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# DynBioSketch: A tool for sketching dynamic visual summaries in biology, and its application to infection phenomena

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# A B S T R A C T

Having simple methods of illustration is essential to scientific thinking. To complement the abstract sketches regularly used in cell biology, we propose DynBioSketch, an easy-to-use digital modeling and animation tool, enabling biologists to resort to less simplified representations when necessary without having to call professional artists. DynBioSketch is an interactive sketching system dedicated to the design and communication of biological phenomena at the cellular scale that can be illustrated in a few minutes of animation. Our model integrates 3D modeling, pattern-based design of 3D shape distributions, and sketch-based animation. These elements can be combined to create complex scenarios such as the infection phenomenon on which we focus, allowing a narrative design adapted to communication between researchers or educational applications in biology. Our results, along with a user study conducted with biology researchers, highlight the potential of DynBioSketch in enabling the direct design of dynamic visual summaries that convey relevant information, as shown in our infection case study. By bridging the gap between abstract representations used by experts and more illustrative depictions, DynBioSketch opens a new avenue for communicating biological concepts.

#### 1. Introduction

Biological phenomena inherently exhibit complexity, as a series of events or chemical reactions disrupt an initially stable environment, triggering transformations toward the next stable state. Biologists observe and experiment to study the causes and consequences of such events on an environment. Despite recent advancements in data visualization, 2D or 3D imagery merely represents events from the actual data, presenting challenges in comprehension and control. Consequently, biologists often resort to simpler representations, ranging from 2D sketches to 10 3D animations, to complement their understanding of a phe-11 nomenon or to communicate about it. While extremely useful, 12 abstract, conceptual sketches may only be understood by ex-13 perts. At the other end of the spectrum, designing full 3D, an-14 imated illustrations typically requires collaboration with a pro-15 fessional artist, a time-consuming process that often involves a 16 series of back-and forth discussions. As a result, many cell biol-17 ogists are in demand for user-friendly creation tools that allow 18 them to design simplified yet expressive digital illustrations of 19 their phenomenon of interest, appropriate to communicate both 20 among peers and in educational contexts. 21

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While current illustration tools are useful for scientific dis-22 semination and education, they often lack the ability to facili-23 tate sketching in volumetric environments. Such environments 24 involve interactions between 3D shapes and dynamic state changes, posing a challenge for representation. Existing meth-26 ods dealing with illustration at the cellular level have mainly focused on modeling vascular networks in 3D using sketch-28 ing conventions, or have explored blood circulation to represent 29 vessel pathologies. However, none of these approaches offers a 30 complete system for creating and animating a complex environ-31 ment in which cells can navigate and evolve dynamically. 32

In this paper, we focus on the cellular scale, particularly ex-33 amining the infection phenomenon. This particular choice en-34 ables us to tailor our objectives accordingly. Specifically, we 35 aim to design a schematic representation of what occurs un-36 der the skin during the infection phenomenon. Our concept 37 involves creating a stable environment composed of collagen fibers and vessels within which cells navigate. Subsequently, a 39 cut triggers the immune response to infection, with white cells deforming and then catching and removing the bacteria while 41 red cells escape through the cut. To facilitate understanding 42 and exploration of this phenomenon, we introduce an interac-43 tive sketching system for 3D modeling, animation, and narra-44 tion. This system is specifically designed to meet the needs of 45 biologists for effectively communicating and exploring this par-46

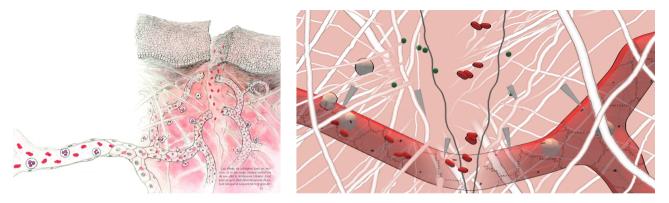


Fig. 1: An immunology illustration from the magazine Globule [1], © Renaud Chabrier (left) served as inspiration for a dynamic visual summary (right), i.e., an animation triggering narrative events, sketched using our system.

ticular phenomenon

This project was initiated through a collaboration with a biologist and a professional artist to help us conduct both a prestudy on depiction and narration in biology. The latter confirmed the need for an authoring tool to help biologists represent their understanding of the dynamic phenomenon of infection in a simplified way. Our solutions build on their feedback. While we use existing solutions for generating implicit surfaces from a sketch [2, 3], our main contributions are:

- A novel sketch-based modeling method to design distributions of 3D shape, enabling the modeling of dense 3D environments such as biological tissues with fiber networks in which cells evolve;
- A solution for sketch-based animation, applicable to both
   2.5D and 3D settings that builds on standard schematic
   conventions in biology to allow the specification of mo tion and deformation;
- An interface for specifying and controlling temporal nar rative scenarios with a series of events triggered over time;
- A final user study with 11 expert biologists, enabling us to validate the relevance of our tool and identify in which use cases 2.5D vs. 3D illustrations are the most relevant.

Throughout this paper, the illustrations focus on the challeng-23 ing case of immunology. We took inspiration from the drawing 24 in Fig. 1 left, expressing the human response to infection at 25 the cell level, and created before this work by one of our ex-26 pert co-authors. Using our tool to create a visual summary of 27 the dynamic phenomenon involved, we achieved similar visual 28 quality (see Fig. 1, right), while enabling the design of anima-29 tions triggered by narrative events positioned along a timeline. 30 This provided a proof of concept of the applicability of our sys-31 tem in a biological context. 32

Lastly, note that the first version of this work appeared in a conference paper focusing on 2.5D animation [4]. The present, fully rewritten paper extends this work by introducing a 3D solution for synthesizing and animating sub-shapes within an implicitly defined container, enabling us to respectively generate and animate blood cells within 3D branching vessels. It also includes the final user study, lacking in the first paper, which enabled us to discuss 2.5D versus 3D solutions with experts in biology. Let us note that although these experts were not directly involved in the development process of the tool, their feedback helped us to improve some aspects of the interface and our resulting toolbox.

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2. Related work

Designing a tool for sketching visual summaries of biolog-46 ical phenomena, such as the one in Fig 1, requires the ability 47 to model 3D shapes, and shape distributions, and to design ani-48 mations. It also requires studying previous illustration and nar-49 ration tools used in biology. We review these different topics 50 below, referring the reader to [5, 6, 7, 8] for comprehensive tu-51 torials and surveys on illustrative rendering techniques and their 52 applications in biology and anatomy. 53

3D modeling from a 2D sketch. Sketch-based modeling tech-54 niques range from methods that take full sketches and annota-55 tions as input to those that allow users to interactively design 56 a 3D model through a series of interleaved sketching and nav-57 igation steps. Among them, a promising approach to sketch-58 ing biological shapes is using implicit surfaces, well known for 59 their ability to represent smooth, organic-looking 3D shapes. 60 Bernhardt et al. proposed in Matisse [2] a general solution 61 for the progressive sketching of such implicit, organic-looking 62 shapes: they compute the medial axes of 2D regions succes-63 sively painted by the user under different viewpoints to use them 64 as skeletons and generate 3D implicit shape parts progressively 65 blended to form a single shape. Zanni et al. later improved this 66 method with Scale Invariant Integral Surfaces (SCALIS)) [3] 67 to capture fine shape details. Less general solutions were ex-68 plored to represent specific biological structures such as muscu-69 loskeletal structures [9] or vascular systems [10, 11, 12]. While 70 it relies on sketching conventions used for teaching anatomy, 71 the method of Pihuit et al. [10] is limited to a single sketch in-72 put representing a network of vessels. Closer to our goals is 73 the work by Saalfeld et al. [11, 12] presenting interactive sys-74 tems aiming at creating and editing vascular systems to explain 75 pathologies. While their first method [11] was restricted to 2D 76 representation and relied on fluid simulation for the blood flow, 77

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their second solution [12] was semi-immersive but required significant user intervention to adjust the weights of the Metaballs composing the vessel and to define blood flows.

