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# **Microfluidics and Nanofluidics in Strong Light–Matter Coupling Systems**

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Abstract: The combination of micro- or nanofluidics and strong light-matter coupling has 19 gained much interest in the past decade, which has led to the development of advanced 20 systems and devices with numerous potential applications in different fields, such as 21 chemistry, biosensing, and material science. Strong light-matter coupling is achieved by 22 placing a dipole (e.g. an atom or a molecule) into a confined electromagnetic field, with 23 molecular transitions being in resonance with the field and the coupling strength exceed-24 ing the average dissipation rate. Despite intense research and encouraging results in this 25 field, some challenges still need to be overcome, related to the fabrication of nano- and 26 microscale optical cavities, stability, scaling up and production, sensitivity, signal-to-noise 27 ratio, and real-time control and monitoring. The goal of this paper is to summarize recent 28 developments of micro- and nanofluidic systems employing strong light-matter coupling. 29 An overview of various methods and techniques used to achieve strong light-matter cou-30 pling in micro- or nanofluidic systems is presented, preceded by a brief outline of the 31 fundamentals of strong light-matter coupling and optofluidics operating in the strong 32 coupling regime. The potential applications of these integrated systems in sensing, opto-33 fluidics, and quantum technologies are explored. The challenges and prospects in this 34 rapidly developing field are discussed. 35

Keywords: Strong light-matter coupling; microfluidics; nanofluidics; polaritons

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#### 1. Introduction

Since the discovery of the Purcell effect in 1946 [1], light–matter interactions have 39 been intensely studied, and considerable advances have been made in both theoretical 40 simulation, experimental research, and practical applications of the effect. Specifically, the 41 so-called cavity quantum electrodynamic (CQED) systems have been developed, which 42 represent optical cavities containing one or several atoms, organic molecules or artificial 43 atoms such as quantum dots (QDs). These and other advances mark the evolution of the 44 strong light–matter coupling (SC) systems, which has led to a number of important 45

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applications in the fields of nanophotonic devices, manipulation of quantum systems,46high-precision measurements [2], and quantum information processing (QIP) [3]. The47timeline of research and developments in strong light–matter coupling systems is shown48in Figure 1.49



**Figure 1.** Strong light–matter coupling (SC) timeline: from cavity quantum electrodynamics (QED) to ultrastrong coupling (USC). The data are from Refs. [1, 4–25].

Some of the most outstanding advances of the last years are related to SC in a liquid 53 environment. The combination of strong light-matter coupling and fluidics has opened 54 up new avenues for interdisciplinary research, providing a fertile ground for innovations 55 in the field of nanophotonics and micro-/nanofluidics. The dynamic interplay between 56 light and matter in a fluid offers a wealth of opportunities for basic research and practical 57 technological developments. By manipulating the interactions of photons and molecules 58 in the liquid or gas phase, researchers have unlocked unprecedented potential for appli-59 cations, spanning from optofluidic sensors to novel light-modulated chemistry. 60

Some of the advantages of combining SC and micro-/nanofluidics are a reduced consumption of expensive materials and reagents, a shortened reaction time and an increased product yield in chemistry-related fields, an improved sensitivity in sensing applications, and portability. Researchers have succeeded in developing a variety of micro- and nanofluidic techniques and technologies using plasmon nanostructures [26,27], whispering gallery resonators [28–30] and photonic crystals (PCs) [31].

Here, we present a comprehensive review of SC in micro-/nanofluidic systems. By summarizing recent research and critically analyzing the emerging trends, we aim to elucidate the fundamental principles behind these systems, experimental techniques used to study light-matter coupling in them, and technological implications of the observed phenomena. The insights presented in this review not only clarify the current state of research, but also inspire future trends and innovations in this field.

#### 2. Fundamentals of Strong Light-Matter Coupling

#### 2.1 Light–Matter Coupling Regimes

When a two-level system (e.g. an atom or a molecule) has a transition that is resonant 75 with the confined electromagnetic (EM) field, two regimes of light-matter interaction 76 could be distinguished: weak coupling and strong coupling. The light-matter coupling 77 strength (g) is then compared with two parameters: the photon decay rate of the cavity 78 ( $\kappa$ ) and the non-resonant decay rate of the molecule ( $\gamma$ ) [12] (Figure 2). 79



**Figure 2.** Schematics of weak (on the left) and strong (on the right) light–matter coupling. 81 Here, a plasmonic resonator is represented by an array of periodically arranged gold rings 82 coupled with emitters. In the weak coupling regime, the emitter decay rate c(t) is enhanced (Purcell enhancement) without a substantial modification of the emitter eigenstate. In the strong coupling regime, the emitter decays at a rate governed by the Rabi frequency  $\Omega_R$ , which leads to the formation of two new eigenstates, exhibiting anticrossing behavior: upper and lower polaritons (UP and LP, respectively). 87

When  $g \ll (\kappa, \gamma)$ , the system is considered to be in the weak coupling regime. This means that the exchange rate is smaller than the loss rates, and, hence, the excitation is lost before it can be shared between the two-level system and the cavity. The weak coupling regime is related to the Purcell enhancement of spontaneous emission (*F*) [32]: 91

$$F = \frac{3Q\lambda^3}{4\pi^2 V n^3},\tag{1}$$

where Q is the quality factor, V is the mode volume,  $\lambda$  is the vacuum wavelength, 92 and n is the refractive index (RI) of the microcavity. As an example, this effect predominates when the molecule is placed next to a metal surface or nanostructure [33], contributing to the tuning of the photophysics and photochemistry (i.e., plasmon chemistry) of the system, which can be used for controlling chemical reactions. 96

When  $g >> (\kappa, \gamma)$ , the system is in the strong coupling regime. This means that the 97 coupling is strong enough to overcome the dissipation. Therefore, it is characterized by a 98 reversible coherent exchange of energy (known as Rabi oscillation) between the field in the 99 cavity and the electronic or vibrational states of the embedded matter, leading to the formation of two polaritonic modes (P<sup>+</sup> and P<sup>-</sup>), which emerge as two distinct peaks in the 101 energy spectrum of the system. In this regime, light–matter interaction cannot be treated 102

perturbatively and requires theoretical methods to describe the interaction between photons, electrons, and nuclei on different length and time scales.

Initially, in the uncoupled state, the transition energy of the two-level system is con-105 stant and has no dispersion, while the energy of the cavity eigenmode exhibits angular 106 dispersion and its minimum coincides with the transition energy at resonance. In the 107 strong coupling regime now both hybridised states exhibit dispersion. It is important to 108 note that the minimum of the cavity energy increases, resulting in a gap between the hy-109 bridised states observed at any angle. This effect is termed anti-crossing because even un-110 der resonance conditions the two branches of the hybridised states diverge. The anti-cross-111 ing of the dispersion curves of polariton states is the most reliable criterion of the strong 112 coupling regime in the system. 113

The difference in energy between the hybrid states is called Rabi splitting and, for a 114 simple two-level system, is given by the equation 115

$$\hbar\Omega_R = \hbar\sqrt{4g^2 - (\kappa - \gamma)^2} , \qquad (2)$$

Considering that  $g >> (\kappa, \gamma)$ , i.e., the light–matter interaction is faster than dissipation processes, the dissipation could be neglected: 117

$$\hbar\Omega_R = 2\hbar g \,, \tag{3}$$

On the basis of the Jaynes–Cummings model, in the absence of dissipation, the Rabi 118 splitting is 119

$$\hbar\Omega_R = 2d \sqrt{\frac{\hbar\omega}{2\varepsilon_0 V}} \sqrt{n_{ph} + 1}, \qquad (4)$$

where  $\hbar\omega$  is the cavity resonance or molecular transition energy,  $\mathcal{E}_0$  is the vacuum permittivity, V is the mode volume, d is the transition dipole moment of the molecule, and  $n_{ph}$  is the number of photons involved in the coupling. Rabi splitting can also occur in the absence of photons in the cavity, when  $n_{ph} = 0$ . This phenomenon is determined by the interaction of matter with zero-point EM fluctuations with the energy of  $\hbar\omega/2$  and is known as vacuum field Rabi splitting [34]. Therefore, in the absence of dissipation and in the vacuum field, the Rabi splitting is

Therefore, in the absence of dissipation and in the vacuum field, the Rabi splitting is 126 determined by the energy of the EM field  $(E_0)$  and the transition dipole moment (d): 127

$$\hbar\Omega_{R} = 2d \cdot E_{0} = 2d \sqrt{\frac{\hbar\omega}{2\varepsilon_{0}V}}, \qquad (5)$$

In addition, strong coupling regime is substantially facilitated by the so-called *collec-* 128 *tive coupling*, where a large number (N) of emitters, e.g., molecules, couple to one optical 129 mode . In this case, Rabi splitting is transformed as follows: 130

$$\hbar\Omega_R = 2d\sqrt{N}\sqrt{\frac{\hbar\omega}{2\varepsilon_0 V}},\tag{6}$$

This expression shows that a small mode volume and a considerable amount of molecules with a large transition dipole moment are required to confine the EM field and increase the coupling strength g and, therefore, the value of the Rabi splitting. Owing to the collective effects of an assemble of vibrational modes, the strong coupling regime can be achieved even at room temperature [19,37,38]. The interaction of the EM field and an assemble of N molecules results in the formation of N + 1 polariton states. However, only two of them can be observed experimentally, e.g., as two peaks in the absorption spectrum, 137 and therefore are termed bright states. The other N – 1 collective states cannot be directly 138 excited with light and, hence, cannot be observed, so they are termed dark states [39]. Thus, 139 the situation remains similar to the case of a single two-level system in terms of observable 140 energy states. 141

In addition, intermolecular interactions are important to consider in collective coupling. If the distance between molecules is small enough, Coulomb interactions between the transition dipole moments of different molecules become significant. Therefore, the transfer of excitation energy from one molecule to another is facilitated; i.e., there is delocalization of the excitation. Consequently, the transfer processes that favor delocalization, are excellent candidates for potential enhancements under strong light-matter coupling [40,41].

In the strong coupling regime, either an electronic or a vibrational transition can be 149 strongly coupled to the optical cavity mode, termed as electronic strong coupling (ESC) 150 and vibrational Strong Coupling (VSC), respectively. VSC is typically easier to obtain be-151 cause vibrational modes are in the infrared range, thus requiring cavity lengths of the order 152 of micrometers. VSC has been demonstrated to considerably modify the chemical reactiv-153 ity of molecules, either enhancing, or reducing it [21,42–45]. The formation of the hybrid 154 states in the VSC regime results in the so-called vibro-polaritonic (VP) modes. ESC occurs 155 in the visible range, which requires cavities smaller in size, making their engineering a real 156 challenge. In the case of ESC, various properties and processes have been found to be al-157 tered, such as work function, charge transport, energy transfer, intersystem crossing, and 158 nonlinear optics, along with the chemical reactivity. 159

The strong coupling regime is not always feasible due to the weak oscillator strength 160 of the vibration to be coupled or the restricted solubility of the active molecules in the 161 solvent used. In this case, the active solute molecules can be dissolved in a solvent or a 162 matrix that has a strong vibrational band interacting and matching with the vibrational 163 mode of the solute. This is called *cooperative coupling* [45,46]. In cooperative coupling, the 164 reactants indirectly couple with the confined EM field through their match with the vibra-165 tional modes of media. If the solvent is involved in VSC, all the solvent molecules sur-166 rounding the solute are vibrating in-phase, which, in turn, drives and couples the solute 167 vibration. As the vibrational energy of a chemical bond being changed by VSC, the ground 168 potential energy surface of the chemical reactions involving this bond is perturbed. In 2016, 169 it has been demonstrated [21] that the relative change in the reaction rate in the VSC regime 170 depends on the Rabi splitting energy, which is proportional to the square root of the con-171 centration of the reactant (*C*) (Figure 3): 172

$$\hbar\Omega_R \propto \sqrt{\frac{N}{V}} = \sqrt{C} , \qquad (7)$$

where N and V are the number of emitters and the cavity volume, respectively. 1



Figure 3. The decrease in the ratio of the reaction rates under vibrational strong coupling 175 and outside the cavity as a function of the Rabi splitting energy. The inset shows the linear 176 dependence of the Rabi splitting on the square root of the 1-phenyl-2-trimethylsilylacety-177 lene (PTA) concentration. Reproduced with permission from Thomas, A. et al. (2016) [21], 178 published by Wiley-VCH GmbH & Co. 179

#### 2.2 Optical Characteristics

The confinement efficiency under the strong light-matter coupling regime is typi-181 cally described in terms of the quality factor (Q-factor), photon lifetime ( $\tau$ ), and mode 182 volume (V). The Q-factor measures the resonant quality or the capacity for confining light 183 inside the microcavity and can be calculated as follows: 184

$$Q = \frac{\lambda_{cf}}{\Delta\lambda},\tag{8}$$

where  $\lambda_{cf}$  is the central wavelength of the resonance mode, and  $\Delta\lambda$  is its spectral lin-185 ewidth, commonly termed the full width at half maximum (FWHM). Microcavities with 186 Q-factors in the range from 10<sup>3</sup> to 10<sup>6</sup> are referred to as high-Q cavities, and those with Q-187 factor higher than 10<sup>7</sup>, as ultrahigh-Q cavities [47]. 188

The photon lifetime is defined as the time that a photon is trapped inside the micro-189 cavity before being dissipated and it is proportional to the Q-factor. Hence, fabrication of 190 structures with a high Q-factor is critically important. The photon lifetime can be calcu-191 lated as follows: 192

$$\tau = \frac{Q}{\omega_{cf}},\tag{9}$$

where  $\mathcal{O}_{cf}$  is the central frequency of the resonance mode.

The mode volume, also referred to as the effective mode volume, in contrast to the 194 geometric volume, which is only relevant to the physical size of the cavity, describes how 195 efficiently the cavity confines the field and is determined from the spatial distribution of 196 the EM field in the cavity: 197

$$V_{eff} = \frac{\int_{V} \varepsilon(r) |E(r)|^2 d^3 r}{\max(\varepsilon(r) |E(r)|^2)},$$
(10)

193



where  $\varepsilon(r)$  is the permittivity at position r, E(r) is the electric field strength at position 198 r, and  $\,V\,$  is the volume of the microcavity. This expression means that the energy density 199 increases as the mode volume decreases. Therefore, a smaller mode volume can lead to a 200 stronger interaction between the light and the system. The mode volume is usually ex-201 pressed in units equal to the cube of the wavelength divided by the RI of the cavity mate-202 rial, i.e.,  $(\lambda/n)^3$ . Since the feasible size of a dielectric cavity cannot be smaller than half the 203 wavelength considered, the minimum available mode volume for a cavity-type resonator 204 is limited by  $V_{eff} \approx 1$  [48]. Therefore, when smaller mode volumes  $V_{eff} < 1$  are desired, 205 nanoscale optical cavities, such as plasmonic cavities, are used, where the diffraction limit 206 is overcome, and the mode volumes can be as small as  $V_{eff} \approx 10^{-6}$ . Yet, in this case the Q 207

values are low because of high losses in the metal  $(Q \approx 10)$  [49]. Figure 4 shows the comparison of Q-factor versus the mode volume for different types of cavities. 208



Figure 4. The quality factor, Q, versus mode volume of different types of cavities. Repro-211duced with permission from Liu, W., et al. (2022) [50], published by Elsevier.212

To describe the light matter-interaction, numerical and computational methodolo-213 gies could be used which can be categorized according to the level of theory complexity 214 (Figure 5). From the computational approach, methods such as Finite difference time do-215 main (FDTD), Finite element method (FEM), and the transfer matrix approach use Max-216 well's equations for the light field. FDTD excels at simulating light propagation and scat-217 tering for time-domain analysis. This method is typically used to determine the mode vol-218 ume [48,51,52] of a system in a cavity, the dispersion diagram, Q-factor, and mode pattern 219 of the cavity. FEM excels in frequency-domain analysis, high-accuracy modeling of com-220 plex geometries, and situations requiring detailed spatial resolution [53]. Transfer matrix 221 method is used to study the reflectivity spectra and the complex frequencies of optical 222 eigenmodes of the structure as functions of the vacuum Rabi frequency [54]. One step 223 further in computational approach is the Density functional theory (DFT) method that 224 solve the electronic Schrödinger equation and often ignore the transverse electromagnetic 225 field effects. Here the matter and light are treated from the quantum and classical ap-226 proach, respectively. To overcome the classical approximation of the electromagnetic field 227 the full complexity of the system must be considered by treating the matter and light on 228 a coherent quantized basis. Therefore, Quantum-electrodynamical density-functional the-229 ory (QEDFT) [55], quantum electro-dynamics coupled cluster theory (QED-CC) [56] and 230 two quantum-electrodynamical coupled-cluster singles-doubles (QED-CCSD) are ab 231

initio methodologies that accurately model correlated electron–photon systems. QEDFT
232
has been used to describe SC however it requires massive computing power. These methodologies have been used recently to calculate local polaritonic effects [57], ionization energies [58], and ground state modifications [59].

The theoretical bases used to describe the SC include classical and quantum models. 236 However, in view of the complexity of the problem, the classical interaction of the EM 237 fields with matter is not sufficient to describe the SC. Therefore, model Hamiltonians are 238 used in most theoretical studies, which are mostly based on the Jaynes–Cummings or 239 Dicke model for single or multiple systems, respectively. These classical and quantum theoretical models are described in the following section. 241



Figure 5. Numerical and computational methodologies for describing light-matter inter-<br/>action. Reproduced with permission from Flick, J., et al. (2018) [60], published by De Gruy-<br/>ter.243<br/>244245

#### 2.3. Classical Approach

In the classical approach, the coupling of an optical mode and matter (quantum sys-247 tems such as atoms, molecules, and solid-state systems) is described in terms of the clas-248 sical EM theory using the analogy of two coupled harmonic oscillators. In an uncoupled 249 system, the behaviors of the harmonic oscillators are independent of each other. When the 250 coupling is strong enough, the two oscillators start to periodically transfer energy to each 251 other, behaving as a single entity. The periodic exchange of energy is represented by a 252 third spring with an interaction spring constant g , which corresponds to the coupling 253 strength of the system. The classical motion description for two dissipative harmonic os-254 cillators can be expressed as [39] 255

$$n_A \ddot{x}_A = -k_A x_A + g(x_B - x_A), \qquad (11)$$

$$m_B \ddot{x}_B = -k_B x_B + g(x_A - x_B), \tag{12}$$

where  $m_i$ ,  $x_i$  and  $k_i$  are the mass, displacement, and spring constant of the two oscillators. These equations assume that the coupling rate is much larger than the dissipation 257

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rate; i.e., the system is in the strong coupling regime. Defining  $\omega = \sqrt{k/m}$  for each oscillator, we obtain two new normal modes: 259

$$\omega_{\pm} = \frac{1}{2} \left( \omega_A + \omega_B \pm \sqrt{(\omega_A - \omega_B)^2 + 4\Omega^2} \right), \tag{13}$$

where  $\omega_A$ ,  $\omega_B$ , and  $\Omega$  are the frequencies of the oscillators A and B, and the frequency 260 splitting, respectively. When the system is at resonance,  $\omega_A = \omega_B \equiv \omega$ , which yields two 261 eigenvalues  $\omega_{\pm} = \omega \pm \Omega$  with two new normal modes differing from each other by 2 $\Omega$  262. As in Equation (3), the splitting ( $\Omega$ ) between the two new modes depends on the coupling strength g. 264

However, it should be noted that the classical approach has limitations, especially in the description of phenomena occurring at the quantum level. For a more accurate description of strong light–matter coupling, a quantum mechanical approach, such as the Jaynes–Cummings model or Dicke model, is often necessary (Table 1, SI) 268

#### 2.4. Quantum Approach

In the quantum approach, both the atom and light are quantized. The main difference 270 of the quantum approach from the classical description is the consideration of vacuum 271 EM fluctuations in empty space. These vacuum fluctuations represent the optical modes 272 with the zero-point energy (ground state energy) of the quantized EM field. In free space, 273 the EM field has a continuum of modes that typically interact weakly with the system. EM 274 fluctuations affect the properties of molecules and materials, such as spontaneous emis-275 sion. One way to enhance this interaction is to place the system inside an optical cavity. 276 In this confined geometry, the EM field presents a discrete spectrum of modes. In addition, 277 the field can be much more localized than in free space. This localized EM field leads to a 278 stronger field enhancement, which increases the emission quantum yield because it 279 strengths the interaction between light and matter, resulting in more efficient energy 280 transfer due to an increase in the excitation and emission rates of the molecules [61,62]. 281

Around 1936, Rabi [63] introduced a model to discuss the effect of a rapidly changing 282 weak magnetic field on an oriented atom that has a nuclear spin. This model considers a 283 two-state system consisting of a single atom (N = 1). The atom is treated as a quantum 284 object, and the field is treated classically as a rotating field. In 1954, Dicke proposed a 285 version for a single quantum field mode with multiple two-level atoms (N > 1). It describes 286 the collective behavior of the atoms and the field, considering the possibility of coopera-287 tive effects, such as superradiance and subradiance. This makes it particularly useful for 288 studying systems with many atoms, where the collective behavior plays an important role 289 (Table 1, SI). 290

Despite its simplicity, the quantum Rabi model was not regarded as exactly solvable. 291 To solve the model, the rotation wave approximation (RWA), in which the counter-rotat-292 ing term (CRT) is neglected, was advanced by Jaynes and Cummings in 1963 [4]. The 293 Jaynes–Cummings (JC) model describes the interaction between a single two-level atom 294 (N = 1) and a single mode of the EM field, taking into account the quantum nature of both 295 the atom and the field. The JC model is readily solved, including the dynamics of the EM 296 field, making it more comprehensive than the Rabi model. The JC model has been suc-297 cessfully used to understand a range of experimental phenomena, such as vacuum Rabi 298 mode splitting [11] and quantum Rabi oscillation [64]. In 1968, Tavis and Cummings pro-299 posed an extension of the JC model for the case of several (N > 1) identical non-interacting 300 two-level systems coupled to a single-mode quantized radiation field. Thus, the Dicke 301 model is a multi-atom version of the quantum Rabi model, whereas the Tavis-Cummings 302 (TC) model is an extension of the JC model for N > 1. Table 1 (SI) summarizes the main 303 characteristics of each model. 304

