

# Early Plasma Interferon- $\beta$ Levels as a Predictive Marker of COVID-19 Severe Clinical Events in Adult Patients Running title: Early IFN- $\beta$ levels in COVID-19 patients

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### Abstract (187 words):

We assessed relationships between early peripheral blood type I Interferons levels, clinical new early warning scores (NEWS) and clinical outcomes in hospitalized COVID-19 adult patients. Early IFN-  $\beta$  levels were lower among patients who further required intensive care unit (ICU) admission than those measured in patients who did not require an ICU admission during SARS-CoV-2 infection. IFN- $\beta$  levels were inversely correlated with NEWS only in the subgroup of patients who further required ICU admission. To assess whether peripheral blood IFN- $\beta$  levels could be a potential relevant biomarker to predict further need for ICU admission, we performed Receiver Operating Characteristic (ROC) curve analyses that showed for all study patients an area under ROC curve of 0.77 growing to 0.86 (P= 0.003) when the analysis was restricted to a subset of patients with NEWS  $\geq$  5 at the time of hospital admission. Overall, our findings indicated that early peripheral blood IFN- $\beta$  levels might be a relevant predictive marker of further need for an intensive care unit admission in hospitalized COVID-19 adult patients, specifically when clinical score (NEWS) was graded as upper than 5 at the time of hospital admission.

Keywords: COVID-19, SARS-CoV-2, type I Interferons, NEWS, intensive care unit

### **BACKGROUND:**

It is now well admitted that SARS-CoV-2 caused coronavirus disease-19 (COVID-19) clinical stages evolve from early infection (stage I) towards a mild-to-moderate pulmonary involvement without or with hypoxia (stage II), which can be followed by an hyperinflammatory phase resulting in potential severe clinical events requiring an intensive care unit (ICU) admission (clinical stage III) <sup>1</sup>. Pathophysiology of SARS-CoV-2 infection is clearly characterized by an early viral replicative phase in respiratory tract tissues associated with low viremia levels (clinical stage I) during 8 days after the first onset of symptoms, followed by an inflammatory phase occurring from 9 to 12 days after the first onset of symptoms (clinical stages II and III).

COVID-19 inflammatory phase is characterized by a dysregulated inflammatory host response with an abnormal release of proinflammatory cytokines mediated by the activation of cell surface or intracellular toll like Receptors (TLRs) <sup>2-6</sup>. Remarkably, a low interferon-Beta (IFN-β) production in the peripheral blood of severe (clinical stage III) COVID-19 patient was evidenced suggesting that type I IFN low production levels could drive the development of clinical stages II and III during the course of the SARS-CoV-2 infection<sup>7</sup>. Type I IFNs influence the development of innate and adaptive immune responses limiting virus replication activities and preventing the development of severe clinical events during viral respiratory infections<sup>8,9</sup>. However, patterns of type I IFNs production during SARS-CoV-2 infection, were investigated only in COVID-19 patients at clinical stages II and III (9-12 days after onset of first symptoms)<sup>7,10</sup>. In the present time, type I IFN expression pattern in the early phase of COVID-19 (clinical stage I) and its potential relationship with further development of severe clinical events remains to be investigated.

In the present retrospective investigation, we assessed peripheral blood type I IFNs and SARS-CoV-2 RNA levels at early phase (1-8 days after the first symptom onset) of COVID-19 in adult patients who further required or not an ICU admission during infection. Among these two subsets of patients, we assessed potential relationships between early blood type I IFNs and SARS-CoV-2 RNA levels and clinical new early warning scores (NEWS) at the time of hospitalisation.

