

A temporal perspective for tumor-associated macrophage identities and functions

Camille Blériot, Garett Dunsmore, Direna Alonso-Curbelo, Florent Ginhoux

▶ To cite this version:

Camille Blériot, Garett Dunsmore, Direna Alonso-Curbelo, Florent Ginhoux. A temporal perspective for tumor-associated macrophage identities and functions. Cancer Cell, 2024, 42 (5), pp.747-758. 10.1016/j.ccell.2024.04.002 . hal-04729136

HAL Id: hal-04729136 https://hal.science/hal-04729136v1

Submitted on 31 Jan 2025 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

A temporal perspective for tumor-associated macrophage identities 1 and functions 2 3 4 Camille Blériot^{1,2}, Garett Dunsmore¹, Direna Alonso Curbelo^{3*} and Florent Ginhoux^{1,4,5,6*} 5 6 ¹Gustave Roussy, INSERM, Villejuif, France ²Institut Necker des Enfants Malades (INEM), INSERM, CNRS, Université Paris Cité, Paris, France ³Institute for Research in Biomedicine (IRB Barcelona), The Barcelona Institute of Science and Technology (BIST), Barcelona, Spain 7 ⁴Singapore Immunology Network (SIgN), Agency for Science, Technology and Research 8 (A*STAR), Singapore 9 ⁵Shanghai Institute of Immunology, Shanghai JiaoTong University School of Medicine, 10 Shanghai, China 11 ⁶Translational Immunology Institute, SingHealth Duke-NUS, Singapore 12 13 14 15 16 *Correspondence: 17 Florent Ginhoux, Ph.D. 18 E-mail : florent.ginhoux@gustaveroussy.fr 19 Institut Gustave Roussy, INSERM U1015, Bâtiment de médecine moléculaire, 114 rue 20 Edouard Vaillant, 94800 Villejuif, France Tel: +33-142115531 21 22 23 24 Direna Alonso-Curbelo, Ph.D. 25 E-mail: direna.alonso-curbelo@irbbarcelona.org 26 Institute for Research in Biomedicine (IRB Barcelona), The Barcelona Institute of 27 Science and Technology, 08028 Barcelona, Spain. 28 Tel: + 34-934037162

1 Abstract

2 Cancer is a progressive disease that can develop and evolve over decades, with 3 inflammation playing a central role at each of its stages, from tumor initiation to 4 metastasis. In this context, macrophages represent well-established bridges 5 reciprocally linking inflammation and cancer via an array of diverse functions that 6 have spurred efforts to classify them into subtypes. Here, we discuss the 7 intertwines between macrophages, inflammation, and cancer with an emphasis on 8 temporal dynamics of macrophage diversity and functions in pre-malignancy and 9 cancer. By instilling temporal dynamism into the more static classic view of tumorassociated macrophage biology, we propose a new framework to better 10 11 contextualize their significance in the inflammatory processes that precede and 12 result from the onset of cancer and shape its evolution.

1 Introduction

2 Perturbations of tissue homeostasis, such as infection or injury, induce a cascade 3 of events resulting in redness, warmth, swelling, and pain that is known as 4 inflammation^{1,2}. Physiologically, inflammation is a protective response of the immune 5 system that facilitates the elimination of the triggering insult and initiates tissue repair. However, when the nature, intensity, and/or duration of this response is altered, it can 6 7 lead to a wide range of pathologies, including heart disease, autoimmune disorders and cancer^{3,4}. Accordingly, cancer commonly arises at inflamed tissue sites, and inflammation 8 9 remains a primary risk factor for tumor development, independent of sexual dimorphism, 10 lifestyle, or exposure to specific mutagens^{5,6}.

11 Among the myriad of cell types involved in physiological and pathological 12 inflammatory responses, the most abundant cell type is often macrophages. In the context 13 of cancer, these are commonly referred to as tumor-associated macrophages (TAMs)^{7,8} 14 to differentiate them from resident tissue macrophages (RTMs) present in normal, nontumoral tissues^{9,10}. TAMs have been long known to be associated with tumor 15 16 development and generally exhibit pro-tumoral functions^{11,12}. However, it is becoming 17 clear that this "TAM" denomination fails to fully reflect the dynamic heterogeneity of 18 macrophage populations^{13,14}. Indeed, single-cell and lineage-tracing technologies have 19 exposed how TAM biology is inherently complex, with the co-existence of distinct 20 subpopulations with a differential ontogeny, spatial tissue localization, and cancer stagedependent functions that evolve over time¹⁵⁻²¹. Moreover, even before the onset of cancer 21 22 initiation, macrophages within inflamed (or previously inflamed) normal tissues can 23 modulate the susceptibility of epithelial cells to subsequent neoplastic transformation by 24 instructing them with an epigenetic memory of inflammation²².

Despite their significance as bridges between inflammation and cancer pathogenesis, the current "TAM" nomenclature only formally refers to macrophages present in *established* tumors, and currently no distinction is made between these and those previously present in the tissue from *before* tumor onset. As both types of macrophages can have different ontogenies and exposure to tissue signals, two wellestablished determinants of macrophage heterogeneity discussed below, an updated framework that integrates the time axis into the biology of TAMs is needed to better

conceptualize their roles orchestrating the inflammatory responses that often precede,
 result from, and fuel cancer.

3 Herein, we discuss recent advances in immunology and oncology that, combined, shape our current vision of TAM biology. We propose a new perspective that captures 4 5 temporal aspects of macrophage evolutionary histories within normal, pre-malignant and malignant tissues, to better describe how their physiological functions are co-opted in 6 cancer²³⁻²⁶ and expose means to interfere with their disease-promoting roles more 7 8 selectively. We argue for a distinction between intra-tumor macrophages that resided in 9 the tissue before cancer onset (PreTAMs), which connect inflammation with cancer 10 initiation and are further reprogrammed within the TME of developing tumors, and 11 macrophages that are recruited after their establishment (TAMs), whose later arrival 12 imposes distinct interactions and features influencing their fate and functions (Figure 1). 13

14 Heterogeneity of macrophages in normalcy, premalignancy, and cancer

Historically, macrophage identities within normal and tumor contexts have been reduced to two antagonist types of 'activation' or polarization states: M1 considered as pro-inflammatory, and M2 as anti-inflammatory^{27,28}. While such classification was convenient for explaining distinct transcriptional and functional states identified *in vitro*, it has become evident that this dichotomy is insufficient to reflect the many levels of macrophage heterogeneity observed *in vivo*, across distinct anatomical sites, tissue niches, and disease contexts^{9,29,30}.

22 A well-established determinant of macrophage heterogeneity is ontogeny, which largely varies according to tissues²⁹. During embryogenesis, fetal macrophage precursors 23 24 seed different tissues and give rise to RTMs, for example, microglia in the brain or Kupffer cells in the liver³¹⁻³⁴. Although RTMs can be maintained by local self-renewal during 25 adulthood^{35,36}, tissues such as the gut lamina propria or the dermis can postnatally recruit 26 27 bone marrow-derived monocytes which also give rise to RTM populations from a different ontogeny^{37,38}. During physiological and pathological inflammation, such as during tissue 28 29 damage or ageing, macrophage tissue diversity is further reprogrammed, with an 30 increase in monocyte-derived macrophages, which can be short-lived or long-lived and

have context-dependent effects that deserve deeper investigation (see³⁹ for an extended
review on macrophage dynamics in normal tissues).

3 Beyond ontogeny, macrophages are highly plastic cells and their identity is further 4 shaped by tissue signals from their niche of residence or arrival that can largely remodel their phenotype, gene expression, and chromatin architecture⁴⁰. For example, within the 5 liver, studies suggest that macrophage identity is influenced by local, sub-tissular 6 interactions with fibroblasts, endothelial cells, or lymphocytes⁴¹⁻⁴³. In the lung, alveolar 7 8 macrophages differ from interstitial macrophages, and even among these lung interstitial 9 macrophages, perivascular and perineural subpopulations have been distinguished with 10 different proximities to endothelial cells and smooth muscle cells, or nerve bundles and endings^{44,45}, respectively 11

12 As in healthy tissues, ontogeny and tissue environment also contribute to the 13 remarkable intra- and inter-tumor heterogeneity of macrophages observed in cancer. 14 Initially, the assumption was that TAMs derived from circulating monocytes that are continuously recruited in response to tumor-induced inflammation⁴⁶. While subsequent 15 16 research has confirmed that monocyte-derived cells often represent the majority of TAMs⁴⁷⁻⁴⁹; these cells co-inhabit the TME with macrophages that resided in the tissue 17 18 before tumor onset. Importantly, embryonic-derived RTMs and adult monocyte-derived 19 TAMs exhibit distinct functions. For example, in a murine model of pancreatitis and 20 pancreatic cancer, embryonic RTMs are potent promoters of fibrotic responses⁵⁰ whereas 21 adult monocyte-derived counterparts regulate adaptive immune responses⁵¹. 22 Regardless, neither ontogeny nor intra-tumoral location of macrophages explain when 23 and how they came to be "TAMs", as monocyte-derived macrophages with TAM-like and 24 tumor-promoting phenotypes can be recruited to tissues before the formation of neoplastic lesions, such as during injury-induced pancreatic metaplasia^{52 53}. State-of-the-25 art fate-mapping⁵⁴, temporally-resolved single-cell profiling⁵⁵⁻⁵⁷ and innovative strategies 26 27 are needed to better define the temporal histories and "birthdates" of distinct types of 28 macrophages within normal, inflamed, pre-malignant and malignant tumor lesions.

Once within a tumor microenvironment, TAMs are influenced by interactions with their neighboring cells that are largely dependent on their precise spatial location within the tumor lesion. For example, specific TAM subpopulations defined by a set of markers

1 and locations have been identified, such as FOLR2⁺ macrophages in perivascular areas of primary breast tumors⁵⁸ or TREM2⁺ macrophages in tumor nests of liver, lung, and 2 3 lymph node metastases⁵⁹. Nevertheless, these identities are not definitive, as TAMs display a high level of plasticity and are influenced by interactions with tumor and non-4 tumor cells that evolve during disease progression (see⁶⁰ for a recent perspective on this 5 topic). As an example, pre-clinical models of hepatocellular carcinomas show that TAMs 6 7 undergo a shift to states reminiscent of embryonic development, termed "oncofetal 8 reprogramming⁶¹⁻⁶³; yet when tumor cells undergo senescence, they can acquire antitumor states that facilitate CD8 T cell-mediated immunity⁶⁴. 9

10 Therefore, origin and environment influence TAM biology, and the relative 11 contribution of each of these determinants is an age-old debate between nature and 12 nurture⁶⁵⁻⁶⁷. We argue that, in addition to these two determinants, time should be 13 integrated into the "TAM" classification framework to reflect the continuum between 14 inflammation and cancer (which starts well before bona fide tumor establishment), and 15 better apprehend how temporal aspects shaping their diversity impact pre-malignant and 16 malignant evolution. Of note, this paradigm shift has begun with the RTM characterization 17 over the last decade and we envision such evolution will also be fruitful in cancer studies. 18

19 A new proposal for defining TAM heterogeneity: time as the protagonist

20 The initiation of cancer can be relatively slow and silent, with genetic mutations, 21 epigenetic aberrations, and/or chronic inflammation being present in tissues even years before the emergence of a full-blown cancer⁶⁸⁻⁷⁰. Yet, the current TAM denomination is 22 23 limited to macrophages present in the tissue *after* the establishment of tumor lesions, 24 which is arguably a late event in the pathogenesis of the disease and obliterates the early 25 steps of cancer development. Indeed, macrophages play key roles in early transitions 26 from tissue normalcy to metaplasia, its reprogramming into pre-malignant neoplastic 27 lesions, and subsequent malignant and metastatic evolution. Moreover, macrophages 28 lifespan can vary from days to months, and even years – with long-lived subpopulations, such as microglial cells, lasting on average 4.2 years old⁷¹. Therefore, considering only 29 30 TAMs as tumor-inhabiting cells without considering their lifespan, history of prior

exposures, and functions prior to tumor onset provides an incomplete picture of their
 relevance interconnecting inflammation and cancer.

