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The BCL2 inhibitor venetoclax mediates anticancer effects

through dendritic cell activation

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Abstract

BCL2 is an apoptosis-inhibitory oncoprotein that also possesses apoptosis-unrelated activities. Pharmacological BCL2 inhibitors have been developed with the scope of driving BCL2-dependent cancer cells into apoptosis, and one BCL2 antagonist, venetoclax, has been clinically approved for the treatment of specific leukemias and lymphomas. Nonetheless, it appears that venetoclax, as well as genetic BCL2 inhibition, can mediate anticancer effects through an indirect action. Such an indirect effect relies on the enhancement of the immunostimulatory function of dendritic cells, hence increasing tumor immunosurveillance. Mechanistically, BCL2 inhibition involves improved antigen presentation by conventional type-1 dendritic cells (cDC1s) due to the activation of an interferon response, leading to a T cell-mediated anticancer immune response that can be further enhanced by PD-1 blockade. These findings support the emerging hypothesis that successful antineoplastic drugs generally mediate their effects indirectly, through the immune system, rather via merely cell-autonomous effects on malignant cells.

Facts

- BCL2 inhibitors suppress cancer growth in an immunocompetent but not in an immunodeficient context.
- BCL2 inhibition improves the capacity of cDC1 to present tumor antigen and to stimulate anticancer immunity.
- BCL2 blockade sensitizes tumors to PD-1/PD-L1-immune checkpoint blockade.

Open Questions

- How does BCL2 inhibition exert cDC1 activation?
- In what way can BCL2 inhibitors be used for improving standard immunotherapy?

Main text

B-cell lymphoma-2 (BCL2) has been initially discovered as an oncogene/oncoprotein that is overexpressed in follicular lymphoma ^{1,2,3}. BCL2 was later found to inhibit apoptosis through its capacity to inhibit mitochondrial permeabilization leading to the release of caspase activators and caspase-independent death effectors ^{4,5,6}. This mitochondrion-sealing effect of BCL2 has been explained by its action on other proteins of an extended family of proteins possessing the so called BCL2 homology (BH1, BH2, BH3, BH4) domains, which together form the BCL2 family of proteins ^{7,8}. In particular, BCL2 was shown to directly interact with other proteins from the BCL2 family, to inhibit pore formation by pro-apoptotic proteins such as BAX and BAK (which both possess BH1, BH2 and BH3 but not BH4 domains) and to intercept (some of) the pro-apoptotic BH3-only proteins as well ^{9,10}. For long, it was believed that the oncogenic function of BCL2 could be completely ascribed to its apoptosis-inhibitory action. Nonetheless, BCL2 has been found to mediate additional, apoptosis-independent cellular effects, including the inhibition of autophagy, inflammation and senescence, as well as broad effects on cellular Ca²⁺ fluxes, redox homeostasis and bioenergetic metabolism ^{11,12,13}.

Small molecule inhibitors of BCL2 have been generated by the pharmacological industry ^{14, 15} and one of them, venetoclax, has been approved for clinical use against BCL2 dependent cancers, in particular chronic lymphocytic leukemia (CLL) and small lymphocytic lymphoma (SLL) ^{16, 17, 18, 19, 20}. Moreover, venetoclax can be combined with other drugs (such as azacitidine, decitabine or cytarabine) for the treatment of acute myeloid leukemia (AML) ²¹. Since these effects reportedly are mediated by direct killing of cancer cells, it has been assumed that the apoptosis-unrelated effects of BCL2 are not relevant for the anticancer effects of BCL2 inhibitors.

