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Metabolites derived from radical oxidation of PUFA: NEO-PUFAs, promising molecules for health?

Anna Abramova^a, Jamie Bride^b, Camille Oger^a, Marie Demion^b, Jean-Marie Galano^a, Thierry Durand $a, **$, Jérôme Roy $c, *$

^a Institut des Biomolécules Max Mousseron, Pôle Recherche Chimie Balard, Université Montpellier, UMR 5247, CNRS, ENSCM, 34293, Montpellier cedex, France

^b *PhyMedExp, Universit*´*e de Montpellier, Inserm U1046, UMR CNRS 9412, Montpellier, France*

^c Universite de Pau et des Pays de l'Adour, INRAE, NUMEA, Aquapôle, 64310, Saint-Pée-sur-Nivelle, France

their potential applications in medical research and treatment.

1. Introduction

Neuroprostanes Bioactive NEO-PUFAs –

In 1956, Denham Harman proposed the idea that "aging and the degenerative diseases associated with it are attributed basically to the deleterious side attacks of free radicals on cell constituents and on the connective tissue" [1]. Since then, numerous findings have emerged connecting oxidative stress to the development of various human diseases, including vascular disorders, inflammatory ailments, cancers, and neurodegenerative conditions [2].

However, certain end-products of oxidative stress, particularly those originating from lipid peroxidation, are not only regarded as sensitive biomarkers of oxidative stress but also often exhibit potent biological activity [3].

The presence of multiple double bonds that are prone to peroxidation, makes polyunsaturated fatty acids (PUFAs), a crucial component of human body and particularly brain cells, vulnerable to the detrimental effects of oxidative stress [4]. Today, several classes of active compounds, resulting from peroxidation of different PUFAs, in other words,

Non-Enzymatic Oxygenated metabolites of PUFAs (newly appointed NEO-PUFAs), have been identified [5]. Following the discovery of prostaglandins, the enzymatic metabolites of arachidonic acid (AA), in 1930s [6], it was commonly believed for many years that NEO-PUFAs either had a deleterious impact or, at best, no significant effect on health. It was not until Morrow et al. [7] identified prostaglandin-like compounds, which they named isoprostanes, in the 1990s that research in the field of non-enzymatic NEO-PUFAs restarted, leading to the subsequent identification and isolation of other compounds such as phytoprostanes (PhytoPs) by Parchmann and Mueller [8] in 1998, isofurans (IsoFs) by Fessel and Roberts [9] in 2004, or dihomo-isoprostanes (dihomo-IsoPs) by VanRollins and colleagues [10] in 2008. Two different nomenclature systems for naming complex NEO-PUFAs were proposed by Taber [11] and by Rokach [12]. According to the latter, NEO-PUFAs are divided in classes based on ω-carbon of fatty acid, which serves as the reference point. Roman numerals I-VIII refer the types of NEO-PUFAs. Noteworthy, this nomenclature is not approved by IUPAC. According to Taber's classification system, NEO-PUFAs are designated

peroxidation of docosahexaenoic acid (DHA), and isoprostanes, resulting from arachidonic acid (AA) and eicosapentaenoic acid (EPA) through the same peroxidation process. These non-enzymatic oxygenated metabolites newly appointed NEO-PUFAs have gained recognition as reliable markers of oxidative stress in neurogenerative and cardiovascular diseases. Moreover, some of them display a wide range of biological activities. The ability to detect and measure these metabolites offers precious insights into the mechanisms of oxidative damage and holds potential therapeutic implications for various health conditions, including neurodegenerative diseases. This review focuses on the role of neuroprostanes as biomarkers for oxidative stress and related diseases, highlighting

* Corresponding author. ** Corresponding author.

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E-mail addresses: thierry.durand@umontpellier.fr (T. Durand), jerome.roy@inrae.fr (J. Roy).

Fig. 1. Formation of isoprostanes (IsoPs) from AA, R = H, phosphatidylcholine (PC), phosphatidylethanolamine (PE), phosphatidylinositol (PI).

Fig. 2. Formation of neuroprostanes (NeuroPs) from DHA, $R = H$, phosphatidylcholine.

based on their precursor PUFA. Additionally, this system describes NEO-PUFAs by their ring substitution pattern (A-H, J), relative orientation of side chains, and the number of double bounds. This system, approved by IUPAC, will be used in this review.

1.1. NEO-PUFA from AA: F2-Isoprostanes

F2-Isoprostanes (IsoPs) are a class of NEO-PUFAs formed by the free radical-catalyzed peroxidation of arachidonic acid (AA). Unlike enzymatically derived eicosanoids, isoprostanes are generated, inside the membranes, through a complex series of reactions involving reactive oxygen species (ROS), which lead to the formation of a variety of

Fig. 3. ROS-mediated oxidation of Polyunsaturated Fatty Acids (PUFAs).

isomeric structures. As a result, their main structural difference compared to prostaglandins include the *cis*-relationship of the side chains and defined number of potential isomers. Besides, it is important to note that all these metabolites are produced *in vivo* as racemic mixture. According to the mechanism of peroxidation of AA proposed by Rokach and co-workers [13], the reaction begins with the abstraction of a hydrogen atom from one of the bis-allylic carbons of AA (C7, C10 ou C13) by reactive oxygen species, such as hydroxyl radicals (•OH). This step generates a radical intermediate (compounds 24–26). The radical

thus formed then reacts with molecular oxygen to yield a peroxy radical that undergoes a cyclization followed by trapping with an additional oxygen molecule to give bicyclic endoperoxide, which is eventually reduced to the final allylic alcohol (compounds **27**–**30**) (**Fig. 1**).

1.2. NEO-PUFA from DHA: F4-Neuroprostanes

F4-Neuroprostanes (NeuroPs) were first described by Roberts and coworkers [14] in 1998. These are NEO-PUFAs formed non-enzymatically *in vivo* by the oxidative attack of free reactive oxygen species (ROS) on docosahexaenoic acid (DHA). According to the biosynthetic mechanism proposed by Yin and colleagues [15], a family of eight possible regioisomeric NeuroPs can be formed. These are derived from five different bis-allylic carbons (C6, C9, C12, C15, C18) of DHA. The biosynthesis of NeuroPs is initiated by hydrogen atom abstraction by HO• at one of these bis-allylic positions to give a resonance stabilized pentadienyl radical (compounds **1**–**5**). Similar to IsoPs, NeuroPs (compounds **6**–**13**) are then formed following the same steps of oxygen trapping, cyclization, and reduction (Fig. 2).

Interestingly, pentadienyl radicals resulting from hydrogen atom abstraction at the C9, C12, or C15 position of DHA give rise in each case to two different NeuroPs regioisomers: the 7/11-series (compounds **6**, **7**), 10/14-series (compounds **8**, **9**), or 13/17-series (compounds **10**, **11**), respectively. Meanwhile, hydrogen abstraction at the C6 or C18 position produces a single regioisomer: the 4-series (compounds **12**) from the C6 position or the 20-series (compounds **13**) from the C18 position. Furthermore, quantitative analysis *in vitro* and *in vivo* has demonstrated that both 4-series and 20-series are produced in greater amounts compared to other regioisomers [16]. Finally, just like IsoPs, NeuroPs

Fig. 4. Synthesis of AdA, DHA, AA, EPA, ALA and DPA metabolites from 1,3-cyclooctadiene **14**.

are produced *in vivo* as racemic mixture, thus doubling the total number of possible products. In the end, it is remarkable how nature can convert such a simple molecule into a vast array of highly complex compounds.

1.3. NEO-PUFA from EPA, ALA, AdA, DPA: F3-IsoPs, dihomo-IsoPs, PhytoPs, F3-NeuroPs

Similar to AA and DHA, other PUFAs such as eicosapentaenoic acid (EPA), adrenic acid (AdA), α-linolenic acid (ALA), and docosapentaenoic acid (DPA) undergo oxidation through analogous metabolic pathways. EPA generates 6 regioisomeric F_3 -isoPs [17] while AdA yields 4 regioisomeric dihomo-Isoprostanes (dihomo-IsoPs) [10]. ALA and DPA oxidation produce 6 regioisomeric phytoprostanes (PhytoPs) [18] and 6 F3-NeuroPs, respectfully (Fig. 3).

1.4. Biological role of DHA

DHA, an ω-3 fatty acid, plays a vital role in maintaining healthy brain function and development. In humans, it is a major component of brain tissue, where it is especially abundant in gray matter, accounting for about 40 % of the total PUFA population and nearly 50 % of the weight of neuronal membranes [19]. Additionally, DHA is highly concentrated in sperm, retinal photoreceptors, and brain synapses $[20,21]$. Given the abundance and significance of DHA in the human body, identification, isolation and quantification of DHA NEO-PUFAs offers unique biomarkers of oxidative stress in the central nervous system. It also provides potential new therapeutic compounds as some of these NEO-PUFAs have demonstrated promising beneficial biological effects [22–25]. This review explores the fascinating journey of DHA metabolites, the neuroprostanes, as well as other NEO-PUFAs. It covers nearly three decades of research, highlighting key developments in synthesis, diagnostic applications, and biological activities, some of which have yet to be fully elucidated. The work outlined in this review results from productive collaborations with chemists, biologists, and clinicians across the globe [26–30].

2. Chemical synthesis

Over the past 30 years, various methods of the total synthesis of NeuroPs and other NEO-PUFAs have been described [31–35]. Durand's group has developed three distinct approaches: one involving radical carbocylization [36], another based on furan ring transformation [37], and a third utilizing a bicyclo-[3.3.0]-octene intermediate [38]. The latter is remarkable because the bicyclo-[3.3.0]-octene intermediate **15** (Fig. 4), prepared in five steps from commercial 1,3-cyclooctadiene **14**, serves as a versatile substrate for the synthesis of NeuroPs (compounds **9** [39], **12** [40], **13** [41], **16** [42]) as well as other NEO-PUFAs, such as IsoPs (compounds **17** [38], **18** [43], **19** [44]), PhytoPs (**20**, **21**), and dihomo-IsoPs (compounds **22** [45], **23** [46]) (Fig. 4). Moreover, intermediate **15** can be synthesized in both enantiomeric forms, enabling acces to each enantiomer.

To access the desired metabolites, lateral chains are introduced via Wittig or Horner-Wadworth-Emmons reactions using the corresponding phosphonium salts or phosphonates.

Fig. 4 outlines a non-exhaustive list of DHA, adrenic acid (AdA), eicosapentaenoic acid (EPA), α-linolenic acid (ALA), arachidonic acid (AA) and docosapentaenoic acid (DPA n-3 or DPA n-6) NEO-PUFAs synthesized by Durand's group using this approach [33–41].

3. NEO-PUFAs as biomarkers

This revue will focus on NEO-PUFAs as biomarkers in diseases and neutraceutics will not be addressed.

The first NEO-PUFAs studied as biomarkers of oxidative stress, are the F_2 -IsoPs derived from AA [47,48]. They became the gold standard for evaluating oxidative stress *in vivo* in human [49] as well as in animal models [50] because these molecules are chemically stable and can be detected in all analyzed biological fluids and tissues [51]. Since the 1990's, the $15-F_{2t}$ -IsoP and other derivatives have been commonly used as the best available standard for measuring the extent of lipid peroxidation in most tissue fluids [52].