### **CLINICAL SAMPLES and METHODS:**

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81 Study design: From November 2020 to March 2021, peripheral blood EDTA-plasma samples taken 82 83 from adult patients had been stored at -80°C in Reims University Hospital (Grand-Est, France) until processing for anti SARS-CoV-2 IgM or IgG antibodies detection assays. 84 This biobank was declared as a biological specimen's research collection and was 85 registered by the French Ministry of High Education and Research (number DC-86 87 2020-4052). From this research biobank samples (n=4704 sampled patients), we retrospectively 88 89 selected samples taken from adult patients (i) hospitalized in medicine wards for 90 COVID-19 (n=1116); (ii) with a time delay less than 9 days between sample collection and onset of COVID-19 symptoms (n= 183); (iii) with full clinical data 91 92 available from a registered clinical data bank (Reims University Hospital GDPR number RMR004-061120) elaborated for an internal purpose between March 1<sup>st</sup> 93 2020 and March 31<sup>rd</sup> 2021 and focusing on COVID-19 adult patients (n=143). 94 95 Patients who required palliative care or hospitalized directly in ICU were excluded from the study. 96 97 Finally, only 43 COVID-19 adult patients with available frozen EDTA-plasma samples 98 and with full registered clinical data were included in this retrospective investigation. 99 100 Demographic, clinical and medical data collection: For each selected patient, the following data had been previously registered in a 101 102 declared database (RMR004-061120) during their hospital stage: sex, age, comorbidities, immunocompromised state, date of symptoms onset, new early 103 warning score (NEWS) graded at the time of hospital admission<sup>11</sup>, computerized 104

tomography chest scan results (CT-chest scan), drugs prescription and further need for intensive care unit admission (ICU) and in-hospital mortality. Demographic and clinical characteristics of study patients are depicted in Table 1.

### Ethical approval:

The Institutional Ethics Committee (IEC), University hospital of Reims (Grand-Est, France) waived the requirement of informed consent and approved all methods and protocols. The study was conducted in accordance with the Declaration of Helsinki Ethical Principles and Good Clinical Practices. Patients did not provide individual consent because of the retrospective and non-interventional nature of this study, in accordance with French legislation. No patient raised an opposition to the use of their recorded medical data or their samples collected at the time of hospitalization for COVID-19. Data confidentiality was preserved during this internal study (Reims University Hospital GDPR register number RMR004-061120), and research collection of biological specimens was registered in French Ministry of Higher Education and Research as number DC-2020-4052.

### Peripheral blood type I interferons and SARS-CoV-2 RNA load

Using our research biological samples (registered research biobank), peripheral blood levels of Interferon-alpha (IFN-α), Beta (IFN-β) were determined on plasma samples using ELISA *(enzyme-linked immunosorbent assay)* (Biotechne, France) according to the manufacturer's instructions. In parallel, SARS-CoV-2 RNA load in EDTA plasma samples were quantified using droplet based digital PCR (dd-PCR) (MOBICYTE technical platform, Reims, France), as previously described<sup>7</sup>, with Bio-Rad SARS-CoV-2 dd-PCR kit (Bio-Rad, France). Viral RNA was extracted using

QIAmp viral RNA mini kit (Qiagen) according to the manufacturer's guidelines. Viral concentration results (copies/ml, cp/ml) were calculated considering the extraction volumes of plasma (140  $\mu$ l).

### Statistical analyses:

Quantitative variables expressed as median [range] were compared using the Mann Whitney U-test. Qualitative variables expressed as percentages were compared using Fischer's exact test or Pearson Chi-2 if applicable. When linear regression was performed, slopes were compared using Spearman test. When Receiver operating characteristic (ROC) curve analysis was performed, Area under ROC Curve, Standard Error and P value were calculated using the trapezoid rule, the Hanley Method and Z ratio respectively. A *P* value<0.05 was considered statistically significant. Statistical analyses were performed using GraphPad Prism 8 (GraphPad) and SAS version 9.4 (SAS Institute Inc).

### Data availability:

The datasets generated and/or analysed during the current study are not publicly available according to French law guidelines and registrations but are available from the corresponding author on reasonable request.

