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1	How environment and genetic
2	architecture of unreduced gametes
3	shape the establishment of
4	autopolyploids
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25 Abstract: It is broadly assumed that polyploidy success is due to an increase in fitness 26 associated with whole genome duplication due to higher tolerance to stressful conditions. In 27 agreement, several theoretical models found that, among other factors, a better tolerance to 28 new environmental conditions can promote polyploidy establishment. These models, 29 however, often made strong hypotheses, for example considering that diploids cannot adapt 30 to new conditions, or that unreduced gametes production is not a limiting factor and that it is 31 of a fixed quantity. In this paper, we challenged some of these hypotheses. We developed a 32 theoretical model in which we modeled the joint evolution of a quantitative trait under 33 selection and the production of unreduced gametes, this trait also being a quantitative trait; 34 both traits were pleiotropically linked. We followed the adaptation of initially diploid 35 populations to a new environment to which neo-tetraploid individuals were directly adapted. 36 The generation of these autotetraploid individuals was enabled by the genetic production of 37 unreduced gametes and by the environmental change modifying the average production of 38 these gametes. We found that for realistic values of unreduced gametes production, 39 adaptation to new environmental conditions was mainly achieved through adaptation of 40 diploids to the new optimum rather than the fixation of newly adapted tetraploid individuals. 41 In broader parameter sets, we found that the adaptation process led to mixed-ploidy 42 populations, except when the populations were swamped with unreduced gametes, and that 43 pleiotropy and environmental effects favored the co-existence of both cytotypes.

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Keywords: unreduced gametes, pleiotropy, autopolyploidy, neo-polyploidy, quantitative
genetics, environment.

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53 Introduction:

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55 Polyploids are organisms known for having more than two chromosome sets compared with 56 their diploid congener. Polyploidy resulting from whole genome duplication (WGD) is 57 frequently accompanied by meiotic abnormalities and alters gene dosage, consequently 58 affecting the phenotypes and long-term evolution of populations (Otto 2007). There is 59 evidence that polyploidization is an important mechanism in speciation and diversification 60 (Soltis et al. 2015), with blooms of diversity associated with ancient WGD events, both in 61 plants, particularly the angiosperms, animals and fungi (Jiao et al. 2011; Van De Peer et al. 62 2017). For example, it has been suggested that all angiosperms descended from an ancient 63 WGD event (Jiao et al., 2011; Amborella Genome Project, 2013), and 25 to 35% of 64 angiosperm species have been estimated to be recently formed polyploid (i.e. neo-polyploid) 65 species (Wood et al., 2009; Mayrose et al., 2011; Barker et al., 2016a). The geographic 66 distribution of neo-polyploids in the angiosperms shows a latitudinal trend, with a higher 67 frequency of polyploids at higher latitudes (Rice et al. 2019). It is an ongoing debate if 68 polyploidy fosters adaptability to stressful environments (Madlung 2013), and if in particular 69 neo-polyploidy is associated with stressful conditions, such as cold and dry conditions, as 70 reviewed by (Van De Peer *et al.* 2021). This environmental association becomes particularly 71 intriguing when considering the mechanisms leading to the formation of neo-polyploids.

Although somatic doubling could potentially play a role in the formation of neopolyploids, it is currently assumed that it usually involves unreduced gametes (UG), which are gametes with the somatic ploidy level, such as diploid gametes in a diploid organism; for the cytological mechanisms and molecular regulatory networks underlying UG formation see (Brownfield & Kohler 2011; De Storme & Geelen 2013). Pathways to polyploidy can be 'one-step' (fusion of two unreduced gametes; rather rare because of the rarity of unreduced gamete formation), 'triploid bridge' (fusion of unreduced and reduced gamete within a diploid population, with triploid offspring producing unreduced gametes that backcross with the reduced gamete parent), and 'hybrid bridge' (same as triploid bridge, only that it refers to interspecific crosses; rather common, as unreduced gamete formation is higher in hybrids) (Mason & Pires 2015). Unreduced gamete formation has been shown to depend on abiotic and biotic factors, such as temperature, moisture, nutrition and herbivory (Kreiner *et al.* 2017a; Mason & Pires 2015; Ramsey & Schemske 1998a).

85 Once polyploids are formed, they face numerous challenges to finally establish in a 86 population, and to form a distinct polyploid lineage that may eventually speciate from its 87 diploid progenitor. Upon emergence, polyploids tend to experience a lack of mating 88 opportunities (minority cytotype exclusion; Levin 1975). The lack of mating opportunities in 89 short-lived neo-polyploid organisms can be theoretically overcome by a strong fitness 90 advantage of neo-polyploids (Husband, 2004), polyploidy-associated mating system 91 transitions (Griswold 2021; Oswald & Nuismer 2011), higher clonal reproduction of 92 polyploids (Van Drunen & Friedman 2022), but also certain spatial components of neo-93 polyploid establishment (Kauai et al. 2023). Unreduced gametes production per se is assumed 94 to come with an immediate fitness disadvantage, due to meiotic mishap and low triploid 95 fitness (Kreiner et al. 2017), but also as a result of the genetic architecture of UG formation. 96 In the formation of unreduced gametes, several genes have been found to be involved 97 (Bretagnolle & Thompson 1995; Brownfield & Kohler 2011; De Storme & Geelen 2013), for 98 which pleiotropic effects (i.e. the fact that several traits are affected by a single locus) can be 99 assumed. Under the assumption of a correlated production of male and female unreduced 100 gametes, polyploid fixation is more likely in combination with mating system transitions, as 101 selfing of an individual that produces unreduced gametes of both sexes will likely result in 102 polyploid progeny. For flowering plants, self-fertilization is indeed assumed to be more

103 common in polyploids compared to diploids (Barringer 2007), although this study did not 104 discriminate between autopolyploids (i.e. multiplied chromosome set within a species) and 105 allopolyploids (i.e. multiplied chromosome set from two species), which does not allow to 106 distinguish between the effect of WGD and hybridization. Later, Husband et al. (2008) found 107 that autopolyploids are predominantly outcrossing, while allopolyploids are largely selfing. 108 However, all above mentioned theoretical considerations strongly depended on key 109 assumptions. The first one is that newly formed polyploid individuals are at least as fit as 110 their diploid progenitors (Husband 2004; Kauai et al. 2023; Oswald & Nuismer 2011; Van 111 Drunen & Friedman 2022). However, neo-polyploid lineages are generally less fit than their 112 diploid progenitors (at least in controlled conditions, see Porturas et al., 2019; Clo and Kolář, 113 2021 for synthetic neo-auto/allopolyploids). Considering such an initial fitness cost strongly 114 affects the theoretical outcome, such as the evolution toward self-fertilization being 115 disfavored in these conditions (Clo et al. 2022; Gaynor et al. 2023). In the longer term, 116 however, autopolyploids may exhibit a fitness advantage over their diploid progenitors (Clo, 117 2022a), especially in scenarios of environmental change (Van De Peer et al. 2021b).

