

The cancer-immune dialogue in the context of stress

Yuting Ma, Guido Kroemer

▶ To cite this version:

Yuting Ma, Guido Kroemer. The cancer-immune dialogue in the context of stress. Nature Reviews Immunology, 2023, 24 (4), pp.264-281. 10.1038/s41577-023-00949-8. hal-04605997

HAL Id: hal-04605997

https://hal.science/hal-04605997

Submitted on 9 Jun 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	The cancer-immune dialogue in the context of stress
2	
3	Yuting Ma ^{1*} , Guido Kroemer ^{1-5*}
4	
5	¹ National Key Laboratory of Immunity and Inflammation, Suzhou Institute of Systems Medicine,
6	Chinese Academy of Medical Sciences & Peking Union Medical College, Suzhou, Jiangsu, China
7	² Equipe labellisée par la Ligue contre le cancer, Université de Paris, Sorbonne Université, INSERM
8	U1138, Centre de Recherche des Cordeliers, Institut Universitaire de France, Paris, France
9	³ Metabolomics and Cell Biology Platforms, Gustave Roussy Cancer Campus, Villejuif, France
10	⁴ Pôle de Biologie, Hôpital Européen Georges Pompidou, AP-HP, Paris, France
11	⁵ Karolinska Institute, Department of Women's and Children's Health, Karolinska University Hospital
12	Stockholm, Sweden
13	
14	All correspondence should be addressed to: Yuting Ma (yuting_ma1984@163.com)
15	*These authors share senior authorship
16	

Abstract

Although there is little direct evidence supporting that stress affects cancer incidence, it does influence the evolution, dissemination and therapeutic outcomes of neoplasia, as shown in human epidemiological analyses and mouse models. The experience of and response to physiological and psychological stressors can trigger neurological and endocrine alterations, which subsequently influence malignant (stem) cells, stromal cells, and immune cells in the tumour microenvironment, as well as systemic factors in the tumour macroenvironment. Importantly, stress-induced neuroendocrine changes that can regulate immune responses have been gradually uncovered. A number of stress-associated immunomodulatory molecules (here termed SAIMs) can reshape natural or therapy-induced anti-tumour responses by engaging their corresponding receptors on immune cells. Moreover, stress can cause systemic or local metabolic reprogramming and change the composition of the gastrointestinal microbiota which can indirectly modulate anti-tumour immunity. Here, we explore the complex circuitries that link stress to perturbations in the cancer-immune dialogue and their implications for therapeutic approaches to cancer.

Introduction

Historically, Western culture has been wedded to the idea that body and mind are separate, often antinomic entities. Despite the rise of 'psychosomatic medicine' during the 20th century, only recently has it been considered that mental and physical health may be inseparable¹ and that psychiatric diseases have a pathophysiological substratum that reaches beyond the central nervous system².

Likewise, until a few decades ago, cancer was thought to be a cell-autonomous disease in which genetic and epigenetic instability drives cells to premalignancy (dysplasia, metaplasia) and then to overt neoplasia. Although it was known that non-transformed cells contribute to tumour progression, for instance by providing trophic support via angiogenesis³, it was only relatively recently discovered that cancer can only arise in the context of failing immunosurveillance and that efficient anticancer therapies rely on the (re)activation of anticancer immune responses⁴. Moreover, emerging evidence also supports the existence of bidirectional regulatory networks between the immune and neuroendocrine systems⁵⁻⁷. Discoveries in this field will help to elucidate the molecular basis of body-mind dialogues in the context of cancer immunosurveillance.

Both the immune and neuroendocrine systems are ubiquitous, with cell types that are part of these systems widely distributed over the body. Consequently, effector molecules produced by both systems can reach any compartment of the body^{8,9}. As a result, it is tempting to assume that neuroendocrine effectors and immunological factors might engage in a crosstalk that has evolved to maintain tissue homeostasis and to minimize or avoid the manifestation of diseases^{5,10}. Indeed, most leukocytes are in close proximity to sympathetic and/or parasympathetic nerve endings in lymphoid organs^{6,11-13}. Nerve fibres in the spleen and lymph nodes can modulate immune responses such as antibody production and inflammation^{7,14,15}. Conversely, manipulation of components of the immune system has been shown to perturb neurobiological and behavioural features in animal models^{16,17}.

Importantly, tumours are generally infiltrated by distinct leukocyte subpopulations¹⁸ and they are sometimes innervated or surrounded by nerve fibres¹⁹. In mouse prostate tumour xenografts, sympathetic and parasympathetic nerve fibres were reported to promote tumour development and dissemination, respectively²⁰. The pro-tumoral effects of sympathetic adrenergic nerve fibres were partially attributed to metabolic shifts in endothelial cells that drive local angiogenesis²¹. High densities of adrenergic and cholinergic nerves also correlated with poor clinical outcomes in patients with prostate cancer²⁰. Conversely, genetically induced stimulation of parasympathetic cholinergic nerve fibres in rodent models of breast cancer effectively blocked tumour outgrowth by downregulating the expression of immune checkpoint molecules²². Although the communication between nerves and immune cells in tumours is starting to be explored^{22,23}, it remains unknown whether stress may influence local neural remodelling and immune regulation in the tumour microenvironment (TME).

Certain environmental and lifestyle factors (such as sunbathing, smoking) have been associated with an increased risk of cancer, as they can lead to genomic damage or instability²⁴. In mice, there is also evidence that chronic stress can accelerate tumorigenesis^{25,26}. However, epidemiological studies failed to provide convincing evidence that objective stressors (such as adverse life events like the death of a spouse,

divorce, or illness) and subjective stress (such as depression or anxiety) increase cancer susceptibility in humans²⁷⁻²⁹. This contrasts with the observation that stress-related conditions such as post-traumatic stress disorder (PTDS) or anxiety show a significant association with obesogenic processes and an elevated risk of autoimmunity^{30,31}. The discrepancy between the observations in humans and mice may reflect inter-species differences, the oversimplification and artificiality of mouse models of stress and tumorigenesis, the vast heterogeneity of human cancer, as well as the low reliability of mental status evaluation in retrospective studies. It is also noteworthy that the vulnerability to stress may vary significantly between different individuals and is affected by variations in genetic risk factors³², neuroanatomic features³³, and metabolic patterns³⁴.

In this Review, we examine how stress affects anticancer immune responses. We summarize evidence showing how stress-evoked neuroendocrine factors (including steroid hormones, neurotransmitters, and neuropeptides) reshape local and systemic immune responses by engaging corresponding receptors on immune cells. We refer to these as stress-associated immunomodulatory molecules (SAIMs). We also depict an interplay between the nervous system, systemic metabolism and the gastrointestinal microbiota upon stress, as well as their effect on the cancer-immune crosstalk (Fig. 1). Based on existing evidence and emerging mechanistic insights, we share our vision of novel immunotherapeutic targets and approaches to treat cancer.

[H1] Links between stress and cancer

Understanding the connection between stress and cancer is particularly important as anxiety and depression are highly prevalent in patients with cancer³⁵. Cancer diagnosis and treatment may cause symptoms that meet the full clinical criteria of PTSD in up to ~20% of total cases and can lead to long-term alterations in neuroendocrine circuitries^{36,37}. Surgery-related stress has been shown to cause an excess release of catecholamines and prostaglandins perioperatively in patients with cancer, which can promote metastatic progression³⁸. Despite the lack of proof that stress causes cancer in humans, there is convincing epidemiological evidence that objective stressors and selfevaluated psychological distress correlate with negative cancer outcomes (progression, metastasis, recurrence, therapeutic failure, and increased mortality)³⁹⁻⁴³. Adverse life circumstances in general have been closely associated with a skewing towards proinflammatory responses and can dampen antiviral activity in circulating leukocytes. These changes have largely been attributed to glucocorticoid-related pathways and the activation of the adrenergic system^{44,45}. Moreover, gene signatures indicative of the activity of the glucocorticoid, adrenergic, dopaminergic, serotoninergic, and muscarinic systems have been closely correlated with unfavourable outcomes in patients with prostate cancer⁴⁶.

The adverse impact of stress on cancer outcomes has been replicated in mouse models, allowing to establish causality and to explore the underlying mechanisms. Repeated restraint, male domination, or unpredictable chronic mild stress (UCMS) are established procedures to induce stress in rodents (**Supplemental Box 1**). These have been shown to cause behavioural, psychological, and biological changes that promote tumour development, metastatic dissemination, and therapeutic failure in mice⁴⁷⁻⁵². Moreover, mice held in standard housing conditions are already stressed and alleviating

stress by shifting them from the hypothermic to thermoneutral conditions or placing them in an 'enriched' environment can significantly reduce tumour growth^{53,54}.

136 137 138

139

140 141

142

143

144

145

146

135

To dissect how stress can influence cancer incidence, evolution, and therapeutic outcomes, the direct effects of neuroendocrine factors on cancer cells and stromal elements have been explored. An abundant literature suggests that stress-induced glucocorticoids and catecholamines can directly regulate cell death and enhance the proliferation, therapeutic resistance, stemness traits, and dissemination of tumour cells in mouse models^{25,55-62} (**Box 1**). Also, these factors can reshape the tumour stroma by promoting angiogenesis, lymphogenesis, axonogenesis, and extracellular matrix formation^{47,49,63-67} (**Box 2**). Here, we focus on stress-induced neuroendocrine effectors, and their direct and indirect effects on immune cells and anti-cancer responses.

147

[H1] The neuroendocrine response to stress

148149150

[H2] Stress-induced neuroendocrine factors

151 152

153

154

155

156157

158

159

160

161

162

163

164

165

166

167168

169

170

171

172

The biosynthesis and secretion of neuroendocrine factors are crucial for stress-induced neuro-endocrine-immune interplay. The sympathetic adrenal-medullary (SAM) and hypothalamic-pituitary-adrenal (HPA) axes constitute two major arms neuroendocrine responses to stress. They generate the paradigmatic 'stress hormones', catecholamines and glucocorticoids, respectively. A close-to-immediate reaction to stress activates the SAM axis, stimulating the production of catecholamines (notably dopamine, noradrenaline (norepinephrine), and adrenaline (epinephrine)) by chromaffin cells of the adrenal medulla and postganglionic fibres of the sympathetic nervous system^{68,69}. Surges of adrenaline and noradrenaline can activate α- or βadrenergic receptors (ARs), which subsequently trigger phospholipase C (PLC)-protein kinase C (PKC) signalling or the cyclic adenosine monophosphate (cAMP)-protein kinase A (PKA) pathway^{70,71}. Following an instant, but transient, response via the SAM axis, a slightly delayed activation of the HPA axis initiates the cascade-like biosynthesis hypothalamic corticotropin-releasing of hormone (CRH) by neurons. adrenocorticotropic hormone (ACTH) by the hypophysis, and finally glucocorticoids (cortisol in humans or corticosterone in rodents) by the adrenal gland⁷². Elevated circulating glucocorticoids can activate the ubiquitously expressed nuclear receptor NR3C1 (also called glucocorticoid receptor, GR), which engages glucocorticoid response elements and initiates transcriptomic reprogramming⁷³. Alternatively, they can engage a high-affinity membrane receptor G-protein coupled receptor 97 (GPR97)⁷⁴ on leukocytes, which modulates the development and inflammatory response of immune cells^{75,76}.

173174175

176

177

178

179 180

181

182

183

184

Stress can stimulate the production of other neuroendocrine factors in the central nervous system. For example, serotonergic neurons in the dorsal raphe nucleus can convert L-tryptophan to serotonin (5-HT), which attenuates stress by engaging multiple postsynaptic 5-HT receptors⁷⁷. Cholinergic neurons in the nucleus basalis of Meynert can synthesize acetylcholine (ACh), which mediates the parasympathetic "rest and digest" response by activating muscarinic or nicotinic ACh receptors^{78,79}. The inhibitory neurotransmitter gamma-aminobutyric acid (GABA) is mainly generated by amygdala GABAergic interneurons, which binds to ionotropic or metabotropic GABA receptors^{80,81} and counterbalances the excitatory neurotransmitter glutamate. Histaminergic neurons in hypothalamic tuberomammillary nucleus can convert L-

histidine to histamine, which can be detected by G protein-coupled histamine receptors 82,83 . Endogenous opioid neuropeptides (known as endorphins, enkephalins, dynorphins) are synthesized by opioidergic neurons in the limbic and paralimbic regions of the brain and the spinal cord, which bind to μ , δ , or κ opioid receptors $^{84-86}$. The above-mentioned receptors have been identified not only on neurons, but also on a diverse array of immune cells 87 .

It is noteworthy that a complex regulatory network exists within each individual stress-responsive module (such as the negative and positive feedback circuits in the HPA axis⁸⁸) and among different neuroendocrine elements (as exemplified by the interplay between the SAM and HPA axes⁶⁸, as well as that between serotonergic neurons and the HPA axis⁸⁹). Mounting evidence suggests that the sources of neuroendocrine factors are not limited to the central nervous system and endocrine glands. They can be also produced locally in peripheral tissues by sympathetic or parasympathetic nerves^{7,9,14,15}, specialized endocrine cells^{90,91}, epithelial cells⁹², immune cells^{14,93}, and even gut microbiota⁹⁴ (**Box 3**). However, how stress may influence the locoregional secretion of neuroendocrine factors and their clearance is yet to be explored.

[H2] Neuroendocrine changes in tumours

In mouse xenograft models, sustained adrenergic signalling favours axonogenesis by stimulating tumour cells to produce brain-derived neurotrophic factor (BDNF)⁶⁷. In return, tumour innervation affects malignant growth and dissemination via locally released neurotransmitters^{19,95}. In mouse breast cancer models, noradrenaline released locally from sympathetic nerve terminals, rather than mildly elevated systemic adrenalin or noradrenaline, is likely to drive stress-induced tumour progression and metastasis⁹⁶. Emerging evidence from mouse models of pancreatic cancer suggest that the sympathetic and sensory nervous systems mainly foster tumour progression, whereas the parasympathetic nervous system appears to inhibit tumour growth. Neurotransmitters that account for these regulatory effects include catecholamines, substance P, and acetylcholine^{97,98}. Accordingly, sympathectomy of the prostate gland or genetic or pharmacological blockade of ARs can reduce BDNF secretion, inhibit tumour innervation and prolong survival of tumour-bearing mice^{20,99}. Indeed, a high density of sympathetic and parasympathetic nerve fibres in tumours and adjacent tissues, as well as elevated levels of intra-tumoral BDNF and noradrenaline, have been associated with poor clinical outcome in patients with cancer^{20,67}. The parasympathetic system and vagal nerves seem to be endowed with anti-tumour properties, since low heart-rate variability (indicating reduced vagal activities) and vagotomy correlate with increased cancer risk or unfavourable clinical outcomes, presumably due to acetylcholine-mediated regulation of the cancer immune contexture 100-102.

[H1] Immune regulation by SAIMs

The cellular, molecular and anatomic basis for the neuroendocrine modulation of local and systemic immunity are starting to be investigated in rodent models¹⁰³. So far, most of this work (often in the context of infection) has focussed on glucocorticoids and catecholamines. These paradigmatic SAIMs exhibit both immunosuppressive or immunostimulatory function, depending on their target cells, the duration and intensity of stress, the type of immune response, and the timing of stress relative to the course of immune response¹⁰⁴⁻¹⁰⁷. For example, in the context of viral infection, a glucocorticoid

surge induced by repeated restraint was shown to reduce the levels of CD8⁺ dendritic cells (DCs) and downregulate their cross-presentation capabilities, thereby weakening the priming of antigen-specific cytotoxic T lymphocytes (CTLs) in mice¹⁰⁸. In a mouse model of moderate stress upon dietary restriction, glucocorticoid-induced upregulation of chemokine receptor CXCR4 on memory CD8⁺T cells facilitated their homing to the bone marrow¹⁰⁹. This stress-induced preconditioning can enhance immune responses to a secondary bacterial infection¹⁰⁹. Indeed, the immunoregulatory effect of glucocorticoid is highly cell type-specific despite the ubiquitous expression of GR¹¹⁰. This can be explained by the diversified transcriptional output and post-translational modifications of GR in different cell types⁷³.

Both acute and chronic stress can affect the normal physiology of the dopaminergic system (such as dopamine metabolism, transmission, turnover, and innervation)¹¹¹ and trigger dopamine release^{112,113}. By engaging different subtypes of metabotropic G protein-coupled dopamine receptors, it can stimulate the homing of naive CD8⁺ T cells¹¹⁴, inhibit NK cell expansion and IFNy production¹¹⁵, and alleviate the activation of the NLRP3 inflammasome¹¹⁶. Surges of noradrenaline can rapidly boost antigen capture by DCs via α2-AR-mediated PI3K and ERK1/2 signalling upon acute stress¹¹⁷. However, stimulating β2-AR on LPS-treated DCs leads to preferential secretion of IL-23 over IL-12p70, therefore augmenting T_H17 response and reducing T_H1 response of CD4⁺ T cells¹¹⁸. β2-adrenergic stimulation through catecholamines can also enhance the suppressive function of regulatory T (T_{reg}) cells by upregulating the expression of cytotoxic T-lymphocyte associated protein 4 (CTLA4)¹¹⁹. Chemogenetic activation of the sympathetic nervous system or administration of AR agonists can mimic stress conditions in mice. These procedures induce vasoconstriction, reduce local blood flow and cause abrupt hypoxia, thus compromising T cell motility and immune responses¹²⁰. In response to acrophobic stress, the noradrenergic signal from splenic sympathetic nerves was shown to stimulate choline acetyltransferase (ChAT)-expressing T cells, which subsequently activated acetylcholine-responding B cells via the α9 nicotinic receptor and enhanced their production of antibodies in response to immunization¹⁴. In all the examples above, the immunomodulatory effects of neuroendocrine factors can be determined by the profile and density of their corresponding receptors on different types of immune cells^{87,121}.

[H1] SAIMs reshape tumour immune contexture

The presence of various leukocyte subpopulations in the TME can reflect either ongoing immunosurveillance or, on the contrary, tumour-promoting inflammation and immunosuppression. The density, composition, organization, and functional state of leukocyte infiltrates, the so-called 'immune contexture', has a decisive impact on cancer evolution and responses to cancer immunotherapy¹⁸. Emerging evidence demonstrates that stress can modulate local and systemic tumour immunosurveillance. For example, repeated restraint was reported to increase the susceptibility of mice to UV-induced squamous cell carcinoma and this correlated with elevated levels of circulating and tumour-infiltrating T_{reg} cells, as well as defective IFNγ production by T cells in the skin⁵⁰. Also, high levels of stress have been associated with impaired NK cell function in patients with ovarian and breast cancer^{122,123}. Hence, SAIMs and their detection by immune cells play a central role in stress-induced immunological changes in individuals with cancer.

