

# Silicon dynamics through the lens of soil-plant-animal interactions: perspectives for agricultural practices

Félix de Tombeur, Philippe Roux, Jean-Thomas Cornelis

#### ▶ To cite this version:

Félix de Tombeur, Philippe Roux, Jean-Thomas Cornelis. Silicon dynamics through the lens of soil-plant-animal interactions: perspectives for agricultural practices. Plant and Soil, 2021, 467 (1-2), pp.1-28. 10.1007/s11104-021-05076-8. hal-04693933

HAL Id: hal-04693933

https://hal.science/hal-04693933

Submitted on 11 Sep 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

#### 1 Marschner Review

### 2 Silicon dynamics through the lens of soil-plant-animal interactions:

## 3 perspectives for agricultural practices

- 4 Félix de Tombeur<sup>1\*</sup>, Philippe Roux<sup>1</sup>, Jean-Thomas Cornelis<sup>2</sup>
- <sup>1</sup>TERRA Teaching and Research Centre, Gembloux Agro-Bio Tech, University of Liege, Gembloux, Belgium
- <sup>2</sup>Faculty of Land and Food Systems, The University of British Columbia, Vancouver, Canada
- 7 \*Corresponding author

8 9

**ORCID** numbers of the authors: Félix de Tombeur (0000-0002-6012-8458), Philippe Roux (0000-0002-0146-0928), Jean-Thomas Cornélis (0000-0003-0205-7345)

10 11 12

#### Acknowledgements

- We sincerely thank Hans Lambers for inviting us to write this review, and for providing helpful and constructive
- comments before the initial submission. J-T.C and F. dT were supported by 'Fonds National de la Recherche
- 15 Scientifique' of Belgium (FNRS; Research Credit Grant for the project SiCliNG CDR J.0117.18).

16

#### 17 Declarations

- 18 Funding
- 19 J-T.C and F.dT were supported by "Fonds National de la Recherche Scientifique" of Belgium (FNRS; Research
- 20 Credit Grant for the project SiCliNG CDR J.0117.18)
- 21 Conflicts of interest/Competing interests
- The authors declare that there is no conflict of interest.
- 23 Availability of data and material
- 24 All data generated or analyzed during this study are included in this published article and its supplementary
- information files.
- 26 *Code availability*
- Not applicable
- 28 Ethics approval
- Not applicable
- 30 Consent to participate
- 31 Not applicable
- 32 Consent for publication
- 33 Not applicable

34

35

36

37

38

#### Abstract

40

#### 41 Background

- 42 Silicon (Si) is increasingly recognized as a pivotal beneficial element for plants in ecology and
- 43 agricultural sciences, but soil-plant Si cycling has been considered mostly through the prism of
- 44 abiotic mineral weathering, whilst numerous biological processes have been overlooked.
- 45 Leveraging ecological processes that impact soil-plant Si cycling in cropping systems might
- improve crop Si status, but this remains hypothetical to date.

#### 47 *Scope*

- We aim to comprehensively compile information about biotic and abiotic processes driving soil-
- 49 plant Si cycling, and translate their potential beneficial effects in agricultural practices. We
- 50 emphasize the fundamental need to consider the effects of agricultural practices on Si mobility in
- 51 soil-plant systems when striving towards sustainable agroecosystems.

#### Conclusions

52

53

54

55

56

57

58

59

60

61

Regarding soil abiotic factors, degree of soil weathering, mineralogy, texture and pH are key predictors of soil Si dynamics, while soil aggregation processes deserve further investigation. The biological processes associated with mycorrhizal associations, silicate-solubilizing bacteria, and soil macrofauna enhance Si mobility in soil-plant systems, while the effect of root exudates is likely, but deserves further studies. Large herbivores strongly affect soil-plant Si mobility by increasing plant-derived Si turnover rates and redistribution, thereby making integrated crop-livestock systems a promising perspective to improve crop Si status. Recycling crop residues and implementing suitable cover crops promotes Si mobility in soil-plant systems by leveraging the

relatively high solubility of plant-derived Si-bearing minerals. The soil-root-microorganism

- 62 interactions facilitated by cereal-legume intercropping systems also contributes to the mobility of
- 63 Si in the soil-plant continuum. The capacity of certain agricultural practices to increase Si
- 64 mobility in soil-plant systems stresses the need to understand complex soil-plant-animal
- interactions when aiming to enhance Si-based plant stress resistance in agroecosystems.
- 66 **Keywords:** large herbivores; cover crops; intercropping; integrated crop-livestock systems;
- 67 nutrient-acquisition strategies; sustainable agroecosystems; recycling crop residues; facilitation;
- 68 biochar

69

77

81

#### Introduction

- 70 Silicon (Si) is taken up by all vascular plants and contributes to a wide range of functions
- 71 (Epstein 1994). It is deposited as hydrated amorphous silica in plant tissues (SiO<sub>2</sub>.nH<sub>2</sub>O;
- 72 phytoliths) and helps mitigate several plant biotic and abiotic stresses (Ma 2004; Liang et al.
- 73 2007; Zhu and Gong 2014; Cooke and Leishman 2016; Hartley and DeGabriel 2016; Debona et
- al. 2017; Coskun et al. 2019), can be used as a cheap plant structural component (Raven 1983),
- and, eventually, increase plant primary productivity and crop yield (Savant et al. 1999; Liang et
- 76 al. 2015b; Tubana et al. 2016; Xu et al. 2020). The essentiality of Si for plants remains debated
  - and challenging to assess (Epstein 1994; Coskun et al. 2019), but overwhelming evidence show
- 78 its importance for plant functioning. It is therefore increasingly considered a key element in plant
- 79 ecology (Cooke and Leishman 2011; Katz 2019) and agriculture, especially considering the
- 80 importance of Si-accumulating species in global food production (e.g., wheat, rice, sugarcane)
  - (Fig. 1) (Meyer and Keeping 2000; Datnoff et al. 2001; Haynes 2014; Liang et al. 2015b; Tubana
- et al. 2016).

Over the last 30 years, soil scientists and biogeochemists studied Si in a wide range of environments to explore the role of soil properties and vegetation on soil-plant Si cycling (Bartoli 1983; Alexandre et al. 1997; Lucas 2001; Derry et al. 2005; Sommer et al. 2006; Henriet et al. 2008b, a; Cornelis et al. 2010; Haynes 2014; Cornelis and Delvaux 2016; Meunier et al. 2018; Vander Linden and Delvaux 2019; de Tombeur et al. 2020c, a, b; Schaller et al. 2021). The processes and factors governing Si release rates from minerals are well documented (Sommer et al. 2006; Churchman and Lowe 2012; Haynes 2014; Cornelis and Delvaux 2016; Schaller et al. 2021), as are the influence of plant-induced mechanisms (bioweathering and Si uptake) on terrestrial Si cycling (Lucas et al. 1993; Alexandre et al. 1997; Street-Perrott and Barker 2008; Haynes 2017; de Tombeur et al. 2020a). However, how certain aspects of the soil-plant-animal continuum influence Si mobility in both natural systems and agroecosystems have been overlooked, especially the contribution of biotic factors. Yet, a detailed understanding of soilplant-animal interactions influencing Si dynamics is paramount if we seek to benefit from Sirelated plant functioning in agriculture (Acevedo et al. 2021). Despite its ubiquity in soils (2<sup>nd</sup> most abundant element of the Earth's crust; Wedepohl 1995), long-term mineral weathering and subsequent desilication (i.e. Si loss by leaching) result in soils with low plant-available Si concentrations in many areas of the world (approximately 3500 million hectares, as estimated by the land area supporting desilicated soils: Ferralsol, Podzol, Arenosol, Lixisol, Plinthosol, Acrisol and Alisol; WRB 2015), particularly in tropical and subtropical regions where soils tend to be older and leaching intensity greater (Savant et al. 1999). In addition, agriculture further enhances desilication by harvesting and exporting large amount of Si (Desplanques et al. 2006; Struyf et al. 2010; Clymans et al. 2011; Guntzer et al.

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

2012; Keller et al. 2012; Vandevenne et al. 2015; Carey and Fulweiler 2016; Tubana et al. 2016; Vander Linden and Delvaux 2019). These days, rock-derived Si fertilizers are routinely applied in some agroecosystems to counterbalance these detrimental effects (Savant et al. 1999; Datnoff et al. 2001; Haynes 2014; Liang et al. 2015b), and intensifying this practice might even be beneficial in less-weathered and desilicated environments (Tubana et al. 2016). However, such a practice relies on non-renewable resources, and low accessibility to common, rock-derived Si fertilizers in some tropical regions can jeopardize the Si benefits in these sensitive agroecosystems. Harnessing ecological processes that increase soil-plant Si mobility by promoting specific agricultural practices may improve the Si status of crops worldwide, while decreasing the need for non-renewable mineral fertilizers (Lambers et al. 2011; Richardson et al. 2011; Mariotte et al. 2018). Indeed, recent evidence demonstrates the positive impact of certain agricultural practices such as intercropping, cover crops or integrated crop-livestock systems, on nutrient management, especially for phosphorus (P) (Hallama et al. 2019; Tang et al. 2020; Carlos et al. 2020). In this review, we aim to compile knowledge about biotic and abiotic factors that govern Si mobility in soil-plant systems and translate their potential benefits in agricultural practices. We specifically emphasize how overlooked ecological/biological processes are pivotal when favoring Si biocycling in agroecosystems, and advocate the permanent need to nurture our understanding of complex interactions between physico-chemical and biological soil processes to develop

#### Biotic and abiotic factors affecting soil-plant Si cycling

sustainable agroecosystems.

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

#### Physico-chemical processes controlling soil Si dynamics

Over the past decade, many studies reported an increase in plant-available Si concentrations with increasing soil pH when multiple soils were considered together (Phonde et al. 2014; Miles et al. 2014; Puppe et al. 2015; Klotzbücher et al. 2018b; Meunier et al. 2018; Schaller et al. 2018, 2021; Haynes 2019; Caubet et al. 2020; de Tombeur et al. 2020b). Prolonged soil acidification during pedogenesis is associated with the loss of reactive Si-bearing minerals (through mineral dissolution and subsequent lixiviation of dissolved Si) and increased desilication (Savant et al. 1999; Chadwick and Chorover 2001; Sommer et al. 2006; Henriet et al. 2008a; Liang et al. 2015a; de Tombeur et al. 2020b). Consequently, plant-available Si concentrations inevitably decrease with the relative enrichment of poorly weatherable minerals and the decrease in soil pH during pedogenesis (Savant et al. 1999; Liang et al. 2015a; Haynes 2019; de Tombeur et al. 2020b). However, increasing soil pH is also associated to a decrease in plant-available Si concentrations, because Si adsorption onto the surface of oxides and silicates increases gradually to about pH 9.5, reflecting the H<sub>4</sub>SiO<sub>4</sub>/H<sub>3</sub>SiO<sub>4</sub> pKa of 9.47 (Jones and Handreck 1963; McKeague and Cline 1963; Beckwith and Reeve 1964; Hingston and Raupach 1967; Hingston et al. 1972; Obihara and Russell 1972; Philippini et al. 2006; Nguyen et al. 2017; Haynes and Zhou 2018). This adsorption process is also dependent on condensation processes (Schaller et al. 2021) as adsorption of polysilicic acid is less reversible than monosilicic acid (Dietzel 2002). On the other hand, a high pH could also increase plant-available Si concentrations via increasing dissolution rates of aluminosilicates from soil pH 7.5/8 (Drever 1994; Kelly et al. 1998), together with increased phytolith dissolution rates with increasing soil pH from 3 to 10 (Fraysse et al. 2006b, 2009). Overall, although the controls of soil weathering, mineralogy and texture on plantavailable Si are now well understood, especially for the silicate weathering domain (Cornelis and

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

Delvaux 2016; de Tombeur et al. 2020b), how high pH and carbonate minerals influence Si plantavailability remains unclear, because they are driven by antagonistic processes (Haynes 2019).

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

Beyond that, the complex physical organization of soil particles and accessibility to roots teach us how challenging it is to make the assessment of nutrient and Si availability accurate and reproducible. For instance, capturing the effect of condensation state of silicic acid on the measurements of plant-available Si by specific extractions (Schaller et al. 2021) is challenging. In particular, the relation between soil pH and mineralogy should be visited through the lens of polymerization and depolymerization processes influencing the equilibrium from monosilicic acid and polysilicic acid to amorphous silica precipitates over very short time scales (hours to days) (Dietzel 2000). Indeed, polymerization rates of monosilicic acid and its precipitation as amorphous silica increase strongly with increasing pH and ionic strength (Icopini et al. 2005). As suggested by Schaller et al. (2021), the activity of monosilicic acid in soil solution, which is directly related to plant-available Si concentrations, is largely dependent on solid-liquid equilibrium that vary on very short time scales (hours to days), because of daily and seasonal variations of water dynamics in soil porosity (Zabowski and Ugolini 1990). The short-term variations of water content are certainly a key driver of polymerization, complexation and adsorption reactions, which govern the availability of monosilicic acid for plants. The polymers of polysilicic acid and submicrometric colloids of amorphous silica precipitating in soil porosity could in turn influence the water holding capacity and aggregation, by clogging the porosity and cementing particles together (Schaller et al. 2021).

Despite a potential role of amorphous silica on soil aggregation (Schaller et al. 2021), how the interactions between soil mineral and organic constituents (Six et al. 2004) impact the extent to

which minerals can be dissolved in aggregates is not understood. In this regard, Li et al. (2020b) showed that soil microaggregates contribute over 60% of the total phytolith stock in a Retisol. The authors proposed that entrapment of phytoliths in aggregates might slow down their dissolution, and increase their persistence in soils and sediments. In addition, soil aggregates host microbial hotspots in pores or at mineral surfaces creating specific soil micro-environments where microbial processes are accelerated as compared to the average rates in the bulk soil, i.e. the hot moments (Kuzyakov and Blagodatskaya 2015). The bio-induced weathering processes in these hotspots most likely also impact soil Si release rates into the soil solution (Uroz et al. 2009), supporting the need to further investigate the role of soil aggregation on soil Si dynamics. This is of special interest since anthropogenic land transformations (e.g., tillage) substantially impact soil aggregation processes, affecting soil structure (Mikha and Rice 2004; Wright and Hons 2005; Fonte et al. 2014; Or et al. 2021). Besides the role of soil solution chemistry on Si availability in the vicinity of roots, it is also recognized that Si compete with other ions via sorption/desorption mechanisms on soil exchangeable sites (Matychenkov and Ammosova 1996; Klotzbücher et al. 2020). In particular, research has long suggested competition between Si and phosphate ions (Smyth and Sanchez 1980; Kundu et al. 1988; Matychenkov and Ammosova 1996; Owino-Gerroh and Gascho 2004; Konhauser et al. 2007; Reithmaier et al. 2017; Hilbrandt et al. 2019; Hömberg et al. 2020; Klotzbücher et al. 2020; Schaller et al. 2020). Such mechanism could explain the benefits of Si addition on plant P nutrition (Hall and Morison 1906; Fisher 1929; Singh and Sarkar 1992; Owino-Gerroh and Gascho 2004; Eneji et al. 2008; Neu et al. 2017), even though other mechanisms were postulated (Ma and Takahashi 1990a, b, 1991a; Kostic et al. 2017; Pavlovic et

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

al. 2021). In addition to P, Si may also compete with dissolved organic matter (Reithmaier et al. 2017; Klotzbücher et al. 2020), selenium (Jordan et al. 2009), arsenic (Christl et al. 2012) and iron (Hömberg et al. 2020). Such sorption/desorption competitive mechanisms are pH-dependent, with rates decreasing with pH given the lower chemical reactivity of monosilicic acid at acidic conditions (Owino-Gerroh and Gascho 2004; Konhauser et al. 2007; Jordan et al. 2009; Christl et al. 2012). It is therefore key to buffer the pH of the sorption/desorption isotherms because it is certainly influenced by the use of alkaline Si fertilizers (e.g., calcium silicates).

Overall, although the controls of soil weathering degree and mineralogy on soil Si dynamics are now well understood (Cornelis and Delvaux 2016), more attention should be given to the process of polymerizing silicic acid from monosilicic and polysilicic acid to amorphous silica precipitates (Schaller et al. 2021). Soil porosity, as well pore tortuosity and connectivity, should also be considered for their influence on monosilicic acid activity in soil solution, as should soil aggregation. Finally, while the size and type of minerals as well as the ionic composition of pore water together control dissolution kinetics, furthering how temporal variations of water content (hourly to seasonally changes) affect plant-available Si is certainly worthwhile goal. Getting closer to the actual plant-availability of Si will therefore require to reconcile microscale soil processes with the extractable solutions used to assess Si concentrations in soil solution.

#### Biological processes controlling soil Si dynamics

*Silicon biocycling: the high reactivity of phytoliths* 

- In addition to the weathering of rock-derived minerals, the dissolution of soil phytogenic silicates
- 313 (phytoliths) also strongly impact Si dynamics (Bartoli 1983; Alexandre et al. 1997; Meunier et al.
- 314 1999; Derry et al. 2005; Farmer et al. 2005; Sommer et al. 2013; de Tombeur et al. 2020a).

Biogeochemical mass-balance calculations have long reported that a significant fraction of Si in the soil solution is derived from the dissolution of the phytogenic Si pool (Bartoli 1983; Alexandre et al. 1997, 2011; Gérard et al. 2008), because of its high solubility compared with that of crystalline Si-bearing minerals (Fraysse et al. 2006b, 2009; Cornelis and Delvaux 2016). This challenged the common view that plant-available Si concentrations were mainly driven by soil parent material, weathering degree, and subsequent soil mineralogy/texture (Savant et al. 1999; Chadwick and Chorover 2001; Henriet et al. 2008a, b; de Tombeur et al. 2020b). To reconcile the control of geochemical and biological processes on Si release in the soil solution, Cornelis and Delvaux (2016) suggested that the biological Si feedback loop (phytolith formation in plants and dissolution in soils) takes over soil litho/pedogenic pools in advanced soil weathering stages. This contention was recently supported by the use of long-term soil chronosequences where plantavailable Si concentrations are mainly governed by soil-derived Si-bearing minerals (clay minerals) in early and intermediate stages of weathering (de Tombeur et al. 2020b), but increasingly by the recycling of phytoliths in old and highly-weathered soils dominated by poorly-soluble quartz minerals (de Tombeur et al. 2020a). The significant effect of vegetation on the soil-plant Si cycle explains why land-use changes and management affect the global Si cycle (Struyf et al. 2010; Clymans et al. 2011; Vandevenne et al. 2015; Carey and Fulweiler 2016). Some soil organisms also accumulate Si to form various siliceous structures (Ehrlich et al. 2010; Puppe 2020). They are classified as zoogenic Si pool (e.g., sponge spicules that get into the soils mostly from freshwaters through aeolian transport or in situ production; Łukowiak 2020), bacterial Si (e.g., Proteus mirabilis, Lauwers and Heinen 1974), fungal Si, protozoic Si (e.g., testate amoeba shells) or protophytic Si (e.g., diatom frustules) (Sommer et al. 2006; Ehrlich et

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

al. 2010; Puppe et al. 2015; Puppe 2020). Recent evidence suggest that these pools are of a similar magnitude as the phytogenic pool, and in turn influence the terrestrial Si cycle (Sommer et al. 2013; Puppe et al. 2014, 2015, 2016). In particular, annual biosilicification from testate amoebae ranges from 17 to 80 kg ha<sup>-1</sup> yr<sup>-1</sup> depending on soil and ecosystem properties which is similar to or even exceeds annual Si uptake by terrestrial vegetation (Sommer et al. 2013; Puppe et al. 2015; Vander Linden and Delvaux 2019). This pioneering work opened new perspectives on the role of Si-based life forms on soil-plant Si cycling (Puppe 2020).