3 To emphasize such early roles, we propose to incorporate a prefix to distinguish 4 macrophages pre-existing in the tissue before tumor onset (PreTAMs), with "Pre" to 5 emphasize they were previously present in the tissue (irrespective of their specific 6 ontogeny) and "TAM" to underline their significance as cellular constituents and drivers 7 of the resulting tumors. As such, PreTAMs would link pre-existing inflammation to cancer 8 pathogenesis, whereas remaining TAMs recruited from circulating monocytes after tumor 9 formation play the reciprocal role linking back this process to tumor-elicited inflammation, 10 in collaboration with PreTAMs that are further reprogrammed within the TME. Of note, 11 this denomination is only based on the time dimension and agnostic of ontogeny, as 12 PreTAMs can be either of embryonic or adult origin, the fundamental point being their 13 presence from before tumor onset.

14 The PreTAM distinction sheds light on an essential yet ill-defined component of 15 macrophage biology: their memory of previous challenges. Pioneering studies in the 16 infectiology field have revealed that macrophages, as other innate immune cells, can 17 keep a "memory" of previous immunogenic challenges and mount a more efficient 18 response to subsequent ones, even from unrelated pathogens⁷²⁻⁷⁴, a phenomenon coined 19 as trained immunity⁷⁵. At the cellular level, this process involves enhancer 20 reprogramming⁷⁶, 3D chromatin topology reorganization⁷⁷, and metabolic rewiring⁷⁸. At a population level, it may also involve changes in the cellular origin of macrophages within 21 22 a given tissue. For example, embryonic alveolar macrophages can be replaced by adult 23 monocyte-derived ones during lung infection and acquire similar phenotypes yet with 24 enhanced protective capacities⁷⁹. Interestingly, while in the context of an infection, such 25 "memory" can increase the resistance of the tissue to new challenges and, as such, be 26 harnessed to enhance trained immunity, increasing evidence suggests this can be 27 associated with the induction of altered inflammatory responses that can both promote or 28 restrain tumor progression⁸⁰⁻⁸².

The underlying mechanisms and implications of time-dependent macrophage features as determinants of cancer susceptibility and evolution are, however, still far from being completely understood. It is tempting to speculate that PreTAMs will be influenced

by ontogeny-related and environment-induced scars of "tissue history" by insults that can be recent or transient or long-established and chronic, depending on the temporal scales linking inflammation and tumorigenesis. Regardless, we envision that time-dependent features will influence macrophage diversity within tissues and disease evolution by various mechanisms that represent open questions to address in future studies.

6 Finally, in addition to the concepts of TME dynamism and macrophages as 7 "guardians of tissue memory", the time parameter is inexorably linked to aging, the top risk factor for cancer development⁸³. As one mechanism, chronic inflammation is 8 9 considered a hallmark of aging⁸⁴. In parallel, aged epithelial cells accumulate (epi)genetic alterations that prime them for clonal expansions⁸⁵ that appear to be unleashed when 10 exposed to inflammatory insults^{86,87}. In parallel, long-lived RTMs or hematopoietic stem 11 12 cell precursors of TAMs can, themselves acquire age-associated alterations that can be directly pathogenic⁸⁸⁻⁹⁰ as observed in neurodegeneration phenotypes driven by 13 BRAF^{V600E} mutation in microglia⁹¹. 14

Therefore, both long-standing and more recent observations coincide in how macrophage biology is highly time-dependent, a notion which should be considered when approaching their relevance as bridges between inflammation and cancer. This constitutes the basis of our proposal of incorporating a temporal mark that distinguishes macrophages that participated in inflammatory processes that preceded (PreTAM) and not only resulted from the tumor development process (TAMs) (**Figure 2**).

21

22 **PreTAM** and **TAM** heterogeneity in cancer initiation, progression and metastasis

23 Pre-existing and tumorigenesis-induced macrophage heterogeneity modulates cancer

24 *initiation*

Cancer initiation is driven by cooperation between genetic and environmental inflammatory insults⁹²⁻⁹⁶. As limiting the exposure to such insults may be challenging, gaining insights into how they impact PreTAMs may expose strategies to prevent or intercept links between inflammation and cancer pathogenesis. In this section, we will present how diverse cancer-initiating insults, such as exposure to environmental pollutants, tissue damage, infection, and genetic mutations, shape PreTAM diversity and functions. Indeed, while appropriately activated, macrophages can recognize and

eliminate damaged and mutant cells and protect the host from tumor development⁹⁷, but
 these physiological functions can be reprogrammed by cancer-initiating insults to trigger
 tumor-promoting inflammation and cancer development.

4

4 Recent perspectives highlight the contribution of non-mutagenic mechanisms of environmental carcinogens to cancer pathogenesis^{93,98}. For example, pollutant 5 particulates can promote cancer through inducing IL-1ß production by macrophages⁹², a 6 7 well-established macrophage pro-inflammatory mechanism targetable by anti-8 inflammatory therapy showing protection against lung cancer incidence and mortality in 9 a randomized clinical trial^{99,100}. Similarly, cigarette smoke triggers a basal level of lung inflammation by modulating the inflammatory state of lung alveolar macrophages¹⁰¹, 10 including IL-1ß release¹⁰², further supporting IL-1ß targeting as a means to prevent 11 12 and/or intercept lung carcinogenesis⁹⁹. Also in the lung, exposure to asbestos triggers 13 apoptosis of otherwise long-lived alveolar macrophages, resulting in the infiltration of 14 monocytes and their differentiation into alveolar macrophage-like cells¹⁰³. Recruited macrophages persist and further enhance inflammation within the fibrotic lesions of 15 16 asbestos-treated mice¹⁰⁴. These responses can also be activated by toxins such as aflatoxin B1, which can activate macrophages directly¹⁰⁵, as well as by triggering 17 18 apoptosis of parenchymal cells and their phagocytosis by macrophages¹⁰⁶. When these 19 inflammatory responses lead to tumors, these ontogenically-distinct populations 20 "educated" within tissues exposed to carcinogens would be considered PreTAMs.

21 Additional non-mutagenic insults such as tissue damage can also unleash tumor-22 promoting inflammation through the rewiring of both RTMs and monocyte-derived 23 macrophages, or their relative abundance. For example, in the pancreas, inflammation 24 triggered by tissue damage, usually referred to as pancreatitis, induces metaplasia and markedly accelerates the development of pancreatic ductal adenocarcinoma in mice¹⁰⁷ in 25 a manner dependent on the presence and functions of macrophages^{52,108,109}. Tumor-26 27 initiating inflammation can also be triggered by chronic viral infections such as Human Papilloma Virus (HPV) or Human Immunodeficiency Virus (HIV)⁵⁷. Despite the lifelong 28 29 treatment of HIV and preventing patients from progressing to acquired immunodeficiency 30 syndrome (AIDS), systemic inflammation is sustained throughout the life of an HIV 31 patient¹¹⁰. Even in antiretroviral therapy-treated patients, viral reservoirs exist in various

tissues including the gastrointestinal tract¹¹¹. As a consequence of these persistent viral 1 2 reservoirs, macrophages are constitutively engaged in inflammatory responses, which 3 impair intestinal barrier function causing bacterial product translocation into the blood¹¹². 4 Beyond such systemic immune activation, the immunosuppression of HIV patients can 5 lead to Kaposi sarcomas driven by co-infection with human herpesvirus 8 (HHV8)¹¹³; and which progress by complex mechanisms involving chemokine homologues encoded by 6 7 the virus and downregulation of the macrophage chemokine scavenger ACKR2 in tumor 8 cells, resulting in cancer-related inflammation by TAMs and monocyte recruitment¹¹⁴. The 9 relative contribution of PreTAMs and TAMs remains to be fully deciphered.

10 Beyond environmental cues, genetic alterations known to predispose to cancer 11 can also create tumor-permissive and -promoting niches by modulating macrophage 12 states directly or indirectly. As an example, TAMs from BRCA1-deficient breast tumors are reprogrammed and can directly mediate immune suppression¹¹⁵. In addition, highly 13 metastatic inflammatory breast cancers (IBC) with BRCA1/2 mutations have a 14 characteristic inflammatory milieu suggested to alter TAM states in favor of cancer 15 16 dissemination^{116,117}. In a different disease context, a phenotypically distinct TME has been noted in BRCA1/2 mutated high-grade serous ovarian cancers as compared to BRCA1/2 17 18 wild-type counterparts, which exhibit a lower abundance of macrophages¹¹⁸. Finally, in 19 thyroid carcinomas, PTEN-inactivating mutations in tumor cells have been associated 20 with a specific macrophage phenotype with high IL-6, TNF- α , IL-8 and MCP-1 expression downstream of hyperactivated mTOR signalling¹¹⁹. It is also tempting to speculate that 21 22 other cancer-predisposing inflammatory disorders associated with Human Leukocyte Antigen (HLA) polymorphisms¹²⁰ involve TAMs, given the specific high expression of HLA 23 24 antigens by macrophages. Indeed, HLA encodes Major Histocompatibility Complex 25 (MHC) molecules that are responsible notably for the communication of the innate 26 immune system such as macrophages and dendritic cells with T cells through antigen 27 presentation. The HLA-B27 expression has been notably associated with an increased risk of inflammatory bowel disease and colorectal cancer development^{121,122}. 28

Overall, it is becoming clear that PreTAMs and TAMs act as key cellular effectors of environmental and genetic cancer-initiating insults. While precise mechanisms, as well as the lifespan and fate of PreTAMs and TAMs within metaplastic, pre-malignant and

malignant tissues need to be further clarified, these examples underline the importance
of PreTAMs as critical players in cancer pathogenesis.

3

4

<u>Tumor-induced rewiring of PreTAMs and TAMs promotes disease progression</u>

5 Once tumors are formed, malignant cells require access to nutrients and complex 6 adaptations to meet their metabolic growth demands. One such adaptation is the 7 promotion of angiogenesis, with TAMs playing a pivotal role by secreting effector 8 molecules such as Vascular Endothelial Growth factor (VEGF) or matrix metalloproteinases^{123,124}. Failure to induce a sufficient blood supply will result in nutrient 9 deprivation and hypoxia in the tumor core which will turn necrotic¹²⁵, a phenomenon that 10 can also lead to inflammation and disease progression¹²⁶⁻¹²⁸. In this context of tumor 11 12 necrosis, macrophages perform one of their most well-described functions: phagocytosis 13 of debris and dying cells, with CD206⁺ macrophages being recognized as the most actively phagocytic populations¹²⁹. Interestingly, CD206⁺ TAMs have also been shown to 14 cross-present tumor antigens to CD8⁺ T cells in tumors^{130,131}, highlighting a complex 15 16 interplay between dynamic inflammatory responses within evolving tumors, 17 macrophages, and adaptive immunity.

18 Our understanding of the crosstalk between myeloid and lymphoid cells in the TME 19 has significantly increased since the appreciation of the process of cancer immunoediting 20 two decades ago⁹⁷. It is well appreciated that interferon gamma produced by innate (NK, ILC1s) and adaptive (CD8⁺ T) immune cells induces a pro-inflammatory phenotype of 21 22 macrophages, which is associated with anti-tumor activities^{132,133}. Such knowledge has 23 led to the design cellular backpacks filled with interferon gamma as a means to prime and 24 maintain anti-tumor responses in TAMs¹³⁴. However, as tumors evolve, macrophage–T 25 cell direct interactions are dynamically reprogrammed and most often ultimately dampen 26 anti-tumor immunity. Consistently, PreTAMs and TAMs can have differential capacities to instruct adaptive responses^{58,61,135}, with the latter promoting recruitment of regulatory 27 T-cells (Treas), for example in ovarian¹³⁶ or liver cancer¹³⁷, and/or promoting CD8 T-cell 28 exhaustion^{138,139}. Therefore, while immunotherapy protocols based on reactivating 29 30 cytotoxic activities of T cells have shown some impressive success in several cancer types¹⁴⁰, a better understanding of when and how PreTAMs and tumor-recruited TAMs 31

enhance vs suppress adaptive immunity is warranted to better understand their dual
potential as igniters of tumor-promoting inflammation and anti-tumor immunity, and tilt the
balance towards the latter.