[H2] Glucocorticoids

285

286287

288

289

290

291

292

293294

295

296

297

298

299

300

301

302

303304

305

306307

308

309 310

311

312313

314315

316

317

318

319

320

321

322323

324

325

326

327

328

329

330

331332

333

334

Although pharmacologic doses of synthetic glucocorticoids are clearly antiinflammatory and immunosuppressive, stress-induced elevation of endogenous glucocorticoids can either inhibit or potentiate immune responses, depending on the dose, timing, and duration of glucocorticoid secretion, target cells and downstream transcriptional changes 105 (Fig. 2). In rodents with metastatic tumours, surgery-induced stress or sustained exposure to a wet cage dramatically reduced the immune-stimulatory effects of a TLR9 agonist CpG-C, whereas blocking GR and ARs with antagonists enhanced the tumoricidal activity of NK cells and favoured long-term survival¹²⁴. In mice under stress from repeated social defeat, the efficacy of chemotherapies, cancer vaccines, and immune checkpoint blockade against primary or transplanted tumours was largely compromised⁵². This stress preconditioning durably elevated the glucocorticoid tonus but decreased the levels of circulating noradrenaline and 5-HT. Mechanistically, stress-induced glucocorticoid surge and GR-mediated transactivation of the transcriptional regulator TSC22D3 impaired type I IFN responses and the antigen presentation machinery in tumour-infiltrating DCs, leading to T cell dysfunction and therapeutic failure⁵². Administration of clinically-relevant doses of synthetic glucocorticoids (without causing a reduction in thymic cellularity) or transgenic overexpression of Tsc22d3 in DCs was sufficient to abolish the efficacy of chemotherapy and prophylactic tumour vaccines in non-stressed mice, whereas systemic administration of the GR antagonist mifepristone or conditional deletion of Tsc22d3 in DCs rendered mice refractory to stress-induced immunosuppression⁵². Indeed, mifepristone showed both therapeutic benefits (by reducing tumour progression) and palliative effects (by alleviating pain and increasing the expectancy and quality of life) in patients with benign, malignant, or even metastatic tumours 125-127. It is noteworthy that antidepressants, such as serotonin/noradrenaline reuptake inhibitors, can reduce the severity of major depression without affecting the survival of patients with cancer^{128,129}.

Within the TME, cells of the monocyte-macrophage lineage can also produce glucocorticoids, which cause CD8+ T cell dysfunction by engaging GR and upregulating the transcription factor TCF1. This in situ glucocorticoid secretion has been associated with immunotherapeutic failure in both preclinical models of melanoma and in patients with melanoma¹³⁰. In patients with lung cancer, single-cell RNA sequencing revealed a positive correlation between TSC22D3 expression by tumour-infiltrating DCs and its expression by peripheral blood mononuclear cells (PBMCs). Moreover, circulating cortisol levels were found to be positively correlated with TSC22D3 expression by PBMCs, as well as the mood state of patients with colorectal or lung cancer^{52,131}. High expression of *TSC22D3* has also been identified as a negative prognostic marker in patients with colorectal, lung, or gastric cancer⁵². Indeed, patients with non-small cell lung cancer who received corticosteroids had poor responses to PD1/PD-L1 blockade¹³². It remains unclear whether this was due to corticosteroid-mediated immune suppression, or simply because patients with cancer at advanced stages are more likely to receive high-doses of the corticosteroid prednisone as a palliative treatment. Although glucocorticoids can abrogate the immunotherapeutic control of peripheral tumours, they do not seem to affect the efficacy of PD1-targeted checkpoint therapy for intracranial gliomas in mice, perhaps because of its unique immune-privileged TME¹³³. In mouse models of mesothelioma, glucocorticoids were

found to promote lymphodepletion in the blood, but not in the TME. This may explain their modest but significant negative impact on the therapeutic benefits of weakly efficacious chemoimmunotherapy¹³⁴. Nevertheless, glucocorticoid treatment did not cause noticeable inhibition in the therapeutic control of mesothelioma in mice treated with highly efficacious cancer therapies¹³⁴. Therefore, the impact of glucocorticoids on tumour immunosurveillance and therapeutic outcomes can be determined by their local production and accessibility in the TME, the anatomic location of tumours, and the immunogenic properties of cancer therapies (**Fig. 2**).

[H2] Catecholamines

335

336

337

338

339

340341

342

343344

345

346

347

348

349

350351

352

353354

355

356357

358

359

360

361362

363

364

365

366

367

368

369370

371

372

373

374

375

376

377

378

379

380

381

382

383

Although catecholamines can be immunostimulatory under some circumstances 135,136, they are generally viewed as immunosuppressive 116,137,138. Aversive stimuli can promote mesolimbic dopamine release in mice and humans 112,113,139, whereas long-term exposure to psychosocial adversity may dampen the dopaminergic activity¹⁴⁰. In the context of malignancy, stress response associated with disease progression may be reflected in significantly elevated levels of plasma dopamine. Increased circulating dopamine in patients with lung cancer is sufficient to inhibit the proliferation and cytotoxicity of T cells via dopamine receptor D1 (DRD1)¹⁴¹. In a mouse model of melanoma, dopamine also exhibited immunosuppressive properties. Genetic or pharmacologic inhibition of DRD3 can boost tumour antigen cross-presentation by DCs and augment CTL responses¹⁴². Nevertheless, the beneficial effects of dopamine signalling on anti-tumour immunity have been demonstrated. For example, in a mouse model of lung cancer, liganded DRD2 on myeloid-derived suppressor cells (MDSCs) decreased their tumour infiltration¹⁴³. By acting on D1-like receptors, dopamine can also attenuate nitric oxide (NO) production by GR1+CD115+ monocytic MDSCs and reverse local immunosuppression in mouse models of lung cancer and melanoma¹⁴⁴.

The immunosuppressive effect of adrenergic signalling in lymphocytes has been well documented. It can rapidly and robustly boost the expression of the orphan nuclear receptor subfamily NR4A145, among which NR4A1 is a key mediator of T cell dysfunction¹⁴⁶. High expression of NR4A1 represses the transcription of several effector molecules, whereas genetic deletion of Nr4a1 in mice boosts immune responses against tumours¹⁴⁶. Catecholamines are very efficient at inhibiting the priming of anti-tumour CD8⁺ T cell responses, which may be explained by the fact that naïve T cells express comparatively high levels of β2-ARs¹⁴⁷. In a mouse model of Bcell lymphoma, chronic stimulation with the non-selective β-AR agonist isoprenaline largely abolished the efficacy of a tumour vaccine targeting NKT cells and immunotherapies targeting costimulatory or coinhibitory molecules. This was found to be largely due to a selective reduction in CD8⁺T cell responses¹⁴⁸. Further studies showed that sustained β-AR signalling in CD8⁺ T cells dampened their metabolic adaptation upon activation, leading to impaired glucose uptake, glycolysis, and mitochondrial function¹⁴⁹. Stress-induced catecholamine surges also favour myeloid cell-mediated immunosuppression. In mouse models of breast cancer, \(\beta 2\)-adrenergic signalling was reported to promote the M2 polarization of macrophages¹⁵⁰ and their infiltration in the primary tumour parenchyma to accelerate metastatic dissemination ¹⁵¹. It also supports the accumulation of MDSCs in the TME and upregulates the expression of arginase-1 and PD-L1¹⁵². In stressed mice with prostate tumours and patients with prostate cancer and depressive symptoms, noradrenaline-stimulated production of neuropeptide Y (NPY) was found to correlate with an increased density of CD68⁺ tumour-associated macrophages (TAMs) and local immunosuppression¹⁵³ (**Fig. 3**).

385 386 387

388

389

390

391

392

393

394

395

396

397398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

384

The genetic deletion or pharmaceutical inhibition of β2-AR (with ICI118.551 or propranolol), but not the blockade of β1-ARs (with metoprolol)¹⁵⁴, enhanced the efficacy of anti-PD1 therapy in mouse models of melanoma and breast cancer^{154,155}. Similarly, blocking adrenergic stress or β2-AR significantly improved the abscopal effect of ionizing radiation therapy by augmenting anti-tumour immunity in mice with melanoma, breast cancer, or colon cancer¹⁵⁶. B2-AR blockade and local injection of TLR2 ligands can increase the efficacy of a tumour antigen-loaded DC vaccine in a mouse model of thymoma¹⁵⁷. Recent clinical studies prove the concept that pharmacological blockade of β -ARs may offer therapeutic benefit to patients with cancer. In a triple-blind placebo-controlled clinical trial for patients with breast cancer, preoperative administration of the nonselective β-AR inhibitor propranolol reduced biomarkers associated with invasive potential, increased gene transcription indicative of M1-polarized macrophages, type 1 and type 2 conventional DCs, and boosted the intra-tumoral accumulation of CD68+ TAMs and CTLs158. In another randomized placebo-controlled phase II clinical trial for patients with breast cancer, perioperative inhibition of β-ARs (with propranolol) and cyclooxygenase-2 (COX-2, with etodolac) substantially inhibited the expression of EMT-related genes and reduced the activity of pro-metastatic and proinflammatory transcription factors¹⁵⁹. This combination therapy also enhanced the production of IL-12, IFNy and CD11a (a NK activation marker), suppressed the secretion of IL-6 and C-reactive protein, and attenuated the mobilization of CD16⁻ monocytes¹⁵⁹. In a prospective study without randomization, blinding, or a placebo control, the off-label use of propranolol at the time of cancer diagnosis appeared to lower the risk of disease recurrence in patients with melanoma¹⁶⁰. In a retrospective clinical study of patients with metastatic melanoma, pan β-AR antagonists, rather than \(\beta 1-AR \) selective blockers, seemed to prolong overall survival following immunotherapy with IL-2 plus CTLA4- and/or PD1-targeted checkpoint therapy¹⁶¹. Future studies may explore predictive biomarkers, optimize therapeutic regimens, and develop combinatory approaches to utilize β-AR antagonists in cancer treatment.

415 416

[H2] Other SAIMs

417 418 419

420

421

422

423

424

425

426

427

The impact of other SAIMs on cancer immunosurveillance has been gradually explored. For example, acute restraint stress was reported to induce tumour immune evasion by increasing the plasma levels of the neuropeptide kisspeptin and the expression of its receptor GPR54 in splenic and tumour-infiltrating T cells. Here, kisspeptin-GPR54 was shown to activate NR4A1 via ERK5 signalling and contribute to T cell dysfunction and exhaustion¹⁶². Endogenous cannabinoids (such as anandamide and 2-arachidonoylglycerol) are actively involved in stress adaptation and regulation of the HPA axis¹⁶³. It was recently discovered that the anandamide can impair the function of tumour-specific T cells through cannabinoid receptor 2 (CB2R), and high levels of plasma anandamide correlated with poor overall survival in patients with cancer¹⁶⁴.

428 429 430

431 432 In response to persistent stress, the stress-relieving activities of the parasympathetic system, vagal responses, and ACh production are often suppressed¹⁶⁵. Interestingly, these responses are positively associated with anti-cancer immune responses, which

may open up new therapeutic approaches. For example, the parasympathetic neurostimulation with genetic approaches reduced the expression of PD1 and FOXP3 on T cells, thereby stimulating IFNy production and inhibiting the progression of breast cancer in mice²². These effects can be counteracted with pirenzepine, an antagonist of cholinergic receptor muscarinic 1, indicating that agonists of this receptor might be beneficial for cancer treatment²². Reportedly, synthetic agonists of α 7-AChR display anti-inflammatory, antiproliferative and tumour suppressive properties in both mice and humans¹⁰⁰. Nicotine (an agonist of nicotinergic AChRs) can upregulate the expression of costimulatory molecules, adhesion molecules, chemokine receptors on DCs, hence facilitating the priming of CTLs and augmenting the preventive and therapeutic effects of DC-based anticancer vaccines¹⁶⁶. Moreover, subdiaphragmatic electrical stimulation designed to increase the vagal tonus in mice also upregulated the secretion of trefoil factor 2 (TFF2) by splenic memory T cells. TFF2 can activate the chemokine receptor CXCR4 and suppress cyclin D1-dependent expansion of MDSCs, and it was shown to reduce the carcinogenic potential of azomethane/dextran sodium sulfate in mouse models of inflammatory colorectal cancer¹⁶⁷ (Fig. 4). Consequently, in addition to inhibiting adrenergic/sympathetic system, the stimulation cholinergic/parasympathetic system might provide novel therapeutic opportunities against cancer. It remains to be investigated whether a combination of cholinergic activation and adrenergic inhibition might yield superior tumour control by local immune infiltrates.

[H1] SAIMs affect the tumour macroenvironment

In addition to the direct regulatory effects on immune cells, SAIMs can induce metabolic reprogramming of immune cells and other cells within the TME and throughout the body^{17,168-172}. Moreover, stress often disrupts the brain-gut axis, leading to intestinal permeabilization, microbial dysbiosis, overt inflammation, and the exposure of microbial components and metabolites¹⁷³. These stress-induced changes in the tumour macroenvironment often cause immune dysregulation and systemic aberrations in anti-tumour immune responses (**Fig. 1**).

[H2] SAIM-induced metabolic reprogramming

Stress can alter the metabolic patterns in the brain^{168,169}, in peripheral organs (such as the liver and the adipose tissue)^{170,171}, as well as in circulating and tissue-resident immune cells¹⁷. A growing body of evidence suggests that metabolic status and fluctuations of metabolites are critical determinants of the viability, differentiation, activation, and function of immune cells¹⁷⁴, which can be mechanistically explained by energy supply, macromolecule synthesis, signal transduction, and post-translational modifications¹⁷⁵.

Stress-induced glucocorticoids and catecholamines can potently mobilize glucose and lipids into the circulation, which support increased demands of energy and biomaterials of the body 176,177 . Accumulating evidence substantiates that SAIMs are closely involved in the metabolic shifts in immune cells, which further influence their responses to tumours. In mouse models of breast cancer, the β 2-AR stress pathway was found to dictate the immunosuppressive function of MDSCs and TAMs by causing a series of metabolic changes 178,179 . It augments oxidative phosphorylation, fatty acid oxidation, and arachidonic acid cycle (to generate an immunosuppressive metabolite

prostaglandin E2 (PGE2) but reduces glycolysis in tumour-infiltrating MDSCs¹⁷⁸. The immunosuppressive effect of \(\beta 2-AR \) signalling in TAMs can be strengthened by inflammatory mediators, mainly through elevating the COX-2/PGE2 system¹⁷⁹. Since metabolic cues can shape T cell fate and effector functions, stress-induced metabolic changes, such as mitochondrial dysfunction and insufficient energy supply, may lead to immune suppression and tumour escape^{180,181}. In mouse models of melanoma and colon cancer, chronic stress was found to dampen T-cell glycolysis and oxidative phosphorylation via β2-AR signalling, thus causing T cell exhaustion in the TME¹⁸². In a mouse model of sarcoma, the administration of corticosteroids decreased the frequency of low-affinity memory T cells (but not their high-affinity counterparts) by suppressing fatty acid metabolism, leading to a compromised efficacy of immune checkpoint blockade¹⁸³. Chronic stress-induced adrenaline was reported to upregulate lactate dehydrogenase A (LDHA) and enhance glycolysis in mouse and human mammary tumours⁶¹. The upregulation of LDHA and elevated levels of lactic acid in the TME favoured immune escape and correlated with poor survival in patients with melanoma and in mouse models of melanoma¹⁸⁴. This can be linked to the reduced expression of nuclear factor of activated T cells (NFAT, a key regulator of T-cell function) and hampered production of IFNy in T cells and NK cells¹⁸⁴. In response to SAIMs such as glucocorticoids, catecholamines, and 5-HT, neutrophils can rapidly release the inflammatory cytokines S100A8 and S100A9185, which are known to mediate immune evasion in cancer¹⁸⁶. Stress-activated neutrophils also generate oxidized lipids and promote the recurrence of dormant lung and ovarian cancer in mice by stimulating the fibroblast growth factor pathway¹⁸⁵. Overall, these findings demonstrate that metabolic changes induced by SAIMs, both on the systemic and on the cellular level, have a negative effect on immune responses against cancer.

[H2] Stress-induced protein modifications

Metabolic reprogramming can modulate immune responses by generating bioactive metabolites that facilitate the post-translational modifications (PTMs) of proteins¹⁸⁷. Several studies showed that stress-induced catecholamines can enhance glycolysis and lactate production^{61,188}. In mouse models of lung cancer and melanoma, the accumulation of exogenous or endogenous lactate in TAMs has been validated¹⁸⁹. This is consistent with a high level of lactylation of histone lysine residues in TAMs, which contributes to their epigenetic switch to the M2-polarized state¹⁸⁹.

The immunoregulatory potential of other metabolite-dependent post-translational modifications have been documented¹⁸⁷, but discoveries regarding their association with stress and SAIMs have just begun to emerge. The addition of O-linked β-N-acetylglucosamine (O-GlcNAc) to serine or threonine residues of target proteins is mediated by O-GlcNAc transferase¹⁹⁰. O-GlcNAcylation can promote the development, differentiation, and activation of T and B cells, support the activation of neutrophils, regulate the inflammatory responses of macrophages, and impede the cytotoxicity of NK cells¹⁹⁰. It is noteworthy that stress-induced release of cortisol and catecholamines often cause hyperglycemia and immune dysregulation^{191,192}. Hyperglycemia was reported to augment O-GlcNAcylation in TAMs, which favours their M2-like polarization and supports the immune evasion of colorectal cancer in mice and humans¹⁷². The covalent attachment of palmitic acid to cysteine residues of target proteins is mainly catalysed by zinc finger DHHC domain-containing (ZDHHC) palmitoyl acyltransferases¹⁹³. These enzymes are expressed in a region- and cell-type

specific pattern in the mouse brain¹⁹⁴, and chronic exposure to restraint stress can significantly alter the synaptic palmitoyl-proteome¹⁹⁵. Interestingly, ZDHHC3-mediated palmitoylation can stabilize PD-L1 by blocking its ubiquitination and degradation¹⁹⁶. Pharmacological inhibition or genetic silencing of ZDHHC3 can decrease PD-L1 expression on tumour cells and enhance anti-tumour T-cell immunity¹⁹⁶. Therefore, stress-induced fluctuations of metabolites and PTMs may enable a fine-tuning of immune responses against cancer.

