

# Apoptotic cell death in disease-Current understanding of the NCCD 2023

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#### ▶ To cite this version:

Ilio Vitale, Federico Pietrocola, Emma Guilbaud, Stuart A Aaronson, John M Abrams, et al.. Apoptotic cell death in disease-Current understanding of the NCCD 2023. Cell Death and Differentiation, 2023, pp.1097-1154. 10.1038/s41418-023-01153-w. hal-04596448

HAL Id: hal-04596448

https://hal.science/hal-04596448

Submitted on 11 Jun 2024

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## **Apoptotic cell death in disease**

# - Current understanding of the NCCD 2023

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- **Running Title:** Apoptosis in physiology and pathology
- 62 **Keywords:** BAX; BCL2; cancer; death receptors; ischemia; mitochondrial outer membrane
- 63 permeabilization; stroke.
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All the Editorial Board Members of Cell Death Differentiation, Cell Death Disease, or Cell Death Discovery are included among the authors. A.D. is shareholder in Denali Therapeutics. A.D.G. has received renumeration, honorarium or consultancy fees from Boehringer Ingelheim, Miltenyi Biotec or IsoPlexis. A.S. and P.E.C. are employees of the Walter and Eliza Hall Institute, which has an agreement with Genentech and AbbVie and receives milestone and royalty payments related to venetoclax. Employees of the Walter and Eliza Hall Institute may be eligible for financial benefits related to these payments. P.E.C. receives such a financial benefit as a result of previous research related to venetoclax. B.D.S. has no COI with the content of this paper. Possible perceived COI: B.D.S.is or has been a consultant for Eli Lilly, Biogen, Janssen Pharmaceutica, Eisai, AbbVie and other companies. B.D.S. is also a scientific founder of Augustine Therapeutics and a scientific founder and stockholder of Muna therapeutics. B.R.S. is an inventor on patents and patent applications involving small molecule drug discovery, ferroptosis, and the antibody 3F3-FMA; co-founded and serves as a consultant to Exarta Therapeutics, and ProJenX Inc.; holds equity in Sonata Therapeutics, and serves as a consultant to Weatherwax Biotechnologies Corporation and Akin Gump Strauss Hauer & Feld LLP, and receives research funding from Sumitomo Dainippon Pharma Oncology. C.M.T. has the following patent applications US20200164026, US20190142915, US20150165061, US20140024597. US2020058683, WO2018013519, WO/2020/223212. D.C.R. serves as a consultant for Alladdin Healthcare Technologies Ltd., Mindrank AI, Nido Biosciences, Drishti Discoveries and PAQ Therapeutics. D.J.M receives funding from the Merck Group (Darmstadt, Germany) & Puma Biotechnology (Los Angeles, CA, USA). D.O. is employed at SANOFI. J.T.O. serves as a consultant for Anji Pharmaceuticals. J.T.O receives research funding from AbbVie. D.W.A. receives funding for research in apoptosis from Amylyx. F.D.V. is Member of the Scientific Advisory Board (SAB) of Biosceptre Ltd (UK), and a consultant with Axxam SpA (Italy). F.M. has financial interest in TLL, The Longevity Labs and Samsara Therapeutics. P.M. is founder and shareholder of NETRIS Pharma. G.A.C. is the scientific founder of Ithax Pharmaceuticals. G.L.K. is an employee of the Walter and Eliza Hall Institute which receives milestone and royalty payments related to venetoclax. G.L.K. has received research funding from Servier. J.R.C.-R. is a scientific consultant for NextRNA Therapeutics, Inc. and Autoimmunity Biologic Solutions, Inc.. K.R. is a founder and MD of KH Biotec GmbH. C.V.R is a scientific founder and member of the Scientific Advisory Board (SAB) of Surface Oncology, a member of Janssen Immunology SAB, and a consultant for the Roche Immunology Incubator. C.V.R has received grant support from Mirati Therapeutics. K.S. is a co-inventor on patent applications for NLRP3 inhibitors which have been licensed to Inflazome Ltd, a company headquartered in Dublin, Ireland. Inflazome is developing drugs that target the NLRP3 inflammasome to address unmet clinical needs in inflammatory disease. K.S. served on the Scientific Advisory Board of Inflazome in 2016–2017, and serves as a consultant to Quench Bio, USA and Novartis, Switzerland. L.A. receives funding for research from Merck and has shares in Epic SRL and CIRCE SRL. M.C. (Michele Carbone) received donations from the UH Foundation through donations from: the Riviera United-4-a Cure, the Melohn Family Endowment, the Honeywell International Inc., the Germaine Hope Brennan Foundation, and the Maurice and Joanna Sullivan Family Foundation. M.C. (Michele Carbone) has a patent issued for BAP1. B.J. is cofounder of CERVO Therapeutics AB. M.C. and two patents issued for HMGB1. M.C. (Michele Carbone) is a board-certified pathologist who provides consultation for pleural pathology, including medical-legal. M.C. (Marcus Conrad) is a co-founder and shareholder of ROSCUE Therapeutics GmbH. M.K. received support from Jansen Pharmaceuticals, Merck and Gossamer Bioscience. M.K. is founder and member of SAB, Elgia Pharmaceuticals. 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BioPharma. S.v.K. is named inventor on patent applications covering some of the therapeutic concepts pertaining to TRAIL-R blockade in disease. Y.H.C. is a member of the board of advisors for Amshenn Inc. and Binde Inc.. T.-D.K. is a consultant for Pfizer. T.V.B. and P.V. hold patents related to ferrostatin-1 analogues. W.S.E-D. is founder and shareholder (no research funding) of Oncoceutics/Chimerix, which is developing ONC201/TIC10 as cancer therapeutic. W.S.E-D. is founder and shareholder (no research funding) of p53-Therapeutics, an early-stage company developing small molecules targeting mutant p53. W.S.E-D. founder, and shareholder (no research funding) of SMURF-Therapeutics, an early-stage company developing small molecules targeting hypoxia. W.S.E-D. is Co-Chair in the Executive Committee for Precision Oncology Alliance (no research funding) of Caris Life Sciences. W.S.E-D. receives support from D&D Pharma and AACR-Novocure. W.S.E-D. is the advisory board of Ocean Biomedical. W.S.E-D. is the advisory board of RAIN Therapeutics. G.K. has been holding research contra.cts with Daiichi Sankyo, Eleor, Kaleido, Lytix Pharma, PharmaMar, Osasuna Therapeutics, Samsara Therapeutics, Sanofi, Tollys, and Vascage. G.K. has been consulting for Reithera. G.K. is on the Board of Directors of the Bristol Myers Squibb Foundation France. G.K. is a scientific co-founder of everImmune, Osasuna Therapeutics, Samsara Therapeutics and Therafast Bio. G.K is the inventor of patents covering therapeutic targeting of aging, cancer, cystic fibrosis and metabolic disorders. G.K.'s brother, Romano Kroemer, was an employee of Sanofi and now consults for Boehringer-Ingelheim. LG is/has been holding research contracts with Lytix Biopharma, Promontory and Onxeo, has received consulting/advisory honoraria from Boehringer Ingelheim, AstraZeneca, OmniSEQ, Onxeo, The Longevity Labs, Inzen, Imvax, Sotio, Promontory, Noxopharm, EduCom, and the Luke Heller TECPR2 Foundation, and holds Promontory stock options. All other authors have no conflicts to declare.

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#### Abstract

Apoptosis is a form of regulated cell death (RCD) that involves proteases of the caspase family. Pharmacological and genetic strategies that experimentally inhibit or delay apoptosis in mammalian systems have elucidated the key contribution of this process not only to (post-)embryonic development and adult tissue homeostasis but also to the etiology of multiple human disorders. Consistent with this notion, while defects in the molecular machinery for apoptotic cell death impair organismal development and promote oncogenesis, the unwarranted activation of apoptosis promotes cell loss and tissue damage in the context of various neurological, cardiovascular, renal, hepatic, infectious, neoplastic and inflammatory conditions. Here, the Nomenclature Committee on Cell Death (NCCD) gathered to critically summarize abundant pre-clinical literature mechanistically linking the core apoptotic apparatus to organismal homeostasis in the context of disease.

#### 391 Facts

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- Intrinsic and extrinsic apoptosis are forms of regulated cell death (RCD) promoting the cellular demise along with the activation of proteases of the caspase family.
  - In mammalian organisms, executioner caspases are activated after cells are already committed to die.
  - Apoptosis can be manipulated by genetic or pharmacological means, and multiple genetically
    engineered animal models and pharmacological tools to modulate apoptosis have been developed.
  - Apoptosis is intimately involved in both (post-)embryonic development and adult tissue homeostasis.
    - Apoptosis deregulation promotes oncogenesis and contributes to the etiology of multiple human disorders, including cardiovascular, hepatic, inflammatory and neurological conditions.
  - To date, venetoclax is the only apoptosis inducer that has received regulatory approval for use in humans.

### **Open Questions**

- Will inhibitors of apoptotic caspases with elevated target specificity become available?
- Will agents specifically conceived to modulate apoptosis enter the clinical practice to treat solid tumors or other human disorders beyond hematological malignancies?
  - Is it conceivable to design combinatorial strategies aimed at inhibiting apoptosis while interrupting compensatory activation of other RCD signaling cascades?
- Will it be possible to specifically inhibit apoptotic signaling without impacting on other processes
  dependent on apoptosis regulators such as differentiation, proliferation, and inflammatory
  reactions?

## Introduction

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The health and homeostasis of multicellular organisms depend on the tight balance between cell proliferation and cell death. In this context, a large body of experimental evidence has demonstrated the existence of a form of regulated cell death (RCD) that is executed by a genetically programmed process, and hence amenable to manipulation by genetic or pharmacological means <sup>1</sup>. Over the past decades, multiple variants of RCD have been characterized at the genetic, biochemical, functional, and immunological level <sup>2, 3, 4, 5, 6, 7, 8</sup>. For instance, programmed cell death (PCD) has been functionally defined as a modality of RCD activated under purely physiological conditions (i.e., in the absence of perturbations of extracellular or intracellular homeostasis) in the context of embryonic/post-embryonic development or adult tissue homeostasis <sup>1,9</sup>. Conversely, pathological RCD is invariably initiated in the context of failure to adapt to shifts in extra-cellular or intra-cellular homeostasis, constituting a de facto organismal program for the elimination of excessively damaged and/or potentially harmful cells, such as cells infected with pathogens 1, 10. From a biochemical perspective, an increasing number of RCD modalities have been defined by the Nomenclature Committee on Cell Death (NCCD) based on the mechanistic involvement of specific molecular components <sup>1, 11</sup>. For instance, apoptotic cell death has been defined as a form of RCD that is mainly executed by proteases of the caspase family, namely caspase 3 (CASP3), CASP6 and CASP7 initiated by CASP8 and CASP9 1, 12, 13. However, in mammalian organisms, with the exception of CASP8, apoptotic caspases simply accelerate RCD because their activation occurs when cells are already committed to die <sup>1, 14, 15, 16</sup>. This means that contrarily to simpler organisms (e.g., C. elegans), in which apoptotic caspase elimination fully rescues cells from death, in mammals, apoptotic cell death can at most be retarded but not prevented by pharmacological or genetic strategies inhibiting the activity of these caspases. Mitochondrial permeability transition (MPT)-driven necrosis, necroptosis, ferroptosis, pyroptosis, parthanatos, entotic cell death, NETotic cell death, lysosome-dependent cell death, and autophagy-dependent cell death represent forms of RCD that involve precise molecular events and hence can also be manipulated with pharmacological or genetic interventions <sup>1, 2, 3, 4, 5, 6, 17, 18, 19</sup>. Other RCD modalities have been recently identified, such as alkaliptosis <sup>20</sup>, cuproptosis <sup>21</sup> and PANoptosis (involving the simultaneous activation of pyroptosis, apoptosis, and necroptosis) <sup>22</sup>, and their signal transduction modules are under investigation. The importance of several of these forms of RCD in health and disease is not yet known.

Along with the identification of key RCD regulators and the advent of modern tools for genetic manipulation, a great experimental effort has been devoted to elucidating the role of RCD in the physiopathology of multi-cellular organisms <sup>23</sup>. Thus, various studies in animals (mostly rodents) genetically altered to be deficient for or over-express components of the apoptotic apparatus (either at the whole-body level or in selected cell/tissue types) have provided formal proof of the relevance, but not always the exquisite requirement, of apoptosis for embryonic and fetal development or adult tissue homeostasis <sup>24, 25, 26</sup>.

Along similar lines, pharmacological and genetic tools aimed at altering apoptotic signaling in preclinical disease models revealed the mechanistic contribution of apoptosis to the etiology of various conditions associated with the loss of post-mitotic or (in certain settings) non-post-mitotic cells, including a panel of neurological, cardiovascular, renal, hepatic, and inflammatory disorders <sup>24</sup>. Extensive studies over the last five decades highlighted the apoptotic machinery as a major target for the development of new therapeutic interventions <sup>27</sup>, not only for the induction of cell death in the context of disrupted tissue homeostasis (e.g., for neoplastic diseases) <sup>28</sup>, but also for the inhibition of cell death in the context of ischemic, degenerative and inflammatory conditions <sup>29, 30</sup>. However, while at least one drug designed to induce apoptosis is currently approved for use in humans, namely the BCL2 apoptosis regulator (BCL2) inhibitor venetoclax <sup>31, 32, 33, 34</sup> which is used alone or in combinatorial regimens for the treatment of

chronic lymphocytic leukemia (CLL), small lymphocytic lymphoma and acute myeloid leukemia (AML) <sup>31, 35, 36, 37, 38</sup>, no other agents specifically conceived to inhibit the apoptotic apparatus have been licensed for clinical practice so far. The broad-spectrum caspase inhibitor emricasan received fast-track designation by the US Food and Drug Administration (FDA) for the treatment of non-alcoholic steatohepatitis in 2016 but demonstrated inconsistent clinical efficacy <sup>39, 40, 41</sup>, and – as of now– is not approved for therapy in humans.

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The lack of clinically approved, selective apoptosis inhibitors and the inconclusive performance of emricasan in recent trials reflect several aspects of (apoptotic and non-apoptotic) RCD that began to emerge only recently (**Figure 1**). First, while detecting cell death as well as biomarkers of specific RCD variants in vitro is relatively straightforward <sup>42</sup>, precise quantification of cell death in vivo in adult tissue remains challenging, at least in part because of rapid disposal of cell corpses by efferocytosis <sup>43, 44, 45, 46</sup>. Thus, the actual contribution of cell death to the etiology of various human disorders is difficult to quantify by observational approaches <sup>47, 48</sup>. Second, while for a long-time, specific forms of RCD were considered virtually independent entities, recently it became clear that the molecular machinery for RCD is composed of highly interconnected modules characterized by substantial redundancy, backup pathways and feedback loops <sup>10, 49, 50</sup>. Thus, molecules that inhibit one specific form of RCD may ultimately be unable to confer actual cyto- and tissue protection instead only altering the kinetic and biochemical manifestations of death by allowing the engagement of a different RCD sub-routine. For instance, while CASP8 is a major signal transducer in death receptor (DR)-driven apoptosis (see below), it intrinsically inhibits necroptosis induced by DR and certain other signaling pathways, such as Toll-like receptor (TLR) signaling 51, 52, 53, suggesting that caspase inhibition in the context of DR signaling may promote necroptotic cell death <sup>54, 55, 56, 57</sup>. Together with a low target specificity and selectivity within the caspase family <sup>57</sup>, this can explain the inadequate efficacy of emricasan observed in pre-clinical and clinical studies. Third, even in the hypothetical scenario of agents capable of simultaneous inhibition of

all (known and unknown) RCD pathways, loss of cellular homeostasis due to failing adaptation to stress generally involve degenerative processes that at some stage cannot be reversed, such as widespread mitochondrial permeabilization and loss of RNA and protein synthesis 4, 58, 59, 60, i.e., even if all RCD modalities could be blocked effectively, cells might undergo uncontrolled necrotic death. In this setting, cell death may occur as a consequence of an irremediable degeneration of cellular functions that can no longer be rescued pharmacologically or even genetically <sup>61</sup>. Supporting these latter notions, accumulating literature indicates that, at least in mammalian systems, perhaps with the exception of CASP8, so-called apoptotic caspases mainly control the kinetics of apoptotic cell death and its immunological manifestations, but not whether cell death ultimately occurs or not, <sup>15, 16</sup>. This points to the caspase family as a major regulator of organismal homeostasis via control of inflammatory responses 62, 63. The simultaneous inhibition of multiple caspases, as for instance by emricasan, may thus also impact inflammation, as was demonstrated for TNF-induced systemic inflammatory respiratory syndrome (SIRS) in vivo for the pan caspase-inhibitor zVAD-fmk <sup>54, 64</sup>. To complicate matters, multiple components of the core apoptotic machinery, including caspases and multiple members of the BCL2 family have been reported to regulate a variety of non-apoptotic functions beyond inflammation, such as mitochondrial energy production, Ca<sup>2+</sup> signaling and terminal differentiation <sup>65, 66, 67, 68, 69, 70, 71, 72</sup>. Structurally, distinguishing between apoptotic and non-apoptotic functions of caspases and the BCL2 family remains challenging. Finally, there is a hitherto unclarified heterogeneity in the regulation of RCD at distinct anatomical sites (possibly linked to micro-environmental features) at distinct stages of cellular differentiation, and in the context of diverse patho-physiological states (e.g., in young vs. adult and aged individuals).

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All these issues should also be kept under consideration in the context of the present review, in which the NCCD aims at critically discussing a large amount of pre-clinical data in support of a key role for the apoptotic machinery in mammalian diseases. Specifically, the interpretation of results of genetic and pharmacological experiments presented herein should place particular attention on the aforementioned connectivity amongst different RCD variants as well as on discriminating between essential *vs.* accessory aspects of cell death <sup>14</sup>. Another issue to be considered is the fact that most conclusions are based on use of knockout/congenic mice which often present other passenger mutations potentially influencing the observed phenotype <sup>73</sup>. Our objective is not only to provide a critical summary of the existing literature, but also to offer an updated framework for interpretation of these findings in view of currently accepted models of RCD signaling.

#### **Intrinsic apoptosis in disease**

There are substantive supporting data from genetic studies to demonstrate that the molecular machinery for intrinsic apoptosis (described in **Box 1** and **Figure 2**) is involved in embryonic and fetal development as well as in adult tissue homeostasis. Numerous preclinical studies in animal models of disease demonstrate that intrinsic apoptosis contributes to etiology in various disorders involving the loss of not only post-mitotic, but also non-post-mitotic tissues, including neurological, cardiac, renal, hepatic, autoimmune/inflammatory, oncological, and infectious conditions. However, as discussed above, the interpretation of these results should be taken with caution given the high interconnectivity of RCD pathways and the crosstalk between RCD and inflammatory response. Moreover, the activation of executioner caspases occurs after cells are already committed to intrinsic apoptosis <sup>15, 16</sup>. Accordingly, caspase inhibition only delays the execution of cell death. In this context, the phenotypes observed under apoptotic caspase-deleted or inhibited conditions may reflect cell-extrinsic effects of caspase activity such as the release of immunomodulatory and cytotoxic signals from dying/dead cells, including damage-associated molecular patterns (DAMPs) or cytokines (this concept is extensively discussed in <sup>14</sup>). These phenotypes may also stem from the lack of processes independent of intrinsic (or extrinsic) apoptosis,

as, for instance, the lack of CASP3-mediated cleavage of gasdermin E (GSDME) leading to impaired pyroptosis and associated inflammatory response <sup>74,75</sup>.

Below, we will provide details of the pro-apoptotic BCL2 proteins, the anti-apoptotic BCL2 proteins, the components of the apoptosome - a platform for the activation of initiator caspases composed of cytochrome c, somatic (CYCS), apoptotic peptidase activating factor 1 (APAF1) and pro-CASP9 - and effector caspases in disease. The instances of involvement encompass participation in the pathogenic mechanisms as well as experimental deletion or inhibition as a means of exploring potential utility as treatment targets. The effects of these regulators and effectors of the intrinsic apoptosis pathway on health are described in **Box 2**, **Box 3** and **Box 4**.

Neurological disorders. Intrinsic apoptotic factors are implicated in the pathophysiology of numerous neurological diseases (Figure 3). In a mouse model of amyotrophic lateral sclerosis (ALS), deletion of BCL2-associated X protein (*Bax*) reduces neuronal cell death coupled to attenuated motor dysfunction and neuromuscular degeneration <sup>76</sup>. Additional ablation of BCL2-antagonist/killer 1 (*Bak1*) further enhances neuroprotection, resulting in improved overall animal survival <sup>77</sup>. Similar protective effects were observed in mice lacking the BH3-only proteins BCL2 like 11 (BCL2L11, best known as BIM) and BCL2 binding component 3 (BBC3, best known as PUMA), as well as in transgenic mice overexpressing BCL2, X-linked inhibitor of apoptosis (XIAP) <sup>78, 79, 80, 81, 82</sup>. Moreover, intra-cerebroventricular administration of the broad-spectrum inhibitor Z-VAD-FMK protects mice from ALS <sup>83</sup>, although whether such protection arises from the inhibition of intrinsic apoptosis was not proven. *Bax* deletion also attenuates neuromuscular dysfunctions in a mouse model of congenital muscular dystrophy (another neurodegenerative disease affecting motoneurons) <sup>84</sup>, while BCL2 overexpression limits neuromuscular disease progression in some (but not all) mouse models of progressive motor neuronopathy and muscular dystrophy <sup>85, 86, 87</sup>. Finally, genetic or pharmacological inhibition of poly (ADP-ribose) polymerase

family, member 1 (PARP1) and PARP2 halts axonal degeneration and improves related motor phenotypes in *Caenorhabditis elegans* models of ALS <sup>88</sup>.

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Multiple components of the molecular machinery for intrinsic apoptosis, including BAX, PUMA, BH3 interacting domain death agonist (BID), Harakiri, BCL2 interacting protein (contains only BH3 domain) (HRK), were shown to drive neuronal death in Alzheimer's disease (AD) and Parkinson's disease (PD) models <sup>89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101</sup>. Thus, overexpression of BCL2 decreases the appearance of early pathological markers of AD, such as amyloid precursor protein (APP) and microtubule-associated protein tau (MAPT, best known as tau) cleavage, which depend on caspases 102, 103, 104, resulting in attenuated neurological defects <sup>105, 106</sup>. Some findings indicate a role of apoptotic caspases in the pathogenesis of AD. However, as discussed above, during intrinsic apoptosis, caspases simply accelerate the course of cell death, and, so, such effects may be linked to the release of cytotoxic and proinflammatory factors from dying cells. In more detail, pharmacological inhibition of CASP3 reduces early synaptic failure in mouse models of AD, ultimately improving cognitive defects <sup>107</sup>. Moreover, expression of a mutated form of amyloid β (an APP cleavage product) or administration of broadspectrum caspase inhibitors attenuates synaptic defects in models of AD, an effect only partially recapitulated by CASP3-specific inhibitors <sup>108</sup>. Along similar lines, genetic deletion of *Casp2* was reported to provide protection from synaptic loss and cognitive decline in a mouse model of AD <sup>109</sup>. Such protection may be linked to the generation of a specific cleavage product (Δtau314) by CASP2, which is reported to impair cognitive and synaptic function by promoting the missorting of tau to dendritic spines <sup>110, 111</sup>. Accordingly, CASP2 inhibitors blocked tau truncation and restored excitatory neurotransmission in mouse models of tauopathies, including AD <sup>112, 113</sup>. Of note, a role for CASP4 in AD pathogenesis is also reported 114, 115. Moreover, studies using senescence-accelerated OXY5 rat model of AD demonstrated that the treatment with mitochondria-targeted antioxidant SkQ1 improved mitochondrial fitness and slowed down the signs of Alzheimer's disease-like pathology in older rats <sup>116</sup>. Lack of BIM

(due to deletion of Bcl2111) also confers protection to dopaminergic neurons in experimental PD imposed by inhibition of mitochondrial complex I, an effect that depends on BAX activation <sup>117</sup>. In addition, genetic deletion or down-regulation of Casp3, as well CASP3 inhibition by transgenic, neuron-restricted expression of XIAP protects mice against pharmacologically induced PD, attenuating both dopaminergic neuron alterations and behavioral deficits <sup>118, 119, 120, 121</sup>. Whether protection arises from the lack of cellintrinsic or cell-extrinsic processes dependent on apoptotic caspases has not been investigated. Finally, pharmacological inhibition of CASP3 confers neuroprotection to rat model of Huntington's disease (HD) 122, 123, 124. That said, the precise mechanisms whereby components of the molecular apparatus for intrinsic apoptosis influence neurodegeneration need to be further explored. Two studies in clear contradiction to each other reported that at sublethal doses, pharmacological inhibition of myeloid cell leukemia sequence 1 (MCL1) improved disease outcome in a mouse model of AD with a mechanism independent of apoptosis induction and involving the stimulation of mitophagy <sup>125</sup>, but that Mcl1 haploinsufficiency accelerated the degeneration and dysfunctionality of motor neurons in mice <sup>126</sup>. Also, there is evidence that necroptosis or ferroptosis rather than apoptosis can be the major contributor in neuronal cell destruction during AD 127, 128. Finally, although Bax deletion prevents the demise of cerebellar granule neurons in a transgenic model of inherited prion disease <sup>129</sup>, the direct contribution of BAX to neurotoxicity during prion disorders is a matter of controversy <sup>130</sup>. BCL2 family proteins have also been reported to contribute to axonal degeneration and neuronal cell death in animal models of brain trauma, degeneration, or neurotoxicity <sup>131, 132, 133</sup>. Thus, BAX- or BIDdeficient mice, as well as transgenic mice overexpressing BCL2, display increased survival of cortical or hippocampal neurons after experimental traumatic brain injury, as compared to wild-type mice <sup>134, 135,</sup> <sup>136, 137</sup>. Moreover, transgenic BCL2 overexpression protects mouse neurons against the detrimental effects of transection of the sciatic nerve <sup>138</sup>. Likewise, BAX deficiency enhances the survival of

oligodendrocytes in mice subjected to spinal cord injury <sup>139</sup>. Both neuroprotection and functional

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improvements were observed in rat or mouse models of traumatic spinal cord injury upon local administration of Z-VAD-FMK) and other caspase inhibitors <sup>140, 141, 142</sup>. However, these findings need to be validated given the low selectivity of these inhibitors among caspases. Of note, in rats, post-traumatic neuroprotection can further be improved by combined inactivation of PARP1 and CASP3 <sup>143</sup>, suggesting a potential involvement for PARP1-dependent parthanatos in the process.

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Deletion of Bax (but not of the genes encoding BIM, PUMA or BID), as well as Bax haploinsufficiency, prevents the death or degeneration of retinal ganglion cells in mice subjected to optic nerve injury 144, 145, <sup>146, 147</sup>. Moreover, the demise of injured retinal ganglion cells is exacerbated in mice with a conditional loss of Bcl211 (leading to lack of BCL-X<sub>L</sub>) <sup>148</sup> and decreased in transgenic mice over-expressing XIAP <sup>149</sup> or BCL-X<sub>L</sub> <sup>150</sup> in the eye, or in rodents treated with an XIAP-derived cell-permeant peptide targeting CASP9 <sup>151</sup>, or a CASP3-targeting small-interfering RNA (siRNA) <sup>152, 153</sup>. Moreover, transgenic or adenovirus-driven XIAP expression protects the retina in various animal models of retinal disease, degeneration, or ischemia 154, 155, 156, 157, 158, 159, while a BCL-X<sub>L</sub> inhibitor alleviated pathogenic neovascularization during diabetic retinopathy <sup>160</sup>. Genetic deletion of *Casp9* from endothelial cells protected retinal ganglion cells from ischemic death, supporting non-cell autonomous functions of CASP9 <sup>151</sup>. Of note, CASP7 seems to play a crucial role in retinal ganglion cell death, as demonstrated in a model of optic injury in Casp7<sup>-/-</sup> mice <sup>161</sup>. However, both pro-survival (BCL2) and pro-apoptotic (BAK1, BAX and BIM) BCL2 family members contribute to retinal neo-vascularization in response to experimental ischemic retinopathy 162, 163, 164. In one of these papers, this effect is linked to an increased survival of endothelial cells in the absence of BAX and BAK1 <sup>164</sup>. Persistent endothelial cells promote rapid tissue re-vascularization, thus preventing the occurrence of a pathogenic excessive neovascularization. Moreover, the inhibition of the intrinsic apoptotic pathway by genetic inhibition of c-Jun N-terminal kinase 1 (Jnk1) or the administration of a broad-spectrum caspase inhibitor led to reduced choroidal neovascularization in the murine model of wet age-related macular degeneration (AMD) <sup>165</sup>. These observations may indicate that factors released by dying cells regulate neo-vascularization in the retina or other eye tissues.

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Deletion of Bax, Hrk or Casp3 as well as transgenic overexpression of XIAP prevents neuronal loss and/or axon degeneration in mouse models of trophic factor deprivation including nerve growth factor (NGF) withdrawal <sup>166, 167, 168</sup>. Conversely, lack of BIM or PUMA does not limit hippocampal neuronal injury upon experimental excitotoxicity 169, 170. Moreover, while in vivo delivery of an XIAP fusion protein protects neurons against death induced by glutamate or kainic acid <sup>171</sup>, kainic acid-mediated neurodegeneration cannot be rescued by the CASP3 inhibitor DEVD-CHO <sup>172</sup>. Conversely, BIM appears to be activated during excitotoxicity <sup>173</sup>, and *Bcl2l11*<sup>-/-</sup> mice (which lack BIM) display attenuated neurodegeneration after experimental seizures induced by administration of kainic acid into the amygdala, at least in part because of decreased neuronal cell death in the hippocampus (but not in the neocortex) <sup>174</sup>. Moreover, data from knockout mice suggest that experimental seizure-induced neuronal death involves BCL2-associated agonist of cell death (BAD), BCL2 interacting killer (BIK), BCL2 modifying factor (BMF), or PUMA <sup>175, 176, 177, 178</sup> and that BCL2-like 2 (BCL2L2; best known as BCL-W) may provide neuroprotective, seizure-suppressive functions <sup>179</sup>. Confirming a certain degree of functional redundancy, phorbol-12-myristate-13-acetate-induced protein 1 (PMAIP1, best known as NOXA) and BID seem dispensable for RCD driven by excitotoxicity, as shown in kainic acid-treated animals <sup>180, 181</sup>. Intrinsic apoptosis is also involved in neuronal apoptosis post-ischemic injury in both developing and adult brains. In a mouse model of neonatal hypoxia-ischemia, neuroprotection was documented upon deletion of Bax <sup>182</sup>, simultaneous absence of BIM and BAD <sup>183</sup>, or transgenic overexpression of XIAP <sup>184</sup>. Conversely, *Xiap*<sup>-/-</sup> mice are sensitized to neonatal hypoxia-ischemia injury <sup>185</sup>. Apparently at odds

with these findings, Casp3<sup>-/-</sup> mice display increased vulnerability to such experimental perturbation,

possibly due to complementary over-activation of CASP3-independent pathways <sup>186</sup>. Of note, the

absence of CASP3, BAX, or PUMA (but not the absence of NOXA, BIM or HRK) also confers neuroprotection to newborn mice acutely exposed to ethanol <sup>187, 188, 189</sup>, while loss of BAX is neuroprotective in newborn mice exposed to isoflurane <sup>190</sup> as well as ionizing radiation <sup>133, 191</sup>. At the same time, it is interesting to note that BAX-dependent neuronal RCD also contributes to reactive microgliosis during the recovery of the developing brain from acute alcohol exposure <sup>192</sup>, pointing to an etiological role for activation of microglial cells by dead neurons. Bax<sup>-/-</sup> mice displayed pronounced neuroprotection when subjected to distinct experimental brain injuries, including middle cerebral artery occlusion <sup>193</sup>. A similar protection against experimental ischemic insults has been observed in mice deficient for BMF <sup>194</sup>, or BID <sup>195, 196, 197</sup>. Conversely, NOXA seems to be dispensable for neuronal damage induced by experimental ischemic stroke <sup>194</sup>. Moreover, the absence of BID fails to protect mice from ischemia-reperfusion, although it limits the associated inflammatory response <sup>198</sup>. Transgenic over-expression of BCL2, BCL-X<sub>L</sub> or XIAP as well as inhibition of apoptotic caspases or genetic deletion of CASP6 ameliorates neuronal survival upon global ischemia, focal ischemia or stroke <sup>199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215</sup>. It should be noted, however, that in these settings, neuroprotection by inhibition or deletion of caspases may be related to the lack of cell-extrinsic or apoptotic-unrelated roles of caspases. Morevoer, various examples of caspaseindependent neuronal death after cerebral ischemia have been reported <sup>216, 217, 218, 219</sup>. In thic context, it is important to note that apoptosis is dynamically regulated during lifespan in the brain <sup>24</sup>. Indeed, while immature brain cells express high levels of many BCL2 proteins <sup>133, 220, 221</sup>, most of these proteins are downregulated in the adult brain, when most post-mitotic neural cells become resistant to apoptosis <sup>131</sup>, <sup>222</sup>. This may help explain the divergent findings on the mechanisms of neural cell death reported above. **Cardiovascular conditions.** While a role for RCD in non-reperfused myocardial infarction remains

questionable, apoptosis and other cell death programs including necroptosis, MPT-driven necrosis,

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ferroptosis, pyroptosis and autosis appear to contribute to cardiomyocyte death and tissue damage during myocardial infarction with reperfusion (also referred to as myocardial ischemia-reperfusion injury). However, the relative importance of the RCD and how they interconnect mechanistically and functionally to produce an integrated response remains poorly understood. For example, Bak1<sup>-/-</sup>mice with a cardiomyocyte-specific deletion of Bax displayed considerably reduced infarct size as compared to their wild-type littermates when subjected to experimental myocardial ischemia-reperfusion, although it remains unclear whether these effects are attributable to reductions in apoptosis or MPT-driven necrosis <sup>223, 224, 225</sup>, a RCD variant shown to participate in the pathogenesis of ischemic stroke <sup>226</sup>. Protection against myocardial ischemia-reperfusion has also been reported in transgenic mice overexpressing BCL2 <sup>227, 228, 229</sup> or a BCL-X<sub>L</sub>-derived peptide <sup>230</sup>. Likewise, deletion of *Bbc3* (leading to lack of PUMA) ameliorates myocardial ischemia-reperfusion injury <sup>231</sup>, ultimately translating into increased survival <sup>232</sup>. Moreover, neurotrophin-3 was reported to confer cardioprotection from schemic and reperfusion injuries by reducing BIM levels <sup>233</sup>. Broad spectrum caspase inhibition <sup>234, 235, 236</sup> and XIAP mimicking peptides <sup>237</sup> were shown to modestly reduce myocardial infarct size. Finally, simultaneous deletion of *Casp3* and Casp7 had no cardioprotective effect during reperfused myocardial infarction <sup>238</sup>, in line with the notion that the absence of caspase only delays cell death. In contrast to the large burst of cell death over several hours characterizing myocardial infarction, cardiomyocytes are lost gradually over months to years during heart failure with reduced ejection fraction <sup>3</sup>. The role of intrinsic apoptosis in these heart conditions is, however, debated. In a mouse model of cardiomyopathy based on the deletion of desmin (Des), the cardiomyocyte-specific over-expression of BCL2 reduces cardiac lesions and hypertrophy coupled to ameliorated cardiac functionality <sup>239</sup>. However, despite improved survival, these mice show increased levels of necrosis due to the activation of alternative cell death pathways <sup>240</sup>. Moreover, Casp3<sup>-/-</sup> mice display enhanced vulnerability to experimental cardiomyopathy, at least in part reflecting the inefficient activation of pro-survival AKT

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serine/threonine kinase 1 (AKT1) signaling <sup>241</sup>. As an alternative explanation, the absence of CASP3 may foster RCD-driven inflammation as a consequence of increased type I interferon (IFN) release <sup>242</sup>, <sup>243, 244</sup>. Indeed, experimental data linking dysregulated type I IFN release and cardiac conditions have recently emerged <sup>245</sup>.

