



**HAL**  
open science

# Topology and Dynamics of Transcriptome (Dys)Regulation

Michel Planat, David Chester

► **To cite this version:**

Michel Planat, David Chester. Topology and Dynamics of Transcriptome (Dys)Regulation. International Journal of Molecular Sciences, 2024, 25 (9), pp.4971. 10.3390/ijms25094971 . hal-04693035

**HAL Id: hal-04693035**

**<https://hal.science/hal-04693035>**

Submitted on 10 Sep 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Article

Not peer-reviewed version

---

# Topology and Dynamics of Transcriptome (Dys)regulation

---

[Michel Planat](#)\* and [David Chester](#)

Posted Date: 28 March 2024

doi: 10.20944/preprints202403.1741.v1

Keywords: group theory; character variety; Painlevé equations; transcriptome; microRNAs; diseases; cancer research



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Article

# Topology and Dynamics of Transcriptome (Dys)Regulation

Michel Planat <sup>1,\*</sup>  and David Chester <sup>2</sup> 

<sup>1</sup> Université de Franche-Comté, Institut FEMTO-ST CNRS UMR 6174, 15 B Avenue des Montboucons, F-25044 Besançon, France

<sup>2</sup> Quantum Gravity Research, Los Angeles, CA 90290, USA; davidc@quantumgravityresearch.org

\* Correspondence: michel.planat@femto-st.fr

**Abstract:** RNA transcripts play a crucial role as witnesses of gene expression health. Identifying disruptive short sequences in RNA transcription and regulation is essential for potentially treating diseases. Let's delve into the mathematical intricacies of these sequences. We've previously devised a mathematical approach for defining a "healthy" sequence (10.14218/GE.2023.00079). This sequence is characterized by having at most four distinct nucleotides (denoted as  $nt \leq 4$ ). It serves as the generator of a group denoted as  $f_p$ . The desired properties of this sequence are as follows:  $f_p$  should be close to a free group of rank  $nt - 1$ , it must be aperiodic,  $f_p$  should not have isolated singularities within its  $SL_2(\mathbb{C})$  character variety (specifically within the corresponding Groebner basis). Now, let's explore the concept of singularities. There are cubic surfaces associated with the character variety of a four-punctured sphere denoted as  $S_2^4$ . When we encounter these singularities, we find ourselves dealing with some algebraic solutions of a dynamical second-order differential (and transcendental) equation known as the Painlevé VI equation (10.3390/dynamics4010001). In certain cases,  $S_2^4$  degenerates, in the sense that two punctures collapse, resulting in a "wild" dynamics governed by the Painlevé equations of an index lower than VI. In our paper, we provide examples of these fascinating mathematical structures within the context of miRNAs. Specifically, we find a clear relationship between decorated character varieties of Painlevé equations and the character variety calculated from the seed of oncomirs. These findings should find applications in cancer research.

**Keywords:** group theory; character variety; Painlevé equations; transcriptome; microRNAs; diseases; cancer research

## 1. Introduction

Self-regulation of an animal organism, also-called homeostasis, depends on several factors. At a macroscopic scale, pH, temperature, the amount of oxygen and carbon dioxide in the blood and so on in living organisms self-adjust to fight against threats present in the environment and keep a stable physiology.

In mammals, the main organs involved in homeostasis are the hypothalamus that controls body temperature and circadian cycles, the pituitary gland that rates the secreting hormones, the lungs that regulate the balance between blood oxygen and the carbon dioxide, the skin that protects against outer pathogens, the muscles in the circulatory, gastrointestinal and blood systems, the kidneys that regulate water and salts in the blood and the amount of urine thanks to hormones, the liver which acts as a factory for bile, glucose, hemoglobin, ammonia and toxins, the pancreas that produces the hormones and enzymes for digestion and finally the brain that controls behavior whether conscious or unconscious. Such macroscopic features of homeostasis are well summarized by Claude Bernard in his 19th-century lectures [1].

A genome-scale homeostasis exists as well. Gene expression (GE) is the process by which information from a gene is used to synthesize end products such as proteins or non-coding RNA. GE is the most fundamental level at which the genotype gives rise to observable traits called the phenotype. At all steps, GE is regulated by transcription, translation and post-transcriptional changes in the production of proteins. Such microscopic homeostasis for creating messenger mRNA is a

complex machinery that makes use of non-coding genes transcribed as precursors that undergo further processing [2].

Rules for genome scale homeostasis similar to the constancy rules of macroscopic homeostasis are not yet firmly established. We mention previous work in this direction [3–5]. In our previous publications we started to unveil some of these rules by using a group theoretical approach. Since the genome is digital with its four nucleotides A, U/T, G and C as letters, there are important short sequences as the consensus sequence of a transcription factor, a TATA box, a polyadenylation signal or a microRNA (miRNA) whose structure should obey group theory and associated character variety rules. Otherwise, a disease is in sight [6–8]. Chemical modifications in the environment also controls the metabolism of GE. For instance, RNA methylation with  $N^6$ -methymadenosine ( $m^6A$ ) is documented in our recent paper [8].

For studying the aforementioned key sequences, we employ infinite (finitely generated) groups denoted by  $f_p$ , and their representations over the matrix group  $SL_2(\mathbb{C})$ , where the entries of matrices are complex numbers [6]. The importance of this group extends across all fields of physics as it represents a space-time-spin group.

Our crucial observation is that an  $f_p$  group associated with a “healthy” sequence usually approximates a free group  $F_r$ , where the rank  $r$  equals the number of distinct nucleotides minus one. A sequence deviating from this may suggest a potential dysregulation leading to a disease. However, an  $f_p$  group closely resembling a free group does not provide sufficient assurance against a disease. Additional examination of the  $SL_2(\mathbb{C})$  representations of  $f_p$  termed the character variety, specifically its Groebner basis  $\mathcal{G}$ , is necessary. The Groebner basis comprises a set of surfaces. A surface within  $\mathcal{G}$  containing isolated singularities indicates a potential disease that can be identified specifically, e.g., relating to an oncogene or a neurological disorder [6] [Figure 6, Tables 2 to 4].

An additional attribute of “healthy” sequences, which leads to a group  $f_p$  approximating the free group  $F_r$  and not mentioned in [6], is their connection to aperiodicity. Schrödinger’s book [9] proposes aperiodicity of living “crystals”. Our papers [7,8] characterize some aperiodic DNA/RNA sequences. But in this paper we do not focus on this aspect of genome scale homeostasis.

Our new goal in the current paper is to feature the dynamical equations sustaining some of the algebraic surfaces contained in the Groebner basis attached to the group  $f_p$  of the considered short sequence. This task was started in [10] using the list of algebraic solutions of Painlevé VI (PVI) equation [11]. We made use of the PVI equation and the cubic algebraic surfaces contained in its character variety in the context of miRNAs. We continue along these lines by looking at cubic surfaces contained in the character variety of Painlevé equations of lower index such as PV and PIV. As explained below, this is a crucial step since the topology of the character variety of PV and PIV is wilder than that of PVI, the singularities of the corresponding surfaces are more irregular than those with PVI [12,13]. The occurrence of surfaces related to PV and PIV may signify a more severe disease than a surface related to PVI.

In Section 2, we investigate the topology of Painlevé equations and their related character variety. We also remind readers how the character variety of a group  $f_p$  may contain simply singular surfaces of the Painlevé type. In Section 3, we apply these techniques to miRNAs recognized as oncomirs or tumor suppressors. Our results are in accordance with our aforementioned view of health versus disease. Section 4 summarizes our approach.

## 2. Materials and Methods

For an introduction to Painlevé equations, readers are encouraged to explore our recent paper [10] and the references therein. Painlevé equations represent nonlinear ordinary differential equations whose solutions are considered transcendental due to their inability to be expressed in terms of familiar special functions, such as elliptic, hyperelliptic, or hypergeometric functions [14]. The hallmark of Painlevé transcendents lies in the Painlevé property, indicating that the only movable singularities are poles [15]. In recent years, there has been a shift in focus towards uncovering explicit algebraic

solutions of PVI, making it a significant subject in 20th-century mathematical research with connections to algebraic topology, algebraic geometry, and representation theory [16,17]. Painlevé equations of lower index than VI exhibit a more complex topological structure, which is elaborated upon below. The topological structure and the associated  $SL_2(\mathbb{C})$  representations of PVI, PV, and PIV are further elucidated. These features prove to be invaluable for understanding diseases at the transcriptome scale.

### 2.1. Painlevé Equations and THEIR associated $SL_2(\mathbb{C})$ Character Variety

At a first step, the concept of a flat connection on a fiber bundle  $M \rightarrow B$  is fully relevant, where the base  $B$  assumes the form of a three-punctured sphere, denoted as  $B = S_2^{(3)} = \mathbb{P}^1 \setminus \{0, 1, \infty\}$ . Over a point  $t \in B$ , a corresponding four-punctured sphere  $P_t = S_2^{(4)} = \mathbb{P}^1 \setminus \{0, 1, t, \infty\}$  exists. Let  $M_t$  denote the fiber of  $M$  over the base point  $t \in B$ , the monodromy action unfolds through the action of the fundamental group of the base on the fiber. This defines a homomorphism  $\pi_1(B) \rightarrow \text{Aut}(M_t)$  [10,16].