3.1. Central nervous system (CNS)

The brain is particularly susceptible to oxidative stress due to its high oxygen requirements for oxidative metabolism and its rich content of ω-3 PUFAs, in particular DHA. One compelling example highlighting the biomarker role of NEO-PUFAs pertains to Rett syndrome (RTT) pathology [53]. This syndrome stems from a neuronal developmental disorder resulting from loss-of-function mutations in the methyl-CpG binding protein 2 (MECP2) gene which leads to autism in females [54], myocardial dysfunction (subclinical biventricular systo-diastolic myocardial dysfunction), and a 300-fold increased risk of sudden death [55]. In this study, increased oxidative stress was observed, which was manifested as significant lipid peroxidation in blood samples taken from RTT patients [53].

 F_4 -NeuroPs emerge as significant biomarkers in RTT, with plasma F_4 -NeuroPs levels correlating with disease severity, neurological symptom severity, mutation type, and clinical presentation. Thus, F₄-NeuroPs likely could play a role in the biochemical pathway from MeCP2 gene mutation to clinical manifestations, indicating the occurrence of a DHA oxidation process [56].

Additionally to F_4 -NeuroPs, clinical evidence indicates the involvement of F_2 -IsoPs in the underlying pathogenetic mechanisms of RTT [57]. Plasma levels of free F_2 -IsoPs are significantly elevated during the early stages of RTT compared to the later stages of typical RTT progression. Prior to 2011, it was believed that primary CNS damage in RTT primarily affected gray matter. However, the abundance of the precursor AdA in myelin and the heightened levels of F_2 -dihomo-IsoPs suggest early and severe damage to white matter in the brain, which supports previous findings from brain MRI studies $[57]$. Therefore, F₂-dihomo-IsoPs could be considered early indicators of lipid peroxidation in RTT. Finally, the research group led by De Felice and Signorini has highlighted the potential importance of NeuroPs and dihomo-IsoPs in sperm capacitation [58] and in various neurological diseases, including Krabbe disease [59] and cerebral adrenoleukodystrophy [60].

Furthermore, conditions such as aneurysmal subarachnoid hemorrhage (aSAH) and traumatic brain injury (TBI) are associated with severe CNS damage. Two case-controlled studies have demonstrated a significant elevation in cerebrospinal fluid (CSF) levels of IsoFs in patients with aSAH and TBI compared to their respective age- and gendermatched controls. Additionally, aSAH patients exhibited notably increased CSF levels of F_4 -NeuroPs and F_2 -IsoPs. In contrast, patients with TBI showed elevated CSF F_4 -NeuroPs levels, while F_2 -IsoPs remained similar to control levels [61]. These findings affirm that CNS injury, whether due to aSAH or TBI, leads to heightened oxidative stress. Given that DHA is a major PUFA in the brain, with further investigation, levels of F4-NeuroPs in CSF may serve as a more specific indicator of neurological dysfunction than F₂-IsoPs.

Moreover, Hsieh et al. demonstrated a correlation between increased F4-NeuroPs levels in CSF of aSAH patients and poor neurological outcomes $[62]$. They suggested that F₄-NeuroPs might offer greater utility than F_2 -IsoPs in CSF for predicting outcomes and interpreting the role of hemorrhage in aSAH. While Farias et al. observed increased F_2 -IsoPs levels during rat brain ischemia, they found that E_2/D_2 -IsoPs were elevated to a greater extent, suggesting that the latter may serve as better markers of oxidative stress in brain ischemia [63].

Some studies also reported the importance of NeuroPs and dihomo-IsoPs in Alzheimer disease (AD) and brain injury in newborn [64,65]. Recognizing the significant role of lipid peroxidation in AD, Garci-Blanco et al. (2018) hypothesized that corresponding NEO-PUFAs detected in urine samples could serve as potential biomarkers [66].

Through the analysis of urine samples from patients early diagnosed with mild cognitive impairment or mild dementia AD, their study revealed differences in total NEO-PUFAs (NeuroPs, IsoPs, IsoFs) between the control and clinical groups. Consequently, with further investigation, these NEO-PUFAs could have the potential to serve as early biomarkers for Alzheimer's disease, indicating the pro-oxidant condition of patients.

Finally, in 2021, this research group delved into studying the relationship between an oxidative stress, inflammatory biomarkers and cerebral lesion in newborns following an hypoxic-ischemic encephalopathy (HIE), a major cause of severe neurological disability and mortality in the perinatal period [67]. Remarkably, the identification of specific biomarkers associated with diagnosis and long-term outcome prediction, as a supplementary tool to standard evaluation, remains an unexplored territory in newborn critical care. To address this gap, they conducted a longitudinal study aiming to identify biomarkers of oxidative stress and inflammation in noninvasive urine samples from newborns with moderate/severe HIE, using liquid chromatography-mass spectrometry. Their findings unveiled significant differences in oxidative stress levels among neonates exhibiting distinct brain lesion patterns as detected by magnetic resonance imaging. Notably, there was an elevation in lipid peroxidation biomarkers (numerous NeuroPs, IsoPs) in newborns with cerebral lesions. This observation leads to the hypothesis that monitoring noninvasive lipid peroxidation biomarkers could contribute to the diagnosis and prediction of long-term outcomes, serving as a complementary tool to standard assessment methods. More recently, in 2023, Raffay et al. confirmed that NeuroPs in urine could serve as biomarkers for hypoxemia events in preterm neonate [68].

3.2. Pregnancy

In 2012, a study has quantified F₄-NeuroPs, F₂-IsoPs, and IsoFs in maternal plasma and cord blood of women with pre-eclampsia and those with normal pregnancies [69,70]. Pre-eclampsia is a pregnancy disorder that combines high blood pressure with the presence of protein in the urine. It results from a dysfunction of the placenta. Women with pre-eclampsia exhibited significantly elevated levels of maternal IsoFs and F_4 -NeuroPs, but no increase in F_2 -IsoPs. Cord blood levels of F4-NeuroPs were also found to be higher among neonates born to mothers with pre-eclampsia. Notably, cord blood levels of IsoFs were approximately five times higher than those detected in maternal plasma. This disparity could be indicative of the oxidative stress encountered during birth, wherein there is a transition from a relatively low intra-uterine oxygen environment to a significantly higher extra-uterine oxygen environment. Interestingly, maternal levels of F4-NeuroPs did not show a significant correlation with cord blood levels of F₄-NeuroPs in both pre-eclamptic and normal pregnancies, suggesting that the origin of cord blood F4-NeuroPs may be independent of maternal plasma levels. In normal pregnancies, there was a negative association between birth weight and maternal levels of F_2 -IsoPs, IsoFs, and F_4 -NeuroPs suggesting them as potential markers in maternal blood at early stages of pregnancy relate to subsequent maternal, fetal, and neonatal complications.

In 2023, Ferreira et al. investigated whether gestational diabetes mellitus (GDM) associated with maternal obesity modifies the placental profile of F_4 -NeuroPs and F_2 -IsoPs [71]. Their results revealed that levels of F_4 -NeuroPs and F_2 -IsoPs were higher in obesity compared to lean controls. However, when obesity was further complicated with GDM, these levels decreased to levels similar to those of lean women. The lower content of F_2 -IsoPs suggests adaptive placental responses in GDM, which may attenuate oxidative stress. However, the quantification of arachidonic acid (AA) is necessary to confirm this hypothesis. Conversely, the reduced levels of placental F4-NeuroPs in GDM may indicate impaired DHA metabolism, which could potentially affect fetal development and offspring health but further experiments are needed.

Interestingly, these results were not correlated with differences in placental content of DHA, AA, and PUFAs status, nor with maternal diet or gestational weight gain. This suggests that, with further investigation, that NeuroPs and IsoPs could serve as potential biomarkers and predictors of altered placental fatty acid metabolism.

3.3. Obesity

Obesity is a chronic disease and a major cause of morbidity and mortality. Numerous evidence has linked this condition to inflammation and oxidative stress [72]. To assess the potential NEO-PUFAs as biomarkers in obesity, recently, Sarajlic et al., have measured an increase of pro-inflammatory markers $5-F_{2c}$ -IsoP and $15-F_{2t}$ -IsoP as well and a decrease of $14-F_{4t}$ -NeuroP and $5-F_{3t}$ -IsoP with age in urine sample from obese patients [73]. This study suggests that ω -3/ ω -6 oxidation ratio may serve as an indicator of inflammation associated with obesity.

3.4. Cancer

Several pieces of evidence have suggested that prostate cancers may be associated with oxidative stress. This hypothesis was investigated by Barocas et al., in 2011, where urine was collected from men before biopsy and diagnosis. An increase in F_2 -IsoPs was highlighted in men diagnosed with high grade prostatic intra-epithelial neoplasia and prostate cancer [74]. This hypothesis was further explored by Brys et al., who measured high level of urinary 8-iso-PGF₂ in patients diagnosed with prostate cancer $[75]$. These results could suggest that F_2 -IsoPs found in urine may be a biomarker of oxidative stress in prostate cancer.

3.5. Renal function

End-stage renal disease is a condition in which the kidneys completely lose their ability to filter blood, requiring dialysis and a kidney transplant. However, after transplantation, there is an increased risk of allograft rejection, which leads to further graft loss and affects patient survival. It is therefore important to identify potential biomarkers that could reflect graft function after transplantation and predict the risk of rejection. In this regard, De las Heras-Gómez et al., showed a trend towards lower levels of 4-*epi*− 4-F3t -NeuroP, *ent*− 7 *(RS)*− 7-F2t-dihomo-IsoP, *ent*− 7*(S)*− 7-F2t-dihomo-IsoP and 15-F2t-IsoP in urine of renal transplant patients, which was associated with an improved kidney function [76]. In conclusion, with further investigation, these metabolites may serve as predictors of rejection.

3.6. Cardiovascular function

NEO-PUFAs has also been investigated as biomarkers in cardiovascular diseases such as atherosclerosis. This condition is a chronic inflammatory disease of vascular system associated with an increase of ROS and consequently lipid peroxidation. In 1997, Praticò et al., showed an increase of the content of two F₂-IsoPs, 8-epi-PGF_{2α} and IPF_{2α}-I, in human atherosclerotic plaque compared to controls [77]. In 2023, Lefort et al. used a mouse model deficient in very-long chain acyl-CoA dehydrogenase (VLCAD), an enzyme which catalyzes the first step of beta-oxidation in the mitochondria to mimics the inherited VLCAD deficiency (VLCADD) causing a predisposition to neonatal arrhythmias [78]. Compared to controls (*VLCAD^{+/+}*), *VLCAD^{-/-}* mouse hearts exhibited lipid alterations characterized by significantly higher levels of F_2 -IsoPs (8- F_{2t} -IsoP, 5- F_{2c} -IsoP and 5-*epi*-5- F_{2t} -IsoP) and lower levels of F4-NeuroPs (10-F4t-NeuroP, 20-F4t-NeuroP). This finding could suggest that, with further investigation, these NEO-PUFAs could serve as intriguing biomolecules for use as biological markers in the development of some cardiovascular pathologies.

