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In-vivo diagnosis of cerebral amyloid angiopathy: an updated review

Nicolas Raposo^{a,b,c}, Charlotte Périole^{a,b} and Mélanie Planton^{a,c}

Purpose of review

Sporadic cerebral amyloid angiopathy (CAA) is a highly prevalent small vessel disease in ageing population with potential severe complications including lobar intracerebral hemorrhage (ICH), cognitive impairment, and dementia. Although diagnosis of CAA was made only with postmortem neuropathological examination a few decades ago, diagnosing CAA without pathological proof is now allowed in living patients. This review focuses on recently identified biomarkers of CAA and current diagnostic criteria.

Recent findings

Over the past few years, clinicians and researchers have shown increased interest for CAA, and important advances have been made. Thanks to recent insights into mechanisms involved in CAA and advances in structural and functional neuroimaging, PET amyloid tracers, cerebrospinal fluid and plasma biomarkers analysis, a growing number of biomarkers of CAA have been identified. Imaging-based diagnostic criteria including emerging biomarkers have been recently developed or updated, enabling accurate and earlier diagnosis of CAA in living patients.

Summary

Recent advances in neuroimaging allow diagnosing CAA in the absence of pathological examination. Current imaging-based criteria have high diagnostic performance in patients presenting with ICH, but is more limited in other clinical context such as cognitively impaired patients or asymptomatic individuals. Further research is still needed to improve diagnostic accuracy.

Keywords

biomarkers, cerebral amyloid angiopathy, diagnosis

INTRODUCTION

Sporadic cerebral amyloid angiopathy (CAA) is a common small vessel disease (SVD) characterized by the accumulation of amyloid- β (A β) peptide in the wall of cortical and leptomeningeal vessels [1]. Advancing age is the main risk factor for developing CAA. CAA pathology is found in 20–40% in general aging population and up to 50–60% in demented patients [2]. CAA is a well known cause of lobar intracerebral hemorrhage (ICH), but its clinical spectrum has been recently shown to be much larger, including transient focal neurological episodes (TFNE), rapidly progressive cognitive and neurological decline related to inflammatory CAA, cognitive impairment and dementia [3].

There is currently no known effective therapy for CAA. Nonetheless, treatments targeting A β production or clearance are currently in development and clinical trial assessing anticoagulant strategy in these patients are ongoing, raising the perspective of future treatments. Accurate and early diagnosis of CAA has therefore important prognostic and potential therapeutic implications.

Although diagnosis of CAA was made only with postmortem neuropathological examination a few decades ago, diagnosing CAA without pathological proof is now allowed in living patients, thanks to imaging-based criteria, which have been recently developed or updated [4^{**},5].

In this review, we provide a comprehensive update on the latest identified biomarkers and current diagnostic criteria of CAA.

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KEY POINTS

- Over the last decades, a growing number of biomarkers of cerebral amyloid angiopathy (CAA) have been identified including hemorrhagic and nonhemorrhagic MRI markers, amyloid PET, ApoE genotype, and circulating biomarkers in plasma and cerebrospinal fluid.
- Neuroimaging-based criteria have been recently developed and updated, enabling accurate diagnosis of CAA in living patients, in the absence of pathological examination.
- Current criteria have high diagnostic performance in intracerebral hemorrhage (ICH) presentations of CAA, but sensitivity is more limited in non-ICH presentations (i.e. cognitive impairment and dementia, asymptomatic individuals).

BIOMARKERS OF CEREBRAL AMYLOID ANGIOPATHY

Initially, diagnosis of CAA required brain tissue neuropathological examination from brain biopsy or necropsy. In the 1990s, diagnosis of CAA was allowed in living patients with the introduction of the Boston criteria when brain imaging demonstrated single or multiple strictly lobar hemorrhages in people aged 55 years or older [6]. During the last decade, important advances in neuroimaging and increased interest from clinicians and researchers for CAA enabled development of novel biomarkers of CAA.

Lobar cerebral microbleeds

Definition and mechanisms: Cerebral microbleeds (CMBs) are silent brain hemorrhages visible on MRI blood-sensitive sequences, characterized by small foci of hemosiderin deposits [7]. CMBs are well known neuroimaging markers of SVD, and their distribution is suggestive of SVD subtype [8[•]]. While lobar CMBs are associated with CAA, nonlobar CMBs indicate arteriolosclerosis (also named hypertensive angiopathy) [9^{••}].

Detection: CMBs appear as small (2–10 mm), hypointense, ovoid or round lesions on T2*weighted gradient recalled echo (T2*-GRE) or susceptibility weighted (SWI). See figure for illustrative images. Guidelines for detection and mimics have been published [10,11^{••}].

Link with CAA: Lobar CMBs are neuroimaging markers of CAA included in the MRI-based Boston criteria [4^{••}]. Importantly, the clinical context (ICH, cognitive impairment, ischemic stroke, healthy population) and other imaging markers of SVD

may affect the diagnostic and prognostic (e.g. risk of ICH or cognitive decline) value of CMBs, and should be therefore considered when evaluating CMBs.