For the setting of the monodromy problem for PVI, the reader may consult [18] [Section 1] where complex constants  $\theta_0, \theta_1, \theta_t$  and  $\theta_\infty$  arise from the eigenvalues of the matrices in the associated Schlesinger's system.

For each  $t \in B$ , the space of conjugacy classes of  $SL_2(\mathbb{C})$  representations for the fundamental group  $\pi_1(P_t) \cong F_3$  is the character variety

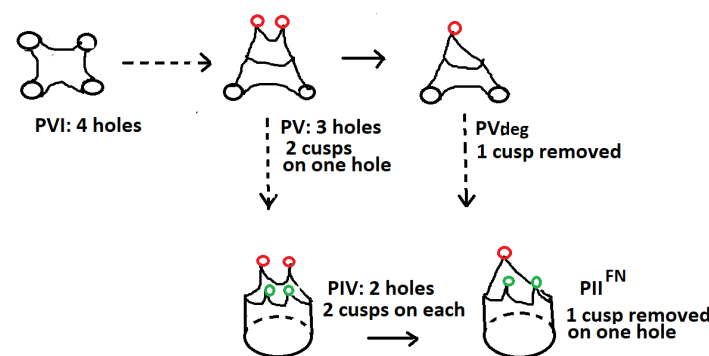
$$\mathcal{C}_t = \text{Hom}(\pi_1(P_t), G)/G, \text{ with } G = SL_2(\mathbb{C}).$$

The connection is flat and described by  $P_{VI}$  equation as follows:

$$y_{tt} = \frac{1}{2} \left( \frac{1}{y} + \frac{1}{y-1} + \frac{1}{y-t} \right) y_t^2 - \left( \frac{1}{t} + \frac{1}{t-1} + \frac{1}{y-t} \right) y_t + \frac{y(y-1)(y-t)}{2t^2(t-1)^2} \left[ \alpha + \beta \frac{t}{y^2} + \gamma \frac{t-1}{(y-1)^2} + \delta \frac{t(t-1)}{(y-t)^2} \right] \quad (PVI) \quad (1)$$

with  $y_t = \frac{dy}{dt}$  and parameters  $\alpha = (\theta_\infty - 1)^2$ ,  $\beta = -\theta_0^2$ ,  $\gamma = \theta_1^2$ ,  $\delta = 1 - \theta_t^2$ .

Further topological steps are illustrated in Figure 1 [19] [Figure 3]. The corresponding Painlevé equations are in [18] as follows



**Figure 1.** A partial view of singularities of Riemann surfaces from the Painlevé perspective [19] [Figure 3]. The dashed arrows correspond to the creation of cusps (red or green bullets) on some holes (black bullets). The full arrows point out a cusp removal. Only Painlevé topologies expected in the context of our model of gene expression are considered. The paths in the diagram correspond to the move  $PVI \rightarrow PV \rightarrow PIV \rightarrow PII^{FN}$  or the move  $PVI \rightarrow PV \rightarrow PV_{deg} \rightarrow PII^{FN}$ .

$$y_{tt} = \left(\frac{1}{2y} + \frac{1}{y-1}\right)y_t^2 - \frac{1}{t}y_t + \frac{(y-1)^2}{t^2}\left(\alpha y + \frac{\beta}{y}\right) + \frac{\gamma y}{t} + \frac{\delta y(y+1)}{y-1} \quad (PV) \quad (2)$$

with parameters  $\alpha = \frac{1}{8}(\theta_0 - \theta_1 + \theta_\infty)^2$ ,  $\beta = -\frac{1}{8}(\theta_0 - \theta_1 - \theta_\infty)^2$ ,  $\gamma = 1 - \theta_0 - \theta_1$ ,  $\delta = -\frac{1}{2}$ .

$$y_{tt} = \frac{1}{2y}y_t^2 + \frac{3}{2}y^3 + 4ty^2 + 2(t^2 - \alpha)y + \frac{\beta}{y} \quad (PIV) \quad (3)$$

with parameters  $\alpha = 2\theta_\infty - 1$ ,  $\beta = -8\theta_0^2$ .

$$y_{tt} = 2y^3 + ty + \alpha \quad (PII) \quad (4)$$

with parameter  $\alpha = \frac{1}{2} - \theta_0$ .

### The Painlevé-Jimbo-Fricke Cubics Attached to Painlevé Equations

In the following, we introduce the boundary traces  $a, b, c, d$  of matrices representing simple loops around the punctures of the corresponding Riemann surface shown in Figure 1. In papers [19,20], it is shown that the  $SL_2(\mathbb{C})$  character variety takes the form of Painlevé-Jimbo-Fricke cubics as follows.

$$\begin{aligned} xyz + x^2 + y^2 + z^2 - \omega_1 x - \omega_2 y - \omega_3 z - \omega_4 &= 0, \\ \omega_1 = ad + bc, \omega_2 = ac + bd, \omega_3 = ab + cd, \omega_4 = a^2 + b^2 + c^2 + d^2 + abcd - 4, \end{aligned} \quad (type PVI),$$

$$\begin{aligned} xyz + x^2 + y^2 - \omega_1 x - \omega_2 y - \omega_3 z - \omega_4 &= 0, \\ \text{with } \omega_4 = -1 - \omega_3^2 - \frac{\omega_3(-\omega_2 + \omega_1\omega_3)(-\omega_1 + \omega_2\omega_3)}{(\omega_3^2 - 1)^2}, \\ \omega_1 = a + bc, \omega_2 = b + ac, \omega_3 = c, \omega_4 = -(abc + c^2 + 1) \end{aligned} \quad (type PV),$$

$$\begin{aligned} xyz + x^2 + y^2 - \omega_1 x - \omega_2 y + 1 &= 0, \\ \omega_1 = a + bc, \omega_2 = b, \end{aligned} \quad (type PV_{deg}),$$

$$\begin{aligned} xyz + x^2 - \omega_1 x - \omega_2(y+z) - \omega_2(-\omega_1 + \omega_2) &= 0, \\ \omega_1 = ab + b^2, \omega_2 = b^2, \end{aligned} \quad (type PIV)$$

$$\begin{aligned} xyz + x^2 - \omega_1 x - y + 1 &= 0. \\ \omega_1 = a, \end{aligned} \quad (type PII^{FN})$$

(5)

The Fricke cubics refers to the case of PVI character variety [21].

### 2.2. The Character Variety Associated to a DNA/RNA Sequence

We can consider the representations of an  $f_p$  group over the space-time-spin group  $SL_2(\mathbb{C})$ , as we did in [6,7].



Representations of  $f_p$  in  $SL_2(\mathbb{C})$  are homomorphisms  $\rho : f_p \rightarrow SL_2(\mathbb{C})$  with character  $\kappa_\rho(g) = \text{tr}(\rho(g))$ ,  $g \in f_p$ . The notation  $\text{tr}(\rho(g))$  signifies the trace of the matrix  $\rho(g)$ . The set of characters is employed to determine an algebraic set by taking the quotient of the set of representations  $\rho$  by the group  $SL_2(\mathbb{C})$ , which acts by conjugation on representations [21,22].

In our paper [6] [Section 2.2], we elaborated that the character variety of  $f_p$  is a set comprised of a sequence  $X$  of multivariate polynomials. A particular basis related to  $X$  is the Groebner basis  $\mathcal{G}(X)$ , whose factors define hypersurfaces.

For a two-generator group  $f_p$ , the factors are three-dimensional surfaces. In general, these surfaces can be classified by mapping them to a rational surface across five categories [6] [Section 3]. Often encountered surfaces are degree  $p$  Del Pezzo surfaces where  $1 \leq p \leq 9$ . A rational surface may either be non-singular, 'almost non-singular', having only isolated singularities, or singular. Almost non-singular surfaces are crucial in our context. A simple singularity is referred to as an A-D-E singularity and must be of the type  $A_n$ ,  $n \geq 1$ ,  $D_n$ ,  $n \geq 4$ ,  $E_6$ ,  $E_7$  or  $E_8$ .

The A-D-E type is mirrored in the notation we employ. For instance,  $S^{(lA_1, mA_2, nA_3, \dots)}$  denotes a surface containing  $l$  type  $A_1$ ,  $m$  type  $A_2$ ,  $n$  type  $A_3$  singularities, etc. A generic surface is the Cayley cubic we encountered in our previous papers, defined as  $S^{(4A_1)} = xyz + x^2 + y^2 + z^2 - 4$  [6] [Figure 5].

For a three-generator group  $f_p$ , the factors of  $\mathcal{G}(X)$  are seven-dimensional surfaces of the form  $S_{a,b,c,d}(x,y,z)$ . Some of them belong to the Fricke family [6] [Equation (3)], which is associated with the four-punctured sphere. But for a chosen set of parameters  $a, b, c, d$ , the hypersurface reduces to an ordinary three-dimensional surface.

For a four-generator group  $f_p$ , the factors of  $\mathcal{G}(X)$  are 14-dimensional surfaces generally containing 4 copies of the form  $S(x,y,z)$ ,  $S(x,u,v)$ ,  $S(y,u,v)$  and  $S(z,v,w)$  for selected choices of 8 parameters.