Beyond their immunomodulatory roles, another targetable trait of macrophages 4 that is reprogrammed in established tumors is their metabolism¹⁴¹⁻¹⁴⁴. While Otto Warburg 5 had described a century ago the higher consumption of glucose by tumor cells¹⁴⁵, the 6 metabolic rewiring of macrophages within TME has been more recently reported^{146,147}. 7 8 yet its specific impact on the course of the disease remains still poorly characterized. For 9 example, an upregulation of REDD1 in hypoxic TAMs has been shown to promote angiogenesis¹⁴⁸ while enhanced tryptophan degradation in interleukin-4-induced-1 10 (IL4I1)⁺ macrophages promotes tumor progression¹⁴⁹. Understanding whether different 11 12 macrophage subpopulations differing in their ontogeny, specific tissue contexts, and/or 13 time of pre-existence in the tumor tissue, are differentially "primed" for such diverse tumor-14 promoting functions will be an important step for predicting and interfering with tumor 15 progression. This will require the use of precise preclinical cancer models allowing for 16 differential targeting of PreTAMs and TAMs in a spatiotemporally-controlled manner.

17

18 Macrophage diversity in primary tumors and secondary localizations shape metastasis

As the ultimate expression of tumor evolution, metastasis defines the process by which tumor cells disseminate away from the originating tumor to ultimately produce a new one in nearby or distant site^{150,151}. While metastasis requires cancer cells to acquire cell-intrinsic properties necessary for cell migration, invasion, immune evasion, survival in circulation and colonization of the new site, it also involves a complex interplay with TME cells that can favor or restrain each of these processes in a non-cell-autonomous manner. Heterogeneous functions of TAMs participate in all these different steps.

In many solid tumors, migration and dissemination of cancer cells involve cellular plasticity, such as full or hybrid epithelial to mesenchymal cell (EMT) transitions, a process which is facilitated by TAMs acquiring transcriptional states that drive secretion of EMT inducers, such as TGF- β^{152} . Interestingly, EMT transitions can happen very early during tumor development, are exacerbated by inflammation¹⁵³, and involve TAMs^{153,154}. Accordingly, spatial single-cell analyses reveal inflammatory cells in proximity with

transitioning mesenchymal-like cells¹⁵⁵. Considering that different subpopulations of TAMs (such as PreTAMs) organize differentially within the TME of primary tumors^{58,153} and that EMT can be restricted to the leading edge or inflamed areas, functional studies involving targeting of specific TAM subpopulations will provide valuable insights into microenvironmental determinants of cancer dissemination, and the influence of macrophage ontogeny, tissue location, and time as regulators of this process (**Figure 3**).

7 Beyond changes in the primary tumor ecosystem, the "seed and soil" theory 8 highlights how a given tumor will preferentially metastasize to predefined tissues "permissive" to this metastasis outgrowth¹⁵⁶. Interestingly, this theory emerged 9 10 concomitantly with the first description of macrophages, and these two seminal 11 discoveries have evolved in parallel for decades but without conceptual relationship^{157,158}. 12 Today, immune cells are recognized as key building components of what has been defined as the pre-metastatic niche¹⁵⁹. Indeed, a seminal study has highlighted that 13 14 specific bone marrow progenitors were homing to pre-metastatic sites and were facilitating arrival of disseminating cancer cells¹⁶⁰. Furthermore, RTMs present in distant 15 16 tissues before metastatic dissemination should also be considered as a specific subset 17 of PreTAMs, considering that they populate the tissue before disseminating cancer cells, 18 but with the noticeable difference that they likely have already been subjected to the 19 systemic effects of the primary tumor. In this regard, pre-metastatic niche PreTAMs can 20 be already reprogrammed, at least partially, and contribute to the priming of the tissue 21 "soil" for metastatic outgrowth. Expanding the concept of "PreTAMs" beyond the primary 22 tumor site, this phenomenon is highly tissue-specific, being orchestrated by both RTMs 23 and bone marrow-derived macrophages present in, or recruited to, the secondary site 24 before the arrival of any disseminating cancer cell. Mechanistically, such remote priming of macrophages at the pre-metastatic niche can involve both extracellular vesicles¹⁶¹⁻¹⁶⁴ 25 and secreted factors¹⁶⁵ produced by primary tumors, as well as, potentially, by tumor-26 27 unrelated cues (eg prior environmental or mutagenic insults) that modulate macrophage 28 identities and functions at distal sites.

After successful metastatic seeding by tumor cells, pre-metastatic niche PreTAMs and TAMs subsequently recruited by seeded tumor cells coexist and favor secondary tumor outgrowth by establishing a reciprocal interplay with cancer cells and other

1 stromal/immune cells in the environment that provides growth factors and facilitates 2 escape from immune surveillance¹⁶⁶. For example, seminal work shows that inhibition of 3 CCL2–CCR2-recruitment of monocyte-derived macrophages reduces metastasis burden in the lung⁴⁹. Such dynamic heterogeneity of macrophages at metastatic sites continues 4 5 to impact (and can help predict) disease progression in highly tissue-specific manner. As 6 a few examples, in the liver, single-cell analyses of metastasis-associated macrophages have revealed subpopulations with distinct morphologies and transcriptional programs¹⁶⁷ 7 8 and identified clinically relevant markers (eg GPNMB¹⁶⁸, C1Q¹⁶⁹, SPP1¹⁷⁰) of poor 9 prognosis. In this line, mouse models of colorectal carcinoma metastasis to liver associate 10 specific subpopulations of recruited macrophages with T cell exclusion and transition from micro- to macro-metastasis¹⁷¹. In the omentum, RTMs play a pivotal role in the 11 establishment and development of metastasis in ovarian cancer¹⁷². Similarly, in the bone, 12 13 metastasis-infiltrating macrophages instruct resistance to androgen deprivation therapies to prostate cancer cell¹⁷³, mediating a highly-tissue specific interplay between 14 15 inflammation and hormones. In the brain, TAM heterogeneity of brain metastasis is 16 distinct from that of primary tumors (gliomas), with an increased abundance of tumoreducated microglia (PreTAMs) with differential immunomodulatory capacities^{174,175}. Thus, 17 just like at the primary tumor, tissue contexture specificities and temporal aspects 18 19 influence macrophage diversity at metastatic sites.

20 Whether organismal-level effects of macrophages dictate metastasis risk and 21 evolutionary patterns deserves further investigation. In a murine model of myocardial 22 infarction, local inflammation/injury can trigger changes in macrophage numbers and 23 states at uninjured distant sites, underscoring a remarkable network of remote, highly tissue-specific, macrophage responses spanning different organs¹⁷⁶. Taking this concept 24 25 into the context of cancer, it is conceivable that pre-existing or tumor-induced systemic 26 inflammatory responses influence metastatic disease progression by impacting the 27 heterogeneity and function of pre-metastatic and/or metastasis-recruited TAMs in a 28 tissue-specific manner.

- 29
- 30
- 31

1 Concluding remarks

2 While macrophages can physiologically exert tissue reparative and anti-tumor 3 immune activities¹⁷⁷, most clinical studies have established a clear correlation between macrophage abundance in tumors and the severity of the prognosis^{7,178-180}, supporting 4 5 the notion of a subversion of these cells in pre-malignancy and cancer. Clinical 6 observations and experimental systems support that their heterogeneity, plasticity, and 7 pleiotropic functions are influenced by multiple parameters, including ontogeny, spatial 8 context, and several temporal determinants, from macrophage individual experience and 9 lifespan to the organism's age and exposome. Herein, we propose to integrate the 10 dimension of time into the current framework to better understand the roles of 11 macrophages inter-connecting inflammation and cancer, and their dynamism within 12 developing and evolving tumoral niches. Such a paradigm shift is occurring in non-cancerrelated fields, such as in neurodegenerative and metabolic diseases, where a specific 13 subpopulation of TREM2⁺ macrophages termed DAM^{181,182} or LAM^{183,184} respectively has 14 15 been shown to act as pathological drivers.

16 Recent innovations in single-cell sequencing technologies, lineage-tracing 17 technologies, and model systems now provide the tools and knowledge base to deeply characterize TAMs at spatial and *temporal* resolution. To parallel these advances, we 18 19 propose to similarly refine macrophage nomenclature by distinguishing between 20 PreTAMs vs TAMs, depending on whether macrophages (irrespective of ontogeny or 21 spatial location) pre-existed in the tissue vs were recruited after tumor or metastasis 22 formation. Of note, distinguishing time and ontogeny-related features are not mutually 23 exclusive choices. We rather envision that combining both nomenclatures will be a 24 powerful and simple way to conceptualize (and refer to) the multi-factorial biology 25 underlying macrophage identity and functions in cancer. Developing and consolidating a 26 framework that considers the impact of developmental origin, tissue contexture, and time 27 in TAM identity will revolutionize our capacity to explore, chart, and therapeutically target 28 their roles in the development, establishment, and progression of cancers.

- 29
- 30
- 31

1 Acknowledgements

2 We thank members of the Ginhoux and Alonso-Curbelo lab for critical discussions. (TO 3 ADD YOUR GRANT/FELLOWSHIP DETAILS HERE). D.A.-C. is supported by the La Caixa Junior Leader Fellowship (award number LCF/BQ/PI20/11760006), FERO-4 ASEICA (award number BFERO2021), the US Department of Defense Pancreatic Cancer 5 6 Research Program (award number W81XWH-22-1-0814), and the Spanish Ministry of 7 Science and Innovation (grant number PID2021-128102OA-I00). We also thank all the 8 researchers who have contributed to the tumor-associated macrophage field and the 9 concepts discussed herein. We apologize to the many investigators whose work could 10 not be referenced in this piece due to space constraints.

1 References

- Medzhitov, R. (2021). The spectrum of inflammatory responses. Science 374, 1070-1075.
 10.1126/science.abi5200.
- 4 2. Medzhitov, R. (2008). Origin and physiological roles of inflammation. Nature 454, 4285 435. 10.1038/nature07201.
- Besic, M., and Greten, F.R. (2016). Inflammation and cancer: tissue regeneration gone awry. Curr. Opin. Cell Biol. 43, 55-61. 10.1016/j.ceb.2016.07.010.
- 8 4. Coussens, L.M., and Werb, Z. (2002). Inflammation and cancer. Nature 420, 860-867.
 9 10.1038/nature01322.
- 10 5. <u>https://www.cancer.gov/about-cancer/causes-prevention/risk/age</u>. (2021).
- 116.Greten, F.R., and Grivennikov, S.I. (2019). Inflammation and Cancer: Triggers,12Mechanisms, and Consequences. Immunity 51, 27-41. 10.1016/j.immuni.2019.06.025.
- Cassetta, L., and Pollard, J.W. (2018). Targeting macrophages: therapeutic approaches in cancer. Nat. Rev. Drug Discov. 17, 887-904. 10.1038/nrd.2018.169.
- 158.Noy, R., and Pollard, J.W. (2014). Tumor-associated macrophages: from mechanisms to16therapy. Immunity 41, 49-61. 10.1016/j.immuni.2014.06.010.
- Park, M.D., Silvin, A., Ginhoux, F., and Merad, M. (2022). Macrophages in health and disease. Cell *185*, 4259-4279. 10.1016/j.cell.2022.10.007.
- 19 10. Wynn, T.A., Chawla, A., and Pollard, J.W. (2013). Macrophage biology in development,
 20 homeostasis and disease. Nature 496, 445-455. 10.1038/nature12034.
- 11. Nowicki, A., Szenajch, J., Ostrowska, G., Wojtowicz, A., Wojtowicz, K., Kruszewski,
 A.A., Maruszynski, M., Aukerman, S.L., and Wiktor-Jedrzejczak, W. (1996). Impaired
 tumor growth in colony-stimulating factor 1 (CSF-1)-deficient, macrophage-deficient
 op/op mouse: evidence for a role of CSF-1-dependent macrophages in formation of tumor
 stroma. Int. J. Cancer 65, 112-119. 10.1002/(SICI)1097-0215(19960103)65:1<112::AID-
 IJC19>3.0.CO;2-I.
- Bottazzi, B., Polentarutti, N., Acero, R., Balsari, A., Boraschi, D., Ghezzi, P., Salmona,
 M., and Mantovani, A. (1983). Regulation of the macrophage content of neoplasms by
 chemoattractants. Science 220, 210-212. 10.1126/science.6828888.
- Ferrante, C.J., and Leibovich, S.J. (2012). Regulation of Macrophage Polarization and
 Wound Healing. Adv Wound Care (New Rochelle) *1*, 10-16. 10.1089/wound.2011.0307.
- Mosser, D.M., and Edwards, J.P. (2008). Exploring the full spectrum of macrophage
 activation. Nat. Rev. Immunol. *8*, 958-969. 10.1038/nri2448.
- Mulder, K., Patel, A.A., Kong, W.T., Piot, C., Halitzki, E., Dunsmore, G., Khalilnezhad,
 S., Irac, S.E., Dubuisson, A., Chevrier, M., et al. (2021). Cross-tissue single-cell landscape
 of human monocytes and macrophages in health and disease. Immunity *54*, 1883-1900
 e1885. 10.1016/j.immuni.2021.07.007.
- Cheng, S., Li, Z., Gao, R., Xing, B., Gao, Y., Yang, Y., Qin, S., Zhang, L., Ouyang, H.,
 Du, P., et al. (2021). A pan-cancer single-cell transcriptional atlas of tumor infiltrating
 myeloid cells. Cell *184*, 792-809 e723. 10.1016/j.cell.2021.01.010.
- Peng, J., Sun, B.F., Chen, C.Y., Zhou, J.Y., Chen, Y.S., Chen, H., Liu, L., Huang, D., Jiang,
 J., Cui, G.S., et al. (2019). Single-cell RNA-seq highlights intra-tumoral heterogeneity and
 malignant progression in pancreatic ductal adenocarcinoma. Cell Res. 29, 725-738.
 10.1038/s41422-019-0195-y.
- 45 18. Giladi, A., and Amit, I. (2018). Single-Cell Genomics: A Stepping Stone for Future
 46 Immunology Discoveries. Cell *172*, 14-21. 10.1016/j.cell.2017.11.011.

