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▶ To cite this version:

Lea Farouil, Muriel Sylvestre, Alain Fournet, Gerardo Cebrián-Torrejón. Review on canthin-6-one alkaloids: Distribution, chemical aspects and biological activities. European Journal of Medicinal Chemistry Reports, 2022, 5, pp.100049. 10.1016/j.ejmcr.2022.100049. hal-04722266

HAL Id: hal-04722266 https://hal.science/hal-04722266v1

Submitted on 13 Nov 2024

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Version of Record: https://www.sciencedirect.com/science/article/pii/S2772417422000218 Manuscript_db37f7bcec65afa966b572946505409c

Review on canthin-6-one alkaloids: distribution, chemical aspects and biological activities

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17 ABSTRACT

18 Canthin-6-one alkaloids have been a topic of research in many phytochemical and 19 pharmacological laboratories because they are a structurally unique and biologically intriguing 20 family. This review summarizes the diversity, biogenesis, chemistry and bioactivity of canthin-6-21 ones.

- 22 **Keywords:** canthin-6-one; β-carboline; alkaloids; synthesis; biological activity
- 23

24 **1. Introduction**

The canthin-6-one (or canthinone) alkaloids are a subclass of β -carboline alkaloids with an additional D-ring (Figure 1). Canthin-6-one (1) was first isolated in 1952 by Haynes [1] from the Australian plant species *Pentaceras australis* (F.Muell.) Benth. (Rutaceae). Since then, more than eighty members of this class of alkaloids have been reported from several plants primarily of the Rutaceae and Simaroubaceae families, but also from Malvaceae, Amaranthaceae, Caryophyllaceae,

- Zygophyllaceae and recently from fungi (*e.g. Boletus curtisii* Berk.). Several of these alkaloids have
 been bioassayed and show interesting pharmacological activities [2].
- 32
- 33



34 35

Figure 1. Canthin-6-one structure.

The biosynthesis of canthinone-type alkaloids was demonstrated by the Guicciardi [3] group by feeding experiments with cell cultures of *Ailanthus altissima* (Mill.) Swingle (Simaroubaceae). Synthetically speaking, there is a paucity of strategies for the synthesis of the basic canthinone scaffold; some of them present a biomimetic basis, starting from the natural precursors, tryptophan (2) or tryptamine (3) or derivatives and some others showing a purely synthetic approach.

42 A continuous increase in the number of scientific publications on "canthinones" has appeared over the 43 course of the last 70 years (Figure 2) showing an undeniable interest of these compounds [4]. Such reports included numerous biological studies that have confirmed the potential value of these natural 44 45 products. The latest review article concerning canthin-6-one alkaloids was published by Dai et al. 46 providing a broad overview of canthin-6-ones. The authors describe the biosynthetic assembly lines, 47 different synthetic pathway and biological activities of this class of alkaloids that have been published from 1952 to 2015 [5]. In most publications, canthinones alkaloids have been identified by classical 48 49 technics as NMR and Mass spectroscopies. However, over the last few years, other ways of 50 characterization have been used such as high-performance liquid chromatography (HPLC) with UV 51 detection [6,7] or innovative application of the voltammetry of microparticles [8] to identify the 52 phytochemical composition of plants extracts.



- 54
- Figure 2.Occurrence of the number of publications related to "canthin-6-one" from 1952 to 2021
 (Source: Scholar (28/10/2021).
- All related available information on canthin-6-one alkaloids and its distribution, chemical aspects and
 biological activities was collected via electronic search (using Google Scholar, Pubmed, Scopus and
- 60 Web of Science) without time limit.
- 61

62 **2. Natural occurrence of canthin-6-one alkaloids**

63 The canthinones are a small group of alkaloids; around ninety canthin-6-one alkaloids have 64 been isolated from more than fifty species. Table 1 illustrates the natural distribution and the diversity 65 of natural canthin-6-one alkaloids and classifies the canthin-6-one alkaloids taxonomically.

 Table 1.Canthin-6-one alkaloids and their natural occurrence.



- Rutaceae : Zanthoxylum dimorphophyllum [18]
- Simaroubaceae : Ailanthus altissima [3,9]

II- Canthin-6-ones mono-substituted by a heteroatom (O or S)

Mono hydrorylatad aarth:	$\begin{array}{c} 11 \\ 10 \\ 9 \\ 8 \\ 8 \\ 9 \\ 8 \\ 5 \\ 5 \\ 5 \\ 5 \\ 5 \\ 5 \\ 6 \\ 6 \\ 8 \\ 5 \\ 5 \\ 5 \\ 5 \\ 5 \\ 6 \\ 7 \\ 7 \\ 7 \\ 7 \\ 7 \\ 7 \\ 7 \\ 7 \\ 7$
wiono-nyuroxyiated cantini	
1-hydroxy-canthin-6-one	- Simaroubaceae : Attaninus attissima, Ailanthus giraldii [9,42], Eurycoma longifolia [27,29,43]
2-hydroxy-canthin-6-one	 Simaroubaceae : Quassia kerstingii [9], Simarouba glauca [31], Simaba bahiensis [44], Ailanthus altissima [3]
4-hydroxy-canthin-6-one	• Simaroubaceae : Ailanthus altissima [3,9]
5-hydroxy-canthin-6-one	• Simaroubaceae : Simarouba amara [9,45], Brucea mollis [24], Ailanthus altíssima [9]
8-hydroxy-canthin-6-one	 Simaroubaceae : Ailanthus excels [23], Odyendea gabonensis, Pierreodendrom kerstingii, Simaba multiflora [9], Picrasma quassioides [46,47]
9-hydroxy-canthin-6-one	 Simaroubaceae : Eurycoma longifolia [27,43,48,49], Eurycoma harmandiana [32], Pricrolemma granatensis [26], Simarouba berteroana [50]
10-hydroxy-canthin-6-one	Simaroubaceae : Eurycoma longifolia, Simaba multiflora [9]
	• Amaranthaceae : Aerva lanata [38,39]
11-hydroxy-canthin-6-one	 Simaroubaceae : Amaroria soulameoides [51], Brucea antydisenterica [23], Quassia kerstingii [9], Brucea mollis, Eurycoma longifolia [28,29,43], Picrasma quassioides [52]

Mono-methovylated canthi	$10 \xrightarrow{11}{10} \xrightarrow{1}{10} \xrightarrow{1}{1$
1-methoxycanthin-6-one	 Simaroubaceae : Ailanthus altissima, Ailanthus excelsa, Soulamea pancheri [23], Ailanthus giraldii, Brucea antidysenterica, Hannoa klaineana [9], Brucea antidysenterica [35], Picrasma quassioides [52] Caryophylaceae : Drymaria diandra [53] Leitneriogeae : Leitneria floridana [54]
	• Lettherlaceae : Lettherla Jortaana [34]
2-methoxycanthin-6-one	• Simaroubaceae : Simarouba glauca [30,31], Brucea antidysenterica [35], Simaba bahiensis [55], Quassia amara [56]
4-methoxycanthin-6-one	 Amaranthaceae : Charpentiana obovata [9] Simaroubaceae : Simaba ferruginea [33], Eurycoma longifolia [29], Picrasma quassioides [52] Caryophylaceae : Drymaria cordata [9], Drymaria diandra [57]
5-methoxycanthin-6-one	 Rutaceae : Pentaceras australis, Zanthoxylum elephantiasis [11], Zanthoxylum chiloperone [14,58], Zanthoxylum rugosum [59] Simaroubaceae : Ailanthus excels [33], Odyendea gabonensis, Picrasma excels [9], Picrasma quassioides [9, 23, 37], Brucea antidysenterica [36], Eurycoma longifolia [27,29], Simarouba amara [45], Leitneria floridana [60] Leitneriaceae : Leitneria floridana [24,54]
8-methoxycanthin-6-one	• Simaroubaceae : Simaba cuspidata [61]
9-methoxycanthin-6-one	 Simaroubaceae : Simaba cuspidata, Simaba multiflora [9,62], Eurycoma longifolia [27- 29,43,48,49], Pricrolemma granatensis [26], Eurycoma harmandiana [32], Simarouba glauca [30,31], Simarouba berteroana [50], Simaba polyphylla [63]
	• Rutaceae : Zanthoxylum paracanthum [22]
10-methoxycanthin-6-one	 Simaroubaceae: Simaba multiflora [9], Eurycoma longifolia [27] Amaranthaceae : Aerva lanata [38,39] Rutaceae : Zanthoxylum paracanthum [21]
11-methoxycanthin-6-one	• Simaroubaceae : Amaroria soulameoides [51]





Thiomethylated canthin-6-ones

1 mathalthic conthin 6 and	 Boletineae : Boletus curtisii [41] 	
4-methyltmocanthin-6-one	• Rutaceae : <i>Pentaceras australis</i> [9]	
5-methylthiocanthin-6-one	Boletineae : Boletus curtisii [41]	
9-methylthiocanthin-6-one	Boletineae : Boletus curtisii [41]	
11-methylthiocanthin-6-one	Boletineae : Boletus curtisii [41]	
4-methylthiocanthin-6-one 5-methylthiocanthin-6-one 9-methylthiocanthin-6-one 11-methylthiocanthin-6-one	 Rutaceae : Pentaceras australis [9] Boletineae : Boletus curtisii [41] Boletineae : Boletus curtisii [41] Boletineae : Boletus curtisii [41] 	



Canthin-6-one-1- O - β -D-xylopyranosyl-(1' \rightarrow 6')- β -D-glucopyranoside	
R ₁ =O-glc	
$R_{2}=H, R_{6}=HO^{(1)}HO^{(1)}HO^{(1)}$	 Simaroubaceae: Ailanthus altissima [65]
Canthin-6-one-5- O - β -D-xylopyranosyl-(1' \rightarrow 6')- β - D-glucopyranoside R ₅ = O -glc HO'' \rightarrow β β \rightarrow	 Simaroubaceae: Ailanthus altissima [65]
Canthin-6-one-1- O -[6- O -(3-hydroxy-3- methylglutaryl)]- β -D-glucopyranoside R ₁ = O -glc R_2 =H, R ₆ = O HO OH	• Simaroubaceae : Ailanthus altissima [66]
canthin-6-one-1- <i>O</i> - β -d-apiofuranosyl-(1' \rightarrow 2')- β -D- glucopyranoside R ₁ = <i>O</i> -glc HO HO HO HO HO $R_{6'}$ = H	• Simaroubaceae : Ailanthus altissima [66]
Canthin-6-one-1- O -[2- β -D-apiofuranosyl-6- O -(3-hydroxy-3-methylglutaryl)]- β -D-glucopyranoside R ₁ = O -glc $R_{2'}$ = HO O C HO O HO O O O O O O O O O	 Simaroubaceae : Ailanthus altissima [66]

III- Polysubstituted canthin-6-ones

	Ŗ ₁₁	R₁ ≻	R_2
R ₁₀			N
R ₉		N,	R_4
	R ₈	\sum	

Polyhydroxylated/methoxylated c	anthin-6-ones c_{R_s}
1-hydroxy-2-methoxycanthin-6-one R_1 =OH, R_2 =OMe, $R_{4\rightarrow 11}$ = H	• Simaroubaceae : Brucea antidysenterica [35]
1,2-dimethoxycanthin-6-one $R_1=R_2=OMe, R_{4\rightarrow 11}=H$	• Simaroubaceae : Picrolemma huberi [67]
1-hydroxy-11-methoxycanthin-6-one R_1 =OH, R_{11} =OMe, $R_{2\rightarrow 10}$ = H	• Simaroubaceae : Brucea antidysenterica [9]
11-hydroxy-1-methoxycanthin-6-one	• Simaroubaceae : Brucea antidysenterica [9],

R_1 =OMe, R_{11} =OH, $R_{2\to 10}$ = H	Soulamea pancheri [9,23]	
1,11-dimethoxycanthin-6-one	• Simaroubaceae: Brucea antidyisenterica	
$R_1 = R_{11} = OMe, R_{2 \to 10} = H$	[9,23,35]	
1-hydroxy-2,11-dimethoxycanthin-6-one	• Simaroubaceae : Brucea antidysenterica	
$R_1=OH, R_2=R_{11}=OMe, R_{4\to 10}=H$	[35]	
2-hydroxy-1,11-dimethoxycanthin-6-one	• Simaroubaceae : Brucea antidysenterica	
$R_1=R_{11}=OMe$, $R_2=OH$, $R_{4\rightarrow 10}=H$	[35]	
10-hydroxy-11-methoxycanthin-6-one	• Simaroubaceae : Eurycoma longifolia	
$R_{10}=OH, R_{11}=OMe, R_{1\to 9}=H$	[29,43], Brucea antidysenterica [36]	
11-hydroxy-10-methoxycanthin-6-one	Simaroubaceae · Eurycoma Ionaifolia [28]	
R_{10} =OMe, R_{11} =OH, $R_{1\to 9}$ = H	- Sima oubaccae . Eurycomu tongtjonu [20]	
1-hydroxy-9-methoxycanthin-6-one	 Simaroubaceae: Eurycoma longifolia [27] 	
$R_1 = OH, R_9 = OMe, R_{2 \to 8,10,11} = H$		
4-hydroxy-5-methoxycanthin-6-one	• Simaroubaceae : Picrasma quassioides	
$R_4=OH, R_5=OMe, R_{1,2,8\to11}=H$	[9,47,68], Eurycoma longifolia [27,48]	
	• Simaroubaceae : Picrasma excelsa [9],	
5-hydroxy-4-methoxycanthin-6-one	Picrasma quassioides [9,23,46,47,69,70], Quassia amara [71,72] Europorta longifolia	
$K_4=OMe, K_5=OH, K_{1,2,8\to11}=H$	[29,43]	
4.5-dihydroxycanthin-6-one		
$R_4=R_5=OH, R_{1,2,8\to11}=H$	• Simaroubaceae : Simarouba glauca [30,31]	
4,5-dimethoxycanthin-6-one R ₄ =R ₅ =OMe, R _{1,2,8\rightarrow11} = H	 Simaroubaceae : Odyendea gabonensis, Quassia africana [9], Pricrolemma granatensis [26], Simarouba glauca [30,31], Picrasma quassioides [23,46,47,69,72], Eurycoma longifolia [27,43] 	
11-hydroxy-5-methoxycanthin-6-one $B_{5}=OMe_{1}B_{1}=OH_{1}B_{1}=A_{2}=H_{1}$	• Simaroubaceae : Picrasma quassioides [73]	
1-hydroxy-4 5-dimethoxycanthin-6-one		
R_1 =OH, R_4 = R_5 =OMe, $R_{2,8\rightarrow11}$ = H	• Simaroubaceae : Picrasma quassioides [47]	
10-hydroxy-4,5-dimethoxycanthin-6-one	Simaroubaceae · Picrasma auassioides [46]	
$R_4 = R_5 = OMe, R_{10} = OH, R_{1,2,8,9,11} = H$		
4,10-dihydroxy-5-methoxycanthin-6-one	• Simaroubaceae : Picrasma auassioides [74]	
$R_4 = R_{10} = OH, R_5 = OMe, R_{1,2,8,9,11} = H$		
4,9-dimethoxycanthin-6-one	• Simaroubaceae : Eurycoma longifolia	
$R_4 = R_9 = OMe, R_{1,2,5,8,10,11} = H$	[29,43]	
9-hydroxy-5-methoxycanthin-6-one	• Simaroubaceae : Odyendea gabonensis	
$R_5 = OMe, R_9 = OH, R_{1 \rightarrow 4,8,10,11} = H$	[75]	
5,9-dimethoxycanthin-6-one	• Simaroubaceae : Eurycoma longifolia	
$K_5 = K_9 = OMe, R_{1 \to 4,8,10,11} = H$	[28,29,43]	
5-hydroxy-4,9-dimethoxycanthin-6-one	• Simaroubaceae : Eurycoma longifolia [29]	
$K_4 = K_9 = OMe, K_5 = OH, K_{1,2,8,10,11} = H$		
8-hydroxy-9-methoxycanthin-6-one	Simaroubaceae : Pricrolemma granatensis Sch Ferrererere here if the 127 20 421	
$K_8=OH, R_9=OMe, K_{1\to 5,10,11}=H$	[20], Eurycoma longifolia [27,29,43]	
8,9-dihydroxycanthin-6-one $R_8=R_9=OH, R_{1\rightarrow 5,10,11}=H$	• Simaroubaceae : Picrasma quassioides [52]	

10-hydroxy-9-methoxycanthin-6-one	• Simaroubaceae : Eurycoma longifolia
R ₉ =OMe, R_{10} =OH, $R_{1\rightarrow8,11}$ = H	[27,28,43]
9,10-dimethoxycanthin-6-one R ₉ =R ₁₀ =OMe, R _{1\rightarrow8,11} = H	• Simaroubaceae : <i>Eurycoma harmandiana</i> [32], <i>Eurycoma longifolia</i> [27-29,43,48]

Polymethoxylated/glucosylated canthin-6-ones		
Canthin-6-one-9-methoxy-5- O - β -D-glucopyranoside R ₅ = O -glc, R ₉ =OMe, R _{1\rightarrow4,8,10,11} = H	 Simaroubaceae : Simarouba berteroana [50], Eurycoma longifolia [29] 	
<i>b</i> =		