Cortical superficial siderosis

Definition and mechanisms: Cortical superficial siderosis (cSS) refers to linear deposits of hemosiderin on the surface of the CNS, restricted to the convexity of the brain hemisphere [12]. Increased evidence suggest that cSS may result from repeated bleeding within the subarachnoid spaces [13–16].

Detection: cSS is defined as a homogeneous hypointense curvilinear signal intensity on T2*-GRE or SWI in the cerebral cortex, within the subarachnoid space, or both [17]. cSS is typically delineated on blood-sensitive sequences as two almost parallel, bilinear hypointensities within the cortex, showing a "tracklike" appearance [18].

Link with CAA: Growing evidence suggests that cSS is a key neuroimaging marker of CAA. Although cSS is detected in 0.7% of the general population [19], prevalence of cSS is 20% in patients with CAA without ICH [20] and 40–60% in CAA patients with ICH [20–22]. In a validation MRI-pathology study, cSS was observed in 23 of 38 patients with pathologically-proven CAA vs. 0 of 22 subjects with ICH not related to CAA [22]. cSS has been linked to transient focal neurological episode [23] and APOE ϵ 2 allele [24], and is included in the MRI-based Boston criteria [4^{••},22].

Convexity subarachnoid hemorrhage

Definition and mechanisms: In patients with CAA, acute convexity subarachnoid hemorrhage (cSAH) can occur in two distinct contexts: (a) in the absence of acute ICH (isolated cSAH), or (b) simultaneously with acute lobar ICH. Two main mechanisms have been advanced to explain cSAH in CAA: (i) leakage from a lobar ICH into the subarachnoid space; (ii) rupture of meningeal vessels directly into the subarachnoid space [17].

Detection: Isolated cSAH corresponds to a subarachnoid bleeding confined to the convexities of the brain that does not extend into the parenchyma, sylvian fissures, ventricles, or basal cisterns [25]. In general, both noncontrast CT and MRI can detect acute subarachnoid hemorrhage with high sensitivity [26–29]. On MRI, acute cSAH appears as linear hyperintense signal on FLAIR (suggestive of acute blood), with a corresponding hypointense signal (but not constant) on T2*GRE or SWI in the subarachnoid space that affected one or more cortical sulci of the cerebral convexities. In patients with CAA, acute cSAH is more frequently observed in association with an acute lobar ICH.

Link with CAA: A growing number of cohort studies have linked CAA and isolated cSAH [30– 34]. CAA is now considered as the main cause of cSAH in subjects older than 60 years [25,35]. The clinical presentation of cSAH related to CAA consisted typically in TFNEs and cSS is observed in up to 90%. Recent studies of patients with acute ICH suggest that presence of cSAH is independently associated with underlying CAA pathology and is now included in the CT-based Edinburgh criteria [5]. cSAH may also be a risk factor for ICH recurrence in patients with CAA-related ICH [36].

White matter hyperintensities

Definition and mechanisms: CAA is also recognized as a significant contributor to white matter hyperintensities (WMH). Growing evidence suggest that beta-amyloid accumulation in the vessel wall can lead to impaired vascular reactivity, resulting in hypoperfusion [37^{*},38].

Detection: The Boston Criteria v2.0 integrate WMH features as novel MRI marker for CAA with multispot spatial distribution, defined as more than ten T2-weighted FLAIR small circular or ovoid hyperintense lesions in the subcortical white matter of both hemispheres [4^{••}].

Link with CAA: Although WMH multisport pattern is associated with CAA-related ICH and other imaging markers of CAA (i.e. lobar CMBs and enlarged perivascular spaces in the centrum semiovale), a WMH peri-basal ganglia pattern is associated with Hypertension-related ICH and imaging markers of arteriolosclerosis (i.e. deep CMBs and enlarged perivascular spaces in the basal ganglia) [39].

Enlarged perivascular spaces

Definition and mechanisms: Perivascular spaces are extensions of the extracerebral fluid space around small cerebral vessels as they course from the brain surface into and through the brain parenchyma [40,41]. They are usually not visible on conventional neuroimaging. However, they become visible on MRI when enlargement of the potential space occurs, possibly secondary to impaired interstitial fluid drainage and are named enlarged perivascular spaces (EPVS) [42].

Detection: EPVS are defined as small (< 3 mm) round or linear CSF isointense lesions (T2-hyper-intense and T1/FLAIR hypointense with respect to brain) along the course of penetrating arteries [43]. They can be detected in the centrum semiovale

(CSO) white matter, the basal ganglia (BG), the hippocampus and the midbrain. Several visual rating scales have been developed and validated [44,45].

Link with CAA: EPVS are emerging markers of SVD, observed both in CAA and hypertension angiopathy (i.e. arteriolosclerosis). Their distribution pattern may suggest the underlying SVD. EPVS in the basal ganglia (BG-EPVS) have been linked to hypertension and imaging markers of hypertensive angiopathy (i.e. lacunes, WMH and deep CMB) whereas EPVS in the CSO (CSO-EPVS) are associated with Alzheimer disease (AD) as well as MRI markers of CAA, amyloid PET positivity and CAA pathology [44,46–54]. Severe CSO-EPVS are now included in the Boston criteria V2.0 [4^{••}].