- Regev, A., Teichmann, S.A., Lander, E.S., Amit, I., Benoist, C., Birney, E., Bodenmiller,
 B., Campbell, P., Carninci, P., Clatworthy, M., et al. (2017). The Human Cell Atlas. Elife
 6. 10.7554/eLife.27041.
- Chung, W., Eum, H.H., Lee, H.O., Lee, K.M., Lee, H.B., Kim, K.T., Ryu, H.S., Kim, S.,
 Lee, J.E., Park, Y.H., et al. (2017). Single-cell RNA-seq enables comprehensive tumour
 and immune cell profiling in primary breast cancer. Nat Commun *8*, 15081.
 10.1038/ncomms15081.
- 8 21. Kirschenbaum, D., Xie, K., Ingelfinger, F., Katzenelenbogen, Y., Abadie, K., Look, T.,
 9 Sheban, F., Phan, T.S., Li, B., Zwicky, P., et al. (2023). Time-resolved single-cell
 10 transcriptomics defines immune trajectories in glioblastoma. Cell.
 11 10.1016/j.cell.2023.11.032.
- Del Poggetto, E., Ho, I.L., Balestrieri, C., Yen, E.Y., Zhang, S., Citron, F., Shah, R., Corti,
 D., Diaferia, G.R., Li, C.Y., et al. (2021). Epithelial memory of inflammation limits tissue
 damage while promoting pancreatic tumorigenesis. Science 373, eabj0486.
 10.1126/science.abj0486.
- Carron, E.C., Homra, S., Rosenberg, J., Coffelt, S.B., Kittrell, F., Zhang, Y., Creighton,
 C.J., Fuqua, S.A., Medina, D., and Machado, H.L. (2017). Macrophages promote the
 progression of premalignant mammary lesions to invasive cancer. Oncotarget *8*, 5073150746. 10.18632/oncotarget.14913.
- 20 24. Ruffell, B., and Coussens, L.M. (2015). Macrophages and therapeutic resistance in cancer.
 21 Cancer Cell 27, 462-472. 10.1016/j.ccell.2015.02.015.
- 22 25. Whiteside, T.L. (2008). The tumor microenvironment and its role in promoting tumor
 23 growth. Oncogene 27, 5904-5912. 10.1038/onc.2008.271.
- 26. Pollard, J.W. (2004). Tumour-educated macrophages promote tumour progression and metastasis. Nat. Rev. Cancer 4, 71-78. 10.1038/nrc1256.
- 26 27. Mills, C.D., Kincaid, K., Alt, J.M., Heilman, M.J., and Hill, A.M. (2000). M-1/M-2
 27 macrophages and the Th1/Th2 paradigm. J. Immunol. 164, 6166-6173.
 28 10.4049/jimmunol.164.12.6166.
- 28. Stein, M., Keshav, S., Harris, N., and Gordon, S. (1992). Interleukin 4 potently enhances
 murine macrophage mannose receptor activity: a marker of alternative immunologic
 macrophage activation. J. Exp. Med. *176*, 287-292. 10.1084/jem.176.1.287.
- Bleriot, C., Chakarov, S., and Ginhoux, F. (2020). Determinants of Resident Tissue
 Macrophage Identity and Function. Immunity 52, 957-970.
 10.1016/j.immuni.2020.05.014.
- 35 30. Ginhoux, F., Schultze, J.L., Murray, P.J., Ochando, J., and Biswas, S.K. (2016). New
 insights into the multidimensional concept of macrophage ontogeny, activation and
 function. Nat. Immunol. 17, 34-40. 10.1038/ni.3324.
- 38 31. Mass, E., Ballesteros, I., Farlik, M., Halbritter, F., Gunther, P., Crozet, L., Jacome-Galarza,
 39 C.E., Handler, K., Klughammer, J., Kobayashi, Y., et al. (2016). Specification of tissue40 resident macrophages during organogenesis. Science 353. 10.1126/science.aaf4238.
- 41 32. Hoeffel, G., Chen, J., Lavin, Y., Low, D., Almeida, F.F., See, P., Beaudin, A.E., Lum, J.,
 42 Low, I., Forsberg, E.C., et al. (2015). C-Myb(+) erythro-myeloid progenitor-derived fetal
 43 monocytes give rise to adult tissue-resident macrophages. Immunity 42, 665-678.
 44 10.1016/j.immuni.2015.03.011.
- 33. Schulz, C., Gomez Perdiguero, E., Chorro, L., Szabo-Rogers, H., Cagnard, N., Kierdorf,
 K., Prinz, M., Wu, B., Jacobsen, S.E., Pollard, J.W., et al. (2012). A lineage of myeloid

- cells independent of Myb and hematopoietic stem cells. Science 336, 86-90.
 10.1126/science.1219179.
- 3 34. Ginhoux, F., Greter, M., Leboeuf, M., Nandi, S., See, P., Gokhan, S., Mehler, M.F.,
 Conway, S.J., Ng, L.G., Stanley, E.R., et al. (2010). Fate mapping analysis reveals that
 adult microglia derive from primitive macrophages. Science 330, 841-845.
 10.1126/science.1194637.
- Sieweke, M.H., and Allen, J.E. (2013). Beyond stem cells: self-renewal of differentiated
 macrophages. Science *342*, 1242974. 10.1126/science.1242974.
- 9 36. Aziz, A., Soucie, E., Sarrazin, S., and Sieweke, M.H. (2009). MafB/c-Maf deficiency
 10 enables self-renewal of differentiated functional macrophages. Science 326, 867-871.
 11 10.1126/science.1176056.
- 37. Bain, C.C., Bravo-Blas, A., Scott, C.L., Gomez Perdiguero, E., Geissmann, F., Henri, S.,
 Malissen, B., Osborne, L.C., Artis, D., and Mowat, A.M. (2014). Constant replenishment
 from circulating monocytes maintains the macrophage pool in the intestine of adult mice.
 Nat. Immunol. 15, 929-937. 10.1038/ni.2967.
- Tamoutounour, S., Guilliams, M., Montanana Sanchis, F., Liu, H., Terhorst, D., Malosse,
 C., Pollet, E., Ardouin, L., Luche, H., Sanchez, C., et al. (2013). Origins and functional
 specialization of macrophages and of conventional and monocyte-derived dendritic cells
 in mouse skin. Immunity *39*, 925-938. 10.1016/j.immuni.2013.10.004.
- 39. Mass, E., Nimmerjahn, F., Kierdorf, K., and Schlitzer, A. (2023). Tissue-specific
 macrophages: how they develop and choreograph tissue biology. Nat. Rev. Immunol. 23,
 563-579. 10.1038/s41577-023-00848-y.
- 40. Lavin, Y., Winter, D., Blecher-Gonen, R., David, E., Keren-Shaul, H., Merad, M., Jung,
 S., and Amit, I. (2014). Tissue-resident macrophage enhancer landscapes are shaped by the
 local microenvironment. Cell *159*, 1312-1326. 10.1016/j.cell.2014.11.018.
- 41. Martinek, J., Lin, J., Kim, K.I., Wang, V.G., Wu, T.C., Chiorazzi, M., Boruchov, H.,
 Gulati, A., Seeniraj, S., Sun, L., et al. (2022). Transcriptional profiling of macrophages in
 situ in metastatic melanoma reveals localization-dependent phenotypes and function. Cell
 Rep Med *3*, 100621. 10.1016/j.xcrm.2022.100621.
- Sakai, M., Troutman, T.D., Seidman, J.S., Ouyang, Z., Spann, N.J., Abe, Y., Ego, K.M.,
 Bruni, C.M., Deng, Z., Schlachetzki, J.C.M., et al. (2019). Liver-Derived Signals
 Sequentially Reprogram Myeloid Enhancers to Initiate and Maintain Kupffer Cell Identity.
 Immunity *51*, 655-670 e658. 10.1016/j.immuni.2019.09.002.
- 34 43. Bonnardel, J., T'Jonck, W., Gaublomme, D., Browaeys, R., Scott, C.L., Martens, L., Vanneste, B., De Prijck, S., Nedospasov, S.A., Kremer, A., et al. (2019). Stellate Cells, 35 36 Hepatocytes, and Endothelial Cells Imprint the Kupffer Cell Identity on Monocytes 37 Colonizing the Liver Macrophage Niche. Immunity 51, 638-654 e639. 38 10.1016/j.immuni.2019.08.017.
- 44. Chakarov, S., Lim, H.Y., Tan, L., Lim, S.Y., See, P., Lum, J., Zhang, X.M., Foo, S.,
 Nakamizo, S., Duan, K., et al. (2019). Two distinct interstitial macrophage populations
 coexist across tissues in specific subtissular niches. Science *363*. 10.1126/science.aau0964.
- 42 45. Lim, H.Y., Lim, S.Y., Tan, C.K., Thiam, C.H., Goh, C.C., Carbajo, D., Chew, S.H.S., See,
 43 P., Chakarov, S., Wang, X.N., et al. (2018). Hyaluronan Receptor LYVE-1-Expressing
 44 Macrophages Maintain Arterial Tone through Hyaluronan-Mediated Regulation of Smooth
 45 Muscle Cell Collagon, Immunity 49, 326, 341, 6227, 10, 1016/j. immuni 2018, 06, 008
- 45 Muscle Cell Collagen. Immunity *49*, 326-341 e327. 10.1016/j.immuni.2018.06.008.