Let us consider the model of a single-mode field interacting with a two-level system. 305 The full JC Hamiltonian takes the form 306

328

$$\widehat{H}_{JC} = \hbar \omega_c \hat{a}^{\dagger} \hat{a} + \frac{\hbar \omega_a}{2} \hat{\sigma}_z + \hbar g (\hat{\sigma}_+ \hat{a} + \hat{\sigma}_- \hat{a}^{\dagger}), \qquad (14)$$

where  $\hbar \omega_c$  is the resonance of the electric field,  $\hbar \omega_a = E_e - E_g$  represents the atomic transition energy,  $\hat{a}^{\dagger}$  and  $\hat{a}$  are the creation and annihilation operators of a single-mode bosonic field satisfying the canonical commutation relation  $[\hat{a}, \hat{a}^{\dagger}] = 1$ ,  $\hat{\sigma}_z = |e\rangle\langle e| - |g\rangle\langle g|$  is the Hermitian inversion operator,  $\hat{\sigma}_+ = |e\rangle\langle g|$  and  $\hat{\sigma}_- = |g\rangle\langle e|$  are the atomic rising and lowering operators, and g is proportional to the dipole moment. The JC Hamiltonian has the following eigenstates:

$$|1n\rangle = \frac{1}{\sqrt{2}} \left( |e,n\rangle + |g,n+1\rangle \right) , \qquad (15)$$

$$\left|2n\right\rangle = \frac{1}{\sqrt{2}}\left(-\left|e,n\right\rangle + \left|g,n+1\right\rangle\right)$$
(16)

The new, hybridized eigenstates (15)–(16) are called *dressed* states of the atom, i.e., the eigenstates of the atom "dressed" by the cavity mode. 314

In summary, the Rabi model is a simplified description of the light-matter interaction 315 for a single two-level atom, with the atom treated as quantized and the field treated as a 316 classically rotating field, whereas the JC model extrapolates this situation to a quantized 317 EM field. The Dicke model, on the other hand, considers the collective behavior of many 318 atoms interacting with a common field. Each model is useful for different types of systems 319 and highlights different aspects of light-matter interaction. Research in fully quantum 320 models is still underway to understand the effects related to strong coupling. Due to the 321 mathematical difficulties entailed by the inclusion of the counter-rotating terms, the Rabi 322 model has been mostly analyzed either numerically or by approximate methods appro-323 priate for each particular situation. Connecting these predictions to experimental obser-324 vations is quite challenging [65-67]. In the following section, we describe experimental 325 models of strong light-matter coupling integrating micro- and nanofluidics. 326

#### 3. Methods and Techniques for Obtaining Strong Light–Matter Coupling in Microand Nanofluidic Systems

Micro- and nanofluidics imply accurate control and manipulation of fluids on the 329 microscale (1-100 µm) and nanoscale (1-100 nm), in so-called microchannels and na-330 nochannels, respectively [68]. The numerous techniques for the fabrication of micro-331 /nanofluidic devices developed to date are described in [69,70]. In terms of fabrication, the 332 earliest devices were engineered using traditional microfabrication methods, such as pho-333 tolithography [71] and soft lithography [72], and were mainly used for academic research. 334 As the field has developed, the range of methods and techniques for fabricating microflu-335 idic devices has expanded to encompass 3D printing, laser machining, and micro-milling, 336 enabling the designing of more complex and diverse devices. Similarly, the evolution of 337 nanofluidics has been driven by advances in nanofabrication techniques, such as electron 338 beam lithography and focused ion beam milling, allowing precise manipulation and con-339 trol of a nanoscale fluid flow. For example, the fabrication of micro- and nanointerfaces 340 with geometrically controlled ~10-nm nanochannels on a single fused-silica substrate us-341 ing the lithography and etching processes has been reported. The system has been used 342 for obtaining a pressure-driven flow of liquids in 50-nm channels [73]. 343

Initially, microfluidic devices were based on silicon and glass. Over time, new materials, such as polymers, paper substrates, metals, hydrogels, and composites have been included. In the case of nanofluidics, the use of nanomaterials, such as carbon nanotubes and graphene, has opened new possibilities for controlling the flow of a fluid. In general, a simple micro-/nanofluidic device allows the storage and handling of fluidic samples or 348 reagents, as well as waste materials, in a single, compact, and disposable unit, which 349 largely facilitates automated sample supply for sensing and minimization of the sample 350 volume. At the dawn of microfluidics, the fluid was supplied by pumps; as the technology 351 advanced, digital microfluidics emerged instead of a continuous flow. This technology allows transporting microdroplets by electrowetting, avoiding various potential problems 353 such as channel clogging, air bubble formation, and sample cross-contamination. 354

The applications of micro- and nanofluidics have also expanded rapidly. In particular, synergy of strong light–matter coupling and micro-/nanofluidics for controlling molecular processes and chemical behavior in a cavity environment is a subject of intense experimental and theoretical research and developments. This combination brings together the advantages of fluidics, such as the capacity for controlling a low sample volume and flexibility in producing multiple channels, with the possible benefits of strong coupling in biosensing and control of chemical reactions. 355 356 357 358 359 360 361

However, studying micro- and nanofluidics is not straightforward because of the numerous phenomena occurring simultaneously at these small scales, including chemical and biological reactive processes; transport of mass, charge, momentum, and energy; crystallization; and molecular interactions, which makes micro-/nanofluidics an inherently multiphysics discipline. The selection of the materials should take into consideration reproducibility, affordability, and the required configuration for SC applications, as detailed below. 368

## 4. Configurations Combining Strong Light–Matter Coupling with Micro- and Nanofluidics

Optical microcavities, also referred to as microresonators, can be subdivided, according to their designs, into Fabry–Pérot microcavities, plasmonic nanocavities, photonic crystal microcavities, and whispering-gallery mode (WGM) microcavities (Figure 6). 373



**Figure 6.** Different setups used for obtaining strong light–matter coupling for the use in micro-/nanofluidics: (A) Fabry–Pérot cavity, (B) plasmon nanocavity, (C) photonic crystal fiber plasmonic sensor, (D) whispering-gallery mode microcavity (microring resonator). 377

#### 4.1. Fabry-Pérot Microcavity

A Fabry–Pérot cavity consists of two mirrors separated by a usually adjustable spacing. The mirrors are aligned parallel to each other so that light can be confined between the mirrors, forming a standing-wave resonator for electromagnetic waves. The cavity is considered to be on-resonance when the cavity length,  $L_{cav}$ , is equal to an integer number of half-wavelengths: 383

$$L_{cav} = m \left(\frac{\lambda_c}{2n}\right),\tag{17}$$

where  $\lambda_c$ , n, and m are the wavelength of light, the RI of the material inside the cavity, 384 and the mode order, respectively. 385

The main elements that constitute a Fabry–Pérot cavity are two reflective surfaces, 386 e.g., metal (Ag, Al, or Au) mirrors. The thickness of the adjustable spacing determines the 387

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resonance frequency of the cavity. The bottom reflective surface is usually supported by a transparent substrate (e.g., glass or quartz in the case of visible light, and CaF<sub>2</sub> or ZnSe in the case of infrared light). An alternative to these reflective surfaces are distributed Bragg reflectors (DBRs), which are multilayer structures comprising alternating quarter-wavelength layers with low and high RIs with a thickness of  $\lambda/4$ .

Generally, a high Q-factor of the cavity is achieved by improving the reflectivity of 393 the mirrors and can be calculated using Equation (8). The use of DBRs helps to increase the 394 Q-factor of the cavity, which in this case depends on the number of layer pairs [74] and the 395 RI contrast between the two dielectric materials. However, they require advanced manu-396 facture methods that are usually time-consuming. At the same time, metal-mirror micro-397 cavities have a relatively low Q-factor, which is limited by the intrinsic reflectivity (R) of 398 the metals ( $R_{A1}$  ~ 90–85%,  $R_{Ag}$  ~ 95–99%, and  $R_{Au}$  ~ 70–90% in the range of 550–700 nm 399 [75,76]) and the thickness of the films, e.g., gold colloid films, as a function of metal volume 400 fractions [77]. Surface roughness of the metal mirrors can lead to a deterioration of the 401 optical response and a shift in the peak wavelengths of resonance modes compared with 402 smooth mirrors [78]. In addition, when metal mirrors are used, surface plasmon resonance 403 (SPR) can be observed as a result of the contact of the metal mirror with the atoms or mol-404 ecules placed inside the cavity. SPR can cause distortion of mode dispersion curves, result-405 ing in an imprecise assessment of the SC impact on the properties of atoms or molecules 406 [79]. Therefore, a thin protection layer is recommended in order to avoid inaccurate eval-407 uation of strong coupling and protect the mirrors from damage. Moreover, a setup that 408 allows the study of different samples and the adjustment of the cavity length depending 409 on the required wavelength is desirable. For this propose, tunable Fabry-Pérot cavities 410 have been developed that allow varying the length of the cavity depending on each par-411 ticular sample, thus making it flexible and more versatile, avoiding the need to design a 412 new setup for each sample [80]. 413

Apart from regular Fabry–Pérot cavities, cavities filled with a liquid medium, which 414 have been considerably less studied, are of much interest. However, as shown in Equation 415 (17), it is technically challenging to fabricate a cavity with the sizes required for ensuring 416 resonance conditions, especially for electronic transitions. The energy of electronic transitions is in the visible range; therefore, a nanosized cavity is required. At the same time, 418 strong coupling to molecular vibrations is more feasible, because it requires a micrometerscale cavity length. 420

Figure 7 shows different setups used to implement micro-/nanofluidics under the 421 conditions of strong coupling. Generally, such systems consist of a Fabry-Pérot cavity with 422 two microfluidic junctions. The first study on strong coupling in a liquid phase was pub-423 lished in 2015 [20] (Figure 7A). There, a tunable microcavity was used to obtain vibrational 424 strong coupling in the infrared range for a variety of functional groups. The setup con-425 sisted of a commercial liquid sample cell unit with ZnSe windows coated with a 10-nm Au 426 layer and separated with Mylar spacers. Different substances were studied, including di-427 phenyl phosphoryl azide (DPPA) with an isolated N=N=N stretching mode at 2169 cm<sup>-1</sup>. 428 The results showed a vacuum Rabi splitting of about 127 cm<sup>-1</sup>, which is larger than both 429 the FWHM of the cavity mode (43 cm<sup>-1</sup>) and the vibrational band (39 cm<sup>-1</sup>). 430



Figure 7. (A) Left panel: Schematics of the flow-cell Fabry–Pérot cavity. Right panel: In-432 frared absorption spectrum of DPPA (black) and its transmission spectrum under strong 433 coupling with the Fabry-Pérot cavity modes (red). (B) Left panel: Optical micrographs 434 showing white light transmission through the nanofluidic Fabry-Pérot cavities before and 435 after filling them with toluene. Central panel: Schematic cross-section of Fabry-Pérot 436 nanocavities of thickness d, made of Ag layers and glass substrates. Right panel: Angle-437 resolved dispersion reflection spectra of a Fabry-Pérot nanocavity containing Chlorin e6 438 in dimethylformamide at a high concentration under strong coupling regime. Adapted 439 with permission from George, J., et al. (2015) [20], published by the American Chemical 440 Society (Panel A), and Bahsoun, H. et al. (2017) [81], published by the American Chemical 441 Society (Panel B). 442

Another example of a microfludic Fabry-Pérot microcavity is presented in [12]. Here, 443 an infrared microfluidic cavity was used to investigate cooperative VSC of the solute (p-444 nitrophenyl benzoate) and solvent (isopropyl acetate) molecules. The reflective surfaces 445 consisted of BaF<sub>2</sub> substrates sputtered with Au (10 nm), which were separated by an elas-446 tic Mylar spacer (12  $\mu$ m). VSC was achieved by coupling the cavity mode to the C=O 447 stretching band (1739 cm<sup>-1</sup>) of isopropyl acetate at a solute concentration 100 times lower 448 than that of the bulk solvent. The cooperative VSC of the transesterification reaction de-449 creases the free energy of activation by 2–5 kJ mol<sup>-1</sup>, thereby catalyzing the reaction. 450

In [82] VSC was achieved using the ferrocyanide ion [Fe(CN)<sub>6</sub>]<sup>4</sup> as a model substance 451 in an aqueous solution. The reflective surfaces of the Fabry-Pérot cavity consisted of two 452 rectangular CaF<sub>2</sub> glasses coated with Au. Both glasses had sizes of l = 38.5 mm, w = 19.0453 mm, and h = 4.0 mm, and one of them had 2-mm through holes drilled to form a micro-454 fluidic channel. The results showed tunable strong vibrational coupling of [Fe(CN)6]<sup>4</sup> at 455 moderate concentrations, with a lower limit of about 15 mM, because of its large molar 456 extinction coefficient and narrow resonance bandwidth. The  $C \equiv N$  stretches at 2038.6 457 cm<sup>-1</sup> split into two polariton peaks ~90 cm<sup>-1</sup> apart in a saturated (0.649 M) K<sub>4</sub>[Fe(CN)<sub>6</sub>] 458 solution, with a cavity length of  $6.84 \mu m$ . 459

By functionalizing the sensing area with receptor molecules, the optical cavity can be adapted for biosensing. In [83], a cavity-based optical biosensor has been adapted for biosensing experiments with streptavidin and C-reactive protein. The sensor detected the local RI change due to the adsorption of target biomolecules on the receptor molecules. When the target biomolecules were adsorbed on the receptors, a shift in the resonant response occurred. The fluid control was performed using a small sample volume of 15 μL

for 30 min. The results showed limits of detection (LODs) of 1.35 nM for streptavidin and 466 377 pM for C-reactive protein. 467

Nanofluidic Fabry-Pérot cavities used to study the photophysical properties of the 468 chlorophyll analog Chlorin e6 (Ce6) in both the weak and strong coupling regimes are 469 shown in Figure 7B. The reflective surfaces consisted of two parallel Ag layers on 2.5 cm × 470 2.5 cm glass substrates. The distance between the mirrors was on the order of 100 nm. The 471 nanofluidic channel (a rectangular area of 10–20 µm × 120 µm) was formed using a focused 472 ion beam. Then, a 30-nm Ag layer was evaporated on the substrate to act as the channel 473 bottom mirror. The two substrates were cold-welded by high pressure (~1.6 tons/cm<sup>2</sup>). The 474 thickness of the top mirror layer depended on the required measurements. For instance, a 475 30-nm top Ag mirror was used in the transmission mode and a 200-nm one in the reflection 476 mode. Here, solutions of Chlorin e6 in dimethylformamide (DMF), at a high concentration 477 (0.18 M) were introduced into the nanocavities by capillary forces. The mode splitting (Fig-478 ure 7B, right panel) was due to the strong light-matter coupling and had a value of about 479 110 meV [81]. With these findings, the research group demonstrated how to obtain na-480 noscale liquid Fabry-Pérot nanocavities to study strong light-matter coupling for molec-481 ular solutions. 482

In addition to the conventional configurations for obtaining strong coupling in 483 Fabry-Pérot cavities, another method has been proposed, which uses self-assembled opti-484 cal Fabry-Pérot microcavities called Casimir cavities. These systems can have cavity 485 lengths down to 100–200 nm, enabling SC in the visible range in a tunable regime. Figure 486 8A shows two possible configurations of Casimir cavities, which are either two parallel 487 nanoflakes floating in an aqueous solution or a nanoflake placed above a static mirror. The 488 system loaded with a several-layer WSe2 flake under strong coupling conditions exhibited 489 anti-crossing (see Figure 2) with a Rabi splitting of  $\Omega_R \approx 110$  meV [23]. In addition, the 490 control of objects with a nanometer precision was performed using the critical Casimir 491 repulsion forces to counteract the friction due to the Casimir-Lifshitz forces preventing the 492 contacting surfaces from moving [84]. These results offer an option to fabricate liquid 493 nanocavities with a nanometer precision where maintaining the parallelism of two flat 494 surfaces at small distances has proven to be difficult. 495

Light-matter coupling has also been observed on a carbon nanotube in a Fabry-Pérot 496 microcavity based on a uniform nanoscale-thick film of aligned, packed, single-chirality 497 nanotubes, as shown in Figure 8B. The system displays ultrastrong coupling (with VRS as 498 high as 329 meV or a coupling strength to transition energy ratio of 13.3%) for polarization 499 parallel to the nanotube axis [85]. The system is tunable from weak coupling to strong and 500 ultrastrong couplings. However, the main applications of the system are related to ultras-501 trong coupling, which is beyond the scope of this review. These systems have been re-502 viewed in detail in Refs. [86-88]. 503



Figure 8. (A) Left panel: Two parallel gold nanoflakes floating in an aqueous solution of 505 a ligand (CTAB). Bright-field image of a self-assembled dimer formed from flake I (trian-506 gular) and flake II (hexagonal). Inset: A SEM image of an exemplary gold nanoflake. Cen-507 tral panel: A self-assembled microcavity coupled to a few layers of WSe<sub>2</sub>. Right panel: 508 Anti-crossing of self-assembled microcavity coupled to a few layers of WSe2. (B) Left 509 panel: Schematics of the exciton-polariton device based on a film (with the nanoscale 510 thickness d) of aligned carbon nanotubes. Right panel: The vacuum Rabi splitting  $(\hbar \Omega_{R})$ 511 (red dots) and the extracted coupling strength  $(2\hbar g)$  (blue triangles) continuously tuned 512 via the polarization angle  $\phi$  from strong to weak coupling. Adapted with permission 513 from Munkhbat, B. et al. (2021) [23], published by Springer Nature (Panel A), and Gao, W. 514 et al. (2018) [85], published by Springer Nature (Panel B). 515

#### 4.2. Photonic-Crystal Fiber Plasmonic Sensor

The design of a photonic-crystal fiber (PCF) plasmonic sensor combines the concepts 517 of PCF and plasmonic materials. A PCF is a type of optical fiber that leverages the proper-518 ties of photonic crystals, periodic structures characterized by a photonic bandgap prevent-519 ing waves of specific lengths from propagating in the cladding, thus confining them in the 520 core. This effect is determined by a characteristic microstructure, which consists of an or-521 ganized pattern of air channels running along the entire fiber, designed to confine and 522 guide light. By incorporating a plasmonic material layer into the PCF structure, surface 523 plasmon resonance is enabled, which improves the sensitivity to environmental changes 524 at the metal-dielectric interface. Typically, this involves the deposition of a metal layer, 525 such as gold or silver, on the inner surface of the air channels or on the outer surface of the 526 fiber, and the dielectric material is the medium surrounding the fiber, such as air or a spe-527 cific analyte solution. 528

As shown in Figure 9, the PCF surface plasmon resonance (SPR) sensor employs the evanescent wave generated by the surface plasmonic wave (SPW) propagating at the metal-dielectric interface. This occurs when the frequency of incident light matches the oscillation frequency of surface electrons. The coupling between the SPW and the evanescent field at a specific wavelength creates the resonance condition, leading to the appearance of the confinement loss peak [89]. The confinement loss is calculated from the imagi-534

nary part of the  $n_{eff}$  (effective refractive index) values of the core-guided mode:

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$$\alpha_{loss} \left[ \frac{dB}{cm} \right] = 8.686 \times \frac{2\pi}{\lambda} \cdot \operatorname{Im} \left[ n_{eff} \right] \times 10^4 \,, \tag{18}$$

where  $\lambda$  is the operating wavelength and  $\text{Im}[n_{eff}]$  is the imaginary part of the  $n_{eff}$  of 536 the core-guided fundamental mode, the primary mode of light propagation in PCF. 537



Figure 9. Schematic representation of a photonic-crystal fiber surface plasmon resonance 539 sensor. Left panel: The sensor with an evanescent wave generated by the surface plasmon 540 wave propagating at the metal-dielectric interface. Right panel: The real part of the effective index for the core and surface plasmon polariton (spp) modes and the loss spectrum of the core mode. The wavelength at which complete mode coupling occurs (1700 nm) is indicated. Reproduce with permission from Luo, W., et al. (2023) [111], published by Elsevier.