Apolipoprotein E genotype

Definition and mechanisms: The apolipoprotein E (ApoE) gene, located in chromosome 19q13.2, comprises 3 alleles ($\epsilon 2$, $\epsilon 3$, and $\epsilon 4$). Compared to the most common APOE $\epsilon 3$ homozygous genotype, ApoE $\epsilon 2$ and/or $\epsilon 4$ allele carriers have increased risk of CAA.

Link with CAA: ApoE genotype is the most important genetic risk factor identified for sporadic CAA. Neuropathological studies suggested that APOE ε 4 may enhance vascular A β deposition in a dose-dependent fashion, while APOE ε 2 promotes vascular breakdown of amyloid-laden vessels [55– 57]. Cohort studies reported that APOE ε 2 was associated with cSS [58] and increased risk of symptomatic ICH [59] whereas APOE ε 4 was linked to higher burden of lobar CMBs [24].

Amyloid positron emission tomography (PET)

Definition and mechanisms: Detecting during life vascular amyloid deposition rather than brain injury may have important implications. Amyloid PET tracers, originally developed to identify amyloid plaques in AD, have been evaluated in CAA.

Detection: Different amyloid PET tracer and methods allow in vivo detection and/or quantification of amyloid deposition in brain parenchyma and vessels. Visual assessment of PET images and quantitative methods are commonly used to analyze amyloid PET.

Link with CAA: Amyloid PET using 11C-PiB and 18F-florbetapir radiotracers has been assessed in cross-sectional studies [60–67]. Most of these studies have shown that amyloid tracer retention was increased in patients with CAA compared to controls [68]. Due to limited sample sizes and heterogenicity across studies with different radiotracer,

PET analysis methods and control groups, the value of amyloid PET for diagnosing CAA remain uncertain. Future large PET studies including subjects with pathological examination for CAA would be of great interest.

Circulating biomarkers

Definition and mechanisms: Detecting amyloid biomarkers – originally developed in AD – in the cerebrospinal fluid (CSF) or in the plasma may help in diagnosing CAA, providing evidence of amyloid pathology.

Detection: Core CSF biomarkers have been explored in CAA patients. However, the number of studies on this topic is limited and the sample sizes of CAA cohorts are small (n < 20). Plasma biomarkers may offer several potential advantages compared to CSF biomarkers (less invasive) or amyloid PET (less expensive). Nonetheless, amyloid biomarkers have low concentrations in blood, constituting an important technical challenge.

Link with CAA: Compared to healthy controls, patients with AD show decreased level of AB42 (without decreased level of $A\beta 40$) with increased level of total tau (t-tau) and phosphorylated tau (p-tau) in CSF. A meta-analysis of CSF biomarkers in sporadic CAA have shown that patients with CAA had decreased level of both AB40 and AB42 with mild increase in t-tau and p-tau levels [69]. This specific pattern of AB40/AB42 is consistent with pathological observations that showed predominantly AB42 in amyloid plaque although vascular amyloid contains substantial amounts of AB40 in addition to AB42 [70]. To date, evidence of a potential role of plasma amyloid biomarkers in CAA are scarce with conflicting data [71,72]. Nonetheless, high-performance plasma amyloid-ß biomarkers have been recently developed in AD, showing high performance in predicting brain amyloid-β burden [73–77]. Their clinical utility in CAA has not yet been assessed.

Other candidate biomarkers

Advances in neuroimaging enabled development of novel techniques that may be of interest as a diagnostic tool and/or a marker of disease severity. Those include small acute ischemic lesions on diffusionweighted imaging (DWI) [78–81] or cortical microinfarcts visible on 7 T MRI, but also on 1.5 or 3 T MRI [82,83]. Evaluating brain network connectivity using diffusion tensor imaging (DTI) – a noninvasive MRI method to quantify microscopic white matter tissue injury – may also be useful, as a marker of CAA severity [84–87]. Techniques assessing cerebrovascular reactivity, which is altered in CAA, might be useful for early diagnosis of CAA. Among them, functional MRI [blood oxygen level-dependent (BOLD) response] may be more sensitive methods to detect cerebrovascular dysfunction in CAA patients, showing altered response to visual stimuli compared to healthy controls [88,89].

DIAGNOSTIC CRITERIA FOR CEREBRAL AMYLOID ANGIOPATHY

Definite diagnosis of CAA requires histopathological evaluation of brain tissue from autopsy or biopsy samples showing characteristic vascular lesions and accumulation of amyloid β peptide in the cerebrovascular wall of cortical and leptomeningeal vessels. During the last decades, important advances in neuroimaging have been made and diagnostic imaging-based criteria have been developed or updated, enabling CAA diagnosis in living patients in the absence of pathological examination.

