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# Home made SM 100bp DNA ladder for agarose gel v1

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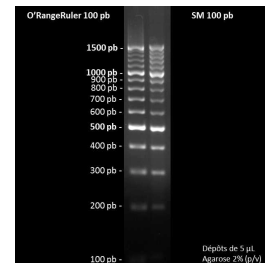
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## 🌐 Home made SM 100bp DNA ladder for agarose gel

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**Protocol status:** Working

**We use this protocol and it's working**

**Created:** August 01, 2024

**Last Modified:** August 01, 2024

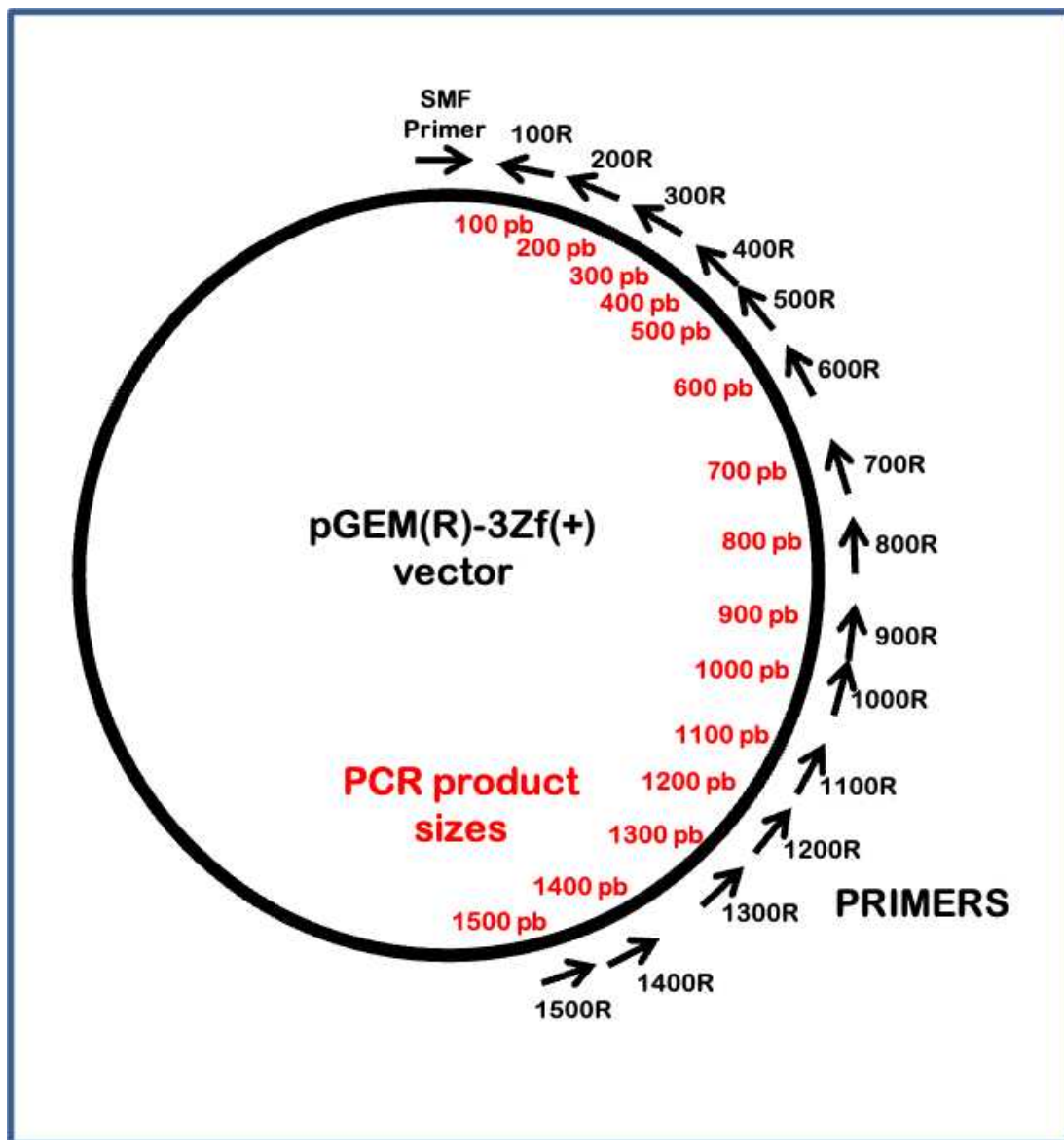