Biological weathering of Si-bearing minerals

In addition to the production of an easily weatherable Si pool, plants are extremely active when it comes to enhancing weathering ability of different soil constituents, either directly or indirectly. Nevertheless, plants are not stand-alone entities in their ability to affect Si cycling, but should be considered as "holobionts", which includes the microbiome associated with their development (Vandenkoornhuyse et al. 2015). There are numerous well-known mechanisms by which the actors of the soil-plant continuum contribute to the dissolution of Si-bearing minerals and extensive reviews can be found elsewhere (Finlay et al. 2020; Dontsova et al. 2020). Briefly, these mechanisms can be divided into two categories: biochemical and biophysical weathering. These processes are not mutually independent; rather, they often play cumulative or synergistic roles.

The main biochemical effect on Si dissolution is the modification of the chemical conditions in the rhizosphere. This occurs through the release of organic acids, either as a by-product of cellular metabolism or as root exudates, plant excretion of H<sup>+</sup> in exchange for cationic nutrients, formation of carbonic acid (Golubev et al. 2005; Brantley 2008) through the release of CO<sub>2</sub> via

root respiration or organic matter mineralization, and the release of inorganic acids from redox reactions (e.g. sulfuric acid via pyrite oxidation). Proton-promoted dissolution is supplemented by ligand-promoted dissolution where organic acids, in addition to their pH altering ability, can act synergistically with strong chelators such as phytosiderophores or carboxylates to further enhance weathering of Si-bearing minerals by destabilizing mineral lattices through the binding of metal cations (Bennett et al. 2001; Buss et al. 2007; Pastore et al. 2020). Biophysical mechanisms include hyphal tunneling, or boring and other mechanisms of penetration by plant roots or fungi along mineral weakness points (Smits et al. 2005; van Schöll et al. 2008; Teodoro et al. 2019). This, in turn, increases substrate porosity and, therefore, increases the mineral surface exposed to chemical weathering agents (Pawlik et al. 2016; Gadd 2017). Finally, plants and associated microorganisms can also affect water movement and retention capacity through uptake and biofilms and therefore strongly influence water residence time and weathering patterns (Lucas 2001; Flemming and Wingender 2010). While the biological impact on weathering is recognized, its contribution to the mobility of nutrients in natural and agricultural systems, including Si, remains poorly understood. In the following, we therefore aim to assess the effects of each biological agent on Si plant-availability. Root exudates Plant roots secrete a wide range of exudates that mobilize poorly-available nutrients in the rhizosphere (Dakora and Phillips 2002; Lambers et al. 2006; Finlay et al. 2020).

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

nutrients in the rhizosphere (Dakora and Phillips 2002; Lambers et al. 2006; Finlay et al. 2020). A long history of experimental studies has shown the increase of silicate dissolution by root exudates, through organic acids (Stillings et al. 1996; Drever and Stillings 1997; Cama and Ganor 2006; Bray et al. 2015) or forest floor extracts (van Hees et al. 2002). Increasing dissolution of silicates or pedogenic oxides in the presence of siderophores was also demonstrated, whether

they are microbial (Liermann et al. 2000; Buss et al. 2007) or root-derived (Reichard et al. 2005). However, despite their importance for plant nutrient acquisition, the role of root exudates in mobilizing Si for plant uptake has been overlooked. Yet root-released carboxylates do increase the mobility of P, K, Fe, Al, Ca, P and numerous micronutrients (Gerke et al. 1994; Ström et al. 2005; Wang et al. 2011; Houben and Sonnet 2012; Abrahão et al. 2014; Colombo et al. 2014; Kabas et al. 2017; Teodoro et al. 2019), and there could be similar effects on soil-plant Si mobility. Early work by Hinsinger et al. (2001) has shown the impact of banana (*Musa paradisiaca*), maize (Zea mays), canola (Brassica napus), and white lupin (Lupinus albus) on the weathering of a basaltic rock. After 36 days of growth, the amount of Si released from basalt in the presence of hydroponically grown plants was increased two-fold compared with the abiotic control. More recently, Burghelea et al. (2015) and Zaharescu et al. (2019) showed that buffalo grass (Bouteloua dactyloides) grown on schist and rhyolite for 124 to 603 days, respectively, increased the mobility of Si compared with that of an abiotic control. Furthermore, Gattullo et al. (2016) showed that Fe-deprived barley (Hordeum vulgare) plants rapidly released more exudates into the rhizosphere to mobilize Fe from amorphous Fe oxides. Then, when the soil-plant contact was extended to 12 days, plants overcame Fe nutritional stress and the exudation of organic ligands mobilized Si from smectite (Gattullo et al. 2016). These results demonstrate that root exudates are primarily influenced by macro- or micronutrients limitation, but that co-solubilization of Si is very likely (de Tombeur et al. 2021b). In support of this claim, recent studies showed an increase in leaf Si concentrations with decreasing soil P concentrations and Si plant-availability along a

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

long-term soil chronosequence (de Tombeur et al. 2020a, 2021c), particularly in old and highly-

weathered environments where carboxylate-releasing strategies are common (Lambers et al. 2008; Zemunik et al. 2015). This result suggests a role for carboxylates in mobilizing soil Si from poorly-soluble forms for plant uptake, as it is the case for manganese (Mn) along the Jurien Bay chronosequence (Hayes et al. 2014; Lambers et al. 2015; de Tombeur et al. 2021b). Future research is required to elucidate to which extent Si is co-mobilized by different nutrient-acquisition strategies. In addition, future studies should test if root exudation patterns are directly influenced by low Si availability.

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

Mycorrhizal associations With nearly 90% of plants harbouring either arbuscular mycorrhizal (AM) or ectomycorrhizal fungi (EM) symbionts (Brundrett 2002; Smith and Read 2008), mycorrhizas exhibit strong control over major ecosystem processes including plant nutrient acquisition (Marschner and Dell 1994; Clark and Zeto 2000; Richardson et al. 2009), biogeochemical cycles (Högberg et al. 2001; van Hees et al. 2006), plant diversity and productivity (Van Der Heijden et al. 1998, 2008) and weathering potential (Leake and Read 2017; Smits and Wallander 2017). Plants enable mycorrhizal fungal growth and activity by translocating various organic compounds (sugars, lipids) into the roots (Jiang et al. 2017; Rich et al. 2017). In exchange of plant photosynthates, fungi develop hyphal networks into the soil and enhance weathering processes for lithogenic nutrient acquisition (Van Breemen et al. 2000; van Schöll et al. 2006, 2008). In 1990, Kothari et al. provided the first evidence of Si mobilization by mycorrhizal fungi by showing an increase in maize (Zea mays) root Si concentrations after inoculation with an arbuscular mycorrhizal fungi (Kothari et al. 1990). Since then, other studies have shown that the presence of mycorrhizal fungi may significantly increase the Si concentrations of different species (maize, sugarcane, banana, chickpea, pigeon pea, soybean),

and in different plant organs (roots, leaves, stems) (Table 1) (Yost and Fox 1982; Clark and Zeto 1996; Garg and Bhandari 2016; Oye Anda et al. 2016; Frew et al. 2017b, a, 2020; Garg and Singh 2018; Gbongue et al. 2019). Root Si concentrations are positively correlated with the degree of arbuscular mycorrhizal colonization, which reduces root herbivory (Frew et al. 2017a). Moreover, mycorrhizal colonization increases root Si concentrations, but uniquely for plants growing on soils with low concentrations of plant-available Si (Frew et al. 2017a). This pattern was confirmed by other studies where the effects of mycorrhizal fungi on plant Si concentrations were less important, or even absent, when Si was supplied to plants (Oye Anda et al. 2016; Frew et al. 2017b). Even if the effects of mycorrhizas on plant Si concentration depend on initial Si availability in soil, these results reveal that root mycorrhizal colonization can be a significant driver of plant Si uptake and concentrations in plants, with a direct impact on herbivory. In some cases, plant Si concentrations have indeed more than doubled after inoculation with mycorrhizal fungi (Oye Anda et al. 2016). More broadly, although phylogenetic variation and the presence or absence of Si transporters remain the main explanations for variation in plant Si accumulation (Hodson et al. 2005; Ma et al. 2006, 2007; Deshmukh and Bélanger 2016; Deshmukh et al. 2020), nutrientacquisition strategies like mycorrhizal associations and root-released carboxylates could play a

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

significant, but so far overlooked role. This is of special interest since both strategies increase

with decreasing P availability (Abbott et al. 1984; Tang et al. 2001; Covacevich et al. 2007; He et

al. 2020), and P-depleted soils are often also Si-depleted, due to high weathering degree.

Plant	Organ	AM fungi	Increase in plant [Si] (%)	Reference
Maize (Zea mays)	Shoot	Glomus sp.	+3% to +66% for acid soils	Clark and Zeto
		•	-28% to -6% for alkaline soils	(1996)
			+78% to +252% of Si content (weight by plant)	
			for acid soils	
			-9% to +25% of Si content (weight by plant) for	
			alkaline soils	
Chickpea (Cicer arietinum)	Shoot	Funneliformis	+17-20%	Garg and Bhandari
D (11	ъ .	mosseae	1.400/ - 1:1	(2016)
Banana (Musa acuminata)	Pseudostem	Rhizophagus	+149% without Si	Oye Anda et al.
Danana (Musa saunin sta)	Leaves	irregularis	+88% with Si +84% without Si	(2016) Oye Anda et al.
Banana (Musa acuminata)	Leaves	Rhizophagus irregularis	+84% without Si +70% with Si	(2016)
Banana (Musa acuminata)	Roots	Rhizophagus	+109% without Si	Oye Anda et al.
Banana (Musa acaminaia)	Roots	irregularis	+30% with Si	(2016)
Sugarcane (Saccharum spp.	Roots	Glomus sp.	+50% to +102% without Si	Frew et al. (2017b)
hybrid)	110010	Gromms sp.	-4% to +22% with Si	110 00 a (20170)
Sugarcane (Saccharum spp.	Roots	Glomus sp.	+42% to +71% for the low Si soil	Frew et al. (2017a)
hybrid)		•	+0% to $+18%$ for the high Si soil (ns)	
Sugarcane (Saccharum spp.	Leaves	Glomus sp.	-29% to -21% for the low Si soil (ns)	Frew et al. (2017a)
hybrid)			-20% to -4% for the low Si soil (ns)	
Pigeon pea (Cajanus cajan)	Leaves	Rhizophagus	+10%	Garg and Singh
		irregularis		(2018)
Banana (Musa acuminata)	Roots	Rhizophagus	+30% (Si content; weight by plant)	Gbongue et al. (2019)
		irregularis		
Banana (Musa acuminata)	Leaves	Rhizophagus irregularis	+14% (Si content; weight by plant)	Gbongue et al. (2019)
Soybean (Glycine max)	Leaves	Unknown	+28% to +208% (depending on P supply)	Yost and Fox (1982)

'ns' stands for "not significant"

**Bacteria** Bacteria may colonize mineral surfaces, initiate or accelerate weathering, and stimulate plant growth (Jackson 1971; Bosecker 1997; Banfield et al. 1999; Bennett et al. 2001; Vessey 2003; Calvaruso et al. 2006; Uroz et al. 2009; Burghelea et al. 2015; Zaharescu et al. 2019;

Finlay et al. 2020; Pastore et al. 2020). Bacteria often associate with fungi in soil to form biofilms on substrate surfaces via excretion of extracellular polymeric substances (EPS) which causes very localized weathering "hotspots" (Flemming and Wingender 2010; Deveau et al. 2018; Guennoc et al. 2018; Finlay et al. 2020). The subsequent dissolution of lithogenic nutrients can therefore be used by all organisms of these hotspots including plants, making the soil-plant continuum a very effective biogeochemical engineer and enhancing overall plant nutrition. Increased rates of weathering and Si release in the presence of certain bacteria have been demonstrated for different mineralogical contents such as feldspar (Barker et al. 1998; Welch and Ullman 1999; Wang et al. 2015), hornblende (Liermann et al. 2000), mica (Barker et al. 1998; Liu et al. 2006; Wang et al. 2015), smectite (Dong et al. 2003; Kim et al. 2004), amorphous silica (diatoms and sponge) (Bidle and Azam 1999; Schröer et al. 2003), granite (Song et al. 2007; Wu et al. 2008), basalt (Pastore et al. 2020), gneiss (Pastore et al. 2020) and saprolite (Brucker et al. 2020). These days, a large number of silicate-solubilizing bacteria (SSB) have been identified, belonging to different genera: Aeromonas, Aminobacter, Azotobacter, Bacillus, Burkholderia, Cellvibrio, Collimonas, Dyella, Ensifer, Enterobacter, Flavobacterium, Frateuria, Janthinobacterium, Kosakonia, Labrys, Microbacterium, Paracoccus, Proteus, Pseudomonas, Rhizobium and Sphingomonas (Uroz et al. 2009; Meena et al. 2014; Hu et al. 2018). Recently, Raturi et al. (2021) reviewed the role of SSB on the Si biogeochemical cycle and their role as potential biofertilizers. In a pioneering work, Zahra et al. (1984) showed that soil inoculation with Bacillus circulans significantly increases Si release from different minerals and subsequent Si uptake by different crop species (barley, maize and clover), demonstrating a key role of bacteria in increasing plant-

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

479

available Si. More recently, an increasing number of studies demonstrated the impact of SSB on

Si dissolution rates, plant Si uptake, and a potential positive effect on plant growth (Table 2 and references therein). In these studies, soluble Si concentrations increased by up to 60%, while plant Si content increased by up to 78% (Table 2). Besides the role of SSB in increasing silicates dissolution, bacteria could also increase phytoliths dissolution and soluble Si through the breaking down and mineralization of phytolith-occluded carbon (Hodson 2019). Future research is needed to identify the abundance, diversity and functions of SSB in different geopedoclimatic contexts, as well as their ability to stimulate soil-plant Si mobility.

**Table 2** – Effect of silicate-solubilizing bacteria on soluble/plant-available Si and leaf Si concentrations from the literature. We used WebPlotDigitizer to extract data published as figures (Rohatgi 2012).

Plant	Organ	Bacteria	Increase in soluble [Si] (%)	Increase in plant [Si] (%)	Reference
Rice (Oryza sativa)	Leaf	Enterobacter ludwigii GAK2	Not determined	+24%	Lee et al. (2019)
Rice (Oryza sativa)	Leaf	Rhizobium sp. (IIRR-1)	from +12.4 to +60.2%, depending on silicates	from +9.0% to +78.5% of Si content (weight by plant), depending on silicates	Chandrakala et al. (2019)
Maize (Zea mays)	Leaf and root	Kosakonia sp.	+10%	+23% for both leaf and root	Hu et al. (2019)
Rice (Oryza sativa)	Leaf	Bacillus amyloliquefaciens	Not determined	+29%	Bist et al. (2020)
Rice (Oryza sativa)	Whole plant	Burkholderia eburnea CS4-2	Not determined	+24%	Kang et al. (2017)
Maize (Zea mays)	Leaf and root	Flavobacterium sp.	+16%	ns for leaves; +20% for roots	Hu et al. (2018)

'ns' stands for "not significant"

491 Soil macrofauna

Soil macrofauna like earthworms, beetles and termites contribute to nutrient cycling, soil formation or primary production (Jouquet et al. 2011; Blouin et al. 2013), but little is known about their effect on soil Si dynamics. Yet, soil macrofauna enhances the mineralization of organic matter (Ingham et al. 1985; Schulmann and Tiunov, Alexei 1999), which might in turn accelerate Si release in soil solution through enhanced phytoliths dissolution (Fraysse et al. 2006a, 2010; Vandevenne et al. 2013). Macrofauna also increase the chemical (Jouquet et al.

2002; Carpenter et al. 2007) and physical (Suzuki et al. 2003) weathering of silicate minerals. Recently, Bityutskii et al. (2016) showed that earthworm casts in a sandy and sandy loam soils had a significantly higher soluble Si concentration than the non-bioturbed soil (up to 12 times for Lumbricus terrestris casts in the sandy soil). Moreover, the concentrations and translocation rates of Si in the xylem sap of maize and cucumber plants significantly increased when plants grew on a soil previously bioturbed by earthworms. Following this research, Hu et al. (2018) isolated SSB from the gut of *Pheretima guillelmi*, and showed that they markedly increased the release of Si from feldspar and quartz powder, enhanced the uptake and accumulation of Si by maize, and promoted seedling growth. In addition, significantly more SSB were found in the earthworm gut than in the surrounding soil. The authors thus demonstrated that the increased soil-to-plant translocation of Si following earthworm activity was at least in part explained by the presence of SSB in earthworm guts (Hu et al. 2018). In accordance with this, Georgiadis et al. (2019) showed that the release of dissolved Si from quartz, which is highly resistant to weathering (Goldich 1938), was much greater after passage through the gut of Eisenia andrei. They discussed this as resulting from a combination of mechanical alteration (Suzuki et al. 2003), and the presence of SSB in the earthworm gut (Hu et al. 2018). In addition, soil macrofauna strongly impact the redistribution of material in the soil profile (Jouquet et al. 2011; Blouin et al. 2013). For instance, Jouquet et al. (2020) showed that termite activity impacted the distribution of phytoliths and clay minerals type (1:1 versus 2:1) in south Indian forest soils which could, in turn, impact the concentration of plant-available Si in soil profiles. Overall, soil macrofauna have a significant but overlooked effect on soil Si dynamics, whose magnitude still needs to be determined.

#### Large herbivores

498

499

500

501

502

503

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

Large herbivores can cause important changes in ecosystem-scale nutrient cycling (Bardgett and Wardle 2003; Veldhuis et al. 2018; Forbes et al. 2019; Hwang and Metcalfe 2021). They can either accelerate nutrient cycling through the conversion of aboveground biomass into labile waste products, or decrease it through selective foraging and subsequent shifts towards species that decompose more slowly (Bardgett and Wardle 2003; Forbes et al. 2019). Compared with N, P and C (e.g., Veldhuis et al. 2018; le Roux et al. 2020; Sitters et al. 2020), the impact of large herbivores on Si dynamics is poorly quantified. Yet faeces of large herbivores exhibit high silica concentrations (from 17 to 163 g silica kg<sup>-1</sup> for large African herbivores; Hummel et al. 2011), as does sheep urine (up to 259 mg silica L<sup>-1</sup>; Nottle and Armstrong 1966). As a consequence, large herbivores strongly impact the land-to-ocean Si transfer by foraging grasses and transporting phytoliths from land ecosystems directly to rivers (Schoelynck et al. 2019). For instance, hippos of south-western Kenya contribute to 32% to the biogenic Si flux and more than 76% to the total Si flux to watercourses (Schoelynck et al. 2019). Given the abundance of Si-accumulating species in grassland ecosystems, the impact of large herbivores on phytolith redistribution within the same ecosystem (not land-to-river transfer; Schoelynck et al. 2019) is probably also significant. We estimate that large herbivores in a savanna ecosystem ingest and displace from  $0.005 \pm 0.002$ kg Si ha<sup>-1</sup> yr<sup>-1</sup> (grey duiker) to  $23.2 \pm 3.8$  kg Si ha<sup>-1</sup> yr<sup>-1</sup> (buffalo) (Table 3). The higher value corresponds to yearly litterfall of a short grass ecosystem of the Central Great Plains, USA (Blecker et al. 2006). This redistribution of phytoliths could, in turn, modify the spatial variability of plant-available Si given their high reactivity in soil environments (Alexandre et al. 1997; Blecker et al. 2006; Sommer et al. 2013; de Tombeur et al. 2020a).