The Boston criteria

The Boston criteria are the most widely used criteria for CAA diagnosis in research and clinical practice. The original Boston criteria (v 1.0) were first introduced in 1995 and validated in a small (n=39) single center cohort study [6]. They included the following categories: (a) definite CAA, based on pathological proof of CAA on full autopsy, (b) probable CAA (with or without pathological proof), defined as evidence of at least 2 brain hemorrhages strictly located to lobar (i.e. cortical and cortico-subcortical) regions, including ICH and CMBs, (c) possible CAA, defined as a single lobar hemorrhage.

In 2010, the Boston criteria were revised with the introduction of cSS, as an additional diagnostic imaging marker, and validated in a single cohort study (n = 60). These revised criteria are known as the modified Boston criteria (v 1.5) [22].

The original Boston criteria have high sensitivity and specificity, and the modified Boston criteria have shown improved sensitivity without lowering specificity compared to the original Boston criteria. However, the validity of these MRI-based Boston criteria suffered from several limitations including the limited sample size (<100) and the monocentric design of imaging-pathological studies [6,22,51]. In addition, included patients had predominantly ICH clinical presentations, where the Boston criteria may have higher diagnostic performance compared to non-ICH presentations (i.e. cognitive impairment, TFNE).

To address these issues, a large international multicenter MRI-pathological study was designed

across the International CAA Association network. This study, published in 2022, allowed to update and validate the Boston criteria v 2.0 in a large multicenter cohort (n = 341). Key changes of these updated criteria include (a) integration of emerging white matter MRI markers of CAA (i.e. severe visible perivascular spaces in the centrum semiovale and white matter hyperintensities in a multispot pattern), (b) patient's age 50 years and older (vs. 55 years and older in the Boston criteria v 1.5), (c) applicable to subjects either with ICH or non-ICH presentations. Compared to the Boston criteria v 1.5, the Boston criteria v2.0 have higher sensitivity (79.8% vs. 70.8%) and comparable specificity (84.7% vs. 88.8%). As observed with criteria v 1.5, sensitivity of the Boston criteria v 2.0 appears to be higher in ICH presentations than non-ICH presentations.

The Edinburg criteria

Since MRI is sometimes contraindicated, not tolerated or unavailable in the setting of acute ICH, the Edinburg CT and genetic diagnostic criteria have been developed to diagnose underlying CAA in patients with acute lobar ICH, in the absence of brain MRI and pathological examination [5]. Those included the presence of ApoE ɛ4 allele and 2 characteristic CT feature of CAA-related ICH including finger-like projections and subarachnoid hemorrhage. In a cohort of 62 patients with severe lobar ICH who died and had postmortem pathological evaluation, the Edinburgh criteria had a 96% (95% CI, 78–100) rule in specificity when at least 2/3 markers were present (high risk), and a 100% (95% CI, 88–100) rule out sensitivity in the absence of these markers (low risk) [8[•]].

CONCLUSION

Over the last decades, a growing number of biomarkers have been identified thanks to recent insights into mechanisms involved in CAA provided by animal models and advances in structural and functional neuroimaging, PET amyloid tracers, circulating biomarkers analysis. Current diagnostic criteria have high sensitivity and specificity in ICH presentations of CAA, but have lower performance in other clinical context such as cognitive



FIGURE 1. MRI markers of cerebral amyloid angiopathy. (a, e) Convexity subarachnoid hemorrhage (white arrow) appearing as linear hypointense signal on T2*GRE (a), with a corresponding hyperintense signal on FLAIR (e) in the subarachnoid space that affected two cortical sulci of the cerebral convexities. (b, f) Cortical superficial siderosis (white arrowhead) delineated on T2*GRE (b) as two almost parallel, bilinear hypointensities within the cortex, showing a tracklike appearance, without corresponding hyperintense signal on FLAIR (f). (c, g) Multiple lobar cerebral microbleeds visible as small round hypointense signal on T2*GRE (c) and SWI (g). (d) Severe enlarged perivascular spaces in the centrum semiovale (black arrows) visible on T2-weighted imaging (i.e., more than 20 visible perivascular spaces in the centrum semiovale of one hemisphere). (e) White matter hyperintensities (black arrowheads) in a multispot pattern (i.e., more than 10 small circular or ovoid T2-weighted FLAIR hyperintense lesions in the bilateral subcortical white matter).

Cerebrovascular disease

presentation or mixed hemorrhages (i.e. presence of both deep and lobar hemorrhages) [90,91]. Further research is needed to evaluate whether novels biomarkers including amyloid PET and circulating biomarkers would improve diagnostic accuracy (Fig. 1).