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119 The second strong assumption is that UG production is high enough for the 120 production of neo-polyploid lineages not to be a limiting factor in polyploid evolution (Baack 121 2005; Husband 2004; Li et al. 2004; Van Drunen & Friedman 2021). However, the 122 production of unreduced gametes is generally much lower in natural populations of non-123 hybrid species (between 0.1 to 2%; Kreiner et al., 2017) compared to what is found in 124 theoretical models (Felber 1991; Husband 2004; Oswald & Nuismer 2011; Van Drunen & 125 Friedman 2022). Few models attempted to model the effect and evolution of UG production 126 on the probability of polyploid evolution (Clo et al. 2022; Gerstner et al. 2024). Clo et al. 127 (2022) treated the evolution of unreduced gametes as a quantitative trait and found that under

128 realistic conditions high enough levels of UG production for autopolyploidy to invade are 129 only reached when genetic drift is strong. Gerstner et al. (2024) modeled the inter-generation 130 variation of unreduced gametes following the empirical data by Kreiner et al. (2017b) and 131 found that such variation can lead to the evolution of autopolyploidy in initially diploid 132 populations. While both models are informative, they are also built on strong assumptions. 133 Clo et al. (2022) made the genetic architecture of UG production too simplistic, omitting, for 134 example, the fact that mutations increasing UG production are pleiotropically linked to 135 fitness and strongly affect fitness components such as pollen and/or ovule production 136 (d'Erfurth et al. 2010; Erilova et al. 2009; Ravi et al. 2008; Wang et al. 2010; see also 137 Brownfield & Kohler 2011 for a review). They also neglected the effects of the environment, 138 while such factors, like temperature, are known to have major effects on UG production rates 139 (De Storme et al. 2012; Mason et al. 2011; Pecrix et al. 2011; Schindfessel et al. 2023; Wang 140 et al. 2024). On the opposite, Gerstner et al. (2024) made the hypothesis that the variation in 141 UG production is only environmental, while it is known that this trait is determined by both 142 environmental and genetic factors (Parrott & Smith 1986; Tavoletti et al. 1991a). Considering 143 a more realistic genetic architecture of UG production could strongly impact the conclusions 144 of the above-mentioned models, and could make us understand the relative contribution of 145 both genetic and environmental effects in UG production and polyploidy evolution.

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In this paper, we investigate the effect of the genetic architecture of UG production in the origin and establishment of autopolyploids in both stable and disturbed environments, by considering both environmental stochasticity and the pleiotropic effect of mutations affecting UG production on fitness. We build an individual-based simulation model extended from Clo *et al.* (2022), in which we test the following parameters: environmental factor and pleiotropy as effects of studying how unreduced gametes as a quantitative trait evolve and affect the origin and establishment of polyploids. In conclusion, we found that for realistic values of UG production, adaptation to new environmental conditions was mainly realized by adaptation of diploids to the new optimum rather than the fixation of newly adapted tetraploid individuals. In broader parameter sets, we found that the adaptation process led to mixedploidy populations, except when the populations were swamped with unreduced gametes, and that pleiotropy and environmental effects favored the co-existence of both cytotypes.

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161 Material and methods:

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163 General assumptions

We considered the joint evolution of a quantitative trait Z and the production of unreduced gametes (UG) in a population of constant size N, initially made of diploid individuals reproducing through obligate random mating. The phenotypic value z of an individual was determined by the additive action of L_Z loci each with an infinite possible number of alleles and was given by

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$$z = g_z + e_z, (1)$$

where g_z was the genetic component of the individual's phenotype and was given by $g_{z-2x} = \sum_{j}^{L} (g_j^M + g_j^P)$, where g_j^M (respectively g_j^P) was the additive allelic effect at locus *j* inherited from the maternal (respectively paternal) gamete in the diploid population. After polyploidization and with tetrasomic inheritance, the genetic component became $g_{4x} = \sum_{j}^{L} d (g_j^{M1} + g_j^{M2} + g_j^{P1} + g_j^{P2})$, where g_j^{M1} and g_j^{M2} (respectively g_j^{P1} and g_j^{P2}) were the additive allelic effects at locus *j* inherited from the maternal (respectively paternal) gametes. The parameter *d* controlled for the dosage effect and determined the effect of 177 polyploidization on the tetraploid genotypic values compared to diploid ones. The additive 178 value of a new mutant allele was drawn from a Gaussian distribution of mean 0 and variance 179 a^2 . The random environmental effect on the phenotypic trait e_z was drawn from a Gaussian 180 distribution of mean 0 and variance V_E and was considered to be independent of the genetic 181 components of fitness.

182 The trait underwent stabilizing selection around an arbitrary optimal phenotypic value, 183 denoted Z_{opt} and being equal to zero in this model. The fitness value W_Z of an individual with 184 phenotype *z* was thus described by the Gaussian function:

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$$W_Z = e^{-\delta^2/2\omega^2}, (2)$$

186 where δ was the distance between the individual's phenotype *z* and the optimum trait value 187 and ω^2 was the width of the fitness function, representing the strength of selection. There was 188 no dominance at the phenotypic scale in this model, but recessivity of mutations arose 189 naturally at the fitness scale due to the non-linearity of the phenotype-fitness function (see 190 Manna *et al.* 2011 for diploids and Clo, 2022 for tetraploids).

191 The trait "unreduced gametes production" was coded for both male and female 192 reproductive functions by $L_{G-M} = L_{G-F}$ freely recombining bi-allelic loci. The average UG 193 production for male and female functions p_{UG-M} and p_{UG-F} of an individual was determined by 194 the additive action of alleles at different loci each with an infinite possible number of alleles 195 and was given by

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$$p_{\text{UG-M}} = g_{\text{UG-M}} + e_{\text{UG}} (3.1)$$

197 and

198 $p_{\text{UG-F}} = g_{\text{UG-F}} + e_{\text{UG}} (3.2)$

199 where g_{UG-M} (respectively g_{UG-F}) was the genetic component of the individual's male (respectively female) UG production and was given by $g_{\text{UG-M-2x}} = \sum_{j=1}^{L} (g_j^{UG-M-M} +$ 200 g_j^{UG-M-P}), (respectively $g_{UG-F-2x} = \sum_j^L (g_j^{UG-F-M} + g_j^{UG-F-P}))$, where g^{UG-M-M}_j 201 (respectively $g^{\text{UG-M-P}}_{j}$) was the additive allelic effect at locus j inherited from the maternal 202 203 (respectively paternal) gamete in the diploid population. After polyploidization and with 204 tetrasomic inheritance, the genetic component became (for male) $g_{UG-M-4x} =$ $\sum_{j}^{L} (g_{j}^{UG-M-M1} + g_{j}^{UG-M-M2} + g_{j}^{UG-M-P1} + g_{j}^{UG-M-P2}), \text{ where } g^{UG-M-M1}_{j} \text{ and } g^{UG-M-M2}_{j}$ 205 (respectively $g^{\text{UG-M-PI}}_{j}$ and $g^{\text{UG-M-P2}}_{j}$) were the additive allelic effects at locus j inherited from 206 207 the maternal (respectively paternal) gametes. The same applied for female (un)reduced 208 gametes production. The additive value of a new mutant allele was drawn from a Gaussian distribution of mean 0 and variance a^2 . The environmental effect e_{UG} was fixed during 209 210 simulations but could vary between the two steps of the simulation process. If by chance p_{UG} 211 _M and/or $p_{\text{UG-F}}$ were < 0 (respectively > 1), then $p_{\text{UG-M}}$ and/or $p_{\text{UG-F}}$ were forced to be equal to 212 0 (respectively 1). In this model, we simulated three levels of pleiotropy: 1) all the different 213 loci of the different traits can be independent, or 2) pleiotropy between loci involved in UG 214 production for male and female functions can happen, or 3) pleiotropy between all kind of 215 loci (phenotypic trait and unreduced gametes for male and female functions) can occur.