### 3. Results and Discussion

The theory developed in the previous section is applied to sequences determined by the seed of a miRNA. Animal miRNAs are able to recognize their target mRNAs by using as few as 6 to 8 nucleotides (the seed region) at the 5' end of the miRNA (for a -5p strand) or at the 3' end of the miRNA (for a -3p strand). A given miRNA may have hundreds of different mRNA targets, and a given target might be regulated by multiple miRNAs. Thus it is not surprising that a simple correlation between a miRNA and a disease cannot be found. In the following, the selected miRNAs occur in the context of cancer research [23]. We mention that there are repositories where useful links between diseases and non coding RNAs such as miRNAs are listed [24,25] [Table 1]. The seeds used for our calculations of the group  $f_p$  are taken from [26,27].

Our results are summarized in Table 1 for miRNAs generally considered as oncomirs and Table 2 for miRNAs generally considered as tumor suppressors. Of course the boundary between the two types is not sharp and depends on the type of disease under examination. We remind readers from the introduction that dysregulation may occur when either the  $f_p$  group generated by the seed sequence is away from the free group  $F_1$  (for a seed with two distinct nucleotides), away from the free group  $F_2$  (for a seed with three distinct nucleotides) or away from the free group  $F_3$  (for a seed with three distinct nucleotides). The number of conjugacy classes of subgroups of a given index for the group  $f_p$  is abbreviated as 'card seq'. The first step is to check if the card seq is close to that of a free group  $F_r$ ,  $r = 1, 2$  or  $3$  or not. In the later case, the sequence may predict a risk of disease.

The second step is to check the structure of the character variety associated to  $f_p$ , more precisely the structure of its Groebner basis  $\mathcal{G}$ . We focus on cubic surfaces belonging to  $\mathcal{G}$  and, if they contain an isolated singularity, we determine the A-D-E type (abbreviated as ADE type). From the introduction we claim that dysregulation may occur if such an isolated singularity exists in a cubic surface in  $\mathcal{G}$ . Finally, we can check if the simply singular surface belongs to the Painlevé class (PVI, PV or PIV). E.g. in Table 1, the notation  $f^{(4A_1)}$ : PVI means that the Cayley cubic of ADE type  $4A_1$  is a solution of equation  $xyz + x^2 + y^2 + z^2 - 4 = 0$  associated to the character variety of PVI whose topology is the

4-punctured sphere (as in Figure 1). In Table 1, the notation  $f^{(A_2)}$ : PV for the surface  $xyz + x^2 + y^2 + 1$  means that the ADE type is  $A_2$  and that the surface is associated to PV equation.

All oncomirs in Table 1 have an  $f_p$  such that the corresponding  $\mathcal{G}$  contains simply singular cubic surfaces whatever the card seq of  $f_p$  is that of a free group or not. Thus a) correlation between our approach and the risk of the disease is well established.

In Table 2, only five miRNAs generally considered as tumor suppressors have an  $f_p$  either away from the free group  $F_2$  or with a  $\mathcal{G}$  containing simply singular cubic surfaces. The remaining miRNAs have an  $f_p$  like the free group  $F_2$  and show no simply singular surface in  $\mathcal{G}$ . It is well known that a microRNA may act as an oncogen for a given type of cancer or as a tumor suppressor for another type. E.g. let-7a-5p is a regulator for tumors in many cancers but may also unfavorable in lung cancer.

We do not investigate miRNA seeds comprising 4 distinct nucleotides. It is straightforward to check if the corresponding  $f_p$  group is close to the free group  $F_3$  or not and if the seed sequence is aperiodic or not. But the Groebner basis is more difficult to obtain and its structure does not allow us to differentiate between 'healthy' and 'disruptive'. Further progress is necessary [10] [Section 5.1].

Details and references for each considered miRNA are given in the subsections below. Most cubics appearing in the Groebner basis attached to the  $f_p$  group of the seed are degree 3 Del Pezzo surfaces (denoted as DP3).

**Table 1.** A partial list of miRNAs generally considered as oncomirs.

ine oncomir	seed	card seq	sing type	Cancer type
ine miR-21-3p	aacacca	not $F_1$	$f^{(A_3)}, f^{(A_2)}$	ubiquitous
miR-204-5p	ucccuuu	not $F_1$	$f^{(A_3)}, f^{(A_2)}$	prostate
miR-126-5p	auuauua	$H_3$	$f^{(A_2)}$	liver
miR-1908-5p	ggcgggg	$F_1$	$f^{(4A_1)}$ : PVI	ubiquitous
miR-1908-3p	cggccgc	$H_3$	$f^{(A_2)}, f^{(A_1A_2)}$	breast
ine miR-155-3p	uccuaca	not $F_2$	$f^{(A_2)}, f^{(A_1A_2)}$	ubiquitous
miR-9-5p	cuuuggu	$F_2$	$f^{(A_2)}$ : PV <sup>deg</sup> , PVI	metastasis
miR-146b-3p	cccugug	not $F_2$	$f^{(A_2)}, f^{(A_1A_1A_2)}$ : PV	pancreatic
miR-19a-5p	guuuugc	$F_2$	$f^{(A_2A_2)}, f^{(A_1A_1)}, PVI_a$	pancreatic
miR-181a-5p	acaucaa	$F_2$	$f^{(A_2)}, f^{(A_4)}$	ubiquitous
miR-15-5p	agcagca	$F_2$	$\{f^{(A_2)}, f^{(A_1A_2)}\}$ : PV	prostate
miR-569	guuaaug	not $F_2$	$f^{(A_2)}, f^{(A_4)}$	lung, breast
miR-133b	uuggucc	not $F_2$	$f^{(A_2)}, f^{(A_1)}, f^{(A_2A_2)}$	ubiquitous
miR-1270	uggagau	$F_2$	$f^{(A_2)}, f^{(A_2A_2)}$	osteosarcoma
ine				



**Table 2.** A partial list of miRNAs generally considered as tumor suppressors. According to our view, as well as the references given in the text, the first five miRNAs behave more likely as oncomirs.

ine tumor suppressor	seed	card seq	sing type	Cancer type
ine miR-let-7a-5p	gagguag	$F_2$	$f^{(A_2A_2)}, f^{(A_1A_4)}$	ubiquitous
miR-let-7a-3p	uauaca	$F_2$	no cubic	ubiquitous
miR-34b-3p	aaucacu	not $F_2$	$f^{(A_2)}, f^{(A_3)}, f^{(A_4)}$	lung, breast
miR-200a-5p	aucuuac	not $F_2$	$f^{(A_1A_3)}, f^{(A_1A_1A_3)}$	skin, colorectal
miR-22-5p	guucuuc	$F_2$	$f^{(D_4)}$	nsc lung
miR-214-5p	gccugu	$F_2$	$f^{(A_2)}$	ubiquitous
.	gccuguc	$F_2$	no cubic	.
miR-214-3p	cagcag	$F_2$	$f^{(A_1A_4)}$	.
.	cagcagg	$F_2$	no cubic	.
ine miR-503-5p	agcagcg	$F_2$	rational scroll	colon
miR-141-5p	aucuucc	$F_2$	no cubic	blood leukemia
miR-31-5p	ggcaaga	$F_2$	rational scroll	oral
miR-122-5p	gagugug	$F_2$	no cubic	ubiquitous
miR-29b-5p	cugguuu	$F_2$	no isol. sing.	head, neck
miR-143-3p	gagauga	$F_2$	no cubic	pancreatic
miR-140-5p	agugguu	$F_2$	no isol. sing.	colorectal
miR-340-5p	uauaaag	$F_2$	no cubic	ubiquitous
miR-340-3p	cguucuc	$F_2$	no cubic	ubiquitous
miR-27-5p	cugucug	$F_2$	no isol sing.	Wilms' tumor
miR-27-3p	acagcaa	$F_2$	no cubic	cervical
ine				

### 3.1. Human microRNAs That Mainly Act as Oncomirs

In this subsection, we provide details about the results summarized in Table 1. The first five miRNAs have two distinct nucleotides in their seed. Their card seq may be that of the free group  $F_1$  of rank one or correspond to a group away from  $F_1$ . All of them have a Groebner basis containing DP3 simply singular cubic surfaces. The other selected oncomirs have three distinct nucleotides in their seed. Their card seq may be that of the free group  $F_2$  of rank one or correspond to a group away from  $F_2$ . All of them have a Groebner basis containing DP3 simply singular cubic surfaces. Some of the surfaces correspond to a Painlevé equation PVI ( $PVI_a$  means that we are dealing with an algebraic solution of PVI), some surfaces correspond to PV equation. These results confirm our approach that a disease is in sight when either the card seq is away from a free group or the Groebner basis contains surfaces with isolated singularities. As already mentioned, PV is a wilder singular form than PV and we can expect a more severe disease in this case.

