



**HAL**  
open science

## Respiratory organoids models from bronchial cells of asthmatic patients: Fabrication and study of viral infection permissivity

Flora Archambault, Fabienne Tombette, Magalie Benard, Ludovic Galas, Caroline Tapparel, Samuel Constant, Jean-Christophe Plantier, Elodie Alessandri-Gradt

### ► To cite this version:

Flora Archambault, Fabienne Tombette, Magalie Benard, Ludovic Galas, Caroline Tapparel, et al.. Respiratory organoids models from bronchial cells of asthmatic patients: Fabrication and study of viral infection permissivity. Journées normandes de recherche biomédicale (JNRB), Jun 2024, Rouen, France. . hal-04615618

**HAL Id: hal-04615618**

**<https://hal.science/hal-04615618v1>**

Submitted on 18 Jun 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Respiratory organoids models from bronchial cells of asthmatic patients : Fabrication and study of viral infection permissivity

Flora Archambault<sup>1</sup>, Fabienne Tombette<sup>2</sup>, Magalie Benard<sup>3</sup>, Ludovic Galas<sup>3</sup>, Caroline Tapparel<sup>4</sup>, Samuel Constant<sup>5</sup>, Jean-Christophe Plantier<sup>2</sup> and Elodie Alessandri-Gradt<sup>2</sup>

<sup>1</sup> Univ Rouen Normandie, Université de Caen Normandie, INSERM, Normandie Univ, DYNAMICURE UMR 1311, F-76000 Rouen, France;

<sup>2</sup> Univ Rouen Normandie, Université de Caen Normandie, INSERM, Normandie Univ, DYNAMICURE UMR 1311, CHU Rouen, Department of virology, F-76000 Rouen, France;

<sup>3</sup> Univ Rouen Normandie, INSERM, CNRS, Normandie Université, HeRaLeS US51 UAR2026, Primacen, F-76000 Rouen, France; <sup>4</sup> Département de Microbiologie et Médecine moléculaire, Université de Genève, 1206 Genève, Suisse; <sup>5</sup> Epithelix SAS

## Introduction

With **4 millions people in France**, asthma is a serious public health issue. The development of new models mimicking lung tissue complexity and functionalities is essential to fight and understand this pathology.

**Respiratory viruses are the first incriminated in asthma exacerbation<sup>1</sup>** with a predominance of Respiratory syncytial viruses (RSV) and human Rhinovirus (RV) even in co-infections (30%)<sup>2</sup>, but aim of viruses in asthma etiology still misunderstood

### NEW MODEL : ORGANOIDS

3D multicellular tissues, aim is to mimic human organs complex functionalities

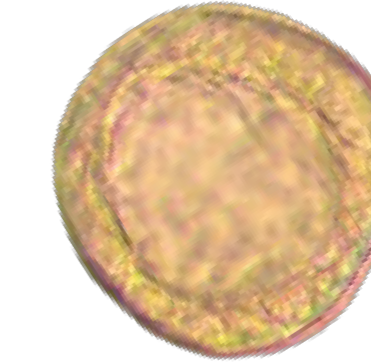
**2022** : development of **healthy respiratory organoids with inverted polarity** (ciliated cells on external face, allow exposure of virus target cells) and infected with RV and Influenza<sup>3</sup>.

## Goals

→ **Develop apical out organoids with bronchial cells of asthmatic patients**

Primary epithelial cells from the respiratory tract of asthmatic donors

Inverted polarity respiratory organoids



1) **Production**

- Characterize the model
- Confirm apical out configuration
- Define cellular composition