Due to the sensitivity of the SPW to the RI of the surrounding medium, even a small 546 change in the RI caused by an analyte leads to a shift in the resonance peak. Therefore, 547 these sensors are suitable for applications in medical diagnosis, biological and chemical 548 sensing, and environmental monitoring based on RI measurement [90]. 549

Due to their microchannel structure allowing strong coupling between light and the 550 liquid medium inside, PCFs can be used for both light guiding and fluid transport. The 551 channels in the cladding region of the PCF offer the possibility to impregnate it with bio-552 logical samples or chemicals in the gaseous or liquid form, which can not only penetrate 553 into the fiber core, but also fill the internal and external sensing channels of the fiber. Ad-554 ditionally, the sensitivity and selectivity of the PCF SPR sensor could be improved by 555 functionalizing the microchannels of the PCF with capture molecules binding biomolecu-556 lar targets. 557

The main benefits of combining PCF with the SPR technology are the ability to detect 558 a broad spectrum of analytes under a variety of conditions (making them suitable for var-559 ious applications), compact size (which is important in nanosensors and remote sensing 560 applications), and an extremely high sensitivity. Moreover, PCF sensing requires very 561 small volumes of the analyte (from several tenths to several tens of microliters) due to the 562 small size of the air channels of the PCF core and cladding, as well as intensified interac-563 tion between the sample and light. This possibility of using extremely small sample vol-564 umes is especially valuable for chemical and biological applications, such as identifying 565 toxic chemicals, antibodies, blood components, cells, bacteria, DNA, and viruses. How-566 ever, a disadvantage of the PCF technology is its extremely high cost and a challenging 567 fabrication process. 568

There are various types of PCF plasmonic sensors, each designed for a specific sens-569 ing application by modifying the structure of the PCF and the plasmonic element config-570 uration. The material of plasmonic element (silver, gold, aluminum, copper, bismuth, pal-571 ladium, etc.) also affects the sensitivity of the sensors [91]. For example, compared to other 572

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metals, gold exhibits a higher chemical stability and provides a greater analyte-induced 573 change in the position of resonant peak; silver provides a sharper resonance peak, but it 574 is unstable chemically [92]; and copper oxidizes in an aqueous environment [93]. PCF en-575 ables both SPR and localized SPR (LSPR) [94-96], which differ in that SPR occurs at con-576 tinuous or periodic metal surfaces, e.g., planar films, gratings, nanohole arrays, etc., 577 whereas LSPR is strongly localized by metal nanostructures: nanospheres, nanodisks, na-578 norods, nanowires, etc. [97]. The parameters of the nanostructures, such as geometry and 579 material, determine the resonant frequency of LSPR. For instance, aluminum-based 580 nanostructures are used to obtain LSPR in the UV and blue spectral ranges; silver-based 581 structures, in the range from blue to green; and gold is used for LSPR in the range from 582 red to near-infrared. LSPR-based systems have been used for surface sensing, enhanced 583 photocatalysis, photochemistry [98]. When combined with PCFs, they become particu-584larly suitable for analyzing biological samples and SERS enhancement. The limit of detec-585 tion (LOD) of LSPR is 0.102 ng/mL for zearalenone in sensors using gold nanoparticles 586 ~25 nm in size [99]. In the case of SPR on a continuous gold film in a sensor chip fabricated 587 on the basis of a self-assembled monolayer, the LOD for zearalenone was 7.07 ng/mL 588 [100]. It has been demonstrated that, with increasing number of deposited nanoparticles, 589 the changes in the LSPR maximum decrease but the wavelength shifts increase [101]. 590 These results show that LSPR is more sensitive to molecular binding events at the surface 591 due to its localized field enhancement. However, it is still difficult to deposit nanoparticles 592 uniformly along the optical fiber. 593

Innovative sensor designs have been proposed that use diverse configurations, e.g., 594 photonic quasicrystal fibers (PQFs) characterized by ordered but nonperiodic arrangement of air channels in the cladding region, unique optical waveguide properties, such as 596 birefringence, a low dispersion, and a high confinement loss. 597

To further improve sensing capabilities, multistructural forms have been used and 598 techniques for coating PCF with thin metal layers (e.g., gold or silver) have been improved 599 for enhancing the performance of PCF SPR sensors. In general, a good sensing perfor-600 mance requires a uniform and thin metal coating for reducing the surface roughness and, 601 hence, ensuring laminar analyte flow. An alternative way to form a plasmonic coating in 602 a PCF sensor is the use of metal nanoparticles synthesized by colloidal methods under the 603 conditions ensuring the control of their size. For example, gold nanoparticles can be syn-604 thesized using the citrate reduction method. By controlling the concentration of citrate 605 and the reaction conditions, the size of the gold nanoparticles can be precisely controlled 606 [102]. In addition, the metal layer can also be coated onto the inner surface of the PCF to 607 create a plasmonic layer. Typically, two methods are used to deposit the sensing metal 608 layer in PCFs: chemical vapor deposition (CVD) [103,104] or high-pressure metal pump-609 ing [105,106]. 610

Because the air holes to be coated with metal are only a few micrometers in size, it is 611 challenging to coat their inner surface with metal while maintaining the homogeneity of 612 the film. To overcome this fabrication difficulties, a D-shaped PCF configuration (Figures 613 10) has been proposed for an external sensing approach. These sensors are fabricated by 614 flattening one side of the PCF (thus creating a D-shaped cross-section) and coating the flat 615 side with a metal layer. In the case of a D-shaped sensor, the "stack and draw" approach 616 or capillary stacking technique could be used for the fabrication [107]. For the formation 617 of air channels, capillaries are stacked to form the core of the PCF, which is then drawn to 618 obtain a PCF cane, after which a protective tube is added to the surface of the PCF to 619 obtain the desired sensor structure. Afterwards, a flat surface is obtained by polishing the 620 upper half of the fiber [108], and a micro-opening channel is formed by focused ion-beam 621 milling or femtosecond laser micromachining. In order to form the SPR film in the chan-622 nel, chemical vapor deposition of metal is used [109]. In addition, atomic layer deposition 623 (ALD) could be considered for applying a gold layer onto curved surfaces. Finally, the 624 bio-analyte is applied onto the metal layer for subsequent measurements [110]. Several D-625



shaped PCF SPR biosensors, including those based on two-dimensional plasmonic materials, such as graphene and MoS<sub>2</sub>, have been reported recently. 627

Figure 10. (A) Schematic representation of a D-shaped photonic-crystal fiber (PCF) surface 629 plasmon resonance biosensor for refractive index sensing. Left panel: PCF cross section. 630 Central panel: PCF general view. Right panel: Schematics of the sensor operation. (B) Left 631 panel: A micrograph of the PCF cross-section before polishing. Central panel: The cross-632 section of the D-shaped sensor. Right panel: A SEM image of the polished and Au-coated 633 D-shaped PCF. Split air holes can be seen. (C) Experimentally detected spectrum at differ-634 ent RIs. Adapted with permission from Luo, W., et al. (2023) [111], published by Elsevier 635 (Panels B and C). 636

Later, the D-shaped sensor was theoretically adapted for simultaneous detection of 637 changes in the RI in two separate wavelength ranges; the adapted sensor is referred to as 638 a dual-band D-shaped PCF SPR sensor [112]. In this case, an Au-TiO2 grating is deposited 639 onto the flat surface of a D-shaped PCF, and then SPRs are excited for detecting the change 640 in the RI of the analyte through analyzing the loss spectra and birefringence. The maxi-641 mum sensitivity of the sensor is 26,700 nm/RIU, and the RI resolution is  $3.75 \times 10^{-6}$  RIU. 642 Furthermore, a Rabi-like loss spectrum splitting (Figure 11) was simulated by making a 643 small hole in the fiber. In this case, Rabi-like splitting results in two hybrid modes (loss 644 peaks) with higher and lower frequencies. It has been shown that the two loss peaks allow 645 the detection of RI changes in the corresponding spectral range, which makes the sensor 646 even more useful. 647



**Figure 11.** Loss spectra of the sensor with different (a–c) diameters of the air holes and (d)  $^{649}$  hole-to-hole spacings ( $\Lambda$ ). Reproduced with permission from Ma, Y., et al. (2023) [112],  $^{650}$  published by Elsevier.  $^{651}$ 

The integration of PCF SPR sensors with microfluidic systems has ensured precise 652 control of sample introduction and enhanced sensor functionality. It should be noted that 653 the sample solution filling the microfluidic channels of the PCF should flow in the same 654 direction as the light propagates during sensing. Therefore, misaligning the PCF device 655 can affect the coupling condition and distort the results. This issue can be addressed by 656 using an external microfluidic channel, which, in addition, simplifies the fabrication. For 657 example, an external microfluidic module can be added to a D-shaped PCF sensor to ob-658 tain a sealed system where the solution on the surface of the metal coating can be replaced 659 without disturbing the internal setup. Moreover, functionalization of PCF microchannels 660 with biomolecules improves the selectivity and sensitivity of PCF SPR sensors for biolog-661 ical applications. The fabrication of PCF SPR sensors is described in detail in Refs. [89, 92]. 662 Recently, PCF-based biosensors with single-channel, semi-channel and multi-channel 663 configurations have been proposed for detecting analytes with different RI values [113-664 115]. In addition, the application of machine learning algorithms to the analysis of PCF 665 SPR sensor signals is used to improve the accuracy and reliability of measurements. 666

Theoretical and computational research is being carried out in order to develop 667 highly sensitive and portable PCF SPR sensors. For example, the finite-element method 668 (FEM) is used to determine the optimal geometric parameters of such systems[116,117]. 669 In [118], two different configurations of RI biosensors based on photonic quasicrystal fi-670 bers (PQF-RIBS) with six- and eightfold air hole patterns have been simulated by the FEM. 671 The results show that, for the sixfold configuration, the maximum sensitivity is 28,000 672 nm/RIU for an analyte RI range of 1.41–1.42, the figure of merit being 491. In the case of 673 the eightfold configuration, the maximum sensitivity is 27,000 nm/RIU for an analyte RI 674 range of 1.40–1.41, the figure of merit being 231. Both systems operate in a wide range of 675 wavelengths, from 600 to 1300 nm. In the system with an eightfold arrangement of the air 676 holes, the *n*<sub>eff</sub> is reduced and confinement loss is increased compared to the system with a 677 sixfold one. This also indicates a strong mode-field coupling in the eightfold PQF-based 678 RIBS. 679 Thus, the combination of biosensors based on PCF or PQF with microfluidics has already been tested experimentally, with theoretical and experimental research still underway in order to further improve the sensing properties of these devices.

#### 4.3. Plasmonic Nanocavity

Surface plasmons are collective electron oscillations that arise at the interface be-684 tween two materials, e.g., metal and dielectric, in such a way that the real part of the die-685 lectric function changes its sign across the interface [119]. Plasmonic nanocavities are opti-686 cal structures that use surface plasmon resonance to confine light at the plasmonic reso-687 nance frequency. Plasmon resonance could be either localized (e.g., in metal nanoparticles) 688 or delocalized (e.g., in thin metal films) [120]. These structures are used to obtain strong 689 enhancement of local electromagnetic fields, required for coupling with target molecules. 690 The EM field is enhanced at certain locations, such as tips, sharp corners, or junctions in 691 plasmonic nanoparticles (NPs), generating focused "hot spots" with sizes similar to those 692 of single molecules. Therefore, although plasmonic nanocavities have low Q-factors and 693 high ohmic losses, these structures are particularly attractive for achieving strong field 694 confinement with a subwavelength effective mode volume (Figure 4), making it possible 695 to obtain strong coupling at the level of one or several molecules [121]. The physical sizes 696 of plasmonic cavities can be smaller than the diffraction limit of light, in contrast to those 697 of conventional cavities, which are limited by the wavelength. However, it is important to 698 note that the coupling strength is determined mainly by the dipole moment of the analyte 699 and its concentration in the mode volume, regardless of the cavity type [39].

Theoretically, plasmonic nanocavities are usually described in terms of the Jaynes-Cummings model considered in the previous section. The coupling strength of a system (e.g., a dipole) to an isolated nanocavity can be expressed as 703

$$2g = \sqrt{(\Gamma_{tot} - \Gamma_0)\kappa} , \qquad (19)$$

where  $\Gamma_{tot}$  is the decay rate of the system coupled to the isolated nanocavity,  $\Gamma_0$  is the 704 decay rate of the system in free space, which is determined as the sum of its radiative and 705 nonradiative decay rates, and  $\kappa$  is the total loss rate of the cavity ( $\kappa = \kappa_{rad} + \kappa_{nrad}$ ). 706

Experimentally, since the demonstration of the confinement of light in nanometer 707 gaps between metal nanoparticles, plasmonic nanocavities have been employed for en-708 hancing the Raman scattering signal [122,123], and the photoluminescence or fluorescence 709 intensity [124,125] thus substantially increasing the sensitivity of plasmonic nanocavities. 710 Later, their performance has been improved by integration with conventional photonics 711 [126]. With the demonstration of strong coupling between light and matter in plasmonic 712 nanocavities and the improvement of fabrication techniques, such as electron beam lithog-713 raphy and focused ion beam milling, more precise and complex plasmonic nanocavity 714 designs became feasible, including the integration with microfluidics. In order to confine 715 light efficiently, a plasmonic nanocavity should match a number of parameters, such as 716 the optimal roughness of metal structures [78], energy loss, reactivity, and resonant region 717 (plasmon tuning range) [119]. There are various configurations combining plasmonic 718 nanocavities with microfluidics, including isolated plasmon cavities, dimers, arrays of na-719 noparticles called plasmonic lattices, and nanoparticle-on-mirror configurations [127,128], 720 which are described below. 721

In [52], an ultrasensitive biomechanical nanosensor was proposed and the coupling 722 of J-aggregates to an isolated cuboid Au@Ag nanocavity in a microfluidic environment 723 was analyzed (Figure 12A). The dye concentration was continuously varied in the nanoscale space adjacent to the plasmon hotspots of the Au@Ag nanocavity to observe the 725 emergence and evolution of light–matter coupling. This tunability supported by microfluidics allowed obtaining a coupled J-aggregate exciton number from 0 to ~3 and Rabi 727 splitting of up to 167 meV. 728

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The fabrication of dimers by placing gold or silver nanoparticles close to one another 729 has been reported. These nanoparticles can interact with one another through their plas-730 monic fields, which leads to unique optical properties that differ from those of individual 731 nanoparticles [129]. In [130] QDs were embedded in plasmonic cavities formed by silver 732 bowties on 18-nm SiO<sub>2</sub> membranes. The QDs are positioned into the gap region of the 733 bowties using interfacial capillary forces. The authors succeeded in increasing the cou-734 pling strength by improving the fabrication of the QD–plasmonic cavity device by reduc-735 ing the thickness of the adhesion layer under the plasmonic cavity. The results have 736 shown coupling strength values in the range of 50–110 meV (Figure 12B). The possibility 737 of further enhancing the coupling to individual QDs could make it possible to control the 738 chemical reactivity, manipulate the excited states, and engineer single-photon sources op-739 erating at room temperature for quantum informatic applications. 740

Moreover, for better tunability and performance, plasmonic nanocavities can be fab-741 ricated in the nanoparticle-on-mirror configuration [131]. For instance, nanoantennas 742 have been used to obtain vibrational strong coupling. These systems also could be inte-743 grated with nanofluidics [132]. Figure 12C shows the strong coupling of gap surface plas-744 mons and molecular vibrations in an ultracompact hybrid system based on a nanogap 745 patch antenna, with a Rabi splitting as high as 108 cm<sup>-1</sup>. Other studies support the possi-746 bility of integrating these configurations with nanofluidics in order to obtain strong cou-747 pling of molecules in the liquid phase [133]. 748

Multifunctional plasmonic nanocavities capable of multiple sensing have been engi-749 neered by fabricating plasmon ensembles to improve optical trapping and enhancement 750 of the electric field intensity in light-matter interaction. For example, a plasmonic biosen-751 sor for the detection of bisphenol A at ultralow concentrations consisting of an array of 752 gold nanoantennas has been fabricated on the basis of an optical fiber with a D-shaped 753 cladding [126] (Figure 12D). The fluidic cell used a programmable syringe driver 754 equipped with two isolation valves, on the delivery and exit ports of the fluidic cell, to 755 hold and immerse the sensor. The maximum volume of solution contained in the cell was 756 1.9 mL. The device could detect a small amount of bisphenol A, providing a wavelength 757 shift of  $0.15 \pm 0.01$  nm in response to the analyte concentration of 1 fM, with a detection 758 limit of  $330 \pm 70$  aM. 759

Moreover, hybrid structures fabricated of plasmonic and 2D materials have been de-760 veloped, e.g., an Ag nanoantenna combined with a layered WS<sub>2</sub> flake [134] and ultra-761 compact plasmonic nanocavities containing WSe2 with a Rabi splitting exceeding 135 meV 762 [135]. Another example is a fluidic system consisting of self-organized gold nanocavities 763 enhanced with graphene nanosheets for plasmon-assisted electrochemical detection of 764 H<sub>2</sub>O<sub>2</sub> released from cancer cells, with an LOD of 1 pM in a linear range of 1 pM–10  $\mu$ M. 765 Here, Au nanoparticles generated strong electromagnetic fields resulting in a high con-766 centration of electric charge carriers at the electrode surface, and graphene further en-767 hanced the electrode properties [136]. Plasmonic nanocavities strongly coupled to excitons 768 in 2D materials consisting of one or several layers are characterized by room-temperature 769 functioning, miniature size, and high stability. 770



Figure 12. (A) Left panel: Schematics of the nanostructured microfluidic device for detec-772 tion of H2O2. Right panel: Dynamic tuning of the plasmon-exciton coupling in the micro-773 fluidic channel: emergence and evolution of hybrid polariton states in scattering spectra. 774 (B) Top panel: STEM images of a device with two quantum dots trapped within plasmonic 775 bowties. Bottom panel: The dark-field scattering (green) and photoluminescence (PL) 776 (red) spectra of the device. (C) Left panel: Schematics of the nanogap patch antenna. Cen-777 tral panel: A SEM image of the fabricated patch antenna (top view). Right panel: Measured 778 reflectance spectra of the nanogap patch antenna cavity for different periodicities of the 779 array. The solid curve shows the measured reflectance from a single nanogap patch an-780 tenna. (D) Schematics of a plasmonic biosensor capable of detecting bisphenol A at ul-781 tralow concentrations. Adapted with permission from Liang, K., et al. (2021), published 782 by American Chemical Society [52] (Panel A); Gupta, S. N., et al. (2021), published by 783 Springer Nature [130] (Panel B); Dayal, G., et al. (2021), published by American Chemical 784 Society [133] (Panel C), and Allsop, T.D.P., et al. (2019), published by Elsevier [126] (Panel 785 D). 786

Research in plasmonic nanocavities and microfluidics continues to bring novel ap-787 plications. However, challenges still remain preventing the use of the full potential of 788 plasmonic cavities, the key ones being the requirements of precisely and controllably po-789 sitioning molecules in nanosized plasmonic mode volumes, determining the best plas-790 monic cavity geometry, finding how to separate useful photons from background ones, 791 dealing with the fluorescence quenching in the vicinity of metal particles [49], and pre-792 cisely determining the plasmon field intensity distribution in the nanogap. In order to 793 prevent molecular aggregation of the emitters and to ensure that the transition dipole 794

moment is perfectly aligned with the gap plasmon (along the electric field), a host-guest 795 chemical approach has been used. An example is the confinement of methylene blue using 796 the macrocyclic cucurbit[n]uril molecules. Cucurbit[7]uril is water-soluble and can host 797 only one methylene blue molecule inside. Without the host, the methylene blue molecules 798 oriented perpendicular to the plasmon field exhibit only a gap plasmon [137]. In contrast, 799 the host enables the alignment of the molecule parallel to the plasmon field, in which case 800 the spectrum exhibits peak splitting due to the strong interaction between the molecule 801 and the plasmon. In addition, residual molecules, such as dyes, in the microfluidic channel 802 may bring ambiguity in analyzing the coupling process. Therefore, to minimize the influ-803 ence of uncoupled dye, the microfluidic device is flushed several times with deionized 804 water before spectral measurements. Simulation of the electric field predicts that only the 805 dye molecules attached closely to the nanotips could contribute to the strong coupling 806 process, while the other molecules should have low impact on the hybrid system [52].In 807 addition, strong coupling has been theoretically predicted between semiconducting sin-808 gle-walled carbon nanotubes and metal nanoelectrodes, with a Rabi splitting of  $\hbar\Omega \approx 127$ 809 meV and an augmented Purcell effect (F = 105) at a source–drain voltage of 1.75 V. These 810 results show that the strong coupling regime and boosted Purcell effect can be reached in 811 electrically driven plasmonic junctions [138], which could be another option for combin-812 ing with fluidics. 813

#### 4.4. Whispering-Gallery Mode Microcavity

In a WGM microcavity, light propagates in the form of circulating waveguide modes 815 confined in a microscale volume through total internal reflection and constructive inter-816 ference on a closed concave surface. This light is trapped into the cavity via an adjacent 817 linear waveguide positioned within a couple of hundred nanometers. Part of the EM field 818 confined on the closed concave surface extends beyond the cavity surface; this part is re-819 ferred to as the evanescent field. Regarding photonic crystal fiber plasmonic sensors, op-820 tical WGMs are highly sensitive to the surrounding medium (analyte solution), and the 821 changes around the evanescent field can lead to changes in the resonance mode. There-822 fore, when an analyte interacts with the evanescent field, it alters the local RI. For example, 823 biomolecule binding (e.g., antibody-antigen interaction) on the surface of the cavity leads 824 to a change in the effective RI and a shift of the resonant wavelength (Figure 13). This 825 effect is used for biochemical, temperature, and mechanical sensings. 826



Figure 13. A schematic diagram of the principle of a whispering-gallery mode microcavity828(microring resonator) used for biosensing and the corresponding optical loss spectrum.829Reproduced with permission from Park, M.K., et al. (2013), published by Elsevier [139].830

Due to the strong light confinement, WGM microcavities have been demonstrated to be excellent candidate devices for obtaining strong coupling between an optical mode and 832

827

a system [140,141]. These cavities are characterized by their high Q-factor (up to 1011), ex-833tremely small mode volume, and high optical energy density and have been used for de-834tection and highly sensitive measurements on the microscale and nanoscale, including835single-molecule detection.836

The main parameters analyzed in WGM microcavities, in addition to the Q-factor 837 (Equation (8)) and mode volume (Equation (10)), are the optical resonance condition and 838 sensitivity. The optical resonance condition for WGMs is the following: 839

$$m\lambda = 2\pi R_b n_{eff} , \qquad (20)$$

where *m* is an integer number,  $\lambda$  is the wavelength,  $R_b$  is the radius of the resonator, 840 and  $n_{eff}$  is the effective RI. WGM microcavities are characterized by a high sensitivity, 841 which is important for numerous applications. When a molecule lands on the surface of 842 the microcavity, the circulating light interacts with it, generating the detection signal with 843 a time resolution of microseconds suitable for label-free detection. 844

WGM microcavities are fabricated with various geometries, sizes, and materials 845 [142,143]. The most common WGM structures for sensing are microbottles [144,145], microdisks [146,147], microspheres [148,149], microtoroids [150], microcapillaries [151,152], 847 and microrings [153,154]. Table 1 summarizes the main methods used for the fabrication 848 of different WGMs. The sensing in these cavities could be based on mode shifting [155], 849 broadening [156] or splitting [141], as shown in Figure 14. 850

Table 1. Fabrication methods for the different whispering-gallery mode microcavity

Whispering- gallery mode micro- cavity	Scheme	Fabrication method	Ref.
Microsphere		Heating of an optical fiber tip with CO2 laser, arc discharge, H2 flame	[140]
Microbubble	*	Local heating of a capillary with CO <sub>2</sub> laser, arc discharge, H <sub>2</sub> flame, while applying internal aerostatic pressure to inflate the SiO <sub>2</sub>	[140]
Microbottle	**	<ul> <li>Heating and stretching two regions into thinner ones.</li> <li>Heating and softening an optical fiber and squeezing it along the axis to build up material in the pinched region</li> </ul>	[157]
Microdisks	+ 	Lithographically forming disks by wet and dry chemical etching	[158]



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Figure 14. Regimes of sensing using whispering-gallery mode (WGM) microcavities:857(A) mode shifting, (B) mode broadening, and (C) mode splitting.858

Mode shifting (Figure 14A) occurs when the effective RI is altered with changes in 859 the surrounding medium. For example, a red shift usually results from an increase in the 860 concentration. The changes in the effective RI can also be influenced by the variations of 861 the temperature and magnetic field due to the thermo-optical effect and magneto-optical 862 effect, respectively. Variation of the geometry of the resonators can be caused by changes 863 in the pressure, flow rate, stress, tension, and thermal expansion [160]. For example, in the 864 case of microbubbles, the wall thickness is selected depending on the practical application: 865 a thin wall is always used for the detection of media inside the microbubble, whereas a 866 thick wall, for physical sensing, such as temperature sensing. In addition, to avoid errors 867 when different substances in a solution cause the same change in the bulk RI, receptor 868 molecules are integrated for selective detection. This method is sufficiently sensitive for 869 monitoring single-molecule biochemical reactions [161]. Because the sensor sensitivity can 870 be affected by external perturbations, packing is used to enhance stability. For example, 871 glass scaffolds and low-RI polymers are used for protecting coupling systems and optical 872 coupling regions, respectively. The process involves a portable, low-cost fiber fusion 873 splicer for electrode-discharge heating [162]. 874

In mode-broadening sensing (Figure 14B), the detection is performed by monitoring 875 the WGM linewidth [156]. The spectral linewidth of the resonance mode is related to the 876 Q-factor, as seen from Equation Ошибка! Источник ссылки не найден., and changes 877 in the Q-factor are caused by variations of the optical loss. Therefore, the spectral lin-878 ewidth increases with increasing cavity losses, such as scattering and radiation losses. In 879 contrast to the wavelength shift, mode-broadening sensing is unaffected by changes in the 880 external medium due to its self-referenced mechanism. Therefore, it has smaller detection 881 limits for small molecules and single nanoparticles. 882

Mode splitting (Figure 14C) occurs in WGM microcavities due to the propagation of 883 degenerate modes clockwise (light circulating clockwise around the cavity) and counter-884 clockwise (light circulating counterclockwise around the cavity) as a result of the scatter-885 ing of light from subwavelength scatterers, such as nanoparticles, entering the resonator 886 mode volume. As a consequence, the eigenstates are transformed into two orthogonal 887 standing waves in the cavity with a mode splitting in the transmission spectrum [141]. 888 The size and number of the scatterers can be determined by measuring the frequency 889 splitting. In contrast to mode broadening, mode splitting requires a cavity with an ultra-890 high Q-factor so that the two modes could be distinguished. 891

Generally, WGM devices based on a silicon photonic platform have been developed for multiplexed biological analysis [163]. For example, Figure 15A shows a double-slotwaveguide-based microring resonator for the detecting hemoglobin and measuring its concentration in anemia. A sensitivity as high as 1024 nm/RIU with the minimum deflection limit of 14.88 × 10<sup>-6</sup> RIU was obtained [154].