### **RESULTS**

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Early peripheral blood IFN-β, SARS-CoV-2 RNA levels and further need for

intensive care unit admission in COVID-19 adult patients

Type I Interferons (IFNs) and SARS-CoV-2 RNA load were assessed in peripheral blood samples taken from adult patients after a median time delay of 5 [1-8 days after the onset of symptoms] (Table1). Among the 43 selected patients, early peripheral blood IFN-B were detected in all the study patients with values ranging from 40 to 929 pg/ml, whereas peripheral blood IFN-α were detectable in only 86% of the study patients with values ranging from non-detectable to 326 pg/ml (Figure 1A). Because low type I IFN production levels could drive the development of COVID-19 induced inflammatory phase, we segregated our patient group into two subsets according for their further need for an ICU admission during infection. Remarkably, levels of IFN-β appeared to be significantly lower among the subset of patients who further required ICU admission than the subset of patients who did not further require ICU admission (P=0.001) (Figure 1B). For IFN- $\alpha$  levels, no significant differences between patient subsets requiring or not ICU admission was evidenced at the time of hospitalization (Figure 1C). In addition, the plasma SARS-CoV-2 RNA load appeared to be significantly higher in patients who further required ICU admission than those measured in samples of patients who did not further required ICU admission (P= 0.03) (Figure 1D).

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### Associations between early peripheral blood IFN-β levels, SARS-CoV-2 RNA

### loads and clinical new early warning scores

Because clinical new early warning scores (NEWS) had been previously associated with further ICU admission events in COVID-19 adult patients<sup>11</sup>, we first sought for a

potential association between NEWS and IFN- $\beta$  concentrations and then between NEWS and SARS-CoV-2 RNA load according to a further need for ICU admission. Among the subset of patients who further required an ICU admission, a significant negative correlation between NEWS and IFN- $\beta$  concentrations (P= 0.045;  $R^2$ = 0.226) was observed (Figure 2A, left panel). By contrast a non-significant positive correlation was observed between NEWS and IFN- $\beta$  concentrations among the subset of patients who did not require further ICU admission (P= 0.182;  $R^2$ = 0.075) (Figure 2A, right panel). No significant correlation was evidenced between NEWS and SARS-CoV-2 RNA load among the two subsets of study patients (Figure 2B). Remarkably, when the analysis was restricted to a subset of patients with a NEWS ≥ 5 at hospital admission we observed a significant negative correlation between blood IFN- $\beta$  level and plasma SARS-CoV-2 RNA load in the subset of COVID-19 patient who did not further require an ICU admission. Such a correlation was not observed among patients who further require ICU admission (supplementary Figure 1).

## Early Blood IFN-β concentrations accuracy to predict the need for a further ICU admission

To assess whether peripheral blood IFN- $\beta$  levels could be a relevant biomarker to predict further need for ICU admission, we performed Receiver Operating Characteristic (ROC) curve analyses (Figure 3). Including all study patients, the sensitivity and specificity were respectively of 88% and 50% with an area under the receiver operating characteristic curve (AUC) of 0.77 (Figure 3A). Remarkably, the area under the ROC curve of blood IFN- $\beta$  level grew to 0.86 when the analysis was restricted to a subset of patients with a NEWS  $\geq$  5 at hospital admission indicating a relevant predictive value of early IFN- $\beta$  levels among the most clinically severe study

patients. Among the subset of patients with NEWS ≥ 5 at hospital admission, peripheral blood IFN-β levels below 107.6 pg/ml were associated to a further ICU requirement with a specificity of 100% and a sensitivity of 60% (Figure 3B). 

### **DISCUSSION**

Investigation of peripheral blood type I IFN production patterns in SARS-CoV-2 infected adult patients is of major interest for understanding COVID-19 pathophysiology and clinical courses of infection  $^{7,10}$ . Herein, we assessed relationships between early peripheral blood Interferon-Beta (IFN- $\beta$ ) levels, clinical new early warning scores (NEWS) and clinical outcomes in SARS-CoV-2 infected adult patients hospitalized from 1 to 8 days after the first symptom onset (clinical stage I).