As for therapeutic interventions, cardioprotective effects have been achieved by inhibition of CASP3 in rodent models of myocardial dysfunction induced by endotoxin <sup>246</sup>, burn injury <sup>247</sup> or hypoxia <sup>248</sup>, although perhaps such effects can be attributed to the lack of cell-extrinsic or apoptosis-unrelated effects of caspase activity. Moreover, inhibition of BAX prevents cardiotoxicity induced by doxorubicin in zebrafish and mice without affecting the anti-neoplastic activity of doxorubicin <sup>249</sup>. Similarly, the endothelial cell-specific expression of B cell leukemia/lymphoma 2 related protein A1a (BCL2A1A) promotes survival in a model of allogeneic heart transplantation <sup>250</sup>.

Finally, the mechanistic links between intrinsic apoptosis and atherosclerosis remain a matter of debate. Indeed, while *Casp3* deletion favors plaque development in mouse models of atherosclerosis <sup>251</sup>, the absence of DNA fragmentation factor subunit beta (DFFB, best known as CAD)) <sup>252</sup> protects mice against the disease. Likewise, while conditional deletion of *Mcl1* in myeloid cells is pro-atherogenic <sup>253</sup>, genetic or pharmacological inhibition of BCL-X<sub>L</sub> reduces atherosclerosis via a mechanism involving the depletion of platelets <sup>254</sup>. Moreover, the macrophage or leukocyte-specific deletion of the gene encoding BIM in mice has modest effects on plaque development, especially in the early phase of atherosclerosis <sup>255, 256</sup>. As the etiology of atherosclerosis involves a major inflammatory component, these apparently discrepant results may reflect (at least in part) the key role of some components of the apoptotic machinery in the control of inflammatory responses.

**Renal disorders.** Germline or kidney-specific deletion of *Bax* attenuates acute kidney damage in mice subjected to experimental renal ischemia/reperfusion <sup>257</sup>. A similar nephron-protection has been observed

in Bid<sup>-/-</sup> mice <sup>258</sup>, as well as in transgenic mice specifically expressing BCL-X<sub>L</sub> in the kidney <sup>259</sup>. Moreover, the simultaneous deletion of Bax and Bak1 in kidney proximal tubules limits tubular apoptosis and ameliorates kidney inflammation and fibrosis in a mouse model of renal fibrosis based on unilateral ureteral obstruction <sup>260, 261</sup>. Apoptotic caspases also appear to contribute to the etiology of renal conditions, although, perhaps, this reflects cell-extrinsic effects of caspase activity. Casp3 deletion reduces microvascular rarefaction and renal fibrosis in mice subjected to experimental ischemiareperfusion injury <sup>262</sup>, resulting in better long-term outcomes <sup>263</sup>. Moreover, the lack of CASP3 increases the survival of mice with chronic kidney disease caused by a congenital mutation in cystin 1 (CysI) <sup>264</sup>. In this setting, CASP3-deficient mice display increased CASP7 and decreased BCL2 expression, which is in line with recent clinical evidence of constitutive BCL2 down-regulation in patients with polycystic kidney disease 265. Administration of broad-spectrum caspase inhibitors limits kidney damage and improves renal functionality after a variety of experimental insults to kidneys, as observed in animal models of renal ischemia <sup>266, 267</sup>, polycystic kidney disease <sup>268</sup>, glomerulonephritis <sup>269</sup>, lupus nephritis <sup>270</sup> and diabetic renal disease <sup>271</sup>. Nonetheless, the specific targeting of apoptotic caspases will reveal whether this effect reflects the inhibition of intrinsic apoptosis. Indeed, these studies do not rule out the involvement of non-apoptotic RCD pathways in the etiology of acute and chronic kidney injury <sup>272, 273</sup>. Moreover, some of the nephron-protective effects of broad-spectrum caspase inhibitors have been linked to decreased post-RCD inflammation rather than the sole inhibition of apoptosis <sup>266, 274</sup>. In this context, Z-VAD-FMK aggravates (rather than ameliorates) renal dysfunction in a mouse model of cisplatin nephrotoxicity, by a mechanism involving the abrogation of cyto-protective autophagy <sup>275</sup>. Similarly, Z-VAD-FMK is ineffective in mouse models of osmotic nephrosis and contrast-induced acute kidney injury <sup>276</sup>, and this may be linked to the ability of Z-VAD-FMK to inhibit CASP8 (and hence promote necroptosis). Finally, acute loss of BCL-X<sub>L</sub> in all tissues of adult mice, except for hematopoietic cells,

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caused severe renal tubular degeneration leading to fatal anemia due to the loss of erythropoietin production <sup>277</sup>.

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**Hepatic diseases.** Abundant evidence highlights pathogenic roles of apoptosis in acute liver injuries, as well as in alcohol-related and alcohol-unrelated chronic liver disorders. Hepatocytes express high levels of BID, which connects DR signaling to mitochondrial outer membrane permeabilization (MOMP) upon CASP8-dependent cleavage <sup>278</sup>, and this complicates distinguishing between the intrinsic and extrinsic pathways. Here, we shall discuss studies performed on animal models of liver injury unrelated to overt signaling engaged by the Fas cell surface death receptor (FAS; also known as CD95 or APO-1) or the TNF receptor superfamily member 1A (TNFRSF1A, best known as TNF-R1) (which instead will be discussed in the next section). Distinct preclinical models of hepatic ischemia-reperfusion injury demonstrated that deletion of Bcl2111 (leading to lack of BIM) and/or Bid as well as over-expression of BCL2 or administration of pharmacological broad-spectrum caspase inhibition mediate robust hepatoprotective effects <sup>279, 280, 281,</sup> <sup>282</sup>. A similar improvement of hepatocyte survival and liver functionality was observed in rodents specifically expressing a mutated variant of BID in the liver and subjected to warm ischemia/reperfusion injury <sup>283</sup>. As for other models of liver injury, BIM-deficient mice are protected against viral hepatitis <sup>284</sup>. Moreover, deletion of the genes encoding BIM or PUMA, but not that of BCL2-related ovarian killer (Bok) limits liver injury in mice exposed to the hepatotoxic agent acetaminophen <sup>285, 286, 287</sup>. Moreover, pre-treatment with Z-VAD-FMK improves the survival of mice subjected to extensive hepatectomy <sup>288</sup>.

There is contrasting evidence on the role of BID in the etiology of liver conditions unrelated to overt FAS and TNF-R1 signaling. In a model of alcohol-related liver disease, the lack of BID confers some protection against ethanol-induced fibrosis, although mice display persisting signs of inflammation and steatosis <sup>289</sup>. Moreover, mice with a hepatocyte-specific deletion of *Bid* present reduced liver

inflammation and fibrosis when subjected to a choline-deficient diet to cause non-alcoholic steatohepatitis (NASH) <sup>290</sup>. Also, administration of BID-targeting antisense oligonucleotides exerted significant hepatoprotective effects <sup>291</sup>. However, BID deficiency fails to ameliorate liver injury and fibrosis upon bile duct ligation (as a model of obstructive cholestasis and chronic liver disease) <sup>292</sup>. Of note, in the same experimental model, the liver-specific overexpression of MCL1 but not BCL2 protects animals from hepatic damage 293, 294, suggesting some specificity for MCL1. To add a layer of complexity, conditional deletion of Xiap in hepatocytes does not result in liver injury, steatosis, or fibrosis, possibly due to compensatory effects of other inhibitor of apoptosis protein (IAPs) isoforms <sup>295</sup>. That said, Xiap<sup>-/-</sup> and Casp3<sup>-/-</sup> mice subjected to diet-induced hepatic steatosis and/or fibrosis, display exacerbated and attenuated liver damage, respectively <sup>296, 297</sup>. These effects have been linked to the modulation of the inflammatory response rather than apoptosis. Finally, genetic co-deletion of Mcl1 and transformation-related protein 53 (Trp53, best known as p53) <sup>298</sup> as well as conditional deletion of the genes encoding BCL-X<sub>L</sub> or MCL1 promote fibrosis and/or carcinogenesis, two common final stages of liver disease <sup>299</sup>. In this latter study, the additional deletion of *Bak1* limited hepatotoxicity, which is in line with evidence indicating that deletion of Bid and/or Bok protects mice against experimentally induced hepatocarcinogenesis <sup>300, 301, 302</sup>. CASP2 was found upregulated in mouse model of NASH and in NASH patients and was implicated in driving lipogenesis and steatohepatitis with a mechanism involving the cleavage of the site-1-protease (S1) followed by the activation of sterol regulatory element binding proteins (SREBP) <sup>303</sup>. In this study, the ablation or pharmacological inhibition of CASP2 prevented diet-induced steatosis and NASH progression. Of note, CASP2 deficiency was also reported to protect mice from diet-induced obesity and

metabolic syndrome <sup>304</sup>. Supporting the etiological contribution of caspase activation to liver disease, the

administration of broad-spectrum caspase inhibitors (e.g., emricasan, VX-166) reduced liver injury,

inflammation and fibrosis in mice fed a diet rich in fat or deficient in methionine and choline <sup>305, 306</sup>.

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Along similar lines, emricasan reportedly decreased portal pressure, fibrogenesis and hepatic inflammation, and preserved liver function in rodent models of chronic carbon tetrachloride (CCl<sub>4</sub>)-mediated cirrhosis or cholestasis driven by bile duct ligation <sup>307, 308, 309</sup>. Preliminary anti-inflammatory effects coupled with improved liver function have also been observed in patients with NASH-related cirrhosis treated with emricasan <sup>39, 310</sup>. However, follow-up, randomized clinical studies failed to observe beneficial effects of this agent on portal pressure and clinical outcome <sup>40, 41, 311</sup>. At least in part, these findings may reflect the complex interconnection between multiple RCD variants involved in the pathogenesis of NASH. Supporting this possibility, the administration of CASP3-specific inhibitors that abrogate both pro-apoptotic and pro-pyroptotic activities of CASP3 protected mice against acute liver injury caused by bile duct ligation <sup>312</sup>. Additional pharmacological and genetic studies specifically targeting intrinsic apoptosis (over other RCD pathways controlled by caspases) are needed to formally ascertain the involvement of this pathway in the etiology of hepatic disorders.

Hematological malignancies and solid cancers. The role of the intrinsic apoptosis pathway in preventing oncogenesis has been demonstrated in multiple animal models of induced hematological and solid tumors. In particular, a wide range of evidence demonstrates that over-expression of BCL2, BCL-X<sub>L</sub> or MCL-1 accelerates the onset of leukemia and lymphoma induced by over-expression of the MYC proto-oncogene, bHLH transcription factor (MYC) <sup>313, 314, 315, 316, 317</sup>. Accordingly, the pharmacological inhibition of anti-apoptotic BCL2 proteins is effective against MYC-driven tumors, even when they lack p53 function <sup>318, 319, 320, 321</sup>. In this context p53 has been shown to exert multiple roles in RCD (e.g., <sup>322, 323, 324</sup>). In particular, it acts as a direct or indirect regulator of the expression of several apoptotic genes <sup>325, 326, 327, 328</sup> and connects apoptosis induction and cell cycle arrest <sup>329</sup>. One main target of p53 in this context is cyclin dependent kinase inhibitor 1A (*CDKNIA*, best known as p21). p53-induced expression of *CDKNIA* leads to the activation of DREAM and RB/E2F transcriptional repressor complexes, in turn promoting cell cycle arrest by downregulating crucial cell cycle regulators such as cyclins and cyclin-

dependent kinases <sup>326, 327, 330</sup>. However, recent finding indicates that the p53-p21-DREAM or p53-p21-RB/E2F axis can also downregulate CASP2 and CASP8-associated protein 2/FLASH (CASP8AP2), generating a feedback loop centered on p53 that limits rather than promoting the induction of apoptosis <sup>326, 327</sup>. Of note, when analyzing the impact of endogenous proteins, it was shown that the absence of BCL-X<sub>L</sub> but not BCL2 limits the development of lymphoma in transgenic mice expressing MYC under the IgH enhancer (Eµ-myc mice) <sup>331, 332</sup>, thus supporting the therapeutic use of BCL-X<sub>L</sub> inhibitors against these blood cancers. Along similar lines, MCL1 overexpression <sup>317</sup> or *Mcl1* ablation <sup>318, 333, 334</sup>, respectively, accelerates and suppresses MYC-driven lymphomagenesis. Lending further support to the relevance of MCL1, prevalence and onset of MYC-driven lymphoma development were reduced by Mcl1 haploinsufficiency <sup>318, 334</sup>, or B cell-specific deletion of *Mcl1* <sup>335</sup>. Of note, loss of one allele of *Mcl1* (but not complete loss of the gene encoding BCL-X<sub>L</sub>) also impairs the development of thymic lymphoma in p53-deficient mice <sup>336</sup>, which possibly explains the limited effect of the BCL-X<sub>L</sub> + BCL2 + BCL-W inhibitor ABT-737 in these models of tumorigenesis <sup>337</sup>. The contribution of pro-survival BCL2 proteins in the development of AML has been demonstrated by using mice reconstituted with genetically modified bone marrow cells overexpressing MYC 338 and in human Burkitt lymphomas and diffuse large B-cell lymphomas (Diepstraten, 2020,31985804). Notably, the acute genetic removal of Mcl1 prevents the sustained survival and proliferation of AML driven by diverse oncogenic fusion proteins <sup>339</sup>. Accordingly, MCL-1 specific BH3 mimetic drugs, such as S63845, are able to potently kill a diverse range of lymphoid and myeloid malignant cells in culture and even in tumor transplanted mice <sup>340</sup>. Finally, ablation of Bcl2l2 (leading to lack of BCL-W) limits the development of MYC-mediated B cell lymphoma <sup>341</sup>. In support of the relevance of the intrinsic apoptosis pathway in tumorigenesis, several studies demonstrated that the development of MYC-driven lymphoma and leukemia is accelerated in mice lacking the genes encoding BAX <sup>342</sup>, BIM <sup>343, 344</sup>, BAD <sup>345</sup>, BMF <sup>345</sup> or PUMA <sup>346, 347, 348</sup>. In particular,

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these studies report that loss of only a single allele of Bcl2111 (encoding BIM) accelerates the development of lymphoma and this effect was reversed following full ablation of Bcl2l1 (leading to lack of BCL-X<sub>L</sub>) <sup>344</sup>. In this context, the presence of all prosurvival BCL2 proteins is shown to limit the impact of BIM in  $E\mu$ -Myc transgenic mice <sup>349</sup>. Instead, the combined ablation of the genes encoding BIM and p53 or PUMA and p53 accelerates MYC-driven lymphomagenesis <sup>350</sup>. This is in line with the evidence that loss of the genes encoding BAX or BIM augmented lymphomagenesis in p53-deficient mice <sup>351, 352</sup>. Of note, PUMA seems to exert a strong tumor-suppressive role in blood cancers, as shown by the evidence that Bbc3 deletion accelerates the development of MYC-driven B-cell lymphomas and that Eu-Myc lymphomas developing in PUMA-proficient mice display downregulated expression of PUMA 347, <sup>348, 353</sup>. On the contrary, the loss of the gene encoding NOXA does not accelerate MYC-driven lymphomagenesis, and the role of BIK in this murine lymphoma model is debated <sup>347, 354</sup>. Along similar lines, while CASP2 suppresses MYC-induced lymphomagenesis in mice 355, the tumor suppressive role of apoptosome components (**Box 1**) is questioned, as shown in lethally irradiated mice reconstituted with Eμ-Myc transgenic APAF1-deficient or CASP9-deficient fetal liver cells which showed no difference in the incidence of lymphoma compared to their wild-type counterparts <sup>356</sup>. This is consistent with the notion that APAF1 and caspase-9 function downstream of the commitment to cell death (MOMP) and therefore do not act as tumor suppressors <sup>15</sup>. Concerning other experimental animal models of induced hematological malignancies, the absence of PUMA (due to ablation of Bbc3) abrogated the development of both myelodysplasia, as shown in transgenic mice expressing a nucleoporin 98 (Nup98)-homeobox D13 (Hoxd13) fusion gene 357, and thymic T cell lymphoma induced by gamma radiation <sup>358, 359</sup>. The explanation for these surprising findings is based on the fact that the absence of PUMA prevents the extensive death of hematopoietic cells caused by gamma radiation, which causes mobilization and extensive proliferation of hematopoietic stem and progenitor cells, resulting in elevated replication stress and genetic instability and

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lymphomagenesis. These findings show that inhibition of apoptosis does not only promote the development of hematological malignancies, but in certain conditions can do the exact opposite and prevent lymphoma development. The absence of NOXA, augments the development of chronic lymphocytic leukemia in T cell lymphoma breakpoint 1 (TCL1) transgenic mice <sup>360</sup> and accelerated the development of thymic T lymphoma induced by gamma radiation <sup>358</sup>. Moreover, conditional deletion of Bcl2111 in B cells (leading to the absence of BIM) accelerates the development of mantle cell lymphoma in mice driven by cyclin D1 (CCND1) over-expression <sup>361</sup>. Over-expression of MCL1 and/or BCL2 promotes the development of acute myeloid leukemia driven by lysine (K)-specific methyltransferase 2A (KMT2A, best known as MLL) fusion proteins <sup>339, 362</sup> and plasmacytoma driven by ABL protooncogene 1, non-receptor tyrosine kinase (ABL1) <sup>363</sup>. Conversely, the loss of one *Mcl1* allele suppresses the development of T cell lymphoma, as shown in models based on sequential low-dose irradiation or the expression of a transgene encoding an IL2 inducible T cell kinase (ITK)-spleen tyrosine kinase (SYK) fusion protein <sup>364</sup>. Finally, the absence of CASP2 accelerates lymphomagenesis in ataxia telangiectasia mutated (ATM)-deficient mice <sup>365</sup>, but this may be due to the loss of the function of caspase-2 in mitotic cell division <sup>366</sup>. Lending support to the role of intrinsic apoptosis in hematologic malignancies, the BCL2 inhibitor venetoclax has entered clinical practice for the treatment of CLL as single agent or more effectively in combination with other therapeutic agents <sup>31, 35, 36, 37</sup>. Combinatorial regimens of BCL2 inhibition with epigenetic modulation have entered center stage in certain settings of AML 38, 367. However, mechanisms of resistance of CLL and AML to venetoclax related to defects in p53 and the apoptotic network or deregulated energy metabolism have been described 368, 369, 370, 371. Venetoclaxbased regimens also display effectiveness in patients with high-risk myelodysplastic syndromes <sup>372</sup>, thus suggesting a potential application in these syndromes <sup>373, 374</sup>.

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Significant work demonstrated a tumor suppressor role of the intrinsic apoptotic pathway in many cancers. For example, BCL2 overexpression accelerates the development of MYC-induced mammary

tumorigenesis <sup>375</sup>. A similar acceleration of tumor development has been described for the loss of genes encoding BAX, BIM, CASP2 or PUMA in distinct models of breast cancer induced by expression or overexpression of C3(1)/SV40 T-antigen, MYC, or erb-b2 receptor tyrosine kinase 2 (ERBB2, best known as HER2) 376, 377, 378, 379. At odds with these results, BCL2 overexpression in the mammary gland suppresses the development of breast tumors driven by the administration of dimethylbenz(a)anthracene <sup>380</sup>. This latter finding may be explained in a similar way as was mentioned for the suppression of radiation-induced thymic T cell lymphoma development by over-expression of BCL-2 or loss of PUMA (see above). Conditional deletion of the genes encoding BCL2 or BCL-X<sub>L</sub> in intestinal epithelial cells delays the development of colorectal cancer driven by inflammation <sup>381, 382</sup>, which is in line with the evidence that the absence of PUMA (due to Bbc3 deletion) exacerbates colorectal tumorigenesis as shown in a mouse model of intestinal oncogenesis driven by colitis or APC, WNT signaling pathway regulator (APC) <sup>383</sup>. Interestingly, doxorubicin-induced intestinal cytotoxicity requires PUMA but not BIM, whereas the reverse is true for MYC-driven apoptosis in the gut, indicative of differential roles for different BH3-only proteins in this tissue 384. Intriguingly, treatment with BCL-X<sub>L</sub>, but not BCL2targeting BH3 mimetics is sufficient to prevent intestinal tumorigenesis, suggesting that BCL-X<sub>L</sub> is the crucial mediator of protection of early neoplastic cells in this model <sup>385</sup>. In agreement, earlier work showed BCL-X<sub>L</sub> dependency in cell cultures derived from both colorectal and non-small cell lung cancers 386, 387. Moreover, a tumor suppressive effect is ascribed to BAX and CASP2 respectively in murine models of brain cancer 388, 389 and lung cancer 390 development. In line with this evidence, pharmacologic/genetic inhibition of MCL1 delayed tumor development in a mouse model of mutant KRas-driven adenoma/adenocarcinoma <sup>391</sup>. In the same model, tumor progression was promoted by the ablation of pro-apoptotic Bok <sup>392</sup>. Of note, there is evidence of a certain tissue-specificity in the epigenetic regulation of Bcl2 and Mcl1, such as the epigenetic mechanism centered on the deubiquitinase BRCA1 associated protein 1 (BAP1) 393 a tumor suppressor that is frequently mutated in certain cancers 394 and

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has been associated with tumor aggressiveness and therapy resistance <sup>395, 396</sup>. Finally, age-related differences in the expression of pro-apoptotic members of the BCL2 family have been linked to the increased sensitivity of neonatal/childhood tissues, relative to adult counterparts, to chemotherapy and radiotherapy. This was causally linked to MYC-dependent expression of genes encoding BAX, BID and BIM, both in mice and humans <sup>133</sup>.

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Cancer-specific contributions were attributed to particular BCL2 protein family members. For example, deletion of *Bax* accelerates the development of MYC-induced pancreatic tumors <sup>397</sup> which was not seen with ablation of Bak1 or Casp3 397, 398 but was achieved by BCL-X<sub>L</sub> overexpression 314, 399. Likewise, BOK seems to be crucial in hepatocarcinogenesis, as demonstrated in a mouse model of diethylnitrosamine-induced liver cancer which was accelerated on a *Bok*-/- genetic background <sup>300</sup>. Using the same mouse model, enhanced hepatic cancer development was also demonstrated for the deletion of the genes encoding PUMA or CASP2 400, 401. Conversely, overexpression of BCL2 was shown to limit transforming growth factor-a (TGFA)-driven hepatic tumorigenesis 402, 403, perhaps because the death of certain cells in the liver causes massive mobilization and proliferation of progenitor cells, leading to acquisition of oncogenic lesions that drive tumorigenesis in a manner similar to radiation-induced thymic lymphoma development (see above). Finally, the transgenic overexpression of BCL-X<sub>L</sub> (but not BCL2) and the keratinocyte-specific deletion of Bcl211 (leading to lack of BCL-X<sub>L</sub>) respectively accelerates or limits chemically- and/or ultraviolet B (UVB)-induced skin tumorigenesis 404, 405, 406, 407. It will be important to investigate and better understand why in certain settings inhibition of apoptotic cell death promotes tumorigenesis whereas it inhibits tumorigenesis in others.

**Autoimmune and inflammatory diseases.** There is substantial evidence linking intrinsic apoptosis to the development and progression of autoimmune diseases. However, the interpretation of these findings

should take into consideration the crosstalk between the apoptotic and inflammatory pathways and the fact that apoptotic caspases accelerate cell death and regulate its immunological manifestation.

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The first evidence that defects in the intrinsic apoptosis pathway can cause the development of autoimmune disease was reported when over-expression of BCL-2 in B lymphocytes 408 or loss of BIM in all tissues 409 was shown to cause fatal systemic lupus erythematosus (SLE)-like disease. Consistent with a critical role for the intrinsic apoptotic pathway in preventing autoimmune disease, the combined loss of the genes encoding BAX and BAK1 in hematopoietic cells, achieved by transplantation of lethally irradiated wild-type mice with hematopoietic stem/progenitor cells from the livers of E14.5 Bax<sup>-/-</sup>Bak1<sup>-/-</sup> embryos also causes fatal SLE-like disease 410. In mouse models of rheumatoid arthritis, ablation of the genes encoding BIM, BID or BAD, but not the loss of Bax and Bak1, accelerated the emergence and increased the duration and severity of this disease 411, 412, 413. Consistent with these findings, administration of a BIM mimetic suppressed inflammatory arthritis in mice 414. Mice deficient for BAX as well as transgenic mice expressing XIAP display increased severity of autoimmune encephalomyelitis induced by immunization with myelin oligodendrocyte glycoprotein (MOG) 415, 416. Similar results have been obtained in mouse models of autoimmune encephalomyelitis genetically engineered for hematopoietic cell-specific deletion of Bcl2111 (leading to BIM deficiency), or the neuron-specific overexpression of BCL2 417, 418. Consistent with the notion that inhibition of apoptosis can promote the development of auto-immune disease, inhibition of BCL-2, BCL-XL and BCL-W using the BH3 mimetic drug ABT-263 substantially reduced pathology in several mouse models of autoimmune disease, including scleroderma <sup>419</sup>. In apparent contrast with these results, studies using models of type 1 (autoimmune) or type 2 (non-autoimmune) diabetes revealed that deletion of *Bax* alone or combined loss of Bax and Bak1 420, 421, deletion of the gene encoding BIM, alone or together with the gene encoding PUMA 417, 422, 423, 424 or loss of BMF 425 protect pancreatic β cells from autoimmune destruction. Moreover, the absence of BIM prevented the emergence of type 1 diabetes in non-obese diabetic (NOD) mice  $^{417,422}$ , while ablation of Trp53 in pancreatic  $\beta$  cells failed to halt cell death in multiple experimental models of diabetes  $^{426}$ .

Based on the studies described above, inhibiting or deleting pro-apoptotic proteins or genes can have conflicting effects on autoimmune disease progression. This may depend on the cell type in which the major effect on apoptosis occurs, e.g., the immune cells (attacking the target cell) or the target cell. Inhibiting cell death in the target cells would provide protection and may improve disease outcome, whereas inhibiting cell death in the immune cell may lead to an accumulation of immune cells and aggravation of the autoimmune disease. The distinction could be explored by studying tissue-specific deletion of apoptosis regulator genes.

In this context, there is evidence that inflammatory and autoimmune disorders may derive from increased survival of specific immune cell population. For instance, elevated levels of cytokines, such as GM-CSF, IL-3 and IL-5 in immune disorders have been associated with prolonged survival of neutrophils, eosinophils or basophils with a mechanism involving the upregulation of anti-apoptotic proteins MCL1, BCL-X<sub>L</sub> and Baculoviral IAP Repeat Containing 2 (BIRC2, best known as cIAP2) promoted by <sup>427, 428, 429, 430, 431, 432, 433, 434</sup>. Apoptosis also plays a relevant role in certain hemopathies, including beta thalassemia <sup>435</sup>, Diamond-Blackfand anemia <sup>436</sup>, and in the Cohen syndrome neutropenia <sup>437</sup>. BIM, BID and BAD have all been shown to influence survival in mouse models of septic shock, as their targeting confer protective effects from tissue damage of multiple organs <sup>438, 439, 440</sup>, as well as in patients with severe sepsis <sup>441</sup>. On the contrary, the role of apoptotic caspase in septic shock is contentious <sup>54, 73, 442, 443</sup>. The precise impact of apoptosis in widespread inflammation during sepsis requires further investigation. Concerning other inflammatory disease, while broad-spectrum caspase inhibition reportedly protected

rats against severe acute pancreatitis 444, activation of intrinsic apoptosis appears to attenuate the severity

of this disease by limiting inflammation, as shown in vivo in a pancreatitis mouse model lacking XIAP

445. These data reinforce the notion that inhibiting (apoptotic) cell death may exacerbate unwarranted inflammatory reactions that contribute to the pathology of various autoimmune and inflammatory disorders. In line with this notion, chronic colitis driven by dextran sulfate sodium in mice manifests with increased (rather than decreased) severity in BID- or BIM-deficient hosts as compared to their wild-type littermates, at least in part owing to immune dysregulation 446, 447. Similarly, inhibition of BCL2 and/or BCL-X<sub>L</sub> reduces inflammation and ameliorates experimental colitis <sup>448, 449</sup>, an effect that was abrogated by concomitant deletion of the gene encoding BIM 449. PUMA-deficient mice displayed reduced levels of apoptosis amongst intestinal epithelial cells but not reduced inflammation in an experimental model of colitis <sup>450</sup>. Corroborating the specific relevance of PUMA for intestinal homeostasis, mice deficient for PUMA but not Bax-/-Bak1-/- mice were protected against the gastrointestinal side effects of radiotherapy, at least in part due to increased survival of intestinal stem/progenitor cells <sup>451, 452</sup>. Moreover, the absence of PUMA conferred protection to intestinal epithelial cells in mouse models of hypertensive gastropathy <sup>453</sup>, ulcerative colitis (UC) <sup>454</sup> and intestinal ischemia/reperfusion <sup>455</sup>. In the latter model, transgenic BCL2 expression limited intestinal epithelial cell death <sup>456</sup>. On the other hand, deficiency in XIAP, an inhibitor of CASP3, CASP7, CASP9, causes X-linked lymphoproliferative syndrome 2 with one-third of these patients suffering from severe and therapy-refractory inflammatory bowel disease <sup>457,</sup> <sup>458, 459, 460</sup>. Absence of XIAP also results in enhanced TNF production and TNF-R1/TNF-R2 targeting of TLR5-expressing Paneth cells and dendritic cells, leading to ileitis and dysbiosis <sup>461</sup>. In this context, it is interesting to note that CASP3- or CASP7-deficient mice display an altered gut microbiome 462, which may play a hitherto unexplored role in multiple autoimmune and inflammatory disorders beyond intestinal conditions. However, recently it was found that under steady state conditions absence of CASP3 and CASP7 in the intestinal epithelial cells apparently does not affect the microbiome neither cause spontaneous inflammation, suggesting that apoptosis may be dispensable for intestinal epithelium turnover and homeostasis at steady state <sup>463</sup>.

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Infectious diseases. Activation of RCD constitutes a protective mechanism against many microbial infections by eliminating infected cells and potentiating the anti-infective immune response. Accordingly, both viruses and bacteria have developed multiple strategies to overcome or disable host intrinsic apoptosis thus improving survival of both the host cells and the infectious organisms 464, 465 Waguia Kontchou, 2022 35397654 Waguia Kontchou, 2022 35397654 Waguia Kontchou, 2022 35397654 Waguia Kontchou, 2022 35397654. Mice with loss of one allele of the genes encoding BCL-X<sub>L</sub> displayed reduced pathology and had improved survival rates when challenged with Japanese encephalitis virus (JEV), as compared with wild-type mice. This was attributed to compromised viral propagation within JEV-infected cells succumbing to intrinsic apoptosis 466. There is also evidence of a contribution of BAX and BAK1 to the response to murine cytomegalovirus (MCMV) infection. In particular, the MCMV genome encodes inhibitors of BAK1 (m41.1 protein) and BAX (m38.5 protein), promoting viral replication by inhibiting the induction of intrinsic apoptosis in infected cells 467 Fleming, 2013, 23468630 Fleming, 2013, 23468630 Fleming, 2013, 23468630 Fleming, 2013, 23468630, 468 Manzur, 2009, 18949000 Manzur, 2009, 18949000 Manzur, 2009, 18949000 Manzur, 2009, 18949000. Supporting the requirement of the inhibition of intrinsic apoptosis for optimal in vivo MCMV dissemination, the titers of m41.1-deficient viruses were higher in salivary glands and other organs in  $Bak1^{-/-}$  mice as compared to wild-type animals  $^{467 \, \mathrm{Fleming}, \, 2013, \, 23468630}$ Fleming, 2013, 23468630 Fleming, 2013, 23468630 Fleming, 2013, 23468630. Intrinsic apoptosis also protects against bacterial infections, as demonstrated by the lethal course of disease in Bbc3<sup>-/-</sup> mice (which lack PUMA) after Streptococcus pneumoniae infection 469. Such an effect has been attributed to insufficient immunemediated bacterial clearance because of an increased neutrophil lifespan in the absence of PUMAmediated apoptosis. However, in certain other contexts, excessive activation of the intrinsic apoptosis pathway has been reported to drive, rather than prevent, microbial disease pathogenesis and lethality. For example, loss of Xiap increased the susceptibility of mice to Shigella infection, manifested with coalescing necrotic areas and a high bacterial burden in the liver and this was associated with an inefficient immune-mediated

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resolution of the bacterial infection <sup>470</sup>. Although at least part of this effect may be due to the requirement for XIAP to activate NOD signaling, rather than its ability to inhibit caspases 458, 470, 471. Moreover, mice lacking the genes encoding BIM and NOXA (i.e., Bcl2l11<sup>-/-</sup>Pmaip1<sup>-/-</sup> mice) displayed high resistance to the challenge with high doses of *Listeria monocytogenes*, as shown by a decreased bacterial burden and low apoptosis induction in the spleen <sup>472</sup>. The overexpression of BCL2 in the hematopoietic compartment increased the survival of mice infected with Ebola virus <sup>473</sup>, while deletion of *Bok* increased resistance of lung epithelial cells to apoptosis induced by SARS-CoV-2 virus membrane (M) protein <sup>474</sup>. Intriguingly, this study showed that the SARS-CoV-2 M protein, induced BOK to trigger apoptosis in the absence of BAX and BAK1 474. In another example, conditional deletion of Casp3 in the murine intestinal epithelium conferred protection from pathogenic Salmonella enterica, and this was attributed to a reduction in cell death-induced nutrients that are critical for sustaining bacterial growth <sup>475</sup>. Finally, Casp3<sup>-/-</sup> mice subjected to intracranial inoculation of reovirus type 3 (strain Dearing) displayed limited injuries in the central nervous system (CNS) and enhanced survival compared to wild-type mice <sup>476</sup>. As discussed above, the interpretation of the infection phenotypes using CASP3-, CASP7- and/or CASP9deficient mice needs particular caution because of the crucial roles of these caspases in modulating immune and inflammatory responses <sup>242, 243, 244</sup>. Notably, there is evidence for a role of specific regulators of apoptosis in the response to infection with human herpes simplex virus 1 (HSV-1). Thus, on the one hand, a significant accumulation in total leukocyte and CD8<sup>+</sup> T cells was observed in mice deficient for BIM and PUMA upon infection with HSV-1 477, which is in line with a role of these BH3-only proteins in controlling the survival of lymphoid and myeloid cells <sup>409, 478, 479</sup>. On the other hand, mice deficient for NOXA, BAD or BID were reported to mount a normal CD8<sup>+</sup> T cell immune response to HSV-1 infection <sup>477</sup>. Some of the contradictory results reported may arise from the divergent effects of inhibition or promotion of apoptosis on immune cells versus other cell types affected by the infectious disease, a distinction that cannot be addressed using mice in which apoptotic regulators have been deleted in the

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germline. In this context it is noteworthy that myeloid cell-specific deletion of the gene encoding BCL- $X_L$  or its inhibition using BH3 mimetic drugs massively reduced bacterial burden in the lung and extended the survival of mice infected with Legionella bacteria  $^{480}$ . This indicates that BH3 mimetic drugs might be effective for the treatment of intra-cellular bacterial infections.

Other diseases. Pro-apoptotic BCL2 proteins and caspases have also been implicated in disorders affecting other tissues/organs, such as skeletal muscle and lungs. For instance, the conditional ablation of *Bax* and *Bak1* protected mouse skeletal muscles against pressure-induced injury <sup>481</sup>. Similar results have been obtained in rats receiving Z-VAD-FMK after being subjected to muscular compression or blunt injury <sup>482, 483</sup>. Moreover, deletion of *Casp3* or CASP3 inhibition with Ac-DEVD-CHO limited muscular damage and atrophy in experimental models of plaster-mediated immobilization <sup>484, 485</sup>. In mouse models of catabolic disorders, muscle wasting due to protein degradation was decreased by lentiviral expression of XIAP <sup>486, 487</sup>, although whether this effect reflects the inhibition of intrinsic apoptosis needs further confirmation. Finally, *Casp3*-/- mice were protected against denervation-induced muscular atrophy <sup>488</sup>, while expression of a dominant-negative variant of CASP9 improved the neuromuscular activity in a transgenic mouse model of slow-channel syndrome <sup>489</sup>.

