

ERR α : unraveling its role as a key player in cell migration

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1	${\sf ERR}\alpha$: Unraveling its Role as a Key Player in Cell Migration
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10	Abstract
11	Cell migration is essential throughout the life of multicellular organisms, and largely depends
12	on the spatial and temporal regulation of cytoskeletal dynamics, cell adhesion and signal
13	transduction. Interestingly, Estrogen-related receptor alpha (ERR α) has been identified as a
14	major regulator of cell migration in both physiological and pathological conditions. ERR $lpha$ is an
15	orphan member of the nuclear hormone receptor superfamily of transcription factors and
16	displays many biological functions. ERR $lpha$ is a global regulator of energy metabolism, and it is
17	also highly involved in bone homeostasis, development, differentiation, immunity and cancer
18	progression. Importantly, in some instances, the regulation of these biological processes relies
19	on the ability to orchestrate cell movements. Therefore, this review describes how $ERRlpha$ -
20	mediated cell migration contributes not only to tissue homeostasis but also to tumorigenesis
21	and metastasis, and highlights the molecular and cellular mechanisms by which ERR $lpha$ finely
22	controls the cell migratory potential.
23	Keywords: ERR α , cell migration, development, homeostasis, cancer

Running title: ERR α , a key player in cell migration

25

26 Introduction

27 Cell migration is a crucial process driving morphogenesis throughout embryonic development. 28 In the adult, cell movements participate in immune surveillance, tissue repair and renewal 29 among other physiological processes (reviewed in ¹). In addition, abnormal cell migration 30 contributes to a wide variety of diseases such as chronic inflammatory diseases, mental 31 retardation and cancer cell dissemination (reviewed in ²). Depending on the context, cells can 32 migrate either individually or collectively in response to guidance cues (reviewed in ^{3,4}). In 33 most cases, cell movements require a polarized organization, an active remodeling of the actin 34 and microtubule cytoskeletons, an interaction with the extracellular matrix (ECM) through 35 integrin-mediated focal adhesions (FA) and the activation of migration-related signaling 36 pathways (reviewed in 5-7) (Fig. 1). Although cell migration has been extensively studied, the 37 complex mechanisms involved in the regulation of these events are still incompletely 38 understood.

39 Estrogen-related receptor alpha (ERRa, encoded by ESRRA) is an orphan nuclear receptor 40 acting as a transcription factor. It exerts multiple essential biological functions through its 41 capacity to control specific target gene expression and, particularly, its involvement in cell 42 metabolism and cancer progression has been widely described (reviewed in ^{8–10}). Importantly, 43 ERR α has been shown to play a crucial role in the regulation of cell movements during both 44 development and adulthood and it also regulates cancer cell migration (Fig. 2). Herein, the 45 review focuses on the ERRa-dependent mechanisms that contribute to cell migration in these 46 different contexts.

47

48 Structure and regulation of ERRα

49 ERR α (NR3B1) has been identified in 1988 based on the high sequence similarity within 50 its DNA binding domain to that of the Estrogen receptor alpha (ER α) ¹¹. ERR α belongs to a 51 nuclear receptor subfamily that comprises two other isoforms in mammals encoded by 52 distinct genes: ERR β (NR3B2) and ERR γ (NR3B3). However, since no natural ligands have been 53 identified so far, the ERRs are considered orphan nuclear receptors (reviewed in ^{12,13}).

54 As other nuclear receptors, ERRa displays a typical conserved structural organization, 55 comprising a highly variable amino-terminal domain (the A/B domain), a centrally located 56 DNA-binding domain (DBD) that includes two Zn fingers, a flexible hinge region and a carboxy-57 terminal ligand-binding domain (LBD) containing a transcription transactivation region (reviewed in ^{14,15}). The ERR α -DBD specifically binds to the consensus DNA sequence 58 59 TnAAGGTCA, referred to as estrogen-related response element (ERRE) ^{16,17}. Strikingly, the 60 sequence-dependent DNA shape of this binding site contributes to the stabilization of the 61 ERRα homodimer and facilitates its asymmetric positioning on DNA ^{18,19}. As shown by 62 structural analysis, the ERRα-LBD includes a putative small binding ligand pocket (LBP) almost 63 completely filled with bulky side chains of amino acids that confers an active conformation to ERR α in a ligand independent-manner ²⁰. However, the existence of ligands regulating ERR α 64 65 activity by inducing conformational changes and/or modulating the recruitment of cofactors, 66 cannot be excluded. In this way, specific ERRa synthetic ligands such as compound A, 67 compound 29 and XCT790 have been identified. These compounds which have the ability to enter the ERR α -LBP by significantly changing its conformation and consequently deactivate 68 the receptor, are called inverse agonists (reviewed in ^{12,13}). Importantly, ERRα activity mostly 69 70 depends on its association with cofactors that are differentially expressed in cells and tissues ^{21,22}. In addition, post-translational modifications such as phosphorylation, sumoylation or 71 72 acetylation of ERRa can modulate its transcriptional activity in a cell type, cofactor and

promoter-dependent manner ^{23–27}. The modulation of ERR α expression is also crucial for the regulation of its functions (reviewed in ¹²).

75

76 **The ERR isoforms**

As mentioned above, the ERR subfamily consists of three ERR isoforms (ERR α , β and γ). All three isoforms show a high sequence identity in their DBD and their LBD is less conserved. The ERRs recognize the same specific DNA sequences and thus exhibit some overlapping functions by regulating similar targets genes in a ligand-independent manner. However, they also display differential activities which may depend on their tissue specificity or different expression levels.

83 Similar to ERR α , ERR γ is strongly expressed in tissues with high energy demand and plays a 84 role in the regulation of energy metabolism (reviewed in ^{15,28}). In addition, analysis of knockout 85 mice has revealed that both ERR α and ERR γ act as major regulators of heart functions ^{29,30} (reviewed in ¹⁵). ERR γ is also involved in the control of the differentiation of mesenchymal 86 87 stem cells and in the maintenance of bone homeostasis (reviewed in ³¹). At first glance, ERRa 88 and ERRy appear to play opposite roles in cancer cells, respectively inducing or inhibiting 89 cancer progression. ERRy is weakly expressed in endometrial, colorectal, prostate and gastric 90 cancers. By contrast, it is highly expressed in breast and ovarian cancers and this correlates with a favorable prognosis ³²(reviewed in ^{21,33}). Specifically, ERR_γ has been shown to inhibit 91 92 breast, gastric and colorectal cancer cell proliferation and tumor growth, in part, by repressing 93 the Wnt signaling pathway or by inducing oxidative metabolism and mesenchymal-toepithelial transition 34,35,32 . However, a pro-tumoral activity of ERR γ has also emerged. 94 Notably, its upregulation promotes cancer cell proliferation and invasion $^{36-40}$ (reviewed in 33). 95

96 Interestingly, in the same way as ERR α , ERR γ can drive cell migration to facilitate cancer 97 progression ⁴⁰.

98 ERR β is mainly expressed in embryonic cells and the ERR β knockout demonstrates a major 99 role of this receptor in the regulation of placental formation, the maintenance of pluripotency 100 of embryonic stem cells and the specification of epithelial cells of the inner ear (reviewed in 101 ^{15,31}). ERR β is globally poorly or not expressed in adult and cancer tissues (reviewed in ²¹). 102 However, ERR β is significantly expressed in the brain and involved in the regulation of stress response, food intake, satiety and whole-body energy balance (reviewed in ¹⁵). Interestingly, 103 104 ERRy appear to have opposite effects on these processes, suggesting a tight tissue-dependent 105 regulation of the ERR isoform activity or expression. Splicing ERR^β isoforms have been 106 identified and shown to be expressed in glioblastoma cell lines and tumors ^{41,42}. In particular, 107 the long ERR_β2 isoform plays an antitumor role by inducing apoptotic cell death and 108 restricting glioblastoma cell growth and migration 42 . ERR β and the ERR β 2 isoform have also 109 been shown to inhibit prostate and breast cancer cell proliferation ⁴³ (reviewed in ²¹).