In this work, we reuse the general solution from [2, 3] for building volumetric shapes from a sketch. 5

Synthesizing shape distributions from a sketch. Although extensive research focused on example-based texture synthesis [13], only a few methods tackled the synthesis of discrete vector textures. We focus on the latter, as the most likely to be extendable to the sketch-based creation of 3D shape distri-10 butions such as fiber networks in a volume representing some 11 human tissue. Note that all the work discussed here only tack-12 led 2D distribution synthesis. Moreover, while they showed 13 promising results for texture synthesis, deep learning meth-14 ods [14, 15, 16] are primarily image-based and have not yet 15 been generalized to handle vector shape distributions effec-16 tively. 17

Landes et al. [17] was the first one to extend example-18 based vector texture synthesis, so far restricted to point dis-19 tributions [18, 19, 20, 21], to the synthesis of shape arrange-20 ments. They simplify the input shapes into proxy geometries 21 before computing spatial relationships between them, such as 22 distances and relative orientations. Then, they use this anal-23 vsis to synthesize a visually similar output texture. However, 24 their method is limited to bounded shapes, in addition to being 25 not real-time, which would make the creation of fiber networks 26 difficult. Subsequently, Roveri et al. [22] presented an example-27 based distribution synthesis method handling both bounded and 28 unbounded shapes. Their model decomposes shapes into point 29 samples encoded in a functional representation and relies on 30 neighborhood matching and energy optimization for the synthe-31 sis. Like other neighborhood-based texture synthesis methods, 32 their approach requires input patterns with a sufficient amount 33 of repetitions. Lastly, Olivier et al. [23] proposed a method for 34 extending a 2D sketch representing a pattern-based vector tex-35 ture while keeping the part drawn by the user unchanged. Their 36 method constructs a hierarchy of support structures used to cap-37 ture and reproduce multi-scale alignments in real time. 38

Our solution builds on the work of Olivier et al. [23], en-39 abling us to achieve the 3D design of arbitrary shape distributions (including anisotropic distributions and unbounded 41 shapes) in real time. We extend the former method to a new 42 problem: the generation of 3D distributions from a single, 2D 43 input sketch. 44

Sketch-based animation. Using sketches to design animations 45 has already been investigated in the context of scientific il-46 lustrations, mostly to depict fluids. Zhu et al.[24] relied on 47 sketches, manipulation, and fluid simulation to design an inter-48 active method for animating fluid systems, enabling visualiza-49 tion and conveying information about cardiac abnormalities or 50 surgical interventions. Kazi et al. proposed Draco [25] an inter-51 active system to create animated illustrations that combine data 52 samples and motion properties using kinetic textures. This was 53 later extended by Kitty [26], allowing the specification of func-54 tional relationships between entities. Milliez et al. introduced 55 Hierarchical Motion Brushes [27] to brush animated contents 56

onto a 3D scene, supporting coarse-to-fine animation thanks to 57 content reuse at different levels of detail. However, it required some precomputation and lacked methods for processing distributions, which the authors later addressed using hierarchical 60 spatio-temporal clustering [28]. Closer to our concerns, the Energy Brushes system [29] focuses on passive, secondary mo-62 tion. Users generate local energy patterns by sketching strokes and defining flow particles. The energy brushes represent the 64 direction of energy and forces that are continuously emitting particles along the sketched trajectory.

As in Draco, Kitty, and Energy Brushes, our system enables users to draw arrows to define velocity fields while we employ gradient triangles to specify force fields. We share the idea of continuously emitting elements similar to their flow particles. As in Energy Brushes, these elements can sketched by the user, but in contrast, we augment arrows by morphing and implicit deformers to capture more general types of deformations.

Storytelling in biology. Biological visualization was exten-74 sively studied for many years, including for illustrative abstrac-75 tion [30] and multi-scale visualization [31]. Subsequent work 76 explored context-based visual preferences of static data [32, 33] 77 and the importance of animated data relative to static one, 78 as in [34] for complex bio-molecular configurations. In par-79 allel, the dynamic nature of biological phenomena triggered 80 a strong interest in interfaces focused on narrative design, 81 whether for general data [35, 36, 37, 38], medical visualiza-82 tion [39, 40, 41, 42], or the narrative illustration of molecular 83 phenomena [43, 44]. More recently, a few software specifically 84 targeted this problem. CellPaint [45, 46] is a user-friendly in-85 teractive digital tool to create illustrations of molecular struc-86 tures such as cells and viruses. Molecumentary [47] enables 87 the asynchronous communication of design ideas by control-88 ling mesoscale animations of 3D models. 80

The work we present here is complementary: rather than reusing existing 3D animated content in the context of narration and addressing their differences of scale, we target the interactive, sketch-based creation of animated 3D content to support new narration ideas. In contrast to previous work, all narrative steps do not need to be known beforehand. Instead, we aim to create an interactive tool enabling biologists to illustrate their vision of their phenomena of interest and explore new narrative options.

### 3. Biological Context and Design Choices

#### 3.1. Preliminary study with biology researchers

To better understand how biologists represent and communi-101 cate the phenomenon they study, we conducted a preliminary 102 study with four cell biology researchers specialized in differ-103 ent subtopics. The participants included: a Research Director 104 with 35 years of experience in cancer research and experimen-105 tal therapeutics, a Professor with 20 years of expertise in cell 106 mechanics, another Research Director with 26 years of experi-107 ence in cell polarity and division, a Research Director holding 108 38 years of expertise in membrane and cytoskeleton dynamics, 109 particularly within the context of breast cancer. They explained 110

to us in simple terms the main steps of their studies, employing visual analogies or visual vocabulary when needed, (e.g., "cut-2 ting similarly to scissors", "cells hooking onto collagen fibers 3 to squeeze through"). They also highlighted the characteristics 4 of their phenomenon of interest in terms of shape types, behav-5 iors, and constraints. We observed that they all presented their 6 studied phenomenon from a narrative angle. In this narrative 7 framework, an initial animated environment undergoes evolu-8 tionary changes in response to specific triggering events. De-9 spite clear oral explanations and some sketches or 2D figures as 10 visual aids (see Fig. 2), there was a notable lack of visual sup-11 port to effectively represent the dynamic events they described. 12 This study highlighted the inadequacy of using only static 2D 13 figures for biologists, as they fail to capture the dynamic, 3D 14 nature of phenomena aligned with their mental vision. This ob-15 servation served as the driving force for the foundation design 16 of the framework we propose. 17

# Transmigration Chemotaxis Polarized PMN Directional sensing IL-8, fMLF.

Fig. 2: Examples of 2D illustrations in cell biology that emphasize the use of schematic visual elements: on the left, ( [48]), arrows illustrating the trajectory of a cell; in the top right corner, ([49]), streamlines depicting the flow within a vessel; in the bottom right corner, ([50]), a blue gradient triangle indicating the attraction of cells outside the vessel.

#### 3.2. Schematic visual vocabulary in cell-biology 18

Our biologist collaborator enriched this study by formalizing 19 the schematic vocabulary frequently used in biological illustra-20 tions to describe dynamic phenomena (see Fig. 2): 21

Arrows (left) depict the kinematic trajectory of an element, 22 giving both direction and speed of displacement; 23

Streamlines (top right) indicate flow within a vessel using 24 three arrows followed by a straight line for homogeneous 25 flows or by a curved line for non-uniform flows (for exam-26 ple, allowing for a lower speed near the vessel membrane 27 due to friction); 28

- Gradient triangles (bottom right) are used to represent the 29 difference of gradient in a medium, (e.g., pressure gradient 30
- or chemical gradient) denoting the attraction of elements. 31

#### 3.3. Findings and design goals for DynBioSketch

Building upon this comprehensive preliminary study and the accompanying illustrations, we have categorized the types of 3D modeling tools, animation mechanisms, and rendering techniques to address in the development of our interactive authoring tool:

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- Variety of shapes observed: volumetric elements such as organic shapes (e.g., vessel, cells, and their interiors); larger surfaces or volumes, particularly those unbounded at the observation scale, (e.g., skin); and linear shapes like fibers. We noticed that organ surface representation often requires the inclusion of textures, such as Voronoi-type tessellations for cell boundaries.
- Shape distributions: applicable to unbounded shapes (like a field of fibers) and nested bounded ones (like cells within a vessel), which also underlines the need for models capable of extrapolating a user-sketched input into a 3D distribution.
- Animation: need to sketch global flows, individual trajectories, and control shape deformations. In biological representation, precise control of animation synchronization is essential for building narrative scenarios. Sequences of triggering events can define such scenarios and effectively convey the dynamics of the phenomenon.
- Expressive rendering: based on transparency and local shape cuts to effectively represent nested environments and movements within the current element.

#### 3.4. Methodology and technical contributions

Drawing inspirations from visual representation and narrative techniques in biology, we conceptualized DynBioSketch, as a new creative medium that seamlessly integrates the progressive modeling of a 3D biological environment with the development of animated narratives. In this interactive sketchbased modeling framework, biologists can convey their understanding of a phenomenon. Additionally, this innovative approach serves as an accessible entry point for non-expert users to explore biological phenomena. In both cases, users initiate the process by designing an initial, stable environment and subsequently triggering its temporal transformation. Let us note that by a stable 3D environment in an animated interface, we refer to a scene configuration that remains reliable and consistent while being animated or not. This setup allows users to iterate and refine environment designs or animation ideas quickly using simple sketches.

