

# Ghrelin proteolysis increases in plasma of men, but not women, with obesity

Antonela S Fittipaldi, Daniel Castrogiovanni, Daniela Lufrano, Camila Saenz, Pablo N de Francesco, Tyler Lalonde, Leonard G Luyt, Sonia Cantel, Jean-Alain Fehrentz, María F Andreoli, et al.

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3	Ghrelin Proteolysis Increases in Plasma of Men, but not Women, with Obesity
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5	Running title: ghrelin proteolysis in human obesity
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34 Abstract

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Aims: Since plasma ghrelin can undergo des-acylation and proteolysis, the aim of this study
 was to investigate the extent to which an enhancement of these reactions is associated to
 the decrease of ghrelin in plasma after food intake or in individuals with obesity.

Main methods: we performed an intervention cross-sectional study, in which levels of
 ghrelin, desacyl-ghrelin (DAG), glucose, insulin, ghrelin des-acylation and ghrelin
 proteolysis were assessed in plasma before and after a test meal in 40 people (n=21 males)
 with normal weight (NW, n=20) or overweight/obesity (OW/OB, n=20).

43 Key findings: Preprandial ghrelin and DAG levels were lower, whereas preprandial ghrelin 44 proteolysis was ~4.6-fold higher in plasma of males with OW/OB. In males, ghrelin 45 proteolysis positively correlated with glycemia. Ghrelin and DAG levels were also lower in 46 females with OW/OB, but preprandial ghrelin proteolysis was not different between females 47 with NW or OW/OB. Ghrelin and DAG levels decreased postprandially in males and females, 48 independently of BMI, and ghrelin proteolysis increased postprandially ~2 folds only in 49 individuals with NW. Ghrelin des-acylation remained unaffected by BMI or feeding status in 50 both sexes.

51 <u>Significance</u>: Current study shows that ghrelin proteolysis increases in males with obesity 52 as well as after meal in lean individuals. Therefore, ghrelin proteolysis may be an important 53 checkpoint and, consequently, a putative pharmacological target to control circulating 54 ghrelin levels in humans.

- 55
- 56 **Key words**: ghrelin proteolysis, ghrelin des-acylation, obesity.

## 57 Introduction

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59 Ghrelin is a 28-residue peptide hormone predominantly secreted by cells of the 60 stomach that displays several unique biochemical and biological features <sup>1</sup>. Chemically, 61 ghrelin is the only peptide known to be acylated with an octanoic moiety, a posttranslational 62 modification that takes place at Ser3 of ghrelin and is catalysed by the enzyme ghrelin-O-63 acyltransferase (GOAT) early in the biosynthetic processing within the ghrelin-producing 64 cells<sup>2</sup>. The biological actions of ghrelin, which are mediated by the growth hormone 65 secretagogue receptor (GHSR), entirely depend on the octanoylation of ghrelin and include 66 the most potent orexigenic effect described to date <sup>3,4</sup>. Ghrelin also induces growth hormone 67 and glucocorticoid secretion, recruits several mechanisms that increase glycemia, increases 68 gastrointestinal motility and affects non-homeostatic aspects of food intake, among other 69 effects <sup>1</sup>. Thus, ghrelin is a pleiotropic hormone that contains an exceptional 70 posttranslational modification that is essential for its bioactivity.

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72 Plasma ghrelin level is tightly regulated by the daily feeding status and by the long-73 term energy balance. In the short term, ghrelin level increases before meals and decreases 74 to baseline levels within the first hour after meals <sup>5</sup>. Ghrelin level increases in prolonged 75 energy deficit conditions, such as fasting, calorie restriction or anorexia nervosa <sup>6,7</sup>, when a 76 rise in plasma ghrelin contributes to seek for food and maintain glycemia. In contrast, ghrelin 77 level decreases under energy surplus conditions, such as obesity<sup>8,9</sup>, and such decrease 78 presumably results as a compensatory adaptation aiming to reduce feeding and glycemia. 79 As other hormones, plasma ghrelin level depends on the equilibrium among its secretion, 80 degradation and clearance rates. A large body of studies characterized the mechanisms 81 controlling the biosynthesis and release of ghrelin from ghrelin cells mainly using animal 82 models or *in vitro* systems <sup>10</sup>. Conversely, the molecular mechanisms controlling the 83 disappearance of ghrelin from the circulation have been less investigated. Ghrelin rapidly 84 disappears from plasma, having a half-life of 9-13 min after a single bolus injection of the 85 hormone <sup>11-13</sup>, and the key chemical reactions affecting plasma ghrelin involve not only des-86 acylation but also proteolysis. Ghrelin des-acylation in plasma results in the generation of 87 desacyl-ghrelin (DAG), which is also secreted from ghrelin cells. DAG does not bind to 88 GHSR at physiological levels, but it has been suggested to display some GHSR-89 independent effects on insulin secretion, osteoblast growth and lipid metabolism <sup>1</sup>. Ghrelin 90 des-acylation in plasma is presumably mediated by different esterases including platelet 91 activating factor acetylhydrolase, paraoxonase, a2-macroglobulin, butyrylcholinesterase

92 and acyl protein thioesterase 1; although it is likely that other esterases can also hydrolyze 93 the lipid moiety from ghrelin <sup>14-19</sup>. Ghrelin also undergoes proteolysis in plasma giving rise to 94 shorter peptides <sup>15,16</sup>. Ghrelin proteolysis mainly occurs at the middle segment of the peptide 95 (i.e. between Arg15 and Lys16) and generates peptides of different length, including 96 peptides containing the N-terminal end of ghrelin that may be able to evoke some activation 97 of GHSR although less potently than full-length ghrelin <sup>16,20,21</sup>. Ghrelin proteolysis was 98 suggested to involve endopeptidase activated protein C in bovine plasma<sup>22</sup>, and insulin-99 degrading enzyme in an *in vitro* system<sup>23</sup>. Still, the molecular identity of the enzymatic 100 system mediating ghrelin inactivation in plasma remains to be fully clarified.

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102 Since its discovery, manipulating ghrelin system emerged as a putative strategy in 103 the treatment of obesity and related diseases. In order to further reduce ghrelin actions in 104 obesity and achieve additional beneficial effects, GHSR antagonists, ghrelin vaccines and 105 GOAT inhibitors have been developed; however, none of these strategies reached the 106 desired effects in human trials<sup>24</sup>. Notably, the inhibition of the extracellular inactivation of 107 incretins has proved to be a critical strategy to improve glycaemic control in patients suffering 108 type 2 diabetes <sup>25</sup>. In this context, understanding the molecular mechanisms controlling the 109 level of ghrelin in plasma emerges as a fundamental step forward to not only better 110 understand the biology of ghrelin but also find new pharmacological strategies to manipulate 111 such levels and achieve beneficial effects. To the best of our knowledge, the extent to which 112 des-acylation or proteolysis of ghrelin in plasma are affected by the daily feeding status or 113 obesity has not been previously investigated. Here, we hypothesized that decrease of 114 plasma ghrelin in individuals with obesity and/or in the postprandial state could be due to an 115 increase in its extracellular processing. In order to test our hypothesis, we performed a single 116 test meal study in individuals with either normal weight (NW) or with overweight or obesity 117 (OW/OB) and assessed not only levels of ghrelin and DAG in plasma but also the specific 118 levels of ghrelin des-acylation and ghrelin proteolysis in plasma.

- 119 Materials and Methods
- 120

2.1 Study design. The study was approved by the Institutional Ethical Committee of
 San Martin Hospital, La Plata (Protocol ID HSMLP2020/0044) and conducted according to
 the Declaration of Helsinki guidelines and Argentinian legal provisions governing clinical
 research in humans. Written informed consents were obtained from all participants.
 Participants were recruited via advertising in social media from August to October 2021.

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2.2 Study participants. This intervention cross-sectional study included 20–45 years old adults, without chronic medical illness, personal history of diabetes, history of medical
 condition, or medication related to obesity or diabetes risk status. The study included 21
 males and 19 females. Pregnant women were excluded from the study. Sample size was
 calculated to unmask differences in plasma ghrelin levels, assuming a standard deviation of
 3.26 fmol/ml <sup>26</sup>, 80% statistical power and 5% significance level in preprandial ghrelin level,
 leading to a minimal sample size of 40 participants (20 with NW and 20 with OW/OB).