Recently, we performed a genome-wide screen to identify genes the knockout of which can enhance antigen presentation by dendritic cells (DCs) ²². DCs are terminally differentiated cells, meaning that genetic screens on such cells are impractical ²³. To circumvent this difficulty, we resorted to an immortalized DC precursor cell lines that expresses the SV40 large-T-cell antigen to inactivate the tumor suppressors TP53 and RB under the control of a doxycyclineinducible promoter ^{24, 25}. If cultured in the presence of doxycycline and the synthetic glucocorticoid dexamethasone, these immortalized DC precursor cell lines can be cultured indefinitively ^{24, 25, 26, 27}, allowing us to stably introduce the CRISP/Cas9 system into the cells and to transfect the cells with a genome-wide collection of barcoded guidance RNAs ²². Upon removal of doxycycline and dexamethasone, the DC precursors lose SV40 expression and differentiate into mature DCs that resemble one particular DC subset, namely conventional type 1 DCs (cDC1s) at the level of the transcriptome. Moreover, the de-immortalized mature DCs generated in vitro ressemble cDC1s with respect to the capacity to sample proteins present in the extracellular fluid, giving them access to the cytosol and to the antigen-presenting machinery. Indeed, such cDC1s are the only cells in the body that, in the presence of extracellular cytochrome c, undergo apoptosis because cytochrome c freely enters the cytosol where it activates apoptosomes containing APAF1 and caspase-9 ²⁸. We thererfore refer to our artificial DCs as cDC1s.

Based on our genome-wide screen performed on cDC1s, we identified BCL2 as a gene the knockout of which confers a gain-of-function phenotype ²². This gain-of-function phenotype could be mimicked by using pharmacological BCL2 inhibitors including the clinically approved drug venetoclax and the experimental drug navitoclax. Importantly, genetic or pharmacological inhibition of BCL2 did not compromise the survival of cDC1s in vitro or in

vivo, suggesting that enhanced antigen presentation occurs independently from the apoptotic pathway ²².

DCs, in particular of the cDC1 type, play a decisive role in the initiation and stimulation of cellular immune responses mediated by T lymphocytes ^{29, 30}. Based on this knowledge, we wondered whether BCL2 inhibition might have a general immunostimulatory effect that might improve cancer immunosurveillance. In the course of this experimental exploration, we obtained multiple results pleading in favor of the idea that BCL2 inhibition can exert antineoplastic effects through an action on cDC1s (**Fig. 1**).

In vitro experiments on murine cancer cell lines. When added to cutaneous fibrosarcoma MCA205 cells or non-small cell lung cancer (NSCLC) TC1 cells, the BCL2 inhibitors navitoclax and venetoclax largely fail to induce apoptosis. This contrasts with the fact that such BCL2 inhibitors can be administered to immunocompetent mice bearing orthotopic MCA205 or TC1 tumors under the skin or in the lung, respectively, and then cause a decrease in tumor growth. This anticancer effect is completely lost when T lymphocytes are depleted by antibodies recognizing CD4 and CD8, demonstrating that they require a cellular immune response ²². A similar result has been previously described for MC38 colorectal cancer cells that can form subcutaneous tumors if inoculated into mice. Such ectopic cancers also respond to venetoclax only in immunocompetent, not in T cell-deficient, mice ³¹. Moreover, MC38 cells do not undergo apoptosis when cultured in the presence of venetoclax ³¹. Hence, cancers that do not respond to BCL2 inhibitors in vitro can respond to such inhibitors in vivo, but only in the context of an intact immune system.

In vitro experiments on cDC1s. Knockout of BCL2 in cDC1s improves their antigenpresenting function in vitro coupled to the upregulation of multiple co-stimulatory signals (such
as CD80 and CD86), cytokines (such as IL-12), as well as a massive Type-1 interferon response

22. Pharmacological inhibition of BCL2 with navitoclax or venetoclax in cDC1s phenocopies
the in vitro effects of the BCL2 knockout, but does not further enhance the antigen-presenting
function of BCL2-deficient cDC1s, demonstrating that this is an on-target effect. Improved
antigen presentation by BCL2-inhibited cDC1 is lost upon neutralization of CD80, CD86 and
the common Type-1 interferon receptor (IFNAR), unraveling the mechanisms of
immunostimulation ²².