As illustrated in Fig. 3, we centered this work around the infection phenomenon. In implementing our prototype, we focused on the most important of our design objectives, aim-78 ing to achieve the right balance between simplicity and richness of representation. We proposed solutions that enable users 80 not only to model individual 3D shapes from a sketch but also 81 to express bounded and unbounded distributions of 3D shapes 82 (Sec. 4). In line with this approach, our sketch-based anima-83 tion mechanisms go beyond rigid motion, facilitating the spec-84 ification of deformations. This functionality is complemented

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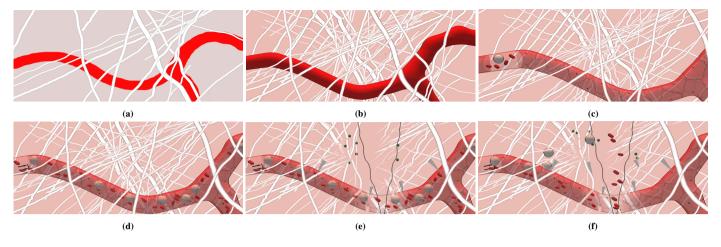


Fig. 3: Overview of DynBioSketch: from creating the 3D biological environment (a-c) to designing a dynamic scenario (d-f).

by a timeline that allows users to navigate and trigger narrative
events (see Sec. 5). Throughout the creative process, users benefit from expressive visualization thanks to the combination of
3D shading, contouring, transparency, and textures (see Sec. 6).
We have designed all the methods mentioned above as real-time
solutions, which is mandatory for their seamless integration into
an interactive authoring tool.

*Case study.* To highlight the usability of our system to create an animated biological story, we selected the bacterial infection phenomenon as our case study: this phenomenon is initiated 10 by a cut in the tissue, impacting the skin, the underlying fibers, 11 and the vascular network, which causes blood cells near the 12 cut to flow through it and allows external elements like bacte-13 ria to enter the environment. Consequently, white cells, as key 14 players in the immune response, are activated to migrate from 15 the vessel to the cut area, contributing to the wound healing 16 process. This migration involves their initial attraction to the 17 vessel's inner membrane, followed by a rolling motion along 18 this endothelium. Ultimately, such cells escape by undergoing 19 shape deformation to pass through small openings between the 20 endothelial cells of the membrane. We selected this particular 21 example to exemplify our technical contributions in this arti-22 cle, as it features an illustration by a professional artist. Biol-23 ogists have also validated this illustration, establishing it as an 24 authoritative visual reference that showcases the capabilities of 25 our system. Fig. 3 provides a comprehensive overview depict-26 ing: (a) the input 2D sketch, (b) the resulting 3D environment, 27 (c) an illustrative example showcasing cell distribution within a 28 vessel, along with the rendering of its surface, (d) the resultant 29 distribution of cells and the use of the flow tool to specify their 30 movement, (e) the cutting of tissues, triggering an infection re-31 sulting in the arrival of bacterial cells (in green) and white cell 32 activation using gradient triangles and (f) the deformation of 33 white cells as they cross the vascular membrane, attracted by 34 the bacteria, while red cells escape through the cut. 35

## 4. Creation of a 3D biological environment

This section introduces three new tools that enable users to progressively create a stable 3D environment, in which the targeted biological phenomenon will later be triggered. Although stable over time, the 3D environment can include animated elements, such as blood cells following a stationary flow predefined within a vessel.

### 4.1. Volumetric background tissues from a 2D sketch

Our inspiration figure (see Fig.1, left) emphasizes that biological environments usually feature distributions of 3D elements in the background (e.g., the field of collagen fibers around the vessel in our case study).

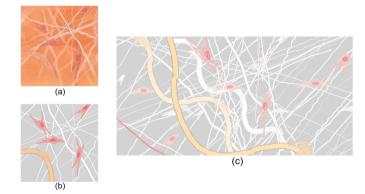
Our goal is to generate a 3D distribution of elements from a 2D input sketch, allowing users to navigate within this generated 3D environment. Our concept involves allowing users to sketch a small representative sample of what they want to visualize from the front. Subsequently, we extend this sample in 2D by a given ratio k before immersing the expanded 2D distribution in 3D by introducing an extended depth of h. The resulting 3D volume of material can be replicated with adjusted rendering to simulate an infinite environment.

The 2D expansion of the user-sketched vector texture follows 57 the methodology outlined by Olivier et al. [23]. This approach 58 efficiently computes a larger 2D distribution of strokes through 59 a multi-resolution alignment analysis, seamlessly extending the 60 initially sketched sample. Specifically, vector sketched strokes are efficiently organized into a hierarchy of linear or curved 62 support structures, each characterized by a main direction and a ribbon containing clustered sub-structures. During synthesis, 6/ the existing supports in the exemplar are progressively extended and duplicated from top to bottom of the hierarchy, incorporat-66 ing some randomization to align with the larger output space. 67 Clones of the input strokes are ultimately distributed, with suit-68 able jittering, along the lowest-level structures (refer to [23]).

In the second stage, we determine depth parameters for each 70 stroke to expand the generated 2D vector texture into a 3D dis-71 tribution. Our approach relies on a common perceptual assump-72 tion regarding general views: the alignment of strokes in the 2D 73 exemplar intentionally represents a 3D alignment rather than 74 arising solely from the projection of a specific viewpoint. Con-75 sequently, we immerse clustered strokes with similar depths 76 through the following two steps. 77

For each cluster, we initially define a midpoint's depth  $d_e$ and a slope angle  $\delta$  relative to the sketching plane (*XY*). These parameters are selected randomly within the ranges of [0, *h*] for  $d_e$  (where *h* is the depth of the output domain) and  $[-\pi/4, \pi/4]$ for the slope (a range informed by perceptual studies [51]). For more details, we let the user refer to the model of Olivier et al. [23].

To simulate an infinite field of 3D strokes, duplicating the 8 resulting volume along the depth axis is crucial. We configure 9 these duplicated instances to overlap randomly in depth, en-10 abling intricate arrangements. To achieve this, we replicate the 11 initial volume and shift each instance in the depth direction by 12 a factor of  $l \cdot h$ , where l is an integer in the range of  $[1, \infty]$ . 13 This process generates a virtually infinite field of anisotropic 14 elements. The visualization of these elements is guided by two 15 perceptual assumptions. Firstly, distant objects gradually fade 16 away, implying that strokes in deeper layers appear less promi-17 nent. Secondly, we conceptualize the 3D space with a toroidal 18 topology in depth, meaning that objects exceeding a certain 19 depth "wrap-around" and reappear at the front. By incorporat-20 ing these perceptual assumptions, we can effectively represent 21 multiple layers and create a visually coherent volumetric dis-22 play (see Fig. 4). 23



**Fig. 4:** (a) Depiction of cells navigating through a 3D arrangement of fibers in a biological context; (b) 2D sketch input inspired by (a); (c) Output generated by our method for synthesizing the 3D fiber and shape distribution.

#### 24 4.2. Sketch-based modeling of nested shapes distributions

Since we target the creation of organic shapes such as ves-25 sels, cells, and internal cellular structures, we rely on an ex-26 isting sketch-based implicit modeling methodology, borrowed 27 from Matisse [2] to ease their creation: users alternate naviga-28 tion and painting 2D regions in a plane facing the camera. The 29 medial axis of each painted region is extracted and simplified 30 into a graph of segments, augmented with radii information. 31 This graph serves as a skeleton to generate a scalar field mod-32 eling a density of matter around it and progressively decreasing 33 with distance. A specific isosurface defines the target implicit 34 surface. Note that while each sketch results in a flat skeleton, 35 and thus an implicit surface with a flat silhouette, sketching 36 such shape parts from different viewpoints and then blending 37 their fields through summation allows the creation of arbitrary 38 3D shapes. In our implementation, we use the field defined in SCALIS [3] to better match the sketched contours than in the original Matisse system.

In addition to sketching isolated organic shapes, the user is invited to sketch distributions of sub-shapes nested within a larger one (see Fig. 5). This can be useful, for instance, to populate a vessel with blood cells. To this end, we developed a second extension of the method of Olivier et al. [23] dedicated to distributions of bounded shapes within a container.

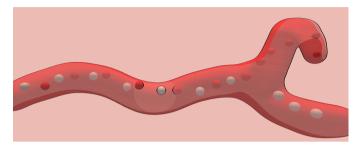


Fig. 5: From an exemplar to a distribution of cells in a 3D vessel.

The user first selects as *container* any volumetric implicit shape previously generated from a sketch. Using a camera facing the plane where a flat part of the skeleton is defined, the user selects a specific section of the container (made transparent) and creates a sample of the desired distribution. This can be done by directly sketching sub-shapes within the container or by instantiating some existing shapes. Note that this input distribution exemplar should only include bounded shapes entirely within the container. We then propose a 2.5D solution for analyzing the exemplar and extending it within the entire container: we analyze the exemplar in the skeletal plane by taking into account the 2D projections of the sub shapes, and after distribution synthesis in 2D, the depth of the newly generated sub-shapes is computed from the original depths in the exemplar, as we detail in the following paragraphs.

Adapting the original 2D distribution analysis and synthesis method from Olivier et al. [23] to free-form planar regions is challenging.

As the synthesis is limited to the interior of the free-form container, we cannot directly extract shape alignments and their directions by a successive grouping of straight lines. However, the scalar field defining the implicit container is an excellent basis for grouping. In our case, we call "quasi-aligned" the shapes positioned around the same isovalue and we define their support structure by the corresponding isoline curve in the scalar field rather than by a straight line. As in the original method, support structures are grouped recursively, with ribbons expressing the variance of the elements they represent. For that, we use the difference of isovalues as clustering distance. These simple adjustments to the original method allow us to apply the existing algorithms to analyze the free-form input sample and then extend the distribution to the full 2D projection of the container shape.

