

Class Ib Ribonucleotide Reductases: Activation of a Peroxido-Mn II Mn III to Generate a Reactive Oxo-Mn III Mn IV Oxidant

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Class Ib Ribonucleotide Reductases: Activation of a Peroxido-Mn^{II}Mn^{III} to Generate a Reactive Oxo-Mn^{III}Mn^{IV} Oxidant

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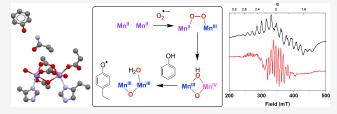
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ABSTRACT: In the postulated catalytic cycle of class Ib Mn₂ ribonucleotide reductases (RNRs), a Mn^{II}₂ core is suggested to react with superoxide (O2 -) to generate peroxido-MnIIMnIII and oxo-Mn^{III}Mn^{IV} entities prior to proton-coupled electron transfer (PCET) oxidation of tyrosine. There is limited experimental support for this mechanism. We demonstrate that $[Mn^{II}]_2(BPMP)$ - $(OAc)_2$ (ClO₄) (1, HBPMP = 2,6-bis (bis(2 pyridylmethyl)amino)methyl]-4-methylphenol) was converted to peroxido-

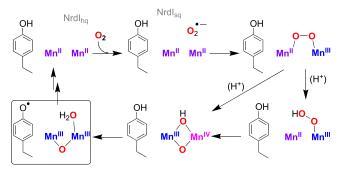


 $Mn^{II}Mn^{III}$ (2) in the presence of superoxide anion that converted to $(\mu$ -O) $(\mu$ -OH) $Mn^{III}Mn^{IV}$ (3) via the addition of an H⁺donor (p-TsOH) or $(\bar{\mu}\text{-O})_2\text{Mn}^{\text{III}}\text{Mn}^{\text{IV}}$ (4) upon warming to room temperature. The physical properties of 3 and 4 were probed using UV-vis, EPR, X-ray absorption, and IR spectroscopies and mass spectrometry. Compounds 3 and 4 were capable of phenol oxidation to yield a phenoxyl radical via a concerted PCET oxidation, supporting the proposed mechanism of tyrosyl radical cofactor generation in RNRs. The synthetic models demonstrate that the postulated O₂/Mn₂/tyrosine activation mechanism in class Ib Mn₂ RNRs is plausible and provides spectral insights into intermediates currently elusive in the native enzyme.

INTRODUCTION

Ribonucleotide reductases (RNRs) are responsible for catalyzing the conversion of ribonucleotides to the corresponding deoxyribonucleotide, providing precursors for deoxyribonucleic acid (DNA) synthesis and repair in all organisms. 1-4 RNRs are an important target for anticancer drugs, and therefore understanding the enzyme's mechanism is important. Each of the three classes of RNRs (I, II, and III) initiates ribonucleotide reduction through the generation of a thiyl radical, which initiates the reductive dehydration of nucleotides. The thiyl radical generation method differs across the three classes in the nature of the metallo-cofactor that facilitates thiyl radical formation. 1-4 The class I metallocofactors contain a dinuclear transition-metal cluster that activates dioxygen to initiate radical production. Classes II and III employ adenosylcobalamin and a 4Fe-4S cluster, respectively, for thiyl radical generation. Class I is further divided into subclasses (a, b, c, and d) depending on the identity of the metallo-cofactor: ¹⁻⁴ an Fe₂ cluster (class Ia), a Mn₂ cluster (Ib, Id), or an FeMn cluster (Ic).

The mechanism of activation of O₂ in class Ia RNRs is wellestablished: 6 a Fe $^{II}_{2}$ core reacts with O_{2} to yield a μ -peroxido-Fe^{III}Fe^{III} entity, followed by electron donation and O-O bond scission to yield a bis- μ -oxo-Fe^{III}Fe^{IV} oxidant that activates a tyrosine group to yield a tyrosyl radical and Fe^{III}₂. The tyrosyl radical is responsible for initiating the thiyl radical formation. Class Ib RNRs,⁷⁻⁹ in contrast, have a considerably less wellunderstood mode of O₂ activation (Scheme 1). The Mn₂ Scheme 1. Proposed Catalytic Cycle for Tyrosyl Radical Cofactor Generation in Class Ib RNRs; NrdI_{hq} = Flavodoxin-Like Protein Hydroquinone Cofactor; NrdI_{sq} = Semiquinone Cofactor; Presence of H in the Hydroxide Bridge Has Not Yet Been Experimentally Verified



cluster does not react with O_2 : superoxide anion (O_2^{-}) is postulated to be generated upon the activation of O2 by a nearby flavodoxin protein (NrdI-hydroquinone (hq) to semiquinone (sq) conversion), followed by the reaction of

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Scheme 2. Reaction of 1 with KO_2 to Yield 2 and the Activation of 2 Using Acids (Top, 2 to 3 Conversion) or via Thermal Decay (Bottom, 2 to 4 Conversion); L-O = BPMP

O2.- with the MnII2 core. 10,11 This is proposed to yield a mixed-valent peroxido-Mn^{II}Mn^{III} adduct, which subsequently undergoes O-O bond cleavage to generate bis-µ-oxo-Mn^{III}Mn^{IV} species. This Mn^{III}Mn^{IV} unit is postulated to be the active oxidant responsible for the oxidation of tyrosine to yield a tyrosyl radical, which then initiates thiyl radical formation. While there is strong evidence of O2 - generation during the first step of the process, there is little support for the identity of the products of the reaction between the Mn^{II}₂ core and O₂...6 The formation of a transient Mn^{III}Mn^{IV} entity is supported by a characteristic 16-line electron paramagnetic resonance (EPR) signal, 11 although little more is known of this species. Furthermore, the role of a proton donor in O-O bond cleavage is poorly understood, and the protonation state of the bis-µ-oxo-MnIIIMnIV core is unknown. Another area of uncertainty is the mechanism of the tyrosyl radical generation by the putative bis-μ-oxo-Mn^{III}Mn^{IV} species. ¹² This protoncoupled electron transfer (PCET) reaction could proceed in a single concerted step or a stepwise process, whereby electron transfer (ET) is followed by proton transfer (PT) or vice versa.¹³

We explored the properties of synthetic Mn2 model complexes in order to understand the postulated mechanism of class Ib RNRs and investigate the structural and electronic properties of the intermediates (Scheme 1). We found that $[(Mn^{II}_{2})(HPTB)(OAc)](ClO_{4})_{2} (HPTB = N,N,N',N'$ tetrakis(2-(benzimidazolyl))-2-hydroxy-1,3-diaminopropane; OAc = acetate) and $[Mn^{II}_{2}(BPMP)(OAc)_{2}](ClO_{4})$, (1, Scheme 2, HBPMP = 2,6-bis[(bis(2 pyridylmethyl)amino)methyl]-4-methylphenol) reacted with O2 (KO2 solubilized with 18-crown-6) to form peroxido-Mn^{II}Mn^{III} species. 14-16 These studies verified the postulated reactivity of a Mn^{II}₂ core in class Ib RNRs with O_2 . and were the first examples of mixed-valent peroxido-Mn^{II}Mn^{III} complexes. As these peroxide species were relatively inert and unable to activate the weak O-H bonds in phenols, we postulated that such species were not directly responsible for the oxidation of tyrosine in class Ib RNRs. Herein, we describe the activation of a peroxido-Mn^{II}Mn^{III} complex to yield Mn^{III}Mn^{IV} adducts that were characterized by spectroscopic and spectrometric analyses and for which their PCET phenol oxidation has been explored.

RESULTS AND DISCUSSION

Compound 1 was prepared according to previous reports. $^{14-16}$ [Mn^{II}Mn^{III}(O₂)(BPMP)]²⁺ (2) was prepared using a modified procedure (Scheme 1): 1 (1.5 mM) was reacted with KO₂ (1 equiv) in 1:9 CH₃CN/tetrahydrofuran (THF) at -90 °C to yield 2. However, the previously used 18-crown-6 (used to

solubilize KO_2) was replaced with 2,2,2-cryptand (4,7,13,16,21,24-hexaoxa-1,10-diazabicyclo[8.8.8]hexacosane), allowing a notable increase in yield of 2 (Figure S1; see Supporting Information for details). The substitution of 2,2,2-cryptand for 18-crown-6 did not have any effect on the spectroscopic properties of 2, as evidenced by the same electronic absorption features (λ = 440, 590 nm, Figures 1 and

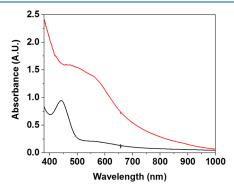


Figure 1. Electronic absorption spectra of **2** (black) and **3** (red trace, formed from the reaction between **2** and *p*-TsOH (2 equiv)).