2) **Successful production**

- Analyse the impact of **RSV subtype A** infection on asthmatic organoids

Characterize Improve Criticize

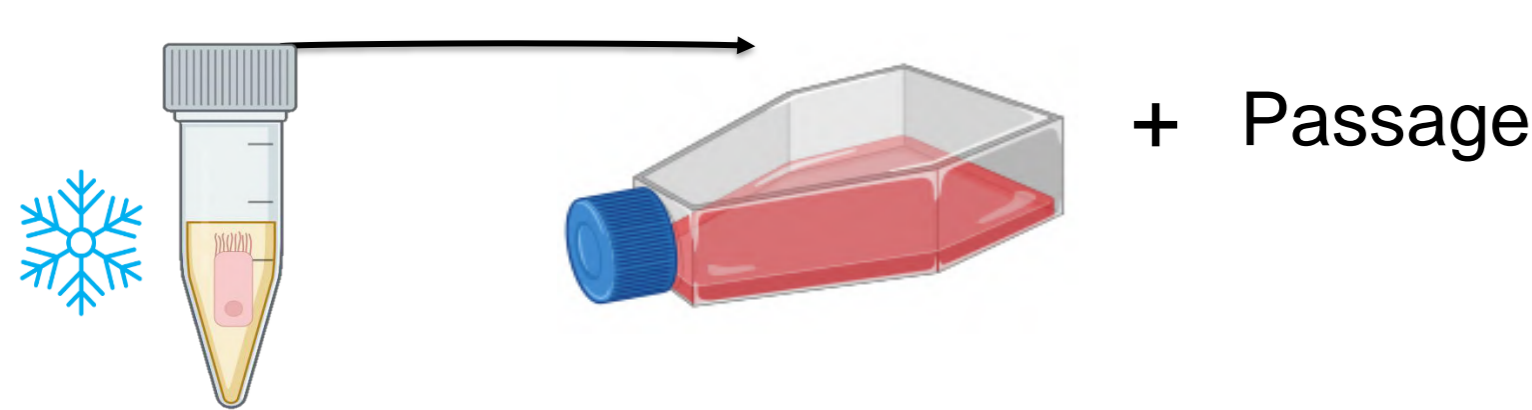
**Pilot study : produce preliminary data on the first asthma organoids model**

Compare to a control (healthy cells)

## Materiels et Methods

### 1 Organoids production :

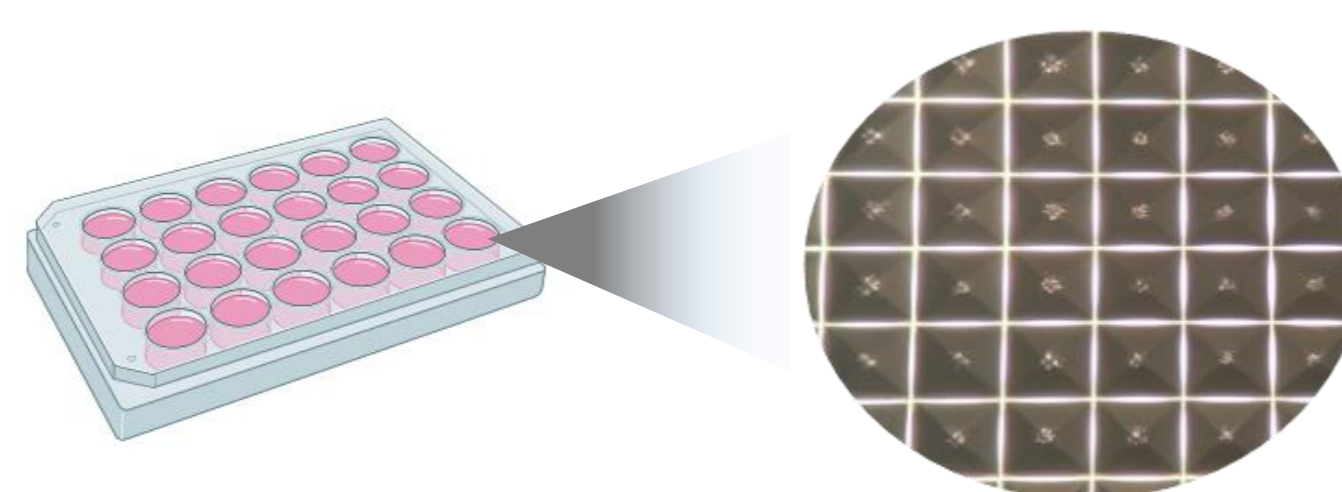
Thawing human airway epithelial cells (hAEC) (healthy +asthmatic donors) and grow in hAEC medium (Epithelix)