It is important to note that isolines do not require explicit computation during the synthesis phase. Instead, we iteratively instantiate the next shape to generate along a given isoline using a series of small displacements from the previous position. Each step involves a translation along the tangent direction fol-

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lowed by a projection along the field gradient back to the target
isovalue. We define the tangent direction as the cross-product
between the field gradient and the depth direction, and the step
size by the local curvature. In addition, a slight perturbation
is introduced in the new position to avoid significant repetition
during the synthesis phase. After extending the planar distribution to the entire container, the generated clones for each subshapes return to their original depth, with some extra jittering,
which allows us to disperse them throughout the 3D container.

We expanded our synthesis model into 3D to accommodate 10 more intricate containers. The primary challenge lies in manag-11 ing the transition in the depth direction when moving between 12 different 3D branches. To tackle this issue, we leverage our definition of trajectories (detailed in Section 4.3) along the isoline 14 direction, which enables us to calculate the subsequent posi-15 tions in a container, where the generated clones follow a spiral-16 ing path around an isosurface. To introduce additional variation 17 to this trajectory, we analyze the local cross-section of the iso-18 surface surrounding the clone's position and rotate this position 19 around the cross-section, introducing variations to the trajec-20 tory. The degree of rotation is determined randomly within pre-21 defined angle ranges. The position of the clone shape serves as 22 the basis for defining its local frame and the direction of its iso-23 line, which in turn determines the subsequent position for the 24 next (clone) shape to create. 25

Figure 5 illustrates the method's robustness in generating distributions of sub-shapes within containers with 3D branches,
facilitated by the implicitly defined isosurfaces.

#### 29 4.3. Creating stationary animations

The last step in creating a representative biological environ-30 ment is to enable animation. We focus here on background 31 motion that remains stationary over time, as opposed to the 32 unique animated events discussed in Section 5.3. The sketch-33 based tools presented next enable users to define velocity fields 34 that guide the motion of sub-shapes within a container, such as 35 blood particles animated within a vessel. To this end, we use 36 the flow to directly set the velocity of kinematic sub-shapes, al-37 though using it to exert viscous frictional forces on physically-38 based objects would also be straightforward. 39

In the following, we take the hypothesis of elongated contain-40 ers such as blood vessels. Implicit shapes being closed surfaces, 41 we truncate the extremities of such a container in the flow di-42 rection and use a toroidal topology to allow the sub-shapes that 43 reach the upstream extremity to be re-generated downstream. 44 During the animation, we detect collisions with the container 45 and re-project sub-shapes in the field's local gradient direction 46 to prevent unwanted leakage of sub-shapes across the container 47 surface. Note that collision detection and response between sub-shapes can be neglected, given that they follow the same 49 velocity field and are usually located at different depths. 50

To allow comparisons between 2.5D and 3D illustrations in the user study, we developed two alternative versions of the flow animation system, presented next: the first one is specific for flat-silhouette containers, which were sketched from a single viewpoint and therefore have a planar skeleton, enabling us to use a planar flow; whereas the second focuses on more general flows on 3D arbitrary containers that may bend out of the plane facing the camera.

3-Arrow flow tool for 2.5D illustrations. In the following, we call **2.5D container** the input implicit container drawn in a plane facing the camera, and f the field used to define it. To create a 2.5D animated illustration, we compute a 2D flow used to animate the planar coordinates of the sub-shapes within the container while their depth is constant.

We inspire from conventions in biological illustrations to propose a fast way to design a global flow spanning the whole container: the user sketches three arrows to indicate the desired direction and strength of the flow and adds either a straight or a curved line near their extremities indicating a possible reduction of velocity, mimicking frictional effects, near the container's surface (see Fig. 6).

To extend this locally-defined flow to a velocity field span-72 ning the entire container, we take the following hypothesis: we 73 orient the flow along the isolines of the container's field f, with 74 a constant velocity along each isoline. Moreover, although the 75 sketched flow may be small relative to the size of the container, 76 we assume that the flow extends over its entire cross section, 77 with a maximal value at the middle and a minimal one close to 78 the surface, which enables us to define the flow as follows. We 79 extract the flow maximum velocity and lateral damping from 80 the user's sketch. We then compute the norm of the velocity 81 field from changes of isovalues of f within the container, as il-82 lustrated in Figure 6. We also register the orientation of the flow 83 relative to the container's skeleton by projecting the central arrow to the closest central skeletal segment. The local velocity 85 at any position P within the container is then defined as a vector perpendicular to the local gradient direction, oriented as to 87 agree with the flow direction propagated along the skeleton, using the norm registered for the isovaluee f(P) of the flow. 80

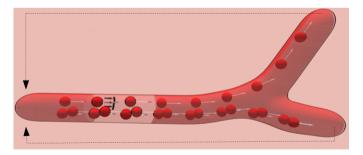


Fig. 6: Use of the triple arrows tool (in black, within the vessel) to create a velocity field within a 2.5D container. The latter, a closed implicit surface, is given a toroidal topology to bring back downstream (long arrows) all the objects advected beyond a predefined upstream limit.

*Frame-based flow tool for 2.5D and 3D illustrations.* To enable the animation of sub-shapes within arbitrary 3D containers in addition to 2.5D containers, we provide a second method for defining stationary flows. We address the lack of parameterization of implicit surfaces by leveraging the local frames defined by the field gradient direction, tangent, and binormal of the container's isosurfaces. Note that contrary to the 3-arrow flow tool, the frame-based tool is not limited to the design of a flow that

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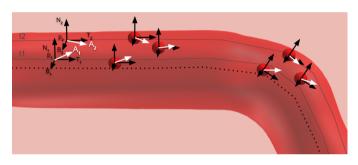
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follows the container's isosurfaces since we can define locally contractive flows that guide the sub-shapes closer to the con-2 tainer's skeleton or dispersive flows that push them towards its 3 outer surface, as well as a combination of both.

The input is a 3D container, implicitly defined as an isosur-5 face of a field f, within which the user has defined sub-shapes. The user then uses an arrow tool to sketch one or several samples of flow velocity vectors within the container shape in a 8 plane facing the current camera:

- If the user sketches only one single arrow, the latter is used 10 to define a steady flow field within the whole container; 11
  - Otherwise, the arrows are interpreted as flow velocity samples at different isovalues, enabling linear interpolation in between (see Figure 7).



**Fig. 7:** The user sketches two arrows, respectively  $A_1$  and  $A_2$  (in white), to specify a flow with dispersive behavior at the center and compressive behavior near the surface of a container. Both arrows indicate a motion to the right at different speeds. We store each arrow  $A_i$  in the local frame defined by the tangent  $(T_i)$ , the normal  $(N_i)$ , and the binormal  $(B_i)$ . These arrows are combined to define velocity vectors (white) anywhere in the container (see the red subshapes), using interpolation according to the local isovalue.

An arrow  $\mathbf{A} = (A_x, A_y, A_z)$  sketched at a point P, is decom-15 posed into local coordinates  $(V_T, V_N, V_B)$  inside a local frame 16 (T, N, B) aligned with the isosurface passing through P, and 17 computed as follows: N is the normal to the local isosurface 18 computed from the gradient of f, B is binormal to the isosur-19 face defined as the unit vector pointing to the camera,  $T = N \times B$ 20 is a unit tangent vector to the isosurface. Therefore,  $V_T$  defines 21 the speed of motion along the isosurface of isovalue f(P), while 22  $V_N$  defines the speed of motion in the field gradient direction, 23 enabling the definition of more general flows such as dispersive 24 or contractive ones. 25

In the case of a unique arrow, we reuse the coordinates 26  $(V_T, V_N, V_B)$  to set the local velocity vector at any point P within 27 the container's shape while being interpreted in a new local 28 frame (see the black arrows near sub-shapes in Figure 7). Con-29 versely, if the user sketched several velocity vectors  $\vec{A_i}$  at differ-30 ent points  $P_i$ , each vector is decomposed as before and attached 31 to the isovalue  $f(P_i)$ . This allows for the linear interpolation 32 of new velocity vectors in local coordinates. Let P a point in-33 side the container, such that  $f(P) \in [f(P_i), f(P_i)]$ , we define 34  $\lambda(P) \in [0, 1]$  and then  $\vec{A}(P)$  as: 35

$$\lambda(P) = \frac{f(P) - f(P_i)}{f(P_j) - f(P_i)}$$
$$\vec{A}(P) = (1 - \lambda(P))\vec{A}_i + \lambda(P)\vec{A}_j$$

For points outside the range of field values of the sketched velocity samples, we use the nearest sample to compute A(P).

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During the animation, the sub-shapes' motion is guided by the flow computed using these interpolated velocity vectors in local-frame coordinates and then re-expressing them in the world frame given the new local frame (T,N,B)—which may have rotated-at the current sub shape's position. The subshape moves along the resulting velocity direction. Note that in the case of flow specified to remain parallel to the isosurfaces ( $V_N = 0$  everywhere), precision can increase by reprojecting each sub-shape, at each timestep, to their initial isosurface along the local normal direction.