Taken together, this non-exhaustive list of publications highlights some NEO-PUFAs as potential biomarkers in several diseases associated with oxidative stress. However, there is considerable evidence that some

of these NEO-PUFAs also display beneficial biological properties. This topic will be discussed in the next chapter.

4. New bioactive lipids

Regarding their bioactivity, NEO-PUFAs are poorly studied, and the idea of attributing biological activity to these peroxidation products remains dogmatically difficult to accept. The lack of studies can be attributed to several reasons: 1*- In vivo*, the concentrations of these NEO-PUFAs may be low in the absence of supplementation, 2- NEO-PUFAs are not commercially available or are scarcely so, and they must be customsynthesized, a skill possessed by few chemistry groups worldwide, 3- Enzymatic synthesis pathways quantitatively limit any other form of production of bioactive NEO-PUFAs.

4.1. Biological activities of ω-6 NEO-PUFAs

F2-IsoPs as « degradation products » induced by oxidative stress have long been considered only as biomarkers in diseases [79–81]. It was not until the research of Morrow and Roberts that the biological activity of these NEO-AA began to be discussed. Indeed, the biological activity of 15-F_{2t}-IsoP (also known as 8-epi-PGF2α) has been studied in the rat kidney and has shown that an infusion into the peripheral vein or the intrarenal artery reduces the glomerular filtration rate and the renal blood flow, indicating that it is a potent renal vasoconstrictor [7].

Since then, investigations have uncovered the vasoconstrictive effects of IsoP from AA in numerous vascular beds (heart, liver, lung, kidney, smooth muscle, retina) via the thromboxane receptor (TP) [82]. Additionally, $15-F_{2t}$ -IsoP can modulate platelet activity through the same receptor $[83]$. Furthermore, 15-E_{2t}-IsoP, an other NEO-AA, acts as a vasoconstrictor via the TP [84]. It also serves as a ligand for the E-series of prostaglandin receptors (EP), contributing to bronchoconstriction in the lungs. Bendorf et al. demonstrated that $15-F_{2t}$ -IsoP, 15- E_{2t} -IsoP, and 15- A_{2t} -IsoP inhibit VEGF-induced migration and tube formation of endothelial cells, thereby collectively inhibiting angiogenesis via activation of TBXA2R $[85]$. 15-J₂-IsoP has been found to elicit an inflammatory response by inhibiting peroxisome proliferator-activated receptor gamma (PPARγ) activation and inducing RAW264.7 cell apoptosis in a PPARγ-independent manner [86]. Another cyclopentenone AA derivative, 15-A₂-IsoP, exhibits anti-inflammatory effects by inhibiting the NF-κB pathway in lipopolysaccharide (LPS)-induced macrophages and human gestational tissues [87]. The overall picture of these cyclopentenone metabolites remains unclear, as recently highlighted for their prostaglandin equivalents, and is likely highly dependent on their structures [88].

4.2. Biological activities of ω-3 NEO-PUFAs

Like ω-6 NEO-PUFAs, ω-3 NEO-PUFAs are also considered as biomarkers in diseases. But for many years, diets enriched with ω-3 PUFAs have been associated with beneficial outcomes when given in diseases.

4.2.1. Anti-inflammatory and anti-atherogenic properties

To assess wheter ω-3 NEO-PUFAs are beneficial in an inflammation context, Sethi's group in 1996 demonstrated that the reduction in monocyte adhesion to endothelial cells is only observed when the cells are exposed to previously oxidized ω -3 PUFA [89], thereby challenging the role of ω-3 PUFA themselves and the importance of considering their NEO-PUFAs in studies.

Also, a first study revealed that, unlike $15-F_{2t}$ -IsoP derived from AA, 15-F3t-IsoP from EPA did not activate platelet aggregation but instead exerted an antiplatelet aggregating effect [90]. These initial data suggest that there is indeed a notable difference in activity between oxygenated products derived from ω-3 and ω-6 PUFAs, most likely linked to their structural differences [49].

In addition, endothelial cells treated with oxidized EPA revealed

inhibition of monocyte chemoattractant protein-1 (MCP-1) and endothelial nuclear factor-κB (NF-κB) through a PPAR-dependent pathway, supporting the anti-inflammatory effects of ω-3 NEO-PUFAs [91].

Among NEO-EPAs, 15-A3t-IsoP revealed a potent biological activity by inhibiting the activity of enzymes involved in inflammation such as iNOS or COX-2 via NF-κB inhibition in mouse macrophages [92]. Thus, many observed effects of ω-3 PUFA on inflammation may be due to their oxidized NEO-PUFAs, as well as their effects on oxidative stress. Indeed, 15-A_{3t}-IsoP and 15-J_{3t}-IsoP derived from EPA exhibit inflammatory activities [92]. They activate the NRF2 pathway (Nuclear factor erythroid 2-related factor 2) by inhibiting one of its negative regulators, Keap1 (Kelch-like ECH-associated protein 1) [93].

In 2017, Bosviel et al. conducted a dose-dependent and comparative study in human primary macrophages aiming to decipher the potential anti-inflammatory activity of NeuroPs (14-A_{4t}- and 4-F_{4t}-NeuroP) [24]. They observed that NeuroPs have anti-inflammatory properties with the highest dose (i.e., 10 μM). Interestingly, the strongest effects were obtained with the $14-A_{4t}$ -NeuroP, which caused substantial decreases of IL-6, TNF-α, MCP-1 mRNA. 4-F_{4t}-NeuroP, also reduced these mRNA levels. Part of the anti-inflammatory properties of the NEO-DHA investigated could be linked to their activation of PPARs.

In contrast, NEO-PUFAs from DHA have been more extensively studied than EPA derivatives, with a greater emphasis on NeuroPs function. Just like AA-derived IsoPs, the group of Morrow and Roberts, pioneers in the *in vivo* identification of IsoPs/NeuroPs, also demonstrated biological effects of NeuroPs, particularly the A4/J4-NeuroPs. These DHA-derived cyclopentenones act as mediators of inflammation in murine cell line macrophages [94] Similar to oxidized EPA NEO-PUFAs, 14-A4-NeuroP reduces the expression of pro-inflammatory mediators in these macrophages. This study confirmed that the inhibition of the NFκB pathway constitutes a mechanism of action for the NEO-PUFAs of EPA and DHA. Additionally, Majkova et al. demonstrated that A4/J4-NeuroPs down-regulated PCB77-induced MCP-1 expression and NRF2 activation in primary pulmonary endothelial cells [95].

In 2022, knowing the abundance of DHA in brain membrane phospholipids and this role in neurological functions Geng et al., examined the effects of 4-F4t-NeuroP on oxidative and inflammatory responses in microglial cells (primary and immortalized microglia (BV2)) treated with LPS $[96]$. They observed that the 4-F_{4t}-NeuroP attenuated LPS-induced production of ROS in both primary and BV2 cells. This NEO-PUFA also attenuated LPS-induced inflammation through suppressing NFκB-p65 and levels of iNOS and TNFα. In addition, 4-F4t-NeuroP attenuated LPS-induced mitochondrial membrane potential loss in BV2 cells. In summary, these findings with microglial cells demonstrated neuroprotective effects of this 4-F4t-NeuroP and shed light into the potential of nutraceutical therapy for neurodegenerative diseases.

The anti-atherogenic effects of ω-3 PUFAs, specifically EPA and DHA are well-established [97], yet the impact of dietary intake of fatty acid precursors on the quantity of newly synthesized NEO-PUFAs profiles remains unknown in specific tissues and animal models studied. Profiling such mediators could provide novel targets for future studies investigating the mechanisms of action of ω-3 fatty acids. Researchers have focused on the inflammatory process resulting from atherosclerosis and the potential role of NEO-PUFAs. However, the initial observation was conducted for F2-IsoPs. In 2007, the *in vivo* study published by Saraswathi [98] demonstrated that fish oil feeding decreased the formation of pro-inflammatory F_2 -isoP, markers of oxidative stress. The increased white adipose tissue lipid storage in fish oil–fed mice was associated with reduced lipid accumulation in liver and decreased atherosclerotic lesion area. Taken together, these data highlight the specific role of dietary fish oil–mediated improvement in systemic lipid homeostasis and atherosclerosis. This hypothesis has been more recently investigated for NEO-PUFAs from ω-3 fatty acids through an integrated approach combined with a DHA supplementation study in *LDLR^{−/−}* (low-density lipoprotein receptor) mice prone to atherosclerosis [25].

Special attention was paid to non-enzymatic NEO-PUFAs due to DHA's susceptibility to free radical-mediated peroxidation and the heightened oxidative stress associated with plaque formation. Atherosclerosis-prone mice (*LDLR^{-/−}*) received DHA over 20 weeks, displayed a dose-dependent reduction in atherosclerosis, triglyceridemia, and cholesterolemia. Moreover, Gladine et al. demonstrated in this atherosclerosis mice model (low-density lipoprotein receptor ^{-/-} or *LDLR*^{-/-}) fed with ω -6 or ω −3 PUFAs enriched diet that the liver content of F₄-NeuroPs was negatively correlated with atherosclerotic plaque extension but further experiments are required to confirm this correlation [25]. These results suggests that F4-NeuroPs could be potential biomarkers of DHA-associated atherosclerosis prevention.

4.2.2. Antiarrhythmic properties

At the cardiac level, few studies have focused on the effects of peroxidized NEO-PUFAs on cardiac pathophysiology. In recent decades, there has been extensive documentation of the cardioprotective effects conferred by a diet enriched in ω-3 PUFA, primarily attributed to EPA and DHA [99]. A significant prospective study highlighted that DHA and EPA supplementation notably reduces the incidence of sudden cardiac death in the months following a heart attack. This reduction has been attributed, at least in part, to a decrease in arrhythmias and systolic cardiac failure. Studies conducted on individual cardiac cells and animals have revealed that EPA and DHA can modulate the activity of ion channels, which are transmembrane proteins crucial for the heart's electrical activity. However, there is also speculation that NEO-PUFAs (from DHA and EPA) may contribute to these effects [100]. Indeed, research has demonstrated that some of DHA's impact on rat cardiac ion channels stems from a NEO-PUFAs.

Roy et al. have tested different F_4 -NeuroPs on isoprenaline-induced arrhythmias on murine isolated ventricular cardiomyocytes. Among them, the 4-F_{4t}-NeuroP displays anti-arrhythmic effects (IC₅₀ \approx 100 nM). The 4-F_{4t}-NeuroP, showed dose-dependent antiarrhythmic properties *in cellulo* and *in vivo* in permanent myocardial infarction mice [22] or after ischemia/reperfusion event in rats [23]. At the cellular level, the mechanism of action can be explained by the Ryanodine Receptor (RyR2)/FKBP12.6 complex stabilization. In conclusion, the researchers proposed that in oxidative stress conditions, such as ischemic events, 4-F4t-NeuroP is product and then responsible for the antiarrhythmic properties of ω-3 PUFAs.