2D cell culture

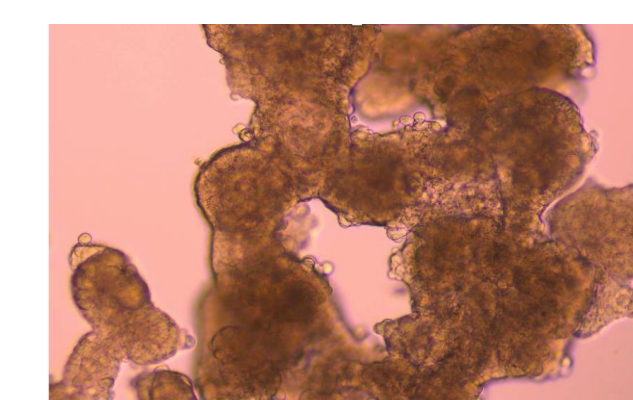
PneumaCult™ Medium + AggreWell™ 400 plates (Stemcell)

Differentiation of hAEC cells to mature apical-out airway organoids, generates organoids in 15 days



Cellular aggregation

Fused asthma organoids at D15



\*Optimization : Determine number of day in Aggrewell based on the model to avoid fused organoids

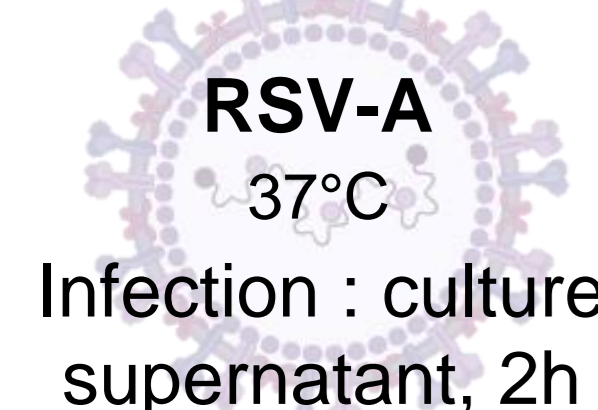
After aggregation time, use a flat bottom plate until D15

Organoids maturation

Immunofluorescent staining : DAPI for nucleus Leica Stellaris 8 (confocal microscopy)

Bioimaging PRIMACEN Plateform

### 2 Successful production: Infection D15



Infection : culture supernatant, 2h

After infection : 100 organoids per well (96w plate) in triplicate

Infection kinetics : Collect at D1, D2, D4, D6 and D8 post infection (pi)