110

111 Main biological functions of ERRα

112 $ERR\alpha$ is a key regulator of cellular metabolism

ERR α is widely expressed in embryos and adult tissues. Of interest, ERR α has been identified as a master regulator of cellular energy metabolism, mostly in tissues with high-energy demand such as the skeletal muscle, kidney, liver, heart and adipose tissue. Importantly, ERR α binds to the promoter regions of many genes involved in oxidative phosphorylation, ATP synthesis, fatty acid oxidation, pyruvate metabolism, tricarboxylic acid (TCA) cycle, glycolysis, mitochondrial biogenesis and mitophagy (reviewed in ^{9,44}). These metabolic functions of ERR α mainly involve the peroxisome proliferator-activated receptor γ (PPAR γ)-coactivator 1 α (PGC-

1 α) and PGC-1 β coactivators (^{45,46} and reviewed in ⁴⁷). However, in some cases, the cofactors 120 RIP140 and NCOR1 can also be involved (reviewed in 15). In addition, a crosstalk between ERRlpha121 122 and the thyroid hormone receptors (THR) has been shown ^{48,49}. Notably, TH induces 123 mitochondrial metabolic pathways such as oxidative phosphorylation, fatty acid oxidation and TCA cycle through ERR α in a THRB1 and PGC-1 α -dependent manner. ERR α also contributes to 124 125 the TH-mediated regulation of mitochondrial fission, biogenesis and mitophagy ⁴⁹. 126 Interestingly, TH has been shown to modulate breast cancer cell migration ^{50,51}. A coordinated 127 ERR α -THR effect may thus be possible, although not demonstrated, in this regulation.

128 Unsurprisingly, the alteration of the metabolic pathways regulated by ERR α is associated with

human metabolic diseases such as heart failure, diabetes and obesity (reviewed in ¹⁵). ERR α

130 also contributes to the specific metabolic adaptations of cancer cells (reviewed in ^{9,44}).

131

132 ERR α modulates the differentiation of multiple mesenchymal lineages

133 ERR α has been shown to control the differentiation of mesenchymal stem cells towards 134 specific cell fates. For instance, the expression of ERRa increases early during myogenesis in 135 parallel with that of the PGC-1 α coactivator, and both factors contribute to the myogenic 136 program *in vitro* and *ex vivo* (^{52–56} and reviewed in ¹⁵). Particularly, ERRα promotes myocyte 137 differentiation through the modulation of mitochondrial metabolism and MAPK signaling (^{30,54} and reviewed in ^{15,57}). Similarly, ERR α may contribute to muscle regeneration in mice by 138 regulating the expression of metabolic genes ⁵⁸. Furthermore, ERRα plays a major role in bone 139 140 homeostasis by repressing osteoblast differentiation both in vitro and in vivo (and reviewed in 15,57). Additionally, a role of ERR α in the differentiation of adipocytes and chondrocytes in 141 culture is supported by several studies (^{59,62,63} and reviewed in ⁵⁷). 142

144 ERR α is involved in cancer progression and invasion

145 Importantly, ERR α is strongly expressed in various types of cancers, and its elevated 146 expression is associated with metastasis and poor prognosis ^{64–74}.

147 Many studies have demonstrated that ERR α plays a role in tumor growth and progression. 148 Notably, ERR α promotes proliferation of colon, pancreatic, lung and breast cancer lines either 149 by acting downstream of the HER2/IGF-1 pathways or by modulating the expression of the p21 cell cycle inhibitor, members of the cyclin family or other cell cycle regulators (74–77 and 150 reviewed in 21,78). In addition, ERR α stimulates angiogenesis by directly inducing the 151 152 expression of the vascular endothelial growth factor (VEGF) in several cancer cell lines ^{79–81}. 153 Furthermore, ERR α enhances Hypoxia-Inducible Factor 1 α (HIF-1 α)-mediated transcription 154 and consequently facilitates tumor and blood vessel growth in mice under hypoxic conditions ^{81,82}. ERR α also contributes to the metabolic switch of cancer cells, also termed the Warburg 155 effect, by stimulating aerobic glycolysis ^{83,84}. Moreover, ERRa contributes to Epithelial-156 157 Mesenchymal Transition (EMT), and promotes cell migration and extracellular matrix (ECM) invasion (^{85–87} and reviewed in ⁹, also see below). 158

- 159
- 160 ERRα-mediated regulation of cell migration

161 ERRα promotes the migration of cells during development and adult tissue homeostasis
162 without interfering with their differentiation

Gastrulation is an early developmental process during which cell migration leads to massive
reorganization of the embryo, allowing the establishment of the three embryonic tissue layers.
Later, cells migrate within layers and differentiate to form tissues and organs (reviewed in ⁸⁸).
Interestingly, ERRα function is not required for cell fate determination in zebrafish embryos,
but it plays a major role in controlling different morphogenetic movements during zebrafish

168 gastrulation ⁸⁹. However, the molecular mechanisms by which ERR α regulates these cellular 169 movements have not yet been characterized. *WNT11* regulates hypoblast cell movements 170 during zebrafish gastrulation and it also has been identified as a transcriptional target of ERR α 171 that promotes cancer cell migration (^{90,91}, also see below). Altogether, this suggests that the 172 *wnt11*-dependent signaling pathway could be involved downstream of ERR α to control 173 orientated cell movements at the onset of gastrulation (Fig. 3).

174 Furthermore, ERR α has been shown to modulate osteoblast differentiation and bone 175 formation (see above). It also regulates the migration of osteoclasts. Indeed, loss of ERR α 176 activity leads to actin filament disorganization, reduced adhesion and inhibition of 177 transmigration of mature osteoclasts in vitro. It also results in a decreased expression of 178 osteopontin, integrin β 3 and altered localization of c-src that may contribute to the adhesion 179 and migration defects observed 92 (Fig. 3). Thus, ERR α may participate in the regulation of 180 bone homeostasis and remodeling by acting on the osteoblast lineage but also by controlling 181 osteoclast migration. In addition, ERR α regulates the migration of macrophages *ex vivo* and *in* 182 vivo; this probably depends on its capacity to modulate the expression of TNFAIP1 and the 183 stability of the RhoA GTPase in these cells ⁸⁵ (Fig. 3). Of interest, inhibition of cell migration 184 resulting from the loss of function of ERR α does not result from an impaired differentiation of the osteoclast precursors and macrophages in these conditions ^{85,92}. 185

186

187 $ERR\alpha$ can behave as a negative or positive modulator of vascular cell migration and 188 angiogenesis

Angiogenesis is a process by which new vessels form; this involves the proliferation, migration and differentiation of endothelial cells (EC). ERR α is a major regulator of angiogenesis under both physiological and pathological conditions (reviewed in ⁹, see also

192 below). Interestingly, this function of ERR α relies in part on its ability to regulate EC migration. 193 ERR α has been shown to be expressed in EC from different tissues and it can either repress or 194 enhance EC migration and angiogenesis, depending on the cellular context and the availability 195 of cofactors. Gene expression analysis of ERR α -deficient EC indicates that the receptor mainly 196 acts as a transcriptional repressor of target genes associated with angiogenesis, cell adhesion 197 and cell migration 93 . In particular, ERR α modulates the expression of the vascular endothelial 198 growth factor (VEGF) and also inhibits the expression of several migratory and cell-adhesion 199 genes such as *Pecam*, *Mmp3*, *Cdh3* and *Esam1* which are all involved in angiogenic regulation 200 (Fig. 3). As a consequence, ERR α plays a role in limiting angiogenesis by negatively controlling 201 EC migration, tube formation and sprouting in response to growth factors in vitro and in vivo 202 93.

203 In contrast, ERR α facilitates angiogenesis through the induction of VEGF expression and secretion from several cell types 79-82,94-96. In response to a deprivation in nutrients and 204 205 oxygen, the cofactor PGC-1 α coactivates ERR α to induce the expression of VEGF, and this may contribute to the physiological response to ischemia ⁹⁴ (Fig. 3). Similarly, flavonoids of 206 207 Scutellaria baicalensis root such as Baicalin stimulate the expression of VEGF through the 208 ERR α -PGC-1 α pathway, and consequently induce EC migration and sprout formation ⁸⁰. In 209 addition, ERR α regulates the neovascularisation process through VEGF to facilitate tumoral 210 growth and progression in mice. ERR α also enhances the expression of osteoprotegerin that can contribute to the angiogenic process 95,97 . Furthermore, ERRlpha interacts with and stabilizes 211 212 HIF-1 α , and stimulates the expression of its target genes including VEGF (Fig. 3). The functional 213 interaction between ERR α and HIF-1 α plays thus an important role in adapting cancer cells to 214 in vitro and in vivo hypoxic conditions, and promotes angiogenesis, tumoral growth and 215 invasion 81,82 . In addition, ERR α induces the expression of eNOS in EC, an enzyme that

generates nitric oxide (NO). As a consequence, ERRα may promote tumor progression through
the NO-mediated regulation of EC migration, vascular permeability, blood vessel remodeling
and angiogenesis ^{98,99} (Fig. 3).