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135 2.3 Study protocol. All participants arrived at the facilities of the Institute in the 136 morning after a 12-h overnight fast and underwent an initial evaluation, including a medical 137 and family history. Weight and body composition were measured with the participants barefoot and wearing minimal clothing, on a bioimpedance scale (Omron HBF514, 138 Argentina) with a resolution of 10 g. Height was measured with a Harpenden-type wall-139 140 mounted stadiometer (Holtain Ltd., United Kingdom) with a resolution of 1 mm. BMI was calculated by dividing weight by height squared (kg/m<sup>2</sup>). As described by the World Health 141 142 Organization, participants were classified as NW when BMI was between 18.5 and 24.99 Kg/m<sup>2</sup> or as OW/OB when BMI≥25 Kg/m<sup>2</sup> <sup>27</sup>. Next, a preprandial blood sample was obtained. 143 144 Participants were given a test meal (breakfast) that provided 20% of the individual's daily 145 energy requirement as estimated using the "Body Weight Planner" digital application of the 146 National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK, USA)<sup>28</sup>. The 147 breakfast composition was 56 kcal% carbohydrates, 12 kcal% protein and 34 kcal% lipids 148 and comprised yogurt, cereals and almonds. Participants were asked to consume the entire 149 breakfast in less than 15 min. Blood sampling was repeated at 60 min after starting breakfast 150 (postprandial), when plasma ghrelin level is significantly reduced, according to a meta-151 analysis study 5. Blood samples were collected in tubes containing either heparin, to assess glucose and insulin, ghrelin proteolysis and ghrelin des-acylation, or EDTA (1 mg/mL final) 152 153 and p-hydroxy-mercuribenzoic acid (0.4 mM final), to assess ghrelin and DAG. For ghrelin

and DAG assays, plasma was immediately acidified with HCl (0.1 N final) <sup>29</sup>. All samples
 were stored frozen at -80°C until analysis.

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157 2.4 Biochemical analyses. Glycemia was measured using an enzymatic procedure 158 in a CM 250 (Wiener Lab, Argentina). Insulin level was assessed using chemiluminescence (Advia Centaur XP, Siemens, Germany). Insulin resistance was estimated by the 159 160 homeostasis model assessment for insulin resistance (HOMA index) <sup>30</sup>. Ghrelin and DAG were assessed using specific enzyme immunoassays from Bertin Bioreagent (A05306 and 161 162 DAG, respectively), and following manufacturer's instructions. Ghrelin/DAG ratio was 163 computed on a molar basis as ghrelin concentration divided by DAG concentration. The food intake-induced change of plasma ghrelin was calculated as preprandial minus postprandial 164 165 ghrelin concentration in molar basis.

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2.5 Assessment of ghrelin des-acylation in plasma. Here, we used a biotinylated 167 analogue of ghrelin consisting in the first 11 residues of ghrelin with the octanoyl moiety 168 linked to Ser3 and a biotin attached to the *ɛ*-amino group of Arg (biotin-ghrelin or 169 170 GSS(Oct)FLSPEHQR-biotin). To assess ghrelin des-acylation in plasma (Fig. 1A), biotin-171 ghrelin (0.01  $\mu$ g/ $\mu$ L) was incubated with each plasma sample (20 % v/v) in phosphate buffer 172 saline at 37 °C. After 60 min, trifluoroacetic acid (20 % final) was added to the tube to stop 173 the reaction. We next used a reverse-phase chromatographic procedure to separate biotin-174 ahrelin from biotin-DAG, based on a previously described protocol<sup>2</sup>. Briefly, reaction mixtures for each plasma sample were applied to a ZipTip with C18 resin, according to 175 manufacturer's instructions, and eluted with a 20% and 100%-CH<sub>3</sub>CN solutions in 0.1% TFA 176 177 in order to elute biotin-DAG and biotin-ghrelin, respectively. After CH<sub>3</sub>CN evaporation, eluted 178 samples were subjected to dot blot using PVDF membranes. Next, blot membranes were 179 incubated with Vectastain Elite ABC kit (Vector Laboratories, cat # PK6200) for 2 h, 180 according to manufacturer's protocols, and the presence of biotin-ghrelin and biotin-DAG was visualized using a 3-3'-diaminobenzidine (DAB)/Nickel solution, which gives a 181 182 black/purple precipitate. DAB signal per spot was registered using a Gel Doc XR (Bio-Rad) and quantified using the software Fiji. Ghrelin des-acylation was calculated as the ratio 183 184 between the intensity of biotin-DAG and biotin-ghrelin and expressed as relative units (r.u.). The accuracy of the separation and quantification system was checked using solutions with 185 186 different concentrations of either intact biotin-ghrelin and biotin-DAG.

188 2.6 Assessment of ghrelin proteolysis in plasma. Here, we used a fluorescent 189 analogue of human ghrelin (F-ghrelin) consisting in the first 19 residues of ghrelin with the 190 octanoyl moiety linked to a diaminopropanoic acid (Dpr) at position 3 and a fluorescein 191 attached to the ε-amino of Lvs19 isothiocyanate group 192 (GSDpr(Oct)FLSPEHQRVQQRKESK(FITC)-NH<sub>2</sub>), which was synthesized as previously described <sup>31</sup>. To assess ghrelin proteolysis in plasma (Fig. 1B), F-ghrelin (0.025  $\mu$ g/ $\mu$ L) was 193 194 incubated with each plasma sample (12.5 % v/v) in phosphate buffer saline at 37 °C. After 195 180 min, loading buffer was added to the tubes and reaction mixtures for each sample were 196 loaded onto 20% polyacrylamide gels prepared according to the Ornstein-Davis 197 discontinuous system. Fluorescence was visualized by trans-UV exposure, registered using a 520DF30 emission filter (Gel Doc XR, Bio-Rad) and guantified using the software Fiji. 198 199 Ghrelin proteolysis was calculated as the ratio between the fluorescence of the shorter 200 peptide products of cleavage and the fluorescence of intact F-ghrelin, and expressed as 201 relative units (r.u.).

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2.7 Statistics and data analyses. Statistical analyses were performed using 203 204 GraphPad Prism 9.0.0 Normal distribution of data was tested using the Shapiro-Wilk test. 205 Preprandial characteristics in males were compared with Student's unpaired t test or Mann 206 Whitney test. Positively skewed variables (ghrelin, DAG, ghrelin/DAG, and ghrelin 207 proteolysis) were log-transformed for all subsequent analyses. Changes in time between groups were evaluated with two-way repeated measures analysis of variance (ANOVA), with 208 group (NW vs. OW/OB) as between factor and feeding condition (preprandial vs. 209 postprandial) as within factor. Linear correlations between log-transformed ghrelin 210 211 proteolysis and other variables were assessed using Pearson's (ghrelin, BMI and glycemia) or Spearman's (DAG) correlation coefficients. Tests used for each comparison were 212 213 indicated in each figure legends. Variables with normal distribution were expressed as the 214 mean ± standard deviation (SD), and variables with non-normal distribution were reported as median (interguartile range [IQR]). Differences were considered statistically significant 215 216 when p<0.05.

- 217 Results
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3.1 Ghrelin level decreases whereas ghrelin proteolysis increases in preprandial 219 plasma of males with OW/OB. Initially, male participants were grouped as NW (n=9) or 220 221 OW/OB (n=12). As shown in Table 1, age did not differ between groups nor correlate with any assessed parameter, whereas BMI, preprandial glycemia, preprandial insulin levels and 222 HOMA index were significantly higher in OW/OB group, as compared to NW group. 223 Conversely, preprandial ghrelin (Fig. 2A) and DAG (Fig. 2B) levels were ~2.2- and ~6.0-fold 224 lower, respectively, in OW/OB group, as compared to NW group. Ghrelin/DAG ratio (Fig. 225 2C) was ~8.5-fold higher in OW/OB group, as compared to NW group, suggesting that the 226 mechanisms controlling plasma ghrelin level are differentially affected in males with OW/OB. 227 Thus, we assessed the capacity of plasma samples to des-acylate a biotinylated ghrelin 228 analogue. We found that ghrelin des-acylation was not different between plasma samples 229 of NW and OW/OB groups (Fig. 2D). Next, we assessed the capacity of plasma samples to 230 proteolyze a fluorescent ghrelin analogue. Here, ghrelin proteolysis was ~4.6-fold higher in 231 232 OW/OB group, as compared to NW group (Fig. 2E). Notably, ghrelin proteolysis negatively correlated with plasma levels of ghrelin and DAG (Fig. 3A-B), it did not correlate with age 233 (r=0.223, p=0.3317, not shown), plasma insulin levels (r=0.269, p=0.239, not shown), HOMA 234 index (r=0.303, p=0.182, not shown) nor BMI (Fig. 3C) and it positively correlated with 235 236 glycemia (Fig. 3D).

