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Validation of rumen sampling via stomach tubing to inoculate dual-effluent mini-fermenters

Berthelot V.¹, Charef-Mansouri M.¹, Giger-Reverdin S.¹, Broudiscou L.P.^{1,2}

¹Université Paris-Saclay, AgroParisTech, INRAE, UMR 971 MoSAR, France

²UMR 1419 NuMeA, INRAE UPPA, France



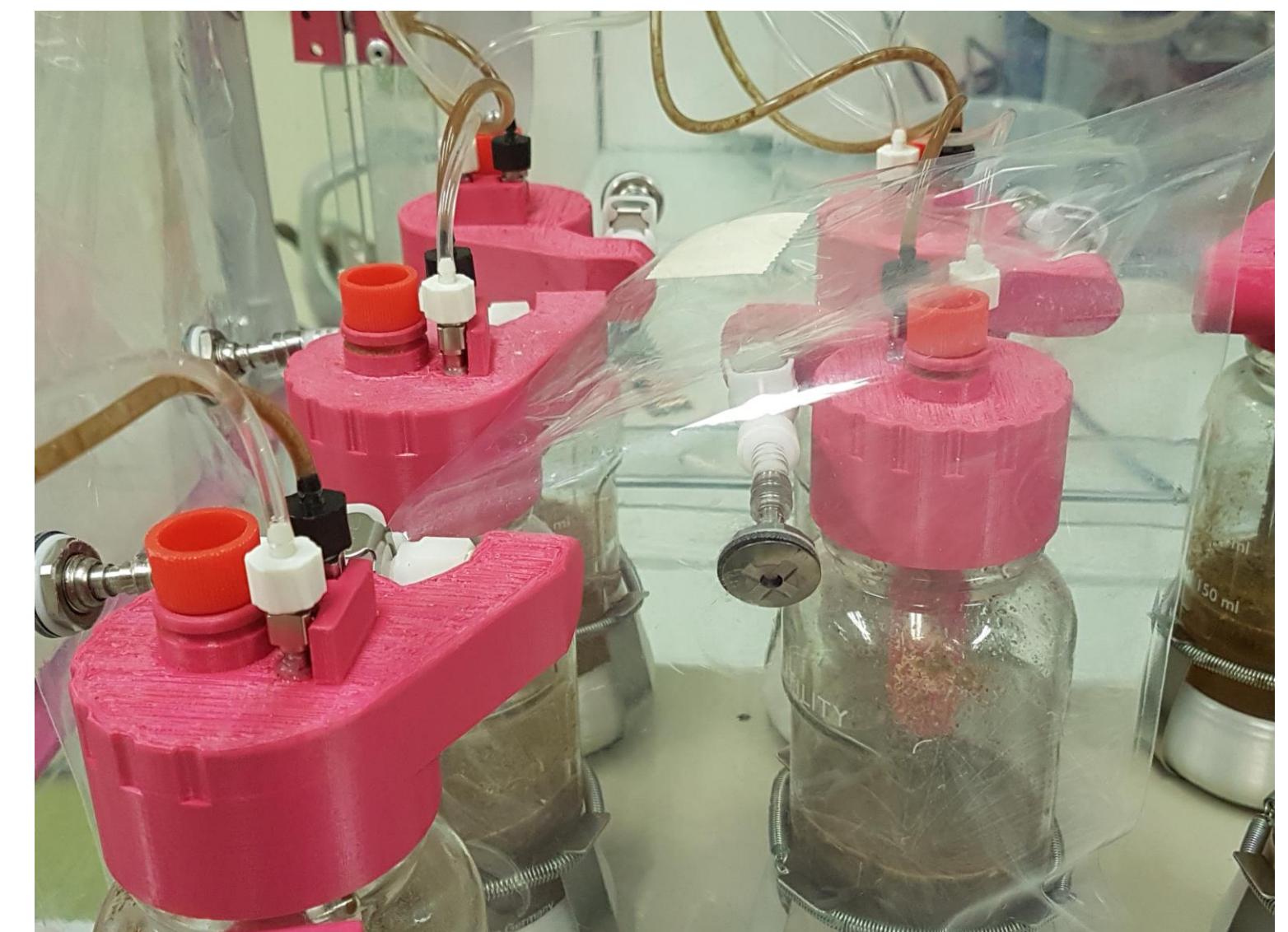
Validating alternatives to rumen cannulation to obtain rumen contents for inoculating *in vitro* devices is necessary due to societal demand for less invasive techniques in animal experimentation

Aim of the study

To compare, in dual-effluent mini-fermenters, stomach tubing (ST) and rumen cannulation (RC) for fermentation parameters methanogenesis microbiota composition