In a mouse model of oxidant-induced lung injury, the tissue-specific ablation of *Bax* and *Bak1* but not that of the genes encoding BID, BIM, NOXA or PUMA protected lung epithelial cells from degeneration <sup>490</sup>. Among the anti-apoptotic BCL2 proteins, BCL2A1 (best known as A1) seems to exert a crucial role in this setting, as *Bcl2a1* deletion aggravated lung injury in mice subjected to hyperoxia <sup>491</sup>, while lung-specific overexpression of BCL2 did not confer protection to mice exposed to excessive oxygen supply <sup>492</sup>. That said, no critical cytoprotective effect of A1 was seen in acute lung inflammation and peritonitis <sup>493</sup>. Intrinsic apoptosis has also been reported to be involved in pulmonary fibrosis <sup>494</sup>. *Bid*<sup>7-</sup> mice displayed decreased levels of pulmonary fibrosis after intra-tracheal bleomycin administration <sup>495</sup>. In

apparent contradiction, in the same model of fibrotic pulmonary damage, similar protection was reported in mice deleted for *Bcl2* <sup>496</sup> or in animals treated with inhibitors of BCL2 <sup>496</sup> or caspases <sup>497, 498</sup>. Along similar lines, ablation of *Bid* limited acute lung injury in mice induced by exposure to lipopolysaccharide <sup>499</sup>. Moreover, CASP3 depletion using short-hairpin RNAs (shRNAs) protected the lungs of mice subjected to pulmonary ischemia/reperfusion <sup>500</sup>, a protection further strengthened when necroptosis was concomitantly also suppressed <sup>501</sup>. BCL2 overexpression or caspase inhibition protected rodents subjected to lung transplantation <sup>502, 503</sup>. This is in line with the notion that delivery of the caspase inhibitor Z-VAD-FMK to rodents ameliorated lung injury developing as a consequence of severe acute pancreatitis or lipopolysaccharide administration <sup>504, 505</sup> but not as a result of pneumovirus infection <sup>506</sup>. In the latter case, lung damage was exacerbated by Z-VAD-FMK, perhaps due to increased inflammation downstream of necroptotic RCD <sup>506</sup>.

The studies briefly summarized above illustrate that components of the intrinsic apoptosis pathway can be part of the pathogenic mechanism of disease, and, in certain cases, this may offer the opportunity for therapeutic intervention. It is important to note though that in many pathogenic processes intrinsic apoptotic cell death is the endpoint, and simply inhibiting it will not be curative. If the cells continue being exposed to the initiating insult, they will likely undergo less regulated forms of cell death. However, inhibiting the intrinsic apoptotic cell death may buy time to remediate the factors that are damaging the cells in first place. Ischemia and hypoxia, in cases where the ensuing cell death has a substantial intrinsic apoptotic component, are examples. If cells in the ischemic region were kept alive until adequate circulation was restored, therapeutic benefits might be achieved. Other examples include metabolic disorders, which may be amenable to correction, and traumatic injury, where healing might be supported by inhibiting apoptosis. It would be worth concentrating on inhibiting intrinsic apoptotic cell death in conditions where the initiating tissue insults can be (at least partially) reversed. In contrast, failure to undergo intrinsic apoptosis is the initial pathogenic step or a contributing factor in certain

malignancies. Here, the induction of apoptosis, for example by using BH3 mimetic drugs <sup>33, 34</sup>, targets the pathogenesis directly.

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## Extrinsic apoptosis in disease

The molecular apparatus for extrinsic apoptosis is described in **Box 5** and illustrated in **Figure 4.** Unlike the intrinsic apoptotic pathway, DR-induced apoptosis is not required for embryonic or fetal development but plays a critical role in adult tissue homeostasis, as detailed in **Box 6** and **Box 7**. Of note, various components of the extrinsic pathway of apoptosis are involved in the etiology of multiple human disorders, although (1) with a considerable degree of context-dependency, and (2) with an effect not necessarily linked to the activation of apoptosis but often due to the role of DR signaling in necroptosis and inflammation, as outlined below. Neurological diseases. Although numerous studies investigated FAS and TNF-R1 signaling in the pathogenesis of multiple neurological diseases, the precise role of extrinsic apoptosis remains unclear (Figure 5). Loss-of-function mutations of FAS ligand (Fasl) as well as Fas silencing prevented motoneuron loss in mouse models of ALS driven by defect in superoxide dismutase 1, soluble (SOD1) 507, 508. Moreover, the lack of TNF did not affect motor neuron loss and mouse survival in this model <sup>509</sup>, while binding of the TNF receptor superfamily member 1B (TNFRSF1B, best known as TNF-R2) appeared to mediate neuroprotective effects <sup>510</sup>. As an additional layer of complexity, TNF mediates neuroprotective functions in wobbler mice - another mouse model of ALS that carries a point mutation in VPS54 GARP complex subunit (Vps54), at least in part by promoting the upregulation of ADAM metallopeptidase domain 8 (ADAM8) <sup>511</sup>. CASP8 has not yet been implicated in the pathogenesis of ALS, and nonapoptotic forms of FAS-driven RCD may play a predominant role in this context. For example, FAS

stimulation reportedly triggered the demise of motoneurons in mouse models of ALS by aggravating 1118 endoplasmic reticulum stress <sup>512</sup>. Similarly, cleavage of BID by CASP1 (and not CASP8) appears to 1119 contribute to neurodegeneration in transgenic mice expressing a mutant form of human SOD1 513. 1120 1121 However, the precise contributions of endoplasmic reticulum stress and CASP1 in ALS and other motoneuron disorders remain to be elucidated. 1122 The ability of TNF-R1 signaling to influence neurodegenerative conditions involves not only the 1123 induction of extrinsic apoptosis but also the activation of an inflammatory response. In distinct murine 1124 1125 models of AD, deletion of *Inf*, modification of its untranslated region (UTR) as well as pharmacological or genetic removal of TNF reduced plaque formation, resulting in attenuated neurological deficits 514, 515, 1126  $^{516,517,518,519,520,521}$ . Mechanistic studies in mice and monkeys revealed that TNF-R1 activation stimulates 1127 the protein activator of interferon-induced protein kinase EIF2AK2 (PRKRA) network <sup>522</sup>, which is 1128 linked to PD in humans <sup>523</sup>. Moreover, TNF-R1 signaling has been shown to favor microglial reactivity 1129 during neurodegeneration, culminating in neuronal loss <sup>524</sup>. Amelioration of disease was seen in mouse 1130 models of AD upon genetic or pharmacological inhibition of TNF-R1 525, 526. AD-associated 1131 neuroinflammation seems to depend on TNF-induced necroptosis rather than extrinsic apoptosis <sup>527, 528</sup>. 1132 1133 Unexpectedly, AD pathogenesis was shown to be enhanced in mice bearing a co-deletion of the TNF receptor superfamily member genes *Tnfrsf1a* and *Tnfrsf1b* 529, a phenotype that appears to impinge on a 1134 complex network of mutual interactions between TNF-R1 and TNF-R2 signaling <sup>530</sup>. Such a network 1135 1136 may also contribute to PD pathogenesis. Genetic ablation of *Tnf* or *Tnfrsf1a* plus *Tnfrsf1b* (leading to the lack of both TNF receptors), as well as pharmacological inhibition of TNF, were reported to protect 1137 dopaminergic neurons in murine models of PD following the administration of 1-metil 4-phenyl 1,2,3,6-1138 tetraidro-piridina (MPTP) or 6-hydroxydopamine 531, 532, 533, 534. Notably, in the above-mentioned 1139 experimental settings, TNF is thought to induce neuronal death in vivo by promoting microglia reactivity 1140

<sup>535</sup> with a complex interaction between TNF-R1 and TNF-R2 signaling <sup>536</sup>. Importantly, clinical evidence 1141 1142 from AD patients subjected to perispinal administration of the TNF blockers infliximab or etanercept suggests that the inhibition of TNF can ameliorate AD 537,538. In contrast, a dominant-negative variant of 1143 TNF failed to protect mice against neuronal degeneration in a model of HD <sup>539</sup>, suggesting that this 1144 approach may not be viable in patients with HD. 1145 TRAIL/TRAIL-R signaling has also been implicated in the onset and progression of AD 540, 541. 1146 Specifically, in a mouse model of AD, neutralization of TNF superfamily member 10 (TNFSF10, best 1147 1148 known as TRAIL) with a monoclonal antibody resulted in decreased neuroinflammation and a reduction in cognitive defects <sup>540</sup>. However, these findings were not extensively validated. Similarly, the impact of 1149 FASL-FAS signaling on neurodegenerative conditions is debated. Indeed, lymphoproliferative (lpr/lpr) 1150 mice, which lack FAS <sup>542</sup> and to a lesser extent generalized lymphoproliferative disease (gld/gld) mice. 1151 which lack FASL 542, are particularly susceptible to neuronal degeneration driven by MPTP 543. However, 1152 contrasting results have been obtained in another study of MPTP-treated mice with FAS deficiency 544, 1153 <sup>545</sup>. In this context, FAS-associated factor 1 (*Faf1*, a FAS binding protein that can initiate or enhance 1154 apoptosis) was found increased in midbrain in murine models of PD <sup>546</sup>. Moreover, a reduction in Faf1 1155 expression reduced MPTP-induced dopaminergic cell loss <sup>547</sup>. Such an apparent discrepancy in results 1156 may originate from the pleiotropic role of FAS in apoptosis and inflammation and other pro-1157 survival/regenerative signals. 1158 CASP8 activation has been detected in the brain of both AD 548 and HD 549 patients as well as in 1159 dopaminergic neurons of MPTP-treated mice and PD patients, a setting in which BID cleavage has also 1160 been documented <sup>119</sup>. This is in line with the ability of the broad-spectrum caspase inhibitor Q-VD-OPH 1161 to inhibit BID cleavage and mediate neuroprotection in MPTP-treated mice and rats <sup>550</sup>. Of note, CASP8 1162 was also reported to promote microglia reactivity potentially leading to neuronal loss <sup>551, 552, 553</sup>. In this 1163

context, genetic loss or pharmacological inhibition of CASP8 attenuated neurotoxicity by reducing microglial reactivity, thus extending survival of neurons, at least in part by stimulating the necroptotic death of activated microglial cells <sup>551, 552, 553</sup>. Consistent with this notion, *Casp8* deletion in myeloid cells protected mice from MPTP-mediated neurotoxicity <sup>554</sup>, suggesting that CASP8 inhibitors may be harnessed for the treatment of neurodegenerative conditions. Corroborating this idea, a pharmacological inhibitor of TNF-R1-associated death domain protein (TRADD) protected mice from disease in a model of AD-like proteinopathy driven by mutant tau <sup>555</sup>. However, pharmacological inhibition of CASP8 only partially prevented neuronal alterations in other models of AD <sup>108</sup> and even exacerbated dopaminergic neuronal necrosis in mice developing PD upon MPTP administration <sup>556</sup>. Moreover, rare CASP8 loss-offunction variants have been associated with AD in a large cohort of patients 557. Thus, the precise contribution of CASP8 signaling to neurodegenerative disorders and whether this relates to its function in driving extrinsic apoptosis, inhibiting necroptosis or promoting inflammatory cytokine production remains to be formally defined. Concerning dependence receptors, Netrin 1 (NTN1) upregulation was shown to confer neuroprotection in murine models of PD, suggesting a potential role of dependence receptors in neurodegenerative disease <sup>558</sup>. DR signaling has also been shown to contribute to neuronal death and inflammation in preclinical models of CNS trauma. In a compression model of spinal cord injury, mice with loss of FAS (i.e., lpr/lpr mice) as well as mice treated with FASL blockers displayed reduced post-traumatic neuronal degeneration and inflammation coupled to considerable functional improvement <sup>559, 560, 561</sup>. This beneficial effect also involved reduced engagement of the intrinsic apoptosis pathway <sup>562</sup>. Myeloid cell-specific deletion of Fast promoted neuronal regeneration and functional recovery in mice subjected to spinal cord injury <sup>563</sup>. A similar functional improvement after spinal injury was observed in mice with conditional deletion of Tr in macrophages and neutrophils but not in microglia <sup>564</sup>. Moreover, neuroprotection and limited

neuroinflammation have been documented in FAS-deficient lpr/lpr mice subjected to traumatic brain

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injury <sup>565</sup> as well as in mice subjected to experimental spondylotic myelopathy and exposed to FASLneutralizing antibodies 566. Studies on mice with loss of Fas and Tnfrsfla revealed at least some redundancy between FAS and TNF-R1 signaling in the context of experimental brain trauma <sup>567, 568, 569,</sup> <sup>570, 571</sup>. Furthermore, TNF inhibition reduced damage in mice or rats experiencing spinal cord injury <sup>572</sup>, <sup>573, 574</sup>, and also reduced the appearance of signs of autonomic dysreflexia, a cardiovascular disease associated with high-level spinal cord injury <sup>572, 575</sup>. Interestingly, some of these studies point to a neuroprotective function for TNF-R2 <sup>567, 569, 571</sup>, which is in line with at least some results from models of ALS <sup>510, 530</sup>. Moreover, several studies question a purely detrimental effect of TNF signaling in these experimental settings 576, 577, 578, 579. In particular, TNF was reported to support, at least in part, regeneration and long-term functional recovery in mice exposed to traumatic brain injury 577, 578, 579. Conversely, TRAIL neutralization stands out as a promising strategy to promote neuronal regeneration and functional recovery based on mice with spinal cord injuries <sup>580, 581</sup>. In this context, injured neurons seem to undergo Fas-associated via death domain (FADD)- and CASP8-dependent RCD <sup>582</sup>. Accordingly, Casp8 deletion or transgenic expression of a FADD inhibitor (the glycoprotein P45) protected mice after spinal cord injury <sup>583, 584</sup>. Similarly, transgenic expression of a dominant negative mutant of FADD (FADD-DN) limited motoneuron loss in mice undergoing axotomy <sup>585</sup>. Components of the molecular apparatus for the extrinsic pathway are associated with disorders of the visual system, again in the context of both exacerbated cell death and inflammation. Thus, in mouse and rat models of optic nerve injury, deletion of *Tnfrsf1a* (encoding TNF-R1) or inhibition of CASP8 with Z-IETD-FMK inhibited the degeneration of retinal ganglion cells <sup>586, 587</sup>. Moreover, the absence of TNF-R1 (but not the absence of TNF-R2) attenuated neurodegeneration in a mouse model of retinal ischemia, despite neuronal survival not being improved <sup>588</sup>. Along similar lines, deletion of *Tnf* <sup>589</sup> as well as inhibition of FAS 590 or TNF 591, 592 protected mice against retinal ganglion cell death in a model of glaucoma. Similar neuroprotective effects were documented for the conditional deletion of Casp8 in

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astrocytes or intra-ocular Z-IETD-FMK administration <sup>593</sup>. In this context, the conditional inducible ablation of *Casp8* from endothelial cells reduced postnatal retinal angiogenesis and pathological neovascularization in a mouse model of oxygen-induced retinopathy <sup>594</sup> (note that ablation of *Casp8* in endothelial cells is embryonically lethal <sup>595</sup>; *see* **Box 7**). Moreover, CASP8 inhibition could prevent experimental neovascularization of the cornea <sup>596</sup>. Finally, TRAIL neutralization protected the retinal tissue from damage associated with AD in a mouse model <sup>597</sup>.

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Experimental models of ischemic stroke and hemorrhage revealed a role of DR signaling in the pathophysiology of brain damage. In models of focal ischemia induced by middle cerebral artery occlusion, lpr/lpr as well as gld/gld mice (deficient for FAS or FAS ligand, respectively) displayed decreased infarct size and neuroinflammation <sup>598, 599, 600</sup>. Robust neuroprotection was also observed in lpr/lpr mice subjected to neonatal hypoxia-ischemia 601, as well as in lpr/lpr and gld/gld mice subjected to hyperoxia 602. Accordingly, inhibition of FAS or FASL exerted neuroprotective effects in an experimental murine model of stroke <sup>603, 604</sup>. Likewise, TRAIL neutralization limited brain injury in rats and mice subjected to middle cerebral artery occlusion 600, 605 or transient ischemia-reperfusion 606. Moreover, despite some contention in this respect 607, 608, 609, 610, abrogation of TNF/TNF-R1 signaling by genetic or pharmacological means prevented brain injury in rodent models of intracerebral hemorrhage 611 and focal cerebral ischemia 612, 613, 614, 615, 616, 617, 618, 619, 620. Further corroborating a pathogenic role of DR signaling, transgene-driven expression of dominant-negative CASP8 mutant and of FADD-like apoptosis regulator (CFLAR; best known as c-FLIP) attenuated brain damage after middle cerebral artery occlusion 621, 622. This is in line with the ability of CASP8 to drive BID activation upon focal cerebral ischemia <sup>196</sup>, as well as with the neuroprotective effects afforded by pharmacological CASP8 inhibitors seen in mice experiencing subarachnoid hemorrhage 623 or mice and rats subjected to focal cerebral ischemia <sup>624, 625</sup>. Importantly, FADD and CASP8 expression and/or activation have also been associated with ischemic stroke in humans 626, 627.

Perhaps surprisingly, TNF appears to protect mice against experimental seizures, not only through the engagement of TNF-R2 but also through TNF-R1 signaling <sup>610, 628, 629, 630, 631, 632, 633</sup> and consequent modulation of NF-κB <sup>634, 635</sup>. Conversely, *lpr/lpr* mice <sup>636</sup>, mice with neuron-specific deletion of the gene encoding TNF-R1 <sup>637</sup> as well as mice and rats treated with Z-IETD-FMK <sup>584, 638, 639</sup> displayed a reduced sensitivity to experimental seizures, pointing to a detrimental role for apoptotic DR signaling in this condition. Precise mechanisms through which TNF-R1 signaling promotes anti-apoptotic and anti-inflammatory effects in the context of excitotoxic insults remain unclear.

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Cardiovascular disorders. Data from preclinical models of ischemic and non-ischemic conditions indicate the involvement of FASL, TRAIL and TNF in the onset and progression of myocardial infarction with reperfusion and other heart diseases. In particular, both *lpr/lpr* mice (lacking FAS), as well as hearts isolated from these animals, displayed reduced cardiomyocyte death and infarct area upon experimental ischemia-reperfusion <sup>640, 641</sup>. Nonetheless, no protection against ischemia-reperfusion was found in hearts from Fas<sup>-/-</sup> or Fasl<sup>-/-</sup> mice <sup>642</sup>. However, supporting the therapeutic potential of the inhibition of DR signaling for the management of myocardial infarction, FASL-neutralizing antibodies conferred cardioprotection, limited inflammation, and improved cardiac function in mice experiencing cardiac ischemia-reperfusion <sup>643, 644, 645</sup>. Likewise, TRAIL blockade protected monkeys, pigs, and rats against experimental infarction by increasing cardiomyocyte survival and reducing inflammation <sup>646</sup>. This is in line with the predictive value of the levels of TRAIL as a biomarker for heart failure in patients <sup>647, 648</sup>. Of note, TRAIL has also been reported to exert apoptosis-independent roles in cardiomyocyte growth and heart hypertrophy <sup>649</sup> as well as in angiogenesis and neovascularization upon experimental hindlimb ischemia <sup>650</sup>. Similar to neurological conditions, while TNF-R2 signaling appears to exert cardioprotective effects, the engagement of TNF-R1 drives cardiac hypertrophy, inflammation and cardiomyocyte loss <sup>651, 652, 653, 654, 655, 656, 657, 658</sup>. The opposite outcome of TNF-R1 vs TNF-R2 signaling has been invoked to explain the clinical failure of TNF blocking agents in patients with chronic heart

failure <sup>659</sup>, despite encouraging preliminary findings <sup>660, 661</sup>, as well as cardiotoxic effects associated with the use of TNF blockers in patients with rheumatoid arthritis <sup>662</sup>. Confirming the involvement of extrinsic apoptosis in cardiac diseases, cardiomyocyte-specific deletion of *Fadd* in mice improved cardiomyocyte survival and heart function after ischemia/reperfusion <sup>663</sup>. Accordingly, haploinsufficiency of the gene encoding c-FLIP increased infarct area and aggravated cardiac dysfunction in mice subjected to myocardial infarction, while the cardiomyocyte-specific overexpression of c-FLIP attenuated pathology <sup>664, 665</sup>. Cardioprotection has been observed in a mouse model of ischemia/reperfusion upon shRNA-mediated CASP8 depletion <sup>666</sup> or treatment with the CASP8 inhibitor Q-LETD-OPh <sup>667</sup>. Moreover, transplantation of *CASP8*<sup>-/-</sup> cells did not increase neovascularization in wild-type mice subjected to hindlimb ischemia <sup>668</sup>, in line with a crucial role of CASP8 in the maintenance of endothelia in healthy conditions <sup>595</sup> (*see Box 7*). That said, combined pharmacological inhibition of apoptosis and necroptosis exerted greater cardioprotection than monotherapy in myocardial ischemia-reperfusion injury <sup>669</sup>, suggesting the involvement of multiple RCD pathways in cardiovascular disorders.

FASL neutralization has been reported to improve cardiomyocyte survival and cardiac function in a model of cirrhotic cardiomyopathy <sup>670</sup>. Conversely, a cardioprotective effect of TRAIL and TNF was observed in mice developing cardiomyopathy upon the deletion of apolipoprotein E (*ApoE*) <sup>671</sup> or desmin (*Des*) <sup>672</sup>, respectively. Both FASL deficiency and administration of CASP8 inhibitors decrease tissue inflammation and aneurysm formation in mice subjected to CaCl<sub>2</sub>-induced abdominal aortic aneurysms <sup>673</sup>. A potential role of extrinsic apoptosis in gradual cardiomyocyte attrition during heart failure with reduced fraction was also reported in a transgenic mouse model of inducible CASP8 overexpression <sup>674</sup>. Concerning TNF receptors, deletion of *Tnfrsf1b* resulted in increased cardiomyocyte death and hypertrophy induced by isoproterenol <sup>675</sup>. In contrast, deletion of *Tnfrsf1a* (but not *Tnfrsf1b*) was shown to be cardioprotective in murine models of vascular thrombosis <sup>676</sup>, and heart failure based on angiotensin II administration <sup>677</sup>. Similar cardioprotection to angiotensin II was reported after silencing of *Tnfrsf1a* 

 $^{678}$ . In line with these findings,  $Cflar^{+/-}$  mice (which lack c-FLIP) displayed increased sensitivity to cardiac injury upon angiotensin II administration  $^{679}$ .

FASL and TNF have also been reported to promote cardiac maladaptation and hypertrophy in models of pressure overload <sup>680, 681, 682, 683, 684</sup>. Consistent with this notion, TNF inhibition <sup>685</sup> or transgenic c-FLIP overexpression <sup>686</sup> limited experimental heart hypertrophy driven by hypertension. Moreover, treatment with etanercept reduced cardiac fibrosis in a diet-induced mouse model of obesity <sup>687</sup>. Conversely, both FAS and TNF receptor superfamily member 10b (TNFRSF10B, best known as TRAIL-R2 or mTRAIL-R) were reported to protect mice against atherosclerosis, at least in part, by modulating TNF superfamily member 11 (TNFSF11, best known as RANKL) signaling <sup>688, 689, 690, 691, 692</sup>, while the impact of TNF on experimental atherosclerosis remains a matter of debate <sup>693, 694, 695, 696</sup>. Finally, pharmacological inhibition of TNF prevented cardiotoxicity induced by doxorubicin in mice <sup>697, 698, 699</sup>

Renal conditions. FASL, TNF and TRAIL have all been implicated in the development of acute kidney injury by driving the activation of both extrinsic apoptosis and inflammation. Loss-of-function mutations in *Fasl*, inhibition or depletion of FASL <sup>700, 701, 702</sup> as well as *Fas* <sup>703</sup> or *Tnf* <sup>704</sup> silencing, TNF neutralization <sup>705,706</sup>, or TRAIL blockade <sup>707</sup> exerted nephron-protective effects in mouse models of renal ischemia/reperfusion. Generation of chimeric mice reconstituted with spleen cells from *gld/gld* mice (lacking FAS ligand) revealed a particular impact of FASL signaling in the hematopoietic compartment on ischemic acute kidney injury <sup>701</sup>. However, some functional overlap between DRs has also been reported. Indeed, while one study suggested that FASL neutralization was more effective than *Tnfrsf1a* deletion (leading to lack of TNF-R1) in preventing renal inflammation and cell death after acute kidney injury <sup>700</sup>, another study reported that the neutralization of TNF but not FASL prevented tubular apoptosis and renal atrophy upon ischemia/reperfusion injury <sup>705</sup>.

TRAIL blockade reportedly protected mice against renal damage after full-thickness scald burn, burn of all layers of the skin including epidermis and dermis <sup>708</sup>, while TNF inhibition limited nephrotoxicity, in mice treated with cisplatin <sup>709</sup>, and acute tubulointerstitial nephritis, in cancer patients administered with immune checkpoint inhibitors <sup>710</sup>. TNF neutralization also reduced tubulointerstitial fibrosis and renal injury in a mouse model of unilateral urethral obstruction <sup>711, 712</sup>. Contesting these findings, *Tnf*<sup>-/-</sup> mice showed increased fibrosis at later stages of ureteral obstruction <sup>713</sup>. This apparent discrepancy may reflect the distinct contribution of TNF-R1 and TNF-R2 signaling to different stages of renal fibrosis driven by urethral obstruction <sup>714</sup>. Conversely, experiments with *lpr/lpr* mice subjected to unilateral urethral ligation demonstrated a limited impact of FAS signaling to pathology <sup>715</sup>. The involvement of CASP8 in acute kidney injury is debated. While Casp8 and Casp3 protected kidneys against damage induced by renal ischemia, increasing the survival of these mice 703, 716, such a nephroprotective effect was not observed after treatment with the broad-spectrum caspase inhibitor Z-VAD-FMK 717, potentially due to caspase inhibition promoting necroptosis after DR stimulation. In line with this notion, chemical inhibitors of receptor-interacting serine/threonine kinase 1 (RIPK1) as well as deletion of Ripk3 exerted robust nephroprotection in mouse models of ischemia/reperfusion <sup>717, 718</sup>. However, combined deletion of Casp8 and Ripk3 did not extend the beneficial effects of the genetic loss of Ripk3 and was associated with a more pronounced demise of tubular epithelial cells by intrinsic apoptosis <sup>719</sup>. DR activation has also been associated with chronic kidney disorders, but evidence involving CASP8mediated apoptotic death is lacking. The conditional deletion of Tnf from macrophages  $^{720}$ , as well as the administration of TNF inhibitors 720, 721, 722, 723, were reported to mediate beneficial effects in murine models of diabetic nephropathy. Conversely, the impact of TRAIL on this renal condition remains unclear <sup>724, 725, 726</sup>, like that of TNF on polycystic kidney disease <sup>727, 728</sup>. As for glomerular inflammation,

gld/gld mice (lacking FAS ligand), as well as wild-type mice treated with TNF blockers, displayed

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reduced tissue damage during crescentic glomerulonephritis <sup>729, 730, 731, 732</sup>. Indeed, balanced TNF-R1 and TNF-R2 signaling appeared to be critical for mice to resist experimentally induced glomerulonephritis <sup>733, 734, 735, 736, 737, 738</sup>. This may explain apparently discrepant findings obtained with TNF-targeting measures.

**Hepatic disorders.** TNF-deficient mice, as well as rodents treated with TNF inhibitors, presented with attenuated liver injury and apoptosis upon experimental ischemia/reperfusion, resulting in improved survival <sup>739, 740, 741</sup>. Of note, this beneficial effect could not always be recapitulated in *lpr/lpr* and *gld/gld* mice, lacking FAS or FAS ligand, respectively <sup>741</sup>. Similarly, FAS inhibition, FASL neutralization, as well as administration of low-dose TNF (as a pre-conditioning maneuver) have been shown to protect the liver against ischemia/reperfusion injury by reducing hepatic cell apoptosis and/or inflammation <sup>742, 743, 744</sup>. Protection of the liver from ischemia/reperfusion has also been observed in mice deficient for TRAIL <sup>745</sup>, as well as upon the conditional knockdown of CASP8 or CASP3, the combined deletion of *Casp8* and *Ripk3*, and the transgenic expression of a BID mutant that cannot be cleaved by CASP8 <sup>283, 746, 747</sup>

*Lpr/lpr* mice lacking FAS <sup>748</sup>, *Tnfsf10*<sup>-/-</sup> mice (which lack TRAIL) <sup>286</sup>, as well as animals exposed to TRAIL blockers <sup>749</sup>, were protected against acetaminophen-induced liver damage, in line with the notion that FAS signaling and TRAIL receptor exacerbate acetaminophen hepatotoxicity <sup>750</sup>. Along similar lines, the hepatocyte-specific deletion of the gene encoding c-FLIP enhances liver injury and fibrosis induced by treatment with CCl<sub>4</sub> or thioacetamide <sup>751</sup>. Moreover, a large body of evidence demonstrates that the abrogation of extrinsic apoptosis protects mice against fulminant hepatitis and hemorrhage in the liver induced by FASL and TNF. This has been achieved with strategies including (but not limited to) FADD blockade <sup>752, 753</sup>, *Casp8* <sup>595, 754, 755</sup> or *Fadd* <sup>756</sup> ablation, and *Casp8* silencing <sup>757</sup>. Accordingly, hepatocyte-specific deletion of *Cflar* augmented liver damage in mouse model of acute hepatic injury

<sup>758</sup>. Consistent with the notion that engagement of the intrinsic apoptotic pathway is critical for DR induced cell killing in the liver, Bid<sup>-/-</sup> mice resist fatal hepatitis and hepatocytes apoptosis induced by FAS or TNF <sup>278, 282, 759, 760</sup>, a protection enhanced by concomitant loss of BIM or CASP8 <sup>282</sup>. Conditional deletion of the genes encoding BAX, BAK1 or PUMA, as well as overexpression of BCL2, can also protect hepatocytes from FAS-induced killing 761,762,763,764. The impact of loss of BAD on TNF-induced hepatitis is controversial <sup>765, 766</sup>. Mice deficient for CASP3 or treated with CASP3 or CASP8 inhibitors have also been shown to be less sensitive to FAS-induced hepatocyte apoptosis <sup>767, 768</sup>. Of note, some degree of functional compensation between caspases and alternative mechanisms of caspase activation have emerged from studies in hepatocytes responding to FAS agonists <sup>769</sup>. Finally, FAS and TNF-R1 signaling, as well as FADD activation, are involved in liver regeneration following partial hepatectomy 770, 771, 772, 773, 774. In this context, liver-specific deletion of *Casp8* resulted in dysregulated hepatocyte proliferation upon partial hepatectomy coupled to the initiation of an inflammatory response <sup>775</sup>. It has been suggested that CASP8 modulates liver regeneration by balancing NF-κB activation and necroptosis rather than by inducing apoptosis <sup>776</sup>. Gld/gld mice (lacking FAS ligand) chronically fed with ethanol displayed reduced liver injury, steatosis and inflammation as compared to wild-type mice, but exhibited signs of incipient fibrosis 777. Some degree of protection against alcohol-induced liver damage has also been documented in mice deficient for the apoptosis-inducing TRAIL receptor mTRAIL-R <sup>778</sup> or TNF-R1 (but not TNF-R2) <sup>779</sup>, as well as in mice receiving a TRAIL-neutralizing antibody <sup>780</sup>. Accordingly, the hepatocyte-specific ablation of Casp8 limited hepatic steatosis in murine models of ethanol administration, although it failed to prevent apoptotic RCD <sup>781</sup>. Conversely, apoptosis driven in hepatocytes by chronic ethanol exposure could be abolished by systemic inhibition of CASP3 with Ac-DEVD-FMK <sup>782</sup>.

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The liver-restricted overexpression of FAS induces hepatic steatosis and insulin resistance in mice subjected to a high-fat diet (HFD) <sup>783</sup>. In the same experimental setting, hepato-protection was observed with the hepatocyte-specific ablation of *Fas* or germline deletion of *Bid* <sup>783</sup>. Moreover, *Tnf* deletion <sup>784</sup>. <sup>785</sup>, whole-body deletion of *Tnfrsf1a* (encoding TNF-R1) alone or in combination with the gene encoding TNF-R2 <sup>786, 787</sup> as well as inhibition of TNF <sup>788, 789, 790</sup> or TNF-R1 <sup>791</sup> significantly reduced hepatic steatosis, fibrosis, damage, and metabolic alterations in several diet-induced or genetic models of non-alcoholic fatty liver disease (NAFLD). In apparent contrast with these findings, the hepatocyte-specific deletion of *Tnfrsf1a* failed to protect mice from diet-driven NASH <sup>792</sup>. Moreover, *Tnfrsf1a* deletion accelerated progression of steatosis to steatohepatitis in mice on HFD <sup>793</sup>. Taken together, these findings underscore the pleiotropic and context-dependent effects of TNF/TNF-R signaling in NAFLD. The impact of TRAIL on NAFLD is also debated. Indeed, contrasting evidence from experiments with mice deficient for TRAIL or treated with recombinant TRAIL suggests either a detrimental or a beneficial role to TRAIL in NAFLD induced by HFD <sup>794, 795, 796</sup>.

The absence of mTRAIL-R promoted hepatic inflammation and fibrosis in a genetic mouse model of cholestasis <sup>797</sup>. Similarly, *lpr/lpr* mice lacking FAS <sup>798, 799, 800</sup> as well as TNF-deficient <sup>801, 802</sup> and TRAIL-deficient <sup>803, 804</sup> mice displayed reduced hepatocyte apoptosis and fibrogenesis after experimental cholestasis induced by bile duct ligation. In line with these results, expression of a phosphorylated FADD mimicking mutant resulted in attenuated HFD-induced hepatomegaly and steatosis <sup>805</sup>. Experiments based on the hepatocyte-specific deletion of *Cflar* (encoding c-FLIP) or transgenic overexpression of c-FLIP revealed a role for this modulator of CASP8 activation as a suppressor of hepatic steatosis and inflammation induced by HFD <sup>806</sup>. Moreover, the hepatocyte-specific deletion of *Cflar* in mice resulted in enhanced cholestatic liver injury and inflammatory responses upon bile duct ligation <sup>807</sup>. Moreover, the hepatocyte-specific deletion of *Casp8* protected mice against liver injury in models of cholestatic hepatitis caused by the administration of 3.5-diethoxycarbonyl-1,4-dihydrocollidine <sup>808</sup>, as well as in

models of steatosis caused by the feeding of a methionine- and choline-deficient diet <sup>809</sup>. A similar hepato-protection against obstructive cholestasis has been documented in mice with hepatocyte-specific *Casp8* deletion <sup>810</sup>. Furthermore, liver parenchymal cell-specific ablation of the gene encoding FADD prevented RIPK1-dependent but not TNF-R1-, FAS-, and TRAIL-R-independent hepatocyte apoptosis, chronic liver inflammation and hepato-carcinogenesis in mice with liver-specific deficiency in Inhibitor Of Nuclear Factor Kappa B Kinase Regulatory Subunit Gamma (IKBKG, best known as NEMO or IKKgamma) <sup>811,812</sup>. Finally, decreased BID cleavage has been associated with attenuated liver injury in mouse models of chronic cholestasis <sup>813</sup>.

## Hematologic malignancies and solid cancers.

Human patients with autoimmune lymphoproliferative syndrome (ALPS) caused by defects in FAS are known to show abnormally increased predisposition to lymphoma development  $^{814}$ . Accordingly, FAS-deficient lpr/lpr mice develop plasmacytoma-like disease in advanced age  $^{815}$ . TRAIL also seems to exert a tumor suppressive function in lymphomagenesis. The ablation of the gene encoding mTRAIL-R accelerated the development of lymphoma in  $E\mu$ -Myc transgenic mice  $^{816}$ . Moreover, deficiency in TRAIL (but not in mTRAIL-R) promoted the development of lymphoma and other tumors in mice with haploinsufficiency for Trp53  $^{817}$ .  $^{818}$ . Interestingly, mice engineered to express exclusively either membrane-bound or secreted FasL showed an increased incidence of spontaneous tumor formation when expressing only soluble FasL which was unable to induce FAS-mediated apoptosis but could exert inflammatory effects  $^{819}$ .