237

3.2 Postprandial ghrelin proteolysis in plasma increases in males with NW, but not 238 with OW/OB, whereas postprandial ghrelin des-acylation remains unaffected. After the 239 single test meal intervention, glycemia was not affected by feeding condition (p condition= 240 0.820), and remained higher in OW/OB group, as compared to NW group (p group=0.012). 241 Plasma levels of ghrelin and DAG changed in a group- and feeding condition-dependent 242 manner, being decreased in OW/OB group and postprandially (Fig. 4A-B). Here, the food 243 intake-induced change of plasma ghrelin level tended to be lower in OW/OB group, as 244 compared to NW group (6.5±5.7 vs. 10.9±7.8 fmol/mL, respectively, P=0.0765). Notably, 245 ghrelin des-acylation remained unaffected by group or feeding condition (Fig. 4C). Ghrelin 246 proteolysis also showed a significant interaction between the effects of group and feeding 247 condition, being ~1.6-fold increased after food intake only in NW group and increased in 248 OW/OB group, independently of the feeding condition (Fig. 4D). 249

251 <u>3.3 Postprandial ghrelin proteolysis increases in NW females but remains unaffected</u>

252 in plasma of females with OW/OB, independently of the feeding condition. Since some 253 evidence indicate that ghrelin level fluctuates in a sex-dependent manner in humans <sup>32,33</sup>, 254 we also studied females under the same study protocol and grouped them as NW (N=11) 255 or OW/OB (n=8). As shown in Table 2, age and preprandial glycemia did not differ between 256 groups whereas BMI, preprandial insulin levels and HOMA index were significantly higher 257 in OW/OB group, as compared to NW group. In preprandial condition, ghrelin and DAG 258 levels were ~2.0- and ~2.6-fold lower, respectively, in OW/OB group, but ghrelin/DAG ratio 259 was not different between both groups, suggesting that the mechanisms controlling plasma 260 ghrelin level in females with OW/OB were less affected than in males with OW/OB. Here, 261 ghrelin des-acylation and ghrelin proteolysis in plasma were not different between NW and 262 OW/OB groups (Table 2). In contrast to males, ghrelin proteolytic activity in plasma did not 263 correlate with glycemia in females (r=0.154, p=0.530).

After the test meal intervention, plasma levels of ghrelin and DAG changed in a group- and feeding condition-dependent manner, being decreased in OW/OB group and postprandially (Fig. 5A-B). Here, the food intake-induced change of plasma ghrelin level was lower in the OW/OB group, as compared to NW group (10.2±8.3 vs. 26.4±16.75 fmol/mL, respectively, P=0.0230). As found in males, ghrelin des-acylation was unaffected by the feeding condition in NW and OW/OB groups (not shown) whereas ghrelin proteolysis was postprandially increased ~2.2 folds only in NW group (Fig. 5C).

272 Discussion

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274 Here, we performed a single test meal study in individuals with NW or OW/OB and 275 investigated levels of ghrelin, ghrelin des-acylation and ghrelin proteolysis in plasma. We 276 confirmed that preprandial ghrelin level is lower in individuals with OW/OB, independently of sex, as previously shown <sup>5,8,9,32,34-38</sup>. After food intake, ghrelin level decreased in males 277 278 and females of NW and OW/OB groups. Interestingly, ghrelin proteolysis was higher in plasma of males, but not females, with OW/OB and correlated with glycemia. Ghrelin 279 280 proteolysis increased after food intake in individuals with NW only. In contrast, ghrelin des-281 acylation was similar in plasma of individuals with NW or OW/OB and remained unchanged 282 after food intake.

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The current study was carefully designed to assess the chemical reactions affecting 284 285 plasma ghrelin at two key time points: in the morning after overnight fasting, when plasma ghrelin is around the highest diurnal level<sup>8</sup>, and 60 min after meal, when plasma ghrelin 286 decreases <sup>5</sup>. Indeed, we detected the expected postprandial decline in ghrelin level in most 287 lean individuals (19 out of 20), as shown in several previous reports <sup>5,32,35-38</sup>, confirming that 288 289 plasma ghrelin level was studied when it undergoes a significant change. It is important to 290 highlight that it is now well established that the inaccurate manipulation of plasma samples 291 results in the conversion of most ghrelin into DAG and a consequent low ghrelin/DAG ratio 292 <sup>39</sup>. Here, plasma ghrelin/DAG ratio in NW individuals was ~0.6, which is similar to the highest 293 reported ghrelin/DAG ratios in similar studies (see for instance <sup>40</sup>), indicating that the 294 conditions of sampling and storage were accurate to preserve ghrelin in plasma despite the 295 complexity involved in clinical studies. Finally, current study was performed in relatively 296 young individuals in order to minimize the putative impact of the age-dependent decline in 297 plasma ghrelin that has been observed older individuals <sup>41</sup>.

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299 In order to specifically assess ghrelin des-acylation or proteolysis in plasma, we set 300 up two reproducible assays whose characteristics should be taken in consideration when 301 interpreting current findings. Importantly, we used two different labelled variants of ghrelin that rely on modifications at its C-terminal end, which is not essential for bioactivity <sup>20,42</sup>. To 302 303 assess ghrelin des-acylation, we used biotin-ghrelin, a 11-residue analogue of ghrelin 304 biotinylated at the Arg11 C-terminus. Biotin-ghrelin is susceptible to hydrolysis by esterases, 305 since the octanoyl moiety is coupled at Ser3 through an ester bond, but presumably resistant 306 to plasma proteases, since ghrelin1-11 is the shorter derivative of ghrelin found after

proteolysis of full-length ghrelin<sup>22</sup>. To assess ghrelin proteolysis, we used F-ghrelin, an 18-307 308 residue analog of ghrelin conjugated to fluorescein isothiocyanate through a Lys added at 309 its C terminus. F-ghrelin contains the entire peptide segment susceptible to proteolysis, 310 according to previous descriptions <sup>22</sup>, but is resistant to esterases because Ser3 was 311 replaced with Dpr, which is coupled with the octanoyl moiety through an amide bond <sup>31</sup>. Of 312 note, F-ghrelin and the fluorescent products of proteolysis were directly visualized and 313 quantified after separation by native gel electrophoresis. In contrast, visualization and 314 quantification of biotin-ghrelin and biotin-DAG required several steps including reverse-315 phase chromatographic procedure for separation, dot-blot and chromogenic staining. Thus, 316 it is possible that the method used to assess ghrelin des-acylation lacked enough sensitivity 317 to unmask small differences in enzymatic activity. Also, reaction times to estimate each 318 enzymatic activity were set up in order to have detectable amounts of substrate and products 319 of each assay. Thus, ghrelin des-acylation and ghrelin proteolysis assays required 60 and 320 180 min of reaction time, respectively. The observation that the hydrolysis of peptide bonds 321 of ghrelin in plasma is a relatively slow process would suggest that changes in ghrelin proteolysis have stronger implications in long-term conditions, as we observed here in males 322 323 with OW/OB. In contrast, the physiological implications of the detected postprandial increase 324 of ghrelin proteolysis in plasma of lean individuals is uncertain given the difference between 325 the timing for the rapid postprandial decrease of plasma ghrelin and the reaction time 326 required to assess ghrelin proteolysis in our experimental conditions.

327

328 To the best of our knowledge, the current study provides the first indication that proteolysis of ghrelin in plasma increases in males with OW/OB. The observation that ghrelin 329 330 proteolysis negatively correlates with plasma levels of ghrelin and DAG in the preprandial 331 state suggests that increased ghrelin proteolysis in males with OW/OB may contribute to 332 reduce plasma levels of these peptides. Also, increased ghrelin proteolysis in obesity may 333 contribute to impair the effects of ghrelin treatment (the so-called ghrelin resistance), reported in obese mice <sup>43</sup>. Strikingly, we found here that ghrelin proteolysis positively 334 335 correlated with glycemia, but did not correlate with insulinaemia or HOMA index. Conversely, preprandial ghrelin proteolysis and glycemia remained unaffected in females with OW/OB. 336 337 despite they showed reduced preprandial plasma ghrelin level and insulin resistance, as compared to females with NW. Thus, preprandial ghrelin proteolysis is somehow associated 338 339 to glucose level itself, via mechanisms that do not seem to involve insulin. The reasons why ghrelin proteolytic activity increases only in males with OW/OB are uncertain, but it is in line 340 341 with the observation that ghrelin level in obesity tends to be lower in males than females, as

found here and reported in previous studies <sup>32,33</sup>. Interestingly, insulin-degrading enzyme level in plasma was found higher in individuals (of undisclosed sex) with metabolic syndrome and strongly correlated with the severity of the symptoms <sup>44</sup>. Also, plasma activated protein C level was higher in individuals (male and female combined) with obesity and reduced after bariatric surgery <sup>45,46</sup>. Thus, some proteases able to cleave ghrelin are strong candidates to mediate increased ghrelin proteolysis in plasma of male individuals with OW/OB.

