

Early Plasma Interferon- β Levels as a Predictive Marker of COVID-19 Severe Clinical Events in Adult Patients Running title: Early IFN- β levels in COVID-19 patients

Fatma Berri, Yohan N'Guyen, Domitille Callon, Anne-laure Lebreil, Marie Glenet, Laetitia Heng, Bach-nga Pham, Firouze Bani-Sadr, Laurent

Andreoletti

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3	Full Title:
4	Early Plasma Interferon-β Levels as a Predictive Marker of COVID-
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9	Berri F ¹ *, N'Guyen Y ^{1,2} *, Callon D ^{1,3} , Lebreil AL ¹ , Glenet M ¹ , Heng L ¹ , Pham BN ⁴ ,
10	Bani-Sadr F ^{1,2} , Andreoletti L ^{1,5} and the Reims COVID-19 Study group
11	
12	¹ EA4684 Cardiovir research laboratory, University of Reims Champagne-Ardenne, Reims
13	France
14	² Internal Medicine, infectious diseases and Clinical Immunology, Robert Debré University
15	Hospital, Reims, France
16	³ CHU Reims, Hôpital Robert Debré, Pathology Department, Reims, France
17	⁴ CHU Reims, Hôpital Robert Debré, Immunology Department, Reims, France
18	⁵ CHU Reims, Hôpital Robert Debré, Virology Department, Reims, France
19	
20	* contributed equally to this work
21	
22	Corresponding author: Laurent Andreoletti, Laboratoire de Virologie médicale et moléculaire and EA-
23	4684 Cardiovir, Hôpital Robert Debré, Avenue du Général Koenig, 51092 REIMS Cedex, France. Tel:
24	(+33) 3 26 78 77 02; Fax: (+33) 3 26 78 41 34; E-mail: landreoletti@chu-reims.fr
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27 Abstract (187 words):

28 We assessed relationships between early peripheral blood type I Interferons levels, clinical new early warning scores (NEWS) and clinical outcomes in hospitalized 29 COVID-19 adult patients. Early IFN- β levels were lower among patients who further 30 required intensive care unit (ICU) admission than those measured in patients who did 31 32 not require an ICU admission during SARS-CoV-2 infection. IFN-B levels were 33 inversely correlated with NEWS only in the subgroup of patients who further required 34 ICU admission. To assess whether peripheral blood IFN-β levels could be a potential relevant biomarker to predict further need for ICU admission, we performed Receiver 35 Operating Characteristic (ROC) curve analyses that showed for all study patients an 36 37 area under ROC curve of 0.77 growing to 0.86 (P= 0.003) when the analysis was restricted to a subset of patients with NEWS \geq 5 at the time of hospital admission. 38 39 Overall, our findings indicated that early peripheral blood IFN- β levels might be a relevant predictive marker of further need for an intensive care unit admission in 40 hospitalized COVID-19 adult patients, specifically when clinical score (NEWS) was 41 42 graded as upper than 5 at the time of hospital admission.

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Keywords: COVID-19, SARS-CoV-2, type I Interferons, NEWS, intensive care unit
 45

46 **BACKGROUND**:

It is now well admitted that SARS-CoV-2 caused coronavirus disease-19 (COVID-19) 47 clinical stages evolve from early infection (stage I) towards a mild-to-moderate 48 pulmonary involvement without or with hypoxia (stage II), which can be followed by 49 an hyperinflammatory phase resulting in potential severe clinical events requiring an 50 intensive care unit (ICU) admission (clinical stage III)¹. Pathophysiology of SARS-51 CoV-2 infection is clearly characterized by an early viral replicative phase in 52 53 respiratory tract tissues associated with low viremia levels (clinical stage I) during 8 days after the first onset of symptoms, followed by an inflammatory phase occurring 54 from 9 to 12 days after the first onset of symptoms (clinical stages II and III). 55

