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Metabolic balance of [3H]-BADGE in rodents:

A spotlight on the complex in vivo fate of the diglycidyl ether analogue of bisphenol A

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Introduction

Bisphenol A diglycidyl ether = BADGE

(2,2-Bis[4-(glycidyloxy)phenyl]propane, CAS N°1675-54-3, MW 340.4)

- Used in the manufacture of epoxy resins and polycarbonates for food packaging
- Synthesized by a condensation reaction between epichlorohydrin and Bisphenol A
- **Protective coatings**: car and marine industry, food packaging, adhesives, dental sealants, etc.^[1, 2]
- Reported human daily intake: approximately 0.16 µg/kg b.w./day^[2, 3]
- Tolerable Daily Intake, BADGE + hydrolysis products = 0.15 mg/kg b.w.^[4]
- Migration of BADGE + hydrolysis products does not exceed: 9 mg/kg in food, 1 mg/kg for chlorinated derivates^[4]
- Possible **endocrine disruptor** (PPARγ antagonist)^[5, 6]

Objectives: Investigate the **fate of BADGE** in male and female Wistar rats Explore BADGE metabolic balance and identify the different metabolic pathways involved in its biotransformation for toxicological purposes **Identify candidate biomarker metabolites**, with the perspective of further human biomonitoring studies

Chemicals: [3H]-BADGE

[³H]-BADGE (radiochemical purity > 97 %, specific activity 10.9 GBq/mmol) was synthesized in-house from radio-labelled [3H]-Bisphenol A (Moravek). Unlabeled BADGE was purchased from Sigma Aldrich.

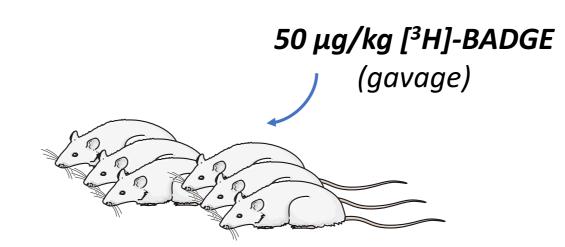
> Animal experiment:

High-performance-liquid-chromatography coupled to on-line radioactivity detection

(Radio-HPLC). Excreted metabolites were separated on an Ultimate 3000

(ThermoScientific, c.) HPLC system, with a Zorbax SB-C18 column (5 μm; 4.6 x 250 mm,

Agilent) coupled to a Flo-One A610TR detector (Perkin Elmer, d.), using 2 mL/min Flo-



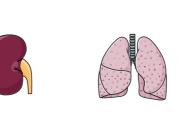
3 male and 3 female 8-week old Wistar rats were force-fed an ajusted single [3H]-BADGE dose of **50** μg/kg b.w. (1.6 MBq/kg b.w.) dissolved in corn oil. After 24h spent in individual stainless steel metabolic cages, with water and food supplied ad libitum, rats were weighed and euthanized.

▶ Metabolic Balance and Tissue Distribution:



Methodology









Urine, feces, blood, liver, genital tract, glands, adipose Tri-Carb 2910TR, with Packard Ultima Gold as the tissue, muscle, digestive tract (and its content), brain, kidney, spleen, lung, heart, bladder, pancreas, skin and the remaining carcass were collected for radioactivity quantification. Extractions were performed on feces and digestive contents. Radioactivity was measured in liquid samples using a Packard scintillation analyzer (a.)

scintillation cocktail (Perkin Elmer). Feces were dried and ground before solubilization in Soluene (Perkin Elmer). Radioactivity in homogenized tissues, organs, blood, and carcasses was determined by complete combustion using an Oxidizer 307 (Perkin Elmer, b.), followed by quantification on a scintillation analyzer. At least 3 replicates were analyzed for each sample.

