. Janeway predicted that such signals must be 40 the result of the recognition of microbial molecules via germline encoded pattern recognition 41 receptors (PRRs), as opposed to the refined antigen receptors of the adaptive immune system. 42 Janeway posited that such microbial components should be i) absent from the host to 43 distinguish between self and non-self, ii) conserved among a large number of pathogens, and, 44 iii) essential components of the microbe's lifecycle, preventing escape from recognition 45 through mutations [1]. The identification of *toll* genes in Drosophila and Toll-like receptor genes 46 47 in mammals opened the gates to understanding an evolutionarily ancient arm of host defense, dubbed "innate immunity" [2,3]. Extensive research following the identification of TLRs 48 deciphered how pathogen-associated molecular patterns (PAMPs) such as bacterial, fungal, 49 viral nucleic acids, proteins or components induce signaling cascades that lead to inflammation 50 in vertebrate and invertebrate animals [4] (Figure 1). 51

52 Archetypal role in bacterial sensing

53 LBP, an extracellular LPS-binding protein, facilitates LPS extraction from bacterial outer 54 membrane forming an LPS-LBP complex. This complex is then recognized by soluble or membrane-bound CD14, which shuttles it to an MD-2-TLR4 complex, leading to TLR4 55 dimerization (Figure 2A) [5,6]. Mechanistically, the contact of lipid A, a hydrophobic 56 component of LPS, with MD-2 induces the formation of TLR4 homodimers, leading to a 57 multilayered signaling cascade that eventually induces NF-kB translocation to the nucleus and 58 the transcription of genes encoding pro-inflammatory cytokines (Box 1) [7]. In addition to LPS, 59 lipo-oligosaccharides and lipoteichoic acid (an anchor for peptidoglycans on some gram-60 positive bacteria) interact with the TLR4-MD-2 complex on the surface of human immune cells 61 [8–10]. 62

63 Genetic evidence of the impact on viral infections

At the turn of the century, Beutler and colleagues linked differences in responses to LPS to *TLR4* single nucleotide polymorphisms (SNPs) [11,12]. These polymorphisms, likely caused by selective pressure from pathogens, are found in the promoter, introns, and exons. Although detailed analyses have not been performed, it is likely that SNPs outside the coding regions modify TLR4 levels in APCs, while those within exons lead to changes in PAMP recognition [13–15] (**Figure 2B**). *TLR4* SNPs, such as RS⁴⁹⁸⁶⁷⁹⁰ and RS⁴⁹⁸⁶⁷⁹¹ cause an aspartic acid to

glycine substitution at position 299, and isoleucine to threonine at 399, respectively, which alter 70 the formation of the TLR4/MD-2 complex [16]. The first epidemiological study that identified 71 SNP-virulence links concerned severe forms of bronchiolitis caused by respiratory syncytial 72 virus (RSV) [17] (see Table 1 for a list of viruses and Table 2 for virulence-associated SNPs). 73 Of recent interest, SARS CoV-2 and RS⁴⁹⁸⁶⁷⁹⁰ or RS⁴⁹⁸⁶⁷⁹¹ were associated with cytokine 74 storms, possibly via increased spike-TLR4 interactions or dysregulation of the TLR4 signaling 75 cascade [18]. To add another layer of complexity, the impact of RS⁴⁹⁸⁶⁷⁹⁰ and RS⁴⁹⁸⁶⁷⁹¹, may 76 be due to dominance of a quanine or cytosine, as the final base in the respective codon [19-77 23]. In the case of human papillomavirus (HPV), transient TLR4 engagement leads to the 78 inhibition of the HPV genome integration, while chronic activation by high-mobility group box 1 79 protein (HMGB1), a by-product of HPV infection, may induce HPV-associated tumorigenesis 80 [24]. A study of Han women (the largest ethnic group in China), and women from Gujarat (one 81 of India's western states) showed that the risk of developing HPV-associated cervical cancer 82 was higher in carriers of RS⁷⁸⁷³⁷⁸⁴-G and RS¹⁹²⁷⁹¹¹, respectively [25,26]. 83

84 Direct or indirect interactions with TLR4/MD-2 complex

The interactions between viruses and TLR4 can be loosely divided into i) direct interactions 85 86 (Figure 2C), ii) interactions via a bridging protein, or iii) detection of viral components during propagation (Figure 3). Virus-receptor interactions are often context dependent, and therefore 87 drawing broad conclusions can be challenging because viruses tend to enter different cell 88 types using different receptors and mechanisms. Considering this, one needs to take into 89 account the possibility that virus can use multiple modes of entry, and therefore blocking one 90 pathway may have a modest impact on some readouts. Another caveat is that studies have 91 tended to focus on TLR4-virus interactions at the cell surface, while downplaying TLR4 92 interactions in endosomal vesicles. 93