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Current study also provides the first indication that ghrelin proteolysis increases in 349 plasma after food intake in lean individuals, in a sex-independent manner, allowing to 350 351 hypothesize that increased postprandial ghrelin proteolysis in lean individuals contributes to 352 decrease plasma ghrelin levels after food intake. Since glycemia is well-known to transiently 353 increase after food intake, it is possible that the postprandial increment of ghrelin proteolysis is also linked to glucose levels. In line with this possibility, a previous study found that a 354 glucose load potently decreases ghrelin levels in lean individuals <sup>31</sup>. Further studies 355 assessing blood samples at earlier time points will be required to test the extent to which 356 357 postprandial increments of glycemia and ghrelin proteolysis correlate. Interestingly, ghrelin 358 proteolysis did not increase after food intake in plasma of individuals with OW/OB, in which 359 plasma ghrelin level shows a smaller decrease after meal intake, as previously shown <sup>5</sup> and 360 confirmed here. Thus, it is likely that the lack of postprandial increase in ghrelin proteolysis in obesity contributes to impair the postprandial reduction of ghrelin levels, an alteration that 361 362 has been proposed to reduce satiety feelings in individuals with obesity <sup>47-49</sup>.

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Although informative, this study is not without limitations. For instance, some aspects 364 365 of our experimental design, such as the postprandial blood sample collection time or the 366 sample size, were based on previous studies investigating the postprandial decline in ghrelin level <sup>5,32,35-38</sup>. However, such experimental design may have prevented us from detecting the 367 368 postprandial change of ghrelin proteolytic activity in individuals with OW/OB, if an increase in such enzymatic activity takes place with a different kinetics or if it requires a higher sample 369 370 size to be unmasked. Also, the time chosen to collect the postprandial blood sample did not 371 allow us to detect the transient meal-induced increase of glycemia and, consequently, we 372 could not evaluate if postprandial hyperglycaemia either was differentially affected in individuals with OW/OB or correlated the postprandial increase of ghrelin proteolytic activity 373 374 in lean individuals. The study did not include "sham-feeding" control groups and, 375 consequently, a postprandial increase of ghrelin proteolytic activity may be related to its

physiological diurnal rhythm. Despite these limitations, the current study was very valuableto gain insights about a novel aspect of the biology of ghrelin in human plasma.

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Ghrelin des-acylation in plasma is fast and affects the circulating ghrelin level <sup>50</sup>. 379 Indeed, intravenous administration of ghrelin in humans results in a peak of ghrelin levels, 380 at 15 min after injection, and is rapidly eliminated <sup>11,40</sup>. Ghrelin des-acylation seems to be 381 the main mechanism mediating the reduction of plasma ghrelin elevation since the referred 382 increase of ghrelin level is followed by a peak of DAG level, which then disappears from 383 plasma more slowly than ghrelin <sup>11,40</sup>. Here, we found that ghrelin des-acylation was 384 unaffected in individuals with OW/OB or postprandially suggesting that des-acylation is not 385 a key mechanism mediating the decrease of plasma ghrelin level in those metabolic 386 conditions. In line with our observations, it was reported that the level of serum 387 butyrylcholinesterase activity, a ghrelin esterase in humans, was not altered during fasting 388 <sup>51</sup>. Since our results indicate that ghrelin proteolysis is an important mechanism affecting the 389 decrease of plasma ghrelin level in males with OW/OB, the mechanisms mediating the 390 decrease of plasma ghrelin level in females with OW/OB remain uncertain. Given the fast 391 rate of ghrelin des-acylation, it can be hypothesized that the observed decrease of plasma 392 ghrelin level in the above referred conditions is due to a reduction of the secretion of ghrelin 393 from the stomach <sup>10</sup>, as it has been shown in obese mice <sup>52</sup>. Also, it is possible that des-394 acylation of circulating ghrelin is unaffected in plasma but regulated in some specific tissues, 395 such as liver. Future studies in humans are needed to test these possibilities. 396

397

### 398 Conclusions

We provide here evidence indicating that ghrelin proteolysis in plasma increases in 399 males with OW/OB and that it is acutely regulated by food intake in lean individuals. Also, 400 we report that ghrelin des-acylation in plasma is not affected by body weight or food intake. 401 Thus, current study helps to better understand the mechanisms controlling the changes of 402 plasma ghrelin under different metabolic conditions in humans. Ghrelin proteolysis may be 403 a putative pharmacological target to control circulating ghrelin level in humans. In particular, 404 enhancement of ghrelin proteolysis in males with OW/OB or after food intake may help to 405 further reduce plasma ghrelin level, with concomitant "beneficial" effects such as appetite 406 reduction and downregulation of mechanisms that increase glycemia. 407

408

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418

## 419 **Conflict of interest**

- 420 The authors declare no conflicts of interest. The sponsors had no role in the design of the
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582 Figure Legends.

583

584 <u>Figure 1</u>. Overview of methods to assess ghrelin des-acylation (A) and ghrelin proteolysis
585 (B). See text for details. Eppendorf tube illustration adapted from scidraw.io
586 (doi.org/10.5281/zenodo.6808872).

587

Figure 2. Preprandial plasma levels of ghrelin (A), DAG (B), ghrelin/DAG ratio (C), ghrelin
des-acylation activity (D) and ghrelin proteolysis (E) in NW and OW/OB male participants.
\*, \*\*, \*\*\* and \*\*\*\* indicate significant difference at p<0.05, p<0.01 p<0.001 p<0.0001 by</li>
Student's t test or Mann Whitney test.

592

593 <u>Figure 3</u>. Correlations of ghrelin proteolysis with ghrelin (A), DAG (B), BMI (C), and glycemia
594 (D) in male participants in preprandial conditions.

595

596 <u>Figure 4</u>. Preprandial and postprandial plasma levels of ghrelin (A), DAG (B), ghrelin des-597 acylation activity (C) and ghrelin proteolysis (D) in males with NW or OW/OB. Coloured 598 points and lines show the geometric mean and each grey line represents the change within 599 an individual participant. Exact P values are shown. Two-way repeated measures analysis 600 of variance (ANOVA) with Bonferroni multiple-comparisons test was performed. \*: significant 601 difference vs. same group but different feeding condition. #: significant difference vs. 602 different group but same feeding condition.

603

<u>Figure 5</u>. Preprandial and postprandial plasma levels of ghrelin (A), DAG (B) and ghrelin
 proteolysis (C) in females with NW or OW/OB. Coloured points and lines show the geometric
 mean and each grey line represents the change within an individual participant. Exact P
 values are shown. Two-way repeated measures analysis of variance (ANOVA) with
 Bonferroni multiple-comparisons test was performed. \*: significant difference vs. same group
 but different feeding condition. #: significant difference vs. different group but same feeding
 condition.

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3	Ghrelin Proteolysis Increases in Plasma of Men, but not Women, with Obesity
4	
5	Running title: ghrelin proteolysis in human obesity
6	
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#### 34 Abstract

35

Aims: Since plasma ghrelin can undergo des-acylation and proteolysis, the aim of this study was to investigate the extent to which an enhancement of these reactions is associated to the decrease of ghrelin in plasma after food intake or in individuals with obesity.

Main methods: we performed an intervention cross-sectional study, in which levels of
 ghrelin, desacyl-ghrelin (DAG), glucose, insulin, ghrelin des-acylation and ghrelin
 proteolysis were assessed in plasma before and after a test meal in 40 people (n=21 males)
 with normal weight (NW, n=20) or overweight/obesity (OW/OB, n=20).