520

521

522

523

524

525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

**Table 3** – Estimation of yearly silicon (Si) inputs into soil by large herbivores in a savanna ecosystem, and their potential effect on Si cycling.

Species	Scientific name	<b>Dung produced</b> <sup>a</sup> g ha <sup>-1</sup> day <sup>-1</sup>	Dung [BSi] <sup>b</sup> g kg <sup>-1</sup>	<b>Dung Si input</b> <sup>c</sup> kg Si ha <sup>-1</sup> yr <sup>-1</sup>	MRT <sub>particle</sub> <sup>d</sup> hours	Effect on Si cycling <sup>e</sup>
Buffalo (ruminant)	Syncerus caffer	1018	$134 \pm 22$	$23.2 \pm 3.8$	49	+++
Elephant	Loxodonta Africana	432	$53 \pm 11$	$3.9 \pm 0.8$	30	++
Grey duiker (ruminant)	Giraffa camelopardalis	1.3	$22 \pm 9$	$0.0 \pm 0.0$	45	+/-
Impala (ruminant)	Aepyceros melampus	200	$123 \pm 49$	$4.2 \pm 1.7$	-	++
Nyala (ruminant)	Tragelaphus angasii	8.6	$38 \pm 13$	$0.1 \pm 0.0$	-	+/-
White rhino	Ceratotherium simum	124	$75 \pm 13$	$1.6 \pm 0.3$	44	+
Wildebeest (ruminant)	Connochaetes taurinus	53	$135 \pm 15$	$1.2 \pm 0.1$	-	+
Zebra	Equus burchellii	77	$126 \pm 18$	$1.6 \pm 0.2$	28	+

<sup>&</sup>lt;sup>a</sup>Data from Veldhuis et al. (2018)

Although silica ingestion can reduce the apparent digestibility of herbage (Shewmaker et al. 1989; Hartley and DeGabriel 2016; Johnson et al. 2021), how phytoliths are processed during digestion remains poorly known. Some herbivores have a neutral to slightly alkaline stomach (e.g., pH 7.3 for *Lama guanicoe*; Beasley et al. 2015) and phytolith dissolution rates increase significantly from pH 5 to pH 8 (Fraysse et al. 2009). In 1971, Blackman & Bailey showed that up to 39% of silica was dissolved after 24 h of ingestion in a cow rumen (Blackman and Bailey 1971). More recently, Vandevenne et al. (2013) showed that phytolith concentrations in cow, sheep, horse and donkey faeces were two to four times higher than those in the plants prior to ingestion and from which the faeces were derived. They also showed that readily-soluble Si concentrations increased in faeces compared with pasture forage (except for horse faeces). Moreover, relative to the initial phytolith content in dungs, 60%, 16% and 8% of Si was mobilized in rain water after 24 h for cow, horse and sheep faeces, respectively, but only 4% for

<sup>&</sup>lt;sup>b</sup>Data from Hummel et al. (2011). Means of wet and dry season data were considered.

<sup>&</sup>lt;sup>c</sup>BSi was converted to Si by dividing by 2.14.

<sup>&</sup>lt;sup>d</sup>Food particles' mean retention time (MRT) comes from Steuer et al. (2011) for Buffalo, Elephant, and Zebra, and from Müller et al. (2011) for Grey duiker and White rhino

<sup>&</sup>lt;sup>e</sup>Estimated through the combination of yearly dung Si input and MRT<sub>particle</sub> control on phytolith turnover rates (Vandevenne et al. 2013).

the corresponding pasture forage. These results demonstrate that herbivores have a strong potential to increase Si mobility in soil-plant systems by releasing dissolved Si in urine and increasing phytolith turnover rates, probably through the degradation of plant tissues and an onset of phytolith dissolution in the digestive tract (Vandevenne et al. 2013).

Vandevenne et al. (2013) suggested that ruminants (sheep and cows), which achieve greater particle size reduction through higher food particles' mean retention time (MRT) (Johnson et al. 2021), have a greater potential to quickly mobilize the highly-soluble fraction of phytoliths, partly via urine, compared to non-ruminants (horse and donkey). Following this idea, ruminants (that have longer food particles' MRT), such as buffalo or duiker, would increase the phytoliths turnover more strongly than non-ruminants such as elephant or zebra do (Table 3). Therefore, ruminants that produce large amounts of dung – which largely depends on body mass (Veldhuis et al. 2018) – will have a greater impact on soil-plant Si dynamics, and eventually on land-to-ocean Si transfer (Vandevenne et al. 2013), compared with non-ruminants that produce moderate amounts of dung (Table 3). Overall, large herbivores play a significant but overlooked role in Si biogeochemistry (but see Hwang and Metcalfe 2021) by affecting phytolith turnover rates and distribution in terrestrial ecosystems.

#### Silicon and agricultural practices

Knowledge acquired from complex natural systems can be used to increase the resource-use efficiency and productivity of modern agroecosystems (Lambers et al. 2011; Mariotte et al. 2018). After highlighting biotic and abiotic factors influencing soil-plant Si mobility (Fig. 2), next we discuss the potential of certain agricultural practices to impact soil Si dynamics and stimulate soil-plant Si cycling.

589

590

591

592

593

594

595

596

597

598

599

600

601

602

603

604

605

606

607

608

#### Leveraging the high reactivity of phytoliths: recycling crop residues

Recycling agricultural residues such as leaves, stems or grain husks is key to improve the crop Si status and limit long-term desilication (Guntzer et al. 2012; Meharg and Meharg 2015; Haynes 2017; Puppe et al. 2021). Annual Si uptake by crop species may be an order of magnitude higher than that in natural ecosystems (Vander Linden and Delyaux 2019) and removing crop residues at harvest is common in some parts of the world (Klotzbücher et al. 2015) which lowers the soil phytolith pool (Desplanques et al. 2006; Guntzer et al. 2012; Keller et al. 2012). The application of different crop residues to soil increases Na<sub>2</sub>CO<sub>3</sub>-extractable Si (up to 37% in Yang et al. 2020), water-soluble Si concentrations (up to 15% in Ma and Takahashi 1991b; up to 50% in Watanabe et al. 2017; up to 44% in Yang et al. 2020), plant Si concentration (up to 17% in Ma and Takahashi 1991b; up to 136% in Sistani et al. 1997; up to 168% in Hossain et al. 2001; up to 57% in Yang et al. 2020), and plant Si uptake (up to 25% in Ma and Takahashi 1991b; up to 212% in Marxen et al. 2016). The same pattern occurs for manure, which increases acetic acidextractable Si and NaOH-extractable Si concentrations by 101% and 32%, respectively, after 10 years of application to a Gleysol (Song et al. 2014). Klotzbücher et al. (2018b) also found that manure application tends to increase acetate-extractable Si concentrations, especially together with liming. These results highlight the benefit of returning phytoliths to topsoil because of their fast dissolution rates to replenish the soil solution in dissolved Si (Wickramasinghe and Rowell 2006; Seyfferth et al. 2013; Marxen et al. 2016; Klotzbücher et al. 2018a; Puppe et al. 2021),

with subsequent positive impacts on plant Si uptake. On the other hand, returning carbon-rich

phytoliths to topsoil could also possibly contribute to C sequestration, even though many questions about the phytoliths ability to sequester C on the long-term remain (Hodson 2019). Recently, the use of pyrolyzed Si-rich crop residues (i.e. Si-rich biochar) as a potential alternative to common Si fertilizers has attracted a lot of attention (Xiao et al. 2014; Li and Delvaux 2019; Wang et al. 2019b for reviews). According to Li and Delvaux (2019), the pyrolysis has the advantage of (1) concentrating Si in biochar compared with unpyrolyzed crop residues; (2) providing the same effect as liming and (3) enhancing the reactive surface area; both (2) and (3) contribute to increasing phytolith dissolution. Numerous recent studies show that the application of different biochars increases soil soluble Si and/or plant Si concentrations (Houben et al. 2014; Liu et al. 2014; Ibrahim et al. 2016; Qian et al. 2016; Koyama and Hayashi 2017; Abbas et al. 2017; Alvarez-Campos et al. 2018; Li et al. 2018; Limmer et al. 2018; Leksungnoen et al. 2019; Li et al. 2019; Seleiman et al. 2019; Wang et al. 2019a; Huang et al. 2020; Wang et al. 2020; de Tombeur et al. 2021a), confirming its potential as a suitable Si fertilizer (Li and Delvaux 2019). The types of biochar used, application rates (on a Si basis), and increase in percentages of soil soluble Si and/or plant Si concentrations are reported in Table S1. Recycling crop residues via direct incorporation, burning or biochar/manure/compost production and subsequent application has therefore a strong potential to increase crop Si uptake. However, the application rates used in most studies largely exceed coherent annual crop yields. For instance, considering biochar, the application of pyrolyzed material to concentrations of 1% (w/w) or more (bulk density of 1.3 g cm<sup>-3</sup>; depth incorporation of 10 cm), which corresponds to yearly crop yields of approximately 43 t ha<sup>-1</sup> (taking into account a pyrolysis yield of 30%) is common. This generally exceeds mean cereal yields worldwide (up to about 13 t ha<sup>-1</sup>; Ritchie and

609

610

611

612

613

614

615

616

617

618

619

620

621

622

623

624

625

626

627

628

629

Roser 2013), except for sugarcane (up to about 120 t ha<sup>-1</sup>; Ritchie and Roser 2013), and the fraction not available for pyrolysis has to be considered. Li and Delvaux (2019) calculated that a realistic application rate of biochar in the tropics would be around 1.7 t ha<sup>-1</sup> yr<sup>-1</sup>, which is about an order of magnitude lower than what was applied in some studies. We, therefore, stress the importance of performing long-term studies in agroecosystems with minimal inputs from external sources and outputs of crop residues (Hughes et al. 2020), to better assess the long-term sustainability of such recycling practices. In these systems, detailed analyses of soil and plant Si pools must be conducted, as well as mass-balance calculations, to study the magnitude of desilication as a function of crop residue management. For example, Hughes et al. (2020) recently showed that enhanced Si accumulation in rice grain (including husks) in highly-weathered soil environments could further contribute to long-term desilication. Furthermore, the effect of crop residue quality on organic matter decomposition rates in contrasting soil, climatic and agricultural contexts should be considered because of its key role in dissolved Si release rates (Fraysse et al. 2006a, 2010; Marxen et al. 2016; Nakamura et al. 2020).

#### Harnessing Si biocycling and recycling using cover crops

Cover crops are grown specifically for covering the soil during the off-season to reduce soil erosion, increase soil organic matter content and microbial diversity, and improve nutrient cycling (Reeves 1994; Adetunji et al. 2020). The positive impact of cover crops on N, P and C cycles has been extensively demonstrated (Abdalla et al. 2019; Hallama et al. 2019), while their effects on soil Si dynamics for different cover crops species are unknown. In the short term, cover crops could have detrimental effects by consuming the pool of soil readily-soluble Si during the winter, lowering Si plant availability for main crops in summer. However, in the long

term, the yearly transfer of Si stored in the cover crop via plant residues will significantly increase the soil phytogenic Si pool, and the cover crop will reduce phytolith losses through erosion (Fig. 3a). This conversion of litho/pedogenic Si-bearing minerals to phytoliths could significantly stimulate Si mobility in agroecosystems since the dissolution rates of phytoliths are an order of magnitude greater than those of typical soil clay minerals (Fraysse et al. 2009). Such positive effect would be particularly significant in highly-desilicated soils, where phytolith dissolution has a major effect on the soil-plant Si cycle (Alexandre et al. 1997; Sommer et al. 2013; de Tombeur et al. 2020a).

Based on shoot biomass data of common cover crop species (Hallama et al. 2019), we estimated Si stocks in aboveground biomass of cover crops (Table 4). For legumes and *Brassica* sp. cover crops, from 2 to 56 kg Si ha<sup>-1</sup> could annually be brought to soil via crop residues. This range, despite being very large, approximately corresponds with the annual Si uptake in major forest ecosystems worldwide, expect bamboo forests (Vander Linden and Delvaux 2019). The use of Poaceae sp. as cover crops would allow an extreme degree of annual Si inputs, up to 360 kg Si ha<sup>-1</sup> for *Lolium* sp., which approximately corresponds to Si uptake in sugarcane agroecosystems (Vander Linden and Delvaux 2019). In addition, cover crops may also impact Si dynamics by providing a legacy of increased mycorrhizal abundance, modifying rhizosphere physico-chemical properties (e.g., pH, soil aggregation, root exudates) or changing soil microbial communities and earthworms abundance (Roarty et al. 2017; Hallama et al. 2019; Adetunji et al. 2020; Euteneuer et al. 2020). Finally, Si-rich cover crops could diminish herbivore populations, with beneficial legacy effects on the main crop (Vernavá et al. 2004).

**Table 4** – Estimation of shoot Si stocks in common cover crop species.

	Family	Shoot biomass <sup>a</sup> (t ha <sup>-1</sup> )	Shoot Si concentration <sup>b</sup> (g kg <sup>-1</sup> )	Shoot Si stocks (kg ha <sup>-1</sup> )
Legume cover crops				
Lupinus sp.	Fabaceae	0.7-12.4 (n=6)	2.8-4.5	2-56
Pisum sativum	Fabaceae	3.7-3.9 (n=3)	2.8-5.6	10-22
Vicia sp.	Fabaceae	4.3-8.0 ( <i>n</i> =7)	2.4-4.8	10-38
Non-legume cover crops				
Avena sp.	Poaceae	7.9-13.2 ( <i>n</i> =6)	11.5-15.1	91-200
Lolium sp.	Poaceae	1.4-10.0 ( <i>n</i> =3)	9.7-36.4	14-364
Secale cereale	Poaceae	1.5-12.7 ( <i>n</i> =11)	12.6	19-160
Brassica sp.	Brassicaceae	2.2-4.1(n=4)	2.3-11.2	5-46

<sup>&</sup>lt;sup>a</sup>Data from Hallama et al. (2019)

#### Facilitative interactions: cereal-legume intercropping systems

Intercropping (Smith and McSorley 2000) has the potential to globally increase yields, reduce fertilizer inputs and save land (Martin-Guay et al. 2018; Li et al. 2020a), while increasing soil C and N content, improving mineral nutrition and reducing effects of pests (Hinsinger et al. 2011; Brooker et al. 2015; Cong et al. 2015; Xue et al. 2016; Tang et al. 2020). To our knowledge, the impact of intercropping systems on Si dynamics has been considered only once, through the study of rice (*Oryza sativa*) intercropped with water spinach (*Ipomoea aquatic*) (Ning et al. 2017). Plant-available Si concentrations in soil were not markedly impacted by the intercropped system compared with rice monoculture, yet they significantly increased when water spinach was cultivated alone, likely because spinach accumulates less Si than rice (Ning et al. 2017). However, leaf Si concentrations and stocks (i.e. Si concentration × leaf dry weight) of rice plants significantly increased in the intercropping system compared with the rice monoculture (up to ~25% for Si concentrations and ~75% for Si stocks) (Ning et al. 2017). Moreover, the disease index of rice sheath and the incidence of leaf folders significantly decreased in the intercropped

<sup>&</sup>lt;sup>b</sup>Data from Hodson et al. (2005) for all species. Additional data from Xiao et al. (2016) for *Pisum sativum*, Hasan et al. (2020) for *Vicia* sp., Soratto et al. (2012) for *Avena* sp. and Song et al. (2009) for *Brassica* sp.

system, which could be explained by (1) a role of physical barrier played by the strip distribution in intercropping system, (2) a more favorable field microclimate in the intercropping system, (3) the release of allelochemicals by water spinach, and/or (4) higher concentrations of silica-based defenses (Ning et al. 2017).

707

708

709

710

711

712

713

714

715

716

717

718

719

720

721

722

723

724

725

726

727

728

The pioneering work of Ning et al. has opened up new compelling directions in intercropping-Si research. Here, we propose different processes that could affect soil-plant Si dynamics in cereallegume intercropping agroecosystems (Fig. 3b). First, more Si would be available for cereal crop uptake since grasses accumulate more Si than legumes. In the long term, cereal-legume intercropping might even slow down soil desilication, even though mass-balance calculations should be performed to estimate Si stocks in biomass and export from harvests. However, the opposite effect might occur for legume crops, for which less Si would be available than in a monoculture system (Ning et al. 2017). This is important to consider since Si has beneficial effects also for legumes, and may promote the symbiotic relationship with nitrogen-fixing bacteria in root nodules (Putra et al. 2020). Second, cereal-legume intercropping might induce a wide range of facilitative interactions (Li et al. 2014). Under conditions of Fe and Zn deficiencies, cereals such as wheat, barley, maize or rice secrete phytosiderophores in the soil solution that mobilize Fe and Zn (Ahmed and Holmström 2014), which can then be transferred to both crops (Zuo et al. 2000; Xue et al. 2016). The same mechanism applies to legumes that may secrete carboxylates to mobilize P, especially under P deficiency (Lambers et al. 2006, 2015; Pang et al. 2018), which could in turn benefit both crops (Xue et al. 2016; Lambers et al. 2018). As discussed above, Si co-solubilization or desorption by root exudates such as phytosiderophores and carboxylates is likely, but this needs to be further assessed to determine if such facilitative interactions also occur for this element. Similarly, facilitation via common mycorrhizal networks and nutrient transfer can occur for N and P (Walder et al. 2012), but has not been addressed for Si. Third, earthworm abundance and biomass can greatly increase in cereal-legume intercropping systems compared with monocultures (Schmidt et al. 2001, 2003) which could increase plant-available Si concentrations and Si soil-plant mobility (Fig. 2). Finally, intercropping might also impact Si dynamics by changing soil microbial diversity and modifying the physico-chemical properties of the rhizosphere (Brooker et al. 2015), but the direction of these processes needs to be elucidated.

#### Grazers as biocatalysts of Si cycling: crop-livestock systems

Despite recent simplification and specialization of agricultural systems worldwide, integrated crop-livestock systems have been employed for millennia and remain the main agriculture model for over two thirds of global farmers, and represent about half of the world's food (Russelle et al. 2007; Herrero et al. 2010; Lemaire et al. 2014). Integrated crop-livestock systems have the potential to improve carbon and nutrient cycling/use efficiency (Alves et al. 2019; Brewer and Gaudin 2020; Carlos et al. 2020). Based on the evidence of Si mobilization by large herbivores (Fig. 2), we suggest several benefits of integrated crop-livestock systems on soil-plant Si dynamics, based on a simple pasture/crop rotation (Fig. 3c). First, large ruminants strongly increase biogenic silica turnover rates (Blackman and Bailey 1971; Vandevenne et al. 2013). For instance, Vandevenne et al. (2013) estimated that a cow-based pasture mobilizes between 18 and 28 kg Si ha<sup>-1</sup> yr<sup>-1</sup>, against 1.3-1.8 kg Si ha<sup>-1</sup> yr<sup>-1</sup> in ungrazed pastures. In the long term, greater Si mobilization potentially accelerates soil desilication through Si leaching (Vandevenne et al. 2013). However, in the short term, it will most likely increase plant-available Si concentrations

for the subsequent crop, especially if soil texture-related leaching potential is low. Second, pasture/crop rotations will reduce long-term soil desilication because Si exports through crop harvest will occur only once every two years. Finally, integrating crops and livestock impacts numerous aspects of soil-plant systems such as soil aggregation, microbial community structure/biomass and annual net primary production (Brewer and Gaudin 2020), which could ultimately also affect the soil-plant Si cycle. Crop-livestock systems are promising to enhance Si mobility in soil-plant systems, particularly through the effects of ruminants on phytolith turnover, and therefore deserve further investigations, including the time scales of different processes.