In vivo effects of BCL2 inhibitors on cDC1s. The intravenous administration of navitoclax or venetoclax to mice results in the activation of cDC1s (but less so cDC2s) with an increased expression of the costimulatory molecules CD80 and CD86, the chemokine receptors CCR2, CCR7 and XCR1, as well as the activation marker MHC class-II. High-dimensional immunofluorescence cytometry confirmed that, in patients with AML treated with venetoclax, CCR2, CCR7, XCR1, CD5, CD86 and HLA-DQ were upregulated in circulating cDC1s (not cDC2s). Similar effects were obtained ex vivo when peripheral blood mononuclear cells from health donors were treated with venetoclax in vitro and the phenotypic properties of cDC1s were then analyzed by immunofluorescence cytometry ²². Hence, pharmacological inhibition of BCL2 results in the activation of cDC1s (not cDC2s) in both human and mouse.

In vivo effects of BCL2-inhibited cDC1s. Intravenous injection of cDC1s that have been primed with tumor antigens in vitro confers some antitumor activity in vivo, in tumor-bearing immunocompetent mice, but this activity is enhanced when BCL2 knockout cDC1s are used or when WT cDC1s are pretreated with navitoclax or venetoclax prior to their injection ²².

These effects of intravenously injected BCL2-inhibited cDC1s are particularly striking in BATF3 knockout cells (which lack the cDC1 subset and hence exhibit a major defect in immunosurveillance) ³² but are lost in mice lacking mature T cells ²². Of note, BCL2 knockout cDC1 acquire the transcriptional phenotype of migratory cDC1s (a phenotype that is not seen in WT cDC1s) and infiltrate lung cancer lesions more efficiently than WT cDC1s ²². Hence, BCL2-inhibited cDC1s acquire a gain-of-function phenotype that improves their capacity to ignite cellular immune responses against tumor-associated antigens.

In vivo effects of BCL2 inhibitors on cancer immunosurveillance. Injection of navitoclax or venetoclax into immunocompetent mice slows down the progression, or causes the regression, of tumors that do not respond to navitoclax or venetoclax in vitro, especially if combined with PD-1 blockade ²². This in vivo anticancer activity of navitoclax or venetoclax is lost when the cellular immune system can be compromised due to (i) knockout of the essential cDC1 lineage-specific transcription factor BATF3 causing the constitutive absence of cDC1s ³²; (ii) repeated intravenous injection of cytochrome *c* for the acquired depletion of cDC1s ²⁸; (iii) antibody-mediated blockade of CD11b, which prevents cDC1s to enter tumor beds or tumor-draining lymph nodes ³³; (iv) antibody-mediated neutralization of IFNAR, which is the common Type-1 interferon receptor; and (v) depletion of T cells by injection of suitable anti-CD4 and anti-CD8 antibodies ²². Of note, it appears that the effects of venetoclax on T cells occur downstream of the effects on cDC1s. Thus, venetoclax upregulates the expression of PD-1 and CTLA-4 on CD4⁺ and CD8⁺ T cells in vivo, in tumor-bearing mice, but this effect is lost upon depletion of cDC1s by cytochrome c injections ²².

Altogether, the aforementioned results (Fig. 1) suggest that BCL2 inhibition can be active against cancers that are not BCL2-dependent (and hence do not undergo apoptosis upon BCL2

knockout or neutralization) through an indirect effect via the immune system that is primarily involving the activation of cDC1s ²². Mechanistically, it appears that genetic or pharmacological inhibition of BCL2 causes the release of DNA from mitochondria in cDC1s, thereby activating the cGAS/STING pathway, leading to the initially cell-autonomous activation of a classical Type-1 interferon pathway. Through autocrine or paracrine effects of Type-1 interferons on IFNAR, cDC1s then undergo transcriptional and functional reprogramming and improve their capacity to migrate into tumors, to present tumor antigens to T cells and to activate them. T cell activation is coupled to the upregulation of PD-1 (and CTLA-4) and specific T cell subsets. As an end result of this cascade of events instigated by BCL2 inhibition, tumor antigen-specific T cells infiltrate the cancer. This improves immunosurveillance and sensitizes the host to immune checkpoint inhibition (ICI) targeting the interaction between PD-1 and PD-L1 (Fig. 2).