S1). In the same vein, the EPR spectrum of **2** prepared with 2,2,2-cryptand displayed the same 22-line EPR signal centered at g = 2, observed when 18-crown-6 was used (Figure S2).

Acid-Mediated Activation of 2. Treatment of 2 in 1:9 CH₃CN/THF (1.5 mM at -90 °C) with para-toluenesulfonic acid (p-TsOH) resulted in the disappearance of the features assigned to 2, followed by the immediate formation of a broad electronic absorption feature assigned to a new species (designated as 3, Figure 1). The maximum yield of 3 was achieved upon the addition of 2 equiv of p-TsOH according to electronic absorption and EPR spectroscopies (Figures S3 and S4). It should be noted that treatment of 3 with more than 2 equiv of TsOH or indeed with stronger acids (HCl, HClO₄, and HBF₄) resulted in spectral features that can be assigned to a [Mn^{II}Mn^{III}(BPMP)]⁴⁺ adduct, comparable to that reported by Hendrikson and co-workers.¹⁷ We surmise that a large excess of acid or stronger acids caused the release of H2O2 in those instances. Compound 3 displayed a broad band with λ_{max} \sim 550 and 590 nm that trailed into the near-infrared (near-IR) region. Complex 3 was found to be metastable, with a half-life $t_{1/2}$ = 1200 s at -90 °C. The electronic absorption features of 3 were red-shifted and more intense in comparison to those of the peroxido- $Mn^{II}Mn^{III}$ complex 2. $Bis-(\mu-O)Mn^{III}Mn^{IV}$ complexes have been found to display similar features. For such complexes, absorption bands typically fall between 520

Table 1. Electronic Absorption and Electron Paramagnetic Resonance Properties of Mn^{III}Mn^{IV} Complexes

complex	λ (nm)	g-value	# of EPR lines (total spectral width [mT])	ref
3	550, 590	2	16 (125)	this work
4	broad 400-800	2	16 (110)	this work
Ib RNR	broad 300-350	2	16 (130)	11
$[\mathrm{Mn^{III}Mn^{IV}}(\mu\text{-O}_2)(\kappa^4\text{-N4py})_2]^{3+}$	565, 667	2	16 (140)	25
$[Mn^{III}Mn^{IV}(\mu-O)_2(terpy)_2(OH_2)_2]^{3+}$	553, 654			23
$[Mn^{III}Mn^{IV}(\mu-O)_2 (bisimMe_2en)_2]^{3+}$	419, 536, 644	2	16 (135)	21
$[{ m Mn^{III}Mn^{IV}}~(\mu ext{-O})_2({ m bpy})_4]^{3+}$	525, 555, 684	2	16 (—)	18
$[Mn^{III}Mn^{IV}(\mu-O)_2(14-aneN_4)_2]^{3+}$	556, 646	2	16 (—)	19
$[Mn^{III}Mn^{IV}(\mu-O)_2(\mu-O_2CMe)(L_2^M)]^{2+}$	259, 400, 440, 548, 640, 800	2	16 (135)	20
$[Mn^{III}Mn^{IV}(2\text{-OH})(3,5\text{-Cl}_2Sal)pn)_2]^+$	400	2	16 (160)	22
$[\mathrm{Mn^{III}Mn^{IV}}(\mu ext{-O})_2(\mathrm{tmpa})_2]^{3+}$	443, 561, 658	2	16 (150)	28
$[\mathrm{Mn^{III}Mn^{IV}}(\mu ext{-O})(\mathrm{L^{H}}_{2})]^{3+}$	344, 408, 570	2	18 (130)	26

and 570 nm (assigned to d-to-d transitions) and 590–700 nm (assigned to ligand-to-metal charge transfer (LMCT), e.g., oxo-to-Mn, Table 1). ^{18–25} For example, [Mn^{III}Mn^{IV}(μ -O₂)(κ ⁴-N4py)₂]³⁺ displayed an intense feature at λ = 565 nm and a less intense one at λ = 667 nm. ²⁵ The μ -O-Mn^{III}Mn^{IV} species, supported by a phenolate ligand, exhibited a broad absorption band at λ _{max} = 570 nm that was attributed to phenolate-to-Mn charge transfer (with oxo-to-Mn at higher energy). ^{26,27} Therefore, we concluded that 3 plausibly represents a μ -oxo-Mn₂ complex based on its electronic absorption features which are consistent with oxo- and phenolate-to-Mn^{IV} transitions.

EPR spectra were recorded on frozen CH₃CN/THF solutions of 3 (Figures 2 and S5). They displayed a 16-line

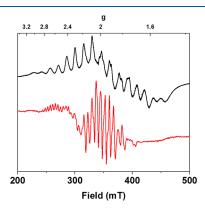


Figure 2. Perpendicular-mode X-band EPR spectra of **2** (black trace) measured at 2 K and of **3** (red trace) measured at 30 K (9.65 GHz, 0.2 mW microwave power).

signal centered at g = 2. The average hyperfine coupling constant (A_{av}) was measured as $A_{av} = 7.4$ mT. Both the number of lines and the $A_{\rm av}$ value were consistent with the synthetic Mn^{III}Mn^{IV} species and, importantly, Mn^{III}Mn^{IV} identified in the studies on class Ib RNRs (Table 1).29-31 This is in contrast to the larger value of $A_{av} = 12.4 \text{ mT}$ measured for 2. Furthermore, the spectral width for the 16-line g = 2 signal of 3 was approximately 125 mT, consistent with that measured for the previously described MnIIIMnIV complexes.^{32,33} In contrast, the spectral width for 2 spans 250 mT. Dismukes and co-workers postulated that the narrow spectral width allowed Mn^{III}Mn^{IV} to be distinguished from Mn^{II}Mn^{III} systems.³² This disparity in spectral width comes from the spin-coupling coefficients that multiply the hyperfine coupling constants (hfc's) that are larger for the Mn^{II}Mn^{III} complexes than for the Mn^{III}Mn^{IV} ones.³⁴ Indeed, the

individual ion hfc's for Mn^{n+} vary little with n for ligands with similar covalency. Moreover, the 16-line g=2 signal showed no temperature dependence, displaying well-resolved signals up to 77 K (Figure S14). 20,35,36 In contrast, the EPR spectra of $Mn^{II}Mn^{III}$ complexes generally need to be recorded below 10 K to be well-resolved. Therefore, we assigned the 16-line g=2 signal of 3 to a $Mn^{III}Mn^{IV}$ complex. The yield of the 16-line species was determined by EPR integration to be \sim 50 \pm 20% (using a Cu^{II} salt as a reference).

A second low-intensity signal was present in the EPR spectrum of 3: an 11-line feature centered at g = 2.6 with an $A_{\rm av}$ value of 4.4 mT, corresponding to approximately half of that determined for 3 (7.8 mT). Such a hyperfine pattern is consistent with a complex containing two equivalent Mn (MnI = 5/2) centers. Interestingly, the previously characterized [M n ^{I I} ₂ (T M E D A) ₂ (O C C H ₃) ₄ (H ₂ O)] and $[Mn^{II}_{2}(Me_{3}TACN)_{2}(\mu\text{-OCCH}_{3})_{3}]BPh_{4}$ complexes also displayed an 11-line signal centered at g = 2.34 with $A_{av} = 4.5$ mT,³⁷ and g = 1.96 with $A_{av} = 4.6$ mT,³⁸ respectively. Therefore, this 11-line EPR feature can be assigned to a Mn_{2}^{II} species that differs from 1. The EPR properties of 1 have been previously investigated by the Hendrickson group and our group, and no such signal has been observed (by us or Hendrickson).¹⁷ Variable-temperature EPR experiments recorded on peroxide complex 2, however, revealed the presence of the 11-line EPR signal but in negligible amounts (Figure S6). Therefore, we postulate that upon the addition of superoxide to 1 to yield 2, a residual Mn_{2}^{II} formed in low yield, which remains in solution upon the acid-mediated conversion of 2 to 3.