219 ERR α also positively controls the migration of vascular smooth muscle cells (VSMCs). VSMCs 220 are important components of blood vessel walls that maintain structural integrity and drive 221 the contraction of the vascular wall. The platelet-derived growth factor (PDGF), mainly 222 produced by VSMCs, vascular endothelial cells, platelets or macrophages in the injured 223 vascular walls is involved in vascular remodeling (reviewed in 100). Interestingly, ERR α is 224 required both in vitro and in vivo for platelet-derived growth factor (PDGF)-induced 225 proliferation and migration of VSMC through the regulation of RhoA/p27/Cdks and Wnt4/βcatenin pathways ¹⁰¹ (Fig. 3). In this way, ERR α acts as an effector of PDGF in controlling 226 227 vascular remodeling, and thus may contribute to the development of cardiovascular diseases such as atherosclerosis and restenosis ^{100,101}. 228

229

230 ERR α plays a role in the induction of Epithelial-Mesenchymal Transition

EMT is a cellular process that leads to the conversion of epithelial cells into mesenchymal cells. EMT occurs during embryonic development and becomes reactivated in pathological contexts such as inflammation and cancer progression. During EMT, epithelial cells lose their polarity and cell-cell junctions, and gain migratory and invasive properties (reviewed in ¹⁰²).

ERRα has been shown to induce EMT in cancer cells and consecutively to promote their
migration. Interestingly, multiple mechanisms are involved in ERRα-induced EMT (Fig. 3).
Depletion of ERRα in a range of cancer cell lines has been shown to increase the expression of
E-cadherin, a marker of epithelial cells, and to decrease the expression of vimentin, fibronectin

and N-cadherin which mark a mesenchymal phenotype ^{74,103–109}. Interestingly, in most of these 240 241 cancer cells, ERR α drives EMT and maintains the mesenchymal phenotype by regulating the 242 expression of the transcription factors SNAIL, SLUG and/or Zinc-finger E-box-binding 1 (ZEB1) ^{74,103,106,107,110,111}. In gallbladder cancer cells, ERR α promotes EMT and cell migration through 243 244 the regulation of the expression of Nectin cell adhesion molecule 4 (Nectin-4) and the 245 activation of PI3K/AKT pathway ¹⁰⁷. In addition, ERR α regulates cell migration *in vitro*, tumor 246 growth and invasion in vivo via the activation of the fibronectin and STAT3 pathways that 247 promote EMT ^{104–106}. In turn, the JAK2/STAT3 pathway positively regulates the expression of 248 ERR α to induce EMT ¹¹¹. ERR α also regulates the expression and the secretion of TGF- β by 249 osteosarcoma and endometrial cancer cell lines, which can activate ERR α expression via a 250 positive feedback mechanism. As a consequence, ERR α mediates TGF- β -induced SNAIL activation, EMT and cell migration ^{106,112,113}. 251

252 Surprisingly, the ERR α -knockout mice exhibit no evident developmental abnormalities which 253 is unexpected given the key role of ERR α in the regulation of EMT (see above). They were 254 initially found to display reduced adiposity, peripheral fat deposits and resistance to high-fat diet-induced obesity ¹¹⁴. In addition, they show a decrease capacity in mitochondrial 255 256 biogenesis and oxidative metabolism in the brown adipose tissue and they are consequently defective in adaptive thermogenesis 115 . ERR α deficiency also produces behavioral 257 disturbances associated with synaptic dysfunction 116,117 . To some extent, ERR α and ERR γ have 258 259 overlapping functions, suggesting that the absence of a phenotype associated with a defective 260 EMT might be attributed to functional redundancy between ERR isoforms during 261 development.

262

263 A crosstalk between ER α , ERBB2 and ERR α in breast cancers

264 Based on the structural homology shared by ERR α and ER α , a potential functional 265 relationship between these receptors has been suggested for years in breast cancers. 266 However, a genome-wide analysis indicated that most of the ERR α target genes are distinct 267 from those regulated by ER α in breast cancer cell lines and tumors samples, demonstrating that ERR α plays a role in breast cancers that is mainly independent of ER α ^{118,119}. Nonetheless, 268 269 ERR α and ER α regulate a small fraction of common target genes either through binding to 270 distinct but closely located estrogen-response elements (EREs) and ERREs or through 271 competitive binding to overlapping ERE / ERRE motifs. Interestingly, several of these common 272 target genes are linked to the ErbB2 signaling ¹¹⁹. Specifically, ERR α and its cofactor PGC-1 β 273 are involved in the transcriptional regulation of *ErbB2* and several co-amplified genes located 274 within the *ErbB2* amplicon that play a role in survival and cell migration *in vitro*, and tumor growth *in vivo* ¹²⁰. In turn, the transcriptional ERR α activity is regulated by the EGF/ErbB2 275 276 signaling pathway in breast cancer cell lines ^{24,64} (Fig. 3). Therefore, reciprocal regulation of 277 ERRa and ErbB2 contributes to breast tumor development, progression and tamoxifen resistance ¹²⁰. 278

279

280 ERR α is a crucial factor regulating cancer cell migration

ERR α has been identified as a major regulator of cancer cell migration and invasion (^{73,85–} 87,91,118,121,122) (Fig. 3). Given the importance of altered metabolism in cancers as well as the fact that ERR α modulates different aspects of energy storage and consumption, it could be argued that all cancers phenotypes in which the receptor in involved are a consequence of its metabolic effects. However, ERR α regulates cancer cell migration in cells that do not express PGC-1 α or $-\beta$ ⁸⁶. Furthermore, RNA-Seq approaches have shown that ERR α does not modulate the expression of metabolic targets in these cells although ChIP-Seq experiments indicate that

288 ERR α is bound to the regulatory regions of these genes ¹²³. Altogether, this points to a critical 289 effect of the coregulators bound by ERR α , that control different transcriptional targets and 290 thus phenotypic outcome ¹²⁴.

291 Several studies demonstrated an interplay between ERR α and the Wnt/ β -catenin signaling 292 ^{91,101,125}. In cancer cells, ERR α and β -catenin interact, and depend on each other to regulate 293 the expression of genes known to be involved in cell migration such as WNT11, MSX1 and N-294 *cadherin*. Specifically, the ERR α/β -catenin/WNT11 axis plays a role in promoting prostate and breast cancer cell migration *in vitro*⁹¹. Moreover, ERRα interacts with LSD1 and induces its 295 296 demethylase activity on the lysine 9 of histone 3 at the promoter of common target genes, 297 resulting in their transcriptional activation. In this way, ERRa and LSD1 promote breast cancer 298 cell invasion *in vitro* through the stimulation of the Matrix Metalloproteinase 1 (MMP1) 299 expression ⁸⁶. Noteworthy, the nuclear respiratory factor 1 (NRF1) is required for the 300 recruitment of LSD1 at the transcriptional start site of the ERR α -LSD1-positively regulated 301 target genes and participates in the regulation of MMP1 expression and breast cancer cell 302 invasion ⁸⁷. Other matrix metalloproteinases, such as MMP13, also regulated by ERRα, could 303 be involved in the degradation of the basement membrane, facilitating cell invasion ⁹⁵. In 304 addition, ERR α promotes the proliferation, migration and invasion of pancreatic cancer cell 305 lines by enhancing the expression of the plasminogen activator inhibitor 1 (PAI1) and 306 consequently activating the mitogen-activated protein kinase/extracellular signal-regulated 307 kinase (MEK/ERK) pathway ⁷⁴.

Surprisingly, ERRα can also have a negative impact on breast cancer cell proliferation and migration *in vitro* and can decrease lung colonization *in vivo* by down-regulating the expression of the 40S ribosomal S6 kinase 1 (S6K1), a downstream effector of the mTOR pathway. As a consequence, ERRα knockdown leads to an upregulation of these cellular

312 processes and sensitizes the cells to mTOR inhibitors under reduced serum or glucose 313 conditions ¹²⁶. Interestingly, a connection between ERRα and mTOR signaling has been found 314 previously, and in particular, ERRa has been implicated in the mTOR-mediated control of 315 energy metabolism in the mouse liver 127 . Therefore, it is possible that, when ERR α contributes 316 to the metabolic adaptation of the cells, it can repress or activate cell proliferation and 317 migration depending on the cellular context. In line with this hypothesis, other studies 318 indicated that the cofactor PGC-1 α requires ERR α to suppress prostate cancer metastasis; this 319 relies on their capacity to modulate metabolic programs and to decrease the expression of 320 specific genes, such as MYC, involved in cytoskeletal remodeling and prostate cancer cell invasion ^{128,129}. On the other hand, ERRα has been shown to promote breast cancer metastasis 321 322 to the bone, and the receptor activator of nuclear factor κ B (RANK) identified as an ERR α 323 target gene may contribute to this process. In this context, ERR α stimulates mTOR/S6K signaling and breast cancer cell migration *in vitro* in a RANK-dependent manner ¹³⁰. 324