The versatility of WGMs allows for the integration of additional components, such 897 as plasmonic nanoparticles, ranging from single particles to arrays to obtain optoplas-898 monic WGM sensors capable of single-molecule detection [164,165]. As mentioned above, 899 localized surface plasmon resonance allows light confinement beyond the diffraction limit 900 [166], boosting the near-field intensity by a factor of over 1000. Figure 15B [167] shows an 901 ultrasensitive optofluidic biosensor representing a microbubble resonator. The resonator 902 enables sensing of biomolecules with a detection limit of 0.3 pg/cm<sup>2</sup> and facilitates single-903 molecule detection by using plasmonic nanoparticles. This sensor exhibits ultrasmall sam-904 ple consumption (down to 10 pL) and provides an automated platform for biomedical 905 analysis. 906

The integration of WGM microcavities with microfluidics enhances their functional-907 ity and opens new research avenues in strong light-matter interactions. For instance, sili-908 con ring resonators for high-throughput sensing of specific interactions of biomolecules 909 have been reported [163,168,169]. At the same time, continuous advancements in photonic 910 integration had led to the development of on-chip WGM systems, paving the way for 911 compact and efficient photonic devices for communication, sensing, and computation. 912 Figure 15C shows an example of on-chip WGM sensors integrated with microfluidic chan-913 nels. In this study, silicon nanoclusters were used as microresonators. The silicon disk has 914 a nanogap structure 25 nm in width where the target biomolecules are selectively detected 915 with the sensitivity enhanced by strongly confined field. The sensitivity confirmed by 916 real-time measurements for the streptavidin-biotin complex is 0.012 nm/nM, which is 917 over 20 times larger than the previously reported WGM sensors with remote readout 918 [170]. 919

Additionally, Figure 15D shows a WGM microdroplet sensor for real-time and highsensitive detection of acetylcholinesterase and its inhibitors, as well as the acetylcholinesterase enzymatic reaction. For example, this device has been demonstrated to detect pesticides. This device provides the limit of detection as low as 0.1 pg/mL for fenobucarb and 1 pg/mL for dimethoate, which is considerably lower than the standard levels of pesticides specified in water quality standards [171].



Figure 15. (A) Schematics of a ring resonator (B) Schematics of an optofluidic microbubble 927 biosensor. (Left panel) Analytes enter the microfluidic channel and are captured by the 928 interior surface of the cavity. (Right panel) Schematics of a gold nanorod adsorbed on the 929 interior surface of the microbubble cavity of the Au-coated microsensor. (C) (a) A SEM 930 image of a silicon disk resonator. (b) A magnified image of the silicon disk, with a 25-nm 931 gap indicated. (c) Simulation of the transverse magnetic fundamental mode in an aqueous 932 environment. (d) A photograph of the on-chip sensor integrated with a microfluidic chan-933 nel. (D) Schematics of a microdroplet sensor. Reproduced with permission from Baha-934 doran, M., et al. (2022), published by Springer Nature [154] (Panel A); Yu, X.C., et al. 935 (2022), published by the National Academy of Sciences of USA [167] (Panel B); Kim, Y. 936 and Lee, H. (2019), published by Optica, formerly known as the Optical Society of America 937 (OSA) [170] (Panel C), and Duan, R., et al. (2020), published by Elsevier [171] (Panel D). 938

Some of the advanced technologies have been already transferred to industry and are 939 commercially available, e.g., EPIC, ResoSens, and Creoptix WAVE for label-free tests, as 940 well as Maverick. The latter, in particular, is based on silicon photonics, with a single unit 941 including 16 microring resonators, so that 16 analytes can be detected in a single test with 942 a low sample consumption (5–10  $\mu$ L) within 10 min [172]. The results obtained with the 943 test are in a good agreement with standard methods. In addition, the system has particular 944 advantage over standard clinical tests: it allows to simultaneously perform 12 autoanti-945 body detection assays in whole blood for clinically diagnosed patients in less than 15 min. 946 However, essential bulky elements like the external light source, fluidic pump, spectrom-947 eter, and data processing units remain unavoidable [169,173]. 948

Ongoing research aims to achieve and use strong coupling regimes in WGM micro-949 cavities for exploring new physical phenomena and improving device performance. Ef-950 forts are also made to optimize protocols for detection of biomolecules. Key challenges 951 include liquid evaporation [28], stability, high sample preparation requirements, clogging 952 of microfluidic channels during experiment, and ensuring precise geometry of the reso-953 nators. A protocol has been developed that encompasses several steps addressing these 954 challenges in the functionalization and detection of protein biomarkers using silicon mi-955 croring resonator chips, specifically for detecting Ebola virus (EBOV) and Sudan virus 956 (SUDV) soluble glycoprotein (sGP). This protocol can be adapted for other analytes, in-957 cluding cytokines, chemokines, nucleic acids, and some viruses [174]. 958

Different computational tools are used in designing and studying WGMs [175], using 959 FDTD [176], FEM [177], and the Monte Carlo approach [178]. In addition, machine 960

972

learning has been implemented to automatically extract the spectrum characteristics, the961average accuracies of single-parameter and double parameter identification being 99.5962and 97.0%, respectively [179].963

In conclusion WGM devices offer high sensitivity for biological analysis and versatility, allowing for adding other components, such as plasmonic nanoparticles, that enhance their detection capacity. WGM sensors combined with microfluidics are efficient biomedical devices, used, e.g., in the EPIC and Maverick technologies. Despite challenges such as liquid evaporation and channel clogging, ongoing research and development of advanced computational tools are expected to improve these sensors for broader applications.

#### 5. Potential Applications of Strong Light–Matter Coupling in Micro- and Nanofluidics

Light-matter interaction, in particular, strong light-matter coupling, can be employed in various fluidic applications, including monitoring of chemical reactions, modification of chemical pathways, sensing, and nanophotonics. In this section, we discuss the state of the art in some of these application niches. 976

Control of chemical reactions using new methods, such as polaritonic chemistry and 977 molecular polaritonics, which fundamentally differ from traditional ones, such as electro-978 chemistry, thermochemistry, and photochemistry, attracts much attention from both ex-979 perimental and theoretical viewpoints. Strong light-matter coupling can modify chemical 980 reactions by altering the energy landscape and the reaction rates. This could be used to 981 improve the efficiency of catalysis, photochemical reactions, and other processes in micro-982 /nanofluidic systems. However, the multiscale nature of polaritonic chemistry poses 983 unique challenges in both experimental and theoretical research. The discovery of new 984 reaction pathways enhanced by light-matter interactions requires comprehensive experi-985 mental study, as well as new theoretical insight and advanced computational models. 986 Dealing with large ensembles of molecules and a large number of degrees of freedom in 987 optical cavity at wavelength scale requires careful approximations. In addition, even in 988 the case of a single molecule or a small number of molecules under study, the theoretical 989 interpretation of polaritonic effects is challenging because of large loss rates and the at-990 omistic details, i.e., the intricate and specific characteristics of individual atoms and mol-991 ecules in the system, including their positions, electronic states, interactions, and dynam-992 ics. Furthermore, the consideration of quantum effects, such as tunneling, at the nanopar-993 ticle-molecule interface requires a multiscale layered technique. This helps to model com-994 plex systems where different scales (e.g., atomic, molecular, and macroscopic) are consid-995 ered simultaneously. This technique allows for the integration of detailed atomic-level 996 interactions with larger-scale phenomena, providing a comprehensive understanding of 997 the system's behavior. 998

Advances in machine learning (ML) models trained on ab initio calculations have sig-999 nificantly promoted the study of strong light-matter coupling by improving the predic-1000 tive accuracy and computational efficiency. These ML models, especially deep learning 1001 networks, can capture complex patterns in ab initio datasets, enabling precise predictions 1002 of the properties of systems experiencing strong coupling. They expedite simulations that 1003 would otherwise be computationally intensive, making the study of large and complex 1004 systems feasible [180,181]. Recently, a theoretical model based on a combination of a ma-1005 chine learning model trained on *ab initio* calculations and modular cavity molecular dy-1006 namics has been proposed to study the effect of strong light-matter coupling on the Si-C 1007 bond in an experimentally investigated  $S_N2$  reaction. It has revealed a frequency depend-1008 ence of the rate constant and identified three distinct regimes of kinetic changes [25]. They 1009 include strong inhibition effect without a clear vibrational contribution, vibrationally sup-1010 ported catalysis, and vibrationally supported inhibition, each affecting enthalpy and en-1011 tropy differently. The findings are consistent with some experimental observations but 1012 highlight discrepancies with previous ab initio calculations, suggesting that the dynamic 1013 change in electronic polarization induced by nuclear motion and mediated by the cavity 1014 is important to be considered in the theoretical model. 1015

Chemistry-related applications of strong coupling typically employ Fabry-Pérot cav-1016 ities and plasmon nanocavities. Photochemical reactions, such as photobleaching, have 1017 been demonstrated to be controlled and suppressed by the degree of plasmon-exciton 1018 coupling and detuning between organic molecules and plasmonic nanocavities [182]. De-1019 tuning refers to the difference in energy between the resonant frequency of the cavity (or 1020 plasmonic nanocavity) and the electronic transition frequency of the molecules. By adjust-1021 ing this energy difference, the strength of interaction between light and matter can be 1022 modified to control the reaction dynamics. When the system is precisely tuned, it results 1023 in strong coupling and the formation of new polaritonic states that can either enhance or 1024 suppress chemical reactions. 1025

Ebbesen's research group has made pioneering contributions to the field of strong 1026 light-matter coupling, elucidating its impact on chemical reactions and molecular prop-1027 erties. In 2012, Ebbesen's group demonstrated that the spiropyran-merocyanine pho-1028 to isomerization could be suppressed through strong coupling to a specific electronic tran-1029 sition of merocyanine, resulting in new polaritonic states that alter the energy landscape 1030 of the reaction [183]. This discovery highlighted the potential of strong coupling to control 1031 reaction dynamics. The group explored vibrational strong coupling (VSC) in ground-state 1032 chemical reactions to show that modes of vibrational coupling of molecules to a cavity 1033 could enable control of the reaction rates, as demonstrated with the silane deprotection 1034 reaction of 1-phenyl-2-trimethylsilylacetylene (PTA) with tetra-n-butylammonium fluo-1035 ride (TBAF) [21,184]. Additionally, they developed Fabry-Pérot microfluidic flow cells to 1036 study reactions in solution under strong coupling conditions [20]. These and other insights 1037 pave the way for applications in catalysis and materials science, offering the potential for 1038 more efficient and selective chemical processes through fine tuning of molecular transi-1039 tions [34,45,185]. Further studies of the effect of VSC on a series of carbonyl reactants un-1040 dergoing Prins cyclization have shown a significant decrease in the reaction rate [186]. 1041 This exploration sheds light on the potential of strong coupling to manipulate and control 1042 chemical reactions, including those involving complex ring-forming processes, such as 1043 Prins cyclization. Recently, resonant suppression of the intracavity alcoholysis reaction 1044 between phenyl isocyanate and cyclohexanol has been reported [187]. Experiments have 1045 also shown that, under USC, the interaction between molecular vibrations and photons is 1046 intensified, which leads to enhanced self-interaction effects within the vibrational modes, 1047 which has been found through analyzing the increase in the effective oscillator strength 1048 of the asymmetric stretching band of CS2. This research provides valuable insights into the 1049 intricate dynamics of light-matter interactions at the molecular level and contributes to 1050 the understanding of USC effects on molecular vibrations [188]. Another promising direc-1051 tion for advancing catalytic processes is the study of cooperative VSC [46]. 1052

Numerous materials have been studied in the strong coupling regime, including J-1053 aggregates [189], organic dyes [190], organic crystals [191], organic semiconducting poly-1054 mers [192], carbon nanotubes (CNTs) [193], proteins [194], and chlorosomes [195]. J-ag-1055 gregates are preferred for their strong interaction with light and cost-effectiveness. Recent 1056 research has demonstrated enhanced coupling with MB J-aggregates in plasmon nanocav-1057 ities, although excessive concentration or increased nanoparticle size reduced the cou-1058 pling strength [196]. Other J-aggregates, such as the JC-1 cyanine dye, have also been stud-1059 ied [197,198]. Some organic dyes (e.g., BODIPY dyes) have high absorption and quantum 1060 yield, but they tend to crystallize, necessitating the use of polymer matrices to prevent 1061 inhomogeneity [199]. Organic single crystals in microcavities are preferable for polariton 1062 Bose-Einstein condensation due to their structural order and high exciton interaction 1063 [200,201]. Organic semiconductor-based microcavities offer high oscillator strengths and 1064 flexibility, which makes it possible to fabricate flexible photonic devices that operate at 1065 room temperature. However, they face challenges such as environmental degradation, 1066 low charge mobility, fabrication difficulties, and thermal instability [192]. CNTs are 1067 promising due to their strong exciton-photon interactions and possibility of functionali-1068 zation, which extends the potential optoelectronic applications. Specifically, single-walled 1069 CNTs (SWCNTs) exhibit narrow-band exciton absorption and strong exciton-photon in-1070 teractions, making them ideal candidates for generating exciton-polaritons at room tem-1071 perature [202,203]. Some organic materials, e.g., enhanced green fluorescent protein, ex-1072 hibit USC, offering superior photonic properties, such as high quantum yield and photo-1073 stability. They enable the formation of stable excitons and polaritons at room temperature, 1074 surpassing the limitations of traditional semiconductor systems [194]. Research in chloro-1075 somes from photosynthetic bacteria suggests potential applications in artificial light-har-1076 vesting devices and the development of "living polariton" systems [195]. 1077

Additionally, strong light-matter coupling enhances the sensitivity of micro- and 1078 nanofluidic systems by monitoring changes in the optical properties of the system, such 1079 as fluorescence, absorbance, and changes in the RI. This enhancement allows the detection 1080 and quantification of analytes with high precision. Nanofluidic systems offer a unique 1081 platform for studying strong light-matter coupling due to their capacity for confining and 1082 manipulating a fluid flow at the nanoscale. By integrating nanofluidic channels with op-1083 tical resonators or cavities, a highly efficient real-time sensing platform can be engineered. 1084 One of the key advantages of strong light-matter coupling in micro-/nanofluidics for sens-1085 ing applications is its capacity for enhancing the interaction between light and analytes, 1086 leading to better sensitivity and lower detection limits. By tuning the optical properties of 1087 the system, researchers can achieve increased signal-to-noise ratio and improved detec-1088 tion capabilities, making strong light-matter coupling a powerful tool for sensing appli-1089 cations with a potential in various fields, including healthcare, in particular, point-of-care 1090 diagnosis [162], as well as biomolecule sensing, environmental monitoring, and security, 1091 where the detection of low concentrations of analytes is critical. However, further research 1092 is needed to fully harness the potential of this technology for practical sensing applica-1093 tions. 1094

Sensor systems employing the strong coupling effects can be based on different con-1095 figurations, including Fabry-Pérot microcavities [83,204] and plasmonic nanocavities 1096 [205]. A novel approach to biomolecular detection employs a combination of Fabry–Pérot 1097 cavities and surface plasmon photodiodes. This method makes it possible to enhance the 1098 sensitivity and specificity of biosensing by leveraging the strong coupling between surface 1099 plasmons and the Fabry-Pérot cavity modes. It has been demonstrated that the device can 1100 detect low concentrations of antigens, such as the SARS-CoV-2 nucleocapsid protein. This 1101 technology holds promise for advancing biosensing applications in medical diagnosis and 1102 environmental monitoring by offering a compact, highly sensitive, and versatile platform 1103 for detecting a wide range of biomolecules [206]. 1104

Recently, a cavity-free configuration was used instead of Fabry-Pérot cavities or plas-1105 monic nanostructures to obtain ultrastrong light-matter coupling [207]. The research ex-1106 plored how photochromic molecules could be used to manipulate phase singularities 1107 through strong light-matter coupling. By embedding spiropyran molecules in a 1108 polymethylmethacrylate host matrix and then converting them to merocyanine by UV 1109 exposure, the researchers have demonstrated a transition from weak to strong coupling 1110 regimes. This transition leads to the formation of pairs of phase singularities, which are 1111 regions where the phase of light is indefinite. These singularities depend on the merocy-1112 anine concentration and the film thickness, which provides a way to control them opti-1113 cally. This method offers a new, simple platform for phase optics and highlights the po-1114 tential of using strong coupling for developing advanced photonic devices with potential 1115 applications in sensing. Phase singularities have already been used for these purposes 1116 [208-210]. These advanced configurations could favor plasmon biosensing. Plasmonic bi-1117 osensing employs surface plasmon resonances in metal films ~50 nm in thickness, where 1118 the metal surfaces are functionalized to achieve selectivity. However, many metals tend 1119 to corrode in biological solutions, which can decrease the performance and sensitivity of 1120 the device. Adding graphene on the metal film increases the sensitivity by three to four 1121 orders of magnitude, because it guarantees stability of the metal in liquid and preserves 1122 the plasmonic resonances under biofunctionalization [211]. 1123

The possibility of using PCF plasmonic sensors in various fields, including environ-1124 mental monitoring, chemical industry, food industry, and biomedicine, is also widely ex-1125 plored [212]. One of the examples is the proposed plasmonic biosensor with gold and 1126 titanium dioxide immobilized on a PCF for analyzing blood composition. This device al-1127 lows the detection of red blood cells, hemoglobin, white blood cells, etc. with a maximum 1128 wavelength sensitivity of 12,400 nm/RIU [213]. Currently, numerous theoretical studies of 1129 this type of devices have been reported, but experimental studies are scarce due to the 1130 challenging fabrication. 1131

WGM optical microcavities have been widely used as biosensors for detecting a va-1132 riety of biomolecules due to their exceptional sensitivity to changes in their microenviron-1133 ment. For example, an optofluidic sensing array for dynamic monitoring of protein aggre-1134 gation has been proposed which consists of microdisks and Au nanoparticles, with the 1135 microdisks serving as WGM microcavities and plasmonic nanoparticles ensuring surface-1136 enhanced Raman scattering. Tests have shown an enhancement factor of the Raman signal 1137 intensity as high as 107–108, ensuring an ultralow detection limit for R6G and methylene 1138 blue (10<sup>-9</sup> M). Owing to real-time recording of the Raman spectra of proteins, the device 1139 seems promising for studying the pathogenesis and pathological treatment of Alzheimer's 1140 disease [214]. Another research group employed a plasma separating microfluidic array 1141 combining LSPR with an asymmetric Fabry-Pérot cavity [215] to detect Dengue virus by 1142 its NS1 biomarker at a concentration of 0.1-10 µg/mL in bovine blood [216]. Thus, WGM 1143 sensors can also be used to detect nanosized viruses, bacteria [217], and protein monolay-1144 ers [218]; to monitor biomolecular interactions [219]; and to study many other objects and 1145 processes [142]. 1146