Electrospray ionization mass spectrometry (ESI-MS) analysis of a just-thawed methanol solution of 3 revealed a peak at m/Z = 335.5837, consistent with the mass of the dication $[(Mn)_2(O)_2(BPMP)]^{2+}$ (Figure S7; expected mass m/Z =335.5682). Other peaks in the mass spectra can be assigned to the free ligand, cryptand, and starting material 1. The m/Z =335 peak was not observed in the ESI-MS spectrum of the peroxide precursor 2. In the ESI-MS spectrum of 2, the m/Z =770.29 monocation corresponding to $[(Mn)_2(O)_2(BPMP)]$ -(ClO₄)]⁺ was observed, which was greatly diminished in the spectrum of 3 (Figure S8). This suggested that the peroxide O-O bond in 2 was cleaved and yielded two oxo-ligands. Preparation of 2 using K¹⁸O₂, followed by activation with p-TsOH to yield the isotopically labeled form of complex 3, caused the mass peak to shift by two mass units (m/Z)337.5788), consistent with the incorporation of two ¹⁸O atoms

in this dicationic ion, consistent with 3 containing two O atoms in a bis- μ -oxo core derived from KO₂.

Mn K-edge X-ray absorption near-edge spectroscopy (XANES) was performed on 2 and 3 in order to gain an understanding of the oxidation states of the metal centers and their local geometry (Figure S9). The XANES spectra of the peroxido-Mn^{II}Mn^{III} 2 displayed a rising edge with the first inflection point calculated to be at 6548.9 eV, consistent with the previously recorded data on the same complex. 14-16 Upon conversion from 2 to 3, the edge energy shifted minimally (0.5 eV). Upon irradiation of 3, we noted a color change of the samples (despite best efforts to irradiate fresh spots with every scan). An increase in the oxidation number of Mn by one unit is typically associated with an edge energy shift of 2-4 eV. 39,40 We attributed the lack of a shift to the following: photoreduction of 3 by incident X-rays during the XAS measurement and a less-than-optimal yield of Mn^{III}Mn^{IV} species 3, only ~50 ± 20% by EPR integration. We concluded that XAS would not provide fruitful insights into the electronic and geometric structures of 3, and thus, we did not pursue further EXAFS measurements.

Thermal Activation of 2. A solution of **2** in CH₃CN/THF (1:9, 1.5 mM) was warmed from -90 to +20 °C, a process that lasted 900 s (Scheme 2). This resulted in the disappearance of the electronic absorption features assigned to **2**, to yield a broad indistinct absorption that trailed from the UV into the near-IR region, associated with the formation of a new species (defined as **4**, Figure 3). Compound **4** displayed a half-life of $t_{1/2} = 2300$ s at 20 °C.

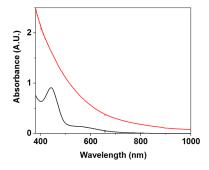


Figure 3. Electronic absorption features for the conversion of 2 (black trace) to 4 (red trace) through thermal activation (-90 to +20 °C) in 1:9 CH₂CN/THF.

EPR analysis of 4 measured at 2 K revealed two new signals, a 16-line signal centered at g = 2 with $A_{av} = 7.9$ mT and a 11line signal centered at g = 2.6 with $A_{av} = 4.4$ mT (Figures 4 and S10). The spectrum obtained was remarkably similar to that measured for 3. For the same reasons outlined above, we assigned the 16-line g = 2.0 signal observed for 4 to a Mn^{III}Mn^{IV} species. The quantification of the 16-line EPR spectrum indicates a conversion from 3 to 4 in \sim 60% \pm 20% yield. As in the case of 3, we postulated that the g = 2.6 signal originates from a Mn^{II}₂ species. The EPR spectra of 3 and 4 display similar signals at g = 2 regarding their line-shape and the number of lines. The g = 2 signals of both 3 and 4 display similar spectral widths of 125 and 110 mT, respectively. Furthermore, both features were resolved at temperatures higher than 10 K (Figure S11). Based on the similarities of the EPR characteristics of complexes 3 and 4, we postulated that they are both $Mn^{III}Mn^{IV}$ species, with a bis- μ -oxo $Mn^{III}Mn^{IV}$ core for 4 and a μ -oxo- μ -hydroxo-Mn^{III}Mn^{IV} core for 3. We

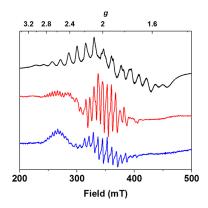


Figure 4. EPR spectra of 2 (black trace) recorded at 2 K, 3 (red trace) at 30 K, and 4 (blue trace) at 7 K (9.65 GHz, 0.2 mW microwave power).

ascribe a difference in the protonation states in 3 and 4 based on the analysis of their absorption spectra and on the reactivity studies described below.

ESI-MS of a just-thawed solution of 4 revealed a peak at m/Z=335.58 that corresponded to the dication $[\mathrm{Mn_2(O)_2(BPMP)}]^{2+}$ (Figure S12). Preparation of 4 using $\mathrm{K^{18}O_2}$ caused this peak to shift by 2 mass units, (m/Z=337.5788) consistent with two $^{18}\mathrm{O}$ oxygen atoms in the oxo moiety in the dicationic molecule. As with 3, this is indicative of the formation of a $\mathrm{Mn^{III}Mn^{IV}}$ species with a bis(μ -oxo) core structure. Both complexes 3 and 4 displayed the same dication peaks in their mass spectra, indicating that both complexes form similar species during the activation of 2.

The broad, tailing UV-vis spectrum observed for 4 has previously been observed for Mn^{III}Mn^{IV} complexes (Table 1), which generally display absorbance features in the visible region, typically derived from the ligand-to-metal charge transfer or d-d transitions. For example, the UV-vis spectrum of a putative $bis(\mu-oxo)Mn^{III}Mn^{IV}$ species in the recently discovered class Id RNRs displayed a similar tailing absorption into the UV region.⁴¹ Likewise, [bis(μ -oxo)-Mn^{III}Mn^{IV}(2-OH(3,5-Cl₂Sal)pn)₂]ClO₄ showed a broad spectrum that tailed into the UV region. 22 Therefore, 4 is plausibly a bis $(\mu$ -oxo)Mn^{III}Mn^{IV} species. Overlay of the electronic absorption spectra of 3 and 4 revealed some similarities (Figure S13). Both 3 and 4 displayed broad absorptions that trail into the near-IR region of the spectra. However, the important difference between the two are the features at λ_{max} = 550, 590 nm, postulated to be oxo- or phenolate-to- $Mn^{\rm IV}$ charge transfer, displayed by 3, that is not present in the spectrum of 4. In studies exploring the protonation of the bis(μ -oxo) cores of Mn^{IV}₂ complexes, conducted by Pecoraro, it was discovered that the addition of H+ resulted in the shifting of the absorption features attributed to O-to-Mn^{IV} charge transfer.²⁷ We surmise that the loss of features in 4 with respect to 3 could be attributed to a similar shifting of chargetransfer bands. This supports the conclusion that a protonation state difference exists in 3 and 4, suggesting that perhaps 3 contains a protonated core, while 4 contains an unprotonated $bis(\mu$ -oxo) core, given that no external protons have been added to the reaction mixture containing 4.

The addition of a base to 3 allowed us to probe its protonation state further. Upon the addition of triethylamine (NEt₃, 100 equiv) to 3, an immediate reaction was observed (Figure S14). The obtained electronic absorption spectrum

was similar to the broad, featureless spectrum of 4. We postulate that a $(\mu$ -oxo) $(\mu$ -hydroxo) core in 3 was deprotonated by the base NEt₃ to yield the bis $(\mu$ -oxo) complex, 4. This is consistent with the fact that 3 is generated from an acid-mediated activation, whereas 4 was prepared in the absence of an acid.

Mn K-edge XANES was performed on 4; however, the results were equally as nonfruitful as those obtained for 3 (Figure S15).