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COVID-19 inflammatory phase is characterized by a dysregulated inflammatory host 57 58 response with an abnormal release of proinflammatory cytokines mediated by the activation of cell surface or intracellular toll like Receptors (TLRs)²⁻⁶. Remarkably, a 59 60 low interferon-Beta (IFN-β) production in the peripheral blood of severe (clinical stage 61 III) COVID-19 patient was evidenced suggesting that type I IFN low production levels could drive the development of clinical stages II and III during the course of the 62 SARS-CoV-2 infection⁷. Type I IFNs influence the development of innate and 63 64 adaptive immune responses limiting virus replication activities and preventing the development of severe clinical events during viral respiratory infections^{8,9}. However, 65 patterns of type I IFNs production during SARS-CoV-2 infection, were investigated 66 only in COVID-19 patients at clinical stages II and III (9-12 days after onset of first 67 symptoms)^{7,10}. In the present time, type I IFN expression pattern in the early phase of 68 69 COVID-19 (clinical stage I) and its potential relationship with further development of severe clinical events remains to be investigated. 70

In the present retrospective investigation, we assessed peripheral blood type I IFNs and SARS-CoV-2 RNA levels at early phase (1-8 days after the first symptom onset) of COVID-19 in adult patients who further required or not an ICU admission during infection. Among these two subsets of patients, we assessed potential relationships between early blood type I IFNs and SARS-CoV-2 RNA levels and clinical new early warning scores (NEWS) at the time of hospitalisation.

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80 CLINICAL SAMPLES and METHODS:

81 Study design:

From November 2020 to March 2021, peripheral blood EDTA-plasma samples taken
from adult patients had been stored at -80°C in Reims University Hospital (Grand-Est,
France) until processing for anti SARS-CoV-2 IgM or IgG antibodies detection assays.
This biobank was declared as a biological specimen's research collection and was
registered by the French Ministry of High Education and Research (number DC2020-4052).

From this research biobank samples (n=4704 sampled patients), we retrospectively 88 89 selected samples taken from adult patients (i) hospitalized in medicine wards for 90 COVID-19 (n=1116); (ii) with a time delay less than 9 days between sample collection and onset of COVID-19 symptoms (n= 183); (iii) with full clinical data 91 92 available from a registered clinical data bank (Reims University Hospital GDPR number RMR004-061120) elaborated for an internal purpose between March 1st 93 2020 and March 31rd 2021 and focusing on COVID-19 adult patients (n=143). 94 95 Patients who required palliative care or hospitalized directly in ICU were excluded from the study. 96

Finally, only 43 COVID-19 adult patients with available frozen EDTA-plasma samples
and with full registered clinical data were included in this retrospective investigation.

100 Demographic, clinical and medical data collection:

For each selected patient, the following data had been previously registered in a declared database (RMR004-061120) during their hospital stage: sex, age, comorbidities, immunocompromised state, date of symptoms onset, new early warning score (NEWS) graded at the time of hospital admission¹¹, computerized tomography chest scan results (CT-chest scan), drugs prescription and further need
 for intensive care unit admission (ICU) and in-hospital mortality. Demographic and
 clinical characteristics of study patients are depicted in Table 1.

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109 *Ethical approval:*

110 The Institutional Ethics Committee (IEC), University hospital of Reims (Grand-Est, France) waived the requirement of informed consent and approved all methods and 111 112 protocols. The study was conducted in accordance with the Declaration of Helsinki 113 Ethical Principles and Good Clinical Practices. Patients did not provide individual 114 consent because of the retrospective and non-interventional nature of this study, in accordance with French legislation. No patient raised an opposition to the use of their 115 116 recorded medical data or their samples collected at the time of hospitalization for 117 COVID-19. Data confidentiality was preserved during this internal study (Reims University Hospital GDPR register number RMR004-061120), and research 118 119 collection of biological specimens was registered in French Ministry of Higher 120 Education and Research as number DC-2020-4052.