- Key findings: Preprandial ghrelin and DAG levels were lower, whereas preprandial ghrelin 43 proteolysis was ~4.6-fold higher in plasma of males with OW/OB. In males, ghrelin 44 proteolysis positively correlated with glycemia. Ghrelin and DAG levels were also lower in 45 females with OW/OB, but preprandial ghrelin proteolysis was not different between females 46 with NW or OW/OB. Ghrelin and DAG levels decreased postprandially in males and females, 47 independently of BMI, and ghrelin proteolysis increased postprandially ~2 folds only in 48 individuals with NW. Ghrelin des-acylation remained unaffected by BMI or feeding status in 49 50 both sexes.
- 51 <u>Significance</u>: Current study shows that ghrelin proteolysis increases in males with obesity 52 as well as after meal in lean individuals. Therefore, ghrelin proteolysis may be an important 53 checkpoint and, consequently, a putative pharmacological target to control circulating 54 ghrelin levels in humans.
- 55
- 56 Key words: ghrelin proteolysis, ghrelin des-acylation, obesity.

#### 57 Introduction

58

59 Ghrelin is a 28-residue peptide hormone predominantly secreted by cells of the 60 stomach that displays several unique biochemical and biological features <sup>1</sup>. Chemically, ghrelin is the only peptide known to be acylated with an octanoic moiety, a posttranslational 61 modification that takes place at Ser3 of ghrelin and is catalysed by the enzyme ghrelin-O-62 63 acyltransferase (GOAT) early in the biosynthetic processing within the ghrelin-producing cells<sup>2</sup>. The biological actions of ghrelin, which are mediated by the growth hormone 64 65 secretagogue receptor (GHSR), entirely depend on the octanoylation of ghrelin and include the most potent orexigenic effect described to date <sup>3,4</sup>. Ghrelin also induces growth hormone 66 and glucocorticoid secretion, recruits several mechanisms that increase glycemia, increases 67 gastrointestinal motility and affects non-homeostatic aspects of food intake, among other 68 effects <sup>1</sup>. Thus, ghrelin is a pleiotropic hormone that contains an exceptional 69 posttranslational modification that is essential for its bioactivity. 70

71

72 Plasma ghrelin level is tightly regulated by the daily feeding status and by the long-73 term energy balance. In the short term, ghrelin level increases before meals and decreases 74 to baseline levels within the first hour after meals <sup>5</sup>. Ghrelin level increases in prolonged 75 energy deficit conditions, such as fasting, calorie restriction or anorexia nervosa <sup>6,7</sup>, when a 76 rise in plasma ghrelin contributes to seek for food and maintain glycemia. In contrast, ghrelin 77 level decreases under energy surplus conditions, such as obesity<sup>8,9</sup>, and such decrease presumably results as a compensatory adaptation aiming to reduce feeding and glycemia. 78 As other hormones, plasma ghrelin level depends on the equilibrium among its secretion, 79 degradation and clearance rates. A large body of studies characterized the mechanisms 80 controlling the biosynthesis and release of ghrelin from ghrelin cells mainly using animal 81 models or *in vitro* systems <sup>10</sup>. Conversely, the molecular mechanisms controlling the 82 83 disappearance of ghrelin from the circulation have been less investigated. Ghrelin rapidly disappears from plasma, having a half-life of 9-13 min after a single bolus injection of the 84 hormone <sup>11-13</sup>, and the key chemical reactions affecting plasma ghrelin involve not only des-85 acylation but also proteolysis. Ghrelin des-acylation in plasma results in the generation of 86 87 desacyl-ghrelin (DAG), which is also secreted from ghrelin cells. DAG does not bind to GHSR at physiological levels, but it has been suggested to display some GHSR-88 89 independent effects on insulin secretion, osteoblast growth and lipid metabolism <sup>1</sup>. Ghrelin des-acylation in plasma is presumably mediated by different esterases including platelet 90 91 activating factor acetylhydrolase, paraoxonase,  $\alpha^2$ -macroglobulin, butyrylcholinesterase

92 and acyl protein thioesterase 1; although it is likely that other esterases can also hydrolyze the lipid moiety from ghrelin <sup>14-19</sup>. Ghrelin also undergoes proteolysis in plasma giving rise to 93 shorter peptides <sup>15,16</sup>. Ghrelin proteolysis mainly occurs at the middle segment of the peptide 94 95 (i.e. between Arg15 and Lys16) and generates peptides of different length, including peptides containing the N-terminal end of ghrelin that may be able to evoke some activation 96 of GHSR although less potently than full-length ghrelin <sup>16,20,21</sup>. Ghrelin proteolysis was 97 98 suggested to involve endopeptidase activated protein C in bovine plasma<sup>22</sup>, and insulindegrading enzyme in an *in vitro* system <sup>23</sup>. Still, the molecular identity of the enzymatic 99 100 system mediating ghrelin inactivation in plasma remains to be fully clarified.

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Since its discovery, manipulating ghrelin system emerged as a putative strategy in 102 103 the treatment of obesity and related diseases. In order to further reduce ghrelin actions in 104 obesity and achieve additional beneficial effects, GHSR antagonists, ghrelin vaccines and GOAT inhibitors have been developed; however, none of these strategies reached the 105 desired effects in human trials <sup>24</sup>. Notably, the inhibition of the extracellular inactivation of 106 incretins has proved to be a critical strategy to improve glycaemic control in patients suffering 107 type 2 diabetes <sup>25</sup>. In this context, understanding the molecular mechanisms controlling the 108 109 level of ghrelin in plasma emerges as a fundamental step forward to not only better 110 understand the biology of ghrelin but also find new pharmacological strategies to manipulate such levels and achieve beneficial effects. To the best of our knowledge, the extent to which 111 des-acylation or proteolysis of ghrelin in plasma are affected by the daily feeding status or 112 obesity has not been previously investigated. Here, we hypothesized that decrease of 113 plasma ghrelin in individuals with obesity and/or in the postprandial state could be due to an 114 115 increase in its extracellular processing. In order to test our hypothesis, we performed a single 116 test meal study in individuals with either normal weight (NW) or with overweight or obesity 117 (OW/OB) and assessed not only levels of ghrelin and DAG in plasma but also the specific 118 levels of ghrelin des-acylation and ghrelin proteolysis in plasma.

- 119 Materials and Methods
- 120

2.1 Study design. The study was approved by the Institutional Ethical Committee of
 San Martin Hospital, La Plata (Protocol ID HSMLP2020/0044) and conducted according to
 the Declaration of Helsinki guidelines and Argentinian legal provisions governing clinical
 research in humans. Written informed consents were obtained from all participants.
 Participants were recruited via advertising in social media from August to October 2021.

126

2.2 Study participants. This intervention cross-sectional study included 20–45 years old adults, without chronic medical illness, personal history of diabetes, history of medical
 condition, or medication related to obesity or diabetes risk status. The study included 21
 males and 19 females. Pregnant women were excluded from the study. Sample size was
 calculated to unmask differences in plasma ghrelin levels, assuming a standard deviation of
 3.26 fmol/ml <sup>26</sup>, 80% statistical power and 5% significance level in preprandial ghrelin level,
 leading to a minimal sample size of 40 participants (20 with NW and 20 with OW/OB).

134

135 2.3 Study protocol. All participants arrived at the facilities of the Institute in the 136 morning after a 12-h overnight fast and underwent an initial evaluation, including a medical 137 and family history. Weight and body composition were measured with the participants 138 barefoot and wearing minimal clothing, on a bioimpedance scale (Omron HBF514, Argentina) with a resolution of 10 g. Height was measured with a Harpenden-type wall-139 mounted stadiometer (Holtain Ltd., United Kingdom) with a resolution of 1 mm. BMI was 140 calculated by dividing weight by height squared (kg/m<sup>2</sup>). As described by the World Health 141 142 Organization, participants were classified as NW when BMI was between 18.5 and 24.99 Kg/m<sup>2</sup> or as OW/OB when BMI≥25 Kg/m<sup>2</sup> <sup>27</sup>. Next, a preprandial blood sample was obtained. 143 144 Participants were given a test meal (breakfast) that provided 20% of the individual's daily 145 energy requirement as estimated using the "Body Weight Planner" digital application of the 146 National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK, USA)<sup>28</sup>. The 147 breakfast composition was 56 kcal% carbohydrates, 12 kcal% protein and 34 kcal% lipids 148 and comprised yogurt, cereals and almonds. Participants were asked to consume the entire 149 breakfast in less than 15 min. Blood sampling was repeated at 60 min after starting breakfast (postprandial), when plasma ghrelin level is significantly reduced, according to a meta-150 151 analysis study 5. Blood samples were collected in tubes containing either heparin, to assess glucose and insulin, ghrelin proteolysis and ghrelin des-acylation, or EDTA (1 mg/mL final) 152 153 and p-hydroxy-mercuribenzoic acid (0.4 mM final), to assess ghrelin and DAG. For ghrelin

and DAG assays, plasma was immediately acidified with HCl (0.1 N final) <sup>29</sup>. All samples
 were stored frozen at -80°C until analysis.