94 Direct interactions

95 **Coronavirus** (e.g., SARS-CoV-2, NL63, 229E, OC43, and HKU1)

The immunopathology of COVID-19, is due, in part, to exacerbated expression of 96 97 proinflammatory cytokines like interleukin-6 (IL-6) and TNF, which are prototypic products of TLR activation. Using in silico studies, Choudhury & Mukherjee suggested that the SARS-CoV-98 2 spike glycoprotein has stronger interactions with TLR4 compared to TLR1 or TLR6 99 (respective binding energy values of -120 vs -57 or -68 kcal/mol) in docked protein complexes 100 [27]. Interestingly, high valued antigenic epitopes were at the interacting interface of the spike 101 protein and the TLR-binding region. Affinity-based interactions using surface plasmon 102 resonance were consistent with spike interacting with TLR4 with a K_d of 300 nM [28]. 103

A handful of labs explored the impact of spike protein on the transcriptome of TLR4-expressing 104 human and rodent APCs [28–31]. RT-qPCR assays showed that SARS-CoV-2 spike induces 105 TLR4-associated transcription. The key assays included the use of TAK-242 or siRNA 106 knockdown of mouse TLR4. In addition to transcriptional responses, biochemical assays 107 showing modification of TNF, phosphorylated NF-κB p65 and JNK p54 levels also supported 108 the conclusion that spike activates TLR4 [30]. In addition to the TLR-4-mediated response in 109 APCs, Li et al. found that seasonal coronaviruses (229E or OC43) seeded an NLRP3 110 inflammasome [32]. An inflammasome is a multiprotein platform containing a PRR that 111 promotes aggregation of ASC (apoptosis-associated speck-like protein containing a caspase 112 activation and recruitment domain). ASC recruits and auto-activates pro-caspase-1, which can 113 be followed by protease cleavage of the N terminus of gasdermin D, which initiates pore-114 formation leading to the loss of plasma membrane integrity [33] (Figure 2D). Canonical 115 NLRP3-based inflammasome formation is typically preceded by a transcriptional priming event 116 117 that produces mRNAs of inflammasome components and cytokines [34]. However, in human monocytes, LPS-TLR4 engagement induces an alternative inflammasome activation 118 characterized by IL-1ß release in the absence of a transcriptional priming event, inflammasome 119 aggregation, and pyroptotic cell death [33]. Li et al. concluded that 229E- and OC43-TLR4 120 priming induced inflammasome activation in PMA-differentiated THP-1 cells (where TAK-242 121 inhibited this cascade) [32]. 122

While Zhao et al. used THP-1 cells (a human monocyte cell line) to identify SARS-CoV-2 spike-123 TLR4 interactions (presumably in liaison with CD14 and MD-2 on the cell surface) and 124 125 signaling [28], van der Donk et al. proposed more nuanced conclusions [35]. Donk et al. found that primary human monocyte-derived dendritic cells (moDCs) respond to SARS-CoV-2 126 nanoparticles (analyzing IP10 & IL-8 protein levels, IL6, IL10, IFNb mRNA, and phenotypic 127 readouts) only when infected by SARS-CoV-2 via ectopically expressed ACE2. These data led 128 129 to the conclusion that innate immune sensing in some cells occurs via cytosolic TLRs [35]. In addition, van der Donk et al. also suggested that SARS-CoV-2 suppresses TLR4-induced 130 immunity via DC-SIGN (a C-type lectin receptor on DCs) via the Raf-1 pathway [36]. Finally, a 131 SARS-CoV2 spike-TLR4 axis was also challenged by Khan et al., who found that all 132 inflammatory pathways were still activated in TIr4^{-/-} murine bone marrow-derived macrophage 133 [37]. Importantly, neither TAK-242 nor TLR4 blocking antibodies were used in these latter 134 studies. Given the differences in species (mouse vs human), cell lines vs. primary cells, 135 agonists (spike proteins, nanoparticles, pseudotyped virions), and readouts, distilling a 136 simplified conclusion is unwarranted. 137