Measurement of the vibrational properties of 3 and 4 by Raman and infrared (IR) spectroscopies provided limited insights. The following limitations of our vibrational analysis were identified: resonances of interest (ν_{O-O}, ν_{Mn-O}) for 3 and 4 could be masked by the solvent (mixture of CH₃CN/THF) and/or 2,2,2-cryptand and/or BPMP signals. Furthermore, the thermal instability of compound 3 prevented IR analysis. The IR analysis of 4 showed a new feature at $\nu_{\rm Mn-O} = 712~{\rm cm}^{-1}$ that we tentatively assign to a Mn₂O₂ diamond core, the socalled breathing mode (Figure S16). Unfortunately, due to an unexpected explosion of our in-house-prepared K¹⁸O₂ (see Supporting Information for details), we were unable to analyze an isotopically labeled sample. The vibrational properties of μ oxo-Mn₂ species have been probed through Raman and IR spectroscopies, ^{42–45} showing features for $\nu_{\rm Mn-O} \sim 690~{\rm cm}^{-1}$ assigned to the Mn₂O₂ diamond core breathing modes. We tentatively conclude that 4 contains a Mn₂O₂ diamond core.

With the combination of electronic absorption, EPR, IR, and mass spectrometry data, we assign the new species 3 to be a $(\mu$ -oxo) $(\mu$ -hydroxo)Mn^{III}Mn^{IV} species and 4 to be a bis $(\mu$ oxo)Mn^{III}Mn^{IV} complex formed during the protonation or thermal decay of peroxido-Mn^{II}Mn^{III} 2, respectively (Scheme 2). We postulate that the addition of the weak acid p-TsOH as a proton source facilitated O-O bond scission in the peroxide complex 2 and promoted conversion to the oxo-bridged Mn^{III}Mn^{IV} species. In close analogy to the conversion of 2 to 3, Que and co-workers recently reported the cleavage of a μ -1,2peroxido-Fe^{III}₂ species upon the addition of the Lewis acid Sc³⁺ to generate a bis- μ -oxo-Fe^{IV}₂core. ⁴⁶ Moreover, with respect to the formation of 4 from 2, Jackson and co-workers reported a mononuclear peroxido-Mn^{III} species ([Mn^{III}(O₂)(κ⁴-N4Py)]²⁺) that thermally decayed into a bis(μ -oxo)Mn^{III}Mn^{IV} species $[Mn^{III}(\mu-O)_2Mn^{IV}(\kappa^4-N4py)_2]^{3+.25}$ Alongside the reactivity studies below, these literature precedents and our collated spectroscopic data support our structural assignments of 3 and 4.

Reactivity of 3 with Phenols. For the catalytic cycle of class Ib RNRs, it has been postulated that a bis- μ -oxo-Mn^{III}Mn^{IV} species is responsible for the oxidation of a nearby tyrosine residue initiating ribonucleotide reduction. To explore this reactivity further, Mn^{III}Mn^{IV} complexes 3 and 4 were reacted with phenols.

The addition of 4-methoxy-2,6-di-*tert*-butylphenol (4-CH₃O-2,6-DTBP, 50 equiv) to 3 at -90 °C resulted in the decay of the electronic absorption features associated with 3 and the simultaneous growth of a sharp feature at $\lambda_{\rm max}=405$ nm (Figure 5) which matched the features attributed to the independently synthesized 4-methoxy-2,6-di-*tert*-butylphenoxyl radical (Figure S17).^{47,48} The EPR analysis of the postreaction mixture of 3 and 4-CH₃O-2,6-DTBP revealed a sharp and isotropic signal centered at g=2, indicative of the formation of an organic radical (Figure 5),⁴⁹ with a yield of 35% \pm 20%. Interestingly, no other signals (or signals displaying Mnhyperfine) were observed in the postreaction EPR. This

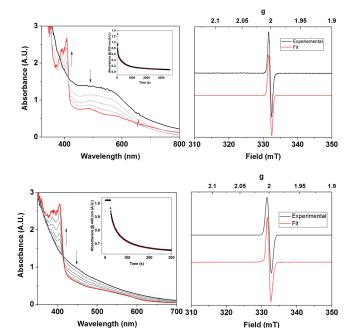


Figure 5. Top left: electronic absorption spectra of the reaction of 3 (black) with 4-CH₃O-2,6-DTBP (50 equiv; red trace is the spectrum at the end of the reaction). Inset: plot of the change of absorption at λ = 550 nm over time. Top right: Perpendicular-mode X-band EPR spectrum of the reaction of 3 and 4-CH₃O-2,6-DTBP measured at 77 K (9.65 GHz, 0.2 mW microwave power, black trace) and simulated spectra (red trace). Bottom left: electronic absorption spectra changes in the reaction of 4 with 4-CH₃O-2,6-DTBP. Inset: plot of the change in absorption at λ = 440 nm over time. Bottom right: perpendicular-mode X-band EPR spectrum of the reaction of 4 with 4-CH₃O-2,6-DTBP measured at 77 K (9.65 GHz, 0.2 mW microwave power, black trace) and the simulated spectrum (red trace).

implies the formation of an EPR-silent Mn product following the reaction of 3 with 4-CH₃O-2,6-DTBP, possibly a Mn^{III} Mnoily. Compound 3 thus facilitated the PCET oxidation of a phenol to yield a phenoxyl radical, mimicking the proposed tyrosyl radical generation process in class Ib RNRs.

Compound 3 also reacted with 9-azabicyclo [3.3.1] nonane-N-hydroxide (ABNO-H) and substrates from the 1-hydroxy-2,2,6,6-tetramethylpiperidine (4-X-TEMPO-H, where X = O, H, and C(O)CH₃) family, allowing us to gain mechanistic insights (Table 2 and Figures S18-S30). These reactions showed the disappearance of the features associated with 3, as determined by electronic absorption spectroscopy. EPR analysis of the postreaction mixture of 3 and TEMPO-H displayed an intense rhombic signal with $g_{av} = 2.00$, consistent with the previously reported spectra for the 2,2,6,6tetramethyl-piperidin-1-yl)oxyl (TEMPO) radical product (Figures S31-S34). 50 The postreaction mixtures for the 3 + ABNO-H, 4-CH₃O-TEMPO-H, and 4-oxo-TEMPOH reactions all displayed similar intense rhombic signals with g_{av} = 2.00. The yields of the radical species were calculated to be 35-75% ($\pm 20\%$) using EPR spectroscopy, assuming a oneelectron oxidation (Table 2). In the ESI-MS spectrum of the postreaction mixture of 3 and TEMPO-H, the mass peak assigned to 3 (m/Z = 335.5837) was no longer present (Figure S35). This provides further evidence for 3 being the active oxidant responsible for the activation of the O-H bonds. In summary, for a series of substrates containing O-H bonds, 3

Table 2. Kinetic Parameters, Yields of Products, and Bond Dissociation Free Energy (BDFE) Values for the Reactions of O-H Bond-Containing Substrates with 3 and 4

	$k_2 \ (\mathrm{M}^{-1} \ \mathrm{s}^{-1})$	$k_1 (s^{-1})$	$K_{\mathrm{M}}~(\mathrm{M}^{-1})$	$\mathrm{BDFE}_{\mathrm{O-H}} \; (\mathrm{kcal} \; \mathrm{mol}^{-1})$	EPR yield of radical (±20%)
3 ^a					
[H]-4-CH ₃ O-2,6-DTBP	0.0072			74.9	35
[D]-4-CH ₃ O-2,6-DTBP	0.0028				
ABNO-H	0.018			71.5	40
ТЕМРО-Н	0.150			66.5	75
4-CH ₃ O-TEMPO-H	0.258			65	60
4-oxo-TEMPO-H	0.134			65.6	75
4^{b}					
4-CN-2,6-DTBP		0.014	0.81	79.4	0
4-H-2,6-DTBP		0.023	0.02	78.3	10
4-CH ₃ -2,6-DTBP		0.025	0.12	76.9	0
4-C ₂ H ₅ -2,6-DTBP		0.058	0.09	75.5	0
2,4,6-TTBP		0.024	0.52	76.7	20
[H]-4-CH ₃ O-2,6-DTBP		0.086	0.002	73.8	70
[D]-4-CH ₃ O-2,6-DTBP		0.032	0.0006		

"For 3: measured at -90 °C; the BDFE_{O-H} values of each substrate are reported in CH₃CN; $^{51-53}$ BDFE_{O-H} values for ABNO-H were not available in CH₃CN, and hence the value was derived using gas-phase bond dissociation enthalpies. Details in Supporting Information. b For 4: measured at +20 °C; the BDFE_{O-H} values for the substrates were not available in 1:9 CH₃CN/THF; hence, we have reported BDFE_{O-H} values for benzene.

facilitated PCET oxidation to yield an organic-oxyl radical product in good- to-high yields.