Altogether, these findings add to the general concept that anticancer drugs generally do not (only) act on malignant cells, in spite of the fact that they have been conceived to kill them as cytotoxic chemotherapeutics or 'targeted' agents interfering with specific survival pathways ³⁴, ³⁵. Rather, successful anticancer drugs have empirically been selected by clinicians because of their capacity of inducing antitumor immune responses that account for treatment responses ³⁶, ³⁷. The major question that emerges from these considerations concerns the mechanisms of venetoclax against AML, CML and SLL. Does venetoclax favor anticancer immune responses against leukemia and lymphoma cells that might explain its true mode action, well beyond its cell-autonomous pro-apoptotic effects? Although there is no direct evidence in favor of this hypothesis, it appears intriguing that patients with AML can respond to immunotherapy with interleukin-2 ³⁸, vaccination with AML cell fused with autologous dendritic cells ³⁹, the PD-1 blocking antibody nivolumab ⁴⁰ or bispecific T cell engager antibodies ⁴¹. Hence, AML cells

are potentially immunogenic. Moreover, the prognosis of AML patients treated with chemotherapy is particularly positive when AML blasts exhibit signs of immunogenicity (such as calreticulin exposure) ^{42, 43} or are recognized by AML-specific antibodies detectable in the plasma ⁴⁴. Thus, it is possible, yet remains to be formally demonstrated, that blood cancers including AML are under therapy-relevant immunosurveillance. If this conjecture is correct, it would be highly plausible that venetoclax-triggered cDC1 activation contributes to the therapeutic success of this BCL2 inhibitor.

Historically, cell death research has been focusing on cell-intrinsic pathway, hoping to identify cell-autonomous pathways that can be targeted for selectively killing cancer cells or preserving normal cells. The senior author of this paper has participated to these bygone - and largely vain - efforts that were based on an over-simplistic view of cell (death) biology. Indeed, in vivo, cell death never occurs in isolation, as this might happen in the artificial setting of cell cultures, but rather within the context of a complex multicellular ecosystem. Such an ecosystem shapes the organismal response to cellular demise consisting in, for example, silent efferocytosis, inflammation and subsequent repair, as well as in innate or acquired immune responses against dead-cell antigens. Hence, the question whether and how cells die in isolation is superseded in importance by the question how cell stress and death are perceived by the organism ^{45, 46, 47, 48, 49, 50}

In this general context, it is not surprising that the clinical success of BCL2 inhibition finally is not (just) attributable to direct cancer cell killing but rather involves favorable effects on the cancer-immunity dialogue. In this vein, it will be interesting to determine whether venetoclax or other BCL2 inhibitors may be repurposed to stimulate anticancer immune responses against a vast spectrum of oncological diseases, likely in the context of immune checkpoint inhibitors

or other modalities of immunotherapy. Hence, the finding that BCL2 inhibitors likely mediate their therapeutic effect not (only) by killing BCL2-dependent malignant cells ultimately might yield a luminous perspective. Thanks to their immunostimulatory action on cDC1s, BCL2 inhibitors might mediate rather broad, target-agnostic anticancer effects. Beyond this consideration, it will be interesting to identify other pharmacological agents that, like BCL2 inhibitors, enhance the functional capabilities of cDC1s. Indeed, such drugs that enhance the function of cDC1s (to which we refer as "ICD enhancers") ⁵¹ might be used as a new class of immune checkpoint inhibitors to treat neoplastic disease.