[H2] Stress-induced microbial alterations

Microbial communities in the gut have been recognized as key regulators of the bidirectional interaction between the enteric and the central nervous system¹⁹⁷. By generating various metabolic and neurochemical factors and regulating the production of inflammatory cytokines, they can build up connections between intestinal events and emotional and cognitive responses in the brain¹⁹⁸. It is noteworthy that stress-induced proinflammatory cytokines and neuroendocrine factors (such as catecholamines, histamine, 5-HT, and CRH) can disrupt the gut-brain axis, cause a leaky gut, and ultimately lead to microbial imbalance^{173,199,200}. Indeed, faecal samples harvested from patients with major depressive disorder contain more *Enterobacteriaceae* and *Allistipes*, but less *Faecalibacterium*^{201,202}, as compared to those from healthy individuals. In the GI tract of patients with generalized anxiety disorder, the prevalence of *Faecalibacterium*, *Eubacterium rectale*, *Lachnospira*, *Butyricicoccus*, and *Sutterella* were significantly reduced²⁰².

A balanced microbiota is vital for immune homeostasis, whereas stress-induced dysbiosis often leads to immune dysregulation^{2,203}. In a mouse model of inflammatory bowel disease, chronic stress can induce GI dysbiosis and augment local inflammation in the colonic lamina propria, thus facilitating the development of colitis²⁰⁴. GI dysbiosis allows bacteria to invade the mucosal barriers and migrate into lymphoid nodes²⁰⁵ and other tissues²⁰⁶, leading to the exposure of pathogen-associated molecular patterns (PAMPs)²⁰³ to the immune system. Thus, bacterial DNA, lipopolysaccharide, double-stranded RNA, polysaccharide A, peptidoglycan motifs, and bacterial cell wallderived muramyl dipeptide can modulate local and systemic immune responses by engaging corresponding pattern recognition receptors on immune cells^{207,208}. Moreover, microbial and tumour-associated antigens may share cross-reactive T cell epitopes^{209,210}, and microbial peptides were reported to boost anti-tumour immunity via antigenic mimicry²¹¹. Importantly, probiotic supplementation with *Bifidobacterium longum*, Lactobacillus farciminis, and Lactobacillus helveticus can help to re-establish a balanced microflora, attenuate stress-induced neuroendocrine circuits, diminish intestinal permeability, and alleviate inflammation in patients with endotoxemia^{212,213}. Dysbiosis of the microflora has been shown to compromise anticancer immunity and the therapeutic benefits of cancer immunotherapy²¹⁴⁻²¹⁶. Conversely, a high abundance of commensal bacteria and probiotics is associated with efficient immunotherapeutic responses in patients with melanoma²¹⁶, supporting the idea that bacteria that promote general health also promote anticancer immunity²¹⁷.

GI microflora-derived metabolites also exhibit immunoregulatory properties^{218,219}. Notably, indole metabolites, tryptophan derivatives, secondary bile acids, and short-chain fatty acids (SCFAs) from the microbiota can modulate local or systemic immune responses by engaging pregnane X receptor (PXR), aryl hydrocarbon receptor (AhR),

farnesoid X receptor (FXR), Takeda G protein-coupled receptor 5 (TGR5), and orphan G protein coupled receptors (GPR41 and GPR43), respectively²²⁰. In the context of cancer, the activation of these receptors on immune cells or tumour cells often leads to immunosuppression. For example, TGR5 signalling in TAMs can promote their M2polarization, which subsequently hamper CTL response against mouse lung cancer²²¹. High expression of FXR in mouse lung cancer cells also creates an immunosuppressive microenvironment and causes CTL inactivation and exhaustion²²². AhR signalling in mouse oral squamous cell carcinoma can upregulate the expression of immune checkpoints, whereas genetic deletion of AhR triggers long-lasting anti-tumour immunity and facilitates tumour regression²²³. Stress-associated enteric dysbiosis can reduce the abundance of SCFA-producing bacteria in patients with generalized anxiety disorder²⁰². In mice, it was shown that SCFAs contribute to the development of CD8⁺ memory T cells²²⁴. Of note, in patients with solid tumours, the outcome of PD1-targeted immune checkpoint inhibitors correlated with the abundance of SCFAs^{225,226}. Altogether, stress-induced metabolic changes in the gut microbiota may have oncoimmunological impacts, which lack in-depth exploration and merit further investigation.

Microbiome-associated biosynthesis of neuroactive factors in the gut and their impact on tumour immunosurveillance are being gradually unravelled. Intestinal flora-derived SCFAs can promote 5-HT production by enterochromaffin cells in the gut epithelium²¹³, and indigenous spore-forming bacteria have been identified as important modulators of this process²¹². Indeed, gut 5-HT accounts for a substantial proportion of the total pool of 5-HT in the body²¹². Depletion of 5-HT cargo or inhibition of 5-HT release can enhance the accumulation and effector function of the CD8+ T cells, decrease PD-L1 expression, and delay the outgrowth of pancreatic and colorectal tumours in mice²²⁷. Dysbiosis and stress-related disorders can be alleviated by some which promote the generation of neurotransmitters (such as endocannabinoids produced by Lactobacillus casei DG^{94}) or influence the expression of their receptors (such as Lactobacillus rhamnosus-dependent regulation of GABA receptors in the brain via the vagus nerve²²⁸). By engaging CB1R or CB2R on immune cells, cannabinoids can reshape the immune landscape of the TME to positively or negatively regulate tumour growth²²⁹. In mouse models of lung and colon cancer, GABA signalling can promote local immunosuppression by excluding T cells from the TME²³⁰ and eliciting IL-10-producing anti-inflammatory TAMs²³¹. It was reported that Staphylococcus pseudintermedius can generate dopamine and 5-HT²³², while Firmicutes (such as Clostridium sporogenes and Ruminococcus gnavus) can decarboxylate tryptophan to produce β-arylamine neurotransmitter tryptamine²³³. The opportunistic pathogen Enterococcus faecalis can generate tyramine (the invertebrate equivalent of noradrenaline²³⁴) via tyrosine decarboxylases²³⁵. Accumulated tyramine can engage trace amine-associated receptors (TAARs) and induce the release of catecholamine stores from nerve terminals in porcines²³⁶. Altogether, we speculate that targeting stress-induced dysbiosis, metabolic or neuroendocrine changes, and related immunological abnormalities may provide novel therapeutic strategies to improve tumour immunosurveillance in individuals under unresolved stress.

Conclusion and perspective

583

584

585

586

587

588

589

590

591

592

593

594

595

596

597

598 599

600

601

602

603

604

605

606

607 608

609

610

611

612

613

614

615

616

617

618

619

620

621

622

623

624

625

626

627 628

629 630

631

632

As discussed above, psychosocial stress affects cancer immunosurveillance. The regulatory network underpinning these interactions is composed of central or peripheral

SAIMs, metabolic changes and bioactive metabolites, dysbiosis and specific microbial components, as well as different cell types that either generate or respond to these factors. There are various known and yet unknown receptors or sensors of SAIMs, and many of the downstream signalling pathways that affect transcriptomic profiles, metabolomic patterns, and PTMs are yet to be elucidated. The disentanglement of this ecosystem has barely begun. Progress in the area will require interdisciplinary collaborations among scientists working across diverse areas, such as neurology, psychology, psychiatry, oncology, immunology, endocrinology, metabolism, and microbiology. Moreover, a scaling up of spatially and temporarily resolved 'omics' approaches as well as the development of reductionist in vitro and in vivo models will be necessary, using single cell- and spatially-resolved transcriptomics, proteomics and metabolomics, live-cell or live-tissue imaging, organoid and 3D culturing, microfluidics and other biophysical methods. Such models will need to reconstruct the complexity of neoplasia by regarding tumours as organized assemblies of different cell types that closely communicate with sympathetic, parasympathetic, and sensory nerve terminations, as well as the microbiota both in the GI tract and in the tumour.

648649650

651

652

653

654 655

656

657

658

659

633

634

635

636

637

638 639

640

641642

643

644

645

646

647

However, it should be noted that cancer directly affects the neuroendocrine and immune systems as well. This is due to the usually traumatic experience of cancer diagnosis, often invasive operations, and major side effects of treatments, but it also concerns the pathophysiology of the disease itself. Thus, advanced cancer with its corollaries (such as pain, inflammation, cachexia) is usually coupled to the 'sickness response', involving a radical rewiring of neuroendocrine circuitries, metabolic switches from anabolism to catabolism, as well as behavioural alterations (insomnia, social isolation) and immune abnormalites¹⁰. This stereotyped response activates an immune-neuroendocrine feedback loop, in which inflammatory mediators produced in peripheral tissues can reach specific areas in the hypothalamus to induce similar changes as those induced by stress.

660661662

663

664

665

666

667

668

669

670

671

672

673

674

675

676

677

678

679

680

681 682

Since neurotransmitters and stress hormones can exhibit different local and systemic effects, pharmacological intervention targeting the synthesis of catecholamines or cortisol steroidogenesis are not feasible to treat stress-related disorders due to their excessive side effects. In contrast, specific antagonists of subclasses of ARs are in medical use and are generally well tolerated²³⁷. Although it appears obvious that, given their immunosuppressive effects, synthetic glucocorticoids should be avoided for the routine management of patients with cancer, it is certainly premature to broadly recommend GR antagonists for cancer treatment. Ongoing clinical trials are testing the anti-cancer potential of the GR antagonist mifepristone as monotherapy or in combination with chemotherapy or immunotherapy for breast cancer^{125,126}. A similar caveat may apply to surgical, electrical, or pharmacological sympathectomy and parasympathetic stimulation. Given the importance of these systems in homeostatic regulation, clinical trials need to be carefully planned to avoid deleterious side effects for such neuroendocrine interventions. Strictly localized approaches (such as intratumoral manipulations of SAIM receptors with antagonists or agonists, sympathetic denervation, or parasympathetic stimulation), used alone or combined with immunotherapies, may be feasible strategies to elicit anti-tumour immunity with an abscopal effect on metastases²³⁸. Traditional approaches of stress management and psychotropic medications have been put into use for a long time. However, their impact on tumour control and antitumour immune response needs further investigation (Box 4).

Here we have aimed to condense limited knowledge on physiological and psychological stress, neuronal/endocrine circuitries, and immuno-oncology into a coherent paradigm involving SAIMs that affect the cancer-immune crosstalk. Rewired metabolic roadmaps, metabolite-mediated cell-cell communication, and microbial dynamics are also emerging as crucial modulators of behavioural patterns and the cancer-immune dialogue^{239,240}. Although this endeavour already appears very ambitious, it might underestimate the impact of other so-far unknown factors. Hence, future research needs to address these ever-expanding complexities to understand the inseparable link between mind and body.

Acknowledgements

683

684

685

686

687

688

689

690

691 692

693

694 695

696

697 698

699

700701

702

703704

705

706 707

708

709710

711712713

714

715

716

717

718

719

720

721 722

723 724

725

726

727

728 729

Y.M. is supported by Science and Technology Innovation 2030 Major Project (STI2030-Major Projects 2022ZD0205700), Natural Science Foundation of China (NSFC grant No. 81972701), CAMS Innovation Fund for Medical Sciences (CIFMS; 2021-I2M-1-074, 2022-I2M-2-004), National special support plan for high-level talents, Suzhou Municipal Key Laboratory (SZS2023005), and Innovative and Entrepreneurial Team Program (Jiangsu Province). G.K. is supported by the Ligue contre le Cancer (équipe labellisée); Agence National de la Recherche (ANR)-Projets blancs; AMMICa US23/CNRS UMS3655; Association pour la recherche sur le cancer (ARC); Cancéropôle Ile-de-France; Fondation pour la Recherche Médicale (FRM); Equipex Onco-Pheno-Screen; European Joint Programme on Rare Diseases; the European Union Horizon 2020 Projects Oncobiome and Crimson; Institut National du Cancer (INCa); Institut Universitaire de France; LabEx Immuno-Oncology (ANR-18-IDEX-0001); High-end Foreign Expert Program in China (GDW20171100085); the RHU Immunolife; Seerave Foundation; SIRIC Stratified Oncology Cell DNA Repair and Tumor Immune Elimination (SOCRATE); and SIRIC Cancer Research and Personalized Medicine (CARPEM).

References

- Braslow, J. T. & Marder, S. R. History of Psychopharmacology. *Annu Rev Clin Psychol* **15**, 25-50, doi:10.1146/annurev-clinpsy-050718-095514 (2019).
- 2 Cruz-Pereira, J. S. *et al.* Depression's Unholy Trinity: Dysregulated Stress, Immunity, and the Microbiome. *Annu Rev Psychol* **71**, 49-78, doi:10.1146/annurev-psych-122216-011613 (2020).
- 3 Hanahan, D. & Weinberg, R. A. The hallmarks of cancer. *Cell* **100**, 57-70, doi:10.1016/s0092-8674(00)81683-9 (2000).
- This review elegantly summarizes current knowledge about the links among depression pathogenesis, immunity and microbiome, as well as possible therapeutic interventions.
- 4 Hanahan, D. & Weinberg, R. A. Hallmarks of cancer: the next generation. *Cell* **144**, 646-674, doi:10.1016/j.cell.2011.02.013 (2011).
- Veiga-Fernandes, H. & Artis, D. Neuronal-immune system cross-talk in homeostasis. *Science* **359**, 1465-1466, doi:10.1126/science.aap9598 (2018).
- Godinho-Silva, C., Cardoso, F. & Veiga-Fernandes, H. Neuro-Immune Cell Units: A New Paradigm in Physiology. *Annu Rev Immunol* **37**, 19-46, doi:10.1146/annurev-immunol-042718-041812 (2019).
- Huang, S. *et al.* Lymph nodes are innervated by a unique population of sensory neurons with immunomodulatory potential. *Cell* **184**, 441-459 e425, doi:10.1016/j.cell.2020.11.028 (2021).
- This study discovered that lymph node-innervating sensory neurons can interact with several predicted cell types and change their transcriptome. And the unexpected sensory neuro-immune circuit exhibits the capacity to monitor the inflammatory state in the lymph node.
- 735 8 Kabata, H. & Artis, D. Neuro-immune crosstalk and allergic inflammation. J Clin Invest 130,

736 1475-1482, doi:10.1172/JCI124609 (2019).

751

752

755

756

757

763

764 765

766

767

768

769

770

774

775

776

777

778

- 737 9 Chen, C.-S., Barnoud, C. & Scheiermann, C. Peripheral neurotransmitters in the immune system.

 738 *Current Opinion in Physiology* **19**, 73-79, doi:https://doi.org/10.1016/j.cophys.2020.09.009

 739 (2021).
- Wang, A., Luan, H. H. & Medzhitov, R. An evolutionary perspective on immunometabolism. Science 363, doi:10.1126/science.aar3932 (2019).
- Guyot, M. *et al.* Apical splenic nerve electrical stimulation discloses an anti-inflammatory pathway relying on adrenergic and nicotinic receptors in myeloid cells. *Brain Behav Immun* **80**, 238-246, doi:10.1016/j.bbi.2019.03.015 (2019).
- 745 12 Al-Shalan, H. A. M., Hu, D., Nicholls, P. K., Greene, W. K. & Ma, B. Immunofluorescent characterization of innervation and nerve-immune cell neighborhood in mouse thymus. *Cell Tissue Res* **378**, 239-254, doi:10.1007/s00441-019-03052-4 (2019).
- Jung, W. C., Levesque, J. P. & Ruitenberg, M. J. It takes nerve to fight back: The significance of neural innervation of the bone marrow and spleen for immune function. *Semin Cell Dev Biol* **61**, 60-70, doi:10.1016/j.semcdb.2016.08.010 (2017).
 - Zhang, X. et al. Brain control of humoral immune responses amenable to behavioural modulation. *Nature* **581**, 204-208, doi:10.1038/s41586-020-2235-7 (2020).
- This well-designed study identifies a specific brain–spleen neural connection in mice that enhances humoral responses in response to an elevated platform regimen.
 - Rosas-Ballina, M. *et al.* Splenic nerve is required for cholinergic antiinflammatory pathway control of TNF in endotoxemia. *Proc Natl Acad Sci U S A* **105**, 11008-11013, doi:10.1073/pnas.0803237105 (2008).
- 758 16 Miyajima, M. *et al.* Metabolic shift induced by systemic activation of T cells in PD-1-deficient mice perturbs brain monoamines and emotional behavior. *Nat Immunol* **18**, 1342-1352, doi:10.1038/ni.3867 (2017).
- This paper reveals how a key immune checkpoint molecule PD-1 is involved in regulating systemic metabolism, the biosynthesis of neurotransmitters, and behaviours.
 - Fan, K. Q. *et al.* Stress-Induced Metabolic Disorder in Peripheral CD4(+) T Cells Leads to Anxiety-like Behavior. *Cell* **179**, 864-879 e819, doi:10.1016/j.cell.2019.10.001 (2019).
 - This interesting study suggests that stress-induced abnormal mitochondrial fission and purine synthesis in CD4⁺T cells influence oligodendrocytes in the amygdala, which is a prerequisite for the onset of anxiety.
 - Fridman, W. H., Zitvogel, L., Sautes-Fridman, C. & Kroemer, G. The immune contexture in cancer prognosis and treatment. *Nat Rev Clin Oncol* **14**, 717-734, doi:10.1038/nrclinonc.2017.101 (2017).
- 771 19 Faulkner, S., Jobling, P., March, B., Jiang, C. C. & Hondermarck, H. Tumor Neurobiology and the War of Nerves in Cancer. *Cancer Discov* **9**, 702-710, doi:10.1158/2159-8290.CD-18-1398 (2019).
 - This comprehensive review summarizes recent progresses that deal with the potential link between tumour-induced innervation with in the TME and cancer initiation, progression, and metastasis.
 - Magnon, C. *et al.* Autonomic nerve development contributes to prostate cancer progression. *Science* **341**, 1236361, doi:10.1126/science.1236361 (2013).
 - Zahalka, A. H. *et al.* Adrenergic nerves activate an angio-metabolic switch in prostate cancer. *Science* **358**, 321-326, doi:10.1126/science.aah5072 (2017).
- Kamiya, A. *et al.* Genetic manipulation of autonomic nerve fiber innervation and activity and its effect on breast cancer progression. *Nat Neurosci* **22**, 1289-1305, doi:10.1038/s41593-019-0430-3 (2019).
- Cervantes-Villagrana, R. D., Albores-Garcia, D., Cervantes-Villagrana, A. R. & Garcia-Acevez, S. J. Tumor-induced neurogenesis and immune evasion as targets of innovative anti-cancer therapies. *Signal Transduct Target Ther* **5**, 99, doi:10.1038/s41392-020-0205-z (2020).
- 786 24 Wei, E. K., Wolin, K. Y. & Colditz, G. A. Time course of risk factors in cancer etiology and progression. *J Clin Oncol* **28**, 4052-4057, doi:10.1200/JCO.2009.26.9324 (2010).
- Feng, Z. *et al.* Chronic restraint stress attenuates p53 function and promotes tumorigenesis. *Proc Natl Acad Sci U S A* **109**, 7013-7018, doi:10.1073/pnas.1203930109 (2012).
- Jang, H. J., Boo, H. J., Lee, H. J., Min, H. Y. & Lee, H. Y. Chronic Stress Facilitates Lung Tumorigenesis by Promoting Exocytosis of IGF2 in Lung Epithelial Cells. *Cancer Res* **76**, 6607-6619, doi:10.1158/0008-5472.CAN-16-0990 (2016).
- Schoemaker, M. J. *et al.* Psychological stress, adverse life events and breast cancer incidence: a cohort investigation in 106,000 women in the United Kingdom. *Breast Cancer Res* **18**, 72, doi:10.1186/s13058-016-0733-1 (2016).