- 46. van Furth, R., Cohn, Z.A., Hirsch, J.G., Humphrey, J.H., Spector, W.G., and Langevoort,
 H.L. (1972). The mononuclear phagocyte system: a new classification of macrophages,
 monocytes, and their precursor cells. Bull World Health Organ 46, 845-852.
- 4 47. Cortez-Retamozo, V., Etzrodt, M., Newton, A., Rauch, P.J., Chudnovskiy, A., Berger, C.,
 5 Ryan, R.J., Iwamoto, Y., Marinelli, B., Gorbatov, R., et al. (2012). Origins of tumorassociated macrophages and neutrophils. Proc Natl Acad Sci U S A *109*, 2491-2496.
 7 10.1073/pnas.1113744109.
- 48. Franklin, R.A., Liao, W., Sarkar, A., Kim, M.V., Bivona, M.R., Liu, K., Pamer, E.G., and
 Li, M.O. (2014). The cellular and molecular origin of tumor-associated macrophages.
 Science 344, 921-925. 10.1126/science.1252510.
- 49. Qian, B.Z., Li, J., Zhang, H., Kitamura, T., Zhang, J., Campion, L.R., Kaiser, E.A., Snyder,
 L.A., and Pollard, J.W. (2011). CCL2 recruits inflammatory monocytes to facilitate breasttumour metastasis. Nature 475, 222-225. 10.1038/nature10138.
- 50. Baer, J.M., Zuo, C., Kang, L.I., de la Lastra, A.A., Borcherding, N.C., Knolhoff, B.L.,
 Bogner, S.J., Zhu, Y., Yang, L., Laurent, J., et al. (2023). Fibrosis induced by resident
 macrophages has divergent roles in pancreas inflammatory injury and PDAC. Nat.
 Immunol. 24, 1443-1457. 10.1038/s41590-023-01579-x.
- 51. Zhu, Y., Herndon, J.M., Sojka, D.K., Kim, K.W., Knolhoff, B.L., Zuo, C., Cullinan, D.R.,
 Luo, J., Bearden, A.R., Lavine, K.J., et al. (2017). Tissue-Resident Macrophages in
 Pancreatic Ductal Adenocarcinoma Originate from Embryonic Hematopoiesis and
 Promote Tumor Progression. Immunity 47, 323-338 e326. 10.1016/j.immuni.2017.07.014.
- Liou, G.Y., Doppler, H., Necela, B., Krishna, M., Crawford, H.C., Raimondo, M., and
 Storz, P. (2013). Macrophage-secreted cytokines drive pancreatic acinar-to-ductal
 metaplasia through NF-kappaB and MMPs. J. Cell Biol. 202, 563-577.
 10.1083/jcb.201301001.
- 53. Liou, G.Y., Fleming Martinez, A.K., Doppler, H.R., Bastea, L.I., and Storz, P. (2023).
 Inflammatory and alternatively activated macrophages independently induce metaplasia
 but cooperatively drive pancreatic precancerous lesion growth. iScience 26, 106820.
 10.1016/j.isci.2023.106820.
- 54. Liu, Z., Gu, Y., Chakarov, S., Bleriot, C., Kwok, I., Chen, X., Shin, A., Huang, W., Dress,
 R.J., Dutertre, C.A., et al. (2019). Fate Mapping via Ms4a3-Expression History Traces
 Monocyte-Derived Cells. Cell *178*, 1509-1525 e1519. 10.1016/j.cell.2019.08.009.
- 55. Kirschenbaum, D., Xie, K., Ingelfinger, F., Katzenelenbogen, Y., Abadie, K., Look, T.,
 Sheban, F., Phan, T.S., Li, B., Zwicky, P., et al. (2024). Time-resolved single-cell
 transcriptomics defines immune trajectories in glioblastoma. Cell 187, 149-165 e123.
 10.1016/j.cell.2023.11.032.
- S6. Caronni, N., La Terza, F., Vittoria, F.M., Barbiera, G., Mezzanzanica, L., Cuzzola, V.,
 Barresi, S., Pellegatta, M., Canevazzi, P., Dunsmore, G., et al. (2023). IL-1beta(+)
 macrophages fuel pathogenic inflammation in pancreatic cancer. Nature 623, 415-422.
 10.1038/s41586-023-06685-2.
- 57. Burdziak, C., Alonso-Curbelo, D., Walle, T., Reyes, J., Barriga, F.M., Haviv, D., Xie, Y.,
 Zhao, Z., Zhao, C.J., Chen, H.A., et al. (2023). Epigenetic plasticity cooperates with cellcell interactions to direct pancreatic tumorigenesis. Science *380*, eadd5327.
 10.1126/science.add5327.
- 45 58. Nalio Ramos, R., Missolo-Koussou, Y., Gerber-Ferder, Y., Bromley, C.P., Bugatti, M.,
 46 Nunez, N.G., Tosello Boari, J., Richer, W., Menger, L., Denizeau, J., et al. (2022). Tissue-

1 2		resident FOLR2(+) macrophages associate with CD8(+) T cell infiltration in human breast cancer. Cell <i>185</i> , 1189-1207 e1125. 10.1016/j.cell.2022.02.021.
3	59.	Molgora, M., Esaulova, E., Vermi, W., Hou, J., Chen, Y., Luo, J., Brioschi, S., Bugatti,
4		M., Omodei, A.S., Ricci, B., et al. (2020). TREM2 Modulation Remodels the Tumor
5		Myeloid Landscape Enhancing Anti-PD-1 Immunotherapy. Cell 182, 886-900 e817.
6		10.1016/j.cell.2020.07.013.
7	60.	Kloosterman, D.J., and Akkari, L. (2023). Macrophages at the interface of the co-evolving
8		cancer ecosystem. Cell 186, 1627-1651, 10.1016/j.cell.2023.02.020.
9	61.	Li, Z., Pai, R., Gupta, S., Currenti, J., Guo, W., Di Bartolomeo, A., Feng, H., Zhang, Z.,
10		Li, Z., Liu, L., et al. (2024). Presence of onco-fetal neighborhoods in hepatocellular
11		carcinoma is associated with relapse and response to immunotherapy. Nat Cancer.
12		10.1038/s43018-023-00672-2.
13	62.	Sharma, A., Bleriot, C., Currenti, J., and Ginhoux, F. (2022). Oncofetal reprogramming in
14		tumour development and progression. Nat. Rev. Cancer 22, 593-602, 10.1038/s41568-022-
15		00497-8.
16	63.	Sharma, A., Seow, J.J.W., Dutertre, C.A., Pai, R., Bleriot, C., Mishra, A., Wong, R.M.M.,
17		Singh, G.S.N., Sudhagar, S., Khalilnezhad, S., et al. (2020). Onco-fetal Reprogramming of
18		Endothelial Cells Drives Immunosuppressive Macrophages in Hepatocellular Carcinoma.
19		Cell 183, 377-394 e321, 10,1016/i.cell.2020.08.040.
20	64	Chen, H.A., Ho, Y.L. Mezzadra, R., Adrover, J.M., Smolkin, R., Zhu, C., Woess, K.,
$\frac{2}{21}$	0	Bernstein, N., Schmitt, G., Fong, L., et al. (2023). Senescence Rewires Microenvironment
22		Sensing to Facilitate Antitumor Immunity, Cancer Discov 13, 432-453, 10,1158/2159-
23		8290 CD-22-0528
24	65	Okabe, Y., and Medzhitov, R. (2014). Tissue-specific signals control reversible program
25	001	of localization and functional polarization of macrophages. Cell 157, 832-844.
26		10.1016/i.cell.2014.04.016.
27	66	Zhou X Franklin R A Adler M Jacox I B Bailis W Shver I A Flavell R A
28	00.	Mayo A Alon U and Medzhitov R (2018) Circuit Design Features of a Stable Two-
29		Cell System Cell 172 744-757 e717 10 1016/j cell 2018 01 015
30	67	Okabe Y and Medzhitov R (2016) Tissue biology perspective on macrophages Nat
31	07.	Immunol 17 9-17 10 1038/ni 3320
32	68	Martincorena L and Campbell P L (2015) Somatic mutation in cancer and normal cells
33	00.	Science 349 1483-1489 10 1126/science aab4082
34	69	Stratton M.R. Campbell P.I. and Futreal P.A. (2009) The cancer genome Nature 458
35	07.	719-724 10 1038/nature07943
36	70	Grivennikov SI Greten FR and Karin M (2010) Immunity inflammation and
37	70.	cancer Cell 140 883-899 10 1016/i cell 2010 01 025
38	71	Reu P Khosravi A Bernard S Mold LF Salehpour M Alkass K Perl S Tisdale
30	/1.	I Possnert G Druid H and Frisen I (2017) The Lifesnan and Turnover of Microglia
40		in the Human Brain Cell Rep 20, 779-784, 10, 1016/i celrep 2017, 07, 004
<u>4</u> 0	72	Netea M.G. Giamarellos-Bourboulis F.I. Dominguez-Andres I. Curtis N. van Crevel
42	12.	R van de Veerdonk F I and Bonten M (2020) Trained Immunity: a Tool for Reducing
<u>⊿</u> ∠ ∕13		Susceptibility to and the Severity of SARS-CoV-2 Infection Cell 181 060-077
$\Delta \Delta$		$10\ 1016/i\ cell\ 2020\ 04\ 042$
тт 45	72	Netea M.G. Dominguez-Andres I. Barreiro I.R. Chavakis T. Divangahi M. Fucha
46	15.	E Josten I. A B van der Meer I W M Mblanga M M Mulder W I M et al (2020)
10		2000000000000000000000000000000000000

- 1Defining trained immunity and its role in health and disease. Nat. Rev. Immunol. 20, 375-2388. 10.1038/s41577-020-0285-6.
- 74. Netea, M.G., Joosten, L.A., Latz, E., Mills, K.H., Natoli, G., Stunnenberg, H.G., O'Neill,
 L.A., and Xavier, R.J. (2016). Trained immunity: A program of innate immune memory in
 health and disease. Science 352, aaf1098. 10.1126/science.aaf1098.
- 75. Netea, M.G., Quintin, J., and van der Meer, J.W. (2011). Trained immunity: a memory for
 innate host defense. Cell Host Microbe 9, 355-361. 10.1016/j.chom.2011.04.006.
- 8 76. Ghisletti, S., Barozzi, I., Mietton, F., Polletti, S., De Santa, F., Venturini, E., Gregory, L.,
 9 Lonie, L., Chew, A., Wei, C.L., et al. (2010). Identification and characterization of
 10 enhancers controlling the inflammatory gene expression program in macrophages.
 11 Immunity 32, 317-328. 10.1016/j.immuni.2010.02.008.
- Fanucchi, S., Fok, E.T., Dalla, E., Shibayama, Y., Borner, K., Chang, E.Y., Stoychev, S.,
 Imakaev, M., Grimm, D., Wang, K.C., et al. (2019). Immune genes are primed for robust
 transcription by proximal long noncoding RNAs located in nuclear compartments. Nat.
 Genet. *51*, 138-150. 10.1038/s41588-018-0298-2.
- 78. Penkov, S., Mitroulis, I., Hajishengallis, G., and Chavakis, T. (2019). Immunometabolic
 Crosstalk: An Ancestral Principle of Trained Immunity? Trends Immunol. 40, 1-11.
 10.1016/j.it.2018.11.002.
- Aegerter, H., Kulikauskaite, J., Crotta, S., Patel, H., Kelly, G., Hessel, E.M., Mack, M.,
 Beinke, S., and Wack, A. (2020). Influenza-induced monocyte-derived alveolar
 macrophages confer prolonged antibacterial protection. Nat. Immunol. 10.1038/s41590019-0568-x.
- 80. Naik, S., and Fuchs, E. (2022). Inflammatory memory and tissue adaptation in sickness
 and in health. Nature 607, 249-255. 10.1038/s41586-022-04919-3.
- 81. Geller, A.E., Shrestha, R., Woeste, M.R., Guo, H., Hu, X., Ding, C., Andreeva, K.,
 Chariker, J.H., Zhou, M., Tieri, D., et al. (2022). The induction of peripheral trained
 immunity in the pancreas incites anti-tumor activity to control pancreatic cancer
 progression. Nat Commun 13, 759. 10.1038/s41467-022-28407-4.
- 82. Priem, B., van Leent, M.M.T., Teunissen, A.J.P., Sofias, A.M., Mourits, V.P., Willemsen,
 L., Klein, E.D., Oosterwijk, R.S., Meerwaldt, A.E., Munitz, J., et al. (2020). Trained
 Immunity-Promoting Nanobiologic Therapy Suppresses Tumor Growth and Potentiates
 Checkpoint Inhibition. Cell 183, 786-801 e719. 10.1016/j.cell.2020.09.059.
- 83. White, M.C., Holman, D.M., Boehm, J.E., Peipins, L.A., Grossman, M., and Henley, S.J.
 (2014). Age and cancer risk: a potentially modifiable relationship. Am. J. Prev. Med. 46,
 S7-15. 10.1016/j.amepre.2013.10.029.
- 84. Lopez-Otin, C., Blasco, M.A., Partridge, L., Serrano, M., and Kroemer, G. (2022).
 Hallmarks of aging: An expanding universe. Cell. 10.1016/j.cell.2022.11.001.
- Solution
 85. Cagan, A., Baez-Ortega, A., Brzozowska, N., Abascal, F., Coorens, T.H.H., Sanders, M.A.,
 Lawson, A.R.J., Harvey, L.M.R., Bhosle, S., Jones, D., et al. (2022). Somatic mutation
 rates scale with lifespan across mammals. Nature *604*, 517-524. 10.1038/s41586-02204618-z.
- 42 86. Vogelstein, B., Papadopoulos, N., Velculescu, V.E., Zhou, S., Diaz, L.A., Jr., and Kinzler,
 43 K.W. (2013). Cancer genome landscapes. Science *339*, 1546-1558.
 44 10.1126/science.1235122.