#### The Oncomir miR-21-3p

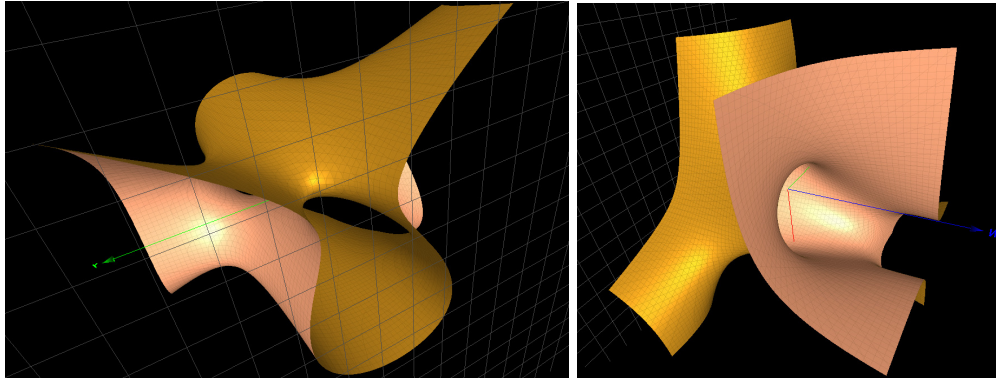
The clinical importance of miR-31-3p in human cancers was investigated in [28]. miR-21 expression is different among cancers and the expression level of miR-21 is primarily increased to promote tumor progression. Upregulation of miR-21 results in drug resistance and radio-resistance in various cancers.

The seed aacacca of miR-21-3p contains two distinct nucleotides a and c. The associated card seq is the sequence

[1, 1, 1, 2, 1, 3, 3, 1, 2, 2,  $\dots$ ] that is away from that of the free group  $F_1$ . According to our approach, this feature may be associated to a disease. The Groebner basis associated to the character variety of the seed is simple enough to merit an explicit display as

$$\mathcal{G}_{miR-21-3p} = (yz^2 - x^2 - z^2 - 2y + 4)(x^3 - y^2z + xy - 3x + z) \\ (z^3 - x + 3z)(xz - z^2 - y + 2)$$

The two cubic surfaces defining  $\mathcal{G}_{miR-21-3p}$  are degree 3 Del Pezzo (DP3) of the ADE type  $f^{(A_3)}$  and  $f^{(A_2)}$  (shown in Figure 2: Left). The third factor is a curve and the last one is a quadric not playing a role in our singularity analysis.



**Figure 2.** Left: the surface  $x^3 - y^2z + xy - 3x + z = 0$  of type  $f^{(A_2)}$  in the Groebner basis for the character variety of  $\mathcal{G}_{miR-21-3p}$ . Right: the surface  $xyz - x^2 - y^2 - yz + x + 2 = 0$  of the ADE type  $f^{(A_2)}$  in the Groebner basis for  $\mathcal{G}_{miR-204-5p}$ .

#### The Oncomir miR-204-5p

Abnormal expression of miR-204-5p plays a contrasting role in cancer [29,30]. A decrease expression often due to hypermethylation favors the progression of tumors while an increase expression may also be associated to prostate, breast or ovarian cancers. But miR-204-5p also has a role in a variety of tumors by regulating metastasis and apoptosis.

The seed uccuuu of miR-204-5p contains two distinct nucleotides u and c. The associated card seq is the sequence  $[1, 1, 2, 5, 5, 25, 43, 123, \dots]$  that is away from that of the free group  $F_1$ . According to our approach, this feature may be associated to a disease. The Groebner basis associated to the character variety of the seed is simple

$$\begin{aligned} \mathcal{G}_{miR-204-5p} &= (x-1)(x+1)(xz^2 - y^2 - z^2 - 2x + 4) \\ &\quad (y^3 - x^2z + xy - 3y + z)(xyz - x^2 - y^2 - yz + x + 2) \end{aligned}$$

The three cubic surfaces defining  $\mathcal{G}_{miR-204-5p}$  are DP3 of the type  $f^{(A_3)}$ ,  $f^{(A_2)}$  and  $f^{(A_2)}$  (shown in Figure 2), respectively.

#### The Oncomir miR-126b-5p

MicroRNA miR-126b-5p promotes tumor cell proliferation, metastasis and invasion by targeting the enzyme TD02 in hepatocellular carcinoma, one of the most common gastrointestinal malignancies, with the third highest mortality rate [31].

The seed auuauua of miR-204-5p contains two distinct nucleotides a and u. The associated card seq is the sequence  $[1, 2, 3, 2, 8, 7, 10, 18, 28, 27, 88, \dots]$  which corresponds to the modular group, denoted  $H_3$ . According to our approach, this feature may be associated to a disease. The Groebner basis associated to the character variety of the seed

$$\begin{aligned} \mathcal{G}_{miR-126b-5p} &= (y^3 - x^2z + xy - 3y + z)(y^2z - xy - xz + y - z) \\ &\quad (xyz - x^2 - y^2 - yz + x + 2)(xz^2 - y^2 + z^2 - 2yz)(yz^2 - xz - y - z) \end{aligned}$$

The first three cubic surfaces defining  $\mathcal{G}_{miR-126b-5p}$  are DP3 of the type  $f^{(A_2)}$  (an isolated singularity of the ADE type  $A_2$ ) or  $f^{(A_1A_2)}$  (an isolated singularity of the ADE type  $A_1A_2$ ). The last two cubic surfaces are rational scrolls devoid of an isolated singularity.

#### *The Oncomir mir-1908*

miR-1908-5p is aberrantly expressed in many diseases, especially cancer, being widely involved in a variety of cellular behaviors, including cell proliferation, cell differentiation, apoptosis, cancer cell invasion and metastasis, and extracellular vesicle secretion [32]. miR-1908-3p promotes the breast cancer cells proliferation and metastasis by suppressing several genes [33]. miR-1908 is also involved in the pathoetiology of bipolar disorder, myocardial infarction, obesity, renal fibrosis, rheumatoid arthritis and scar formation [34].

The seed ggcgggg of miR-1908-5p contains two distinct nucleotides c and g. The associated card seq is that of the free group  $F_1$  with a single generator. The attached Groebner basis contains the Cayley cubic  $f^{(4A_1)}$ : PVI.

The seed cggccgc of miR-1908-3p contains two distinct nucleotides c and g. The associated card seq is that of the group  $H_3$  already encountered for mmiR-126-5p. The attached Groebner basis contains DP3 surfaces of ADE type  $A_2$  and  $A_1A_2$ .

#### *The Oncomir miR-155-3p*

mir-155 is a key miRNA in both immunity and cancer and its miRNA strand, miR-155-3p, has been functionally implicated in both areas despite its low abundance (about 2%) compared to the -5p strand. An increased miR-155-3p strand percentage in cancerous tissues may indicate a specific dysregulation of mir-155 [35].

The seed uccuaca of miR-155-3p contains three distinct nucleotides a, c and u. The associated card seq is the sequence [3, 10, 51, 164, 1230, 7829, 59835, 491145 ···] that is away from that of the free group  $F_2$  of rank 2. According to our approach, this feature may be associated to a dysregulation and a disease.

The character variety is much more complicated than in the case of seeds with two distinct nucleotides. But it is not difficult to identify that the surface  $xyz + y^2 + z^2 - d = 0$ , with  $d = 4$  or  $6$ , of ADE type  $A_2$ , is part of the game.

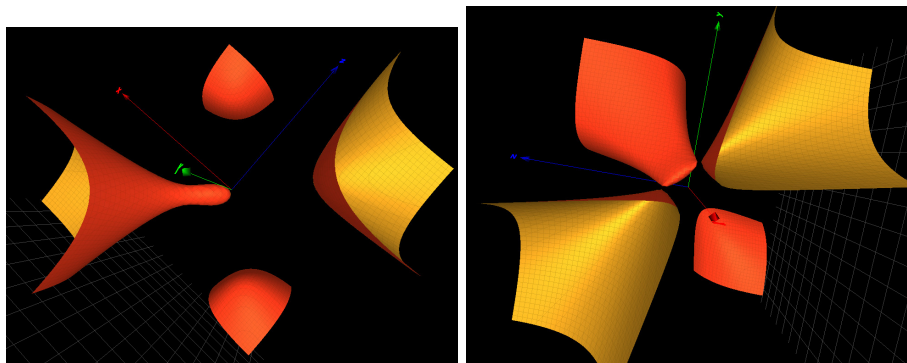
Additionally, the Groebner basis contains the surface  $2xyz - 2xz - 2x + y^2 - z^2 - 2z - 1 = 0$  whose ADE type is  $A_1A_2$ .

#### *The Oncomir miR-9-5p*

miR-9-5p is a well known promotor or suppressor of cancer metastasis. mir-9 mainly act as a promotor of metastasis capability in breast cancer, osteosarcoma, prostate cancer and bladder cancer, while it plays the opposite roles in other cancers such as colorectal, nasopharyngeal carcinoma, melanoma, gastric and brain cancer [36] [Table 1].

The seed cuuuggu of miR-9-5p contains three distinct nucleotides c, u and g. The associated card seq is that of the free group  $F_2$ .

The Groebner basis for the character variety attached to the seed contains the cubic surface  $xyz + x^2 + z^2 + 10 = 0$  which is of the type  $f^{(A_2)}$ :  $PV^{deg}$ . The Painlevé type is obtained with  $\omega_1 = \omega_2 = 0$  and the change of variables  $x \rightarrow \sqrt{10}x, z \rightarrow \sqrt{10}z$ . The Groebner basis also contains the DP3 cubic surface:  $xyz + x^2 + y^2 + z^2 - 2y + z + 1 = 0$  (see Figure 3: Left) and similar surfaces related to the solutions of PVI equation. Such surfaces are non algebraic that is not in the list provided in [11] [Table 4].