The role of FAS and TRAIL-R in the development of colorectal cancer is controversial. For instance, the loss of FAS was reported to enhance APC mutation induced but not inflammation induced intestinal tumorigenesis <sup>820,821,822</sup>. Along similar lines, while the ablation of *Tnfrsf10b* (leading to lack of mTRAIL-R) in mice did not impact tumorigenesis induced by *Apc* mutations <sup>818</sup>, the administration of TRAIL

suppressed tumorigenesis in a mouse model of colitis-associated colon cancer 823. Despite some contention in this respect 824, 825, 826, 827, TNF seems to contribute to the development of colorectal cancer, although whether such effects depend on the apoptotic function of TNF needs further demonstration. The administration of TNF blockers 828, 829, 830, 831, 832 or ablation of Tnf 833 or Tnfrsfla 833, 834 limited tumor development, as shown in animal models of colorectal cancer induced by colitis, chemicals, or a mutation in Apc. Finally, loss of the dependence receptor DCC netrin 1 receptor (Dcc) accelerated cancer progression in a mouse model of Apc mutation driven colorectal oncogenesis 835. A tumor suppressor role in colorectal cancer is also described for the dependence neurotrophic tyrosine kinase, receptor, type 3 (Ntrk3, best known as TrkC) <sup>836</sup>. Of note, the association between gain of dependence receptors ligands (e.g., NTN1) with tumor progression 837, may make their targeting a promising anti-cancer approach 838 (https://clinicaltrials.gov). With regard to other tumor types, both TNF-R1 and FAS displayed a pro-oncogenic role in hepatic and ovarian oncogenesis. Thus, conditional deletion of Fas in hepatocytes delayed chemically-induced hepato-carcinogenesis, while Fas ablation suppressed the development of ovarian tumors in phosphatase and tensin homolog (PTEN)-deficient/Kirsten rat sarcoma viral oncogene (KRAS) mutated mice 839. Likewise, TNF neutralization limited the onset of hepatic cancer driven by experimentally induced cholestatic hepatitis 840. Consistent with these findings, Casp8-/- mice are protected against the development of inflammation-driven liver cancer <sup>754</sup>. Hyperactivation of CASP8 in the context of RIPK1 and TNF receptor-associated factor 2 (TRAF2) deficiency has been implicated in the development of hepatocellular carcinoma 841 although such effects may be independent of apoptosis induction 842, 843. In contrast, recent studies show a tumor-suppressive function of CASP8 in the liver and certain other tissues 844, 845, 846, 847. In particular, there is evidence of a role of CAPS8 in early tumorigenesis (but not tumor progression) exerted by modulating the DNA damage response 844 or the level of chromosomal instability (CIN) 845.

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Consistent with a pro-tumorigenic effect of TNF, the ablation of Tnf or Tnfrsf1a or the blockade of TNF in mice conferred some protection against chemically-induced skin cancer development <sup>848, 849, 850, 851, 852,</sup>  $^{853}$ . In contrast, the impact of genetic and pharmacological inhibition of TNF in UVB-induced skin cancer is debated 854, 855. Of note, the comparison between TNF-R1- vs. TNF-R2-deficient mice revealed a primary role of TNF-R1 in chemically induced skin oncogenesis <sup>850</sup>. Furthermore, TNF-R1 deficiency suppressed the development of skin cancer induced by NF-κB inhibition <sup>856</sup>. A similar role for TNF-R1 in supporting tumorigenesis was described in murine models of N-methyl-N-nitrosourea/testosteroneinduced prostate cancer 857 and methylcholanthrene (MCA)-induced fibrosarcoma 858. As opposed to TNF-R1, TNF-R2 shows tumor-suppressive functions in mouse models of tumorigenesis, such as the development of fibrosarcoma triggered by MCA 858 and of breast cancer induced by transgenic expression of wingless-type MMTV integration site family, member 1 (Wnt1) 859. Moreover, the absence of TNF impaired tumor growth in HER2-driven mammary tumorigenesis in mice 860 and TNF neutralization suppressed chemically-induced oral <sup>861</sup> and urethane-induced pulmonary tumorigenesis. Conversely, TNF overexpression in the airway epithelium enhanced mutant KRASdriven lung cancer development <sup>863</sup>. Pre-clinical evidence indicates some tumor type-specificity for the role of TRAIL and its receptor(s) in tumorigenesis. Transgenic expression of TRAIL in the skin delayed chemically induced carcinogenesis <sup>864</sup>. This effect was recapitulated in mice lacking TRADD <sup>865</sup> but, curiously, not in mTRAIL-R-deficient mice 866, with the latter actually showing enhanced lymph node metastasis. In support of an anti-tumor function for the TRAIL/TRAIL-R system, TRAIL-deficient mice as well as mice treated with TRAIL blockers displayed increased susceptibility to MCA-induced fibrosarcoma 867, 868. In a recent study the combined treatment with recombinant TRAIL and inhibition of cyclin-dependent kinase 9 (CDK9) was effective in a wide range of cancers <sup>869</sup>. Yet in contrast to this and in support of a tumor-supportive role

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of endogenous TRAIL, deficiency in mTRAIL-R limited tumor growth and improved survival in a mouse model of mutant KRAS-driven lung and pancreatic tumorigenesis <sup>870</sup>. Moreover, malignant cell-specific ablation of genes encoding mTRAIL-R or FADD reduced lung cancer growth and tumor-protective inflammation <sup>871</sup>, while systemic ablation of *Tnfsf10* (leading to lack of TRAIL) had no impact on HER-2 driven breast oncogenesis <sup>817</sup>. Interestingly, KRAS mutations have been shown to promote the switch of FAS and TRAIL receptors from a predominantly death-inducing into a metastasis promoting function <sup>872</sup>. Since TRAIL as well as FASL can trigger either apoptosis, necroptosis, inflammatory or pro-invasive signaling, cancer-specific preferences for one or the other of these signaling outputs likely accounts for the pleiotropic effects observed in various cancer models.

Autoimmune and inflammatory diseases. The interpretation of results on the impact of extrinsic apoptosis in the etiology of autoimmune and inflammatory disease should consider the fact that DR engagement can also result in the initiation of an inflammatory response not related to RCD (see Box 6 and **Box 7**), meaning that DR deregulation may lead to inflammatory diseases independently of the induction of extrinsic apoptosis. The notion that defects in DR signaling can cause autoimmune disease is supported by the observation that lpr/lpr as well as gld/gld mutant mice, deficient for FAS or FAS ligand, respectively, as well as humans with defects in FAS develop systemic lupus erythematosus (SLE)-like autoimmune disease accompanied by lymphadenopathy, splenomegaly and hepatomegaly <sup>873</sup>, <sup>874</sup>. A critical role for loss of caspase-CASP8 mediated apoptosis in this disease was demonstrated by the observation that similar autoimmune disease is seen in mice lacking CASP8 and also RIPK3 or MLKL (to prevent necroptosis) 51, 52, 875. However, the roles of DRs in autoimmune disease are complex. TRAIL/TRAIL-R signaling was reported to protect mice and rats against autoimmune encephalomyelitis 876, 877, 878, 879, 880, 881, autoimmune arthritis 882, 883, 884, 885, 886 and type I diabetes 689, 882, 887, 888, 889, 890. Perhaps surprisingly, the presence of FAS and TNF-R1 is associated with the development of certain autoimmune conditions. Indeed, both lpr/lpr lacking FAS and gld/gld mice lacking FAS ligand, as well as TNF-R1-

deficient mice, were reported to be protected against experimental encephalomyelitis 891, 892, 893, 894. Similar results were obtained in mice with *Tnf* deletion in monocytes and macrophages but not in mice lacking TNF in microglial cells <sup>895</sup>. Protection against experimentally induced autoimmune conditions were also found in mice subjected to neutralization of TNF or TNF-R1 inhibition 896, 897, 898, 899, 900, 901, 902, 903. FAS-independent mechanisms also appear to support the pathogenesis of experimental autoimmune encephalomyelitis 891,904, with some studies pointing to a protective role for FAS-induced RCD amongst lymphocytes in this disease model <sup>905</sup>. Moreover, FAS engagement was reported to differentially contribute to the initiation vs. the recovery from autoimmune encephalomyelitis 906, 907. In particular, FASL expression in astrocytes appears to promote recovery from experimental autoimmune encephalomyelitis, as shown by persisting demyelination and paralysis of mice with an astrocyte restricted deletion of the Fasl gene 906. Finally, at least in some studies, Tnf deletion or TNF neutralization failed to attenuate the severity of autoimmune encephalomyelitis once the disease was established <sup>908, 909</sup>. Mice with defects in FASL or TNF signaling are protected against arthritis induced by immunization with xenogeneic type II collagen in complete Freund's adjuvant <sup>910, 911, 912, 913</sup>. Similar protection was observed in mice transplanted with mesenchymal stem cells engineered to express TNF inhibitors <sup>914</sup>. In keeping with this evidence, the myeloid cell specific deletion of Fas or the administration of antibodies that target both TNF and chemokine (C-X-C motif) ligand 10 (CXCL10) resulted in accelerated disease resolution in a model of rheumatoid arthritis induced by K/BxN serum transfer 915, 916. Genetic loss of Fas or pharmacological inhibition of FAS conferred protection against autoimmune diabetes in certain animal models, including NOD mice 917, 918, 919, 920, 921, 922. However, whether the impact of FAS on the pathogenesis of autoimmune diabetes depends on its role in the death of pancreatic β-cell <sup>917</sup> or its role in inflammation (e.g., in the context of insulitis) remains a matter of debate <sup>920</sup>. Conversely, other studies found no role for FAS in diabetes 923, 924, 925. TNF neutralization is effective only in a limited sub-group of patients with inflammatory bowel disease 926, 927. This is in line with the finding that deletion of the

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gene encoding TNF-R1 exacerbated colitis in interleukin 10 (IL10)-deficient mice <sup>928</sup>. Similar protection was ascribed to TRAIL/TRAIL-R signaling in a dextran sodium sulfate-induced model of colitis model <sup>929, 930</sup>. Finally, it has been suggested that FASL and TNF signaling contribute to the pathogenesis of acute pancreatitis <sup>931, 932</sup>. A similar detrimental role has been proposed for TNF in autoimmune neuritis 933, 934, 935, although there is also some contention 936, as well as in spondylarthritis 937 and psoriasis 938. Conversely, mTRAIL-R appears to mediate beneficial effects in autoimmune thyroiditis <sup>939, 940, 941, 942, 943</sup> At least in part, these findings reflect the pleiotropic effects of whole-body/systemic inhibition of DRs signaling, which concomitantly impacts both the target (i.e., parenchymal) and the perpetrator (i.e., immune cells) of damage. Some experimental evidence links CASP8 activation to autoimmune and inflammatory disorders. In a recent study using a chemically-induced model of intestinal inflammation, the selective absence of CASP8 in intestinal epithelial cells decreased their survival, also resulting in gut barrier dysfunction and chronic inflammation <sup>944</sup>. Of note, in these settings, inflammation can occur via a mechanism independent of the induction of necroptosis (which is inhibited by CASP8) and involving the activation of RIPK1 and the RNA Sensor RIG-I pathway 945, 946. Along similar lines, chronic proliferative dermatitis in mice deficient for components of the linear ubiquitin chain assembly complex (LUBAC) was associated with an increased keratinocyte apoptosis mediated by the engagement of TN-FR1 and the activation of the RIPK1- and/or FADD-CASP8 cascade 947, 948, 949, 950, 951. Importantly in this mouse model of an inflammatory disease the relevant contributions of cell death versus inflammatory signalling from TNF-R1 were genetically dissected demonstrating that excess apoptosis/necroptosis drove different elements of the inflammatory response depending upon the affected tissue. Concerning autoimmunity, in a mouse model of autoimmune encephalomyelitis, the oligodendrocyte-specific deletion of Fadd reduced demyelination and this was accompanied by limited immune cell infiltration in the spinal cord 952. Likewise, experimental autoimmune encephalomyelitis could be prevented by transgenic expression of

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FADD-DN (dominant negative form of FADD) in T cells  $^{953}$  but it must be noted that this kills antigen receptor activated T cells  $^{954}$ . Therefore, this protective effect is due to the removal of the T cells that would cause tissue destruction. Activation of CASP8 was identified in the microglia of patients with multiple sclerosis  $^{955}$ . Moreover, transgenic expression of FADD-DN or *Casp8* ablation in pancreatic  $\beta$  cells protected mice from autoimmune diabetes  $^{956}$ . This indicates that the killing of these cells is mediated by death receptor induced apoptosis. BID appears to be dispensable for the development of diabetes in NOD mice  $^{957}$ .

There are also contrasting observations on the impact of DR-induced apoptosis on the development and resolution of autoimmune rheumatoid arthritis. The absence of c-FLIP (due to *Cflar* deletion) resulted in increased disease severity but limited disease resolution in mice experiencing arthritis upon intraperitoneal injection of serum from mice expressing both the T cell receptor transgene KRN and the MHC class II molecule A(g7) (K/BxN mice) <sup>958</sup>. In the same model, deletion of *Casp8* in all myeloid cells enhanced disease resolution, while deletion of *Casp8* selectively in dendritic cells accelerated disease onset <sup>959</sup>. Further experiments are required to unveil the reasons for such cell type specificity for the role of CASP8 to help more clearly understand the role of extrinsic apoptosis in this and other autoimmune disorders.

Infectious diseases. Extrinsic apoptosis is reported to act as an anti-infective mechanism. FAS deficient *lpr/lpr*, FAS ligand deficient *gld/gld* and *Bid*<sup>-/-</sup> mice exhibit delayed clearance of *Citrobacter rodentium* and increased intestinal pathology <sup>960</sup>. Confirming the importance of DR-induced apoptosis, this pathogen was shown to inhibit extrinsic apoptosis of infected enterocytes by expressing specific virulence proteins, such as N-acetylglucosamine transferase NleB1, which prevents FADD-mediated recruitment and activation of CASP8 <sup>960</sup> Li, <sup>2013</sup>, <sup>23955153</sup> Li, <sup>2013</sup>, <sup>2395515</sup>

monocytogenes, succumbing to neurolisteriosis. This was proposed to be promoted by an impaired loss of monocytes due to upregulated expression of c-FLIP by the bacterial protein InlB <sup>961</sup>. In support of this result, conditional deletion of Cflar in myeloid cells improved Listeria monocytogenes clearance and animal survival <sup>962</sup>. FAS signaling also conferred protection from infection with (i) human herpes simplex virus 2 (HSV-2), as demonstrated by a decrease in the loss of monocyte and immune cell recruitment at the infection site in  $Fas^{-/-}$  and  $Fasl^{-/-}$  mice  $^{963}$ , and (ii) Citrobacter rodentium or lymphocytic choriomeningitis virus, as demonstrated by an increased neutrophil fraction in mice with conditional deletion of Fas in the myeloid compartment <sup>964</sup>. Supporting an anti-infection role of CASP8, mice lacking RIPK1 kinase activity failed to control systemic Yersinia infection, rapidly dying because of excess apoptosis driven by a kinase independent function of RIPK1 <sup>965, 966</sup>. In line with this finding, *Ripk3*<sup>-/-</sup> Casp8<sup>-/-</sup> but not *Ripk3*<sup>-/-</sup> mice died from Toxoplasma gondii infection due to acute toxoplasmosis, an observation supporting the anti-infection role of CASP8-mediated apoptosis <sup>967</sup>. Moreover, hepatocyte-specific deficiency for CASP8 facilitated liver infection of mice by Listeria monocytogenes, resulting in inflammation and development of necrotic lesions in the liver <sup>775</sup>. These results also suggest an interconnection of multiple RCD pathways in controlling infection. Accordingly, the deletion of Z-DNA binding protein 1 (Zbp1), an essential cytoplasmic sensor of Influenza A virus (IAV) Z-RNA required for the activation of mixed lineage kinase domain like pseudokinase (MLKL)-dependent necroptosis, RIPK1/FADD-dependent apoptosis and NLRP3 inflammasome-dependent pyroptosis, as well as co-deletion of the genes encoding MLKL and FADD, caused a defect in the control of Influenza A virus (IAV) infection, with these mutant mice succumbing to lethal respiratory failure. These findings support an essential role of apoptosis, necroptosis and pyroptosis in IAV clearance 968, 969 Oltean, 2021, 33976111, Nogusa, 2016, 27321907, Zhang, 2020, 32200799 Oltean, 2021, 33976111, Nogusa, 2016, 27321907, Zhang, 2020, 32200799 Oltean, 2021, 33976111, Nogusa, 2016, 27321907, Zhang, 2020, 32200799 Oltean, 2021,

33976111, Nogusa, 2016, 27321907, Zhang, 2020, 32200799. Similarly, combined activation of apoptosis and other RCD

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pathways contribute to the response of mice to *Burkholderia thailandensis* infection <sup>970</sup>. Finally, pharmacological or tissue specific genetic deletion of cIAP1 and cIAP2 results in better control of hepatitis B virus and liver stage malaria parasites due to increased TNF induced death of infected cells (Ebert, 2015 25902529; Ebert 2015, 25902530; Ebert 2020, 32234472).

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Experimental evidence also suggests a detrimental role of extrinsic apoptosis during certain infections. Mice deficient for both TNF-R1 and TNF-R2 displayed decreased sensitivity to lipopolysaccharide, suggesting a critical role for TNF in tissue injury during gram-negative bacterial infection <sup>971</sup>. Along similar lines, TNF-R1-deficient mice were more resistant than wild-type mice to the cytopathic effects of TNF during Sindbis virus infection, as evidenced by reduced mortality and delayed paralysis 972. Moreover, ablation of Ripk1 protected mice from acute liver injury after infection with Listeria monocytogenes <sup>973</sup>, while knockout of Fas or Fasl reduced the effect of toxin A-induced enteritis in mice infected with *Clostridium difficile*, which has been attributed to a reduction in enterocyte loss <sup>974</sup>. Additionally, the infectious spleen and kidney necrosis virus (ISKNV) induced tissue damage in zebrafish by activation of DR-induced apoptosis by a viral protein encoding a TRADD interactor <sup>975</sup>. Of note, in this study, the absence of CASP8 protected zebrafish from ISKNV infection. Finally, Ripk3<sup>-/-</sup>Casp8<sup>-/-</sup> mice exhibited high levels of protection from LPS-induced septic shock <sup>976</sup> or a lethal cytokine shock and tissue damage driven by TNF and IFN-γ, mirroring that of SARS-CoV-2 977. This evidence suggests that the combination of several types of RCD can also mediate infection-associated pathogenesis, as demonstrated for infection with Salmonella <sup>50</sup>.

**Other diseases.** TNF is reported to impair myogenesis in a mouse model of skeletal muscle regeneration after hindlimb immobilization (hindlimb suspension) <sup>978</sup>. Moreover, silencing of TRAIL improved muscle regeneration in mice with acute skeletal muscle injury due to local injection of BaCl2 <sup>979</sup>. An inhibitory role in myogenesis was also ascribed to FADD, at least in response to freezing-induced muscle

injury 980. In apparent contrast with this result, combined deletion of the genes encoding TNF-R1 and 1610 TNF-R2 limited skeletal muscle regeneration after cardiotoxin-induced injury <sup>981, 982</sup>, suggesting the 1611 relevance of a balance between TNF-R1 and TNF-R2 signaling in this model. TRAIL neutralization 1612 increased muscular strength in a mouse model of Duchenne muscular dystrophy 983, while other studies 1613 associated TRAIL and FASL to myositis <sup>984, 985</sup>. 1614 Activation of DRs has also been implicated in the pathogenesis of acute lung injury. Fas silencing as 1615 well as TNF neutralization protected mice from lung injury induced by ischemia-reperfusion 986, 987. 1616 Similarly, deletion of *Tnfrsf1a* (encoding TNF-R1) or pharmacological inhibition of TNF-R1 or CASP8 1617 1618 attenuated pulmonary edema formation and improved alveolar epithelial function in a murine model of acute lung injury induced by acid inhalation <sup>988, 989</sup>. A similar protective effect was provided by 1619 1620 pharmacological inhibition or genetic deletion of FASL or TNF in a lipopolysaccharide-induced mouse model of acute lung injury 990, 991, 992, 993, 994, 995, 996. However, in one study FAS signaling was shown to 1621 contribute to the resolution of acute lung injury by promoting the depletion of macrophages <sup>997</sup>. Using 1622 1623 distinct mouse models of acute lung damage following sepsis, it was shown that the abrogation of FAS and TNF-R1 signaling, including the silencing of Fadd, decreased pulmonary apoptosis and ameliorated 1624 pathology, and in some cases this led to a survival benefit for the animals (e.g., 998, 999, 1000, 1001, 1002, 1003, 1625 <sup>1004</sup>). Hyperoxia-induced lung injury and bleomycin-induced pulmonary fibrosis, a model for cancer 1626 therapy-induced lung injury, are also impacted by the DR pathway. FAS and TNF deficiency exacerbated 1627 hyperoxia-induced lung injury and/or inflammation in newborn mice 1005, 1006. In contrast, TNF inhibition 1628 conferred protection against hyperoxia-induced lung damage in a murine model 1007, 1008, 1009. Moreover, 1629 the absence of TNF-R1 (but not the absence of TNF-R2) improved survival in mice subjected to 1630 excessive oxygen supply, although without decreasing inflammation <sup>1010</sup>. In support of these results, 1631 specific ablation of Fas in murine fibroblasts or T cells exacerbated pulmonary fibrosis induced by 1632

bleomycin <sup>1011, 1012</sup>. However, the level of bleomycin-induced pulmonary fibrosis was diminished in FAS

deficient *lpr/lpr* or FAS ligand deficient *gld/gld* mice <sup>1013</sup> and remained unchanged in mice treated with FAS neutralizing agents <sup>1014</sup>. Likewise, contrasting findings support or refute a role for TNF <sup>1015, 1016, 1017</sup> and TRAIL <sup>1018, 1019</sup> in both the onset and resolution of pulmonary fibrosis after administration of bleomycin. TNF neutralization has been reported to attenuate and enhance interstitial pulmonary fibrosis induced by nitrogen mustard <sup>1020</sup> or rituximab <sup>1021</sup>. Finally, FASL, TNF and/or TRAIL have been implicated in infectious or non-infectious lung disorders, including (but not limited to) infection with respiratory syncytial virus (RSV) <sup>1022, 1023, 1024, 1025, 1026, 1027, 1028</sup>, adenovirus type 1 respiratory disease <sup>1029, 1030</sup>, allergic reaction and asthma <sup>1031, 1032, 1033, 1034, 1035, 1036, 1037, 1038, 1039, 1040, 1041, 1042</sup> and idiopathic pneumonia syndrome <sup>1043</sup>, as well as to chronic lung diseases (e.g., chronic obstructive pulmonary disease) <sup>863, 1044, 1045</sup>.

The studies discussed above illustrate that DR-induced apoptosis is at the heart of many disorders either promoting recovery or exacerbating disease. The active involvement in disease severity and progression makes this pathway a potentially tractable target for therapeutic interventions in a wide range of diseases, typically those with an inflammatory component. However, this effect may be llinked to the role of DR signaling in other RCD pathways and in inflammation. Moreover, there is little consensus on the roles of FASL, TNF and/or TRAIL, highlighting a high complexity of the systems which needs further investigation.

## **Concluding remarks**

Abundant preclinical evidence demonstrates that the intrinsic and the extrinsic pathways of apoptosis not only contribute to adult tissue homeostasis and, in the case of the intrinsic pathway, to embryonic development – the implication of CASP8 in normal development is mainly linked to its role as necroptosis inhibitor (*see* **Box 6** and **Box 7**) - but also contribute to the pathogenesis of multiple diseases, including various cardiovascular, hepatic, neurological and renal disorders as well as multiple infectious,

autoimmune, inflammatory and oncological conditions. However, despite great potential as targets for therapeutic interventions and a considerable research effort dedicated to developing effective approaches, the success of intrinsic or extrinsic apoptosis-targeting agents in clinical settings is so far limited to BH3 mimetic drugs, SMAC mimetics, caspase inhibitors as well as activators or inhibitors of DR signaling, with only one compound, the BCL-2 inhibitor venetoclax (BH3 mimetic drug), approved for routine treatment of patients with CLL or AML.

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Rather than mitigating the enthusiasm about the clinical potential of modulators of apoptosis, this challenge suggests the need for a substantial change in the experimental design and re-interpretation of results, at different levels (**Figure 1**). One major issue is that studies evaluating the impact of apoptotic cell death on disease have not always addressed the connections between the core components of the intrinsic and extrinsic apoptotic machinery or their potential interaction and functional overlap with other RCD pathways. Also, the potential activation of alternative RCD modalities as a mechanism to compensate for the inhibition of apoptotic RCD has not always been explored and thus it has not been tried to prevent or overcome these alternative forms of RCD to achieve superior outcomes. The importance of independent replication of findings that suggest success from targeting a pathway in the treatment of a disease cannot be emphasized enough. Only then can the costly process of clinical translation be approached with confidence and with an increased chance of success. For example, the findings that overexpression of BCL2 or its pro-survival relatives can promote tumorigenesis and can render malignant cells resistant to diverse anti-cancer therapeutics had been reproduced hundreds of times before BH3 mimetic drug development was started. This is not yet the case for many of the other studies discussed here, as best demonstrated by the fact that for certain experiments diametrically opposing results were reported by different groups. These questions must be resolved before considering drug development programs.

Moreover, certain regulators of apoptosis and signaling cascades have been reported to exert a variety of functions beyond cell death control, including (but not limited to) inflammation (e.g., multiple activated caspases and IAPs), cell differentiation (e.g., pro-and anti-apoptotic BCL2 proteins), cell proliferation and survival (e.g., DR engagement). The relevance of these functions is often dependent on cell/tissue type (as it is related to variable expression levels and activation status of other regulators of RCD) and the intensity and duration of the initiating stimulus (as they can direct to a distinct biological outcome, as exemplified by DR ligation). Of note, some of these cell death unrelated functions of bona fide cell death regulators are highly controversial and much more work must be done to verify or discard these notions. On the one hand, this pleiotropy may result in a variable (even including an antagonistic protective vs. promoting) impact of apoptosis on distinct human diseases, also explaining the considerable degree of context-dependency (e.g., effect of stromal and immune cells) observed for its experimental modulation. On the other, the pathogenic effect of core components of the apoptotic machinery is often mediated by such apoptosis-unrelated functions including inflammation, which may point to unexplored targets for the development of new therapeutic agents or approaches.

In our opinion, investigating the molecular cascade of apoptotic cell death in the context of the functional inter-connection between apoptotic and non-apoptotic pathways, for instance by interrupting some of the molecular connections between different RCD signaling cascades, may instigate new advances, ultimately leading to clinical use of specific apoptosis-modulatory agents for the treatment of many diseases.

**Author's contributions.** L.G. and I.V. conceived the review. I.V., L.G. and F.P. wrote the first version of the manuscript with constructive input from all authors. E.G., C.G and G.M. prepared display items under the supervision of L.G. and I.V.

L.G., I.V. and F.P. addressed requests from the Reviewers and Editors of *Cell Death and Differentiation*. All authors approved
 the final version of the article and figures.

Funding and acknowledgments: I.V. is and has been supported by the Associazione Italiana per la Ricerca sul Cancro (AIRC, IG 2017 #20417 and IG 2022 #27685) and a startup grant from the Italian Institute for Genomic Medicine (Candiolo, Turin, Italy) and Compagnia di San Paolo (Torino, Italy). M.P., G.M., S.M., G.C. are supported by the Ministro dell'Università (Italy) progetto Heal Italia PE6. L.G. (as a PI unless otherwise indicated) is or has been supported by two Breakthrough Level 2 grants from the US DoD BCRP (#BC180476P1; #BC210945), by a Transformative Breast Cancer Consortium Grant from the US DoD BCRP (#W81XWH2120034, PI: Formenti), by a U54 grant from NIH/NCI (#CA274291, PI: Deasy, Formenti, Weichselbaum), by a STARR Cancer Consortium grant (#I16-0064), by the 2019 Laura Ziskin Prize in Translational Research (#ZP-6177, PI: Formenti) from the Stand Up to Cancer (SU2C), by a Mantle Cell Lymphoma Research Initiative (MCL-RI, PI: Chen-Kiang) grant from the Leukemia and Lymphoma Society (LLS), by a Rapid Response Grant from the Functional Genomics Initiative (New York, US), by startup funds from the Dept. of Radiation Oncology at Weill Cornell Medicine (New York, US), by industrial collaborations with Lytix Biopharma (Oslo, Norway), Promontory (New York, US) and Onxeo (Paris, France), as well as by donations from Promontory (New York, US), the Luke Heller TECPR2 Foundation (Boston, US), Sotio a.s. (Prague, Czech Republic), Lytix Biopharma (Oslo, Norway), Onxeo (Paris, France), Ricerchiamo (Brescia, Italy), and Noxopharm (Chatswood, Australia). G.K. is supported by the Ligue contre le Cancer (équipe labellisée); Agence National de la Recherche (ANR) – Projets blancs; AMMICa US23/CNRS UMS3655; Association pour la recherche sur le cancer (ARC); Cancéropôle Ile-de-France; European Research Council Advanced Investigator Grand "ICD-Cancer", Fondation pour la Recherche Médicale (FRM); a donation by Elior; Equipex Onco-Pheno-Screen; European Joint Programme on Rare Diseases (EJPRD); European Research Council (ICD-Cancer), European Union Horizon 2020 Projects Oncobiome and Crimson; Fondation Carrefour; Institut National du Cancer (INCa); Institut Universitaire de France; LabEx Immuno-Oncology (ANR-18-IDEX-0001); a Cancer Research ASPIRE Award from the Mark Foundation; the RHU Immunolife; Seerave Foundation; SIRIC Stratified Oncology Cell DNA Repair and Tumor Immune Elimination (SOCRATE); and SIRIC Cancer Research and Personalized Medicine (CARPEM). This study contributes to the IdEx Université de Paris ANR-18-IDEX-0001.

Ethics delaration: conflicts of interest. See above.

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**Data Availability.** No data are included in the article

## References

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- 1731 1. Galluzzi L, Vitale I, Aaronson SA, Abrams JM, Adam D, Agostinis P, et al. Molecular mechanisms of cell death: recommendations of the Nomenclature Committee on Cell Death 2018. 1732
- *Cell death and differentiation* 2018, **25**(3): 486-541. 1733
- 1735 2. Jiang X, Stockwell BR, Conrad M. Ferroptosis: mechanisms, biology and role in disease. *Nature* reviews Molecular cell biology 2021, 22(4): 266-282. 1736
- 1738 3. Del Re DP, Amgalan D, Linkermann A, Liu Q, Kitsis RN. Fundamental Mechanisms of Regulated Cell Death and Implications for Heart Disease. Physiological reviews 2019, 99(4): 1739 1765-1817. 1740
- 1742 4. Bock FJ, Tait SWG. Mitochondria as multifaceted regulators of cell death. Nature reviews *Molecular cell biology* 2020, **21**(2): 85-100. 1743
- 5. Broz P, Pelegrín P, Shao F. The gasdermins, a protein family executing cell death and 1745 1746 inflammation. *Nature reviews Immunology* 2020, **20**(3): 143-157.
- 1748 6. Weinlich R, Oberst A, Beere HM, Green DR. Necroptosis in development, inflammation and disease. Nature reviews Molecular cell biology 2017, 18(2): 127-136. 1749
- 7. Galluzzi L, Buqué A, Kepp O, Zitvogel L, Kroemer G. Immunogenic cell death in cancer and 1751 infectious disease. *Nature reviews Immunology* 2017, **17**(2): 97-111. 1752
- 8. Tang D, Kang R, Berghe TV, Vandenabeele P, Kroemer G. The molecular machinery of regulated 1754 1755 cell death. Cell research 2019, **29**(5): 347-364.
- 9. Gudipaty SA, Conner CM, Rosenblatt J, Montell DJ. Unconventional Ways to Live and Die: Cell 1757 Death and Survival in Development, Homeostasis, and Disease. Annual review of cell and 1758 1759 *developmental biology* 2018, **34:** 311-332.
- 10. Bedoui S, Herold MJ, Strasser A. Emerging connectivity of programmed cell death pathways and 1761 its physiological implications. *Nature reviews Molecular cell biology* 2020, **21**(11): 678-695. 1762
- 1764 11. Galluzzi L, Vitale I, Abrams JM, Alnemri ES, Baehrecke EH, Blagosklonny MV, et al. Molecular definitions of cell death subroutines: recommendations of the Nomenclature Committee on Cell 1765 1766 Death 2012. *Cell death and differentiation* 2012, **19**(1): 107-120.

1767 12. 1768 Kesavardhana S, Malireddi RKS, Kanneganti TD. Caspases in Cell Death, Inflammation, and Pyroptosis. *Annual review of immunology* 2020, **38:** 567-595. 1769 1770 13. Kumar S, Dorstyn L, Lim Y. The role of caspases as executioners of apoptosis. Biochemical 1771 *Society transactions* 2022, **50**(1): 33-45. 1772 1773 1774 14. Galluzzi L, Bravo-San Pedro JM, Vitale I, Aaronson SA, Abrams JM, Adam D, et al. Essential versus accessory aspects of cell death: recommendations of the NCCD 2015. Cell death and 1775 differentiation 2015, 22(1): 58-73. 1776 1777 1778 15. Marsden VS, O'Connor L, O'Reilly LA, Silke J, Metcalf D, Ekert PG, et al. Apoptosis initiated by Bcl-2-regulated caspase activation independently of the cytochrome c/Apaf-1/caspase-9 1779 apoptosome. Nature 2002, 419(6907): 634-637. 1780 1781 1782 16. Cheng EH, Wei MC, Weiler S, Flavell RA, Mak TW, Lindsten T, et al. BCL-2, BCL-X(L) sequester BH3 domain-only molecules preventing BAX- and BAK-mediated mitochondrial 1783 1784 apoptosis. *Molecular cell* 2001, **8**(3): 705-711. 1785 1786 17. Pandian N, Kanneganti TD. PANoptosis: A Unique Innate Immune Inflammatory Cell Death Modality. *Journal of immunology (Baltimore, Md: 1950)* 2022, **209**(9): 1625-1633. 1787 1788 18. Bonora M, Giorgi C, Pinton P. Molecular mechanisms and consequences of mitochondrial 1789 1790 permeability transition. *Nature reviews Molecular cell biology* 2022, **23**(4): 266-285. 1791 19. Chen X, Zeh HJ, Kang R, Kroemer G, Tang D. Cell death in pancreatic cancer: from pathogenesis 1792 to therapy. Nat Rev Gastroenterol Hepatol 2021, **18**(11): 804-823. 1793 1794 1795 20. Song X, Zhu S, Xie Y, Liu J, Sun L, Zeng D, et al. JTC801 Induces pH-dependent Death Specifically in Cancer Cells and Slows Growth of Tumors in Mice. Gastroenterology 2018, 1796 **154**(5): 1480-1493. 1797 1798 1799 21. Tsvetkov P, Coy S, Petrova B, Dreishpoon M, Verma A, Abdusamad M, et al. Copper induces 1800 cell death by targeting lipoylated TCA cycle proteins. Science (New York, NY) 2022, 375(6586): 1801 1254-1261.

Malireddi RKS, Kesavardhana S, Kanneganti TD. ZBP1 and TAK1: Master Regulators of NLRP3 Inflammasome/Pyroptosis, Apoptosis, and Necroptosis (PAN-optosis). *Front Cell Infect* 

1802 1803

1804

1805

22.

Microbiol 2019, 9: 406.