Finally, strong light–matter coupling can be used in nanofluidics to tune the optical 1147 properties of the system, thereby enabling the development of novel devices in the fields 1148 of nanofluidic photonics and optoelectronics [220]. In particular, the development of these 1149 devices will make it possible to further explore and understand the mechanisms of chemical reactions involved in photocatalysis and biological processes. Table 2 summarizes all 1151 the ways of combining strong light–matter coupling with micro-/nanofluidics discussed above. 1153

#### 6. Future Prospects and Concluding Remarks

The large variety of devices combining microfluidics and strong coupling presented 1155 in this review indicates their broad potential research value and future practical applica-1156 tions. Some of the benefits expected from using microfluidics in combination with strong 1157 coupling are reduced consumption of materials and reagents due to precise control of the 1158 sample volume, enhanced sensitivity, increased product yields, improved portability, as 1159 well as faster and less expensive techniques of the detection of chemical and biomedical 1160 entities compared to the existing approaches handling biological samples on the mac-1161 roscale. Micro-/nanofluidic architectures further amplify these benefits by providing tight 1162 fluidic confinement and precise fluidic control at the micro- and nanoscale. This precise 1163 control is ideally matched with the integrated optical characteristics of micro- and 1164 nanocavities, making the combination highly effective for various applications. Over the 1165 years, a growing number of emerging technologies have begun to be used in microfluid-1166 ics, which involves next-generation materials and detection methods making it possible 1167 to develop multifunctional microfluidic chips. However, further improvements are 1168 needed to select the optimal conditions for integrating fluid flows on both micro- and 1169 nanoscales with the strong light-matter coupling regime. In this regard, different config-1170 urations have been studied in order to design this type of integrated devices. 1171

On the one hand, Fabry–Pérot microcavities based on metal mirrors are relatively 1172 easy to fabricate and have a large surface area, suitable for spectroscopic analysis of collective strong coupling and for integrating with fluidic devices. However, they may suffer 1174

from incorrect detection of the strong coupling signal, and they may also be expensive 1175 because they require high-quality mirrors and are limited in size because they should 1176 match half the wavelength, making it especially difficult to engineer devices that operate 1177 in the visible range. Porous silicon has been proposed as a material for Fabry-Pérot mi-1178 crocavities based on distributed Bragg reflectors (DBRs). However, further developments 1179 are required to integrate them with fluidics, because the embedment of the molecules for 1180 subsequent detection is limited by the geometry and size of the pores. Small nanopores 1181 may restrict the flow of the analyte, which leads to the formation of an unwanted layer on 1182 the top of the porous silicon microcavity. Another type of cavities, Casimir cavities, rep-1183 resents an approach to nanoscale separation of the reflective surfaces. However, they are 1184 at early stages of development, and more experimental research is needed. 1185

An alternative way to deal with subwavelengths are plasmonic cavities, which are 1186 nanoscale objects often requiring microscopy techniques to be probed, but they offer the 1187 possibility of direct contact with the formed polaritons because the cavity is open. It 1188should be noted that this possibility is important not only for detection, but also for con-1189 trolling chemical reactions. In addition, the possibility of VSC between a single nanogap 1190 antenna and several molecules is important not only from the practical point of view, due 1191 to potential applications in ultrasensitive spectroscopy, site-selective chemistry, and com-1192 pact molecular polariton-based devices, but also from the theoretical one, because it could 1193 help solving many-body physics problems. 1194

PCF SPR is also an interesting option for detection applications. However, the sensors 1195 based on it are still at the early stage of development. Most studies in this field are either 1196 at the proof-of-concept level or represent theoretical and computational simulations be-1197 cause of the technical challenges in fabricating these devices. Therefore, further experi-1198 mental research is needed to validate these concepts and demonstrate the potential future 1199 applications in detection of analytes for wider range of chemical and biological samples. 1200 In addition, it is necessary to deal with some problems, such as the lack of low-cost tech-1201 nology for fabricating metal nanostructures with a large surface area and a high uni-1202 formity. In the case of LSPR sensors, the main problem is that no tests have been carried 1203 out *in situ*; i.e., there might be a discrepancy between the test results and the real situation. 1204 Moreover, there is lack of reliable methods of fixing the nanoparticles. 1205

Finally, WGM sensors are ideal for RI detection. This is a particularly attractive de-1206 tection methodology because the signal increases with an increase in analyte concentra-1207 tion rather than volume, making it suitable for small volume applications. One of the ad-1208 vantages of employing WGM and PCF SPR in fluorescent sensors is the possibility of la-1209 bel-free detection, which relies on the intrinsic properties of the target substance and, 1210 hence, requires fewer reagents and detection steps. In addition, these sensors can be engi-1211 neered to be reusable, which helps in detecting different types of analytes. Other ad-1212 vantages of WGM technology are low-cost manufacturing and simple packaging tech-1213 niques that protect the device from damage and that can be integrated with microfluidics. 1214 The possibility of analyzing samples *in situ* in real time makes these sensors suitable for 1215 rapid, portable, lab-on-a-chip assays to be used in point-of-care diagnosis. The perfor-1216 mance could be further improved by testing several samples on a single chip. It remains 1217 an ongoing challenge to improve LOD, because the target biomarkers are often contained 1218 in body fluids at trace concentrations. Additionally, noise should be reduced for the de-1219 tection of smaller quantities of the analyte. This requires dealing with various sources of 1220 noise, including environmental fluctuations (related to temperature, RI, particles, etc.), 1221 mechanical vibrations, and laser noise. Since ring WGM resonators detect only a shift of 1222 the resonance wavelength but have no mechanism to distinguish all sources of the asso-1223 ciated dielectric disturbances that could contribute to the shift, it is important to provide 1224 1225 selectivity in order to ensure that the shift is related only to the analyte of interest. Therefore, the surface should be treated using some blocking method to make it inert to non-1226 specific contaminations. Point-of-care sensors should be capable of operating at the loca-1227 tion of the patient without a permanent dedicated space or highly trained personnel. In 1228 the particular case of microbubble resonators, additional optimization is required due to the difficulty of rapid mass-scale production, using the conventional semiconductor processing technology. Integration of a tunable laser is also difficult because of the size restrictions and could compromise stability. It is important to maintain the same sensor performance outside the laboratory environment for its applicability in real-world measurements.

The main challenges entailed with the integration of these optical devices with fluid-1235 ics are the following. (1) Control and stability which require careful design and fabrication 1236 of devices, as well as the development of robust control systems. (2) Integration of differ-1237 ent technologies requires interdisciplinary collaboration and expertise, as well as the de-1238 velopment of novel fabrication and integration methods. (3) The development of scalable 1239 fabrication methods and the optimization of device performance are needed. (4) The de-1240 vice parameters should be optimized, with the necessary volumes of fluids minimized 1241 and new sensitive detection methods developed. (5) Theoretical models and experimental 1242 techniques should be developed to study and manipulate fundamental interactions be-1243 tween light and matter in microfluidic systems. (6) Real-time control and monitoring of 1244 microfluidic devices and strong light-matter coupling systems are challenging in terms of 1245 the development of fast and precise control technologies, as well as the integration of sens-1246 ing and feedback systems. (7) The cavity parameters, such as the cavity length, mirror 1247 reflectivity, and material properties should be optimized to obtain the desired strength of 1248 light-matter coupling. This may involve iterative simulations and experimental adjust-1249 ments. (8) The cavity should be installed in a controlled environment, such as a vacuum 1250 chamber or inert gas atmosphere, to minimize contamination and unwanted interactions 1251 with the surrounding medium. (9) The flatness (in the case of a nanofluidic cavity), vis-1252 cosity, concentration, adsorption, and other parameters should be controlled. 1253

Recent studies have demonstrated that confined electromagnetic fields required for 1254 the strong coupling regime can be obtained without the use of external structures (such 1255 as planar microcavities or plasmonic nanostructures). This means that electronic or vibra-1256 tional excitations in bulk or nanostructured materials can self-couple to an optical mode 1257 sustained by its own geometry. This occurs because polaritonic states are natural and 1258 ubiquitous to bulk materials and nanostructures that can be described by a generic Lo-1259 rentz resonance(s). Here, the boundaries of the Lorentz medium play the role of the mir-1260 rors that allow formation of well-defined optical modes, which, in turn, couple to the res-1261 onant transition [24,221-223]. 1262

VSC in a monolithic self-coupled photonic crystal operating in the terahertz range 1263 has also been reported. In this structure, the confinement is due to total internal reflection. 1264 Therefore, its Q-factor is only limited by the surface roughness and the optical out-coupling. This system is fabricated in a simpler way than DBRs (e.g., by physical or chemical 1266 vapor deposition, lithography, or etching) and could be more suitable for polaritonic 1267 chemistry because no additional external materials are used [222] . 1268

Configuration	Scheme	Principle of oper- ation	Sensing	Sample studied	Performance	Limitations	Ref.
Photonic		Guided light	Label-free	-	D-shaped PCF	Difficult to fabricate:	
crystal fiber		confined in a			biosensor based on a	Poor attachment of the	
(PCF)		periodic dielectric			plasmonic layer with a	bio-analyte to the sensor	[224]
nlasmonic		structure and			resolution of	surface and a large	
plasmonic	PML Liquid Si Gold	enhanced by			9.53×10 <sup>-6</sup> RIU and a	surface and a large	
sensor	Air	plasmonic effects				distance between the	

Table 2. Configurations that integrate strong light-matter coupling regime with micro- and nanofluidics

					maximum sensitivity of 10,493 nm/RIU	core and the plasmonic layer; Mostly simulations	
Fabry–Pérot (F–P) microcavity	Montude Connections	Multiple reflections between two	Label	K4[Fe(CN)6] solutions at concentrations within the water solubility range of the compound	15 mM with a Rabi splitting of ~20 cm <sup>-1</sup>	Large mode volumes	[82]
	intet + Outlet Input port + Outlet Silver + Glass Microfluidic charnel	parallel mirrors		Streptavidin and human C-reactive protein (CRP)	1.35 nM for streptavidin, 377 pM for CRP		[83]
Plasmon	- 	Localized surface	Label	Poly-γ-D- glutamic acid (PGA)	100 pg/mL		[225]
cavity		in metal nanostructures	-	Molecular glue cucurbit[5]uril	Coupling strength g ~ 100 cm <sup>-1</sup>	Low quality factors	[226]
Whispering- gallery-mode (WGM) microring resonator	Bus waveguide	Total internal reflection at a curved surface	Label-free	Carcinoembryoni c antigen, a pancreatic cancer biomarker	2.5 ng/mL	The fabrication and	[163]
Whispering- gallery-mode (WGM)		‡ leading to a circumnavigationa l path	Label-free	GR-5 DNAzyme	15 fM; dynamic detection range from 0.1 to 100 pM	nor straightforward	[227]
microbubble resonator				D-biotins	0.41 pM		[228]

resonator Reproduced with permission from \*Casey, S.R. and Sparks, J.R. (2016), published by the American Chemical Society [82]; \*\*Rho, D. and Kim, S. (2020), published by MDPI [83]; \*Arul, R., et al. (2020), published by Springer Nature [226]; \*Fu, L., et al. (2020), published 1270 by Elsevier [227]. 1272

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Ref	erences		1290
1.	Purcell, E.M.; Torrey, H.	C.; Pound, R.V. Resonance Absorption by Nuclear Magnetic Moments in a Solid. <i>Phys. Rev.</i>	1291
	<b>1946</b> , <i>69</i> , 37–38, doi:10.11	103/PhysRev.69.37.	1292
2.	Degen, C.L.; Reinhar	d, F.; Cappellaro, P. Quantum Sensing. <i>Rev. Mod. Phys.</i> <b>2017</b> , <i>89</i> , 035002,	1293
	doi:10.1103/RevModPhy	rs.89.035002.	1294
3.	Wendin, G. Quantum In	formation Processing with Superconducting Circuits: A Review. Rep. Prog. Phys. 2017, 80,	1295
	106001, doi:10.1088/1361	-6633/aa7e1a.	1296
4.	Jaynes, E.T.; Cummings,	, F.W. Comparison of Quantum and Semiclassical Radiation Theories with Application to	1297
	the Beam Maser. Proc. IE	EEE <b>1963</b> , <i>51</i> , 89–109, doi:10.1109/PROC.1963.1664.	1298
5.	Agranovich, V.M.; Malsł	nukov, A.G. Surface Polariton Spectra If the Resonance with the Transition Layer Vibrations	1299
	Exist. Optics Communicat	tions <b>1974</b> , 11, 169–171, doi:10.1016/0030-4018(74)90211-9.	1300
6.	Yakovlev, V.A.; Nazin, V	<i>I.G.;</i> Zhizhin, G.N. The Surface Polariton Splitting Due to Thin Surface Film LO Vibrations.	1301
	Optics Communications 1	<b>975</b> , <i>15</i> , 293–295, doi:10.1016/0030-4018(75)90306-5.	1302
7.	Pockrand, I.; Brillante,	A.; Möbius, D. Exciton-Surface Plasmon Coupling: An Experimental Investigation. The	1303
	Journal of Chemical Physic	<i>cs</i> <b>1982</b> , 77, 6289–6295, doi:10.1063/1.443834.	1304
8.	Kaluzny, Y.; Goy, P.; Gr	oss, M.; Raimond, J.M.; Haroche, S. Observation of Self-Induced Rabi Oscillations in Two-	1305
	Level Atoms Excited In	side a Resonant Cavity: The Ringing Regime of Superradiance. <i>Phys. Rev. Lett.</i> <b>1983</b> , <i>51</i> ,	1306
	1175–1178, doi:10.1103/l	'hysRevLett.51.1175.	1307
9.	Meschede, D.; Walth	ner, H.; Müller, G. One-Atom Maser. <i>Phys. Rev. Lett.</i> <b>1985</b> , 54, 551–554,	1308
	doi:10.1103/PhysRevLet	t.54.551.	1309
10.	Weisbuch, C.; Nishioka,	M.; Ishikawa, A.; Arakawa, Y. Observation of the Coupled Exciton-Photon Mode Splitting	1310
	in a Semiconductor Qua	ntum Microcavity. <i>Phys. Rev. Lett.</i> <b>1992</b> , <i>69</i> , 3314–3317, doi:10.1103/PhysRevLett.69.3314.	1311
11.	I nompson, K.J.; Kempe,	G.; KIMDIE, H.J. Observation of Normal-Mode Splitting for an Atom in an Optical Cavity.	1312
10	rnys. кеv. Lett. <b>1992</b> , 68,	1132–1133, doi:10.1103/PhyskevLett.68.1132.	1313
12.	Rempe, G.; Inompson,	K.J.; KIMDIE, H.J. CAVITY Quantum Electrodynamics with Strong Coupling in the Optical	1314
	Domain. Phys. Scr. 1994,	151, 07–77, 001:10.1060/0031-6949/1994/151/009.	1315

13.	Agranovich, V.; Benisty, H.; Weisbuch, C. Organic and Inorganic Quantum Wells in a Microcavity: Frenkel-	1316
	Wannier-Mott Excitons Hybridization and Energy Transformation. Solid State Communications 1997, 102, 631–636,	1317
	doi:10.1016/S0038-1098(96)00433-4.	1318
14.	Lidzey, D.G.; Bradley, D.D.C.; Skolnick, M.S.; Virgili, T.; Walker, S.; Whittaker, D.M. Strong Exciton-Photon	1319
	Coupling in an Organic Semiconductor Microcavity. Nature 1998, 395, 53–55, doi:10.1038/25692.	1320
15.	Boca, A.; Miller, R.; Birnbaum, K.M.; Boozer, A.D.; McKeever, J.; Kimble, H.J. Observation of the Vacuum Rabi	1321
	Spectrum for One Trapped Atom. Phys. Rev. Lett. 2004, 93, 233603, doi:10.1103/PhysRevLett.93.233603.	1322
16.	Ciuti, C.; Bastard, G.; Carusotto, I. Quantum Vacuum Properties of the Intersubband Cavity Polariton Field. Phys.	1323
	<i>Rev. B</i> <b>2005</b> , <i>72</i> , 115303, doi:10.1103/PhysRevB.72.115303.	1324
17.	Anappara, A.A.; De Liberato, S.; Tredicucci, A.; Ciuti, C.; Biasiol, G.; Sorba, L.; Beltram, F. Signatures of the	1325
	Ultrastrong Light-Matter Coupling Regime. Phys. Rev. B 2009, 79, 201303, doi:10.1103/PhysRevB.79.201303.	1326
18.	Lodahl, P.; Mahmoodian, S.; Stobbe, S. Interfacing Single Photons and Single Quantum Dots with Photonic	1327
	Nanostructures. Rev. Mod. Phys. 2015, 87, 347–400, doi:10.1103/RevModPhys.87.347.	1328
19.	Shalabney, A.; George, J.; Hutchison, J.; Pupillo, G.; Genet, C.; Ebbesen, T.W. Coherent Coupling of Molecular	1329
	Resonators with a Microcavity Mode. Nat Commun 2015, 6, 5981, doi:10.1038/ncomms6981.	1330
20.	George, J.; Shalabney, A.; Hutchison, J.A.; Genet, C.; Ebbesen, T.W. Liquid-Phase Vibrational Strong Coupling. J.	1331
	<i>Phys. Chem. Lett.</i> <b>2015</b> , <i>6</i> , 1027–1031, doi:10.1021/acs.jpclett.5b00204.	1332
21.	Thomas, A.; George, J.; Shalabney, A.; Dryzhakov, M.; Varma, S.J.; Moran, J.; Chervy, T.; Zhong, X.; Devaux, E.;	1333
	Genet, C.; et al. Ground-State Chemical Reactivity under Vibrational Coupling to the Vacuum Electromagnetic	1334
	Field. Angewandte Chemie <b>2016</b> , 128, 11634–11638, doi:10.1002/ange.201605504.	1335
22.	Gu, X.; Kockum, A.F.; Miranowicz, A.; Liu, Y.; Nori, F. Microwave Photonics with Superconducting Quantum	1336
	Circuits. Physics Reports 2017, 718–719, 1–102, doi:10.1016/j.physrep.2017.10.002.	1337
23.	Munkhbat, B.; Canales, A.; Küçüköz, B.; Baranov, D.G.; Shegai, T.O. Tunable Self-Assembled Casimir Microcavities	1338
	and Polaritons. <i>Nature</i> <b>2021</b> , 597, 214–219, doi:10.1038/s41586-021-03826-3.	1339
24.	Canales, A.; Baranov, D.G.; Antosiewicz, T.J.; Shegai, T. Abundance of Cavity-Free Polaritonic States in Resonant	1340
	Materials and Nanostructures. The Journal of Chemical Physics 2021, 154, 024701, doi:10.1063/5.0033352.	1341
25.	Schäfer, C.; Fojt, J.; Lindgren, E.; Erhart, P. Machine Learning for Polaritonic Chemistry: Accessing Chemical	1342
	Kinetics. J. Am. Chem. Soc. 2024, 146, 5402–5413, doi:10.1021/jacs.3c12829.	1343
26.	Fedutik, Y.; Temnov, V.V.; Schöps, O.; Woggon, U.; Artemyev, M.V. Exciton-Plasmon-Photon Conversion in	1344
	Plasmonic Nanostructures. Phys. Rev. Lett. 2007, 99, 136802, doi:10.1103/PhysRevLett.99.136802.	1345
27.	Liu, D.; Xue, C. Plasmonic Coupling Architectures for Enhanced Photocatalysis. Advanced Materials 2021, 33,	1346
	2005738, doi:10.1002/adma.202005738.	1347
28.	Wang, Y.; Zeng, S.; Humbert, G.; Ho, H. Microfluidic Whispering Gallery Mode Optical Sensors for Biological	1348
	Applications. <i>Laser &amp; amp; Photonics Reviews</i> <b>2020</b> , <i>14</i> , 2000135, doi:10.1002/lpor.202000135.	1349
29.	Gubin, A.I.; Barannik, A.A.; Cherpak, N.T.; Protsenko, I.A.; Pud, S.; Offenhausser, A.; Vitusevich, S.A. Whispering-	1350
	Gallery-Mode Resonator Technique With Microfluidic Channel for Permittivity Measurement of Liquids. IEEE	1351
	<i>Trans. Microwave Theory Techn.</i> <b>2015</b> , <i>63</i> , 2003–2009, doi:10.1109/TMTT.2015.2423289.	1352
30.	Zhao, X.; Guo, Z.; Zhou, Y.; Guo, J.; Liu, Z.; Li, Y.; Luo, M.; Wu, X. Optical Whispering-Gallery-Mode Microbubble	1353
<u> </u>	Sensors. <i>Micromachines</i> <b>2022</b> , <i>13</i> , 592, doi:10.3390/mi13040592.	1354
31.	Luo, W.; Li, X.; Meng, J.; Wang, Y.; Hong, X. Surface Plasmon Resonance Sensor Based on Side-Polished D-Shaped	1355
	Photonic Crystal Fiber With Split Cladding Air Holes. <i>IEEE Trans. Instrum. Meas.</i> 2021, 70, 1–11,	1356
	doi:10.1109/11M.2021.3054003.	1357