- Wang, Y. H. *et al.* Depression and anxiety in relation to cancer incidence and mortality: a systematic review and meta-analysis of cohort studies. *Mol Psychiatry* **25**, 1487-1499, doi:10.1038/s41380-019-0595-x (2020).
- A systematic review and meta-analysis containing 51 eligible cohort studies involving 2,611,907 participants, which tested the association between "anxiety and depression" and the risk of cancer incidence, cancer-specific mortality, and all-cause mortality in cancer patients.
- 802 29 Butow, P. et al. Does stress increase risk of breast cancer? A 15-year prospective study. 803 Psychooncology 27, 1908-1914, doi:10.1002/pon.4740 (2018).
- 804 30 Tomiyama, A. J. Stress and Obesity. *Annu Rev Psychol* **70**, 703-718, doi:10.1146/annurev-psych-010418-102936 (2019).
- Song, H. *et al.* Association of Stress-Related Disorders With Subsequent Autoimmune Disease. *JAMA* **319**, 2388-2400, doi:10.1001/jama.2018.7028 (2018).
- Buchanan, T. W. & Lovallo, W. R. The role of genetics in stress effects on health and addiction. Curr Opin Psychol 27, 72-76, doi:10.1016/j.copsyc.2018.09.005 (2019).
- Anacker, C. *et al.* Neuroanatomic Differences Associated With Stress Susceptibility and Resilience. *Biol Psychiatry* **79**, 840-849, doi:10.1016/j.biopsych.2015.08.009 (2016).
- Misiewicz, Z. *et al.* Multi-omics analysis identifies mitochondrial pathways associated with anxiety-related behavior. *PLoS Genet* **15**, e1008358, doi:10.1371/journal.pgen.1008358 (2019).
- Mitchell, A. J. *et al.* Prevalence of depression, anxiety, and adjustment disorder in oncological, haematological, and palliative-care settings: a meta-analysis of 94 interview-based studies. *Lancet Oncol* **12**, 160-174, doi:10.1016/S1470-2045(11)70002-X (2011).
- 817 36 Cordova, M. J., Riba, M. B. & Spiegel, D. Post-traumatic stress disorder and cancer. *Lancet Psychiatry* **4**, 330-338, doi:10.1016/S2215-0366(17)30014-7 (2017).
- Mehnert, A. & Koch, U. Prevalence of acute and post-traumatic stress disorder and comorbid mental disorders in breast cancer patients during primary cancer care: a prospective study.

 Psychooncology 16, 181-188, doi:10.1002/pon.1057 (2007).
- Horowitz, M., Neeman, E., Sharon, E. & Ben-Eliyahu, S. Exploiting the critical perioperative period to improve long-term cancer outcomes. *Nat Rev Clin Oncol* **12**, 213-226, doi:10.1038/nrclinonc.2014.224 (2015).
- Batty, G. D., Russ, T. C., Stamatakis, E. & Kivimaki, M. Psychological distress in relation to site specific cancer mortality: pooling of unpublished data from 16 prospective cohort studies. BMJ 356, j108, doi:10.1136/bmj.j108 (2017).
- 828 40 Palesh, O. *et al.* Stress history and breast cancer recurrence. *J Psychosom Res* **63**, 233-239, doi:10.1016/j.jpsychores.2007.05.012 (2007).
- Wang, X. *et al.* Prognostic value of depression and anxiety on breast cancer recurrence and mortality: a systematic review and meta-analysis of 282,203 patients. *Mol Psychiatry* **25**, 3186-3197, doi:10.1038/s41380-020-00865-6 (2020).
- 833 42 Moreno-Smith, M., Lutgendorf, S. K. & Sood, A. K. Impact of stress on cancer metastasis. 834 *Future Oncol* **6**, 1863-1881, doi:10.2217/fon.10.142 (2010).
- Chida, Y., Hamer, M., Wardle, J. & Steptoe, A. Do stress-related psychosocial factors contribute to cancer incidence and survival? *Nat Clin Pract Oncol* **5**, 466-475, doi:10.1038/ncponc1134 (2008).
- Powell, N. D. *et al.* Social stress up-regulates inflammatory gene expression in the leukocyte transcriptome via beta-adrenergic induction of myelopoiesis. *Proc Natl Acad Sci U S A* **110**, 16574-16579, doi:10.1073/pnas.1310655110 (2013).
- Irwin, M. R. & Cole, S. W. Reciprocal regulation of the neural and innate immune systems. *Nat Rev Immunol* 11, 625-632, doi:10.1038/nri3042 (2011).
- Lu, D. *et al.* Stress-Related Signaling Pathways in Lethal and Nonlethal Prostate Cancer. *Clin Cancer Res* **22**, 765-772, doi:10.1158/1078-0432.CCR-15-0101 (2016).
- Thaker, P. H. *et al.* Chronic stress promotes tumor growth and angiogenesis in a mouse model of ovarian carcinoma. *Nat Med* **12**, 939-944, doi:10.1038/nm1447 (2006).
- Zhang, X. *et al.* Chronic stress promotes gastric cancer progression and metastasis: an essential role for ADRB2. *Cell Death Dis* **10**, 788, doi:10.1038/s41419-019-2030-2 (2019).
- Le, C. P. *et al.* Chronic stress in mice remodels lymph vasculature to promote tumour cell dissemination. *Nat Commun* **7**, 10634, doi:10.1038/ncomms10634 (2016).
- Saul, A. N. *et al.* Chronic stress and susceptibility to skin cancer. *J Natl Cancer Inst* **97**, 1760-1767, doi:10.1093/jnci/dji401 (2005).
- Ben-Eliyahu, S., Page, G. G., Yirmiya, R. & Shakhar, G. Evidence that stress and surgical interventions promote tumor development by suppressing natural killer cell activity. *Int J Cancer* **80**, 880-888, doi:10.1002/(sici)1097-0215(19990315)80:6<880::aid-ijc14>3.0.co;2-y

- 856 (1999).
- 857 52 Yang, H. *et al.* Stress-glucocorticoid-TSC22D3 axis compromises therapy-induced antitumor immunity. *Nat Med* **25**, 1428-1441, doi:10.1038/s41591-019-0566-4 (2019).
- This study provides strong evidence that psychological stress compromises the efficacy of immunedependent cancer therapies, by elevating the endogenous glucocorticoid tonus and stimulating
 TSC22D3 expression in tumour-infiltrating DC.
- Kokolus, K. M. *et al.* Baseline tumor growth and immune control in laboratory mice are significantly influenced by subthermoneutral housing temperature. *Proc Natl Acad Sci U S A* **110**, 20176-20181, doi:10.1073/pnas.1304291110 (2013).
- Cao, L. *et al.* Environmental and genetic activation of a brain-adipocyte BDNF/leptin axis causes cancer remission and inhibition. *Cell* **142**, 52-64, doi:10.1016/j.cell.2010.05.029 (2010).
- He, L. *et al.* Glucocorticoid Receptor Signaling Activates TEAD4 to Promote Breast Cancer Progression. *Cancer Res* **79**, 4399-4411, doi:10.1158/0008-5472.CAN-19-0012 (2019).
- 869 56 Obradovic, M. M. S. *et al.* Glucocorticoids promote breast cancer metastasis. *Nature* **567**, 540-870 544, doi:10.1038/s41586-019-1019-4 (2019).
- Melhem, A. *et al.* Administration of glucocorticoids to ovarian cancer patients is associated with expression of the anti-apoptotic genes SGK1 and MKP1/DUSP1 in ovarian tissues. *Clin Cancer Res* **15**, 3196-3204, doi:10.1158/1078-0432.CCR-08-2131 (2009).
- Petrella, A. *et al.* Dexamethasone inhibits TRAIL-induced apoptosis of thyroid cancer cells via Bcl-xL induction. *Eur J Cancer* **42**, 3287-3293, doi:10.1016/j.ejca.2006.07.018 (2006).
- Sorrentino, G. *et al.* Glucocorticoid receptor signalling activates YAP in breast cancer. *Nat Commun* **8**, 14073, doi:10.1038/ncomms14073 (2017).
- Flaherty, R. L. *et al.* Glucocorticoids induce production of reactive oxygen species/reactive nitrogen species and DNA damage through an iNOS mediated pathway in breast cancer. *Breast Cancer Res* **19**, 35, doi:10.1186/s13058-017-0823-8 (2017).
- Cui, B. *et al.* Stress-induced epinephrine enhances lactate dehydrogenase A and promotes breast cancer stem-like cells. *J Clin Invest* **129**, 1030-1046, doi:10.1172/JCI121685 (2019).
- Sastry, K. S. *et al.* Epinephrine protects cancer cells from apoptosis via activation of cAMP-dependent protein kinase and BAD phosphorylation. *J Biol Chem* **282**, 14094-14100, doi:10.1074/jbc.M611370200 (2007).
- Nagaraja, A. S. *et al.* Adrenergic-mediated increases in INHBA drive CAF phenotype and collagens. *JCI Insight* **2**, doi:10.1172/jci.insight.93076 (2017).
- Yang, E. V. *et al.* Norepinephrine up-regulates the expression of vascular endothelial growth factor, matrix metalloproteinase (MMP)-2, and MMP-9 in nasopharyngeal carcinoma tumor cells. *Cancer Res* **66**, 10357-10364, doi:10.1158/0008-5472.CAN-06-2496 (2006).
- 891 65 Sood, A. K. *et al.* Stress hormone-mediated invasion of ovarian cancer cells. *Clin Cancer Res* 12, 369-375, doi:10.1158/1078-0432.CCR-05-1698 (2006).
- Kilpatrick, L. E. *et al.* Complex Formation between VEGFR2 and the beta2-Adrenoceptor. *Cell Chem Biol* **26**, 830-841 e839, doi:10.1016/j.chembiol.2019.02.014 (2019).
- Allen, J. K. *et al.* Sustained Adrenergic Signaling Promotes Intratumoral Innervation through BDNF Induction. *Cancer Res* **78**, 3233-3242, doi:10.1158/0008-5472.CAN-16-1701 (2018).
- Ulrich-Lai, Y. M. & Herman, J. P. Neural regulation of endocrine and autonomic stress responses. *Nat Rev Neurosci* **10**, 397-409, doi:10.1038/nrn2647 (2009).
- Picard, M. et al. Mitochondrial functions modulate neuroendocrine, metabolic, inflammatory,
 and transcriptional responses to acute psychological stress. Proc Natl Acad Sci U S A 112,
 E6614-6623, doi:10.1073/pnas.1515733112 (2015).
- 902 70 Cole, S. W. & Sood, A. K. Molecular pathways: beta-adrenergic signaling in cancer. *Clin Cancer* 903 *Res* **18**, 1201-1206, doi:10.1158/1078-0432.CCR-11-0641 (2012).
- Wu, D., Katz, A., Lee, C. H. & Simon, M. I. Activation of phospholipase C by alpha 1adrenergic receptors is mediated by the alpha subunits of Gq family. *J Biol Chem* **267**, 25798-25802 (1992).
- 907 72 Wick, G., Hu, Y., Schwarz, S. & Kroemer, G. Immunoendocrine communication via the hypothalamo-pituitary-adrenal axis in autoimmune diseases. *Endocr Rev* **14**, 539-563, doi:10.1210/edrv-14-5-539 (1993).
- 910 73 Weikum, E. R., Knuesel, M. T., Ortlund, E. A. & Yamamoto, K. R. Glucocorticoid receptor control of transcription: precision and plasticity via allostery. *Nat Rev Mol Cell Biol* **18**, 159-174, doi:10.1038/nrm.2016.152 (2017).
- 913 74 Ping, Y. Q. *et al.* Structures of the glucocorticoid-bound adhesion receptor GPR97-Go complex. *Nature* **589**, 620-626, doi:10.1038/s41586-020-03083-w (2021).
- 915 75 Wang, J. J. et al. Gpr97 is essential for the follicular versus marginal zone B-lymphocyte fate

- 916 decision. Cell Death Dis 4, e853, doi:10.1038/cddis.2013.346 (2013).
- 917 76 Chu, T. Y. *et al.* GPR97 triggers inflammatory processes in human neutrophils via a macromolecular complex upstream of PAR2 activation. *Nat Commun* **13**, 6385, doi:10.1038/s41467-022-34083-1 (2022).
- 920 77 Graeff, F. G., Guimaraes, F. S., De Andrade, T. G. & Deakin, J. F. Role of 5-HT in stress, anxiety, and depression. *Pharmacol Biochem Behav* **54**, 129-141, doi:10.1016/0091-3057(95)02135-3 (1996).
- 923 78 Mineur, Y. S. *et al.* Cholinergic signaling in the hippocampus regulates social stress resilience 924 and anxiety- and depression-like behavior. *Proc Natl Acad Sci U S A* **110**, 3573-3578, 925 doi:10.1073/pnas.1219731110 (2013).
- Picciotto, M. R., Higley, M. J. & Mineur, Y. S. Acetylcholine as a neuromodulator: cholinergic signaling shapes nervous system function and behavior. *Neuron* **76**, 116-129, doi:10.1016/j.neuron.2012.08.036 (2012).
- 929 80 Lydiard, R. B. The role of GABA in anxiety disorders. *J Clin Psychiatry* **64 Suppl 3**, 21-27 (2003).
- Nuss, P. Anxiety disorders and GABA neurotransmission: a disturbance of modulation. Neuropsychiatr Dis Treat 11, 165-175, doi:10.2147/NDT.S58841 (2015).
- Panula, P. & Nuutinen, S. The histaminergic network in the brain: basic organization and role in disease. *Nat Rev Neurosci* **14**, 472-487, doi:10.1038/nrn3526 (2013).
- de Almeida, D. O., Ferreira, H. S., Pereira, L. B. & Fregoneze, J. B. Hypertensive response to stress: the role of histaminergic H1 and H2 receptors in the medial amygdala. *Physiol Behav* 144, 95-102, doi:10.1016/j.physbeh.2015.03.009 (2015).
- 938 84 Bali, A., Randhawa, P. K. & Jaggi, A. S. Stress and opioids: role of opioids in modulating stress-939 related behavior and effect of stress on morphine conditioned place preference. *Neurosci Biobehav Rev* **51**, 138-150, doi:10.1016/j.neubiorev.2014.12.018 (2015).
- 941 85 Valentino, R. J. & Van Bockstaele, E. Endogenous Opioids: The Downside of Opposing Stress. 942 *Neurobiol Stress* 1, 23-32, doi:10.1016/j.ynstr.2014.09.006 (2015).
- Pecina, M. *et al.* Endogenous opioid system dysregulation in depression: implications for new therapeutic approaches. *Mol Psychiatry* **24**, 576-587, doi:10.1038/s41380-018-0117-2 (2019).
- 945 87 Oshaghi, M., Kourosh-Arami, M. & Roozbehkia, M. Role of neurotransmitters in immune-946 mediated inflammatory disorders: a crosstalk between the nervous and immune systems. *Neurol* 947 *Sci* 44, 99-113, doi:10.1007/s10072-022-06413-0 (2023).
- 948 88 Herman, J. P. *et al.* Regulation of the Hypothalamic-Pituitary-Adrenocortical Stress Response. 949 *Compr Physiol* **6**, 603-621, doi:10.1002/cphy.c150015 (2016).
- 950 89 Lanfumey, L., Mongeau, R., Cohen-Salmon, C. & Hamon, M. Corticosteroid-serotonin 951 interactions in the neurobiological mechanisms of stress-related disorders. *Neurosci Biobehav* 952 *Rev* **32**, 1174-1184, doi:10.1016/j.neubiorev.2008.04.006 (2008).
- 953 90 Sui, P. *et al.* Pulmonary neuroendocrine cells amplify allergic asthma responses. *Science* **360**, doi:10.1126/science.aan8546 (2018).
- 955 91 Bellono, N. W. *et al.* Enterochromaffin Cells Are Gut Chemosensors that Couple to Sensory Neural Pathways. *Cell* **170**, 185-198 e116, doi:10.1016/j.cell.2017.05.034 (2017).
- 957 92 Sidler, D. *et al.* Colon cancer cells produce immunoregulatory glucocorticoids. *Oncogene* **30**, 2411-2419, doi:10.1038/onc.2010.629 (2011).
- 959 93 Honke, N. *et al.* Endogenously produced catecholamines improve the regulatory function of TLR9-activated B cells. *PLoS Biol* **20**, e3001513, doi:10.1371/journal.pbio.3001513 (2022).
- 961 94 Guida, F. *et al.* Antibiotic-induced microbiota perturbation causes gut endocannabinoidome changes, hippocampal neuroglial reorganization and depression in mice. *Brain Behav Immun* 67, 230-245, doi:10.1016/j.bbi.2017.09.001 (2018).
- 964 95 Monje, M. *et al.* Roadmap for the Emerging Field of Cancer Neuroscience. *Cell* **181**, 219-222, doi:10.1016/j.cell.2020.03.034 (2020).
- 966 96 Walker, A. K. *et al.* Circulating epinephrine is not required for chronic stress to enhance metastasis. *Psychoneuroendocrinology* **99**, 191-195, doi:10.1016/j.psyneuen.2018.09.012 (2019).
- 969 97 Renz, B. W. *et al.* beta2 Adrenergic-Neurotrophin Feedforward Loop Promotes Pancreatic Cancer. *Cancer Cell* 33, 75-90 e77, doi:10.1016/j.ccell.2017.11.007 (2018).
- 971 98 Renz, B. W. *et al.* Cholinergic Signaling via Muscarinic Receptors Directly and Indirectly 972 Suppresses Pancreatic Tumorigenesis and Cancer Stemness. *Cancer Discov* **8**, 1458-1473, doi:10.1158/2159-8290.CD-18-0046 (2018).
- 974 99 Renz, B. W. *et al.* beta2 Adrenergic-Neurotrophin Feedforward Loop Promotes Pancreatic Cancer. *Cancer Cell* **34**, 863-867, doi:10.1016/j.ccell.2018.10.010 (2018).

