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2	Unraveling the liver antiviral immunity in functional cure of chronic
3	hepatitis B using scRNASeq
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25 Editorial

26 More than 250 million people are infected with hepatitis B virus (HBV) worldwide, despite 27 availability of an effective prophylactic vaccine[1,2]. Although current standard of care such as 28 nucleos(t)ide analogs (NA) and interferon-alfa (PEG-IFN) are successful in controlling viral 29 replication, these therapies very rarely result in complete cure due to persistence of HBV 30 covalently closed circular DNA (cccDNA) in infected hepatocytes[3]. Thus, CHB patients remain 31 at high risk of developing cirrhosis and hepatocellular carcinoma[2]. Functional cure (FC), which 32 is characterized by the loss of viral surface antigen (HBsAg), has emerged as a more feasible 33 treatment endpoint[4,5]. Indeed, most adults with acute HBV infection can successfully control 34 the virus. Moreover a fraction of patients with CHB can reach FC following a course of antiviral 35 therapy[4,6]. Functional cure has been associated with lower risk of disease progression and 36 HCC [2].

The antiviral immune response plays a crucial role for the control of HBV infection. In CHB, immune cells become dysfunctional due to prolonged exposure to high loads of antigens[6]. Previous studies have pinpointed the important role for virus-specific T cells as a potential key cellular component of FC[7–9]. Others have highlighted a possible role for innate immune cells, such as Kupffer and NK cells[10–12]. However, the composition of the liver immune microenvironment in patients with FC remains largely elusive.

43 To address this important question, an elegant study from Drs. Balakrishnan C. Narmada, 44 Atefeh Khakpoor, Niranjan Shirgaonkar and colleagues [13], led by senior authors Profs. Seng 45 Gee Lim, from the National University Hospital, Singapore, and Ramanuj DasGupta, from the 46 Genome Institute of Singapore, performed a comprehensive analysis of the liver 47 microenvironment and peripheral immune responses in patients achieving FC. The authors used 48 single-cell RNA sequencing and multiparametric flow-cytometry analysis of core needle liver 49 biopsies and matched-peripheral blood samples to capture 26 distinct major cell types/states, 50 including hepatocytes, from 29 patients with CHB and 9 patients with FC (patients presenting

51 clearance of viral HBsAg, either upon treatment (n=7) or spontaneously (n=2), with biopsies being 52 performed between 7 to 84 months after HBsAg loss). The study unraveled a unique liver 53 microenvironment associated with FC characterized by altered innate and adaptive immune cell 54 compositions and functional states, as compared to CHB.

55 The first key finding of the study was the association of remodeling of the liver innate 56 immune microenvironment with FC (Fig. 1). Single cell transcriptomic analysis of hepatic needle 57 biopsies revealed an altered innate immune signature in FC. In particular, the authors identified 58 an increase in conventional and liver-resident NK cells in FC, marked by the emergence of a 59 CXCR6+ CD69+ NK population expressing high level of activating receptors (NKp46, CD38, 60 NKG2D and TRAIL) and lower level of inhibitory receptors (PD-1 and TIM-3)[13]. This was 61 accompanied by a remodeling of the myeloid liver compartment, characterized by increased 62 frequencies of pro-inflammatory CD14+ monocytes, a subset of conventional dendritic cells (DC2) 63 as well as FOLR2/VSIG4 Kupffer cells, in FC compared to CHB. The authors also reported a 64 higher number of marginated neutrophils in FC. Interestingly, computational analysis of inferred-65 interactions identified high inferred-interactions between neutrophils and endothelial cells, 66 through chemokine/receptor interactions, as a potential mechanism for increased neutrophil 67 trafficking in the liver of FC patients [13]. Importantly, most of the above findings were validated 68 at the protein level using flow cytometry and by comparison with a single-cell transcriptomic 69 dataset of healthy liver tissues [14] suggesting an activation state of the intrahepatic innate 70 immune microenvironment in FC. In addition, computational analysis of inferred-interactions 71 highlighted potential interactions between innate cells, particularly DC2 and Kupffer cells, with 72 CD4+ and CD8+ T cells, suggesting an important role for myeloid cells in eliciting T cell immune 73 responses [13]. Interestingly, the authors pinpointed the similarities between the observed 74 FOLR2/VSIG4 Kupffer cell gene profile and an IL2-dependent "KC2" subtype of Kupffer cells, 75 recently reported by De Simone et al, with the ability to cross-present antigens and rescue 76 exhausted T cells in mouse models of HBV infection[12].