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157 2.4 Biochemical analyses. Glycemia was measured using an enzymatic procedure 158 in a CM 250 (Wiener Lab, Argentina). Insulin level was assessed using chemiluminescence (Advia Centaur XP, Siemens, Germany). Insulin resistance was estimated by the 159 160 homeostasis model assessment for insulin resistance (HOMA index) <sup>30</sup>. Ghrelin and DAG were assessed using specific enzyme immunoassays from Bertin Bioreagent (A05306 and 161 162 DAG, respectively), and following manufacturer's instructions. Ghrelin/DAG ratio was 163 computed on a molar basis as ghrelin concentration divided by DAG concentration. The food intake-induced change of plasma ghrelin was calculated as preprandial minus postprandial 164 165 ghrelin concentration in molar basis.

166

2.5 Assessment of ghrelin des-acylation in plasma. Here, we used a biotinylated 167 analogue of ghrelin consisting in the first 11 residues of ghrelin with the octanoyl moiety 168 linked to Ser3 and a biotin attached to the *ɛ*-amino group of Arg (biotin-ghrelin or 169 170 GSS(Oct)FLSPEHQR-biotin). To assess ghrelin des-acylation in plasma (Fig. 1A), biotin-171 ghrelin (0.01  $\mu$ g/ $\mu$ L) was incubated with each plasma sample (20 % v/v) in phosphate buffer 172 saline at 37 °C. After 60 min, trifluoroacetic acid (20 % final) was added to the tube to stop 173 the reaction. We next used a reverse-phase chromatographic procedure to separate biotin-174 ahrelin from biotin-DAG, based on a previously described protocol<sup>2</sup>. Briefly, reaction mixtures for each plasma sample were applied to a ZipTip with C18 resin, according to 175 manufacturer's instructions, and eluted with a 20% and 100%-CH<sub>3</sub>CN solutions in 0.1% TFA 176 177 in order to elute biotin-DAG and biotin-ghrelin, respectively. After CH<sub>3</sub>CN evaporation, eluted 178 samples were subjected to dot blot using PVDF membranes. Next, blot membranes were 179 incubated with Vectastain Elite ABC kit (Vector Laboratories, cat # PK6200) for 2 h, 180 according to manufacturer's protocols, and the presence of biotin-ghrelin and biotin-DAG was visualized using a 3-3'-diaminobenzidine (DAB)/Nickel solution, which gives a 181 182 black/purple precipitate. DAB signal per spot was registered using a Gel Doc XR (Bio-Rad) and quantified using the software Fiji. Ghrelin des-acylation was calculated as the ratio 183 184 between the intensity of biotin-DAG and biotin-ghrelin and expressed as relative units (r.u.). The accuracy of the separation and quantification system was checked using solutions with 185 186 different concentrations of either intact biotin-ghrelin and biotin-DAG.

188 2.6 Assessment of ghrelin proteolysis in plasma. Here, we used a fluorescent 189 analogue of human ghrelin (F-ghrelin) consisting in the first 19 residues of ghrelin with the 190 octanoyl moiety linked to a diaminopropanoic acid (Dpr) at position 3 and a fluorescein 191 attached to the ε-amino of Lvs19 isothiocyanate group 192 (GSDpr(Oct)FLSPEHQRVQQRKESK(FITC)-NH<sub>2</sub>), which was synthesized as previously described <sup>31</sup>. To assess ghrelin proteolysis in plasma (Fig. 1B), F-ghrelin (0.025  $\mu$ g/ $\mu$ L) was 193 194 incubated with each plasma sample (12.5 % v/v) in phosphate buffer saline at 37 °C. After 195 180 min, loading buffer was added to the tubes and reaction mixtures for each sample were 196 loaded onto 20% polyacrylamide gels prepared according to the Ornstein-Davis 197 discontinuous system. Fluorescence was visualized by trans-UV exposure, registered using a 520DF30 emission filter (Gel Doc XR, Bio-Rad) and guantified using the software Fiji. 198 199 Ghrelin proteolysis was calculated as the ratio between the fluorescence of the shorter 200 peptide products of cleavage and the fluorescence of intact F-ghrelin, and expressed as 201 relative units (r.u.).

202

2.7 Statistics and data analyses. Statistical analyses were performed using 203 204 GraphPad Prism 9.0.0 Normal distribution of data was tested using the Shapiro-Wilk test. 205 Preprandial characteristics in males were compared with Student's unpaired t test or Mann 206 Whitney test. Positively skewed variables (ghrelin, DAG, ghrelin/DAG, and ghrelin 207 proteolysis) were log-transformed for all subsequent analyses. Changes in time between groups were evaluated with two-way repeated measures analysis of variance (ANOVA), with 208 group (NW vs. OW/OB) as between factor and feeding condition (preprandial vs. 209 postprandial) as within factor. Linear correlations between log-transformed ghrelin 210 211 proteolysis and other variables were assessed using Pearson's (ghrelin, BMI and glycemia) or Spearman's (DAG) correlation coefficients. Tests used for each comparison were 212 213 indicated in each figure legends. Variables with normal distribution were expressed as the 214 mean ± standard deviation (SD), and variables with non-normal distribution were reported as median (interguartile range [IQR]). Differences were considered statistically significant 215 216 when p<0.05.

- 217 Results
- 218

219 3.1 Ghrelin level decreases whereas ghrelin proteolysis increases in preprandial 220 plasma of males with OW/OB. Initially, male participants were grouped as NW (n=9) or 221 OW/OB (n=12). As shown in Table 1, age did not differ between groups nor correlate with any assessed parameter, whereas BMI, preprandial glycemia, preprandial insulin levels and 222 223 HOMA index were significantly higher in OW/OB group, as compared to NW group. 224 Conversely, preprandial ghrelin (Fig. 2A) and DAG (Fig. 2B) levels were ~2.2- and ~6.0-fold 225 lower, respectively, in OW/OB group, as compared to NW group. Ghrelin/DAG ratio (Fig. 2C) was ~8.5-fold higher in OW/OB group, as compared to NW group, suggesting that the 226 mechanisms controlling plasma ghrelin level are differentially affected in males with OW/OB. 227 228 Thus, we assessed the capacity of plasma samples to des-acylate a biotinylated ghrelin 229 analogue. We found that ghrelin des-acylation was not different between plasma samples of NW and OW/OB groups (Fig. 2D). Next, we assessed the capacity of plasma samples to 230 proteolyze a fluorescent ghrelin analogue. Here, ghrelin proteolysis was ~4.6-fold higher in 231 232 OW/OB group, as compared to NW group (Fig. 2E). Notably, ghrelin proteolysis negatively 233 correlated with plasma levels of ghrelin and DAG (Fig. 3A-B), it did not correlate with age (r=0.223, p=0.3317, not shown), plasma insulin levels (r=0.269, p=0.239, not shown), HOMA 234 235 index (r=0.303, p=0.182, not shown) nor BMI (Fig. 3C) and it positively correlated with 236 glycemia (Fig. 3D).

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3.2 Postprandial ghrelin proteolysis in plasma increases in males with NW, but not with OW/OB, whereas postprandial ghrelin des-acylation remains unaffected. After the 239 240 single test meal intervention, glycemia was not affected by feeding condition (p condition= 241 0.820), and remained higher in OW/OB group, as compared to NW group (p group=0.012). Plasma levels of ghrelin and DAG changed in a group- and feeding condition-dependent 242 243 manner, being decreased in OW/OB group and postprandially (Fig. 4A-B). Here, the food 244 intake-induced change of plasma ghrelin level tended to be lower in OW/OB group, as 245 compared to NW group (6.5±5.7 vs. 10.9±7.8 fmol/mL, respectively, P=0.0765). Notably, ghrelin des-acylation remained unaffected by group or feeding condition (Fig. 4C). Ghrelin 246 proteolysis also showed a significant interaction between the effects of group and feeding 247 condition, being ~1.6-fold increased after food intake only in NW group and increased in 248 249 OW/OB group, independently of the feeding condition (Fig. 4D).