Conflict of interest statement

OK and GK have been holding research contracts with Daiichi Sankyo, Eleor, Kaleido, Lytix Pharma, PharmaMar, Osasuna Therapeutics, Samsara Therapeutics, Sanofi, Tollys, and Vascage. LZ has held research contracts with Glaxo Smyth Kline, Incyte, Lytix, Kaleido, Innovate Pharma, Daiichi Sankyo, Pilege, Merus, Transgene, 9 m, Tusk and Roche, was on the on the Board of Directors of Transgene, is a cofounder of everImmune, and holds patents covering the treatment of cancer and the therapeutic manipulation of the microbiota. GK is on the Board of Directors of the Bristol Myers Squibb Foundation France. GK is a scientific co-founder of everImmune, Osasuna Therapeutics, Samsara Therapeutics and Therafast Bio. OK is a scientific co-founder of Samsara Therapeutics. GK is in the scientific advisory boards of Hevolution, Institut Servier and Longevity Vision Funds. GK is the inventor of patents covering therapeutic targeting of aging, cancer, cystic fibrosis and metabolic disorders. GK's brother, Romano Kroemer, was an employee of Sanofi and now consults for Boehringer-Ingelheim. The funders had no role in the design of the study; in the writing of the manuscript, or in the decision to publish the results.

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Author Contributions

PL and LZ summarized date and designed display items, LZ, OK and GK wrote the manuscript.

References

- 1. Pegoraro L, Palumbo A, Erikson J, Falda M, Giovanazzo B, Emanuel BS, et al. A 14;18 and an 8;14 chromosome translocation in a cell line derived from an acute B-cell leukemia. *Proc Natl Acad Sci U S A* 1984, **81**(22): 7166-7170.
- 2. Reed JC, Tsujimoto Y, Alpers JD, Croce CM, Nowell PC. Regulation of bcl-2 protooncogene expression during normal human lymphocyte proliferation. *Science* 1987, **236**(4806): 1295-1299.
- 3. Tsujimoto Y, Croce CM. Analysis of the structure, transcripts, and protein products of bcl-2, the gene involved in human follicular lymphoma. *Proc Natl Acad Sci U S A* 1986, **83**(14): 5214-5218.
- 4. Susin SA, Zamzami N, Castedo M, Hirsch T, Marchetti P, Macho A, et al. Bcl-2 inhibits the mitochondrial release of an apoptogenic protease. *J Exp Med* 1996, **184**(4): 1331-1341.
- 5. Yang J, Liu X, Bhalla K, Kim CN, Ibrado AM, Cai J, et al. Prevention of apoptosis by Bcl-2: release of cytochrome c from mitochondria blocked. *Science* 1997, **275**(5303): 1129-1132.
- 6. Kluck RM, Bossy-Wetzel E, Green DR, Newmeyer DD. The release of cytochrome c from mitochondria: a primary site for Bcl-2 regulation of apoptosis. *Science* 1997, **275**(5303): 1132-1136.
- 7. Chao DT, Korsmeyer SJ. BCL-2 family: regulators of cell death. *Annu Rev Immunol* 1998, **16:** 395-419.
- 8. Marsden VS, Strasser A. Control of apoptosis in the immune system: Bcl-2, BH3-only proteins and more. *Annu Rev Immunol* 2003, **21**: 71-105.
- 9. Singh R, Letai A, Sarosiek K. Regulation of apoptosis in health and disease: the balancing act of BCL-2 family proteins. *Nat Rev Mol Cell Biol* 2019, **20**(3): 175-193.
- 10. Czabotar PE, Garcia-Saez AJ. Mechanisms of BCL-2 family proteins in mitochondrial apoptosis. *Nat Rev Mol Cell Biol* 2023.
- 11. Pattingre S, Tassa A, Qu X, Garuti R, Liang XH, Mizushima N, et al. Bcl-2 antiapoptotic proteins inhibit Beclin 1-dependent autophagy. *Cell* 2005, **122**(6): 927-939.
- 12. Gross A, Katz SG. Non-apoptotic functions of BCL-2 family proteins. *Cell Death Differ* 2017, **24**(8): 1348-1358.
- 13. Chong SJF, Marchi S, Petroni G, Kroemer G, Galluzzi L, Pervaiz S. Noncanonical Cell Fate Regulation by Bcl-2 Proteins. *Trends Cell Biol* 2020, **30**(7): 537-555.