4.2.3. Anti-proliferative properties

Another study by same team revealed that DHA possesses antiproliferative properties, which were inhibited by α-tocopherol (an antioxidant) and enhanced by the pro-oxidant hydrogen peroxide $(H₂O₂)$. This confirms that DHA needs to be oxidized to exert its antiproliferative effects $[101]$. They also found that certain F_{4t} -NeuroPs exhibited antiproliferative effects on the human breast cancer cell line (MDA-MB-231), while no inhibitory effect was observed on a human mammary epithelial normal cell line (MCF-10A). These results suggest that NeuroPs aims to enhance stability against degradation, increase cellular availability, and control the release of bioactive fatty acids based on their lipophilic properties. Building on this study and the encapsulation methods used, Abawi et al., in 2023, demonstrated that integrating F4-NeuroP into liposomes with a fluid character enabled selective targeting of the metastatic prostate cell line (PC-3) *versus* fibroblast controls. A significant decrease in viability was observed for the PC-3 cancer line in the presence of F4-NeuroP fluid liposomes, whereas rigid F4t-NeuroP liposomes did not alter PC-3 cell viability. These findings confirm the antiproliferative role of F_4 -NeuroP and suggest that liposomes encapsulating 4-F4t-NeuroP or other related molecules may serve as promising drug carriers based on membrane fluidity [102].

In their 2021 study, Bilodeau et al. studied the relationship between NEO-PUFAs and tumor growth. They conducted experiments using immune-competent eugonadal and castrated C57BL/6 male mice

injected with TRAMP-C2 prostate tumor cells, while being fed diets rich in either ω -3 or ω -6 PUFAs [103]. Their findings revealed a significant inverse correlation between EPA and its NEO-EPA (F_3 -NeuroPs) and tumor volume. Conversely, F_2 - IsoPs showed a positive correlation with tumor volume. Notably, NEO-DHA (F4-NeuroPs) exhibited a specific increase in TRAMP-C2 tumors of castrated mice compared to those of eugonadal mice. The study concluded that the observed reduction in tumor growth under the ω-3 diet regimen could be partially attributed to elevated levels of NEO-PUFAs.

4.2.4. Neuroprotective properties

Jamil et al. investigated the ability of $5-F_{3t}$ -IsoP to regulate glutamatergic neurotransmission [104]. This product could have significant pharmacological implications as glutamate serves as an excitatory neurotransmitter in several vertebrate retinal cells, as well as in ganglion cells. These researchers then examined the modulatory role of 5-*epi*-5--F3t-IsoP on potassium efflux associated with glutamate release from isolated bovine retina [104]. In this study, 5-epi-5-F_{3t}-IsoP attenuates potassium efflux in a dose-dependent manner through mechanisms that are partially dependent on the activation of so-called "EP1-type prostanoid receptors." These results demonstrate a beneficial effect of this NEO-PUFA in attenuating the release of an excitatory neurotransmitter, as well as slowing the progression of ocular neuropathic diseases. In 2020, Lee et al. reported for the first time that native $4-F_{4t}$ -NeuroP has a regulatory role in neurons for cell survival on human SH-SYSY neuroblastoma cells $[105]$. They showed that 4-F_{4t}-NeuroP can cross the blood brain barrier into rat brain tissue and alter brain ω-3 and ω-6 PUFA profiles, where anti-inflammatory and pro-resolvin lipid biomarkers were significantly elevated.

4.2.5. Pro-fertility properties

Recently, F4-NeuroPs showed a biological activity in sperm function and was able to induce capacitation via increasing AMPK phosphorylation, and its role at ryanodine receptors is currently being explored [58]. Later, the same group investigated the biological role of 4-F4t-NeuroP on sperm capacitation (sperm hyperactivation and acrosomal reaction) *in vitro* [106]. After incubating swim-up selected human sperm with 7 ng of 4- F_{4t} -NeuroP for 2 and 4 h, various parameters including motility, membrane mitochondrial potential, percentage of reacted acrosome, and chromatin integrity were evaluated. The results indicated an increase in progressive sperm motility and hyperactivation, suggesting that F_4 -NeuroPs could influence the physiological behavior of sperm. Mitochondrial functionality is closely linked to sperm quality. Finally, Moretti et al. reported that $4-F_{4t}$ -NeuroP impacts human sperm, particularly affecting sperm capacitation mediated once again by a ryanodine receptor. In conclusion, a specific concentration of 4-F4t-NeuroP in seminal fluid may induce sperm capacitation through the modulation of ion channel activity present in sperm cells. Considering that infertile men often struggle with capacitation, these molecules could potentially assist in sperm preparation *in vitro* for some patients, thereby enhancing the success rate of assisted fertilization procedures.

4.2.6. Other species

Recently, Roy et al. unveiled the presence of certain NEO-PUFAs (including NeuroPs and IsoPs) in the brain of rainbow trout, along with the modulation of brain lipid content by ω-3 PUFAs such as DHA and EPA in a farmed fish species [107]. They observed that NEO-DHA (but not NEO-EPA) increased in brain tissues of rainbow trout having a diet enriched in ω-3 PUFAs. In summary, this study represents the first documentation of NEO-PUFAs presence in fish brains and their regulation by dietary DHA and EPA levels, suggesting their potential as nutritional biomarkers but also potentiel biological activity in fish farmed. Given the known biological activities of ω-3 PUFAs in mammals brain but also trout brain [108], further investigations are needed to fully characterize the impact of NEO-PUFAs on brain function and physiological effects in trout model.

Fig. 5. Graphical abstract.

5. Conclusion

The free radical-induced autoxidation of PUFAs occurs in numerous pathophysiological conditions, ranging from cardiovascular disorders to cancers and neurodegenerative diseases (Fig. 5). Our understanding of thse processes and their role still needs to be improved. However, leveraging our expertise in organic chemistry, we can contribute significantly to both clinical and basic research endeavors by developing new synthetic approaches and providing samples for biological and analytical investigations. Several innovative methods for the chiral synthesis of NeuroPs and other isoprostanoids have emerged. Some of these synthesized products hold potential as diagnostic and prognostic markers for patients, necessitating precise and accurate measurement. It is imperative even if dreamy at the moment to thoroughly ascertain the contribution of each unique NeuroP to tissue and organ damage within the intricate network of signaling molecules and mediators. Despite the considerable advancements in this field, there remains much to uncover, underscoring the critical importance of collaborations between chemists and biologists. Such interdisciplinary partnerships are paramount for pushing the boundaries of knowledge and driving progress in unraveling the complexities of PUFAs peroxidation-related diseases but also could allow the identification of key molecules displaying beneficial properties.

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CRediT authorship contribution statement

Anna Abramova: Conceptualization, Methodology, Writing – original draft, Writing – review & editing. **Jamie Bride:** Conceptualization, Methodology, Writing – original draft, Writing – review $\&$ editing. **Camille Oger:** Visualization, Investigation, Writing – original draft.