Chikungunya is a disease caused by the Chikungunya virus (CHIKV). CHIKV is transmitted 138 by Aedes mosquitoes, and therefore referred to as an arbovirus (arthropod-borne virus). 139 CHIKV can bind to cells by using several cell surface proteins including Eps15, TIM-1, DC-140 SIGN, PHB1 & 2, and Mxra8, which may contain glycosaminoglycans as post-translational 141 modifications. Using in vivo and in vitro murine models, as well as human PBMCs, Mahish et 142 al. showed that TAK-242, anti-TLR4 Abs, or knockout of TLR4 prevented CHIKV-induced 143 increase in inflammatory cytokine production (TNF, IL-6, MCP-1) and reduced activation of 144 p38, JNK and NF-kB signaling pathways and cell maturation, leading to decreased CHIKV 145 infection and prolonged survival of mice [38]. To complement the functional data, Mahish et al. 146 also used molecular docking to propose eleven polar interactions between the extracellular 147 domain of TLR4 (from aa 454-592) and CHIKV E2 protein (primarily between aa 303-315). 148

Respiratory syncytial virus (RSV): Upon infection by RSV, syncytia (large multinucleated "cell-like structures") arise due to the effect of RSV fusion protein (F) causing neighboring cell membranes to merge. More than 20 years ago, Kurt-Jones et al. reported TLR4 involvement in RSV sensing [39] by using recombinant F protein and neutralizing antibodies binding F protein in TLR4^{-/-} and CD14^{-/-} mice [39]. Using *in silico* molecular docking simulations, Akagawa *et al.* proposed that TLR4 binding affinity increases during pre- to post-fusion conformational changes [40].

156 Indirect TLR4 interactions via bridging molecules

One complex challenge faced when exploring virus-receptor interactions is modelling and/or understanding an *in vivo* environment. Relatively simplified *in vitro* assays, using cultured cells in defined media, poorly take into account the impact of extra-cellular factors that could interact with viral particles and influence binding and uptake.

161 The Adenoviridae family contains greater than 300 different types [41]. Much of the impetus for characterizing adenovirus-induced immune responses has been their use as gene transfer 162 vectors and vaccines [42]. An initial clue that some adenovirus types interact with TLR4 came 163 from Amalfitano and colleagues [43]. They used transgenic mice to identify TRIF, and TRIF-164 interacting TLRs, that differentially modulate vector-induced immune responses. In a 165 syngeneic approach, Doronin et al. show that factor X (FX) binding to some human adenovirus 166 types leads to activation of the TLR4/Myd88-TRIF pathway in murine macrophages [44]. Using 167 modified capsids that do not bind FX, they concluded that FX acts as a bridging molecule 168 between the capsid and TLR4 to induce an innate immune response. Paradoxically, human 169 adenovirus type 5, a benchmark of adenovirus-induced immunogenicity, poorly induces human 170 DC maturation and cytokine production [45]. 171

In addition to coagulation factors, some adenovirus types interact with antimicrobial peptides 172 (AMPs). AMPs are innate immune effector molecules that can also promote the maturation of 173 APCs and amplify or synergize with innate responses [46]. Neutrophils are a rich source of 174 AMPs and, following a breach in tissue homeostasis, release their cytoplasmic content which 175 can be up to 20% AMPs. Lactoferrin and α -defensins are alarmins (a subset of AMPs) that 176 also impact innate and adaptive immune responses by activating PRRs in APCs [47]. In 177 epithelial cells, α -defensing impairs human adenovirus type 5, 12, and 35 infection by 178 stabilizing their vertices, thus preventing the disassembly of the capsid in endocytic vesicles 179 [48]. In two studies, Eichholz, Cheneau, Tran et al. found that lactoferrin and human neutrophil 180 defensin 1 bind, with micro- to nanomolar affinity, to the capsid of adenovirus type 5, 26, and 181 182 35. Then, via a bridging mechanism, the AMPs bind to TLR4 on the surface of human APCs 183 to facilitate AdV uptake [33,49]. Downstream of the adenovirus-AMP interactions with TLR4, the APCs undergo maturation via the induction of an NLRP3-associated inflammasome. 184

185 Detection of secreted viral proteins

Similar to bacteria, virus propagation often involves active or passive release of intracellular
 pathogen components into the extracellular environment. Consequently, viral proteins that may
 not be exposed on the surface of viral particles can still act as TLR4 agonists.

The five **Dengue virus** (DENV) serotypes cause a spectrum of disease. During propagation, a polyprotein precursor of ~3,400 aa is cleaved into three structural (C, prM and E) and seven nonstructural proteins (NS1, 2A & B, 3, 4A & B, and 5). In 2015, Modhiran *et al.* demonstrated that NS1, released from infected cells as soluble lipid-associated hexamer, activates TLR4 on the surface of endothelial cells to increase vasculature leakage [50]. In addition to the interaction with APCs, NS1 appears to be linked with thrombocytopenia by triggering TLR4mediated platelet activation [51].