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122 Peripheral blood type I interferons and SARS-CoV-2 RNA load

Using our research biological samples (registered research biobank), peripheral blood levels of Interferon-alpha (IFN-α), Beta (IFN-β) were determined on plasma samples using ELISA *(enzyme-linked immunosorbent assay)* (Biotechne, France) according to the manufacturer's instructions. In parallel, SARS-CoV-2 RNA load in EDTA plasma samples were quantified using droplet based digital PCR (dd-PCR) (MOBICYTE technical platform, Reims, France), as previously described⁷, with Bio-Rad SARS-CoV-2 dd-PCR kit (Bio-Rad, France). Viral RNA was extracted using QIAmp viral RNA mini kit (Qiagen) according to the manufacturer's guidelines. Viral
concentration results (copies/ml, cp/ml) were calculated considering the extraction
volumes of plasma (140 µl).

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134 Statistical analyses:

135 Quantitative variables expressed as median [range] were compared using the Mann 136 Whitney U-test. Qualitative variables expressed as percentages were compared 137 using Fischer's exact test or Pearson Chi-2 if applicable. When linear regression was performed, slopes were compared using Spearman test. When Receiver operating 138 139 characteristic (ROC) curve analysis was performed, Area under ROC Curve, Standard Error and P value were calculated using the trapezoid rule, the Hanley 140 141 Method and Z ratio respectively. A P value<0.05 was considered statistically 142 significant. Statistical analyses were performed using GraphPad Prism 8 (GraphPad) 143 and SAS version 9.4 (SAS Institute Inc).

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145 Data availability:

The datasets generated and/or analysed during the current study are not publicly available according to French law guidelines and registrations but are available from the corresponding author on reasonable request.

150 **RESULTS**

Early peripheral blood IFN-β, SARS-CoV-2 RNA levels and further need for intensive care unit admission in COVID-19 adult patients

Type I Interferons (IFNs) and SARS-CoV-2 RNA load were assessed in peripheral 153 154 blood samples taken from adult patients after a median time delay of 5 [1-8 days after the onset of symptoms] (Table1). Among the 43 selected patients, early 155 peripheral blood IFN-B were detected in all the study patients with values ranging 156 157 from 40 to 929 pg/ml, whereas peripheral blood IFN-α were detectable in only 86% of the study patients with values ranging from non-detectable to 326 pg/ml (Figure 1A). 158 159 Because low type I IFN production levels could drive the development of COVID-19 160 induced inflammatory phase, we segregated our patient group into two subsets according for their further need for an ICU admission during infection. Remarkably, 161 162 levels of IFN-β appeared to be significantly lower among the subset of patients who 163 further required ICU admission than the subset of patients who did not further require 164 ICU admission (P=0.001) (Figure 1B). For IFN- α levels, no significant differences 165 between patient subsets requiring or not ICU admission was evidenced at the time of hospitalization (Figure 1C). In addition, the plasma SARS-CoV-2 RNA load appeared 166 to be significantly higher in patients who further required ICU admission than those 167 168 measured in samples of patients who did not further required ICU admission (P= 169 0.03) (Figure 1D).

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Associations between early peripheral blood IFN-β levels, SARS-CoV-2 RNA loads and clinical new early warning scores

Because clinical new early warning scores (NEWS) had been previously associated
with further ICU admission events in COVID-19 adult patients¹¹, we first sought for a

potential association between NEWS and IFN-β concentrations and then between 175 NEWS and SARS-CoV-2 RNA load according to a further need for ICU admission. 176 177 Among the subset of patients who further required an ICU admission, a significant negative correlation between NEWS and IFN- β concentrations (*P*= 0.045; R²= 0.226) 178 179 was observed (Figure 2A, left panel). By contrast a non-significant positive 180 correlation was observed between NEWS and IFN-ß concentrations among the 181 subset of patients who did not require further ICU admission (P= 0.182; R²= 0.075) 182 (Figure 2A, right panel). No significant correlation was evidenced between NEWS and SARS-CoV-2 RNA load among the two subsets of study patients (Figure 2B). 183 184 Remarkably, when the analysis was restricted to a subset of patients with a NEWS ≥ 185 5 at hospital admission we observed a significant negative correlation between blood IFN-ß level and plasma SARS-CoV-2 RNA load in the subset of COVID-19 patient 186 187 who did not further require an ICU admission. Such a correlation was not observed 188 among patients who further require ICU admission (supplementary Figure 1).