Materials and methods

- ▶ Donors of rumen inoculum : 2 rumen cannulated dry goats
- ▶ Dual-effluent fermenters : working volume 80 mL; liquid (7%/h) and solid (3%/h) turnover rates; artificial saliva at 7 mL/h
- ▶ Four 7-days periods with each condition tested in duplicate
- ▶ Diet (hay 0.6; barley 0.2; soybean meal 0.2) given twice a day at 8:30 and 15:30
- ▶ At the end of each period (days 6 and 7), all 8 fermenters sampled for
 - ▶ fermentation end-products production (CH_4 and SCFA production)
 - ▶ microbiota composition using 16S-rDNA sequencing of V4-V5 regions, FROGS pipeline (Escudé F, Auer L, Bernard M, Mariadassou M, Cauquil L, Vidal K, Maman S, Hernandez-Raquet G, Combes S, Pascal G. 2018. Bioinformatics, 4, 1287–1294).



Results

Fermentation end-products

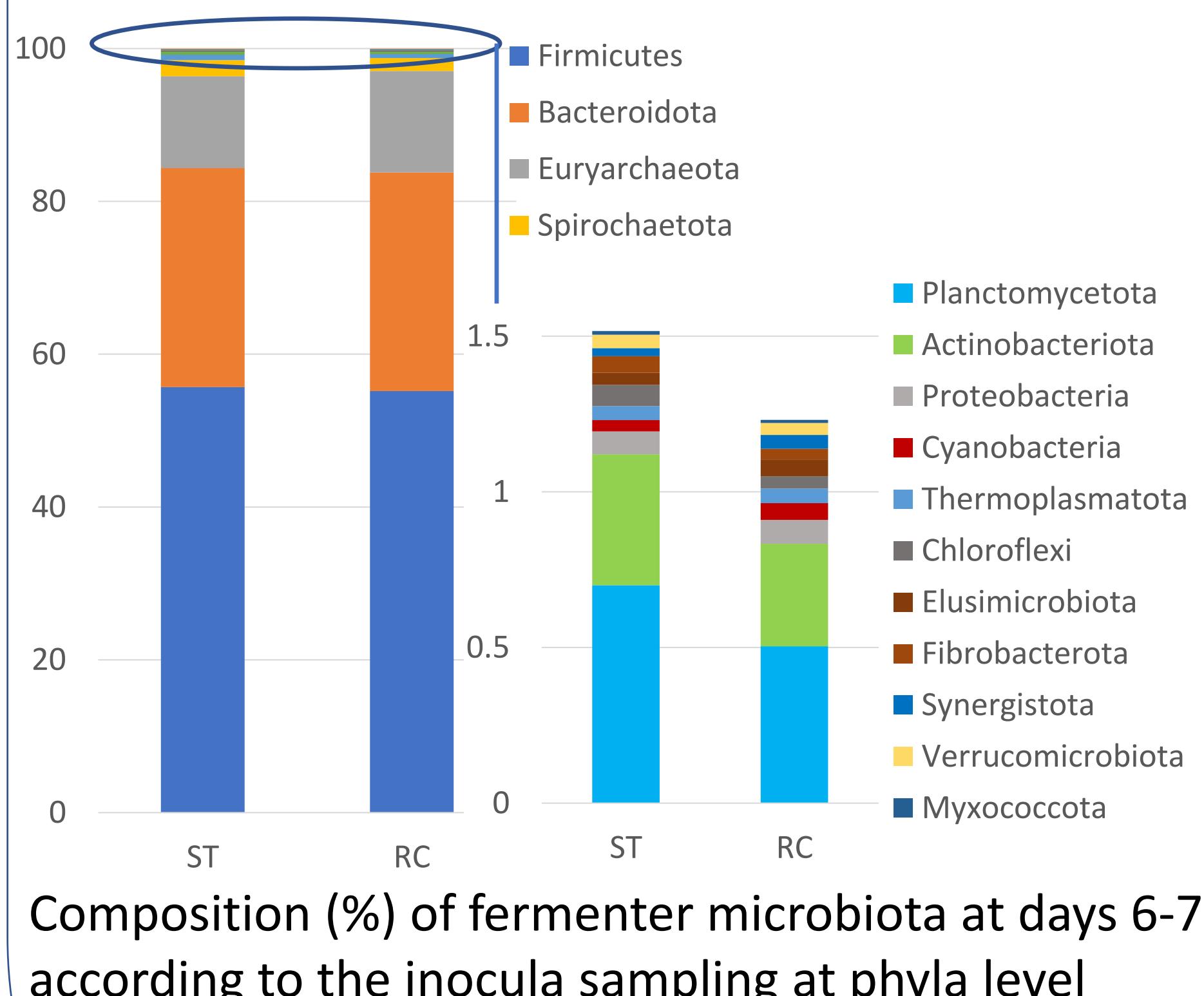
SCFA pattern in inocula, amounts of hexoses fermented (HF) and fermentation pattern in fermenters at days 6-7