#### Liming and soil-plant Si dynamics: a gap between theory and practice

The effect of liming on soil pH in agroecosystems may affect Si dynamics through antagonistic processes (Fig. 3d), as discussed above. Therefore, the liming effect is still unclear because it strongly depends on the initial pedological context in terms of soil pH, mineralogy, buffering capacity and phytolith content (Haynes 2019; Vander Linden and Delvaux 2019). Previous studies showed either an increase (Castro and Crusciol 2013; Klotzbücher et al. 2018b) or a decrease (Kostic et al. 2017; Keeping et al. 2017; Haynes and Zhou 2018) in soil Si availability with liming, while others found no significant effect (Mathews et al. 2009; Bhat et al. 2010). However, different extractants were used to estimate plant-available Si concentrations in these studies (e.g., CaCl<sub>2</sub>, acetic acid). In fact, liming is expected to increase the pool of adsorbed Si, often associated with acetate and acetic acid extractants, while decreasing the soluble Si pool, often associated with CaCl<sub>2</sub> or water extractants (Sauer et al. 2006; Georgiadis et al. 2013). Haynes and Zhou (2018) confirmed this pattern by showing an increase of Si-acetic acid by about 75-110% and a decrease of Si-CaCl<sub>2</sub> by about 25-35% for the pH range 5.0-6.5 in limed Podzol

and Gleysol. The authors also found a decrease of alkali-extractable Si, suggesting a loss of phytoliths following increasing dissolution rates (Haynes and Zhou 2018). More recently, Caubet et al. (2020) showed that French agricultural soils had higher Si-CaCl<sub>2</sub> concentrations than noncultivated soils. The authors interpreted this difference as resulting from the pH increase after liming that could modify clay mineralogy (Cornu et al. 2012) and increase phytolith dissolution for soils with clay size mineral contents ranging between 5 and 32%. In their study, soils were classified by parent material types, where soils developed on sediments were separated into two groups: carbonated soils (>1% carbonates) and non-carbonated soils (<1% carbonates). In the non-carbonated group, Si-CaCl<sub>2</sub> also correlates with the 2-um fraction cation exchange capacity, used as a proxy of the nature of clay-size minerals. This supports a possible effect of clay mineralogy on Si availability that can be superimposed on the liming effect in cultivated land compared with non-cultivated land (forests, wetlands, pastures, parks). Indeed, soils with higher weatherable mineral reserves and subsequently higher Si availability are preferred for agriculture. As soil extractants are only proxies for plant-available Si concentrations (e.g., Wu et al. 2020), the liming effect on soil Si dynamics should be addressed by quantifying plant Si concentrations. Although some studies showed that liming had no significant effect on plant Si uptake (Bhat et al. 2010; Castro and Crusciol 2013; Keeping et al. 2017), others found a marked decrease (Mathews et al. 2009; Tavakkoli et al. 2011). In particular, Mathews et al. (2009) noted a decrease in Si concentrations of Pennisetum clandestinum of about 30% with increasing calcium carbonate application, and in the pH range of 5.2-6.2. This study could highlight that raising soil pH above 6.0 may reduce plant Si uptake, possibly because of increased Si adsorption and subsequent decline of Si plant-availability. In addition, raising pH will increase polymerization rates of

773

774

775

776

777

778

779

780

781

782

783

784

785

786

787

788

789

790

791

792

793

monosilicic acid and its precipitation as amorphous silica, which could also decrease Si plant-availability (Icopini et al. 2005; Schaller et al. 2021). Besides, phytolith dissolution rates strongly increase in this pH range (Fraysse et al. 2009) which possibly enhances plant Si uptake (Guntzer et al. 2012), but the size of the phytogenic Si pool is low compared with the litho/pedogenic-Si pool (Alexandre et al. 1997, 2011; Sommer et al. 2013). Phytoliths can, therefore, not be the main factor of Si plant-availability and subsequent plant uptake in certain pedological contexts (Keller et al. 2021). Overall, the effect of liming on soil-plant Si dynamics is still unclear because numerous antagonistic processes occur on different time scales (Fig. 3d) (Haynes 2019; Vander Linden and Delvaux 2019).

#### **Conclusions and perspectives**

Soil-plant Si cycling is mainly studied through the prism of abiotic mineral weathering or plant Si uptake followed by soil phytoliths dissolution (e.g., Bartoli 1983; Lucas et al. 1993; Alexandre et al. 1997; de Tombeur et al. 2020a), while biotic factors tend to be overlooked. Besides, numerous studies on biological weathering have been conducted, on different scales (Barker et al. 1998; Banfield et al. 1999; Lucas 2001; Uroz et al. 2009; Finlay et al. 2020), but rarely in the framework of plant Si nutrition and subsequent positive influence on plant performance or crop yield. We have stressed the importance of biotic factors such as mycorrhizal associations, SSB, soil macrofauna, large herbivores and root exudates on soil-plant Si mobility, and suggest different mechanisms by which these processes may affect Si dynamics and stimulate soil-plant Si cycling in agroecosystems. The time has come to seriously consider biotic factors, both in natural and agricultural systems, because Si is involved in numerous functions that contribute to

plant performance and stress regulation, which may impact plant productivity and crop yield. We suggest the following aspects to be considered in the near future:

- 1. What is the abundance and diversity of SSB in soils? In which soil types/mineralogy are they found most? To what extent do SSB contribute to the plant-available Si pool and subsequent plant Si uptake compared with abiotic mineral weathering, and by what mechanisms? How do agricultural practices and cropping systems impact SSB abundance, diversity, and functions?
  - 2. By which mechanisms do mycorrhizal fungi increase plant Si concentrations? Can plant Si nutrition be improved through common mycorrhizal networks? Does an increase in plant Si concentrations as affected by mycorrhizal fungi mostly occur in nutrient-depleted soils?
  - 3. To what extent do non-mycorrhizal nutrient-acquisition strategies influence soil Si mobilization and plant Si uptake? By which mechanisms? If root exudates mobilize Si from soil minerals, does this occur only in P-depleted soils and without P fertilization? Can we exploit these strategies to improve crop Si nutrition in cropping systems such as cereal-legume intercropping?
  - 4. Is the earthworms-related increase in soil-plant Si mobility significant at soil profile scales? Do cover crops, intercropping or integrated crop-livestock systems influence this process by modifying earthworm abundance and diversity?
  - 5. What is the impact of large herbivores on global Si biogeochemistry? To what extent do they stimulate soil-plant Si mobility in natural ecosystems and integrated crop-livestock

- systems? Do integrated crop-livestock systems influence Si dynamics though changes in microbial community and biomass, soil aggregation or organic matter dynamics?
- 6. Does the long-term recycling of crop residues significantly increase plant-available and plant Si concentrations, and slow down long-term soil desilication? What are the long-term advantages and disadvantages of different residue management practices (i.e. direct incorporation, burning, or biochar/manure/compost production and subsequent application)? How does decomposition dynamics influence Si release from phytolith dissolution in different geopedoclimatic contexts, and for different plant species?
- 7. What are the short- and long-term effects of cover crops on soil-plant Si mobility? Can legume cover crops significantly stimulate Si mobility compared with grass cover crops?

  Does the absence of cover crops significantly increase soil phytolith losses through soil erosion?
- 8. How do soil aggregation and soil structure impact soil-plant Si dynamics? Do *microbial hotspots* in soil aggregate fractions and at surfaces of minerals affect soil Si dynamics and release into the soil solution? Is it significant at the soil profile scale? How do modifications of soil structure and aggregation by anthropogenic land transformations (ploughing, soil pH-induced modification by liming, clay dispersion associated with high sodium content in saline-sodic soils) influence these processes?

## References

860	Abbas T, Rizwan M, Ali S, et al (2017) Effect of biochar on cadmium bioavailability and uptake
861	in wheat (Triticum aestivum L.) grown in a soil with aged contamination. Ecotoxicol
862	Environ Saf 140:37–47. https://doi.org/10.1016/j.ecoenv.2017.02.028
863	Abbott LK, Robson AD, De Boer G (1984) The effect of phosphorus on the formation of hyphae
864	in soil by the vesicular-arbuscular mycorrhizal fungus, Glomus Fasciculatum. New Phytol
865	97:437–446
866	Abdalla M, Hastings A, Cheng K, et al (2019) A critical review of the impacts of cover crops on
867	nitrogen leaching, net greenhouse gas balance and crop productivity. Glob Chang Biol
868	25:2530–2543. https://doi.org/10.1111/gcb.14644
869	Abrahão A, Lambers H, Sawaya ACHF, et al (2014) Convergence of a specialized root trait in
870	plants from nutrient-impoverished soils: phosphorus-acquisition strategy in a
871	nonmycorrhizal cactus. Oecologia 176:345–355. https://doi.org/10.1007/s00442-014-3033-4
872	Acevedo FE, Peiffer M, Ray S, et al (2021) Silicon-mediated enhancement of herbivore
873	resistance in agricultural crops. Front Plant Sci 12:631824.
874	https://doi.org/10.3389/fpls.2021.631824
875	Adetunji AT, Ncube B, Mulidzi R, Lewu FB (2020) Management impact and benefit of cover
876	crops on soil quality: A review. Soil Tillage Res 204:104717.
877	https://doi.org/10.1016/j.still.2020.104717

878	Ahmed E, Holmström SJM (2014) Siderophores in environmental research: Roles and
879	applications. Microb Biotechnol 7:196–208. https://doi.org/10.1111/1751-7915.12117
880	Alexandre A, Bouvet M, Abbadie L (2011) The role of savannas in the terrestrial Si cycle: A
881	case-study from Lamto, Ivory Coast. Glob Planet Change 78:162-169.
882	https://doi.org/10.1016/j.gloplacha.2011.06.007
883	Alexandre A, Meunier J-D, Colin F, Koud J-M (1997) Plant impact on the biogeochemical cycle
884	of silicon and related weathering processes. Geochim Cosmochim Acta 61:677–682.
885	https://doi.org/10.1016/S0016-7037(97)00001-X
886	Alvarez-Campos O, Lang TA, Bhadha JH, et al (2018) Biochar and mill ash improve yields of
887	sugarcane on a sand soil in Florida. Agric Ecosyst Environ 253:122-130.
888	https://doi.org/10.1016/j.agee.2017.11.006
889	Alves LA, Denardin LG de O, Martins AP, et al (2019) Soil acidification and P, K, Ca and Mg
890	budget as affected by sheep grazing and crop rotation in a long-term integrated crop-
891	livestock system in southern Brazil. Geoderma 351:197–208.
892	https://doi.org/10.1016/j.geoderma.2019.04.036
893	Banfield JF, Barker WW, Welch SA, Taunton A (1999) Biological impact on mineral
894	dissolution: Application of the lichen model to understanding mineral weathering in the
895	rhizosphere. Proc Natl Acad Sci U S A 96:3404-3411.
896	https://doi.org/10.1073/pnas.96.7.3404
897	Bardgett RD, Wardle DA (2003) Herbivore-mediated linkages between aboveground and
898	belowground communities. Ecology 84:2258–2268

899	Barker WW, Welch SA, Chu S, Banfield JF (1998) Experimental observations of the effects of
900	bacteria on aluminosilicate weathering. Am Mineral 83:1551–1563.
901	https://doi.org/10.2138/am-1998-11-1243
902	Bartoli F (1983) The biogeochemical cycle of silicon in two temperate forest ecosystems. Ecol
903	Bull 35:469–476
904	Beasley DE, Koltz AM, Lambert JE, et al (2015) The evolution of stomach acidity and its
905	relevance to the human microbiome. PLoS One 10:1–12.
906	https://doi.org/10.1371/journal.pone.0134116
907	Beckwith RS, Reeve R (1964) Studies on soluble silica in soils. II. The release of monosilicic
908	acid from soils. Aust J Soil Res 2:33–45
909	Bennett PC, Rogers JR, Choi WJ, Hiebert FK (2001) Silicates, silicate weathering, and microbial
910	ecology. Geomicrobiol J 18:3–19. https://doi.org/10.1080/01490450151079734
911	Bhat JA, Kundu MC, Hazra GC, et al (2010) Rehabilitating acid soils for increasing crop
912	productivity through low-cost liming material. Sci Total Environ 408:4346–4353.
913	https://doi.org/10.1016/j.scitotenv.2010.07.011
914	Bidle KD, Azam F (1999) Accelerated dissolution of diatom silica by marine bacterial
915	assemblages. Nature 397:508–512. https://doi.org/10.1038/17351
916	Bist V, Niranjan A, Ranjan M, et al (2020) Silicon-solubilizing media and its implication for
917	characterization of bacteria to mitigate biotic stress. Front Plant Sci 11:28.
918	https://doi.org/10.3389/fpls.2020.00028

919	Bityutskii N, Kaidun P, Yakkonen K (2016) Earthworms can increase mobility and
920	bioavailability of silicon in soil. Soil Biol Biochem 99:47–53.
921	https://doi.org/10.1016/j.soilbio.2016.04.022
922	Blackman E, Bailey CB (1971) Dissolution of silica from dried grass in nylon bags placed in the
923	rumen of a cow. Can J Anim Sci 51:327–332
924	Blecker SW, McCulley RL, Chadwick OA, Kelly EF (2006) Biologic cycling of silica across a
925	grassland bioclimosequence. Global Biogeochem Cycles 20:1–11.
926	https://doi.org/10.1029/2006GB002690
927	Blouin M, Hodson ME, Delgado EA, et al (2013) A review of earthworm impact on soil function
928	and ecosystem services. Eur J Soil Sci 64:161–182. https://doi.org/10.1111/ejss.12025
929	Bosecker K (1997) Bioleaching: metal solubilization by microorganisms. FEMS Microbiol Lett
930	20:591–604
931	Brantley SL (2008) Kinetics of mineral dissolution. In: Brantley SL, Kubicki JD, White AF (eds)
932	Kinetics of water-rock interaction. New York: Springer Science & Business Media., pp
933	151–210
934	Bray AW, Oelkers EH, Bonneville S, et al (2015) The effect of pH, grain size, and organic
935	ligands on biotite weathering rates. Geochim Cosmochim Acta 164:127–145.
936	https://doi.org/10.1016/j.gca.2015.04.048
937	Brewer KM, Gaudin ACM (2020) Potential of crop-livestock integration to enhance carbon
938	sequestration and agroecosystem functioning in semi-arid croplands. Soil Biol Biochem

939	149:107936. https://doi.org/10.1016/j.soilbio.2020.107936
940	Brooker RW, Bennett AE, Cong WF, et al (2015) Improving intercropping: A synthesis of
941	research in agronomy, plant physiology and ecology. New Phytol 206:107-117.
942	https://doi.org/10.1111/nph.13132
943	Brucker E, Kernchen S, Spohn M (2020) Release of phosphorus and silicon from minerals by soi
944	microorganisms depends on the availability of organic carbon. Soil Biol Biochem
945	143:107737. https://doi.org/10.1016/j.soilbio.2020.107737
946	Brundrett MC (2002) Coevolution of roots and mycorrhizas of land plants. New Phytol 154:275-
947	304. https://doi.org/10.1046/j.1469-8137.2002.00397.x
948	Burghelea C, Zaharescu DG, Dontsova K, et al (2015) Mineral nutrient mobilization by plants
949	from rock: influence of rock type and arbuscular mycorrhiza. Biogeochemistry 124:187-
950	203. https://doi.org/10.1007/s10533-015-0092-5
951	Buss HL, Lüttge A, Brantley SL (2007) Etch pit formation on iron silicate surfaces during
952	siderophore-promoted dissolution. Chem Geol 240:326–342.
953	https://doi.org/10.1016/j.chemgeo.2007.03.003
954	Calvaruso C, Turpault MP, Frey-Klett P (2006) Root-associated bacteria contribute to mineral
955	weathering and to mineral nutrition in trees: A budgeting analysis. Appl Environ Microbiol
956	72:1258–1266. https://doi.org/10.1128/AEM.72.2.1258-1266.2006
957	Cama J, Ganor J (2006) The effects of organic acids on the dissolution of silicate minerals: A
958	case study of oxalate catalysis of kaolinite dissolution. Geochim Cosmochim Acta 70:2191-

959	2209. https://doi.org/10.1016/j.gca.2006.01.028
960	Carey JC, Fulweiler RW (2016) Human appropriation of biogenic silicon – the increasing role of
961	agriculture. Funct Ecol 30:1331–1339. https://doi.org/10.1111/1365-2435.12544
962	Carlos FS, Oliveira Denardin LG, Martins AP, et al (2020) Integrated crop-livestock systems in
963	lowlands increase the availability of nutrients to irrigated rice. L Degrad Dev 31:2962–2972.
964	https://doi.org/10.1002/ldr.3653
965	Carpenter D, Hodson ME, Eggleton P, Kirk C (2007) Earthworm induced mineral weathering:
966	Preliminary results. Eur J Soil Biol 43:176–183. https://doi.org/10.1016/j.ejsobi.2007.08.053
967	Castro GSA, Crusciol CAC (2013) Yield and mineral nutrition of soybean, maize, and congo
968	signal grass as affected by limestone and slag. Pesqui Agropecu Bras 48:673-681.
969	https://doi.org/10.1590/S0100-204X2013000600013
970	Caubet M, Cornu S, Saby NPA, Meunier J-D (2020) Agriculture increases the bioavailability of
971	silicon, a beneficial element for crop, in temperate soils. Sci Rep 10:19999.
972	https://doi.org/10.1038/s41598-020-77059-1
973	Chadwick OA, Chorover J (2001) The chemistry of pedogenic thresholds. Geoderma 100:321–
974	353. https://doi.org/10.1016/S0016-7061(01)00027-1
975	Chandrakala C, Voleti SR, Bandeppa S, et al (2019) Silicate solubilization and plant growth
976	promoting potential of <i>Rhizobium</i> Sp. isolated from rice rhizosphere. Silicon 11:2895–2906.
977	https://doi.org/10.1007/s12633-019-0079-2
978	Christl I, Brechbühl Y, Graf M, Kretzschmar R (2012) Polymerization of silicate on hematite