**Figure 3.** Left: the surface  $xyz + x^2 + y^2 + z^2 - 2y + z + 1 = 0$  of type PVI in the Groebner basis for the seed of miR-9-5p. Right: the surface  $xyz + y^2 + z^2 - 2y - 2z + 1 = 0$  of type  $f^{(A_2A_1A_1)} : PV$  in the Groebner basis for the seed of miR-146b-3p.

#### The Oncomir miR-146b-3p

It is known that miR-146b-3p is one important regulator of the proliferation of pancreatic cancer stem cells [37].

The seed ccugug of miR-146b-3p contains three distinct nucleotides c, g and u. The associated card seq is the sequence [3, 13, 92, 499, 5955, 77873 · · ·] that is away from that of the free group  $F_2$ . This feature may be associated to a dysregulation and a disease.

The Groebner basis attached to the seed contains DP3 cubic surfaces such as the surface  $xyz + y^2 + z^2 - 2 = 0$  of ADE type  $A_2$  and the surface  $xyz + y^2 + z^2 - 2y - 2z + 1 = 0$  the ZDE type  $A_2A_1A_1$  and Painlevé form PV, shown in Figure 3: Right.

#### The Oncomir miR-19a-5p

miR-19a-5p is part of the mir-17-92 cluster: a NF- $\kappa$ B regulated family of miRNAs that promotes cell proliferation, oncogenic transformation and evasion of apoptosis. High levels of miR-19a-5p correlate with poor prognosis in pancreatic cancer [38].

The seed guuugc of miR-19a-5p contains three distinct nucleotides c, g and u. The associated card seq is that of the free group  $F_2$ .

The Groebner basis attached to the seed contains the DP3 cubic surface  $xyz + x^2 + y^2 + z^2 - 6 = 0$  which corresponds to a solution of the PVI equation devoid of an isolated singularity. This solution is the (octahedral) algebraic solution 30 in [11]. A parametric plot for the modulus of this solution is in [10, Figure 5].

Other DP3 cubic surfaces in the Groebner basis are  $xyz + y^2 - 8 = 0$  of type  $f^{(A_2A_2)}$  and  $xyz + y^2 - xy - yz - 2xz + z - 1 = 0$  of type  $f^{(A_1A_1)}$ .

#### The Oncomir miR-181a-5p

The dysregulation of miR-181a-5p has been implicated in various types of cancer and functions as an oncomir or tumor inhibitor [39].

The seed acauca of miR-181a-5p contains three distinct nucleotides a, c and u. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed contains DP3 cubic surfaces such as the surface  $xyz + x^2 + z^2 - 2 = 0$ ,  $xyz + x^2 + z^2 - 4 = 0$  and  $xyz + x^2 + z^2 - x - 2z - 2 = 0$  which are all the ADE type  $A_2$ . It also contains the surface  $xyz + x^2 + z^2 - x = 0$  of ADE type  $A_4$ .

#### The Oncomir miR-15-5p

miR-15-5p participates in the pathogenesis of several cancers as well as non-malignant conditions, such as abdominal aortic aneurysm, Alzheimer's and Parkinson's diseases. Dysregulation of miR-15b-5p in clinical samples has been associated with poor outcome in different kinds of cancers [40].

The seed agcagca of miR-15-5p contains three distinct nucleotides a, c and g. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed contains a DP3 cubic surface  $xyz + x^2 + y^2 - 4y + 1 = 0$  which is of type  $f^{(A_2)}$ : PV as well of other surfaces of the PV type.

#### *The Oncomir miR-569*

miR-569 expression levels are markedly downregulated in lung cancer cells. miR-569 also contributes to ovarian and breast cancer cell survival and proliferation [41].

The seed guuaaug of miR-569 contains three distinct nucleotides a, g and u. The associated card seq is the sequence [3, 10, 51, 164, 1230, 7829, ...] that is away from that of the free group  $F_2$ . This feature may be associated to a dysregulation and a disease. The Groebner basis attached to the seed contains the DP3 cubic surfaces  $xyz + x^2 + y^2 - 4 = 0$  which is of the ADE type  $A_2$  and  $y^2z + 2xy - 2z^2 - 2x + z = 0$  which is of type  $A_4$ .

#### *The Oncomir miR-133b*

miR-133b originally defined as a canonical muscle-specific microRNA. It has been found to show abnormal expression in various kinds of human cancer and its complex complicated regulatory networks affects the tumorigenicity and development of malignant tumors [42].

The seed uuggucc of miR-133b-3p contains three distinct nucleotides c, g and u. The associated card seq is the sequence [3, 7, 34, 139, 931, 5208, ...] that is away from that of the free group  $F_2$ . The Groebner basis attached to the seed contains the DP3 cubic surfaces  $xyz + x^2 + z^2 - d = 0$ ,  $d=1$  or  $2$ , characteristic of the ADE type  $A_2$ , surfaces such as  $xyz - yz + x^2 + 3 = 0$  of type  $A_2A_2$  and surfaces of type  $A_1$ .

#### *The Oncomir miR-1270b*

miR-1270 plays a crucial role in the initiation and development of osteosarcoma, which is the most common bone malignancy worldwide, characterized by high morbidity and mortality [43]. Incidentally, miR-1270 is also differentially expressed in Meniere's disease, a chronic debilitating disorder of the inner ear, characterized by fluctuating episodes of vertigo and hearing loss [44].

The seed uggagau of miR-1270 contains three distinct nucleotides a, u and g. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed contains DP3 surfaces such as  $4xyz - 4z^2 + 3y^2 - 2yz = 0$  of ADE type  $A_4$ ,  $4xyz + 4y^2 + 4z^2 - z - 2 = 0$  of type  $A_2$  and  $4xyz + 3y^2 - 2yz - 6 = 0$  of type  $A_2A_2$ . None of them belong to the Painlevé class.

### *3.2. Human microRNAs That Mainly Act as Tumor Suppressors*

Table 2 provide a short list of miRNAs generally considered as tumor suppressors. As already mentioned, the boundary between an oncomir and a tumor suppressor is not tight. The first five miRNAs either have a card seq away from the free group  $F_2$  or have a Groebner basis containing simply singular surfaces in one of their -5-p or 3-p strands. But many other miRNAs in Table 2 have the card seq of  $F_2$  and no simply singular surface is found in the Groebner basis. This is in accordance of our approach.

#### *MicroRNA let-7a-5p*

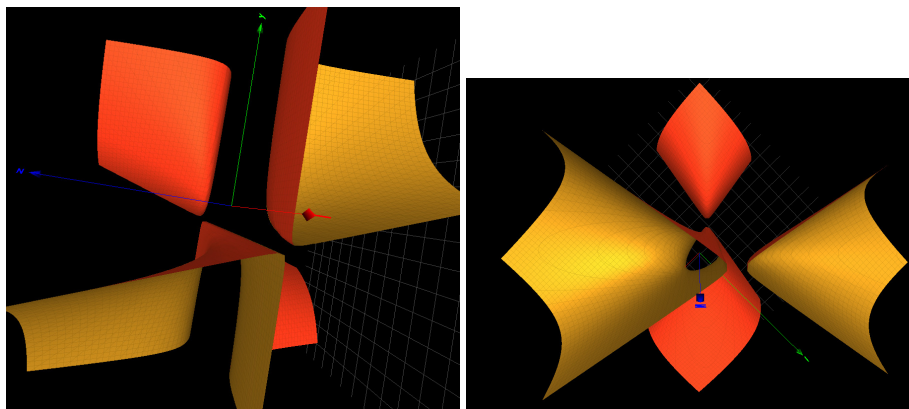
The let-7 family of microRNAs, first identified in *C. elegans* but functionally conserved from worms to human, is an important class of regulators for diverse cellular functions ranging from cell proliferation, differentiation and pluripotency to cancer development and progression [45,46]. let-7a is abundant in glioma and play a role in prostate and lung cancer development. Lower expression of miR-let-7a in patients with lung cancer brain metastasis was closely related to unfavorable efficacy and prognosis of radiotherapy [47].

The seed gagguaug of miR-let-7a-5p contains three distinct nucleotides a, g and u. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed contains DP3 cubic surfaces  $xyz + 2z^3 + 3 = 0$  and  $xyz + 2z^2 + xz + y - 1 = 0$  of type  $f^{(A_1A_2)}$  and  $xyz + 2z^2 + xz + y + 1 = 0$  of



type  $f^{(A_1A_4)}$ . The later surface is pictured in Figure 4: Left. None of these surfaces are found of the Painlevé type.

The seed uauacaa of miR-let-7a-5p contains three distinct nucleotides a, c and u. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed does not contain cubics and higher order surfaces.



**Figure 4.** Left: the surface  $xyz + 2z^2 + xz + y + 1 = 0$  of type  $f^{(A_1A_4)}$  in the Groebner basis for the seed of miR-let-7a. Right: the surface  $xyz + x^2 + y^2 + yz - 1 = 0 = 0$  of type  $f^{(A_4)}$  in the Groebner basis for the seed of miR-34b-3p.

#### MicroRNA 34b-3p

The overexpression of miR-34b-3p has been shown to suppress lung cancer cell growth, including proliferation inhibition, cell cycle arrest and increased apoptosis [48]. Downregulation of miR-34b-3p is significantly hypermethylated and consequently downregulated, particularly in neuroblastoma patients at high risk of progression [49].