1806 23. Green DR. The Coming Decade of Cell Death Research: Five Riddles. Cell 2019, 177(5): 1094-1807 1808 1107. 1809 24. Singh R, Letai A, Sarosiek K. Regulation of apoptosis in health and disease: the balancing act of 1810 BCL-2 family proteins. *Nature reviews Molecular cell biology* 2019, **20**(3): 175-193. 1811 1812 25. Ke FFS, Brinkmann K, Voss AK, Strasser A. Some mice lacking intrinsic, as well as death 1813 1814 receptor induced apoptosis and necroptosis, can survive to adulthood. Cell death & disease 2022, **13**(4): 317. 1815 1816 26. Ke FFS, Vanyai HK, Cowan AD, Delbridge ARD, Whitehead L, Grabow S, et al. Embryogenesis 1817 1818 and Adult Life in the Absence of Intrinsic Apoptosis Effectors BAX, BAK, and BOK. Cell 2018, **173**(5): 1217-1230.e1217. 1819 1820 27. 1821 Spetz J, Galluzzi L. Preface: Life through death-Key role of cellular suicide for colonial and organismal homeostasis. *International review of cell and molecular biology* 2020, **352:** xi-xv. 1822 1823 Carneiro BA, El-Deiry WS. Targeting apoptosis in cancer therapy. Nature reviews Clinical 1824 28. 1825 oncology 2020, **17**(7): 395-417. 1826 1827 29. Anderton H, Wicks IP, Silke J. Cell death in chronic inflammation: breaking the cycle to treat rheumatic disease. *Nature reviews Rheumatology* 2020, **16**(9): 496-513. 1828 1829 30. Li K, van Delft MF, Dewson G. Too much death can kill you: inhibiting intrinsic apoptosis to 1830 treat disease. *The EMBO journal* 2021, **40**(14): e107341. 1831 1832 31. Jain N, Keating M, Thompson P, Ferrajoli A, Burger J, Borthakur G, et al. Ibrutinib and 1833 Venetoclax for First-Line Treatment of CLL. The New England journal of medicine 2019, 1834 **380**(22)**:** 2095-2103. 1835 1836 1837 32. Souers AJ, Leverson JD, Boghaert ER, Ackler SL, Catron ND, Chen J, et al. ABT-199, a potent and selective BCL-2 inhibitor, achieves antitumor activity while sparing platelets. Nature 1838 medicine 2013, **19**(2): 202-208. 1839 1840 33. Diepstraten ST, Anderson MA, Czabotar PE, Lessene G, Strasser A, Kelly GL. The manipulation 1841 1842 of apoptosis for cancer therapy using BH3-mimetic drugs. Nature reviews Cancer 2022, 22(1):

45-64.

1843

- Merino D, Kelly GL, Lessene G, Wei AH, Roberts AW, Strasser A. BH3-Mimetic Drugs: Blazing the Trail for New Cancer Medicines. *Cancer cell* 2018, **34**(6): 879-891.
- Fischer K, Al-Sawaf O, Bahlo J, Fink AM, Tandon M, Dixon M, et al. Venetoclax and Obinutuzumab in Patients with CLL and Coexisting Conditions. The New England journal of medicine 2019, **380**(23): 2225-2236.

1851

1855

1859

1863

1868

1872

1876

1880

- 1852 36. Roberts AW, Davids MS, Pagel JM, Kahl BS, Puvvada SD, Gerecitano JF, *et al.* Targeting BCL2 with Venetoclax in Relapsed Chronic Lymphocytic Leukemia. *The New England journal of medicine* 2016, **374**(4): 311-322.
- Seymour JF, Kipps TJ, Eichhorst B, Hillmen P, D'Rozario J, Assouline S, *et al.* Venetoclax-Rituximab in Relapsed or Refractory Chronic Lymphocytic Leukemia. *The New England journal* of medicine 2018, **378**(12): 1107-1120.
- DiNardo CD, Jonas BA, Pullarkat V, Thirman MJ, Garcia JS, Wei AH, *et al.* Azacitidine and Venetoclax in Previously Untreated Acute Myeloid Leukemia. *The New England journal of medicine* 2020, **383**(7): 617-629.
- Frenette CT, Morelli G, Shiffman ML, Frederick RT, Rubin RA, Fallon MB, *et al.* Emricasan Improves Liver Function in Patients With Cirrhosis and High Model for End-Stage Liver Disease Scores Compared With Placebo. *Clinical gastroenterology and hepatology : the official clinical practice journal of the American Gastroenterological Association* 2019, **17**(4): 774-783.e774.
- Garcia-Tsao G, Bosch J, Kayali Z, Harrison SA, Abdelmalek MF, Lawitz E, *et al.* Randomized placebo-controlled trial of emricasan for non-alcoholic steatohepatitis-related cirrhosis with severe portal hypertension. *Journal of hepatology* 2020, **72**(5): 885-895.
- Harrison SA, Goodman Z, Jabbar A, Vemulapalli R, Younes ZH, Freilich B, *et al.* A randomized, placebo-controlled trial of emricasan in patients with NASH and F1-F3 fibrosis. *Journal of hepatology* 2020, **72**(5): 816-827.
- Galluzzi L, Aaronson SA, Abrams J, Alnemri ES, Andrews DW, Baehrecke EH, et al. Guidelines for the use and interpretation of assays for monitoring cell death in higher eukaryotes. Cell death and differentiation 2009, 16(8): 1093-1107.
- Boada-Romero E, Martinez J, Heckmann BL, Green DR. The clearance of dead cells by efferocytosis. *Nature reviews Molecular cell biology* 2020, **21**(7): 398-414.

- 1884 44. Rothlin CV, Hille TD, Ghosh S. Determining the effector response to cell death. *Nature reviews Immunology* 2021, **21**(5): 292-304.
- Morioka S, Maueroder C, Ravichandran KS. Living on the Edge: Efferocytosis at the Interface of Homeostasis and Pathology. *Immunity* 2019, **50**(5): 1149-1162.

1889

1893

1897

1900

1903

1907

1911

1914

1918

- 1890 46. Raymond MH, Davidson AJ, Shen Y, Tudor DR, Lucas CD, Morioka S, *et al.* Live cell tracking of macrophage efferocytosis during Drosophila embryo development in vivo. *Science (New York, NY)* 2022, **375**(6585): 1182-1187.
- Nonomura K, Yamaguchi Y, Hamachi M, Koike M, Uchiyama Y, Nakazato K, *et al.* Local apoptosis modulates early mammalian brain development through the elimination of morphogen-producing cells. *Developmental cell* 2013, **27**(6): 621-634.
- Li MO, Sarkisian MR, Mehal WZ, Rakic P, Flavell RA. Phosphatidylserine receptor is required for clearance of apoptotic cells. *Science (New York, NY)* 2003, **302**(5650): 1560-1563.
- 1901 49. Kist M, Vucic D. Cell death pathways: intricate connections and disease implications. *The EMBO journal* 2021, **40**(5): e106700.
- Doerflinger M, Deng Y, Whitney P, Salvamoser R, Engel S, Kueh AJ, *et al.* Flexible Usage and Interconnectivity of Diverse Cell Death Pathways Protect against Intracellular Infection. *Immunity* 2020, **53**(3): 533-547.e537.
- 1908 51. Oberst A, Dillon CP, Weinlich R, McCormick LL, Fitzgerald P, Pop C, *et al.* Catalytic activity of the caspase-8-FLIP(L) complex inhibits RIPK3-dependent necrosis. *Nature* 2011, **471**(7338): 363-367.
- 1912 52. Kaiser WJ, Upton JW, Long AB, Livingston-Rosanoff D, Daley-Bauer LP, Hakem R, *et al.* RIP3 mediates the embryonic lethality of caspase-8-deficient mice. *Nature* 2011, **471**(7338): 368-372.
- 1915 53. O'Donnell MA, Perez-Jimenez E, Oberst A, Ng A, Massoumi R, Xavier R, *et al.* Caspase 8 1916 inhibits programmed necrosis by processing CYLD. *Nature cell biology* 2011, **13**(12): 1437-1917 1442.
- 1919 54. Cauwels A, Janssen B, Waeytens A, Cuvelier C, Brouckaert P. Caspase inhibition causes hyperacute tumor necrosis factor-induced shock via oxidative stress and phospholipase A2.

  Nature immunology 2003, 4(4): 387-393.

- Vercammen D, Brouckaert G, Denecker G, Van de Craen M, Declercq W, Fiers W, *et al.* Dual signaling of the Fas receptor: initiation of both apoptotic and necrotic cell death pathways. *The Journal of experimental medicine* 1998, **188**(5): 919-930.
- Vercammen D, Beyaert R, Denecker G, Goossens V, Van Loo G, Declercq W, *et al.* Inhibition of caspases increases the sensitivity of L929 cells to necrosis mediated by tumor necrosis factor. *The Journal of experimental medicine* 1998, **187**(9): 1477-1485.

1930

1934

1938

1941

1944

1947

1950

1953

1957

- Brumatti G, Ma C, Lalaoui N, Nguyen NY, Navarro M, Tanzer MC, *et al.* The caspase-8 inhibitor emricasan combines with the SMAC mimetic birinapant to induce necroptosis and treat acute myeloid leukemia. *Science translational medicine* 2016, **8**(339): 339ra369.
- Saelens X, Kalai M, Vandenabeele P. Translation inhibition in apoptosis: caspase-dependent PKR activation and eIF2-alpha phosphorylation. *The Journal of biological chemistry* 2001, **276**(45): 41620-41628.
- Tait SW, Green DR. Mitochondria and cell death: outer membrane permeabilization and beyond.
   Nature reviews Molecular cell biology 2010, 11(9): 621-632.
- 1942 60. Chipuk JE, Mohammed JN, Gelles JD, Chen Y. Mechanistic connections between mitochondrial biology and regulated cell death. *Developmental cell* 2021, **56**(9): 1221-1233.
- 1945 61. Green DR, Victor B. The pantheon of the fallen: why are there so many forms of cell death? *Trends in cell biology* 2012, **22**(11): 555-556.
- Davidovich P, Kearney CJ, Martin SJ. Inflammatory outcomes of apoptosis, necrosis and necroptosis. *Biological chemistry* 2014, **395**(10): 1163-1171.
- 1951 63. Galluzzi L, López-Soto A, Kumar S, Kroemer G. Caspases Connect Cell-Death Signaling to Organismal Homeostasis. *Immunity* 2016, **44**(2): 221-231.
- Duprez L, Takahashi N, Van Hauwermeiren F, Vandendriessche B, Goossens V, Vanden Berghe T, *et al.* RIP kinase-dependent necrosis drives lethal systemic inflammatory response syndrome. *Immunity* 2011, **35**(6): 908-918.
- 1958 65. Glab JA, Cao Z, Puthalakath H. Bcl-2 family proteins, beyond the veil. *International review of cell and molecular biology* 2020, **351:** 1-22.

- 1961 66. Gross A, Katz SG. Non-apoptotic functions of BCL-2 family proteins. *Cell death and differentiation* 2017, **24**(8): 1348-1358.
- Hollville E, Deshmukh M. Physiological functions of non-apoptotic caspase activity in the nervous system. *Seminars in cell & developmental biology* 2018, **82:** 127-136.

1966

1969

1972

1976

1980

1985

1989

1992

- 1967 68. Nakajima YI, Kuranaga E. Caspase-dependent non-apoptotic processes in development. *Cell death and differentiation* 2017, **24**(8): 1422-1430.
- Aram L, Yacobi-Sharon K, Arama E. CDPs: caspase-dependent non-lethal cellular processes.
   Cell death and differentiation 2017, 24(8): 1307-1310.
- Feinstein-Rotkopf Y, Arama E. Can't live without them, can live with them: roles of caspases during vital cellular processes. *Apoptosis : an international journal on programmed cell death* 2009, **14**(8): 980-995.
- Perciavalle RM, Stewart DP, Koss B, Lynch J, Milasta S, Bathina M, *et al.* Anti-apoptotic MCLlocalizes to the mitochondrial matrix and couples mitochondrial fusion to respiration. *Nature cell biology* 2012, **14**(6): 575-583.
- Wu L, Tan JL, Wang ZH, Chen YX, Gao L, Liu JL, *et al.* ROS generated during early reperfusion contribute to intermittent hypobaric hypoxia-afforded cardioprotection against postischemia-induced Ca(2+) overload and contractile dysfunction via the JAK2/STAT3 pathway. *Journal of molecular and cellular cardiology* 2015, **81:** 150-161.
- Vanden Berghe T, Hulpiau P, Martens L, Vandenbroucke RE, Van Wonterghem E, Perry SW, et
   al. Passenger Mutations Confound Interpretation of All Genetically Modified Congenic Mice.
   Immunity 2015, 43(1): 200-209.
- Wang Y, Gao W, Shi X, Ding J, Liu W, He H, *et al.* Chemotherapy drugs induce pyroptosis through caspase-3 cleavage of a gasdermin. *Nature* 2017, **547**(7661): 99-103.
- 1993 75. Rogers C, Fernandes-Alnemri T, Mayes L, Alnemri D, Cingolani G, Alnemri ES. Cleavage of DFNA5 by caspase-3 during apoptosis mediates progression to secondary necrotic/pyroptotic cell death. *Nature communications* 2017, **8:** 14128.
- Gould TW, Buss RR, Vinsant S, Prevette D, Sun W, Knudson CM, *et al.* Complete dissociation of motor neuron death from motor dysfunction by Bax deletion in a mouse model of ALS. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2006, **26**(34): 8774-8786.

2001
2002
77. Reyes NA, Fisher JK, Austgen K, VandenBerg S, Huang EJ, Oakes SA. Blocking the mitochondrial apoptotic pathway preserves motor neuron viability and function in a mouse model of amyotrophic lateral sclerosis. *The Journal of clinical investigation* 2010, **120**(10): 3673-3679.

78. Kostic V, Jackson-Lewis V, de Bilbao F, Dubois-Dauphin M, Przedborski S. Bcl-2: prolonging life in a transgenic mouse model of familial amyotrophic lateral sclerosis. *Science (New York, NY)* 1997, 277(5325): 559-562.

2005

2009

2014

2018

2022

2027

2031

2035

- Vukosavic S, Stefanis L, Jackson-Lewis V, Guégan C, Romero N, Chen C, et al. Delaying caspase activation by Bcl-2: A clue to disease retardation in a transgenic mouse model of amyotrophic lateral sclerosis. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2000, **20**(24): 9119-9125.
- Inoue H, Tsukita K, Iwasato T, Suzuki Y, Tomioka M, Tateno M, *et al.* The crucial role of caspase-9 in the disease progression of a transgenic ALS mouse model. *The EMBO journal* 2003, **22**(24): 6665-6674.
- Wootz H, Hansson I, Korhonen L, Lindholm D. XIAP decreases caspase-12 cleavage and calpain activity in spinal cord of ALS transgenic mice. *Experimental cell research* 2006, **312**(10): 1890-1898.
- Kieran D, Woods I, Villunger A, Strasser A, Prehn JH. Deletion of the BH3-only protein puma protects motoneurons from ER stress-induced apoptosis and delays motoneuron loss in ALS mice. *Proceedings of the National Academy of Sciences of the United States of America* 2007, **104**(51): 20606-20611.
- 2028 83. Li M, Ona VO, Guégan C, Chen M, Jackson-Lewis V, Andrews LJ, *et al.* Functional role of caspase-1 and caspase-3 in an ALS transgenic mouse model. *Science (New York, NY)* 2000, **288**(5464): 335-339.
- Girgenrath M, Dominov JA, Kostek CA, Miller JB. Inhibition of apoptosis improves outcome in a model of congenital muscular dystrophy. *The Journal of clinical investigation* 2004, **114**(11): 1635-1639.
- Davies JE, Rubinsztein DC. Over-expression of BCL2 rescues muscle weakness in a mouse model of oculopharyngeal muscular dystrophy. *Human molecular genetics* 2011, **20**(6): 1154-1163.

- Dominov JA, Kravetz AJ, Ardelt M, Kostek CA, Beermann ML, Miller JB. Muscle-specific BCL2 expression ameliorates muscle disease in laminin {alpha}2-deficient, but not in dystrophin-deficient, mice. *Human molecular genetics* 2005, **14**(8): 1029-1040.
- Sagot Y, Dubois-Dauphin M, Tan SA, de Bilbao F, Aebischer P, Martinou JC, *et al.* Bcl-2 overexpression prevents motoneuron cell body loss but not axonal degeneration in a mouse model of a neurodegenerative disease. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 1995, **15**(11): 7727-7733.

2048

2052

2055

2059

2064

2068

2072

2075

- Tossing G, Livernoche R, Maios C, Bretonneau C, Labarre A, Parker JA. Genetic and pharmacological PARP inhibition reduces axonal degeneration in C. elegans models of ALS. *Human molecular genetics* 2022, **31**(19): 3313-3324.
- 89. Kudo W, Lee HP, Smith MA, Zhu X, Matsuyama S, Lee HG. Inhibition of Bax protects neuronal cells from oligomeric Aβ neurotoxicity. *Cell death & disease* 2012, **3**(5): e309.
- 2056 90. Bové J, Martínez-Vicente M, Dehay B, Perier C, Recasens A, Bombrun A, *et al.* BAX channel activity mediates lysosomal disruption linked to Parkinson disease. *Autophagy* 2014, **10**(5): 889-900.
- Vila M, Jackson-Lewis V, Vukosavic S, Djaldetti R, Liberatore G, Offen D, *et al.* Bax ablation prevents dopaminergic neurodegeneration in the 1-methyl- 4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. *Proceedings of the National Academy of Sciences of the United States of America* 2001, **98**(5): 2837-2842.
- 2065 92. Kim TW, Moon Y, Kim K, Lee JE, Koh HC, Rhyu IJ, *et al.* Dissociation of progressive dopaminergic neuronal death and behavioral impairments by Bax deletion in a mouse model of Parkinson's diseases. *PloS one* 2011, **6**(10): e25346.
- 2069 93. Ma C, Pan Y, Yang Z, Meng Z, Sun R, Wang T, *et al.* Pre-administration of BAX-inhibiting peptides decrease the loss of the nigral dopaminergic neurons in rats. *Life sciences* 2016, **144:** 113-120.
- Jiang H, He P, Adler CH, Shill H, Beach TG, Li R, *et al.* Bid signal pathway components are identified in the temporal cortex with Parkinson disease. *Neurology* 2012, **79**(17): 1767-1773.
- Biswas SC, Ryu E, Park C, Malagelada C, Greene LA. Puma and p53 play required roles in death evoked in a cellular model of Parkinson disease. *Neurochemical research* 2005, **30**(6-7): 839-845.

- 2080 96. Akhter R, Saleem S, Saha A, Biswas SC. The pro-apoptotic protein Bmf co-operates with Bim and Puma in neuron death induced by  $\beta$ -amyloid or NGF deprivation. *Molecular and cellular neurosciences* 2018, **88:** 249-257.
- Imaizumi K, Morihara T, Mori Y, Katayama T, Tsuda M, Furuyama T, *et al.* The cell death-promoting gene DP5, which interacts with the BCL2 family, is induced during neuronal apoptosis following exposure to amyloid beta protein. *The Journal of biological chemistry* 1999, **274**(12): 7975-7981.

2088

2092

2096

2101

2104

2108

2112

- 2089 98. Louneva N, Cohen JW, Han LY, Talbot K, Wilson RS, Bennett DA, *et al.* Caspase-3 is enriched in postsynaptic densities and increased in Alzheimer's disease. *The American journal of pathology* 2008, **173**(5): 1488-1495.
- 2093 99. Rohn TT, Rissman RA, Davis MC, Kim YE, Cotman CW, Head E. Caspase-9 activation and caspase cleavage of tau in the Alzheimer's disease brain. *Neurobiology of disease* 2002, **11**(2): 341-354.
- Hartmann A, Hunot S, Michel PP, Muriel MP, Vyas S, Faucheux BA, *et al.* Caspase-3: A vulnerability factor and final effector in apoptotic death of dopaminergic neurons in Parkinson's disease. *Proceedings of the National Academy of Sciences of the United States of America* 2000, **97**(6): 2875-2880.
- 2102 101. Zhang L, Qian Y, Li J, Zhou X, Xu H, Yan J, *et al.* BAD-mediated neuronal apoptosis and neuroinflammation contribute to Alzheimer's disease pathology. *iScience* 2021, **24**(9): 102942.
- 2105 102. Rissman RA, Poon WW, Blurton-Jones M, Oddo S, Torp R, Vitek MP, *et al.* Caspase-cleavage of tau is an early event in Alzheimer disease tangle pathology. *The Journal of clinical investigation* 2004, **114**(1): 121-130.
- 2109 103. Gervais FG, Xu D, Robertson GS, Vaillancourt JP, Zhu Y, Huang J, *et al.* Involvement of caspases in proteolytic cleavage of Alzheimer's amyloid-beta precursor protein and amyloidogenic A beta peptide formation. *Cell* 1999, **97**(3): 395-406.
- 2113 104. Chu J, Lauretti E, Praticò D. Caspase-3-dependent cleavage of Akt modulates tau phosphorylation via GSK3β kinase: implications for Alzheimer's disease. *Molecular psychiatry* 2115 2017, 22(7): 1002-1008.
- 2117 105. Rohn TT, Vyas V, Hernandez-Estrada T, Nichol KE, Christie LA, Head E. Lack of pathology in a triple transgenic mouse model of Alzheimer's disease after overexpression of the anti-apoptotic protein Bcl-2. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2120 2008, **28**(12): 3051-3059.

2121 106. Kumasaka DK, Galvan V, Head E, Rohn TT. Caspase cleavage of the amyloid precursor protein 2122 is prevented after overexpression of bcl-2 in a triple transgenic mouse model of Alzheimer's 2123 2124 disease. International journal of physiology, pathophysiology and pharmacology 2009, 1(1): 48-56. 2125 2126 107. 2127 D'Amelio M, Cavallucci V, Middei S, Marchetti C, Pacioni S, Ferri A, et al. Caspase-3 triggers early synaptic dysfunction in a mouse model of Alzheimer's disease. Nature neuroscience 2011, 2128 **14**(1): 69-76. 2129 2130 2131 108. Park G, Nhan HS, Tyan SH, Kawakatsu Y, Zhang C, Navarro M, et al. Caspase Activation and Caspase-Mediated Cleavage of APP Is Associated with Amyloid β-Protein-Induced Synapse 2132 Loss in Alzheimer's Disease. Cell reports 2020, 31(13): 107839. 2133 2134 109. Pozueta J, Lefort R, Ribe EM, Troy CM, Arancio O, Shelanski M. Caspase-2 is required for 2135 2136 dendritic spine and behavioural alterations in J20 APP transgenic mice. Nature communications 2137 2013, **4:** 1939. 2138 110. Troy CM, Shelanski ML. Caspase-2 and tau-a toxic partnership? *Nature medicine* 2016, **22**(11): 2139 1207-1208. 2140 2141 111. Zhao X, Kotilinek LA, Smith B, Hlynialuk C, Zahs K, Ramsden M, et al. Caspase-2 cleavage of 2142 tau reversibly impairs memory. *Nature medicine* 2016, **22**(11): 1268-1276. 2143 2144 2145 112. Steuer EL, Kemper LJ, Hlynialuk CJW, Leinonen-Wright K, Montonye ML, Lapcinski IP, et al. Blocking Site-Specific Cleavage of Human Tau Delays Progression of Disease-Related 2146 2147 Phenotypes in Genetically Matched Tau-Transgenic Mice Modeling Frontotemporal Dementia. The Journal of neuroscience: the official journal of the Society for Neuroscience 2022, 42(23): 2148 4737-4754. 2149 2150 2151 Bresinsky M, Strasser JM, Vallaster B, Liu P, McCue WM, Fuller J, et al. Structure-Based Design and Biological Evaluation of Novel Caspase-2 Inhibitors Based on the Peptide AcVDVAD-CHO 2152 and the Caspase-2-Mediated Tau Cleavage Sequence YKPVD314. ACS pharmacology & 2153 2154 *translational science* 2022, **5**(1): 20-40.

Kajiwara Y, McKenzie A, Dorr N, Gama Sosa MA, Elder G, Schmeidler J, et al. The human-specific CASP4 gene product contributes to Alzheimer-related synaptic and behavioural deficits.

Human molecular genetics 2016, **25**(19): 4315-4327.

21552156

2157

2158

2159

114.

- 2160 115. Lee JH, Won SM, Suh J, Son SJ, Moon GJ, Park UJ, *et al.* Induction of the unfolded protein response and cell death pathway in Alzheimer's disease, but not in aged Tg2576 mice. *Experimental & molecular medicine* 2010, **42**(5): 386-394.
- 116. Kolosova NG, Tyumentsev MA, Muraleva NA, Kiseleva E, Vitovtov AO, Stefanova NA.
   2165 Antioxidant SkQ1 Alleviates Signs of Alzheimer's Disease-like Pathology in Old OXYS Rats by
   2166 Reversing Mitochondrial Deterioration. Current Alzheimer research 2017, 14(12): 1283-1292.

2167

2172

2176

2181

2185

2189

2193

- Perier C, Bové J, Wu DC, Dehay B, Choi DK, Jackson-Lewis V, et al. Two molecular pathways initiate mitochondria-dependent dopaminergic neurodegeneration in experimental Parkinson's disease. Proceedings of the National Academy of Sciences of the United States of America 2007, 104(19): 8161-8166.
- 2173 118. Yamada M, Kida K, Amutuhaire W, Ichinose F, Kaneki M. Gene disruption of caspase-3 prevents
  2174 MPTP-induced Parkinson's disease in mice. *Biochemical and biophysical research*2175 communications 2010, **402**(2): 312-318.
- 119. Viswanath V, Wu Y, Boonplueang R, Chen S, Stevenson FF, Yantiri F, *et al.* Caspase-9 activation results in downstream caspase-8 activation and bid cleavage in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinson's disease. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2001, **21**(24): 9519-9528.
- 2182 120. Crocker SJ, Liston P, Anisman H, Lee CJ, Smith PD, Earl N, *et al.* Attenuation of MPTP-induced neurotoxicity and behavioural impairment in NSE-XIAP transgenic mice. *Neurobiology of disease* 2003, **12**(2): 150-161.
- Liu Y, Guo Y, An S, Kuang Y, He X, Ma H, *et al.* Targeting caspase-3 as dual therapeutic benefits by RNAi facilitating brain-targeted nanoparticles in a rat model of Parkinson's disease. *PloS one* 2013, **8**(5): e62905.
- Toulmond S, Tang K, Bureau Y, Ashdown H, Degen S, O'Donnell R, *et al.* Neuroprotective effects of M826, a reversible caspase-3 inhibitor, in the rat malonate model of Huntington's disease. *British journal of pharmacology* 2004, **141**(4): 689-697.
- Leyva MJ, Degiacomo F, Kaltenbach LS, Holcomb J, Zhang N, Gafni J, *et al.* Identification and evaluation of small molecule pan-caspase inhibitors in Huntington's disease models. *Chemistry* & *biology* 2010, **17**(11): 1189-1200.
- 2198 124. Chen M, Ona VO, Li M, Ferrante RJ, Fink KB, Zhu S, *et al.* Minocycline inhibits caspase-1 and caspase-3 expression and delays mortality in a transgenic mouse model of Huntington disease.

  2200 *Nature medicine* 2000, **6**(7): 797-801.

2201
 2202 125. Cen X, Chen Y, Xu X, Wu R, He F, Zhao Q, et al. Pharmacological targeting of MCL-1 promotes
 2203 mitophagy and improves disease pathologies in an Alzheimer's disease mouse model. Nature
 2204 communications 2020, 11(1): 5731.

Ekholm-Reed S, Baker R, Campos AR, Stouffer D, Henze M, Wolf DA, et al. Reducing Mcl-1 gene dosage induces dopaminergic neuronal loss and motor impairments in Park2 knockout mice.
 Communications biology 2019, 2: 125.

2205

2209

2213

2217

2222

2227

2230

2234

2238

127. Koper MJ, Van Schoor E, Ospitalieri S, Vandenberghe R, Vandenbulcke M, von Arnim CAF, *et al.* Necrosome complex detected in granulovacuolar degeneration is associated with neuronal loss in Alzheimer's disease. *Acta neuropathologica* 2020, **139**(3): 463-484.

- Hambright WS, Fonseca RS, Chen L, Na R, Ran Q. Ablation of ferroptosis regulator glutathione peroxidase 4 in forebrain neurons promotes cognitive impairment and neurodegeneration. *Redox biology* 2017, **12:** 8-17.
- Chiesa R, Piccardo P, Dossena S, Nowoslawski L, Roth KA, Ghetti B, et al. Bax deletion prevents neuronal loss but not neurological symptoms in a transgenic model of inherited prion disease.
   Proceedings of the National Academy of Sciences of the United States of America 2005, 102(1): 238-243.
- 2223 130. Steele AD, King OD, Jackson WS, Hetz CA, Borkowski AW, Thielen P, *et al.* Diminishing apoptosis by deletion of Bax or overexpression of Bcl-2 does not protect against infectious prion toxicity in vivo. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2007, **27**(47): 13022-13027.
- Pemberton JM, Pogmore JP, Andrews DW. Neuronal cell life, death, and axonal degeneration as regulated by the BCL-2 family proteins. *Cell death and differentiation* 2021, **28**(1): 108-122.
- 2231 132. Ray SK, Samantaray S, Smith JA, Matzelle DD, Das A, Banik NL. Inhibition of cysteine proteases in acute and chronic spinal cord injury. *Neurotherapeutics : the journal of the American Society for Experimental NeuroTherapeutics* 2011, **8**(2): 180-186.
- 2235 133. Sarosiek KA, Fraser C, Muthalagu N, Bhola PD, Chang W, McBrayer SK, *et al.* Developmental 2236 Regulation of Mitochondrial Apoptosis by c-Myc Governs Age- and Tissue-Specific Sensitivity 2237 to Cancer Therapeutics. *Cancer cell* 2017, **31**(1): 142-156.
- Tehranian R, Rose ME, Vagni V, Pickrell AM, Griffith RP, Liu H, *et al.* Disruption of Bax protein prevents neuronal cell death but produces cognitive impairment in mice following traumatic brain injury. *Journal of neurotrauma* 2008, **25**(7): 755-767.

Tehranian R, Rose ME, Vagni V, Griffith RP, Wu S, Maits S, et al. Transgenic mice that overexpress the anti-apoptotic Bcl-2 protein have improved histological outcome but unchanged behavioral outcome after traumatic brain injury. Brain research 2006, 1101(1): 126-135.

2247 136. Bermpohl D, You Z, Korsmeyer SJ, Moskowitz MA, Whalen MJ. Traumatic brain injury in mice 2248 deficient in Bid: effects on histopathology and functional outcome. *Journal of cerebral blood* 2249 *flow and metabolism : official journal of the International Society of Cerebral Blood Flow and* 2250 *Metabolism* 2006, **26**(5): 625-633.

2246

2251

2256

2260

2265

2269

2273

2277

2281

2252 137. Raghupathi R, Fernandez SC, Murai H, Trusko SP, Scott RW, Nishioka WK, *et al.* BCL-2 2253 overexpression attenuates cortical cell loss after traumatic brain injury in transgenic mice. *Journal* 2254 *of cerebral blood flow and metabolism : official journal of the International Society of Cerebral* 2255 *Blood Flow and Metabolism* 1998, **18**(11): 1259-1269.

Farlie PG, Dringen R, Rees SM, Kannourakis G, Bernard O. bcl-2 transgene expression can protect neurons against developmental and induced cell death. *Proceedings of the National Academy of Sciences of the United States of America* 1995, 92(10): 4397-4401.

Dong H, Fazzaro A, Xiang C, Korsmeyer SJ, Jacquin MF, McDonald JW. Enhanced oligodendrocyte survival after spinal cord injury in Bax-deficient mice and mice with delayed Wallerian degeneration. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2003, **23**(25): 8682-8691.

2266 140. Barut S, Unlü YA, Karaoğlan A, Tunçdemir M, Dağistanli FK, Oztürk M, *et al.* The neuroprotective effects of z-DEVD.fmk, a caspase-3 inhibitor, on traumatic spinal cord injury in rats. *Surgical neurology* 2005, **64**(3): 213-220; discussion 220.

2270 141. Colak A, Karaoğlan A, Barut S, Köktürk S, Akyildiz AI, Taşyürekli M. Neuroprotection and functional recovery after application of the caspase-9 inhibitor z-LEHD-fmk in a rat model of traumatic spinal cord injury. *Journal of neurosurgery Spine* 2005, **2**(3): 327-334.

2274 142. Li M, Ona VO, Chen M, Kaul M, Tenneti L, Zhang X, *et al.* Functional role and therapeutic implications of neuronal caspase-1 and -3 in a mouse model of traumatic spinal cord injury.

Neuroscience 2000, **99**(2): 333-342.

2278 Zhao W, Li H, Hou Y, Jin Y, Zhang L. Combined Administration of Poly-ADP-Ribose 2279 Polymerase-1 and Caspase-3 Inhibitors Alleviates Neuronal Apoptosis After Spinal Cord Injury 2280 in Rats. *World neurosurgery* 2019, **127**: e346-e352.

- Donahue RJ, Maes ME, Grosser JA, Nickells RW. BAX-Depleted Retinal Ganglion Cells Survive and Become Quiescent Following Optic Nerve Damage. *Molecular neurobiology* 2020, 57(2): 1070-1084.
- Libby RT, Li Y, Savinova OV, Barter J, Smith RS, Nickells RW, *et al.* Susceptibility to neurodegeneration in a glaucoma is modified by Bax gene dosage. *PLoS genetics* 2005, **1**(1): 17-228

2289

2292

2295

2299

2303

2307

2311

2315

- Harder JM, Libby RT. BBC3 (PUMA) regulates developmental apoptosis but not axonal injury induced death in the retina. *Molecular neurodegeneration* 2011, **6:** 50.
- Harder JM, Libby RT. Deficiency in Bim, Bid and Bbc3 (Puma) do not prevent axonal injury induced death. *Cell death and differentiation* 2013, **20**(1): 182.
- Harder JM, Ding Q, Fernandes KA, Cherry JD, Gan L, Libby RT. BCL2L1 (BCL-X) promotes survival of adult and developing retinal ganglion cells. *Molecular and cellular neurosciences* 2012, **51**(1-2): 53-59.
- Visuvanathan S, Baker AN, Lagali PS, Coupland SG, Miller G, Hauswirth WW, et al. XIAP gene therapy effects on retinal ganglion cell structure and function in a mouse model of glaucoma.
   Gene therapy 2022, 29(3-4): 147-156.
- Donahue RJ, Fehrman RL, Gustafson JR, Nickells RW. BCLX(L) gene therapy moderates neuropathology in the DBA/2J mouse model of inherited glaucoma. *Cell death & disease* 2021, **12**(8): 781.
- 2308 151. Avrutsky MI, Ortiz CC, Johnson KV, Potenski AM, Chen CW, Lawson JM, *et al.* Endothelial activation of caspase-9 promotes neurovascular injury in retinal vein occlusion. *Nature communications* 2020, **11**(1): 3173.
- Ishikawa S, Hirata A, Nakabayashi J, Iwakiri R, Okinami S. Neuroprotective effect of small interfering RNA targeted to caspase-3 on rat retinal ganglion cell loss induced by ischemia and reperfusion injury. *Current eye research* 2012, **37**(10): 907-913.
- Tawfik M, Zhang X, Grigartzik L, Heiduschka P, Hintz W, Henrich-Noack P, *et al.* Gene therapy with caspase-3 small interfering RNA-nanoparticles is neuroprotective after optic nerve damage.

  Neural regeneration research 2021, **16**(12): 2534-2541.

- 2320 154. Wassmer SJ, De Repentigny Y, Sheppard D, Lagali PS, Fang L, Coupland SG, *et al.* XIAP Protects Retinal Ganglion Cells in the Mutant ND4 Mouse Model of Leber Hereditary Optic Neuropathy. *Investigative ophthalmology & visual science* 2020, **61**(8): 49.
- Wassmer SJ, Leonard BC, Coupland SG, Baker AN, Hamilton J, Hauswirth WW, et al. Overexpression of the X-Linked Inhibitor of Apoptosis Protects Against Retinal Degeneration in a Feline Model of Retinal Detachment. *Human gene therapy* 2017, **28**(6): 482-492.

