

Unraveling the liver antiviral immunity in functional cure of chronic hepatitis B using scRNAseq

Pierre Tonnerre, Thomas F. Baumert

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Editorial

 More than 250 million people are infected with hepatitis B virus (HBV) worldwide, despite availability of an effective prophylactic vaccine[1,2]. Although current standard of care such as nucleos(t)ide analogs (NA) and interferon-alfa (PEG-IFN) are successful in controlling viral replication, these therapies very rarely result in complete cure due to persistence of HBV covalently closed circular DNA (cccDNA) in infected hepatocytes[3]. Thus, CHB patients remain at high risk of developing cirrhosis and hepatocellular carcinoma[2]. Functional cure (FC), which is characterized by the loss of viral surface antigen (HBsAg), has emerged as a more feasible treatment endpoint[4,5]. Indeed, most adults with acute HBV infection can successfully control the virus. Moreover a fraction of patients with CHB can reach FC following a course of antiviral therapy[4,6]. Functional cure has been associated with lower risk of disease progression and HCC [2].

 The antiviral immune response plays a crucial role for the control of HBV infection. In CHB, immune cells become dysfunctional due to prolonged exposure to high loads of antigens[6]. Previous studies have pinpointed the important role for virus-specific T cells as a potential key cellular component of FC[7–9]. Others have highlighted a possible role for innate immune cells, such as Kupffer and NK cells[10–12]. However, the composition of the liver immune microenvironment in patients with FC remains largely elusive.

 To address this important question, an elegant study from Drs. Balakrishnan C. Narmada, Atefeh Khakpoor, Niranjan Shirgaonkar and colleagues [13], led by senior authors Profs. Seng Gee Lim, from the National University Hospital, Singapore, and Ramanuj DasGupta, from the Genome Institute of Singapore, performed a comprehensive analysis of the liver microenvironment and peripheral immune responses in patients achieving FC. The authors used single-cell RNA sequencing and multiparametric flow-cytometry analysis of core needle liver biopsies and matched-peripheral blood samples to capture 26 distinct major cell types/states, including hepatocytes, from 29 patients with CHB and 9 patients with FC (patients presenting

 clearance of viral HBsAg, either upon treatment (n=7) or spontaneously (n=2), with biopsies being performed between 7 to 84 months after HBsAg loss). The study unraveled a unique liver microenvironment associated with FC characterized by altered innate and adaptive immune cell compositions and functional states, as compared to CHB.

 The first key finding of the study was the association of remodeling of the liver innate immune microenvironment with FC (Fig. 1). Single cell transcriptomic analysis of hepatic needle biopsies revealed an altered innate immune signature in FC. In particular, the authors identified an increase in conventional and liver-resident NK cells in FC, marked by the emergence of a CXCR6+ CD69+ NK population expressing high level of activating receptors (NKp46, CD38, NKG2D and TRAIL) and lower level of inhibitory receptors (PD-1 and TIM-3)[13]. This was accompanied by a remodeling of the myeloid liver compartment, characterized by increased frequencies of pro-inflammatory CD14+ monocytes, a subset of conventional dendritic cells (DC2) as well as FOLR2/VSIG4 Kupffer cells, in FC compared to CHB. The authors also reported a higher number of marginated neutrophils in FC. Interestingly, computational analysis of inferred- interactions identified high inferred-interactions between neutrophils and endothelial cells, through chemokine/receptor interactions, as a potential mechanism for increased neutrophil trafficking in the liver of FC patients [13]. Importantly, most of the above findings were validated at the protein level using flow cytometry and by comparison with a single-cell transcriptomic dataset of healthy liver tissues [14] suggesting an activation state of the intrahepatic innate immune microenvironment in FC. In addition, computational analysis of inferred-interactions highlighted potential interactions between innate cells, particularly DC2 and Kupffer cells, with CD4+ and CD8+ T cells, suggesting an important role for myeloid cells in eliciting T cell immune responses [13]. Interestingly, the authors pinpointed the similarities between the observed FOLR2/VSIG4 Kupffer cell gene profile and an IL2-dependent "KC2" subtype of Kupffer cells, recently reported by De Simone *et al*, with the ability to cross-present antigens and rescue exhausted T cells in mouse models of HBV infection[12].