	RC	ST	RMSE	P
SCFA in inocula				
Total (mM)	46.9	33.4	6.84	0.008
Acetate, C2 (%)	71.3	70.8	0.92	0.73
Propionate, C3 (%)	19.9	20.1	1.12	0.79
Butyrate, C4 (%)	8.7	9.1	1.39	0.99
HF (mmol/day)	4.52	4.54	0.333	0.57
Specific productions (mol/100mol HF)				
C2	115.7	116.7	3.41	0.45
C3	41.6	41.2	2.15	0.70
C4	21.4	21.1	1.88	0.64
CH ₄	55.5	55.5	0.98	0.98

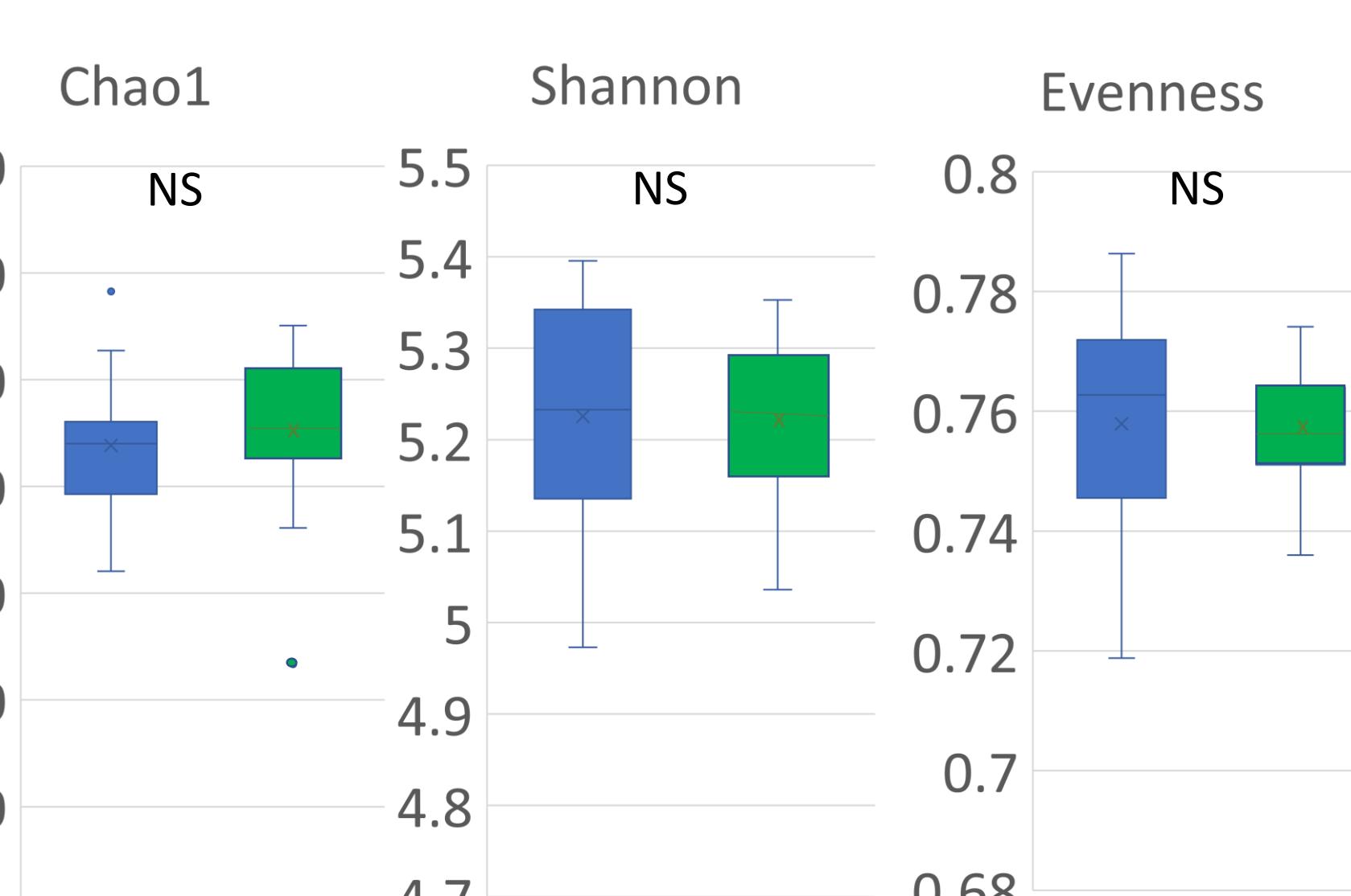
The SCFA concentration of the rumen inocula was lower with ST than with RC because of saliva dilution but with similar proportions of individual SCFA