- 87. Hodis, E., Watson, I.R., Kryukov, G.V., Arold, S.T., Imielinski, M., Theurillat, J.P.,
 Nickerson, E., Auclair, D., Li, L., Place, C., et al. (2012). A landscape of driver mutations
 in melanoma. Cell *150*, 251-263. 10.1016/j.cell.2012.06.024.
- 4 88. Morrison, S.J., Wandycz, A.M., Akashi, K., Globerson, A., and Weissman, I.L. (1996).
 5 The aging of hematopoietic stem cells. Nat. Med. 2, 1011-1016. 10.1038/nm0996-1011.
- 6 89. Chambers, S.M., Shaw, C.A., Gatza, C., Fisk, C.J., Donehower, L.A., and Goodell, M.A.
 7 (2007). Aging hematopoietic stem cells decline in function and exhibit epigenetic dysregulation. PLoS Biol. 5, e201. 10.1371/journal.pbio.0050201.
- 9 90. Sun, D., Luo, M., Jeong, M., Rodriguez, B., Xia, Z., Hannah, R., Wang, H., Le, T., Faull,
 10 K.F., Chen, R., et al. (2014). Epigenomic profiling of young and aged HSCs reveals
 11 concerted changes during aging that reinforce self-renewal. Cell Stem Cell 14, 673-688.
 12 10.1016/j.stem.2014.03.002.
- Mass, E., Jacome-Galarza, C.E., Blank, T., Lazarov, T., Durham, B.H., Ozkaya, N.,
 Pastore, A., Schwabenland, M., Chung, Y.R., Rosenblum, M.K., et al. (2017). A somatic
 mutation in erythro-myeloid progenitors causes neurodegenerative disease. Nature 549,
 389-393. 10.1038/nature23672.
- Hill, W., Lim, E.L., Weeden, C.E., Lee, C., Augustine, M., Chen, K., Kuan, F.C.,
 Marongiu, F., Evans, E.J., Jr., Moore, D.A., et al. (2023). Lung adenocarcinoma promotion
 by air pollutants. Nature *616*, 159-167. 10.1038/s41586-023-05874-3.
- 20 93. Hanahan, D. (2022). Hallmarks of Cancer: New Dimensions. Cancer Discov 12, 31-46.
 21 10.1158/2159-8290.CD-21-1059.
- 94. Alonso-Curbelo, D., Ho, Y.J., Burdziak, C., Maag, J.L.V., Morris, J.P.t., Chandwani, R.,
 Chen, H.A., Tsanov, K.M., Barriga, F.M., Luan, W., et al. (2021). A gene-environmentinduced epigenetic program initiates tumorigenesis. Nature *590*, 642-648. 10.1038/s41586020-03147-x.
- 26 95. Tlsty, T.D., and Gascard, P. (2019). Stromal directives can control cancer. Science *365*,
 27 122-123. 10.1126/science.aaw2368.
- 28 96. Modi, B.G., Neustadter, J., Binda, E., Lewis, J., Filler, R.B., Roberts, S.J., Kwong, B.Y., 29 Reddy, S., Overton, J.D., Galan, A., et al. (2012). Langerhans cells facilitate epithelial 30 DNA damage and squamous cell carcinoma. Science 335. 104-108. 31 10.1126/science.1211600.
- 97. Dunn, G.P., Old, L.J., and Schreiber, R.D. (2004). The immunobiology of cancer
 immunosurveillance and immunoediting. Immunity 21, 137-148.
 10.1016/j.immuni.2004.07.017.
- Balmain, A. (2020). The critical roles of somatic mutations and environmental tumorpromoting agents in cancer risk. Nat. Genet. 52, 1139-1143. 10.1038/s41588-020-007275.
- 38 99. Garlanda, C., and Mantovani, A. (2021). Interleukin-1 in tumor progression, therapy, and
 39 prevention. Cancer Cell *39*, 1023-1027. 10.1016/j.ccell.2021.04.011.
- Ridker, P.M., MacFadyen, J.G., Thuren, T., Everett, B.M., Libby, P., Glynn, R.J., and
 Group, C.T. (2017). Effect of interleukin-1beta inhibition with canakinumab on incident
 lung cancer in patients with atherosclerosis: exploratory results from a randomised, doubleblind, placebo-controlled trial. Lancet *390*, 1833-1842. 10.1016/S0140-6736(17)32247-X.
- 43 blind, placebo-controlled trial. Lancet *390*, 1833-1842. 10.1016/S0140-6736(17)32247-X.
 44 101. Bazzan, E., Turato, G., Tine, M., Radu, C.M., Balestro, E., Rigobello, C., Biondini, D.,
- 44 101. Bazzan, E., Turato, G., Tine, W., Radu, C.W., Baresto, E., Rigobeno, C., Biondini, D., 45 Schiavon, M., Lunardi, F., Baraldo, S., et al. (2017). Dual polarization of human alveolar

1 macrophages progressively increases with smoking and COPD severity. Respir Res 18, 40. 2 10.1186/s12931-017-0522-0. 3 102. Churg, A., Wang, R.D., Tai, H., Wang, X., Xie, C., Dai, J., Shapiro, S.D., and Wright, J.L. 4 (2003). Macrophage metalloelastase mediates acute cigarette smoke-induced inflammation 5 via tumor necrosis factor-alpha release. Am. J. Respir. Crit. Care Med. 167, 1083-1089. 6 10.1164/rccm.200212-1396OC. 7 Hamilton, R.F., Iyer, L.L., and Holian, A. (1996). Asbestos induces apoptosis in human 103. 8 alveolar macrophages. Am J Physiol 271, L813-819. 10.1152/ajplung.1996.271.5.L813. 9 Joshi, N., Watanabe, S., Verma, R., Jablonski, R.P., Chen, C.I., Cheresh, P., Markov, N.S., 104. 10 Reyfman, P.A., McQuattie-Pimentel, A.C., Sichizya, L., et al. (2020). A spatially restricted 11 fibrotic niche in pulmonary fibrosis is sustained by M-CSF/M-CSFR signalling in 12 monocyte-derived alveolar macrophages. Eur. Respir. J. 55. 10.1183/13993003.00646-13 2019. 14 105. Ma, J., Liu, Y., Guo, Y., Ma, Q., Ji, C., and Zhao, L. (2021). Transcriptional Profiling of 15 Aflatoxin B1-Induced Oxidative Stress and Inflammatory Response in Macrophages. 16 Toxins (Basel) 13. 10.3390/toxins13060401. 17 Fishbein, A., Wang, W., Yang, H., Yang, J., Hallisey, V.M., Deng, J., Verheul, S.M.L., 106. 18 Hwang, S.H., Gartung, A., Wang, Y., et al. (2020). Resolution of eicosanoid/cytokine 19 storm prevents carcinogen and inflammation-initiated hepatocellular cancer progression. 20 Proc Natl Acad Sci U S A 117, 21576-21587. 10.1073/pnas.2007412117. Storz, P. (2017). Acinar cell plasticity and development of pancreatic ductal 21 107. 22 adenocarcinoma. Nat Rev Gastroenterol Hepatol 14, 296-304. 10.1038/nrgastro.2017.12. 23 Zhang, Y., Yan, W., Collins, M.A., Bednar, F., Rakshit, S., Zetter, B.R., Stanger, B.Z., 108. 24 Chung, I., Rhim, A.D., and di Magliano, M.P. (2013). Interleukin-6 is required for 25 pancreatic cancer progression by promoting MAPK signaling activation and oxidative 26 stress resistance. Cancer Res. 73, 6359-6374. 10.1158/0008-5472.CAN-13-1558-T. 27 109. Lesina, M., Kurkowski, M.U., Ludes, K., Rose-John, S., Treiber, M., Kloppel, G., 28 Yoshimura, A., Reindl, W., Sipos, B., Akira, S., et al. (2011). Stat3/Socs3 activation by 29 IL-6 transsignaling promotes progression of pancreatic intraepithelial neoplasia and 30 development of pancreatic cancer. Cancer Cell 19, 456-469. 10.1016/j.ccr.2011.03.009. 31 110. Deeks, S.G., Tracy, R., and Douek, D.C. (2013). Systemic effects of inflammation on 32 health during chronic HIV infection. Immunity 39, 633-645. 33 10.1016/j.immuni.2013.10.001. 34 Belmonte, L., Olmos, M., Fanin, A., Parodi, C., Bare, P., Concetti, H., Perez, H., de Bracco, 111. 35 M.M., and Cahn, P. (2007). The intestinal mucosa as a reservoir of HIV-1 infection after 36 successful HAART. AIDS 21, 2106-2108. 10.1097/QAD.0b013e3282efb74b. 37 112. Sandler, N.G., and Douek, D.C. (2012). Microbial translocation in HIV infection: causes, 38 consequences and treatment opportunities. Nat. Rev. Microbiol. 10, 655-666. 39 10.1038/nrmicro2848. 40 113. Yarchoan, R., and Uldrick, T.S. (2018). HIV-Associated Cancers and Related Diseases. N 41 Engl J Med 378, 1029-1041. 10.1056/NEJMra1615896. 42 114. Bonecchi, R., Savino, B., Caronni, N., Celesti, G., Mantovani, A., and Locati, M. (2014). 43 Atypical chemokine receptor 2: a brake against Kaposi's sarcoma aggressiveness. 44 Oncoimmunology 3, e955337. 10.4161/21624011.2014.955337. 45 115. Wang, Q., Bergholz, J.S., Ding, L., Lin, Z., Kabraji, S.K., Hughes, M.E., He, X., Xie, S., 46 Jiang, T., Wang, W., et al. (2022). STING agonism reprograms tumor-associated