251 3.3 Postprandial ghrelin proteolysis increases in NW females but remains unaffected 252 in plasma of females with OW/OB, independently of the feeding condition. Since some evidence indicate that ghrelin level fluctuates in a sex-dependent manner in humans <sup>32,33</sup>, 253 254 we also studied females under the same study protocol and grouped them as NW (N=11) 255 or OW/OB (n=8). As shown in Table 2, age and preprandial glycemia did not differ between 256 groups whereas BMI, preprandial insulin levels and HOMA index were significantly higher 257 in OW/OB group, as compared to NW group. In preprandial condition, ghrelin and DAG levels were ~2.0- and ~2.6-fold lower, respectively, in OW/OB group, but ghrelin/DAG ratio 258 259 was not different between both groups, suggesting that the mechanisms controlling plasma 260 ghrelin level in females with OW/OB were less affected than in males with OW/OB. Here, 261 ghrelin des-acylation and ghrelin proteolysis in plasma were not different between NW and 262 OW/OB groups (Table 2). In contrast to males, ghrelin proteolytic activity in plasma did not 263 correlate with glycemia in females (r=0.154, p=0.530).

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After the test meal intervention, plasma levels of ghrelin and DAG changed in a group- and feeding condition-dependent manner, being decreased in OW/OB group and postprandially (Fig. 5A-B). Here, the food intake-induced change of plasma ghrelin level was lower in the OW/OB group, as compared to NW group (10.2±8.3 vs. 26.4±16.75 fmol/mL, respectively, P=0.0230). As found in males, ghrelin des-acylation was unaffected by the feeding condition in NW and OW/OB groups (not shown) whereas ghrelin proteolysis was postprandially increased ~2.2 folds only in NW group (Fig. 5C). 272 Discussion

273

274 Here, we performed a single test meal study in individuals with NW or OW/OB and 275 investigated levels of ghrelin, ghrelin des-acylation and ghrelin proteolysis in plasma. We 276 confirmed that preprandial ghrelin level is lower in individuals with OW/OB, independently of sex, as previously shown <sup>5,8,9,32,34-38</sup>. After food intake, ghrelin level decreased in males 277 278 and females of NW and OW/OB groups. Interestingly, ghrelin proteolysis was higher in 279 plasma of males, but not females, with OW/OB and correlated with glycemia. Ghrelin 280 proteolysis increased after food intake in individuals with NW only. In contrast, ghrelin des-281 acylation was similar in plasma of individuals with NW or OW/OB and remained unchanged 282 after food intake.

283

284 The current study was carefully designed to assess the chemical reactions affecting 285 plasma ghrelin at two key time points: in the morning after overnight fasting, when plasma ghrelin is around the highest diurnal level<sup>8</sup>, and 60 min after meal, when plasma ghrelin 286 decreases <sup>5</sup>. Indeed, we detected the expected postprandial decline in ghrelin level in most 287 lean individuals (19 out of 20), as shown in several previous reports <sup>5,32,35-38</sup>, confirming that 288 289 plasma ghrelin level was studied when it undergoes a significant change. It is important to 290 highlight that it is now well established that the inaccurate manipulation of plasma samples 291 results in the conversion of most ghrelin into DAG and a consequent low ghrelin/DAG ratio 292 <sup>39</sup>. Here, plasma ghrelin/DAG ratio in NW individuals was ~0.6, which is similar to the highest reported ghrelin/DAG ratios in similar studies (see for instance <sup>40</sup>), indicating that the 293 294 conditions of sampling and storage were accurate to preserve ghrelin in plasma despite the 295 complexity involved in clinical studies. Finally, current study was performed in relatively 296 young individuals in order to minimize the putative impact of the age-dependent decline in plasma ghrelin that has been observed older individuals <sup>41</sup>. 297

298

In order to specifically assess ghrelin des-acylation or proteolysis in plasma, we set 299 300 up two reproducible assays whose characteristics should be taken in consideration when interpreting current findings. Importantly, we used two different labelled variants of ghrelin 301 that rely on modifications at its C-terminal end, which is not essential for bioactivity <sup>20,42</sup>. To 302 303 assess ghrelin des-acylation, we used biotin-ghrelin, a 11-residue analogue of ghrelin 304 biotinylated at the Arg11 C-terminus. Biotin-ghrelin is susceptible to hydrolysis by esterases, 305 since the octanoyl moiety is coupled at Ser3 through an ester bond, but presumably resistant 306 to plasma proteases, since ghrelin1-11 is the shorter derivative of ghrelin found after

proteolysis of full-length ghrelin<sup>22</sup>. To assess ghrelin proteolysis, we used F-ghrelin, an 18-307 308 residue analog of ghrelin conjugated to fluorescein isothiocyanate through a Lys added at 309 its C terminus. F-ghrelin contains the entire peptide segment susceptible to proteolysis, 310 according to previous descriptions <sup>22</sup>, but is resistant to esterases because Ser3 was 311 replaced with Dpr, which is coupled with the octanoyl moiety through an amide bond <sup>31</sup>. Of 312 note, F-ghrelin and the fluorescent products of proteolysis were directly visualized and 313 quantified after separation by native gel electrophoresis. In contrast, visualization and 314 quantification of biotin-ghrelin and biotin-DAG required several steps including reverse-315 phase chromatographic procedure for separation, dot-blot and chromogenic staining. Thus, 316 it is possible that the method used to assess ghrelin des-acylation lacked enough sensitivity 317 to unmask small differences in enzymatic activity. Also, reaction times to estimate each 318 enzymatic activity were set up in order to have detectable amounts of substrate and products 319 of each assay. Thus, ghrelin des-acylation and ghrelin proteolysis assays required 60 and 320 180 min of reaction time, respectively. The observation that the hydrolysis of peptide bonds 321 of ghrelin in plasma is a relatively slow process would suggest that changes in ghrelin proteolysis have stronger implications in long-term conditions, as we observed here in males 322 323 with OW/OB. In contrast, the physiological implications of the detected postprandial increase 324 of ghrelin proteolysis in plasma of lean individuals is uncertain given the difference between 325 the timing for the rapid postprandial decrease of plasma ghrelin and the reaction time 326 required to assess ghrelin proteolysis in our experimental conditions.

327

328 To the best of our knowledge, the current study provides the first indication that proteolysis of ghrelin in plasma increases in males with OW/OB. The observation that ghrelin 329 330 proteolysis negatively correlates with plasma levels of ghrelin and DAG in the preprandial 331 state suggests that increased ghrelin proteolysis in males with OW/OB may contribute to 332 reduce plasma levels of these peptides. Also, increased ghrelin proteolysis in obesity may 333 contribute to impair the effects of ghrelin treatment (the so-called ghrelin resistance), reported in obese mice <sup>43</sup>. Strikingly, we found here that ghrelin proteolysis positively 334 335 correlated with glycemia, but did not correlate with insulinaemia or HOMA index. Conversely, preprandial ghrelin proteolysis and glycemia remained unaffected in females with OW/OB. 336 337 despite they showed reduced preprandial plasma ghrelin level and insulin resistance, as compared to females with NW. Thus, preprandial ghrelin proteolysis is somehow associated 338 339 to glucose level itself, via mechanisms that do not seem to involve insulin. The reasons why ghrelin proteolytic activity increases only in males with OW/OB are uncertain, but it is in line 340 341 with the observation that ghrelin level in obesity tends to be lower in males than females, as

found here and reported in previous studies <sup>32,33</sup>. Interestingly, insulin-degrading enzyme level in plasma was found higher in individuals (of undisclosed sex) with metabolic syndrome and strongly correlated with the severity of the symptoms <sup>44</sup>. Also, plasma activated protein C level was higher in individuals (male and female combined) with obesity and reduced after bariatric surgery <sup>45,46</sup>. Thus, some proteases able to cleave ghrelin are strong candidates to mediate increased ghrelin proteolysis in plasma of male individuals with OW/OB.

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Current study also provides the first indication that ghrelin proteolysis increases in 349 plasma after food intake in lean individuals, in a sex-independent manner, allowing to 350 351 hypothesize that increased postprandial ghrelin proteolysis in lean individuals contributes to 352 decrease plasma ghrelin levels after food intake. Since glycemia is well-known to transiently 353 increase after food intake, it is possible that the postprandial increment of ghrelin proteolysis 354 is also linked to glucose levels. In line with this possibility, a previous study found that a glucose load potently decreases ghrelin levels in lean individuals <sup>31</sup>. Further studies 355 assessing blood samples at earlier time points will be required to test the extent to which 356 postprandial increments of glycemia and ghrelin proteolysis correlate. Interestingly, ghrelin 357 358 proteolysis did not increase after food intake in plasma of individuals with OW/OB, in which 359 plasma ghrelin level shows a smaller decrease after meal intake, as previously shown <sup>5</sup> and 360 confirmed here. Thus, it is likely that the lack of postprandial increase in ghrelin proteolysis in obesity contributes to impair the postprandial reduction of ghrelin levels, an alteration that 361 362 has been proposed to reduce satiety feelings in individuals with obesity <sup>47-49</sup>.