2327

2330

2334

2338

2342

2346

2349

2353

- 2328 156. Renwick J, Narang MA, Coupland SG, Xuan JY, Baker AN, Brousseau J, *et al.* XIAP-mediated neuroprotection in retinal ischemia. *Gene therapy* 2006, **13**(4): 339-347.
- McKinnon SJ, Lehman DM, Tahzib NG, Ransom NL, Reitsamer HA, Liston P, et al. Baculoviral IAP repeat-containing-4 protects optic nerve axons in a rat glaucoma model. Molecular therapy
   the journal of the American Society of Gene Therapy 2002, 5(6): 780-787.
- Zadro-Lamoureux LA, Zacks DN, Baker AN, Zheng QD, Hauswirth WW, Tsilfidis C. XIAP effects on retinal detachment-induced photoreceptor apoptosis [corrected]. *Investigative ophthalmology & visual science* 2009, **50**(3): 1448-1453.
- Yao J, Feathers KL, Khanna H, Thompson D, Tsilfidis C, Hauswirth WW, *et al.* XIAP therapy increases survival of transplanted rod precursors in a degenerating host retina. *Investigative ophthalmology & visual science* 2011, **52**(3): 1567-1572.
- 2343 160. Crespo-Garcia S, Tsuruda PR, Dejda A, Ryan RD, Fournier F, Chaney SY, *et al.* Pathological angiogenesis in retinopathy engages cellular senescence and is amenable to therapeutic elimination via BCL-xL inhibition. *Cell metabolism* 2021, **33**(4): 818-832.e817.
- 2347 161. Choudhury S, Liu Y, Clark AF, Pang IH. Caspase-7: a critical mediator of optic nerve injury-2348 induced retinal ganglion cell death. *Molecular neurodegeneration* 2015, **10:** 40.
- Wang S, Sorenson CM, Sheibani N. Attenuation of retinal vascular development and neovascularization during oxygen-induced ischemic retinopathy in Bcl-2-/- mice. *Developmental biology* 2005, **279**(1): 205-219.
- Wang S, Park S, Fei P, Sorenson CM. Bim is responsible for the inherent sensitivity of the developing retinal vasculature to hyperoxia. *Developmental biology* 2011, **349**(2): 296-309.
- 2357 164. Grant ZL, Whitehead L, Wong VH, He Z, Yan RY, Miles AR, *et al.* Blocking endothelial apoptosis revascularizes the retina in a model of ischemic retinopathy. *The Journal of clinical investigation* 2020, **130**(8): 4235-4251.

Du H, Sun X, Guma M, Luo J, Ouyang H, Zhang X, et al. JNK inhibition reduces apoptosis and neovascularization in a murine model of age-related macular degeneration. Proceedings of the National Academy of Sciences of the United States of America 2013, 110(6): 2377-2382.

2364
2365 166. Deckwerth TL, Elliott JL, Knudson CM, Johnson EM, Jr., Snider WD, Korsmeyer SJ. BAX is required for neuronal death after trophic factor deprivation and during development. *Neuron* 1996, **17**(3): 401-411.

Unsain N, Higgins JM, Parker KN, Johnstone AD, Barker PA. XIAP regulates caspase activity in degenerating axons. *Cell reports* 2013, **4**(4): 751-763.

2368

2371

2376

2380

2385

2389

2393

2397

Imaizumi K, Benito A, Kiryu-Seo S, Gonzalez V, Inohara N, Lieberman AP, *et al.* Critical role for DP5/Harakiri, a Bcl-2 homology domain 3-only Bcl-2 family member, in axotomy-induced neuronal cell death. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2004, **24**(15): 3721-3725.

Theofilas P, Bedner P, Hüttmann K, Theis M, Steinhäuser C, Frank S. The proapoptotic BCL-2 homology domain 3-only protein Bim is not critical for acute excitotoxic cell death. *Journal of neuropathology and experimental neurology* 2009, **68**(1): 102-110.

Bunk EC, König HG, Prehn JHM, Kirby BP. p53 upregulated mediator of apoptosis (Puma) deficiency increases survival of adult neural stem cells generated physiologically in the hippocampus, but does not protect stem cells generated in surplus after an excitotoxic lesion.

Journal of basic and clinical physiology and pharmacology 2020, 32(2): 57-66.

Li T, Fan Y, Luo Y, Xiao B, Lu C. In vivo delivery of a XIAP (BIR3-RING) fusion protein containing the protein transduction domain protects against neuronal death induced by seizures. Experimental neurology 2006, **197**(2): 301-308.

Tzeng TT, Tsay HJ, Chang L, Hsu CL, Lai TH, Huang FL, *et al.* Caspase 3 involves in neuroplasticity, microglial activation and neurogenesis in the mice hippocampus after intracerebral injection of kainic acid. *Journal of biomedical science* 2013, **20**(1): 90.

2394 173. Concannon CG, Tuffy LP, Weisová P, Bonner HP, Dávila D, Bonner C, *et al.* AMP kinasemediated activation of the BH3-only protein Bim couples energy depletion to stress-induced apoptosis. *The Journal of cell biology* 2010, **189**(1): 83-94.

Murphy BM, Engel T, Paucard A, Hatazaki S, Mouri G, Tanaka K, *et al.* Contrasting patterns of Bim induction and neuroprotection in Bim-deficient mice between hippocampus and neocortex after status epilepticus. *Cell death and differentiation* 2010, **17**(3): 459-468.

Foley J, Burnham V, Tedoldi M, Danial NN, Yellen G. BAD knockout provides metabolic seizure resistance in a genetic model of epilepsy with sudden unexplained death in epilepsy. *Epilepsia* 2018, 59(1): e1-e4.

Moran C, Sanz-Rodriguez A, Jimenez-Pacheco A, Martinez-Villareal J, McKiernan RC,
 Jimenez-Mateos EM, *et al.* Bmf upregulation through the AMP-activated protein kinase pathway
 may protect the brain from seizure-induced cell death. *Cell death & disease* 2013, 4(4): e606.

2405

2409

2414

2417

2421

2425

2429

2433

- 2410 177. Engel T, Murphy BM, Hatazaki S, Jimenez-Mateos EM, Concannon CG, Woods I, et al. Reduced 2411 hippocampal damage and epileptic seizures after status epilepticus in mice lacking proapoptotic 2412 Puma. FASEB journal: official publication of the Federation of American Societies for 2413 Experimental Biology 2010, 24(3): 853-861.
- 2415 178. Engel T, Hatazaki S, Tanaka K, Prehn JH, Henshall DC. Deletion of Puma protects hippocampal neurons in a model of severe status epilepticus. *Neuroscience* 2010, **168**(2): 443-450.
- 2418 179. Murphy B, Dunleavy M, Shinoda S, Schindler C, Meller R, Bellver-Estelles C, *et al.* Bcl-w protects hippocampus during experimental status epilepticus. *The American journal of pathology* 2007, **171**(4): 1258-1268.
- 180. Ichikawa N, Alves M, Pfeiffer S, Langa E, Hernández-Santana YE, Suzuki H, *et al.* Deletion of the BH3-only protein Noxa alters electrographic seizures but does not protect against hippocampal damage after status epilepticus in mice. *Cell death & disease* 2017, **8**(1): e2556.
- 2426 181. Engel T, Caballero-Caballero A, Schindler CK, Plesnila N, Strasser A, Prehn JH, *et al.* BH3-only protein Bid is dispensable for seizure-induced neuronal death and the associated nuclear accumulation of apoptosis-inducing factor. *Journal of neurochemistry* 2010, **115**(1): 92-101.
- 2430 182. Gibson ME, Han BH, Choi J, Knudson CM, Korsmeyer SJ, Parsadanian M, *et al.* BAX contributes to apoptotic-like death following neonatal hypoxia-ischemia: evidence for distinct apoptosis pathways. *Molecular medicine (Cambridge, Mass)* 2001, **7**(9): 644-655.
- Ness JM, Harvey CA, Strasser A, Bouillet P, Klocke BJ, Roth KA. Selective involvement of BH3-only Bcl-2 family members Bim and Bad in neonatal hypoxia-ischemia. *Brain research* 2006, **1099**(1): 150-159.
- 2438 184. Wang X, Zhu C, Wang X, Hagberg H, Korhonen L, Sandberg M, *et al.* X-linked inhibitor of apoptosis (XIAP) protein protects against caspase activation and tissue loss after neonatal hypoxia-ischemia. *Neurobiology of disease* 2004, **16**(1): 179-189.

West T, Stump M, Lodygensky G, Neil JJ, Deshmukh M, Holtzman DM. Lack of X-linked inhibitor of apoptosis protein leads to increased apoptosis and tissue loss following neonatal brain injury. *ASN neuro* 2009, **1**(1).

2445

2446 186. West T, Atzeva M, Holtzman DM. Caspase-3 deficiency during development increases vulnerability to hypoxic-ischemic injury through caspase-3-independent pathways. *Neurobiology* of disease 2006, **22**(3): 523-537.

2449

2450 187. Ghosh AP, Walls KC, Klocke BJ, Toms R, Strasser A, Roth KA. The proapoptotic BH3-only, 2451 Bcl-2 family member, Puma is critical for acute ethanol-induced neuronal apoptosis. *Journal of neuropathology and experimental neurology* 2009, **68**(7): 747-756.

2453 2454

2454 188. Young C, Klocke BJ, Tenkova T, Choi J, Labruyere J, Qin YQ, et al. Ethanol-induced neuronal apoptosis in vivo requires BAX in the developing mouse brain. Cell death and differentiation 2003, **10**(10): 1148-1155.

2457

Young C, Roth KA, Klocke BJ, West T, Holtzman DM, Labruyere J, *et al.* Role of caspase-3 in ethanol-induced developmental neurodegeneration. *Neurobiology of disease* 2005, **20**(2): 608-614.

2461

2462 190. Slupe AM, Villasana L, Wright KM. GABAergic neurons are susceptible to BAX-dependent apoptosis following isoflurane exposure in the neonatal period. *PloS one* 2021, **16**(1): e0238799.

2464

2465 191. Chong MJ, Murray MR, Gosink EC, Russell HR, Srinivasan A, Kapsetaki M, *et al.* Atm and Bax cooperate in ionizing radiation-induced apoptosis in the central nervous system. *Proceedings of the National Academy of Sciences of the United States of America* 2000, **97**(2): 889-894.

2468

2469 192. Ahlers KE, Karaçay B, Fuller L, Bonthius DJ, Dailey ME. Transient activation of microglia 2470 following acute alcohol exposure in developing mouse neocortex is primarily driven by BAX-2471 dependent neurodegeneration. *Glia* 2015, **63**(10): 1694-1713.

2472

2473 193. D'Orsi B, Kilbride SM, Chen G, Perez Alvarez S, Bonner HP, Pfeiffer S, et al. Bax regulates 2474 neuronal Ca2+ homeostasis. *The Journal of neuroscience : the official journal of the Society for* 2475 *Neuroscience* 2015, **35**(4): 1706-1722.

2476

2477 194. Pfeiffer S, Anilkumar U, Chen G, Ramírez-Peinado S, Galindo-Moreno J, Muñoz-Pinedo C, *et al.* Analysis of BH3-only proteins upregulated in response to oxygen/glucose deprivation in cortical neurons identifies Bmf but not Noxa as potential mediator of neuronal injury. *Cell death & disease* 2014, **5**(10): e1456.

2482 195. Plesnila N, Zinkel S, Le DA, Amin-Hanjani S, Wu Y, Qiu J, et al. BID mediates neuronal cell death after oxygen/glucose deprivation and focal cerebral ischemia. Proceedings of the National Academy of Sciences of the United States of America 2001, **98**(26): 15318-15323.

2485

2486 196. Yin XM, Luo Y, Cao G, Bai L, Pei W, Kuharsky DK, *et al.* Bid-mediated mitochondrial pathway is critical to ischemic neuronal apoptosis and focal cerebral ischemia. *The Journal of biological chemistry* 2002, **277**(44): 42074-42081.

2489

2490 197. Plesnila N, Zinkel S, Amin-Hanjani S, Qiu J, Korsmeyer SJ, Moskowitz MA. Function of BID 2491 - a molecule of the bcl-2 family -- in ischemic cell death in the brain. *European surgical research*2492 *Europaische chirurgische Forschung Recherches chirurgicales europeennes* 2002, **34**(1-2): 372493 41.

2494

Martin NA, Bonner H, Elkjær ML, D'Orsi B, Chen G, König HG, *et al.* BID Mediates Oxygen-Glucose Deprivation-Induced Neuronal Injury in Organotypic Hippocampal Slice Cultures and Modulates Tissue Inflammation in a Transient Focal Cerebral Ischemia Model without Changing Lesion Volume. *Frontiers in cellular neuroscience* 2016, **10:** 14.

2499

2500 199. Kitagawa K, Matsumoto M, Tsujimoto Y, Ohtsuki T, Kuwabara K, Matsushita K, *et al.*2501 Amelioration of hippocampal neuronal damage after global ischemia by neuronal overexpression of BCL-2 in transgenic mice. *Stroke* 1998, **29**(12): 2616-2621.

2503

2504 200. Cao G, Pei W, Ge H, Liang Q, Luo Y, Sharp FR, *et al.* In Vivo Delivery of a Bcl-xL Fusion Protein Containing the TAT Protein Transduction Domain Protects against Ischemic Brain Injury and Neuronal Apoptosis. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2002, **22**(13): 5423-5431.

2508

2509 201. Kilic E, Hermann DM, Kügler S, Kilic U, Holzmüller H, Schmeer C, *et al.* Adenovirus-mediated Bcl-X(L) expression using a neuron-specific synapsin-1 promoter protects against disseminated neuronal injury and brain infarction following focal cerebral ischemia in mice. *Neurobiology of disease* 2002, **11**(2): 275-284.

2513

Akpan N, Serrano-Saiz E, Zacharia BE, Otten ML, Ducruet AF, Snipas SJ, *et al.* Intranasal delivery of caspase-9 inhibitor reduces caspase-6-dependent axon/neuron loss and improves neurological function after stroke. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2011, **31**(24): 8894-8904.

2518

2519 203. Fan YF, Lu CZ, Xie J, Zhao YX, Yang GY. Apoptosis inhibition in ischemic brain by intraperitoneal PTD-BIR3-RING (XIAP). *Neurochemistry international* 2006, **48**(1): 50-59.

- Trapp T, Korhonen L, Besselmann M, Martinez R, Mercer EA, Lindholm D. Transgenic mice 2522 204. overexpressing XIAP in neurons show better outcome after transient cerebral ischemia. 2523
- *Molecular and cellular neurosciences* 2003, **23**(2): 302-313. 2524

Zhu C, Xu F, Fukuda A, Wang X, Fukuda H, Korhonen L, et al. X chromosome-linked inhibitor 2526 205. 2527 of apoptosis protein reduces oxidative stress after cerebral irradiation or hypoxia-ischemia through up-regulation of mitochondrial antioxidants. The European journal of neuroscience 2528 2007, **26**(12): 3402-3410. 2529

2530

2531 206. Zhao H, Yenari MA, Cheng D, Sapolsky RM, Steinberg GK. Biphasic cytochrome c release after 2532 transient global ischemia and its inhibition by hypothermia. Journal of cerebral blood flow and metabolism: official journal of the International Society of Cerebral Blood Flow and Metabolism 2533 2005, **25**(9): 1119-1129. 2534

2535

Chen J, Nagayama T, Jin K, Stetler RA, Zhu RL, Graham SH, et al. Induction of caspase-3-like 2536 207. 2537 protease may mediate delayed neuronal death in the hippocampus after transient cerebral 2538 ischemia. The Journal of neuroscience: the official journal of the Society for Neuroscience 1998, **18**(13): 4914-4928. 2539

2540

Gao Y, Liang W, Hu X, Zhang W, Stetler RA, Vosler P, et al. Neuroprotection against hypoxic-2541 208. ischemic brain injury by inhibiting the apoptotic protease activating factor-1 pathway. Stroke 2542 2010, **41**(1): 166-172. 2543

2544

209. Karatas H, Aktas Y, Gursoy-Ozdemir Y, Bodur E, Yemisci M, Caban S, et al. A nanomedicine 2545 transports a peptide caspase-3 inhibitor across the blood-brain barrier and provides 2546 2547 neuroprotection. The Journal of neuroscience: the official journal of the Society for Neuroscience 2009, **29**(44): 13761-13769. 2548

2549

2550 210. Endres M, Namura S, Shimizu-Sasamata M, Waeber C, Zhang L, Gómez-Isla T, et al. Attenuation of delayed neuronal death after mild focal ischemia in mice by inhibition of the 2551 caspase family. Journal of cerebral blood flow and metabolism: official journal of the 2552 2553 *International Society of Cerebral Blood Flow and Metabolism* 1998, **18**(3): 238-247.

2554

2555 211. Gottron FJ, Ying HS, Choi DW. Caspase inhibition selectively reduces the apoptotic component of oxygen-glucose deprivation-induced cortical neuronal cell death. Molecular and cellular 2556 neurosciences 1997, **9**(3): 159-169. 2557

2558

Shibata M, Hisahara S, Hara H, Yamawaki T, Fukuuchi Y, Yuan J, et al. Caspases determine the 2559 212. 2560 vulnerability of oligodendrocytes in the ischemic brain. The Journal of clinical investigation 2000, **106**(5): 643-653. 2561

- 213. Sung JH, Zhao H, Roy M, Sapolsky RM, Steinberg GK. Viral caspase inhibitor p35, but not crmA, is neuroprotective in the ischemic penumbra following experimental stroke. *Neuroscience* 2007, **149**(4): 804-812.
- 2567 214. Braun JS, Prass K, Dirnagl U, Meisel A, Meisel C. Protection from brain damage and bacterial infection in murine stroke by the novel caspase-inhibitor Q-VD-OPH. *Experimental neurology* 2007, **206**(2): 183-191.

2570

2573

2577

2580

2586

2589

2594

2598

- 215. Sun Y, Xu Y, Geng L. Caspase-3 inhibitor prevents the apoptosis of brain tissue in rats with acute cerebral infarction. *Experimental and therapeutic medicine* 2015, **10**(1): 133-138.
- 2574 216. Lapchak PA, Araujo DM, Weir CJ, Wei J, Zivin JA. Effects of intrathecal administration of a cell permeant caspase inhibitor, boc-D-fluoromethylketone (BDFMK), on behavioral deficits following spinal cord ischemia: a dose-response analysis. *Brain research* 2003, **959**(2): 183-190.
- 2578 217. Osman AM, Neumann S, Kuhn HG, Blomgren K. Caspase inhibition impaired the neural stem/progenitor cell response after cortical ischemia in mice. *Oncotarget* 2016, **7**(3): 2239-2248.
- 218. Zhan RZ, Wu C, Fujihara H, Taga K, Qi S, Naito M, et al. Both caspase-dependent and caspase-independent pathways may be involved in hippocampal CA1 neuronal death because of loss of cytochrome c From mitochondria in a rat forebrain ischemia model. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* 2001, **21**(5): 529-540.
- 2587 219. Moujalled D, Strasser A, Liddell JR. Molecular mechanisms of cell death in neurological diseases. *Cell death and differentiation* 2021, **28**(7): 2029-2044.
- 2590 220. Crowther AJ, Gama V, Bevilacqua A, Chang SX, Yuan H, Deshmukh M, *et al.* Tonic activation of Bax primes neural progenitors for rapid apoptosis through a mechanism preserved in medulloblastoma. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2013, **33**(46): 18098-18108.
- 2595 221. Nakaya K, Hasegawa T, Flickinger JC, Kondziolka DS, Fellows-Mayle W, Gobbel GT. Sensitivity to radiation-induced apoptosis and neuron loss declines rapidly in the postnatal mouse neocortex. *Int J Radiat Biol* 2005, **81**(7): 545-554.
- 2599 222. Kole AJ, Annis RP, Deshmukh M. Mature neurons: equipped for survival. *Cell death & disease* 2600 2013, 4(6): e689.

- 223. Whelan RS, Konstantinidis K, Wei AC, Chen Y, Reyna DE, Jha S, *et al.* Bax regulates primary necrosis through mitochondrial dynamics. *Proceedings of the National Academy of Sciences of the United States of America* 2012, **109**(17): 6566-6571.
- 2606 224. Karch J, Kwong JQ, Burr AR, Sargent MA, Elrod JW, Peixoto PM, *et al.* Bax and Bak function as the outer membrane component of the mitochondrial permeability pore in regulating necrotic cell death in mice. *eLife* 2013, **2:** e00772.

2609

2613

2616

2620

2625

2629

2634

- 2610 225. Hochhauser E, Cheporko Y, Yasovich N, Pinchas L, Offen D, Barhum Y, *et al.* Bax deficiency reduces infarct size and improves long-term function after myocardial infarction. *Cell biochemistry and biophysics* 2007, **47**(1): 11-20.
- Vaseva AV, Marchenko ND, Ji K, Tsirka SE, Holzmann S, Moll UM. p53 opens the mitochondrial permeability transition pore to trigger necrosis. *Cell* 2012, **149**(7): 1536-1548.
- 227. Brocheriou V, Hagège AA, Oubenaïssa A, Lambert M, Mallet VO, Duriez M, *et al.* Cardiac functional improvement by a human Bcl-2 transgene in a mouse model of ischemia/reperfusion injury. *The journal of gene medicine* 2000, **2**(5): 326-333.
- 2621 228. Kristen AV, Ackermann K, Buss S, Lehmann L, Schnabel PA, Haunstetter A, *et al.* Inhibition of apoptosis by the intrinsic but not the extrinsic apoptotic pathway in myocardial ischemia-reperfusion. *Cardiovascular pathology : the official journal of the Society for Cardiovascular Pathology* 2013, **22**(4): 280-286.
- 2626 229. Chen Z, Chua CC, Ho YS, Hamdy RC, Chua BH. Overexpression of Bcl-2 attenuates apoptosis and protects against myocardial I/R injury in transgenic mice. *American journal of physiology Heart and circulatory physiology* 2001, **280**(5): H2313-2320.
- 230. Ono M, Sawa Y, Ryugo M, Alechine AN, Shimizu S, Sugioka R, et al. BH4 peptide derivative from Bcl-xL attenuates ischemia/reperfusion injury thorough anti-apoptotic mechanism in rat hearts. European journal of cardio-thoracic surgery: official journal of the European Association for Cardio-thoracic Surgery 2005, 27(1): 117-121.
- 2635 231. Toth A, Jeffers JR, Nickson P, Min JY, Morgan JP, Zambetti GP, et al. Targeted deletion of Puma
   2636 attenuates cardiomyocyte death and improves cardiac function during ischemia-reperfusion.
   2637 American journal of physiology Heart and circulatory physiology 2006, 291(1): H52-60.
- 2639 232. Gao J, Zhang L, Wang WL, Ma Q, Chu HC. Post-conditioning anti-PUMA treatment protects mice against mice heart I/R injury. *European review for medical and pharmacological sciences* 2016, **20**(8): 1623-1627.

2642
2643
2644
Bi W, Wang J, Jiang Y, Li Q, Wang S, Liu M, *et al.* Neurotrophin-3 contributes to benefits of human embryonic stem cell-derived cardiovascular progenitor cells against reperfused myocardial infarction. *Stem cells translational medicine* 2021, **10**(5): 756-772.

234. Mersmann J, Zacharowski PA, Schmitz I, Zacharowski K. Caspase inhibitor zVAD.fmk reduces infarct size after myocardial ischaemia and reperfusion in rats but not in mice. *Resuscitation* 2008, **79**(3): 468-474.

2646

2650

2653

2657

2661

2666

2670

2673

2676

- Yaoita H, Ogawa K, Maehara K, Maruyama Y. Attenuation of ischemia/reperfusion injury in rats by a caspase inhibitor. *Circulation* 1998, **97**(3): 276-281.
- Huang JQ, Radinovic S, Rezaiefar P, Black SC. In vivo myocardial infarct size reduction by a caspase inhibitor administered after the onset of ischemia. *European journal of pharmacology* 2000, **402**(1-2): 139-142.
- 2658 237. Souktani R, Pons S, Guegan C, Bouhidel O, Bruneval P, Zini R, et al. Cardioprotection against
   2659 myocardial infarction with PTD-BIR3/RING, a XIAP mimicking protein. Journal of molecular
   2660 and cellular cardiology 2009, 46(5): 713-718.
- Inserte J, Cardona M, Poncelas-Nozal M, Hernando V, Vilardosa Ú, Aluja D, *et al.* Studies on the role of apoptosis after transient myocardial ischemia: genetic deletion of the executioner caspases-3 and -7 does not limit infarct size and ventricular remodeling. *Basic research in cardiology* 2016, **111**(2): 18.
- 239. Weisleder N, Taffet GE, Capetanaki Y. Bcl-2 overexpression corrects mitochondrial defects and ameliorates inherited desmin null cardiomyopathy. *Proceedings of the National Academy of Sciences of the United States of America* 2004, **101**(3): 769-774.
- 240. Maloyan A, Sayegh J, Osinska H, Chua BH, Robbins J. Manipulation of death pathways in desmin-related cardiomyopathy. *Circulation research* 2010, **106**(9): 1524-1532.
- 241. Khalil H, Peltzer N, Walicki J, Yang JY, Dubuis G, Gardiol N, *et al.* Caspase-3 protects stressed organs against cell death. *Molecular and cellular biology* 2012, **32**(22): 4523-4533.
- 242. Rodriguez-Ruiz ME, Buqué A, Hensler M, Chen J, Bloy N, Petroni G, *et al.* Apoptotic caspases inhibit abscopal responses to radiation and identify a new prognostic biomarker for breast cancer patients. *Oncoimmunology* 2019, **8**(11): e1655964.

- 243. White MJ, McArthur K, Metcalf D, Lane RM, Cambier JC, Herold MJ, *et al.* Apoptotic caspases suppress mtDNA-induced STING-mediated type I IFN production. *Cell* 2014, **159**(7): 1549-1562.
- 2685 244. Rongvaux A, Jackson R, Harman CC, Li T, West AP, de Zoete MR, *et al.* Apoptotic caspases prevent the induction of type I interferons by mitochondrial DNA. *Cell* 2014, **159**(7): 1563-1577.

2687

2690

2694

2698

2702

2706

2710

2714

- 2688 245. King KR, Aguirre AD, Ye YX, Sun Y, Roh JD, Ng RP, Jr., *et al.* IRF3 and type I interferons fuel a fatal response to myocardial infarction. *Nature medicine* 2017, **23**(12): 1481-1487.
- 246. Fauvel H, Marchetti P, Chopin C, Formstecher P, Nevière R. Differential effects of caspase inhibitors on endotoxin-induced myocardial dysfunction and heart apoptosis. *American journal of physiology Heart and circulatory physiology* 2001, **280**(4): H1608-1614.
- 247. Carlson DL, Maass DL, White J, Sikes P, Horton JW. Caspase inhibition reduces cardiac myocyte dyshomeostasis and improves cardiac contractile function after major burn injury. *Journal of applied physiology (Bethesda, Md : 1985)* 2007, **103**(1): 323-330.
- 248. Araki T, Shibata M, Takano R, Hisahara S, Imamura S, Fukuuchi Y, *et al.* Conditional expression of anti-apoptotic protein p35 by Cre-mediated DNA recombination in cardiomyocytes from loxP-p35-transgenic mice. *Cell death and differentiation* 2000, **7**(5): 485-492.
- 2703 249. Amgalan D, Garner TP, Pekson R, Jia XF, Yanamandala M, Paulino V, *et al.* A small-molecule allosteric inhibitor of BAX protects against doxorubicin-induced cardiomyopathy. *Nature cancer* 2020, **1**(3): 315-328.
- 2707 250. Smyth LA, Meader L, Xiao F, Woodward M, Brady HJ, Lechler R, *et al.* Constitutive expression of the anti-apoptotic Bcl-2 family member A1 in murine endothelial cells leads to transplant tolerance. *Clinical and experimental immunology* 2017, **188**(2): 219-225.
- 2711 251. Grootaert MO, Schrijvers DM, Hermans M, Van Hoof VO, De Meyer GR, Martinet W. Caspase 2712 3 Deletion Promotes Necrosis in Atherosclerotic Plaques of ApoE Knockout Mice. Oxidative
   2713 medicine and cellular longevity 2016, 2016: 3087469.
- 2715 252. Chao ML, Guo J, Cheng WL, Zhu XY, She ZG, Huang Z, et al. Loss of Caspase-Activated DNase
   2716 Protects Against Atherosclerosis in Apolipoprotein E-Deficient Mice. Journal of the American
   2717 Heart Association 2016, 5(12).

2719 253. Fontaine MAC, Westra MM, Bot I, Jin H, Franssen A, Bot M, *et al.* Low human and murine Mcl-1 expression leads to a pro-apoptotic plaque phenotype enriched in giant-cells. *Scientific reports* 2721 2019, **9**(1): 14547.

2722

254. Lee MKS, Kraakman MJ, Dragoljevic D, Hanssen NMJ, Flynn MC, Al-Sharea A, et al. Apoptotic
 Ablation of Platelets Reduces Atherosclerosis in Mice With Diabetes. Arteriosclerosis,
 thrombosis, and vascular biology 2021, 41(3): 1167-1178.

2726

2727 255. Temmerman L, Westra MM, Bot I, van Vlijmen BJM, van Bree N, Bot M, *et al.* Leukocyte Bim deficiency does not impact atherogenesis in ldlr (-/-) mice, despite a pronounced induction of autoimmune inflammation. *Scientific reports* 2017, **7**(1): 3086.

2730

2731 256. Thorp E, Li Y, Bao L, Yao PM, Kuriakose G, Rong J, *et al.* Brief report: increased apoptosis in advanced atherosclerotic lesions of Apoe-/- mice lacking macrophage Bcl-2. *Arteriosclerosis, thrombosis, and vascular biology* 2009, **29**(2): 169-172.

2734

2735 257. Wei Q, Dong G, Chen JK, Ramesh G, Dong Z. Bax and Bak have critical roles in ischemic acute kidney injury in global and proximal tubule-specific knockout mouse models. *Kidney international* 2013, **84**(1): 138-148.

2738

2739 258. Wei Q, Yin XM, Wang MH, Dong Z. Bid deficiency ameliorates ischemic renal failure and delays animal death in C57BL/6 mice. *American journal of physiology Renal physiology* 2006, **290**(1): F35-42.

2742

2743 259. Chien CT, Shyue SK, Lai MK. Bcl-xL augmentation potentially reduces ischemia/reperfusion induced proximal and distal tubular apoptosis and autophagy. *Transplantation* 2007, **84**(9): 1183-1190.

2746

2747 260. Mei S, Li L, Wei Q, Hao J, Su Y, Mei C, *et al.* Double knockout of Bax and Bak from kidney proximal tubules reduces unilateral urethral obstruction associated apoptosis and renal interstitial fibrosis. *Scientific reports* 2017, **7:** 44892.

2750

2751 261. Jang HS, Padanilam BJ. Simultaneous deletion of Bax and Bak is required to prevent apoptosis and interstitial fibrosis in obstructive nephropathy. *American journal of physiology Renal physiology* 2015, **309**(6): F540-550.

2754

Yang B, Lan S, Dieudé M, Sabo-Vatasescu JP, Karakeussian-Rimbaud A, Turgeon J, et al.
 Caspase-3 Is a Pivotal Regulator of Microvascular Rarefaction and Renal Fibrosis after Ischemia Reperfusion Injury. *Journal of the American Society of Nephrology: JASN* 2018, 29(7): 1900 1916.

2759
 2760 263. Lan S, Yang B, Migneault F, Turgeon J, Bourgault M, Dieudé M, et al. Caspase-3-dependent peritubular capillary dysfunction is pivotal for the transition from acute to chronic kidney disease after acute ischemia-reperfusion injury. American journal of physiology Renal physiology 2021,

2763 **321**(3): F335-f351.

2768

2772

2776

2780

2785

2789

2794

2798

2764
 2765 264. Tao Y, Zafar I, Kim J, Schrier RW, Edelstein CL. Caspase-3 gene deletion prolongs survival in polycystic kidney disease. *Journal of the American Society of Nephrology : JASN* 2008, 19(4): 749-755.

265. Duplomb L, Droin N, Bouchot O, Thauvin-Robinet C, Bruel AL, Thevenon J, et al. A constitutive
 BCL2 down-regulation aggravates the phenotype of PKD1-mutant-induced polycystic kidney
 disease. Human molecular genetics 2017, 26(23): 4680-4688.

2773 266. Daemen MA, van 't Veer C, Denecker G, Heemskerk VH, Wolfs TG, Clauss M, *et al.* Inhibition of apoptosis induced by ischemia-reperfusion prevents inflammation. *The Journal of clinical investigation* 1999, **104**(5): 541-549.

267. Bral M, Pawlick R, Marfil-Garza B, Dadheech N, Hefler J, Thiesen A, *et al.* Pan-caspase inhibitor F573 mitigates liver ischemia reperfusion injury in a murine model. *PloS one* 2019, **14**(11): e0224567.

Tao Y, Kim J, Faubel S, Wu JC, Falk SA, Schrier RW, et al. Caspase inhibition reduces tubular apoptosis and proliferation and slows disease progression in polycystic kidney disease.
 Proceedings of the National Academy of Sciences of the United States of America 2005, 102(19): 6954-6959.

2786 269. Yang B, Johnson TS, Haylor JL, Wagner B, Watson PF, El Kossi MM, *et al.* Effects of caspase inhibition on the progression of experimental glomerulonephritis. *Kidney international* 2003, **63**(6): 2050-2064.

279. Seery JP, Cattell V, Watt FM. Cutting edge: amelioration of kidney disease in a transgenic mouse model of lupus nephritis by administration of the caspase inhibitor carbobenzoxy-valyl-alanyl-aspartyl-(beta-o-methyl)-fluoromethylketone. *Journal of immunology (Baltimore, Md : 1950)* 2001, **167**(5): 2452-2455.

Wen S, Wang ZH, Zhang CX, Yang Y, Fan QL. Caspase-3 Promotes Diabetic Kidney Disease
 Through Gasdermin E-Mediated Progression to Secondary Necrosis During Apoptosis. *Diabetes, metabolic syndrome and obesity : targets and therapy* 2020, 13: 313-323.

- 2799 272. Belavgeni A, Meyer C, Stumpf J, Hugo C, Linkermann A. Ferroptosis and Necroptosis in the Kidney. *Cell chemical biology* 2020, **27**(4): 448-462.
- 2801
   2802 273. von Mässenhausen A, Tonnus W, Linkermann A. Cell Death Pathways Drive Necroinflammation during Acute Kidney Injury. *Nephron* 2018, **140**(2): 144-147.
- 2805 274. Guo R, Wang Y, Minto AW, Quigg RJ, Cunningham PN. Acute renal failure in endotoxemia is dependent on caspase activation. *Journal of the American Society of Nephrology : JASN* 2004, **15**(12): 3093-3102.

2808

2812

2816

2820

2823

2827

2831

- 2809 275. Herzog C, Yang C, Holmes A, Kaushal GP. zVAD-fmk prevents cisplatin-induced cleavage of autophagy proteins but impairs autophagic flux and worsens renal function. *American journal of physiology Renal physiology* 2012, **303**(8): F1239-1250.
- 2813 276. Linkermann A, Heller JO, Prókai A, Weinberg JM, De Zen F, Himmerkus N, *et al.* The RIP1-2814 kinase inhibitor necrostatin-1 prevents osmotic nephrosis and contrast-induced AKI in mice. 2815 *Journal of the American Society of Nephrology : JASN* 2013, **24**(10): 1545-1557.
- 2817 277. Brinkmann K, Waring P, Glaser SP, Wimmer V, Cottle DL, Tham MS, *et al.* BCL-XL exerts a protective role against anemia caused by radiation-induced kidney damage. *The EMBO journal* 2020, **39**(24): e105561.
- 278. Yin XM, Wang K, Gross A, Zhao Y, Zinkel S, Klocke B, *et al.* Bid-deficient mice are resistant to Fas-induced hepatocellular apoptosis. *Nature* 1999, **400**(6747): 886-891.
- DuBray BJ, Jr., Conzen KD, Upadhya GA, Gunter KL, Jia J, Knolhoff BL, *et al.* BH3-only proteins contribute to steatotic liver ischemia-reperfusion injury. *The Journal of surgical research* 2015, **194**(2): 653-658.
- 280. Selzner M, Rüdiger HA, Selzner N, Thomas DW, Sindram D, Clavien PA. Transgenic mice overexpressing human Bcl-2 are resistant to hepatic ischemia and reperfusion. *Journal of hepatology* 2002, **36**(2): 218-225.
- 283. Cursio R, Gugenheim J, Ricci JE, Crenesse D, Rostagno P, Maulon L, *et al.* Caspase inhibition protects from liver injury following ischemia and reperfusion in rats. *Transplant international : official journal of the European Society for Organ Transplantation* 2000, **13 Suppl 1:** S568-572.
- 282. Kaufmann T, Jost PJ, Pellegrini M, Puthalakath H, Gugasyan R, Gerondakis S, *et al.* Fatal hepatitis mediated by tumor necrosis factor TNFalpha requires caspase-8 and involves the BH3-only proteins Bid and Bim. *Immunity* 2009, **30**(1): 56-66.