Alkylated canthin-6-ones		
5-methyl-9-methoxy-canthin-6-one R ₅ =Me, R ₉ =OMe, $R_{1\rightarrow4,8,10,11}$ = H	• Zygophyllaceae : <i>Peganum nigellastrum</i> [76]	
5-ethyl-9-methoxy-canthin-6-one R ₅ =Et, R ₉ =OMe, $R_{1\rightarrow4,8,10,11}$ = H	• Zygophyllaceae : <i>Peganum nigellastrum</i> [76]	

Alkoxylated canthin-6-ones	
5-hydroxymethylcanthin-6-one	• Simaroubaceae : <i>Ailanthus altissima</i> [9,77],
$\mathbf{R}_5 = \mathbf{CH}_2\mathbf{OH}, \mathbf{R}_{1 \to 4, 8 \to 11} = \mathbf{H}$	Eurycoma longifolia [27]
(R/S)5-(1'-hydroxyethyl)canthin-6-one	 Simaroubaceae : Eurycoma longifolia [29],
$R_5 = CH(OH)CH_3, R_{1 \rightarrow 4,8 \rightarrow 11} = H$	Ailanthus altissima [78]
5-hydroxymethyl-9-methoxy-canthin-6-one	• Simononhaaaaa European laasifalia [27]
$R_5 = CH_2OH, R_9 = OMe, R_{1 \rightarrow 4,8,10,11} = H$	- Simaroubaceae : Eurycoma tongijotta [27]
8-hydroxymethyl-1-methoxy-canthin-6-one	• Euphorbiaceae : Euphorbia bivonae [79]
R_8 = CH ₂ OH, R_1 =OMe, $R_{2\rightarrow 5,9\rightarrow 11}$ = H	
5,8-dihydroxymethylcanthin-6-one	• Euphorbiaceae : Euphorbia bivonae [79]
$R_5 = R_8 = CH_2OH, R_{1 \to 4,9 \to 11} = H$	
(R/S)5-(1'-hydroxyethyl)-9-methoxycanthin-6-one	• Simonoubaceae • Europeana longifelia [70]
$R_5 = CH(OH)CH_3, R_9 = OMe, R_{1 \rightarrow 4,8,10,11} = H$	- Sinai oubaccae . Eurycomu longijoliu [79]

Polyhydroxylated/methylsulfoxide	canthin-6-ones $R_1 \rightarrow R_1 \rightarrow R_2 \rightarrow R_2$
nethylsulfoxidecanthin-6-one	• Boletineae : Boletus curtisii [41]

1-hydroxy-4-methylsulfoxidecanthin-6-one R_1 =OH, $R_{2\rightarrow 11}$ = H	• Boletineae : Boletus curtisii [41]
1,9-dihydroxy-4-methylsulfoxidecanthin-6-one	• Boletineae : Boletus curtisii [41]
$R_1 = R_9 = OH, R_{2,5,8,10,11} = H$	• Rutaceae : Pentaceras australis [9]

$R_{10} \xrightarrow{R_{11}} \xrightarrow{R_{1}} \xrightarrow{R_{2}} \xrightarrow{K_{10}} \xrightarrow{R_{10}} \xrightarrow{R_{10}}$	
Canthin-6-one-3- <i>N</i> -oxide $R_{1\rightarrow 11}=H$	 Simaroubaceae : Ailanthus altissima [9], Brucea mollis [24], Eurycoma harmandiana [32], Simarouba berteroana [50], Eurycoma longifolia [27,28], Simaba bahiensis [44-45], Rutaceae : Zanthoxylum chiloperone [80], Zanthoxylum paracanthum [22]
1-methoxycanthin-6-one-3- <i>N</i> -oxide R_1 =OMe, $R_{2\rightarrow 11}$ = H	• Simaroubaceae : Ailanthus altissima [81]
2-methoxycanthin-6-one-3- <i>N</i> -oxide R ₂ =OMe, $R_{1,4\rightarrow11}$ = H	• Simaroubaceae : Simaba bahiensis [55]
9-hydroxycanthin-6-one-3- <i>N</i> -oxide R ₉ =OH, $R_{1\rightarrow8,10,11}$ = H	 Simaroubaceae : Eurycoma longifolia [28,49], Simarouba berteroana [50], Eurycoma longifolia [27]
9-methoxycanthin-6-one-3- <i>N</i> -oxide	Simaroubaceae : Eurycoma longifolia
$R_{11}=OH, R_{1\rightarrow 0}=H$ $R_{11}=OH, R_{1\rightarrow 10}=H$	 [43,49], Pricrolemma granatensis [26] Simaroubaceae : Brucea mollis [24], Simarouba berteroana [50]

IV- Canthinediones	
2,6-diones	R_{10} R
Canthin-2,6-dione	• Simaroubaceae : Eurycoma harmandiana
$R_{1 \to 11} = H$	[9], Simaba multiflora [82]
5-methoxy-canthin-2,6-dione	Simproubaceae · Samadara indica [0]
$R_5 = OMe, R_{1 \to 4,8 \to 11} = H$	- Sinal oubaceae . Sumuler a matca [9]
3-methylcanthin-2,6-dione	• Simaroubaceae: Picrasma quassioides [83]
$R_3=Me, R_{1,4\to11}=H$	
3 methovycanthin 2.6 dione	 Simaroubaceae : Simaba multiflora [9,82], Simaba cuspidata [61], Simaba bahiensis [44]
$\mathbf{R}_{3}=\mathbf{OMe}, \mathbf{R}_{1,4\rightarrow11}=\mathbf{H}$	
10-hydroxy-3-methoxycanthin-2,6-dione	• Simaroubaceae : Simaba multiflora [9,82]
$R_3 = OMe, R_{10} = OH, R_{1,4 \rightarrow 9,11} = H$	

5,6-diones	R_{10} R_{9} R_{8} R_{9} R_{8} R_{9} R_{11} R_{2} R_{2} R_{3} R_{4}
3-methylcanthin-5,6-dione	• Simaroubaceae : Picrasma quassioides
$R_3=Me, R_{1,2,4\to11}=H$	[9,46,47,72,84,85], Eurycoma longifolia [28]
4-hydroxy-3-methylcanthin-5,6-dione	• Simaroubaceae : Picrasma quassioides
$R_3=Me, R_4=OH, R_{1,2,8\to11}=H$	[9,84]
4-methoxy-3-methylcanthin-5,6-dione	• Simaroubaceae : Picrasma quassioides
$R_3=Me, R_4=OMe, R_{1,2,8\to11}=H$	[9,68,84]
9-methoxy-3-methylcanthin-5,6-dione	Simarouhaceae: Eurocoma longifolia [28]
$R_3=Me, R_9=OH, R_{1,2,4\to8,10,11}=H$	Siniai Subaccae. Eurycomu iongifoliu [26]

V- Other derivatives	
Furancanthin O OH	• Simaroubaceae : Picrasma quassioides [73]
$\begin{array}{c} & & & R_{4} \\ & & & R_{3} \\ & & & & R_{1} \\ & & & & R_{2} \\ \end{array}$ Amarastelline A O Picrasidine N R_{1}=R_{2}=R_{4}=H, R_{3}=OCH_{3} \\ & & & R_{1}=R_{4}=H, R_{2}=R_{3}=OCH_{3} \\ Picrasidine M Picrasidine U R_{1}=R_{3}=H, R_{2}=R_{4}=OCH_{3} \\ & & R_{1}=R_{2}=R_{4}=OCH_{3}, R_{3}=H \\ \end{array}	• Simaroubaceae : Picrasma quasinoides [68,85,86], Quassia amara [87]
Quassidine E OCH_3 HN HN O N	• Simaroubaceae : Picrasma quasinoides [47]
Hyrtimomine D R = H Hyrtimomine E R = COOH HO HO HO HO HO HO HO HO HO	 Sponges: Hyrtios sp. [88]



72

67 Large amounts of these alkaloids occur in the root bark and leaves of Rutaceae and Simaroubaceae and lesser amounts are found in their heartwood. Other genus belonging to Malvaceae, 68 69 Amaranthaceae, Zygophyllaceae, Caryophylaceae and Leitneriaceae contain canthin-6-one alkaloids. More recently, several S-containing derivatives have been identified from Boletineae [41]. The natural 70 occurrence of canthin-6-one alkaloids is illustrated in Figure 3. 71



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Figure 3. Distribution of canthin-6-one alkaloids cited in table 1 in the nature. a) Recurrence and b) diversity of canthin-6-one alkaloids for each family.

- Figure 3a represents the canthin-6-one derivatives molecules recurrence for each family whereas 77 78 Figure 3b shows the average number of canthin-6-one alkaloids per species belonging to the different families. It is observed that the main families, Simaroubaceae and Rutaceae, have an important number 79 80 of molecules derivatives from canthin-6-one among their species, 174 and 30 respectively. However, same families show a poor diversity of molecules with an average number of 2.4 and 0.4 per species 81 respectively. A reversed trend is observed for Boletineae and Sponges families, having an only species 82 containing different canthin-6-one alkaloids, 7 and 3 respectively. 83
- 84

85 3. Biosynthesis of canthin-6-ones

86

87 Despite the utility of this family of compounds, in the eighties, canthin-6-one (1) and its 88 derivatives had no real commercial interest. However they had been studied as a model system for the production of alkaloids by plant cell cultures, due to the success of high yields of canthinones from 89 90 cell suspensions of Ailanthus altissima Swingle (Simaroubaceae) [90].

91 The reported biosynthetic hypotheses for formation of canthin-6-one (1) building blocks, originated 92 from the proteinogenic aminoacid tryptophan (2) was first predicted by Anderson et al. [91]. They investigated the uptake of L-[methylene 14 C] tryptophan (2) by Ailanthus altissima cells and 93 demonstrated the incorporation of the radio-label into canthin-6-one (1), 1-hydroxycanthin-6-one (6) 94 95 and 1-methoxycanthin-6-one (7) (Schema 1). Similar studies with tryptamine (3) have also shown incorporation of the radio-label into canthinone alkaloids [92]. The postulated key steps rely on several 96 97 presumed precursors isolated from cell cultures of Ailanthus altissima by Crespi-Perellino and 98 coworkers in 1986 [3,93]. In these experiments cells were fed radio-labeled tryptophan (2) and various 99 alkaloids products were extracted and isolated according to their radiochromatographic peak. This allowed the synthesis of labeled canthin-6-one by the addition of each labeled intermediate to the 100 101 cellular culture. As result of these experiments they proposed the synthesis of canthin-6-one scaffold 102 by condensation of tryptophan and suggested an origin from tricarboxylic acid cycle (TCA) for the carbon atoms C-3, C-4, C-5 and C-6, resulting in a series of β-carboline intermediates in various states 103 of oxidation (Schema 1). 183

Several publications support the Crespi-Perellino *et al.* [3] hypothesis. For example the biogenic
 studies performed by the Phillipson group, where they had shown the origin from methionine of the
 methyl group of 1-methoxycanthin-6-one (7) [92] (Schema 1).

- Following the first experimental biosynthetic study of canthinones performed by Crespi-Perellino *et al.*, [3]. Aragozinni and coworkers proposed ketoglutarate as an early precursor for the carbon atoms C-3, C-4, C-5 and C-6 of canthin-6-one (Schema 1) by feeding (¹³C) sodium acetate to cell cultures of
- 112 *Ailanthus altissima* and evaluating the labeled sites by ¹³C NMR spectrometry [94] (Schema 1).

Schema 1. Biosynthetic building blocks, including the tryptophan (2), the ketoglutarate (4) and the methylmethionine (5) with the respective labels employed by Crespi-Perellino (¹⁴C), Aragozzini (¹³C) and Anderson (¹⁴C).

116



119 A study of Movassaghi's alternative conditions for the Bischler–Napieralski reaction allowed to 120 formulate a general strategy for the synthesis of all intermediates in the canthin-6-one biosynthetic 121 pathway and their chemical interconversions [95]. The results with a nanoparticular mimic system of 122 the relevant biosynthetic assembly lines have been disclosed and merit further developments. Finally, 123 biological evaluations using *C. elegans* revealed new findings that may contribute to a better 124 understanding of the role of canthin-6-one alkaloids in chemical ecology.

A filamentous fungus, *Cunninghamella blakesleeana* CGMCC 3.970, was used as a microbial system to mimic mammalian metabolism of 4,5-dimethoxy-canthin-6-one, a canthin-6-one type alkaloids, which is a major bioactive constituent of a traditional Chinese medicine (the stems of *Picrasma quassioides*). The metabolic pathways of 4,5-dimethoxyl-canthin-6-one were proposed, and the metabolic processes involved phase I (*O*-demethylation, dehydroxylation, demethoxylation, *N*oxidation, hydroxylation, and oxidative ring cleavage) and phase II (glycosylation) reactions [96].

131

132 **4. Synthesis of canthin-6-ones**

133

The various strategies utilized for synthesis of canthinone alkaloids employ two well know reactions: the Pictet-Spengler and Bischler-Napieralski cyclizations. In the case of canthinones, the starting reagent utilized is tryptamine (3). There are also a few original synthetic schemes in that are not derived from tryptamine (3) or indole-precursors.

138 For clarity, we have divided the different approaches into several groups depending on the 139 number of carbons presented by each reactive. The strategies used in most of the work are based on the 140 cyclization of Bischler-Napieralski as well as the Pictet-Spengler to obtain a compound derived from 141 combinations of intermediates $C_8N_2 + C_6$, $C_{12}N_2 + C_2$, $C_{10}N_2 + C_2 + C_2$ or $C_{10}N_2 + C_4$. Sometimes these intermediates may be involved in a cyclization inter-or intramolecular to form the canthinone 142 143 skeleton. Most of these syntheses are facing final steps of oxidation, which in general are the cause of 144 low to very low yields [97-101]. In schema 2, we show the four different approaches that will be 145 developed for the synthesis of the basic structure of canthinones [80,97-123].

146

147 **Schema 2.** Different synthetic strategies to reach the canthin-6-one scaffold.



4.1. Synthesis based on the strategy $C_{10}N_2 + C_2 + C_2$ $131 \\ 32$

153

There are many examples of $C_{10}N_2 + C_2 + C_2$ synthesis strategy starting from the methyl ester 154 of tryptophan (2) including the work of Settimi and colleagues [106]. They reported that the Perkin 155 reaction of β -carboline 3-carbomethoxy derivatives (8) with acetic anhydride under basic conditions 156 gave methyl canthin-6-one-2-carboxylate (9) (Schema 3, i). A similar approach was followed by the 157 group of Murakami [121,122], employing stronger basic conditions (t-BuOK in DMSO). The work 158 developed by Condie and coworkers [103-104] demonstrate another strategy to reach the synthesis of 159 the canthin-6-one core. In this case, they propose the conversion of the 1-fomyl- β -carboline (8) to 160 tetracyclic compounds possessing the canthin-parent structure by treatment of the aldehyde (8) with 161 phosphorane (Schema 3, ii). Another synthesis with similar features was published by the Singh group 162 [119], this time additional carbons were introduced from acrylates (10) via Baylis-Hilman reaction 163 followed by intramolecular cyclization resulting in the canthinone scaffold as shown in molecule (11) 164 (Schema 3, iii et iv). 165

Schema 3. Synthesis based on the strategy $C_{10}N_2 + C_2 + C_2$. 169



168 169 Reagents and conditions : (i) (CH₃CO)₂O (excess), reflux, 3 h, (72 %) ; (ii) Ph₃PCHCO₂Et, toluene, reflux 170 (29%); (iii) DABCO, t.a.; (iv) K₂CO₃, DMF, t.a.

171

172 With similar reagents and conditions, Fang et al. proposed a way to synthesis 4-methoxycanthin-6-173 one, named cordatanine (14), in four steps via a Pictet-Spengler reaction using tryptamine (3) and 174 ethyl glyoxylate (12) with a total yield of 8% (Schema 4) [124].





- 178 *Reagents and conditions:* (i) HCl, EtOH, Pd/C, toluene, reflux; (ii) NaHDMS, EtOAc, 13, THF, -78 °C; (iii)
 179 Cs₂CO₃, MeOMs; (iv) NaH, THF (8%)[124]
- 180

181 Ma *et al* [76] proposed a way to synthesis alkylated canthin-6-one in two steps from harmine (**15**) via 182 1-formyl-7-methoxy- β -carboline (**16**) by slightly modified procedures of Settimj and colleagues [106]. 183 The reaction between compound (**16**) and propionic anhydride or butyric anhydride afforded luotonins 184 C (**17**) and D (**18**) with a yield of 15% and 13% respectively (schema 5).

185

186 Schema 5. Synthetic steps to luotonins C (17) and D (18)



187

188 **Reagents and conditions**: (i) EtOH, SeO₂, reflux, 3h; (ii) Et₃N, $(C_2H_5CO)_2O$, reflux, 2 h, (15%) or (iii) Et₃N, 189 $(C_3H_7CO)_2O$, reflux, 2 h, (13%)[76]

190

191 Starting from the same type of intermediate, Tremmel group developed a versatile approach to the 192 canthin-4-one ring system via a condensation of 1-acyl- β -carbolines (19) with Bredereck's reagent 193 (tert-butoxy-bis(dimethylamino)methane) or amide acetals to give enaminoketones (20), which 194 undergo cyclization under the reaction conditions resulting the compounds (23) and (24) with a yield 195 of 80% and 52% respectively [125] (Schema 6, a route). They proposed also a new approach based on 196 a Claisen type condensation of 1-acyl- β -carbolines (19) with a carbonyl derivative (21) in basic 197 medium giving an intermediate (22) which cyclizes to the canthin-4-one ring system in a similar 198 manner. Via this pathway, compounds (24) and (25) are obtained with a yield of 80% and 52% respectively [126] (Schema 6, b route). 199

- 200
- 201 Schema 6. Synthetic steps to canthin-4-one derivatives (23), (24) and (25).