Marie Demion: Visualization, Investigation, Writing – original draft. **Jean-Marie Galano:** Visualization, Investigation, Writing – original draft. **Thierry Durand:** Supervision, Visualization, Investigation, Validation, Writing – original draft, Writing – review $\&$ editing, Funding acquisition. **Jérôme Roy:** Supervision, Visualization, Investigation, Validation, Writing – original draft, Writing – review $\&$ editing, Funding acquisition.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- [1] Denham Harman, Aging: a theory based on free radical and radiation chemistry, J. Gerontol. 11 (1956) 298–300,<https://doi.org/10.1093/geronj/11.3.298>.
- [2] D. Giustarini, I. Dalle-Donne, D. Tsikas, R. Rossi, Oxidative stress and human diseases: origin, link, measurement, mechanisms, and biomarkers, Crit. Rev. Clin. Lab Sci. 46 (2009) 241–281, [https://doi.org/10.3109/10408360903142326.](https://doi.org/10.3109/10408360903142326)
- [3] J. Roy, J.-M. Galano, T. Durand, J.-Y. Le Guennec, J.C.-Y. Lee, Physiological role of reactive oxygen species as promoters of natural defenses, FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol. 31 (2017) 3729–3745, [https://doi.org/10.1096/](https://doi.org/10.1096/fj.201700170R) [fj.201700170R.](https://doi.org/10.1096/fj.201700170R)
- [4] J.-M. Galano, E. Mas, A. Barden, T.A. Mori, C. Signorini, C. De Felice, A. Barrett, C. Opere, E. Pinot, E. Schwedhelm, R. Benndorf, J. Roy, J.-Y. Le Guennec, C. Oger, T. Durand, Isoprostanes and neuroprostanes: total synthesis, biological activity and biomarkers of oxidative stress in humans, Prostag. Other Lipid Mediat. 107 (2013) 95–102, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.prostaglandins.2013.04.003) [prostaglandins.2013.04.003](https://doi.org/10.1016/j.prostaglandins.2013.04.003).
- [5] U. Jahn, J. Galano, T. Durand, Beyond prostaglandins—chemistry and biology of cyclic oxygenated metabolites formed by free-radical pathways from polyunsaturated fatty acids, Angew. Chem. Int. Ed. 47 (2008) 5894–5955, <https://doi.org/10.1002/anie.200705122>.
- [6] R.B. Zurier, Prostaglandins, in: Mediators Inflamm, Springer, Boston, MA, 1974, pp. 163-180, https://doi.org/10.1007/978-1-4684-0745
- [7] J.D. Morrow, K.E. Hill, R.F. Burk, T.M. Nammour, K.F. Badr, L.J. Roberts, A series of prostaglandin F2-like compounds are produced in vivo in humans by a noncyclooxygenase, free radical-catalyzed mechanism, Proc. Natl. Acad. Sci. U.S.A. 87 (1990) 9383–9387, [https://doi.org/10.1073/pnas.87.23.9383.](https://doi.org/10.1073/pnas.87.23.9383)
- [8] S. Parchmann, M.J. Mueller, Evidence for the Formation of dinor isoprostanes E1from α-linolenic acid in plants, J. Biol. Chem. 273 (1998) 32650–32655, [https://doi.org/10.1074/jbc.273.49.32650.](https://doi.org/10.1074/jbc.273.49.32650)
- [9] Joshua P. Fessel, L. Jackson Roberts, Isofurans: novel products of lipid peroxidation that define the occurrence of oxidant injury in settings of elevated oxygen tension, Antioxidants Redox Signal. 7 (2004), [https://doi.org/10.1089/](https://doi.org/10.1089/ars.2005.7.202) 005.7.202
- [10] M. VanRollins, R.L. Woltjer, H. Yin, J.D. Morrow, T.J. Montine, F2-Dihomoisoprostanes arise from free radical attack on adrenic acid, J. Lipid Res. 49 (2008) 995–1005, [https://doi.org/10.1194/jlr.M700503-JLR200.](https://doi.org/10.1194/jlr.M700503-JLR200)
- [11] D.F. Taber, J.D. Morrow, L. Jackson Roberts, A nomenclature system for the isoprostanes, Prostaglandins 53 (1997) 63–67, [https://doi.org/10.1016/S0090-](https://doi.org/10.1016/S0090-6980(97)00005-1) [6980\(97\)00005-1](https://doi.org/10.1016/S0090-6980(97)00005-1).
- [12] J. Rokach, S.P. Khanapure, S.-W. Hwang, M. Adiyaman, J.A. Lawson, G. A. FitzGerald, Nomenclature of isoprostanes: a proposal, Prostaglandins 54 (1997) 853–873, [https://doi.org/10.1016/S0090-6980\(97\)00184-6](https://doi.org/10.1016/S0090-6980(97)00184-6).
- [13] J. Rokach, S. Kim, S. Bellone, J.A. Lawson, D. Praticò, W.S. Powell, G. A. FitzGerald, Total synthesis of isoprostanes: discovery and quantitation in biological systems, Chem. Phys. Lipids 128 (2004) 35–56, [https://doi.org/](https://doi.org/10.1016/j.chemphyslip.2003.09.011) [10.1016/j.chemphyslip.2003.09.011.](https://doi.org/10.1016/j.chemphyslip.2003.09.011)
- [14] L.J. Roberts, T.J. Montine, W.R. Markesbery, A.R. Tapper, P. Hardy, S. Chemtob, W.D. Dettbarn, J.D. Morrow, Formation of isoprostane-like compounds (neuroprostanes) in vivo from docosahexaenoic acid, J. Biol. Chem. 273 (1998) 13605–13612, [https://doi.org/10.1074/jbc.273.22.13605.](https://doi.org/10.1074/jbc.273.22.13605)
- [15] H. Yin, C.M. Havrilla, L. Gao, J.D. Morrow, N.A. Porter, Mechanisms for the Formation of isoprostane endoperoxides from arachidonic acid, J. Biol. Chem. 278 (2003) 16720–16725,<https://doi.org/10.1074/jbc.M300604200>.
- [16] H. Yin, E.S. Musiek, L. Gao, N.A. Porter, J.D. Morrow, Regiochemistry of neuroprostanes generated from the peroxidation of docosahexaenoic acid in vitro and in vivo, J. Biol. Chem. 280 (2005) 26600–26611, [https://doi.org/10.1074/](https://doi.org/10.1074/jbc.M503088200) [jbc.M503088200](https://doi.org/10.1074/jbc.M503088200).
- [17] J.-M. Galano, C. Oger, V. Bultel-Poncé, G. Reversat, A. Guy, J. Vercauteren, [C. Vigor, T. Durand, F3-Isoprostanes and F4-Neuroprostanes: Non-enzymatic](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref17) [Cyclic Oxygenated Metabolites of Omega-3 Polyunsaturated Fatty Acids:](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref17) [Biomarkers and Bioactive Lipids, 2017](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref17).
- [18] J. Collado-González, T. Durand, F. Ferreres, S. Medina, A. Torrecillas, Á. Gil-Izquierdo, Phytoprostanes, Lipid Technol. 27 (2015) 127–130, [https://doi.org/](https://doi.org/10.1002/lite.201500020) [10.1002/lite.201500020.](https://doi.org/10.1002/lite.201500020)
- [19] [M. Singh, Essential fatty acids, DHA and human brain, Indian J. Pediatr. 72](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref19) [\(2005\) 239](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref19)–242.
- [20] W.J. Lukiw, N.G. Bazan, Docosahexaenoic acid and the aging brain, J. Nutr. 138 (2008) 2510–2514, <https://doi.org/10.3945/jn.108.096016>.
- [21] [C.Y. Chang, J.Y. Chen, Essential Fatty Acids and Human Brain, 2009,](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref21) pp. 231–[241. PMID 20329590 18.](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref21)
- [22] J. Roy, C. Oger, J. Thireau, J. Roussel, O. Mercier-Touzet, D. Faure, E. Pinot, C. Farah, D.F. Taber, J.-P. Cristol, J.C.Y. Lee, A. Lacampagne, J.-M. Galano, T. Durand, J.-Y. Le Guennec, Nonenzymatic lipid mediators, neuroprostanes, exert the antiarrhythmic properties of docosahexaenoic acid, Free Radic. Biol. Med. 86 (2015) 269–278, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2015.04.014) [freeradbiomed.2015.04.014](https://doi.org/10.1016/j.freeradbiomed.2015.04.014).
- [23] J. Roy, J. Fauconnier, C. Oger, C. Farah, C. Angebault-Prouteau, J. Thireau, P. Bideaux, V. Scheuermann, V. Bultel-Poncé, M. Demion, J.-M. Galano, T. Durand, J.C.-Y. Lee, J.-Y. Le Guennec, Non-enzymatic oxidized metabolite of DHA, 4(RS)-4-F4t-neuroprostane protects the heart against reperfusion injury, Free Radic. Biol. Med. 102 (2017) 229–239, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2016.12.005) [freeradbiomed.2016.12.005](https://doi.org/10.1016/j.freeradbiomed.2016.12.005).
- [24] R. Bosviel, L. Joumard-Cubizolles, G. Chinetti-Gbaguidi, D. Bayle, C. Copin, N. Hennuyer, I. Duplan, B. Staels, G. Zanoni, A. Porta, L. Balas, J.-M. Galano, C. Oger, A. Mazur, T. Durand, C. Gladine, DHA-derived oxylipins, neuroprostanes and protectins, differentially and dose-dependently modulate the inflammatory response in human macrophages: putative mechanisms through PPAR activation, Free Radic. Biol. Med. 103 (2017) 146–154, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2016.12.018) [freeradbiomed.2016.12.018](https://doi.org/10.1016/j.freeradbiomed.2016.12.018).
- [25] C. Gladine, J.W. Newman, T. Durand, T.L. Pedersen, J.-M. Galano, C. Demougeot, O. Berdeaux, E. Pujos-Guillot, A. Mazur, B. Comte, Lipid profiling following intake of the omega 3 fatty acid DHA identifies the peroxidized metabolites F4 neuroprostanes as the best predictors of atherosclerosis prevention, PLoS One 9 (2014) e89393, [https://doi.org/10.1371/journal.pone.0089393.](https://doi.org/10.1371/journal.pone.0089393)
- [26] C. Signorini, C. De Felice, T. Durand, J.-M. Galano, C. Oger, S. Leoncini, L. Ciccoli, M. Carone, M. Ulivelli, C. Manna, A. Cortelazzo, J.C.-Y. Lee, J. Hayek, Relevance of 4-F4t-neuroprostane and 10-F4t-neuroprostane to neurological diseases, Free Radic. Biol. Med. 115 (2018) 278–287, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2017.12.009) [freeradbiomed.2017.12.009](https://doi.org/10.1016/j.freeradbiomed.2017.12.009).
- [27] J.-M. Galano, J.C.-Y. Lee, C. Gladine, B. Comte, J.-Y. Le Guennec, C. Oger, T. Durand, Non-enzymatic cyclic oxygenated metabolites of adrenic, docosahexaenoic, eicosapentaenoic and α-linolenic acids; bioactivities and potential use as biomarkers, Biochim. Biophys. Acta BBA - Mol. Cell Biol. Lipids 1851 (2015) 446–455, <https://doi.org/10.1016/j.bbalip.2014.11.004>.
- [28] J.-M. Galano, J. Roy, T. Durand, J.C.-Y. Lee, J.-Y. Le Guennec, C. Oger, M. Demion, Biological activities of non-enzymatic oxygenated metabolites of polyunsaturated fatty acids (NEO-PUFAs) derived from EPA and DHA: new antiarrhythmic compounds? Mol. Aspect. Med. 64 (2018) 161–168, [https://doi.org/](https://doi.org/10.1016/j.mam.2018.03.003) [10.1016/j.mam.2018.03.003.](https://doi.org/10.1016/j.mam.2018.03.003)
- [29] C. Signorini, C. De Felice, J.-M. Galano, C. Oger, S. Leoncini, A. Cortelazzo, L. Ciccoli, T. Durand, J. Hayek, J. Lee, Isoprostanoids in clinical and experimental

neurological disease models, Antioxidants 7 (2018) 88, [https://doi.org/10.3390/](https://doi.org/10.3390/antiox7070088) $\arctan 707008$