- Reijmen, E., Vannucci, L., De Couck, M., De Greve, J. & Gidron, Y. Therapeutic potential of the vagus nerve in cancer. *Immunol Lett* **202**, 38-43, doi:10.1016/j.imlet.2018.07.006 (2018).
- 978 101 De Couck, M., Caers, R., Spiegel, D. & Gidron, Y. The Role of the Vagus Nerve in Cancer 979 Prognosis: A Systematic and a Comprehensive Review. *J Oncol* **2018**, 1236787, 980 doi:10.1155/2018/1236787 (2018).
- 981 102 Husby, A., Wohlfahrt, J. & Melbye, M. Vasectomy and Prostate Cancer Risk: A 38-Year 982 Nationwide Cohort Study. *J Natl Cancer Inst* 112, 71-77, doi:10.1093/jnci/djz099 (2020).
- 983 103 Webster, J. I., Tonelli, L. & Sternberg, E. M. Neuroendocrine regulation of immunity. *Annu Rev Immunol* **20**, 125-163, doi:10.1146/annurev.immunol.20.082401.104914 (2002).
- 985 104 Taves, M. D. & Ashwell, J. D. Glucocorticoids in T cell development, differentiation and function. *Nat Rev Immunol*, doi:10.1038/s41577-020-00464-0 (2020).
- 987 105 Cain, D. W. & Cidlowski, J. A. Immune regulation by glucocorticoids. *Nat Rev Immunol* 17, 233-247, doi:10.1038/nri.2017.1 (2017).
- 989 106 Dhabhar, F. S. & McEwen, B. S. Enhancing versus suppressive effects of stress hormones on skin immune function. *Proc Natl Acad Sci U S A* **96**, 1059-1064, doi:10.1073/pnas.96.3.1059 (1999).
- 992 107 Sharma, D. & Farrar, J. D. Adrenergic regulation of immune cell function and inflammation. 993 *Semin Immunopathol* **42**, 709-717, doi:10.1007/s00281-020-00829-6 (2020).
- 994 108 Hunzeker, J. T. *et al.* A marked reduction in priming of cytotoxic CD8+ T cells mediated by stress-induced glucocorticoids involves multiple deficiencies in cross-presentation by dendritic cells. *J Immunol* **186**, 183-194, doi:10.4049/jimmunol.1001737 (2011).
- 997 109 Collins, N. *et al.* The Bone Marrow Protects and Optimizes Immunological Memory during 998 Dietary Restriction. *Cell* **178**, 1088-1101 e1015, doi:10.1016/j.cell.2019.07.049 (2019).
- Franco, L. M. *et al.* Immune regulation by glucocorticoids can be linked to cell type-dependent transcriptional responses. *J Exp Med* **216**, 384-406, doi:10.1084/jem.20180595 (2019).
- 1001 111 Pani, L., Porcella, A. & Gessa, G. L. The role of stress in the pathophysiology of the dopaminergic system. *Mol Psychiatry* **5**, 14-21, doi:10.1038/sj.mp.4000589 (2000).
- 1003 112 Soliman, A. *et al.* Stress-induced dopamine release in humans at risk of psychosis: a 1004 [11C]raclopride PET study. *Neuropsychopharmacology* 33, 2033-2041, doi:10.1038/sj.npp.1301597 (2008).
- 1006 113 Pruessner, J. C., Champagne, F., Meaney, M. J. & Dagher, A. Dopamine release in response to a psychological stress in humans and its relationship to early life maternal care: a positron emission tomography study using [11C]raclopride. *J Neurosci* 24, 2825-2831, doi:10.1523/JNEUROSCI.3422-03.2004 (2004).
- Watanabe, Y. *et al.* Dopamine selectively induces migration and homing of naive CD8+ T cells via dopamine receptor D3. *J Immunol* **176**, 848-856, doi:10.4049/jimmunol.176.2.848 (2006).
- Mikulak, J. *et al.* Dopamine inhibits the effector functions of activated NK cells via the upregulation of the D5 receptor. *J Immunol* **193**, 2792-2800, doi:10.4049/jimmunol.1401114 (2014).
- 1015 116 Yan, Y. *et al.* Dopamine controls systemic inflammation through inhibition of NLRP3 inflammasome. *Cell* **160**, 62-73, doi:10.1016/j.cell.2014.11.047 (2015).
- 1017 Yanagawa, Y., Matsumoto, M. & Togashi, H. Enhanced dendritic cell antigen uptake via alpha2 1018 adrenoceptor-mediated PI3K activation following brief exposure to noradrenaline. *J Immunol* 1019 **185**, 5762-5768, doi:10.4049/jimmunol.1001899 (2010).
- 1020 118 Takenaka, M. C. *et al.* Norepinephrine Controls Effector T Cell Differentiation through beta2-1021 Adrenergic Receptor-Mediated Inhibition of NF-kappaB and AP-1 in Dendritic Cells. *J Immunol* **196**, 637-644, doi:10.4049/jimmunol.1501206 (2016).
- 1023 119 Guereschi, M. G. *et al.* Beta2-adrenergic receptor signaling in CD4+ Foxp3+ regulatory T cells enhances their suppressive function in a PKA-dependent manner. *Eur J Immunol* **43**, 1001-1012, doi:10.1002/eji.201243005 (2013).
- Devi, S. *et al.* Adrenergic regulation of the vasculature impairs leukocyte interstitial migration and suppresses immune responses. *Immunity* **54**, 1219-1230 e1217, doi:10.1016/j.immuni.2021.03.025 (2021).
- 1029 121 Schiller, M., Ben-Shaanan, T. L. & Rolls, A. Neuronal regulation of immunity: why, how and where? *Nat Rev Immunol* **21**, 20-36, doi:10.1038/s41577-020-0387-1 (2021).
- Lutgendorf, S. K. *et al.* Social support, psychological distress, and natural killer cell activity in ovarian cancer. *J Clin Oncol* **23**, 7105-7113, doi:10.1200/JCO.2005.10.015 (2005).
- Varker, K. A. *et al.* Impaired natural killer cell lysis in breast cancer patients with high levels of psychological stress is associated with altered expression of killer immunoglobin-like receptors. *J Surg Res* **139**, 36-44, doi:10.1016/j.jss.2006.08.037 (2007).

- Levi, B. *et al.* Stress impairs the efficacy of immune stimulation by CpG-C: Potential neuroendocrine mediating mechanisms and significance to tumor metastasis and the perioperative period. *Brain Behav Immun* **56**, 209-220, doi:10.1016/j.bbi.2016.02.025 (2016).
- 1039 125 Koide, S. S. Mifepristone. Auxiliary therapeutic use in cancer and related disorders. *J Reprod* 1040 *Med* 43, 551-560 (1998).
- 1041 126 Check, J. H., Dix, E., Cohen, R., Check, D. & Wilson, C. Efficacy of the progesterone receptor antagonist mifepristone for palliative therapy of patients with a variety of advanced cancer types.

 1043 Anticancer Res 30, 623-628 (2010).
- 1044 127 Check, J. H., Check, D., Wilson, C. & Lofberg, P. Long-term High-quality Survival with Single-1045 agent Mifepristone Treatment Despite Advanced Cancer. *Anticancer Res* **36**, 6511-6513, 1046 doi:10.21873/anticanres.11251 (2016).
- 1047 128 Cronin-Fenton, D. *et al.* Concurrent new drug prescriptions and prognosis of early breast cancer: studies using the Danish Breast Cancer Group clinical database. *Acta Oncol* **57**, 120-128, doi:10.1080/0284186X.2017.1407040 (2018).
- This study identifies a particular myeloid cell population in the TME that produces endogenous glucocorticoid to induce dysfunctional CD8⁺T cells and failure of cancer immunotherapy.
- Mulick, A. *et al.* Is improvement in comorbid major depression associated with longer survival in people with cancer? A long-term follow-up of participants in the SMaRT Oncology-2 and 3 trials. *J Psychosom Res* **116**, 106-112, doi:10.1016/j.jpsychores.2018.11.008 (2019).
- 1055 130 Acharya, N. *et al.* Endogenous Glucocorticoid Signaling Regulates CD8(+) T Cell Differentiation and Development of Dysfunction in the Tumor Microenvironment. *Immunity* **53**, 658-671 e656, doi:10.1016/j.immuni.2020.08.005 (2020).
- 1058 131 Yi, L. & Zheng, C. The emerging roles of ZDHHCs-mediated protein palmitoylation in the antiviral innate immune responses. *Crit Rev Microbiol* 47, 34-43, doi:10.1080/1040841X.2020.1835821 (2021).
- 1061 132 Arbour, K. C. *et al.* Impact of Baseline Steroids on Efficacy of Programmed Cell Death-1 and Programmed Death-Ligand 1 Blockade in Patients With Non-Small-Cell Lung Cancer. *J Clin Oncol* **36**, 2872-2878, doi:10.1200/JCO.2018.79.0006 (2018).
- 1064 133 Maxwell, R. *et al.* Contrasting impact of corticosteroids on anti-PD-1 immunotherapy efficacy for tumor histologies located within or outside the central nervous system. *Oncoimmunology* 7, e1500108, doi:10.1080/2162402X.2018.1500108 (2018).
- 1067 134 Aston, W. J. *et al.* Dexamethasone differentially depletes tumour and peripheral blood lymphocytes and can impact the efficacy of chemotherapy/checkpoint blockade combination treatment. *Oncoimmunology* **8**, e1641390, doi:10.1080/2162402X.2019.1641390 (2019).
- 1070 135 Papa, I. *et al.* T(FH)-derived dopamine accelerates productive synapses in germinal centres. Nature **547**, 318-323, doi:10.1038/nature23013 (2017).
- 1072 136 van der Heijden, C. *et al.* Catecholamines Induce Trained Immunity in Monocytes In Vitro and In Vivo. *Circ Res* **127**, 269-283, doi:10.1161/CIRCRESAHA.119.315800 (2020).
- 1074 137 Elenkov, I. J. in NeuroImmune Biology Vol. 7 189-206 (Elsevier, 2007).
- 1075 138 Arreola, R. *et al.* Immunomodulatory Effects Mediated by Dopamine. *J Immunol Res* **2016**, 1076 3160486, doi:10.1155/2016/3160486 (2016).
- 1077 139 Baik, J. H. Stress and the dopaminergic reward system. *Exp Mol Med* **52**, 1879-1890, doi:10.1038/s12276-020-00532-4 (2020).
- Bloomfield, M. A., McCutcheon, R. A., Kempton, M., Freeman, T. P. & Howes, O. The effects of psychosocial stress on dopaminergic function and the acute stress response. *Elife* **8**, doi:10.7554/eLife.46797 (2019).
- 1082 141 Saha, B., Mondal, A. C., Basu, S. & Dasgupta, P. S. Circulating dopamine level, in lung carcinoma patients, inhibits proliferation and cytotoxicity of CD4+ and CD8+ T cells by D1 dopamine receptors: an in vitro analysis. *Int Immunopharmacol* 1, 1363-1374, doi:10.1016/s1567-5769(01)00068-6 (2001).
- Figueroa, C. *et al.* Inhibition of dopamine receptor D3 signaling in dendritic cells increases antigen cross-presentation to CD8(+) T-cells favoring anti-tumor immunity. *J Neuroimmunol* **303**, 99-107, doi:10.1016/j.jneuroim.2016.12.014 (2017).
- Hoeppner, L. H. *et al.* Dopamine D2 receptor agonists inhibit lung cancer progression by reducing angiogenesis and tumor infiltrating myeloid derived suppressor cells. *Mol Oncol* **9**, 270-281, doi:10.1016/j.molonc.2014.08.008 (2015).
- 1092 144 Wu, J. *et al.* Dopamine inhibits the function of Gr-1+CD115+ myeloid-derived suppressor cells through D1-like receptors and enhances anti-tumor immunity. *J Leukoc Biol* **97**, 191-200, doi:10.1189/jlb.5A1113-626RR (2015).
- 1095 145 Myers, S. A., Eriksson, N., Burow, R., Wang, S. C. & Muscat, G. E. Beta-adrenergic signaling

- regulates NR4A nuclear receptor and metabolic gene expression in multiple tissues. *Mol Cell Endocrinol* **309**, 101-108, doi:10.1016/j.mce.2009.05.006 (2009).
- 1098 146 Liu, X. *et al.* Genome-wide analysis identifies NR4A1 as a key mediator of T cell dysfunction. 1099 *Nature* **567**, 525-529, doi:10.1038/s41586-019-0979-8 (2019).
- Daher, C. *et al.* Blockade of beta-Adrenergic Receptors Improves CD8(+) T-cell Priming and Cancer Vaccine Efficacy. *Cancer Immunol Res* 7, 1849-1863, doi:10.1158/2326-6066.CIR-18-0833 (2019).
- 1103 148 Nissen, M. D., Sloan, E. K. & Mattarollo, S. R. beta-Adrenergic Signaling Impairs Antitumor 1104 CD8(+) T-cell Responses to B-cell Lymphoma Immunotherapy. *Cancer Immunol Res* **6**, 98-109, 1105 doi:10.1158/2326-6066.CIR-17-0401 (2018).
- 1106 149 Qiao, G. *et al.* beta-Adrenergic signaling blocks murine CD8(+) T-cell metabolic reprogramming during activation: a mechanism for immunosuppression by adrenergic stress. 1108 *Cancer Immunol Immunother* **68**, 11-22, doi:10.1007/s00262-018-2243-8 (2019).
- 1109 Qin, J. F. *et al.* Adrenergic receptor beta2 activation by stress promotes breast cancer progression 1110 through macrophages M2 polarization in tumor microenvironment. *BMB Rep* **48**, 295-300, 1111 doi:10.5483/bmbrep.2015.48.5.008 (2015).
- 1112 151 Sloan, E. K. *et al.* The sympathetic nervous system induces a metastatic switch in primary breast cancer. *Cancer Res* **70**, 7042-7052, doi:10.1158/0008-5472.CAN-10-0522 (2010).
- Mohammadpour, H. *et al.* beta2 adrenergic receptor-mediated signaling regulates the immunosuppressive potential of myeloid-derived suppressor cells. *J Clin Invest* **129**, 5537-5552, doi:10.1172/JCI129502 (2019).
- 1117 153 Cheng, Y. *et al.* Depression-Induced Neuropeptide Y Secretion Promotes Prostate Cancer 1118 Growth by Recruiting Myeloid Cells. *Clin Cancer Res* **25**, 2621-2632, doi:10.1158/1078-1119 0432.CCR-18-2912 (2019).
- 1120 154 Kokolus, K. M. *et al.* Beta blocker use correlates with better overall survival in metastatic melanoma patients and improves the efficacy of immunotherapies in mice. *Oncoimmunology* 7, e1405205, doi:10.1080/2162402X.2017.1405205 (2018).
- Bucsek, M. J. *et al.* beta-Adrenergic Signaling in Mice Housed at Standard Temperatures Suppresses an Effector Phenotype in CD8(+) T Cells and Undermines Checkpoint Inhibitor Therapy. *Cancer Res* 77, 5639-5651, doi:10.1158/0008-5472.CAN-17-0546 (2017).
- 1126 Chen, M. *et al.* Adrenergic stress constrains the development of anti-tumor immunity and abscopal responses following local radiation. *Nat Commun* **11**, 1821, doi:10.1038/s41467-020-128 15676-0 (2020).
- 1129 157 Botta, F. & Maestroni, G. J. Adrenergic modulation of dendritic cell cancer vaccine in a mouse model: role of dendritic cell maturation. *J Immunother* 31, 263-270, doi:10.1097/CJI.0b013e318160995e (2008).
- Hiller, J. G. *et al.* Pre-operative beta-blockade with propranolol reduces biomarkers of metastasis in breast cancer: a Phase II randomized trial. *Clin Cancer Res*, doi:10.1158/1078-0432.CCR-19-2641 (2019).
- Shaashua, L. *et al.* Perioperative COX-2 and beta-Adrenergic Blockade Improves Metastatic Biomarkers in Breast Cancer Patients in a Phase-II Randomized Trial. *Clin Cancer Res* **23**, 4651-4661, doi:10.1158/1078-0432.CCR-17-0152 (2017).
- De Giorgi, V. *et al.* Propranolol for Off-label Treatment of Patients With Melanoma: Results From a Cohort Study. *JAMA Oncol* **4**, e172908, doi:10.1001/jamaoncol.2017.2908 (2018).
- 1140 161 Phadke, S. & Clamon, G. Beta blockade as adjunctive breast cancer therapy: A review. *Crit Rev Oncol Hematol* 138, 173-177, doi:10.1016/j.critrevonc.2019.04.006 (2019).
- Thang, S. *et al.* Neuroendocrine Regulation of Stress-Induced T Cell Dysfunction during Lung Cancer Immunosurveillance via the Kisspeptin/GPR54 Signaling Pathway. *Adv Sci (Weinh)* **9**, e2104132, doi:10.1002/advs.202104132 (2022).
- Hill, M. N. *et al.* Endogenous cannabinoid signaling is essential for stress adaptation. *Proc Natl Acad Sci U S A* **107**, 9406-9411, doi:10.1073/pnas.0914661107 (2010).
- 1147 164 Xiong, X. *et al.* Cannabis suppresses antitumor immunity by inhibiting JAK/STAT signaling in T cells through CNR2. *Signal Transduct Target Ther* 7, 99, doi:10.1038/s41392-022-00918-y (2022).
- Won, E. & Kim, Y. K. Stress, the Autonomic Nervous System, and the Immune-kynurenine Pathway in the Etiology of Depression. *Curr Neuropharmacol* **14**, 665-673, doi:10.2174/1570159x14666151208113006 (2016).
- 1153 166 Gao, F. G., Wan da, F. & Gu, J. R. Ex vivo nicotine stimulation augments the efficacy of therapeutic bone marrow-derived dendritic cell vaccination. *Clin Cancer Res* **13**, 3706-3712, doi:10.1158/1078-0432.CCR-07-0028 (2007).