Reagents and conditions: a : with Bredereck's reagent (R= H) (i) Bredereck's reagent wet DMF, reflux; (ii)
tert.-BuOK (cat.), anhydrous DMF, reflux (80%). With amide acetal (R=CH₃) (i) N,N-dimethylacetamide
dimethylacetal anhydrous DMF, 130°C (52%)[125] b : i) MgBr₂·OEt₂, i-Pr₂NEt, methylene chloride,
microwave 100 W, 70 °C ii) LDA, THF, 0 °C to rt,(34 and 16%) [126].

202

209 In 1999, an innovative way of canthin-6-one synthesis was proposed by Röbler et al. using a single 210 electron transfer (SET) induced radical cationic hetero [4+2] cycloaddition as high yielding key step [127]. They used harmalan (26) as starting reagent. The SET induced cycloaddition between 211 212 intermediate (27) and methyl E-3-(*N*,*N*-dimethylamino)-acrylate (28) was carried out 213 electrochemically at a potential of 400 mV giving an high yield of 87% of compound (29). Few more 214 steps are needed to obtain canthin-6-one (1) with an overall synthetic yield of 18% at that time 215 (schema 7).

216

217 Schema 7. Total synthesis of canthin-6-one using a SET induced hetero [4+2] cycloaddition



219**Reagents and conditions**: With $R = SO_2CF_3$ i) $(CF_3SO_2)_2O$, Et_3N , CH_2Cl_2 at 0°C (94%); ii) ratio (27) 5 : 1220(28), E = 400 mV in CH_3CN/CH_2Cl_2 (1/4:0.1 M LiClO₄), rt (87%); iii) Na/naphthalene, DME at 0°C, MnO₂,221 CH_2Cl_2 , rt (74%); iv) HCl, Cu/pyridine (50%).

Instead of using a carbonyl intermediates with tryptamine (**3**), Dighe *et al.* have developed an alternative route to the synthesis of canthin-6-one aryl derivatives (**31**) via iodine-mediated oxidative Pictet-Spengler reaction choosing terminal alkynes as the 2-oxoaldehyde surrogate [128]. Lower loading of iodine and better yields of products as compared to the reported methodologies where acetophenones have been used as the 2-oxoaldehyde source are attractive attributes of this protocol. The protocol is amenable to aryl terminal alkynes (**30**) whereas alkyl-based terminal alkynes failed to react. This way of synthesis lead to a high yield of 83% (schema 8).

230

218

222

231 Schema 8. Synthetic steps to 4-arylcanthin-6-ones



Reagents and conditions: (i) I₂ (0.5 equiv), DMSO, 100°C, 4-5h (ii) Ac₂O (1.5 equiv), NaH (2.0 equiv), THF,
 70°C, 12h (83%)

 $\frac{235}{236}$

232

237 4.2. Synthesis based on the strategy C₁₀N₂ + C₄

238

The synthetic approach based on intermediates type $C_{10}N_2 + C_4$ to yield the canthinone scaffold, employ as starting material compounds that have the same number of carbons as the canthin-6-one (1). The group of Rosenkranz [118] reported the synthesis of canthin-6-one (1) in two steps from tryptophan (2) through the formation of the intermediate (31) (Schema 9, a route). In 2006, Soriano-

243 Agatón *et al.* designed a synthetic pathway utilizing a similar strategy, based on the addition of the 244 four carbon atoms from succinic anhydride, through a four steps reaction sequence to tryptamine (3) 245 [80]. In the presence of succinic anhydride, tryptamine (3) is easily converted into tryptamide (32), 246 protecting the carboxylic acid form of methyl ester (33), and employing this molecule as substrate of 247 the Bischler-Napieralski cyclization resulting in a β -carboline that will undergo an intramolecular 248 cyclization. A final spontaneous oxidation of the resulting tetracyclic system led directly to the 249 canthin-6-one (1) with an overall yield of about 60% (Schema 9, a route). Shelar et al. described a 250 similar pathway of 4-methoxycanthin-6-one (14) synthesis starting from tryptamine (3) and 251 methoxymaleic anhydride [129]. They proposed an efficient total synthesis of cordatanine (14) via 252 regioselective reduction of methoxymaleimide, acid-catalyzed intramolecular cyclization of the formed 253 lactamol, in situ stepwise oxidations leading to aromatization, and intramolecular cyclization with the 254 exchange of N-regioselectivity. Cook and coworkers were able to synthesize similar compounds by 255 two different synthetic routes [99,102,108-110,114]. They also synthesized canthin-6-one (1) in four 256 steps from tryptamine (3). A final oxidation step showed very low yields (Schema 9, b route). Few 257 years later, Czerwinski and collaborators reported a methodology that would improve the yield of this 258 oxidation step [105] (Schema 9, c route) synthesizing 1-methoxycanthin-6-one (38). 259



260 Schema 9. Synthesis based on the strategy $C_{10}N_2 + C_4$.

Reactives and conditions: *a*: (*i*) Succinic acid (1.1 equiv.), sol. 2N K_2CO_3 , 30 min, t.a., (60 %); (*ii*) polyphosphoric acid, V_2O_5 , $POCl_3$ (excess), 1 h, 115 °C, (25 %); (*iii*) succinic anhydrid (1.1 equiv.), CH_2Cl_2 , 18 h, r.t., (>97 %); (*iv*) Amberlyst[®] H-15 (20 % m/m), MeOH reflux, 18 h, (>97 %); (*v*) $POCl_3$ (3 equiv.), benzene, 1 h, reflux; (*vi*) DBU (3 equiv.), CH_2Cl_2 , 18 h, r.t., (60 % + 80 %). *b*: (*i*) Na /NH₃, MeI, 30 min, -70 °C, (80 %); (*ii*) PhCHO, MeOH, 6 h, r.t.; (*iii*) NaBH₄, MeOH, 0-5 °C, 3 h, (two steps, 95 %); (*iv*) 2-

cetoglutaric acid (1.1 equiv.), benzene, 7 days, reflux, (82 %); (v) SeO₂ (1.5 equiv.), dioxane, reflux, 24 h, (30 %). **c** : (i) Dimethyl 2-ketoglutarate (1.5 equiv.), MeOH, 20 h, reflux, (92 %) ; (ii) DDQ (excess), THF /H₂O, r.t., (75 %) ; (iii) HCl, AcOH, reflux, 3 h, (88 %) ; (iv) MeOH, CH(OCH₃)₃, PTSA, reflux 24 h, (82 %) ; (v) DDQ (excess), dioxane, 10 h, reflux, (70 %).

Between 2006 and 2009 the laboratories of Corey [117] and Nicolaou [115,116] described the synthesis of (+)-haplophytine (**39**), based on a similar approach. (+)-haplophytine (**39**) is a potent insecticidal alkaloid which derive from two indoles, aspidophytine (**41**) coupling with a canthiphytine (**40**) derivative which is very similar in structure to canthin-6-one (Schema 10).

Schema 10. Retrosynthesis of (+)-haplophytine (39).



Synthetic approach type $C_{10}N_2 + C_4$ may also be employed for building benzocanthin-6-one (42) and derivatives. This type of approach included Bischler–Napieralski cyclization of tryptamine (3) and phthalic anhydride used by Soriano-Agatón *et al.* [80] or Pictet–Spengler cyclization of tryptophan (2) and *o*-formylbenzoic acid (43) [130] both followed by intramolecular *N*-acylation of the pyrrole N and dehydration (schema 11, a route). The same year, Zimmerman's group published a versatile route to synthesize benzocanthinone (42) by Suzuki coupling as well [113]. This compound were prepared by radical-induced cyclizations of halo N-aroyl derivatives of β -carboline (45). 1-chloro- β -carboline (44) was converted to compound (45), which afforded the benzocanthinone (42) as the only observed product (Scheme 11, b route).

Schema 11. Synthetic steps to benzocanthin-6-one (42).



Reagents and conditions: **a** : (i) AcOH,100°C, reflux, 30-40h; (ii) O₂(35-56%)¹³⁰. **b** : (i) NaH, DMF, r.t., benzoyl chloride, DMAP, DMF, 120°C; (ii) Bu₃SnH, ACN, toluene, reflux (74%)[113]

4.3. Synthesis based on the strategy $C_8N_2 + C_6$

Koutentis group presented a total synthesis of canthin-6-one (1) using a strategy totally independent of the biosynthesis of the β -carboline alkaloids [107,111]. This newest pathway did not exploit starting materials derived from tryptophan (2) and instead used novel original strategy type $C_8N_2 + C_6$. This work reported a revolutionary synthesis of canthin-6-one (1) in two steps from 8-bromo-2-methoxynaphthyridine (46) and 2-chlorophenylboronic acid (47) by one-pot reaction sequence using Pd-catalyzed Suzuki-Miyaura coupling followed by Cu-catalyzed C-N coupling [122] (Schema 12).

Schema 12. Synthesis based on the strategy $C_8N_2 + C_6$.



Reagents and conditions: (i) ArB(OH)₂, Pd(dppf)Cl₂, K₂CO₃, dioxane:H₂O, reflux ; (ii) HCl_(aq), dioxane, reflux; (iii) CuI, DMEDA, Cs₂CO₃, H₂O, dioxane, reflux, (99%)[107]

More recently, Broumidis and Koutentis still used this strategy by developing a one-pot, two-step

synthesis of 3-deazacanthin-4-one (**48**) involving readily available 2-chloro-arylboronic acids (**49**) and 8-iodoquinolin-4(1H)-one (**50**) (Schema 13). The procedure takes advantage of transition metalcatalyzed Suzuki-Miyaura and Buchwald-Hartwig coupling strategies to construct the 3-deazacanthin-4-one central B ring [131]. The route successfully provides a rapid route towards 8-, 9-, and 10substituted 3-deazacanthin-4-ones.

Schema 13. Retrosynthetic analysis of 3-deazacanthin-6-one



261 4.4. Synthesis of unnatural canthine alkaloids based on the strategy C₁₂N₂ + C₂

263 The synthetic approach based on intermediates type $C_{12}N_2 + C_2$ to obtain the basic canthine 264 scaffold (53) was put for the by the Snyder group [120,132] disclosing an elegant entry to canthine 265 skeleton utilizing indole as dienophile in an intramolecular inverse electron demand Diels-Alder 266 reaction (Schema 14). An additional oxidation step is necessary to obtain the canthin-6-one alkaloid 267 [133]. Treatment of acyl-hydrazine-tethered indole (51) with a 1,2-diketone and excess of ammonium acetate in refluxing acetic acid for several hours provided the triazine-tethered indole (52) which was 268 269 refluxed in triisopropylbenzene to provide the basic canthine skeleton (53). Recently, Lindsley et al. 270 reported the synthesis of the basic canthine core by the Snyder synthesis on one pot microwave-271 assisted protocol [112,123].

272

262

273 Schema 14. Synthesis of unnatural canthine alkaloids based on the strategy $C_{12}N_2 + C_2$.



275 *Reagents and conditions : a : (i)* NH4OAc (excess), AcOH, reflux 6-4 h, (50-60 %); (ii) triisopropylbenzene,
276 reflux, 232 °C, (72-92 %) ; (iii) NH4OAc (excess), AcOH, 180 °C, 5 min, MWI, (6 %).

- 277 278
- 279 5. Pharmacological and toxicological study
- 280
- 281 **5.1. Antileishmanial activity**
- 282

The antileihsmanial activity demonstrated by *Zanthoxylum chiloperone* is attributed to the presence of canthin-6-one and 5-methoxycanthin-6-one [14]. The effect of canthin-6-one *in vivo* is stronger than the molecule of reference, the meglumine antimoniate, used as a positive control. The 5methoxycanthin-6-one showed activity, but less than canthin-6-one [14,134].

Silva's group exposed antileishmanial activity of synthetic benzocanthin-6-one derivatives [135]. Compound bearing an N-methylpiperazylcarboxamide at C-2, showed the highest activity towards *Leishmania amazonensis* parasite with the half maximal inhibitory concentration (IC₅₀) of $0.90 \pm 0.13 \mu$ M.

291

292 **5.2.** Antimalarial activity

293

294 The antimalarial activities of canthin-6-one and some derivatives (9-hydroxycanthin-6-one, 5-295 methoxycanthin-6-one) were studied [27,49,136,137], with mixed results. Kuo et al. showed that 296 canthin-6-one and 9-hydroxycanthin-6-one exhibited low antimalarial activity against the W2 297 *Plasmodium falciparum* clone with IC₅₀ values 2238 and 2336 ng/mL, respectively [27]. We report a 298 good antiplasmodial effect on chloroquine resistant and sensitive strains of *Plasmodium falciparum* for 299 the canthinone alkaloids with IC₅₀ of 2.0-5.3 µg/mL and 5.1-10.4 µg/mL for canthin-6-one and 5-300 methoxycanthin-6-one, respectively [136]. In vivo data (unpublished) in our hands did not match the in 301 vitro activity. This discrepancy is likely due to in vivo metabolism of the canthinones.

302

5.3. Antifungal

304

The activity against *Candida albicans* is the first activity attributed to the canthin-6-one, this activity was described in *Zanthoxylum elephanthiasis* [9]. Recently, bark and root extracts of *Zanthoxylum usambarense* and *Zanthoxylum chiloperone* antifungal activities against different mycelia and yeasts were evaluated with promising results [15,138].

The antifungal activity of canthin-6-one and some analogues against *Candida albicans*, *Aspergillus fumigatus, Trichophyton mentagrophytes, Cryptococcus neoformans and Saccharomyces cerevisiae*, among others, is not so far away to the reference drug, ketoconazole [80]. Later, a mechanism of action of antifungal canthin-6-one *N*-oxide was investigated in *Saccharomyces cerevisiae* and found to affect fatty acid metabolism [139].

Dejos *et al.* screened a yeast genomic library in *Saccharomyces cerevisiae* to isolate high-copynumber suppressors of canthin-6-one sensitivity [140]. Among the genes selected with this strategy, they isolated the yeast AP1 homologue (*YAP1*), a gene involved in cell response to various stress, more particularly oxidative stress and pleiotropic drug resistance in high-copy number. In this study they showed that plasmid-borne overexpression of *YAP1* leads to canthin-6-one resistance via the major-facilitator-superfamily (MFS) transporter Flr1 involved in the efflux of a broad range of cytotoxic molecules.

Recently, Gazoni *et al.* presented the phytochemical analysis by high performance liquid chromatography (HPLC), toxicological potential of canthin-6-one, using the cytotoxicity, genotoxicity assays with CHOK1 cells and in vivo acute test in mice [141]. They evaluated potent in vitro antifungal activity of canthin-6-one against clinically important *Aspergillus niger* and *Candida* species showing a minimal fungicidal concentration between 3.12 to 25 μ g/mL. They also studied the antifungal mode of action on *Saccharomyces cerevisiae* and *Neurospora crassa* highlighting that canthin-6-one inhibited growth and altered cell membrane respectively.

In Zhao *et al.* researches, a novel series of ester derivatives of 10-hydroxycanthin-6-one were synthesized and evaluated for their bioactivity against three phytopathogenic fungi (*Alternaria solani*, *Fusarium graminearum*, and *Fusarium solani*) and four bacterial strains (*Bacillus cereus*, *Bacillus subtilis*, *Ralstonia solanacearum*, and *Pseudomonas syringae*) [142]. The aliphatic ester derivatives of 10-hydroxycanthin-6-one had much higher antifungal activity than did the aromatic ester derivatives.

333

334 **5.4. Antitumor activity**

335

Canthin-6-one and derivatives (hydroxylated and methoxylated) were tested for cytotoxic activity against different cell lines exhibiting weaker antitumor activity than the reference compound, cisplatin [31,82,143-145].

Peduto *et al.* described some β -carboline derivatives which are potent agent against prostate cancer [146]. In this work, the authors highlight the relationship between structure and cytotoxic activities. The in vitro experiments showed that the shift of methoxy group in position 4 of harmine led to enhanced cytotoxic activity and substituents at position 1 were essential for high activity towards specific tumor types. 3N substitution of 1-methoxycanthin-6-one with benzyl group increased activity against most of the cell lines tested.

Leite silva's group confirmed the previous results by synthesizing benzo[4,5] canthin-6-ones derivatives substituted on position 3 with N-alkylcarboxamide and analyzing the antitumor activities [135]. Several compounds exhibited moderated to significant activity especially the Nmethylpiperazylcarbohydrazide at C-2 derivative, which displayed IC₅₀ values in the range of 1.15– 8.46 μ M for all cell lines tested. The highest cytotoxicity was observed towards ovarian cancer cell lines (OVCAR-3) with IC₅₀ of 1.15 μ M. It seems that the interaction with DNA, enzymatic or receptor systems are the most commonly mode of action observed for antitumor β -carboline compounds.

352 Interestingly, some canthin-5,6-dione derivatives showed antitumor activity against P-388 353 murine leukemia cells and P-6 human lung carcinoma cells [147]. The antiproliferative effect of a commercially available canthin-6-one was examined dose-dependently on six cancer cell lines (human 354 prostate, PC-3; human colon, HT-29; human lymphocyte, Jurkat; human cervix, HeLa; rat glioma, C6; 355 and mouse embryonic fibroblasts, NIH-3T3)¹⁴⁸. Strong antiproliferative effects of the compound were 356 observed in all cell lines, whereas cytotoxic effects were very dependent on cell type. The data 357 358 suggested that the primary effect of canthin-6-one is antiproliferative, possibly by interfering with the 359 G2/M transition.