Cyclic-frames versus Flow-frames:. Computing consistent local frames during animation is essential for the above method to work efficiently.

Upon creation, we initialize the sub-shape's local frame at its centroid by computing (T, N, B) as above, using the camera's view direction for setting B. At each time step i, the sub-shape is moved to its new position  $P_i$ , computed from the local frame and interpolated flow velocity at  $P_{i-1}$ . We then use the new normal direction and the previous binormal direction to update the local frame:

$$\vec{N}_{i}(\mathbf{x}) = -\vec{\nabla}f(\mathbf{x})$$
$$\vec{T}_{i}(\mathbf{x}) = \vec{N}_{i}(\mathbf{x}) \times \vec{B}_{i-1}(\mathbf{x})$$
$$\vec{B}_{i}(\mathbf{x}) = \vec{T}_{i}(\mathbf{x}) \times \vec{N}_{i}(\mathbf{x})$$

This algorithm directly generates *cyclic-frames*, i.e., frames 58 used to compute periodic motions of sub-shapes turning within 59 a container, e.g., along isosurfaces. Indeed, the normal vector 60 always points outwards, so even a global flow defined using a 61 single arrow is interpreted as two different orientations at the 62 top and bottom of the container. To define directional flows 63 spanning the entire cross-section of the container (such as the 64 flows of blood cells within a vessel), the local frame must be 65 transformed into aa flow-frame, as follows: any sub-shape ini-66 tialized below the centroid of the container in the flat drawing 67 region facing the camera has its local frame defined using -N68 instead of N, which in turn orients T back to the consistent flow 69 direction. 70

See Figure 8 and the second supplemental video for a visual comparison between animations created using cyclic vs. flow frames. Our method ensures temporal coherence. It is robust to containers of arbitrary topological genus (top), possibly with 74 branchings (bottom), including 3D containers drawn from different viewpoints, which lack a fully connected skeleton. Note that due to the recomputation of local frames, sub-shapes typically follow a twisting motion within such containers, as if they were forced back in by a collision with the outer surface. In our experiments, this was considered a positive feature of the 80 method. 81

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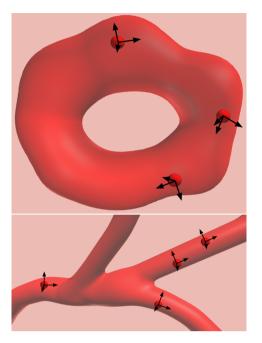


Fig. 8: We provide two categories of local frames: cyclic frames (top) defined by the local normal direction and enabling the generation of cyclic motions along isosurfaces, versus flow-frames (bottom) where the normal axis is flipped (i.e., point to the inside instead of the outside) for particles initialized below the center-line of the container. Then, all the particles passing through a given cross-section move in the same direction.

#### 5. Design of the main biological phenomenon

At this stage, the users have created the initial, stable animated 3D environment as a basis for the phenomenon they want to represent. Users can now design their phenomena of interest by placing triggering events along a timeline and sketching the resulting consequences in the 3D scene.

#### 5.1. Triggering events

In our scenario, the triggering event is the incision made in the environment, which affects the tissue, fibers field, and vessel network. We maintain real-time reactivity after the cut by us-10 ing expressive rendering techniquess (See Sec. 6 for the imple-11 mentation details). Such incision disrupts blood flow as neigh-12 boring cells migrate through it. We describe this behavior by 13 assigning cells an elliptical trajectory towards the center of the 14 incision, followed by random oscillations as they leave the en-15 vironment through the cut. Moreover, bacteria can emerge in 16 the environment. Users can introduce new bacterial shapes us-17 ing our Organic Shape Creation tool (see Sec.4). These shapes 18 exhibit localized random movement, reinforcing their living ap-19 pearance. Such a triggering event is marked on the timeline as 20 an important event and is essential for narrative scenarios (see 21 Sec. 5.3). 22

#### 5.2. Expressing the consequences 23

Setting new events in the narrative dynamic scenario implies 24 some consequences on the environment that need to be modeled 25 and integrated into our system and along the timeline. We let 26 users specify the following consequences: 27

Sketching motion. We propose a series of sketch-based anima-28 tion tools to let users express the consequences of this event on the environment. To seamlessly integrate animation design into 20 our sketch-based creative system, we draw on visual vocabulary derived from the analysis of biological illustrations (see 32 Section 3). This approach enables us to establish a direct link 33 between the familiar schematic representations in biology and 34 the motion or deformation predicted in our system. While de-35 scriptions are initially provided in the plane facing the camera 36 and applied to shapes at a constant depth, these movements and 37 deformations can extend to a full 3D animation. We introduce motion design tools from different viewpoints for users to com-39 bine their effects. We propose two sketch-based animation tools 40 to express such consequences: arrow and gradient triangles. 41

The user initiates a trajectory by sketching a single arrow from the designated target object (see Fig. 9). We meticulously 43 sample this trajectory based on the drawing time, ensuring that the speed of the gesture directly influences the speed of the sub-45 sequent movement. The displacement between two consecutive points along the timeline is then incorporated to streamline the 47 synchronization of various motions. This approach enhances the fluidity and coherence of the animation as a whole.

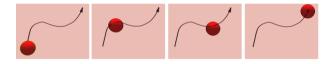


Fig. 9: The cell is following the user's arrow.

On the other hand, the user can position gradient triangles 50 in the 2D plane facing the camera to specify directional force fields due to chemical gradients (see Fig. 1, right). These forces 52 are applied to neighboring shapes of a given category with their 53 intensity decreasing with distance. In our case study, we use such gradients to model the chemical procedure that attracts 55 white blood cells to the site of infection. Additionally, we intro-56 duced a novel representation of the attractive forces exerted by 57 cells escaping the vessel toward bacteria using an implicit type 58 of gradient triangle. We chose an ellipse-based path directed 59 toward the nearest available bacteria to model these forces. 60

Specifying shape deformation. Enabling shape deformation 61 and potentially altering their topology through fusion or fis-62 sion poses a significant challenge in biological animation. For example, in our specific case study, the deformation of white 64 blood cells is essential to let them navigate tight spaces between vessel membrane cells and thread their way through complex 66 fibers. To meet this challenge, we have designed two mech-67 anisms that allow users to precisely specify and control the 68 desired deformations: shape morphing and shape deformation 69 through a hole. 70

For shape morphing, we employ sketch-based modeling to define the initial and final shapes, positioning them at specific moments along the timeline. To generate the intermediate shapes, we utilize a morphing algorithm based on Minkowski sums [52], as illustrated in Fig. 10.

The alternative model proposed here is specific to shape deformation through a hole. Considering a target shape T tasked

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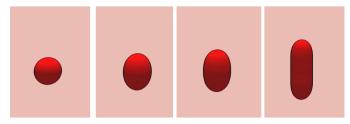


Fig. 10: Morphing between the user-specified initial (left) and final (right) states.

- with traversing a hole H within a surface S. Let  $R_t$  and  $R_h$  be the radii of the target and the hole-bounding spheres, respectively. We seek to model the following behavior:
- <sup>3</sup> we seek to model the following behavior.
- If  $R_t < R_h$ , the target escapes without undergoing any deformation.
- If  $R_t > 2R_h$ , excessive compression of the target will occur, rendering it unable to escape.
- If none of the above conditions apply, the target will deform while retaining an almost constant volume.

In our system, the target cell T and the surface S (a vessel) 10 are implicit surfaces. The hole is an implicit primitive generated 11 by a negative field created from a skeleton point. To simulate 12 the impact of passing through the hole on the shape of the tar-13 get, which must compress and elongate before returning to its 14 original shape on the other side of the surface, we introduce a 15 deformation field in addition to the one that defines T, as fol-16 17 lows.

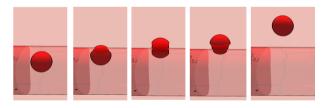


Fig. 11: Cell deforming to cross a membrane through a hole.

We position a deformation primitive on top of the hole, di-18 19 viding its region of influence into three distinct zones: an outer ring, a central ring, and the remaining spherical zone around 20 the center. In the outer ring, we apply a negative field to lo-21 cally compress the target shape and ensure it fits correctly into 22 the available space of the hole. In the central ring (correspond-23 ing to the size of the hole), a positive field is used to preserve 24 the volume of the target during deformation. In this way, the 25 target's shape stretches along its trajectory as it approaches the 26 hole and the same applies when it exits from the other side. We 27 set the deformation field to zero in the spherical inner region. 28

Fig. 11 illustrates the impact of applying this deformation primitive to a passing target. In our specific case study, this approach allows us to efficiently simulate the behavior of white blood cells crossing the vessel membrane.



Fig. 12: Illustration of a narrative scenario in DynBioSketch: the user sketched a trajectory arrow that saved an event at time 4.8 and returned to this event to replay the displacement.