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218 Simulation process:

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220 The simulations took place in two steps: a first one of stabilizing selection, and a 221 second one of directional selection (adaptation process). For this step, we initiated each

222 simulation from a population of genetically identical diploid individuals that are homozygous 223 for the ancestral allele at all loci ($z = 0 \& p_{UG-M} = p_{UG-F} = 0$ at t = 0). The life cycle can be 224 summarized in five successive events. First, there is a fitness-dependent choice of the first 225 parent (selection), followed by the choice of the second parent, which is also fitness-226 dependent. Selection takes place as follows: an individual is sampled randomly among the N227 individuals, but its sampling probability is weighted by its fitness. Once the two parents are 228 chosen, the type of gametes (reduced or unreduced) they will produce is chosen. For each 229 reproducer, one number is sampled in a uniform distribution between 0 and 1. If $p_{\rm UG}$ (male or 230 female) is higher than the sampled value, the reproducer generates an unreduced gamete. For 231 reduced (n) gametes (n = x and n = 2x gametes for diploid and tetraploid individuals, 232 respectively), one allele per locus of each sister chromatid is sampled randomly for each trait. 233 This phase is then followed by the introduction of mutations for each trait, the number of 234 which is sampled from a Poisson distribution with parameter U, the haploid genomic 235 mutation rate (with $U = \mu L$, where μ is the per-locus mutation rate and L is the number of loci 236 underlying the trait under study). If the resulting offspring is not diploid or tetraploid, it will 237 not contribute to the next generation (i.e. triploid individuals are considered inviable or 238 sterile). The reproduction phase stops when N viable offspring are formed. The first step 239 stops when the population is at Mutation-Selection-Drift (M-S-D) equilibrium. The 240 population is considered to be at M-S-D equilibrium when the average population fitness 241 value calculated over the last thousand generations does not differ by more than one percent 242 from the mean fitness calculated over the previous thousand generations.

The second step of the simulations began once the population reached M-S-D equilibrium, by introducing an environmental change (done by changing the phenotypic optimum Z_{opt} from 0 to 2.5). After the environmental change, we assumed the polyploids are adapted to their new conditions (while before the change the fitness of each cytotype is

247 approximately the same) and have a fitness advantage over diploids. In addition, the 248 environmental change can induce or not a change in UG production (i.e. there is a positive 249 non-zero effect of the environment on UG production). The population adapted either by (i) 250 making diploid individuals to adapt to the new conditions thanks to standing genetic diversity 251 and/or *de novo* mutations, (ii) increasing the number of newly adapted tetraploid individuals 252 thanks to genetic diversity for unreduced gametes and the potential effect of the environment, 253 or (iii) a mix of the two solutions (mixed-ploidy populations). The population evolved during 254 500 generations after the change, and the frequency and variance of unreduced gametes, the 255 average fitness of the population, and the frequency of tetraploids in the population were 256 inferred at each generation.

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258 Parameter sets:

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260 Simulations were run for several parameter sets. The values chosen for the mutation rate U261 range from 0.005 to 0.1, reflecting the per-trait haploid genomic mutation rate found in the 262 literature (Halligan & Keightley 2009). We used parameter set values similar to those 263 explored in similar theoretical models (Bürger et al. 1989; Ronce et al. 2009; Oswald & 264 Nuismer 2011), with the number of freely recombining loci under selection L = 100, and $a^2 =$ 265 0.05, var(e_Z) = var(e_{UG}) = 1. The strength of stabilizing selection is set to $\omega^2 = 9$ to fit 266 empirical observations (Clo & Opedal 2021). We considered a population size of N = 200267 individuals. The dosage parameter d is equal to 0.65 ($g_{Z-4x} = 1.3*g_{Z-2x}$, which is found in neo-268 polyploid and established polyploid populations on average; see Porturas et al., 2019 and Clo 269 & Kolář, 2021 for meta-analyses). When pleiotropy is considered, the number of pleiotropic 270 loci can be equal to 0 (no pleiotropy), 40 (moderate pleiotropy), or 80 (high pleiotropy). As

271 mentioned earlier, before the environmental change, the fitness of neo-polyploids was close 272 to the one of diploids (slightly lower because of the gigas phenotypic effect) but was initially 273 higher at the beginning of the environmental change. Before the change, e_{UG} equals 0 but can 274 be equal to 0, 0.1, or 0.3 after the change when considering that the environmental stress 275 (such as change in temperature) also affects UG production. Finally, Z_{opt} switched from 0 to 276 2.5 after the environmental change. A summary of the parameters used and their values can 277 be found in Table 1.

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Table 1. Description of the model parameters, their abbreviations, and tested values insimulations.

Parameter	Abbreviation	Value(s)
Population size	N	200
Number of loci	L	100
Haploid genomic mutation rate	U	0.005, 0.05 or 0.1
Variance of mutational effects	a^2	0.05
Dosage effect	d	0.65
$V_{\rm E}$ of environmental effects	$V_{ m E}$	1
Strength of stabilizing selection	ω^2	9

284 **Results**:

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286 **Description of populations at Mutation-Selection-Drift equilibrium:**

288 Under stable environmental conditions (i.e. $Z_{opt} = 0$), the main factors affecting population 289 metrics were pleiotropy and the mutation rate (Figure 1; Supplementary Figures 1 and 2). 290 Under a fixed mutation rate (U = 0.005; see below for details), pleiotropy on the quantitative 291 trait and on UG production generally led to more efficient purging of deleterious mutations 292 (both on the quantitative trait under stabilizing selection and on mutations increasing UG 293 production), resulting in lower UG production (median 1% with pleiotropy vs. median 2% 294 without pleiotropy; Figure 1A), lower genetic diversity for UG production (median 0.18% 295 with pleiotropy vs. median 0.30% without pleiotropy; Figure 1B) and slightly increased population fitness (median 94.4% with pleiotropy vs. median 94.2% without pleiotropy; 296 297 Figure 1C). Higher mutation rates (U = 0.05 and 0.1) did not change the patterns 298 qualitatively, but generally led to higher average UG production (median 18% without 299 pleiotropy for U = 0.05 vs. median 35% for U = 0.1; Supplementary Figures 1 and 2), higher 300 genetic variance of unreduced gametes (median 18% without pleiotropy for U = 0.05 vs. 301 median 36% for U = 0.1; Supplementary Figures 1 and 2) and lower population fitness 302 (median 91% without pleiotropy for U = 0.05 vs. median 85% for U = 0.1; Supplementary 303 Figures 1 and 2). Under low mutation rates, we did not find mixed-ploidy populations at the 304 M-S-D equilibrium (0% of tetraploids, the populations were purely diploid; Figure 1D), while 305 intermediate and high mutation rates can lead to stable mixed-ploidy populations (between 1 306 to 10% of tetraploids, the higher when no pleiotropic effects were modeled; Supplementary 307 Figures 1 and 2). Because the simulations with low mutation rates gave the most realistic 308 pattern of UG production for empirical rates of UG production (1-2%), we described 309 separately the evolution of polyploidy with this biologically realistic scenario in the first time 310 and other scenarios in the second time.



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314 Figure 1: Effect of pleiotropy type (no, all traits, unreduced gametes; number of pleiotropic loci = 80) on 315 unreduced gametes production and its variance, population fitness and tetraploid fixation, simulated from a 316 population of genetically identical diploid individuals under the mutation rate U = 0.005. Pleiotropy on all traits 317 means that pleiotropy affected both the quantitative trait under directional selection and unreduced gametes 318 production. (A) Proportion of unreduced gametes production in diploid individuals; (B) variance of unreduced 319 gametes production in diploid individuals; (C) population fitness; (D) frequency of tetraploids in the population. 320

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322 The evolution of polyploidy under environmental change for realistic parameter sets

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324 Here we focused on simulations with low mutation rates (U = 0.005) and realistic 325 environmental conditions (directional selection, Z_{opt} shifting from 0 to 2.5, and an 326 environmental effect on UG production of 10%). It was observed that environmental 327 conditions did not significantly alter the population cytotype structure and fitness after the 328 directional selection episode (Figure 2). Both UG production and variance remained similar 329 (UG production: median 2% with(out) environmental effect, no pleiotropy; Figure 2A; UG

330 variance: median 0,3% with(out) environmental effect, no pleiotropy; Figure 2B). Mean 331 population fitness also remained comparable after 500 generations when pleiotropy is absent (median 94%; Figure 2C), but with both environmental effect and pleiotropy population 332 333 fitness decreased substantially, the highest being when pleiotropy only affected UG 334 production (median 85%; Figure 2C). At the end of the simulations, the populations were 335 only made of diploid individuals (Figure 2D), suggesting that the recovery of fitness is made 336 by diploid individuals adapting to new conditions rather than the invasion of newly adapted 337 tetraploids. Environmental effects on UG production rarely led to mixed-ploidy populations 338 (Figure 2D). We attributed this to a conflict between environmental effect and new mutations 339 that could have favored the selection of newly adapted tetraploids (but that had a negative 340 pleiotropic effect on the trait under directional selection) to the detriment of the evolution of 341 diploid individuals to the new optimum (Figure 3B; see Figure 3A for a comparison without 342 environmental effect). Indeed, one can see that the production of unreduced gametes slightly 343 increased at the beginning of the environmental change (generally doubling in the first 20 344 generations, shifting for example from 2% to 4% when no pleiotropy is modeled; Figure 3B). 345 The production then decreased during the 500 generations of adaptation (Figure 3B).