The activity of *Z. chiloperone* var. *angustifolium* alkaloids (canthin-6-one, 5-methoxycanthin-6-one and canthin-6-one N-oxide) and some close analogues ((4,5-dihydrocanthin-6-one, 10methoxycanthin-6-one and 10-benzyloxycanthin-6-one) were evaluated against malignant glioblastoma cells [149]. 10-benzyloxycanthin-6-one revealed an effect of 40% of maximal viability loss in TG1 cells and 85% of maximal viability loss in TG10 cells when at the same time no viability changes were observed on NSC24 making this compound the most selective of the series. The de Feo group utilizing *in vitro* bioassay studies, investigated the antiproliferative and cytotoxic effects of the 1-methoxycanthin-6-one isolated from *Ailanthus altissima* (Tree-of-Heaven) [150,151]. They also showed that 1-methoxy-canthin-6-one induced apoptosis *via* a *c*-jun NH₂terminal kinase (JNK) [152].

The AKT is a serine/threonine kinase that recently garnered attention as a promising target for cancer therapy due to its critical role as a regulator of the cell's apoptotic machinery. A series of synthetic canthine alkaloids were shown to be potent and selective AKT kinase inhibitors that induced apoptosis in LnCaP cells [153].

374 Recently, quassidine E, a unique derivative of canthin-6-one, showed potent inhibitory activity 375 on the production of nitric oxide (NO), tumor necrosis factor α (TNF α), and interleukin 6 (IL-6) in 376 mouse monocyte-macrophage RAW264.7 cells stimulated by lipopolysaccharide (LPS) [64].

A bioassay-guided purification on the root extract yielded two alkaloids canthin-6-one and bruceolline J which were further investigated for their bioactivity in representative human cancer lines and normal phenotypic counterparts [154]. Canthin-6-one was able to synergize the cytotoxic effect of cisplatin when used in combination, suggesting the potential of combination therapy for those less responsive lesions to standard chemotherapy.

382 Recently, a biochemical study showed that canthin-6-one inhibited basal soluble guanylate 383 cyclase (sGC) activity in a dose dependent manner at micromolar range [155]. The electrochemical 384 analysis of sGC-canthin-6-one interactions was achieved by the solution-phase and solid-state 385 voltammetry. The electrochemical data suggest that deactivation of sGC by canthin-6-one operates via a coordinating pathway rather than a direct oxidation of the metal center, the canthin-6-one 386 387 coordination possibly does not affect the Fe-His bond, but involves binding to the sGC active site in 388 different orientations. Such mechanisms appear to be strongly influenced by structural/conformational constraints allowing for the occurrence of that oxidative pathway under determined circumstances. 389

Vieira Torquato *et al.* showed that canthin-6-one could affect Kasumi-1 cells by promoting cell death, cell cycle arrest and cell differentiation depending on concentration used [156]. Canthin-6-one also promoted differentiation of Kasumi-1, evidenced by an increase in the expression of myeloid markers (CD11 b and CD15) and the transcription factor PU.1.

The voltammetry of immobilized particles (VIMP) was used to study a possible correlation between the solid-state electrochemistry of canthin-6-one compounds and their anti-proliferative activity against cancer stem cells [157,158]. There was no single, direct correlation between anti-proliferation activity, which is a complex biochemical process, and solid state electrochemistry, where interfacial electron transfer processes are involved, current data suggest that the ability to experience the oneelectron reduction of the naphthyridone motif present in the canthin-6-one structure could be a crucial aspect potentially related to biochemical activity.

401

402 **5.5. Antibacterial activity**

403

404 Over twenty canthin-6-one alkaloids have been bioassayed for their antimicrobial activity.
 405 Activities of various canthinones against a number of bacterial strains were minor compared to that of
 406 streptomycin. Activity was observed to increase with a hydroxyl group on the canthin-6-one nucleus

407 and decrease when the hydroxyl group was substituted for a methoxyl group. Some canthinones, 408 however, were found to inhibit the growth of *Staphyloccocus aureus* and its drug-resistant strains [40].

- 409 The bacterial type II topoisomerase DNA gyrase (GyrA) and topoisomerase IV are validated targets
- 410 for clinically useful antimicrobial drugs. The canthin-6-one was proposed as highest-scoring GyrA
- 411 targeted compounds identified by molecular docking [159].
- Recently, the *in vitro* screening for antimicrobial activity revealed that two 5-substituted canthin-4ones (3-pyridyl, 2-bromophenyl) exhibit significant activity against *Streptococcus entericus*, coupled with high selectivity and the lack of cytotoxicity against mammalian cells. The intact canthin-4-one ring system was demonstrated to be essential for antibacterial activity [160].
- 416 Dai *et al.* have synthesized hydrophilic analogs of canthin-6-one via an easier synthetic route and 417 evaluated their *in vitro* antibacterial activities against Gram-positive (*Staphylococcus aureus* and 418 *Bacillus cereus*) and Gram-negative bacteria (*Ralstonia solanacearum* and *Pseudomonas* 419 *solanacearum*) [161]. The data clearly indicated that some compounds such as 3-*N*-benzylated 420 derivatives, were more potent than canthin-6-one and may become lead compounds for further 421 diseases-relevant studies even if the mechanism of these compounds' antibacterial activity is still 422 ongoing to explore.
- 423 In accordance with previous studies, Li et al. developed twenty-two new 3-N-benzylated 10-methoxy 424 canthin-6-ones synthesized through quaternization reaction [162]. They discussed the in vitro 425 antibacterial activity against three bacteria R. solanacearum, P. syringae and B. cereus. Moreover, the 426 structure-activity relationships (SARs) were carefully summarized in order to guide the development of antibacterial canthin-6-one agents. Two highly active compounds (CH2Ph(3,4-dichloro) and 427 428 CH₂Ph(*m*-I) 3-N substituents) displayed 8-fold superiority (MIC = $3.91 \mu g/mL$) against agricultural 429 pathogenic bacteria R. solanacearum and P. syringae compared to agrochemical streptomycin sulfate, 430 and exhibited potential activity against B. cereus while showing good "drug-like" properties, low cytotoxicity, and no inhibition on seed germination. 431
- 432 Dai and coworkers synthesized, in one hand, C₂, C₅ modified and D rejiggered ring canthin-6-ones, on the other hand, C₁₀ and truncated canthin-6-ones, in order to study their antibacterial activity and 433 434 structure-activity relationships [163,164]. The C_2 substituted and D rejiggered ring (54) compound 435 showed a broad spectrum of antibacterial activity, being active against all the tested bacteria such as S. 436 aureus, B. subtilis (MIC = 12.5 nmol/mL), MRSA, B. cereus, R. solanacearum (MIC = 25 nmol/mL) except E. coli. (MIC >200 nmol/mL) [163]. Moreover, the C ring-truncated canthin-6-one (55) 437 438 exhibited also a broad spectrum of antibacterial effect (8-fold superiority against Staphylococcus 439 aureus and 2-fold superiority against Ralstonia solanacearum) than fosfomycin sodium and propineb 440 with a minimum inhibitory concentration value of 2 µg/mL (Schema 15). For both compounds, their 441 bactericidal effects occur by damaging bacterial cell membranes, disrupting the function of DNA 442 topoisomerases or influencing the membrane formation, both of which could lead to cell death.
- 443

444 **Schema 15**. Structure of C_2 substituted and D rejiggered ring (54) and the C ring-truncated canthin-6-445 one (55) [163,164].



448 **5.6. Antiviral activity**

449

450 Different canthin-6-one analogs were tested against HIV, 1-methoxycanthin-6-one was the first 451 example of a β -carboline exhibiting anti-HIV activity, its isomer 5-methoxy-canthin-6-one presents a 452 higher activity [27,53,54].

The antiherpes activity of four kinds of canthin-6-one alkaloids was also evaluated. Among these compounds, some of them have activity on a level with that of acyclovir, the control. It is noticed however, that the therapeutic index was small [165].

In Chen *et al.* works, crude extracts from the wood of *Picrasma quassioides* and all isolated compounds have been screened for their inhibitory activities against Tobacco Mosaic Virus (TMV) [166]. Among these compounds, 4, 5-dimethoxycanthin-6-one inhibited TMV accumulation more effectively than ningnanmycin with the EC₅₀ value of 53.0 μ g/mL and mixed with the quassinoid, nigakilactone B (25 μ g/mL each), the canthin-6-one derivative reached a lower EC₅₀ value of 36.4 μ g/mL. This study highlighted the synergistic effects between isolated β -carboline alkaloids and the quassinoid nigakilactone B against TMV.

463 Compounds of the three plants *viz. E. harmandiana*, *S. flavescens* and *A. paniculata* were screened for 464 antiviral activity against the proteases of the SARS CoV-2 by Verma *et al.* [167]. The molecular 465 docking and ADMET analysis suggested that canthin-6-one 9-O- β -glucopyranoside showed a potential 466 binding to the Mpro/3CLpro and PLpro and tended to inhibit the replication and transcription of the 467 virus stopping the virus multiplication.

468

469 **5.7. Trypanocidal activity**

470

The canthin-6-one and some of its analogs isolated from *Z. chiloperone* stem bark exhibited interesting trypanocidal activity *in vivo* in the mouse model of acute or chronic infection. In addition, the total extract of alkaloids of *Z. chiloperone* stem bark and leaves led to high levels of parasitological clearance [6,168].

To improve the pharmacological effect of the antiprotozoal and antifungal canthin-6-one, biodegradable poly(octylcyanoacrylate) matrix loaded with the alkaloid nanoparticles (170 nm average size) have been proposed by Arias *et al.* [169]. The canthin-6-one loading to the polymeric matrix during the polymerization process yielded very appropriate drug-loading values and a biphasic (controlled) drug release profile.

Silva's group discussed also antiprotozoal activities of synthetic benzocanthin-6-one derivatives with 481 the N-methylpiperazylcarboxamide at C-2 compound showing the best IC₅₀ of $0.40 \pm 0.01 \mu$ M against 482 Trypanosoma cruzi parasite [135].

483

485

484 **5.8.** Cyclic AMP phosphodiesterase inhibitory activity

486 The inhibitory activities on cyclic AMP phosphodiesterase of canthinones found in Ailanthus altissima and Simaba cuspidate were examined. They showed relatively high inhibitory activities 487 488 stronger than the control compound, the papaverine [170,171].

489

490 5.9. Other activities

491

492 Several quassinoids isolated from Ailanthus altissima and Eurycoma longifolia showed 493 amoebicidal activity determined by bioassay using Entamoeba histolytica [172-174].

494 Some canthinones manifested phototoxicity on bacteria and fungi on irradiation with near-UV light 495 (320-400 nm). These compounds inhibited mitosis of chinese hamster ovary cells and induced 496 chromosomal changes. It was apparent that the two compounds could be photosensitizers, though their 497 activities were weaker than 8-methoxypsoralen, the control [175].

- 498 The rate of blood flow in the stomach intestine of rabbits was assayed in vivo by the hydrogen 499 clearance method. Canthin-6-one and some derivatives (nigakinone and 4, 5-dihydrocanthinone) 500 increased the blood flow rate in the intestine by 15, 25 and 35 %, respectively. In the stomach, 501 however, canthin-6-one increase the rate only 10 % and nigakinone decreased it [165].
- 502 The pharmacological mechanism of action of the antiulcer effects of the methanol extract of Simaba ferruginea and its canthin-6-ones were studied. The results of this study indicate that canthinones are 503 the compounds responsible of the antiulcer action of the extract [176]. 504
- 505 In a recent study, Yuan *et al.* identified canthin-6-one as a potent ubiquin-proteasome-system (UPS) 506 activity enhancer which promoted alpha-synuclein (α -syn) degradation, a drug target used in the 507 pathology of Parkinson disease [177].
- 508 Picrasma quassioides stems extract with EtOAc revealed potential neuroprotective activities in L-509 glutamate-stimulated PC12 and A\u00f325-35-stimulated SH-SY5Y cell models and improved memory and
- 510 cognitive abilities in Alzheimer disease mice induced by amyloid- β peptide in Guo *et al.* research
- 511 [178]. Among the Picrasma quassioides stems extract, nearly twenty compounds have been isolated
- including seven canthin-6-ones alkaloids. Except for canthin-6-one, all derivatives showed potential 512 513 neuroprotective activities in vitro with a cell proliferation rate of 1.06 % - 56.25 % at the dosage
- 514 ranging from 10 μ M to 40 μ M in above cell models.
- 515 Sasaki et al. have screened natural or chemical synthetic canthinone alkaloids and β -carboline
- 516 alkaloids for their cerebral protective effects at 100 mg/kg (p.o.) in ischemic animal, which provided a
- 517 disease model of dementia [179]. Picrasidine O was improving learning and memory performance,
- 518 inhibiting delayed neuronal cell death induced by cerebral ischemia, (reducing neuronal cell death
- 519 induced by the excitatory neurotransmitter, glutamic acid and the excitotoxin, kainic acid, and
- 520 decreasing the extent of brain tissue damage. Moreover, picrasidine O was shown to have no effect on 521 heart rate or mean blood pressure.

522 Screening of a natural compound library resulted in six canthinone alkaloids, namely, picrasidine L (3-523 methylcanthin-5,6-dione), 3,4-dimethyl-canthin-5,6-dione, 4-ethyl-3-methyl-canthin-5,6-dione, 524 eurycomine E (3-methyl-9-methoxycanthin-5,6-dione), 5-methoxy-canthin-6-one, and 5-acethoxycanthin-6-one, as novel protein tyrosine phosphatase 1B (PTP1B) inhibitors [180]. It is expressed 525 526 ubiquitously in the classical insulin-targeted tissues and plays critical roles in negatively regulating 527 both insulin and leptin signaling cascades [181]. Picrasidine L was found to be the competitive PTP1B inhibitor with the best inhibitory selectivity between PTP1B and other PTPs. 528

- 529 Two molecules derivatives from canthin-5,6-dione, amarastelline A and nigakinone, the latter being a tautomer of 5-hydroxy-canthin-6-one, were isolated as fluorescent natural products from the bark of 530 531 Quassia amara [182]. They showed respectively strong fluorescence that could stain cytoplasm of 532 living cells and a solvent-dependent fluorescence properties. By designing and synthesizing canthin-5.6-dione derivatives, Yokoo *et al.* found that the combination of N^3 - and 10-substituted groups can 533 regulate the fluorescence properties, and showed that this behavior could be utilized to obtain a 534 fluorescent sensor responding to a specific range of pH. These findings indicated that the canthin-5,6-535 536 dione moiety could be a valuable scaffold for a range of fluorescent sensors and probes [182].
- 537 Cho et al. investigated the anti-inflammatory effects of canthin-6-one and its derivative 5-(1'hydroxyethyl)-canthin-6-one both isolated from the stem barks of Ailanthus altissima in 538 539 lipopolysaccharide (LPS)-stimulated macrophages [183]. These two alkaloids significantly inhibited 540 the LPS-induced expression of inducible nitric oxide synthase, markedly suppressed the production of 541 prostaglandin E2 (PGE2) and expression of cyclooxygenase-2, a key enzyme in PGE2 synthesis and inhibited the phosphorylation of inhibitor kappa B and transcriptional activation of nuclear factor 542 543 kappa B (NF-kB) in LPS-stimulated macrophages. Unlike its derivative, canthin-6-one reduced 544 monocyte chemotactic protein-1 (MCP-1) and tumor necrosis factor-a (TNF-a) expression and 545 reduced Akt phosphorylation making it a potential anti-inflammatory agent in LPS-stimulated macrophages through the downregulation of both the NF-kB and the Akt pathway. 546
- Similarly, Zhang *et al.* studied canthin-6-one alkaloids from Thailand *E. longifolia* roots conducted for
 searching anti-inflammatory agents [29]. All the isolated alkaloids were evaluated for their inhibitory
 effects on lipopolysaccharide (LPS)-induced nitric oxide (NO) production in RAW 264.7 cells at a non
 toxic concentration of 50 µM and exhibited inhibitory effects on NO release significantly comparing
- 551 with the control group.
- 552 Table 2 displays the structure of some natural and synthetic canthin-6-ones derivatives which presents
- 553 biological activities.
- 554 **Table 2.** Structure of few canthin-6-one alkaloids with their biological activities.

Molecules	Biological activities
*5-methoxycanthin-6-one	antileishmanial [14, 134], antimalarial [136], antiviral [54]





555 * Natural molecules from plants extract; ** synthetic molecules.

6. Quantification and optimization of production

558

559 Classical methods such as NMR and Mass spectroscopies are used to identify canthinones alkaloids 560 from plants extracts, however, analytical method in separation and quantification of canthin-6-one 561 alkaloids are limited.