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Among the 43 selected patients, anti-SARS-CoV-2 total antibodies were not detected and interleukin 6 (IL6) and tumour necrosis factor-alpha (TNF-α) were not measured at significant levels in peripheral blood (data not shown). These results were associated with detectable SARS-CoV-2 RNA load confirming that study patients were at the viral phase of COVID-19 (Figure 1D). Interestingly, early peripheral blood IFN-β were detected in all the study patients, whereas IFN-α were detectable only in 86% of the study patients (Figure 1A). Because low type I IFN production levels could drive the development of COVID-19 induced inflammatory phase (clinical stages II and III), we segregated our patient group into two subsets according for their further need for an ICU admission. Interestingly, our findings indicated for the first time that patients who further required ICU admission demonstrated significant lower peripheral blood IFN-β levels and higher SARS-CoV-2 RNA load than those observed in patients who did not require ICU admission (Figure 1). Low early IFN-β concentration levels in patient could be related to inter-individual sequences variability in human genes implicated into the regulation of type I IFN innate immunity<sup>12</sup>, but also to the translation of SARS-CoV-2 nonstructural proteins

inhibiting the IFN-β promoter activation<sup>13</sup> or to preexisting peripheral blood autoantibodies directed against human type I IFNs<sup>14</sup>.

Among the subset of COVID-19 patients who further required ICU admission, a negative correlation between peripheral blood IFN- $\beta$  concentrations and clinical NEWS was evidenced, whereas no significant association was observed between clinical scores and SARS-CoV-2 RNA levels (Figure 2). These findings were completed by ROC analyses and results indicated that early peripheral blood IFN- $\beta$ , but not SARS-CoV-2 RNA load, might be a relevant biomarker of a further ICU admission, specifically among patients with a NEWS  $\geq$  5 at the time of hospital admission (Figure 3B). Overall, these findings supported the concept that an early IFN- $\beta$  treatment in COVID-19 adult patients hospitalized at phase I of SARS-CoV-2 infection might be of therapeutic benefit improving the clinical outcomes. However, these findings remain to be fully investigated in further prospective large multicenter clinical investigations.

### Conclusions

Even if our study is a limited retrospective monocentric investigation conducted before large anti-SARS-CoV-2 vaccination campaigns in general population and before the emergence of new Delta or Omicron viral variants, our original findings revealed a significant association between early low peripheral blood IFN-β levels and high clinical scores (NEWS) during SARS-CoV-2 clinical phase I. Overall, our findings indicated that early peripheral blood IFN-β levels might be a relevant predictive marker of further need for an intensive care unit admission in COVID-19 adult patients, specifically when clinical score (NEWS) was graded as upper than 5 at hospital admission.

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### Author's contribution

- 297 L.A, Y.N'G and F.B designed the study. L.A, F.B and Y.N'G wrote the manuscript.
- 298 L.A, BN. P and Y.N'G established the research biobank, the clinical data bank and
- supervised the data collection. F.B and Y.N'G analyzed the data and designed the
- table and the figures. D.C helped for data collection. F.B, Y.N'G, L.A, D.C, A-L.L,
- 301 M.G, L.H, F.B-S and BN. P provided valuable input to interpretation of the data and
- 302 critically reviewed the manuscript. All these authors approved the final version of the
- 303 present manuscript.