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Conflicts of interest

Dr Raposo has received consulting fees from Novartis. The other authors have no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

of special interest

- of outstanding interest
- Charidimou A, Gang O, Werring DJ. Sporadic cerebral amyloid angiopathy revisited: recent insights into pathophysiology and clinical spectrum. J Neurol Neurosurg Psychiatry 2012; 83:124–137.
- Keage HÅ, Carare ŘO, Friedland RP, et al. Population studies of sporadic cerebral amyloid angiopathy and dementia: a systematic review. BMC Neurol 2009; 9:3.
- Wermer MJH, Greenberg SM. The growing clinical spectrum of cerebral amyloid angiopathy. Curr Opin Neurol 2018; 31:28–35.
- **4.** Charidimou A, Boulouis G, Frosch MP, *et al.* The Boston criteria version 2.0 ■ for cerebral amyloid angiopathy: a multicentre, retrospective, MRI-neuro-

pathology diagnostic accuracy study. Lancet Neurol 2022; 21:714-725. This international MRI-pathological study allowed development and validation of the updated version V2.0 of the MRI-based Boston criteria which are the most widely used criteria for diagnosing CAA.

- Rodrigues MA, Samarasekera N, Lerpiniere C, et al. The Edinburgh CT and genetic diagnostic criteria for lobar intracerebral haemorrhage associated with cerebral amyloid angiopathy: model development and diagnostic test accuracy study. Lancet Neurol 2018; 17:232-240.
- Knudsen KA, Rosand J, Karluk D, Greenberg SM. Clinical diagnosis of cerebral amyloid angiopathy: validation of the Boston criteria. Neurology 2001; 56:537-539.
- Fazekas F, Kleinert R, Roob G, et al. Histopathologic analysis of foci of signal loss on gradient-echo T2*-weighted MR images in patients with spontaneous intracerebral hemorrhage: evidence of microangiopathy-related microbleeds. AJNR Am J Neuroradiol 1999; 20:637–642.
- 8. Puy L, Pasi M, Rodrigues M, et al. Cerebral microbleeds: from depiction to
- interpretation. J Neurol Neurosurg Psychiatry 2021; 92:598-607.

This comprehensive review focuses on mechanisms, detection and diagnostic value of cerebral microbleeds.

9. Raposo N, Zanon Zotin MC, Seiffge DJ, *et al.* A causal classification system
for intracerebral hemorrhage subtypes. Ann Neurol 2022; 5:16–28.

This is a novel causal classification for ICH subtypes, named CLAS-ICH, based on the most recent advances in neuroimaging, ICH epidemiology, and cerebral small vessel disease phenotyping.

- Greenberg SM, Vernooij MW, Cordonnier C, et al. Cerebral microbleeds: a guide to detection and interpretation. Lancet Neurol 2009; 8:165–174.
- Duering M, Biessels GJ, Brodtmann A, et al. Neuroimaging standards for research into small vessel disease-advances since. Lancet Neurol 2023; 22:602-618.

This comprehensive review focuses on recent insight into small vessel disease mechanisms and imaging, and provide updates on international guidelines for small vessel disease neuroimaging (STRIVE).

- Wilson D, Chatterjee F, Farmer SF, et al. Infratentorial superficial siderosis: Classification, diagnostic criteria, and rational investigation pathway. Ann Neurol 2017; 81:333–343.
- Koeppen AH, Dickson AC, Chu RC, Thach RE. The pathogenesis of superficial siderosis of the central nervous system. Ann Neurol 1993; 34:646–653.
- Calviere L, Cuvinciuc V, Raposo N, et al. Acute convexity subarachnoid hemorrhage related to cerebral amyloid angiopathy: clinicoradiological features and outcome. J Stroke Cerebrovasc Dis 2016; 25:1009–1016.