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Figure 2: Effect of pleiotropy type (no, all traits, unreduced gametes; number of pleiotropic loci = 80) and environmental factor (0.1) on the evolution of polyploidy after 500 generations of directional selection, simulated from a population of genetically identical diploid individuals under the mutation rate U = 0.005. Pleiotropy on all traits means that pleiotropy affected both the quantitative trait under directional selection and unreduced gametes production. (A) Proportion of unreduced gametes production in diploid individuals; (B) variance of unreduced gametes production in diploid individuals; (C) population fitness; (D) frequency of tetraploids in the population.





367 The general effects of environment and genetic architecture on the evolution of 368 polyploidy:

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370 At the beginning of the directional selection episode, the frequency of unreduced gametes, 371 due to genetic effects, increased in the first generations and then decreased back to an 372 equilibrium value (Figures 3B and 4). This steep initial increase was due to the fact that at the 373 beginning of the adaptation process, selection favored both the formation and the increase in 374 frequency of newly adapted tetraploid individuals. Once diploid individuals were closer to 375 the new phenotypic optimum, formation of tetraploids was either selected against and 376 tetraploids remained in the minority within the population due to minority cytotype exclusion 377 (between 0 to 10% of tetraploids when no environmental effect was modeled; Figure 5A; 378 between 0 to 55% of tetraploids with a realistic environmental effect of 0.1 on UG 379 production; Figure 5B), or tetraploid formation was continuously favored until fixation of 380 tetraploidy (100% of tetraploids when the effect of the environment on UG production is 381 strong, independent of the mutation rate; Figure 5C). As previously described, the overall 382 effect of increased mutation rates was to increase both the average production of unreduced 383 gametes and of tetraploids within populations (Figures 4 and 5). It was, however, notable that 384 the mutation rate has a synergistic effect with the environmental effect on unreduced 385 gametes: the higher the environmental effect, the higher the genetic response to increase UG 386 production (Figure 4). This is because the production of unreduced gametes is under 387 frequency dependence: the higher UG production is in the population, the less it is counter-388 selected. Thus, the stronger the effect of the environment, the easier it is for a mutation 389 increasing UG production to be maintained in the population. Pleiotropy had a generally 390 negative effect by decreasing the frequency of unreduced gametes and tetraploids (Figures 4 391 and 5). Interestingly, the effect of pleiotropy on limiting both UG production and the 392 frequency of tetraploids was stronger when pleiotropy was limited to UG production of both

393 male and female functions, rather than affecting both UG production and the quantitative trait 394 under directional selection (Figures 4B and 5B). This was due to the fact that if the mutation 395 affecting both UG production and the phenotypic trait had a positive effect on fitness (i.e. the 396 mutation brought the phenotype closer to the new optimum), the mutation was selected 397 despite its costly effect on UG production.

398 Overall, the fixation of polyploidy remained rare (Figures 5 and 6). With low 399 mutation rates, populations adapted to new conditions either by remaining diploid, with the 400 individuals adapting to the new conditions, or by forming mixed-ploidy populations of 401 adapted diploids and tetraploids, notably when the effect of the environment on UG 402 formation is high (Figures 5 and 6). With higher mutation rates, the formation of a mixed-403 ploidy population was much more frequent, with tetraploids reaching 10 to 55% in the 404 populations (Figure 5B). The fixation of polyploidy was frequent when unreduced gametes 405 swamped the population, i.e. with a high mutation rate and a strong effect of the environment 406 (Figures 5C and 6).

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Figure 4: Effect of pleiotropy type (no, all traits, unreduced gametes; number of pleiotropic loci = 40) and the environmental factor (0, 0.1, 0.3) on unreduced gametes production under different mutation rates (U = 0.005, 0.05, 0.1; differently colored lines), simulated from a population of genetically identical diploid individuals for 500 generations. Pleiotropy on all traits means that pleiotropy affected both the quantitative trait under directional selection and unreduced gametes production. (A) No environmental factor; (B) environmental factor 0.1; (C) environmental factor 0.3.



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Figure 5: Effect of pleiotropy type (no, all traits, unreduced gametes; number of pleiotropic loci = 40) and the environmental factor (0, 0.1, 0.3) on the frequency of tetraploids in the population under different mutation rates (U = 0.005, 0.05, 0.1; differently colored lines). simulated from a population of genetically identical diploid individuals for 500 generations. Pleiotropy on all traits means that pleiotropy affected both the quantitative trait under directional selection and unreduced gametes production. (A) No environmental factor; (B) environmental factor 0.1; (C) environmental factor 0.3.

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Figure 6: Effect of pleiotropy type (no, all traits, unreduced gametes; number of pleiotropic loci = 40) and the environmental factor (0, 0.1, 0.3) on the (A) fixation and (B) frequency of tetraploids in the population under different mutation rates (U = 0.005, 0.05, 0.1; differently colored lines). simulated from a population of genetically identical diploid individuals for 500 generations. Pleiotropy on all traits means that pleiotropy affected both the quantitative trait under directional selection and unreduced gametes production.

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441 Discussion & Conclusion:

442 We simulated the interplay between stabilizing selection and directional selection in response 443 to an environmental factor during the origin and fixation of autopolyploids. In this process, 444 the pleiotropic effect of mutations on UG production and the quantitative trait affecting 445 population fitness were tested. Under stabilizing selection, pleiotropy had a negative effect on 446 the production and genetic variation of unreduced gametes. Under a realistic low mutation 447 rate in the focal genes, we found no fixation of tetraploidy. Higher mutation rates led to 448 higher heritable rates of UG production and to the formation of stable mixed-ploidy 449 populations, however, with tetraploids remaining rare (< 10% in frequency). Under 450 directional selection, the environmental factor had a complex effect on UG production and 451 population cytotype structure. Taken as an isolated parameter, the higher the effect of the 452 environment on UG production, the higher the frequency of polyploidy at the end of the 500 453 simulated generations following the environmental change. In addition, environment and 454 genetic architecture had a synergistic effect: the stronger the effect of the environment, the 455 higher the genetic component of UG production. This is mainly because the production of 456 unreduced gametes is frequency-dependent, i.e. the higher it is, the less costly it is.