The seed aaucacu of miR-34b-3p contains three distinct nucleotides a, c and u. The associated card seq is the sequence  $[3, 10, 51, 164, 1230, 7829, \dots]$  that is away from that of the free group  $F_2$ . The Groebner basis attached to the seed contains DP3 surfaces  $xyz + x^2 + y^2 - 2 = 0$  which is of type  $A_2$ , as well as  $xyz + x^2 + y^2 + yz - 2y - 1 = 0$  which is of type  $A_3$  and  $xyz + x^2 + y^2 + yz - 1 = 0$  which is of type  $A_4$ . The later surface is pictured in Figure 4: Right.

#### MicroRNA 200a-5p

Expression in melanoma cells may be controlled by miR-200a-5p leading to a downregulation of surface expression for the human leukocyte antigen class I, which is linked to a reduced survival of patients [50]. The mir-200 family acts as an oncogene in colorectal cancer [51]. MicroRNAs from the mir-200 family are commonly associated with the inhibition of the metastatic potential of cancer cells, following inhibition of ZEB transcription factors expression and epithelial-to-mesenchymal transition [52].

The seed aucuuac of miR-200a-5p contains three distinct nucleotides a, c and u. The associated card seq is the sequence  $[3, 10, 35, 140, 921, 5778, \dots]$  that is away from that of the free group  $F_2$ . The Groebner basis attached to the seed contains DP3 cubic surfaces of the ADE type  $f^{(A_1A_3)}$  and  $f^{(A_1A_1A_3)}$  which are not in the Painlevé class.

#### MicroRNA 22-5p

A study revealed that miR-22-5p downregulation contributed to the malignant progression of non-small-cell lung cancer by targeting transcription factor TWIST2 [53]. miR-22-5p may also inhibit tumor progression by sponging DNA topoisomerase II-alpha [54].



The seed guucuuc of miR-22-5p contains three distinct nucleotides u, c and g. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed contains a cubic surface which is the DP3 surface  $y^2z + x^2 - 2y^2 + 2xy + 1 = 0$  of the ADE type  $D_4$ .

#### *MicroRNA 214*

miR-214 is one of the fastest evolving microRNAs [55] [Table 3]. First mir-214 was reported to promote apoptosis in HeLa cells. Presently, mir-214 is implicated in an extensive range of conditions such as cardiovascular diseases, cancers, bone formation and cell differentiation [56].

For miR-214-5p and the short seed sequence GCCUGU, we find the DP3 surface  $xyz + y^2 + z^2 - 4 = 0$  of type  $A_2$ . The reader can look at [57] [Figure 5] for a picture. A surface  $xyz + y^2 + z^2y - 2z - 1 = 0$  of  $A_2$  is also found. For a longer seed GCCUGUC, surfaces are not found to contain a cubic surface.

For miR-214-3p and the short seed sequence cagcag, we find a DP3 surface  $xyz + 2xy - y^2 + z + 2 = 0$  of ADE type  $f^{(A_1A_4)}$  in the corresponding Groebner basis. With the longer seed cagcagg, no cubic surface is found within the Groebner basis.

#### *MiRNA 503-5p*

The slowest evolving miRNA gene in the human species is hsa-miR-503. It regulates gene expression in various pathological processes of diseases, including carcinogenesis, angiogenesis, tissue fibrosis, and colon cancer [58,59].

The seed region for miR-503-5p is agcagcg. The associated card seq is that of the free group  $F_2$ . The Groebner basis is a rational scroll devoid of isolated singularities For a longer seed agcagcgg singular surfaces are found [57, Section 5].

#### *MicroRNA 141-5p*

miR-141-5p, an important member of the mir-200 family, has been reported to be involved in cellular proliferation, migration, invasion, and drug resistance in different kinds of human malignant tumors. It may act as a tumor suppressor via targeting gene RAB32 in the development of chronic myeloid leukemia [60].

The seed aucuucc of miR-141-5p contains three distinct nucleotides a, u and c. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed do not contain a cubic.

#### *MicroRNA 31-5p*

miR-31 was shown to regulate a number of metastasis-related genes involved in the proliferation, cell-cycle regulation and apoptosis [61]. miR-31-5p Is a potential circulating biomarker and therapeutic target for oral cancer [62].

The seed ggcaaga of miR-31-5p contains three distinct nucleotides a, c and g. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed contains a cubic surface  $xy^2 - 2xy - x + z = 0$  which is rational scroll devoid of isolated singularity and not of the Painlevé type.

#### *MicroRNA 122-5p*

Use of miR-122 as a cutting-edge tool for cancer treatment. Cancer cell lines revealed that mir-122 functions as an oncogenic or tumor-suppressive miRNA and examination of patient tumor samples furthermore supported the idea that mir-122 dysregulation is a crucial component in the growth of malignancies [63].

The seed gagugug of miR-122-5p contains three distinct nucleotides a, u and g. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed do not contain a cubic.

#### *MicroRNA 29b-5p*

miR-29b-5p is a key oncomiR and therefore a target for molecular diagnosis and treatment of patients with oral squamous cell carcinoma [64].

The seed cugguuu of miR-29b-5p contains three distinct nucleotides c, u and g. The associated card seq is that of the free group  $F_2$ . The character variety attached to the seed contains cubics of a DP3 type, a rational scroll and a ruled surface over a genus 1 curve but none of them has an isolated singularity and they are not of the Painlevé type.

#### *MicroRNA 143-3p*

Brain metastasis is one of the secondary mortality causes for patients. It was found that miR-143-3p is upregulated in the paired brain metastasis tissues in contrast with primary cancer tissues. It may be used as a prognostic factor for in vivo progression, invasion, metastasis and survival rate of lung cancer [65]. miR-143-3p may suppress tumorigenesis in pancreatic duct adenocarcinoma by targeting KRAS gene [66].

The seed gagauga of miR-143-3p contains three distinct nucleotides a, u and g. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed does not contain a cubic.

#### *MicroRNA 140-5p*

The low expression of miR-140-5p is related to tumor stage or metastasis. The miR-140-5p overexpression suppressed cell proliferation and invasion in colorectal carcinoma [67].

The seed agugguu of miR-140-5p contains three distinct nucleotides a, u and g. The associated card seq is that of the free group  $F_2$ . The Groebner basis attached to the seed contains a ruled surface over a genus 1 curve. There is none isolated singularity neither a Painlevé type cubic.

### 3.3. *MicroRNA 340*

mir-340 may act either as an oncogene or a tumor suppressor by targeting genes related to proliferation, apoptosis, and metastasis, as well as those associated with diagnosis, treatment, chemoresistance, and prognosis [68]. mir-340 restricts the development of breast cancer cells by targeting numerous oncogenes. mir-340 has been shown to play a suppressive role in lung, colorectal ovarian and prostate cancers, laryngeal squamous cell carcinoma and osteosarcoma. mir-340 plays a critical role in in non-small cell lung cancer, and its overexpression restricts the growth and invasion of the corresponding cells [69].

The seed of miR-340-5p is uauaaag and that of miR-340-3p is cguucuc. Both of them have a card seq which is that of the free group  $F_2$ . For both of them the Groebner basis attached to the seed does not contain cubic surfaces.

### 3.4. *MicroRNA 27*

Wilms' tumor is the most common type of renal tumor in children. There is tumor-suppressive effect of miR-27a-5p in Wilms' tumor cells [70]. Cervical cancer is the fourth most frequently occurring cancer and the fourth leading cause of cancer-associated mortality among women [71].

For both cases, the card seq associated to the group  $f_p$  defined from the seed is  $F_2$ . No isolated singularity is found.

## 4. Conclusions

In this paper, we proposed a group theoretical definition of homeostasis at the level of the transcriptome. We applied this research to miRNAs controlling the (dys)regulation of gene expression in cancer. In our approach, the seed of a miRNA serves as a generator for an infinite group  $f_p$  whose  $SL_2(\mathbb{C})$  character variety, more precisely its Groebner basis  $\mathcal{G}$ , contains cubic surfaces that are a signature of a possible disease. We found a correlation between oncomirs and either (i)  $f_p$  is away from the free group  $F_r$ , or (ii)  $\mathcal{G}$  contains at least a simply singular cubic  $f^{(ADE)}$ , or (iii) the dynamics of  $f^{(ADE)}$  is a non linear differential equation of the Painlevé type.

We emphasized the connection of Painlevé VI equation to the topology of the 4-punctured sphere and its breaking when two punctures collapse. Mathematically, the character variety of PVI is described

by a fundamental group. In contrast, the character variety of PV equation is a fundamental groupoid that rules the ‘Stokes phenomenon’ of breaking the 4-punctured sphere [13,72].

The model is applied to many mirRNAs known to play a role in cancer. In the future, this approach may be applied to other nucleotide sequences that play a role in the regulation of gene expression.

**Author Contributions:** Conceptualization, M.P.; methodology, M.P., D.C.; software, M.P.; validation, M.P., D.C.; formal analysis, M.P.; investigation, M.P., D.C.; writing–original draft preparation, M.P.; writing–review and editing, M.P.; visualization, D.C.; supervision, M.P.; project administration, M.P. Both authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Data are available from the authors after a reasonable demand.