Ferreira and coworkers characterized compounds of different organs of Zanthoxylum chiloperone by 562 HPLC-UV-MS qualitatively and quantitatively [6]. They investigated the ethanol extracts of stem 563 564 bark, leaves, fruits, root bark and resin as well as the dichloromethane extracts of stem bark and leaves. Qualitatively, canthin-6-one was identified as the major compound in the stem bark and root bark, 565 while 5-methoxy-canthin-6-one was the major compound in the leaves, fruits and resin, lastly canthin-566 567 6-one N-oxide was present in the resin and in the stem bark. Quantitative analysis was performed by 568 external standard method, using a calibration curve constructed utilizing solutions containing different 569 concentrations of the reference samples. Results showed relative compounds concentrations in the 570 different organs. Canthin-6-one concentrations in the ethanol extract of stem and the root bark are 64.7 571 and 8.2 mg/g extract respectively, while 5-methoxycanthin-6-one concentrations in the ethanol extract of the leaves, the fruits and the resin are 10.8, 3.5 and 5mg/g extracts respectively. Concerning 572 573 canthin-6-one N-oxide into the ethanol extracts of the stem bark and resin they found concentrations of 574 8.2 and 1.9 mg/g of extract respectively.

575 Similarly, Chaingam et al. [17] achieved analytical quantification on Eurycoma longifolia Jack and Eurvcoma harmandiana Pierre using high-performance liquid chromatography (HPLC) with UV 576 577 detection at 354 nm. E. harmandiana species contained high amount of total canthin-6-one alkaloids 578 (1.72±0.06 mg/g) compared to *E. longifolia* (0.16±0.01 mg/g). In addition, they tested different solvent 579 extracts (methanol, hydro-methanol (50%), ethanol, and hydro-ethanol (50%)), and showed maximum of canthin-6-one content was observed from 50% methanolic extract followed by 50% ethanolic, 580 methanol and ethanol extracts, respectively. The HPLC method seemed to be a simple, accurate and 581 582 selective analytical technique for separation and quantification of canthin-6-one alkaloids.

583 Cebrian and coworkers used another analytical method to estimation amounts of canthin-6-ones 584 alkaloids via cyclic voltammetry [8]. They studied leaves, fruits, seeds, stem barks and roots parts of 585 Zanthoxylum chiloperone var. angustifolium and estimated the relative amounts of canthin-6-one, 5-586 methoxycanthin-6-one and trans-avicennol by discussing the peak current values for the corresponding 587 cathodic peaks. Canthin-6-one was the most recurrent compound followed by 5-methoxycanthin-6-one then trans-avicennol in the different parts of the Zanthoxylum chiloperone species. Without resorting 588 589 to sample specific pre-treatment, voltammetry of microparticles methodology appeared to be a relevant 590 tool for phytochemical analysis.

A singular method has been employed by Parikrama *et al.* to increase secondary metabolites content from plant callus culture [184]. Elicitation technique was carried out to evaluate *Eurycoma longifolia* callus culture growth and to analyze its secondary metabolites production after elicitation. Ultraviolet light radiation slightly changed the structure of *Eurycoma longifolia* callus culture without decrease biomass growth. Alkaloids production from *E. longifolia* callus culture, particularly pyrrolidine and canthin-6-one could be increased by 18 hours of ultraviolet exposure daily. GCMS analysis showed that *E. longifolia* callus culture in Murashige-Skoog medium with naphthalene acetic acid and elicited

598 by UV light produced canthin-6-one alkaloid 3.5 times more and pyrrolidine 1.5 times more compared

599 to unelicited culture. It can be concluded that UV elicitation is a useful technique to enhance secondary 600 metabolites production from *E. longifolia* callus culture, by generating protein involved in alkaloids 601 and terpenoids synthesis pathway.

602

603 **7. Conclusions**

604 A thorough update over the last seventy years have been done on the canthin-6-one alkaloids 605 family. All the canthinone derivatives known to date have been listed and classified according to their 606 taxonomy. Their recurrence and diversity have been compared showing the poor diversity of the largest families. Biosynthesis and synthetic routes have been discussed. The latest were differentiated 607 by $C_8N_2 + C_6$, $C_{12}N_2 + C_2$, $C_{10}N_2 + C_2 + C_2$ and $C_{10}N_2 + C_4$ strategies. Biological activities of canthin-608 609 6-one alkaloids compounds were also enumerated with other activities such as their fluorescence 610 sensor capacity. Finally, recent relevant quantitative methods were quoted as well as a technique 611 allowing production of canthinone derivatives by stimulated plants. Further research, based on 612 calculation methods could be employed for a better understanding of structure-activity relationships 613 and mechanisms.

614 Acknowledgments

The authors thank the European Community for financial support (Marie Curie Early Stage Training
Fellowship of the European Community's Sixth Framework Programme: contract BioMedChem (for
G. C-T.)). The authors would like to acknowledge for financial support to the project
"AgroEcoDiv"and "CambioNET" from European Union Fund (FEDER and INTERREG).
We would also like to thank Prof. Dr. Erwan Poupon for fruitful discussions.

- 620 **References**
- 621
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623 References and Notes

- 624 [1] H. Haynes, E. Nelson, J. Price, Alkaloids of the Australian Rutaceae: *Pentaceras australis* Hook.
- F. I. Isolation of the alkaloids and identification of canthin-6-one. Aust. J. Chem. 5 (1952), 387
- 626 https://doi.org/10.1071/CH9520387.
- 627 [2] F. Bracher, T. Tremmel, The chemistry of the canthin-4-ones. MROC 14 (2017) 92–98,
 628 https://doi.org/10.2174/1570193X14666161110114103.
- 629 [3] N. Crespi-Perellino, A. Guicciardi, G. Malyszko, A. Minghetti, Occurrence of indole alkaloids in
- 630 *Ailanthus altissima* cell cultures. J. Nat. Prod. 49 (1986) 1010–1014,
 631 https://doi.org/10.1021/np50048a007.
- [4] H.D.H. Showalter, Progress in the synthesis of canthine alkaloids and ring-truncated congeners. J.
- 633 Nat. Prod. 76 (2013) 455–467, https://doi.org/10.1021/np300753z.

- [5] J. Dai, N. Li, J. Wang, U. Schneider, Fruitful decades for canthin-6-ones from 1952 to 2015:
 biosynthesis, chemistry, and biological activities. Molecules 21 (2016) 493,
 https://doi.org/10.3390/molecules21040493.
- [6] M.E. Ferreira, G. Cebrián-Torrejón, A. Segovia Corrales, N. Vera de Bilbao, M. Rolón, C. Vega
 Gomez, K. Leblanc, G. Yaluf, A. Schinini, S. Torres, E. Serna, A. Rojas de Arias, E. Poupon, A.
- Fournet, *Zanthoxylum chiloperone* leaves extract: First sustainable Chagas disease treatment. J.
 Ethnopharmacol. 133 (2011) 986–993, https://doi.org/10.1016/j.jep.2010.11.032.
- [7] J. Chaingam, T. Juengwatanatrakul, G. Yusakul, T. Kanchanapoom, W. Putalun, Simultaneous 641 determination of three canthin-6-one alkaloids in different extracts of Eurycoma longifolia and 642 643 Eurycoma harmandiana using HPLC-UV. Planta Med. 85 (2019)1441-1442, 644 https://doi.org/10.1055/s-0039-3399775.
- 645 [8] G. Cebrián-Torrejón, A. Doménech-Carbó, B. Figadère, E. Poupon, A. Fournet,
 646 Phytoelectrochemical analysis of *Zanthoxylum chiloperone*: An Electrochemical Approach to
 647 Phytochemical Screening. Phytochem. Anal. 28 (2017) 171–175, https://doi.org/10.1002/pca.2657.
- [9] T. Ohmoto, K. Koike, Canthin-6-one alkaloids, in: A. Brossi (Ed.) vol. 36, The Alkaloids:
 Chemistry and Pharmacology, Academic Press, Inc., New York, 1990, pp. 135–170.
- [10] K. González-García, H. Vélez-Castro, L. N. Paz, L. M. Montiel, J. A. González-Lavaut,
 Aislamiento y efecto antiinflamatorio de un alcaloide de *Zanthoxylum elephantiasis* Macf
 (Rutaceae) introducida en Cuba. Bol. Latinoam. Caribe Plant. Med. Aromaticas Vol. 7 (5) (2008)
 264 267.
- [11] A. T. Awad, J. L. Beal, S. K. Talapatra, M. P. Cava, Isolation and identification of three alkaloids
 from the bark of *Zanthoxylum elephantiasis*. J. Pharm. Sc. 56 (1967) 279–281,
 https://doi.org/10.1002/jps.2600560230.
- [12] C. W. Halstead, P. I. Forster, P. G. Waterman, Alkaloids from the stem bark of an Australian
 population of *Zanthoxylum ovalifolium*. Nat. Prod. Res. 20 (2006) 940–945,
 https://doi.org/10.1080/14786410500462710.
- [13] E. E. Diehl, G. L. von Poser, A. T. Henriques, Constituents of *Zanthoxylum rugosum* St.-Hil &
 Tul. Biochem. System. Ecol. 28 (2000) 275–277, https://doi.org/10.1016/S0305-1978(99)00030-7.

- [14] M. E. Ferreira, A. Rojas de Arias, S. Torres de Ortiz, A. Inchausti, H. Nakayama, C. Thouvenel
 R. Hocquemiller, A. Fournet, Leishmanicidal activity of two canthin-6-one alkaloids, two major
 constituents of *Zanthoxylum chiloperone* var. *angustifolium*. J. Ethnopharmacol. 80 (2002) 199202, https://doi.org/10.1016/S0378-8741(02)00025-9.
- [15] W. He, L. Van Puyvelde, N. De Kimpe, L. Verbruggen, K. Anthonissen, M. Van der Flaas, J.
 Bosselaers, S. G. Mathenge, F. P. Mudida, Chemical constituents and biological activities of
 Zanthoxylum usambarense. Phytother. Res. 16 (2002) 66–70, https://doi.org/10.1002/ptr.849.
- [16] J. J. Chen, H. Y. Fang, C. Y. Duh, I. S. Chen, New Indolopyridoquinazoline, benzo[*c*[70] [phenanthridines and cytotoxic constituents from *Zanthoxylum integrifoliolum*. Planta med. 71
 (2005) 470–475, https://doi.org/10.1055/s-2005-864144.
- [17] M. M. Rahman, M. A. Islam, P. Khondkar, A. I. Gray, Alkaloids and lignans from *Zanthoxylum budrunga* (Rutaceae). Biochem. Systematic. Ecol. 33 (2005) 91–96,
 https://doi.org/10.1016/j.bse.2004.04.016.
- [18] H. D. T. Mai, H. T. Van-Dufat, S. Michel, F. Tillequin, D. Bastien, T. Sévenet, New Diprenyl
 Coumarin and Alkaloids from the Bark of *Zanthoxylum dimorphophyllum* (Rutaceae). Zeitschrift
 für Naturforschung *C* 56 (2001) 492–494, https://doi.org/10.1515/znc-2001-7-802.
- [19] A. Ikuta, T. Nakamura, H. Urabe, Indolopyridoquinazoline, furoquinoline and canthinone type
 alkaloids from *Phellodendron amurense* callus tissues. Phytochem. 48 (1998) 285–291,
 https://doi.org/10.1016/S0031-9422(97)01130-8.
- [20] Y. D. Min, N. Y. Kim, E. J. Choi, K. H. Kim, J. O. Lee, Isolation of limonoids and alkaloids
 from *Phellodendron amurense* and their multidrug resistance (MDR) reversal activity. Arch.
 Pharm. Res. 30 (2007) 58–63, https://doi.org/10.1007/BF02977779.
- [21] M.M. Kaigongi, C.W. Lukhoba, S. Yaouba, N.P. Makunga, J. Githiomi, A. Yenesew, In Vitro
 Antimicrobial and antiproliferative activities of the root bark extract and isolated chemical
 constituents of *Zanthoxylum paracanthum* Kokwaro (Rutaceae). Plants 9 (2020) 920,
 https://doi.org/10.3390/plants9070920.
- 688 [22] L.K. Omosa, G.M. Mbogo, E. Korir, R. Omole, E.J. Seo, A. Yenesew, M. Heydenreich, J. O.
- 689 Midiwo, T. Efferth, Cytotoxic alkaloids from the root of Zanthoxylum paracanthum (mildbr)
- 690 Kokwaro. Nat. Prod. Res. 35 (2021) 579-586, https://doi.org/10.1080/14786419.2021.1913586.

- [23] N. Fukamiya, T. Aratani, K. Negoro, A.T. McPhail, M. Ju-Ichi, K.H. Lee, Antitumor agents, 79.
 Cytotoxic antileukemic alkaloids from *Brucea antidysenterica*. J. Nat. Prod. 49 (1986) 428–434, https://doi.org/10.1021/np50045a007.
- [24] Y. Ouyang, K. Koike, T. Ohmoto, Canthin-6-one alkaloids from *Brucea mollis* var. *tonkinensis*.
 Phytochem. 36 (1994) 1543–1546, https://doi.org/10.1016/S0031-9422(00)89758-7.
- 696 [25] M. A. Wagih, G. Alam, S. Wiryowidagdo, K. Attia, Improved production of the indole alkaloid
 697 canthin-6-one from cell suspension culture of *Brucea javanica* (L.) Merr. Indian J.Sci.Technol. 1
 698 (2008) 1-6, http://dx.doi.org/10.17485/ijst/2008/v1i7/29591.
- [26] E.R. Fo, J.B. Fernandes, C. Vieira, M.F. Silva, M. F. Canthin-6-one alkaloids from *Picrolemma granatensis*. Phytochem. 31 (1992) 2499–2501, https://doi.org/10.1016/0031-9422(92)83309-M.
- [27] P.C. Kuo, L.S. Shi, A.G. Damu, C.R. Su, C.H. Huang, C.H. Ke, J.B. Wu, K.F. Bastow, K.H. Lee,
 T.S. Wu, Cytotoxic and antimalarial β-carboline alkaloids from the roots of *Eurycoma longifolia*. J.
 Nat. Prod. 66 (2003) 1324–1327, https://doi.org/10.1021/np030277n.
- [28] K. Mitsunaga, K.K. Tomoko, Y. Ohkawa, Y. Kobayashi, T. Sawaguchi, T. Ohmoto, Canthin-6one alkaloids from *Eurycoma longifolia*. Phytochem. 35 (1994) 799–802,
 https://doi.org/10.1016/S0031-9422(00)90609-5.
- 707 [29] Y. Zhang, W. Zhao, J. Ruan, N. Wichai, Z. Li, L. Han, Y. Zhang, T. Wang, Anti-inflammatory
 708 canthin-6-one alkaloids from the roots of Thailand *Eurycoma longifolia* Jack. J. Nat. Med. 74
 709 (2020) 804–810, https://doi.org/10.1007/s11418-020-01433-6.
- [30] I. Ribeiro, A.M. Sevcsik, F. Alves, G. Diap, R. Don, M. O. Harhay, S. Chang, B. Pecoul, New,
 Improved Treatments for Chagas Disease: From the R&D Pipeline to the Patients. PLoS Negl
 Trop. Dis. 3 (2009) e484, https://doi.org/10.1371/journal.pntd.0000484.
- 713 [31] J.F. Rivero-Cruz, R. Lezutekong, T. Lobo-Echeverri, A. Ito, Q. Wi, H.B. Chai, D.D. Soejarto,
- 714 G.A. Cordell, J.M. Pezzuto, S.M. Swanson, I. Morelli, A.D. Kinghorn, Cytotoxic constituents of
- the twigs of Simarouba glauca collected from a plot in Southern Florida. Phytother. Res. 19
- 716 (2005) 136–140, https://doi.org/10.1002/ptr.1642.
- [32] T. Kanchanapoom, R. Kasai, P. Chumsri, Y. Hiraga, K. Yamasaki, Canthin-6-one and β-carboline
 alkaloids from *Eurycoma harmandiana*. Phytochem. 56 (2001) 383–386,
 https://doi.org/10.1016/S0031-9422(00)00363-0.

- [33] V.F. Noldin, D.T. Oliveira Martins, C.M. Marcello, J.C. Silva Lima, D.D. Monache, V.C. Filho,
 Phytochemical and antiulcerogenic properties of rhizomes from *Simaba ferruginea* St. Hill.
 (Simaroubaceae). Zeitschrift für Naturforschung C 60 (2005) 701–706,
 https://doi.org/10.1515/znc-2005-9-1007.
- [34] J. Ag Nuddin, A. Adam, A.S. Hamzah, Suggestive placement of *Quassia borneensis* Noot.
 (Simaroubaceae) as indicated by isolation of its quassinoids.. Jurnal Teknologi 77 (2015), https://doi.org/10.11113/jt.v77.5980.
- [35] Y.S. Makong, G. Mouthé Happi, J.L. Djouaka Bavoua, J.D. Wansi, L. Nahar, A.F. Kamdem
 Waffo, C. Martin, N. Sewald, S.D. Sarker, Cytotoxic stilbenes and canthinone alkaloids from *Brucea antidysenterica* (Simaroubaceae). Molecules 24 (2019) 4412,
 https://doi.org/10.3390/molecules24234412.
- [36] A. Harris, L. Anderson, J. Phillipson, R. Brown, Canthin-6-one alkaloids from *Brucea antidysenterica* root bark. Planta Med 51 (1985) 151–153, https://doi.org/10.1055/s-2007-969432.
- [37] T. Ohmoto, K. Koike, Studies on the constituents of *Picrasma quassioides* Bennet. II. On the
 alkaloidal constituents. Chem. Pharm. Bull. 31 (1983) 3198–3204,
 https://doi.org/10.1248/cpb.31.3198.
- [38] G.G. Zapesochnaya, L.N. Pervykh, V.A. Kurkin, A study of the herb *Aerva lanata*. III. Alkaloids.
 Chem. Nat. Compd. 27 (1991) 336–340, https://doi.org/10.1007/BF00630321.
- [39] G. Zapesochnaya, V. Kurkin, V. Okhanov, A. Miroshnikov, Canthin-6-one and β-carboline
 alkaloids from *Aerva lanata*. Planta Med. 58 (1992) 192–196, https://doi.org/10.1055/s-2006961427.
- [40] G. O'Donnell, S. Gibbons, S. Antibacterial activity of two canthin-6-one alkaloids from *Allium neapolitanum*. Phytother. Res. 21 (2007) 653–657, https://doi.org/10.1002/ptr.2136.
- 743 [41] M.G. Bröckelmann, J. Dasenbrock, B. Steffan, W. Steglich, Y. Wang, G. Raabe, J. Fleischhauer,
- An unusual series of thiomethylated canthin-6-ones from the North American mushroom *Boletus curtisii*. Eur. J. Org. Chem. (2004) 4856–4863, https://doi.org/10.1002/ejoc.200400519.
- 746 [42] S.A. Khan, K.M. Shamsuddin, 1-Hydroxycanthin-6-one, an alkaloid from Ailanthus giraldii.
- 747 Phytochem. 20 (1981) 2062–2063, https://doi.org/10.1016/0031-9422(81)84080-0.