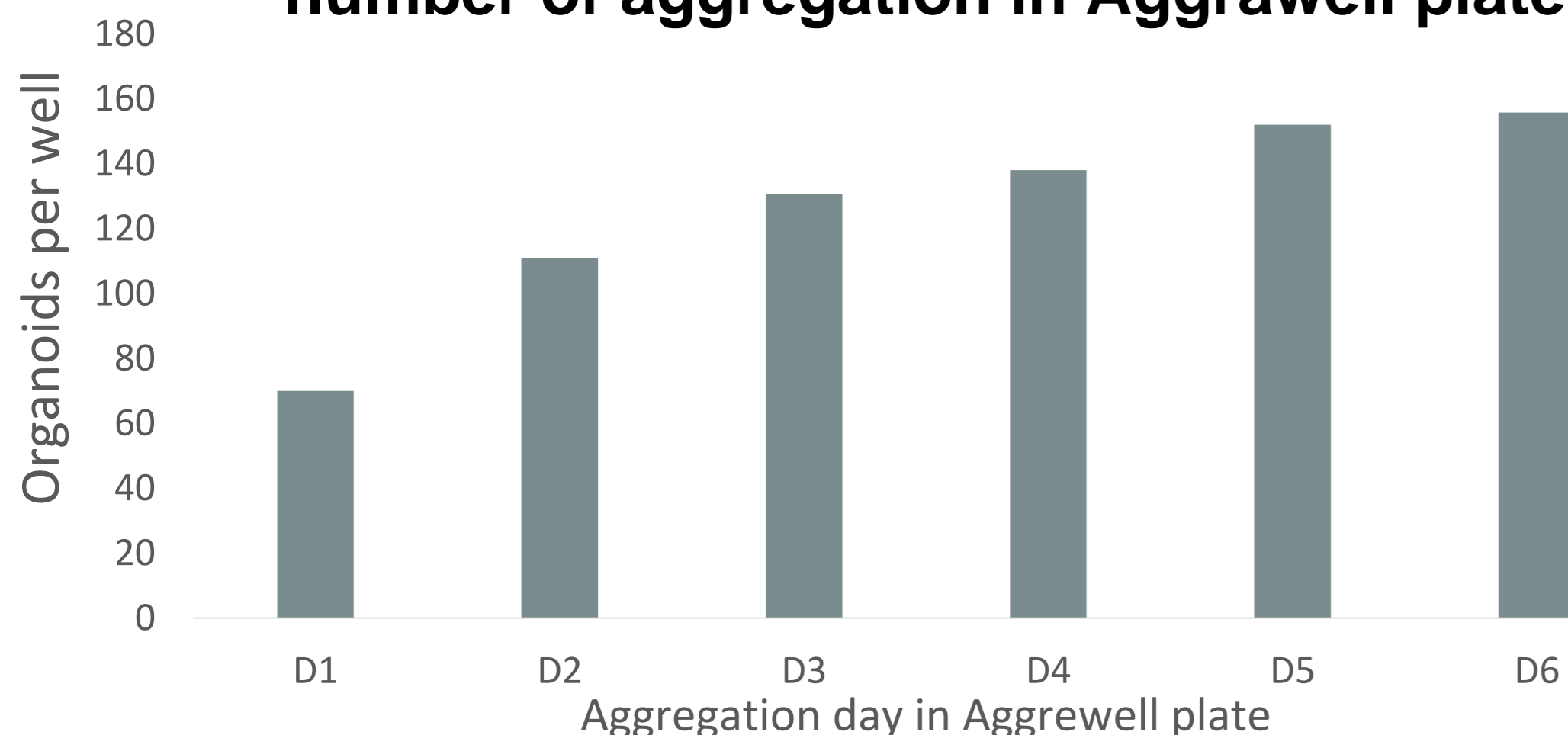
RNA Extraction : kit EZ1 DSP virus Qiagen

Viral quantification qRT-PCR (CFX96 Biorad)

## Results

### 1 Organoids production

1) Asthma organoids count at D15 according to number of aggregation in Aggrewell plate



→ Day (D) 4, D5 and D6 selected (lot of fused organoids on D1, D2 and D3)  
→ D3 selected for healthy organoids