- Beitzke M, Enzinger C, Wunsch G, et al. Contribution of convexal subarachnoid hemorrhage to disease progression in cerebral amyloid angiopathy. Stroke 2015; 46:1533–1540.
- Charidimou A, Perosa V, Frosch MP, et al. Neuropathological correlates of cortical superficial siderosis in cerebral amyloid angiopathy. Brain 2020; 143:3343–3351.
- Charidimou A, Linn J, Vernooij MW, et al. Cortical superficial siderosis: detection and clinical significance in cerebral amyloid angiopathy and related conditions. Brain 2015; 138(Pt 8):2126–2139.
- Linn J, Herms J, Dichgans M, et al. Subarachnoid hemosiderosis and superficial cortical hemosiderosis in cerebral amyloid angiopathy. AJNR Am J Neuroradiol 2008; 29:184–186.
- Vernooij MW, Ikram MA, Hofman A, et al. Superficial siderosis in the general population. Neurology 2009; 73:202–205.
- Charidimou A, Martinez-Ramirez S, Shoamanesh A, et al. Cerebral amyloid angiopathy with and without hemorrhage: evidence for different disease phenotypes. Neurology 2015; 84:1206–1212.
- Charidimou A, Jager RH, Fox Z, et al. Prevalence and mechanisms of cortical superficial siderosis in cerebral amyloid angiopathy. Neurology 2013; 81:626-632.
- Linn J, Halpin A, Demaerel P, et al. Prevalence of superficial siderosis in patients with cerebral amyloid angiopathy. Neurology 2010; 74:1346–1350.
- 23. Charidimou A, Peeters A, Fox Z, et al. Spectrum of transient focal neurological episodes in cerebral amyloid angiopathy: multicentre magnetic resonance imaging cohort study and meta-analysis. Stroke 2012; 43:2324–2330.
- Shoamanesh A, Martinez-Ramirez S, Oliveira-Filho J, et al. Interrelationship of superficial siderosis and microbleeds in cerebral amyloid angiopathy. Neurology 2014; 83:1838–1843.
- Kumar S, Goddeau RP Jr, Selim MH, et al. Atraumatic convexal subarachnoid hemorrhage: clinical presentation, imaging patterns, and etiologies. Neurology 2010; 74:893–899.
- Wiesmann M, Mayer TE, Medele R, Bruckmann H. Diagnosis of acute subarachnoid hemorrhage at 1.5 Tesla using proton-density weighted FSE and MRI sequences. Radiologe 1999; 39:860–865.
- Yuan MK, Lai PH, Chen JY, et al. Detection of subarachnoid hemorrhage at acute and subacute/chronic stages: comparison of four magnetic resonance imaging pulse sequences and computed tomography. J Chin Med Assoc 2005; 68:131-137.
- Mohamed M, Heasly DC, Yagmurlu B, Yousem DM. Fluid-attenuated inversion recovery MR imaging and subarachnoid hemorrhage: not a panacea. AJNR Am J Neuroradiol 2004; 25:545–550.
- Noguchi K, Seto H, Kamisaki Y, et al. Comparison of fluid-attenuated inversion-recovery MR imaging with CT in a simulated model of acute subarachnoid hemorrhage. AJNR Am J Neuroradiol 2000; 21:923–927.
- Brunot S, Fromont A, Ricolfi F, et al. Focal subarachnoid hemorrhage and cerebral amyloid angiopathy: a nonfortuitous association. Rev Neurol (Paris) 2010; 166:83–9.
- Raposo N, Viguier A, Cuvinciuc V, et al. Cortical subarachnoid haemorrhage in the elderly: a recurrent event probably related to cerebral amyloid angiopathy. Eur J Neurol 2011; 18:597–603.
- 32. Apoil M, Cogez J, Dubuc L, et al. Focal cortical subarachnoid hemorrhage revealed by recurrent paresthesias: a clinico-radiological syndrome strongly associated with cerebral amyloid angiopathy. Cerebrovasc Dis 2013; 36:139-144.
- Ly JV, Singhal S, Rowe CC, et al. Convexity subarachnoid hemorrhage with PiB positive pet scans: clinical features and prognosis. J Neuroimaging 2015; 18:420–429.
- Martinez-Lizana E, Carmona-Iragui M, Alcolea D, et al. Cerebral amyloid angiopathy-related atraumatic convexal subarachnoid hemorrhage: an ARIA before the tsunami. J Cereb Blood Flow Metab 2015; 35:710–717.
- Beitzke M, Gattringer T, Enzinger C, et al. Clinical presentation, etiology, and long-term prognosis in patients with nontraumatic convexal subarachnoid hemorrhage. Stroke 2011; 42:3055–3060.
- Raposo N, Charidimou A, Roongpiboonsopit D, et al. Convexity subarachnoid hemorrhage in lobar intracerebral hemorrhage: a prognostic marker. Neurology 2020; 94:e968–e977.
- Gokcal E, Horn MJ, Becker JA, et al. Effect of vascular amyloid on white matter disease is mediated by vascular dysfunction in cerebral amyloid angiopathy. J Cereb Blood Flow Metab 2022; 42:1272–1281.

This recent multimodal study (structural MRI, functional MRI and amyloid PET) provides novel insights into the relationship between amyloid burden, cerebrovascular reactivity and WMH volume in patients with CAA.

- Gurol ME, Viswanathan A, Gidicsin C, et al. Cerebral amyloid angiopathy burden associated with leukoaraiosis: a positron emission tomography/magnetic resonance imaging study. Ann Neurol 2013; 73:529–536.
- Charidimou A, Boulouis G, Haley K, et al. White matter hyperintensity patterns in cerebral amyloid angiopathy and hypertensive arteriopathy. Neurology 2016; 86:505-511.
- Weller RO, Hawkes CA, Kalaria RN, et al. White matter changes in dementia: role of impaired drainage of interstitial fluid. Brain Pathol 2015; 25:63–78.
- Wardlaw JM, Smith C, Dichgans M. Mechanisms of sporadic cerebral small vessel disease: insights from neuroimaging. Lancet Neurol 2013; 12:483–497.
- Wardlaw JM, Benveniste H, Nedergaard M, et al. Perivascular spaces in the brain: anatomy, physiology and pathology. Nat Rev Neurol 2020; 16:137–153.