- 748 [43] K. Miyake, Y. Tezuka, S. Awale, F. Li, S. Kadota, Canthin-6-one alkaloids and a tirucallanoid 749 from Eurycoma longifolia and their cytotoxic activity against a Human HT-1080 Fibrosarcoma 5 17–22, 750 cell line. Nat. Vol. (1)(2010)Prod. Comm. https://doi.org/10.1177/1934578X1000500105. 751
- 752 [44] R. dos Santos Costa, R.F. do Espirito-Santo, L. Silva Abreu, L. Oliveira Aguiar, D. Leites Fontes, 753 J. Fechine Tavares, M. Sobral da Silva, M. Botelho Pereira Soares, E. Silva Velozo, C. F. 754 Villareal, Fluorescent canthin-6-one alkaloids from Simaba bahiensis: isolation, identification, ChemPlusChem 84 (2019)755 and cell-labeling properties. 260-267. 756 https://doi.org/10.1002/cplu.201800591.
- [45] E.V. Lassak, J. Polonsky, H.Jacquemin, 5-hydroxycanthin-6-one from *Simarouba amara*.
 Phytochem. 16 (1977) 1126–1127, https://doi.org/10.1016/S0031-9422(00)86773-4.
- [46] M.X. Jiang, Y. J. Zhou, Canthin-6-one alkaloids from *Picrasma quassioides* and their cytotoxic
 activity. J. Asian Nat. Prod. Res. 10 (2008) 1009–1012,
 https://doi.org/10.1080/10286020802277956.
- [47] M. Y. Jiang, T. Feng, J. K. N. Liu, N-Containing compounds of macromycetes. Nat. Prod. Rep.
 28 (2011) 783, https://doi.org/10.1039/c0np00006j.
- [48] T.V.A. Tran, C. Malainer, S. Schwaiger, A.G. Atanasov, E.H. Heiss, V.M. Dirsch, H. Stuppner,
 NF-κB Inhibitors from Eurycoma longifolia. J. Nat. Prod. 77 (2014) 483–488,
 https://doi.org/10.1021/np400701k.
- 767 [49] L.B.S. Kardono, C.K. Angerhofer, S. Tsauri, K. Padmawinata, J.M. Pezzuto, A.D. Kinghorn,
- Cytotoxic and antimalarial constituents of the roots of Eurycoma longifolia. J. Nat. Prod. 54 (1991)
 1360–1367, https://doi.org/10.1021/np50077a020.
- [50] K.P. Devkota, J.A. Wilson Curtis, J. Henrich, J.B. McMahon, K.M. Reilly, J. Beutler,
 Compounds from *Simarouba berteroana* which inhibit proliferation of NF1-defective cancer cells.
 Phytochem. Lett. 7 (2014) 42–45, https://doi.org/10.1016/j.phytol.2013.09.007.
- [51] P.J. Clarke, K. Jewers, H.F. Jones, Crystal and molecular structure of amarorine monohydrate, a
- new phenolic alkaloid from Amaroria soulameoides. J. Chem. Soc., Perkin Trans. 1 (1980) 1614–
- 775 1617, https://doi.org/10.1039/p19800001614.

- [52] W.Y. Zhao, Y.S. Shang, L. Zhao, G.D. Yao, Z. Sun, X.X. Huang, S.J. Song, Bioactivity-guided
 isolation of β-carboline alkaloids with potential anti-hepatoma effect from *Picrasma quassioides*(D. Don) Benn. Fitoter. 130 (2018), 66–72, https://doi.org/10.1016/j.fitote.2018.08.004.
- [53] P.W. Hsieh, F.R. Chang, K.H. Lee, T.L. Hwang, S.M. Chang, Y.C. Wu, A new anti-HIV alkaloid,
- drymaritin, and a new C-glycoside flavonoid, diandraflavone, from Drymaria diandra. J. Nat.
 Prod. 67 (2004) 1175–1177, https://doi.org/10.1021/np0400196.
- [54] Z. Xu, F.R. Chang, H.K. Wang, Y. Kashiwada, A.T. McPhail, K.F. Bastow, Y. Tachinabana, M.
 Cosentino, K.H. Lee, Anti-HIV agents 45 and antitumor agents 205. Two new sesquiterpenes,
 leitneridanins A and B, and the cytotoxic and anti-HIV principles from *Leitneria f loridana*. J.
 Nat. Prod. 63 (2000) 1712–1715, https://doi.org/10.1021/np000260u.
- [55] R. Costa, D. Fontes, L. Abreu, J. Tavares, E. Velozo, NMR analysis of a new canthin-6-one
 alkaloid from Simaba bahiensis (Simaroubaceae), in Proceedings of MOL2NET 2017, *International Conference on Multidisciplinary Sciences, 3rd edition* 5056 (MDPI, 2017),
 https://doi.org/10.3390/mol2net-03-05056.
- [56] V. Njar, T. Alao, J. Okogun, H. Holland, 2-methoxycanthin-6-one: A new alkaloid from the stem
 wood of *Quassia amara*. Planta Med. 59 (1993) 259–261, https://doi.org/10.1055/s-2006-959664.
- [57] I. Wetzel, L. Allmendinger, F. Bracher, Revised structure of the alkaloid drymaritin. J. Nat.
 Prod. 72 (2009) 1908–1910, https://doi.org/10.1021/np900515b.
- 794 [58] G. Cebrián-Torrejón, L. Kablan, M.E. Ferreira, D. Rodríguez de la Cruz, A. Doménech-Carbó, N. 795 Vera de Bilbao, A. Rojas de Arias, B. Figadère, E. Poupon, A. Fournet A. Harvesting canthinones: 796 identification of the optimal seasonal point of harvest of Zanthoxylum chiloperone leaves as a 797 29 (2015)source of 5-methoxycanthin-6-one. Nat. Prod. Res. 2054-2058, 798 https://doi.org/10.1080/14786419.2015.1022774.
- 799 [59] X. Wang, Y. Lee, Efficient synthesis of substituted pyranoquinolinones from 2,4800 dihydroxy-quinoline: total synthesis of zanthosimuline, *cis* -3 ',4 ' -dihydroxy-3 ',4 ' 801 di-hydroflindersine, and orixalone D. Synthesis (2007) 3044–3050, https://doi.org/10.1055/s-2007802 983893.

- [60] K.E. Readel, D.S. Seigler, D.A. Young, 5-methoxycanthin-6-one from *Leitneria floridana*(Simaroubaceae). Biochem. System. Ecol. 31 (2003) 167–170, https://doi.org/10.1016/S03051978(02)00074-1.
- [61] A.M. Giesbrecht, H.E. Gottlieb, O.R. Gottlieb, M.O.F. Goulart, R.A. De Lima, A.E.G. Sant'ana,
 Canthinones from *Simaba cuspidata*. Phytochem. 19 (1980) 313–315,
 https://doi.org/10.1016/S0031-9422(00)81981-0.
- [62] J. Polonsky, J. Gallas, J. Varenne, T. Prangé, C. Pascard, H. Jacquemin, C. Moretti, Isolation and
 structure (x-ray analysis) of karinolide, a new quassinoid from *Simaba multiflora*. Tetrahedron
 Lett. 23 (1982) 869–872, https://doi.org/10.1016/S0040-4039(00)86970-1.
- [63] R. de C. Saraiva, A.C. Pinto, S.M. Nunomura, A.M. Pohlit, Triterpenes and a canthinone alkaloid
 from the stems of *Simaba polyphylla* (Cavalcante) W.W. Thomas (Simaroubaceae). Quím. Nova
 29 (2006) 264–268, https://doi.org/10.1590/S0100-40422006000200017.
- [64] W.H. Jiao, H. Gao, F. Zhao, H.W. lin, Y.M. Pan, G.X. Zhou, X.S. Yao, Anti-inflammatory
 alkaloids from the stems of *Picrasma quassioides* Bennet. Chem. Pharm. Bull. 59 (2011) 359–364,
 https://doi.org/10.1248/cpb.59.359.
- [65] L.P. Zhang, J.Y. Wang, W. Wang, Y.X. Cui, D.L. Cheng, Two new alkaloidal glycosides from
 the root bark of *Ailanthus altissima*. J. Asian Nat. Prod. Res. 9 (2007) 253–259,
 https://doi.org/10.1080/10286020600604286.
- [66] H.M. Kim, S.J. Kim, H.Y. Kim, B. Ryu, H. Kwak, J. Hur, J.H. Choi, D.S. Jang, Constituents of
 the stem barks of *Ailanthus altissima* and their potential to inhibit LPS-induced nitric oxide
 production. Bioorg. Med. Chem. Lett. 25 (2015) 1017–1020,
 https://doi.org/10.1016/j.bmcl.2015.01.034.
- [67] C. López, M. Pastrana, A. Ríos, A. Cogollo, A. Pabón, Huberine, a new canthin-6-one alkaloid
 from the bark of *Picrolemma huberi*. Molecules 23 (2018) 934,
 https://doi.org/10.3390/molecules23040934.
- 828 [68] T. Ohmoto, K. Koike, Studies on the alkaloids from Picrasma quassioides Bennet. VI. Structures 829 N. 0. Chem. Pharm. Bull. 33 (1985)4901-4905, of picrasidines О. and 830 https://doi.org/10.1248/cpb.33.4901.

- [69] T. Ohmoto, K. Koike, Studies on the constituents of *Picrasma quassioides* Bennet. III. The
 alkaloidal constituents. Chem. Pharm. Bull. 32 (1984) 3579–3583,
 https://doi.org/10.1248/cpb.32.3579.
- [70] Y. Kimura, M. Takido, S. Koizumi, Studies on the standardization of crude drugs. XIX.: The
 constituents of *Picrasma ailanthoides*. I.: The structure of nigakinone. Yakugaku Zasshi 87 (1967)
 1371–1373, https://doi.org/10.1248/yakushi1947.87.11_1371.
- [71] P. Barbetti, G. Grandolini, G. Fardella, I. Chiappini, A. Mastalia, New canthin-6-one alkaloids
 from *Quassia amara*. Planta Med. 56 (1990) 216–217, https://doi.org/10.1055/s-2006-960928.
- 839 [72] Z.Q. Lai, W.H. Liu, S.P. Ip, H.J. Liao, Y.Y. Yi, Z. Qin, X.P. Lai, Z.R. Su, Z.X. Lin, Wei-Hai Liu,
- Seven alkaloids from *Picrasma quassioides* and their cytotoxic activities. Chem. Nat. Compd. 50
 (2014) 884–888, https://doi.org/10.1007/s10600-014-1106-6.
- [73] G. Gong, Q. Lin, J. Xu, F. Ye, L. Jiang, W. Liu, M.F. He, F. Feng, W. Qu, N. Xie, In vivo SAR
 and STR analyses of alkaloids from *Picrasma quassioides* identify 1-hydroxymethyl-8-hydroxy-βcarboline as a novel natural angiogenesis inhibitor. RSC Adv. 6 (2016) 9484–9494,
 https://doi.org/10.1039/C5RA22391A.
- [74] H.Y. Li, K. Koike, T. Ohmoto, New alkaloids, picrasidines W, X and Y, from *Picrasma quassioides* and X-ray crystallographic analysis of picrasidine Q. Chem. Pharm. Bull. 41 (1993)
 1807–1811, https://doi.org/10.1248/cpb.41.1807.
- 849 [75] S. Moladje Donkwe, E.N. Happi, J.D. Wansi, B.N. Lenta, K.P. Devkota, B. Neumann, H.G.
 850 Stammler, N. Sewald, Oxidative burst inhibitory and cytotoxic activity of constituents of the fruits
- of *Odyendyea gabonensis*. Planta Med. 78 (2012) 1949–1956, https://doi.org/10.1055/s-00321327878.
- [76] Z.Z. Ma, Y. Hano, T. Nomura, Y.J. Chen, Alkaloids and phenylpropanoids from *Peganum nigellastrum*. Phytochem. 53 (2000) 1075–1078, https://doi.org/10.1016/S0031-9422(99)00440-9.
- [77] T. Ohmoto, K. Koike, Studies on the constituents of *Ailanthus altissima* Swingle. III. The
 alkaloidal constituents. Chem. Pharm. Bull. 32 (1984) 170–173,
 https://doi.org/10.1248/cpb.32.170.

- [78] H. Kim, J.S. Lee, J. Sezirahiga, J. Kwon, M. Jeong, D. Lee, J.H. Choi, D.S.A. Jang, New
 canthinone-type alkaloid isolated from *Ailanthus altissima* Swingle. Molecules 21 (2016), 642,
 https://doi.org/10.3390/molecules21050642.
- [79] H.I.A. El-Moaty, Investigation of some active constituents of *Euphorbia bivonae* Steud. IAJPS 3
 (2016) 1309–1315, https://doi.org/10.5281/ZENODO.179177.
- [80] F. Soriano-Agatón, D. Lagoutte, E. Poupon, F. Roblot, A. Fournet, J.C. Gantier, R. Hocquemiller,
 Extraction, hemisynthesis, and synthesis of canthin-6-one analogues. Evaluation of their antifungal
 activities. J. Nat. Prod. 68 (2005) 1581–1587, https://doi.org/10.1021/np050250z.
- [81] T. Ohmoto, K. Koike, Y. Sakamoto, Studies on the constituents of Ailanthus altissima Swingle.
- 867 II. Alkaloidal constituents. Chem. Pharm. Bull. 29 (1981) 390–395,
 868 https://doi.org/10.1248/cpb.29.390.
- 869 [82] M. Arisawa, A.D. Kinghorn, G.A. Cordell, N.R. Farnsworth, Plant anticancer agents. XXIV. 870 multiflora. J. Prod. Alkaloid constituents of Simaba Nat. 46 (1983)222-225, 871 https://doi.org/10.1021/np50026a016.
- [83] T. Ohmoto, K. Koike, Studies on the constituents of *Picrasma quassioides* Bennet. I. On the
 alkaloidal constituents. Chem. Pharm. Bull. 30 (1982) 1204–1209,
 https://doi.org/10.1248/cpb.30.1204.
- 875 [84] J. Liu, R.S. Davidson, R.V.D. Heijden, R. Verpoorte, O.W. Howarth, Isolation of 4-Hydroxy-5-
- 876 methoxycanthin-6-one from *Picrasma quassioides* and revision of a previously reported structure.
- ktps://doi.org/10.1002/jlac.1992199201162.
- 878 [85] T. Ohmoto, K. Koike, Studies on the alkaloids from Picrasma quassioides Bennet V. Structures 879 of picrasidines Bull. (1985)L.M. and P. Chem. Pharm. 33 3847-3851. 880 https://doi.org/10.1248/cpb.33.3847.
- [86] K. Koike, T. Ohmoto, Picrasidine-U, dimeric alkaloid from *Picrasma quassioides*. Phytochem. 27
 (1988) 3029–3030, https://doi.org/10.1016/0031-9422(88)80725-8.
- [87] K. Taniguchi, S. Takizawa, T. Hirano, S. Murata, H. Kagechika, A. Kishida, A. Ohsaki*K.
 Amarastelline A: A fluorescent alkaloid from *Quassia amara* and its properties in living cells.
 ChemPlusChem 77 (2012) 427–431, https://doi.org/10.1002/cplu.201200016.