- 32. Purcell, E.M. Spontaneous Emission Probabilities at Radio Frequencies. In *Confined Electrons and Photons*; Burstein, 1358
  E., Weisbuch, C., Eds.; NATO ASI Series; Springer US: Boston, MA, 1995; Vol. 340, pp. 839–839 ISBN 978-1-4613- 1359
  5807-7. 1360
- 33. Kelly, K.L.; Coronado, E.; Zhao, L.L.; Schatz, G.C. The Optical Properties of Metal Nanoparticles: The Influence of 1361
   Size, Shape, and Dielectric Environment. J. Phys. Chem. B 2003, 107, 668–677, doi:10.1021/jp026731y.
   1362
- Nagarajan, K.; Thomas, A.; Ebbesen, T.W. Chemistry under Vibrational Strong Coupling. J. Am. Chem. Soc. 2021, 1363 143, 16877–16889, doi:10.1021/jacs.1c07420.
   1364
- Tavis, M.; Cummings, F.W. Exact Solution for an \$N\$-Molecule---Radiation-Field Hamiltonian. *Phys. Rev.* 1968, 170, 1365
   379–384, doi:10.1103/PhysRev.170.379. 1366
- 36. Tavis, M.; Cummings, F.W. Exact Solution for an N -Molecule Radiation-Field Hamiltonian. *Phys. Rev.* 1968, 170, 1367
   379–384, doi:10.1103/PhysRev.170.379.
- Bisht, A.; Cuadra, J.; Wersäll, M.; Canales, A.; Antosiewicz, T.J.; Shegai, T. Collective Strong Light-Matter Coupling 1369 in Hierarchical Microcavity-Plasmon-Exciton Systems. *Nano Lett.* 2019, 19, 189–196, 1370 doi:10.1021/acs.nanolett.8b03639.
- Yadav, R.K.; Bourgeois, M.R.; Cherqui, C.; Juarez, X.G.; Wang, W.; Odom, T.W.; Schatz, G.C.; Basu, J.K. Room
   Temperature Weak-to-Strong Coupling and the Emergence of Collective Emission from Quantum Dots Coupled to
   Plasmonic Arrays. ACS Nano 2020, 14, 7347–7357, doi:10.1021/acsnano.0c02785.
- Hertzog, M.; Wang, M.; Mony, J.; Börjesson, K. Strong Light–Matter Interactions: A New Direction within Chemistry. *Chem. Soc. Rev.* 2019, 48, 937–961, doi:10.1039/C8CS00193F.
- 40. Haugland, T.S.; Schäfer, C.; Ronca, E.; Rubio, A.; Koch, H. Intermolecular Interactions in Optical Cavities: An *Ab* 1377 *Initio* QED Study. *The Journal of Chemical Physics* 2021, 154, 094113, doi:10.1063/5.0039256.
- Orgiu, E.; George, J.; Hutchison, J.A.; Devaux, E.; Dayen, J.F.; Doudin, B.; Stellacci, F.; Genet, C.; Schachenmayer, J.;
   Genes, C.; et al. Conductivity in Organic Semiconductors Hybridized with the Vacuum Field. *Nature Mater* 2015, 1380
   14, 1123–1129, doi:10.1038/nmat4392.
- Zeng, H.; Pérez-Sánchez, J.B.; Eckdahl, C.T.; Liu, P.; Chang, W.J.; Weiss, E.A.; Kalow, J.A.; Yuen-Zhou, J.; Stern, N.P. 1382 Control of Photoswitching Kinetics with Strong Light–Matter Coupling in a Cavity. J. Am. Chem. Soc. 2023, 145, 1383 19655–19661, doi:10.1021/jacs.3c04254.
- 43. Sau, A.; Nagarajan, K.; Patrahau, B.; Lethuillier-Karl, L.; Vergauwe, R.M.A.; Thomas, A.; Moran, J.; Genet, C.; 1385
  Ebbesen, T.W. Modifying Woodward–Hoffmann Stereoselectivity Under Vibrational Strong Coupling. *Angew* 1386 *Chem Int Ed* 2021, 60, 5712–5717, doi:10.1002/anie.202013465.
- Lather, J.; George, J. Improving Enzyme Catalytic Efficiency by Co-Operative Vibrational Strong Coupling of Water.
   *J. Phys. Chem. Lett.* 2021, *12*, 379–384, doi:10.1021/acs.jpclett.0c03003.
- Lather, J.; Bhatt, P.; Thomas, A.; Ebbesen, T.W.; George, J. Cavity Catalysis by Cooperative Vibrational Strong
   Coupling of Reactant and Solvent Molecules. *Angew Chem Int Ed* 2019, *58*, 10635–10638, doi:10.1002/anie.201905407.
   1391
- Lather, J.; Thabassum, A.N.K.; Singh, J.; George, J. Cavity Catalysis: Modifying Linear Free-Energy Relationship under Cooperative Vibrational Strong Coupling. *Chem. Sci.* 2022, *13*, 195–202, doi:10.1039/D1SC04707H.
- 47. Vahala, K.J. Optical Microcavities. Nature 2003, 424, 839–846, doi:10.1038/nature01939.
- Boroditsky, M.; Coccioli, R.; Yablonovitch, E.; Rahmat-Samii, Y.; Kim, K.W. Smallest Possible Electromagnetic Mode
   Volume in a Dielectric Cavity. *IEE Proceedings Optoelectronics* 1998, 145, 391–397, doi:10.1049/ip-opt:19982468.
- Hugall, J.T.; Singh, A.; Van Hulst, N.F. Plasmonic Cavity Coupling. ACS Photonics 2018, 5, 43–53, 1397 doi:10.1021/acsphotonics.7b01139.

- Liu, W.; Tang, S.-J.; Xiao, Y.-F. Nonlinear Optical Microcavities Towards Single-Molecule Sensing. In *Single Molecule* 1399 *Sensing Beyond Fluorescence*; Bowen, W., Vollmer, F., Gordon, R., Eds.; Springer International Publishing: Cham, 1400 2022; pp. 97–123 ISBN 978-3-030-90338-1. 1401
- Huang, Y.; Wu, F.; Yu, L. Rabi Oscillation Study of Strong Coupling in a Plasmonic Nanocavity. *New J. Phys.* 2020, 1402 22, 063053, doi:10.1088/1367-2630/ab9222.
- Liang, K.; Guo, J.; Wu, F.; Huang, Y.; Yu, L. Dynamic Control of Quantum Emitters Strongly Coupled to the Isolated Plasmon Cavity by the Microfluidic Device. *J. Phys. Chem. C* 2021, *125*, 17303–17310, doi:10.1021/acs.jpcc.1c03583.
- Patra, A.; Caligiuri, V.; Krahne, R.; De Luca, A. Strong Light–Matter Interaction and Spontaneous Emission 1406 Reshaping via Pseudo-Cavity Modes. *Advanced Optical Materials* 2021, *9*, 2101076, doi:10.1002/adom.202101076.
- Cao, J.; De Liberato, S.; Kavokin, A.V. Strong Light–Matter Coupling in Microcavities Characterised by Rabi-Splittings Comparable to the Bragg Stop-Band Widths. *New J. Phys.* 2021, 23, 113015, doi:10.1088/1367-2630/ac3260.
- 55. Ruggenthaler, M.; Flick, J.; Pellegrini, C.; Appel, H.; Tokatly, I.V.; Rubio, A. Quantum-Electrodynamical Density1410
  Functional Theory: Bridging Quantum Optics and Electronic-Structure Theory. *Phys. Rev. A* 2014, *90*, 012508, 1411
  doi:10.1103/PhysRevA.90.012508.
- 56. Haugland, T.S.; Ronca, E.; Kjønstad, E.F.; Rubio, A.; Koch, H. Coupled Cluster Theory for Molecular Polaritons:
   1413 Changing Ground and Excited States. *Phys. Rev. X* 2020, *10*, 041043, doi:10.1103/PhysRevX.10.041043.
   1414
- 57. Sidler, D.; Schäfer, C.; Ruggenthaler, M.; Rubio, A. Polaritonic Chemistry: Collective Strong Coupling Implies 1415
   Strong Local Modification of Chemical Properties. *J. Phys. Chem. Lett.* 2021, 12, 508–516, 1416
   doi:10.1021/acs.jpclett.0c03436.
- 58. DePrince, A.E. Cavity-Modulated Ionization Potentials and Electron Affinities from Quantum Electrodynamics 1418
   Coupled-Cluster Theory. *The Journal of Chemical Physics* 2021, 154, 094112, doi:10.1063/5.0038748.
- 59. Latini, S.; Shin, D.; Sato, S.A.; Schäfer, C.; De Giovannini, U.; Hübener, H.; Rubio, A. The Ferroelectric Photo Ground 1420
   State of SrTiO 3: Cavity Materials Engineering. *Proc. Natl. Acad. Sci. U.S.A.* 2021, 118, e2105618118, 1421
   doi:10.1073/pnas.2105618118. 1422
- 60. Flick, J.; Rivera, N.; Narang, P. Strong Light-Matter Coupling in Quantum Chemistry and Quantum Photonics. 1423 Nanophotonics 2018, 7, 1479–1501, doi:10.1515/nanoph-2018-0067.
- Kang, T.; Bahk, Y.-M.; Kim, D.-S. Terahertz Quantum Plasmonics at Nanoscales and Angstrom Scales. *Nanophotonics* 1425 2020, 9, 435–451, doi:10.1515/nanoph-2019-0436.
- Marlow, E.; Murphy, A.; Pollard, R. Investigating Electromagnetic Field Enhancements from Gold Nanostructured 1427 Arrays for Plasmon Enhanced Fluorescence. In Proceedings of the Plasmonics: Design, Materials, Fabrication, 1428 Characterization, and Applications XVII; Tanaka, T., Tsai, D.P., Eds.; SPIE: San Diego, United States, September 9 1429 2019; p. 54.
- 63. Rabi, I.I. On the Process of Space Quantization. *Phys. Rev.* **1936**, *49*, 324–328, doi:10.1103/PhysRev.49.324.
- Brune, M.; Schmidt-Kaler, F.; Maali, A.; Dreyer, J.; Hagley, E.; Raimond, J.M.; Haroche, S. Quantum Rabi Oscillation: 1432 A Direct Test of Field Quantization in a Cavity. *Phys. Rev. Lett.* **1996**, *76*, 1800–1803, doi:10.1103/PhysRevLett.76.1800. 1433
- 65. Galego, J.; Climent, C.; Garcia-Vidal, F.J.; Feist, J. Cavity Casimir-Polder Forces and Their Effects in Ground-State
   1434 Chemical Reactivity. *Phys. Rev. X* 2019, *9*, 021057, doi:10.1103/PhysRevX.9.021057.
   1435
- 66. Herrera, F. Photochemistry with Quantum Optics from a Non-Adiabatic Quantum Trajectory Perspective. *Chem* 1436
   2020, 6, 7–9, doi:10.1016/j.chempr.2019.12.025.
- 67. Vurgaftman, I.; Simpkins, B.S.; Dunkelberger, A.D.; Owrutsky, J.C. Negligible Effect of Vibrational Polaritons on 1438 Chemical Reaction Rates via the Density of States Pathway. J. Phys. Chem. Lett. 2020, 11, 3557–3562, 1439 doi:10.1021/acs.jpclett.0c00841.

- Prakash, S.; Yeom, J. Nanofluidics and Microfluidics: Systems and Applications; William Andrew, 2014; ISBN 978-1-4377-4470-5.
- 69. Gale, B.K.; Jafek, A.R.; Lambert, C.J.; Goenner, B.L.; Moghimifam, H.; Nze, U.C.; Kamarapu, S.K. A Review of 1443
   Current Methods in Microfluidic Device Fabrication and Future Commercialization Prospects. *Inventions* 2018, 3, 1444
   60, doi:10.3390/inventions3030060. 1445
- Xie, Y.; Dai, L.; Yang, Y. Microfluidic Technology and Its Application in the Point-of-Care Testing Field. *Biosensors* 1446 and *Bioelectronics*: X 2022, 10, 100109, doi:10.1016/j.biosx.2022.100109.
- 71. Zhu, J.M.; Shi, Y.; Zhu, X.Q.; Yang, Y.; Jiang, F.H.; Sun, C.J.; Zhao, W.H.; Han, X.T. Optofluidic Marine Phosphate 1448
   Detection with Enhanced Absorption Using a Fabry–Pérot Resonator. *Lab Chip* 2017, 17, 4025–4030, 1449
   doi:10.1039/C7LC01016H. 1450
- 72. Xia, Y.; Whitesides, G.M. SOFT LITHOGRAPHY. Annu. Rev. Mater. Sci. **1998**, 28, 153–184, 1451 doi:10.1146/annurev.matsci.28.1.153.
- Morikawa, K.; Kazoe, Y.; Takagi, Y.; Tsuyama, Y.; Pihosh, Y.; Tsukahara, T.; Kitamori, T. Advanced Top-Down
   Fabrication for a Fused Silica Nanofluidic Device. *Micromachines* 2020, *11*, 995, doi:10.3390/mi11110995.
- 74. Do, T.C.; Bui, H.; Nguyen, T.V.; Nguyen, T.A.; Nguyen, T.H.; Pham, V.H. A Microcavity Based on a Porous Silicon
   Multilayer. Adv. Nat. Sci: Nanosci. Nanotechnol. 2011, 2, 035001, doi:10.1088/2043-6262/2/3/035001.
- 75. Panagopoulos, C.N.; Markaki, A.; Hontzopoulos, E. Excimer Laser Treatment of Copper-Coated Mild Steel. *Journal* 1457 of Materials Science 1997, 32, 1425–1430, doi:10.1023/A:1018589514262.
- 76. Han, P.; Tsai, C.-M.; Lee, H.-B. The Proposed Necessary and Sufficient Condition for Spectral Switches with 1459 Concave Reflectance of Aluminum Metal. *Advances in Mechanical Engineering* 2017, *9*, 168781401774406, 1460 doi:10.1177/1687814017744068.
- 77. Ung, T.; Liz-Marzán, L.M.; Mulvaney, P. Gold Nanoparticle Thin Films. Colloids and Surfaces A: Physicochemical and 1462 Engineering Aspects 2002, 202, 119–126, doi:10.1016/S0927-7757(01)01083-4.
- 78. Wang, Z.; Liu, L.; Zhang, D.; Krasavin, A.V.; Zheng, J.; Pan, C.; He, E.; Wang, Z.; Zhong, S.; Li, Z.; et al. Effect of Mirror Quality on Optical Response of Nanoparticle-on-Mirror Plasmonic Nanocavities. *Advanced Optical Materials* 1465 2023, 11, 2201914, doi:10.1002/adom.202201914.
- 79. Hayashi, S.; Ishigaki, Y.; Fujii, M. Plasmonic Effects on Strong Exciton-Photon Coupling in Metal-Insulator-Metal Microcavities. *Phys. Rev. B* 2012, *86*, 045408, doi:10.1103/PhysRevB.86.045408.
- 80. Dovzhenko, D.; Mochalov, K.; Vaskan, I.; Kryukova, I.; Rakovich, Y.; Nabiev, I. Polariton-Assisted Splitting of 1469 Broadband Emission Spectra of Strongly Coupled Organic Dye Excitons in Tunable Optical Microcavity. *Opt.* 1470 *Express* 2019, 27, 4077, doi:10.1364/OE.27.004077. 1471
- Bahsoun, H.; Chervy, T.; Thomas, A.; Börjesson, K.; Hertzog, M.; George, J.; Devaux, E.; Genet, C.; Hutchison, J.A.;
   Ebbesen, T.W. Electronic Light–Matter Strong Coupling in Nanofluidic Fabry–Pérot Cavities. ACS Photonics 2018,
   5, 225–232, doi:10.1021/acsphotonics.7b00679.
- Casey, S.R.; Sparks, J.R. Vibrational Strong Coupling of Organometallic Complexes. J. Phys. Chem. C 2016, 120, 1475 28138–28143, doi:10.1021/acs.jpcc.6b10493.
   1476
- Rho, D.; Kim, S. Demonstration of a Label-Free and Low-Cost Optical Cavity-Based Biosensor Using Streptavidin and C-Reactive Protein. *Biosensors* 2020, *11*, 4, doi:10.3390/bios11010004.
- Schmidt, F.; Callegari, A.; Daddi-Moussa-Ider, A.; Munkhbat, B.; Verre, R.; Shegai, T.; Käll, M.; Löwen, H.; 1479 Gambassi, A.; Volpe, G. Tunable Critical Casimir Forces Counteract Casimir–Lifshitz Attraction. *Nat. Phys.* 2022, 1480 doi:10.1038/s41567-022-01795-6. 1481

- 86. Forn-Díaz, P.; Lamata, L.; Rico, E.; Kono, J.; Solano, E. Ultrastrong Coupling Regimes of Light-Matter Interaction. 1485 *Rev. Mod. Phys.* 2019, *91*, 025005, doi:10.1103/RevModPhys.91.025005. 1486
- 87. Frisk Kockum, A.; Miranowicz, A.; De Liberato, S.; Savasta, S.; Nori, F. Ultrastrong Coupling between Light and Matter. *Nat Rev Phys* 2019, *1*, 19–40, doi:10.1038/s42254-018-0006-2.
  1488
- 88. Le Boité, A. Theoretical Methods for Ultrastrong Light–Matter Interactions. *Adv Quantum Tech* 2020, *3*, 1900140, 1489 doi:10.1002/qute.201900140.
- Liu, Q.; Chen, J.; Hou, S.; Lei, J. Investigation into Micro-Polishing Photonic Crystal Fibers for Surface Plasmon 1491 Resonance Sensing. *Crystals* 2022, 12, 1106, doi:10.3390/cryst12081106.
- 90. Chaudhary, V.S.; Kumar, D.; Pandey, B.P.; Kumar, S. Advances in Photonic Crystal Fiber-Based Sensor for 1493
   Detection of Physical and Biochemical Parameters—A Review. *IEEE Sensors J.* 2023, 23, 1012–1023, 1494
   doi:10.1109/JSEN.2022.3222969.
- Dash, J.N.; Jha, R. SPR Biosensor Based on Polymer PCF Coated With Conducting Metal Oxide. *IEEE Photon. Technol.* 1496 Lett. 2014, 26, 595–598, doi:10.1109/LPT.2014.2301153.
- 92. Caucheteur, C.; Guo, T.; Albert, J. Review of Plasmonic Fiber Optic Biochemical Sensors: Improving the Limit of Detection. *Anal Bioanal Chem* 2015, 407, 3883–3897, doi:10.1007/s00216-014-8411-6.
   1499
- P3. Rifat, A.A.; Ahmed, R.; Yetisen, A.K.; Butt, H.; Sabouri, A.; Mahdiraji, G.A.; Yun, S.H.; Adikan, F.R.M. Photonic 1500
  Crystal Fiber Based Plasmonic Sensors. *Sensors and Actuators B: Chemical* 2017, 243, 311–325, 1501
  doi:10.1016/j.snb.2016.11.113. 1502
- 94. Cao, S.; Shao, Y.; Wang, Y.; Wu, T.; Zhang, L.; Huang, Y.; Zhang, F.; Liao, C.; He, J.; Wang, Y. Highly Sensitive 1503 Surface Plasmon Resonance Biosensor Based on a Low-Index Polymer Optical Fiber. *Opt. Express* 2018, 26, 3988, 1504 doi:10.1364/OE.26.003988.
- Lu, J.; Li, Y.; Han, Y.; Liu, Y.; Gao, J. D-Shaped Photonic Crystal Fiber Plasmonic Refractive Index Sensor Based on Gold Grating. *Appl. Opt.* 2018, *57*, 5268, doi:10.1364/AO.57.005268.
- 96. Quero, G.; Zito, G.; Managò, S.; Galeotti, F.; Pisco, M.; De Luca, A.; Cusano, A. Nanosphere Lithography on Fiber: 1508
   Towards Engineered Lab-On-Fiber SERS Optrodes. *Sensors* 2018, 18, 680, doi:10.3390/s18030680. 1509
- 97. Aćimović, S.S.; Ortega, M.A.; Sanz, V.; Berthelot, J.; Garcia-Cordero, J.L.; Renger, J.; Maerkl, S.J.; Kreuzer, M.P.;
  1510 Quidant, R. LSPR Chip for Parallel, Rapid, and Sensitive Detection of Cancer Markers in Serum. *Nano Lett.* 2014, 14, 1511 2636–2641, doi:10.1021/nl500574n.
- Xu, D.; Xiong, X.; Wu, L.; Ren, X.-F.; Png, C.E.; Guo, G.-C.; Gong, Q.; Xiao, Y.-F. Quantum Plasmonics: New Opportunity in Fundamental and Applied Photonics. *Adv. Opt. Photon.* 2018, *10*, 703, doi:10.1364/AOP.10.000703.
- Xu, Y.; Xiong, M.; Yan, H. A Portable Optical Fiber Biosensor for the Detection of Zearalenone Based on the Localized Surface Plasmon Resonance. *Sensors and Actuators B: Chemical* 2021, 336, 129752, 1516 doi:10.1016/j.snb.2021.129752.
- 100. Wei, T.; Ren, P.; Huang, L.; Ouyang, Z.; Wang, Z.; Kong, X.; Li, T.; Yin, Y.; Wu, Y.; He, Q. Simultaneous Detection
   of Aflatoxin B1, Ochratoxin A, Zearalenone and Deoxynivalenol in Corn and Wheat Using Surface Plasmon
   Resonance. *Food Chemistry* 2019, 300, 125176, doi:10.1016/j.foodchem.2019.125176.
- 101. Calatayud-Sanchez, A.; Ortega-Gomez, A.; Barroso, J.; Zubia, J.; Benito-Lopez, F.; Villatoro, J.; Basabe-Desmonts,
   L. A Method for the Controllable Fabrication of Optical Fiber-Based Localized Surface Plasmon Resonance Sensors.
   *Sci Rep* 2022, *12*, 9566, doi:10.1038/s41598-022-13707-y.