There are six *Ebolavirus* (EBOV) species: Bundibugyo, Reston, Sudan, Taï Forest, Bombali, 196 and Zaire, with the latter causing the largest number of outbreaks and highest mortality rate. 197 EBOVs are filamentous-shaped particles with genomes that encode seven proteins: NP 198 (nucleoprotein), VP30, -35, -40 & 42 (matrix proteins), L (an RNA-dependent RNA 199 polymerase), and GP (which encodes two glycosylated spike proteins). NP, L, VP30 and -35 200 form the nucleocapsid containing the genome. During an active EBOV infection, GP, the key 201 protein in EBOV virulence, is shed from both cells and virions (the EBOV envelope is decorated 202 by the membrane-bound trimeric GP [52]) due to cleavage by ADAM17 (A disintegrin and 203 metalloprotease 17). Due to reading errors during transcription and error-prone post-204 translational processes due to seven uracil residues, three GP variants are produced: i) full-205

length type I transmembrane GP (pre-GP1,2), ii) a furin-cleaved pre-GP that creates GP1 and 206 GP2 to form a heterodimer that assembles into the trimeric 450-kDa spike at the surface [53], 207 and iii) a dimeric pre-shed GP, which is generated by "correct" transcription and post-208 209 translational processes. Shed GP (sGP) molecules diffuse through the bloodstream and can activate TLR4-expressing cells at distal locations [54]. sGP interacts directly with TLR4/MD-2 210 211 through the hydrophobic internal fusion loop to activate the signaling pathways leading to a storm of pro-inflammatory and antiviral cytokines [55]. Harty and colleagues used EBOV-like 212 particles, in which they could selectively incorporate proteins, and analyzed cytokine 213 production from human cell lines to screen for an interaction with TLR4 [56]. 214 Immunoprecipitation assays suggested an interaction between GP and TLR4, which was 215 consistent with TLR4-associated signaling cascade. Of note, the 150-residue mucin-like 216 domain of GP1.2 is heavily glycosylated, which may be associated with its ability to induce 217 cytokine storms and increase virulence [56]. Volchkov and colleagues found that extensive 218 enzymatic-based deglycosylation decreased GP-induced cytokine production [57], a finding 219 however recently challenged by Scherm et al. using non-glycosylated mutants of GP [55]. 220 Using mice and recombinant GP as a ligand, Lai et al. also found a direct involvement of GP 221 in TLR4 activation [58]. 222

Similar to EBOV, **Marburg virus** can cause hemorrhagic fever. Younan *et al.* showed that the TLR4 antagonist eritoran prevented the death of mice following challenge with EBOV and Marburg virus via reduced pro-inflammatory cytokine production and decreased systemic levels of infiltrating neutrophils and CD4⁺ and CD8⁺ T cells [59]. Eritoran also protects EBOVinfected mice by reducing the inflammatory effects of oxidized phospholipids, which interfere with maturation of DCs, resulting in limited lymphocyte activation.

229 Cellular environments and cellular partners

230 Cellular environments

Although TLR4 is canonically found on a subset of immune myeloid cells, some tissue-resident 231 epithelial cells express low levels of TLR4 on their surface, which may contribute to organ and 232 233 tissue immunosurveillance. Bulk transcriptome analyses detect **TLR4** mRNA in many organs and tissues [60], and data suggest that some tissues and cell types modulate TLR4 expression 234 levels in pathophysiological conditions [61]. Of note, the pattern of TLR4 mRNA and protein 235 236 expression in situ parallels those of CD14 and LY96 (MD-2). TLR4 location and levels are relevant when considering the cellular environment and the impact on host inflammation and 237 viral infections. As such, TLR4 engagement by PAMPs or DAMPs can induce a MyD88-238 dependent inflammatory response, as well as a MyD88-independent antiviral environment, 239

both of which impact host immune responses and virulence [62]. Massive engagement of TLR4
 complexes should lead to a deleterious response (cytokine storm), while a fine-tuned TLR4 mediated IFN-dependent response favors the host by abrogating virus propagation [63].

When TLR4 engagement increases virulence, then strategies to prevent TLR4 signaling might be beneficial [63,64]. For example, blocking TLR4 signaling prior to infection by Tembusu virus (TMUV), a flavivirus causing egg drop disease syndrome in birds, led to replication inhibition [65]. Also, HTLV-I protein p30 may directly impact TLR4-dependent host immune tolerance or anergy by interfering with activation and expression in macrophages [66]. Additionally, Wilks and colleagues showed that mouse mammary tumor viruses (MMTV) can incorporate TLR4 into their envelope, which leads to an LPS-dependent enhancement of transmission [67].