2840 283. Riddle-Taylor E, Nagasaki K, Lopez J, Esquivel CO, Martinez OM, Krams SM. Mutations to bid cleavage sites protect hepatocytes from apoptosis after ischemia/reperfusion injury.

Transplantation 2007, 84(6): 778, 785

2842 *Transplantation* 2007, **84**(6): 778-785.

2843

284. Lauer C, Brunner T, Corazza N. The proapoptotic Bcl-2 family member Bim plays a central role during the development of virus-induced hepatitis. *Journal of immunology (Baltimore, Md: 1950)* 2012, **188**(2): 916-922.

2847

2848 285. Chen D, Ni HM, Wang L, Ma X, Yu J, Ding WX, *et al.* p53 Up-regulated Modulator of Apoptosis Induction Mediates Acetaminophen-Induced Necrosis and Liver Injury in Mice. *Hepatology* (*Baltimore*, *Md*) 2019, **69**(5): 2164-2179.

2851

286. Badmann A, Keough A, Kaufmann T, Bouillet P, Brunner T, Corazza N. Role of TRAIL and the pro-apoptotic Bcl-2 homolog Bim in acetaminophen-induced liver damage. *Cell death & disease* 2011, **2**(6): e171.

2855

287. Naim S, Fernandez-Marrero Y, de Brot S, Bachmann D, Kaufmann T. Loss of BOK Has a Minor Impact on Acetaminophen Overdose-Induced Liver Damage in Mice. *International journal of molecular sciences* 2021, **22**(6).

2859

288. Yoshida N, Iwata H, Yamada T, Sekino T, Matsuo H, Shirahashi K, *et al.* Improvement of the survival rate after rat massive hepatectomy due to the reduction of apoptosis by caspase inhibitor. *Journal of gastroenterology and hepatology* 2007, **22**(11): 2015-2021.

2863

289. Roychowdhury S, Chiang DJ, Mandal P, McMullen MR, Liu X, Cohen JI, *et al.* Inhibition of apoptosis protects mice from ethanol-mediated acceleration of early markers of CCl4 -induced fibrosis but not steatosis or inflammation. *Alcoholism, clinical and experimental research* 2012, **36**(7): 1139-1147.

2868

2869 290. Eguchi A, De Mollerat Du Jeu X, Johnson CD, Nektaria A, Feldstein AE. Liver Bid suppression for treatment of fibrosis associated with non-alcoholic steatohepatitis. *Journal of hepatology* 2016, **64**(3): 699-707.

2872

291. Higuchi H, Miyoshi H, Bronk SF, Zhang H, Dean N, Gores GJ. Bid antisense attenuates bile acidinduced apoptosis and cholestatic liver injury. *The Journal of pharmacology and experimental therapeutics* 2001, **299**(3): 866-873.

2876

2877 292. Nalapareddy P, Schüngel S, Hong JY, Manns MP, Jaeschke H, Vogel A. The BH3-only protein bid does not mediate death-receptor-induced liver injury in obstructive cholestasis. *The American journal of pathology* 2009, **175**(3): 1077-1085.

283. Kahraman A, Mott JL, Bronk SF, Werneburg NW, Barreyro FJ, Guicciardi ME, *et al.*2882 Overexpression of mcl-1 attenuates liver injury and fibrosis in the bile duct-ligated mouse.
2883 *Digestive diseases and sciences* 2009, **54**(9): 1908-1917.

2884

2885 294. Mitchell C, Mahrouf-Yorgov M, Mayeuf A, Robin MA, Mansouri A, Fromenty B, *et al.*2886 Overexpression of Bcl-2 in hepatocytes protects against injury but does not attenuate fibrosis in
2887 a mouse model of chronic cholestatic liver disease. *Laboratory investigation; a journal of*2888 *technical methods and pathology* 2011, **91**(2): 273-282.

2889

2890 295. He L, Sehrawat TS, Verma VK, Navarro-Corcuera A, Sidhu G, Mauer A, et al. XIAP Knockdown
 2891 in Alcohol-Associated Liver Disease Models Exhibits Divergent in vitro and in vivo Phenotypes
 2892 Owing to a Potential Zonal Inhibitory Role of SMAC. Frontiers in physiology 2021, 12: 664222.

2893

2894 296. Zilu S, Qian H, Haibin W, Chenxu G, Deshuai L, Qiang L, *et al.* Effects of XIAP on high fat dietinduced hepatic steatosis: a mechanism involving NLRP3 inflammasome and oxidative stress. *Aging* 2019, **11**(24): 12177-12201.

2897

2898 297. Thapaliya S, Wree A, Povero D, Inzaugarat ME, Berk M, Dixon L, *et al.* Caspase 3 inactivation protects against hepatic cell death and ameliorates fibrogenesis in a diet-induced NASH model. *Digestive diseases and sciences* 2014, **59**(6): 1197-1206.

2901

298. Weng SY, Yang CY, Li CC, Sun TP, Tung SY, Yen JJ, *et al.* Synergism between p53 and Mcl-1 in protecting from hepatic injury, fibrosis and cancer. *Journal of hepatology* 2011, **54**(4): 685-694.

2905

299. Hikita H, Kodama T, Shimizu S, Li W, Shigekawa M, Tanaka S, *et al.* Bak deficiency inhibits liver carcinogenesis: a causal link between apoptosis and carcinogenesis. *Journal of hepatology* 2012, **57**(1): 92-100.

2909

2910 300. Rabachini T, Fernandez-Marrero Y, Montani M, Loforese G, Sladky V, He Z, et al. BOK promotes chemical-induced hepatocarcinogenesis in mice. *Cell death and differentiation* 2018, 25(4): 708-720.

2913

Wree A, Johnson CD, Font-Burgada J, Eguchi A, Povero D, Karin M, *et al.* Hepatocyte-specific Bid depletion reduces tumor development by suppressing inflammation-related compensatory proliferation. *Cell death and differentiation* 2015, **22**(12): 1985-1994.

2917

Orlik J, Schüngel S, Buitrago-Molina LE, Marhenke S, Geffers R, Endig J, *et al.* The BH3-only protein BID impairs the p38-mediated stress response and promotes hepatocarcinogenesis during chronic liver injury in mice. *Hepatology (Baltimore, Md)* 2015, **62**(3): 816-828.

2921
2922 303. Kim JY, Garcia-Carbonell R, Yamachika S, Zhao P, Dhar D, Loomba R, *et al.* ER Stress Drives
2923 Lipogenesis and Steatohepatitis via Caspase-2 Activation of S1P. *Cell* 2018, 175(1): 1332924 145.e115.

2926 304. Machado MV, Michelotti GA, Jewell ML, Pereira TA, Xie G, Premont RT, *et al.* Caspase-2 2927 promotes obesity, the metabolic syndrome and nonalcoholic fatty liver disease. *Cell death & disease* 2016, **7**(2): e2096.

2925

2929

2934

2938

2943

2947

2951

2955

2959

2930 305. Barreyro FJ, Holod S, Finocchietto PV, Camino AM, Aquino JB, Avagnina A, *et al.* The pan-2931 caspase inhibitor Emricasan (IDN-6556) decreases liver injury and fibrosis in a murine model of 2932 non-alcoholic steatohepatitis. *Liver international : official journal of the International* 2933 *Association for the Study of the Liver* 2015, **35**(3): 953-966.

2935 306. Witek RP, Stone WC, Karaca FG, Syn WK, Pereira TA, Agboola KM, *et al.* Pan-caspase inhibitor VX-166 reduces fibrosis in an animal model of nonalcoholic steatohepatitis. *Hepatology* (*Baltimore, Md*) 2009, **50**(5): 1421-1430.

2939 307. Gracia-Sancho J, Manicardi N, Ortega-Ribera M, Maeso-Díaz R, Guixé-Muntet S, Fernández-1940 Iglesias A, *et al.* Emricasan Ameliorates Portal Hypertension and Liver Fibrosis in Cirrhotic Rats 2941 Through a Hepatocyte-Mediated Paracrine Mechanism. *Hepatology communications* 2019, **3**(7): 2942 987-1000.

2944 308. Eguchi A, Koyama Y, Wree A, Johnson CD, Nakamura R, Povero D, *et al.* Emricasan, a pan-2945 caspase inhibitor, improves survival and portal hypertension in a murine model of common bileduct ligation. *Journal of molecular medicine (Berlin, Germany)* 2018, **96**(6): 575-583.

2948 309. Canbay A, Feldstein A, Baskin-Bey E, Bronk SF, Gores GJ. The caspase inhibitor IDN-6556 attenuates hepatic injury and fibrosis in the bile duct ligated mouse. *The Journal of pharmacology and experimental therapeutics* 2004, **308**(3): 1191-1196.

2952 310. Garcia-Tsao G, Fuchs M, Shiffman M, Borg BB, Pyrsopoulos N, Shetty K, *et al.* Emricasan (IDN-6556) Lowers Portal Pressure in Patients With Compensated Cirrhosis and Severe Portal Hypertension. *Hepatology (Baltimore, Md)* 2019, **69**(2): 717-728.

Frenette C, Kayali Z, Mena E, Mantry PS, Lucas KJ, Neff G, *et al.* Emricasan to prevent new decompensation in patients with NASH-related decompensated cirrhosis. *Journal of hepatology* 2021, **74**(2): 274-282.

- 2960 312. Xu WF, Zhang Q, Ding CJ, Sun HY, Che Y, Huang H, *et al.* Gasdermin E-derived caspase-3 2961 inhibitors effectively protect mice from acute hepatic failure. *Acta pharmacologica Sinica* 2021, 2962 **42**(1): 68-76.
- Högstrand K, Hejll E, Sander B, Rozell B, Larsson LG, Grandien A. Inhibition of the intrinsic but not the extrinsic apoptosis pathway accelerates and drives MYC-driven tumorigenesis towards acute myeloid leukemia. *PloS one* 2012, **7**(2): e31366.

2967

2971

2975

2978

2982

2986

2990

2994

- Finch A, Prescott J, Shchors K, Hunt A, Soucek L, Dansen TB, *et al.* Bcl-xL gain of function and p19 ARF loss of function cooperate oncogenically with Myc in vivo by distinct mechanisms. *Cancer cell* 2006, **10**(2): 113-120.
- 2972 315. Swanson PJ, Kuslak SL, Fang W, Tze L, Gaffney P, Selby S, *et al.* Fatal acute lymphoblastic leukemia in mice transgenic for B cell-restricted bcl-xL and c-myc. *Journal of immunology* (*Baltimore, Md* : 1950) 2004, **172**(11): 6684-6691.
- 2976 316. Strasser A, Harris AW, Bath ML, Cory S. Novel primitive lymphoid tumours induced in transgenic mice by cooperation between myc and bcl-2. *Nature* 1990, **348**(6299): 331-333.
- 2979 317. Campbell KJ, Bath ML, Turner ML, Vandenberg CJ, Bouillet P, Metcalf D, *et al.* Elevated Mcl-2980 1 perturbs lymphopoiesis, promotes transformation of hematopoietic stem/progenitor cells, and 2981 enhances drug resistance. *Blood* 2010, **116**(17): 3197-3207.
- 2983 318. Kelly GL, Grabow S, Glaser SP, Fitzsimmons L, Aubrey BJ, Okamoto T, *et al.* Targeting of MCL-1 kills MYC-driven mouse and human lymphomas even when they bear mutations in p53.

  2985 *Genes & development* 2014, **28**(1): 58-70.
- Vandenberg CJ, Cory S. ABT-199, a new Bcl-2-specific BH3 mimetic, has in vivo efficacy against aggressive Myc-driven mouse lymphomas without provoking thrombocytopenia. *Blood* 2013, **121**(12): 2285-2288.
- 320. Kelly PN, Grabow S, Delbridge AR, Adams JM, Strasser A. Prophylactic treatment with the BH3
   2992 mimetic ABT-737 impedes Myc-driven lymphomagenesis in mice. *Cell death and differentiation* 2993 2013, 20(1): 57-63.
- 2995 321. Mason KD, Vandenberg CJ, Scott CL, Wei AH, Cory S, Huang DC, et al. In vivo efficacy of the Bcl-2 antagonist ABT-737 against aggressive Myc-driven lymphomas. Proceedings of the National Academy of Sciences of the United States of America 2008, **105**(46): 17961-17966.

- Yin K, Lee J, Liu Z, Kim H, Martin DR, Wu D, *et al.* Mitophagy protein PINK1 suppresses colon tumor growth by metabolic reprogramming via p53 activation and reducing acetyl-CoA production. *Cell death and differentiation* 2021, **28**(8): 2421-2435.
- 3003 323. Bowen ME, Mulligan AS, Sorayya A, Attardi LD. Puma- and Caspase9-mediated apoptosis is dispensable for p53-driven neural crest-based developmental defects. *Cell death and differentiation* 2021, **28**(7): 2083-2094.

3006

3009

3012

3015

3018

3022

3024

3027

3030

3033

- 3007 324. Liang J, Niu Z, Zhang B, Yu X, Zheng Y, Wang C, *et al.* p53-dependent elimination of aneuploid mitotic offspring by entosis. *Cell death and differentiation* 2021, **28**(2): 799-813.
- 3010 325. Fischer M, Steiner L, Engeland K. The transcription factor p53: not a repressor, solely an activator. *Cell cycle (Georgetown, Tex)* 2014, **13**(19): 3037-3058.
- 3013 326. Engeland K. Cell cycle arrest through indirect transcriptional repression by p53: I have a DREAM. *Cell death and differentiation* 2018, **25**(1): 114-132.
- 3016 327. Engeland K. Cell cycle regulation: p53-p21-RB signaling. *Cell death and differentiation* 2022, **29**(5): 946-960.
- 3019 328. Aubrey BJ, Kelly GL, Janic A, Herold MJ, Strasser A. How does p53 induce apoptosis and how does this relate to p53-mediated tumour suppression? *Cell death and differentiation* 2018, **25**(1): 104-113.
- 3023 329. Vogelstein B, Lane D, Levine AJ. Surfing the p53 network. *Nature* 2000, **408**(6810): 307-310.
- 3025 330. Uxa S, Castillo-Binder P, Kohler R, Stangner K, Muller GA, Engeland K. Ki-67 gene expression. *Cell death and differentiation* 2021, **28**(12): 3357-3370.
- 3028 331. Kelly PN, Grabow S, Delbridge AR, Strasser A, Adams JM. Endogenous Bcl-xL is essential for Myc-driven lymphomagenesis in mice. *Blood* 2011, **118**(24): 6380-6386.
- 3031 332. Kelly PN, Puthalakath H, Adams JM, Strasser A. Endogenous bcl-2 is not required for the development of Emu-myc-induced B-cell lymphoma. *Blood* 2007, **109**(11): 4907-4913.
- 3034 333. Grabow S, Delbridge AR, Aubrey BJ, Vandenberg CJ, Strasser A. Loss of a Single Mcl-1 Allele Inhibits MYC-Driven Lymphomagenesis by Sensitizing Pro-B Cells to Apoptosis. *Cell reports* 2016, **14**(10): 2337-2347.

- 3038 334. Xiang Z, Luo H, Payton JE, Cain J, Ley TJ, Opferman JT, *et al.* Mcl1 haploinsufficiency protects mice from Myc-induced acute myeloid leukemia. *The Journal of clinical investigation* 2010, **120**(6): 2109-2118.
- 3042 335. Grabow S, Kelly GL, Delbridge AR, Kelly PN, Bouillet P, Adams JM, *et al.* Critical B-lymphoid cell intrinsic role of endogenous MCL-1 in c-MYC-induced lymphomagenesis. *Cell death & disease* 2016, **7**(3): e2132.

3045

3049

3053

3056

3060

3064

3068

3072

- 336. Grabow S, Delbridge AR, Valente LJ, Strasser A. MCL-1 but not BCL-XL is critical for the development and sustained expansion of thymic lymphoma in p53-deficient mice. *Blood* 2014, **124**(26): 3939-3946.
- 3050 337. Grabow S, Waring P, Happo L, Cook M, Mason KD, Kelly PN, *et al.* Pharmacological blockade of Bcl-2, Bcl-x(L) and Bcl-w by the BH3 mimetic ABT-737 has only minor impact on tumour development in p53-deficient mice. *Cell death and differentiation* 2012, **19**(4): 623-632.
- 3054 338. Beverly LJ, Varmus HE. MYC-induced myeloid leukemogenesis is accelerated by all six members of the antiapoptotic BCL family. *Oncogene* 2009, **28**(9): 1274-1279.
- 339. Glaser SP, Lee EF, Trounson E, Bouillet P, Wei A, Fairlie WD, *et al.* Anti-apoptotic Mcl-1 is essential for the development and sustained growth of acute myeloid leukemia. *Genes & development* 2012, **26**(2): 120-125.
- 3061 340. Kotschy A, Szlavik Z, Murray J, Davidson J, Maragno AL, Le Toumelin-Braizat G, *et al.* The MCL1 inhibitor S63845 is tolerable and effective in diverse cancer models. *Nature* 2016, 538(7626): 477-482.
- 341. Adams CM, Kim AS, Mitra R, Choi JK, Gong JZ, Eischen CM. BCL-W has a fundamental role in B cell survival and lymphomagenesis. *The Journal of clinical investigation* 2017, **127**(2): 635-650.
- 3069 342. Eischen CM, Roussel MF, Korsmeyer SJ, Cleveland JL. Bax loss impairs Myc-induced apoptosis and circumvents the selection of p53 mutations during Myc-mediated lymphomagenesis.

  3071 *Molecular and cellular biology* 2001, **21**(22): 7653-7662.
- 3073 343. Egle A, Harris AW, Bouillet P, Cory S. Bim is a suppressor of Myc-induced mouse B cell leukemia. *Proceedings of the National Academy of Sciences of the United States of America* 2004, **101**(16): 6164-6169.

- 3077 344. Delbridge AR, Grabow S, Bouillet P, Adams JM, Strasser A. Functional antagonism between pro-apoptotic BIM and anti-apoptotic BCL-XL in MYC-induced lymphomagenesis. *Oncogene* 2015, **34**(14): 1872-1876.
- 3081 345. Frenzel A, Labi V, Chmelewskij W, Ploner C, Geley S, Fiegl H, *et al.* Suppression of B-cell lymphomagenesis by the BH3-only proteins Bmf and Bad. *Blood* 2010, **115**(5): 995-1005.

3083

3087

3091

3095

3098

3102

3106

3109

- 3084 346. Hemann MT, Zilfou JT, Zhao Z, Burgess DJ, Hannon GJ, Lowe SW. Suppression of tumorigenesis by the p53 target PUMA. *Proceedings of the National Academy of Sciences of the United States of America* 2004, **101**(25): 9333-9338.
- 3088 347. Michalak EM, Jansen ES, Happo L, Cragg MS, Tai L, Smyth GK, *et al.* Puma and to a lesser extent Noxa are suppressors of Myc-induced lymphomagenesis. *Cell death and differentiation* 2009, **16**(5): 684-696.
- 3092 348. Garrison SP, Jeffers JR, Yang C, Nilsson JA, Hall MA, Rehg JE, *et al.* Selection against PUMA gene expression in Myc-driven B-cell lymphomagenesis. *Molecular and cellular biology* 2008, **28**(17): 5391-5402.
- 3096 349. Mérino D, Strasser A, Bouillet P. Bim must be able to engage all pro-survival Bcl-2 family members for efficient tumor suppression. *Oncogene* 2012, **31**(28): 3392-3396.
- 350. Shang Q, Zhang D, Guo C, Lin Q, Guo Z, Deng C. Potential synergism of Bim with p53 in mice with Myc-induced lymphoma in a mouse lymphoma model. *Molecular medicine reports* 2012, 5(6): 1401-1408.
- 3103 351. Delbridge AR, Pang SH, Vandenberg CJ, Grabow S, Aubrey BJ, Tai L, *et al.* RAG-induced DNA lesions activate proapoptotic BIM to suppress lymphomagenesis in p53-deficient mice. *The Journal of experimental medicine* 2016, **213**(10): 2039-2048.
- 352. Knudson CM, Johnson GM, Lin Y, Korsmeyer SJ. Bax accelerates tumorigenesis in p53-deficient mice. *Cancer research* 2001, **61**(2): 659-665.
- 353. Valente LJ, Grabow S, Vandenberg CJ, Strasser A, Janic A. Combined loss of PUMA and p21 accelerates c-MYC-driven lymphoma development considerably less than loss of one allele of p53. *Oncogene* 2016, **35**(29): 3866-3871.
- 354. Happo L, Phipson B, Smyth GK, Strasser A, Scott CL. Neither loss of Bik alone, nor combined loss of Bik and Noxa, accelerate murine lymphoma development or render lymphoma cells resistant to DNA damaging drugs. *Cell death & disease* 2012, **3**(5): e306.

3117 355. 3118 Ho LH, Taylor R, Dorstyn L, Cakouros D, Bouillet P, Kumar S. A tumor suppressor function for caspase-2. Proceedings of the National Academy of Sciences of the United States of America 3119 3120 2009, **106**(13): 5336-5341. 3121 Scott CL, Schuler M, Marsden VS, Egle A, Pellegrini M, Nesic D, et al. Apaf-1 and caspase-9 3122 356. do not act as tumor suppressors in myc-induced lymphomagenesis or mouse embryo fibroblast 3123 transformation. *The Journal of cell biology* 2004, **164**(1): 89-96. 3124 3125 3126 357. Guirguis AA, Slape CI, Failla LM, Saw J, Tremblay CS, Powell DR, et al. PUMA promotes 3127 apoptosis of hematopoietic progenitors driving leukemic progression in a mouse model of myelodysplasia. Cell death and differentiation 2016, 23(6): 1049-1059. 3128 3129 358. 3130 Michalak EM, Vandenberg CJ, Delbridge AR, Wu L, Scott CL, Adams JM, et al. Apoptosispromoted tumorigenesis: gamma-irradiation-induced thymic lymphomagenesis requires Puma-3131 3132 driven leukocyte death. Genes & development 2010, 24(15): 1608-1613. 3133 3134 359. Labi V, Erlacher M, Krumschnabel G, Manzl C, Tzankov A, Pinon J, et al. Apoptosis of leukocytes triggered by acute DNA damage promotes lymphoma formation. Genes & 3135 development 2010, 24(15): 1602-1607. 3136 3137 360. Slinger E, Wensveen FM, Guikema JE, Kater AP, Eldering E. Chronic lymphocytic leukemia 3138 development is accelerated in mice with deficiency of the pro-apoptotic regulator NOXA. 3139 Haematologica 2016, **101**(9): e374-377. 3140 3141 Katz SG, Labelle JL, Meng H, Valeriano RP, Fisher JK, Sun H, et al. Mantle cell lymphoma in 3142 361. cyclin D1 transgenic mice with Bim-deficient B cells. *Blood* 2014, **123**(6): 884-893. 3143 3144 362. Anstee NS, Bilardi RA, Ng AP, Xu Z, Robati M, Vandenberg CJ, et al. Impact of elevated anti-3145 apoptotic MCL-1 and BCL-2 on the development and treatment of MLL-AF9 AML in mice. Cell 3146 death and differentiation 2019, **26**(7): 1316-1331. 3147 3148 3149 363. Vandenberg CJ, Waring P, Strasser A, Cory S. Plasmacytomagenesis in Eμ-v-abl transgenic mice is accelerated when apoptosis is restrained. *Blood* 2014, **124**(7): 1099-1109. 3150 3151

Spinner S, Crispatzu G, Yi JH, Munkhbaatar E, Mayer P, Höckendorf U, et al. Re-activation of

mitochondrial apoptosis inhibits T-cell lymphoma survival and treatment resistance. Leukemia

3152

3153

3154

3155

364.

2016, **30**(7): 1520-1530.

- 3156 365. Puccini J, Shalini S, Voss AK, Gatei M, Wilson CH, Hiwase DK, *et al.* Loss of caspase-2 augments lymphomagenesis and enhances genomic instability in Atm-deficient mice.
- 3158 Proceedings of the National Academy of Sciences of the United States of America 2013, **110**(49):
- 3159 19920-19925.

366. Fava LL, Schuler F, Sladky V, Haschka MD, Soratroi C, Eiterer L, et al. The PIDDosome activates p53 in response to supernumerary centrosomes. *Genes & development* 2017, **31**(1): 34-45.

3164

3165 367. Lachowiez C, DiNardo CD, Konopleva M. Venetoclax in acute myeloid leukemia - current and future directions. *Leukemia & lymphoma* 2020, **61**(6): 1313-1322.

3167

368. Nechiporuk T, Kurtz SE, Nikolova O, Liu T, Jones CL, D'Alessandro A, *et al.* The TP53 Apoptotic Network Is a Primary Mediator of Resistance to BCL2 Inhibition in AML Cells. *Cancer discovery* 2019, **9**(7): 910-925.

3171

369. Bosc C, Saland E, Bousard A, Gadaud N, Sabatier M, Cognet G, *et al.* Mitochondrial inhibitors circumvent adaptive resistance to venetoclax and cytarabine combination therapy in acute myeloid leukemia. *Nature cancer* 2021, **2**(11): 1204-1223.

3175

370. Thijssen R, Diepstraten ST, Moujalled D, Chew E, Flensburg C, Shi MX, *et al.* Intact TP-53 function is essential for sustaining durable responses to BH3-mimetic drugs in leukemias. *Blood* 2021, **137**(20): 2721-2735.

3179

3180 371. Thomalla D, Beckmann L, Grimm C, Oliverio M, Meder L, Herling CD, *et al.* Deregulation and epigenetic modification of BCL2-family genes cause resistance to venetoclax in hematologic malignancies. *Blood* 2022, **140**(20): 2113-2126.

3183

372. Jilg S, Reidel V, Müller-Thomas C, König J, Schauwecker J, Höckendorf U, *et al.* Blockade of BCL-2 proteins efficiently induces apoptosis in progenitor cells of high-risk myelodysplastic syndromes patients. *Leukemia* 2016, **30**(1): 112-123.

3187

373. Ganan-Gomez I, Yang H, Ma F, Montalban-Bravo G, Thongon N, Marchica V, *et al.* Stem cell architecture drives myelodysplastic syndrome progression and predicts response to venetoclax-based therapy. *Nature medicine* 2022, **28**(3): 557-567.

3191

374. Jilg S, Hauch RT, Kauschinger J, Buschhorn L, Odinius TO, Dill V, *et al.* Venetoclax with azacitidine targets refractory MDS but spares healthy hematopoiesis at tailored dose. *Experimental hematology & oncology* 2019, **8:** 9.

375. Jager R, Herzer U, Schenkel J, Weiher H. Overexpression of Bcl-2 inhibits alveolar cell apoptosis 3196 during involution and accelerates c-myc-induced tumorigenesis of the mammary gland in 3197 transgenic mice. Oncogene 1997, **15**(15): 1787-1795. 3198

3199

3200 376. Shibata MA, Liu ML, Knudson MC, Shibata E, Yoshidome K, Bandey T, et al. Haploid loss of bax leads to accelerated mammary tumor development in C3(1)/SV40-TAg transgenic mice: 3201 reduction in protective apoptotic response at the preneoplastic stage. The EMBO journal 1999, 3202 **18**(10): 2692-2701. 3203

3204

3205 377. Jamerson MH, Johnson MD, Korsmeyer SJ, Furth PA, Dickson RB. Bax regulates c-Mycinduced mammary tumour apoptosis but not proliferation in MMTV-c-myc transgenic mice. 3206 *British journal of cancer* 2004, **91**(7): 1372-1379. 3207

3208

3209 378. Bean GR, Ganesan YT, Dong Y, Takeda S, Liu H, Chan PM, et al. PUMA and BIM are required for oncogene inactivation-induced apoptosis. Science signaling 2013, 6(268): ra20. 3210

3211

379. Parsons MJ, McCormick L, Janke L, Howard A, Bouchier-Hayes L, Green DR. Genetic deletion 3212 3213 of caspase-2 accelerates MMTV/c-neu-driven mammary carcinogenesis in mice. Cell death and differentiation 2013, **20**(9): 1174-1182. 3214

3215

380. Murphy KL, Kittrell FS, Gay JP, Jäger R, Medina D, Rosen JM. Bcl-2 expression delays 3216 mammary tumor development in dimethylbenz(a)anthracene-treated transgenic mice. Oncogene 3217 1999, **18**(47): 6597-6604. 3218

3219

3220 381. van der Heijden M, Zimberlin CD, Nicholson AM, Colak S, Kemp R, Meijer SL, et al. Bcl-2 is 3221 a critical mediator of intestinal transformation. *Nature communications* 2016, **7:** 10916.

3222

3223 382. Scherr AL, Gdynia G, Salou M, Radhakrishnan P, Duglova K, Heller A, et al. Bcl-xL is an oncogenic driver in colorectal cancer. Cell death & disease 2016, 7(8): e2342. 3224

3225

Qiu W, Carson-Walter EB, Kuan SF, Zhang L, Yu J. PUMA suppresses intestinal tumorigenesis 3226 383. in mice. Cancer research 2009, 69(12): 4999-5006. 3227

3228

3229 384. Muthalagu N, Junttila MR, Wiese KE, Wolf E, Morton J, Bauer B, et al. BIM is the primary 3230 mediator of MYC-induced apoptosis in multiple solid tissues. Cell reports 2014, 8(5): 1347-1353. 3231

3232

3233 385. Ramesh P, Lannagan TRM, Jackstadt R, Atencia Taboada L, Lansu N, Wirapati P, et al. BCL-XL is crucial for progression through the adenoma-to-carcinoma sequence of colorectal cancer. 3234 3235 *Cell death and differentiation* 2021, **28**(12): 3282-3296.

3236 Zeuner A, Francescangeli F, Contavalli P, Zapparelli G, Apuzzo T, Eramo A, et al. Elimination 3237 386. of quiescent/slow-proliferating cancer stem cells by Bcl-XL inhibition in non-small cell lung 3238

cancer. Cell death and differentiation 2014, 21(12): 1877-1888.

3240

3239

Colak S, Zimberlin CD, Fessler E, Hogdal L, Prasetyanti PR, Grandela CM, et al. Decreased 3241 387. mitochondrial priming determines chemoresistance of colon cancer stem cells. Cell death and 3242 differentiation 2014, **21**(7): 1170-1177. 3243

3244

3245 388. Garcia I, Crowther AJ, Gama V, Miller CR, Deshmukh M, Gershon TR. Bax deficiency prolongs 3246 cerebellar neurogenesis, accelerates medulloblastoma formation and paradoxically increases both malignancy and differentiation. Oncogene 2013, 32(18): 2304-2314. 3247

3250

3248

389. Yin C, Knudson CM, Korsmeyer SJ, Van Dyke T. Bax suppresses tumorigenesis and stimulates 3249 apoptosis in vivo. *Nature* 1997, **385**(6617): 637-640.

3251

390. Terry MR, Arya R, Mukhopadhyay A, Berrett KC, Clair PM, Witt B, et al. Caspase-2 impacts 3252 3253 lung tumorigenesis and chemotherapy response in vivo. Cell death and differentiation 2015, **22**(5): 719-730. 3254

3255

391. Munkhbaatar E, Dietzen M, Agrawal D, Anton M, Jesinghaus M, Boxberg M, et al. MCL-1 gains 3256 occur with high frequency in lung adenocarcinoma and can be targeted therapeutically. Nature 3257 communications 2020, **11**(1): 4527. 3258

3259

392. Meinhardt AL, Munkhbaatar E, Höckendorf U, Dietzen M, Dechant M, Anton M, et al. The BCL-3260 3261 2 family member BOK promotes KRAS-driven lung cancer progression in a p53-dependent manner. Oncogene 2022, **41**(9): 1376-1382. 3262

3263

393. He M, Chaurushiya MS, Webster JD, Kummerfeld S, Reja R, Chaudhuri S, et al. Intrinsic 3264 apoptosis shapes the tumor spectrum linked to inactivation of the deubiquitinase BAP1. Science 3265 3266 (New York, NY) 2019, **364**(6437): 283-285.

3267

3268 394. Carbone M, Harbour JW, Brugarolas J, Bononi A, Pagano I, Dey A, et al. Biological Mechanisms and Clinical Significance of BAP1 Mutations in Human Cancer. Cancer discovery 2020, 10(8): 3269 1103-1120. 3270

3271

3272 395. Novelli F, Bononi A, Wang Q, Bai F, Patergnani S, Kricek F, et al. BAP1 forms a trimer with 3273 HMGB1 and HDAC1 that modulates gene × environment interaction with asbestos. *Proceedings* of the National Academy of Sciences of the United States of America 2021, 118(48). 3274

- 396. Bononi A, Giorgi C, Patergnani S, Larson D, Verbruggen K, Tanji M, *et al.* BAP1 regulates IP3R3-mediated Ca(2+) flux to mitochondria suppressing cell transformation. *Nature* 2017, **546**(7659): 549-553.
- 397. Dansen TB, Whitfield J, Rostker F, Brown-Swigart L, Evan GI. Specific requirement for Bax, not Bak, in Myc-induced apoptosis and tumor suppression in vivo. *The Journal of biological chemistry* 2006, **281**(16): 10890-10895.

3283

3287

3291

3295

3298

3302

3305

3308

- 398. Radziszewska A, Schroer SA, Choi D, Tajmir P, Radulovich N, Ho JC, *et al.* Absence of caspase-3 protects pancreatic {beta}-cells from c-Myc-induced apoptosis without leading to tumor formation. *The Journal of biological chemistry* 2009, **284**(16): 10947-10956.
- 3288 399. Evan GI, Christophorou M, Lawlor EA, Ringshausen I, Prescott J, Dansen T, *et al.* Oncogenedependent tumor suppression: using the dark side of the force for cancer therapy. *Cold Spring* 3290 *Harbor symposia on quantitative biology* 2005, **70:** 263-273.
- 3292 400. Shalini S, Nikolic A, Wilson CH, Puccini J, Sladojevic N, Finnie J, *et al.* Caspase-2 deficiency accelerates chemically induced liver cancer in mice. *Cell death and differentiation* 2016, **23**(10): 1727-1736.
- Qiu W, Wang X, Leibowitz B, Yang W, Zhang L, Yu J. PUMA-mediated apoptosis drives chemical hepatocarcinogenesis in mice. *Hepatology (Baltimore, Md)* 2011, **54**(4): 1249-1258.
- Herce RH, Vail ME, Ralph L, Campbell JS, Fausto N. Bcl-2 expression inhibits liver carcinogenesis and delays the development of proliferating foci. *The American journal of pathology* 2002, **160**(5): 1555-1560.
- Vail ME, Pierce RH, Fausto N. Bcl-2 delays and alters hepatic carcinogenesis induced by transforming growth factor alpha. *Cancer research* 2001, **61**(2): 594-601.
- 3306 404. Pena JC, Rudin CM, Thompson CB. A Bcl-xL transgene promotes malignant conversion of chemically initiated skin papillomas. *Cancer research* 1998, **58**(10): 2111-2116.
- 3309 405. Schenkel J, Weiher H, Fürstenberger G, Jäger R. Suprabasal BCL-2 expression does not sensitize to chemically-induced skin cancer in transgenic mice. *Anticancer research* 2008, **28**(5a): 2825-2829.
- 3313 406. Rossiter H, Beissert S, Mayer C, Schön MP, Wienrich BG, Tschachler E, *et al.* Targeted expression of bcl-2 to murine basal epidermal keratinocytes results in paradoxical retardation of ultraviolet- and chemical-induced tumorigenesis. *Cancer research* 2001, **61**(9): 3619-3626.