After 5 days of adaptation, fermentation parameters were not significantly different between ST and RC

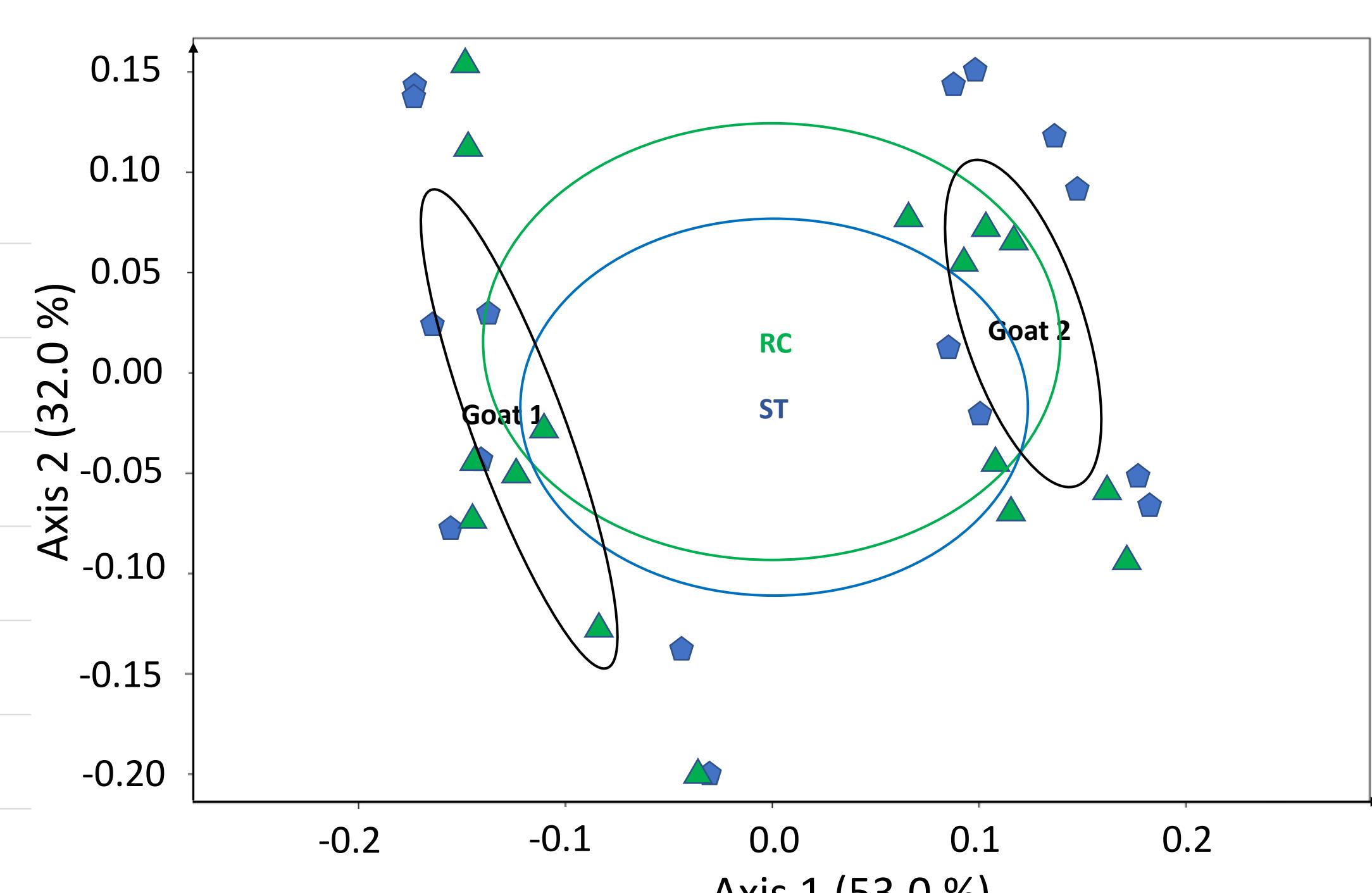
Microbiota composition and diversity



The microbiota composition between ST and RC is similar at the phyla level



Alpha diversity of the fermenter microbiota not significantly different between ST and RC, with indexes closed to *in vivo* measurements



PCoA of inocula microbiota samples based on ASVs community dissimilarities (Bray Curtis) (♦ ST; ▲ RC)

Microbiota community structures similar between SR and RT with a clear clustering by goat

Conclusion

The activity and structure of microbiota in dual-effluent mini-fermenters inoculated with ST and RC were similar. Stomach tubing sampling is a good alternative to rumen cannulation