979	surfaces and its influence on arsenic sorption. Environ Sci Technol 46:13235–13243.
980	https://doi.org/10.1021/es303297m
981	Churchman GJ, Lowe D. (2012) Alteration, formation, and occurrence of minerals in soils.
982	Handb Soil Sci Prop Process 1:20–72. https://doi.org/doi:10.1201/b11267-24
983	Clark RB, Zeto SK (2000) Mineral acquisition by arbuscular mycorrhizal plants. J Plant Nutr
984	23:867–902. https://doi.org/10.1080/01904160009382068
985	Clark RB, Zeto SK (1996) Mineral acquisition by mycorrhizal maize grown on acid and alkaline
986	soil. Soil Biol Biochem 28:1495–1503. https://doi.org/10.1016/S0038-0717(96)00163-0
987	Clymans W, Struyf E, Govers G, et al (2011) Anthropogenic impact on amorphous silica pools in
988	temperate soils. Biogeosciences 8:2281–2293. https://doi.org/10.5194/bg-8-2281-2011
989	Colombo C, Palumbo G, He JZ, et al (2014) Review on iron availability in soil: Interaction of Fe
990	minerals, plants, and microbes. J Soils Sediments 14:538-548.
991	https://doi.org/10.1007/s11368-013-0814-z
992	Cong WF, Hoffland E, Li L, et al (2015) Intercropping enhances soil carbon and nitrogen. Glob
993	Chang Biol 21:1715–1726. https://doi.org/10.1111/gcb.12738
994	Cooke J, Leishman MR (2016) Consistent alleviation of abiotic stress with silicon addition: a
995	meta-analysis. Funct Ecol 30:1340–1357. https://doi.org/10.1111/1365-2435.12713
996	Cooke J, Leishman MR (2011) Is plant ecology more siliceous than we realise? Trends Plant Sci
997	16:61–68. https://doi.org/10.1016/j.tplants.2010.10.003
998	Cornelis J-T, Delvaux B (2016) Soil processes drive the biological silicon feedback loop. Funct

999	Ecol 30:1298–1310. https://doi.org/10.1111/1365-2435.12704
1000	Cornelis J-T, Ranger J, Iserentant A, Delvaux B (2010) Tree species impact the terrestrial cycle
1001	of silicon through various uptakes. Biogeochemistry 97:231-245.
1002	https://doi.org/10.1007/s10533-009-9369-x
1003	Cornu S, Montagne D, Hubert F, et al (2012) Evidence of short-term clay evolution in soils under
1004	human impact. Comptes Rendus - Geosci 344:747–757.
1005	https://doi.org/10.1016/j.crte.2012.09.005
1006	Coskun D, Deshmukh R, Sonah H, et al (2019) The controversies of silicon's role in plant
1007	biology. New Phytol 221:67–85. https://doi.org/10.1111/nph.15343
1008	Covacevich F, Echeverría HE, Aguirrezabal LAN (2007) Soil available phosphorus status
1009	determines indigenous mycorrhizal colonization of field and glasshouse-grown spring wheat
1010	from Argentina. Appl Soil Ecol 35:1–9. https://doi.org/10.1016/j.apsoil.2006.06.001
1011	Dakora FD, Phillips DA (2002) Root exudates as mediators of mineral acquisition in low-nutrient
1012	environments. Plant Soil 245:35-47. https://doi.org/10.1023/A:1020809400075
1013	Datnoff LE, Snyder GH, Korndörfer GH (2001) Silicon in Agriculture, Volume 8
1014	de Tombeur F, Cooke J, Collard L, et al (2021a) Biochar affects silicification patterns and
1015	physical traits of rice leaves cultivated in a desilicated soil (Ferric Lixisol). Plant Soil
1016	460:375–390. https://doi.org/10.1007/s11104-020-04816-6
1017	de Tombeur F, Cornélis J-T, Lambers H (2021b) Silicon mobilisation by root-released
1018	carboxylates. Trends Plant Sci accepted pending revisions

1019	de Tombeur F, Laliberté E, Lambers H, et al (2021c) A shift from phenol to silica-based leaf
1020	defences during long-term soil and ecosystem development. Ecol Lett 24:984-995.
1021	https://doi.org/10.1111/ele.13713
1022	de Tombeur F, Turner BL, Laliberté E, et al (2020a) Plants sustain the terrestrial silicon cycle
1023	during ecosystem retrogression. Science (80- ) 369:1245–1248.
1024	https://doi.org/10.1126/science.abc0393
1025	de Tombeur F, Turner BL, Laliberté E, et al (2020b) Silicon dynamics during 2 million years of
1026	soil development in a coastal dune chronosequence under a Mediterranean climate.
1027	Ecosystems 23:1614–1630. https://doi.org/10.1007/s10021-020-00493-9
1028	de Tombeur F, Vander Linden C, Cornélis J-T, et al (2020c) Soil and climate affect foliar
1029	silicification patterns and silica-cellulose balance in sugarcane (Saccharum officinarum).
1030	Plant Soil 452:529–546. https://doi.org/https://doi.org/10.1007/s11104-020-04588-z
1031	Debona D, Rodrigues FA, Datnoff LE (2017) Silicon's role in abiotic and biotic plant stresses.
1032	Annu Rev Phytopathol 55:85–107. https://doi.org/10.1146/annurev-phyto-080516-035312
1033	Derry LA, Kurtz AC, Ziegler K, Chadwick OA (2005) Biological control of terrestrial silica
1034	cycling and export fluxes to watersheds. Nature 433:728–731.
1035	https://doi.org/10.1038/nature03299
1036	Deshmukh R, Bélanger RR (2016) Molecular evolution of aquaporins and silicon influx in plants
1037	Funct Ecol 30:1277–1285. https://doi.org/10.1111/1365-2435.12570
1038	Deshmukh R, Sonah H, Belanger RR (2020) New evidence defining the evolutionary path of

1039	aquaporins regulating silicon uptake in land plants. J Exp Bot 71:6775-6788.
1040	https://doi.org/10.1093/jxb/eraa342
1041	Desplanques V, Cary L, Mouret JC, et al (2006) Silicon transfers in a rice field in Camargue
1042	(France). J Geochemical Explor 88:190–193. https://doi.org/10.1016/j.gexplo.2005.08.036
1043	Deveau A, Bonito G, Uehling J, et al (2018) Bacterial-fungal interactions: Ecology, mechanisms
1044	and challenges. FEMS Microbiol Rev 42:335–352. https://doi.org/10.1093/femsre/fuy008
1045	Dietzel M (2002) Interaction of polysilicic and monosilicic acid with mineral surfaces. In: Stober
1046	I, Bucher K (eds) Water-Rock Interaction. Springer, Dordrecht, pp 207–235
1047	Dietzel M (2000) Dissolution of silicates and the stability of polysilicic acid. Geochim
1048	Cosmochim Acta 64:3275–3281
1049	Dong H, Kostka JE, Kim J (2003) Microscopic evidence for microbial dissolution of smectite.
1050	Clays Clay Miner 51:502–512. https://doi.org/10.1346/CCMN.2003.0510504
1051	Dontsova K, Balogh-Brunstad Z, Le Roux G (2020) Biogeochemical Cycles: Ecological Drivers
1052	and Environmental Impact. Wiley
1053	Drever JI (1994) The effect of land plants on weathering rates of silicate minerals. Geochim
1054	Cosmochim Acta 58:2325–2332. https://doi.org/10.1016/0016-7037(94)90013-2
1055	Drever JI, Stillings LL (1997) The role of organic acids in mineral weathering. Colloids Surfaces
1056	A Physicochem Eng Asp 120:167–181. https://doi.org/10.1016/S0927-7757(96)03720-X
1057	Ehrlich H, Demadis KD, Pokrovsky OS, Koutsoukos PG (2010) Modern views on
1058	desilicification: Biosilica and abiotic silica dissolution in natural and artificial environments.

1059	Chem Rev 110:4656–4689. https://doi.org/10.1021/cr900334y
1060	Eneji AE, Inanaga S, Muranaka S, et al (2008) Growth and nutrient use in four grasses under
1061	drought stress as mediated by silicon fertilizers. J Plant Nutr 31:355–365.
1062	https://doi.org/10.1080/01904160801894913
1063	Epstein E (1994) The anomaly of silicon in plant biology. Proc Natl Acad Sci 91:11–17.
1064	https://doi.org/10.1073/pnas.91.1.11
1065	Euteneuer P, Wagentristl H, Steinkellner S, et al (2020) Contrasting effects of cover crops on
1066	earthworms: Results from field monitoring and laboratory experiments on growth,
1067	reproduction and food choice. Eur J Soil Biol 100:103225.
1068	https://doi.org/10.1016/j.ejsobi.2020.103225
1069	Farmer VC, Delbos E, Miller JD (2005) The role of phytolith formation and dissolution in
1070	controlling concentrations of silica in soil solutions and streams. Geoderma 127:71–79.
1071	https://doi.org/10.1016/j.geoderma.2004.11.014
1072	Finlay RD, Mahmood S, Rosenstock N, et al (2020) Reviews and syntheses: Biological
1073	weathering and its consequences at different spatial levels - From nanoscale to global scale
1074	Biogeosciences 17:1507–1533. https://doi.org/10.5194/bg-17-1507-2020
1075	Fisher RA (1929) A preliminary note on the effect of sodium silicate in increasing the yield of
1076	barley. J Agric Sci 19:132–139. https://doi.org/10.1017/S0021859600011217
1077	Flemming HC, Wingender J (2010) The biofilm matrix. Nat Rev Microbiol 8:623–633.
1078	https://doi.org/10.1038/nrmicro2415

1079	Fonte SJ, Nesper M, Hegglin D, et al (2014) Pasture degradation impacts soil phosphorus storage
1080	via changes to aggregate-associated soil organic matter in highly weathered tropical soils.
1081	Soil Biol Biochem 68:150–157. https://doi.org/10.1016/j.soilbio.2013.09.025
1082	Forbes ES, Cushman JH, Burkepile DE, et al (2019) Synthesizing the effects of large, wild
1083	herbivore exclusion on ecosystem function. Funct Ecol 33:1597–1610.
1084	https://doi.org/10.1111/1365-2435.13376
1085	Fraysse F, Cantais F, Pokrovsky OS, et al (2006a) Aqueous reactivity of phytoliths and plant
1086	litter: Physico-chemical constraints on terrestrial biogeochemical cycle of silicon. J
1087	Geochemical Explor 88:202–205. https://doi.org/10.1016/j.gexplo.2005.08.039
1088	Fraysse F, Pokrovsky OS, Meunier JD (2010) Experimental study of terrestrial plant litter
1089	interaction with aqueous solutions. Geochim Cosmochim Acta 74:70-84.
1090	https://doi.org/10.1016/j.gca.2009.09.002
1091	Fraysse F, Pokrovsky OS, Schott J, Meunier JD (2009) Surface chemistry and reactivity of plant
1092	phytoliths in aqueous solutions. Chem Geol 258:197–206.
1093	https://doi.org/10.1016/j.chemgeo.2008.10.003
1094	Fraysse F, Pokrovsky OS, Schott J, Meunier JD (2006b) Surface properties, solubility and
1095	dissolution kinetics of bamboo phytoliths. Geochim Cosmochim Acta 70:1939–1951.
1096	https://doi.org/10.1016/j.gca.2005.12.025
1097	Frew A, Powell JR, Allsopp PG, et al (2017a) Arbuscular mycorrhizal fungi promote silicon
1098	accumulation in plant roots, reducing the impacts of root herbivory. Plant Soil 419:423-433.
1099	https://doi.org/10.1007/s11104-017-3357-z

1100	Frew A, Powell JR, Hiltpold I, et al (2017b) Host plant colonisation by arbuscular mycorrhizal
1101	fungi stimulates immune function whereas high root silicon concentrations diminish growth
1102	in a soil-dwelling herbivore. Soil Biol Biochem 112:117–126
1103	Frew A, Powell JR, Johnson SN (2020) Aboveground resource allocation in response to root
1104	herbivory as affected by the arbuscular mycorrhizal symbiosis. Plant Soil 447:463–473.
1105	https://doi.org/10.1007/s11104-019-04399-x
1106	Gadd GM (2017) Fungi, rocks, and minerals. Elements 13:171–176.
1107	https://doi.org/10.2113/gselements.13.3.171
1108	Garg N, Bhandari P (2016) Silicon nutrition and mycorrhizal inoculations improve growth,
1109	nutrient status, K+/Na+ ratio and yield of Cicer arietinum L. genotypes under salinity stress.
1110	Plant Growth Regul 78:371–387. https://doi.org/10.1007/s10725-015-0099-x
1111	Garg N, Singh S (2018) Mycorrhizal inoculations and silicon fortifications improve rhizobial
1112	symbiosis, antioxidant defense, trehalose turnover in pigeon pea genotypes under cadmium
1113	and zinc stress. Plant Growth Regul 86:105-119. https://doi.org/10.1007/s10725-018-0414-
1114	4
1115	Gattullo CE, Allegretta I, Medici L, et al (2016) Silicon dynamics in the rhizosphere:
1116	Connections with iron mobilization. J Plant Nutr Soil Sci 179:409-417.
1117	https://doi.org/10.1002/jpln.201500535
1118	Gbongue LR, Lalaymia I, Zeze A, et al (2019) Increased silicon acquisition in bananas colonized
1119	by Rhizophagus irregularis MUCL 41833 reduces the incidence of Pseudocercospora
1120	fijiensis. Front Plant Sci 9:1977. https://doi.org/10.3389/fpls.2018.01977

1121	Georgiadis A, Marhan S, Lattacher A, et al (2019) Do earthworms affect the fractionation of
1122	silicon in soil? Pedobiologia (Jena) 75:1–7. https://doi.org/10.1016/j.pedobi.2019.05.001
1123	Georgiadis A, Sauer D, Herrmann L, et al (2013) Development of a method for sequential Si
1124	extraction from soils. Geoderma 209–210:251–261.
1125	https://doi.org/10.1016/j.geoderma.2013.06.023
1126	Gérard F, Mayer KU, Hodson MJ, Ranger J (2008) Modelling the biogeochemical cycle of
1127	silicon in soils: Application to a temperate forest ecosystem. Geochim Cosmochim Acta
1128	72:741–758. https://doi.org/10.1016/j.gca.2007.11.010
1129	Gerke J, Römer W, Jungk A (1994) The excretion of citric and malic acid by proteoid roots of
1130	Lupinus albus L.; effects on soil solution concentrations of phosphate, iron, and aluminum
1131	in the proteoid rhizosphere in samples of an oxisol and a luvisol. J Plant Nutr Soil Sci
1132	157:289–294. https://doi.org/10.1002/jpln.19941570408
1133	Goldich SS (1938) A study in rock-weathering. J Geol 46:17–58
1134	Golubev S V., Pokrovsky OS, Schott J (2005) Experimental determination of the effect of
1135	dissolved CO2 on the dissolution kinetics of Mg and Ca silicates at 25 °C. Chem Geol
1136	217:227–238. https://doi.org/10.1016/j.chemgeo.2004.12.011
1137	Guennoc CM, Rose C, Labbé J, Deveau A (2018) Bacterial biofilm formation on the hyphae of
1138	ectomycorrhizal fungi: A widespread ability under controls? FEMS Microbiol Ecol 94:1-14
1139	https://doi.org/10.1093/femsec/fiy093
1140	Guntzer F, Keller C, Poulton PR, et al (2012) Long-term removal of wheat straw decreases soil

1141	amorphous silica at Broadbalk, Rothamsted. Plant Soil 352:173-184.
1142	https://doi.org/10.1007/s11104-011-0987-4
1143	Hall AD, Morison CGT (1906) On the function of silica in the nutrition of cereals.—Part I. Proc
1144	R Soc London Ser B 77:455–477. https://doi.org/10.1098/rspb.1906.0035
1145	Hallama M, Pekrun C, Lambers H, Kandeler E (2019) Hidden miners – the roles of cover crops
1146	and soil microorganisms in phosphorus cycling through agroecosystems. Plant Soil 434:7-
1147	45. https://doi.org/10.1007/s11104-018-3810-7
1148	Hartley SE, DeGabriel JL (2016) The ecology of herbivore-induced silicon defences in grasses.
1149	Funct Ecol 30:1311–1322. https://doi.org/10.1111/1365-2435.12706
1150	Hasan KA, Soliman H, Baka Z, Shabana YM (2020) Efficacy of nano-silicon in the control of
1151	chocolate spot disease of Vicia faba L. caused by Botrytis fabae. Egypt J Basic Appl Sci
1152	7:53–66. https://doi.org/10.1080/2314808x.2020.1727627
1153	Hayes P, Turner BL, Lambers H, Laliberté E (2014) Foliar nutrient concentrations and resorption
1154	efficiency in plants of contrasting nutrient-acquisition strategies along a 2-million-year dune
1155	chronosequence. J Ecol 102:396-410. https://doi.org/10.1111/1365-2745.12196
1156	Haynes RJ (2014) A contemporary overview of silicon availability in agricultural soils. J Plant
1157	Nutr Soil Sci 177:831–844. https://doi.org/10.1002/jpln.201400202
1158	Haynes RJ (2017) The nature of biogenic Si and its potential role in Si supply in agricultural
1159	soils. Agric Ecosyst Environ 245:100–111. https://doi.org/10.1016/j.agee.2017.04.021
1160	Haynes RJ (2019) What effect does liming have on silicon availability in agricultural soils?