- 43. Wardlaw JM, Smith EE, Biessels GJ, et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. Lancet Neurol 2013; 12:822–838.
- Doubal FN, MacLullich AM, Ferguson KJ, et al. Enlarged perivascular spaces on MRI are a feature of cerebral small vessel disease. Stroke 2010; 41:450-454.
- Maclullich AM, Wardlaw JM, Ferguson KJ, et al. Enlarged perivascular spaces are associated with cognitive function in healthy elderly men. J Neurol Neurosurg Psychiatry 2004; 75:1519–1523.
- Charidimou A, Boulouis G, Pasi M, et al. MRI-visible perivascular spaces in cerebral amyloid angiopathy and hypertensive arteriopathy. Neurology 2017; 88:1157–1164.
- 47. Zhu YC, Tzourio C, Soumare A, et al. Severity of dilated Virchow-Robin spaces is associated with age, blood pressure, and MRI markers of small vessel disease: a population-based study. Stroke 2010; 41:2483–2490.
- Banerjee G, Kim HJ, Fox Z, et al. MRI-visible perivascular space location is associated with Alzheimer's disease independently of amyloid burden. Brain 2017; 140:1107–1116.
- Martinez-Ramirez S, Pontes-Neto OM, Dumas AP, et al. Topography of dilated perivascular spaces in subjects from a memory clinic cohort. Neurology 2013; 80:1551–1556.
- 50. Charidimou A, Meegahage R, Fox Z, et al. Enlarged perivascular spaces as a marker of underlying arteriopathy in intracerebral haemorrhage: a multicentre MRI cohort study. J Neurol Neurosurg Psychiatry 2013; 84:624–629.
- Charidimou A, Jaunmuktane Z, Baron JC, et al. White matter perivascular spaces: an MRI marker in pathology-proven cerebral amyloid angiopathy? Neurology 2014; 82:57–62.
- Martinez-Ramirez S, van Rooden S, Charidimou A, et al. Perivascular spaces volume in sporadic and hereditary (Dutch-type) cerebral amyloid angiopathy. Stroke 2018; 49:1913–1919.
- Charidimou A, Hong YT, Jager HR, et al. White matter perivascular spaces on magnetic resonance imaging: marker of cerebrovascular amyloid burden? Stroke 2015; 46:1707–1709.
- Raposo N, Planton M, Payoux P, et al. Enlarged perivascular spaces and florbetapir uptake in patients with intracerebral hemorrhage. Eur J Nucl Med Mol Imaging 2019; 46:2339–2347.
- Rannikmae K, Samarasekera N, Martinez-Gonzalez NA, et al. Genetics of cerebral amyloid angiopathy: systematic review and meta-analysis. J Neurol Neurosurg Psychiatry 2013; 84:901–908.
- Greenberg SM, Briggs ME, Hyman BT, et al. Apolipoprotein E epsilon 4 is associated with the presence and earlier onset of hemorrhage in cerebral amyloid angiopathy. Stroke 1996; 27:1333–1337.
- McCarron MO, Nicoll JA, Stewart J, *et al.* The apolipoprotein E epsilon2 allele and the pathological features in cerebral amyloid angiopathy-related hemorrhage. J Neuropathol Exp Neurol 1999; 58:711–718.
- Charidimou A, Zonneveld HI, Shams S, et al. APOE and cortical superficial siderosis in CAA: meta-analysis and potential mechanisms. Neurology 2019; 93:e358-e371.
- O'Donnell HC, Rosand J, Knudsen KA, *et al.* Apolipoprotein E genotype and the risk of recurrent lobar intracerebral hemorrhage. N Engl J Med 2000; 342:240–245.
- Johnson KA, Gregas M, Becker JA, et al. Imaging of amyloid burden and distribution in cerebral amyloid angiopathy. Ann Neurol 2007; 62:229–234.
- Ly JV, Donnan GA, Villemagne VL, et al. 11C-PIB binding is increased in patients with cerebral amyloid angiopathy-related hemorrhage. Neurology 2010; 74:487–493.
- Gurol ME, Viswanathan A, Gidicsin C, et al. Cerebral amyloid angiopathy burden associated with leukoaraiosis: a positron emission tomography/magnetic resonance imaging study. Ann Neurol 2013; 13:529–536.
- Tsai HH, Tsai LK, Chen YF, et al. Correlation of cerebral microbleed distribution to amyloid burden in patients with primary intracerebral hemorrhage. Sci Rep 2017; 7:44715.
- Tsai HH, Pasi M, Tsai LK, et al. Microangiopathy underlying mixed-location intracerebral hemorrhages/microbleeds: a PiB-PET study. Neurology 2019; 92:e774-e781.
- Baron JC, Farid K, Dolan E, et al. Diagnostic utility of amyloid PET in cerebral amyloid angiopathy-related symptomatic intracerebral hemorrhage. J Cereb Blood Flow Metab 2014; 34:753–758.
- Gurol ME, Becker JA, Fotiadis P, et al. Florbetapir-PET to diagnose cerebral amyloid angiopathy: a prospective study. Neurology 2016; 87:2043-2049.