- [88] N. Tanaka, R. Momose, Y. Takahashi, T. Kubota, A. Takahashi-Nakaguchi, T. Gonoi, J.
 Fromont, J. Kobayshi, Hyrtimomines D and E, bisindole alkaloids from a marine sponge *Hyrtios*sp. Tetrahedron Lett. 54 (2013) 4038–4040, https://doi.org/10.1016/j.tetlet.2013.05.073.
- [89] H. Takahashi, S. Kurimoto, J. Kobayashi, T. Kubota, Ishigadine A, a new canthin-6-one alkaloid
 from an Okinawan marine sponge *Hyrtios* sp. Tetrahedron Lett. 59 (2018) 4500–4502,
 https://doi.org/10.1016/j.tetlet.2018.11.019.
- 892 [90] L.A. Anderson, A. Harris, J.D. Phillipson, Production of cytotoxic canthin-6-one alkaloids by 893 cell 46 Ailanthus altissima plant cultures. J. Nat. Prod. (1983)374-378. 894 https://doi.org/10.1021/np50027a014.
- [91] L.A. Anderson, C.A., Hay, M.F. Roberts, J.D. Phillipson, Studies on *Ailanthus altissima* cell
 suspension cultures: Precursor feeding of L-[methylene-14C]tryptophan and L-tryptophan. Plant
 Cell Reports. 5 (1986) 387–390, https://doi.org/10.1007/BF00268609.
- [92] L.A. Anderson, C.A. Hay, J.D. Phillipson, M.F. Roberts, Studies on *Ailanthus altissima* cell
 suspension cultures. Uptake of L-[methyl-14C]methionine and incorporation of label into 1methoxycanthin-6-one. Plant Cell Reports 6 (1987) 242–243,
 https://doi.org/10.1007/BF00268490.
- 902 [93] N. Crespi-Perellino, A. Guicciardi, G. Malyszko, A. Minghetti, Biosynthetic relationship between
 903 indole alkaloids produced by cell cultures of *Ailanthus altissima*. J. Nat. Prod. 49 (1986) 814–822,
 904 https://doi.org/10.1021/np50047a009.
- 905 [94] F. Aragozzini, E. Maconi, R. Gualandris, Evidence for involvement of ketoglutarate in the
 906 biosynthesis of canthin-6-one from cell cultures of *Ailanthus altissima*. Plant Cell Reports 7 (1988)
 907 213–215, https://doi.org/10.1007/BF00269327.
- 908 [95] G. Cebrián-Torrejón, N. Mackiewicz, R.P. Vazquez-Manrique, A. Fournet, B. Figadère, J.
 909 Nicolas, E. Poupon, Solution phase and nanoparticular biosynthetically inspired interconnections
 910 in the canthin-6-one β-carboline series and study of phenotypic properties on *C. elegans*:
 911 generation of β-carboline analogs and biological study. Eur. J. Org. Chem. (2013) 5821–5828,
 912 https://doi.org/10.1002/ejoc.201300770.
- 913 [96] H.X. Fan, Z.Q. Zhou, J. Peng, B.J. Wu, H.R. Chen, X.F. Bao, Z.Q. Mu, W.H. Jiao, X.S. Yao,
- 914 H.G. Gao, A microbial model of mammalian metabolism: biotransformation of 4,5-dimethoxyl-

- 915 canthin-6-one using *Cunninghamella blakesleeana* CGMCC 3.970. Xenobiotica 47 (2017) 284–
 916 289, https://doi.org/10.1080/00498254.2016.1184774.
- 917 [97] J. Kobayashi, Y Ohizumi, M.R. Walchli, H. Nakamura, Y. Hirata, T. Sasaki, J. Kobayashi,
 918 Eudistomidins B, C, and D: novel antileukemic alkaloids from the Okinawan marine tunicate
 919 *Eudistoma glaucus*. J. Org. Chem. 55 (1990) 3666–3670, https://doi.org/10.1021/jo00298a056.
- 920 [98] F. Hamaguchi, S. Ohki, S. Synthesis of 1,3-Bis(hydroxymethyl)-b-carboline Derivatives.
 921 Heterocycles 8 (1977) 383, https://doi.org/10.3987/S(S)-1977-01-0383.
- 922 [99] D. Soerens, J. Sandrin, G. Ungemach, P. Mokry, G.S. Wu, E. Yamanaka, L. Hutchins, M. Di Pierro, J.M. Cook, Study of the Pictet-Spengler reaction in aprotic media: synthesis of the .beta.-923 924 galactosidase inhibitor, pyridindolol. J. Org. Chem. 44 (1979)535-545, https://doi.org/10.1021/jo01318a014. 925
- [100] B.C. VanWagenen, J.H. Cardellina, Short, efficient syntheses of the antibiotic eudistomins I and
 T. Tetrahedron Lett. 30 (1989) 3605–3608, https://doi.org/10.1016/S0040-4039(01)80460-3.
- [101] K.L. Rinehart, J. Kobayashi, G.C. Harbour, J. Gilmore, M. Mascal, T.G. Holt, L.S. Shield, F.
 Lafargue, Eudistomins A-Q, .beta.-carbolines from the antiviral Caribbean tunicate *Eudistoma olivaceum*. J. Am. Chem. Soc. 109 (1987) 3378–3387, https://doi.org/10.1021/ja00245a031.
- 931 [102] M. Cain, O. Campos, F. Guzman, J.M. Cook, Selenium dioxide oxidations in the indole area. 932 **Synthesis** of alkaloids. Soc. 105 (1983)907-913, β-carboline J. Am. Chem. 933 https://doi.org/10.1021/ja00342a045.
- [103] G.C. Condie, J. Bergman, Synthesis of some fused β-carbolines including the first example of
 the pyrrolo[3,2- c]-β-carboline system. J. Heterocyclic Chem. 41 (2004) 531–540,
 https://doi.org/10.1002/jhet.5570410409.
- 937 [104] G.C. Condie, J. Bergman, Reactivity of β-carbolines and cyclopenta[b]indolones prepared from
 938 the intramolecular cyclization of 5(4H)-oxazolones derived from L-tryptophan. Eur. J. Org. Chem.
 939 (2004) 1286–1297, https://doi.org/10.1002/ejoc.200300673.
- 940 [105] K.M. Czerwinski, C.A. Zificsak, J. Stevens, M. Oberbeck, C. Randlett, M. King, S. Mennen,
- An Improved Synthesis of Canthin-6-one. Synthetic Communications 33 (2003), 1225–1231,
 https://doi.org/10.1081/SCC-120017199.

- 943 [106] M.R.D. Giudice, F. Gatta, G. Settimj, New tetracyclic compounds containing the β-carboline
 944 moiety. J. Heterocyclic Chem. 27 (1990) 967–973, https://doi.org/10.1002/jhet.5570270427.
- [107] A. Gollner, P.A. Koutentis, Two-step total syntheses of canthin-6-one alkaloids: new one-pot
 sequential Pd-catalyzed Suzuki–Miyaura coupling and Cu-catalyzed amidation reaction. Org. Lett.
- 947 12 (2010) 1352–1355, https://doi.org/10.1021/ol100300s.
- [108] F. Guzman, M. Cain, P. Larscheid, T. Hagen, J.M. Cook, M. Schweri, P. Skolnick, S.M.
 Paul, Biomimetic approach to potential benzodiazepine receptor agonists and antagonists. J. Med.
 Chem. 27 (1084) 564–570, https://doi.org/10.1021/jm00371a002.
- [109] T.J. Hagen, J.M. Cook, Synthesis of 1-methoxy canthine-6-one. Tetrahedron Lett. 29 (1988)
 2421–2424, https://doi.org/10.1016/S0040-4039(00)87897-1.
- [110] T.J. Hagen, K. Narayanan, J. Names, J.M. Cook, DDQ oxidations in the indole area. Synthesis
 of 4-alkoxy-.beta.-carbolines including the natural products crenatine and 1-methoxycanthin-6one. J. Org. Chem. 54 (1989) 2170–2178, https://doi.org/10.1021/jo00270a029.
- 956 [111] H.A. Ioannidou, A. Martin, A. Gollner, P.A. Koutentis, Three-step synthesis of ethyl canthinone-
- 957 3-carboxylates from ethyl 4-bromo-6-methoxy-1,5-naphthyridine-3-carboxylate via a Pd-catalyzed
- 958 Suzuki–Miyaura coupling and a Cu-catalyzed amidation reaction. J. Org. Chem. 76 (2011) 5113–
- 959 5122, https://doi.org/10.1021/jo200824b.
- 960 [112] C.W. Lindsley, D.D. Wisnoski, Y. Wang, W.H. Leister, Z.A. Zhao, A 'one pot' microwave961 mediated synthesis of the basic canthine skeleton: expedient access to unnatural β-carboline
 962 alkaloids. Tetrahedron Lett. 44 (2003) 4495–4498, https://doi.org/10.1016/S0040-4039(03)01019963 0.
- 964 [113] J.H. Markgraf, A.A. Dowst, L.A. Hensley, C.E. Jakobsche, C.J. Kaltner, P.J. Webb, P.W.
 965 Zimmerman, A versatile route to benzocanthinones. Tetrahedron 61 (2005) 9102–9110, 966 https://doi.org/10.1016/j.tet.2005.07.034.
- [114] K. Narayanan, L. Schindler, J.M. Cook, Carboxyl-mediated Pictet-Spengler reaction. Direct
 synthesis of 1,2,3,4-tetrahydro-.beta.-carbolines from tryptamine-2-carboxylic acids. J. Org. Chem.
 56 (1991) 359–365, https://doi.org/10.1021/jo00001a066.
- 970 [115] K.C. Nicolaou, U. Majumder, S.P. Roche, D.Y.K. Chen, Construction of the "Left Domain" of
- 971 haplophytine. Angew. Chem. 119 (2007) 4799–4802, https://doi.org/10.1002/ange.200701947.

- [116] K.C. Nicolaou, T. Lister, R.M. Denton, C.F. Gelin, Cascade reactions involving formal [2+2]
 thermal cycloadditions: total synthesis of artochamins F, H, I, and J. Angew. Chem. Int. Ed. 46
 (2007) 7501–7505, https://doi.org/10.1002/anie.200702363.
- [117] P.D. Rege, Y. Tian, E.J. Corey, Studies of new indole alkaloid coupling methods for the
 synthesis of haplophytine. Org. Lett. 8 (2006) 3117–3120, https://doi.org/10.1021/ol061319c.
- [118] H.J. Rosenkranz, G. Botyos, H. Schmid, Synthese von tuboflavin, 4-äthyl-canthin-6-on und
 canthin-6-on. Justus Liebigs Ann. Chem. 691 (1966) 159–164,
 https://doi.org/10.1002/jlac.19666910120.
- 980 [119] V. Singh, S. Hutait, S. Batra, Baylis–Hillman reaction of 1-formyl-β-carboline: one-step
 981 synthesis of the canthin-6-one framework by an unprecedented cascade cyclization reaction. Eur.
 982 J. Org. Chem. (2009) 6211–6216, https://doi.org/10.1002/ejoc.200900962.
- [120] H.R. Snyder, F.X. Werber, Polyphosphoric acid as a dehydrating Agent. I. The cyclodehydration
 of some α-acylamino-β-arylpropionic acids. J. Am. Chem. Soc. 72 (1950) 2962–2965,
 https://doi.org/10.1021/ja01163a041.
- [121] H. Suzuki, K. Sakurai, K. Tokumoto, H. Takahashi, M. Hanada, Y. Yokoyama, Y. Murakami, A
 general synthetic route for 1-substituted 4-oxygenated β-carbolines (Synthetic studies on indoles
 and related compounds 41). Tetrahedron 53 (1997) 1593–1606, https://doi.org/10.1016/S00404020(96)01112-X.
- [122] H. Suzuki, M. Adachi, Y. Ebihara, H. Gyoutoku, H. Furuya, Y. Murakami, H. Okuno*et*, A total
 synthesis of 1-methoxycanthin-6-one: An efficient one-pot synthesis of the canthin-6-one skeleton
 from β-carboline-1-carbaldehyde. Synthesis 1 (2005) 28–32, https://doi.org/10.1055/s-2004834915.
- W.D. Shipe, Y. Feng, Z. Zhijiang, Convenient and general microwave-assisted protocols for
 the expedient synthesis of heterocycles. Heterocycles 70 (2006) 655, https://doi.org/10.3987/REV 06-SR(W)1.
- 997 [124] H.W. Fang, Y.R. Liao, T.L. Hwang, P.C. Shieh, K.H. Lee, H.Y. Hung, T.S. Wu, Total synthesis
 998 of cordatanine, structural reassignment of drymaritin, and anti-inflammatory activity of synthetic
 999 precursors. Bioorg. Med. Chem. Lett. 25 (2015) 3822–3824,
 1000 https://doi.org/10.1016/j.bmcl.2015.07.074.

- [125] A. Puzik, F. Bracher, A convenient approach to the canthin-4-one ring system: Total synthesis of
 the alkaloids tuboflavine and norisotuboflavine. J. Heterocyclic Chem. 46 (2009) 770–773,
 https://doi.org/10.1002/jhet.126.
- 1004 [126] T. Tremmel, F. Bracher, New approaches to the synthesis of canthin-4-one alkaloids and 1005 synthetic analogues. Tetrahedron 71 (2015) 4640–4646, https://doi.org/10.1016/j.tet.2015.05.002.
- 1006 [127] U. Rößler, S. Blechert, E. Steckhan, E. Single electron transfer induced total synthesis of
 1007 canthin-6-one. Tetrahedron Lett. 40 (1999) 7075–7078, https://doi.org/10.1016/S00401008 4039(99)01472-0.
- [128] S.U. Dighe, S.K. Samanta, S. Kolle, S. Batra, Iodine-mediated oxidative Pictet-Spengler
 reaction using terminal alkyne as the 2-oxoaldehyde surrogate for the synthesis of 1-aroyl-βcarbolines and fused-nitrogen heterocycles. Tetrahedron 73 (2017) 2455–2467,
 https://doi.org/10.1016/j.tet.2017.03.031.
- [129] S.V. Shelar, N.P. Argade, Total synthesis of bioactive canthine alkaloid cordatanine comprising
 in situ double oxidative aromatization of tetrahydrocarbazole. ACS Omega 2 (2017) 3945–3950,
 https://doi.org/10.1021/acsomega.7b00609.
- [130] O.A. Sen'ko, A.G. Dybenko, M.M. Garazd, V.G. Kartsev, Synthesis of benzoannelated canthin6-one analogs. Chem. Nat. Compd. 53 (2017) 523–528, https://doi.org/10.1007/s10600-017-20379.
- [131] E. Broumidis, P.A. Koutentis, A one-pot, two-step synthesis of 3-deazacanthin-4-ones via
 sequential Pd-catalyzed Suzuki-Miyaura and Cu-catalyzed Buchwald-Hartwig reactions.
 Tetrahedron Lett. 58 (2017) 2661–2664, https://doi.org/10.1016/j.tetlet.2017.05.076.
- [132] S.C. Benson, J.H. Li, J.K. Snyder, Indole as a dienophile in inverse electron demand Diels-Alder
 reactions. 3. Intramolecular reactions with 1,2,4-triazines to access the canthine skeleton. J. Org.
 Chem. 57 (1992) 5285–5287, https://doi.org/10.1021/jo00046a005.
- Selective oxidation of canthines 1025 [133] J.H. Li, J.K. Snyder, to canthin-6-ones with 1026 triethylbenzylammonium permanganate. Tetrahedron Lett. 35 (1994)1485-1488, https://doi.org/10.1016/S0040-4039(00)76738-4. 1027
- 1028 [134] L.F. Domingues Passero, M.D. Laurenti, G. Santos-Gomes, B.L. Soares Campos, P. Sartorelli,
- 1029 J.H. Lago, In Vivo Antileishmanial Activity of Plant-Based Secondary Metabolites, in: M. K. Rai

- and K. V. Kon (Eds.), Fighting Multidrug Resistance with Herbal Extracts, Essential Oils and
 Their Components, Elsevier Inc., London, 2013, pp. 95-107.
- [135] C.M.B. Leite Silva, F. Pelegrin Garcia, J.H. da Silva Rodrigues, C.V. Nakamura, T. Ueda-1032 1033 Nakamura, E. Meyer, A.L. Tasca Gois Ruiz, M.A. Foglio, J.E. de Carvalho, W. Ferreira da Costa, M.H. Sarragiotto, Synthesis, antitumor, antitrypanosomal and antileishmanial activities of 1034 1035 benzo[4,5] canthin-6-ones bearing the N'-(substituted benzylidene)-carbohydrazide and N-(2012)1372-1379. 1036 alkylcarboxamide groups C-2. Chem. Pharm. Bull. 60 at https://doi.org/10.1248/cpb.c12-00356. 1037
- 1038 [136] G. Cebrian-Torrejon, K. Spelman, K. Leblanc, K. Muñoz-Durango, S. Torijano Gutierrez , M.E.
- 1039 Ferreira M.E., A. Rojas de Arias, B. Figadère, A. Fournet, A. Maciuk, P. Greller, N.B. Cech, E.
- Poupon, The antiplasmodium effects of a traditional South American remedy: *Zanthoxylum chiloperone* var. *angustifolium* against chloroquine resistant and chloroquine sensitive strains of *Plasmodium falciparum*. Rev. Bras. Farmacogn. 21 (2011) 652–661,
 https://doi.org/10.1590/S0102-695X2011005000104.
- 1044 [137] K. Kaur, M. Jain, T. Kaur, R. Jain, Antimalarials from nature. Bioorg. Med. Chem. 17 (2009)
 1045 3229–3256, https://doi.org/10.1016/j.bmc.2009.02.050.
- [138] C. Thouvenel, J.C. Gantier, P. Duret, C. Fourneau, R. Hocquemiller, M.E. Ferreira, A. Rojas de
 Arias, A. Fournet, Antifungal compounds from *Zanthoxylum chiloperone* var. *angustifolium*.
 Phytother. Res. 17 (2003) 678–680, https://doi.org/10.1002/ptr.1137.
- [139] D. Lagoutte, V. Nicolas, E. Poupon, A. Fournet, R. Hocquemiller, D. Libong, P. Chaminade, P.
 Loiseau, Antifungal canthin-6-one series accumulate in lipid droplets and affect fatty acid
 metabolism in *Saccharomyces cerevisiae*. Biomed. Pharmacother. 62 (2008) 99–103,
 https://doi.org/10.1016/j.biopha.2007.07.014.
- 1053 [140] C. Dejos, M. Régnacq, M. Bernard, P. Voisin, T. Bergès, The MFS-type efflux pump Flr1
- induced by Yap1 promotes canthin-6-one resistance in yeast. FEBS Lett. 587 (2013) 3045–3051,
 https://doi.org/10.1016/j.febslet.2013.07.040.
- 1056 [141] V.F. Gazoni, S.O. Balogun, K. Arunachalam, D.M. Oliveira, V.C. Filho, S.R. Lima, E.M.
- 1057 Colodel, I. Mendes Soares, S. Donizetti Ascencio, D.T.D. Oliveira Martins, Assessment of toxicity
- 1058 and differential antimicrobial activity of methanol extract of rhizome of Simaba ferruginea A. St.-