1161	Geoderma 337:375–383. https://doi.org/10.1016/j.geoderma.2018.09.026
1162	Haynes RJ, Zhou YF (2018) Effect of pH and added slag on the extractability of Si in two Si-
1163	deficient sugarcane soils. Chemosphere 193:431–437.
1164	https://doi.org/10.1016/j.chemosphere.2017.10.175
1165	He H, Wu M, Guo L, et al (2020) Release of tartrate as a major carboxylate by alfalfa (Medicago
1166	sativa L.) under phosphorus deficiency and the effect of soil nitrogen supply. Plant Soil
1167	449:169–178. https://doi.org/10.1007/s11104-020-04481-9
1168	Henriet C, Bodarwé L, Dorel M, et al (2008a) Leaf silicon content in banana ( <i>Musa</i> spp.) reveals
1169	the weathering stage of volcanic ash soils in Guadeloupe. Plant Soil 313:71–82.
1170	https://doi.org/10.1007/s11104-008-9680-7
1171	Henriet C, De Jaeger N, Dorel M, et al (2008b) The reserve of weatherable primary silicates
1172	impacts the accumulation of biogenic silicon in volcanic ash soils. Biogeochemistry 90:209-
1173	223. https://doi.org/10.1007/s10533-008-9245-0
1174	Herrero M, Thronton PK, Notenbaert AM, et al (2010) Smart investments in sustainable
1175	production: Revisiting mixed crop-livestock systems. Science (80- ) 327:822–825
1176	Hilbrandt I, Lehmann V, Zietzschmann F, et al (2019) Quantification and isotherm modelling of
1177	competitive phosphate and silicate adsorption onto micro-sized granular ferric hydroxide.
1178	RSC Adv 9:23642–23651. https://doi.org/10.1039/c9ra04865k
1179	Hingston FJ, Posner AM, Quirk JP (1972) Anion adsoprtion by goethite and gibbsite. I. The role
1180	of the proton in determining adsorption envelopes. J Soil Sci 23:177–192

1181	Hingston FJ, Raupach M (1967) The reaction between monosilicic acid and aluminium
1182	hydroxide. Aust J Soil Res 5:295–309
1183	Hinsinger P, Betencourt E, Bernard L, et al (2011) P for two, sharing a scarce resource: Soil
1184	phosphorus acquisition in the rhizosphere of intercropped species. Plant Physiol 156:1078-
1185	1086. https://doi.org/10.1104/pp.111.175331
1186	Hinsinger P, Fernandes Barros ON, Benedetti MF, et al (2001) Plant-induced weathering of a
1187	basaltic rock: Experimental evidence. Geochim Cosmochim Acta 65:137–152.
1188	https://doi.org/10.1016/S0016-7037(00)00524-X
1189	Hodson MJ (2019) The relative importance of cell wall and lumen phytoliths in carbon
1190	sequestration in soil: A hypothesis. Front Earth Sci 7:167.
1191	https://doi.org/10.3389/feart.2019.00167
1192	Hodson MJ, White PJ, Mead A, Broadley MR (2005) Phylogenetic variation in the silicon
1193	composition of plants. Ann Bot 96:1027–1046. https://doi.org/10.1093/aob/mci255
1194	Högberg P, Nordgren A, Buchmann N, et al (2001) Large-scale forest girdling shows that current
1195	photosynthesis drives soil respiration. Nature 411:789–792.
1196	https://doi.org/10.1038/35081058
1197	Hömberg A, Obst M, Knorr KH, et al (2020) Increased silicon concentration in fen peat leads to a
1198	release of iron and phosphate and changes in the composition of dissolved organic matter.
1199	Geoderma 374:114422. https://doi.org/10.1016/j.geoderma.2020.114422
1200	Hossain KA, Horiuchi T, Miyagawa S (2001) Effects of silicate materials on growth and grain

1201	yield of rice plants grown in clay loam and sandy loam soils. J Plant Nutr 24:1–13.
1202	https://doi.org/10.1081/PLN-100000308
1203	Houben D, Sonnet P (2012) Zinc mineral weathering as affected by plant roots. Appl
1204	Geochemistry 27:1587–1592. https://doi.org/10.1016/j.apgeochem.2012.05.004
1205	Houben D, Sonnet P, Cornelis JT (2014) Biochar from Miscanthus: A potential silicon fertilizer
1206	Plant Soil 374:871–882. https://doi.org/10.1007/s11104-013-1885-8
1207	Hu L, Xia M, Lin X, et al (2018) Earthworm gut bacteria increase silicon bioavailability and
1208	acquisition by maize. Soil Biol Biochem 125:215–221.
1209	https://doi.org/10.1016/j.soilbio.2018.07.015
1210	Hu L, Xu CC, Wang J, et al (2019) Application of bryophyte rhizoid-associated bacteria
1211	increases silicon accumulation and growth in maize (Zea mays L.) seedlings. Appl Ecol
1212	Environ Res 17:13423–13433. https://doi.org/10.15666/aeer/1706_1342313433
1213	Huang F, Gao LY, Wu RR, et al (2020) Qualitative and quantitative characterization of
1214	adsorption mechanisms for Cd2+ by silicon-rich biochar. Sci Total Environ 731:139163.
1215	https://doi.org/10.1016/j.scitotenv.2020.139163
1216	Hughes HJ, Hung DT, Sauer D (2020) Silicon recycling through rice residue management does
1217	not prevent silicon depletion in paddy rice cultivation. Nutr Cycl Agroecosystems 118:75-
1218	89. https://doi.org/10.1007/s10705-020-10084-8
1219	Hummel J, Findeisen E, Südekum KH, et al (2011) Another one bites the dust: Faecal silica
1220	levels in large herbivores correlate with high-crowned teeth. Proc R Soc B Biol Sci

1221	278:1742–1747. https://doi.org/10.1098/rspb.2010.1939
1222	Hwang BC, Metcalfe DB (2021) Reviews and syntheses: Impacts of plant-silica-herbivore
1223	interactions on terrestrial biogeochemical cycling. Biogeosciences 18:1259–1268.
1224	https://doi.org/10.5194/bg-18-1259-2021
1225	Ibrahim M, Khan S, Hao X, Li G (2016) Biochar effects on metal bioaccumulation and arsenic
1226	speciation in alfalfa (Medicago sativa L.) grown in contaminated soil. Int J Environ Sci
1227	Technol 13:2467–2474. https://doi.org/10.1007/s13762-016-1081-5
1228	Icopini GA, Brantley SL, Heaney PJ (2005) Kinetics of silica oligomerization and nanocolloid
1229	formation as a function of pH and ionic strength at 25°C. Geochim Cosmochim Acta
1230	69:293–303. https://doi.org/10.1016/j.gca.2004.06.038
1231	Ingham RE, Trofymow JA, Ingham ER, Coleman DC (1985) Interactions of bacteria, fungi, and
1232	their nematode grazers: Effects on nutrient cycling and plant growth. Ecol Monogr 55:119-
1233	140. https://doi.org/10.2307/1942528
1234	Jackson TA (1971) Biochemical weathering of calcium-bearing minerals by rhizosphere micro-
1235	organisms, and its influence on calcium accumulation in trees. Plant Soil 35:655-658
1236	Jiang Y, Wang W, Xie Q, et al (2017) Plants transfer lipids to sustain colonization by mutualistic
1237	mycorrhizal and parasitic fungi. Science (80- ) 356:1172–1173.
1238	https://doi.org/10.1126/science.aam9970
1239	Johnson SN, Hartley SE, Moore BD (2021) Silicon defence in plants: Does herbivore identity
1240	matter? Trends Plant Sci 26:99-101. https://doi.org/10.1016/j.tplants.2020.10.005

1241	soils. Nature 198:852–853
1243	Jordan N, Marmier N, Lomenech C, et al (2009) Competition between selenium (IV) and silicic
1244	acid on the hematite surface. Chemosphere 75:129–134.
1245	https://doi.org/10.1016/j.chemosphere.2008.11.018
1246	Jouquet P, Jamoteau F, Majumdar S, et al (2020) The distribution of Silicon in soil is influenced
1247	by termite bioturbation in South Indian forest soils. Geoderma 372:114362.
1248	https://doi.org/10.1016/j.geoderma.2020.114362
1249	Jouquet P, Mamou L, Lepage M, Velde B (2002) Effect of termites on clay minerals in tropical
1250	soils: Fungus-growing termites as weathering agents. Eur J Soil Sci 53:521–528.
1251	https://doi.org/10.1046/j.1365-2389.2002.00492.x
1252	Jouquet P, Traoré S, Choosai C, et al (2011) Influence of termites on ecosystem functioning.
1253	Ecosystem services provided by termites. Eur J Soil Biol 47:215–222.
1254	https://doi.org/10.1016/j.ejsobi.2011.05.005
1255	Kabas S, Saavedra-Mella F, Huynh T, et al (2017) Metal uptake and organic acid exudation of
1256	native <i>Acacia</i> species in mine tailings. Aust J Bot 65:357–367.
1257	https://doi.org/10.1071/BT16189
1258	Kang SM, Waqas M, Shahzad R, et al (2017) Isolation and characterization of a novel silicate-
1259	solubilizing bacterial strain Burkholderia eburnea CS4-2 that promotes growth of japonica
1260	rice (Oryza sativa L. cv. Dongjin). Soil Sci Plant Nutr 63:233–241.
1261	https://doi.org/10.1080/00380768.2017.1314829

1262	Katz O (2019) Silicon content is a plant functional trait: implications in a changing world. Flora
1263	254:88–94. https://doi.org/https://doi.org/10.1016/j.flora.2018.08.007
1264	Keeping MG, Miles N, Rutherford RS (2017) Liming an acid soil treated with diverse silicon
1265	sources: Effects on silicon uptake by sugarcane (Saccharum spp. hybrids). J Plant Nutr
1266	40:1417–1436. https://doi.org/10.1080/01904167.2016.1267751
1267	Keller C, Guntzer F, Barboni D, et al (2012) Impact of agriculture on the Si biogeochemical
1268	cycle: Input from phytolith studies. Comptes Rendus - Geosci 344:739–746.
1269	https://doi.org/10.1016/j.crte.2012.10.004
1270	Keller C, Rizwan M, Meunier J-D (2021) Are clay minerals a significant source of Si for crops?
1271	A comparison of amorphous silica and the roles of the mineral type and pH. Silicon In press
1272	https://doi.org/10.1007/s12633-020-00877-5
1273	Kelly EF, Chadwick OA, Hilinski TE (1998) The effect of plants on mineral weathering.
1274	Biogeochemistry 42:21–53
1275	Kim J, Dong H, Seabaugh J, et al (2004) Role of microbes in the smectite-to-illite reaction.
1276	Science (80- ) 303:830–832. https://doi.org/10.1126/science.1093245
1277	Klotzbücher A, Klotzbücher T, Jahn R, et al (2018a) Effects of Si fertilization on Si in soil
1278	solution, Si uptake by rice, and resistance of rice to biotic stresses in Southern Vietnam.
1279	Paddy Water Environ 16:243–252. https://doi.org/10.1007/s10333-017-0610-2
1280	Klotzbücher T, Klotzbücher A, Kaiser K, et al (2018b) Impact of agricultural practices on plant-
1281	available silicon. Geoderma 331:15–17. https://doi.org/10.1016/j.geoderma.2018.06.011

1282	Klotzbücher T, Marxen A, Vetterlein D, et al (2015) Plant-available silicon in paddy soils as a
1283	key factor for sustainable rice production in Southeast Asia. Basic Appl Ecol 16:665-673.
1284	https://doi.org/10.1016/j.baae.2014.08.002
1285	Klotzbücher T, Treptow C, Kaiser K, et al (2020) Sorption competition with natural organic
1286	matter as mechanism controlling silicon mobility in soil. Sci Rep 1–11.
1287	https://doi.org/10.1038/s41598-020-68042-x
1288	Konhauser KO, Lalonde S V., Amskold L, Holland HD (2007) Was there really an archean
1289	phosphate crisis? Science (80- ) 315:1234. https://doi.org/10.1126/science.1136328
1290	Kostic L, Nikolic N, Bosnic D, et al (2017) Silicon increases phosphorus (P) uptake by wheat
1291	under low P acid soil conditions. Plant Soil 419:447–455. https://doi.org/10.1007/s11104-
1292	017-3364-0
1293	Kothari SK, Marschner H, Römheld V (1990) Direct and indirect effects of VA mycorrhizal
1294	fungi and rhizosphere microorganisms on acquisition of mineral nutrients by maize (Zea
1295	mays L.) in a calcareous soil. New Phytol 116:637-645. https://doi.org/10.1111/j.1469-
1296	8137.1990.tb00549.x
1297	Koyama S, Hayashi H (2017) Rice yield and soil carbon dynamics over three years of applying
1298	rice husk charcoal to an Andosol paddy field. Plant Prod Sci 20:176-182.
1299	https://doi.org/10.1080/1343943X.2017.1290506
1300	Kundu S, Kamath MB, Goswami NN (1988) Effect of sulphate, silicate and fluoride anions - I.
1301	Phosphate fixation in soils. J Indian Soc Soil Sci 36:43–47

1302	Kuzyakov Y, Blagodatskaya E (2015) Microbial hotspots and hot moments in soil: Concept &
1303	review. Soil Biol Biochem 83:184–199. https://doi.org/10.1016/j.soilbio.2015.01.025
1304	Lambers H, Albornoz F, Kotula L, et al (2018) How belowground interactions contribute to the
1305	coexistence of mycorrhizal and non-mycorrhizal species in severely phosphorus-
1306	impoverished hyperdiverse ecosystems. Plant Soil 424:11–33.
1307	https://doi.org/10.1007/s11104-017-3427-2
1308	Lambers H, Finnegan PM, Laliberté E, et al (2011) Phosphorus nutrition of proteaceae in
1309	severely phosphorus-impoverished soils: Are there lessons to be learned for future crops?
1310	Plant Physiol 156:1058–1066. https://doi.org/10.1104/pp.111.174318
1311	Lambers H, Hayes PE, Laliberté E, et al (2015) Leaf manganese accumulation and phosphorus-
1312	acquisition efficiency. Trends Plant Sci 20:83-90.
1313	https://doi.org/10.1016/j.tplants.2014.10.007
1314	Lambers H, Raven JA, Shaver GR, Smith SE (2008) Plant nutrient-acquisition strategies change
1315	with soil age. Trends Ecol Evol 23:95–103. https://doi.org/10.1016/j.tree.2007.10.008
1316	Lambers H, Shane MW, Cramer MD, et al (2006) Root structure and functioning for efficient
1317	acquisition of phosphorus: Matching morphological and physiological traits. Ann Bot
1318	98:693–713. https://doi.org/10.1093/aob/mcl114
1319	Lauwers AM, Heinen W (1974) Bio-degradation and utilization of silica and quartz. Arch
1320	Microbiol 95:67–78. https://doi.org/10.1007/BF02451749
1321	le Roux E, van Veenhuisen LS, Kerley GIH, Cromsigt JPGM (2020) Animal body size

1322	distribution influences the ratios of nutrients supplied to plants. Proc Natl Acad Sci U S A
1323	117:22256–22263. https://doi.org/10.1073/pnas.2003269117
1324	Leake JR, Read DJ (2017) Mycorrhizal Symbioses and Pedogenesis Throughout Earth's History.
1325	In: Mycorrhizal Mediation of Soil: Fertility, Structure, and Carbon Storage. Elsevier Inc., pp
1326	9–33
1327	Lee KE, Adhikari A, Kang SM, et al (2019) Isolation and characterization of the high silicate and
1328	phosphate solubilizing novel strain enterobacter ludwigii GAK2 that promotes growth in
1329	rice plants. Agronomy 9:. https://doi.org/10.3390/agronomy9030144
1330	Leksungnoen P, Wisawapipat W, Ketrot D, et al (2019) Biochar and ash derived from silicon-rich
1331	rice husk decrease inorganic arsenic species in rice grain. Sci Total Environ 684:360–370.
1332	https://doi.org/10.1016/j.scitotenv.2019.05.247
1333	Lemaire G, Franzluebbers A, Carvalho PC de F, Dedieu B (2014) Integrated crop-livestock
1334	systems: Strategies to achieve synergy between agricultural production and environmental
1335	quality. Agric Ecosyst Environ 190:4–8. https://doi.org/10.1016/j.agee.2013.08.009
1336	Li C, Hoffland E, Kuyper TW, et al (2020a) Syndromes of production in intercropping impact
1337	yield gains. Nat Plants 6:653-660. https://doi.org/10.1038/s41477-020-0680-9
1338	Li L, Tilman D, Lambers H, Zhang FS (2014) Plant diversity and overyielding: Insights from
1339	belowground facilitation of intercropping in agriculture. New Phytol 203:63-69.
1340	https://doi.org/10.1111/nph.12778
1341	Li Z, de Tombeur F, Linden C Vander, et al (2020b) Soil microaggregates store phytoliths in a

1342	sandy loam. Geoderma 360:114037. https://doi.org/10.1016/j.geoderma.2019.114037
1343	Li Z, Delvaux B (2019) Phytolith- rich biochar: A potential Si fertilizer in desilicated soils. GCB
1344	Bioenergy 11:. https://doi.org/10.1111/gcbb.12635
1345	Li Z, Delvaux B, Yans J, et al (2018) Phytolith-rich biochar increases cotton biomass and silicon-
1346	mineralomass in a highly weathered soil. J Plant Nutr Soil Sci.
1347	https://doi.org/10.1002/jpln.201800031
1348	Li Z, Unzué-Belmonte D, Cornelis J-T, et al (2019) Effects of phytolithic rice-straw biochar, soil
1349	buffering capacity and pH on silicon bioavailability. Plant Soil.
1350	https://doi.org/10.1007/s11104-019-04013-0
1351	Liang Y, Nikolic M, Bélanger R, et al (2015a) Silicon biogeochemistry and bioavailability in
1352	soil. In: Silicon in Agriculture. pp 45–68
1353	Liang Y, Nikolic M, Bélanger RR, et al (2015b) Silicon in Agriculture. From theory to practice.
1354	Liang Y, Sun W, Zhu YG, Christie P (2007) Mechanisms of silicon-mediated alleviation of
1355	abiotic stresses in higher plants: A review. Environ Pollut 147:422–428.
1356	https://doi.org/10.1016/j.envpol.2006.06.008
1357	Liermann LJ, Kalinowski BE, Brantley SL, Ferry JG (2000) Role of bacterial siderophores in
1358	dissolution of hornblende. Geochim Cosmochim Acta 64:587-602.
1359	https://doi.org/10.1016/S0016-7037(99)00288-4
1360	Limmer MA, Mann J, Amaral DC, et al (2018) Silicon-rich amendments in rice paddies: Effects
1361	on arsenic uptake and biogeochemistry. Sci Total Environ 624:1360–1368.

1362	https://doi.org/10.1016/j.scitotenv.2017.12.207
1363	Liu W, Xu X, Wu X, et al (2006) Decomposition of silicate minerals by Bacillus mucilaginosus
1364	in liquid culture. Environ Geochem Health 28:133–140. https://doi.org/10.1007/s10653-005-
1365	9022-0
1366	Liu X, Li L, Bian R, et al (2014) Effect of biochar amendment on soil-silicon availability and rice
1367	uptake. J Plant Nutr Soil Sci 177:91–96. https://doi.org/10.1002/jpln.201200582
1368	Lucas Y (2001) The role of plants in controlling rates and products of weathering: Importance of
1369	biological pumping. Annu Rev Earth Planet Sci 29:135-163.
1370	https://doi.org/10.1002/fut.10088
1371	Lucas Y, Luizão FJ, Chauvel A, et al (1993) The relation between biological activity of the rain
1372	forest and mineral composition of soils. Science (80- ) 260:521–523.
1373	https://doi.org/10.1126/science.260.5107.521
1374	Łukowiak M (2020) Utilizing sponge spicules in taxonomic, ecological and environmental
1375	reconstructions: a review. Peer J e10601. https://doi.org/https://doi.org/10.7717/peerj.10601
1376	Ma J, Takahashi E (1991a) Effect of silicate on phosphate availability for rice in a P-deficient
1377	soil. Plant Soil 133:151–155. https://doi.org/10.1007/BF00009187
1378	Ma J, Takahashi E (1990a) The effect of silicic acid on rice in a P-deficient soil. Plant Soil
1379	126:121–125. https://doi.org/10.1007/BF00041377
1380	Ma J, Takahashi E (1991b) Availability of rice straw Si to rice plants. Soil Sci Plant Nutr 37:111-
1381	116. https://doi.org/10.1080/00380768.1991.10415016

1382	Ma JF (2004) Role of silicon in enhancing the resistance of plants to biotic and abiotic stresses.
1383	Soil Sci Plant Nutr 50:11–18. https://doi.org/10.1080/00380768.2004.10408447
1384	Ma JF, Takahashi E (1990b) Effect of silicon on the growth and phosphorus uptake of rice. Plant
1385	Soil 126:115–119. https://doi.org/10.1007/BF00041376
1386	Ma JF, Tamai K, Yamaji N, et al (2006) A silicon transporter in rice. Nature 440:688-691.
1387	https://doi.org/10.1038/nature04590
1388	Ma JF, Yamaji N, Mitani N, et al (2007) An efflux transporter of silicon in rice. Nature 448:209–
1389	212. https://doi.org/10.1038/nature05964
1390	Mariotte P, Mehrabi Z, Bezemer TM, et al (2018) Plant-Soil Feedback: Bridging Natural and
1391	Agricultural Sciences. Trends Ecol Evol 33:129–142.
1392	https://doi.org/10.1016/j.tree.2017.11.005
1393	Marschner H, Dell B (1994) Nutrient uptake in mycorrhizal symbiosis. Plant Soil 159:89–102.
1394	https://doi.org/10.1007/BF00000098
1395	Martin-Guay MO, Paquette A, Dupras J, Rivest D (2018) The new Green Revolution:
1396	Sustainable intensification of agriculture by intercropping. Sci Total Environ 615:767–772.
1397	https://doi.org/10.1016/j.scitotenv.2017.10.024
1398	Marxen A, Klotzbücher T, Jahn R, et al (2016) Interaction between silicon cycling and straw
1399	decomposition in a silicon deficient rice production system. Plant Soil 398:153–163.
1400	https://doi.org/10.1007/s11104-015-2645-8
1401	Mathews BW, Carpenter JR, Sollenberger LE (2009) In vitro digestibility and chemical