325 Upregulated expression of the long non-coding RNA (IncRNA) SPRY4 intronic transcript 1 326 (SPRY4-T1) has been found to be associated with a poor prognosis in hepatocellular carcinoma 327 (HCC). In vitro, the knock-down of SPRY4-T1 leads to the downregulation of the ERRa 328 expression at the mRNA level and the effects of SPRY4-T1 on HCC cell proliferation, colony 329 formation, cell migration and invasion involve ERRa, suggesting a functional interaction 330 between SPRY4-T1 and ERRa¹²¹. Furthermore, ectopic expression of the microRNA 137 (miR-331 137) has been shown to reduce the expression of ERR α by directly targeting the 3'UTR region 332 of its mRNA in breast cancer cell lines. As a consequence, miR-137 decreases the proliferative 333 and invasive capacities of the cells, at least in part, through the negative regulation of the ERRα target genes *CCNE1* and *WNT11*⁷⁷. Similarly, miR-135a and miR-497 down-regulate the 334 335 expression of ERRα and decrease cancer cell migration and invasion *in vitro*^{131,132}. Notably,

miR-497 can control cell behavior by reducing the expression of the macrophage migration
inhibitory factor (MIF) and the activation of MMP9, two factors also regulated by ERRα ¹³².
Importantly, deregulated expression of these miRNA has been reported in various types of
cancers (^{77,131,132}). This may thus contribute to the aberrant expression of ERRα in cancers,
thereby facilitating cell proliferation, migration and invasion.

341

342 ERR α coordinates the basic processes of cell migration

343 Cell migration mainly depends on the dynamic organization of the actin and microtubule 344 cytoskeletons, and requires adhesion to the ECM (reviewed in ^{5,7}) (Fig. 1). Although numerous 345 studies have reported a role of ERR α in cell migration, its involvement in cell adhesion has 346 only been recently suggested. Zou and colleagues previously noticed that ERR α facilitates the 347 attachment of prostate cancer cells to fibronectin, an ECM protein, but they did not explore the underlying mechanisms 81 . As mentioned above, another report showed that ERR α 348 349 depletion in ECs leads to the upregulation of genes linked to cell adhesion and cell migration 350 ⁹³. Furthermore, bioinformatics analyses revealed that the RANK-ERRα regulatory interactome 351 contains proteins associated with ECM regulation, cell adhesion and cell migration, some of 352 which being encoded by putative ERR α target genes ¹³⁰. Altogether, these studies have 353 proposed new potential effectors of ERRa involved in cell adhesion and migration that need 354 further investigation.

Recent evidences indicated that ERRα plays an important role in regulating both the actin cytoskeleton and cell adhesion (Fig. 3). In that way, ERRα has been shown to regulate the actin cytoskeletal organization, the adhesion and migratory capacities of osteoclasts *in vitro* ⁹². In addition, ERRα promotes orientated migration of breast cancer cells by modulating the stability of the RhoA GTPase *in vitro*. Interestingly, increased RhoA activity resulting from ERRα

360 depletion induce cell disorientation that may result from impaired actin cytoskeletal 361 organization and defective lamellipodium formation ⁸⁵. Thereafter, it has been demonstrated 362 that ERR α controls actin polymerisation in these cells through the modulation of the RhoA-ROCK-LIMK-cofilin pathway. In addition, ERR α tightly regulates the expression of mitogen-363 364 activated protein kinase kinase kinase kinase (MAP4K4), and consequently facilitates the 365 turnover of focal adhesions, the main adhesive structures of the cells ¹³³. Thus, these data 366 establish a role for ERR α in coordinating focal adhesions and actin dynamics, that may 367 contribute to cell migration in both physiological and pathological conditions.

368

369 **Concluding remarks**

370 ERR α has been shown to play a role in various biological processes that contribute to 371 embryonic development, physiology and pathologies. Interestingly, as summarized here, 372 ERR α is a key regulator of cell migration in all of these contexts (Fig. 2). ERR α controls the 373 morphogenetic movement of cells during gastrulation in the embryo and also tightly modulate 374 the migration of different cell types in the adult organism, thereby ensuring tissue 375 homeostasis. In addition, ERR α has been widely implicated in cancer cell migration and 376 invasion. Consequently, ERR α has often been proposed as a therapeutic target to inhibit 377 cancer cell dissemination. However, global pharmacological modulation of ERR α activity or 378 expression could cause undesired side-effects as they would impact on the physiological 379 effects of the receptor. This strategy may thus be reconsidered. Recently, new cofactors of 380 ERR α have been shown to be involved in the modulation of distinct sets of ERR α -target genes ¹²³, suggesting that the capacity of ERR α to regulate a given cellular process depends on its 381 382 association with specific cofactors in a context-dependent manner. In that way, cofactors such 383 as LSD1 or SET7 regulate a precise subset of ERR α -target genes involved in cancer cell

migration ^{86,123}. Targeting the complex comprising ERR α and one of these cofactors may thus constitute a promising alternative approach to limit cancer spread. However, further investigations are required to achieve a better understanding of the cofactor-dependent mode of action of ERR α in tumors and other pathological contexts. Besides, it appears also important to dig more deeply into the factors and the mechanisms involved in the ERR α mediated cell movements in physiological environments.

390

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396

Author contributions

398 CF drafted the manuscript and prepare the figures. JMV provide critical feedbacks and 399 contributed to the revisions of the manuscript. JM and CF approved the final version of the 400 manuscript.

401

402 **Conflict of Interest**

403 The authors declare that the research was conducted in the absence of any commercial or404 financial relationships that could be construed as a potential conflict of interest.

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408 References

- 409 Mayor R, Etienne-Manneville, S. The front and rear of collective cell migration. Nat 1 410 *Rev Mol Cell Biol* 2016; 17: 97–109.
- Bravo-Cordero JJ, Hodgson L, Condeelis J. Directed cell invasion and migration 411 2
- 412 during metastasis. Current Opinion in Cell Biology 2012; 24: 277-283.
- 413 De Pascalis C, Etienne-Manneville S. Single and collective cell migration: the 3 414 mechanics of adhesions. Mol Biol Cell 2017; 28: 1833-1846.
- 415 SenGupta S, Parent CA, Bear JE. The principles of directed cell migration. Nat Rev 4 416 Mol Cell Biol 2021; 22: 529-547.
- Gardel ML, Schneider IC, Aratyn-Schaus Y, Waterman CM. Mechanical Integration 417 5 of Actin and Adhesion Dynamics in Cell Migration. Annu Rev Cell Dev Biol 2010; 26: 315-418 419 333.
- 420 6 Lawson CD, Ridley AJ. Rho GTPase signaling complexes in cell migration and 421 invasion. J Cell Biol 2018; 217: 447-457.
- 422 Seetharaman S, Etienne-Manneville S. Cytoskeletal Crosstalk in Cell Migration. 7 423 Trends in Cell Biology 2020; 30: 720–735.
- 424 Villena JA, Kralli A. ERRa: a metabolic function for the oldest orphan. Trends 8 425 Endocrinol Metab 2008; 19: 269–276.
- 426 Tam IS, Giguère V. There and back again: The journey of the estrogen-related 9
- 427 receptors in the cancer realm. The Journal of Steroid Biochemistry and Molecular Biology 428 2016; 157: 13-19.
- 429 Ranhotra HS. The estrogen-related receptors in metabolism and cancer: newer 10 430 insights. Journal of Receptors and Signal Transduction 2018; 38: 95-100.
- 431 Giguère V, Yang N, Segui P, Evans RM. Identification of a new class of steroid 11 432 hormone receptors. Nature 1988; 331: 91-94.
- 433 Crevet L, Vanacker J-M. Regulation of the expression of the estrogen related receptors 12 (ERRs). Cell Mol Life Sci 2020; 77: 4573-4579. 434
- 435 Tripathi M, Yen PM, Singh BK. Estrogen-Related Receptor Alpha: An Under-13
- Appreciated Potential Target for the Treatment of Metabolic Diseases. IJMS 2020; 21: 1645. 436
- 437 Sever R, Glass CK. Signaling by Nuclear Receptors. Cold Spring Harbor Perspectives 14 in Biology 2013; 5: a016709-a016709. 438
- 439 15
- Huss JM, Garbacz WG, Xie W. Constitutive activities of estrogen-related receptors: 440 Transcriptional regulation of metabolism by the ERR pathways in health and disease.
- 441 Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease 2015; 1852: 1912–1927.
- 442 16 Sladek R, Bader JA, Giguère V. The orphan nuclear receptor estrogen-related receptor
- 443 alpha is a transcriptional regulator of the human medium-chain acyl coenzyme A 444 dehydrogenase gene. Mol Cell Biol 1997; 17: 5400-5409.
- 445 Vanacker J-M. Transcriptional Activities of the Orphan Nuclear Receptor ERR 17
- 446 (Estrogen Receptor-Related Receptor-). Molecular Endocrinology 1999; 13: 764-773.
- 447 Mohideen-Abdul K, Tazibt K, Bourguet M, Hazemann I, Lebars I, Takacs M et al. 18
- 448 Importance of the Sequence-Directed DNA Shape for Specific Binding Site Recognition by 449 the Estrogen-Related Receptor. Front Endocrinol 2017; 8: 140.
- 450 Patel AKM, Vilela P, Shaik TB, McEwen AG, Hazemann I, Brillet K et al. 19
- Asymmetric dimerization in a transcription factor superfamily is promoted by allosteric 451 452 interactions with DNA. Nucleic Acids Research 2023; 51: 8864-8879.
- 453 Kallen J, Schlaeppi J-M, Bitsch F, Filipuzzi I, Schilb A, Riou V et al. Evidence for 20
- 454 Ligand-independent Transcriptional Activation of the Human Estrogen-related Receptor a
- 455 (ERRα). J Biol Chem 2004; 279: 49330–49337.
- 456 Bianco S, Sailland J, Vanacker J-M. ERRs and cancers: Effects on metabolism and on 21