- macrophages and overcomes resistance to PARP inhibition in BRCA1-deficient models of
 breast cancer. Nat Commun *13*, 3022. 10.1038/s41467-022-30568-1.
- 116. Rana, H.Q., Sacca, R., Drogan, C., Gutierrez, S., Schlosnagle, E., Regan, M.M., Speare,
 V., LaDuca, H., Dolinsky, J., Garber, J.E., and Overmoyer, B.A. (2019). Prevalence of
 germline variants in inflammatory breast cancer. Cancer 125, 2194-2202.
 10.1002/cncr.32062.
- 7 117. <u>https://www.cancer.gov/types/breast/ibc-fact-sheet</u>. (2016).
- 8 118. Launonen, I.M., Lyytikainen, N., Casado, J., Anttila, E.A., Szabo, A., Haltia, U.M.,
 9 Jacobson, C.A., Lin, J.R., Maliga, Z., Howitt, B.E., et al. (2022). Single-cell tumor-immune
 10 microenvironment of BRCA1/2 mutated high-grade serous ovarian cancer. Nat Commun
 11 13, 835. 10.1038/s41467-022-28389-3.
- 119. Sloot, Y.J.E., Rabold, K., Netea, M.G., Smit, J.W.A., Hoogerbrugge, N., and Netea-Maier,
 R.T. (2019). Effect of PTEN inactivating germline mutations on innate immune cell
 function and thyroid cancer-induced macrophages in patients with PTEN hamartoma tumor
 syndrome. Oncogene *38*, 3743-3755. 10.1038/s41388-019-0685-x.
- Solberg, O.D., Mack, S.J., Lancaster, A.K., Single, R.M., Tsai, Y., Sanchez-Mazas, A.,
 and Thomson, G. (2008). Balancing selection and heterogeneity across the classical human
 leukocyte antigen loci: a meta-analytic review of 497 population studies. Hum. Immunol.
 69, 443-464. 10.1016/j.humimm.2008.05.001.
- 121. Hammer, R.E., Richardson, J.A., Simmons, W.A., White, A.L., Breban, M., and Taurog,
 J.D. (1995). High prevalence of colorectal cancer in HLA-B27 transgenic F344 rats with
 chronic inflammatory bowel disease. J Investig Med *43*, 262-268.
- 23 Alcalay, M., Bontoux, D., Maire, P., Matuchansky, C., Alcalay, D., and Tanzer, J. (1982). 122. 24 Ν J HLA-B27 and colorectal cancer. Engl Med 307, 443-444. 25 10.1056/NEJM198208123070718.
- Lin, E.Y., Li, J.F., Gnatovskiy, L., Deng, Y., Zhu, L., Grzesik, D.A., Qian, H., Xue, X.N.,
 and Pollard, J.W. (2006). Macrophages regulate the angiogenic switch in a mouse model
 of breast cancer. Cancer Res. *66*, 11238-11246. 10.1158/0008-5472.CAN-06-1278.
- Harmey, J.H., Dimitriadis, E., Kay, E., Redmond, H.P., and Bouchier-Hayes, D. (1998).
 Regulation of macrophage production of vascular endothelial growth factor (VEGF) by
 hypoxia and transforming growth factor beta-1. Ann Surg Oncol 5, 271-278.
 10.1007/BF02303785.
- Vakkila, J., and Lotze, M.T. (2004). Inflammation and necrosis promote tumour growth.
 Nat. Rev. Immunol. 4, 641-648. 10.1038/nri1415.
- Ling, Y.H., Chen, J.W., Wen, S.H., Huang, C.Y., Li, P., Lu, L.H., Mei, J., Li, S.H., Wei,
 W., Cai, M.Y., and Guo, R.P. (2020). Tumor necrosis as a poor prognostic predictor on
 postoperative survival of patients with solitary small hepatocellular carcinoma. BMC
 Cancer 20, 607. 10.1186/s12885-020-07097-5.
- Langner, C., Hutterer, G., Chromecki, T., Leibl, S., Rehak, P., and Zigeuner, R. (2006).
 Tumor necrosis as prognostic indicator in transitional cell carcinoma of the upper urinary
 tract. J Urol *176*, 910-913; discussion 913-914. 10.1016/j.juro.2006.04.019.
- 42 128. Swinson, D.E., Jones, J.L., Richardson, D., Cox, G., Edwards, J.G., and O'Byrne, K.J.
 43 (2002). Tumour necrosis is an independent prognostic marker in non-small cell lung 44 cancer: correlation with biological variables. Lung Cancer *37*, 235-240. 10.1016/s0169-45 5002(02)00172-1.

- 129. Gonzalez, A.N., Quintana, J.A., Garcia-Silva, S., Mazariegos, M., Gonzalez de la Aleja,
 A., Nicolas-Avila, J.A., Walter, W., Adrover, J.M., Crainiciuc, G., Kuchroo, V.K., et al.
 (2017). Phagocytosis imprints heterogeneity in tissue-resident macrophages. J. Exp. Med.
 214, 1281-1296. 10.1084/jem.20161375.
- Modak, M., Mattes, A.K., Reiss, D., Skronska-Wasek, W., Langlois, R., Sabarth, N.,
 Konopitzky, R., Ramirez, F., Lehr, K., Mayr, T., et al. (2022). CD206+ tumor-associated
 macrophages cross-present tumor antigen and drive anti-tumor immunity. JCI Insight.
 10.1172/jci.insight.155022.
- 9 131. Jaynes, J.M., Sable, R., Ronzetti, M., Bautista, W., Knotts, Z., Abisoye-Ogunniyan, A., Li,
 10 D., Calvo, R., Dashnyam, M., Singh, A., et al. (2020). Mannose receptor (CD206)
 11 activation in tumor-associated macrophages enhances adaptive and innate antitumor
 12 immune responses. Sci Transl Med *12*. 10.1126/scitranslmed.aax6337.
- 13 132. Ikeda, H., Old, L.J., and Schreiber, R.D. (2002). The roles of IFN gamma in protection against tumor development and cancer immunoediting. Cytokine Growth Factor Rev. 13, 95-109. 10.1016/s1359-6101(01)00038-7.
- 16 133. O'Sullivan, T., Saddawi-Konefka, R., Vermi, W., Koebel, C.M., Arthur, C., White, J.M.,
 17 Uppaluri, R., Andrews, D.M., Ngiow, S.F., Teng, M.W., et al. (2012). Cancer
 18 immunoediting by the innate immune system in the absence of adaptive immunity. J. Exp.
 19 Med. 209, 1869-1882. 10.1084/jem.20112738.
- Shields, C.W.t., Evans, M.A., Wang, L.L., Baugh, N., Iyer, S., Wu, D., Zhao, Z., Pusuluri,
 A., Ukidve, A., Pan, D.C., and Mitragotri, S. (2020). Cellular backpacks for macrophage
 immunotherapy. Sci Adv 6, eaaz6579. 10.1126/sciadv.aaz6579.
- 135. Kawasaki, T., Ikegawa, M., Yunoki, K., Otani, H., Ori, D., Ishii, K.J., Kuroda, E.,
 Takamura, S., Kitabatake, M., Ito, T., et al. (2022). Alveolar macrophages instruct CD8(+)
 T cell expansion by antigen cross-presentation in lung. Cell Rep 41, 111828.
 10.1016/j.celrep.2022.111828.
- 27 136. Curiel, T.J., Coukos, G., Zou, L., Alvarez, X., Cheng, P., Mottram, P., Evdemon-Hogan,
 28 M., Conejo-Garcia, J.R., Zhang, L., Burow, M., et al. (2004). Specific recruitment of
 29 regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced
 30 survival. Nat. Med. 10, 942-949. 10.1038/nm1093.
- 137. Ren, L., Yu, Y., Wang, L., Zhu, Z., Lu, R., and Yao, Z. (2016). Hypoxia-induced CCL28
 promotes recruitment of regulatory T cells and tumor growth in liver cancer. Oncotarget 7,
 75763-75773. 10.18632/oncotarget.12409.
- Nixon, B.G., Kuo, F., Ji, L., Liu, M., Capistrano, K., Do, M., Franklin, R.A., Wu, X.,
 Kansler, E.R., Srivastava, R.M., et al. (2022). Tumor-associated macrophages expressing
 the transcription factor IRF8 promote T cell exhaustion in cancer. Immunity 55, 2044-2058
 e2045. 10.1016/j.immuni.2022.10.002.
- 139. Kersten, K., Hu, K.H., Combes, A.J., Samad, B., Harwin, T., Ray, A., Rao, A.A., Cai, E.,
 Marchuk, K., Artichoker, J., et al. (2022). Spatiotemporal co-dependency between
 macrophages and exhausted CD8(+) T cells in cancer. Cancer Cell 40, 624-638 e629.
 10.1016/j.ccell.2022.05.004.
- Iwai, Y., Ishida, M., Tanaka, Y., Okazaki, T., Honjo, T., and Minato, N. (2002).
 Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor
 immunotherapy by PD-L1 blockade. Proc Natl Acad Sci U S A *99*, 12293-12297.
 10.1073/pnas.192461099.

- 141. Wculek, S.K., Dunphy, G., Heras-Murillo, I., Mastrangelo, A., and Sancho, D. (2022).
 Metabolism of tissue macrophages in homeostasis and pathology. Cell Mol Immunol 19, 384-408. 10.1038/s41423-021-00791-9.
- 4 142. Hotamisligil, G.S. (2017). Inflammation, metaflammation and immunometabolic disorders. Nature *542*, 177-185. 10.1038/nature21363.
- Hotamisligil, G.S. (2017). Foundations of Immunometabolism and Implications for
 Metabolic Health and Disease. Immunity 47, 406-420. 10.1016/j.immuni.2017.08.009.
- 8 144. O'Neill, L.A., Kishton, R.J., and Rathmell, J. (2016). A guide to immunometabolism for
 9 immunologists. Nat. Rev. Immunol. *16*, 553-565. 10.1038/nri.2016.70.
- 10 145. Warburg, O. (1924). Über den Stoffwechsel der Carcinomzelle. Naturwissenschaften
 11 1131-1137.
- 12 146. Vitale, I., Manic, G., Coussens, L.M., Kroemer, G., and Galluzzi, L. (2019). Macrophages
 13 and Metabolism in the Tumor Microenvironment. Cell Metab 30, 36-50.
 14 10.1016/j.cmet.2019.06.001.
- 15 147. Goossens, P., Rodriguez-Vita, J., Etzerodt, A., Masse, M., Rastoin, O., Gouirand, V., Ulas,
 T., Papantonopoulou, O., Van Eck, M., Auphan-Anezin, N., et al. (2019). Membrane
 Cholesterol Efflux Drives Tumor-Associated Macrophage Reprogramming and Tumor
 Progression. Cell Metab 29, 1376-1389 e1374. 10.1016/j.cmet.2019.02.016.
- Wenes, M., Shang, M., Di Matteo, M., Goveia, J., Martin-Perez, R., Serneels, J., Prenen,
 H., Ghesquiere, B., Carmeliet, P., and Mazzone, M. (2016). Macrophage Metabolism
 Controls Tumor Blood Vessel Morphogenesis and Metastasis. Cell Metab 24, 701-715.
 10.1016/j.cmet.2016.09.008.
- Sadik, A., Somarribas Patterson, L.F., Ozturk, S., Mohapatra, S.R., Panitz, V., Secker, P.F.,
 Pfander, P., Loth, S., Salem, H., Prentzell, M.T., et al. (2020). IL411 Is a Metabolic Immune
 Checkpoint that Activates the AHR and Promotes Tumor Progression. Cell *182*, 1252-1270
 e1234. 10.1016/j.cell.2020.07.038.
- Lambert, A.W., Pattabiraman, D.R., and Weinberg, R.A. (2017). Emerging Biological
 Principles of Metastasis. Cell *168*, 670-691. 10.1016/j.cell.2016.11.037.
- Massague, J., and Obenauf, A.C. (2016). Metastatic colonization by circulating tumour cells. Nature 529, 298-306. 10.1038/nature17038.
- 152. Bonde, A.K., Tischler, V., Kumar, S., Soltermann, A., and Schwendener, R.A. (2012).
 Intratumoral macrophages contribute to epithelial-mesenchymal transition in solid tumors.
 BMC Cancer *12*, 35. 10.1186/1471-2407-12-35.
- 153. Casanova-Acebes, M., Dalla, E., Leader, A.M., LeBerichel, J., Nikolic, J., Morales, B.M.,
 Brown, M., Chang, C., Troncoso, L., Chen, S.T., et al. (2021). Tissue-resident
 macrophages provide a pro-tumorigenic niche to early NSCLC cells. Nature 595, 578-584.
 10.1038/s41586-021-03651-8.
- Rhim, A.D., Mirek, E.T., Aiello, N.M., Maitra, A., Bailey, J.M., McAllister, F., Reichert,
 M., Beatty, G.L., Rustgi, A.K., Vonderheide, R.H., et al. (2012). EMT and dissemination
 precede pancreatic tumor formation. Cell *148*, 349-361. 10.1016/j.cell.2011.11.025.
- Pastushenko, I., Mauri, F., Song, Y., de Cock, F., Meeusen, B., Swedlund, B., Impens, F.,
 Van Haver, D., Opitz, M., Thery, M., et al. (2021). Fat1 deletion promotes hybrid EMT
 state, tumour stemness and metastasis. Nature *589*, 448-455. 10.1038/s41586-020-030461.
- 45 156. Paget, S. (1889). The distribution of secondary growths in cancer of the breast. The Lancet
 46 133, 571-573.