77 The second important finding was the discovery of restored liver CD4+ T cell responses 78 as a hallmark of FC (Fig. 1). Investigating T cell populations and associated transcriptomic 79 signatures in patients with FC, the authors observed a marked reduction in FOXP3+ regulatory T 80 cells (Tregs) and a lower frequency of exhausted CD8+ T cells, which were associated with 81 decreased expression of inhibitory receptors encoding genes[13]. Importantly, the authors 82 described an increased frequency of CD4+ T cells with an effector/cytotoxic gene signature 83 (PRF1, GZMA, GZMK TNF, NCR3), referred as "CD4-CTLs" by the authors[13]. Again, these 84 findings were further validated at the protein level by flow cytometry analysis of liver samples. It 85 has to be noted that these signatures were not present in patient's paired-blood samples, 86 highlighting the local compartmentalization of the immune alterations into the liver of patients with 87 FC, or the dilution of these alterations in the peripheral blood. Nevertheless, using peptide pools 88 covering the different proteins of the virus (HBx, Core, Polymerase and HBs), the authors 89 successfully identified HBV-specific T populations in the peripheral blood of patients with CHB 90 and FC. Strikingly, in contrast to CHB, FC associated with an increase in the frequency of 91 Polymerase- and HBs-specific T cell responses, but not Core and HBx, which were dominated by 92 a CD4+ polyfunctional (IFNg+TNFa+) phenotype[13]. These results are in line with recent findings 93 from Hoogeveen et al[9] and highlight the heterogeneity and functional plasticity of HBV-specific 94 T cells, depending on the targeted viral protein. Concomitant with these findings, the authors 95 identified a subset of hepatocytes with higher expression levels of MCH-II genes predominantly 96 present in patients with FC. These results suggest that liver CD4-CTLs or HBV-specific CD4 T 97 cells could directly recognize a subset of hepatocytes through TCR recognition, contributing to T 98 cell priming and viral control in FC. Interestingly, using droplet digital PCR and single cell cDNA 99 amplification of viral sequences, the authors provided evidence of transcriptionally active cccDNA 100 in FC, albeit at much lower levels than in CHB, supporting low-level viral replication, hence T cell 101 antigen stimulation.

The strength of the study is its reasonable numbers of patient samples, the study of the liver microenvironment as its focus area, including the analysis of the liver virological features in in patients with FC, the use of scRNASeq to study the cellular phenotypes, providing an invaluable patient-derived data resource for additional studies, and the phenotypic validation at the protein level of key findings. A potential limitation is the pending mechanistic and functional studies to understand the impact of the findings for the biology of FC.

108 Collectively, this study elegantly highlights the intrinsic link between intrahepatic immune 109 cell activation and virus control, thereby providing novel biological insights into the liver immune 110 composition and cell states in FC. Interestingly, since most of these changes - as compared to 111 CHB - were not observed in paired-PBMCs, the study again confirms the need for more studies 112 investigating the human hepatic microenvironment in FC and the complex interactions and cell-113 cell communication between infected hepatocytes and the liver immune ecosystem. Novel 114 technologies such as spatial transcriptomics[15,16] or multiplexed imaging[17,18] will provide 115 additional contributions to the questions in the future. The next steps of investigation will be the 116 study of the molecular and cellular mechanisms leading to the re-emergence of HBV-specific 117 CD4+ T cell responses in the liver, and whether CD4+ T cells directly target infected-hepatocytes 118 expressing HLA class II-restricted antigens.

What are the clinical implications of the study by Narmada, Khakpoor and colleagues? A detailed understanding of the liver immunobiology associated or leading to FC, will be useful to contribute to develop urgently needed antiviral strategies for FC. The study may provide opportunities to identify blood-based biomarkers associated with FC. In combination with additional mechanistic data, the study of Narmada, Khakpoor colleagues will open new insights informing the design and development of immunotherapies harnessing innate immune response as well as the therapeutic potential of CD4+ T cells to achieve FC.

126

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138	The authors declare no competing interests.
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140	Author contributions:
141	P.T. and T.F.B. conceptualized and wrote the manuscript.
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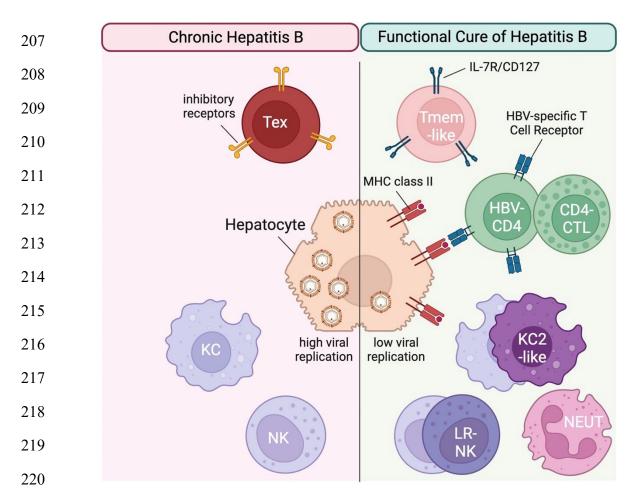
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221 Figure 1: Remodeling of the liver antiviral immunity in functional cure (FC) of chronic 222 hepatitis B unraveled using single cell RNASeg of patient derived liver tissues [13]. Patients 223 with FC are characterized by in situ low-levels of HBV viral replication, the presence of MHC 224 class-II expressing hepatocytes, decreased frequency of intrahepatic exhausted T cells (Tex), the 225 presence of memory-like T cells expressing IL7R/CD127 (Tmem-like) and CD4 T cells with cytotoxic functions (CD4-CTL), and the re-emergence of HBV-specific CD4 T cells targeting 226 227 Polymerase and HBs epitopes. This altered T cell response in FC is accompanied by a re-228 enforced innate immune response characterized by increased Liver Resident-NK cells expressing 229 CXCR6 and CD69 (LR-NK), FOLR2/VSIG4 Kupffer cells (KC2-like) and marginated neutrophils 230 (NEUT) as described by Narmada, Khakpoor and colleagues [13]. Figure created with 231 BioRender.com