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458 Evolution of tetraploidy during stressful conditions

Environmental stress is considered to be important to the origin of polyploids, as it affects UG production (Kreiner *et al.* 2017a; Mason & Pires 2015; Ramsey & Schemske 1998a). In particular, high and low temperatures have been shown to impact male UG formation (De Storme *et al.* 2012; Mason *et al.* 2011; Pecrix *et al.* 2011; Schindfessel *et al.* 2023; Wang *et al.* 2017), although the plants used therein were mainly of hybrid origin, which makes them generally more prone to form unreduced gametes (Ramsey & Schemske 1998a). Treatment 465 with high temperatures is commonly used to generate unreduced male gametes for polyploid 466 breeding (Mai *et al.* 2019). In our simulations, tetraploids got fixed in a population either 467 under extreme environmental effects or through higher genetic mutation rates that increase 468 UG production to ca. 20 to 40% at equilibrium (U = 0.05, 0.1; Figure 5C; Supplementary 469 Figure 2). These results are in line with the recent Gerstner et al. (2024) model which found 470 that environmental stochasticity for UG production can lead to the evolution of polyploidy. 471 However, as UG production in natural populations of non-hybrid species is typically much 472 lower, ranging from 0.1 to 2% (Kreiner et al. 2017), our high mutation rates seem very 473 unrealistic. If we assumed a realistic mutation rate for an average of 2% of UG production, 474 tetraploids only got fixed in a population under a strong environmental effect in our 475 simulations, while a moderate environmental effect resulted in a mixed-ploidy population 476 with rare tetraploids (< 10%; Figure 5C). However, we found that the higher the effect of the 477 environment on UG production, the higher the genetic increase of UG formation. If the 478 environmental stress also affects the mutation rate (like U.V. exposure that could affect both 479 temperature and alter the mutation rate; Gengyo-Ando & Mitani 2000), it is possible that 480 higher rates of polyploids could be found.

481 In the first simulated generations after the onset of the directional selection episode, which 482 represents the environmental change, we observed increased UG production and an increased 483 frequency of newly adapted tetraploid individuals in the population compared to later 484 generations (Figure 4). Under directional selection, mutations leading to UG formation might 485 be beneficial for tetraploid establishment, while counteracting diploid adaptation. Polyploids 486 may initially benefit from phenotypic shifts associated with WGD (Clo & Kolář 2021; 487 Porturas et al. 2019). However, both of these meta-analyses are based on studies of 488 established natural populations, which prevents filtering out the effects of selection and 489 genetic drift that are known to have shaped population dynamics of established polyploids,

490 but have less affected the first generations after a WGD event. The immediate, short-term 491 effect of a WGD can only be assessed using experimentally induced, synthetic 492 autopolyploids. For example, Westermann et al. (2024) showed that pollen tube tip growth is 493 drastically challenged in immediate response to a WGD event: in synthetic autotetraploids of 494 Arabidopsis arenosa and Arabidopsis thaliana pollen tubes grew slowly and abnormally, 495 while in established tetraploids of A. arenosa pollen tube physiology and morphology 496 returned to normal. This suggests initially reduced polyploid formation and fitness shortly 497 after a WGD event - a scenario not modeled here. Empirical frequencies of neo-polyploids in 498 natural populations might thus be lower. Decreased polyploid origin and fitness may also 499 stem from the formation of triploid offspring, which have low viability due to the triploid 500 block (Burton & Husband 2000a; Ramsey & Schemske 1998b). However, as we did not 501 account for triploid formation in our model, this does not affect the simulations.

502 Our findings are in contrast to that of (Clo 2022b) in that the author found neo-tetraploidy to 503 be associated with an initial decrease in their adaptive potential compared to their diploid 504 progenitors, while within less than 100 generations of the simulation they reached or 505 exceeded the additive variance of the diploids. In our simulations, tetraploids were directly 506 adapted and selected for an initial around 50 generations, but then were selected against once 507 diploid individuals were closer to the new phenotypic optimum, and they remained in the 508 minority within populations, resulting in mixed-ploidy populations under moderate 509 environmental effects. Even if the link between polyploidy and adaptive potential is probably 510 highly multifactorial (Clo 2022a), modeling non-adapted tetraploids with smaller genetic 511 diversity might have favored the diploid cytotype in mixed-ploidy populations.

512 Especially for plants, mixed-ploidy populations have been frequently reported in 513 multiple diploid-autotetraploid species, with varying proportions of diploids and their

514 polyploid derivatives (Kolář et al. 2017). The strength of selection against polyploid 515 cytotypes in these populations further depends on the mating system and clonal propagation, 516 although their interactions are not fully understood, based on both empirical data (Barringer 517 2007; Hojsgaard 2018; Van Drunen & Husband 2019) and theoretical modeling (Griswold 518 2021; Oswald & Nuismer 2011; Van Drunen & Friedman 2022). Eventually, gene flow 519 between ploidy levels may transfer novel adaptive alleles (e.g., in A. arenosa, Baduel et al. 520 2018; Australian burrowing frogs of the genus *Neobatrachus*, Novikova et al. 2020) and may 521 contribute to tetraploid fixation in the long-term, but we did not model this scenario. In our 522 simulations, only under a strong environmental effect tetraploids came to fixation. This is in 523 line with the theoretical framework of Oswald & Nuismer (2011) who showed that neo-524 tetraploid populations can adapt more efficiently than their diploid progenitors if the 525 populations undergo an environmental change, and that this change is aligned with the 526 phenotypic modifications found in tetraploid individuals. The duplication of entire genomes 527 immediately creates redundant informational 'modules' in the genome, offering possibilities 528 for a wider exploration of genotypic and phenotypic space (Yao *et al.* 2019). It also supports 529 the hypothesis that polyploids are expected to face higher extinction rates than their diploid 530 progenitors (Levin 2019) in the absence of environmental change, due to their smaller 531 geographical ranges upon emergence. In contrast, polyploids may thrive in stressful 532 environments and broaden their ranges, leading to their prevalence in such environments and 533 during historical periods of climate change (Bomblies 2020; Rice et al. 2019; Van De Peer et 534 al. 2021b). In sum, while our model did not include post-WGD environmental adaptation, it 535 shows that environmental change may support polyploid establishment by directly affecting 536 the rate of UG formation, moreover, with a further positive feedback effect on its underlying 537 genes.

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539 The consequences of UG genetic architecture on polyploidy evolution

540 The production of unreduced gametes is genetically determined (Parrott & Smith 1986; 541 Tavoletti et al. 1991b). It varies among individuals within and between populations, such that 542 most individuals in natural populations of non-hybrid species produce male unreduced 543 gametes at zero or low frequency and only a small number of individuals produce 544 substantially more (Kreiner et al. 2017a). Unreduced gamete mutations affect various 545 cytological stages (Bretagnolle & Thompson 1995; Brownfield & Kohler 2011; De Storme & 546 Geelen 2013), and they involve pre-meiotic, meiotic and post-meiotic pathways. Key proteins 547 comprise DYAD, AtPS1, JAS, TES, MPK4 (meiotic) and INCENP, RBR (post-meiotic). 548 Some of these genes are polygenic (Bretagnolle & Thompson 1995; Brownfield & Kohler 549 2011; De Storme & Geelen 2013), which may account for the variation in UG frequencies in 550 natural populations and the variance of UG production found in our simulations. A clear 551 theoretical and empirical framework for the effect of genetic architecture of UG production, 552 in particular pleiotropic effects, on the fixation of polyploidy is lacking. It is unclear if a few 553 major effect mutations or numerous small effect mutations are involved, and how they affect 554 the further trajectory of polyploid adaptive evolution.

555 In our simulations, one of the main findings is that pleiotropy always has a negative effect on 556 UG production and its variance. For example, a higher number of pleiotropic loci 557 significantly reduces UG production (Figure 3). This indicates that pleiotropy constrains UG 558 production by increasing the cost of complexity, a general effect of pleiotropy (Bomblies & 559 Peichel 2022; Orr 2000). While UG production and variance under pleiotropic effects is 560 lowered during stabilizing selection in our simulations, the overall fitness of the population is 561 often increased and thus seems to benefit from pleiotropic effects (Figure 1). However, we 562 found that during directional selection, if there is pleiotropy between UG formation and fitness-related loci, a mutation having both a positive effect for diploid adaptation and increasing UG production can be temporarily selected. It is known that mutations affecting UG formation in plants are often linked to male and/or female fitness (Brownfield & Kohler 2011). If this kind of pleiotropic mutations can increase in frequency during an environmental change, as explored by us, polyploidy could evolve by chance at least at low frequency, highlighting once again the role of stochasticity in polyploid evolution (Clo *et al.* 2022; Gerstner *et al.* 2024; Kauai *et al.* 2023).