102. Bastús, N.G.: Comenge, L.: Puntes, V. Kinetically Controlled Seeded Growth Synthesis of Citrate-Stabilized Gold	1524
Nanoparticles of up to 200 Nm: Size Focusing versus Ostwald Ripening, Langmuir <b>2011</b> , 27, 11098–11105.	1525
doi:10.1021/la201938u	1526
103 Khalek Md A $\cdot$ Chakma S $\cdot$ Ahmed K $\cdot$ Paul BK $\cdot$ Vigneswaran D $\cdot$ Zakaria R Materials Effect in Sensing	1527
Performance Based on Surface Plasmon Resonance Using Photonic Crystal Fiber Plasmonics 2019 14 861–867	1528
doi:10.1007/s11468-018-0867-3	1520
104 Luan N. Zhao I. Lian V. Lou S. A High Refractive Index Plasmonic Sensor Based on D-Shaped Photonic	1520
Crystal Fiber With Laterally Accessible Hollow-Core IEEE Photonics I <b>2018</b> 10 1-7	1531
doi:10.1109/JPHOT 2018 2873826	1532
105 Lee HW: Schmidt MA: Tyagi HK: Sempere LP: Russell PSt L Polarization-Dependent Coupling to	1533
Plasmon Modes on Submicron Cold Wire in Photonic Crystal Fiber Amlied Physics Letters 2008 93 111102	1534
doi:10.1063/1.2982083	1535
106 Lee HW Schmidt MA Russell R F. Joly NY Tyagi HK Uebel P. Russell PSt I Pressure-Assisted Melt-	1536
Filling and Optical Characterization of Au Nano-Wires in Microstructured Fibers <i>Ont Express</i> <b>2011</b> 19 12180	1537
doi 10 1364/OE 19 012180	1538
107. Russell, P. Photonic Crystal Fibers. <i>Science</i> <b>2003</b> , <i>299</i> , 358–362, doi:10.1126/science.1079280.	1539
108. Ashok, N.; Shin, W. Effective D-Shape Fiber with Air Hole Assistant Design for Birefringence Analysis. <i>Optik</i> <b>2018</b> ,	1540
<i>162</i> , 27–34, doi:10.1016/j.ijleo.2018.02.045.	1541
109. Boehm, J.; François, A.; Ebendorff-Heidepriem, H.; Monro, T.M. Chemical Deposition of Silver for the Fabrication	1542
of Surface Plasmon Microstructured Optical Fibre Sensors. <i>Plasmonics</i> <b>2011</b> , <i>6</i> , 133–136, doi:10.1007/s11468-010-	1543
9178-z.	1544
110. Srivastava, R.; Prajapati, Y.K.; Pal, S.; Kumar, S. Micro-Channel Plasmon Sensor Based on a D-Shaped Photonic	1545
Crystal Fiber for Malaria Diagnosis With Improved Performance. IEEE Sensors J. 2022, 22, 14834–14841,	1546
doi:10.1109/JSEN.2022.3181198.	1547
111. Luo, W.; Li, X.; Abbasi, S.A.; Zhu, S.; Ho, HP.; Yuan, W. Analysis of the D-Shaped PCF-Based SPR Sensor Using	1548
Resonance Electron Relaxation and Fourier Domain Method. Optics and Lasers in Engineering 2023, 166, 107588,	1549
doi:10.1016/j.optlaseng.2023.107588.	1550
112. Ma, Y.; Liu, F.; Ren, Q.; Zhang, H.; Zhang, A. Dual-Band Highly-Sensitive SPR Photonic Crystal Fiber Sensor	1551
Based on Birefringence Analysis. Optics Communications 2023, 532, 129253, doi:10.1016/j.optcom.2022.129253.	1552
113. Islam, N.; Faizul Huq Arif, Md.; Abu Yousuf, M.; Asaduzzaman, S. Highly Sensitive Open Channel Based PCF-	1553
SPR Sensor for Analyte Refractive Index Sensing. Results in Physics 2023, 46, 106266, doi:10.1016/j.rinp.2023.106266.	1554
114. Mawa Nijhum, J.; Ahmed, T.; Hossain, Md.A.; Atai, J.; Hai, N.H. Microchannel-Embedded D-Shaped Photonic	1555
Crystal Fiber-Based Highly Sensitive Plasmonic Biosensor. <i>Applied Sciences</i> <b>2022</b> , <i>12</i> , 4122, doi:10.3390/app12094122.	1556
115. Chen, N.; Chang, M.; Lu, X.; Zhou, J.; Zhang, X. Photonic Crystal Fiber Plasmonic Sensor Based on Dual	1557
Optofluidic Channel. Sensors 2019, 19, 5150, doi:10.3390/s19235150.	1558
116. Sharif, V.; Pakarzadeh, H. High-Performance Surface Plasmon Resonance Fiber Sensor Based on Cylindrical	1559
Vector Modes. Sci Rep 2023, 13, 4563, doi:10.1038/s41598-023-31524-9.	1560
117. Zhang, J.; Yuan, J.; Qu, Y.; Qiu, S.; Mei, C.; Zhou, X.; Yan, B.; Wu, Q.; Wang, K.; Sang, X.; et al. A Novel Surface	1561
Plasmon Resonance-Based Photonic Crystal Fiber Refractive Index Sensor with an Ultra-Wide Detection Range.	1562
<i>Optik</i> <b>2022</b> , 259, 168977, doi:10.1016/j.ijleo.2022.168977.	1563

- 118. Aruna Gandhi, M.S.; Senthilnathan, K.; Babu, P.R.; Li, Q. Visible to near Infrared Highly Sensitive Microbiosensor
   1564
   Based on Surface Plasmon Polariton with External Sensing Approach. *Results in Physics* 2019, 15, 102590,
   1565
   doi:10.1016/j.rinp.2019.102590.
- 119. Liu, J.; Jalali, M.; Mahshid, S.; Wachsmann-Hogiu, S. Are Plasmonic Optical Biosensors Ready for Use in Point-of Need Applications? *Analyst* 2020, *145*, 364–384, doi:10.1039/C9AN02149C.
   1568
- Huang, C.-B.; Huang, J.-S. Integrated Plasmonics Nanocircuits. In *Plasmonic Materials and Metastructures*; Elsevier, 1569 2024; pp. 245–283 ISBN 978-0-323-85379-8.
- Mondal, M.; Semenov, A.; Ochoa, M.A.; Nitzan, A. Strong Coupling in Infrared Plasmonic Cavities. J. Phys. Chem. 1571
   Lett. 2022, 13, 9673–9678, doi:10.1021/acs.jpclett.2c02304. 1572
- 122. Rastogi, R.; Dogbe Foli, E.A.; Vincent, R.; Adam, P.-M.; Krishnamoorthy, S. Engineering Electromagnetic Hot 1573 Spots in Nanoparticle Cluster Arrays on Reflective Substrates for Highly Sensitive Detection of (Bio)Molecular
   1574 Analytes. ACS Appl. Mater. Interfaces 2021, 13, 32653–32661, doi:10.1021/acsami.1c01953.
- 123. Fan, M.; Andrade, G.F.S.; Brolo, A.G. A Review on the Fabrication of Substrates for Surface Enhanced Raman
   1576 Spectroscopy and Their Applications in Analytical Chemistry. *Analytica Chimica Acta* 2011, 693, 7–25, 1577 doi:10.1016/j.aca.2011.03.002.
- 124. Russell, K.J.; Liu, T.-L.; Cui, S.; Hu, E.L. Large Spontaneous Emission Enhancement in Plasmonic Nanocavities. 1579 *Nature Photon* 2012, 6, 459–462, doi:10.1038/nphoton.2012.112.
- 125. Wertz, E.; Isaacoff, B.P.; Flynn, J.D.; Biteen, J.S. Single-Molecule Super-Resolution Microscopy Reveals How Light 1581
   Couples to a Plasmonic Nanoantenna on the Nanometer Scale. *Nano Lett.* 2015, 15, 2662–2670, 1582
   doi:10.1021/acs.nanolett.5b00319.
- 126. Allsop, T.D.P.; Neal, R.; Wang, C.; Nagel, D.A.; Hine, A.V.; Culverhouse, P.; Ania Castañón, J.D.; Webb, D.J.;
   1584 Scarano, S.; Minunni, M. An Ultra-Sensitive Aptasensor on Optical Fibre for the Direct Detection of Bisphenol A.
   *Biosensors and Bioelectronics* 2019, 135, 102–110, doi:10.1016/j.bios.2019.02.043.
- 127. Di Martino, G.; Turek, V.A.; Lombardi, A.; Szabó, I.; De Nijs, B.; Kuhn, A.; Rosta, E.; Baumberg, J.J. Tracking
   1587
   Nanoelectrochemistry Using Individual Plasmonic Nanocavities. Nano Lett. 2017, 17, 4840–4845, 1588
   doi:10.1021/acs.nanolett.7b01676.
- 128. Katzen, J.M.; Tserkezis, C.; Cai, Q.; Li, L.H.; Kim, J.M.; Lee, G.; Yi, G.-R.; Hendren, W.R.; Santos, E.J.G.; Bowman, 1590
  R.M.; et al. Strong Coupling of Carbon Quantum Dots in Plasmonic Nanocavities. ACS Appl. Mater. Interfaces 2020, 1591
  12, 19866–19873, doi:10.1021/acsami.0c03312. 1592
- 129. Bitton, O.; Haran, G. Plasmonic Cavities and Individual Quantum Emitters in the Strong Coupling Limit. Acc. 1593 Chem. Res. 2022, 55, 1659–1668, doi:10.1021/acs.accounts.2c00028.
   1594
- 130. Gupta, S.N.; Bitton, O.; Neuman, T.; Esteban, R.; Chuntonov, L.; Aizpurua, J.; Haran, G. Complex Plasmon-Exciton
   1595
   Dynamics Revealed through Quantum Dot Light Emission in a Nanocavity. Nat Commun 2021, 12, 1310,
   1596
   doi:10.1038/s41467-021-21539-z.
- 131. Lin, Q.-Y.; Li, Z.; Brown, K.A.; O'Brien, M.N.; Ross, M.B.; Zhou, Y.; Butun, S.; Chen, P.-C.; Schatz, G.C.; Dravid, 1598
  V.P.; et al. Strong Coupling between Plasmonic Gap Modes and Photonic Lattice Modes in DNA-Assembled Gold 1599
  Nanocube Arrays. *Nano Lett.* 2015, 15, 4699–4703, doi:10.1021/acs.nanolett.5b01548.
- 132. Le, T.H.H.; Tanaka, T. Plasmonics–Nanofluidics Hydrid Metamaterial: An Ultrasensitive Platform for Infrared 1601
   Absorption Spectroscopy and Quantitative Measurement of Molecules. ACS Nano 2017, 11, 9780–9788, 1602
   doi:10.1021/acsnano.7b02743.
- 133. Dayal, G.; Morichika, I.; Ashihara, S. Vibrational Strong Coupling in Subwavelength Nanogap Patch Antenna at the Single Resonator Level. J. Phys. Chem. Lett. 2021, 12, 3171–3175, doi:10.1021/acs.jpclett.1c00081.
   1605

M.; Olsson, E.; Shegai, T. Visualizing Spatial Variations of Plasmon-Exciton Polaritons at the Nanoscale Using	1607
Electron Microscopy. Nano Lett. 2019, 19, 8171–8181, doi:10.1021/acs.nanolett.9b03534.	1608
135. Kleemann, ME.; Chikkaraddy, R.; Alexeev, E.M.; Kos, D.; Carnegie, C.; Deacon, W.; De Pury, A.C.; Große, C.; De	1609
Nijs, B.; Mertens, J.; et al. Strong-Coupling of WSe2 in Ultra-Compact Plasmonic Nanocavities at Room Temperature.	1610
Nat Commun 2017, 8, 1296, doi:10.1038/s41467-017-01398-3.	1611
136. Del Real Mata, C.; Siavash Moakhar, R.; Hosseini, I.I.; Jalali, M.; Mahshid, S. A Nanostructured Microfluidic	1612
Device for Plasmon-Assisted Electrochemical Detection of Hydrogen Peroxide Released from Cancer Cells.	1613
<i>Nanoscale</i> <b>2021</b> , <i>13</i> , 14316–14329, doi:10.1039/D0NR07608B.	1614
137. Chikkaraddy, R.; De Nijs, B.; Benz, F.; Barrow, S.J.; Scherman, O.A.; Rosta, E.; Demetriadou, A.; Fox, P.; Hess, O.;	1615
Baumberg, J.J. Single-Molecule Strong Coupling at Room Temperature in Plasmonic Nanocavities. Nature 2016, 535,	1616
127–130, doi:10.1038/nature17974.	1617
138. Ahmadivand, A. Tunneling Plasmonics: Vacuum Rabi Oscillations in Carbon Nanotube Mediated	1618
Electromigrated Nanojunctions. J. Phys. Chem. C 2021, 125, 782–791, doi:10.1021/acs.jpcc.0c09325.	1619
139. Park, M.K.; Kee, J.S.; Quah, J.Y.; Netto, V.; Song, J.; Fang, Q.; La Fosse, E.M.; Lo, GQ. Label-Free Aptamer Sensor	1620

134. Yankovich, A.B.; Munkhbat, B.; Baranov, D.G.; Cuadra, J.; Olsén, E.; Lourenço-Martins, H.; Tizei, L.H.G.; Kociak,

- Based on Silicon Microring Resonators. *Sensors and Actuators B: Chemical* **2013**, 176, 552–559, 1621 doi:10.1016/j.snb.2012.08.078.
- Yu, D.; Humar, M.; Meserve, K.; Bailey, R.C.; Chormaic, S.N.; Vollmer, F. Whispering-Gallery-Mode Sensors for Biological and Physical Sensing. *Nat Rev Methods Primers* 2021, *1*, 83, doi:10.1038/s43586-021-00079-2.
- 141. Zhu, J.; Ozdemir, S.K.; Xiao, Y.-F.; Li, L.; He, L.; Chen, D.-R.; Yang, L. On-Chip Single Nanoparticle Detection and
   1625
   Sizing by Mode Splitting in an Ultrahigh-Q Microresonator. *Nature Photon* 2010, 4, 46–49, 1626
   doi:10.1038/nphoton.2009.237.
- 142. Jiang, X.; Qavi, A.J.; Huang, S.H.; Yang, L. Whispering-Gallery Sensors. *Matter* 2020, *3*, 371–392, 1628 doi:10.1016/j.matt.2020.07.008.
- 143. Cai, L.; Pan, J.; Zhao, Y.; Wang, J.; Xiao, S. Whispering Gallery Mode Optical Microresonators: Structures and Sensing Applications. *Physica Status Solidi (a)* 2020, 217, 1900825, doi:10.1002/pssa.201900825.
- 144. Ashadi Md Johari, M.; Hafiz Bin Jali, M.; Helmi Bin Mohd Yusof, H.; Rafis Bin Abdul Rahim, H.; Binti Ahmad, A.; 1632
   Imran Mustafa Abdul Khudus, M.; Wadi Harun, S. Polyvinyl Alcohol Coating Microbottle Resonator on 1633
   Whispering Gallery Modes for Ethanol Liquid Sensor. *Optics & Laser Technology* 2021, 143, 107379, 1634
   doi:10.1016/j.optlastec.2021.107379. 1635
- 145. Zhang, Y.; Wang, M.; Zhu, N.; Han, B.; Liu, Y. Optical Fiber Hydrogen Sensor Based on Self-Assembled PDMS/Pd WO3 Microbottle Resonator. *Sensors and Actuators B: Chemical* 2023, 375, 132866, doi:10.1016/j.snb.2022.132866.
   1637
- 146. Mehta, S.; V., N.N.B.; Singh, M. Engineering Porous Silicon-Based Plasmonic Microdisk Resonator for Highly
   Sensitive Methanol Sensing. *IEEE Sensors J.* 2024, 24, 12304–12311, doi:10.1109/JSEN.2024.3373907.
- 147. Guo, Z.; Qin, Y.; Chen, P.; Hu, J.; Zhou, Y.; Zhao, X.; Liu, Z.; Fei, Y.; Jiang, X.; Wu, X. Hyperboloid-Drum Microdisk
   Laser Biosensors for Ultrasensitive Detection of Human IgG. *Small* 2020, *16*, 2000239, doi:10.1002/smll.202000239.
   1641
- 148. Vollmer, F.; Arnold, S.; Keng, D. Single Virus Detection from the Reactive Shift of a Whispering-Gallery Mode.
   *Proc. Natl. Acad. Sci. U.S.A.* 2008, 105, 20701–20704, doi:10.1073/pnas.0808988106.
- 149. Brice, I.; Grundsteins, K.; Draguns, K.; Atvars, A.; Alnis, J. Whispering Gallery Mode Resonator Temperature 1644
   Compensation and Refractive Index Sensing in Glucose Droplets. *Sensors* 2021, 21, 7184, doi:10.3390/s21217184.

11878, doi:10.1021/acs.analchem.9b02630.

1646

1647

1648

1649

]	Performance of Whispering Gallery Modes in a Microcapillary Resonator Filled with Nematic Liquid Crystal. Optics	1650
(	Communications <b>2023</b> , 537, 129442, doi:10.1016/j.optcom.2023.129442.	1651
152.	Wan, H.; Chen, J.; Wan, C.; Zhou, Q.; Wang, J.; Zhang, Z. Optofluidic Microcapillary Biosensor for Label-Free,	1652
]	Low Glucose Concentration Detection. Biomed. Opt. Express 2019, 10, 3929, doi:10.1364/BOE.10.003929.	1653
153.	Schuler, S.; Muench, J.E.; Ruocco, A.; Balci, O.; Thourhout, D.V.; Sorianello, V.; Romagnoli, M.; Watanabe, K.;	1654
-	Taniguchi, T.; Goykhman, I.; et al. High-Responsivity Graphene Photodetectors Integrated on Silicon Microring	1655
]	Resonators. <i>Nat Commun</i> <b>2021</b> , 12, 3733, doi:10.1038/s41467-021-23436-x.	1656
154.	Bahadoran, M.; Seyfari, A.K.; Sanati, P.; Chua, L.S. Label Free Identification of the Different Status of Anemia	1657
]	Disease Using Optimized Double-Slot Cascaded Microring Resonator. Sci Rep 2022, 12, 5548, doi:10.1038/s41598-	1658
(	022-09504-2.	1659
155.	Vollmer, F.; Braun, D.; Libchaber, A.; Khoshsima, M.; Teraoka, I.; Arnold, S. Protein Detection by Optical Shift of	1660
ć	a Resonant Microcavity. Applied Physics Letters 2002, 80, 4057–4059, doi:10.1063/1.1482797.	1661
156.	Shao, L.; Jiang, X.; Yu, X.; Li, B.; Clements, W.R.; Vollmer, F.; Wang, W.; Xiao, Y.; Gong, Q. Detection of Single	1662
I	Nanoparticles and Lentiviruses Using Microcavity Resonance Broadening. Advanced Materials 2013, 25, 5616–5620,	1663
(	doi:10.1002/adma201302572.	1664
157.	Zhu, N.; Shi, B.; Guo, Y.; Han, B.; Zhang, Y. Polydimethylsiloxane Self-Assembled Whispering Gallery Mode	1665
1	Microbottle Resonator for Ethanol Sensing. Optical Materials 2020, 107, 110024, doi:10.1016/j.optmat.2020.110024.	1666
158.	Ghulinyan, M.; Navarro-Urrios, D.; Pitanti, A.; Lui, A.; Pucker, G.; Pavesi, L. Whispering-Gallery Modes and Light	1667
]	Emission from a Si-Nanocrystal-Based Single Microdisk Resonator. Opt. Express 2008, 16, 13218,	1668
(	doi:10.1364/OE.16.013218.	1669
159.	Lin, S.; Schonbrun, E.; Crozier, K. Optical Manipulation with Planar Silicon Microring Resonators. Nano Lett. 2010,	1670
-	<i>10</i> , 2408–2411, doi:10.1021/nl100501d.	1671
160.	Liao, J.; Qavi, A.; Dong, R.; Yang, L. Packaging of Optofluidic Microbubble Resonator Sensors. In Proceedings of	1672
t	the Chemical, Biological, Radiological, Nuclear, and Explosives (CBRNE) Sensing XX; Guicheteau, J.A., Howle, C.R.,	1673
]	Eds.; SPIE: Baltimore, United States, May 17 2019; p. 46.	1674
161.	Arnold, S.; Keng, D.; Shopova, S.I.; Holler, S.; Zurawsky, W.; Vollmer, F. Whispering Gallery Mode Carousel – a	1675
]	Photonic Mechanism for Enhanced Nanoparticle Detection in Biosensing. Opt. Express 2009, 17, 6230,	1676
(	doi:10.1364/OE.17.006230.	1677
162.	Riesen, N.; Peterkovic, Z.Q.; Guan, B.; François, A.; Lancaster, D.G.; Priest, C. Caged-Sphere Optofluidic Sensors:	1678
J	Whispering Gallery Resonators in Wicking Microfluidics. Sensors 2022, 22, 4135, doi:10.3390/s22114135.	1679
163.	Bailey, R.C.; Washburn, A.L.; Qavi, A.J.; Iqbal, M.; Gleeson, M.; Tybor, F.; Gunn, L.C. A Robust Silicon Photonic	1680
]	Platform for Multiparameter Biological Analysis. In Proceedings of the Silicon Photonics IV; SPIE, February 17 2009;	1681
	Vol. 7220, pp. 147–152.	1682
164.	Chen, Y.; Yin, Y.; Ma, L.; Schmidt, O.G. Recent Progress on Optoplasmonic Whispering-Gallery-Mode	1683
l	Microcavities. <i>Advanced Optical Materials</i> <b>2021</b> , 9, 2100143, doi:10.1002/adom.202100143.	1684

150. Ozgur, E.; Roberts, K.E.; Ozgur, E.O.; Gin, A.N.; Bankhead, J.R.; Wang, Z.; Su, J. Ultrasensitive Detection of Human

151. Wang, Z.; Mallik, A.K.; Wei, F.; Wang, Z.; Rout, A.; Wu, Q.; Semenova, Y. Enhancing the Thermo-Optic Tuning

Chorionic Gonadotropin Using Frequency Locked Microtoroid Optical Resonators. Anal. Chem. 2019, 91, 11872-

165. Kang, T.Y.; Lee, W.; Ahn, H.; Shin, D.-M.; Kim, C.-S.; Oh, J.-W.; Kim, D.; Kim, K. Plasmon-Coupled Whispering
 Gallery Modes on Nanodisk Arrays for Signal Enhancements. *Sci Rep* 2017, *7*, 11737, doi:10.1038/s41598-017-12053 8.