#### 5.3. Narrative scenarios

As shown in Fig. 12, the interface of our creative system includes a toolbox and a timeline. When the user launches an animation, the timeline becomes active. We manage two categories of animation: 1. periodic animations that persist throughout the session, such as the flow in a container or the random movement of bacteria; 2. non-periodic animations triggered by specific events (for example, the appearance of a chemical gradient due to a cut) are the only events displayed on the timeline, as are the animations that follow them.

Our system offers two modes: 1. creation mode, which lets users navigate and create or modify shapes and generate animations and position-triggering events on the timeline; 2. playback mode that lets users view the current animated DynBioSketch from a specific point in time.

When instantiating a new event, we establish an event state. This state records both the time of the event and the current state of the elements in the environment. When the user returns to this event, we reuse these saved states. Fig. 12 illustrates an example at time 4.8, where the user uses the arrow tool to draw a white blood cell out of a vessel through a hole. Switching from one mode to the other enables users, especially biologists, to systematically illustrate the desired phenomenon and to design and record an associated narrative if needed.

#### 6. User interface and Visualization choices

#### 6.1. Interface of DynBioSketch

In addition to a timeline located below the main window, we provide the user with a main design interface at the left, composed of 14 widgets, and described below from top to bottom and left to right (see Figure 13):

- Pencil: A pencil tool to draw annotations;
- Letter A: Annotation tool to create a text box for annotations; 64
- Brush: A 2D painting tool to create or edit organic shapes;
- Scissors: A cutting tool to virtually cut the membrane of a shape, enabling one to see inside (cutting gesture within a shape) or to cut through the shape, as we did in Fig. 1, right to model the cut through tissues (cutting gesture from the outside, orthogonal to the shape); 71

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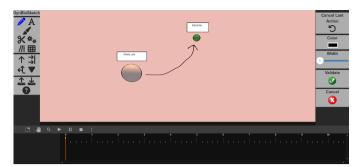


Fig. 13: The DynBioSketch's interface: the creation tools for 3D geometry and for sketching the volumetric environment are at the left of the main window (showing here with a white cell and an arrow to make it move to a bacteria and two annotations). The timeline for triggering events and playing the animation is at the bottom. The interface at the right contains standard tools such as setting the size and color of the brushes and undoing/redoing

- Duo of cells: Shape distribution tool to select a sample of distribution (a few elements such as cells) and then trigger the synthesis of the full distribution within the container containing these elements;
- Vertical lines: 2D sketch distribution tool to open a window for the 2D sketching of the environment texture (fibers in our main example) and then call for the synthesis of a 3D environment in an extended domain;
- Grid: Tesselation tool to sketch 2D black dots, used to define Voronoi regions to texture the membrane of an organic shape;

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- Single arrow: Trajectory arrow tool to define a trajectory arrow for an organic shape;
- Double arrow and vertical line: 3-arrow flow tool to sketch the 3-arrows flow used to animate 2.5D illustrations:
- Fork arrow: Frame-based flow tool to sketch the framebased flow used for 2.5D and 3D illustrations: 17
  - Triangle: Gradient triangle tool to create gradient triangles;
  - Upload: : Upload tool to retrieve an existing DynBioSketch model:
- Download: Download tool to save a DynBioSketch model; 22
- Help: Help tool to display a window with a reminder of 23 the different tools; 24

In addition to the modeling and animation tools we described 25 throughout this paper, the DynBioSketch interface allows users 26 to annotate their sketches by selecting the pencil tool from the 27 left menu and choosing the color and size of the stroke from the 28 right menu. Additionally, users can add text boxes and insert 29 descriptive text to explain what is happening in the scenario. 30

#### 6.2. Illustration-inspired rendering

The user benefits from an expressive, non-photorealistic rendering throughout the creation process. This rendering aims at facilitating the overall perception of the 3D scene and animated events, presenting the entire scene as a shaded 3D sketch reminiscent of the artistic illustration in Fig. 1 (left).

We render each shape generated by our sketch-based modeling system with a default material incorporating Phong shading with procedural value noise, resulting in a paper-like effect. In addition, we apply a black outline effect using a pen-and-ink shader (see the vessel and fibers in Fig. 1). This rendering approach is versatile and can be adapted to the requirements of the animation scenario.

Organic tesselation of surfaces. To give surfaces, such as vessels, the appearance of an organic membrane, we offer an option for texturing a surface with Voronoi-like regions representing the membrane cells. We marked these regions with black dots representing their so-called kernels.

To generate such textures, the user clicks at the desired positions to place the kernels on the selected surface. With each click, we use ray scattering from the camera's point of view to identify P1, the first intersection with the implicit surface, chosen as the kernel point. Instead of using the second ray-surface intersection point P2 as the kernel on the other side of the surface, we introduce randomness in the ray direction before calculating the next intersection point, starting with P1. This randomization avoids unrealistic cell alignments. Automatic point pattern generation techniques on surfaces may be used as well.

Once we position all the kernels, we calculate an approx-59 imate Voronoi diagram on the free-form surface, directly en-60 coded in the surface's geometric attributes to facilitate 3D nav-61 igation. As a first step, we classify the mesh's vertices accord-62 ing to their proximity to the nearest core. As a trade-off be-63 tween efficiency and precision, we chose Euclidean distance as 64 a rough approximation to geodesic distance. We identify and 65 display in black the closest vertex to each kernel and then tra-66 verse the mesh faces to locate triangles whose vertices are asso-67 ciated with three different kernels. These faces mark the corners 68 of the desired Voronoi regions. 69

We then introduce their barycenter, together with its interpolated normal, as a new vertex of the mesh. This vertex is used to tessellate the face into three sub-triangles, connecting it to the original vertices. Similarly, we add a new vertex to the barycenter of each edge between two vertices linked to distinct cores, inserting it into the mesh by dividing the triangle into two segments. This process produces distinct Voronoi-type regions delimited by dotted lines with a shape error proportional to the length of the triangles' edges in the input mesh (see Fig. 14).

Visualization of nested shapes. Thanks to the artistic inspiration of Fig. 1 (left), particularly in the representation of the ves-80 sel's local transparency to reveal the interior at the far left, we 81 have incorporated a similar approach to simplify the design of 82 nested shapes such as cells within a vessel. While all surfaces 83 are partially transparent by default, users can effectively "erase" 84 a section of the membrane with a brush or by intersecting it 85

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**Fig. 14:** Organic-looking tessellation of surfaces: (a) shape to texture; (b) user-selected kernels; (c) the resulting tessellation.

with a box. This action makes the interior fully visible, simplifying the sketching of sub-forms. In addition, as shown in
the accompanying video, we have simplified the phagocytosis
phenomenon by progressively reducing the transparency of the
bacteria, then of the white blood cell, which has "absorbed" the
bacteria, rather than precisely simulating the absorption itself.

*Faking a cut through tissues.* Modeling and animating a crosssection of biological tissue presents a significant challenge, particularly due to its anisotropic behavior influenced by fibers and
vessels. To tackle this challenge, we used a visualization trick
to symbolize a cut-through tissue (see Fig.15) associated with a
temporal event that triggers changes in animation modes.

We provide the user with a cutting tool inspired by a tradi-13 tional cutter to sketch a cutting trajectory in the 2D visualization 14 plane. Assuming that due to tissue tension, a "real" cut would 15 result in a triangular opening, we simplify the visualization by 16 deactivating the display of all elements inside a triangular prism 17 with an isosceles triangle as a basis (with an angle of  $\frac{\pi}{12}$  in our 18 implementation) whose vertex lies at the end of the cut trajec-19 tory. To do this, we register all shapes (implicit surfaces and 20 3D textures such as fibers) intersecting the user's stroke in the 21 projection. We then add other features in the triangular region 22 to this list, using ray scattering at grid-based sample points on 23 24 the triangular shape, letting us remove all parts projecting into the cut. 25

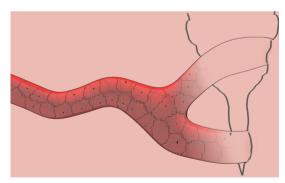


Fig. 15: Rendering effect faking a vertical cut through organs.

#### <sup>26</sup> 6.3. Visualization and navigation within stroke distributions

As mentioned in Sec. 4, most of the static 3D environment, which serves as a background, is constructed using 3D features. We use separate materials for their inner and outer areas for optimal visualization. The central inner area adopts the original color of the stroke, while the contour color changes smoothly from light gray to black, creating a seamless blend with a darker background. To enhance the three-dimensional appearance of elongated features such as fibers, we create a harmonious blend of inner zone and contour colors, giving the feature a cylindrical appearance.