570 According to empirical data, UG production is a deleterious phenotype, maintained at a low 571 frequency in the population (Kreiner *et al.* 2017b). In general, pleiotropy on quantitative traits 572 can reduce the selection efficacy in natural populations (Fraïsse et al. 2019; Zhang 2023). 573 Consequently, in our simulations individuals with UG production faced strong purifying 574 selection but were still maintained in the population. Pleiotropy is discussed to have an 575 antagonistic effect between fitness and reproduction, in accordance with the theory of 576 pleiotropy constraints on adaptive traits (Hughes & Leips 2017). Antagonistic pleiotropy or 577 the trade-off between traits has also been found for reproduction versus aging/longevity 578 (Austad & Hoffman 2018; Wu et al. 2022). Empirical work has shown that intermediate 579 pleiotropy is a driver of adaptation in A. thaliana (Frachon et al. 2017). Besides, through 580 evolution experiments of yeast (Saccharomyces cerevisiae), pleiotropy resulting from a 581 changing environment contributed to molecular adaptation (Chen & Zhang 2020). In our 582 simulations, environment and genetic architecture have a synergistic effect: with both an 583 environmental effect and pleiotropy, population fitness decreased. Similarly, under 584 environmental change, higher pleiotropic effects on single cell morphology in yeast were 585 observed compared to a stable environment (Geiler-Samerotte et al. 2020). Reduction of 586 population fitness resulting from pleiotropy could eventually lead to a reduction of the 587 population size, and the population could then be more affected by genetic drift. This

scenario would be in accordance with previous models showing that the fixation of
polyploids relies on strong genetic drift more than on natural selection (Clo *et al.* 2022),
favoring polyploids under environmental change.

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592 *Limitations of the model*

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594 If our model is informative, we had to avoid some genetic and ecological aspects 595 related to ploidy for the model to remain understandable. A first simplification is that we 596 hypothesized triploid and higher polyploid levels to be non-viable. Triploids may play a 597 crucial role in mixed-ploidy population dynamics (Husband 2004; Kauai et al. 2024). 598 However, triploids are typically very rare in natural populations of autopolyploids, possibly 599 being abundant only during the earliest stage of polyploid formation in a population (Burton 600 & Husband 2000b; Husband & Sabara 2004; Kolář et al. 2017; Ramsey & Schemske 1998a). 601 Also, we decided to model the fact that polyploidy is adaptive in the short-term, which can be 602 true in some conditions (Jiang et al. 2022; Maherali et al. 2009; Wang et al. 2019; Wu et al. 603 2019), but not in general (Clo & Kolář 2021; Porturas et al. 2019). Considering equivalent 604 fitness for diploids and tetraploids, or a decrease in fitness in polyploids, would not have 605 drastically changed the results. If the population is swamped with unreduced gametes (high 606 mutation rate and strong environmental effect), tetraploids would fix in a population 607 independently of their fitness (see Clo et al. 2022 for an example). However, for most 608 simulations for which we found mixed-ploidy populations, it is likely that with no fitness 609 advantage the populations would have remained diploid most of the time.

610 We modeled obligately outcrossing populations, while any form of assortative mating 611 or spatial structure favoring mating within cytotypes could increase the likelihood of fixation 612 of polyploidy (Otto & Whitton 2000). The first reason is that the aim of our model was to 613 simulate a more realistic architecture of UG production, and that this novelty should not 614 modify the outcomes of previous models (Husband 2004; Van Drunen & Friedman 2022). 615 Second, association between ploidy and different forms of assortative mating remains 616 controversial (Clo 2022b). If the association between clonality and polyploidy is widespread 617 (Van Drunen & Husband 2019), no clear patterns emerge for the effect of ploidy on the 618 evolution of selfing (Husband et al. 2008) or plant-pollinator interactions (Rezende et al. 619 2020). Finally, we did not model demographical events following the environmental change. 620 One of our major results is that, for realistic simulations (with UG production fitting 621 empirical data), the adaptation of populations to new conditions was more likely by diploid 622 individuals reaching the new phenotypic optimum, rather than by adapted tetraploids 623 reaching fixation. We could have simulated fluctuating population size, with low-fitness 624 individuals (diploids in our case) going extinct before reaching the reproductive stage for 625 example. With a lower population size, any increase in tetraploid frequency would have 626 lowered the strength of frequency-dependent selection more efficiently than in our 627 simulations, possibly leading to mixed-ploidy or tetraploid populations more often.

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630 Data availability: Simulation code is available at

631 https://github.com/JosselinCLO/Model_UG_pleiotropy_environment.

632

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644 Reference:

- Austad, S.N. & Hoffman, J.M. (2018). Is antagonistic pleiotropy ubiquitous in aging biology?
 Evolution, Medicine, and Public Health, 2018, 287–294.
- Baack, E.J. (2005). To succeed globally, disperse locally: effects of local pollen and seed
 dispersal on tetraploid establishment. *Heredity*, 94, 538–546.
- Baduel, P., Hunter, B., Yeola, S. & Bomblies, K. (2018). Genetic basis and evolution of rapid
 cycling in railway populations of tetraploid Arabidopsis arenosa. *PLoS Genet*, 14,
 e1007510.
- 652 Barringer, B.C. (2007). Polyploidy and self□fertilization in flowering plants. *American J of* 653 *Botany*, 94, 1527–1533.
- Bomblies, K. (2020). When everything changes at once: finding a new normal after genome duplication. *Proc. R. Soc. B.*, 287, 20202154.
- Bomblies, K. & Peichel, C.L. (2022). Genetics of adaptation. *Proc. Natl. Acad. Sci. U.S.A.*,
 119, e2122152119.
- Bretagnolle, F. & Thompson, J.D. (1995). Gametes with the somatic chromosome number:
 mechanisms of their formation and role in the evolution of autopolyploid plants. *New Phytologist*, 129, 1–22.
- 661 Brownfield, L. & Kohler, C. (2011). Unreduced gamete formation in plants: mechanisms and 662 prospects. *Journal of Experimental Botany*, 62, 1659–1668.
- Bürger, R., Wagner, G.P. & Stettinger, F. (1989). How much heritable variation can be
 maintained in finite populations by mutation–selection balance? *Evolution*, 43, 1748–
 1766.
- 666 Burton, T.L. & Husband, B. (2000a). Fitness differences among diploids, tetraploids, and 667 their triploid progeny in Chamerion angustifolium: mechanisms of inviability and 668 implications for polyploid evolution. *Evolution*, 54, 1182–1191.
- Burton, T.L. & Husband, BrianC. (2000b). FITNESS DIFFERENCES AMONG DIPLOIDS,
 TETRAPLOIDS, AND THEIR TRIPLOID PROGENY IN CHAMERION
 ANGUSTIFOLIUM: MECHANISMS OF INVIABILITY AND IMPLICATIONS FOR
 POLYPLOID EVOLUTION. *Evolution*, 54, 1182–1191.
- Čertner, M., Sudová, R., Weiser, M., Suda, J. & Kolář, F. (2019). Ploidy□altered phenotype
 interacts with local environment and may enhance polyploid establishment in *Knautia serpentinicola* (Caprifoliaceae). *New Phytologist*, 221, 1117–1127.