77 The second important finding was the discovery of restored liver CD4+ T cell responses as a hallmark of FC (Fig. 1). Investigating T cell populations and associated transcriptomic signatures in patients with FC, the authors observed a marked reduction in FOXP3+ regulatory T cells (Tregs) and a lower frequency of exhausted CD8+ T cells, which were associated with decreased expression of inhibitory receptors encoding genes[13]. Importantly, the authors described an increased frequency of CD4+ T cells with an effector/cytotoxic gene signature (PRF1, GZMA, GZMK TNF, NCR3), referred as "CD4-CTLs" by the authors[13]. Again, these findings were further validated at the protein level by flow cytometry analysis of liver samples. It has to be noted that these signatures were not present in patient's paired-blood samples, highlighting the local compartmentalization of the immune alterations into the liver of patients with FC, or the dilution of these alterations in the peripheral blood. Nevertheless, using peptide pools covering the different proteins of the virus (HBx, Core, Polymerase and HBs), the authors successfully identified HBV-specific T populations in the peripheral blood of patients with CHB and FC. Strikingly, in contrast to CHB, FC associated with an increase in the frequency of Polymerase- and HBs-specific T cell responses, but not Core and HBx, which were dominated by 92 a CD4+ polyfunctional (IFNg+TNFa+) phenotype[13]. These results are in line with recent findings from Hoogeveen *et al*[9] and highlight the heterogeneity and functional plasticity of HBV-specific T cells, depending on the targeted viral protein. Concomitant with these findings, the authors identified a subset of hepatocytes with higher expression levels of MCH-II genes predominantly present in patients with FC. These results suggest that liver CD4-CTLs or HBV-specific CD4 T cells could directly recognize a subset of hepatocytes through TCR recognition, contributing to T cell priming and viral control in FC. Interestingly, using droplet digital PCR and single cell cDNA amplification of viral sequences, the authors provided evidence of transcriptionally active cccDNA 100 in FC, albeit at much lower levels than in CHB, supporting low-level viral replication, hence T cell antigen stimulation.

 The strength of the study is its reasonable numbers of patient samples, the study of the liver microenvironment as its focus area, including the analysis of the liver virological features in in patients with FC, the use of scRNASeq to study the cellular phenotypes, providing an invaluable patient-derived data resource for additional studies, and the phenotypic validation at the protein level of key findings. A potential limitation is the pending mechanistic and functional studies to understand the impact of the findings for the biology of FC.

 Collectively, this study elegantly highlights the intrinsic link between intrahepatic immune cell activation and virus control, thereby providing novel biological insights into the liver immune composition and cell states in FC. Interestingly, since most of these changes - as compared to CHB - were not observed in paired-PBMCs, the study again confirms the need for more studies investigating the human hepatic microenvironment in FC and the complex interactions and cell- cell communication between infected hepatocytes and the liver immune ecosystem. Novel technologies such as spatial transcriptomics[15,16] or multiplexed imaging[17,18] will provide additional contributions to the questions in the future. The next steps of investigation will be the study of the molecular and cellular mechanisms leading to the re-emergence of HBV-specific CD4+ T cell responses in the liver, and whether CD4+ T cells directly target infected-hepatocytes expressing HLA class II-restricted antigens.

 What are the clinical implications of the study by Narmada, Khakpoor and colleagues? A detailed understanding of the liver immunobiology associated or leading to FC, will be useful to contribute to develop urgently needed antiviral strategies for FC. The study may provide opportunities to identify blood-based biomarkers associated with FC. In combination with additional mechanistic data, the study of Narmada, Khakpoor colleagues will open new insights 124 informing the design and development of immunotherapies harnessing innate immune response as well as the therapeutic potential of CD4+ T cells to achieve FC.

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 Figure 1: Remodeling of the liver antiviral immunity in functional cure (FC) of chronic hepatitis B unraveled using single cell RNASeq of patient derived liver tissues [13]. Patients with FC are characterized by *in situ* low-levels of HBV viral replication, the presence of MHC class-II expressing hepatocytes, decreased frequency of intrahepatic exhausted T cells (Tex), the presence of memory-like T cells expressing IL7R/CD127 (Tmem-like) and CD4 T cells with 226 cytotoxic functions (CD4-CTL), and the re-emergence of HBV-specific CD4 T cells targeting Polymerase and HBs epitopes. This altered T cell response in FC is accompanied by a re- enforced innate immune response characterized by increased Liver Resident-NK cells expressing CXCR6 and CD69 (LR-NK), FOLR2/VSIG4 Kupffer cells (KC2-like) and marginated neutrophils (NEUT) as described by Narmada, Khakpoor and colleagues [13]. Figure created with BioRender.com