1402	composition of kikuyugrass as influenced by soil silicon, liming, and genotype. Commun
1403	Soil Sci Plant Anal 40:2855–2873. https://doi.org/10.1080/00103620903173855
1404	Matychenkov VV, Ammosova YM (1996) Effect of amorphous silica on some properties of a
1405	sod-podzolic soil. Eurasian Soil Sci 28:87–99
1406	McKeague JA, Cline MG (1963) Silica in soil solutions. II. The adsorption of monosilicic acid by
1407	soil and by other substances. Can J Soil Sci 43:83–96
1408	Meena VD, Dotaniya ML, Coumar V, et al (2014) A case for silicon fertilization to improve crop
1409	yields in tropical soils. Proc Natl Acad Sci India Sect B - Biol Sci 84:505-518.
1410	https://doi.org/10.1007/s40011-013-0270-y
1411	Meharg C, Meharg AA (2015) Silicon, the silver bullet for mitigating biotic and abiotic stress,
1412	and improving grain quality, in rice? Environ Exp Bot 120:8–17.
1413	https://doi.org/10.1016/j.envexpbot.2015.07.001
1414	Meunier JD, Colin F, Alarcon C (1999) Biogenic silica storage in soils. Geology 27:835–838.
1415	https://doi.org/10.1130/0091-7613(1999)027<0835:BSSIS>2.3.CO;2
1416	Meunier JD, Sandhya K, Prakash NB, et al (2018) pH as a proxy for estimating plant-available
1417	Si? A case study in rice fields in Karnataka (South India). Plant Soil 432:143-155.
1418	https://doi.org/10.1007/s11104-018-3758-7
1419	Meyer JH, Keeping MG (2000) Review of Research Into the Role of Silicon for Sugarcane
1420	Production. Proc South African Sugar Technol Assoc 74:29–40
1421	Mikha MM, Rice CW (2004) Tillage and Manure Effects on Soil and Aggregate-Associated

1422	Carbon and Nitrogen. Soil Sci Soc Am J 68:809. https://doi.org/10.2136/sssaj2004.0809
1423	Miles N, Manson AD, Rhodes R, et al (2014) Extractable silicon in soils of the South African
1424	sugar industry and relationships with crop uptake. Commun Soil Sci Plant Anal 45:2949-
1425	2958. https://doi.org/10.1080/00103624.2014.956881
1426	Müller DWH, Caton J, Codron D, et al (2011) Phylogenetic constraints on digesta separation:
1427	Variation in fluid throughput in the digestive tract in mammalian herbivores. Comp
1428	Biochem Physiol Part A Mol Integr Physiol 160:207–220
1429	Nakamura R, Cornelis J, de Tombeur F, et al (2020) Diversity of silicon release rates among
1430	tropical tree species during leaf-litter decomposition. Geoderma 368:.
1431	https://doi.org/10.1016/j.geoderma.2020.114288
1432	Neu S, Schaller J, Dudel EG (2017) Silicon availability modifies nutrient use efficiency and
1433	content, C:N:P stoichiometry, and productivity of winter wheat (Triticum aestivum L.). Sci
1434	Rep 7:1–8. https://doi.org/10.1038/srep40829
1435	Nguyen MN, Picardal F, Dultz S, et al (2017) Silicic acid as a dispersibility enhancer in a Fe-
1436	oxide-rich kaolinitic soil clay. Geoderma 286:8–14.
1437	https://doi.org/10.1016/j.geoderma.2016.10.029
1438	Ning C, Qu J, He L, et al (2017) Improvement of yield, pest control and Si nutrition of rice by
1439	rice-water spinach intercropping. F Crop Res 208:34–43.
1440	https://doi.org/10.1016/j.fcr.2017.04.005
1///1	Nottle MC Armstrong IM (1966) Urinary excretion of silica by grazing sheen. Aust I Agric Re

1442	17:165–173
1443	Obihara CH, Russell EW (1972) Specific adsorption of silicate and phosphate by soils. J Soil Sci
1444	23:105–117
1445	Or D, Keller T, Schlesinger WH (2021) Natural and managed soil structure: On the fragile
1446	scaffolding for soil functioning. Soil Tillage Res 208:104912.
1447	https://doi.org/10.1016/j.still.2020.104912
1448	Owino-Gerroh C, Gascho GJ (2004) Effect of silicon on low pH soil phosphorus sorption and on
L449	uptake and growth of maize. Commun Soil Sci Plant Anal 35:2369-2378.
1450	https://doi.org/10.1081/LCSS-200030686
1451	Oye Anda CC, Opfergelt S, Declerck S (2016) Silicon acquisition by bananas (c.V. Grande
1452	Naine) is increased in presence of the arbuscular mycorrhizal fungus Rhizophagus
1453	irregularis MUCL 41833. Plant Soil 409:77–85. https://doi.org/10.1007/s11104-016-2954-6
L454	Pang J, Bansal R, Zhao H, et al (2018) The carboxylate-releasing phosphorus-mobilizing strategy
1455	can be proxied by foliar manganese concentration in a large set of chickpea germplasm
1456	under low phosphorus supply. New Phytol 219:518–529. https://doi.org/10.1111/nph.15200
1457	Pastore G, Kernchen S, Spohn M (2020) Microbial solubilization of silicon and phosphorus from
1458	bedrock in relation to abundance of phosphorus-solubilizing bacteria in temperate forest
1459	soils. Soil Biol Biochem 151:108050. https://doi.org/10.1016/j.soilbio.2020.108050
1460	Pavlovic J, Kostic L, Bosnic P, et al (2021) Interactions of silicon with essential and beneficial
1/161	elements in plants. Front Plant Sci 12:697592, https://doi.org/10.3389/fpls.2021.697592

1462	Pawlik Ł, Phillips JD, Samonil P (2016) Roots, rock, and regolith: Biomechanical and
1463	biochemical weathering by trees and its impact on hillslopes—A critical literature review.
1464	Earth-Science Rev 159:142–159. https://doi.org/10.1016/j.earscirev.2016.06.002
1465	Philippini V, Naveau A, Catalette H, Leclercq S (2006) Sorption of silicon on magnetite and
1466	other corrosion products of iron. J Nucl Mater 348:60-69.
1467	https://doi.org/10.1016/j.jnucmat.2005.09.002
1468	Phonde DB, Deshmukh PS, Banerjee K, Adsule PG (2014) Plant available silicon in sugarcane
1469	soils and its relationship with soil properties, leaf silicon and cane yield. An Asian J Soil Sci
1470	9:176–180. https://doi.org/10.15740/has/ajss/9.2/176-180
1471	Puppe D (2020) Review on protozoic silica and its role in silicon cycling. Geoderma 365:114224
1472	https://doi.org/10.1016/j.geoderma.2020.114224
1473	Puppe D, Ehrmann O, Kaczorek D, et al (2015) The protozoic Si pool in temperate forest
1474	ecosystems - Quantification, abiotic controls and interactions with earthworms. Geoderma
1475	243-244:196-204. https://doi.org/10.1016/j.geoderma.2014.12.018
1476	Puppe D, Höhn A, Kaczorek D, et al (2016) As time goes by-Spatiotemporal changes of biogenic
1477	Si pools in initial soils of an artificial catchment in NE Germany. Appl Soil Ecol 105:9–16.
1478	https://doi.org/10.1016/j.apsoil.2016.01.020
1479	Puppe D, Kaczorek D, Schaller J, et al (2021) Crop straw recycling prevents anthropogenic
1480	desilication of agricultural soil-plant systems in the temperate zone - Results from a long-
1481	term field experiment in NE Germany. Geoderma 403:115187.
1482	https://doi.org/10.1016/j.geoderma.2021.115187

1483	Puppe D, Kaczorek D, Wanner M, Sommer M (2014) Dynamics and drivers of the protozoic Si
1484	pool along a 10-year chronosequence of initial ecosystem states. Ecol Eng 70:477-482.
1485	https://doi.org/10.1016/j.ecoleng.2014.06.011
1486	Putra R, Powell JR, Hartley SE, Johnson SN (2020) Is it time to include legumes in plant silicon
1487	research? Funct Ecol 1–16. https://doi.org/10.1111/1365-2435.13565
1488	Qian L, Chen B, Chen M (2016) Novel Alleviation Mechanisms of Aluminum Phytotoxicity via
1489	Released Biosilicon from Rice Straw-Derived Biochars. Sci Rep 6:29346.
1490	https://doi.org/10.1038/srep29346
1491	Raturi G, Sharma Y, Rana V, et al (2021) Exploration of silicate solubilizing bacteria for
1492	sustainable agriculture and silicon biogeochemical cycle. Plant Physiol Biochem In press:
1493	https://doi.org/https://doi.org/10.1016/j.plaphy.2021.06.039
1494	Raven JA (1983) The transport and function of silicon in plants. Biol Rev 58:179–207
1495	Reeves DW (1994) Cover crops and rotations. In: Crops Residue Management. pp 125–172
1496	Reichard PU, Kraemer SM, Frazier SW, Kretzschmar R (2005) Goethite dissolution in the
1497	presence of phytosiderophores: rates, mechanisms, and the synergistic effect of oxalate.
1498	Plant Soil 276:115–132. https://doi.org/10.1007/s11104-005-3504-9
1499	Reithmaier G-MS, Knorr K-H, Arnhold S, et al (2017) Enhanced silicon availability leads to
1500	increased methane production, nutrient and toxicant mobility in peatlands. Sci Rep 7:8728.
1501	https://doi.org/10.1038/s41598-017-09130-3
1502	Rich MK, Nouri E, Courty PE, Reinhardt D (2017) Diet of arbuscular mycorrhizal fungi: bread

1503	and butter? Trends Plant Sci 22:652–660. https://doi.org/10.1016/j.tplants.2017.05.008
1504	Richardson AE, Barea JM, McNeill AM, Prigent-Combaret C (2009) Acquisition of phosphorus
1505	and nitrogen in the rhizosphere and plant growth promotion by microorganisms. Plant Soil
1506	321:305–339. https://doi.org/10.1007/s11104-009-9895-2
1507	Richardson AE, Lynch JP, Ryan PR, et al (2011) Plant and microbial strategies to improve the
1508	phosphorus efficiency of agriculture. Plant Soil 349:121-156.
1509	https://doi.org/10.1007/s11104-011-0950-4
1510	Ritchie H, Roser M (2013) "Crop Yields." In: Publ. online OurWorldInData.org. Retrieved from
1511	"https://ourworldindata.org/crop-yields"
1512	Roarty S, Hackett RA, Schmidt O (2017) Earthworm populations in twelve cover crop and weed
1513	management combinations. Appl Soil Ecol 114:142–151.
1514	https://doi.org/10.1016/j.apsoil.2017.02.001
1515	Rohatgi A (2012) WebPlotDigitalizer: HTML5 based online tool to extract numerical data from
1516	plot images. Version 4.2
1517	Russelle MP, Entz MH, Franzluebbers AJ (2007) Reconsidering integrated crop-livestock
1518	systems in North America. Agron J 99:325–334. https://doi.org/10.2134/agronj2006.0139
1519	Sauer D, Saccone L, Conley DJ, et al (2006) Review of methodologies for extracting plant-
1520	available and amorphous Si from soils and aquatic sediments. Biogeochemistry 80:89-108.
1521	https://doi.org/10.1007/s10533-005-5879-3
1522	Savant NK, Korndörfer GH, Datnoff LE, Snyder GH (1999) Silicon nutrition and sugarcane

1523	production: A review. J Plant Nutr 22:1853–1903.
1524	https://doi.org/10.1080/01904169909365761
1525	Schaller J, Frei S, Rohn L, Gilfedder BS (2020) Amorphous Silica Controls Water Storage
1526	Capacity and Phosphorus Mobility in Soils. Front Environ Sci 8:94.
1527	https://doi.org/10.3389/fenvs.2020.00094
1528	Schaller J, Puppe D, Kaczorek D, et al (2021) Silicon cycling in soils revisited. Plants 10:295.
1529	https://doi.org/10.3390/plants10020295
1530	Schaller J, Turner BL, Weissflog A, et al (2018) Silicon in tropical forests: large variation across
1531	soils and leaves suggests ecological significance. Biogeochemistry 140:161-174.
1532	https://doi.org/10.1007/s10533-018-0483-5
1533	Schmidt O, Clements RO, Donaldson G (2003) Why do cereal-legume intercrops support large
1534	earthworm populations? Appl Soil Ecol 22:181–190. https://doi.org/10.1016/S0929-
1535	1393(02)00131-2
1536	Schmidt O, Curry JP, Hackett RA, et al (2001) Earthworm communities in conventional wheat
1537	monocropping and low-input wheat-clover intercropping systems. Ann Appl Biol 138:377-
1538	388. https://doi.org/10.1111/j.1744-7348.2001.tb00123.x
1539	Schoelynck J, Subalusky AL, Struyf E, et al (2019) Hippos (Hippopotamus amphibius): The
1540	animal silicon pump. Sci Adv 5:eaav0395
1541	Schröer HC, Krasko A, Le Pennec G, et al (2003) Silicase, an Enzyme Which Degrades
1542	Biogenous Amorphous Silica: Contribution to the Metabolism of Silica Deposition in the

1543	Demosponge Suberites domuncula. In: Springer (ed) Silicon Biomineralization. Progress in
1544	Molecular and Subcellular Biology, vol 33. Berlin, pp 249–268
1545	Schulmann OP, Tiunov, Alexei V (1999) Leaf litter fragmentation by the earthworm <i>Lumbricus</i>
1546	terrestris L. Pedobiologia (Jena) 43:453–458
1547	Seleiman MF, Refay Y, Al-Suhaibani N, et al (2019) Integrative Effects of Rice-Straw Biochar
1548	and Silicon on Oil and Seed Quality, Yield and Physiological Traits of Helianthus annuus L
1549	. Grown under Water Deficit Stress. Agronomy 9:1–21
1550	Seyfferth AL, Kocar BD, Lee JA, Fendorf S (2013) Seasonal dynamics of dissolved silicon in a
1551	rice cropping system after straw incorporation. Geochim Cosmochim Acta 123:120-133.
1552	https://doi.org/10.1016/j.gca.2013.09.015
1553	Shewmaker GE, Mayland HF, Rosenau RC, Asay KH (1989) Silicon in C-3 Grasses: Effects on
1554	Forage Quality and Sheep Preference. J Range Manag 42:122.
1555	https://doi.org/10.2307/3899308
1556	Singh KP, Sarkar MC (1992) Phosphorus availability in soils as affected by fertilizer phosphorus,
1557	sodium silicate and farmyeard manure. J Indian Soc Soil Sci 40:762-767
1558	Sistani KR, Savant NK, Reddy KC (1997) Effect of rice hull ash silicon on rice seedling growth.
1559	J Plant Nutr 20:195–201. https://doi.org/10.1080/01904169709365242
1560	Sitters J, Kimuyu DM, Young TP, et al (2020) Negative effects of cattle on soil carbon and
1561	nutrient pools reversed by megaherbivores. Nat Sustain 3:360–366.
1562	https://doi.org/10.1038/s41893-020-0490-0

1563	Six J, Bossuyt H, Degryze S, Denef K (2004) A history of research on the link between
1564	(micro)aggregates, soil biota, and soil organic matter dynamics. Soil Tillage Res 79:7–31.
1565	https://doi.org/10.1016/j.still.2004.03.008
1566	Smith HA, McSorley R (2000) Intercropping and pest management: A Review of major concepts
1567	Am Entomol 46:154–161. https://doi.org/10.1093/ae/46.3.154
1568	Smith S, Read D (2008) Mycorrhizal Symbiosis. Elsevier
1569	Smits MM, Hoffland E, Jongmans AG, van Breemen N (2005) Contribution of mineral tunneling
1570	to total feldspar weathering. Geoderma 125:59-69.
1571	https://doi.org/10.1016/j.geoderma.2004.06.005
1572	Smits MM, Wallander H (2017) Role of Mycorrhizal Symbiosis in Mineral Weathering and
1573	Nutrient Mining from Soil Parent Material. In: Mycorrhizal Mediation of Soil: Fertility,
1574	Structure, and Carbon Storage. Elsevier Inc., pp 35-46
1575	Smyth TJ, Sanchez PA (1980) Effect of lime, silicate, and phosphorus applications to an Oxisol
1576	on phosphorus sorption and ion retention. Soil Sci Soc Am J 44:500-505
1577	Sommer M, Jochheim H, Höhn A, et al (2013) Si cycling in a forest biogeosystem-the
1578	importance of transient state biogenic Si pools. Biogeosciences 10:4991-5007.
1579	https://doi.org/10.5194/bg-10-4991-2013
1580	Sommer M, Kaczorek D, Kuzyakov Y, Breuer J (2006) Silicon pools and fluxes in soils and
1581	landscapes—a review. J Plant Nutr Soil Sci 169:310–329.
1582	https://doi.org/10.1002/jpln.200521981

1583	Song A, Li Z, Zhang J, et al (2009) Silicon-enhanced resistance to cadmium toxicity in Brassica
1584	chinensis L. is attributed to Si-suppressed cadmium uptake and transport and Si-enhanced
1585	antioxidant defense capacity. J Hazard Mater 172:74-83.
1586	https://doi.org/10.1016/j.jhazmat.2009.06.143
1587	Song W, Ogawa N, Oguchi CT, et al (2007) Effect of <i>Bacillus subtilis</i> on granite weathering: A
1588	laboratory experiment. Catena 70:275–281. https://doi.org/10.1016/j.catena.2006.09.003
1589	Song Z, Wang H, Strong PJ, Shan S (2014) Increase of available soil silicon by Si-rich manure
1590	for sustainable rice production. Agron Sustain Dev 34:813-819.
1591	https://doi.org/10.1007/s13593-013-0202-5
1592	Soratto RP, Crusciol CAC, Castro GSA, et al (2012) Leaf application of silicic acid to white oat
1593	and wheat. Rev Bras Ciência do Solo 36:1538–1544. https://doi.org/10.1590/s0100-
1594	06832012000500018
1595	Steuer P, Südekum K-H, Müller DWH, et al (2011) Is there an influence of body mass on digesta
1596	mean retention time in herbivores? A comparative study on ungulates. Comp Biochem
1597	Physiol Part A Mol Integr Physiol 160:355–364
1598	Stillings LL, Drever JI, Brantley SL, et al (1996) Rates of feldspar dissolution at pH 3–7 with 0–8
1599	mM oxalic acid. Chem Geol 132:79–89. https://doi.org/10.1016/S0009-2541(96)00043-5
1600	Street-Perrott FA, Barker PA (2008) Biogenic silica: a neglected component of the coupled
1601	global continental biogeochemical cycles of carbon and silicon. Earth Surf Process
1602	Landforms 33:1436–1457, https://doi.org/10.1002/esp.1712