676 677	Chen, P. & Zhang, J. (2020). Antagonistic pleiotropy conceals molecular adaptations in changing environments. <i>Nat Ecol Evol.</i> 4, 461–469.
678	Clo, J. (2022a). Polyploidization: Consequences of genome doubling on the evolutionary
680	Clo 1 (2022b) The evolution of the additive variance of a trait under stabilizing selection
681	after autopolyploidization .1 of Evolutionary Biology 35, 891–897
682	Clo. J. & Kolář, F. (2021). Short and long term consequences of genome doubling: a
683	meta analysis. American J of Botany, 108, 2315–2322.
684	Clo, J. & Opedal, Ø.H. (2021). Genetics of quantitative traits with dominance under
685	stabilizing and directional selection in partially selfing species. Evolution, 75, 1920-
686	1935.
687	Clo, J., Padilla García, N. & Kolář, F. (2022). Polyploidization as an opportunistic mutation:
688	The role of unreduced gametes formation and genetic drift in polyploid establishment.
689	J of Evolutionary Biology, 35, 1099–1109. De Storme, N., Conenhaver, C.B. & Coelen, D. (2012). Braduction of Diploid Male Cometee
690 601	in Arabidansis by Cold Induced Destabilization of Destabilization of Destabilization
692	Arrays Plant Physiology 160, 1808–1826
693	De Storme N & Geelen D (2013) Sexual polyploidization in plants – cytological
694	mechanisms and molecular regulation. New Phytologist 198, 670–684
695	d'Erfurth. I., Cromer, L., Jolivet, S., Girard, C., Horlow, C., Sun, Y., <i>et al.</i> (2010). The
696	CYCLIN-A CYCA1;2/TAM Is Required for the Meiosis I to Meiosis II Transition and
697	Cooperates with OSD1 for the Prophase to First Meiotic Division Transition. PLoS
698	Genet, 6, e1000989.
699	Erilova, A., Brownfield, L., Exner, V., Rosa, M., Twell, D., Scheid, O.M., et al. (2009).
700	Imprinting of the Polycomb Group Gene MEDEA Serves as a Ploidy Sensor in
701	Arabidopsis. PLoS Genet, 5, e1000663.
702	Felber, F. (1991). Establishment of a tetraploid cytotype in a diploid population: Effect of
703	relative fitness of the cytotypes. J of Evolutionary Biology, 4, 195–207.
704	Frachon, L., Libourei, C., Villoutreix, R., Carrere, S., Giorieux, C., Huard-Chauveau, C., <i>et al.</i>
705	(2017). Intermediate degrees of synergistic pleiotropy drive adaptive evolution in
700	Eraïsse C. Ruiveu Sala G. & Viceso B. (2010) Pleiotropy Modulates the Efficacy of
708	Selection in Drosophila melanogaster Molecular Biology and Evolution 36, 500–515
709	Gavnor MI Kortessis N Soltis D.F. Soltis P.S. & Ponciano J.M. (2023) Dynamics of
710	mixed-ploidy populations under demographic and environmental stochasticities.
711	BioRxiv, 2023.03. 29.534764.
712	Geiler-Samerotte, K.A., Li, S., Lazaris, C., Taylor, A., Ziv, N., Ramjeawan, C., et al. (2020).
713	Extent and context dependence of pleiotropy revealed by high-throughput single-cell
714	phenotyping. PLoS Biol, 18, e3000836.
715	Gengyo-Ando, K. & Mitani, S. (2000). Characterization of Mutations Induced by Ethyl
716	Methanesulfonate, UV, and Trimethylpsoralen in the Nematode Caenorhabditis
/1/	elegans. Biochemical and Biophysical Research Communications, 269, 64–69.
718	Gerstner, B.P., wearing, H.J. & whitney, K.D. (2024). why so many polypiolog? Accounting
719	barriers to polyploid establishment <i>Journal of Evolutionary Biology</i> 37, 325–335
720	Griswold C.K. (2021) The effects of migration load selfing inbreeding depression and the
722	denetics of adaptation on autotetraploid versus diploid establishment in peripheral
723	habitats. Evolution. 75. 39–55.
724	Halligan, D.L. & Keightley, P.D. (2009). Spontaneous mutation accumulation studies in
725	evolutionary genetics. Annual Review of Ecology, Evolution, and Systematics, 40,
726	151–172.
727	Hojsgaard, D. (2018). Transient Activation of Apomixis in Sexual Neotriploids May Retain
728	Genomically Altered States and Enhance Polyploid Establishment. Front. Plant Sci.,
729	9, 230.
730	Hughes, K.A. & Leips, J. (2017). Pleiotropy, constraint, and modularity in the evolution of life

731	histories: insights from genomic analyses. Annals of the New York Academy of
732	Sciences, 1389, 76–91.
733	Husband, B.C. (2004). The role of triploid hybrids in the evolutionary dynamics of mixed-
734	ploidy populations: TRIPLOIDS IN MIXED-PLOIDY POPULATIONS. Biological
735	Journal of the Linnean Society, 82, 537–546.
736	Husband, B.C., Ozimec, B., Martin, S.L. & Pollock, L. (2008). Mating Consequences of
737	Polyploid Evolution in Flowering Plants: Current Trends and Insights from Synthetic
738	Polyploids. International Journal of Plant Sciences, 169, 195–206.
739	Husband, B.C. & Sabara, H.A. (2004). Reproductive isolation between autotetraploids and
740	their diploid progenitors in fireweed. Chamerion angustifolium (Onagraceae). New
741	Phytologist, 161, 703–713.
742	Jiang, J., Yang, N., Li, L., Qin, G., Ren, K., Wang, H., et al. (2022). Tetraploidy in Citrus
743	wilsonii Enhances Drought Tolerance via Synergistic Regulation of Photosynthesis
744	Phosphorylation and Hormonal Changes Front Plant Sci 13 875011
745	Jiao Y Wickett NJ Avvampalavam S Chanderbali AS Landherr I Ralph PF et
746	al (2011) Ancestral polyploidy in seed plants and angiosperms. Nature 473, 97–
747	100
748	Kauai E Bafort O Mortier E Van Montagu M Bonte D & Van De Peer Y (2024)
749	Interspecific transfer of genetic information through polyploid bridges. Proc. Natl
750	Acad Sci U.S.A. 121 e2400018121
751	Kauai F Mortier F Milosavlievic S Van De Peer Y & Bonte D (2023) Neutral
752	processes underlying the macro eco-evolutionary dynamics of mixed-ploidy systems
753	Proc R Soc B 290 20222456
754	Kolář F. Čertner M. Suda J. Schönswetter P. & Husband B.C. (2017) Mixed-Ploidy
755	Species: Progress and Opportunities in Polyploid Research Trends in Plant Science
756	22 1041 - 1055
757	Kreiner IM Kron P & Husband B C (2017a) Evolutionary Dynamics of Unreduced
758	Gametes Trends in Genetics 33, 583–593
750	Kreiner IM Kron P & Husband B C (2017b) Frequency and maintenance of unreduced
760	cametes in natural plant populations: associations with reproductive mode life
761	history and genome size New Phytologist 214, 870–889
762	Levin D.A. (1975) MINORITY CYTOTYPE EXCLUSION IN LOCAL PLANT
763	
764	Levin D.A. (2010) Why polyploid exceptionalism is not accompanied by reduced extinction
765	rates Plant Syst Evol 305 1–11
766	Li B \Box H X χ \Box M & Ridout M S (2004) Modelling the establishment and spread of
767	autotetranloid plants in a spatially beterogeneous environment. Lof Evolutionary
768	Biology 17 562-573
760	Madlung A (2013) Polyploidy and its effect on evolutionary success: old questions revisited
703	with new tools. Heredity 110, 99–104
771	Maharali H. Waldan A E & Husband B C (2000) Canoma duplication and the evolution
772	of physiological responses to water stress. New Phytologist 184, 721–731
773	Mai V Li H Suo V Eu I Sun P Han W et al (2010) High temperature treatment
774	apportes unreduced pollen in persimmen (Diospures keki Thunh). Scientia
775	Horticulturae 258 108774
776	Manna E. Martin G. & Lenormand T. (2011) Fitness landscapes: an alternative theory for
777	the dominance of mutation. Constice 180, 022, 027
778	Mason A S. Nelson MN. Van G & Cowling WA (2011) Production of viable male
770	unreduced gametes in Brassica interspecific hybrids is genotype specific and
780	stimulated by cold temperatures <i>BMC Plant Biol</i> 11, 103
781	Mason A S & Pires I.C. (2015) Unreduced gametes: meiotic mishan or evolutionary
782	mechanism? Trends in Genetics 31 5-10
783	Novikova PYu Brennan I.G. Booker W. Mahony M. Doughty P. Lemmon Δ.P. et al.
784	(2020) Polyploidy breaks speciation barriers in Australian burrowing frogs
785	Neobatrachus, <i>PLoS Genet</i> , 16, e1008769.