**Protocol Integer ID:** 104413

**Keywords:** double-stranded DNA sizing, agarose gel, 100pb ladder

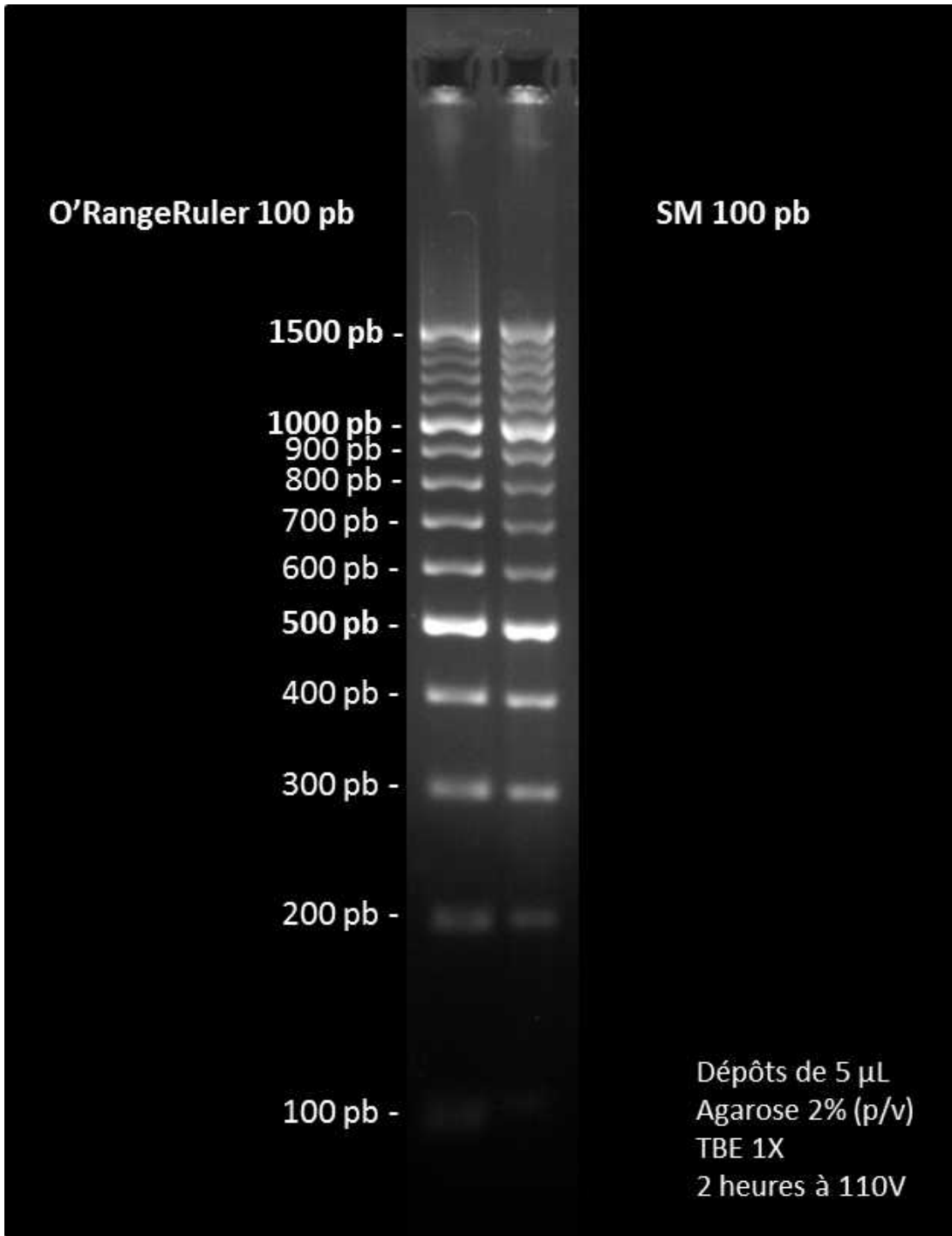
## Abstract

The SM 100pb DNA Ladder is a standard size marker equivalent to the Fisher's O'RangeRuler<sup>®</sup> 100 pb DNA Ladder (#SM0623). The SM 100pb Ladder allows to determine the size of double-stranded DNA fragments between 100 bp and 1500 bp and it is composed of 15 double-stranded DNA fragments of 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000, 1100, 1200, 1300, 1400 and 1500 bp. Like the O'RangeRuler 100 bp, the SM 100 bp gives more intense bands at 500, 1000 and 1500 bp.