- 67. Raposo N, Planton M, Peran P, *et al.* Florbetapir imaging in cerebral amyloid angiopathy-related hemorrhages. Neurology 2017; 89:697–704.
- Charidimou A, Farid K, Baron JC. Amyloid-PET in sporadic cerebral amyloid angiopathy: a diagnostic accuracy meta-analysis. Neurology 2017; 89:1490-1498.
- Charidimou A, Friedrich JO, Greenberg SM, Viswanathan A. Core cerebrospinal fluid biomarker profile in cerebral amyloid angiopathy: a meta-analysis. Neurology 2018; 90:e754-e762.
- 70. Iwatsubo T, Odaka A, Suzuki N, et al. Visualization of A beta 42(43) and A beta 40 in senile plaques with end-specific A beta monoclonals: evidence that an initially deposited species is A beta 42(43). Neuron 1994; 13:45–53.
- Hernandez-Guillamon M, Delgado P, Penalba A, *et al.* Plasma beta-amyloid levels in cerebral amyloid angiopathy-associated hemorrhagic stroke. Neurodegener Dis 2012; 10:320–323.
- Greenberg SM, Cho HS, O'Donnell HC, et al. Plasma beta-amyloid peptide, transforming growth factor-beta 1, and risk for cerebral amyloid angiopathy. Ann N Y Acad Sci 2000; 903:144–149.
- Nakamura A, Kaneko N, Villemagne VL, et al. High performance plasma amyloid-beta biomarkers for Alzheimer's disease. Nature 2018; 554:249-254.
- Schindler SE, Bollinger JG, Ovod V, *et al.* High-precision plasma beta-amyloid 42/40 predicts current and future brain amyloidosis. Neurology 2019; 93: e1647-e1659.
- Verberk IMW, Slot RE, Verfaillie SCJ, et al. Plasma amyloid as prescreener for the earliest Alzheimer pathological changes. Ann Neurol 2018; 84:648–658.
- Vergallo A, Megret L, Lista S, et al. Plasma amyloid beta 40/42 ratio predicts cerebral amyloidosis in cognitively normal individuals at risk for Alzheimer's disease. Alzheimers Dement 2019; 15:764–775.
- 77. de Wolf F, Ghanbari M, Licher S, et al. Plasma tau, neurofilament light chain and amyloid-beta levels and risk of dementia; a population-based cohort study. Brain 2020; 143:1220–1232.
- Kimberly WT, Gilson A, Rost NS, et al. Silent ischemic infarcts are associated with hemorrhage burden in cerebral amyloid angiopathy. Neurology 2009; 72:1230-1235.
- Auriel E, Gurol ME, Ayres A, *et al.* Characteristic distributions of intracerebral hemorrhage-associated diffusion-weighted lesions. Neurology 2012; 79:2335-2341.
- van Veluw SJ, Lauer A, Charidimou A, et al. Evolution of DWI lesions in cerebral amyloid angiopathy: Evidence for ischemia. Neurology 2017; 89:2136-2142.
- Revel-Mouroz P, Viguier A, Cazzola V, *et al.* Acute ischaemic lesions are associated with cortical superficial siderosis in spontaneous intracerebral hemorrhage. Eur J Neurol 2018; 26:660–666.
- van Veluw SJ, Charidimou A, van der Kouwe AJ, *et al.* Microbleed and microinfarct detection in amyloid angiopathy: a high-resolution MRI-histopathology study. Brain 2016; 139(Pt 12):3151–3162.
- van Veluw SJ, Shih AY, Smith EE, et al. Detection, risk factors, and functional consequences of cerebral microinfarcts. Lancet Neurol 2017; 16:730–740.
- Reijmer YD, Fotiadis P, Martinez-Ramirez S, et al. Structural network alterations and neurological dysfunction in cerebral amyloid angiopathy. Brain 2015; 138(Pt 1):179–188.
- Raposo N, Zanon Zotin MC, Schoemaker D, et al. Peak width of skeletonized mean diffusivity as neuroimaging biomarker in cerebral amyloid angiopathy. AJNR Am J Neuroradiol 2021; 42:875–881.
- McCreary CR, Beaudin AE, Subotic A, *et al.* Cross-sectional and longitudinal differences in peak skeletonized white matter mean diffusivity in cerebral amyloid angiopathy. Neuroimage Clin 2020; 27:102280.
- Zanon Zotin MC, Schoemaker D, Raposo N, et al. Peak width of skeletonized mean diffusivity in cerebral amyloid angiopathy: Spatial signature, cognitive, and neuroimaging associations. Front Neurosci 2022; 16:1051038.
- Dumas A, Dierksen GA, Gurol ME, et al. Functional magnetic resonance imaging detection of vascular reactivity in cerebral amyloid angiopathy. Ann Neurol 2012; 72:76–81.
- Peca S, McCreary CR, Donaldson E, et al. Neurovascular decoupling is associated with severity of cerebral amyloid angiopathy. Neurology 2013; 81:1659-1665.
- Blanc C, Viguier A, Calviere L, et al. Underlying small vessel disease associated with mixed cerebral microbleeds. Front Neurol 2019; 10:1126.
- Martinez-Ramirez S, Romero JR, Shoamanesh A, et al. Diagnostic value of lobar microbleeds in individuals without intracerebral hemorrhage. Alzheimers Dement 2015; 11:1480–1488.