- Hil. and its isolate canthin-6-one. J. Ethnopharmacol. 223 (2018) 122–134,
 https://doi.org/10.1016/j.jep.2018.05.014.
- [142] F. Zhao, J.K. Dai, D. Liu, S.J. Wang, J.R. Wang, Synthesis and evaluation of ester derivatives of
 10-hydroxycanthin-6-one as potential antimicrobial agents. Molecules 21 (2016) 390,
 https://doi.org/10.3390/molecules21030390.
- [143] C. Murakami, N. Fukamiya, S. Tamura, M. Okano, K.F. Bastow, H. Tokuda, T. Mukainaka, H.
 Nishino, K.H. Lee, Multidrug-resistant cancer cell susceptibility to cytotoxic quassinoids, and
 cancer chemopreventive effects of quassinoids and canthin alkaloids. Bioorg. Med. Chem. 12
 (2004) 4963–4968, https://doi.org/10.1016/j.bmc.2004.06.045.
- [144] N. Fukamiya, M. Okano, T. Aratani, K. Negoro, Y.M. Lin, K.H. Lee, Antitumor Agents:
 LXXXVII. Cytotoxic antileukemic canthin-6-one alkaloids from *Brucea antidysenterica* and the
 structure activity relationships of their related derivatives ¹. Planta Med. 53 (1987) 140–143,
 https://doi.org/10.1055/s-2006-962656.
- 1072 [145] S.S. Handa, A.D. Kinghorn, G.A. Cordell, N.R. Farnsworth, Plant anticancer agents XXV.
 1073 Constituents of *Soulamea soulameoides*. J. Nat. Prod. 46 (1983) 359–364,
 1074 https://doi.org/10.1021/np50027a011.
- 1075 [146] A. Peduto, V. More, P. de Caprariis, M. Festa, A. Capasso, S. Piacente, L. de Martino, V. de
 1076 Feo, R. Filosa, Synthesis and cytotoxic activity of new β-carboline derivatives. *MRMC* 11 (2011)
 1077 486–491, https://doi.org/10.2174/138955711795843383.
- 1078 [147] K. Koike, H. Yoshino, T. Nikaido, Synthesis and antitumor activity of canthin-5,6-dione
 1079 derivatives. Heterocycles 51 (1999) 315, https://doi.org/10.3987/COM-98-8394.
- [148] C. Dejos, P. Voisin, M. Bernard, M. Régnacq, T. Bergès, Canthin-6-one displays
 antiproliferative activity and causes accumulation of cancer cells in the G2/M phase. J. Nat. Prod.
 77 (2014) 2481–2487, https://doi.org/10.1021/np500516v.
- 1083 [149] G. Cebrián-Torrejón, S. Assad Kahn, M.E. Ferreira, C. Thirant, A. Rojas de Arias, B. Figadère,
- 1084 A. Fournet, H. Chneweiss, E. Poupon, Alkaloids from Rutaceae: activities of canthin-6-one.
 1085 alkaloids and synthetic analogues on glioblastoma stems cells. Med. Chem. Commun. 3 (2012)
- 1086 771, https://doi.org/10.1039/c2md20047c.

- [150] V. De Feo, L. De Martino, E. Quaranta, C. Pizza, Isolation of phytotoxic compounds from Treeof-Heaven (*Ailanthus altissima* Swingle). J. Agric. Food Chem. 51 (2003) 1177–1180,
 https://doi.org/10.1021/jf020686+.
- [151] V. De Feo, L. de Martino, A. Santoro, A. Leone, C. Pizza, S. Franceschelli, M. Pascale,
 Antiproliferative effects of tree-of-heaven (*Ailanthus altissima* Swingle). Phytother. Res. 19
 (2005) 226–230, https://doi.org/10.1002/ptr.1670.
- [152] M. Ammirante, R. Di Giacomo, L. De Martino, A. Rosati, M. Festa, A. Gentilella, M. C.
 Pascale, M.A. Belisario, A. Leone, M.C. Turco, V. De Feo, 1-methoxy-canthin-6-one induces c Jun NH 2 -terminal kinase-dependent apoptosis and synergizes with tumor necrosis factor-related
 apoptosis-inducing ligand activity in human neoplastic cells of hematopoietic or endodermal
 origin. Cancer Res. 66 (2006) 4385–4393, https://doi.org/10.1158/0008-5472.CAN-05-3895.
- [153] C.W. Lindsley, M.J. Bogusky, W.H. Leister, R.T. McClain, R.G. Robinson, S.F. Barnett, D.
 Defeo-Jones, C.W. RossIII, G.D. Hartman, Synthesis and biological evaluation of unnatural
 canthine alkaloids. Tetrahedron Lett. 46 (2005) 2779–2782,
 https://doi.org/10.1016/j.tetlet.2005.02.143.
- [154] N. Samat, M. Fong Nga, H.M. Leea, S.K. Ling, P.J. Tana, V. Patel, Canthin-6-one isolated from
 Brucea javanica root blocks cancer cells in the G₂ /M phase and synergizes with cisplatin. Nat.
 Prod. Comm.12 (2017) 771-778, https://doi.org/10.1177/1934578X1701200522.
- [155] A. Doménech-Carbó, R. Rodrigo, J.D. Maréchal, E. Poupon, A. Fournet, B. Figadère, G. 1105 1106 Cebrian-Torrejon, Bioelectrochemical monitoring of soluble guanylate cyclase inhibition by the 1107 natural β-carboline canthin-6-one. J. Mol. Struct. 1134 (2017),661-667, https://doi.org/10.1016/j.molstruc.2016.12.016. 1108
- [156] H.F. Vieira Torquato, A.C. Ribeiro-Filho, M.V. Buri, R.T. Araujo Jr, R. Pimenta, J.S.D.
 Oliveira, V.C. Filho, A. Macho, E.J. Paredes-Gamero, D.T. Oliveira Martin, Canthin-6-one
 induces cell death, cell cycle arrest and differentiation in human myeloid leukemia cells.
 Biochimica et Biophysica Acta (BBA) General Subjects 1861 (2017) 958–967,
 https://doi.org/10.1016/j.bbagen.2017.01.033.
- [157] G. Cebrián-Torrejón, A. Doménech-Carbó, M.T. Scotti, A. Fournet, B. Figadère, E. Poupon,
 Experimental and theoretical study of possible correlation between the electrochemistry of

- canthin-6-one and the anti-proliferative activity against human cancer stem cells. J. Mol. Structure
 1102 (2015) 242–246, https://doi.org/10.1016/j.molstruc.2015.08.042.
- 1118 [158] A. Doménech-Carbó, G. Cebrián-Torrejón, L. de Miguel, V. Tordera, D. Rodrigues-Furtado, S.
- 1120 quadruplex DNA, and nucleosomal DNA electrochemical screening using canthin-6-one alkaloid-

Assad-Kahn, A. Fournet, B. Figadère, R. Vázquez-Manrique, E. Poupon, dsDNA, ssDNA, G-

- 1121modifiedelectrodes.ElectrochimicaActa115(2014)546–552,1122https://doi.org/10.1016/j.electacta.2013.11.025.
- [159] D.A. Ostrov, J.A. Hernandez Prada, P.E. Corsino, K.A. Finton, N. Le, T.C. Rowe, Discovery of
 novel DNA gyrase inhibitors by high-throughput virtual screening. Antimicrob. Agents
 Chemother. 51 (2007) 3688–3698, https://doi.org/10.1128/AAC.00392-07.
- [160] T. Tremmel, A. Puzik, A.P. Gehring, F. Bracher, Canthin-4-ones as novel antibacterial agents.
 Arch. Pharm. Chem. Life Sci. 349 (2016) 710–723, https://doi.org/10.1002/ardp.201600137.
- [161] J.K. Dai, W.J. Dan, N. Li, H.T. Du, J.W. Zhang, J.R. Wang, Synthesis, in vitro antibacterial
 activities of a series of 3- N -substituted canthin-6-ones. Bioorg. Med. Chem. Lett. 26 (2016) 580–
 583, https://doi.org/10.1016/j.bmcl.2015.11.070.
- [162] N. Li, D. Liu, J.K. Dai, J.Y. Wang, J.R. Wang, Synthesis and In Vitro Antibacterial Activity of
 Quaternized 10-Methoxycanthin-6-one Derivatives. Molecules 24 (2019), 1553,
 https://doi.org/10.3390/molecules24081553.
- [163] J. Dai, W. Dan, N. Li, R.X. Wang, Y. Zhang, N.N. Li, R.Z. Wang, J.R. Wang, Synthesis and
 antibacterial activity of C2 or C5 modified and D ring rejiggered canthin-6-one analogues. Food
 Chem. 253 (2018) 211–220, https://doi.org/10.1016/j.foodchem.2018.01.166.
- [164] J. Dai, W. Dan, Y. Zhang, M. He, J. Wang, Design and synthesis of C10 modified and ringtruncated canthin-6-one analogues as effective membrane-active antibacterial agents. Bioorg. Med.
 Chem. Lett. 28 (2018) 3123–3128, https://doi.org/10.1016/j.bmcl.2018.06.001.
- [165] T. Ohomoto, Y. I. Sung, K. Koike, T. Nikaido, Effect of alkaloids of Simaroubaceous plants on
 the local blood flow rate. Jap. J. Pharmacognosy 39 (1985) 28–34.
- 1142 [166] J. Chen, X. H. Yan, J.H. Dong, P. Sang, X. Fang, Y.T. Di, Z.K. Zhang, X.J. Hao, Tobacco
- 1143 mosaic virus (TMV) inhibitors from *Picrasma quassioides* Benn. J. Agric. Food Chem. 57 (2009)
- 1144 6590–6595, https://doi.org/10.1021/jf901632j.

- 1145 [167] D. Verma, D. Mitra, M. Paul, P. Chaudhary, A. Kamboj, H. Thatoi, P. Janmeda, D. Jain, P.
- 1146 Panneerselvam, R. Shrivastav, K. Pant, P. K. D. Mohapatra, Potential inhibitors of SARS-CoV-
- 2 (COVID 19) proteases PLpro and Mpro/ 3CLpro: molecular docking and simulation studies of
 three pertinent medicinal plant natural components. Current Research in Pharmacology and Drug
 Discovery 2 (2021), 100038, https://doi.org/10.1016/j.crphar.2021.100038.
- [168] M.E. Ferreira, H. Nakayama, A. Rojas de Arias, A. Schinini, N. Vera de Bilbao, E. Serna, S.
 Torres de Ortiz, L. Sanabria, D. Lagoutte, F. Soriano-Agatón, E. Poupon, R. Hocquemiller, A.
 Fournet, Effects of canthin-6-one alkaloids from *Zanthoxylum chiloperone* on *Trypanosoma cruzi*-
- 1153 infected mice. J. Ethnopharmacol. 109 (2007) 258–263, https://doi.org/10.1016/j.jep.2006.07.028.
- [169] J.L. Arias, G. Cebrián-Torrejón, E. Poupon, A. Fournet, P. Couvreur, Biodegradable polymeric
 nanoformulation based on the antiprotozoal canthin-6-one. J. Nanopart. Res. 13 (2011) 6737–
 6746, https://doi.org/10.1007/s11051-011-0580-z.
- [170] T. Ohmoto, T. Nikaido, K. Koike, K. Kohda, U. Sankawa, Inhibition of adenosine 3',5'-cyclic
 monophosphate phosphodiesterase by alkaloids. II. Chem. Pharm. Bull. 36 (1888) 4588–4592,
 https://doi.org/10.1248/cpb.36.4588.
- [171] Y. Sung, K. Koike, T. Nikaido, T. Ohmoto, U. Sankawa, Inhibitors of cyclic AMP
 phosphodiesterase in *Picrasma quassioides* Bennet, and inhibitory activities of related .BETA.carboline alkaloids. Chem. Pharm. Bull. 32 (1984) 1872–1877,
 https://doi.org/10.1248/cpb.32.1872.
- [172] D.H. Bray, P. Boardman, M.J. O'Neill, K.L. Chan, J.D. Phillipson, D.C. Warhurst, M. Suffness, 1164 1165 Plants as a source of antimalarial drugs 5. Activities of Ailanthus altissima stem constituents and 1166 of Phytother. (1987)22 - 24. some related quassinoids. Res. 1 1167 https://doi.org/10.1002/ptr.2650010105.
- [173] K. Chan, M. O'Neill, J.D. Phillipson, D. Warhurst, Plants as Sources of Antimalarial Drugs. Part
 3¹ Eurycoma longifolia. Planta Med. 52 (1986) 105–107, https://doi.org/10.1055/s-2007-969091.
- [174] A. Keene, A. Harris, J.D. Phillipson, D. Warhurst, *In Vitro* amoebicidal testing of natural
 products; Part I. methodology. Planta Med. 52 (1986) 278–285.

- [175] G.H.N. Towers, Z. Abramowski, UV-mediated genotoxicity of furanoquinoline and of certain
 tryptophan-derived alkaloids. J. Nat. Prod. 46 (1983) 576–581, https://doi.org/10.1055/s-2007969152.
- [176] E.S. de Souza Almeida, V.C. Filho, B.K. Clasen, S.O. Balogun, D. Tabajarade, O. Martins,
 Pharmacological mechanisms underlying the anti-ulcer activity of methanol extract and canthin-6one of *Simaba ferruginea* A. St-Hil. in animal models. J. Ethnopharmacol. 134 (2011) 630–636,
- 1178 https://doi.org/10.1016/j.jep.2011.01.009.
- 1179 [177] N.N. Yuan, C.Z. Cai, M.Y. Wu, Q. Zhu, H.X. Su, M. Li, J.Y. Ren, J.Q. Tan, J.H. Lu, Canthin-6-
- 1180 one accelerates alpha-synuclein degradation by enhancing UPS activity: Drug target identification
- by CRISPR-cas9 whole genome-wide screening technology. Front. Pharmacol. 10 (2019), 16,
 https://doi.org/10.3389/fphar.2019.00016.
- [178] E. Guo, Y.W. Hu, T. Du, H.L. Zhu, L. Chen, W. Qu, J. Zhang, N. Xie, W.Y. Liu, F. Feng, J. Xu,
 Effects of *Picrasma quassioides* and its active constituents on Alzheimer's disease in vitro and in
 vivo. Bioorg. Chem. 92 (2019) 103258, https://doi.org/10.1016/j.bioorg.2019.103258.
- [179] T. Sasaki, W. Li, T. Ohmoto, K. Koike, Evaluation of canthinone alkaloids as cerebral protective
 agents. Bioor. Med. Chem. Lett. 26 (2016) 4992–4995,
 https://doi.org/10.1016/j.bmcl.2016.09.006.
- [180] P. Liu, H. Li, R. Luan, G. Huang, Y. Liu, M. Wang, Q. Chao, L. Wang, D. Li, H. Fan, D. Chen,
 L. Li, K. Matsuzaki, W. Li, K. Koike, F. Zhao, Identification of β-carboline and canthinone
 alkaloids as anti-inflammatory agents but with different inhibitory profile on the expression of
 iNOS and COX-2 in lipopolysaccharide-activated RAW 264.7 macrophages. J. Nat. Med. 73
 (2019), 124–130, https://doi.org/10.1007/s11418-018-1251-5.
- [181] T. Sasaki, W. Li, K. Higai, K. Koike, Canthinone alkaloids are novel protein tyrosine
 phosphatase 1B inhibitors. Bioorg. Med. Chem. Lett. 25 (2015) 1979–1981,
 https://doi.org/10.1016/j.bmcl.2015.03.014.
- 1197 [182] H. Yokoo, H. Ohsaki, H. Kagechika, T. Hirano, Structural development of canthin-5,6-dione
 1198 moiety as a fluorescent dye and its application to novel fluorescent sensors. Tetrahedron 72 (2016)
- 1199 5872–5879, https://doi.org/10.1016/j.tet.2016.08.014.

- [183] S.K. Cho, M. Jeong, D. Jang, J.H. Choi, Anti-inflammatory effects of canthin-6-one alkaloids
 from *Ailanthus altissima*. Planta Med. 84 (2018) 527–535, https://doi.org/10.1055/s-0043-123349.
- 1202 [184] R. Parikrama, R.R. Esyanti, Effect of UV elicitation on callus growth, alkaloid and terpenoid
- 1203 contents in *Eurycoma longifolia* Jack. IJACEBS 1,
- 1204 https://doi.org/10.15242/IJACEBS.C1113054.

(2014),