- 457 proliferation and migration capacities. *The Journal of Steroid Biochemistry and Molecular*
- 458 *Biology* 2012; **130**: 180–185.
- 459 22 Deblois G, Giguère V. Functional and physiological genomics of estrogen-related
- receptors (ERRs) in health and disease. *Biochimica et Biophysica Acta (BBA) Molecular Basis of Disease* 2011; 1812: 1032–1040.
- 462 23 Ariazi EA, Kraus RJ, Farrell ML, Jordan VC, Mertz JE. Estrogen-Related Receptor 1
- 463 Transcriptional Activities Are Regulated in Part via the ErbB2/HER2 Signaling Pathway.
 464 *Molecular Cancer Research* 2007; **5**: 71–85.
- 465 24 Barry JB, Giguere V. Epidermal Growth Factor–Induced Signaling in Breast Cancer
- 466 Cells Results in Selective Target Gene Activation by Orphan Nuclear Receptor Estrogen467 Related Receptor A. *Cancer Res* 2005; 65: 6120–6129.
- 468 25 Tremblay AM, Wilson BJ, Yang X-J, Giguère V. Phosphorylation-Dependent
- 469 Sumoylation Regulates Estrogen-Related Receptor- α and - γ Transcriptional Activity through 470 a Synergy Control Motif. *Molecular Endocrinology* 2008; **22**: 570–584.
- 471 26 Vu EH, Kraus RJ, Mertz JE. Phosphorylation-Dependent Sumoylation of Estrogen472 Related Receptor α1. *Biochemistry* 2007; 46: 9795–9804.
- 473 27 Wilson BJ, Tremblay AM, Deblois G, Sylvain-Drolet G, Giguère V. An Acetylation
- 474 Switch Modulates the Transcriptional Activity of Estrogen-Related Receptor α. *Mol*
- 475 *Endocrinol* 2010; **24**: 1349–1358.
- 476 28 Audet-walsh É, Giguére V. The multiple universes of estrogen-related receptor α and γ 477 in metabolic control and related diseases. *Acta Pharmacol Sin* 2015; **36**: 51–61.
- 478 29 Alaynick WA, Kondo RP, Xie W, He W, Dufour CR, Downes M et al. ERRγ Directs
- and Maintains the Transition to Oxidative Metabolism in the Postnatal Heart. *Cell Metabolism* 2007; 6: 13–24.
- 481 30 Dufour CR, Wilson BJ, Huss JM, Kelly DP, Alaynick WA, Downes M *et al.* Genome-482 wide Orchestration of Cardiac Functions by the Orphan Nuclear Receptors ERR α and γ . *Cell* 483 *Metabolism* 2007; **5**: 345–356.
- 484 31 Carnesecchi J, Vanacker J-M. Estrogen-Related Receptors and the control of bone cell
 485 fate. *Molecular and Cellular Endocrinology* 2016; 432: 37–43.
- 486 32 Guo X, Yue L, Li M, Dai A, Sun J, Fang L *et al.* Nuclear receptor estrogen-related
 487 receptor gamma suppresses colorectal cancer aggressiveness by regulating Wnt/β-catenin
 488 signaling. *Carcinogenesis* 2022; **43**: 865–873.
- 489 33 Gulwani D, Upadhyay P, Goel R, Sarangthem V, Debraj Singh T. Unfolding of
- 490 Imminent Bio-Signatures in the Prognosis of Thyroid Cancer; The Emergence of Estrogen
- 491 Related Receptor Gamma (ERRγ) as a Hurricane. *Asian Pac J Cancer Prev* 2023; 24: 375–
 492 387.
- 493 34 Tiraby C, Hazen BC, Gantner ML, Kralli A. Estrogen-Related Receptor gamma
- 494 promotes mesenchymal-to-epithelial transition and suppresses breast tumor growth. *Cancer* 495 *Res* 2011; **71**: 2518–2528.
- 496 35 Kang M-H, Choi H, Oshima M, Cheong J-H, Kim S, Lee JH et al. Estrogen-related
- receptor gamma functions as a tumor suppressor in gastric cancer. *Nat Commun* 2018; 9:
 1920.
- An Argen Stephen Ste
- 502 37 Ijichi N, Shigekawa T, Ikeda K, Horie-Inoue K, Fujimura T, Tsuda H et al. Estrogen-
- related receptor γ modulates cell proliferation and estrogen signaling in breast cancer. *The Journal of Steroid Biochemistry and Molecular Biology* 2011; **123**: 1–7.
- 38 Heckler MM, Thakor H, Schafer CC, Riggins RB. ERK/MAPK regulates ERRγ
 506 expression, transcriptional activity, and receptor-mediated Tamoxifen resistance in ER+

- 507 breast cancer. *FEBS J* 2014; **281**: 2431–2442.
- 508 39 Kim J-H, Choi Y-K, Byun J-K, Kim M-K, Kang YN, Kim SH *et al.* Estrogen-related 509 receptor γ is upregulated in liver cancer and its inhibition suppresses liver cancer cell
- proliferation via induction of p21 and p27. *Exp Mol Med* 2016; **48**: e213–e213.
- 511 40 Hua T, Wang X, Chi S, Liu Y, Feng D, Zhao Y *et al.* Estrogen-related receptor γ
- promotes the migration and metastasis of endometrial cancer cells by targeting S100A4. *Oncol Rep* 2018; 40: 823–832.
- 514 41 Zhou W, Liu Z, Wu J, Liu J, Hyder SM, Antoniou E *et al.* Identification and
- 515 Characterization of Two Novel Splicing Isoforms of Human Estrogen-Related Receptor β.
 516 *The Journal of Clinical Endocrinology & Metabolism* 2006; **91**: 569–579.
- 517 42 Tiek DM, Khatib SA, Trepicchio CJ, Heckler MM, Divekar SD, Sarkaria JN *et al.* 518 Estrogen-related receptor β activation and isoform shifting by cdc2-like kinase inhibition 519 restricts migration and intracranial tumor growth in glioblastoma. *FASEB J* 2019; **33**: 13476–
- 520 13491.
- 43 Heckler MM, Zeleke TZ, Divekar SD, Fernandez AI, Tiek DM, Woodrick J *et al.*522 Antimitotic activity of DY131 and the estrogen-related receptor beta 2 (ERRβ2) splice variant
 523 in broast concert concertained 2016; 7: 47201, 47220
- 523 in breast cancer. *Oncotarget* 2016; 7: 47201–47220.
- 44 Ranhotra HS. Estrogen-related receptor alpha in select host functions and cancer: new
 frontiers. *Mol Cell Biochem* 2022; 477: 1349–1359.
- 526 45 Huss JM, Kopp RP, Kelly DP. Peroxisome Proliferator-activated Receptor
- 527 Coactivator-1α (PGC-1α) Coactivates the Cardiac-enriched Nuclear Receptors Estrogen-
- 528 related Receptor-α and -γ. *J Biol Chem* 2002; **277**: 40265–40274.
- 529 46 Schreiber SN, Knutti D, Brogli K, Uhlmann T, Kralli A. The Transcriptional
- 530 Coactivator PGC-1 Regulates the Expression and Activity of the Orphan Nuclear Receptor
- 531 Estrogen-Related Receptor α (ERRα). *J Biol Chem* 2003; **278**: 9013–9018.
- 532 47 Deblois G, Giguère V. Oestrogen-related receptors in breast cancer: control of cellular
 533 metabolism and beyond. *Nature Reviews Cancer* 2013; 13: 27–36.
- Vanacker J-M, Bonnelye E, Delmarre C, Laudet V. Activation of the thyroid hormone
 receptor α gene promoter by the orphan nuclear receptor ERRα. *Oncogene* 1998; 17: 2429–
 2435.
- 537 49 Singh BK, Sinha RA, Tripathi M, Mendoza A, Ohba K, Sy JAC et al. Thyroid
- bormone receptor and ERRα coordinately regulate mitochondrial fission, mitophagy,
 biogenesis, and function. *Sci Signal* 2018; 11: eaam5855.
- 540 50 Flamini MI, Uzair ID, Pennacchio GE, Neira FJ, Mondaca JM, Cuello-Carrión FD et
- 541 *al.* Thyroid Hormone Controls Breast Cancer Cell Movement via Integrin
- 542 αV/β3/SRC/FAK/PI3-Kinases. *Horm Cancer* 2017; **8**: 16–27.
- 543 51 Uzair ID, Conte Grand J, Flamini MI, Sanchez AM. Molecular Actions of Thyroid
- Hormone on Breast Cancer Cell Migration and Invasion via Cortactin/N-WASP. Front
 Endocrinol (Lausanne) 2019; 10: 139.
- 546 52 Bonnelye E, Vanacker JM, Spruyt N, Alric S, Fournier B, Desbiens X *et al.*
- 547 Expression of the estrogen-related receptor 1 (ERR-1) orphan receptor during mouse
- 548 development. *Mechanisms of Development* 1997; **65**: 71–85.
- 549 53 Huss JM, Torra IP, Staels B, Giguère V, Kelly DP. Estrogen-Related Receptor α
- 550 Directs Peroxisome Proliferator-Activated Receptor α Signaling in the Transcriptional
- 551 Control of Energy Metabolism in Cardiac and Skeletal Muscle. *Mol Cell Biol* 2004; 24:
 552 9079–9091.
- 553 54 Murray J, Huss JM. Estrogen-related receptor α regulates skeletal myocyte
- differentiation via modulation of the ERK MAP kinase pathway. *American Journal of Physiology-Cell Physiology* 2011; **301**: C630–C645.
- 556 55 Shao D, Liu Y, Liu X, Zhu L, Cui Y, Cui A et al. PGC-1β-Regulated mitochondrial