Fitting of the decay of 3 by following the feature at $\lambda = 550$ nm after the addition of substrate (>10 equiv to ensure pseudo-first-order conditions) yielded a rate of decay (k_{obs}). A plot of the $k_{\rm obs}$ values versus substrate concentration revealed a linear relationship for all substrates, the slope of which was determined to obtain second-order reaction rate constants (k_2) . The free energies of activation of the substrates (ΔG^{\ddagger}) were calculated using the Arrhenius equation, and using these values, a Bell-Evans-Polyani (BEP) plot was constructed (Figure S36). A linear fit of the BEP plot for complex 3 afforded a slope of 0.16, which is within the range (0.15-0.7)predicted by the Marcus theory for concerted PCET mechanisms (thus, hydrogen atom transfer (HAT) or concerted proton and electron transfer (CPET)). \$1-53 Furthermore, a stepwise reaction (thus, PT, followed by ET, or vice versa) with TEMPO-H is thermodynamically unlikely because there is a high barrier to the initial deprotonation or one-electron oxidation of TEMPO-H (p $K_a = 41$ and $E^o = 0.71$ V vs ferrocene/ferrocenium in CH₃CN).⁵¹⁻⁵³ A kinetic isotope effect (KIE, $k_{\rm H}/k_{\rm D}$) of 2.7 was determined for the reaction of 3 with O-H bonds using H/D-4-CH₃O-2,6-DTBP (Figure S37). This KIE value falls within the classical range (2-7) and indicates that PT or HAT was rate-limiting and is in agreement with the KIE of 3.5 determined for the similar complex $(\mu$ -oxo) $(\mu$ -hydroxo)Mn^{III}Mn^{IV} (L^{Mepy}) that performs HAT.⁵⁴ The similarities offer further indication that 3 utilizes a concerted PCET mechanism during the oxidation of O-H bond-containing substrates.

Reactivity of 4 with Phenols. Addition of 4-CH₃O-2,6-DTBP to a solution of 4 (1.5 mM, 1:9 CH₃CN/THF, 10 equiv) resulted in an immediate decrease in the absorption features associated with 4, concomitant with an increase in the new feature at λ = 405 nm (Figure 5). As described above, this sharp peak is associated with the formation of the 4-methoxy-2,6-di-*tert*-butyl-phenoxyl radical (Figure S38). A frozen EPR sample of the postreaction mixture showed a sharp isotropic signal centered at g = 2, confirming the formation of the radical

(Figure 5). The yield of 4-methoxy-2,6-di-*tert*-butyl-phenoxyl radical was determined to be $75 \pm 20\%$ by EPR. Thus, as with 3, 4 was capable of PCET oxidation of phenols to yield the phenoxyl radical, also mimicking the postulated tyrosyl radical generation process in class Ib RNRs.

Complex 4 was reacted with a family of phenols 4-X-2,6-DTBP, where $X = (CH_3O, C(CH_3)_3, C_2H_5, CH_3, H, and CN)$ to gain mechanistic insights (Table 2 and Figures S39-S59). The reactions between 4 and ABNO-H or TEMPO-H and its derivatives were too fast for accurate kinetic analysis. This does not suggest that 4 was more potent than 3 because the reactivity difference can more likely be ascribed to the different temperatures at which the reactions were performed (-90 °C for 3 and + 20 °C for 4). The postreaction mixtures were analyzed via EPR spectroscopy (Figures S60 and S61). 4-CH₃O-2,6-DTBP, 2,4,6-TTBP, and 2,6-DTBP revealed the formation of the corresponding phenoxyl radical species in 75, 20, and 10% (\pm 20%) yields, respectively. The postreaction mixtures for 4-C₂H₅-2,6-DTBP, 4-CH₃-2,6-DTBP, and 4-CN-2,6-DTBP displayed no EPR signals at 77 K, indicating that any formed phenoxyl radicals had decayed further. Gas chromatography (GC) analysis of the postreaction mixtures for $X = CH_3O$, $C(CH_3)_3$, CN, and C_2H_5 showed the formation of 2,6-di-tert-butyl-1,4-benzoquinone (2,6-DTBQ), a product of the decay of the parent phenoxyl radicals. Interestingly, as with 3, in the postreaction EPR spectra for all substrates, no Mn signals were observed. Overall, 4 reacted with the phenolic substrates to yield phenoxyl radicals, which then decayed further in some instances.

Plotting the measured $k_{\rm obs}$ values against the substrate concentration ([S]) for the reaction between 4 and 4-CH₃-2,6-DTBP revealed a nonlinear behavior (Figure 6), a trend which was observed for the reactions between 4 and all substrates. This trend indicates the presence of an equilibrium prior to an irreversible substrate transformation. In order to gain a deeper understanding of how 4 was reacting, the kinetic plots of the phenols were linearized according to the equation, $\frac{1}{k_{\rm obs}} = \frac{K}{k_1} \frac{1}{[S]} + \frac{1}{k_1}$, allowing us to calculate k_1 , the maximum rate of reaction, and $K_{\rm M}$, the [S] at half the maximum rate.

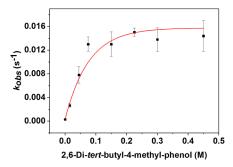


Figure 6. Plot of substrate concentration versus $k_{\rm obs}$ for the reaction of 4 and 4-CH₃-2,6-DTBP.

Using k_1 , ΔG^{\ddagger} was calculated for each substrate. Plotting ΔG^{\ddagger} versus BDFE_{O-H} yielded a plot with a slope of 0.15 (Figure S62). As with 3, 4 falls in the range of slopes reported for systems ascribed to concerted PCET (0.15-0.7), $^{31-3}$ indicating that concerted PCET was the mechanism 4 employed to oxidize phenols. A plot of the logs of k_1 or $K_{\rm M}$ versus the p K_a 's, or K_M versus E_{OX} of the substrates revealed no discernible relationship (Figures S63-S65). In contrast, a Marcus plot of (RT/\hat{F}) $\ln(\tilde{k}_1)$ versus E_{OX} was linear with a slope of -0.36 (Figure S66). This value is on the borderline between those associated with the rate-limiting concerted PCET and rate-limiting ET. 55-58 A KIE of 2.7 was determined for the reaction of 4 with H/D-4-CH₃O-2,6-DTBP (Figures S67 and S68), emblematic of the PT or HAT involved in the rate-limiting step. This further supports a concerted PCET mechanism during the oxidation of phenols of 4. Previous studies into the reactions of bis-µ-oxo-Mn2 complexes with hydrocarbons and phenols provided similar insights—the oxidants abstracted H atoms via a concerted PCET mechanism. 59,60 The KIE value was identical to that obtained for 3, suggesting that both complexes follow a similar pathway. Based on the collected evidence, we tentatively conclude that 3 and 4 oxidized substrates via a concerted PCET reaction mechanism.

We postulate that the source of the nonlinear kinetics in the reactivity of 4 is an initial association of the phenol O–H moiety to the bis- μ -oxo-Mn^{III}Mn^{IV} core (Scheme 3). Jackson and co-workers reported Mn^{III}–OH supported by the bispyridylamine monocarboxamidate ligand dpaq (dpaq = 2-[bis(pyridin-2-ylmethyl)]amino-N-quinolin-8-yl-acetamidate) that showed a similar saturation kinetic behavior in its reaction with the same family of phenols. They postulated that Mn^{III}–OH formed a hydrogen-bonded adduct with phenol in the

Scheme 3. Graphical Explanation of the Influence of Protons in PCET Oxidation by 3 and 4

initial step, followed by an irreversible oxidation of the phenol. The structures of 3 and 4 differ only by the availability of protons that could reside on a μ -oxo-ligand in 3, which are presumed absent in 4 due to the nonavailability of protons in the reaction mixture containing 4. The absence of saturation kinetics for 3 supports this structural assignment and is consistent with our observations of linear kinetics from 3 in its reactions with the O–H bond-containing substrates.