786 787 788	Oberlander, K.C., Dreyer, L.L., Goldblatt, P., Suda, J. & Linder, H.P. (2016). Species and polyploid poor: Insights into the evolutionary role of whole genome duplication from the Cape flora biodiversity botspot. <i>American Journal of Botany</i> , 103, 1336–
789	1347
790	Orr H A (2000) ADAPTATION AND THE COST OF COMPLEXITY Evolution 54 13-20
701	Oswald B.P. & Nuismer S.L. (2011) A Unified Model of Autopolyploid Establishment and
792	Evolution The American Naturalist 178, 687–700
703	Otto S.P. (2007) The Evolutionary Consequences of Polyploidy. Cell 131, 452–462
794	Otto S.P. & Whitton I (2000) Polyploid incidence and evolution Annual review of genetics
795	34 401–437
796	Parrott W & Smith R (1986) Recurrent selection for 2n pollen formation in red clover 1
797	Cron Science 26 1132–1135
798	Pecrix Y Rallo G Folzer H Cigna M Gudin S & Le Bris M (2011) Polyploidization
799	mechanisms: temperature environment can induce diploid gamete formation in Rosa
800	sp. Journal of Experimental Botany 62, 3587–3597
801	Porturas I D Anneberg T I Curé A F Wang S Althoff D M & Segraves K A (2019)
802	A meta analysis of whole genome duplication and the effects on flowering traits in
803	plants American J of Botany 106 469–476
804	Ramsey J & Schemske D W (1998a) PATHWAYS MECHANISMS AND RATES OF
805	POLYPLOID FORMATION IN FLOWERING PLANTS Annu Rev Ecol Syst 29
806	467–501
807	Ramsey J & Schemske D W (1998b) Pathways mechanisms and rates of polyploid
808	formation in flowering plants. Annual review of ecology and systematics 29, 467–
809	501.
810	Ravi, M., Marimuthu, M.P. & Siddigi, I. (2008). Gamete formation without meiosis in
811	Arabidopsis, <i>Nature</i> , 451, 1121–1124.
812	Rezende, L., Suzigan, J., Amorim, F.W. & Moraes, A.P. (2020). Can plant hybridization and
813	polyploidy lead to pollinator shift? Acta Bot. Bras., 34, 229–242.
814	Rice, A., Šmarda, P., Novosolov, M., Drori, M., Glick, L., Sabath, N., et al. (2019), The global
815	biogeography of polyploid plants. <i>Nat Ecol Evol.</i> 3, 265–273.
816	Ronce, O., Shaw, F.H., Rousset, F. & Shaw, R.G. (2009). Is inbreeding depression lower in
817	maladapted populations? A quantitative genetics model. <i>Evolution</i> , 63, 1807–1819.
818	Schindfessel, C., De Storme, N., Trinh, H.K. & Geelen, D. (2023). Asynapsis and meiotic
819	restitution in tomato male meiosis induced by heat stress. Front. Plant Sci., 14,
820	1210092.
821	Soltis, P.S., Marchant, D.B., Van De Peer, Y. & Soltis, D.E. (2015). Polyploidy and genome
822	evolution in plants. Current Opinion in Genetics & Development, 35, 119–125.
823	Tavoletti, S., Mariani, A. & Veronesi, F. (1991a). Cytological Analysis of Macro and
824	Microsporogenesis of a Diploid Alfalfa Clone Producing Male and Female 2 n
825	Gametes. Crop Science, 31, 1258–1263.
826	Tavoletti, S., Mariani, A. & Veronesi, F. (1991b). Phenotypic recurrent selection for 2n pollen
827	and 2n egg production in diploid alfalfa. <i>Euphytica</i> , 57, 97–102.
828	Trávníček, P., Dočkalová, Z., Rosenbaumová, R., Kubátová, B., Szeląg, Z. & Chrtek, J.
829	(2011). Bridging global and microregional scales: ploidy distribution in Pilosella
830	echioides (Asteraceae) in central Europe. Annals of Botany, 107, 443–454.
831	Van De Peer, Y., Ashman, TL., Soltis, P.S. & Soltis, D.E. (2021a). Polyploidy: an
832	evolutionary and ecological force in stressful times. The Plant Cell, 33, 11–26.
833	Van De Peer, Y., Ashman, TL., Soltis, P.S. & Soltis, D.E. (2021b). Polyploidy: an
834	evolutionary and ecological force in stressful times. <i>The Plant Cell</i> , 33, 11–26.
835	Van De Peer, Y., Mizrachi, E. & Marchal, K. (2017). The evolutionary significance of
836	polyploidy. <i>Nat Rev Genet</i> , 18, 411–424.
837	Van Drunen, W.E. & Friedman, J. (2022). Autopolyploid establishment depends on
838	life history strategy and the mating outcomes of clonal architecture. <i>Evolution</i> , 76,
839	1953–1970.
840	Van Drunen, W.E. & Husband, B.C. (2019). Evolutionary associations between polyploidy,

841	clonal reproduction, and perenniality in the angiosperms. New Phytologist, 224,
842	1266–1277.
843	Wang, C., Ge, W., Yin, H., Zhang, Y. & Li, J. (2024). MicroRNAs and their targets involved in
844	unreduced pollen formation induced by heat stress in Camellia nitidissima. Scientia
845	Horticulturae, 323, 112462.
846	Wang, CJ., Zhang, LQ., Dai, SF., Zheng, YL., Zhang, HG. & Liu, DC. (2010).
847	Formation of unreduced gametes is impeded by homologous chromosome pairing in
848	tetraploid Triticum turgidum × Aegilops tauschii hybrids. <i>Euphytica</i> , 175, 323–329.
849	Wang, J., Li, D., Shang, F. & Kang, X. (2017). High temperature-induced production of
850	unreduced pollen and its cytological effects in Populus. Scientific Reports, 7, 5281.
851	Wang, Z., Zhao, Z., Fan, G., Dong, Y., Deng, M., Xu, E., et al. (2019). A comparison of the
852	transcriptomes between diploid and autotetraploid Paulownia fortunei under salt
853	stress. Physiol Mol Biol Plants, 25, 1–11.
854	Westermann, J., Srikant, T., Gonzalo, A., Tan, H.S. & Bomblies, K. (2024). Defective pollen
855	tube tip growth induces neo-polyploid infertility. Science, 383, eadh0755.
856	Wu, D., Wang, Z., Huang, J., Huang, L., Zhang, S., Zhao, R., et al. (2022). An antagonistic
857	pleiotropic gene regulates the reproduction and longevity tradeoff. Proc. Natl. Acad.
858	<i>Sci. U.S.A.</i> , 119, e2120311119.
859	Wu, GQ., Lin, LY., Jiao, Q. & Li, SJ. (2019). Tetraploid exhibits more tolerant to salinity
860	than diploid in sugar beet (Beta vulgaris L.). Acta Physiol Plant, 41, 52.
861	Yao, Y., Carretero-Paulet, L. & Van De Peer, Y. (2019). Using digital organisms to study the
862	evolutionary consequences of whole genome duplication and polyploidy. PLoS ONE,
863	14, e0220257.
864	Zhang, J. (2023). Patterns and Evolutionary Consequences of Pleiotropy. Annu. Rev. Ecol.
865	Evol. Syst., 54, 1–19.
866	