305 Financial support 306 This work was supported by a research grant URCA-2020-EA4684 dedicated to 307 research teams from the University of Reims Champagne-Ardenne. 308 309 Conflict of interest 310 The authors report no conflict of interest. 311 312 Data availability: 313 The datasets generated and/or analyzed during the current study are not publicly 314 available according to French law guidelines and registrations but are available from the corresponding author on reasonable request. 315 316 317 Reims Covid-19 Study group. Collaborators included the following: Dr Ezekiel 318 Bankole, Dr Loïs Bolko, Dr Michel Bonnivard, Dr Nelly Bourse, Dr Aurélie Brunet, Dr 319 Sandy Carazo Mendez, Dr Clement Chopin, Dr Joël Cousson, Dr Sophie Demotier, 320 Pr Gaetan Deslee, Dr Vincent Dupont, Dr Thierry Floch, Dr Julien Gautier, Dr Guillaume Giordano Orsini, Dr Sophie Giordano Orsini Dr Antoine Goury, Dr Marie 321 Laure Gross, Dr Valentin Houannou, Dr Claire Lepouse, Dr Gauthier Lejeune, Dr 322 323 Natacha Noël, Pr Jeanne Marie Perotin, Dr Juliette Romaru, Dr Jerome Tassain. Pr. 324 Bruno Mourvillier.

### 325 TABLE AND FIGURES

 Table 1: Demographic and clinical characteristics COVID-19 adult patients according to their further need for ICU admission.

Characteristics of COVID-19 patients	No further need for ICU admission (n=25)	Further need for ICU admission (n=18)	P-Value
Male gender n (%)	13 (52.0)	10(55.5)	0.81
Median age (years) [range]	68 [28-92]	63 [44-78]	0.14
Comorbidities n (%)#	20 (80.0)	12 (66.6)	0.32*
Immunocompromised state n (%)##	6 (24.0)	4 (22.2)	0.89*
Median time delay between symptoms/admission in Covid-19 medicine ward (days) [range]	5 [0-10]	5 [0-11]	0.22
Median New Early Warning Score [range] <sup>(11)</sup>	3 [0-11]	7 [2-11]	0.0005
Median maximal lung involvement observed on CT- chest scan (%) [range]	25 [0-75]	50 [25-75]	0.001
Corticosteroids treatment n (%)	14 (56.0)	17 (94.4)	0.01*
Antibiotics n (%)	17 (68.0)	18 (100.0)	0.02*
Anticoagulation therapy n (%)	24 (96.0)	18 (100.0)	0.99*
Isocoagulant heparin therapy n (%)	10 (40.0)	7 (38.8)	0.99*
Optiflow nasal high flow therapy n (%)	0 (0.0)	6 (33.3)	0.007*
Mechanical ventilation n (%)	0 (0.0)	11 (61.1)	<0.0001*

In-hospital mortality n (%)	0 (0.0)	6 (33.3)	0.007*

\* Fischer's exact test; SARS-CoV-2: Severe Acute Respiratory Syndrome Coronavirus type 2; RT-PCR: Reverse Transcriptase Polymerase Chain Reaction: CT-chest Scan: Computerized Tomography chest scan; \* Comorbidities: Arterial Hypertension, Diabetes mellitus, Coronary artery disease, Heart Failure, Obesity, Chronic Kidney Failure, Chronic obstructive Pulmonary Disease or unbalanced Asthma; \*\* Immunocompromised state: cases of neoplasia (cancers), antineoplastic chemotherapy or previous corticosteroids or immunosuppressive treatments.

(11): Covino, M. et al. Predicting intensive care unit admission and death for COVID-19 patients in the emergency department using early warning scores. *Resuscitation* 156, 84–91 (2020).

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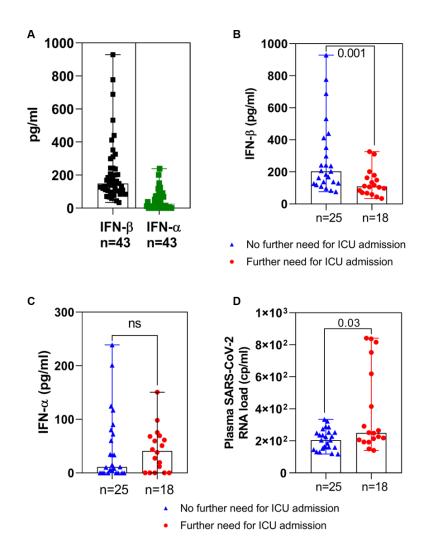
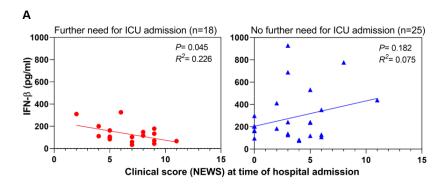