As stated in Introduction, the role of a H⁺-donor in O–O bond cleavage in RNRs is poorly understood, and the protonation state of the bis- μ -oxo-Mn^{III}Mn^{IV} core is unknown. The current evidence demonstrates that an H⁺ can facilitate O–O bond cleavage in a peroxide-Mn^{II}Mn^{III} complex to yield a bis- μ -oxo-Mn^{III}Mn^{IV} entity, 3. However, provision of an H⁺ is not a prerequisite for bis- μ -oxo-Mn^{III}Mn^{IV} formation, as evidenced by the thermally induced formation of 4. Second, the protonation state has an impact on the reactivity properties of bis- μ -oxo-Mn^{III}Mn^{IV} cores, where the current evidence demonstrates that the phenolic substrate displays a preequilibrium hydrogen-bonding step prior to its oxidation, in its interactions with a bis- μ -oxo-Mn^{III}Mn^{IV} oxidant.

CONCLUSIONS AND MECHANISTIC IMPLICATIONS FOR CLASS IB RNRS

In the postulated catalytic cycle of class Ib RNRs (Scheme 1), a Mn_{2}^{II} center is suggested to react with superoxide (O_{2}^{-}) , rather than dioxygen, to generate an intermediate peroxido-Mn^{II}Mn^{III} adduct. The peroxido-Mn^{II}Mn^{III} adduct is believed to rapidly convert to a putative Mn^{III}Mn^{IV} entity, where the involvement/role of a H+-donor is poorly understood. The Mn^{III}Mn^{IV} entity, identified through EPR analysis, is understood to be responsible for the tyrosyl radical/Mn^{III}₂ cofactor generation via PCET oxidation of tyrosine. In this work, a Mn^{II}₂ complex (1) was reacted with O₂⁻ to form peroxido-Mn^{II}Mn^{III} (2). Importantly, 1 did not react with atmospheric O_2 . 2 was converted to $(\mu$ -O) $(\mu$ -OH)Mn^{III}Mn^{IV} (3) via the addition of an H⁺-donor (p-TsOH) or $(\mu$ -O)₂Mn^{III}Mn^{IV} (4) upon warming to room temperature. The physical properties of 3 and 4 were probed using UV-vis, EPR, X-ray absorption, and IR spectroscopies and mass spectrometry. Interestingly, 3 and 4 displayed EPR spectra similar to those of the implicated active Mn^{III}Mn^{IV} species identified in the studies of class Ib RNRs, where a 16-line signal consistent with a Mn^{III}Mn^{IV} oxidant was also observed. The synthetic model thus provides experimental support for the postulated biochemical O₂ activation and high-valent oxidant generation pathways proposed for class Ib Mn₂ RNRs. Critically, the trapped synthetic Mn^{III}Mn^{IV} complexes 3 and 4 were capable of phenol oxidation to yield a phenoxyl radical, supporting the proposed mechanism of tyrosyl radical/Mn^{III}₂ cofactor generation in RNRs. Kinetic analysis revealed that 3 and 4 were oxidizing phenolic substrates through a concerted PCET reaction mechanism, albeit with different kinetic profiles consistent with their different protonation states. The synthetic models reported herein thus support the entire postulated catalytic cycle of the enzyme and provide insights into the spectral and reactivity properties of the as-yet unidentified intermediates in the RNR cycle.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.inorgchem.3c04163.

Experimental and physical methods, NMR, electronic absorption, EPR, IR, XAS spectra, mass spectrometry data, kinetics data, and analysis (PDF)

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Notes

The authors declare no competing financial interest. Perchlorate salts of metal complexes and KO_2 are potentially explosive and must be handled with care and in small quantities. We limited our preparation of 1 to a maximum of 0.2 g per reaction. Caution: preparation and storage of $K^{18}O_2$: we had a minor explosion, inside our glovebox, when handling (using a metal spatula) a small quantity (20–40 mg) of our inhouse-prepared $K^{18}O_2$ in a glass Schlenk flask. Details are provided in the Supporting Information.

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REFERENCES

- (1) Eriksson, M.; Jordan, A.; Eklund, H. Structure of *Salmonella typhimurium nrdF* Ribonucleotide Reductase in Its Oxidized and Reduced Forms. *Biochemistry* **1998**, *37*, 13359–13369.
- (2) Stubbe, J.; Nocera, D. G.; Yee, C. S.; Chang, M. C. Y. Radical Initiation in the Class I Ribonucleotide Reductase: Long-Range Proton-Coupled Electron Transfer? *Chem. Rev.* **2003**, *103*, 2167–2201
- (3) Kolberg, M.; Strand, K. R.; Graff, P.; Andersson, K. K. Structure, function, and mechanism of ribonucleotide reductases. *Biochim. Biophys. Acta (BBA) Proteins Proteomics* **2004**, *1699* (1), 1–34.
- (4) Cotruvo, J. A.; Stubbe, J. Class I Ribonucleotide Reductases: Metallocofactor Assembly and Repair In Vitro and In Vivo. *Annu. Rev. Biochem.* **2011**, *80* (1), 733–767.
- (5) Aye, Y.; Li, M.; Long, M. J. C.; Weiss, R. S. Ribonucleotide reductase and cancer: biological mechanisms and targeted therapies. *Oncogene* **2015**, 34 (16), 2011–2021.
- (6) Ruskoski, T. B.; Boal, A. K. The periodic table of ribonucleotide reductases. *J. Biol. Chem.* **2021**, 297 (4), No. 101137.
- (7) Boal, A. K.; Cotruvo, J. A.; Stubbe, J.; Rosenzweig, A. C. Structural Basis for Activation of Class Ib Ribonucleotide Reductase. *Science* **2010**, 329 (5998), 1526–1530.
- (8) Cotruvo, J. A., Jr.; Stubbe, J. Escherichia coli Class Ib Ribonucleotide Reductase Contains a Dimanganese(III)-Tyrosyl Radical Cofactor in Vivo. *Biochemistry* **2011**, *50* (10), 1672–1681.
- (9) Boal, A. K.; Cotruvo, J. A., Jr; Stubbe, J.; Rosenzweig, A. C. The Dimanganese(II) Site of Bacillus subtilis Class Ib Ribonucleotide Reductase. *Biochemistry* **2012**, *51* (18), 3861–3871.
- (10) Cox, N.; Ogata, H.; Stolle, P.; Reijerse, E.; Auling, G.; Lubitz, W. A Tyrosyl-Dimanganese Coupled Spin System is the Native Metalloradical Cofactor of the R2F Subunit of the Ribonucleotide Reductase of Corynebacterium ammoniagenes. *J. Am. Chem. Soc.* **2010**, *132* (32), 11197–11213.
- (11) Cotruvo, J. A., Jr.; Stich, T. A.; Britt, R. D.; Stubbe, J. Mechanism of Assembly of the Dimanganese-Tyrosyl Radical Cofactor of Class Ib Ribonucleotide Reductase: Enzymatic Generation of Superoxide Is Required for Tyrosine Oxidation via a Mn(III)Mn(IV) Intermediate. *J. Am. Chem. Soc.* **2013**, *135* (10), 4027–4039.
- (12) Kang, G.; Taguchi, A. T.; Stubbe, J.; Drennan, C. L. Structure of a trapped radical transfer pathway within a ribonucleotide reductase holocomplex. *Science* **2020**, *368* (6489), 424–427.
- (13) Minnihan, E. C.; Nocera, D. G.; Stubbe, J. Reversible, Long-Range Radical Transfer in E. coli Class Ia Ribonucleotide Reductase. *Acc. Chem. Res.* **2013**, *46* (11), 2524–2535.
- (14) Magherusan, A. M.; Nelis, D. N.; Twamley, B.; McDonald, A. R. Catechol oxidase activity of comparable dimanganese and dicopper complexes. *Dalton Trans.* **2018**, 47 (43), 15555–15564.
- (15) Magherusan, A. M.; Zhou, A.; Farquhar, E. R.; García-Melchor, M.; Twamley, B.; Que, L.; McDonald, A. R. Mimicking Class I b Mn_2 Ribonucleotide Reductase: A Mn_1^{II} Complex and Its Reaction with Superoxide. *Angew. Chem., Int. Ed.* **2018**, *57* (4), 918–922.
- (16) Magherusan, A. M.; Kal, S.; Nelis, D. N.; Doyle, L. M.; Farquhar, E. R.; Que, L.; McDonald, A. R. A Mn^{II}Mn^{III}-Peroxide Complex Capable of Aldehyde Deformylation. *Angew. Chem., Int. Ed.* **2019**, *58* (17), *5718–5722*.
- (17) Cooper, S. R.; Dismukes, G. C.; Klein, M. P.; Calvin, M. Mixed Valence Interactions in Di- μ -oxo Bridged Manganese Complexes. Electron Paramagnetic Resonance and Magnetic Susceptibility Studies. *J. Am. Chem. Soc.* **1978**, *100* (23), 7248–7252.
- (18) Brewer, K. J.; Calvin, M.; Lumpkin, R. S.; Otvos, J. W.; Spreer, L. O. Synthesis, structure, and characterization of a mixed-valence