> Analytical conditions:

Flow rate	1 mL/min	Column temperature	25 °C	Analysis time	55 m
Phases	A: Ammonium acetate buffer 20 mM pH 3,5 / Acetonitrile (95:5 v/v) B: Ammonium acetate buffer 20 mM pH 3,5 / Acetonitrile (10:90 v/v)				
Gradient	0-4 min: 100 % A, 4-6 min: 100-75 % A, 6-16 min: 75 % A, 16-18 min: 75-70 % A,				
	18-25 min: 70 % A, 25-27 min: 70-30 % A, 27-38 min: 30 % A, 38-40 min: 30-0 % A,				
	40-50 min: 0 % A, 50-51 min: 0-100 % A, 51-55 min: 100 % A				

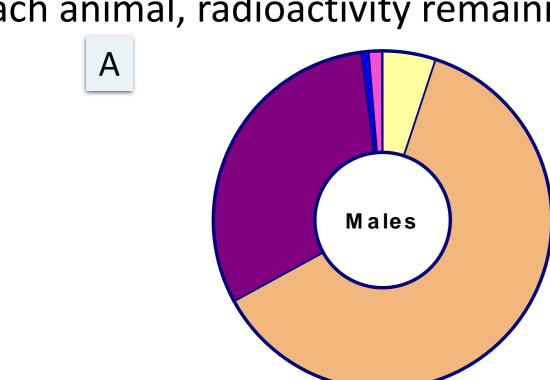
Preliminary results

Scint II (Perkin Elmer) as the scintillation cocktail.

Metabolic profiling and identification:

Metabolic balance of [3H]-BADGE following a single oral dose administered to male and female Wistar rats

> For each animal, radioactivity remaining in the feeding material was quantified to calculate the nominal [3H]-BADGE dose administered: 46.8 ± 2.2 μg/kg b.w. (1.5 ± 0.07 MBq/kg b.w.)



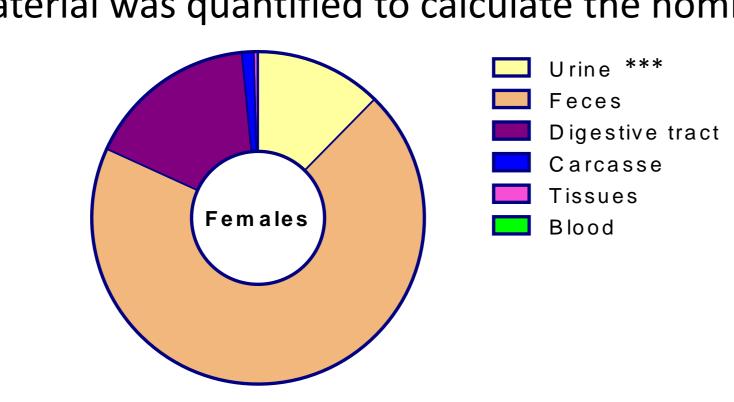


Fig. 1: 24h metabolic balance in male (A) and female (B) Wistar rats. Results (mean, n = 3 for each group) are expressed as % of the administered radioactivity (t-test, *** p-value \leq 0.001, Graphpad®)

- \gt 92.1 ± 9.4 % of the administered radioactivity recovered at 24h following BADGE administration (n = 6).
- \triangleright Feces was the main route of excretion (55.1 ± 16.8 % in males, 66.2 ± 6.7 % in females), and the digestive tract accounted for $27.5 \pm 6.7 \%$ of the administered radioactivity in males and $15.8 \pm 4.1 \%$ in females, respectively.
- > Significant sex-related differences were observed for radioactivity recovery in urine: 4.5 ± 0.9 % in males and 11.8 ± 0.9 % in females, respectively.