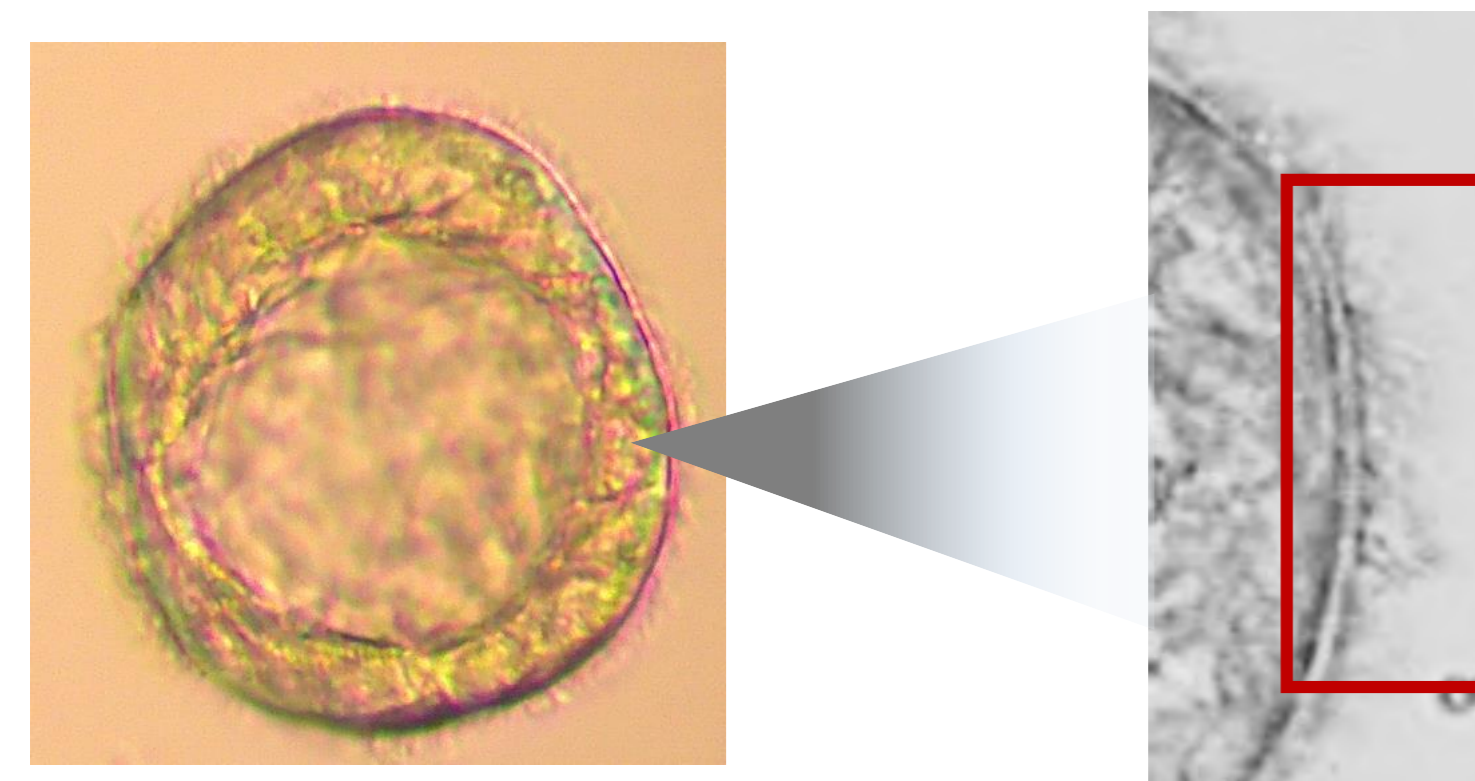
2) Organoids size

Interval of 50µM - 150 µM for healthy and asthma models

Expected mean Diameter<sup>3</sup> : 104.2 µM by day 15

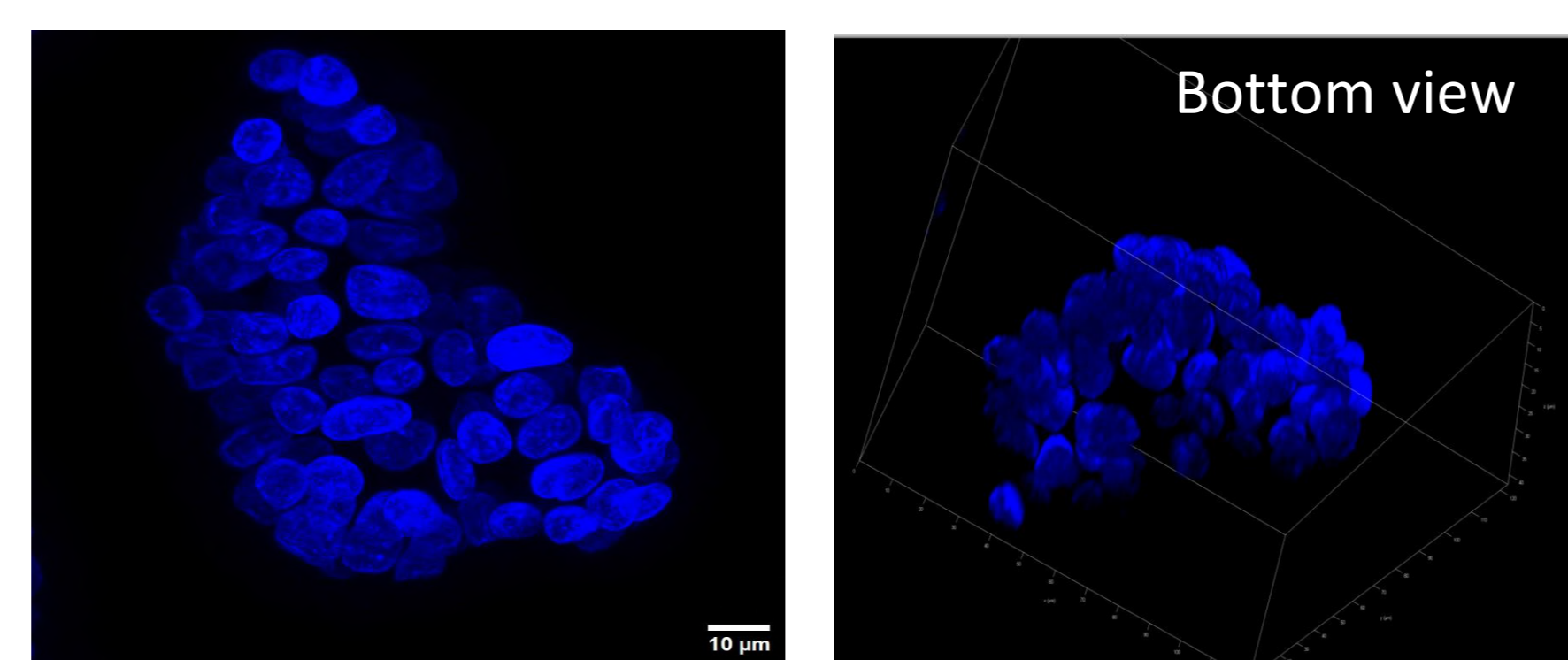
→ Results consistent with the literature

3) Ciliated and mobile organoids



Mobile organoids with cilium on external face (observed with inverted microscopy) : **inverted polarity confirmed**  
→ Organoids can be infected with RSV-A