- 14. Oltersdorf T, Elmore SW, Shoemaker AR, Armstrong RC, Augeri DJ, Belli BA, et al. An inhibitor of Bcl-2 family proteins induces regression of solid tumours. *Nature* 2005, **435**(7042): 677-681.
- 15. Diepstraten ST, Anderson MA, Czabotar PE, Lessene G, Strasser A, Kelly GL. The manipulation of apoptosis for cancer therapy using BH3-mimetic drugs. *Nat Rev Cancer* 2022, **22**(1): 45-64.
- 16. Roberts AW, Davids MS, Pagel JM, Kahl BS, Puvvada SD, Gerecitano JF, et al. Targeting BCL2 with Venetoclax in Relapsed Chronic Lymphocytic Leukemia. *N Engl J Med* 2016, **374**(4): 311-322.
- 17. Stilgenbauer S, Eichhorst B, Schetelig J, Coutre S, Seymour JF, Munir T, et al. Venetoclax in relapsed or refractory chronic lymphocytic leukaemia with 17p deletion: a multicentre, open-label, phase 2 study. *Lancet Oncol* 2016, **17**(6): 768-778.
- 18. Deeks ED. Venetoclax: First Global Approval. *Drugs* 2016, **76**(9): 979-987.
- 19. DiNardo CD, Pratz KW, Letai A, Jonas BA, Wei AH, Thirman M, et al. Safety and preliminary efficacy of venetoclax with decitabine or azacitidine in elderly patients with previously untreated acute myeloid leukaemia: a non-randomised, open-label, phase 1b study. *Lancet Oncol* 2018, **19**(2): 216-228.
- 20. Eichhorst B, Niemann CU, Kater AP, Furstenau M, von Tresckow J, Zhang C, et al. First-Line Venetoclax Combinations in Chronic Lymphocytic Leukemia. *N Engl J Med* 2023, **388**(19): 1739-1754.
- 21. DiNardo CD, Jonas BA, Pullarkat V, Thirman MJ, Garcia JS, Wei AH, et al. Azacitidine and Venetoclax in Previously Untreated Acute Myeloid Leukemia. N Engl J Med 2020, **383**(7): 617-629.
- 22. Zhao L, Liu P, Mao M, Zhang S, Bigenwald C, Dutertre CA, et al. BCL2 inhibition reveals a dendritic cell-specific immune checkpoint that controls tumor immunosurveillance. *Cancer Discov* in press.
- 23. Steinman RM. Decisions about dendritic cells: past, present, and future. *Annu Rev Immunol* 2012, **30:** 1-22.
- 24. Richter C, Thieme S, Bandola J, Laugsch M, Anastassiadis K, Brenner S. Generation of inducible immortalized dendritic cells with proper immune function in vitro and in vivo. *PLoS One* 2013, **8**(4): e62621.
- 25. Zhao L, Liu P, Xie W, Zhang S, Thieme S, Zitvogel L, et al. A genotype-phenotype screening system using conditionally immortalized immature dendritic cells. *STAR Protoc* 2021, **2**(3): 100732.