250 Extra- and intracellular partners

An underappreciated aspect of TLR4 versatility is its different subcellular locations. While the 251 recognition of PAMPs by TLR4 certainly occurs at the cell surface, TLR4 can also traffic 252 through endosomal compartments to initiate interactions or signaling events that regulate 253 antiviral or pro-viral pathways via structural and spatiotemporal parameters [35,68]. The TLR4 254 complex can also interact with extracellular alarmins, leading to the amplification of 255 surrounding inflammation upon viral infections. Indeed, alarmins are passively or actively 256 secreted upon cellular stress or viral infections and can interact with inflammatory receptors 257 such as TLR4 or RAGE [69]. Interestingly, viral infections leading to severe inflammatory 258 disorders often rely on uncontrolled and excessive release of alarmins, like HMGB1 or 259 members of the S100 proteins family (which may aid as biomarkers of disease severity) [69]. 260 This is the case of viral myocarditis caused by Coxsackievirus B3 and for which HMGB1 and 261 262 S100A8/S100A9 are involved [70]. Similarly, acute respiratory disease caused by RSV, SARS-CoV1/2, or influenza virus A is associated with elevated levels of HMGB1 [71] and/or 263 264 S100A8/S100A9 [72], which can exacerbate TLR4-mediated cytokine production. There is therefore a clinical interest to better detect and control the alarmin-TLR4 axis upon virus-265 induced inflammatory disorders. Therefore, it is not only the TLR4-virus interaction, but also 266 associated elements and parameters such as TLR4 localization and conformation, protein 267 partners, and antigen presentation, that impact the inflammatory response. 268

269 Impact on T- and B-cell responses

270 TLR4 and host antiviral cellular immunity

TLR4-mediated immune responses are intimately linked to the adaptive immune system of vertebrates. The involvement of TLR4 in adaptive immune responses was thought to mainly rely on the adjuvant capacity of TLR4 activation that enhances functional maturation and migratory capacities of APCs [73]. It became clear that TLR4-mediated control of CD4⁺ T cell priming impacts inflammatory cytokines expression and overcoming suppressive effects from regulatory T cells [74]. Interestingly, the strength of TLR4 stimulation might differentially regulate the profile of T helper 1 cells (T_{h1}) towards T_{h2} responses, which could become deleterious in a context of viral infection [75]. Furthermore, the engagement of TLR4 with natural or engineered ligands influences adaptive immunity against HIV-1 [76,77] and LCMV [78] infections.

Importantly, a better understanding of the dynamics between viral infections, TLR4 complex, 281 and host cellular immunity, will help identify strategies to enhance antiviral immunity and design 282 effective vaccines. TLR4 targeting is a potent option in vaccine development strategies against 283 viral infections. Promising ex vivo and in vivo data suggest that TLR4 engagement potentiates 284 285 humoral and cellular immune responses upon HBc-AdV-B7-VLPs immunization [79], and confers resistance to HBV [80], and Marek's disease virus [81] infections - as well as inducing 286 robust mucosal immune responses [82] and even improving immunotherapeutic approaches 287 [83]. 288

289 TLR4 and host antiviral humoral immunity

The interaction between TLR4 and B-lymphocyte responses is multifaceted. A seminal study 290 by Pasare and Medzhitov reported that the addition to CD4⁺ T-cell help, the generation of T 291 cell-dependent antigen-specific antibody responses require TLR4 activation in B cells [84]. 292 Similarly, TLR4 is an intermediate immune effector between host innate and adaptive immune 293 responses against RSV [39]. This was demonstrated in a study uncovering that optimal specific 294 antibody responses were dependent on CD4⁺ T-cell help and triggering of TLR4 and TLR7 on 295 296 B cells and DCs [85]. Indeed, immunization with nanoparticles containing Yellow fever virusvaccine, plus ligands signaling through TLR4 and TLR7, protected birds from lethal avian, 297 298 swine influenza virus strains in mice, and induced robust immunity against pandemic H1N1 influenza in rhesus macaques [85]. 299