363

Although informative, this study is not without limitations. For instance, some aspects 364 365 of our experimental design, such as the postprandial blood sample collection time or the 366 sample size, were based on previous studies investigating the postprandial decline in ghrelin level <sup>5,32,35-38</sup>. However, such experimental design may have prevented us from detecting the 367 368 postprandial change of ghrelin proteolytic activity in individuals with OW/OB, if an increase 369 in such enzymatic activity takes place with a different kinetics or if it requires a higher sample 370 size to be unmasked. Also, the time chosen to collect the postprandial blood sample did not 371 allow us to detect the transient meal-induced increase of glycemia and, consequently, we 372 could not evaluate if postprandial hyperglycaemia either was differentially affected in individuals with OW/OB or correlated the postprandial increase of ghrelin proteolytic activity 373 374 in lean individuals. The study did not include "sham-feeding" control groups and, 375 consequently, a postprandial increase of ghrelin proteolytic activity may be related to its

physiological diurnal rhythm. Despite these limitations, the current study was very valuableto gain insights about a novel aspect of the biology of ghrelin in human plasma.

378

379 Ghrelin des-acylation in plasma is fast and affects the circulating ghrelin level <sup>50</sup>. 380 Indeed, intravenous administration of ghrelin in humans results in a peak of ghrelin levels, at 15 min after injection, and is rapidly eliminated <sup>11,40</sup>. Ghrelin des-acylation seems to be 381 382 the main mechanism mediating the reduction of plasma ghrelin elevation since the referred 383 increase of ghrelin level is followed by a peak of DAG level, which then disappears from plasma more slowly than ghrelin <sup>11,40</sup>. Here, we found that ghrelin des-acylation was 384 unaffected in individuals with OW/OB or postprandially suggesting that des-acylation is not 385 386 a key mechanism mediating the decrease of plasma ghrelin level in those metabolic 387 conditions. In line with our observations, it was reported that the level of serum butyrylcholinesterase activity, a ghrelin esterase in humans, was not altered during fasting 388 <sup>51</sup>. Since our results indicate that ghrelin proteolysis is an important mechanism affecting the 389 390 decrease of plasma ghrelin level in males with OW/OB, the mechanisms mediating the decrease of plasma ghrelin level in females with OW/OB remain uncertain. Given the fast 391 392 rate of ghrelin des-acylation, it can be hypothesized that the observed decrease of plasma 393 ghrelin level in the above referred conditions is due to a reduction of the secretion of ghrelin from the stomach <sup>10</sup>, as it has been shown in obese mice <sup>52</sup>. Also, it is possible that des-394 acylation of circulating ghrelin is unaffected in plasma but regulated in some specific tissues, 395 396 such as liver. Future studies in humans are needed to test these possibilities.

397

#### 398 **Conclusions**

399 We provide here evidence indicating that ghrelin proteolysis in plasma increases in 400 males with OW/OB and that it is acutely regulated by food intake in lean individuals. Also, 401 we report that ghrelin des-acylation in plasma is not affected by body weight or food intake. 402 Thus, current study helps to better understand the mechanisms controlling the changes of plasma ghrelin under different metabolic conditions in humans. Ghrelin proteolysis may be 403 404 a putative pharmacological target to control circulating ghrelin level in humans. In particular, enhancement of ghrelin proteolysis in males with OW/OB or after food intake may help to 405 406 further reduce plasma ghrelin level, with concomitant "beneficial" effects such as appetite 407 reduction and downregulation of mechanisms that increase glycemia.

408

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## 419 **Conflict of interest**

- 420 The authors declare no conflicts of interest. The sponsors had no role in the design of the
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582 Figure Legends.

583

584 <u>Figure 1</u>. Overview of methods to assess ghrelin des-acylation (A) and ghrelin proteolysis
585 (B). See text for details. Eppendorf tube illustration adapted from scidraw.io
586 (doi.org/10.5281/zenodo.6808872).

587

Figure 2. Preprandial plasma levels of ghrelin (A), DAG (B), ghrelin/DAG ratio (C), ghrelin
des-acylation activity (D) and ghrelin proteolysis (E) in NW and OW/OB male participants.
\*, \*\*, \*\*\* and \*\*\*\* indicate significant difference at p<0.05, p<0.01 p<0.001 p<0.0001 by</li>
Student's t test or Mann Whitney test.

592

593 <u>Figure 3</u>. Correlations of ghrelin proteolysis with ghrelin (A), DAG (B), BMI (C), and glycemia
594 (D) in male participants in preprandial conditions.

595

596 <u>Figure 4</u>. Preprandial and postprandial plasma levels of ghrelin (A), DAG (B), ghrelin des-597 acylation activity (C) and ghrelin proteolysis (D) in males with NW or OW/OB. Coloured 598 points and lines show the geometric mean and each grey line represents the change within 599 an individual participant. Exact P values are shown. Two-way repeated measures analysis 600 of variance (ANOVA) with Bonferroni multiple-comparisons test was performed. \*: significant 601 difference vs. same group but different feeding condition. #: significant difference vs. 602 different group but same feeding condition.

603

<u>Figure 5</u>. Preprandial and postprandial plasma levels of ghrelin (A), DAG (B) and ghrelin
 proteolysis (C) in females with NW or OW/OB. Coloured points and lines show the geometric
 mean and each grey line represents the change within an individual participant. Exact P
 values are shown. Two-way repeated measures analysis of variance (ANOVA) with
 Bonferroni multiple-comparisons test was performed. \*: significant difference vs. same group
 but different feeding condition. #: significant difference vs. different group but same feeding
 condition.

	Participants with NW	Participants with OW/OB	p-value
Age	34.7 ± 9.1	37.1 ± 5.0	N.S.
BMI	23.6 ± 0.9	$30.9 \pm 4.6$	0.0001
Glycemia (g L <sup>-1</sup> )	$0.99 \pm 0.04$	1.08 ± 0.07	0.0042
Insulin (µIU mL <sup>-1</sup> )	6.29 ± 3.21	14.28 ± 8.35	0.0139
HOMA index	1.55 ± 0.81	3.83 ± 2.31	0.0114

## Table 1: Baseline features of male participants

Abbreviations: BMI: body mass index; HOMA: homeostatic model assessment. Data are presented as mean  $\pm$  SD. Data analysis performed by t-test.

	Participants with NW	Participants with OW/OB	p-value
Age	34.5 ± 8.1	30.3 ± 7.9	N.S.
BMI	22.3 ± 2.1	29.1 ± 2.4	<0.0001
Glycemia (g L <sup>-1</sup> )	$0.96 \pm 0.05$	$0.98 \pm 0.04$	N.S.
Insulin (µIU mL⁻¹)	5.15 ± 1.41	12.87 ± 4.42	<0.0001
HOMA index	1.22 ± 0.35	3.13 ± 1.11	<0.0001
Ghrelin (fmol mL <sup>-1</sup> )	40.26 ± 20.56	$19.47 \pm 9.60$	0.0180
DAG (fmol mL <sup>-1</sup> )	63.77 ± 27.16	24.45 ± 9.27	0.0011
Ghrelin/DAG ratio	0.61 ± 0.21	$0.77 \pm 0.22$	N.S.
Proteolytic activity (r.u.)	0.04 (0.02; 0.10)	0.06 (0.03; 0.21)	N.S.
Desacylase activity (r.u.)	1.31 ± 0.77	$0.94 \pm 0.12$	N.S.

## Table 2: Baseline features of female participants

Abbreviations: BMI: body mass index; HOMA: homeostatic model assessment; DAG: desacyl-ghrelin.

Data are presented as mean  $\pm$  SD and analysed by t-test for all parameters except for proteolytic activity, which is presented as median (IQR) and analysed by Mann Whitney test.











## Credit author statement

Conceptualization, formal analysis, original draft preparation, supervision, overall responsibility for the project: MFA, MP.

Methodology, validation, formal analysis, investigation, resources and data curation; ASF, DC, DL, CS, PNDF, CRT, TL, LGL, SC, JAF, MFA, MP.

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