In our implementation, we use a traveling zoom to move 37 seamlessly from the interactive sketching session, where the 38 user draws the input 2D vector texture to the exploration of 39 the output 3D scene. The volumetric feature distribution lay-40 ers are presented in each frame with increasing transparency 41 and decreasing feature thickness relative to camera distance. 42 This process is carried out stroke by stroke, depending on the 43 distance between the stroke center point and the camera. In 44 addition, we modify the color of distant strokes by converting 45 their RGB values to HSL format and adjusting the luminance 46 parameter from its original value to 1, thus transforming them 47 into white. When a stroke is located behind the camera, we 48 move it in depth towards the deepest visible volume, creating 49 the illusion of infinite space in depth. This approach delivers 50 superior visual results during exploration compared to applying 51 the same transformation to volumetric layers one by one while 52 maintaining real-time performance (see Fig. 4). 53

#### 7. Results & Validation

**Performances.** We implemented our prototype system in WebGL and generated the table below by measuring the runtime performance of Google Chrome on an Intel(R) Core(TM) i7-7920HQ CPU operating at a frequency of 3.10 GHz, with our case study serving as a reference at each stage. Although not optimized, our prototype is thus already convenient for interactive use.

Process	Execution time in sec.		
3D field of fibers	0.098		
Vessel creation	0.791		
Surface texturing	1.13		
Cut (on the shader)	Instant		
Cell creation (per cell)	0.090		
Cell distribution	2.44s		
Flow	0.015		
Vertical cut event	3		
Cell deformation	0.058		

User Study. We evaluated our prototype through a user study among experts. The objective was twofold: firstly, after a short 65 user experience, to assess participants' feelings about the rel-66 evance of DynBioSketch to facilitate communication and ex-67 ploration of dynamic biological phenomena; and secondly, to 68 allow them to express themselves on its advantages and dis-69 advantages compared to the illustration tools they are currently 70 using. The second part was challenging as we could not identify 71 any common baseline for informal, interactive illustration and 72 communication among biology experts. Each biologist seemed 73 to have become accustomed to using a different tool, sometimes 74 diverted from its primary use to meet their objectives. Unable to 75 set up a comparative test accessible to all participants, we for-76 mulated this part of the experiment as open questions, allowing 77 everyone to compare DynBioSketch with their favorite tools. 78

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*Participants:*. They included 6 women and 5 men aged 30 to
 67 years, all working in biological research. Their experience
 in the field ranged from 5 to 45 years. None of them had partic ipated in our preliminary study or the system design.

Protocol:. We first provided a brief introduction to the tool, followed by a 5-minute interactive demonstration in the example of the infection phenomenon. Participants then engaged in hands on experimentation with basic examples (see Fig. 16), allowing them to become familiar with our prototype before exploring their specific target phenomena. These testing ses-10 sions were conducted on a 2.5 GHz Windows 10 laptop with 11 8 Go RAM and lasted approximately 20 minutes. Due to time 12 constraints, the participants only experimented with the 2.5D 13 version of the tool, but we presented them a short demo video 14 of the 3D version in the introduction. Participants concluded 15 their session by completing a survey using the questions listed 16 below. 17

Questions:. Parts of them involved rating, while others were 18 open questions, enabling us to gather more information about 10 the participant's usages and opinions. The first questions 20 (Q1-Q4) focused on evaluating our model independently, while 21 the subsequent ones (Q5-Q8) involved comparing our proto-22 type with the participant's usual practice. To allow that, we 23 asked each participant to specify first which tool(s) they cur-24 rently used for scientific illustration and communication among 25 peers and to use them in the comparison. The exact list of ques-26 tions was: 27

- Q1: Please rate the time it takes to be familiar with Dyn-BioSketch on a scale of 1 (several weeks) to 5 (a few hours).
- Q2: Please rate the ease of creating a full biological environment in DynBioSketch, including modeling individual cells, creating a distribution of cells within a container, sketching a distribution of fibers, and extending it to 3D, on a scale of 1 (very difficult) to 5 (very easy).
  - Q3: Please rate the ability of DynBioSketch to convey motion and deformation for the infection phenomenon case, on a scale from 1 (very far from expectations) to 5 (very close to expectations).

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Q4: : Please rate the ability provided by DynBioSketch to
 specify a triggering event and replay the animation from
 the event creation time, on a scale of 1 (very difficult) to 5
 (very easy).

After having listed the illustration tools you use daily for your
thinking or communication with peers, please compare DynBioSketch with your favorite tool(s) by rating the following
questions from 1 (strongly disagree) to 5 (strongly agree). Is
DynBioSketch better than your favorite tool for:

- Q5: Representing a complex biological environment?
- Q6: Depicting a biological phenomenon (including the narrative part: triggering event and consequences)?

- Q7: Communicating about a biological phenomenon?
- Q8: Exploring ideas about a biological phenomenon (such as what might occur between two states, etc.)?

We concluded the survey with an open question allowing participants to share their thoughts on DynBioSketch 2.5D versus the full 3D modeling and animation for conveying biological phenomena, as well as any additional comments they wished to provide. 59

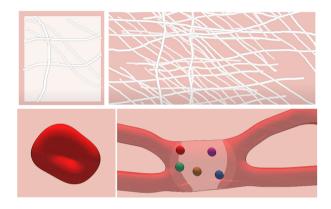


Fig. 16: Some designs from our participants during the user study.

**Results of the user-study.** We computed the mean and standard deviations of the responses gathered for each question.

Evaluation of the tool itself (Q1-Q4):. Fig.17 illustrates the results of questions Q1–Q4. All users expressed satisfaction with DynBioSketch's user-friendly modeling interface (see the results of Q1 in Fig. 18), describing it as "an easy-to-use tool". The participants also stressed the importance of integrating 3D shape representations to enhance understanding of biological environments (Q2). Moreover, two users specifically advocated for introducing more randomness in synthesizing shape distributions.

The users generally expressed approval of the symbols used to represent motions and were pleased with the resulting movements, yielding a mean score of 4.30 with a standard deviation of 0.6 for Q3 (see Fig.17). They especially valued the kinetic aspect offered by our prototype and highlighted "the ability to specify gradients" as particularly useful. Additionally, one user suggested incorporating collision processing with background fibers could enhance the fidelity between symbols and generated motion. Furthermore, enabling animation within such schematic representations was noted to "help sustain attention" by participants.

The participants successfully managed the design of a biological phenomenon and replayed the sequence from the triggering event (Q4). One user praised DynBioSketch for enabling them to "express temporal processes to facilitate discussion around a problem" and 'communicate their vision of a phenomenon". Additionally, three users identified a notable weakness in the perceived simplicity of our deformation tool.

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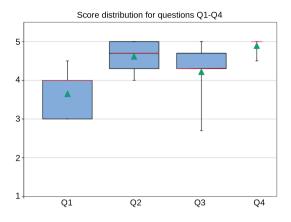


Fig. 17: Score distribution for questions Q1–Q4 evaluating DynBioSketch: the mean is indicated by the green triangle and the median by the red line.

Comparisons with tools commonly used by biologists (Q5-Q7):. Fig. 18 presents the comparisons of DynBioSketch with the 2 tools commonly used by biologists. Ranked by frequency of 3 use, their current favorite tools are as follows: PowerPoint (8 votes), Paper and pencil (5 votes), 3D Movies (3 votes), BioRender (2 votes), Adobe Photoshop (2 votes), Adobe Illustrator (1 vote), Microsoft Paint (1 vote), and Artist Illustration (1 vote). 8

The ability of DynBioSketch to integrate 3D shape repre-9 sentations to represent a complex biological environment was 10 highly praised (Q5) by the participants, particularly noting the 11 limitations of 2D representations or fixed perspectives inherent 12 in traditional methods like paper and pencil, as well as soft-13 ware such as PowerPoint, Photoshop, and Illustrator. Despite 14 the availability of pre-defined shapes or built-in functionalities 15 in such software, users highlighted features missing in our pro-16 totype, such as calibration and spatial scale. 17

Participants found that offering a 3D environment, anima-18 tion, and narration in DynBioSketch (O6) surpassed the capa-19 bilities of traditional tools like paper and pencil, PowerPoint, or 20 BioRender, restricting the results to 2D and static representa-21 tions. 22

The most noteworthy findings from participants' free com-23 ments revealed that biologists found the tool to be "extremely 24 useful when communicating with each other" and "convenient 25 to communicate how they see a phenomenon". Despite having 26 fewer features than professional software, our prototype was 27 considered more effective in communicating biological phe-28 nomena (see Q7 with yellow bars in Fig.18) than commonly 29 used tools by 82% of users as it enabled the creation of more 30 complex events than PowerPoint or traditional paper-and-pencil 31 methods. However, two users suggested that a professionally 32 produced video might be a better alternative for communica-33 tion. 34

Biologists praised DynBioSketch for "helping to express 35 what they imagined simply", "helping detect inconsistencies in 36 initial mental models" and "prompting questions that would not 37 have arisen with flat representations". As illustrated with green 38 bars in Fig.18, DynBioSketch was considered more convenient 39 to explore ideas about a biological phenomenon (Q8) than tra-40 ditional tools and software. 41

Finally, biologists generally expressed that our tool ad-42 dressed a current gap in their toolkit: an interactive platform for 43 interactively creating 3D animated environments accompanied 44 by narration. They viewed this concept as a valuable addition 45 to their resources and particularly highlighted DynBioSketch's 46 unique ability to represent easily a 3D biological phenomenon. 47 In the following table (Table 1), we evaluated the following 48 features: Interactive interface (I), Creation of 3D environment 49 (3D), Animation (A), and Narration (N). 50

	Ι	3D	A	N
DynBioSketch	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
PowerPoint	$\checkmark$	Р	X	Р
Paper and pencil	$\checkmark$	Р	X	Р
Movie	Х	$\checkmark$	$\checkmark$	$\checkmark$
BioRender	$\checkmark$	Р	X	Р
Adobe Photoshop	$\checkmark$	Р	X	Р
Adobe Illustrator	$\checkmark$	Р	X	Р
Microsoft Paint	$\checkmark$	Р	X	Р
Artist's illustration	Х	Р	X	Р

Table 1: Evaluation of DynBioSketch and the tools used by biologists with respect to features. "X" means not handled, "P" means partly handled.