These studies highlighted the concept that viral infections and virus-derived vaccines can 300 301 leverage TLR4-mediated signaling to enhance B-cell activation and promote the production of protective antibodies. This was indeed the case in the context of HBc-HAdV7-VLPs 302 immunization [79], Enterovirus-71 virus-like particles [86], and adenovirus-based anthrax 303 304 vaccine [87]. A recent study showed that inflammatory signals upon immunization with influenza A virus + LPS functioned as B-cell fate-determinants for the rapid generation of 305 protective antiviral extrafollicular responses [88]. TLR4-B-cell receptor co-engagement also 306 induces T cell-independent hypermutated, class-switched, neutralizing antibody responses 307

- which could also be beneficial for antiviral vaccines [89]. Noteworthy, TLR4 effect on B-cell responses could also be indirect and cell-mediated as concluded by Chen *et al.* when finding that the recruitment of conventional type 2 DCs into secondary lymph organs initiated follicular helper T differentiation, thus promoting the proliferation of germinal center B cells and the
- 312 production of RABV-specific IgGs and neutralizing antibodies [90].

313 Concluding remarks

TLR4 has yet to reveal all its functional secrets. Future work will need to i) characterize the impact of SNPs on virulence, ii) explore its involvement in detecting other viruses (either directly, indirectly or via bridging factors), iii) untangle the in vivo role DAMPs versus PAMPs plays in inducing an antiviral environment, iv) identify the mechanism by which viral proteins activate a MyD88-dependent or -independent response, and v) determine the most effective ways to stimulate TLR4 with adjuvants in vaccines to generate broad and long-term T_{h1} responses against emerging viruses, thus helping to tackle future pandemics [91].

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544

546 Figure 1. toll and the Toll-like receptors milestones

547 In the early 1980's Nüsslein-Volhard and colleagues found that the toll is involved in the differentiation of the dorso-548 ventral axis of the drosophila embryo. In 1989, Janeway posited the "pattern recognition hypothesis," suggesting 549 that the immune system distinguishes between self and non-self through the recognition of conserved molecular 550 patterns on pathogens through PRRs. In the mid-1990's, Hoffmann and colleagues showed that toll mutant flies 551 succumb rapidly to challenge with Aspergillus fumigatus, an opportunistic fungus. The first mammalian Toll-like gene was described in 1997. Shortly thereafter, Beutler and colleagues demonstrated that TLRs carry immune 552 functions, linking them to the sensing of PAMPs. Beutler and colleagues showed that mice that harbor missense 553 mutations in TLR4 are unable to sense LPS. A year later, further generation and characterization of knockout mutant 554 555 mice for TLR4 by Akira and colleagues showed that mammalian TLRs induce NF-κB-dependent responses to 556 different PAMPs. In the decade that followed, the "innate immunity" field accelerated the pace of discovery, as the 557 microbial ligands of several mammalian TLRs were identified. This decade included the seminal study by Kurt-Jones and colleagues showing that TLR4 is involved in the detection of RSV. The Nobel prize in Physiology and 558 559 Medicine in 2011 "for the discoveries concerning the activation of innate immunity" recognized not only fundamental research, but also its far-reaching implications concerning vaccine design against infectious agents and 560 development of therapies against cancer and immune-related diseases. Parallel to functional data, the atomic 561 structure led to a molecular understanding of the different modes of PAMP binding to the ectodomains of TLRs, 562 563 their effect on the conformation of the cytoplasmic TIR domain, and subsequent signaling cascades. Over the last 10 years, TLR4 interactions with viruses (including Dengue, Ebola, SARS-CoV-2 and adenovirus) have helped us 564 565 understand virulence and pathogenicity.



566 Figure 2. TLR4/MD-2 interactions and their antagonists



567

568 **A)** The archetypal role of TLR4: LPS detection is initiated by LBP and then LPS is transferred to membrane-bound 569 CD14, prior to engaging the TLR4-MD2 complex and inducing TLR4 dimerization, which activates the TIRAP-570 MyD88 signaling cascade. This cytoplasmic signaling pathway (from IRAK4-p65) eventually induces the 571 translocation of NF-κB and the expression of proinflammatory cytokines and, in some cases, **D)** inflammasome 572 components (e.g., NLRP3, caspase-1, ASC).