- manganese(III)-manganese(IV) bis- μ -oxo complex with a macrocyclic tetraaza ligand. *Inorg. Chem.* **1989**, 28 (25), 4446–4451.
- (19) Mahapatra, S.; Das, P.; Mukherjee, R. A new mixed-valence binuclear complex containing the $[Mn^{IV}(\mu\text{-O})_2(\mu\text{-O}_2\text{CMe})Mn^{III}]^{2+}$ core: synthesis, magnetism, electron paramagnetic resonance and redox properties. *J. Chem. Soc., Dalton Trans.* 1993, 2, 217–220.
- (20) Frapart, Y.-M.; Boussae, A.; Albach, R.; Anxolabéhére-Mallart, E.; Delroisse, M.; Verlhac, J.-B.; Blondin, G.; Girerd, J.-J.; Guilhem, J.; Cesario, M.; et al. Chemical Modeling of the Oxygen-Evolving Center in Plants. Synthesis, Structure, and Electronic and Redox Properties of a New Mixed Valence Mn-Oxo Cluster: [Mn₂^{III,IV}O₂(bisimMe₂en)₂]³⁺ (bisimMe₂en = *N,N'*-Dimethyl-*N,N'*-bis(imidazol-4-ylmethyl)ethane-1,2-diamine). EPR Detection of an Imidazole Radical Induced by UV Irradiation at Low Temperature. *J. Am. Chem. Soc.* **1996**, *118*, 2669–2678.
- (21) Gelasco, A.; Kirk, M. L.; Kampf, J. W.; Pecoraro, V. L. The $[Mn_2(2\text{-OHsalpn})_2]^{2\gamma,0,+}$ System: Synthesis, Structure, Spectroscopy, and Magnetism of the First Structurally Characterized Dinuclear Manganese Series Containing Four Distinct Oxidation States. *Inorg. Chem.* **1997**, 36 (9), 1829–1837.
- (22) Collomb, M.-N.; Deronzier, A.; Richardot, A. L.; Pe'caut, J. Synthesis and characterization of a new kind of $Mn_2^{III,IV}$ μ -oxo complex: $[Mn_2O_2(terpy)_2(H_2O)_2](NO_3)_3$ ·6 H_2O , terpy = 2,2':6',2"-terpyridine. *New. J. Chem.* **1999**, 23 (4), 351–354.
- (23) Yamazaki, H.; Igarashi, S.; Nagata, T.; Yagi, M. Substituent Effects on Core Structures and Heterogeneous Catalytic Activities of $\mathrm{Mn^{III}}(\mu\text{-O})_2\mathrm{Mn^{IV}}$ Dimers with 2,2':6',2"-Terpyridine Derivative Ligands for Water Oxidation. *Inorg. Chem.* **2012**, *51* (3), 1530–1539.
- (24) Leto, D. F.; Chattopadhyay, S.; Day, V. W.; Jackson, T. A. Reaction landscape of a pentadentate N5-ligated Mn^{II} complex with O_2^- and H_2O_2 includes conversion of a peroxomanganese(III) adduct to a bis(μ -oxo)dimanganese(III,IV) species. *Dalton Trans.* **2013**, 42 (36), 13014–13025.
- (25) Horner, O.; Anxolabéhère-Mallart, E.; Charlot, M.-F.; Tchertanov, L.; Guilhem, J.; Mattioli, T. A.; Boussac, A.; Girerd, J.-J. A New Manganese Dinuclear Complex with Phenolate Ligands and a Single Unsupported Oxo Bridge. Storage of Two Positive Charges within Less than 500 mV. Relevance to Photosynthesis. *Inorg. Chem.* 1999, 38 (6), 1222–1232.
- (26) Baldwin, M. J.; Stemmler, T. L.; Riggs-Gelasco, P. J.; Kirk, M. L.; Penner-Hahn, J. E.; Pecoraro, V. L. Structural and Magnetic Effects of Successive Protonations of Oxo Bridges in High-Valent Manganese Dimers. J. Am. Chem. Soc. 1994, 116, 11349–11356.
- (27) Suzuki, M.; Tokura, S.; Suhara, M.; Uehara, A. Dinuclear Manganese (III,IV) and Manganese (IV, IV) Complexes with Tris(2-pyridylmethyl)amine. *Chem. Lett.* **1988**, *17*, 477–480.
- (28) Goodson, P. A.; Hodgson, D. J. Synthesis and Characterization of a Bis-Oxo-Bridged Mn^{III}Mn^{III} Complex, Di-μ-oxobis[N,N'-bis(2,6-dimethylpyridyl)ethane-1,2-diamine]dimanganese(III,III) Perchlorate. *Inorg. Chem.* **1989**, 28, 3606–3608.
- (29) Goodson, P. A.; Oki, A. R.; Glerup, J.; Hodgson, D. J. Design, Synthesis, and Characterization of Bis(μ -oxo)dimanganese(III,III) Complexes. Steric and Electronic Control of Redox Potentials. *J. Am. Chem. Soc.* **1990**, *112*, 6248–6254.
- (30) Goodson, P. A.; Glerup, J.; Hodgson, D. J.; Michelsen, K.; Pedersen, E. Binuclear bis(μ -oxo)dimanganese(III,IV) and -(IV,IV) complexes with N,N'-bis(2-pyridylmethyl)-1,2-ethanediamine. *Inorg. Chem.* **1990**, 29 (3), 503–508.
- (31) Dismukes, G. C.; Sheats, J. E.; Smegal, J. A. Mn²⁺/Mn³⁺ and Mn³⁺/Mn⁴⁺ Mixed Valence Binuclear Manganese Complexes of Biological Interest. *J. Am. Chem. Soc.* **1987**, *109*, 7202–7203.
- (32) Messinger, J.; Robblee, J. H.; Yu, W. O.; Sauer, K.; Rachandra, V. K.; Klein, M. P. The S₀ State of the Oxygen-Evolving Complex in Photosystem II Is Paramagnetic: Detection of an EPR Multiline Signal. J. Am. Chem. Soc. 1997, 119, 11349–11350.
- (33) Teutloff, C.; Schäfer, K.-O.; Sinnecker, S.; Barynin, V.; Bittl, R.; Wieghardt, K.; Lendzian, F.; Lubitz, W. High-field EPR investigations of Mn^{III}Mn^{IV} and Mn^{II}Mn^{III} states of dimanganese catalase and related model systems. *Magn. Reson. Chem.* **2005**, *43* (S1), S51–S64.