- 1 157. Metchnikov, E. (1891). Lecture on Phagocytosis and Immunity. Br. Med. J. 1, 213-217.
- 158. Metchnikov, E. (1883). Untersuchungen ueber die mesodermalen Phagocyten einiger
 Wirbeltiere. Biologisches Centralblatt 3, 560-565.
- Patras, L., Shaashua, L., Matei, I., and Lyden, D. (2023). Immune determinants of the premetastatic niche. Cancer Cell *41*, 546-572. 10.1016/j.ccell.2023.02.018.
- Kaplan, R.N., Riba, R.D., Zacharoulis, S., Bramley, A.H., Vincent, L., Costa, C.,
 MacDonald, D.D., Jin, D.K., Shido, K., Kerns, S.A., et al. (2005). VEGFR1-positive
 haematopoietic bone marrow progenitors initiate the pre-metastatic niche. Nature 438, 820827. 10.1038/nature04186.
- 161. Hoshino, A., Costa-Silva, B., Shen, T.L., Rodrigues, G., Hashimoto, A., Tesic Mark, M.,
 Molina, H., Kohsaka, S., Di Giannatale, A., Ceder, S., et al. (2015). Tumour exosome
 integrins determine organotropic metastasis. Nature 527, 329-335. 10.1038/nature15756.
- 13 162. Costa-Silva, B., Aiello, N.M., Ocean, A.J., Singh, S., Zhang, H., Thakur, B.K., Becker, A.,
 14 Hoshino, A., Mark, M.T., Molina, H., et al. (2015). Pancreatic cancer exosomes initiate
 15 pre-metastatic niche formation in the liver. Nat. Cell Biol. *17*, 816-826. 10.1038/ncb3169.
- 16 163. Wortzel, I., Dror, S., Kenific, C.M., and Lyden, D. (2019). Exosome-Mediated Metastasis:
 17 Communication from a Distance. Dev. Cell 49, 347-360. 10.1016/j.devcel.2019.04.011.
- 18 164. Peinado, H., Aleckovic, M., Lavotshkin, S., Matei, I., Costa-Silva, B., Moreno-Bueno, G.,
 19 Hergueta-Redondo, M., Williams, C., Garcia-Santos, G., Ghajar, C., et al. (2012).
 20 Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic
 21 phenotype through MET. Nat. Med. *18*, 883-891. 10.1038/nm.2753.
- Kitamura, T., Doughty-Shenton, D., Cassetta, L., Fragkogianni, S., Brownlie, D., Kato, Y.,
 Carragher, N., and Pollard, J.W. (2017). Monocytes Differentiate to Immune Suppressive
 Precursors of Metastasis-Associated Macrophages in Mouse Models of Metastatic Breast
 Cancer. Front Immunol 8, 2004. 10.3389/fimmu.2017.02004.
- Baumann, Z., Auf der Maur, P., and Bentires-Alj, M. (2022). Feed-forward loops between
 metastatic cancer cells and their microenvironment-the stage of escalation. EMBO Mol
 Med 14, e14283. 10.15252/emmm.202114283.
- 167. Donadon, M., Torzilli, G., Cortese, N., Soldani, C., Di Tommaso, L., Franceschini, B.,
 Carriero, R., Barbagallo, M., Rigamonti, A., Anselmo, A., et al. (2020). Macrophage
 morphology correlates with single-cell diversity and prognosis in colorectal liver
 metastasis. J. Exp. Med. 217. 10.1084/jem.20191847.
- 168. Cortese, N., Carriero, R., Barbagallo, M., Putignano, A.R., Costa, G., Giavazzi, F., Grizzi,
 F., Pasqualini, F., Peano, C., Basso, G., et al. (2023). High-Resolution Analysis of
 Mononuclear Phagocytes Reveals GPNMB as a Prognostic Marker in Human Colorectal
 Liver Metastasis. Cancer Immunol Res 11, 405-420. 10.1158/2326-60666.CIR-22-0462.
- Yang, J., Lin, P., Yang, M., Liu, W., Fu, X., Liu, D., Tao, L., Huo, Y., Zhang, J., Hua, R.,
 et al. (2021). Integrated genomic and transcriptomic analysis reveals unique characteristics
 of hepatic metastases and pro-metastatic role of complement C1q in pancreatic ductal
 adenocarcinoma. Genome Biol 22, 4. 10.1186/s13059-020-02222-w.
- Sathe, A., Mason, K., Grimes, S.M., Zhou, Z., Lau, B.T., Bai, X., Su, A., Tan, X., Lee, H.,
 Suarez, C.J., et al. (2023). Colorectal Cancer Metastases in the Liver Establish
 Immunosuppressive Spatial Networking between Tumor-Associated SPP1+ Macrophages
 and Fibroblasts. Clin. Cancer. Res. 29, 244-260. 10.1158/1078-0432.CCR-22-2041.
- 45 171. Canellas-Socias, A., Cortina, C., Hernando-Momblona, X., Palomo-Ponce, S., Mulholland,
 46 E.J., Turon, G., Mateo, L., Conti, S., Roman, O., Sevillano, M., et al. (2022). Metastatic

- recurrence in colorectal cancer arises from residual EMP1(+) cells. Nature *611*, 603-613.
 10.1038/s41586-022-05402-9.
- 172. Etzerodt, A., Moulin, M., Doktor, T.K., Delfini, M., Mossadegh-Keller, N., Bajenoff, M.,
 Sieweke, M.H., Moestrup, S.K., Auphan-Anezin, N., and Lawrence, T. (2020). Tissueresident macrophages in omentum promote metastatic spread of ovarian cancer. J. Exp.
 Med. 217. 10.1084/jem.20191869.
- 173. Li, X.F., Selli, C., Zhou, H.L., Cao, J., Wu, S., Ma, R.Y., Lu, Y., Zhang, C.B., Xun, B.,
 Lam, A.D., et al. (2023). Macrophages promote anti-androgen resistance in prostate cancer
 bone disease. J. Exp. Med. 220. 10.1084/jem.20221007.
- 174. Klemm, F., Maas, R.R., Bowman, R.L., Kornete, M., Soukup, K., Nassiri, S., Brouland,
 J.P., Iacobuzio-Donahue, C.A., Brennan, C., Tabar, V., et al. (2020). Interrogation of the
 Microenvironmental Landscape in Brain Tumors Reveals Disease-Specific Alterations of
 Immune Cells. Cell 181, 1643-1660 e1617. 10.1016/j.cell.2020.05.007.
- 14 175. Friebel, E., Kapolou, K., Unger, S., Nunez, N.G., Utz, S., Rushing, E.J., Regli, L., Weller,
 M., Greter, M., Tugues, S., et al. (2020). Single-Cell Mapping of Human Brain Cancer
 Reveals Tumor-Specific Instruction of Tissue-Invading Leukocytes. Cell *181*, 1626-1642
 e1620. 10.1016/j.cell.2020.04.055.
- 18 176. Hoyer, F.F., Naxerova, K., Schloss, M.J., Hulsmans, M., Nair, A.V., Dutta, P., Calcagno,
 D.M., Herisson, F., Anzai, A., Sun, Y., et al. (2019). Tissue-Specific Macrophage
 Responses to Remote Injury Impact the Outcome of Subsequent Local Immune Challenge.
 Immunity. 10.1016/j.immuni.2019.10.010.
- 177. Kruse, B., Buzzai, A.C., Shridhar, N., Braun, A.D., Gellert, S., Knauth, K., Pozniak, J.,
 Peters, J., Dittmann, P., Mengoni, M., et al. (2023). CD4(+) T cell-induced inflammatory
 cell death controls immune-evasive tumours. Nature *618*, 1033-1040. 10.1038/s41586023-06199-x.
- 178. DeNardo, D.G., Brennan, D.J., Rexhepaj, E., Ruffell, B., Shiao, S.L., Madden, S.F.,
 Gallagher, W.M., Wadhwani, N., Keil, S.D., Junaid, S.A., et al. (2011). Leukocyte
 complexity predicts breast cancer survival and functionally regulates response to
 chemotherapy. Cancer Discov 1, 54-67. 10.1158/2159-8274.CD-10-0028.
- 179. Chen, X., Chen, J., Zhang, W., Sun, R., Liu, T., Zheng, Y., and Wu, Y. (2017). Prognostic
 value of diametrically polarized tumor-associated macrophages in multiple myeloma.
 Oncotarget *8*, 112685-112696. 10.18632/oncotarget.22340.
- Shen, H., Liu, J., Chen, S., Ma, X., Ying, Y., Li, J., Wang, W., Wang, X., and Xie, L.
 (2021). Prognostic Value of Tumor-Associated Macrophages in Clear Cell Renal Cell
 Carcinoma: A Systematic Review and Meta-Analysis. Front Oncol 11, 657318.
 10.3389/fonc.2021.657318.
- 181. Deczkowska, A., Keren-Shaul, H., Weiner, A., Colonna, M., Schwartz, M., and Amit, I.
 (2018). Disease-Associated Microglia: A Universal Immune Sensor of Neurodegeneration.
 Cell 173, 1073-1081. 10.1016/j.cell.2018.05.003.
- Keren-Shaul, H., Spinrad, A., Weiner, A., Matcovitch-Natan, O., Dvir-Szternfeld, R.,
 Ulland, T.K., David, E., Baruch, K., Lara-Astaiso, D., Toth, B., et al. (2017). A Unique
 Microglia Type Associated with Restricting Development of Alzheimer's Disease. Cell *169*, 1276-1290 e1217. 10.1016/j.cell.2017.05.018.
- Remmerie, A., Martens, L., Thone, T., Castoldi, A., Seurinck, R., Pavie, B., Roels, J.,
 Vanneste, B., De Prijck, S., Vanhockerhout, M., et al. (2020). Osteopontin Expression

- Identifies a Subset of Recruited Macrophages Distinct from Kupffer Cells in the Fatty
 Liver. Immunity 53, 641-657 e614. 10.1016/j.immuni.2020.08.004.
 I84. Jaitin, D.A., Adlung, L., Thaiss, C.A., Weiner, A., Li, B., Descamps, H., Lundgren, P.,
- 3 184. Jaitin, D.A., Adlung, L., Thaiss, C.A., Weiner, A., Li, B., Descamps, H., Lundgren, P.,
 4 Bleriot, C., Liu, Z., Deczkowska, A., et al. (2019). Lipid-Associated Macrophages Control
 5 Metabolic Homeostasis in a Trem2-Dependent Manner. Cell *178*, 686-698 e614.
 6 10.1016/j.cell.2019.05.054.
- 7

1 Figure legends

2

3 Figure 1: Time is a key determinant of TAM biology

4 Macrophages can be found in healthy tissues where they are known as Resident Tissue 5 Macrophages (RTMs). They contribute to tissue homeostasis but can also be involved at 6 the stage of premalignancy influencing and being influenced by tumor onset. In this 7 context, they should be considered as PreTAMs. Once the tumor is formed, resulting 8 inflammation will promote monocyte recruitment which will differentiate into TAMs, the 9 specificity of these cells being to experience only the tumor microenvironment. Of note, 10 respective half-life and fate of these two populations need to be refined, especially during 11 the three different phases of tumor development, elimination, equilibrium and escape.

12

13 Figure 2: PreTAMs and TAMs are different populations of macrophages

14 As pre-existing in the healthy tissue, PreTAMs can have a tissue-dependent mixed origin, 15 with the co-existence of embryonic and monocyte-derived cells, a unique feature not 16 shared with the later recruited TAMs deriving from monocytes in response to tumor-17 elicited inflammation. PreTAMs have their own history, inhabiting the tissue before tumor 18 onset and can therefore be impacted by prior insults. This is in contrast with monocyte-19 derived TAMs which are only exposed to the tumor niche and have therefore a different 20 imprinting. In light of this, crosstalk with other cells from the tumor microenvironment has 21 to be considered separately.

22

23 Figure 3: Spatial distribution of RTMs and TAMs

PreTAMs and TAMs coexist in established primary tumors and metastasis. But macrophages are sessile cells and their location is different. Later recruited TAMs are mostly found within the core of the tumor whereas PreTAMs are mostly excluded to the periphery, resulting in different interactions with other cells from the TME and different functions. Of note, macrophages from the pre-metastatic niche constitute an example of PreTAMs which prepare the seeding of disseminating tumor cells before their arrival.