- [30] O.S. Ahmed, J.-M. Galano, T. Pavlickova, J. Revol-Cavalier, C. Vigor, J.C.-Y. Lee, C. Oger, T. Durand, Moving forward with isoprostanes, neuroprostanes and phytoprostanes: where are we now? Essays Biochem. 64 (2020) 463–484, [https://](https://doi.org/10.1042/EBC20190096) doi.org/10.1042/EBC20190096.
- [31] D.F. Taber, P.G. Reddy, K.O. Arneson, A potential route to neuroprostanes and isoprostanes: preparation of the four enantiomerically pure diastereomers of 13- F4t-neuroP, J. Org. Chem. 73 (2008) 3467–3474, [https://doi.org/10.1021/](https://doi.org/10.1021/jo702600v) [jo702600v](https://doi.org/10.1021/jo702600v).
- [32] G. Zanoni, E.M. Brunoldi, A. Porta, G. Vidari, Asymmetric synthesis of 14-A _{4t} -neuroprostane: hunting for a suitable biomarker for neurodegenerative diseases, J. Org. Chem. 72 (2007) 9698-9703, https://doi.org/10.1021/jo701719.
- [33] A. Porta, M. Pasi, E. Brunoldi, G. Zanoni, G. Vidari, Biology and chemistry of neuroprostanes. First total synthesis of 17-A4-NeuroP: Validation of a convergent strategy to a number of cyclopentenone neuroprostanes, Chem. Phys. Lipids 174 (2013) 64–74, [https://doi.org/10.1016/j.chemphyslip.2013.07.002.](https://doi.org/10.1016/j.chemphyslip.2013.07.002)
- [34] L.G. Quan, J.K. Cha, Recent advances in the stereoselective synthesis of isoprostanes and neuroprostanes, Chem. Phys. Lipids 128 (2004) 3–14, [https://](https://doi.org/10.1016/j.chemphyslip.2003.10.005) [doi.org/10.1016/j.chemphyslip.2003.10.005.](https://doi.org/10.1016/j.chemphyslip.2003.10.005)
- [35] A. Roland, T. Durand, B. Rondot, J.-P. Vidal, J.-C. Rossi, A practical asymmetric synthesis of ent-12-epi-PGF2α methyl ester, Bull. Soc. Chim. Fr. 133 (1996) 1149–1154, <https://doi.org/10.1002/chin.199718266>.
- [36] A. Roland, T. Durand, B. Rondot, J.-P. Vidal, J.-C. Rossi, A practical asymmetric synthesis of ent-12-epi-PGF2α methyl ester, Bull. Soc. Chim. Fr. 133 (1996) 1149–1154, <https://doi.org/10.1002/chin.199718266>.
- [37] E. Pinot, A. Guy, A. Fournial, L. Balas, J.-C. Rossi, T. Durand, Total synthesis of the four enantiomerically pure diasteroisomers of the phytoprostanes E_1 type II and of the 15-E $_{2t}$ -isoprostanes, J. Org. Chem. 73 (2008) 3063–3069, https://doi. [org/10.1021/jo702455g](https://doi.org/10.1021/jo702455g).
- [38] C. Oger, Y. Brinkmann, S. Bouazzaoui, T. Durand, J.-M. Galano, Stereocontrolled access to isoprostanes via a bicyclo[3.3.0]octene framework, Org. Lett. 10 (2008) 5087–5090, [https://doi.org/10.1021/ol802104z.](https://doi.org/10.1021/ol802104z)
- [39] A. Guy, J. Merad, T. Degrange, G. Reversat, V. Bultel-Poncé, T. Durand, J.-M. Galano, C. Oger, Total synthesis of DHA and DPAn-3 non-enzymatic oxylipins, Synthesis 54 (2022) 1431–1445, [https://doi.org/10.1055/a-1654-4111.](https://doi.org/10.1055/a-1654-4111)
- [40] C. Oger, V. Bultel-Poncé, A. Guy, L. Balas, J.-C. Rossi, T. Durand, J.-M. Galano, The handy use of Brown's P2-Ni catalyst for a skipped diyne deuteration: application to the synthesis of a [D4]-labeled F4t-neuroprostane, Chem. Weinh. Bergstr. Ger. 16 (2010) 13976–13980, [https://doi.org/10.1002/](https://doi.org/10.1002/chem.201002304) [chem.201002304](https://doi.org/10.1002/chem.201002304).
- [41] T. Pavlíčková, V. Bultel-Poncé, A. Guy, A. Rocher, G. Reversat, C. Vigor, T. Durand, J.-M. Galano, U. Jahn, C. Oger, First total syntheses of novel nonenzymatic polyunsaturated fatty acid metabolites and their identification in edible oils, Chem. Weinh. Bergstr. Ger. 26 (2020) 10090-10098, https://doi.org/ [10.1002/chem.202002138.](https://doi.org/10.1002/chem.202002138)
- [42] C. Oger, Z. Marton, Y. Brinkmann, V. Bultel-Poncé, T. Durand, M. Graber, J.-M. Galano, Lipase-catalyzed regioselective monoacetylation of unsymmetrical 1,5-primary diols, J. Org. Chem. 75 (2010) 1892–1897, [https://doi.org/10.1021/](https://doi.org/10.1021/jo902541c) $0902541c$.
- [43] Y. Brinkmann, C. Oger, A. Guy, T. Durand, J.-M. Galano, Total synthesis of 15-D _{2t} - and 15- *epi* -15-E 2t -isoprostanes, J. Org. Chem. 75 (2010) 2411–2414, [https://](https://doi.org/10.1021/jo1000274) doi.org/10.1021/jo1000274.
- [44] A. Guy, C. Oger, J. Heppekausen, C. Signorini, C. De Felice, A. Fürstner, T. Durand, J. Galano, Oxygenated metabolites of *n* -3 polyunsaturated fatty acids as potential oxidative stress biomarkers: total synthesis of 8-F $_{\rm 3t}$ -IsoP, 10-F $_{\rm 4t}$ -NeuroP and [D 4]-10-F 4t -NeuroP, Chem. Eur J. 20 (2014) 6374–6380, [https://](https://doi.org/10.1002/chem.201400380) doi.org/10.1002/chem.201400380.
- [45] C. Vigor, L. Balas, A. Guy, V. Bultel-Poncé, G. Reversat, J. Galano, T. Durand, C. Oger, Isoprostanoids, isofuranoids and isoketals – from synthesis to lipidomics, Eur. J. Org. Chem. 2022 (2022) e202101523, [https://doi.org/10.1002/](https://doi.org/10.1002/ejoc.202101523) [ejoc.202101523.](https://doi.org/10.1002/ejoc.202101523)
- [46] C. Oger, V. Bultel-Poncé, A. Guy, T. Durand, J. Galano, Total synthesis of isoprostanes derived from adrenic acid and EPA, Eur. J. Org. Chem. 2012 (2012) 2621–2634, <https://doi.org/10.1002/ejoc.201200070>.
- [47] L.J. Roberts, G.L. Milne, Isoprostanes, J. Lipid Res. 50 (Suppl) (2009) S219–S223, <https://doi.org/10.1194/jlr.R800037-JLR200>.
- [48] D.B. Clayton, H.A. Stephany, C.B. Ching, S.A. Rahman, S.T. Tanaka, J.C. Thomas, J.C. Pope, M.C. Adams, J.W. Brock, P.E. Clark, S.W. Hayward, R.J. Matusik, G. L. Milne, F2-isoprostanes as a biomarker of oxidative stress in the mouse bladder, J. Urol. 191 (2014) 1597–1601, [https://doi.org/10.1016/j.juro.2013.09.027.](https://doi.org/10.1016/j.juro.2013.09.027)
- [49] G.L. Milne, H. Yin, K.D. Hardy, S.S. Davies, L.J. Roberts, Isoprostane generation and function, Chem. Rev. 111 (2011) 5973–5996, [https://doi.org/10.1021/](https://doi.org/10.1021/cr200160h) [cr200160h.](https://doi.org/10.1021/cr200160h)
- [50] M.B. Kadiiska, B.C. Gladen, D.D. Baird, D. Germolec, L.B. Graham, C.E. Parker, A. Nyska, J.T. Wachsman, B.N. Ames, S. Basu, N. Brot, G.A. Fitzgerald, R. A. Floyd, M. George, J.W. Heinecke, G.E. Hatch, K. Hensley, J.A. Lawson, L. J. Marnett, J.D. Morrow, D.M. Murray, J. Plastaras, L.J. Roberts, J. Rokach, M. K. Shigenaga, R.S. Sohal, J. Sun, R.R. Tice, D.H. Van Thiel, D. Wellner, P. B. Walter, K.B. Tomer, R.P. Mason, J.C. Barrett, Biomarkers of oxidative stress study II: are oxidation products of lipids, proteins, and DNA markers of CCl4 poisoning? Free Radic. Biol. Med. 38 (2005) 698–710, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2004.09.017) reeradbiomed.2004.09.017
- [51] J.D. Morrow, J.A. Awad, H.J. Boss, I.A. Blair, L.J. Roberts, Non-cyclooxygenasederived prostanoids (F2-isoprostanes) are formed in situ on phospholipids, Proc.

Natl. Acad. Sci. U.S.A. 89 (1992) 10721–10725, [https://doi.org/10.1073/](https://doi.org/10.1073/pnas.89.22.10721) [pnas.89.22.10721.](https://doi.org/10.1073/pnas.89.22.10721)