573 B) While TLR4-virus (e.g., DENV) interactions also activate the TIRAP-MyD88 signaling cascade, the bivalent TRAM-TRIF pathway also leads to the upregulation of type-I IFNs upon IRF3 activation, which may be more relevant 574 for antiviral responses. The formation of an inflammasome leads to caspase-1 autoactivation, cleavage of pro-575 GSDMD to GSDMD to create oligomeric pores, and cleavage of pro-IL1 β to release IL-1 β via the pores. 576 Pharmacological inhibitors (in red) can be used to identify and inhibit TLR4 interactions or the downstream signaling 577 cascade. TAK-242 is a cyclohexane derivative that efficiently inhibits the TLR4 pathway. TAK-242 binds cysteine 578 residue 747, located intracellularly, and prevents TIRAP and TRAM engagement. Other TLR4 antagonists include 579 acetylated LPS and LPS-RS from 0111:B4, which bind to the hydrophobic pocket of MD-2. Similarly, eritoran 580 tetrasodium is a lipid A analog that binds to MD-2, blocking the interaction of LPS with the TLR4-MD-2 complex 581 582 and subsequent dimerization and activation. OxPAPC competes with LPS-binding protein (LBP), CD14, and MD-2. Finally, pepinh-TRIF interferes with TLR4-TRIF binding. 583

C) Single nucleotide polymorphisms (SNPs) in *TLR4*, located in the enhancer/promoter, coding, or noncoding regions, can influence TLR4 expression levels, its conformation, and binding partners. As examples, RS⁴⁹⁸⁶⁷⁹⁰ (blue) and RS⁴⁹⁸⁶⁷⁹¹ (orange) are illustrated because their locations in the coding region have been linked to altered virulence of some microbes.

588 Box 1. Divergent versus conventional pathways

589 TLR4-mediated signaling can transit through two distinct pathways involving TIRAP/MyD88 and TRIF/TRAM (TICAM1/2) adaptor proteins, which induce the production of proinflammatory cytokines and antiviral type-I interferon responses respectively. A simplistic interpretation suggests that the conventional TLR4-TIRAP-MyD88 pathway leads to uncontrolled NF-κB-dependent inflammation, while the TLR4-TRIF-TRAM pathway initiated through endosomal trafficking leads to IRF3-dependent transcription of antiviral genes.

594 The TLR4-TIRAP-MyD88 pathway

Upon ligand binding to TLR4, MyD88 is recruited via the bridging factor TIRAP (Mal) to the TLR4 TIR domain which 595 associates and transiently activates dynamic and concerted signalling through factors like interleukin-1 receptor-596 597 associated kinase 4 (IRAK1) and IRAK1/2, nucleating the formation of a supramolecular organizing center (SMOC) 598 coined "myddosome". The myddosomes then co-ordinate TLR-mediated inflammatory response upon association 599 with TNF receptor-associated factor 6 (TRAF6). Then, polyubiquitination allows TRAF6 to activate the Ikk complex 600 and transforming growth factor-β-activated kinase 1 (TAK1) leading to the activation and nuclear translocation of 601 NF-KB and AP-1 transcription factor subunits Jun/Fos and subsequent induction of expression of proinflammatory cytokines and other immune response-related genes. Conventional TLR4-mediated signaling pathway has been 602 most often tagged for its implication in virus-associated severe pathogenesis whether due to genetic regulations or 603 604 polymorphisms of myddosome-associated factors [92], or to a direct effect of viral proteins on this pathway [93].

605 The TLR4-TRIF-TRAM pathway

TLR4 can also drive a MyD88-independent TRAM/TRIF-mediated pathway upon endocytosis and alternative dimerization state of the receptor for which CD14 is essential. The involvement of the TRIF-related adaptor molecule (TRAM) is essential to recruit the adapter protein TRIF to the TLR4-TIR domain and form another SMOC called "Triffosome". The particularity of the TRIF-dependent pathway relies on its connection to other signaling kinases like TANK-binding kinase 1 (TBK1) and inducible IkB kinases (IKKα, IkKβ and Ikkγ) leading to the phosphorylation and nuclear translocation of the activated interferon regulatory factor 3 (IRF3) required to induce type-I IFN thus establishing an antiviral state [94].

613 **Overlapping pathways**

These pathways are not mutually exclusive and some viruses impact both (e.g., Tat of HIV-1 [120]) or induce non-614 615 canonical or alternative pathways: e.g., VSV glycoprotein-G induces a TRIF-independent IFN-I response in DCs 616 and macrophages, and SARS-CoV M protein promotes IFN-β response in immortalized epithelial cells in a TRAF3independent manner. Interestingly, because TRAM and TRIF may regulate the NF-KB pathway and control pro-617 inflammatory cytokines expression upon TLR4 stimulation, this pathway appears central in fine-tuning the amplitude 618 of inflammatory and antiviral responses. As such, the TLR4-TRIF-TRAM pathway needs to be tightly regulated as 619 620 reported for cellular factors like Sterile Alpha and TIR Motif Containing 1 (SARM1), or TRAM adaptor with GOLD domain (TAG). Given its critical role in establishing the antiviral response, this pathway is also impacted during viral 621 infections as shown for hCMV, HCV [95] or the vaccinia virus [96]. 622