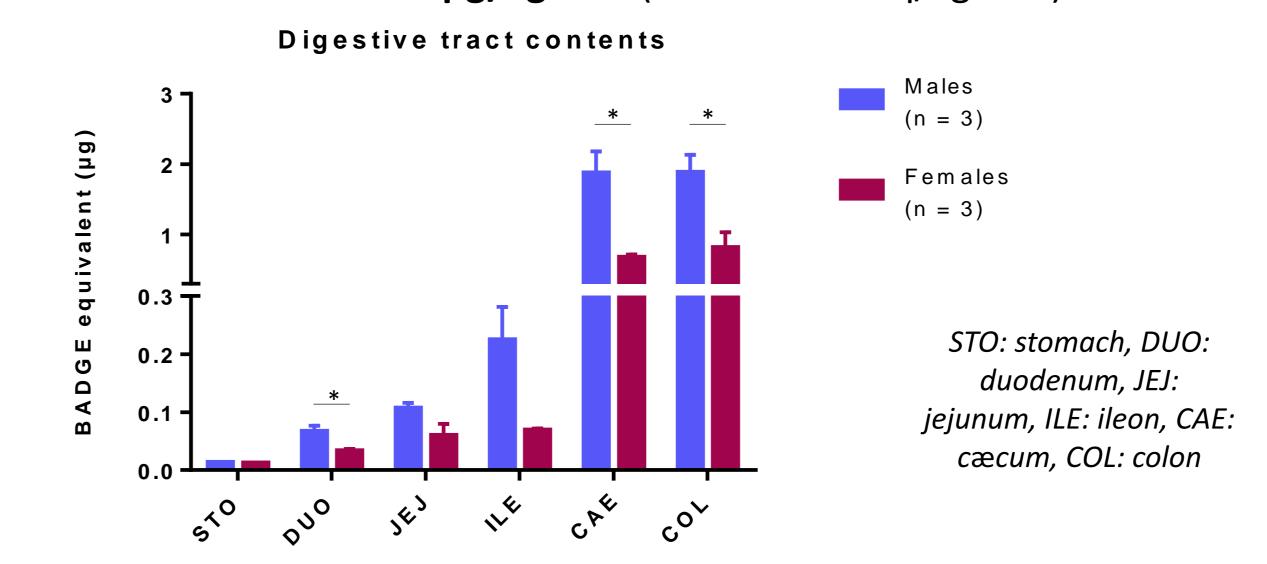


Fig. 2: 24h metabolic balance: residual radioactivity in digestive tract contents of male and female Wistar rats. Results (mean ± SEM) are expressed in μg of BADGE equivalent (t-test, * p-value ≤ 0.05 , Graphpad®)