**Acknowledgments:** The first author would like to acknowledge the contribution of the COST Action CA21169, supported by COST (European Cooperation in Science and Technology).

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Bernard C. *Lectures on the phenomena common to animals and plants* 1974. Charles C Thomas Pub Ltd.: Springfield, IL, USA, 1974.
2. Gene expression, available at [https://en.wikipedia.org/wiki/Gene\\_expression](https://en.wikipedia.org/wiki/Gene_expression) (accessed on 1 January 2023).
3. Hyde, J. S. Genetic homeostasis and behavior: analysis, data, and theory. *Behavior Genetics* **1973**, *3*, 233–245.
4. Cowley Jr, A. W. Genomics and homeostasis. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2002**, *284*, R 611–27.
5. Genome homeostasis defects drive enlarged cells into senescence. Manohar, S.; Estrada, M. E.; Uliana, F.; Vuina, K.; Alvarez, P. M.; de Bruin, R. A.M.; Neurohr, G. E. *Molec. Cell* **2023**, *83*, 4032–4046.
6. Planat, M.; Amaral, M. M.; Irwin, K. Algebraic morphology of DNA–RNA transcription and regulation. *Symmetry* **2023**, *15*, 770.
7. Planat, M.; Amaral, M. M.; Fang, F.; Chester, D.; Aschheim R.; Irwin, K. DNA sequence and structure under the prism of group theory and algebraic surfaces. *Int. J. Mol. Sci.* **2022**, *3*, 13290.
8. Planat, M.; Amaral, M.; Chester, D.; Fang, F.; Aschheim, R.; Irwin, K. Group theory of messenger RNA metabolism and disease. *Gene Expression* **2024**. DOI: 10.14218/GE.2023.00079.
9. Schrödinger, E. *What Is Life? The Physical Aspect of the Living Cell*; Cambridge University Press: **1944**, Cambridge, UK.
10. Planat, M.; Chester, D.; Irwin, K. Dynamics of Fricke–Painlevé VI surfaces. *Dynamics* **2024**, *4*, 1–13.
11. Lisovyy O.; Tykhyy Y. Algebraic solutions of the sixth Painlevé equation. *J. Geom. Phys.*, **2014**, 124–163.
12. Klimeš M. Wild monodromy of the fifth Painlevé equation and its action on the wild character variety: approach of confluence. *arXiv* **2021**, arXiv:math.CA /1609.05185.
13. Paul, E.; Ramis, J. P. Dynamics of the fifth Painlevé foliation. *arXiv* **2023**, arXiv:math.DS /2301.08597.
14. Clarkson, P. A.; Joshi, N.; Mazzocco, M.; Nijhoff, F. W.; Noumi, M. One hundred years of PVI, the Fuchs–Painlevé equation. *J. Phys. A: Math. Gen.* **2006**, *39*, E01.
15. Isomonodromic deformation. Available online: [https://en.wikipedia.org/wiki/Isomonodromic\\_deformation](https://en.wikipedia.org/wiki/Isomonodromic_deformation) (accessed on 1 August 2023).
16. Boalch, P. From Klein to Painlevé via Fourier, Laplace and Jimbo. *Proc. Lond. Math. Soc.* **2005**, *90*, 167–208.
17. Cantat, S. Bers and Hénon, Painlevé and Schrödinger. *Duke Math. J.* **2009**, *149*, 411–460.
18. Jimbo, M.; Miwa, T. Monodromy preserving deformation of linear ordinary differential equations with rational coefficients. II. *Physica* **1981**, *2D*, 407–448.
19. Chekhov, L.; Mazzocco, M.; Rubtsov, V. Painlevé monodromy manifolds, decorated character varieties and cluster algebras. *Int. Math. Res. Not.* **2017**, *24*, 7639–7691.
20. Van Der Put, M.; Saito, M. H. Moduli spaces for linear differential equations and Painlevé equations. *Ann. Inst. Fourier* **2009**, *59*, 2611–2667.

21. Goldman, W. M. Trace coordinates on Fricke spaces of some simple hyperbolic surfaces. In *Handbook of Teichmüller theory*, Eur. Math. Soc., Zürich, **2009**, *13*, 611-684.
22. Ashley, C.; Burelle J.P.; Lawton, S. Rank 1 character varieties of finitely presented groups. *Geom. Dedicata* **2018**, *192*, 1–19.
23. Markopoulos, G. S.; Roupakia, E.; Tokamani, M.; Chavdoula, E.; Hatziapostolou, M.; Polytarchou, C.; Marcu, K. B.; Papavassiliou, A. G. Sandaltzopoulos, R.; Kolettas, E. A step-by-step microRNA guide to cancer development and metastasis. *Cell Oncol.* **2017**, *40*, 303–339.
24. Lei, X.; Mudiyansele, T. B.; Zhang, Y.; Bian, C.; Lan, W.; Yu, N.; Pan, Y. A comprehensive survey on computational methods of non-coding RNA and disease association prediction. *Briefings in Bioinformatics* **2021**, *22*, 1–31.
25. Xie, B.; Ding, Q.; Han, H.; Wu, D. miRCancer: a microRNA-cancer association database constructed by text mining on literature. *Bioinformatics* **2013**, *29*, 638–644
26. Kozomara, A.; Birgaonu, M.; Griffiths-Jones, S. miRBase: from microRNA sequences to function. *Nucl. Acids Res.* **2019**, *47*, D155–D162.
27. miRBase: the microRNA database. Available online: <https://www.mirbase.org/> (accessed on 1 November 2022).
28. Hashemi, M.; Mirdamadi, M. S.A.; Talebi, Y.; Khaniabad, N.; Banaei, G.; Daneii, P.; Gholami, S.; Ghorbani, A.; Tavakolpournegari, A.; Farsani, Z. M.; Zarrabi, A.; Nabavi, N.; Zandieh, M. A.; Rashidi, M.; Taheriazam, A.; Entezari, M.; Khan, H. Pre-clinical and clinical importance of miR-21 in human cancers: umorigenesis, therapy response, delivery approaches and targeting agents. *Pharm. Res.* **2023**, *187*, 106568.
29. Li, T.; Pan, H.; Li, R. The dual regulatory role of miR-204 in cancer. *Tumor Biol.* **2016**, *37*, 11667–11677.
30. Yang, F.; Bian, Z.; Xu, P.; Sun, S.; Huang, Z. MicroRNA-204-5p: a pivotal tumor suppressor. *Cancer Medicine* **2023**, *12*, 3185–3200.
31. Ai, Y.; Luo, S.; Wang, B.; Xiao S.; Wang, Y. miR-126-5p promotes tumor cell proliferation, metastasis and invasion by targeting TDO2 in hepatocellular carcinoma. *Molecules* **2022**, *27*, 443.
32. Shen, J.; Wu, Y.; Ruan, W.; Zhu, F.; Duan, S. miR-1908 Dysregulation in human cancers. *Front. Oncol.* **2022**, *12*, 857743.
33. Zhu, Y.; Wang, Q.; Xia, Y.; Xiong, X.; Weng, S.; Ni, H.; Ye, Y.; Chen, L.; Lin, J.; Chen, Y.; Niu, H.; Chen, X.; Lin, Y. Evaluation of miR-1908-3p as a novel serum biomarker for breast cancer and analysis its oncogenic function and target genes. *BMC Cancer* **2020**, *20*, 644.
34. Ghafouri-Fard, S.; Khoshbakht, T.; Hussein, B. M.; Thaheri, M.; Samsami, M. miR-1908: a microRNA with diverse functions in cancers and non-malignant conditions. *Cancer Cell Int.* **2022**, *22*, 281.
35. Dawson, O.; Piccinini, A. M. miR-155-3p: processing by-product or rising star in immunity and cancer? *Open Biol.* **2022**, *12*, 220070.
36. Liu, Y.; Zhao, Q.; Xi, T.; Zheng, L.; Li, X. MicroRNA-9 as a paradoxical but critical regulator of cancer metastasis: Implications in personalized medicine. *Genes & Diseases* **2021**, *8*, 759–768.
37. Zhou, M.; Gao, Y.; Wang, M.; Guo, X.; Li, X.; Zhu, F.; Xu, S.; Qin, R. miR-146b-3p regulates proliferation of pancreatic cancer cells with stem cell-like properties by targeting MAP3K10. *J. of Cancer* **2021**, *12*, 3726–3740.
38. Ardizzone, A.; Calabrese, G.; Campolo, M.; Filippone, A.; Giuffrida, D.; Esposito, F.; Colarossi, C.; Cuzzocrea, S.; Esposito, E.; Paterniti, I. Role of miRNA-19a in Cancer Diagnosis and Poor Prognosis. *Int. J. Mol. Sci.* **2021**, *22*, 4697.
39. Li, J.; Shen, J.; Zhao, Y.; Du, F.; Li, M.; Wu, X.; Chen, Y.; Wang, S.; Xiao, Z.; Wu, Z. V. Role of miR-81a-5p in cancer (Review). *Int. J. Oncol.* **2023**, *63*, 108.
40. Ghafouri-Fard, S.; Khoshbakht, T.; Hussein, B. M.; Jamal, H. H.; Taheri, M.; Hajiesmaeili, M. A comprehensive review on function of miR-15b-5p in malignant and non-malignant Disorders. *Front. Oncol.* **2022**, *12*, 870996.
41. Zheng, Y. P.; Wu, L.; Gao, J.; Wang, Y. Tumor suppressive role of miR-569 in lung cancer. *Oncol. Lett.* **2018**, *15*, 4087-4092.
42. Li, D.; Xia, L.; Chen, M.; Lin, C.; Wu, H.; Zhang, Y.; Pan, S. Li, X. miR-133b, a particular member of myomiRs, coming into playing its unique pathological role in human cancer. *Oncotarget* **2017**, *8*, 50193-50208.
43. Liu, Y.; Guo, W.; Fang, S.; He, B.; Li, X.; Fan, L. miR-1270 enhances the proliferation, migration, and invasion of osteosarcoma via targeting cingulin. *Eur. J. Histochem.* **2021**, *65*, 3237.
44. Shew, M.; Wichova, H.; Peter, M. St; Warnecke, A.; Staecker, H. Distinct microRNA profiles in the perilymph and serum of patients with Menière's disease. *Front. Neur.* **2021**, *12*, 646928.