- Dubeykovskaya, Z. *et al.* Neural innervation stimulates splenic TFF2 to arrest myeloid cell expansion and cancer. *Nat Commun* 7, 10517, doi:10.1038/ncomms10517 (2016).
- Liu, L. *et al.* Hippocampal metabolic differences implicate distinctions between physical and psychological stress in four rat models of depression. *Transl Psychiatry* **8**, 4, doi:10.1038/s41398-017-0018-1 (2018).
- 1161 169 Czeh, B. *et al.* Stress-induced changes in cerebral metabolites, hippocampal volume, and cell proliferation are prevented by antidepressant treatment with tianeptine. *Proc Natl Acad Sci U S* 1163 *A* **98**, 12796-12801, doi:10.1073/pnas.211427898 (2001).
- Peckett, A. J., Wright, D. C. & Riddell, M. C. The effects of glucocorticoids on adipose tissue lipid metabolism. *Metabolism* **60**, 1500-1510, doi:10.1016/j.metabol.2011.06.012 (2011).
- Jia, H. M. *et al.* Chronic unpredictive mild stress leads to altered hepatic metabolic profile and gene expression. *Sci Rep* **6**, 23441, doi:10.1038/srep23441 (2016).
- 1168 172 Rodrigues Mantuano, N. *et al.* Hyperglycemia Enhances Cancer Immune Evasion by Inducing
 1169 Alternative Macrophage Polarization through Increased O-GlcNAcylation. *Cancer Immunol*1170 *Res* **8**, 1262-1272, doi:10.1158/2326-6066.CIR-19-0904 (2020).
- This study explores how hyperglycemia drives tumour progression, by increasing O-GlcNAcylation in tumour-associated macrophages to switch on the M2-like phenotype and favour cancer immune evasion.
- 1174 173 Brzozowski, B. *et al.* Mechanisms by which Stress Affects the Experimental and Clinical Inflammatory Bowel Disease (IBD): Role of Brain-Gut Axis. *Curr Neuropharmacol* **14**, 892-1176 900, doi:10.2174/1570159x14666160404124127 (2016).
- Patel, C. H., Leone, R. D., Horton, M. R. & Powell, J. D. Targeting metabolism to regulate immune responses in autoimmunity and cancer. *Nat Rev Drug Discov* **18**, 669-688, doi:10.1038/s41573-019-0032-5 (2019).
- 1180 175 Voss, K. *et al.* A guide to interrogating immunometabolism. *Nat Rev Immunol* **21**, 637-652, doi:10.1038/s41577-021-00529-8 (2021).
- Nonogaki, K. & Iguchi, A. Stress, acute hyperglycemia, and hyperlipidemia role of the autonomic nervous system and cytokines. *Trends Endocrinol Metab* **8**, 192-197, doi:10.1016/s1043-2760(97)00038-6 (1997).
- Maduka, I. C., Neboh, E. E. & Ufelle, S. A. The relationship between serum cortisol, adrenaline, blood glucose and lipid profile of undergraduate students under examination stress. *Afr Health Sci* **15**, 131-136, doi:10.4314/ahs.v15i1.18 (2015).
- 1188 178 Mohammadpour, H., MacDonald, C. R., McCarthy, P. L., Abrams, S. I. & Repasky, E. A. beta2-1189 adrenergic receptor signaling regulates metabolic pathways critical to myeloid-derived 1190 cell function within the TME. Cellsuppressor Rep 37. 1191 doi:10.1016/j.celrep.2021.109883 (2021).
- Muthuswamy, R. *et al.* Epinephrine promotes COX-2-dependent immune suppression in myeloid cells and cancer tissues. *Brain Behav Immun* **62**, 78-86, doi:10.1016/j.bbi.2017.02.008 (2017).
- 1195 180 Pearce, E. L., Poffenberger, M. C., Chang, C. H. & Jones, R. G. Fueling immunity: insights into metabolism and lymphocyte function. *Science* **342**, 1242454, doi:10.1126/science.1242454 (2013).
- 1198 181 Picard, M. & McEwen, B. S. Psychological Stress and Mitochondria: A Systematic Review. 1199 *Psychosom Med* **80**, 141-153, doi:10.1097/PSY.00000000000545 (2018).
- 1200 182 Qiao, G. *et al.* Chronic Adrenergic Stress Contributes to Metabolic Dysfunction and an 1201 Exhausted Phenotype in T Cells in the Tumor Microenvironment. *Cancer Immunol Res* **9**, 651-1202 664, doi:10.1158/2326-6066.CIR-20-0445 (2021).
- Tokunaga, A. *et al.* Selective inhibition of low-affinity memory CD8(+) T cells by corticosteroids. *J Exp Med* **216**, 2701-2713, doi:10.1084/jem.20190738 (2019).
- Brand, A. *et al.* LDHA-Associated Lactic Acid Production Blunts Tumor Immunosurveillance by T and NK Cells. *Cell Metab* **24**, 657-671, doi:10.1016/j.cmet.2016.08.011 (2016).
- 1207 185 Perego, M. *et al.* Reactivation of dormant tumor cells by modified lipids derived from stress-1208 activated neutrophils. *Sci Transl Med* **12**, doi:10.1126/scitranslmed.abb5817 (2020).
- 1209 186 Bresnick, A. R., Weber, D. J. & Zimmer, D. B. S100 proteins in cancer. *Nat Rev Cancer* **15**, 96-1210 109, doi:10.1038/nrc3893 (2015).
- 1211 187 Diskin, C., Ryan, T. A. J. & O'Neill, L. A. J. Modification of Proteins by Metabolites in Immunity. *Immunity* **54**, 19-31, doi:10.1016/j.immuni.2020.09.014 (2021).
- 1213 188 Barth, E. *et al.* Glucose metabolism and catecholamines. *Crit Care Med* **35**, S508-518, doi:10.1097/01.CCM.0000278047.06965.20 (2007).
- 1215 189 Zhang, D. et al. Metabolic regulation of gene expression by histone lactylation. Nature 574,

- 1216 575-580, doi:10.1038/s41586-019-1678-1 (2019).
- 1217 190 Chang, Y. H., Weng, C. L. & Lin, K. I. O-GlcNAcylation and its role in the immune system. *J* 1218 *Biomed Sci* **27**, 57, doi:10.1186/s12929-020-00648-9 (2020).
- 1219 Xiu, F., Stanojcic, M., Diao, L. & Jeschke, M. G. Stress hyperglycemia, insulin treatment, and innate immune cells. *Int J Endocrinol* **2014**, 486403, doi:10.1155/2014/486403 (2014).
- 1221 192 Picard, M., Juster, R. P. & McEwen, B. S. Mitochondrial allostatic load puts the 'gluc' back in glucocorticoids. *Nat Rev Endocrinol* **10**, 303-310, doi:10.1038/nrendo.2014.22 (2014).
- 1223 193 Jiang, H. *et al.* Protein Lipidation: Occurrence, Mechanisms, Biological Functions, and 1224 Enabling Technologies. *Chem Rev* **118**, 919-988, doi:10.1021/acs.chemrev.6b00750 (2018).
- 1225 194 Wild, A. R. *et al.* Exploring the expression patterns of palmitoylating and de-palmitoylating enzymes in the mouse brain using the curated RNA-seq database BrainPalmSeq. *Elife* 11, 1227 doi:10.7554/eLife.75804 (2022).
- 1228 195 Zareba-Koziol, M. *et al.* Stress-induced Changes in the S-palmitoylation and S-nitrosylation of Synaptic Proteins. *Mol Cell Proteomics* **18**, 1916-1938, doi:10.1074/mcp.RA119.001581 1230 (2019).
- 1231 196 Yao, H. *et al.* Inhibiting PD-L1 palmitoylation enhances T-cell immune responses against tumours. *Nat Biomed Eng* **3**, 306-317, doi:10.1038/s41551-019-0375-6 (2019).
- 1233 197 Cryan, J. F. *et al.* The Microbiota-Gut-Brain Axis. *Physiol Rev* **99**, 1877-2013, doi:10.1152/physrev.00018.2018 (2019).
- 1235 198 Morais, L. H., Schreiber, H. L. t. & Mazmanian, S. K. The gut microbiota-brain axis in behaviour and brain disorders. *Nat Rev Microbiol* **19**, 241-255, doi:10.1038/s41579-020-00460-1237 0 (2021).
- 1238 199 Mittal, R. *et al.* Neurotransmitters: The Critical Modulators Regulating Gut-Brain Axis. *J Cell Physiol* **232**, 2359-2372, doi:10.1002/jcp.25518 (2017).
- Wu, M. *et al.* Associations between disordered gut microbiota and changes of neurotransmitters and short-chain fatty acids in depressed mice. *Transl Psychiatry* **10**, 350, doi:10.1038/s41398-020-01038-3 (2020).
- 1243 201 Jiang, H. *et al.* Altered fecal microbiota composition in patients with major depressive disorder. 1244 *Brain Behav Immun* **48**, 186-194, doi:10.1016/j.bbi.2015.03.016 (2015).
- 1245 202 Jiang, H. Y. *et al.* Altered gut microbiota profile in patients with generalized anxiety disorder. *J* 1246 *Psychiatr Res* **104**, 130-136, doi:10.1016/j.jpsychires.2018.07.007 (2018).
- Hollins, S. L. & Hodgson, D. M. Stress, microbiota, and immunity. *Current Opinion in Behavioral Sciences* **28**, 66-71, doi:https://doi.org/10.1016/j.cobeha.2019.01.015 (2019).
- 1249 204 Gao, X. *et al.* Chronic stress promotes colitis by disturbing the gut microbiota and triggering 1250 immune system response. *Proc Natl Acad Sci U S A* **115**, E2960-E2969, 1251 doi:10.1073/pnas.1720696115 (2018).
- Viaud, S. *et al.* The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. *Science* **342**, 971-976, doi:10.1126/science.1240537 (2013).
- Yu, L. X. & Schwabe, R. F. The gut microbiome and liver cancer: mechanisms and clinical translation. *Nat Rev Gastroenterol Hepatol* **14**, 527-539, doi:10.1038/nrgastro.2017.72 (2017).
- 1256 207 Lebeer, S., Vanderleyden, J. & De Keersmaecker, S. C. Host interactions of probiotic bacterial surface molecules: comparison with commensals and pathogens. *Nat Rev Microbiol* **8**, 171-184, doi:10.1038/nrmicro2297 (2010).
- Ochoa-Reparaz, J. *et al.* Central nervous system demyelinating disease protection by the human commensal Bacteroides fragilis depends on polysaccharide A expression. *J Immunol* **185**, 4101-4108, doi:10.4049/jimmunol.1001443 (2010).
- 1262 209 Zitvogel, L., Ayyoub, M., Routy, B. & Kroemer, G. Microbiome and Anticancer 1263 Immunosurveillance. *Cell* **165**, 276-287, doi:10.1016/j.cell.2016.03.001 (2016).
- Fluckiger, A. *et al.* Cross-reactivity between tumor MHC class I-restricted antigens and an enterococcal bacteriophage. *Science* **369**, 936-942, doi:10.1126/science.aax0701 (2020).
- This study reports that naturally processed cancer antigens and microbial peptides may share crossreactive T cell epitopes, which can be targeted for cancer immunotherapy.
- Ruff, W. E. & Kriegel, M. A. Autoimmune host-microbiota interactions at barrier sites and beyond. *Trends Mol Med* **21**, 233-244, doi:10.1016/j.molmed.2015.02.006 (2015).
- 1270 212 Ait-Belgnaoui, A. *et al.* Prevention of gut leakiness by a probiotic treatment leads to attenuated HPA response to an acute psychological stress in rats. *Psychoneuroendocrinology* **37**, 1885-1272 1895, doi:10.1016/j.psyneuen.2012.03.024 (2012).
- 1273 213 Ait-Belgnaoui, A. *et al.* Bifidobacterium longum and Lactobacillus helveticus Synergistically
 1274 Suppress Stress-related Visceral Hypersensitivity Through Hypothalamic-Pituitary-Adrenal
 1275 Axis Modulation. *J Neurogastroenterol Motil* **24**, 138-146, doi:10.5056/jnm16167 (2018).

- Routy, B. *et al.* Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science* **359**, 91-97, doi:10.1126/science.aan3706 (2018).
- 1278 215 Gopalakrishnan, V. *et al.* Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. *Science* **359**, 97-103, doi:10.1126/science.aan4236 (2018).
- Matson, V. *et al.* The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients. *Science* **359**, 104-108, doi:10.1126/science.aao3290 (2018).
- 1282 217 Routy, B. *et al.* The gut microbiota influences anticancer immunosurveillance and general health.

 1283 Nat Rev Clin Oncol 15, 382-396, doi:10.1038/s41571-018-0006-2 (2018).
- 1284 218 Merchak, A. & Gaultier, A. Microbial metabolites and immune regulation: New targets for major depressive disorder. *Brain Behav Immun Health* **9**, 100169, doi:10.1016/j.bbih.2020.100169 (2020).
- 1287 219 Sittipo, P., Choi, J., Lee, S. & Lee, Y. K. The function of gut microbiota in immune-related neurological disorders: a review. *J Neuroinflammation* 19, 154, doi:10.1186/s12974-022-02510-1 (2022).
- 1290 220 Kim, C. H. Immune regulation by microbiome metabolites. *Immunology* **154**, 220-229, doi:10.1111/imm.12930 (2018).
- Zhao, L. et al. TGR5 deficiency activates antitumor immunity in non-small cell lung cancer via
 restraining M2 macrophage polarization. Acta Pharm Sin B 12, 787-800,
 doi:10.1016/j.apsb.2021.07.011 (2022).
- 1295 You, W. *et al.* Farnesoid X Receptor Constructs an Immunosuppressive Microenvironment and Sensitizes FXR(high)PD-L1(low) NSCLC to Anti-PD-1 Immunotherapy. *Cancer Immunol Res* 7, 990-1000, doi:10.1158/2326-6066.CIR-17-0672 (2019).
- 1298 223 Kenison, J. E. *et al.* The aryl hydrocarbon receptor suppresses immunity to oral squamous cell carcinoma through immune checkpoint regulation. *Proc Natl Acad Sci U S A* **118**, 1300 doi:10.1073/pnas.2012692118 (2021).
- Bachem, A. *et al.* Microbiota-Derived Short-Chain Fatty Acids Promote the Memory Potential of Antigen-Activated CD8(+) T Cells. *Immunity* **51**, 285-297 e285, doi:10.1016/j.immuni.2019.06.002 (2019).
- This study shows that microbiota-derived metabolites, short-chain fatty acids, can guide the metabolic rewiring that increases the memory potential of antigen-specific CD8⁺ T cells.
- Botticelli, A. *et al.* Gut metabolomics profiling of non-small cell lung cancer (NSCLC) patients under immunotherapy treatment. *J Transl Med* **18**, 49, doi:10.1186/s12967-020-02231-0 (2020).
- Nomura, M. *et al.* Association of Short-Chain Fatty Acids in the Gut Microbiome With Clinical Response to Treatment With Nivolumab or Pembrolizumab in Patients With Solid Cancer Tumors. *JAMA Netw Open* **3**, e202895, doi:10.1001/jamanetworkopen.2020.2895 (2020).
- Schneider, M. A. *et al.* Attenuation of peripheral serotonin inhibits tumor growth and enhances immune checkpoint blockade therapy in murine tumor models. *Sci Transl Med* **13**, eabc8188, doi:10.1126/scitranslmed.abc8188 (2021).
- Bravo, J. A. *et al.* Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc Natl Acad Sci U S A* **108**, 16050-16055, doi:10.1073/pnas.1102999108 (2011).
- Kienzl, M., Kargl, J. & Schicho, R. The Immune Endocannabinoid System of the Tumor Microenvironment. *Int J Mol Sci* **21**, doi:10.3390/ijms21238929 (2020).
- Huang, D. *et al.* Cancer-cell-derived GABA promotes beta-catenin-mediated tumour growth and immunosuppression. *Nat Cell Biol* **24**, 230-241, doi:10.1038/s41556-021-00820-9 (2022).
- 1321 Zhang, B. *et al.* B cell-derived GABA elicits IL-10(+) macrophages to limit anti-tumour immunity. *Nature* **599**, 471-476, doi:10.1038/s41586-021-04082-1 (2021).
- 1323 232 Luqman, A., Nega, M., Nguyen, M. T., Ebner, P. & Gotz, F. SadA-Expressing Staphylococci in 1324 the Human Gut Show Increased Cell Adherence and Internalization. *Cell Rep* 22, 535-545, 1325 doi:10.1016/j.celrep.2017.12.058 (2018).
- Williams, B. B. *et al.* Discovery and characterization of gut microbiota decarboxylases that can produce the neurotransmitter tryptamine. *Cell Host Microbe* **16**, 495-503, doi:10.1016/j.chom.2014.09.001 (2014).
- Bauknecht, P. & Jekely, G. Ancient coexistence of norepinephrine, tyramine, and octopamine signaling in bilaterians. *BMC Biol* **15**, 6, doi:10.1186/s12915-016-0341-7 (2017).
- Connil, N. *et al.* Identification of the Enterococcus faecalis tyrosine decarboxylase operon involved in tyramine production. *Appl Environ Microbiol* **68**, 3537-3544, doi:10.1128/aem.68.7.3537-3544.2002 (2002).
- 1334 236 Lameris, T. W. *et al.* Catecholamine handling in the porcine heart: a microdialysis approach. *Am* 1335 *J Physiol* **277**, H1562-1569, doi:10.1152/ajpheart.1999.277.4.H1562 (1999).