- [52] D. Acevedo-León, S.Á. Gómez-Abril, P. Sanz-García, N. Estañ-Capell, C. Bañuls, G. Sáez, The role of oxidative stress, tumor and inflammatory markers in colorectal cancer patients: a one-year follow-up study, Redox Biol. 62 (2023) 102662, <https://doi.org/10.1016/j.redox.2023.102662>.
- [53] C. De Felice, F. Della Ragione, C. Signorini, S. Leoncini, A. Pecorelli, L. Ciccoli, F. Scalabrì, F. Marracino, M. Madonna, G. Belmonte, L. Ricceri, B. De Filippis, G. Laviola, G. Valacchi, T. Durand, J.-M. Galano, C. Oger, A. Guy, V. Bultel-Poncé, J. Guy, S. Filosa, J. Hayek, M. D'Esposito, Oxidative brain damage in Mecp2 mutant murine models of Rett syndrome, Neurobiol. Dis. 68 (2014) 66–77, [https://doi.org/10.1016/j.nbd.2014.04.006.](https://doi.org/10.1016/j.nbd.2014.04.006)
- [54] S. Leoncini, C. Signorini, L. Boasiako, V. Scandurra, J. Hayek, L. Ciccoli, M. Rossi, R. Canitano, C. De Felice, Breathing abnormalities during sleep and wakefulness in Rett syndrome: clinical relevance and paradoxical relationship with circulating pro-oxidant markers, Front. Neurol. 13 (2022) 833239, https://doi.org/10.338 eur.2022.833239
- [55] C. De Felice, S. Maffei, C. Signorini, S. Leoncini, S. Lunghetti, G. Valacchi, M. D'Esposito, S. Filosa, F. Della Ragione, G. Butera, R. Favilli, L. Ciccoli, J. Hayek, Subclinical myocardial dysfunction in Rett syndrome, Eur. Heart J. Cardiovasc. Imaging 13 (2012) 339–345, [https://doi.org/10.1093/ejechocard/](https://doi.org/10.1093/ejechocard/jer256) [jer256](https://doi.org/10.1093/ejechocard/jer256).
- [56] A. Cortelazzo, C. De Felice, R. Guerranti, C. Signorini, S. Leoncini, A. Pecorelli, F. Scalabrì, M. Madonna, S. Filosa, C. Della Giovampaola, A. Capone, T. Durand, C. Mirasole, L. Zolla, G. Valacchi, L. Ciccoli, J. Guy, M. D'Esposito, J. Hayek, Abnormal N-glycosylation pattern for brain nucleotide pyrophosphatase-5 (NPP-5) in Mecp2-mutant murine models of Rett syndrome, Neurosci. Res. 105 (2016) 28–34, <https://doi.org/10.1016/j.neures.2015.10.002>.
- [57] C. De Felice, C. Signorini, T. Durand, C. Oger, A. Guy, V. Bultel-Poncé, J.-M. Galano, L. Ciccoli, S. Leoncini, M. D'Esposito, S. Filosa, A. Pecorelli, G. Valacchi, J. Hayek, F2-dihomo-isoprostanes as potential early biomarkers of lipid oxidative damage in Rett syndrome, J. Lipid Res. 52 (2011) 2287–2297, <https://doi.org/10.1194/jlr.P017798>.
- [58] C. Signorini, E. Moretti, D. Noto, S. Mattioli, C. Castellini, N.A. Pascarelli, T. Durand, C. Oger, J.-M. Galano, C. De Felice, J.C.-Y. Lee, G. Collodel, F4- Neuroprostanes: a role in sperm capacitation, Life. Basel. Switz. 11 (2021) 655, [https://doi.org/10.3390/life11070655.](https://doi.org/10.3390/life11070655)
- [59] C. Signorini, V. Cardile, G. Pannuzzo, A.C.E. Graziano, T. Durand, J.-M. Galano, C. Oger, S. Leoncini, A. Cortelazzo, J.C.-Y. Lee, J. Hayek, C. De Felice, Increased isoprostanoid levels in brain from murine model of Krabbe disease - relevance of isoprostanes, dihomo-isoprostanes and neuroprostanes to disease severity, Free Radic. Biol. Med. 139 (2019) 46–54, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2019.05.014) eradbiomed.2019.05.014.
- [60] C. Signorini, C. De Felice, T. Durand, J.-M. Galano, C. Oger, S. Leoncini, J. Hayek, J.C.-Y. Lee, T.C. Lund, P.J. Orchard, Isoprostanoid plasma levels are relevant to cerebral adrenoleukodystrophy disease, Life. Basel. Switz. 12 (2022) 146, [https://](https://doi.org/10.3390/life12020146) [doi.org/10.3390/life12020146.](https://doi.org/10.3390/life12020146)
- [61] T.B. Corcoran, E. Mas, A.E. Barden, T. Durand, J.-M. Galano, L.J. Roberts, M. Phillips, K.M. Ho, T.A. Mori, Are isofurans and neuroprostanes increased after subarachnoid hemorrhage and traumatic brain injury? Antioxidants Redox Signal. 15 (2011) 2663-2667, https://doi.org/10.1089/ars.2011.412
- [62] Y.-P. Hsieh, C.-L. Lin, A.-L. Shiue, H. Yin, J.D. Morrow, J.-C. Hsu, T.-C. Hsieh, H.- J. Wei, H.-C. Yen, Correlation of F4-neuroprostanes levels in cerebrospinal fluid with outcome of aneurysmal subarachnoid hemorrhage in humans, Free Radic. Biol. Med. 47 (2009) 814–824, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2009.06.026) [freeradbiomed.2009.06.026](https://doi.org/10.1016/j.freeradbiomed.2009.06.026).
- [63] S.E. Farias, M. Basselin, L. Chang, K.A. Heidenreich, S.I. Rapoport, R.C. Murphy, Formation of eicosanoids, E2/D2 isoprostanes, and docosanoids following decapitation-induced ischemia, measured in high-energy-microwaved rat brain, J. Lipid Res. 49 (2008) 1990–2000, [https://doi.org/10.1194/jlr.M800200-](https://doi.org/10.1194/jlr.M800200-JLR200) [JLR200.](https://doi.org/10.1194/jlr.M800200-JLR200)
- [64] Y.-C. Kao, P.-C. Ho, Y.-K. Tu, I.-M. Jou, K.-J. Tsai, Lipids and alzheimer's disease, Int. J. Mol. Sci. 21 (2020) 1505, <https://doi.org/10.3390/ijms21041505>.
- [65] M.A. Bradley-Whitman, M.A. Lovell, Biomarkers of lipid peroxidation in Alzheimer disease (AD): an update, Arch. Toxicol. 89 (2015) 1035–1044, [https://](https://doi.org/10.1007/s00204-015-1517-6) doi.org/10.1007/s00204-015-1517-6.
- [66] A. García-Blanco, C. Pena-Bautista, ˜ C. Oger, C. Vigor, J.-M. Galano, T. Durand, N. Martín-Ibáñez, M. Baquero, M. Vento, C. Cháfer-Pericás, Reliable determination of new lipid peroxidation compounds as potential early Alzheimer Disease biomarkers, Talanta 184 (2018) 193–201, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.talanta.2018.03.002) [talanta.2018.03.002](https://doi.org/10.1016/j.talanta.2018.03.002).
- [67] M.M. Cascant-Vilaplana, Á. Sánchez-Illana, J.D. Piñeiro-Ramos, R. Llorens-Salvador, G. Quintás, C. Oger, J.-M. Galano, C. Vigor, T. Durand, J. Kuligowski, M. Vento, HYPOTOP study group, do levels of lipid peroxidation biomarkers reflect the degree of brain injury in newborns? Antioxidants Redox Signal. 35 (2021) 1467–1475, [https://doi.org/10.1089/ars.2021.0168.](https://doi.org/10.1089/ars.2021.0168)
- [68] T.M. Raffay, J.M. Di Fiore, Z. Chen, Á. Sánchez-Illana, M. Vento, J.D. Piñeiro-Ramos, J. Kuligowski, R.J. Martin, C. Tatsuoka, N.M. Minich, P.M. MacFarlane, A. M. Hibbs, Hypoxemia events in preterm neonates are associated with urine oxidative biomarkers, Pediatr. Res. 94 (2023) 1444–1450, [https://doi.org/](https://doi.org/10.1038/s41390-023-02646-7) [10.1038/s41390-023-02646-7](https://doi.org/10.1038/s41390-023-02646-7).
- [69] A.E. Barden, T.B. Corcoran, E. Mas, T. Durand, J.-M. Galano, L.J. Roberts, M. Paech, N.A. Muchatuta, M. Phillips, T.A. Mori, Is there a role for isofurans and neuroprostanes in pre-eclampsia and normal pregnancy? Antioxidants Redox Signal. 16 (2012) 165–169, [https://doi.org/10.1089/ars.2011.4214.](https://doi.org/10.1089/ars.2011.4214)
- [70] A.E. Barden, T.A. Mori, J.A. Dunstan, A.L. Taylor, C.A. Thornton, K.D. Croft, L. J. Beilin, S.L. Prescott, Fish oil supplementation in pregnancy lowers F2 isoprostanes in neonates at high risk of atopy, Free Radic. Res. 38 (2004) 233–239, <https://doi.org/10.1080/10715760310001656722>.
- [71] C.S. Ferreira, G.D.A. Pinto, D.L. Reis, C. Vigor, V.A. Goes, D. de A.B. Guimarães, D.B. Mucci, L. Belcastro, M.A. Saraiva, C. Oger, J.-M. Galano, F.L.C. Sardinha, A. G. Torres, T. Durand, G.J. Burton, T. El-Bacha, Placental F4-Neuroprostanes and F2-Isoprostanes are altered in gestational diabetes mellitus and maternal obesity, Prostaglandins Leukot. Essent. Fatty Acids 189 (2023) 102529, [https://doi.org/](https://doi.org/10.1016/j.plefa.2022.102529) [10.1016/j.plefa.2022.102529.](https://doi.org/10.1016/j.plefa.2022.102529)
- [72] L. Marseglia, S. Manti, G. D'Angelo, A. Nicotera, E. Parisi, G. Di Rosa, E. Gitto, T. Arrigo, Oxidative stress in obesity: a critical component in human diseases, Int. J. Mol. Sci. 16 (2014) 378–400, [https://doi.org/10.3390/ijms16010378.](https://doi.org/10.3390/ijms16010378)
- [73] P. Sarajlic, C. Vigor, A. Avignon, B. Zhou, C. Oger, J.-M. Galano, T. Durand, A. Sultan, M. Bäck, Omega-3 to omega-6 fatty acid oxidation ratio as a novel inflammation resolution marker for metabolic complications in obesity, Nutr. Metab. Cardiovasc. Dis. NMCD 33 (2023) 1206–1213, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.numecd.2023.03.007) [numecd.2023.03.007](https://doi.org/10.1016/j.numecd.2023.03.007).
- [74] D.A. Barocas, S. Motley, M.S. Cookson, S.S. Chang, D.F. Penson, Q. Dai, G. Milne, L.J. Roberts, J. Morrow, R.S. Concepcion, J.A. Smith, J.H. Fowke, Oxidative stress measured by urine F2-isoprostane level is associated with prostate cancer, J. Urol. 185 (2011) 2102–2107, [https://doi.org/10.1016/j.juro.2011.02.020.](https://doi.org/10.1016/j.juro.2011.02.020)
- [75] M. Brys, A. Morel, E. Forma, A. Krzeslak, J. Wilkosz, W. Rozanski, B. Olas, Relationship of urinary isoprostanes to prostate cancer occurrence, Mol. Cell. Biochem. 372 (2013) 149–153, [https://doi.org/10.1007/s11010-012-1455-z.](https://doi.org/10.1007/s11010-012-1455-z)
- [76] I. De las Heras-Gómez, S. Medina, T. Casas-Pina, L. Marín-Soler, A. Tomás, P. Martínez-Hernández, C. Oger, J.-M. Galano, T. Durand, L. Jimeno, S. Llorente, E. Lozoya, F. Ferreres, Á. Gil-Izquierdo, Potential applications of lipid peroxidation products – F4-neuroprostanes, F3-neuroprostanesn-6 DPA, F2 dihomo-isoprostanes and F2-isoprostanes - in the evaluation of the allograft function in renal transplantation, Free Radic. Biol. Med. 104 (2017) 178–184, <https://doi.org/10.1016/j.freeradbiomed.2017.01.019>.
- [77] D. Praticò, L. Iuliano, A. Mauriello, L. Spagnoli, J.A. Lawson, J. Rokach, J. Maclouf, F. Violi, G.A. FitzGerald, Localization of distinct F2-isoprostanes in human atherosclerotic lesions, J. Clin. Invest. 100 (1997) 2028–2034, [https://](https://doi.org/10.1172/JCI119735) [doi.org/10.1172/JCI119735.](https://doi.org/10.1172/JCI119735)
- [78] B. Lefort, R. Gélinas, A. Forest, B. Bouchard, C. Daneault, I. Robillard Frayne, J. Roy, C. Oger, K. Greffard, J.-M. Galano, T. Durand, F. Labarthe, J.-F. Bilodeau, M. Ruiz, C. Des Rosiers, Remodeling of lipid landscape in high fat fed very-long chain acyl-CoA dehydrogenase null mice favors pro-arrhythmic polyunsaturated fatty acids and their downstream metabolites, Biochim. Biophys. Acta, Mol. Basis Dis. 1869 (2023) 166843, <https://doi.org/10.1016/j.bbadis.2023.166843>.
- [79] R.J. Mark, K.S. Fuson, P.C. May, Characterization of 8-epiprostaglandin F2alpha as a marker of amyloid beta-peptide-induced oxidative damage, J. Neurochem. 72
(1999) 1146–1153, <u>https://doi.org/10.1046/j</u>.1471-4159.1999.0721146.x.
- [80] A. Steensberg, J. Morrow, A.D. Toft, H. Bruunsgaard, B.K. Pedersen, Prolonged exercise, lymphocyte apoptosis and F2-isoprostanes, Eur. J. Appl. Physiol. 87 (2002) 38–42, [https://doi.org/10.1007/s00421-002-0584-6.](https://doi.org/10.1007/s00421-002-0584-6)