166. Willets, K.A.; Van Duyne, R.P. Localized Surface Plasmon Resonance Spectroscopy and Sensing. Annu. Rev. Phys.	1688
<i>Chem.</i> <b>2007</b> , <i>58</i> , 267–297, doi:10.1146/annurev.physchem.58.032806.104607.	1689

- 167. Yu, X.-C.; Tang, S.-J.; Liu, W.; Xu, Y.; Gong, Q.; Chen, Y.-L.; Xiao, Y.-F. Single-Molecule Optofluidic Microsensor
   1690 with Interface Whispering Gallery Modes. *Proc. Natl. Acad. Sci. U.S.A.* 2022, 119, e2108678119, 1691 doi:10.1073/pnas.2108678119.
- 168. Qavi, A.J.; Bailey, R.C. Multiplexed Detection and Label-Free Quantitation of MicroRNAs Using Arrays of Silicon
   Photonic Microring Resonators. *Angew Chem Int Ed* 2010, 49, 4608–4611, doi:10.1002/anie.201001712.
- 169. Wang, J.; Sanchez, M.M.; Yin, Y.; Herzer, R.; Ma, L.; Schmidt, O.G. Silicon-Based Integrated Label-Free Optofluidic
   1695
   Biosensors: Latest Advances and Roadmap. *Adv Materials Technologies* 2020, *5*, 1901138, doi:10.1002/admt.201901138.
   1696
- 170. Kim, Y.; Lee, H. On-Chip Label-Free Biosensing Based on Active Whispering Gallery Mode Resonators Pumped
   by a Light-Emitting Diode. *Opt. Express* 2019, 27, 34405, doi:10.1364/OE.27.034405.
- 171. Duan, R.; Hao, X.; Li, Y.; Li, H. Detection of Acetylcholinesterase and Its Inhibitors by Liquid Crystal Biosensor
   1699
   Based on Whispering Gallery Mode. Sensors and Actuators B: Chemical 2020, 308, 127672, 1700
   doi:10.1016/j.snb.2020.127672.
- Mudumba, S.; De Alba, S.; Romero, R.; Cherwien, C.; Wu, A.; Wang, J.; Gleeson, M.A.; Iqbal, M.; Burlingame, R.W.
   Photonic Ring Resonance Is a Versatile Platform for Performing Multiplex Immunoassays in Real Time. *Journal of Immunological Methods* 2017, 448, 34–43, doi:10.1016/j.jim.2017.05.005.
- 173. Soni, V.; Chang, C.-W.; Xu, X.; Wang, C.; Yan, H.; D Agati, M.; Tu, L.-W.; Chen, Q.Y.; Tian, H.; Chen, R.T. Portable
   1705 Automatic Microring Resonator System Using a Subwavelength Grating Metamaterial Waveguide for High Sensitivity Real-Time Optical-Biosensing Applications. *IEEE Trans. Biomed. Eng.* 2021, 68, 1894–1902, 1707 doi:10.1109/TBME.2020.3029148.
- 174. Meserve, K.; Qavi, A.J.; Aman, M.J.; Vu, H.; Zeitlin, L.; Dye, J.M.; Froude, J.W.; Leung, D.W.; Yang, L.; Holtsberg, 1709
  F.W.; et al. Detection of Biomarkers for Filoviral Infection with a Silicon Photonic Resonator Platform. *STAR* 1710 *Protocols* 2022, 3, 101719, doi:10.1016/j.xpro.2022.101719. 1711
- 175. Balac, S. WGMode : A Matlab Toolbox for Whispering Gallery Modes Volume Computation in Spherical Optical
   Micro-Resonators. *Computer Physics Communications* 2019, 243, 121–134, doi:10.1016/j.cpc.2019.05.002.
- 176. Khani, S.; Hayati, M. An Ultra-High Sensitive Plasmonic Refractive Index Sensor Using an Elliptical Resonator
   1714 and MIM Waveguide. *Superlattices and Microstructures* 2021, 156, 106970, doi:10.1016/j.spmi.2021.106970.
   1715
- 177. Butt, M.A.; Khonina, S.N.; Kazanskiy, N.L. Highly Sensitive Refractive Index Sensor Based on Hybrid Plasmonic
   1716
   Waveguide Microring Resonator. Waves in Random and Complex Media 2020, 30, 292–299, 1717
   doi:10.1080/17455030.2018.1506191.
- 178. Yu, D.; Vollmer, F. Microscale Whispering-Gallery-Mode Light Sources with Lattice-Confined Atoms. *Sci Rep* 2021, 1719
   11, 13899, doi:10.1038/s41598-021-93295-5.
- 179. Shen, H.; Zhao, C.Y. On-Chip Fluid Information Detection Based on Micro-Ring Optical Frequency Comb
   1721
   Technology and Machine Learning 2024.
   1722
- 180. Jacobson, L.D.; Bochevarov, A.D.; Watson, M.A.; Hughes, T.F.; Rinaldo, D.; Ehrlich, S.; Steinbrecher, T.B.;
  1723 Vaitheeswaran, S.; Philipp, D.M.; Halls, M.D.; et al. Automated Transition State Search and Its Application to
  1724 Diverse Types of Organic Reactions. J. Chem. Theory Comput. 2017, 13, 5780–5797, doi:10.1021/acs.jctc.7b00764.
  1725
- 181. Behler, J. Perspective: Machine Learning Potentials for Atomistic Simulations. *The Journal of Chemical Physics* 2016, 1726 145, 170901, doi:10.1063/1.4966192.

182. Munkhbat, B.; Wersäll, M.; Baranov, D.G.; Antosiewicz, T.J.; Shegai, T. Suppression of Photo-Oxidation of Organic	1728
Chromophores by Strong Coupling to Plasmonic Nanoantennas. Sci. Adv. 2018, 4, eaas9552,	1729
doi:10.1126/sciadv.aas9552.	1730
183. Hutchison, J.A.; Schwartz, T.; Genet, C.; Devaux, E.; Ebbesen, T.W. Modifying Chemical Landscapes by Coupling	1731
to Vacuum Fields. <i>Angew Chem Int Ed</i> 2012, <i>51</i> , 1592–1596, doi:10.1002/anie.201107033.	1732
184. George, J.; Wang, S.; Chervy, T.; Canaguier-Durand, A.; Schaeffer, G.; Lehn, JM.; Hutchison, J.A.; Genet, C.;	1733
Ebbesen, T.W. Ultra-Strong Coupling of Molecular Materials: Spectroscopy and Dynamics. Faraday Discuss. 2015,	1734
178, 281–294, doi:10.1039/C4FD00197D.	1735
185. George, J.; Chervy, T.; Shalabney, A.; Devaux, E.; Hiura, H.; Genet, C.; Ebbesen, T.W. Multiple Rabi Splittings	1736
under Ultrastrong Vibrational Coupling. Phys. Rev. Lett. 2016, 117, 153601, doi:10.1103/PhysRevLett.117.153601.	1737
186. Hirai, K.; Takeda, R.; Hutchison, J.A.; Uji-i, H. Modulation of Prins Cyclization by Vibrational Strong Coupling.	1738
Angew Chem Int Ed <b>2020</b> , 59, 5332–5335, doi:10.1002/anie.201915632.	1739
187. Ahn, W.; Triana, J.F.; Recabal, F.; Herrera, F.; Simpkins, B.S. Modification of Ground-State Chemical Reactivity via	1740
Light-Matter Coherence in Infrared Cavities. Science 2023, 380, 1165–1168, doi:10.1126/science.ade7147.	1741
188. Kadyan, A.; Shaji, A.; George, J. Boosting Self-Interaction of Molecular Vibrations under Ultrastrong Coupling	1742
Condition. J. Phys. Chem. Lett. 2021, 12, 4313-4318, doi:10.1021/acs.jpclett.1c00552.	1743
189. Lidzey, D.G.; Bradley, D.D.C.; Virgili, T.; Armitage, A.; Skolnick, M.S.; Walker, S. Room Temperature Polariton	1744
Emission from Strongly Coupled Organic Semiconductor Microcavities. Phys. Rev. Lett. 1999, 82, 3316–3319,	1745
doi:10.1103/PhysRevLett.82.3316.	1746
190. Holmes, R.J.; Forrest, S.R. Strong Exciton-Photon Coupling and Exciton Hybridization in a Thermally Evaporated	1747
Polycrystalline Film of an Organic Small Molecule. Phys. Rev. Lett. 2004, 93, 186404,	1748
doi:10.1103/PhysRevLett.93.186404.	1749
191. Kéna-Cohen, S.; Davanço, M.; Forrest, S.R. Strong Exciton-Photon Coupling in an Organic Single Crystal	1750
Microcavity. Phys. Rev. Lett. 2008, 101, 116401, doi:10.1103/PhysRevLett.101.116401.	1751
192. Tropf, L.; Dietrich, C.P.; Herbst, S.; Kanibolotsky, A.L.; Skabara, P.J.; Würthner, F.; Samuel, I.D.W.; Gather, M.C.;	1752
Höfling, S. Influence of Optical Material Properties on Strong Coupling in Organic Semiconductor Based	1753
Microcavities. Applied Physics Letters 2017, 110, 153302, doi:10.1063/1.4978646.	1754
193. Graf, A.; Tropf, L.; Zakharko, Y.; Zaumseil, J.; Gather, M.C. Near-Infrared Exciton-Polaritons in Strongly Coupled	1755
Single-Walled Carbon Nanotube Microcavities. <i>Nat Commun</i> 2016, 7, 13078, doi:10.1038/ncomms13078.	1756
194. Dietrich, C.P.; Steude, A.; Schubert, M.; Ohmer, J.; Fischer, U.; Höfling, S.; Gather, M.C. Strong Coupling in Fully	1757
Tunable Microcavities Filled with Biologically Produced Fluorescent Proteins. Advanced Optical Materials 2017, 5,	1758
1600659, doi:10.1002/adom.201600659.	1759
195. Coles, D.M.; Yang, Y.; Wang, Y.; Grant, R.T.; Taylor, R.A.; Saikin, S.K.; Aspuru-Guzik, A.; Lidzey, D.G.; Tang, J.K	1760
H.; Smith, J.M. Strong Coupling between Chlorosomes of Photosynthetic Bacteria and a Confined Optical Cavity	1761
Mode. Nat Commun 2014, 5, 5561, doi:10.1038/ncomms6561.	1762
196. He, Z.; Li, F.; Zuo, P.; Xu, C.; He, W.; He, J.; Zhou, Y.; Zhang, Q.; Chen, K.; Huang, H.; et al. Strong Coupling in a	1763
Hybrid System of Silver Nanoparticles and J-Aggregates at Room Temperature. J. Phys. Chem. C 2022, 126, 17141-	1764
17151, doi:10.1021/acs.jpcc.2c02739.	1765
197. Krivenkov, V.; Samokhvalov, P.; Nabiev, I.; Rakovich, Y.P. pH-Sensing Platform Based on Light–Matter Coupling	1766
in Colloidal Complexes of Silver Nanoplates and J-Aggregates. J. Phys. Chem. C 2021, 125, 1972–1979,	1767
doi:10.1021/acs.jpcc.0c10602.	1768

- 198. Melnikau, D.; Samokhvalov, P.; Sánchez-Iglesias, A.; Grzelczak, M.; Nabiev, I.; Rakovich, Y.P. Strong Coupling 1769
  Effects in a Plexciton System of Gold Nanostars and J-Aggregates. *Journal of Luminescence* 2022, 242, 118557, 1770
  doi:10.1016/j.jlumin.2021.118557. 1771
- 199. Schäfer, C.; Hultmark, S.; Yang, Y.; Müller, C.; Börjesson, K. Room Temperature Dye Glasses: A Guideline Toward the Fabrication of Amorphous Dye Films with Monomeric Absorption and Emission. *Chem. Mater.* 2022, 34, 9294– 1773 9302, doi:10.1021/acs.chemmater.2c02761.
- 200. Tang, J.; Zhang, J.; Lv, Y.; Wang, H.; Xu, F.F.; Zhang, C.; Sun, L.; Yao, J.; Zhao, Y.S. Room Temperature Exciton–
   Polariton Bose–Einstein Condensation in Organic Single-Crystal Microribbon Cavities. *Nat Commun* 2021, *12*, 3265,
   doi:10.1038/s41467-021-23524-y.
- 201. Ren, J.; Liao, Q.; Huang, H.; Li, Y.; Gao, T.; Ma, X.; Schumacher, S.; Yao, J.; Bai, S.; Fu, H. Efficient Bosonic 1778 Condensation of Exciton Polaritons in an H-Aggregate Organic Single-Crystal Microcavity. *Nano Lett.* 2020, 20, 1779 7550–7557, doi:10.1021/acs.nanolett.0c03009.
- 202. Mischok, A.; Lüttgens, J.; Berger, F.; Hillebrandt, S.; Tenopala-Carmona, F.; Kwon, S.; Murawski, C.; Siegmund, 1781
  B.; Zaumseil, J.; Gather, M.C. Spectroscopic Near-Infrared Photodetectors Enabled by Strong Light–Matter 1782
  Coupling in (6,5) Single-Walled Carbon Nanotubes. *The Journal of Chemical Physics* 2020, 153, 201104, 1783
  doi:10.1063/5.0031293. 1784
- 203. Shahnazaryan, V.A.; Saroka, V.A.; Shelykh, I.A.; Barnes, W.L.; Portnoi, M.E. Strong Light-Matter Coupling in 1785 Nanotubes a Route to Exciton Brightening. ACS Photonics 2019, 6, 904–914, Carbon as 1786 doi:10.1021/acsphotonics.8b01543. 1787
- 204. Lasave, L.C.; Urteaga, R.; Koropecki, R.R.; Gonzalez, V.D.; Arce, R.D. Real-Time Study of Protein Adsorption 1788
   Kinetics in Porous Silicon. *Colloids and Surfaces B: Biointerfaces* 2013, 111, 354–359, doi:10.1016/j.colsurfb.2013.06.024. 1789
- 205. Liang, L.; Zheng, P.; Zhang, C.; Barman, I. A Programmable DNA-Silicification-Based Nanocavity for Single Molecule Plasmonic Sensing. *Advanced Materials* 2021, 33, 2005133, doi:10.1002/adma.202005133.
   1791
- 206. Allison, G.; Sana, A.K.; Ogawa, Y.; Kato, H.; Ueno, K.; Misawa, H.; Hayashi, K.; Suzuki, H. A Fabry-Pérot Cavity
   Coupled Surface Plasmon Photodiode for Electrical Biomolecular Sensing. *Nat Commun* 2021, 12, 6483, 1793
   doi:10.1038/s41467-021-26652-7.
- 207. Thomas, P.A.; Menghrajani, K.S.; Barnes, W.L. All-Optical Control of Phase Singularities Using Strong Light Matter Coupling. *Nat Commun* 2022, *13*, 1809, doi:10.1038/s41467-022-29399-x.
- 208. Wang, Y.; Zeng, S.; Crunteanu, A.; Xie, Z.; Humbert, G.; Ma, L.; Wei, Y.; Brunel, A.; Bessette, B.; Orlianges, J.-C.;
  et al. Targeted Sub-Attomole Cancer Biomarker Detection Based on Phase Singularity 2D Nanomaterial-Enhanced
  Plasmonic Biosensor. *Nano-Micro Lett.* 2021, *13*, 96, doi:10.1007/s40820-021-00613-7.
- 209. Sreekanth, K.V.; Sreejith, S.; Han, S.; Mishra, A.; Chen, X.; Sun, H.; Lim, C.T.; Singh, R. Biosensing with the Singular
   Phase of an Ultrathin Metal-Dielectric Nanophotonic Cavity. *Nat Commun* 2018, *9*, 369, doi:10.1038/s41467-018 02860-6.
- 210. Ermolaev, G.; Voronin, K.; Baranov, D.G.; Kravets, V.; Tselikov, G.; Stebunov, Y.; Yakubovsky, D.; Novikov, S.;
  1803
  Vyshnevyy, A.; Mazitov, A.; et al. Topological Phase Singularities in Atomically Thin High-Refractive-Index
  1804
  Materials. *Nat Commun* 2022, 13, 2049, doi:10.1038/s41467-022-29716-4.
- 211. Wu, F.; Thomas, P.A.; Kravets, V.G.; Arola, H.O.; Soikkeli, M.; Iljin, K.; Kim, G.; Kim, M.; Shin, H.S.; Andreeva, 1806
   D.V.; et al. Layered Material Platform for Surface Plasmon Resonance Biosensing. *Sci Rep* 2019, *9*, 20286, 1807
   doi:10.1038/s41598-019-56105-7. 1808
- Jain, S.; Choudhary, K.; Kumar, S. Photonic Crystal Fiber-Based SPR Sensor for Broad Range of Refractive Index
   Sensing Applications. *Optical Fiber Technology* 2022, *73*, 103030, doi:10.1016/j.yofte.2022.103030.

8481, doi:10.1109/JSEN.2022.3160482.

1811

1812

1813

1814

	Monitoring in Microdisk Optofluidic Sensor through Microcavity Enhanced Raman Scattering. Colloids and Surfaces	1815
	A: Physicochemical and Engineering Aspects 2023, 679, 132561, doi:10.1016/j.colsurfa.2023.132561.	1816
215	. Vázquez-Guardado, A.; Smith, A.; Wilson, W.; Ortega, J.; Perez, J.M.; Chanda, D. Hybrid Cavity-Coupled	1817
	Plasmonic Biosensors for Low Concentration, Label-Free and Selective Biomolecular Detection. Opt. Express 2016,	1818
	24, 25785, doi:10.1364/OE.24.025785.	1819
216	. Vázquez-Guardado, A.; Mehta, F.; Jimenez, B.; Biswas, A.; Ray, K.; Baksh, A.; Lee, S.; Saraf, N.; Seal, S.; Chanda,	1820
	D. DNA-Modified Plasmonic Sensor for the Direct Detection of Virus Biomarkers from the Blood. Nano Lett. 2021,	1821
	21, 7505–7511, doi:10.1021/acs.nanolett.1c01609.	1822
217	. Anderson, M.; O'Brien, E.; Grayek, E.; Hermansen, J.; Hunt, H. The Detection of Helicobacter Hepaticus Using	1823
	Whispering-Gallery Mode Microcavity Optical Sensors. Biosensors 2015, 5, 562–576, doi:10.3390/bios5030562.	1824
218	. Wilson, K.A.; Finch, C.A.; Anderson, P.; Vollmer, F.; Hickman, J.J. Whispering Gallery Mode Biosensor	1825
	Quantification of Fibronectin Adsorption Kinetics onto Alkylsilane Monolayers and Interpretation of Resultant	1826
	Cellular Response. Biomaterials 2012, 33, 225–236, doi:10.1016/j.biomaterials.2011.09.036.	1827
219	. Frustaci, S.; Vollmer, F. Whispering-Gallery Mode (WGM) Sensors: Review of Established and WGM-Based	1828
	Techniques to Study Protein Conformational Dynamics. Current Opinion in Chemical Biology 2019, 51, 66–73,	1829
	doi:10.1016/j.cbpa.2019.05.003.	1830
220	. Granizo, E.; Knysh, A.; Sokolov, P.; Samokhvalov, P.; Nabiev, I. Polaritonic Photocatalysis and Polariton-Driven	1831
	Control of Energy Relaxation Pathways in a Tunable Microcavity. Phys. Atom. Nuclei 2023, 86, 2454-2458,	1832
	doi:10.1134/S1063778823110145.	1833
221	. Thomas, P.A.; Menghrajani, K.S.; Barnes, W.L. Cavity-Free Ultrastrong Light-Matter Coupling. J. Phys. Chem. Lett.	1834
	<b>2021</b> , <i>12</i> , 6914–6918, doi:10.1021/acs.jpclett.1c01695.	1835
222	. Kaeek, M.; Damari, R.; Roth, M.; Fleischer, S.; Schwartz, T. Strong Coupling in a Self-Coupled Terahertz Photonic	1836
	Crystal. ACS Photonics 2021, 8, 1881–1888, doi:10.1021/acsphotonics.1c00309.	1837
223	. Munkhbat, B.; Baranov, D.G.; Stührenberg, M.; Wersäll, M.; Bisht, A.; Shegai, T. Self-Hybridized Exciton-	1838
	Polaritons in Multilayers of Transition Metal Dichalcogenides for Efficient Light Absorption. ACS Photonics 2019, 6,	1839
	139–147, doi:10.1021/acsphotonics.8b01194.	1840
224	. An, G.; Hao, X.; Li, S.; Yan, X.; Zhang, X. D-Shaped Photonic Crystal Fiber Refractive Index Sensor Based on	1841
	Surface Plasmon Resonance. <i>Appl. Opt.</i> <b>2017</b> , <i>56</i> , 6988, doi:10.1364/AO.56.006988.	1842
225	. Gao, R.; Ko, J.; Cha, K.; Ho Jeon, J.; Rhie, G.; Choi, J.; deMello, A.J.; Choo, J. Fast and Sensitive Detection of an	1843
	Anthrax Biomarker Using SERS-Based Solenoid Microfluidic Sensor. <i>Biosensors and Bioelectronics</i> 2015, 72, 230–236,	1844
	doi:10.1016/j.bios.2015.05.005.	1845
226	. Arul, R.; Grys, DB.; Chikkaraddy, R.; Mueller, N.S.; Xomalis, A.; Miele, E.; Euser, T.G.; Baumberg, J.J. Giant Mid-	1846
	IR Resonant Coupling to Molecular Vibrations in Sub-Nm Gaps of Plasmonic Multilayer Metafilms. <i>Light Sci Appl</i>	1847
	<b>2022</b> , <i>11</i> , 281, doi:10.1038/s41377-022-00943-0.	1848
227	. Fu, L.; Lu, Q.; Liu, X.; Chen, X.; Wu, X.; Xie, S. Combining Whispering Gallery Mode Optofluidic Microbubble	1849
	Resonator Sensor with GR-5 DNAzyme for Ultra-Sensitive Lead Ion Detection. Talanta 2020, 213, 120815,	1850
	doi:10.1016/j.talanta.2020.120815.	1851

213. Chaudhary, V.S.; Kumar, D.; Mishra, G.P.; Sharma, S.; Kumar, S. Plasmonic Biosensor With Gold and Titanium

214. Wang, W.; Zhang, J.; Ye, P.; Liu, W.; Mao, W.; Shi, Z.; Cui, Q.; Wang, M.; Zhu, G.; Xu, C. Protein Aggregation

Dioxide Immobilized on Photonic Crystal Fiber for Blood Composition Detection. IEEE Sensors J. 2022, 22, 8474-

228. 1	Li, Z.; Zhu, C.; Guo, Z.; Wang, B.; Wu, X.; Fei, Y. Highly Sensitive Label-Free Detection of Small Molecules with	1852
ar	n Optofluidic Microbubble Resonator. Micromachines 2018, 9, 274, doi:10.3390/mi9060274.	1853