1603	Ström L, Owen AG, Godbold DL, Jones DL (2005) Organic acid behaviour in a calcareous soil
1604	implications for rhizosphere nutrient cycling. Soil Biol Biochem 37:2046–2054.
1605	https://doi.org/10.1016/j.soilbio.2005.03.009
1606	Struyf E, Smis A, Van Damme S, et al (2010) Historical land use change has lowered terrestrial
1607	silica mobilization. Nat Commun 1:129. https://doi.org/10.1038/ncomms1128
1608	Suzuki Y, Matsubara T, Hoshino M (2003) Breakdown of mineral grains by earthworms and
1609	beetle larvae. Geoderma 112:131–142. https://doi.org/10.1016/S0016-7061(02)00300-2
1610	Tang F, White JA, Charvat I (2001) The effect of phosphorus availability on arbuscular
1611	mycorrhizal colonization of Typha angustifolia. Mycologia 93:1042–1047
1612	Tang X, Zhang C, Yu Y (2020) Intercropping legumes and cereals increases phosphorus use
1613	efficiency; a meta-analysis. Plant Soil In press:
1614	Tavakkoli E, Lyons G, English P, Guppy CN (2011) Silicon nutrition of rice is affected by soil
1615	pH, weathering and silicon fertilisation. J Plant Nutr Soil Sci 174:437–446.
1616	https://doi.org/10.1002/jpln.201000023
1617	Teodoro GS, Lambers H, Nascimento DL, et al (2019) Specialized roots of Velloziaceae weather
1618	quartzite rock while mobilizing phosphorus using carboxylates. Funct Ecol 33:762-773.
1619	https://doi.org/10.1111/1365-2435.13324
1620	Tubana BS, Babu T, Datnoff LE (2016) A Review of Silicon in Soils and Plants and Its Role in
1621	US Agriculture. Soil Sci 181:1. https://doi.org/10.1097/SS.000000000000179
1622	Uroz S, Calvaruso C, Turpault MP, Frey-Klett P (2009) Mineral weathering by bacteria: ecology,

1623	actors and mechanisms. Trends Microbiol 17:378–387.
1624	https://doi.org/10.1016/j.tim.2009.05.004
1625	Van Breemen N, Finlay R, Lundström U, et al (2000) Mycorrhizal weathering: A true case of
1626	mineral plant nutrition? Biogeochemistry 49:53-67.
1627	https://doi.org/10.1023/A:1006256231670
1628	Van Der Heijden MGA, Bardgett RD, Van Straalen NM (2008) The unseen majority: Soil
1629	microbes as drivers of plant diversity and productivity in terrestrial ecosystems. Ecol Lett
1630	11:296–310. https://doi.org/10.1111/j.1461-0248.2007.01139.x
1631	Van Der Heijden MGA, Klironomos JN, Ursic M, et al (1998) Mycorrhizal fungal diversity
1632	determines plant biodiversity, ecosystem variability and productivity. Nature 396:69-72
1633	van Hees PAW, Lundström US, Mörth CM (2002) Dissolution of microcline and labradorite in a
1634	forest O horizon extract: The effect of naturally occurring organic acids. Chem Geol
1635	189:199–211. https://doi.org/10.1016/S0009-2541(02)00141-9
1636	van Hees PAW, Rosling A, Lundström US, Finlay RD (2006) The biogeochemical impact of
1637	ectomycorrhizal conifers on major soil elements (Al, Fe, K and Si). Geoderma 136:364-377
1638	https://doi.org/10.1016/j.geoderma.2006.04.001
1639	van Schöll L, Hoffland E, Van Breemen N (2006) Organic anion exudation by ectomycorrhizal
1640	fungi and Pinus sylvestris in response to nutrient deficiencies. New Phytol 170:153-163.
1641	https://doi.org/10.1111/j.1469-8137.2006.01649.x
1642	van Schöll L, Kuyper TW, Smits MM, et al (2008) Rock-eating mycorrhizas: their role in plant

1643	nutrition and biogeochemical cycles. Plant Soil 303:35-47. https://doi.org/10.1007/s11104-
1644	007-9513-0
1645	Vandenkoornhuyse P, Quaiser A, Duhamel M, et al (2015) The importance of the microbiome of
1646	the plant holobiont. New Phytol 206:1196–1206. https://doi.org/10.1111/nph.13312
1647	Vander Linden C, Delvaux B (2019) The weathering stage of tropical soils affects the soil-plant
1648	cycle of silicon, but depending on land use. Geoderma 351:209-220.
1649	https://doi.org/10.1016/j.geoderma.2019.05.033
1650	Vandevenne FI, Barão AL, Schoelynck J, et al (2013) Grazers: Biocatalysts of terrestrial silica
1651	cycling. Proc R Soc B Biol Sci 280:1–9. https://doi.org/10.1098/rspb.2013.2083
1652	Vandevenne FI, Barão L, Ronchi B, et al (2015) Silicon pools in human impacted soils of
1653	temperate zones. Global Biogeochem Cycles 29:1439–1450.
1654	https://doi.org/10.1002/2014GB005049.Received
1655	Veldhuis MP, Gommers MI, Olff H, Berg MP (2018) Spatial redistribution of nutrients by large
1656	herbivores and dung beetles in a savanna ecosystem. J Ecol 106:422-433.
1657	https://doi.org/10.1111/1365-2745.12874
1658	Vernavá MN, Phillips-Aalten PM, Hughes LA, et al (2004) Influences of preceding cover crops
1659	on slug damage and biological control using Phasmarhabditis hermaphrodita. Ann Appl
1660	Biol 145:279–284. https://doi.org/10.1111/j.1744-7348.2004.tb00384.x
1661	Vessey JK (2003) Plant growth promoting rhizobacteria as biofertilizers. Plant Soil 255:571–586
1662	https://doi.org/10.1023/A:1026037216893

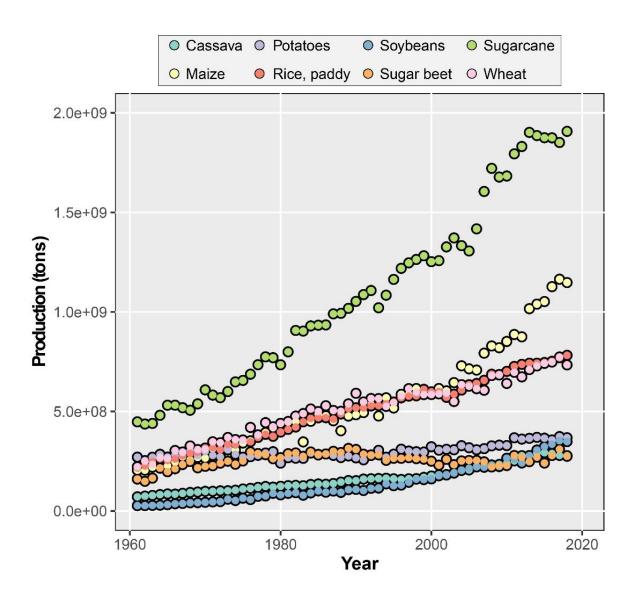
1663	Walder F, Niemann H, Natarajan M, et al (2012) Mycorrhizal networks: Common goods of
1664	plants shared under unequal terms of trade. Plant Physiol 159:789-797.
1665	https://doi.org/10.1104/pp.112.195727
1666	Wang HY, Shen QH, Zhou JM, et al (2011) Plants use alternative strategies to utilize
1667	nonexchangeable potassium in minerals. Plant Soil 343:209–220.
1668	https://doi.org/10.1007/s11104-011-0726-x
1669	Wang M, Wang JJ, Tafti ND, et al (2019a) Effect of alkali-enhanced biochar on silicon uptake
1670	and suppression of gray leaf spot development in perennial ryegrass. Crop Prot 119:9-16.
1671	https://doi.org/10.1016/j.cropro.2019.01.013
1672	Wang RR, Wang Q, He LY, et al (2015) Isolation and the interaction between a mineral-
1673	weathering Rhizobium tropici Q34 and silicate minerals. World J Microbiol Biotechnol
1674	31:747–753. https://doi.org/10.1007/s11274-015-1827-0
1675	Wang Y, Xiao X, Xu Y, Chen B (2019b) Environmental Effects of Silicon within Biochar
1676	(Sichar) and Carbon-Silicon Coupling Mechanisms: A Critical Review. Environ Sci Technol
1677	53:13570–13582. https://doi.org/10.1021/acs.est.9b03607
1678	Wang Y, Zhang K, Lu L, et al (2020) Novel insights into effects of silicon-rich biochar (Sichar)
1679	amendment on cadmium uptake, translocation and accumulation in rice plants. Environ
1680	Pollut 265:114772. https://doi.org/10.1016/j.envpol.2020.114772
1681	Watanabe T, Luu HM, Inubushi K (2017) Effects of the continuous application of rice straw
1682	compost and chemical fertilizer on soil carbon and available silicon under a double rice
1683	cropping system in the Mekong Delta, Vietnam. Japan Agric Res Q 51:233–239.

1684	https://doi.org/10.6090/jarq.51.233
1685	Wedepohl KH (1995) The composition of the continental crust. Geochim Cosmochim Acta
1686	59:1217–1232. https://doi.org/10.1016/0016-7037(95)00038-2
1687	Welch SA, Ullman WJ (1999) The effect of microbial glucose metabolism on bytownite feldspan
1688	dissolution rates between 5° and 35°C. Geochim Cosmochim Acta 63:3247–3259.
1689	https://doi.org/10.1016/S0016-7037(99)00248-3
1690	Wickramasinghe DB, Rowell DL (2006) The release of silicon from amorphous silica and rice
1691	straw in Sri Lankan soils. Biol Fertil Soils 42:231–240. https://doi.org/10.1007/s00374-005-
1692	0020-2
1693	WRB (2015) World reference base for soil resources 2014 International soil classification system
1694	for naming soils and creating legends for soil maps. FAO, Rome, Italy
1695	Wright AL, Hons FM (2005) Soil Carbon and Nitrogen Storage in Aggregates from Different
1696	Tillage and Crop Regimes. Soil Sci Soc Am J 69:141–147.
1697	https://doi.org/10.2136/sssaj2005.0141a
1698	Wu L, Jacobson AD, Hausner M (2008) Characterization of elemental release during microbe-
1699	granite interactions at $T=28^{\circ}\text{C}$ . Geochim Cosmochim Acta 72:1076–1095.
1700	https://doi.org/10.1016/j.gca.2007.11.025
1701	Wu W, Limmer MA, Seyfferth AL (2020) Quantitative assessment of plant- available silicon
1702	extraction methods in rice paddy soils under different management. Soil Sci Soc Am J
1703	84:618–626 https://doi.org/10.1002/sai2.20013

1/04	Xiao W, Yuqiao L, Qiang Z, et al (2016) Efficacy of Si fertilization to modulate the heavy metal
1705	absorption by barley (Hordeum vulgare L.) and pea (Pisum sativum L.). Environ Sci Pollut
1706	Res 23:20402–20407. https://doi.org/10.1007/s11356-016-7182-3
1707	Xiao X, Chen B, Zhu L (2014) Transformation, morphology, and dissolution of silicon and
1708	carbon in rice straw-derived biochars under different pyrolytic temperatures. Environ Sci
1709	Technol 48:3411–3419. https://doi.org/10.1021/es405676h
1710	Xu D, Gao T, Fang X, et al (2020) Silicon addition improves plant productivity and soil nutrient
1711	availability without changing the grass:legume ratio response to N fertilization. Sci Rep
1712	10:1–9. https://doi.org/10.1038/s41598-020-67333-7
1713	Xue Y, Xia H, Christie P, et al (2016) Crop acquisition of phosphorus, iron and zinc from soil in
1714	cereal/legume intercropping systems: a critical review. Ann Bot 117:363-377.
1715	https://doi.org/10.1093/aob/mcv182
1716	Yang X, Song Z, Qin Z, et al (2020) Phytolith-rich straw application and groundwater table
1717	management over 36 years affect the soil-plant silicon cycle of a paddy field. Plant Soil
1718	454:343–358. https://doi.org/10.1007/s11104-020-04656-4
1719	Yost RS, Fox RL (1982) Influence of mycorrhizae on the mineral contents of cowpea and
1720	soybean grown in an Oxisol. Agron J 74:475–481
1721	Zabowski D, Ugolini FC (1990) Lysimeter and centrifuge soil solutions: Seasonal differences
1722	between methods. Soil Sci Soc Am J 54:1130–1135
1723	Zaharescu DG, Burghelea CI, Dontsova K, et al. (2019) Ecosystem-bedrock interaction changes

1/24	nutrient compartmentalization during early oxidative weathering. Sci Rep 9:15006.
1725	https://doi.org/10.1038/s41598-019-51274-x
1726	Zahra MK, Monib M, Abdel-Al SI, Heggo A (1984) Significance of Soil Inoculation with
1727	Silicate Bacteria. Zentralbl Mikrobiol 139:349–357. https://doi.org/10.1016/s0232-
1728	4393(84)80013-x
1729	Zemunik G, Turner BL, Lambers H, Laliberté E (2015) Diversity of plant nutrient-acquisition
1730	strategies increases during long-term ecosystem development. Nat Plants 1:1-4.
1731	https://doi.org/10.1038/nplants.2015.50
1732	Zhu Y, Gong H (2014) Beneficial effects of silicon on salt and drought tolerance in plants. Agron
1733	Sustain Dev 34:455–472. https://doi.org/10.1007/s13593-013-0194-1
1734	Zuo Y, Zhang F, Li X, Cao Y (2000) Studies on the improvement in iron nutrition of peanut by
1735	intercropping with maize on a calcareous soil. Plant Soil 220:13-25.
1736	https://doi.org/10.1023/a:1004724219988
1737	
1738	
1739	
1740	
1741	
1742	

**Figures** 



**Fig. 1** – Global production of the eight most important crops in 2018, from 1961 to 2018 (source = FAOSTAT).

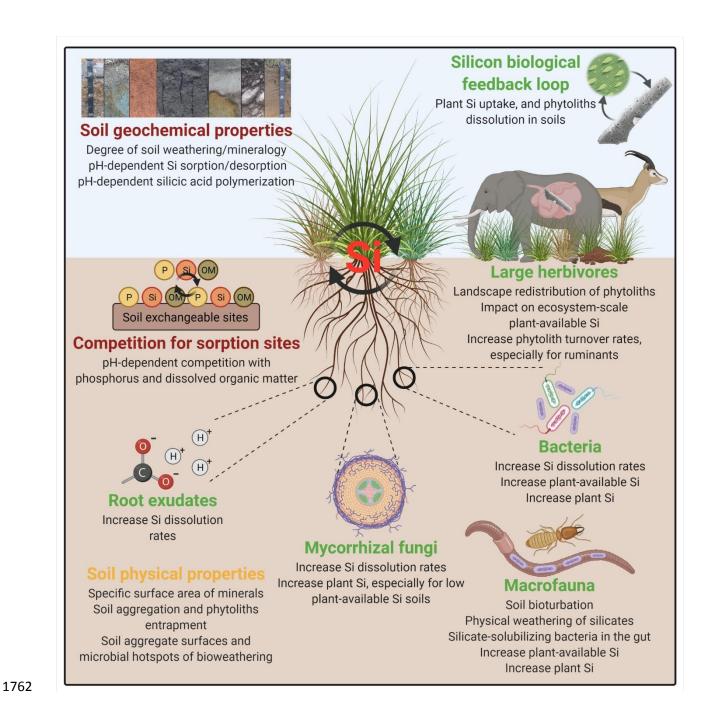
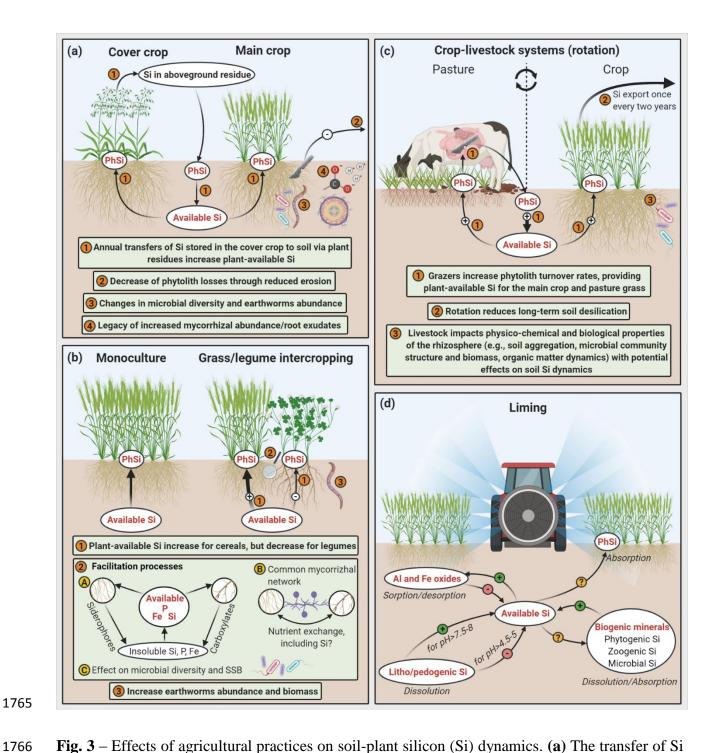


Fig. 2 – Biotic and abiotic factors influencing soil-plant Si dynamics. This figure was created
 using BioRender.



**Fig. 3** – Effects of agricultural practices on soil-plant silicon (Si) dynamics. (a) The transfer of Si stored in the cover crop to soil via plant residues may substantially increase the soil phytogenic Si pool (PhSi), thus contributing to increase plant-available Si concentrations (available Si) (# 1).

Cover crops could also lower phytolith losses though reduced erosion (Adetunji et al. 2020) (#2), modify microbial diversity and earthworm abundance (Roarty et al. 2017; Euteneuer et al. 2020) (# 3), and provide a legacy of increased mycorrhizal abundance/root exudates (Hallama et al. 2019) (# 4), thereby impacting soil-plant Si dynamics. (b) Cereal-legume intercropping may increase plant-available Si concentrations for the cereal crops, but decrease it for the legume crops (Ning et al. 2017) (# 1). Numerous facilitation processes with beneficial effects on soilplant Si mobility may also occur (# 2): taking advantage of different nutrient-acquisition strategies (Li et al. 2014; Xue et al. 2016), sharing nutrients via common mycorrhizal network (Walder et al. 2012), or modifying microbial diversity (Brooker et al. 2015), including silicatesolubilizing bacteria (SSB). Cereal-legume intercropping may also increase plant-available Si concentrations and plant Si uptake through enhanced earthworm abundance and biomass (Schmidt et al. 2001, 2003) (# 3). (c) Integrating crops and livestock will increase Si release from phytoliths in animal dung (Vandevenne et al. 2013), which will provide Si for the main crop and pasture grass (# 1). Pasture/crop rotation could also reduce long-term soil desilication by reducing Si export from harvest (once every two year) (# 2). Crop-livestock systems modify physical, chemical and biological properties of the rhizosphere (Brewer and Gaudin 2020), that could, in turn, influence soil-plant Si dynamics (# 3). (d) Increase in soil pH after liming increases Si adsorption on soil colloids, which, in turn, reduces Si concentrations in the soil solution. Raising pH above 4.5-5.0 decreases aluminosilicate dissolution rates, while raising pH above 7.5-8.0 increases those rates (Drever 1994; Kelly et al. 1998; Haynes 2019). Raising pH in the range 3.0-9.0 increases phytolith dissolution rates (Fraysse et al. 2009). The pH/liming effects

1769

1770

1771

1772

1773

1774

1775

1776

1777

1778

1779

1780

1781

1782

1783

1784

1785

1786

1787

1788

- on Si absorption by soil living organisms and plants is unknown. This figure was created using
- 1791 BioRender.