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Early Blood IFN-β concentrations accuracy to predict the need for a further ICU admission

192 To assess whether peripheral blood IFN-β levels could be a relevant biomarker to 193 predict further need for ICU admission, we performed Receiver Operating 194 Characteristic (ROC) curve analyses (Figure 3). Including all study patients, the 195 sensitivity and specificity were respectively of 88% and 50% with an area under the 196 receiver operating characteristic curve (AUC) of 0.77 (Figure 3A). Remarkably, the 197 area under the ROC curve of blood IFN-β level grew to 0.86 when the analysis was 198 restricted to a subset of patients with a NEWS \geq 5 at hospital admission indicating a 199 relevant predictive value of early IFN-β levels among the most clinically severe study patients. Among the subset of patients with NEWS ≥ 5 at hospital admission, peripheral blood IFN-β levels below 107.6 pg/ml were associated to a further ICU requirement with a specificity of 100% and a sensitivity of 60% (Figure 3B).

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206 **DISCUSSION**

Investigation of peripheral blood type I IFN production patterns in SARS-CoV-2 infected adult patients is of major interest for understanding COVID-19 pathophysiology and clinical courses of infection^{7,10}. Herein, we assessed relationships between early peripheral blood Interferon-Beta (IFN- β) levels, clinical new early warning scores (NEWS) and clinical outcomes in SARS-CoV-2 infected adult patients hospitalized from 1 to 8 days after the first symptom onset (clinical stage I).

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215 Among the 43 selected patients, anti-SARS-CoV-2 total antibodies were not detected 216 and interleukin 6 (IL6) and tumour necrosis factor-alpha (TNF- α) were not measured 217 at significant levels in peripheral blood (data not shown). These results were 218 associated with detectable SARS-CoV-2 RNA load confirming that study patients 219 were at the viral phase of COVID-19 (Figure 1D). Interestingly, early peripheral blood 220 IFN- β were detected in all the study patients, whereas IFN- α were detectable only in 221 86% of the study patients (Figure 1A). Because low type I IFN production levels could drive the development of COVID-19 induced inflammatory phase (clinical 222 stages II and III), we segregated our patient group into two subsets according for 223 224 their further need for an ICU admission. Interestingly, our findings indicated for the 225 first time that patients who further required ICU admission demonstrated significant lower peripheral blood IFN-β levels and higher SARS-CoV-2 RNA load than those 226 227 observed in patients who did not require ICU admission (Figure 1). Low early IFN-β 228 concentration levels in patient could be related to inter-individual sequences variability in human genes implicated into the regulation of type I IFN innate 229 immunity¹², but also to the translation of SARS-CoV-2 nonstructural proteins 230

inhibiting the IFN- β promoter activation¹³ or to preexisting peripheral blood autoantibodies directed against human type I IFNs¹⁴.

233 Among the subset of COVID-19 patients who further required ICU admission, a negative correlation between peripheral blood IFN-B concentrations and clinical 234 235 NEWS was evidenced, whereas no significant association was observed between clinical scores and SARS-CoV-2 RNA levels (Figure 2). These findings were 236 237 completed by ROC analyses and results indicated that early peripheral blood IFN- β , 238 but not SARS-CoV-2 RNA load, might be a relevant biomarker of a further ICU 239 admission, specifically among patients with a NEWS \geq 5 at the time of hospital 240 admission (Figure 3B). Overall, these findings supported the concept that an early 241 IFN-β treatment in COVID-19 adult patients hospitalized at phase I of SARS-CoV-2 242 infection might be of therapeutic benefit improving the clinical outcomes. However, 243 these findings remain to be fully investigated in further prospective large multicenter 244 clinical investigations.