- Weis, W. I. & Kobilka, B. K. The Molecular Basis of G Protein-Coupled Receptor Activation.

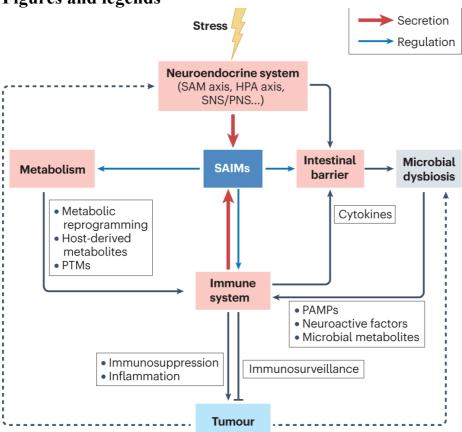
 Annu Rev Biochem 87, 897-919, doi:10.1146/annurev-biochem-060614-033910 (2018).
- 1338 238 Kepp, O., Marabelle, A., Zitvogel, L. & Kroemer, G. Oncolysis without viruses inducing systemic anticancer immune responses with local therapies. *Nat Rev Clin Oncol* 17, 49-64, doi:10.1038/s41571-019-0272-7 (2020).
- Foster, J. A., Rinaman, L. & Cryan, J. F. Stress & the gut-brain axis: Regulation by the microbiome. *Neurobiol Stress* 7, 124-136, doi:10.1016/j.ynstr.2017.03.001 (2017).
- 1343 240 Vuong, H. E., Yano, J. M., Fung, T. C. & Hsiao, E. Y. The Microbiome and Host Behavior. *Annu Rev Neurosci* **40**, 21-49, doi:10.1146/annurev-neuro-072116-031347 (2017).
- 1345 241 Sidler, D. *et al.* Colon cancer cells produce immunoregulatory glucocorticoids. 1346 *Oncoimmunology* 1, 529-530, doi:10.4161/onci.19459 (2012).
- Verhoeven, G. T. *et al.* Glucocorticoids hamper the ex vivo maturation of lung dendritic cells from their low autofluorescent precursors in the human bronchoalveolar lavage: decreases in allostimulatory capacity and expression of CD80 and CD86. *Clin Exp Immunol* **122**, 232-240, doi:10.1046/j.1365-2249.2000.01354.x (2000).
- Rea, D. *et al.* Glucocorticoids transform CD40-triggering of dendritic cells into an alternative activation pathway resulting in antigen-presenting cells that secrete IL-10. *Blood* **95**, 3162-3167 (2000).
- 1354 244 Kim, K. D., Choe, Y. K., Choe, I. S. & Lim, J. S. Inhibition of glucocorticoid-mediated, caspase-1355 independent dendritic cell death by CD40 activation. *J Leukoc Biol* **69**, 426-434 (2001).
- Herold, M. J., McPherson, K. G. & Reichardt, H. M. Glucocorticoids in T cell apoptosis and function. *Cell Mol Life Sci* **63**, 60-72, doi:10.1007/s00018-005-5390-y (2006).
- Talaber, G. *et al.* Mitochondrial translocation of the glucocorticoid receptor in double-positive thymocytes correlates with their sensitivity to glucocorticoid-induced apoptosis. *Int Immunol* **21**, 1269-1276, doi:10.1093/intimm/dxp093 (2009).
- Lck and Fyn. *Blood* **106**, 1703-1710, doi:10.1182/blood-2004-12-4790 (2005).
- Wu, Y. *et al.* The disbalance of LRP1 and SIRPalpha by psychological stress dampens the clearance of tumor cells by macrophages. *Acta Pharm Sin B* **12**, 197-209, doi:10.1016/j.apsb.2021.06.002 (2022).
- 1366 249 Xie, Y. *et al.* Glucocorticoids inhibit macrophage differentiation towards a pro-inflammatory phenotype upon wounding without affecting their migration. *Dis Model Mech* **12**, doi:10.1242/dmm.037887 (2019).
- 1369 250 Lu, Y. *et al.* Glucocorticoid receptor promotes the function of myeloid-derived suppressor cells by suppressing HIF1alpha-dependent glycolysis. *Cell Mol Immunol* **15**, 618-629, doi:10.1038/cmi.2017.5 (2018).
- 1372 251 Xiang, Z. *et al.* Dexamethasone suppresses immune evasion by inducing GR/STAT3 mediated downregulation of PD-L1 and IDO1 pathways. *Oncogene* **40**, 5002-5012, doi:10.1038/s41388-1374 021-01897-0 (2021).
- Eddy, J. L., Krukowski, K., Janusek, L. & Mathews, H. L. Glucocorticoids regulate natural killer cell function epigenetically. *Cell Immunol* **290**, 120-130, doi:10.1016/j.cellimm.2014.05.013 (2014).
- 1378 253 Quatrini, L. *et al.* Endogenous glucocorticoids control host resistance to viral infection through the tissue-specific regulation of PD-1 expression on NK cells. *Nature Immunology* **19**, 954-962, doi:10.1038/s41590-018-0185-0 (2018).
- Cavalcanti, D. M. *et al.* Endogenous glucocorticoids control neutrophil mobilization from bone marrow to blood and tissues in non-inflammatory conditions. *Br J Pharmacol* **152**, 1291-1300, doi:10.1038/sj.bjp.0707512 (2007).
- Nadkarni, S. *et al.* Investigational analysis reveals a potential role for neutrophils in giant-cell arteritis disease progression. *Circ Res* **114**, 242-248, doi:10.1161/CIRCRESAHA.114.301374 (2014).
- Walther, A., Riehemann, K. & Gerke, V. A novel ligand of the formyl peptide receptor: annexin I regulates neutrophil extravasation by interacting with the FPR. *Mol Cell* 5, 831-840, doi:10.1016/s1097-2765(00)80323-8 (2000).
- 1390 257 Saffar, A. S., Ashdown, H. & Gounni, A. S. The molecular mechanisms of glucocorticoids-1391 mediated neutrophil survival. *Current drug targets* 12, 556-562, 1392 doi:10.2174/138945011794751555 (2011).
- Staedtke, V. *et al.* Disruption of a self-amplifying catecholamine loop reduces cytokine release syndrome. *Nature* **564**, 273-277, doi:10.1038/s41586-018-0774-y (2018).
- 1395 259 Cao, M. et al. Chronic restraint stress promotes the mobilization and recruitment of myeloid-

- derived suppressor cells through beta-adrenergic-activated CXCL5-CXCR2-Erk signaling cascades. *Int J Cancer* **149**, 460-472, doi:10.1002/ijc.33552 (2021).
- 1398 260 Karvonen, H. *et al.* Glucocorticoids induce differentiation and chemoresistance in ovarian cancer by promoting ROR1-mediated stemness. *Cell Death Dis* **11**, 790, doi:10.1038/s41419-020-03009-4 (2020).
- Hara, M. R. *et al.* A stress response pathway regulates DNA damage through beta2-adrenoreceptors and beta-arrestin-1. *Nature* **477**, 349-353, doi:10.1038/nature10368 (2011).
- Sood, A. K. *et al.* Adrenergic modulation of focal adhesion kinase protects human ovarian cancer cells from anoikis. *J Clin Invest* **120**, 1515-1523, doi:10.1172/JCI40802 (2010).
- Dwyer, A. R. *et al.* Glucocorticoid Receptors Drive Breast Cancer Cell Migration and Metabolic Reprogramming via PDK4. *Endocrinology* **164**, doi:10.1210/endocr/bqad083 (2023).
- 1407 264 Karra, A. G. *et al.* Increased Expression of the Mitochondrial Glucocorticoid Receptor Enhances
 1408 Tumor Aggressiveness in a Mouse Xenograft Model. *Int J Mol Sci* **24**,
 1409 doi:10.3390/ijms24043740 (2023).
- Tiwari, R. K. *et al.* Epinephrine facilitates the growth of T cell lymphoma by altering cell proliferation, apoptosis, and glucose metabolism. *Chem Biol Interact* **369**, 110278, doi:10.1016/j.cbi.2022.110278 (2023).
- Talaber, G., Jondal, M. & Okret, S. Local glucocorticoid production in the thymus. *Steroids* **103**, 58-63, doi:10.1016/j.steroids.2015.06.010 (2015).
- 1415 267 Croft, A. P. *et al.* Effects of minor laboratory procedures, adrenalectomy, social defeat or acute alcohol on regional brain concentrations of corticosterone. *Brain Res* **1238**, 12-22, doi:10.1016/j.brainres.2008.08.009 (2008).
- Cima, I. *et al.* Intestinal epithelial cells synthesize glucocorticoids and regulate T cell activation. *J Exp Med* **200**, 1635-1646, doi:10.1084/jem.20031958 (2004).
- 1420 269 Nagashima, H. *et al.* Neuropeptide CGRP Limits Group 2 Innate Lymphoid Cell Responses and Constrains Type 2 Inflammation. *Immunity* 51, 682-695 e686, doi:10.1016/j.immuni.2019.06.009 (2019).
- Reigstad, C. S. *et al.* Gut microbes promote colonic serotonin production through an effect of short-chain fatty acids on enterochromaffin cells. *FASEB J* **29**, 1395-1403, doi:10.1096/fj.14-259598 (2015).
- Yano, J. M. *et al.* Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* **161**, 264-276, doi:10.1016/j.cell.2015.02.047 (2015).
- This study highlights that certain bacterial species can act as important modulators of host 5-HT biosynthesis and its downstream bioactivities.
- Finocchiaro, L. M. *et al.* Serotonin and melatonin synthesis in peripheral blood mononuclear cells: stimulation by interferon-gamma as part of an immunomodulatory pathway. *J Interferon Res* **8**, 705-716, doi:10.1089/jir.1988.8.705 (1988).
- Kushnir-Sukhov, N. M., Brown, J. M., Wu, Y., Kirshenbaum, A. & Metcalfe, D. D. Human mast
 cells are capable of serotonin synthesis and release. *J Allergy Clin Immunol* 119, 498-499,
 doi:10.1016/j.jaci.2006.09.003 (2007).
- Leon-Ponte, M., Ahern, G. P. & O'Connell, P. J. Serotonin provides an accessory signal to enhance T-cell activation by signaling through the 5-HT7 receptor. *Blood* **109**, 3139-3146, doi:10.1182/blood-2006-10-052787 (2007).
- 1439 275 Brenner, B. *et al.* Plasma serotonin levels and the platelet serotonin transporter. *J Neurochem* 1440 102, 206-215, doi:10.1111/j.1471-4159.2007.04542.x (2007).
- 1441 276 Cox, M. A. *et al.* Beyond neurotransmission: acetylcholine in immunity and inflammation. *J Intern Med*, doi:10.1111/joim.13006 (2019).
- Pirzgalska, R. M. *et al.* Sympathetic neuron-associated macrophages contribute to obesity by importing and metabolizing norepinephrine. *Nat Med* **23**, 1309-1318, doi:10.1038/nm.4422 (2017).
- 1446 278 Kiecolt-Glaser, J. K. *et al.* Yoga's impact on inflammation, mood, and fatigue in breast cancer survivors: a randomized controlled trial. *J Clin Oncol* **32**, 1040-1049, doi:10.1200/JCO.2013.51.8860 (2014).
- Stagl, J. M. *et al.* A randomized controlled trial of cognitive-behavioral stress management in breast cancer: survival and recurrence at 11-year follow-up. *Breast Cancer Res Treat* **154**, 319-328, doi:10.1007/s10549-015-3626-6 (2015).
- Teo, I., Krishnan, A. & Lee, G. L. Psychosocial interventions for advanced cancer patients: A systematic review. *Psychooncology* **28**, 1394-1407, doi:10.1002/pon.5103 (2019).
- Dethlefsen, C. *et al.* Exercise-Induced Catecholamines Activate the Hippo Tumor Suppressor Pathway to Reduce Risks of Breast Cancer Development. *Cancer Res* 77, 4894-4904,

1456 1457 1458 1459	282 283	doi:10.1158/0008-5472.CAN-16-3125 (2017). Huang, C. W. <i>et al.</i> Irisin, an exercise myokine, potently suppresses tumor proliferation, invasion, and growth in glioma. <i>FASEB J</i> 34 , 9678-9693, doi:10.1096/fj.202000573RR (2020). Sarkar, D. K., Murugan, S., Zhang, C. & Boyadjieva, N. Regulation of cancer progression by
1460		beta-endorphin neuron. Cancer Res 72, 836-840, doi:10.1158/0008-5472.CAN-11-3292 (2012).
1461	284	Harber, V. J. & Sutton, J. R. Endorphins and exercise. Sports Med 1, 154-171,
1462		doi:10.2165/00007256-198401020-00004 (1984).
1463	285	Hojman, P. et al. Exercise-induced muscle-derived cytokines inhibit mammary cancer cell
1464		growth. Am J Physiol Endocrinol Metab 301, E504-510, doi:10.1152/ajpendo.00520.2010
1465		(2011).
1466	286	Hamy, A. S. et al. Comedications influence immune infiltration and pathological response to
1467		neoadjuvant chemotherapy in breast cancer. Oncoimmunology 9, 1677427,
1468		doi:10.1080/2162402X.2019.1677427 (2020).
1469		
1470		

1472 Figures and legends

1471



1473 1474 1475

1476

1477

1478

1479

1480

1481

1482

1483

1484

1485

1486

1487

1488

1489

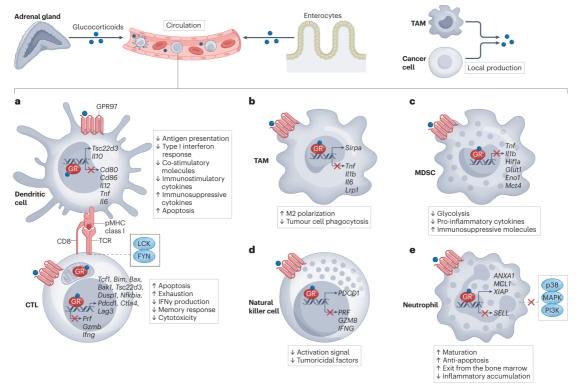
1490

1491 1492

1493

1494

Fig. 1. The effects of stress on cancer. Overview of the intricate relationships between the neuroendocrine system, immune system, metabolic process, gut microbiota, and cancer under conditions of stress. Stress can provoke the SAM and HPA axes for systemic adaptations and stimulate the peripheral nervous system (including sympathetic and parasympathetic nervous system (SNS and PNS)) for localized adjustments. This leads to altered secretion of SAIMs, which engage their corresponding receptors on immune cells and directly reshape the immune landscape of tumours. It is noteworthy that immune cells can also generate SAIMs. Stresstriggered systemic changes, including metabolic reprogramming and dysbiosis of microbiota, can sculpture the tumour macroenvironment. SAIMs are closely involved in the metabolic fluctuations and functional alterations of circulating and tissue-resident immune cells, which can be attributed to changes in their energy supply, macromolecule synthesis, signal transduction, and transcriptional networks. Moreover, some bioactive metabolites generated upon stress can induce PTMs (such as lactylation, O-GlcNAcylation and palmitoylation) of target proteins to enable immunoregulation. Importantly, stress-induced proinflammatory cytokines and neuroendocrine factors often compromise the intestinal integrity and cause microbial imbalance. Structural components from the gastrointestinal microflora (such as PAMPs), immunoregulatory metabolites, and neuroactive factors can stimulate corresponding receptors or sensors on immune cells and modulate their response to cancer. Besides, cancer diagnosis, tumour progression and treatment may lead to long-term alterations in neuroendocrine circuitries, which may associate with dysbiosis and immune dysregulation.