**At the end, the SM 100 pb DNA Ladder is 25 times cheaper than the O'RangeRuler 100 pb.**



Map of the universal cloning vector pGEM<sup>®</sup>-3Zf(+) showing the position of the 16 primers



Comparaison between O'rangeRuler 100pb (Thermofisher) and SM100bp ladder on 2% agarose gel


## Protocol materials


 GoTaq® Flexi DNA Polymerase **Promega Catalog #M8291** Step 3


 pGEM®-3Zf( ) Vectors **Promega Catalog #P2271** Step 2



## Primers and pGEM®-3Zf(+) Vectors preparation

### 1 Oligo primers dilution


In  1.5 mL microtubes, dilute 1:10 each oligo primer at 100µM to final concentration 10µM, **see the SM100pb Primers.xlsx file below.**

 50 µL oligo primer (100µM)


 450 µL nuclease free water


Homogenize and store at  4 °C (or at  -20 °C for a long-term storage).


SM100pb oligo primers sequences (salt free purification):



 SM100pb Primers.xlsx 12KB

### 2 pGEM®-3Zf(+) Vectors dilution

In  1.5 mL microtube, dilute 1:100 pGEM®-3Zf(+) vector at 1µg/µL to final concentration 10ng/µL.

 5 µL pGEM®-3Zf(+) (1µg/µL)

 495 µL nuclease free water


Homogenize and store at  4 °C (or at  -20 °C for a long-term storage).


 pGEM®-3Zf(+) Vectors **Promega Catalog #P2271**

## PCR amplification to generate 100pb-1500pb double-stranded DNA fragments 10s

### 3 PCR mix preparation 10s

SM100pb is produced using **28 PCRs amplifications** in a total of  50 µL reaction volume:

1 x  50 µL for 600pb, 700pb, 800pb, 900pb, 1100pb, 1200pb, 1300pb and 1400pb fragments

2 x  50 µL for 200pb, 300pb and 400pb fragments

3 x  50 µL for 1000pb and 1500pb fragments

4 x  50 µL for 100pb and 500pb fragments

Defreeze and vortex all reagents, except enzymes (stored at  $-20\text{ }^{\circ}\text{C}$ ), for approximately

00:00:05

Spin down all reagents for approximately 00:00:05 and place On ice.

In 1.5  $\mu\text{L}$  microtube, prepare the PCR mix according to the following table :

A	B	C	D	E
	Initial concentration	Final concentration	n=1	n=28
SMF forward primer	10 $\mu\text{M}$	400mM	2 $\mu\text{L}$	56 $\mu\text{L}$
Green GoTaq buffer	5X	1X	10 $\mu\text{L}$	280 $\mu\text{L}$
MgCl <sub>2</sub>	25mM	1mM	2 $\mu\text{L}$	56 $\mu\text{L}$
pGEM®-3Zf(+) vecto	10ng/ $\mu\text{L}$	20ng	2 $\mu\text{L}$	56 $\mu\text{L}$
dNTP mix	2.5mM	150 $\mu\text{M}$ each	3 $\mu\text{L}$	84 $\mu\text{L}$
GoTaq polymerase	5 u/ $\mu\text{L}$	1.75U	0.35 $\mu\text{L}$	9.8 $\mu\text{L}$
nuclease free water			28.65 $\mu\text{L}$	802.2 $\mu\text{L}$
TOTAL			48 $\mu\text{L}$	1344 $\mu\text{L}$

PCR mix composition

GoTaq® Flexi DNA Polymerase **Promega Catalog #M8291**


#### 4 Reverse primers and mix combinaison

Defreeze and vortex all the 15 reverse oligo primers at 10 $\mu\text{M}$

In a 96-well plate PCR, transfer 2  $\mu\text{L}$  of each reverse oligo primers (10 $\mu\text{M}$ ) according to the following map :

A	B	C	D	E	F	G	H
600R	200R	1000R	500R				
700R	200R	1500R	500R				
800R	300R	1500R	500R				
900R	300R	1500R	500R				
1100R	400R	100R					
1200R	400R	100R					
1300R	1000R	100R					
1400R	1000R	100R					

Map of the PCR plate

Vortex and spin down the PCR mix tube, transfer  48  $\mu\text{L}$  in each of the 28 wells.

Seal the PCR plate.