245 **Conclusions**

246 Even if our study is a limited retrospective monocentric investigation conducted before large anti-SARS-CoV-2 vaccination campaigns in general population and 247 before the emergence of new Delta or Omicron viral variants, our original findings 248 249 revealed a significant association between early low peripheral blood IFN-β levels 250 and high clinical scores (NEWS) during SARS-CoV-2 clinical phase I. Overall, our 251 findings indicated that early peripheral blood IFN-ß levels might be a relevant predictive marker of further need for an intensive care unit admission in COVID-19 252 253 adult patients, specifically when clinical score (NEWS) was graded as upper than 5 at 254 hospital admission.

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295

296 Author's contribution

L.A, Y.N'G and F.B designed the study. L.A, F.B and Y.N'G wrote the manuscript.
L.A, BN. P and Y.N'G established the research biobank, the clinical data bank and
supervised the data collection. F.B and Y.N'G analyzed the data and designed the
table and the figures. D.C helped for data collection. F.B, Y.N'G, L.A, D.C, A-L.L,
M.G, L.H, F.B-S and BN. P provided valuable input to interpretation of the data and
critically reviewed the manuscript. All these authors approved the final version of the

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- 308

309 **Conflict of interest**

- 310 The authors report no conflict of interest.
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312 Data availability:

313 The datasets generated and/or analyzed during the current study are not publicly

314 available according to French law guidelines and registrations but are available from

315 the corresponding author on reasonable request.

316

317 Reims Covid-19 Study group. Collaborators included the following: Dr Ezekiel 318 Bankole, Dr Loïs Bolko, Dr Michel Bonnivard, Dr Nelly Bourse, Dr Aurélie Brunet, Dr 319 Sandy Carazo Mendez, Dr Clement Chopin, Dr Joël Cousson, Dr Sophie Demotier, 320 Pr Gaetan Deslee, Dr Vincent Dupont, Dr Thierry Floch, Dr Julien Gautier, Dr Guillaume Giordano Orsini, Dr Sophie Giordano Orsini Dr Antoine Goury, Dr Marie 321 Laure Gross, Dr Valentin Houannou, Dr Claire Lepouse, Dr Gauthier Lejeune, Dr 322 323 Natacha Noël, Pr Jeanne Marie Perotin, Dr Juliette Romaru, Dr Jerome Tassain. Pr. 324 Bruno Mourvillier.

TABLE AND FIGURES

326	Table 1: Demographic and clinical	characteristics COVID-	19 adult patients accordi	ing to their further need for I	CU admission.
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Characteristics of COVID-19 patients	No further need for ICU admission (n=25)	Further need for ICU admission (n=18)	P-Value
Male gender n (%)	13 (52.0)	10(55.5)	0.81
Median age (years) [range]	68 [28-92]	63 [44-78]	0.14
Comorbidities n (%) [#]	20 (80.0)	12 (66.6)	0.32*
Immunocompromised state n (%)##	6 (24.0)	4 (22.2)	0.89*
Median time delay between symptoms/admission in Covid-19 medicine ward (days) [range]	5 [0-10]	5 [0-11]	0.22
Median New Early Warning Score [range] ⁽¹¹⁾	3 [0-11]	7 [2-11]	0.0005
Median maximal lung involvement observed on CT- chest scan (%) [range]	25 [0-75]	50 [25-75]	0.001
Corticosteroids treatment n (%)	14 (56.0)	17 (94.4)	0.01*
Antibiotics n (%)	17 (68.0)	18 (100.0)	0.02*
Anticoagulation therapy n (%)	24 (96.0)	18 (100.0)	0.99*
Isocoagulant heparin therapy n (%)	10 (40.0)	7 (38.8)	0.99*
Optiflow nasal high flow therapy n (%)	0 (0.0)	6 (33.3)	0.007*
Mechanical ventilation n (%)	0 (0.0)	11 (61.1)	<0.0001*