- (34) Hureau, C.; Sabater, L.; Anxolabéhère-Mallart, E.; Nierlich, M.; Charlot, M.-F.; Gonnet, F.; Rivière, E.; Blondin, G. Synthesis, Structure, and Characterisation of a New Phenolato-Bridged Manganese Complex $[Mn_2(mL)_2]^{2+}$: Chemical and Electrochemical Access to a New Mono- μ -Oxo Dimanganese Core Unit. *Chem.—Eur. J.* 2004, 10 (8), 1998–2010.
- (35) Lessa, J. A.; Horn, A., Jr; Bull, É. S.; Rocha, M. R.; Benassi, M.; Catharino, R. R.; Eberlin, M. N.; Casellato, A.; Noble, C. J.; Hanson, G. R.; et al. Catalase vs Peroxidase Activity of a Manganese(II) Compound: Identification of a $Mn(III)-(\mu-O)_2-Mn(IV)$ Reaction Intermediate by Electrospray Ionization Mass Spectrometry and Electron Paramagnetic Resonance Spectroscopy. *Inorg. Chem.* **2009**, 48 (10), 4569–4579.
- (36) Howard, T.; Telser, J.; DeRose, V. J. An Electron Paramagnetic Resonance Study of $Mn_2(H_2O)(OAc)_4(tmeda)_2$ (tmeda = N,N,N',N'-Tetramethylethylenediamine): A Model for Dinuclear Manganese Enzyme Active Sites. *Inorg. Chem.* **2000**, 39 (15), 3379–3385.
- (37) Golombek, A. P.; Hendrich, M. P. Quantitative analysis of dinuclear manganese(II) EPR spectra. *J. Magn. Reson.* **2003**, *165* (1), 33–48.
- (38) Diril, H.; Chang, H.-R.; Nilges, M. J.; Zhang, X.; Potenza, J. A.; Schugar, H. J.; Isied, S. S.; Hendrickson, D. N. Simulation Strategies for Unusual EPR Spectra of Binuclear Mixed-Valence Manganese Complexes: Synthesis, Properties, and X-ray Structures of the Mn^{II}Mn^{III} Complexes [Mn₂(bpmp)(μ-OAc)₂](ClO₄)·H₂O and [Mn₂(bcmp)(m-OAc)₂](ClO₄)·CH₂Cl₂. *J. Am. Chem. Soc.* **1989**, *111*, 5102–5114.
- (39) Stemmler, T. L.; Sossong, J.; T, M.; Goldstein, J. I.; Ash, D. E.; Elgren, T. E.; Kurtz, D. M., Jr.; Penner-Hahn, J. E. EXAFS Comparison of the Dimanganese Core Structures of Manganese Catalase, Arginase, and Manganese-Substituted Ribonucleotide Reductase and Hemerythrin. *Biochemistry* 1997, 36, 9847–9858.
- (40) Schreiber, R. E.; Cohen, H.; Leitus, G.; Wolf, S. G.; Zhou, A.; Que, L., Jr; Neumann, R. Reactivity and O₂ Formation by Mn(IV)-and Mn(V)-Hydroxo Species Stabilized within a Polyfluoroxometalate Framework. *J. Am. Chem. Soc.* **2015**, *137* (27), 8738–8748.
- (41) Rose, H. R.; Ghosh, M. K.; Maggiolo, A. O.; Pollock, C. J.; Blaesi, E. J.; Hajj, V.; Wei, Y.; Rajakovich, L. J.; Chang, W.-C.; Han, Y.; et al. Structural Basis for Superoxide Activation of Flavobacterium johnsoniae Class I Ribonucleotide Reductase and for Radical Initiation by Its Dimanganese Cofactor. *Biochemistry* **2018**, *57* (18), 2679–2693.
- (42) Cua, A.; Vrettos, J. S.; de Paula, J. C.; Brudvig, G. W.; Bocian, D. F. Raman spectra and normal coordinate analyses of low-frequency vibrations of oxo-bridged manganese complexes. *J. Bioinorg. Chem.* **2003**, 8 (4), 439–451.
- (43) Cooper, S. R.; Calvin, M. Mixed valence interactions in di- μ -oxo bridged manganese complexes. *J. Am. Chem. Soc.* **1977**, 99 (20), 6623–6630.
- (44) Manchanda, R.; Brudvig, G. W.; Crabtree, R. H. High-Valent Oxomanganese Clusters: Structural and Mechanistic Work Relevant to the Oxygen Evolving Center in Photosystem II. *Coord. Chem. Rev.* 1995, 144, 1–38.
- (45) Dubois, L.; Pécaut, J.; Charlot, M.-F.; Baffert, C.; Collomb, M.-N.; Deronzier, A.; Latour, J.-M. Carboxylate Ligands Drastically Enhance the Rates of Oxo Exchange and Hydrogen Peroxide Disproportionation by Oxo Manganese Compounds of Potential Biological Significance. *Chem.—Eur. J.* **2008**, *14* (10), 3013–3025.
- (46) Banerjee, S.; Draksharapu, A.; Crossland, P. M.; Fan, R.; Guo, Y.; Swart, M.; Que, L., Jr Sc³⁺-Promoted O–O Bond Cleavage of a $(\mu$ -1,2-Peroxo)Diiron(III) Species Formed from an Iron(II) Precursor and O₂ to Generate a Complex with an Fe^{IV}₂ $(\mu$ -O)₂ Core. *J. Am. Chem. Soc.* **2020**, *142* (9), 4285–4297.
- (47) Wittman, J. M.; Hayoun, R.; Kaminsky, W.; Coggins, M. K.; Mayer, J. M. A C-C Bonded Phenoxyl Radical Dimer with a Zero Bond Dissociation Free Energy. *J. Am. Chem. Soc.* **2013**, *135* (35), 12956–12959.

- (48) Spedalotto, G.; Lovisari, M.; McDonald, A. R. Reactivity Properties of Mixed- and High-Valent $Bis(\mu$ -Hydroxide)-Dinickel Complexes. *ACS Omega* **2021**, *6* (42), 28162–28170.
- (49) Lovisari, M.; McDonald, A. R. Hydrogen Atom Transfer Oxidation by a Gold-Hydroxide Complex. *Inorg. Chem.* **2020**, *6* (59), 3659–3665.
- (50) Wijeratne, G. B.; Corzine, B.; Day, V. W.; Jackson, T. A. Saturation Kinetics in Phenolic O–H Bond Oxidation by a Mononuclear Mn(III)–OH Complex Derived from Dioxygen. *Inorg. Chem.* **2014**, *53* (14), 7622–7634.
- (51) Warren, J. J.; Tronic, T. A.; Mayer, J. M. Thermochemistry of Proton-Coupled Electron Transfer Reagents and its Implications. *Chem. Rev.* **2010**, *110* (12), 6961–7001.
- (52) Darcy, J. W.; Koronkiewicz, B.; Parada, G. A.; Mayer, J. M. A Continuum of Proton-Coupled Electron Transfer Reactivity. *Acc. Chem. Res.* **2018**, *51* (10), 2391–2399.
- (53) Agarwal, R. G.; Coste, S. C.; Groff, B. D.; Heuer, A. M.; Noh, H.; Parada, G. A.; Wise, C. F.; Nichols, E. M.; Warren, J. J.; Mayer, J. M. Free Energies of Proton-Coupled Electron Transfer Reagents and Their Applications. *Chem. Rev.* **2022**, *122* (1), 1–49.
- (54) Blakely, M. N.; Dedushko, M. A.; Yan Poon, P. C.; Villar-Acevedo, G.; Kovacs, J. A. Formation of a Reactive, Alkyl Thiolate-Ligated Fe^{III}-Superoxo Intermediate Derived from Dioxygen. *J. Am. Chem. Soc.* **2019**, *141* (5), 1867–1870.
- (55) Ram, M.; Hupp, J. T. Linear free energy relations for multielectron transfer kinetics: a brief look at the Broensted/Tafel analogy. *J. Phys. Chem. A* **1990**, *94* (6), 2378–2380.
- (56) Weatherly, S. C.; Yang, I. V.; Thorp, H. H. Proton-coupled electron transfer in duplex DNA: driving force dependence and isotope effects on electrocatalytic oxidation of guanine. *J. Am. Chem. Soc.* **2001**, *123* (6), 1236–1237.
- (57) Osako, T.; Ohkubo, K.; Taki, M.; Tachi, Y.; Fukuzumi, S.; Itoh, S. Oxidation mechanism of phenols by dicopper—dioxygen (Cu₂/O₂) complexes. *J. Am. Chem. Soc.* **2003**, *125* (36), 11027–11033.
- (58) Kundu, S.; Miceli, E.; Farquhar, E. R.; Ray, K. Mechanism of phenol oxidation by heterodinuclear NiCu-bis-(μ -oxo) complexes involving nucleophilic oxo groups. *Dalton Trans.* **2014**, 43 (11), 4264–4267.
- (59) Wang, K.; Mayer, J. M. Oxidation of Hydrocarbons by $[(phen)_2Mn(\mu-O)_2Mn(phen)_2]^{3+}$ via Hydrogen Atom Abstraction. J. Am. Chem. Soc. Society **1997**, 119 (6), 1470–1471.
- (60) Baldwin, M. J.; Pecoraro, V. L. Energetics of Proton-Coupled Electron Transfer in High-Valent $\mathrm{Mn_2}(\mu\text{-O})_2$ Systems: Models for Water Oxidation by the Oxygen-Evolving Complex of Photosystem II. *J. Am. Chem. Soc.* **1996**, *118*, 11325–11326.