- biogenesis and function in myotubes is mediated by NRF-1 and ERRα. *Mitochondrion* 2010;
 10: 516–527.
- 559 56 Wei W, Schwaid AG, Wang X, Wang X, Chen S, Chu Q *et al.* Ligand Activation of
- 560 ERRα by Cholesterol Mediates Statin and Bisphosphonate Effects. *Cell Metabolism* 2016; 23:
 561 479–491.
- 562 57 Carnesecchi J, Vanacker J-M. Estrogen-Related Receptors and the control of bone cell
 563 fate. *Molecular and Cellular Endocrinology* 2016; 432: 37–43.
- 564 58 LaBarge S, McDonald M, Smith-Powell L, Auwerx J, Huss JM. Estrogen-related
- receptor-α (ERRα) deficiency in skeletal muscle impairs regeneration in response to injury.
 The FASEB Journal 2014; 28: 1082–1097.
- 567 59 Delhon I, Gutzwiller S, Morvan F, Rangwala S, Wyder L, Evans G *et al.* Absence of
 568 Estrogen Receptor-Related-α Increases Osteoblastic Differentiation and Cancellous Bone
 569 Mineral Density. *Endocrinology* 2009; **150**: 4463–4472.
- 570 60 Teyssier C, Gallet M, Rabier B, Monfoulet L, Dine J, Macari C et al. Absence of
- 571 ERRα in Female Mice Confers Resistance to Bone Loss Induced by Age or Estrogen572 Deficiency. *PLoS ONE* 2009; 4: e7942.
- 573 61 Gallet M, Saïdi S, Haÿ E, Photsavang J, Marty C, Sailland J et al. Repression of
- 574 osteoblast maturation by ERRα accounts for bone loss induced by estrogen deficiency. *PLoS*575 *ONE* 2013; 8: e54837.
- 576 62 Bonnelye E, Zirngibl RA, Jurdic P, Aubin JE. The Orphan Nuclear Estrogen
 577 Receptor-Related Receptor-α Regulates Cartilage Formation *in Vitro* : Implication of Sox9.
 578 *Endocrinology* 2007; **148**: 1195–1205.
- 63 Rajalin A-M, Pollock H, Aarnisalo P. ERRα regulates osteoblastic and adipogenic
 580 differentiation of mouse bone marrow mesenchymal stem cells. *Biochemical and Biophysical*
- 581 *Research Communications* 2010; **396**: 477–482.
- 582 64 Ariazi EA, Clark GM, Mertz JE. Estrogen-related Receptor alpha and Estrogen-related
 583 Receptor gamma Associate with Unfavorable and Favorable Biomarkers, Respectively, in
 584 Human Breast Cancer. 2002; 62: 6510–6518.
- 585 65 Suzuki T, Miki Y, Moriya T, Shimada N, Ishida T, Hirakawa H *et al.* Estrogen586 Related Receptor Alpha in Human Breast Carcinoma as a Potent Prognostic Factor. 2004; 64:
 587 4670–4676.
- 588 66 Cavallini A, Notarnicola M, Giannini R, Montemurro S, Lorusso D, Visconti A *et al.*
- 589 Oestrogen receptor-related receptor alpha (ERRα) and oestrogen receptors (ERα and ERβ)
 590 exhibit different gene expression in human colorectal tumour progression. *European Journal*
- 591 *of Cancer* 2005; **41**: 1487–1494.
- 592 67 Sun P, Sehouli J, Denkert C, Mustea A, Könsgen D, Koch I et al. Expression of
- estrogen receptor-related receptors, a subfamily of orphan nuclear receptors, as new tumor
 biomarkers in ovarian cancer cells. *J Mol Med* 2005; 83: 457–467.
- 595 68 Cheung CP, Yu S, Wong KB, Chan LW, Lai FMM, Wang X *et al.* Expression and 596 Functional Study of Estrogen Receptor-Related Receptors in Human Prostatic Cells and
- 597 Tissues. *The Journal of Clinical Endocrinology & Metabolism* 2005; **90**: 1830–1844.
- 598 69 Fujimura T, Takahashi S, Urano T, Kumagai J, Ogushi T, Horie-Inoue K *et al.*599 Increased expression of estrogen-related receptor α (ERRα) is a negative prognostic predictor
 600 in human prostate cancer. *International Journal of Cancer* 2007; **120**: 2325–2330.
- in human prostate cancer. *International Journal of Cancer* 2007; 120: 2325–2330.
 Fujimoto J, Alam SM, Jahan I, Sato E, Sakaguchi H, Tamaya T. Clinical implication
- 602 of estrogen-related receptor (ERR) expression in ovarian cancers. *The Journal of Steroid*
- 603 Biochemistry and Molecular Biology 2007; 104: 301–304.
- 604 71 Fujimoto J, Sato E. Clinical implication of estrogen-related receptor (ERR) expression
- 605 in uterine endometrial cancers. *The Journal of Steroid Biochemistry and Molecular Biology*
- 606 2009; **116**: 71–75.