- 26. Le Naour J, Liu P, Zhao L, Adjemian S, Sztupinszki Z, Taieb J, et al. A TLR3 Ligand Reestablishes Chemotherapeutic Responses in the Context of FPR1 Deficiency. *Cancer Discov* 2021, **11**(2): 408-423.
- 27. Zhang S, Zhao L, Guo M, Liu P, Li S, Xie W, et al. Anticancer effects of ikarugamycin and astemizole identified in a screen for stimulators of cellular immune responses. *J Immunother Cancer* 2023, **11**(7).
- 28. Lin ML, Zhan Y, Proietto AI, Prato S, Wu L, Heath WR, et al. Selective suicide of cross-presenting CD8+ dendritic cells by cytochrome c injection shows functional heterogeneity within this subset. *Proc Natl Acad Sci U S A* 2008, **105**(8): 3029-3034.
- 29. Steinman RM, Banchereau J. Taking dendritic cells into medicine. *Nature* 2007, **449**(7161): 419-426.
- 30. Cabeza-Cabrerizo M, Cardoso A, Minutti CM, Pereira da Costa M, Reis e Sousa C. Dendritic Cells Revisited. *Annu Rev Immunol* 2021, **39:** 131-166.
- 31. Kohlhapp FJ, Haribhai D, Mathew R, Duggan R, Ellis PA, Wang R, et al. Venetoclax Increases Intratumoral Effector T Cells and Antitumor Efficacy in Combination with Immune Checkpoint Blockade. *Cancer Discov* 2021, **11**(1): 68-79.
- 32. Hildner K, Edelson BT, Purtha WE, Diamond M, Matsushita H, Kohyama M, et al. Batf3 deficiency reveals a critical role for CD8alpha+ dendritic cells in cytotoxic T cell immunity. *Science* 2008, **322**(5904): 1097-1100.
- 33. Ma Y, Adjemian S, Mattarollo SR, Yamazaki T, Aymeric L, Yang H, et al. Anticancer chemotherapy-induced intratumoral recruitment and differentiation of antigen-presenting cells. *Immunity* 2013, **38**(4): 729-741.
- 34. Galluzzi L, Buque A, Kepp O, Zitvogel L, Kroemer G. Immunological Effects of Conventional Chemotherapy and Targeted Anticancer Agents. *Cancer Cell* 2015, **28**(6): 690-714.
- 35. Petroni G, Buque A, Zitvogel L, Kroemer G, Galluzzi L. Immunomodulation by targeted anticancer agents. *Cancer Cell* 2021, **39**(3): 310-345.
- 36. Zitvogel L, Apetoh L, Ghiringhelli F, Andre F, Tesniere A, Kroemer G. The anticancer immune response: indispensable for therapeutic success? *J Clin Invest* 2008, **118**(6): 1991-2001.
- 37. Galluzzi L, Humeau J, Buque A, Zitvogel L, Kroemer G. Immunostimulation with chemotherapy in the era of immune checkpoint inhibitors. *Nat Rev Clin Oncol* 2020, **17**(12): 725-741.