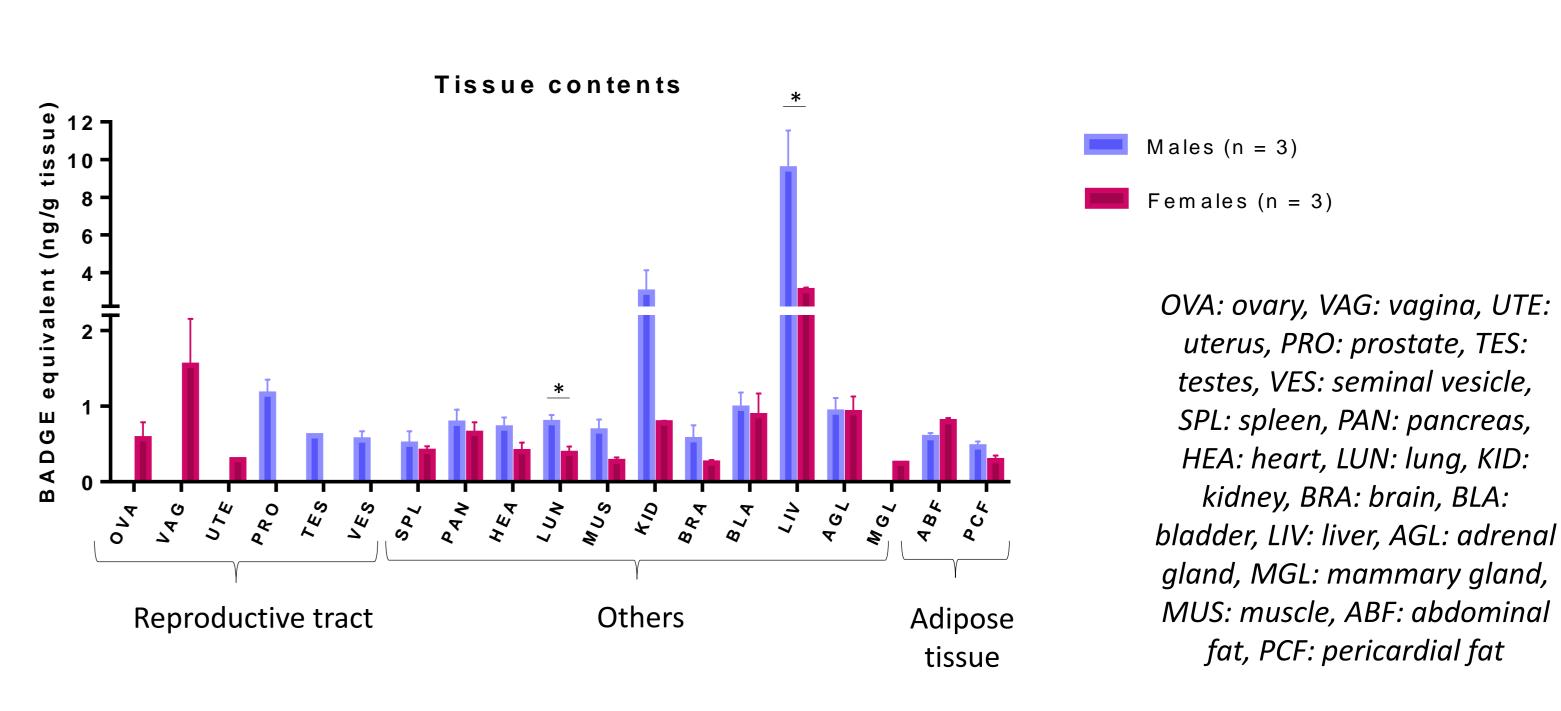
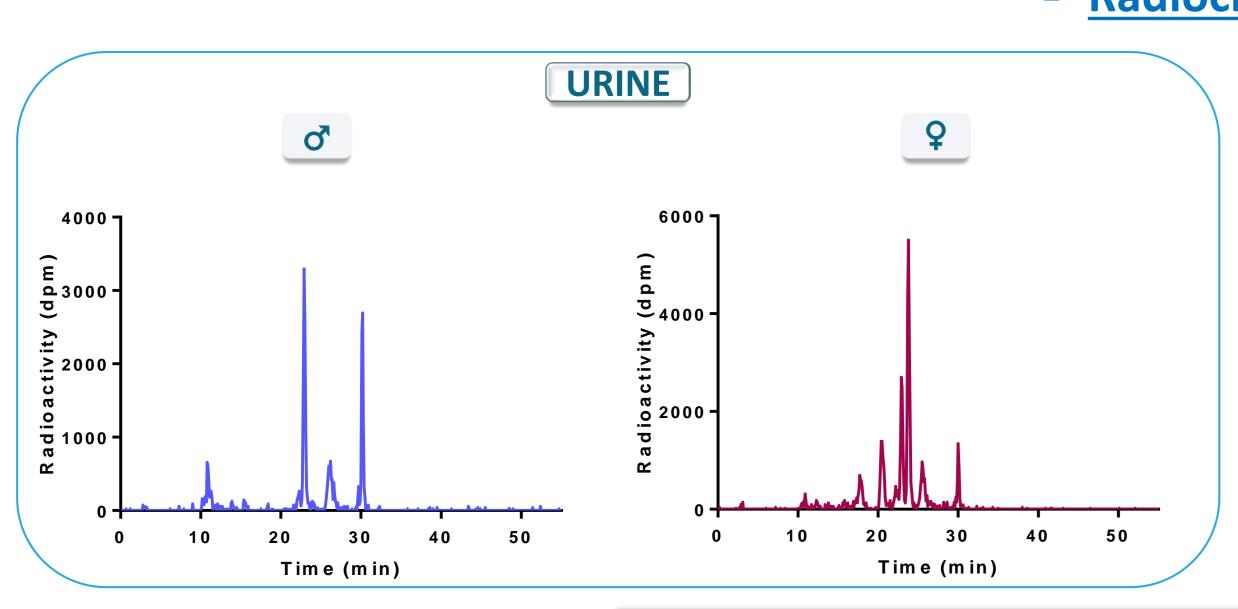
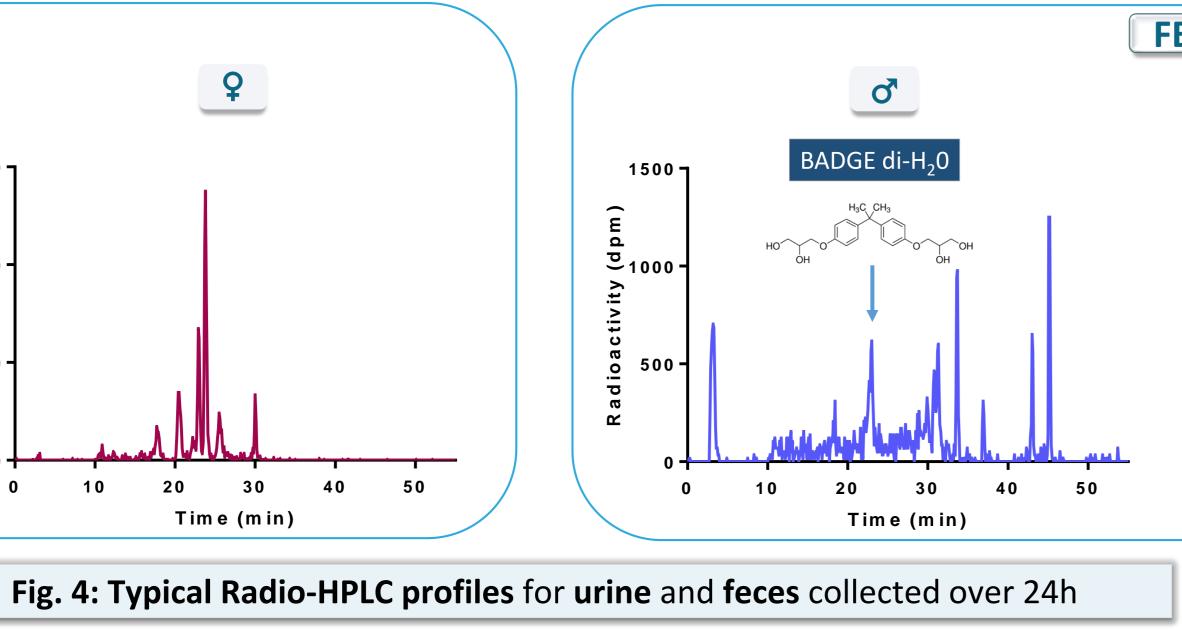
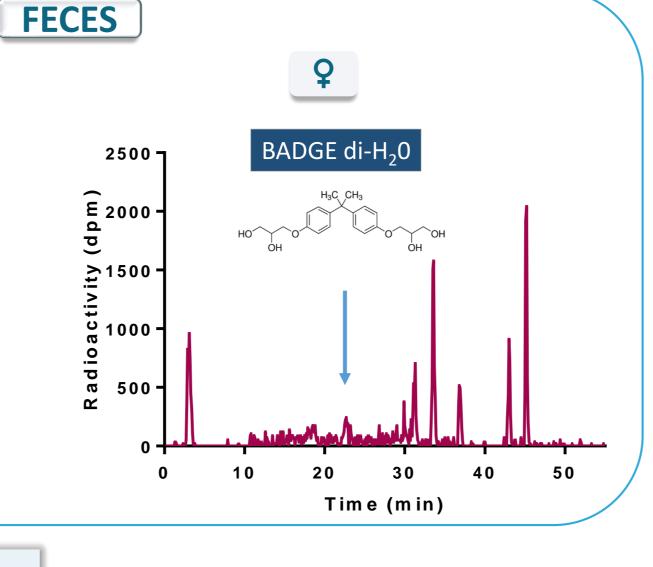