The balance between TLR4-mediated proinflammatory and antiviral signaling is crucial to mount effective host defense, but also to limit virus-driven immunopathologies caused by excessive inflammation. It remains to better determine the mechanisms at play upon viral infections leading to selective modulation of TLR4-mediated signaling hubs as well as potential viral components able to modify one pathway over the other and therefore impacting innate and adaptive antiviral responses. In this context, Bonham *et al.* concluded that the promiscuity of lipid binding of the sorting adaptor TIRAP could explain the diversity in subcellular localization of TLR4 signaling hubs and virus detection via endosomal TLRs [97].

630 Table 1. Glossary of viruses

Genus	Family	Genome*	~size (kb)	Structure
Adenovirus	Adenoviridae	ds DNA	28 to 42	non-enveloped
Chikungunya virus	Togaviridae	+ss RNA	11.7	enveloped
Coxsackievirus B3	Picornaviridae	+ss RNA	7.3	non-enveloped
Dengue virus	Flaviviridae	+ss RNA	11	enveloped
Ebolavirus	Filoviridae	-ss RNA	19	enveloped
Enterovirus A71	Picornaviridae	+ss RNA	7.5	non-enveloped
Foot & mouth disease virus	Picornaviridae	+ss RNA	8.3	non-enveloped
Human T-lymphotropic virus 1	Retroviridae	+ss RNA	9	enveloped
Hepatitis B virus	Hepadnaviridae	ds DNA	3.2	enveloped
Hepatitis C virus	Flaviviridae	+ss RNA	9.6	enveloped
Hepatitis E virus	Hepeviridae	+ss RNA	7.2	non-enveloped
Human cytomegalovirus	Herpesviridae	ds DNA	235	enveloped
Human immunodeficiency virus 1	Retroviridae	2 copies +ss RNA	9.7	enveloped
Human papillomavirus	Papillomaviridae	ds DNA	8	non-enveloped
Influenza H1N1	Orthomyxoviridae	segmented -ss RNA	13.5	enveloped
Marburg virus	Filoviridae	-ss RNA	19	enveloped
Marek's disease virus	Orthoherpesviridae	ds DNA	177	enveloped
Mouse mammary tumor virus	Retroviridae	+ss RNA	10	enveloped
Murine norovirus	Caliciviridae	+ss RNA	7.5	non-enveloped
Rabies virus	Rhabdoviridae	-ss RNA	12	enveloped
Respiratory syncytial virus	Pneumoviridae	-ss RNA	15.2	enveloped
SARS-CoV-2, NL63, 229E, OC43 & HKU1	Coronaviridae	+ss RNA	26 to 32	enveloped
Tembusu virus	Flaviviridae	+ss RNA	10.9	enveloped
Yellow fever virus	Flaviviridae	+ss RNA	11	enveloped

*ds = double stranded, ss = single stranded, "+" = positive sense, "-" = negative sense

Table 2. Examples of TLR4 Single nucleotide polymorphisms (SNPs) in selected populations that impact
virulence

SNP	Population	Virulence	Viruses	References
RS ⁷⁸⁷³⁷⁸⁴	Chinese	1	HPV	[25]
RS ¹⁰⁷⁵⁹⁹³²	Chinese	7	EV71	[98]
	Indian	4	HPV; DENV; HIV	[21,99–101]
RS ⁴⁹⁸⁶⁷⁹⁰	Egyptian	~	SARS-CoV2	[18]
	Caucasian		HIV	[99]
	Saudi Arabian		HCV	[102]
	Tunisian		HCV; HBV	[103]
	Polish		HCMV	[23]
	Indian		HEV; DENV	[21,22]
	Saudi Arabian		HCV	[102]
RS ⁴⁹⁸⁶⁷⁹¹	Egyptian		SARS-CoV2	[18]
	Mexican		DENV	[20]
	Polish		HCMV	[19]
RS ¹⁹²⁷⁹¹¹	Indian	1	HPV	[26]
RS ¹¹⁵³⁶⁸⁶⁵	Mexican	×	DENV	[20]
RS ²⁷³⁷¹⁹⁰	Mexican	X	DENV	[20]

- **Figure 3.** Indirect and direct TLR4 engagement
- A schema of the means by which TLR4 is engaged by viruses: the left panel depicts direct detection of intact SARCoV-2, the middle panel depicts the detection of an adenovirus capsid via an AMP bridge (lactoferrin in this case), and the in the right panel the detection of viral proteins.