- [81] D. Labb´e, M.-A. Teranishi, A. Hess, W. Bloch, O. Michel, Activation of caspase-3 is associated with oxidative stress in the hydropic Guinea pig cochlea, Hear. Res. 202 (2005) 21–27, <https://doi.org/10.1016/j.heares.2004.10.002>.
- [82] J.-L. Cracowski, T. Durand, Cardiovascular pharmacology and physiology of the isoprostanes, Fundam. Clin. Pharmacol. 20 (2006) 417–427, [https://doi.org/](https://doi.org/10.1111/j.1472-8206.2006.00435.x) [10.1111/j.1472-8206.2006.00435.x.](https://doi.org/10.1111/j.1472-8206.2006.00435.x)
- [83] X. Hou, F. Gobeil, K. Peri, G. Speranza, A.M. Marrache, P. Lachapelle, J. Roberts, D.R. Varma, S. Chemtob, E.F. Ellis, Augmented vasoconstriction and thromboxane formation by 15-F(2t)-isoprostane (8-iso-prostaglandin F(2alpha)) in immature pig periventricular brain microvessels, Stroke 31 (2000) 516–524, [https://doi.org/10.1161/01.str.31.2.516.](https://doi.org/10.1161/01.str.31.2.516) ; discussion 525.
- [84] J.D. Morrow, The isoprostanes unique products of arachidonate peroxidation: their role as mediators of oxidant stress, Curr. Pharmaceut. Des. 12 (2006) 895–902, <https://doi.org/10.2174/138161206776055985>.
- [85] R.A. Benndorf, E. Schwedhelm, A. Gnann, R. Taheri, G. Kom, M. Didié, A. Steenpass, S. Ergün, R.H. Böger, Isoprostanes inhibit vascular endothelial growth factor-induced endothelial cell migration, tube formation, and cardiac vessel sprouting in vitro, as well as angiogenesis in vivo via activation of the thromboxane A(2) receptor: a potential link between oxidative stress and impaired angiogenesis, Circ. Res. 103 (2008) 1037–1046, [https://doi.org/](https://doi.org/10.1161/CIRCRESAHA.108.184036) [10.1161/CIRCRESAHA.108.184036](https://doi.org/10.1161/CIRCRESAHA.108.184036).
- [86] J. Bauer, A. Ripperger, S. Frantz, S. Ergün, E. Schwedhelm, R.A. Benndorf, Pathophysiology of isoprostanes in the cardiovascular system: implications of isoprostane-mediated thromboxane A2 receptor activation, Br. J. Pharmacol. 171 (2014) 3115-3131, https://doi.org/10.1111/bph.1267
- [87] E.S. Musiek, L. Gao, G.L. Milne, W. Han, M.B. Everhart, D. Wang, M.G. Backlund, R.N. DuBois, G. Zanoni, G. Vidari, T.S. Blackwell, J.D. Morrow, Cyclopentenone isoprostanes inhibit the inflammatory response in macrophages, J. Biol. Chem. 280 (2005) 35562–35570,<https://doi.org/10.1074/jbc.M504785200>.
- [88] M. Lappas, M. Permezel, S.J. Holdsworth, G. Zanoni, A. Porta, G.E. Rice, Antiinflammatory effects of the cyclopentenone isoprostane 15-A(2)-IsoP in human gestational tissues, Free Radic. Biol. Med. 42 (2007) 1791–1796, [https://](https://doi.org/10.1016/j.freeradbiomed.2007.03.009) doi.org/10.1016/j.freeradbiomed.2007.03.009.
- [89] S. Sethi, A.Y. Eastman, J.W. Eaton, Inhibition of phagocyte-endothelium interactions by oxidized fatty acids: a natural anti-inflammatory mechanism? J. Lab. Clin. Med. 128 (1996) 27–38, [https://doi.org/10.1016/s0022-2143\(96\)](https://doi.org/10.1016/s0022-2143(96)90111-0) [90111-0.](https://doi.org/10.1016/s0022-2143(96)90111-0)
- [90] D. Praticò, E.M. Smyth, F. Violi, G.A. FitzGerald, Local amplification of platelet function by 8-Epi prostaglandin F2alpha is not mediated by thromboxane receptor isoforms, J. Biol. Chem. 271 (1996) 14916–14924, [https://doi.org/](https://doi.org/10.1074/jbc.271.25.14916) [10.1074/jbc.271.25.14916](https://doi.org/10.1074/jbc.271.25.14916).
- [91] A. Mishra, A. Chaudhary, S. Sethi, Oxidized omega-3 fatty acids inhibit NFkappaB activation via a PPARalpha-dependent pathway, Arterioscler. Thromb. Vasc. Biol. 24 (2004) 1621–1627, [https://doi.org/10.1161/01.](https://doi.org/10.1161/01.ATV.0000137191.02577.86) [ATV.0000137191.02577.86.](https://doi.org/10.1161/01.ATV.0000137191.02577.86)
- [92] J.D. Brooks, E.S. Musiek, T.R. Koestner, J.N. Stankowski, J.R. Howard, E. M. Brunoldi, A. Porta, G. Zanoni, G. Vidari, J.D. Morrow, G.L. Milne, B. McLaughlin, The fatty acid oxidation product 15-A3t-isoprostane is a potent inhibitor of NFκB transcription and macrophage transformation, J. Neurochem. 119 (2011) 604–616, <https://doi.org/10.1111/j.1471-4159.2011.07422.x>.
- [93] L. Gao, J. Wang, K.R. Sekhar, H. Yin, N.F. Yared, S.N. Schneider, S. Sasi, T. P. Dalton, M.E. Anderson, J.Y. Chan, J.D. Morrow, M.L. Freeman, Novel n-3 fatty acid oxidation products activate Nrf2 by destabilizing the association between Keap1 and Cullin3, J. Biol. Chem. 282 (2007) 2529–2537, [https://doi.org/](https://doi.org/10.1074/jbc.M607622200) [10.1074/jbc.M607622200.](https://doi.org/10.1074/jbc.M607622200)
- [94] E.S. Musiek, J.D. Brooks, M. Joo, E. Brunoldi, A. Porta, G. Zanoni, G. Vidari, T. S. Blackwell, T.J. Montine, G.L. Milne, B. McLaughlin, J.D. Morrow, Electrophilic cyclopentenone neuroprostanes are anti-inflammatory mediators formed from the peroxidation of the omega-3 polyunsaturated fatty acid docosahexaenoic acid, J. Biol. Chem. 283 (2008) 19927–19935, [https://doi.org/10.1074/jbc.](https://doi.org/10.1074/jbc.M803625200) [M803625200](https://doi.org/10.1074/jbc.M803625200).
- [95] Z. Majkova, J. Layne, M. Sunkara, A.J. Morris, M. Toborek, B. Hennig, Omega-3 fatty acid oxidation products prevent vascular endothelial cell activation by coplanar polychlorinated biphenyls, Toxicol. Appl. Pharmacol. 251 (2011) 41–49, [https://doi.org/10.1016/j.taap.2010.11.013.](https://doi.org/10.1016/j.taap.2010.11.013)
- [96] X. Geng, J.-M. Galano, C. Oger, G.Y. Sun, T. Durand, J.C. Lee, Neuroprotective effects of DHA-derived peroxidation product 4(RS)-4-F4t-neuroprostane on microglia, Free Radic. Biol. Med. 185 (2022) 1–5, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2022.04.002) [freeradbiomed.2022.04.002](https://doi.org/10.1016/j.freeradbiomed.2022.04.002).
- [97] M. Massaro, E. Scoditti, M.A. Carluccio, M.R. Montinari, R. De Caterina, Omega-3 fatty acids, inflammation and angiogenesis: nutrigenomic effects as an explanation for anti-atherogenic and anti-inflammatory effects of fish and fish oils, J. Nutrigenetics Nutrigenomics 1 (2008) 4–23, [https://doi.org/10.1159/](https://doi.org/10.1159/000109871) [000109871](https://doi.org/10.1159/000109871).
- [98] V. Saraswathi, L. Gao, J.D. Morrow, A. Chait, K.D. Niswender, A.H. Hasty, Fish oil increases cholesterol storage in white adipose tissue with concomitant decreases in inflammation, hepatic steatosis, and atherosclerosis in mice, J. Nutr. 137 (2007) 1776–1782, [https://doi.org/10.1093/jn/137.7.1776.](https://doi.org/10.1093/jn/137.7.1776)
- [99] [Dietary supplementation with n-3 polyunsaturated fatty acids and vitamin E after](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref99) [myocardial infarction: results of the GISSI-Prevenzione trial. Gruppo Italiano per](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref99) lo Studio della Sopravvivenza nell'[Infarto miocardico, Lancet Lond. Engl. 354](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref99) [\(1999\) 447](http://refhub.elsevier.com/S0021-9150(24)01172-9/sref99)–455.
- [100] S. Judé, S. Bedut, S. Roger, M. Pinault, P. Champeroux, E. White, J.-Y. Le Guennec, Peroxidation of docosahexaenoic acid is responsible for its effects on I TO and I SS in rat ventricular myocytes, Br. J. Pharmacol. 139 (2003) 816–822, [https://doi.org/10.1038/sj.bjp.0705308.](https://doi.org/10.1038/sj.bjp.0705308)
- [101] J. Roy, L.T. Oliveira, C. Oger, J.-M. Galano, V. Bultel-Poncé, S. Richard, A. G. Guimaraes, J.M.C. Vilela, M.S. Andrade, T. Durand, P. Besson, V.C. F. Mosqueira, J.-Y. Le Guennec, Polymeric nanocapsules prevent oxidation of core-loaded molecules: evidence based on the effects of docosahexaenoic acid and neuroprostane on breast cancer cells proliferation, J. Exp. Clin. Cancer Res. CR 34 (2015) 155, <https://doi.org/10.1186/s13046-015-0273-z>.
- [102] A. Abawi, C. Thomann, G. Lollo, T. Granjon, E. Petiot, A. Bérot, C. Oger, V. Bultel-Poncé, A. Guy, J.-M. Galano, T. Durand, A. Girard-Egrot, O. Maniti, Carrier-tumor cell membrane interactions for optimized delivery of a promising drug, 4(RS)-4- F4t-neuroprostane, Pharmaceutics 15 (2023) 2739, [https://doi.org/10.3390/](https://doi.org/10.3390/pharmaceutics15122739) [pharmaceutics15122739](https://doi.org/10.3390/pharmaceutics15122739).
- [103] J.-F. Bilodeau, N. Gevariya, J. Larose, K. Robitaille, J. Roy, C. Oger, J.-M. Galano, A. Bergeron, T. Durand, Y. Fradet, P. Julien, V. Fradet, Long chain omega-3 fatty acids and their oxidized metabolites are associated with reduced prostate tumor growth, Prostaglandins Leukot. Essent. Fatty Acids 164 (2021) 102215, [https://](https://doi.org/10.1016/j.plefa.2020.102215) doi.org/10.1016/j.plefa.2020.102215.
- [104] J. Jamil, P. Bankhele, A. Salvi, J.E. Mannix, C. Oger, A. Guy, J.-M. Galano, T. Durand, Y.F. Njie-Mbye, S.E. Ohia, C.A. Opere, Role of the non-enzymatic metabolite of eicosapentaenoic acid, 5-epi-5-F3t-isoprostane in the regulation of [(3)H]D-aspartate release in isolated bovine retina, Neurochem. Res. 39 (2014) 2360–2369, [https://doi.org/10.1007/s11064-014-1436-6.](https://doi.org/10.1007/s11064-014-1436-6)
- [105] Y.Y. Lee, J.-M. Galano, H.H. Leung, L. Balas, C. Oger, T. Durand, J.C.-Y. Lee, Nonenzymatic oxygenated metabolite of docosahexaenoic acid, 4(RS)-4-F4t -neuroprostane, acts as a bioactive lipid molecule in neuronal cells, FEBS Lett. 594 (2020) 1797–1808, <https://doi.org/10.1002/1873-3468.13774>.
- [106] E. Moretti, C. Signorini, D. Noto, R. Corsaro, L. Micheli, T. Durand, C. Oger, J. M. Galano, G. Collodel, F4-Neuroprostane effects on human sperm, Int. J. Mol. Sci. 24 (2023) 935, <https://doi.org/10.3390/ijms24020935>.
- [107] J. Roy, C. Vigor, J. Vercauteren, G. Reversat, B. Zhou, A. Surget, L. Larroquet, A. Lanuque, F. Sandres, F. Terrier, C. Oger, J.-M. Galano, G. Corraze, T. Durand, Characterization and modulation of brain lipids content of rainbow trout fed with 100% plant based diet rich in omega-3 long chain polyunsaturated fatty acids DHA and EPA, Biochimie 178 (2020) 137–147, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.biochi.2020.06.010) [biochi.2020.06.010.](https://doi.org/10.1016/j.biochi.2020.06.010)
- [108] J. Roy, L. Larroquet, A. Surget, A. Lanuque, F. Sandres, F. Terrier, G. Corraze, J. Chung-Yung Lee, S. Skiba-Cassy, Impact on cerebral function in rainbow trout fed with plant based omega-3 long chain polyunsaturated fatty acids enriched with DHA and EPA, Fish Shellfish Immunol. 103 (2020) 409–420, [https://doi.](https://doi.org/10.1016/j.fsi.2020.05.044) [org/10.1016/j.fsi.2020.05.044](https://doi.org/10.1016/j.fsi.2020.05.044).