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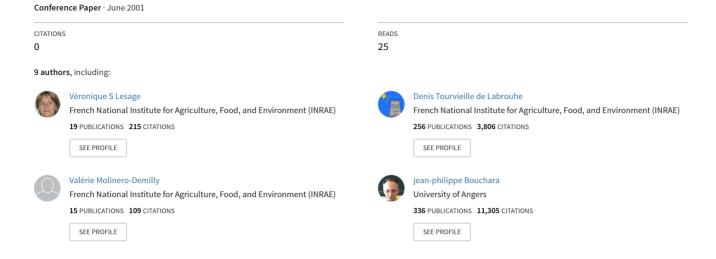
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Development of a sandwich ELISA and a PCR -based assays for the diagnosis of downy mildew in sunflower seed



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for the diagnosis of downy mildew in sunflower seed

Bouterige S.*1, Says-Lesage V.*2, Meliala C.3, Tourvieille de Labrouhe D.3, Molinéro-Demilly V.3, Bouchara J.P.1, Nicolas P.3, Roeckel-Drevet P.2, Robert R.1

*The contributions of the two first authors are equivalent

Introduction



Downy mildew repartition in France during the year 1999. (Source SEPV Mid-Pyrenees)

Race 710 Races 710 and 703 Race 703

The downy mildew of sunflower, caused by Plasmopara halstedii (Farlow) Berlese et de Toni, is a widespread and destructive pathogen known to occur in almost all areas of intensive sunflower cultivation. Typical symptoms include chlorosis of the leaves, often associated with sever stunting of systemically infected plants, and various types and degrees of leaf distortion. Infected plants were shown to have higher respiration. rate and higher enzyme activity as compared with healthy plants. The effect of this pathogen has considerable economic implications. Owing to the risk of propagation of the disease by infected seeds and to the regular description of metalaxyl-resistant isolates of the fungus, there is a greater need for efficient methods to detect the fungus in seeds or plant tissues. Therefore, we developed two detection methods, one based on monoclonal antibodies, the other on Polymerase Chain Reaction.

In the first method, we produced two monoclonal antibodies (MAbs) by immunization of mice with a partially purified extract of P. halstedii race 100. Using ELISA technique, both MAbs detected all races of P. halstedii present in France, but not the other molds pathogenic to sunflower or encountered as saprophytes on the seeds. The specificity of these MAbs for P. halstedii and their recognition of distinct epitopes on the

fungal antigens were used to develop a sandwich ELISA detection system. Incubation of seed or tissue homogenates from infected plants in wells coated with MAb 18E2 resulted in the trapping of P. halstedii antigens that were identified with biotinylated MAb 12C9.

The second method using PCR was developed using specific primers designed from the nucleotide sequence of one cloned random amplified polymorphic DNA fragment. Agarose gel electrophoresis stained with ethidium bromide revealed a 1038-bp amplification product for various races of P. halstedii. No staining was seen with DNA from other saprophytic or pathogenic molds, nor with the host plant DNA.

We analyzed thirty seed lots: fifteen were checked for the presence of P. halstedii and fifteen others for the absence of the pathogen with optical microscopy. The aim of this study was to compare the immunological and the PCR-based techniques for the detection of downy mildew in sunflower.

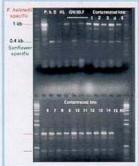
ELISA procedure

Coating the well with the Mab 12C9 in PBS-Addition of a chromogenic PAL-substrate 0

PCR procedure

DNA was extracted from 500 mg of whole seed, with a combined

method using Dellaporta (Plant Mol Biol Rep. 1983) and Amersham (RPN8511) procedures, and a further purification step to eliminate PCR inhibitors present in the seed. Thirty amplification cycles were conducted with an annealing temperature of 60 °C, and two sets of primers: one set was specific of all races of P. halstedii, the other one was sunflower specific, for an internal amplification control



P. h.: Plasmopara halstedii S: Sunflower (healthy leaves) HL: Healthy seed Lot GM: contaminated lot as NC: Negative Control (water)

For PCR, results showed two amplified fragments for all the contaminated lots. The highest, around 1 kb long, and the lowest, around 400 bp, were P. halstedii and sunflower specific respectively. The 400-bp fragment was the only one amplified both in healthy seeds and leaves. The negative control showed no amplification.

Results of the two methods

	ELISA Test		PCR Test
	DO value	Results	Results
Seed lots	Contaminated lots		
Lot I	0.279		++
Lot 2	0.382		+++
Lot 3	0,353	+	++
Lot 4	0,261	+	911
Lot 5	0,870	+++	+
Lot 6	0,556	++	**
Lot 7	0,508	44	++
Lot 8	0,510	- ++	1 .+:
Lot 9	0,416	**	1.4.1
Lo: 10	1,613	***	+++
Lot 11	0,354	. +	
Lot 12	0,373		14.5
Ln: 13	1,234	111	+
Lo: 14	1,074	+++	++
Lat 15	0,402	++	+
	Healthy lots		
15 lots	< 0.195		
Negative control	0,195		1 55

For the ELISA test, the results showed that the lots corresponding to healthy plants presented a negative response (inferior to the negative control). Conversely, the lots corresponding to contaminated plants showed, for all of them, a positive response (superior to the negative control). The statistical study, performed by the non-parametric test of Mann-Whitney, showed that the difference obtained between lots, coming from infected and healthy plants, are highly significant for p < 0.01.

Discussion

Finally, both methods allowed the detection of the fungus in all infected seed lots. However, contamination in these lots varied from 70 to 100 %, which was probably much higher than

that we can expect in commercial seed lots. We have made, therefore, further PCR studies on various lots provided by the phytosanitary administration of the Midi-Pyrénées region. Downy mildew was detected in lots containing 10 % of seeds taken from plants showing downy mildew symptoms. In conclusion, these methods may be used to guarantee the phytosanitary quality of seeds, and therefore may help reduce the incidence of seed-borne disease transmission.

Groupe d'Étude des iInteractions Hôtes-Parasites Laboratoire d'Immunologie-Parisitologie Faculté de Pharmacie, 16 Bd Daviers, 49100 Angers

INRA - Université Blaise Pascal UMR Amélioration et Santé des Plantes 24 Avenue des Landais, 63177 Aubière, France

^{III} GEVES-SNES Station Nationale d'Essais de Semences rue G. Morel, BP 24, 49071 Beaucouzé Cedex, France