- 607 Tiwari A, Swamy S, Gopinath KS, Kumar A. Genomic amplification upregulates 72 608 estrogen-related receptor alpha and its depletion inhibits oral squamous cell carcinoma tumors 609 in vivo. Sci Rep 2015; 5: 17621. Li P, Wang J, Wu D, Ren X, Wu W, Zuo R et al. ERRa is an aggressive factor in lung 610 73 611 adenocarcinoma indicating poor prognostic outcomes. Cancer Manag Res 2019; 11: 8111-612 8123. 613 74 Liu S-L, Wu X-S, Li F-N, Yao W-Y, Wu Z-Y, Dong P et al. ERRa promotes pancreatic cancer progression by enhancing the transcription of PAI1 and activating the 614 615 MEK/ERK pathway. Am J Cancer Res 2020; 10: 3622-3643. 616 Bernatchez G, Giroux V, Lassalle T, Carpentier AC, Rivard N, Carrier JC. ERRa 75 617 metabolic nuclear receptor controls growth of colon cancer cells. *Carcinogenesis* 2013; 34: 618 2253-2261. 619 76 Bianco S, Lanvin O, Tribollet V, Macari C, North S, Vanacker J-M. Modulating 620 estrogen receptor-related receptor-alpha activity inhibits cell proliferation. J Biol Chem 2009; 621 **284**: 23286–23292. Zhao Y, Li Y, Lou G, Zhao L, Xu Z, Zhang Y et al. MiR-137 Targets Estrogen-622 77 623 Related Receptor Alpha and Impairs the Proliferative and Migratory Capacity of Breast 624 Cancer Cells. PLoS One 2012; 7: e39102. 625 78 Mukherjee T, Malik P, Hoidal J. The emerging role of estrogen related receptora in 626 complications of non-small cell lung cancers (Review). Oncol Lett 2021; 21: 258. 627 Stein RA, Gaillard S, McDonnell DP. Estrogen-related receptor alpha induces the 79 628 expression of vascular endothelial growth factor in breast cancer cells. J Steroid Biochem Mol 629 Biol 2009; 114: 106–112. 630 80 Zhang K, Lu J, Mori T, Smith-Powell L, Synold TW, Chen S et al. Baicalin increases 631 VEGF expression and angiogenesis by activating the ERR{alpha}/PGC-1{alpha} pathway. 632 Cardiovasc Res 2011; 89: 426-435. 633 Zou C, Yu S, Xu Z, Wu D, Ng C-F, Yao X et al. ERRα augments HIF-1 signalling by 81 634 directly interacting with HIF-1a in normoxic and hypoxic prostate cancer cells. The Journal 635 of Pathology 2014; 233: 61-73. 636 Ao A, Wang H, Kamarajugadda S, Lu J. Involvement of estrogen-related receptors in 82 637 transcriptional response to hypoxia and growth of solid tumors. Proc Natl Acad Sci USA 638 2008; 105: 7821–7826. 639 Tennessen JM, Baker KD, Lam G, Evans J, Thummel CS. The Drosophila estrogen-83 640 related receptor directs a metabolic switch that supports developmental growth. Cell Metab 641 2011; 13: 139–148. 642 Cai Q, Lin T, Kamarajugadda S, Lu J. Regulation of glycolysis and the Warburg effect 84 643 by estrogen-related receptors. Oncogene 2013; 32: 2079-2086. 644 Sailland J, Tribollet V, Forcet C, Billon C, Barenton B, Carnesecchi J et al. Estrogen-85 645 related receptor a decreases RHOA stability to induce orientated cell migration. Proc Natl 646 Acad Sci USA 2014; 111: 15108–15113. 647 Carnesecchi J, Forcet C, Zhang L, Tribollet V, Barenton B, Boudra R et al. ERRa 86 648 induces H3K9 demethylation by LSD1 to promote cell invasion. Proc Natl Acad Sci USA 649 2017; 114: 3909–3914. 650 87 Zhang L, Carnesecchi J, Cerutti C, Tribollet V, Périan S, Forcet C et al. LSD1-ERRa 651 complex requires NRF1 to positively regulate transcription and cell invasion. Sci Rep 2018; 8: 10041. 652 653 88 Weng S, Wallingford JB. May the force be with you. *eLife* 2018; 7: e36662. 654 89 Bardet P-L, Horard B, Laudet V, Vanacker J-M. The ERRa orphan nuclear receptor 655 controls morphogenetic movements during zebrafish gastrulation. Developmental Biology
- 656 2005; **281**: 102–111.

- 657 90 Ulrich F, Concha ML, Heid PJ, Voss E, Witzel S, Roehl H *et al.* Slb/Wnt11 controls
 658 hypoblast cell migration and morphogenesis at the onset of zebrafish gastrulation.
- 659 Development 2003; **130**: 5375–5384.
- Dwyer MA, Joseph J, Wade HE, Eaton ML, Kunder RS, Kazmin D et al. WNT11

661 expression is induced by ERR α and β -catenin and acts in an autocrine manner to increase 662 cancer cell migration. *Cancer Res* 2010; **70**: 9298–9308.

Bonnelye E, Saltel F, Chabadel A, Zirngibl RA, Aubin JE, Jurdic P. Involvement of
 the orphan nuclear estrogen receptor-related receptor α in osteoclast adhesion and

transmigration. J Mol Endocrinol 2010; 45: 365–377.

- 666 93 Likhite N, Yadav V, Milliman EJ, Sopariwala DH, Lorca S, Narayana NP *et al.* Loss
 667 of Estrogen-Related Receptor Alpha Facilitates Angiogenesis in Endothelial Cells. *Mol Cell*668 *Biol* 2019; **39**: e00411-18.
- 669 94 Arany Z, Foo S-Y, Ma Y, Ruas JL, Bommi-Reddy A, Girnun G et al. HIF-
- 670 independent regulation of VEGF and angiogenesis by the transcriptional coactivator PGC-1α.
 671 *Nature* 2008; **451**: 1008–1012.
- 672 95 Fradet A, Sorel H, Bouazza L, Goehrig D, Depalle B, Bellahcene A *et al.* Dual
- Function of ERR in Breast Cancer and Bone Metastasis Formation: Implication of VEGF and
 Osteoprotegerin. *Cancer Research* 2011; **71**: 5728–5738.
- 675 96 Zhang L-D, Chen L, Zhang M, Qi H-J, Chen L, Chen H-F et al. Downregulation of
- 676 ERR α inhibits angiogenesis in human umbilical vein endothelial cells through regulating
- VEGF production and PI3K/Akt/STAT3 signaling pathway. *European Journal of Pharmacology* 2015; **769**: 167–176.
- 679 97 Benslimane-Ahmim Z, Pereira J, Lokajczyk A, Dizier B, Galy-Fauroux I, Fischer A-
- 680 M *et al.* Osteoprotegerin regulates cancer cell migration through SDF-1/CXCR4 axis and 681 promotes tumour development by increasing neovascularization. *Cancer Letters* 2017; **395**:
- 682 11–19.
- 683 98 Sumi D, Ignarro LJ. Estrogen-related receptor 1 up-regulates endothelial nitric oxide
- synthase expression. *Proceedings of the National Academy of Sciences* 2003; 100: 14451–
 14456.
- 686 99 Xia W, Yin J, Zhang S, Guo C, Li Y, Zhang Y *et al.* Parkin Modulates ERRα/eNOS
 687 Signaling Pathway in Endothelial Cells. *Cell Physiol Biochem* 2018; **49**: 2022–2034.
- 688 100 Hu W, Huang Y. Targeting the platelet-derived growth factor signalling in
- 689 cardiovascular disease. *Clin Exp Pharmacol Physiol* 2015; **42**: 1221–1224.
- 690 101 Li Q, Zhu L, Zhang L, Chen H, Zhu Y, Du Y et al. Inhibition of estrogen related
- 691 receptor α attenuates vascular smooth muscle cell proliferation and migration by regulating
- 692 RhoA/p27 Kip1 and β-Catenin/Wnt4 signaling pathway. *European Journal of Pharmacology*693 2017; **799**: 188–195.
- Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial–mesenchymal
 transition. *Nat Rev Mol Cell Biol* 2014; 15: 178–196.
- Huang J, Guan B, Yin L, Liu F, Hu B, Zheng Q et al. Effects of estrogen-related
- receptor alpha (ERRα) on proliferation and metastasis of human lung cancer A549 cells. J *Huazhong Univ Sci Technol [Med Sci]* 2014; 34: 875–881.
- Wu Y-M, Chen Z-J, Liu H, Wei W-D, Lu L-L, Yang X-L *et al.* Inhibition of ERRα
 suppresses epithelial mesenchymal transition of triple negative breast cancer cells by directly
 targeting fibronectin. *Oncotarget* 2015; 6: 25588–25601.
- 702 105 Chen H, Pan J, Zhang L, Chen L, Qi H, Zhong M et al. Downregulation of estrogen-
- related receptor alpha inhibits human cutaneous squamous cell carcinoma cell proliferation
- and migration by regulating EMT via fibronectin and STAT3 signaling pathways. *European*
- 705 *Journal of Pharmacology* 2018; **825**: 133–142.
- 706 106 Yoriki K, Mori T, Kokabu T, Matsushima H, Umemura S, Tarumi Y et al. Estrogen-