Fig. 3: Residual levels of radioactivity in tissues of male and female Wistar rats. Results (mean ± SEM) are expressed in ng of BADGE equivalent/g tissue (ppb) (t-test, * p-value ≤ 0.05, Graphpad®)









- > Sex-related differences were highlighted for urine and fecal metabolite profiles
- Complex radio-chromatographic profiles underline the chemical reactivity of BADGE

Conclusions

In this first in vivo 24h-study of [³H]-BADGE carried out in male and female Wistar rats, radioactivity was found to be **excreted** mainly in **feces**, with residues found in all tissues, including reproductive organs.

Sex-related differences were highlighted for urine, liver, lung as well as duodenum, cæcum and colon contents. Qualitative differences are also suggested by first Radio-HPLC analyzes carried out for urine and

BADGE elimination over 24h (urine + feces) appears to be faster in female rats compared to males (77.9 \pm 6.2 % in females vs. 59.6 ± 16.2 % in males).

Preliminary HPLC Radio-profiling analyzes strongly suggest a complex metabolism of this molecule. Further HPLC-MS/MS investigations are being carried out, with the aim to identify unknown BADGE metabolites.

References

[1] Xue J. *et al.*, 2018, Environ Res. 162:35-40. [2] Poole A. et al., 2004, Food Addit Contam. 21(9):905-19. [3] Wang L. et al., 2015, Environ Sci Technol. 49(5):3150-7. [4] Commission regulation N°1895/2005 on the restriction of use of certain epoxy derivatives in materials and articles intended to come into contact with food, 2005. [5] Wright HM. et al., 2000, J Biol Chem. 275(3):1873-7. [6] Satoh K. et al., 2004, Food Chem Toxicol. 42(6):983-93.