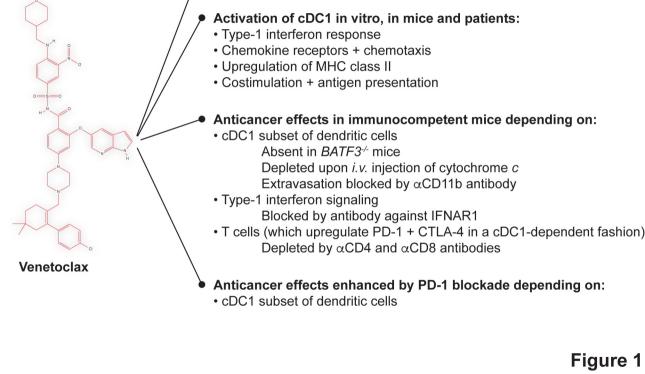
- 38. Brune M, Castaigne S, Catalano J, Gehlsen K, Ho AD, Hofmann WK, et al. Improved leukemia-free survival after postconsolidation immunotherapy with histamine dihydrochloride and interleukin-2 in acute myeloid leukemia: results of a randomized phase 3 trial. *Blood* 2006, **108**(1): 88-96.
- 39. Rosenblatt J, Stone RM, Uhl L, Neuberg D, Joyce R, Levine JD, et al. Individualized vaccination of AML patients in remission is associated with induction of antileukemia immunity and prolonged remissions. *Sci Transl Med* 2016, **8**(368): 368ra171.
- 40. Daver N, Garcia-Manero G, Basu S, Boddu PC, Alfayez M, Cortes JE, et al. Efficacy, Safety, and Biomarkers of Response to Azacitidine and Nivolumab in Relapsed/Refractory Acute Myeloid Leukemia: A Nonrandomized, Open-Label, Phase II Study. *Cancer Discov* 2019, **9**(3): 370-383.
- 41. Uy GL, Aldoss I, Foster MC, Sayre PH, Wieduwilt MJ, Advani AS, et al. Flotetuzumab as salvage immunotherapy for refractory acute myeloid leukemia. *Blood* 2021, **137**(6): 751-762.
- 42. Wemeau M, Kepp O, Tesniere A, Panaretakis T, Flament C, De Botton S, et al. Calreticulin exposure on malignant blasts predicts a cellular anticancer immune response in patients with acute myeloid leukemia. *Cell Death Dis* 2010, **1:** e104.
- 43. Truxova I, Kasikova L, Salek C, Hensler M, Lysak D, Holicek P, et al. Calreticulin exposure on malignant blasts correlates with improved natural killer cell-mediated cytotoxicity in acute myeloid leukemia patients. *Haematologica* 2020, **105**(7): 1868-1878.
- 44. Gillissen MA, Kedde M, Jong G, Moiset G, Yasuda E, Levie SE, et al. AML-specific cytotoxic antibodies in patients with durable graft-versus-leukemia responses. *Blood* 2018, **131**(1): 131-143.
- 45. Zitvogel L, Kepp O, Kroemer G. Decoding cell death signals in inflammation and immunity. *Cell* 2010, **140**(6): 798-804.
- 46. Henson PM. Cell Removal: Efferocytosis. *Annu Rev Cell Dev Biol* 2017, **33:** 127-144.
- 47. Nagata S. Apoptosis and Clearance of Apoptotic Cells. *Annu Rev Immunol* 2018, **36**: 489-517.
- 48. Galluzzi L, Yamazaki T, Kroemer G. Linking cellular stress responses to systemic homeostasis. *Nat Rev Mol Cell Biol* 2018, **19**(11): 731-745.
- 49. Kroemer G, Galassi C, Zitvogel L, Galluzzi L. Immunogenic cell stress and death. *Nat Immunol* 2022, **23**(4): 487-500.
- 50. Mehrotra P, Ravichandran KS. Drugging the efferocytosis process: concepts and opportunities. *Nat Rev Drug Discov* 2022, **21**(8): 601-620.

51.	Liu P, Zhao L, Zitvogel L, Kepp O, Kroemer G. Immunogenic cell death (ICD) enhancers- Drugs that enhance the perception of ICD by dendritic cells. <i>Immunol Rev</i> 2023.

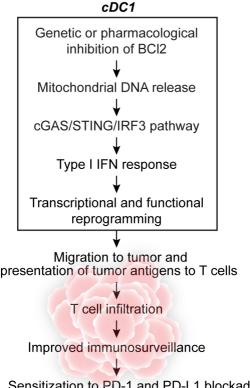
Figure legends

Figure 1: Mechanisms of anticancer effects of venetoclax against solid cancers. For details consult main text. The chemical structure of venetoclax was adapted from Pubchem.

Figure 2: Cascade of molecular, cellular and immunological events linking the inhibition of BCL2 to the stimulation of anticancer immune responses in vivo. For details consult main text.



No direct effects on most solid cancers (e.g. MC38, MCA205, TC1)



Sensitization to PD-1 and PD-L1 blockade

Figure 2