- 707 related receptor alpha induces epithelial-mesenchymal transition through cancer-stromal
- 708 interactions in endometrial cancer. Sci Rep 2019; 9: 6697.
- 709 107 Wang L, Yang M, Guo X, Yang Z, Liu S, Ji Y et al. Estrogen-related receptor-a
- 710 promotes gallbladder cancer development by enhancing the transcription of Nectin-4. Cancer 711 Sci 2020; 111: 1514–1527.
- 712 108 Mao X, Lei H, Yi T, Su P, Tang S, Tong Y et al. Lipid reprogramming induced by the
- 713 TFEB-ERRα axis enhanced membrane fluidity to promote EC progression. J Exp Clin Cancer 714 *Res* 2022; **41**: 28.
- 715 Li F-N, Zhang Q-Y, Li O, Liu S-L, Yang Z-Y, Pan L-J et al. ESRRA promotes gastric 109
- 716 cancer development by regulating the CDC25C/CDK1/CyclinB1 pathway via DSN1. Int J 717 Biol Sci 2021; 17: 1909–1924.
- 718 Lam SS, Mak AS, Yam JW, Cheung AN, Ngan HY, Wong AS. Targeting Estrogen-110
- 719 Related Receptor Alpha Inhibits Epithelial-to-Mesenchymal Transition and Stem Cell 720 Properties of Ovarian Cancer Cells. Mol Ther 2014; 22: 743-751.
- 721 Ma J-H, Qi J, Lin S-Q, Zhang C-Y, Liu F, Xie W-D et al. STAT3 Targets ERR-α to 111
- 722 Promote Epithelial-Mesenchymal Transition, Migration, and Invasion in Triple-Negative 723 Breast Cancer Cells. Mol Cancer Res 2019; 17: 2184–2195.
- 724 Chen Y, Zhang K, Li Y, He Q. Estrogen-related receptor a participates transforming 112
- 725 growth factor- β (TGF- β) induced epithelial-mesenchymal transition of osteosarcoma cells.
- 726 *Cell Adh Migr* 2016; **11**: 338–346.
- 727 Huang X, Wang X, Shang J, Zhaang Z, Cui B, Lin Y et al. Estrogen related receptor 113 728 alpha triggers the migration and invasion of endometrial cancer cells via up regulation of 729 TGFB1. Cell Adh Migr 2018; 12: 538–547.
- 730 Luo J, Sladek R, Carrier J, Bader J-A, Richard D, Giguere V. Reduced Fat Mass in 114 731 Mice Lacking Orphan Nuclear Receptor Estrogen-Related Receptor Alpha. Mol Cell Biol 732 2003; 23: 7947–7956.
- 733 Villena JA, Hock MB, Chang WY, Barcas JE, Giguère V, Kralli A. Orphan nuclear 115
- 734 receptor estrogen-related receptor α is essential for adaptive thermogenesis. *Proc Natl Acad* 735 *Sci USA* 2007; **104**: 1418–1423.
- 736 Cui H, Lu Y, Khan MZ, Anderson RM, McDaniel L, Wilson HE et al. Behavioral 116 737 Disturbances in Estrogen-Related Receptor alpha-Null Mice. Cell Reports 2015; 11: 344-350.
- 738 De Jesús-Cortés H, Lu Y, Anderson RM, Khan MZ, Nath V, McDaniel L et al. Loss 117
- 739 of estrogen-related receptor alpha disrupts ventral-striatal synaptic function in female mice. 740 Neuroscience 2016: 329: 66-73.
- 741 118 Stein RA, Chang C, Kazmin DA, Way J, Schroeder T, Wergin M et al. Estrogen-
- 742 Related Receptor a Is Critical for the Growth of Estrogen Receptor–Negative Breast Cancer.
- 743 Cancer Res 2008; 68: 8805-8812.
- 744 Deblois G, Hall JA, Perry M-C, Laganiere J, Ghahremani M, Park M et al. Genome-119
- 745 Wide Identification of Direct Target Genes Implicates Estrogen-Related Receptor as a
- 746 Determinant of Breast Cancer Heterogeneity. Cancer Research 2009; 69: 6149-6157. 747 Deblois G, Chahrour G, Perry M-C, Sylvain-Drolet G, Muller WJ, Giguère V. 120
- 748
- Transcriptional Control of the ERBB2 Amplicon by ERRα and PGC-1β Promotes Mammary 749 Gland Tumorigenesis. Cancer Res 2010; 70: 10277–10287.
- 750 121 Yu G, Lin J, Liu C, Hou K, Liang M, Shi B. Long non-coding RNA SPRY4-IT1
- 751 promotes development of hepatic cellular carcinoma by interacting with ERRa and predicts 752 poor prognosis. Sci Rep 2017; 7. doi:10.1038/s41598-017-16781-9.
- 753 Avena P, De Luca A, Chimento A, Nocito MC, Sculco S, La Padula D et al. Estrogen 122
- 754 Related Receptor Alpha (ERRa) a Bridge between Metabolism and Adrenocortical Cancer 755 Progression. Cancers (Basel) 2022; 14: 3885.
- 756 123 Cerutti C, Zhang L, Tribollet V, Shi J-R, Brillet R, Gillet B et al. Computational

- 757 identification of new potential transcriptional partners of ERR α in breast cancer cells: specific partners for specific targets. Sci Rep 2022; 12: 3826. 758
- 759 124 Cerutti C, Shi J-R, Vanacker J-M. Multifaceted Transcriptional Network of Estrogen-760 Related Receptor Alpha in Health and Disease. Int J Mol Sci 2023; 24: 4265.
- Auld KL, Berasi SP, Liu Y, Cain M, Zhang Y, Huard C et al. Estrogen-related 761 125
- 762 receptor α regulates osteoblast differentiation via Wnt/β-catenin signaling. Journal of 763 Molecular Endocrinology 2012; 48: 177–191.
- Berman AY, Manna S, Schwartz NS, Katz YE, Sun Y, Behrmann CA et al. ERRa 764 126
- 765 regulates the growth of triple-negative breast cancer cells via S6K1-dependent mechanism. 766 Sig Transduct Target Ther 2017; 2: 17035.
- Chaveroux C, Eichner LJ, Dufour CR, Shatnawi A, Khoutorsky A, Bourque G et al. 767 127 768 Molecular and Genetic Crosstalks between mTOR and ERRa Are Key Determinants of
- 769 Rapamycin-Induced Nonalcoholic Fatty Liver. Cell Metabolism 2013; 17: 586–598.
- 770 Torrano V, Valcarcel-Jimenez L, Cortazar AR, Liu X, Urosevic J, Castillo-Martin M 128 771 et al. The metabolic co-regulator PGC1a suppresses prostate cancer metastasis. Nat Cell Biol
- 772 2016; 18: 645-656.
- 773 Valcarcel-Jimenez L, Macchia A, Crosas-Molist E, Schaub-Clerigué A, Camacho L, 129
- 774 Martín-Martín N et al. PGC1a Suppresses Prostate Cancer Cell Invasion through ERRa
- 775 Transcriptional Control. Cancer Res 2019; 79: 6153-6165.
- 776 130 Vargas G, Bouchet M, Bouazza L, Reboul P, Boyault C, Gervais M et al. ERRa promotes breast cancer cell dissemination to bone by increasing RANK expression in primary 777 778 breast tumors. Oncogene 2019; 38: 950-964.
- 779 Tribollet V, Barenton B, Kroiss A, Vincent S, Zhang L, Forcet C et al. miR-135a 131
- 780 Inhibits the Invasion of Cancer Cells via Suppression of ERRa. PLoS ONE 2016; 11: 781 e0156445.
- 782 132 Han L, Liu B, Jiang L, Liu J, Han S. MicroRNA-497 downregulation contributes to
- 783 cell proliferation, migration, and invasion of estrogen receptor alpha negative breast cancer by targeting estrogen-related receptor alpha. Tumor Biol 2016; 37: 13205-13214.
- 784
- 785 133 Tribollet V, Cerutti C, Géloën A, Berger E, De Mets R, Balland M et al. ERRa
- 786 coordinates actin and focal adhesion dynamics. Cancer Gene Ther 2022; 29: 1429-1438. 787

788 Figure legends

789

790 Figure 1: Core mechanisms of cell migration

791 Cell migration, whether during embryonic development or in adulthood, plays a crucial role in 792 various biological processes. At the onset of movement, cells undergo polarization and extend 793 protrusions in the direction of migration. These protrusions are predominantly driven by the 794 actin cytoskeleton and are stabilized by adhering to the extracellular matrix. Subsequently, 795 these adhesions generate traction stresses, which enable the cells to propel forward, and 796 disassembly of adhesion at the rear facilitates cell detachment. In addition, the microtubule 797 cytoskeleton, which governs intracellular trafficking and signaling also participates in 798 regulating the different steps of cell migration.

799

800 Figure 2: ERRα regulates cell migration in both physiological and pathological conditions

801 ERR α is involved in regulating morphogenetic movements during gastrulation and it also 802 influences the movements of various cell types that participate to tissue remodelling and 803 homeostasis. Furthermore, ERR α -mediated cell migration contributes to cardiovascular 804 diseases and cancer progression.

805

806 Figure 3: Molecular mechanisms through which ERRα finely controls migration potential

807 ERRα orchestrates cell movements through diverse mechanisms. ERRα engages in specific
808 cofactor interactions, regulates gene expression and signaling pathways. As a result, these
809 actions culminate in the modulation of cell migration and determine cellular responses based
810 on the context. Figure adapted from images created with <u>BioRender.com</u>.