45. Yazarlou, F.; Kadkhoda, S.; Ghafouri-Fard, S. Emerging role of let-7 family in the pathogenesis of hematological malignancies. *Biomed. & Pharmac.* **2021**, *144*, 112334.
46. Ma, Y.; Snen, N.; Wicha, M. S.; Luo, M. The Roles of the Let-7 Family of MicroRNAs in the Regulation of Cancer Stemness. *Cells* **2021**, *10*, 2415.
47. Liu, J. K.; Liu, H. F.; Ding, Y.; Gao, G. D. Predictive value of microRNA let-7a expression for efficacy and prognosis of radiotherapy in patients with lung cancer brain metastasis. *Medicine (Baltimore)* **2018**, *97*, e12847.
48. Feng, H.; Ge, F.; Du, L.; Zhang, Z.; Liu, D. miR-34b-3p represses cell proliferation, cell cycle progression and cell apoptosis in non-small-cell lung cancer (NSCLC) by targeting CDK4. *J. Cell Mol. Med.* **2019**, *23*, 5282-5291.
49. Aravindan, N.; Subramanian, K.; Somasundaran, D. B.; Herman, T. S.; Aravindan, S. MicroRNAs in neuroblastoma tumorigenesis, therapy resistance, and disease evolution. *Cancer Drug Resist* **2019**, *2*, 1086.
50. Lazaridou, M. F.; Gonschorek, E.; Massa, C.; Friedrich, M.; Handke, D.; Mueller, A.; Jasinski-Bergner, S.; Dummer, R.; Koelblinger, P.; Seliger, B. Identification of miR-200a-5p targeting the peptide transporter TAP1 and its association with the clinical outcome of melanoma patients. *Oncoimmunology* **2020**, *9*, 1774323.
51. Carter, J. V.; O'Brien, S. J.; Burton, J. F.; Oxford, B. G.; Stephen, V.; Hallion, J.; Bishop, C.; Galbraith, N. J.; Eichenberger, M. R.; Sarojini, H.; Hattab, E.; Galandiuk, S. The microRNA-200 family acts as an oncogene in colorectal cancer by inhibiting the tumor suppressor RASSF2. *Oncol. Lett.* **2019**, *18*, 3994-4007.
52. Diaz-Riascos, Z. V.; Ginesta, M. M.; Fabrejat, J.; Buscai, L.; Cordelier, P.; Capellá, G. Expression and role of microRNAs from the miR-200 Family in the tumor formation and metastatic propensity of pancreatic cancer. *Mol. Ther. Nucleic Acids* **2019**, *17*, 491-503.
53. Han, X.; Li, H.; Liu, S.; Ehao, Z. Study on the potential mechanism of miR-22-5p in non-small-cell lung cancer. *Dis. Markers* **2022**, *Sep. 6*, 3750734.
54. Zhao, H.; Chen, C.; Song, H.; Qin, R.; Wang, X.; He, Q.; Li, F.; Zhao, H.; Li, Y. DNA topoisomerase II- $\alpha$  regulated by miR-22-5p promotes hepatocellular carcinoma invasion and migration through the hippo pathway. *Oxid Med. Cell Longev* **2022**, *Oct. 17*, 4277254.
55. Fromm, B.; Billipp, T.; Peck, L. E.; Johansen, M.; Tarver, J. E.; King, B. L.; Newcomb, J. M.; Sempere, L. F.; Flatmark, K.; Hovig, E.; Peterson, K. J. A uniform system for the annotation of human microRNA genes and the evolution of the human microRNAome. *Annu. Rev. Genet.* **2015**, *23*, 213-242.
56. Amin, M. M.J.; Trevelyan, C. J.; Tyturner, N. A. MicroRNA-214 in health and disease. *Cells* **2021**, *23*, 3274.
57. Planat, M.; Amaral, M. M.; Chester, D.; Irwin, K.  $SL(2, \mathbb{C})$  scheme processing of singularities in quantum computing and genetics. *Axioms* **2023**, *12*, 233.
58. He, Y.; Cai, Y.; Pai, P. M.; Ren, X.; Xia, Z. The causes and consequences of miR-503 dysregulation and its impact on cardiovascular disease and cancer. *Front. Pharmacol.* **2021**, *12*, 629611.
59. Wei, L.; Sun, C.; Zhang, Y.; Han, N.; Sun, S. miR-503-5p inhibits colon cancer tumorigenesis, angiogenesis, and lymphangiogenesis by directly downregulating VEGF-A. *Gene Ther.* **2022**, *1-2*, 28-40.
60. Bao, J.; Li, X.; Li, Y.; Huang, C.; Meng, X.; Li, J. MicroRNA-141-5p acts as a tumor suppressor via targeting RAB32 in chronic myeloid leukemia. *Front. Pharmacol.* **2019**, *10*, 1545.
61. Schmittgen, T. D. mir-31: a master regulator of metastasis? *Future Oncol.* **2009**, *6*, 17-20.
62. Lu, Z.; He, Q.; Liang, J.; Li, W.; Su, Q.; Chen, Z.; Wan, Q.; Zhou, X.; Cao, L.; Sun, J.; Wu, Y.; Liu, L.; Wu, X.; Hou, J.; Lian, K.; Wang, A. miR-31-5p is a potential circulating biomarker and therapeutic target for oral cancer. *Mol. Ther. Nucleic Acids* **2019**, *16*, 471-480.
63. Lashkarian, M. F.; Hashemipour, N.; Niaraki, N.; Soghala, S.; Moradi, A.; Sarhangi S.; Hatami, M.; Aghaei-Zarch, F.; Khosravifar, M.; Mohhammadzadeh, A.; Najafi, S.; Majidpoor, J.; Farnia, P.; Aghaei-Zarch, S. M. MicroRNA-122 in human cancers: from mechanistic to clinical perspectives. *Cancer Cell Int* **2023**, *23*, 20.
64. Kurihara-Shimomura, M.; Sasahira, T.; Shimomura, H.; Nakashima, C.; Kirita, T. The oncogenic activity of miR-29b-1-5p induces the epithelial-mesenchymal transition in oral squamous cell carcinoma. *J. Clin. Med.* **2019**, *8*, 273.
65. Asghariazar, V.; Kadkhodayi, M.; Sarailoo, M.; Jolfayi, A. G.; Baradaran, B. MicroRNA-143 as a potential tumor suppressor in cancer: An insight into molecular targets and signaling pathways. *Patho. Res. and Pract.* **2023**, *250*, 154792.
66. Xie, F.; Li, C.; Zhang, X.; Peng, W.; Wen, T. miR-143-3p suppresses tumorigenesis in pancreatic ductal adenocarcinoma by targeting KRAS. *Biomed. & Pharmac.* **2019**, *119*, 109424.



67. Zhao, Z.; Liu, W.; Li, J. miR-140-5p inhibits cell proliferation and invasion in colorectal carcinoma by targeting SOX4. *Oncol. Lett.* **2019**, *17*, 2215–2220.
68. Huang, Z.; Xu, Y.; Wan, M.; Zeng, X.; Wu, J. miR-340: A multifunctional role in human malignant diseases. *Int. J. Biol. Sci.* **2021**, *17*, 236–246.
69. Zhu, X.; Tian, G.; Quan, J.; He, P.; Liu, J. Effects of miR-340 overexpression and knockdown on the proliferation and metastasis of NSCLC cell lines. *Int. J. Mol. Med.* **2019**, *44*, 643–651.
70. Guo, Z. T.; Yu, Q.; Miao, C.; Ge, W.; Li, P. MicroRNA-27a-5p inhibits proliferation, migration, and invasion and promotes apoptosis of Wilms' tumor cell by targeting PBOV1. *Mol. Cell. Biog.* **2022**, *42*, e0039721.
71. Li, M.; Gao, Z.; Wang, S. Zhao, Y.; Xie, H. miR-27a-3p upregulation by p65 facilitates cervical tumorigenesis by increasing TAB3 expression and is involved in the positive feedback loop of NF- $\kappa$ B signaling. *Oncol. Rep.* **2023**, *50*, 1791–2431.
72. Boalch, P. Topology of the Stokes phenomenon. *Proc. Symp. Pure Math.* **2021**, *103.1*, 01832.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.