The industrial software tools highlighted above (PowerPoint, 51 BioRender, Adobe Photoshop, Adobe Illustrator, and Microsoft 52 Paint) are confined to a 2D environment and restrict users to 53 creating, at most, 3D objects viewed from a fixed perspective (as denoted by 'P' in Table 1). Furthermore, as these tools lack animation capabilities, narration is typically limited by labels, text, or arrows. While movies provide immersion into an envi-57 ronment and animation, they lack interactivity.

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Final open questions and identified shortcomings:. Through its interactivity and combination of modeling and animation, Dyn-BioSketch has unanimously been considered a very promising tool for exploring ideas on biological phenomena.

However, several participants emphasized the importance of incorporating calibration options similar to those found in professional software like BioRender to broaden the utility of Dyn-BioSketch for a wider range of scenarios. Two of them also noted that enabling progressive refinement, possibly at other space and time scales, would have been a great addition to the current tool.

In addition, we were surprised and deceived, to be honest, 70 by the participants' answers to the last question about their 71 preference between DynBioSketch in 2.5D and a 3D version. 72 While we had put a lot of effort into developing our 2.5D ver-73 sion, based on non-photorealistic yet expressive modeling, ani-74 mation, and rendering, thinking we were offering a good com-75 promise between expressivity and simplicity of control, 8 users 76 among 11 expressed that they preferred the 3D version. They 77 said that such 3D version was needed to communicate their 78 phenomenon of study. All, however, noted that the 2.5D ver-79 sion was an excellent way to generate simplified representa-80 tions used when teaching or to produce the visual summary im-81 ages required for each publication in biology. While we respect 82 their appeal to full 3D to reach more realism, we did not have

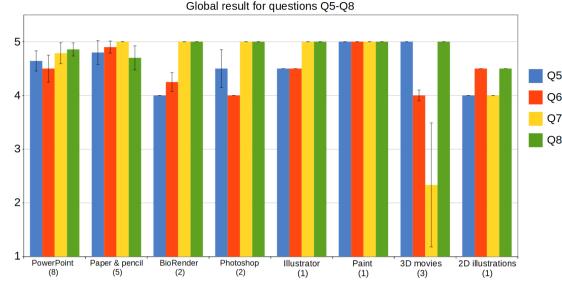


Fig. 18: User-study results for DynBioSketch comparison with former tools or usages biologists were familiar with: Each participant was allowed several comparisons, so the number of participants mentioning a given tool is indicated in the bottom line: mean (color) and standard deviation (black) for the answers to Q5-Q8. Note that in addition to the tools they were themselves using, biologists also compared the illustration they get when they ask an artist to design a 3D movie or a 2D illustration for them (two last columns).

enough time during the user study to make them manipulate the full 3D version. Although we think that participants could have changed their minds if they were aware of the difficulty of manually controlling the camera trajectory and zoom factor, in addition to designing a full animated 3D scene, this unexpected result encouraged us to pursue, in future work, the extension our illustration tool to full 3D animated illustrations-probably to be enhanced with some automated camera control.

#### **Discussion & Limitations** 8.

We presented a complete pipeline for creating, animating, 10 and rendering interactive depictions of biological scenes with 11 a particular focus on representing the phenomenon of infec-12 tion. Drawing inspiration from illustrations in cell biology, our 13 design choices emulate biological sketching techniques. How-14 ever, we have also paid attention that our user interface is user 15 friendly for experts in biology and novices alike. Moreover, our system offers users considerable flexibility in modeling various 17 shapes at different scales, thanks to our sketch based system for 18 the 3D implicit modeling system and cutting tool. In terms of 19 animation, we recognize that different phenomena may require 20 incorporating additional motion or deformation behaviors. 21

While our aim was not to produce exact replicas of the in-22 spiration figure, our tool aimed at complementing existing il-23 lustration tools by providing an intermediate between abstract, 24 conceptual sketches, and full 3D animation reproducing or sim-25 ply visualizing the real phenomena with precision. The images 26 and animations created by our prototype are intended to bring a 27 clear understanding of a phenomenon, which users may enrich 28 with additional annotations, text, or labels. 29

While not covering every aspect, our designed goals (Sec-30 tion 3.3) were sufficient for the cell biology scenario we inves-31 tigated, enabling users to recognize DynBioSketch's potential 32

to create dynamic visual illustrations. However, we identified the following three limitations of our current solutions:

Firstly, as mentioned above, our solution for animating subshapes within a 3D container automatically induces spiraling trajectories. While this was considered quite natural by most users, we are not offering any control of this feature, which only depends on the timestep used and the local amount of bending of the container's shape. This twisting motion could be a problem for some applications.

Secondly, while implicit surfaces are a suitable model to create interactively and deform organic-looking shapes, their lack of parametrization made texture mapping problematic. While we resorted to explicit mesh tessellation to create our Voronoi membrane cells, our method is not general and leaves room for improvement. With a goal of better Voronoi-like texture of freeform surfaces, more precision would be needed in terms of sampling kernel positions, and the tessellation computation should use the geodesic distance along the surface, as was done by Meuschke et al. [32].

Lastly, to facilitate real-time tissue cutting, we rendered objects transparent instead of simulating any complex change in their geometry. In future work, representing some local deformations occurring during a "real" cutting event would enable us to convey the phenomenon more convincingly. This would necessitate physics-based volumetric simulation in anisotropic materials, accounting for the tension exerted by fibers and vessels in biological tissues and the subsequent relaxation of these tissues upon cutting. Although realizing real-time animation at this level of detail is challenging, this would be an interesting extension of our work.

At a more conceptual level, biological phenomena are multiscale, from our scale to molecules through organs and cells. Inspiring from recent work in visualization, enabling multi-scale representation at the design stage, such as letting biologist stop 66

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the animation, zooming on a part of their design, and adding details at other temporal or spatial scales, would be an ambitious

yet exciting avenue for future work.

## 9. Conclusion

We presented DynBioSketch, an interactive platform intended to help biologists express their conceptual understanding of infectious phenomena through animated 3D sketches. It also serves as a tool for non-specialists to explore immune re-8 sponses to infection. After a prestudy involving biology experts to identify relevant animation forms and tools, we developed a 10 complete pipeline to achieve this goal and implemented a pro-11 totype system. We tested the system on a complex scenario: 12 modeling the immune response of tissues to a skin cut. Users 13 managed to create a DynBioSketch accompanied by a narrative 14 within a few minutes. Our user study underscores the need for 15 a system like DynBioSketch as a valuable addition to presen-16 tations. Although not comprehensive at present, we anticipate 17 that our innovative system and its future improved extensions 18 will create new opportunities for visual communication, educa-19 tion, and research, extending beyond biology. 20

In particular, we would like to extend DynBioSketch's capa-21 bilities to the representation of other phenomena, with a partic-22 ular focus on multi-resolution sketching. This capability would 23 empower users to dynamically create and zoom into animated 24 scientific illustrations with virtually unlimited levels of detail. 25 Similar to the approach proposed by Wither and al. [51], inte-26 grating machine learning techniques would help facilitate the 27 reproduction of sketch-based detail in different sections of a 28 media form. This functionality is essential for seamless navi-29 gation and zoom-out functions. 30

#### **CRediT** authorship contribution statement 31

Pauline Olivier: Conceptualization, Methodology, Soft-32 ware, Validation, Formal Analysis, Writing - Original draft, 33 Writing - Review and editing. Tara Butler: Methodology, 34 Writing - Original draft . Pascal Guehl: Methodology, Soft-35 ware, Writing - Review and editing, Jean-Luc Coll: Valida-36 tion, Writing - Review and editing Renaud Chabrier: Concep-37 tualization, Validation, Pooran Memari: Conceptualization, 38 Methodology, Formal Analysis, Writing - Review and editing, 39 Marie-Paule Cani: Conceptualization, Methodology, Supervi-40 sion, Project administration, Writing - Review and editing

#### **Declaration of competing interest** 42

The authors assert that they do not have any identifiable fi-43 nancial conflicts of interest or personal relationships that might 44 have been perceived to influence the work reported in this paper. 45

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