1498

1499

1500

1501

1502

1503

1504

1505

1506

1507

1508

1509 1510

1511

1512

1513

1514

1515

1516

1517

1518

1519

1520

1521

1522

1523

1524

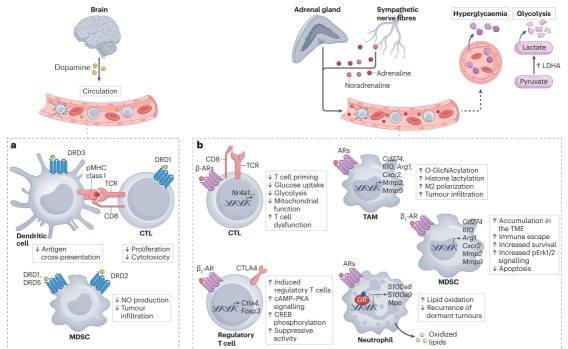
1525

1526

1527

Fig. 2. The effect of glucocorticoids on the immune system and anti-cancer immune responses. Stress-induced elevation of circulating glucocorticoids can be largely attributed to the activation of the HPA axis and increased secretion from the adrenal glands. Moreover, glucocorticoids can be produced by enterocytes in the gut and diffuse into the circulation. During malignant progression, local synthesis of glucocorticoids by tumour cells and tumour-infiltrating myeloid cells is often enhanced^{130,241}. Elevated glucocorticoids in the circulation and in the TME can engage their nuclear receptor (GR) or membrane receptor (GPR97) on immune cells^{73,74}. This leads to transcriptional reprogramming and altered signal transduction¹⁰⁵, thus influencing the viability, differentiation, migration, metabolic patterns, and function of different immune cell populations. a) Glucocorticoids confer immunosuppressive properties on DCs by down-regulating co-stimulatory molecules (such as CD80 and CD86)²⁴², immunostimulatory cytokines (such as IL-12, IL-6 and TNF) and type I IFN response, whilst augmenting the secretion of immunosuppressive cytokines (such as IL-10)²⁴³. Stress-induced GR signalling in DCs can elevate the transcription of Tsc22d3, which impairs type I IFN response and MHC class I/II antigen presentation pathways, thus compromising anti-tumour immune responses⁵². Glucocorticoid-induced downregulation of anti-apoptotic molecules leads to apoptosis of DCs and this process is independent of caspase activation²⁴⁴. By stimulating the expression of BCL-2 interacting mediator of cell death (Bim), which in turn activates BCL2 antagonist/killer 1 (Bak1) and BCL2associated X protein (Bax) in T cells, glucocorticoids can elicit the mitochondrial pathway of apoptosis²⁴⁵. Moreover, translocation of ligand-bound GR to the mitochondria can disrupt their membrane integrity and activate the intrinsic apoptotic pathway in T cells²⁴⁶. Glucocorticoids can rapidly attenuate TCR signalling by abrogating the recruitment and activation of LCK/FYN and blocking the phosphorylation of their downstream signalling intermediates 105,247. They also dampen the effector programmes of T cells by upregulating immunosuppressive genes (Tsc22d3, Dusp1, and Nfkbia) and co-inhibitory molecules (Pdcd1, Ctla4, Lag3), while downregulating multiple co-stimulatory molecules, cytokines, and

chemokines¹⁰⁴. Ligand-engaged GR can drive various transcriptomic and epigenetic 1529 changes that favour the differentiation of memory precursor T cells and T_{reg} cells, 1530 whereas strongly restraining T_H1 differentiation and the effector phenotype of CTLs 1531 (via upregulating TCF1)^{104,130}. b) Liganded GR can disturb TAM-mediated clearance 1532 of tumour cells by the trans-repression of "eat me" signal receptor (low-density 1533 lipoprotein receptor-related protein-1, Lrp1) and the elevation of "don't eat me" signal 1534 receptor (signal regulatory protein α, Sirpa)²⁴⁸. Glucocorticoids can also drive TAMs 1535 towards M2-like phenotype and inhibit the expression of inflammatory cytokines 1536 (such as TNF, IL-1β and IL-6)²⁴⁹. c) GR signalling can promote the 1537 immunosuppressive activity of MDSCs by inhibiting the expression of hypoxia-1538 1539 inducible factor 1-α (Hifla), glycolysis-related genes (such as Glut1, Eno1 and Mct4)²⁵⁰ and pro-inflammatory cytokines (such as Tnf and Il1b)²⁵¹. d) Glucocorticoids 1540 1541 dampen the cytolytic activity of NK cells by reducing histone acetylation at the 1542 promoter or enhancer regions of tumoricidal factors such as perforin (PRF), granzyme B (GZMB), and IFN γ (IFNG)²⁵². They also limit the production of IFN γ by 1543 upregulating checkpoint receptor PD1²⁵³. e) Glucocorticoids can accelerate neutrophil 1544 exit from the bone marrow by reducing the expression of adhesion molecules (such as 1545 PECAM-1, ICAM-1 and VCAM-1) on endothelial cells²⁵⁴ (not shown). They also 1546 block the accumulation of neutrophils in inflamed tissues by downregulating L-1547 selectin (SELL) and upregulating Annexin A1 (ANXA1, which mediates the shedding 1548 of SELL)^{255,256}. Moreover, glucocorticoids exhibit an anti-apoptotic effect on 1549 neutrophils by directly activating p38 MAPK and PI3K and upregulating myeloid cell 1550 leukemia-1 (MCL1) and X-linked inhibitor of apoptosis proteins (XIAP)²⁵⁷. 1551



1553 1554

1555

1556

1557

1558

1559

1560

1561

15621563

1564

1565

1566

1567

1568

15691570

1571

1572

1573

1574

1575

1576

1577

15781579

1580

1581

1582

1583

Fig. 3. The effect of catecholamines on the immune system and anti-cancer immune responses. Stress-induced hyperactivation of the SAM axis can generate high levels of catecholamines, such as dopamine produced in the brain and adrenaline and noradrenaline released from the adrenal glands and sympathetic nerve fibres^{68,69}. Moreover, cancer immunotherapy with oncolytic bacteria or CAR-T cells can trigger a self-amplifying catecholamine loop in myeloid cells, which leads to cytokine release syndrome²⁵⁸ (not shown). Catecholamines can either promote (sometimes) or suppress (generally) tumor immunosurveillance by engaging diversified catecholamine receptors on immune cells. For example, elevation of plasma dopamine can engage DRD1 and increase intracellular cAMP in T cells, hence blocking their proliferation and cytotoxicity¹⁴¹. It can attenuate CTL response against tumours by eliciting DRD3 signalling in DCs and inhibiting their antigen cross-presentation¹⁴². However, dopamine can also stimulate DRD1, DRD5 and DRD2 on MDSCs and unleash antitumour immunity by reducing their accumulation in the TME and production of $NO^{143,144}$. Surges of adrenaline and noradrenaline can stimulate α- or β-adrenaline receptors (ARs) on immune cells. Activation of these G-protein-coupled receptors often leads to impaired priming and metabolic defects of T cells (such as impaired glucose uptake, glycolysis, and mitochondrial function), resulting in T cell dysfunction¹⁴⁵⁻¹⁴⁹. β2-AR activation can facilitate the conversion of naïve T cells to FOXP3⁺ T_{reg} cells and enhance their suppressive activities by increasing intracellular cAMP, PKA-dependent CREB phosphorylation, and CTLA4 expression¹¹⁹. Adrenaline and noradrenaline also promote tumour infiltration and M2-polarization of TAMs, thus accelerating the progression and metastatic spreading of tumours^{150,151}. Stress-induced catecholamines can boost hyperglycemia and glycolysis, which augment O-GlcNAcylation of target proteins and lactylation of histones in TAMs. This favours their M2-like polarization and supports the immune evasion of colorectal cancer in mice and humans ^{172,189}. β2-AR signalling in MDSCs leads to downregulation of apoptosis-related genes and upregulation of arginase-1 and PD-L1, thus favouring their survival and immunosuppressive properties¹⁵². The β2-AR axis also promotes tumour progression via the CXCL5-CXCR2 axis and ERK pathway, which enhance the mobilization of MDSCs and local immunosuppression²⁵⁹. Upon

1585	stimulation with norepinephrine, neutrophils rapidly release S100A8/A9 and activate
1586	myeloperoxidase (MPO) to generate oxidized lipids, which further reactivate dormant
1587	tumour cells ¹⁸⁵ .
1588	
1589	

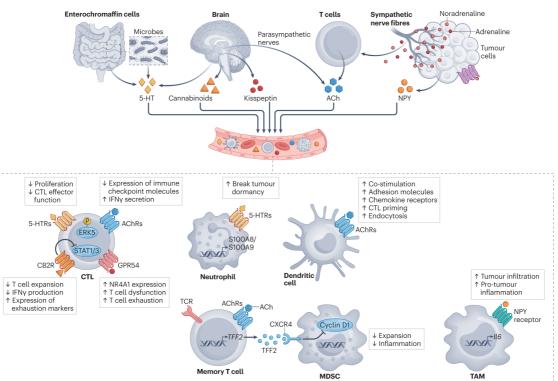


Fig. 4. The effect of other SAIMs on the immune system and anti-cancer immune responses. Acute stress and chronic stress adaptation responses result in the release of SAIMs other than glucocorticoids and catecholamines. For example, 5-HT generated from the brain, enterochromaffin cells, or gut microbes can block the accumulation of CTLs in the TME and reduce their production of IFNy and GZMB²²⁷. Once engaged by endogenous cannabinoids, cannabinoid receptor 2 (CBR2) can bind to JAK1 and inhibit the phosphorylation of STAT1 and STAT3. This causes impeded expansion and antitumor function of CTLs¹⁶⁴. Stress can augment the level of plasma kisspeptin and the expression of its receptor GPR54 on tumor-infiltrating T cells. This leads to T cell dysfunction and exhaustion, due to ERK5-mediated NR4A1 activation ¹⁶². Persistent stress can block the parasympathetic system, vagal responses, and ACh production¹⁶⁵. However, parasympathetic neurostimulation of tumours can significantly reduce the expression of immune checkpoint molecules (such as PD1 and FOXP3) on tumour-infiltrating CTLs and augment their capability to produce IFN₂². 5-HT-triggered rapid release of S100A8/A9 from neutrophils can break tumour dormancy¹⁸⁵ and facilitate tumour immune evasion¹⁸⁶. Agonistic stimulation of nicotinic AChRs on DCs can upregulate the expression of costimulatory molecules, adhesion molecule CD11b, and chemokine receptor CCR7, which boost their capacity of endocytosis, CTL priming and IL-12 secretion 166. Moreover, vagal nerve stimulation can potentiate the secretion of TFF2 from ACh-producing splenic memory T cells, which inhibits the development of colorectal cancer in mice by engaging CXCR4 on MDSCs and inhibits their expansion¹⁶⁷. Besides, noradrenaline-induced production of NPY from tumour cells can recruit macrophages to the TME and boost their secretion of pro-tumoural cytokine IL-6¹⁵³.

Boxes

1590

1591

1592

1593

1594

1595

1596

1597

1598

1599

1600

1601

1602

1603

1604

1605

1606

1607

1608

1609

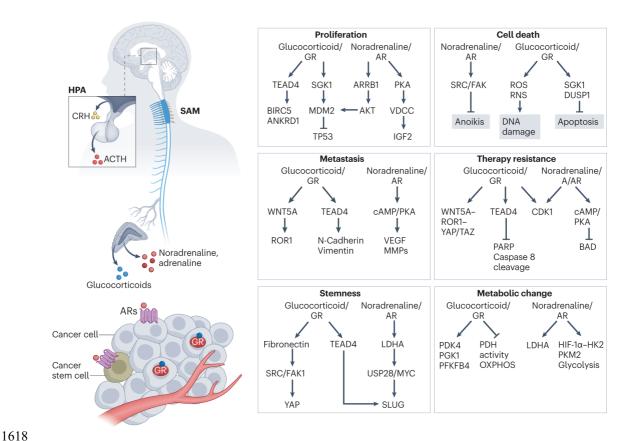
1610

1611

1612

1613

16141615



1619

1620

1621

1622

1623

1624

1625

1626

1627

1628

1629

1630

1631

1632

1633

1634

1635

1636

1637

1638

1639

1640

1641

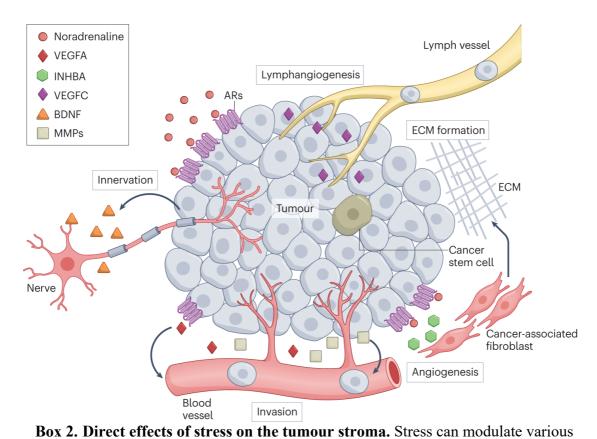
1642

1643

1644

Box 1. Direct effects of stress on cancer cells. Biological features of tumour cells, such as proliferation, cell death, metastasis, therapy resistance, stemness and metabolic changes can be affected by stress-induced neuroendocrine factors. For example, a glucocorticoid surge during chronic restraint can accelerate radiationinduced tumorigenesis by activating the kinase SGK1 and the E3 protein ubiquitin ligase MDM2, which subsequently attenuate the expression and tumour-suppressive function of TP53²⁵. Glucocorticoids can activate the transcription factor TEAD4 in breast cancer cells, which facilitates tumour growth and metastasis by upregulating genes related to proliferation (such BIRC5 and ANKRD1), epithelial-mesenchymal transition, and cancer stem cell traits⁵⁵. Cancer progression-induced increase of glucocorticoids can activate GR and escalate metastatic colonization of breast cancer cells in distant organs by upregulating the expression of receptor tyrosine kinase like orphan receptor 1 (ROR1) and its ligand WTN5A⁵⁶. Glucocorticoid administration also favours the evasion of cell death by elevating the expression of pro-survival and anti-apoptotic proteins (such as SGK1 and DUSP1) on cancer cells⁵⁷. In contrast, stress-induced glucocorticoids and their synthetic mimetics can promote cell death by augmenting the production of reactive oxygen and nitrogen species, which cause DNA damage and impaired DNA repair in human and mouse breast cancer cells⁶⁰. Additionally, glucocorticoids can promote cancer stemness and chemoresistance by activating TEAD4⁵⁵ and YAP^{59,260}. Likewise, catecholamines are closely involved in the initiation, progression, dissemination, and therapeutic resistance of tumours. For example, chronic catecholamine stimulation leads to excessive DNA damage and genomic instability in the thymus and testes by triggering TP53 degradation via β-arrestin-1 (ARRB1)/AKT-mediated MDM2 activation²⁶¹. Noradrenaline can induce PKA-dependent phosphorylation of voltagedependent calcium channels (VDCC), which facilitates the secretion of insulin-like growth factor 2 (IGF2) and lung epithelial cell transformation in mice²⁶. Stress-

induced adrenaline elevates LDHA and enhances stem-like traits of human breast 1646 cancer cells through metabolic rewiring and USP28/MYC/SLUG signalling⁶¹. Also, 1647 noradrenaline exhibits antiapoptotic effects on human prostate and breast cancer 1648 cells, by inactivating the apoptosis promoter BAD⁶². It can also activate SRC-1649 dependent phosphorylation of the kinase FAK and render human ovarian cancer 1650 cells resistant to anoikis²⁶². Glucocorticoid- and catecholamine-induced metabolic 1651 reprogramming also favour tumour growth and metastasis, owing to the 1652 transcriptional regulation genes related to glycolysis and oxidative phosphorylation 1653 (OXPHOS), such aspyruvate dehydrogenase kinase 4 (PDK4), phosphoglycerate 1654 kinase 1 (PGK1), (6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 4) 1655 PFKFB4²⁶³, pyruvate dehydrogenase (PDH)²⁶⁴, hexokinase 2 (HK2), pyruvate 1656 kinase M2 (PKM2)²⁶⁵, and LDHA⁶¹. 1657 1658



stromal components of tumours. In preclinical models of ovarian, breast, and colon cancers, noradrenaline-stimulated secretion of inhibin β A (INHBA) from cancer cells can activate cancer-associated fibroblasts (CAFs), leading to increased collagen deposition and extracellular matrix formation 63 . In mouse models of ovarian and nasopharyngeal cancer, stress-induced activation of β 2-AR signalling can increase the expression of vascular endothelial growth factor-A (VEGF-A) and matrix metalloproteinases (MMPs) on tumour cells, which promote angiogenesis and malignant outgrowth 47,64 . The catecholamine receptor β 2-AR on vascular endothelial cells can also promote angiogenesis by directly forming a complex with VEGFR2 66 . Moreover, noradrenaline can stimulate local lymphogenesis and

enhance metastatic progression by inducing the secretion of PGE2 and VEGF-C from TAMs and malignant cells⁴⁹. It is noteworthy that sustained adrenergic

signalling can stimulate the production of brain-derived neurotrophic factor (BDNF) from mouse ovarian and colon cancer cells, which promotes axonogenesis in the TME and favours tumour growth and metastasis⁶⁷.

Box 3. Peripheral sources of neuroendocrine factors

Steroidogenesis and glucocorticoid production has been observed in the thymus²⁶⁶, brain²⁶⁷, and in colon epithelial cells^{92,268}. Glucocorticoid production by tumour-infiltrating myeloid cells has been reported to facilitate the emergence of dysfunctional CD8⁺ T cells¹³⁰. Myeloid cells expressing tyrosine hydroxylase largely contribute to the catecholamine surge in response to T-cell-activating therapies, which augments pro-inflammatory circuitries and can cause cytokine release syndrome²⁵⁸. In the lung, pulmonary neuroendocrine cells are rich sources of GABA

and calcitonin gene-related peptide (CGRP), which can stimulate neighbouring group 2 innate lymphoid cells (ILC2s) and modulate allergic responses^{90,269}. In the GI tract, catecholamines from sympathetic nerves can engage basolateral \alpha2-adrenoreceptors and trigger 5-HT release from adjacent chromogranin A⁺ enterochromaffin cells⁹¹. Enterochromaffin cell-mediated production of 5-HT can be enhanced by short-chain fatty acids (SCFAs) from the gut microbiota²⁷⁰, as well as specific metabolites generated by indigenous spore-forming bacteria in the gut (such as α-tocopherol, paminobenzoate, and tyramine)²⁷¹. Moreover, 5-HT biosynthesis has been documented in various immune cell subsets, including mast cells, monocytes, and T cells²⁷²⁻²⁷⁴. Platelets can store large quantities of 5-HT in their dense granules and secret these upon activation²⁷⁵. Moreover, T cells can generate ACh, which induces local vasodilation and allows for T cell extravasation²⁷⁶. Histamine released by mast cells can modulate allergic and inflammatory responses⁸². Intriguingly, immune cells may also enable the local clearance of neuroendocrine factors, as exemplified by adipose-tissue associated macrophages and sympathetic neurons that express the noradrenaline transporter as well as enzymes that degrade noradrenaline²⁷⁷.

Box 4. Stress management affects cancer immunosurveillance

1688

1689

1690

1691

1692

1693

1694

1695

1696

1697

1698

1699

1700

1701

1702

17031704

1705

1706

1707

1708

1709

1710

1711

1712

1713

1714

1715

1716

1717

1718

1719

1720

1721

1722

17231724

Stress management with physical, mindfulness-based, or cognitive-behavioural approaches (such as yoga, Tai-Chi, other stress-reducing exercises, meditation, and relaxation) has been associated with immunological changes, such as reduced inflammation, increased T_H1 responses, and enhanced NK cell activity²⁷⁸. Psychological interventions were shown to significantly reduce the risk of mortality and cancer recurrence in patients with early-stage breast cancer, when provided postsurgery and prior to adjuvant therapies²⁷⁹. These regimens also improve the quality of life and subjective well-being of patients with cancer²⁸⁰. Of note, running-induced catecholamines (which cannot be categorized as a 'psychological stress') can activate YAP/TAZ phosphorylation in breast cancer cells and inhibit tumour outgrowth in immunodeficient mice²⁸¹. This indicates that the short-term peaks of catecholamines, endogenous opioids, and myokines following exercise may prevent the development and progression of cancer²⁸²⁻²⁸⁵. Intriguingly, some psychiatric drugs can exhibit immunostimulatory properties (such as influencing the density of tumour-infiltrating lymphocytes). For example, in patients with triple negative breast cancer, the usage of psycholeptics or psychoanaleptics was associated with higher rate of pathological complete responses²⁸⁶. At present, it remains unclear which cell subsets and receptors respond to these drugs and whether their impact on the TME and cancer outcome is on-target or off-target.