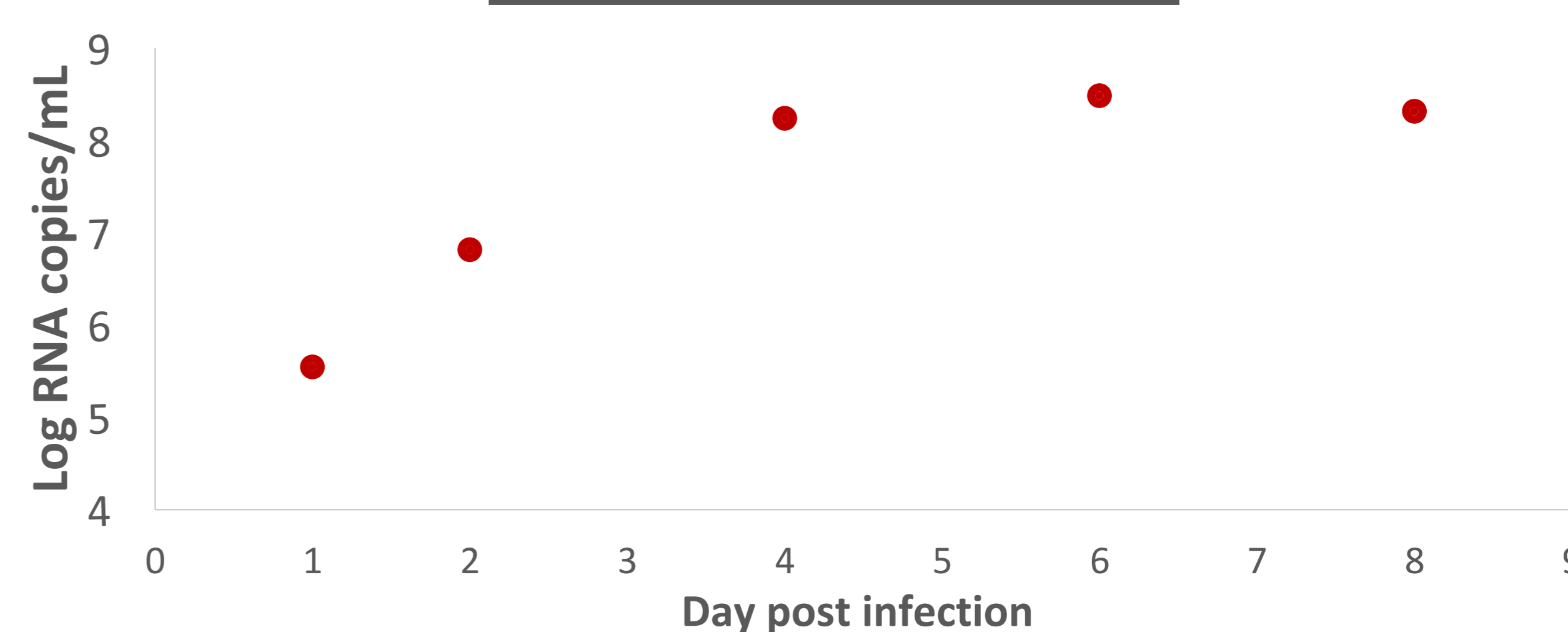
4) Immunofluorescent staining PRIMACEN collaboration



Nucleus in asthma organoid marked with DAPI : at least 61 nucleus/organoid  
→ Limit : fluorescent signal loss (DAPI hard to reveal with Stellaris)