In thermocycler, run PCR amplification with cycles follows:


A	B	C	D
Cycles step	Temperature	Time	Cycles
Initial denaturation	94°C	5 min	1
Denaturation	94°C	30 sec	40
Annealing	60°C	30 sec	40
Extension	72°C	30 sec	40
Final extension	72°C	60 min	1
Hold	4°C		


PCR program



After PCR, pool and dilute 1:2 the PCR amplification



In a  5 mL tube:

 1400  $\mu\text{L}$  of PCR amplification (28 x  50  $\mu\text{L}$  )

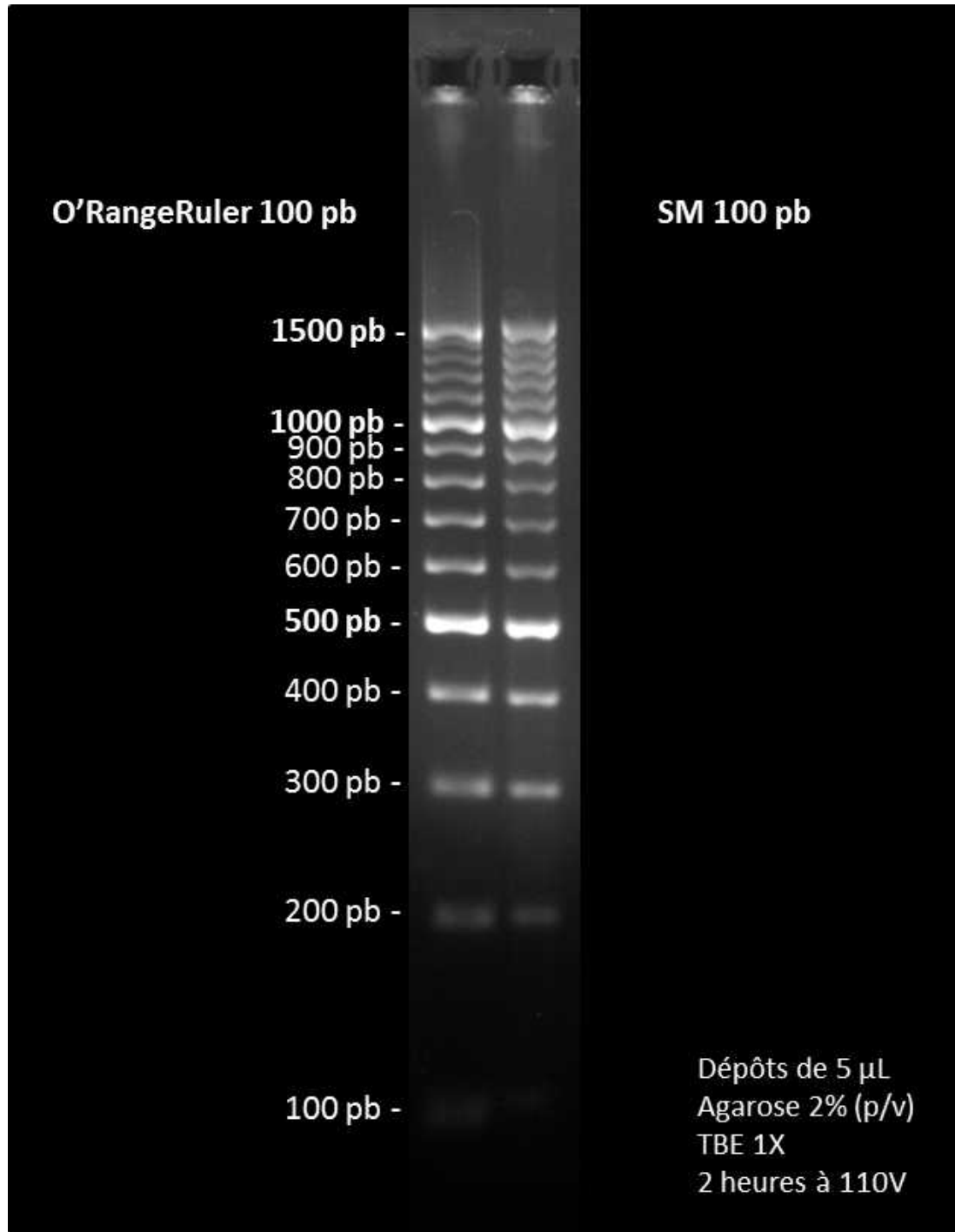
 420  $\mu\text{L}$  5X green GoTag buffer

 980  $\mu\text{L}$  nuclease free water

Homogenize and store at  4 °C (or at  -20 °C for a long-term storage).

Load  5  $\mu\text{L}$  to  10  $\mu\text{L}$  per line in a agarose gel.





Comparaison between O'rangeRuler 100pb (Thermofisher) and SM100bp ladder on 2% agarose gel