Figure 1: Early peripheral blood type I IFN levels, SARS-CoV-2 RNA load and clinical new early warning scores in COVID-19 adult patients requiring or not further intensive care unit admission

At the time of hospital admission type I interferon (IFN) and SARS-CoV-2 RNA load were measured in the peripheral blood of patients who further required (red circle, n=18) or not (blue triangle, n= 25) further Intensive Unit Care (ICU) admission; **A)** IFN- $\beta$  (black square) and IFN- $\alpha$  (green square) were measured using ELISA (*enzyme-linked immunosorbent assay*) in the peripheral blood of 43 COVID-19 patients. **B, C)** IFN- $\beta$  and IFN- $\alpha$  levels were measured using a referenced ELISA; **D)** Plasma SARS-CoV-2 RNA loads were assessed by a referenced digital PCR assay. Data indicated median values. *P* values were determined using Mann-Whitney test by two-way comparisons. ns: non-significant.



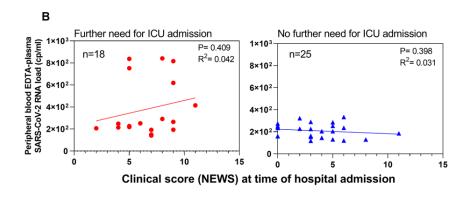


Figure 2: Association of early peripheral blood type I IFN levels, SARS-CoV-2 RNA load with clinical new early warning scores in COVID-19 adult patients requiring or not further intensive care unit admission

**A)** Correlation analyses between clinical scores (NEWS: New Early Warning Score) and IFN-β levels in the peripheral blood of Covid-19 adult patients at the time of hospital admission: left panel with red circles (n=18) represent patients who further required an ICU admission; right panel with blue triangles (n=25) represent patients who did not further require an ICU admission. Linear regression analyses were performed using Spearman tests, P=0.045; R<sup>2</sup>= 0.226 and P=0.182; R<sup>2</sup>= 0.075, respectively. **B)** Correlation analyses between clinical scores (NEWS: New Early Warning Score) and peripheral blood plasma SARS-CoV-2 RNA load in COVID-19 patients at the time of hospital admission: left panel with red circles (n=18) represent patients who further required an ICU admission and right panel with blue triangles (n=25) represent patients who did not further require an ICU admission. Linear regression analyses were performed using Spearman tests, P= 0.409; R<sup>2</sup>= 0.042 and P= 0.398; R<sup>2</sup>= 0.031, respectively.

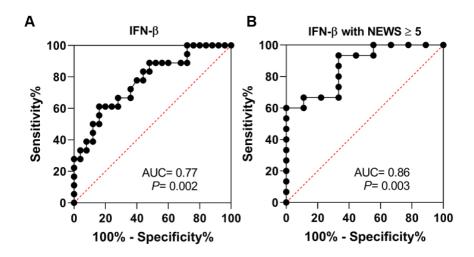


Figure 3: Receiver Operating Characteristic (ROC) curve of IFN- $\beta$  for relevance to predict coronavirus disease 2019 (COVID-19) severity.

**A)** ROC curve of blood IFN-β levels among study Covid-19 patients who further require or not an ICU admission; Area under the receiver operating characteristic (AUC)= 0.77, P= 0.002. **B)** ROC curve Blood IFN-β levels of Covid-19 patients who further required or not an ICU admission when the analysis was restricted to patients with a NEWS  $\geq$  5 at admission. Area under the receiver operating characteristic (AUC)= 0.86, P= 0.003.