### 2 Infection reveal : kinetics

RSV-A kinetic of infection (D1 to D8) on asthma organoids in assay 1

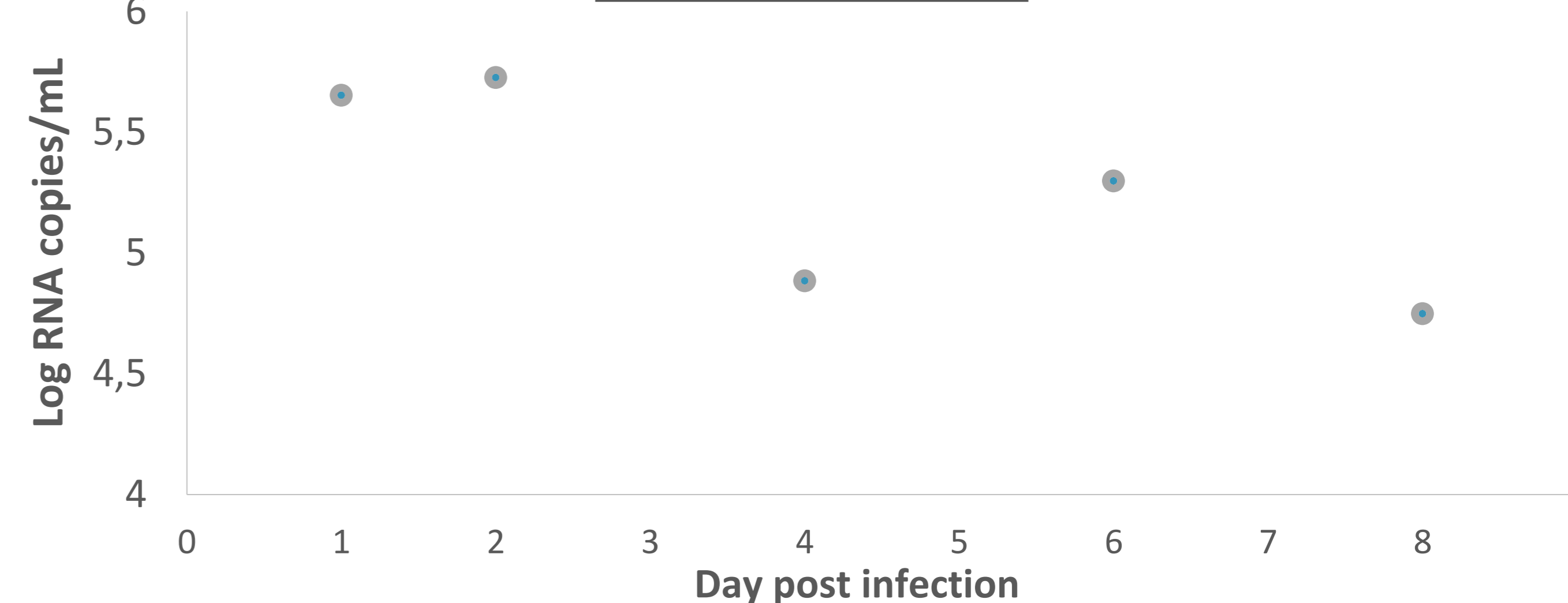


Asthma organoids :

Plateau : 1.73x10<sup>8</sup> RNA copies/mL at D4 (or 8,23 log)

→ Viral replication for asthmatic organoids, non reproducible results in assay

RSV-A kinetic infection (D1 to D8) on healthy organoids



Healthy organoids :

7.67x10<sup>4</sup> RNA copies/mL at D4 (or 4,88 log) : **NO viral replication** over the days (same results in assay 2)

## Conclusions

## Bibliography and Acknowledgements

### 1) Organoids production

- Aggregation time for healthy cells (3 days) and asthmatic cells (4 at 6 days)
- Range size (between 50µM and 150 µM) for non-fused organoids
- Mobile organoids with cilium on the external face → inverted polarity confirmed
- Immunofluorescent staining (IS) : successful with DAPI should test another microscope for others marked antibodies (Leica Thunder Tissue 3D)
- Prospect : Check asthma phenotype with IS on the 4 main bronchial cell types

### 2) Infection

- Successful asthma organoids infection with RSV : decrease of Cq over the days post infection for asthmatic cells but not reproducible → hypothesis : work on infectious dose
- Prospect : test RV infection on organoids and work on co-infection RV/RSV

<sup>1</sup> Jackson D. J. *et al* (2014). IL-33-Dependent type 2 inflammation during rhinovirus-induced asthma exacerbations in vivo.

<sup>2</sup> Petat H *et al* (2021). High Frequency of Viral Co-Detections in Acute Bronchiolitis.

<sup>3</sup> Stroulios, G. *et al* (2022). Apical-out airway organoids as a platform for studying viral infections and screening for antiviral drugs.

**Acknowledgments** : Dr Meriadeg Le Gouil for sending us a plasmid essential to quantify RSV , Dr Sophie Candon for her wise advices about immunology and Stéphane Rosset for his rapidity and help about RT-qPCR.