In-hospital mortality n (%) 0 (0.0) 6 (33.3) 0.00)07*
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* Fischer's exact test; SARS-CoV-2: Severe Acute Respiratory Syndrome Coronavirus type 2; RT-PCR: Reverse Transcriptase
 Polymerase Chain Reaction: CT-chest Scan: Computerized Tomography chest scan; [#] Comorbidities: Arterial Hypertension,
 Diabetes mellitus, Coronary artery disease, Heart Failure, Obesity, Chronic Kidney Failure, Chronic obstructive Pulmonary Disease
 or unbalanced Asthma; ^{##} Immunocompromised state: cases of neoplasia (cancers), antineoplastic chemotherapy or previous
 corticosteroids or immunosuppressive treatments.

⁽¹¹⁾: Covino, M. *et al.* Predicting intensive care unit admission and death for COVID-19 patients in the emergency department using
 early warning scores. *Resuscitation* 156, 84–91 (2020).



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Figure 1: Early peripheral blood type I IFN levels, SARS-CoV-2 RNA load and clinical new early warning scores in COVID-19 adult patients requiring or not further intensive care unit admission

339 At the time of hospital admission type I interferon (IFN) and SARS-CoV-2 RNA load 340 were measured in the peripheral blood of patients who further required (red circle, 341 n=18) or not (blue triangle, n= 25) further Intensive Unit Care (ICU) admission; A) 342 IFN- β (black square) and IFN- α (green square) were measured using ELISA (enzyme-linked immunosorbent assay) in the peripheral blood of 43 COVID-19 343 patients. **B**, **C**) IFN- β and IFN- α levels were measured using a referenced ELISA; **D**) 344 345 Plasma SARS-CoV-2 RNA loads were assessed by a referenced digital PCR assay. Data indicated median values. P values were determined using Mann-Whitney test 346 by two-way comparisons. ns: non-significant. 347

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Figure 2: Association of early peripheral blood type I IFN levels, SARS-CoV-2 RNA load with clinical new early warning scores in COVID-19 adult patients requiring or not further intensive care unit admission

355 **A)** Correlation analyses between clinical scores (NEWS: New Early Warning Score) 356 and IFN-B levels in the peripheral blood of Covid-19 adult patients at the time of 357 hospital admission: left panel with red circles (n=18) represent patients who further 358 required an ICU admission; right panel with blue triangles (n=25) represent patients 359 who did not further require an ICU admission. Linear regression analyses were performed using Spearman tests, P=0.045; $R^2=0.226$ and P=0.182; $R^2=0.075$, 360 respectively. B) Correlation analyses between clinical scores (NEWS: New Early 361 362 Warning Score) and peripheral blood plasma SARS-CoV-2 RNA load in COVID-19 363 patients at the time of hospital admission: left panel with red circles (n=18) represent 364 patients who further required an ICU admission and right panel with blue triangles 365 (n=25) represent patients who did not further require an ICU admission. Linear regression analyses were performed using Spearman tests, P=0.409; $R^2=0.042$ and 366 $P=0.398; R^2=0.031$, respectively. 367

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Figure 3: Receiver Operating Characteristic (ROC) curve of IFN-β for relevance to predict coronavirus disease 2019 (COVID-19) severity.

A) ROC curve of blood IFN-β levels among study Covid-19 patients who further require or not an ICU admission; Area under the receiver operating characteristic (AUC)= 0.77, P= 0.002. B) ROC curve Blood IFN-β levels of Covid-19 patients who further required or not an ICU admission when the analysis was restricted to patients with a NEWS ≥ 5 at admission. Area under the receiver operating characteristic (AUC)= 0.86, P= 0.003.