3317 407. Kim DJ, Kataoka K, Sano S, Connolly K, Kiguchi K, DiGiovanni J. Targeted disruption of Bcl-3318 xL in mouse keratinocytes inhibits both UVB- and chemically induced skin carcinogenesis. 3319 *Molecular carcinogenesis* 2009, **48**(10): 873-885.

3320

3321 408. Strasser A, Whittingham S, Vaux DL, Bath ML, Adams JM, Cory S, *et al.* Enforced BCL2 expression in B-lymphoid cells prolongs antibody responses and elicits autoimmune disease. *Proceedings of the National Academy of Sciences of the United States of America* 1991, **88**(19): 8661-8665.

3325

3326 409. Bouillet P, Metcalf D, Huang DC, Tarlinton DM, Kay TW, Köntgen F, *et al.* Proapoptotic Bcl-2 relative Bim required for certain apoptotic responses, leukocyte homeostasis, and to preclude autoimmunity. *Science (New York, NY)* 1999, **286**(5445): 1735-1738.

3329

3330 410. Mason KD, Lin A, Robb L, Josefsson EC, Henley KJ, Gray DH, et al. Proapoptotic Bak and Bax
 3331 guard against fatal systemic and organ-specific autoimmune disease. Proceedings of the National
 3332 Academy of Sciences of the United States of America 2013, 110(7): 2599-2604.

3333

Scatizzi JC, Bickel E, Hutcheson J, Haines GK, 3rd, Perlman H. Bim deficiency leads to exacerbation and prolongation of joint inflammation in experimental arthritis. *Arthritis and rheumatism* 2006, **54**(10): 3182-3193.

3337

Li J, Zhang L, Zheng Y, Shao R, Liang Q, Yu W, *et al.* BAD inactivation exacerbates rheumatoid arthritis pathology by promoting survival of sublining macrophages. *eLife* 2020, **9**.

3340

Scatizzi JC, Hutcheson J, Bickel E, Haines GK, 3rd, Perlman H. Pro-apoptotic Bid is required for the resolution of the effector phase of inflammatory arthritis. *Arthritis research & therapy* 2007, **9**(3): R49.

3344

3345 414. Scatizzi JC, Hutcheson J, Pope RM, Firestein GS, Koch AE, Mavers M, *et al.* Bim-Bcl-2 3346 homology 3 mimetic therapy is effective at suppressing inflammatory arthritis through the 3347 activation of myeloid cell apoptosis. *Arthritis and rheumatism* 2010, **62**(2): 441-451.

3348

Moore CS, Hebb AL, Blanchard MM, Crocker CE, Liston P, Korneluk RG, *et al.* Increased X-linked inhibitor of apoptosis protein (XIAP) expression exacerbates experimental autoimmune encephalomyelitis (EAE). *Journal of neuroimmunology* 2008, **203**(1): 79-93.

3352

Lev N, Barhum Y, Melamed E, Offen D. Bax-ablation attenuates experimental autoimmune encephalomyelitis in mice. *Neuroscience letters* 2004, **359**(3): 139-142.

Ludwinski MW, Sun J, Hilliard B, Gong S, Xue F, Carmody RJ, *et al.* Critical roles of Bim in T cell activation and T cell-mediated autoimmune inflammation in mice. *The Journal of clinical investigation* 2009, **119**(6): 1706-1713.

3359

3360 418. Offen D, Kaye JF, Bernard O, Merims D, Coire CI, Panet H, *et al.* Mice overexpressing Bcl-2 in their neurons are resistant to myelin oligodendrocyte glycoprotein (MOG)-induced experimental autoimmune encephalomyelitis (EAE). *Journal of molecular neuroscience : MN* 2000, **15**(3): 167-176.

3364

Lagares D, Santos A, Grasberger PE, Liu F, Probst CK, Rahimi RA, et al. Targeted apoptosis of myofibroblasts with the BH3 mimetic ABT-263 reverses established fibrosis. Science translational medicine 2017, 9(420).

3368

3369 420. Sun J, Mao LQ, Polonsky KS, Ren DC. Pancreatic β-Cell Death due to Pdx-1 Deficiency Requires
 3370 Multi-BH Domain Protein Bax but Not Bak. *The Journal of biological chemistry* 2016, **291**(26):
 13529-13534.

3372

White SA, Zhang LS, Pasula DJ, Yang YHC, Luciani DS. Bax and Bak jointly control survival and dampen the early unfolded protein response in pancreatic β-cells under glucolipotoxic stress.
 Scientific reports 2020, 10(1): 10986.

3376

Krishnamurthy B, Chee J, Jhala G, Trivedi P, Catterall T, Selck C, et al. BIM Deficiency Protects
 NOD Mice From Diabetes by Diverting Thymocytes to Regulatory T Cells. Diabetes 2015, 64(9):
 3229-3238.

3380

Ren D, Sun J, Wang C, Ye H, Mao L, Cheng EH, et al. Role of BH3-only molecules Bim and Puma in β-cell death in Pdx1 deficiency. Diabetes 2014, 63(8): 2744-2750.

3383

3384 424. Ren D, Sun J, Mao L, Ye H, Polonsky KS. BH3-only molecule Bim mediates β-cell death in IRS2 deficiency. *Diabetes* 2014, 63(10): 3378-3387.

3386

Pfeiffer S, Halang L, Düssmann H, Byrne MM, Prehn J. BH3-Only protein bmf is required for the maintenance of glucose homeostasis in an in vivo model of HNF1α-MODY diabetes. *Cell death discovery* 2015, 1: 15041.

3390

3391 426. Uhlemeyer C, Muller N, Rieck M, Kuboth J, Schlegel C, Griess K, *et al.* Selective ablation of P53 in pancreatic beta cells fails to ameliorate glucose metabolism in genetic, dietary and pharmacological models of diabetes mellitus. *Mol Metab* 2023, **67:** 101650.

- Rohner L, Reinhart R, Hagmann B, Odermatt A, Babirye A, Kaufmann T, *et al.* FcɛRI cross-linking and IL-3 protect human basophils from intrinsic apoptotic stress. *The Journal of allergy and clinical immunology* 2018, **142**(5): 1647-1650.e1643.
- Reinhart R, Kaufmann T. IL-4 enhances survival of in vitro-differentiated mouse basophils through transcription-independent signaling downstream of PI3K. *Cell death & disease* 2018, **9**(7): 713.

3402

3405

3409

3414

3417

3421

3424

3428

- 3403 429. Didichenko SA, Spiegl N, Brunner T, Dahinden CA. IL-3 induces a Pim1-dependent antiapoptotic pathway in primary human basophils. *Blood* 2008, **112**(10): 3949-3958.
- 3406 430. Vassina EM, Yousefi S, Simon D, Zwicky C, Conus S, Simon HU. cIAP-2 and survivin contribute to cytokine-mediated delayed eosinophil apoptosis. *European journal of immunology* 2006, **36**(7): 1975-1984.
- Hasegawa T, Suzuki K, Sakamoto C, Ohta K, Nishiki S, Hino M, *et al.* Expression of the inhibitor of apoptosis (IAP) family members in human neutrophils: up-regulation of cIAP2 by granulocyte colony-stimulating factor and overexpression of cIAP2 in chronic neutrophilic leukemia. *Blood* 2003, **101**(3): 1164-1171.
- Moulding DA, Quayle JA, Hart CA, Edwards SW. Mcl-1 expression in human neutrophils: regulation by cytokines and correlation with cell survival. *Blood* 1998, **92**(7): 2495-2502.
- 3418 433. Dibbert B, Daigle I, Braun D, Schranz C, Weber M, Blaser K, *et al.* Role for Bcl-xL in delayed eosinophil apoptosis mediated by granulocyte-macrophage colony-stimulating factor and interleukin-5. *Blood* 1998, **92**(3): 778-783.
- 434. Liew PX, Kubes P. The Neutrophil's Role During Health and Disease. *Physiological reviews* 2019, **99**(2): 1223-1248.
- 435. Arlet JB, Ribeil JA, Guillem F, Negre O, Hazoume A, Marcion G, et al. HSP70 sequestration by
   free α-globin promotes ineffective erythropoiesis in β-thalassaemia. Nature 2014, 514(7521):
   242-246.
- 3429 436. Gastou M, Rio S, Dussiot M, Karboul N, Moniz H, Leblanc T, *et al.* The severe phenotype of Diamond-Blackfan anemia is modulated by heat shock protein 70. *Blood advances* 2017, **1**(22): 1959-1976.

Duplomb L, Rivière J, Jego G, Da Costa R, Hammann A, Racine J, *et al.* Serpin B1 defect and increased apoptosis of neutrophils in Cohen syndrome neutropenia. *Journal of molecular medicine (Berlin, Germany)* 2019, **97**(5): 633-645.

3436

3437 438. Schwulst SJ, Muenzer JT, Peck-Palmer OM, Chang KC, Davis CG, McDonough JS, *et al.* Bim siRNA decreases lymphocyte apoptosis and improves survival in sepsis. *Shock (Augusta, Ga)* 2008, **30**(2): 127-134.

3440

3441 439. Chung CS, Venet F, Chen Y, Jones LN, Wilson DC, Ayala CA, *et al.* Deficiency of Bid protein reduces sepsis-induced apoptosis and inflammation, while improving septic survival. *Shock* (*Augusta, Ga*) 2010, **34**(2): 150-161.

3444

Yan J, Zhang H, Xiang J, Zhao Y, Yuan X, Sun B, *et al.* The BH3-only protein BAD mediates
 TNFα cytotoxicity despite concurrent activation of IKK and NF-κB in septic shock. *Cell research* 2018, 28(7): 701-718.

3448

3449 441. Weber SU, Schewe JC, Lehmann LE, Müller S, Book M, Klaschik S, *et al.* Induction of Bim and Bid gene expression during accelerated apoptosis in severe sepsis. *Critical care (London, England)* 2008, **12**(5): R128.

3452

Oberholzer C, Tschoeke SK, Moldawer LL, Oberholzer A. Local thymic caspase-9 inhibition improves survival during polymicrobial sepsis in mice. *Journal of molecular medicine (Berlin, Germany)* 2006, **84**(5): 389-395.

3456

Lamkanfi M, Moreira LO, Makena P, Spierings DC, Boyd K, Murray PJ, *et al.* Caspase-7 deficiency protects from endotoxin-induced lymphocyte apoptosis and improves survival. *Blood* 2009, **113**(12): 2742-2745.

3460

Yasuda T, Takeyama Y, Ueda T, Shinzeki M, Kishi S, Sawa H, *et al.* Protective effect of caspase inhibitor on intestinal integrity in experimental severe acute pancreatitis. *The Journal of surgical research* 2007, **138**(2): 300-307.

3464

Liu Y, Chen XD, Yu J, Chi JL, Long FW, Yang HW, et al. Deletion Of XIAP reduces the severity
 of acute pancreatitis via regulation of cell death and nuclear factor-κB activity. Cell death & disease 2017, 8(3): e2685.

3468

Leucht K, Caj M, Fried M, Rogler G, Hausmann M. Impaired removal of Vβ8(+) lymphocytes aggravates colitis in mice deficient for B cell lymphoma-2-interacting mediator of cell death (Bim). Clinical and experimental immunology 2013, 173(3): 493-501.

- 3473 447. Wicki S, Gurzeler U, Corazza N, Genitsch V, Wong WW, Kaufmann T. Loss of BID Delays
   3474 FASL-Induced Cell Death of Mouse Neutrophils and Aggravates DSS-Induced Weight Loss.
   3475 International journal of molecular sciences 2018, 19(3).
- Weder B, Mozaffari M, Biedermann L, Mamie C, Moncsek A, Wang L, *et al.* BCL-2 levels do not predict azathioprine treatment response in inflammatory bowel disease, but inhibition induces lymphocyte apoptosis and ameliorates colitis in mice. *Clinical and experimental immunology* 2018, **193**(3): 346-360.

3481

3485

3489

3493

3497

3500

3504

3507

- Lutz C, Mozaffari M, Tosevski V, Caj M, Cippà P, McRae BL, *et al.* Increased lymphocyte apoptosis in mouse models of colitis upon ABT-737 treatment is dependent upon BIM expression. *Clinical and experimental immunology* 2015, **181**(2): 343-356.
- 3486 450. Dirisina R, Katzman RB, Goretsky T, Managlia E, Mittal N, Williams DB, *et al.* p53 and PUMA independently regulate apoptosis of intestinal epithelial cells in patients and mice with colitis. *Gastroenterology* 2011, **141**(3): 1036-1045.
- 3490 451. Qiu W, Carson-Walter EB, Liu H, Epperly M, Greenberger JS, Zambetti GP, *et al.* PUMA regulates intestinal progenitor cell radiosensitivity and gastrointestinal syndrome. *Cell stem cell* 2008, **2**(6): 576-583.
- 3494 452. Kirsch DG, Santiago PM, di Tomaso E, Sullivan JM, Hou WS, Dayton T, *et al.* p53 controls radiation-induced gastrointestinal syndrome in mice independent of apoptosis. *Science (New York, NY)* 2010, **327**(5965): 593-596.
- Tan S, Wei X, Song M, Tao J, Yang Y, Khatoon S, *et al.* PUMA mediates ER stress-induced apoptosis in portal hypertensive gastropathy. *Cell death & disease* 2014, **5**(3): e1128.
- 3501 454. Qiu W, Wu B, Wang X, Buchanan ME, Regueiro MD, Hartman DJ, *et al.* PUMA-mediated intestinal epithelial apoptosis contributes to ulcerative colitis in humans and mice. *The Journal of clinical investigation* 2011, **121**(5): 1722-1732.
- Wu B, Qiu W, Wang P, Yu H, Cheng T, Zambetti GP, *et al.* p53 independent induction of PUMA mediates intestinal apoptosis in response to ischaemia-reperfusion. *Gut* 2007, **56**(5): 645-654.
- 3508 456. Coopersmith CM, O'Donnell D, Gordon JI. Bcl-2 inhibits ischemia-reperfusion-induced apoptosis in the intestinal epithelium of transgenic mice. *The American journal of physiology* 1999, **276**(3): G677-686.

Damgaard RB, Fiil BK, Speckmann C, Yabal M, zur Stadt U, Bekker-Jensen S, *et al.* Disease-causing mutations in the XIAP BIR2 domain impair NOD2-dependent immune signalling. *EMBO molecular medicine* 2013, **5**(8): 1278-1295.

3515

Damgaard RB, Nachbur U, Yabal M, Wong WW, Fiil BK, Kastirr M, *et al.* The ubiquitin ligase XIAP recruits LUBAC for NOD2 signaling in inflammation and innate immunity. *Molecular cell* 2012, **46**(6): 746-758.

3519

3520 459. Yang X, Kanegane H, Nishida N, Imamura T, Hamamoto K, Miyashita R, *et al.* Clinical and genetic characteristics of XIAP deficiency in Japan. *Journal of clinical immunology* 2012, **32**(3): 411-420.

3523

3524 460. Salzer U, Hagena T, Webster DB, Grimbacher B. Sequence analysis of BIRC4/XIAP in male patients with common variable immunodeficiency. *International archives of allergy and immunology* 2008, **147**(2): 147-151.

3527

Wahida A, Muller M, Hiergeist A, Popper B, Steiger K, Branca C, *et al.* XIAP restrains TNF-driven intestinal inflammation and dysbiosis by promoting innate immune responses of Paneth and dendritic cells. *Sci Immunol* 2021, **6**(65): eabf7235.

3531

Brinkman BM, Hildebrand F, Kubica M, Goosens D, Del Favero J, Declercq W, et al. Caspase deficiency alters the murine gut microbiome. *Cell death & disease* 2011, **2**(10): e220.

3534

Ghazavi F, Huysentruyt J, De Coninck J, Kourula S, Martens S, Hassannia B, *et al.* Executioner caspases 3 and 7 are dispensable for intestinal epithelium turnover and homeostasis at steady state. *Proceedings of the National Academy of Sciences of the United States of America* 2022, **119**(6).

3539

3540 464. Galluzzi L, Brenner C, Morselli E, Touat Z, Kroemer G. Viral control of mitochondrial apoptosis. 3541 *PLoS pathogens* 2008, **4**(5): e1000018.

3542

Günther SD, Fritsch M, Seeger JM, Schiffmann LM, Snipas SJ, Coutelle M, *et al.* Cytosolic Gram-negative bacteria prevent apoptosis by inhibition of effector caspases through lipopolysaccharide. *Nature microbiology* 2020, **5**(2): 354-367.

3546

Suzuki T, Okamoto T, Katoh H, Sugiyama Y, Kusakabe S, Tokunaga M, *et al.* Infection with flaviviruses requires BCLXL for cell survival. *PLoS pathogens* 2018, **14**(9): e1007299.

Handke W, Luig C, Popovic B, Krmpotic A, Jonjic S, Brune W. Viral inhibition of BAK promotes murine cytomegalovirus dissemination to salivary glands. *Journal of virology* 2013, **87**(6): 3592-3596.

**67**(0)**.** 3392-3390

3553

3557

3561

3565

3569

3573

3577

3581

3584

- Fleming P, Kvansakul M, Voigt V, Kile BT, Kluck RM, Huang DC, *et al.* MCMV-mediated inhibition of the pro-apoptotic Bak protein is required for optimal in vivo replication. *PLoS pathogens* 2013, **9**(2): e1003192.
- Garrison SP, Thornton JA, Häcker H, Webby R, Rehg JE, Parganas E, *et al.* The p53-target gene puma drives neutrophil-mediated protection against lethal bacterial sepsis. *PLoS pathogens* 2010, **6**(12): e1001240.
- 3562 470. Andree M, Seeger JM, Schüll S, Coutelle O, Wagner-Stippich D, Wiegmann K, *et al.* BID-3563 dependent release of mitochondrial SMAC dampens XIAP-mediated immunity against Shigella. 3564 *The EMBO journal* 2014, **33**(19): 2171-2187.
- 3566 471. Stafford CA, Lawlor KE, Heim VJ, Bankovacki A, Bernardini JP, Silke J, *et al.* IAPs Regulate Distinct Innate Immune Pathways to Co-ordinate the Response to Bacterial Peptidoglycans. *Cell reports* 2018, **22**(6): 1496-1508.
- 3570 472. Margaroli C, Oberle S, Lavanchy C, Scherer S, Rosa M, Strasser A, *et al.* Role of proapoptotic BH3-only proteins in Listeria monocytogenes infection. *European journal of immunology* 2016, **46**(6): 1427-1437.
- 3574 473. Bradfute SB, Swanson PE, Smith MA, Watanabe E, McDunn JE, Hotchkiss RS, *et al.*3575 Mechanisms and consequences of ebolavirus-induced lymphocyte apoptosis. *Journal of immunology (Baltimore, Md : 1950)* 2010, **184**(1): 327-335.
- 3578 474. Yang Y, Wu Y, Meng X, Wang Z, Younis M, Liu Y, et al. SARS-CoV-2 membrane protein causes the mitochondrial apoptosis and pulmonary edema via targeting BOK. Cell death and differentiation 2022, **29**(7): 1395-1408.
- Anderson CJ, Medina CB, Barron BJ, Karvelyte L, Aaes TL, Lambertz I, *et al.* Microbes exploit death-induced nutrient release by gut epithelial cells. *Nature* 2021, **596**(7871): 262-267.
- 3585 476. Beckham JD, Tuttle KD, Tyler KL. Caspase-3 activation is required for reovirus-induced encephalitis in vivo. *Journal of neurovirology* 2010, **16**(4): 306-317.

- Fischer SF, Belz GT, Strasser A. BH3-only protein Puma contributes to death of antigen-specific T cells during shutdown of an immune response to acute viral infection. *Proceedings of the National Academy of Sciences of the United States of America* 2008, **105**(8): 3035-3040.
- Villunger A, Michalak EM, Coultas L, Müllauer F, Böck G, Ausserlechner MJ, *et al.* p53- and drug-induced apoptotic responses mediated by BH3-only proteins puma and noxa. *Science (New York, NY)* 2003, **302**(5647): 1036-1038.

3595

3599

3602

3605

3609

3612

3616

3620

3623

- Pellegrini M, Bouillet P, Robati M, Belz GT, Davey GM, Strasser A. Loss of Bim increases T cell production and function in interleukin 7 receptor-deficient mice. *The Journal of experimental medicine* 2004, **200**(9): 1189-1195.
- 3600 480. Speir M, Lawlor KE, Glaser SP, Abraham G, Chow S, Vogrin A, *et al.* Eliminating Legionella by inhibiting BCL-XL to induce macrophage apoptosis. *Nature microbiology* 2016, **1:** 15034.
- Tam BT, Yu AP, Tam EW, Monks DA, Wang XP, Pei XM, *et al.* Ablation of Bax and Bak protects skeletal muscle against pressure-induced injury. *Scientific reports* 2018, **8**(1): 3689.
- 3606 482. Stratos I, Li Z, Rotter R, Herlyn P, Mittlmeier T, Vollmar B. Inhibition of caspase mediated apoptosis restores muscle function after crush injury in rat skeletal muscle. *Apoptosis : an international journal on programmed cell death* 2012, **17**(3): 269-277.
- Teng BT, Tam EW, Benzie IF, Siu PM. Protective effect of caspase inhibition on compression-induced muscle damage. *The Journal of physiology* 2011, **589**(Pt 13): 3349-3369.
- 3613 484. Talbert EE, Smuder AJ, Min K, Kwon OS, Powers SK. Calpain and caspase-3 play required roles 3614 in immobilization-induced limb muscle atrophy. *Journal of applied physiology (Bethesda, Md*: 3615 1985) 2013, **114**(10): 1482-1489.
- Zhu S, Nagashima M, Khan MA, Yasuhara S, Kaneki M, Martyn JA. Lack of caspase-3 attenuates immobilization-induced muscle atrophy and loss of tension generation along with mitigation of apoptosis and inflammation. *Muscle & nerve* 2013, **47**(5): 711-721.
- Wang XH, Hu J, Du J, Klein JD. X-chromosome linked inhibitor of apoptosis protein inhibits muscle proteolysis in insulin-deficient mice. *Gene therapy* 2007, **14**(9): 711-720.
- Hu J, Du J, Zhang L, Price SR, Klein JD, Wang XH. XIAP reduces muscle proteolysis induced by CKD. *Journal of the American Society of Nephrology : JASN* 2010, **21**(7): 1174-1183.

- Hant PJ, Bain JR, Correa JE, Woo M, Batt J. Absence of caspase-3 protects against denervation-induced skeletal muscle atrophy. *Journal of applied physiology (Bethesda, Md : 1985)* 2009, **107**(1): 224-234.
- 3631 489. Zhu H, Pytel P, Gomez CM. Selective inhibition of caspases in skeletal muscle reverses the apoptotic synaptic degeneration in slow-channel myasthenic syndrome. *Human molecular genetics* 2014, **23**(1): 69-77.

3634

3638

3642

3647

3651

3656

3660

- 3635 490. Budinger GR, Mutlu GM, Urich D, Soberanes S, Buccellato LJ, Hawkins K, *et al.* Epithelial cell death is an important contributor to oxidant-mediated acute lung injury. *American journal of respiratory and critical care medicine* 2011, **183**(8): 1043-1054.
- He CH, Waxman AB, Lee CG, Link H, Rabach ME, Ma B, *et al.* Bcl-2-related protein A1 is an endogenous and cytokine-stimulated mediator of cytoprotection in hyperoxic acute lung injury. *The Journal of clinical investigation* 2005, **115**(4): 1039-1048.
- Métrailler-Ruchonnet I, Pagano A, Carnesecchi S, Khatib K, Herrera P, Donati Y, *et al.* Bcl-2 overexpression in type II epithelial cells does not prevent hyperoxia-induced acute lung injury in mice. *American journal of physiology Lung cellular and molecular physiology* 2010, **299**(3): L312-322.
- Gangoda L, Schenk RL, Best SA, Nedeva C, Louis C, D'Silva DB, *et al.* Absence of pro-survival A1 has no impact on inflammatory cell survival in vivo during acute lung inflammation and peritonitis. *Cell death and differentiation* 2022, **29**(1): 96-104.
- Kang HR, Cho SJ, Lee CG, Homer RJ, Elias JA. Transforming growth factor (TGF)-betal stimulates pulmonary fibrosis and inflammation via a Bax-dependent, bid-activated pathway that involves matrix metalloproteinase-12. *The Journal of biological chemistry* 2007, **282**(10): 7723-7732.
- 3657 495. Budinger GR, Mutlu GM, Eisenbart J, Fuller AC, Bellmeyer AA, Baker CM, et al. Proapoptotic Bid is required for pulmonary fibrosis. Proceedings of the National Academy of Sciences of the United States of America 2006, **103**(12): 4604-4609.
- 3661 496. Gu L, Surolia R, Larson-Casey JL, He C, Davis D, Kang J, *et al.* Targeting Cpt1a-Bcl-2 interaction modulates apoptosis resistance and fibrotic remodeling. *Cell death and differentiation* 2022, **29**(1): 118-132.
- Kuwano K, Kunitake R, Maeyama T, Hagimoto N, Kawasaki M, Matsuba T, *et al.* Attenuation of bleomycin-induced pneumopathy in mice by a caspase inhibitor. *American journal of physiology Lung cellular and molecular physiology* 2001, **280**(2): L316-325.

Wang R, Ibarra-Sunga O, Verlinski L, Pick R, Uhal BD. Abrogation of bleomycin-induced
 epithelial apoptosis and lung fibrosis by captopril or by a caspase inhibitor. *American journal of physiology Lung cellular and molecular physiology* 2000, 279(1): L143-151.

Wang HL, Akinci IO, Baker CM, Urich D, Bellmeyer A, Jain M, *et al.* The intrinsic apoptotic pathway is required for lipopolysaccharide-induced lung endothelial cell death. *Journal of immunology (Baltimore, Md : 1950)* 2007, **179**(3): 1834-1841.

3672

3676

3680

3684

3688

3693

3698

3702

- 3677 500. Zhang YX, Fan H, Shi Y, Xu ST, Yuan YF, Zheng RH, *et al.* Prevention of lung ischemia-3678 reperfusion injury by short hairpin RNA-mediated caspase-3 gene silencing. *The Journal of thoracic and cardiovascular surgery* 2010, **139**(3): 758-764.
- Wang L, Chen B, Xiong X, Chen S, Jin L, Zhu M. Necrostatin-1 Synergizes the Pan Caspase Inhibitor to Attenuate Lung Injury Induced by Ischemia Reperfusion in Rats. *Mediators of inflammation* 2020, **2020**: 7059304.
- Cooke DT, Hoyt EG, Robbins RC. Overexpression of human Bcl-2 in syngeneic rat donor lungs
   preserves posttransplant function and reduces intragraft caspase activity and interleukin-1beta
   production. *Transplantation* 2005, 79(7): 762-767.
- Quadri SM, Segall L, de Perrot M, Han B, Edwards V, Jones N, et al. Caspase inhibition improves ischemia-reperfusion injury after lung transplantation. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons 2005, 5(2): 292-299.
- Liu M, Shi L, Zou X, Zheng X, Zhang F, Ding X, et al. Caspase inhibitor zVAD-fmk protects against acute pancreatitis-associated lung injury via inhibiting inflammation and apoptosis.

  Pancreatology: official journal of the International Association of Pancreatology (IAP) [et al] 2016, 16(5): 733-738.
- Kawasaki M, Kuwano K, Hagimoto N, Matsuba T, Kunitake R, Tanaka T, et al. Protection from lethal apoptosis in lipopolysaccharide-induced acute lung injury in mice by a caspase inhibitor.
   The American journal of pathology 2000, 157(2): 597-603.
- van den Berg E, Bal SM, Kuipers MT, Matute-Bello G, Lutter R, Bos AP, *et al.* The caspase inhibitor zVAD increases lung inflammation in pneumovirus infection in mice. *Physiological reports* 2015, **3**(3).

- 507. Locatelli F, Corti S, Papadimitriou D, Fortunato F, Del Bo R, Donadoni C, *et al.* Fas small interfering RNA reduces motoneuron death in amyotrophic lateral sclerosis mice. *Annals of neurology* 2007, **62**(1): 81-92.
- 508. Petri S, Kiaei M, Wille E, Calingasan NY, Flint Beal M. Loss of Fas ligand-function improves survival in G93A-transgenic ALS mice. *Journal of the neurological sciences* 2006, **251**(1-2): 44-49.

3714

3718

3723

3728

3733

3737

3741

- 3715 509. Gowing G, Dequen F, Soucy G, Julien JP. Absence of tumor necrosis factor-alpha does not affect motor neuron disease caused by superoxide dismutase 1 mutations. *The Journal of neuroscience* : the official journal of the Society for Neuroscience 2006, **26**(44): 11397-11402.
- Tortarolo M, Vallarola A, Lidonnici D, Battaglia E, Gensano F, Spaltro G, *et al.* Lack of TNFalpha receptor type 2 protects motor neurons in a cellular model of amyotrophic lateral sclerosis and in mutant SOD1 mice but does not affect disease progression. *Journal of neurochemistry* 2015, **135**(1): 109-124.
- 3724 511. Bartsch JW, Wildeboer D, Koller G, Naus S, Rittger A, Moss ML, *et al.* Tumor necrosis factor-3725 alpha (TNF-alpha) regulates shedding of TNF-alpha receptor 1 by the metalloprotease-disintegrin 3726 ADAM8: evidence for a protease-regulated feedback loop in neuroprotection. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2010, **30**(36): 12210-12218.
- 3729 512. Bernard-Marissal N, Moumen A, Sunyach C, Pellegrino C, Dudley K, Henderson CE, *et al.*Reduced calreticulin levels link endoplasmic reticulum stress and Fas-triggered cell death in
  motoneurons vulnerable to ALS. *The Journal of neuroscience : the official journal of the Society*for Neuroscience 2012, **32**(14): 4901-4912.
- 3734 513. Guégan C, Vila M, Teismann P, Chen C, Onténiente B, Li M, *et al.* Instrumental activation of bid by caspase-1 in a transgenic mouse model of ALS. *Molecular and cellular neurosciences* 2002, **20**(4): 553-562.
- Kalovyrna N, Apokotou O, Boulekou S, Paouri E, Boutou A, Georgopoulos S. A 3'UTR modification of the TNF-α mouse gene increases peripheral TNF-α and modulates the Alzheimer-like phenotype in 5XFAD mice. *Scientific reports* 2020, **10**(1): 8670.
- Paouri E, Tzara O, Zenelak S, Georgopoulos S. Genetic Deletion of Tumor Necrosis Factor-α
   Attenuates Amyloid-β Production and Decreases Amyloid Plaque Formation and Glial Response
   in the 5XFAD Model of Alzheimer's Disease. *Journal of Alzheimer's disease : JAD* 2017, 60(1):
   165-181.

Paouri E, Tzara O, Kartalou GI, Zenelak S, Georgopoulos S. Peripheral Tumor Necrosis Factor-Alpha (TNF-α) Modulates Amyloid Pathology by Regulating Blood-Derived Immune Cells and Glial Response in the Brain of AD/TNF Transgenic Mice. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2017, 37(20): 5155-5171.

3751

Tweedie D, Ferguson RA, Fishman K, Frankola KA, Van Praag H, Holloway HW, *et al.* Tumor necrosis factor-α synthesis inhibitor 3,6'-dithiothalidomide attenuates markers of inflammation,
 Alzheimer pathology and behavioral deficits in animal models of neuroinflammation and
 Alzheimer's disease. *Journal of neuroinflammation* 2012, 9: 106.

3756

3757 518. McAlpine FE, Lee JK, Harms AS, Ruhn KA, Blurton-Jones M, Hong J, *et al.* Inhibition of soluble TNF signaling in a mouse model of Alzheimer's disease prevents pre-plaque amyloid-associated neuropathology. *Neurobiology of disease* 2009, **34**(1): 163-177.

3760

MacPherson KP, Sompol P, Kannarkat GT, Chang J, Sniffen L, Wildner ME, *et al.* Peripheral administration of the soluble TNF inhibitor XPro1595 modifies brain immune cell profiles, decreases beta-amyloid plaque load, and rescues impaired long-term potentiation in 5xFAD mice. *Neurobiology of disease* 2017, **102:** 81-95.

3765

520. Gabbita SP, Johnson MF, Kobritz N, Eslami P, Poteshkina A, Varadarajan S, *et al.* Oral TNFα
 3767 Modulation Alters Neutrophil Infiltration, Improves Cognition and Diminishes Tau and Amyloid
 3768 Pathology in the 3xTgAD Mouse Model. *PloS one* 2015, **10**(10): e0137305.

3769

Gabbita SP, Srivastava MK, Eslami P, Johnson MF, Kobritz NK, Tweedie D, et al. Early intervention with a small molecule inhibitor for tumor necrosis factor-α prevents cognitive deficits in a triple transgenic mouse model of Alzheimer's disease. *Journal of neuroinflammation* 2012, 9: 99.

3774

522. Lourenco MV, Clarke JR, Frozza RL, Bomfim TR, Forny-Germano L, Batista AF, *et al.* TNF-α
 3776 mediates PKR-dependent memory impairment and brain IRS-1 inhibition induced by Alzheimer's
 3777 β-amyloid oligomers in mice and monkeys. *Cell metabolism* 2013, **18**(6): 831-843.

3778

523. Camargos S, Scholz S, Simón-Sánchez J, Paisán-Ruiz C, Lewis P, Hernandez D, *et al.* DYT16, a novel young-onset dystonia-parkinsonism disorder: identification of a segregating mutation in the stress-response protein PRKRA. *The Lancet Neurology* 2008, **7**(3): 207-215.

3782

Bhaskar K, Maphis N, Xu G, Varvel NH, Kokiko-Cochran ON, Weick JP, et al. Microglial derived tumor necrosis factor-α drives Alzheimer's disease-related neuronal cell cycle events.
 Neurobiology of disease 2014, 62: 273-285.

- 525. Steeland S, Gorlé N, Vandendriessche C, Balusu S, Brkic M, Van Cauwenberghe C, *et al.*Counteracting the effects of TNF receptor-1 has therapeutic potential in Alzheimer's disease.

  EMBO molecular medicine 2018, **10**(4).
- He P, Zhong Z, Lindholm K, Berning L, Lee W, Lemere C, *et al.* Deletion of tumor necrosis factor death receptor inhibits amyloid beta generation and prevents learning and memory deficits in Alzheimer's mice. *The Journal of cell biology* 2007, **178**(5): 829-841.

3794

3798

3802

3807

3812

3816

3821

- Jayaraman A, Htike TT, James R, Picon C, Reynolds R. TNF-mediated neuroinflammation is linked to neuronal necroptosis in Alzheimer's disease hippocampus. *Acta Neuropathol Commun* 2021, **9**(1): 159.
- 3799 528. Xu C, Wu J, Wu Y, Ren Z, Yao Y, Chen G, *et al.* TNF-alpha-dependent neuronal necroptosis regulated in Alzheimer's disease by coordination of RIPK1-p62 complex with autophagic UVRAG. *Theranostics* 2021, **11**(19): 9452-9469.
- Montgomery SL, Mastrangelo MA, Habib D, Narrow WC, Knowlden SA, Wright TW, *et al.*Ablation of TNF-RI/RII expression in Alzheimer's disease mice leads to an unexpected enhancement of pathology: implications for chronic pan-TNF-α suppressive therapeutic strategies in the brain. *The American journal of pathology* 2011, **179**(4): 2053-2070.
- Montgomery SL, Narrow WC, Mastrangelo MA, Olschowka JA, O'Banion MK, Bowers WJ. Chronic neuron- and age-selective down-regulation of TNF receptor expression in triple-transgenic Alzheimer disease mice leads to significant modulation of amyloid- and Tau-related pathologies. *The American journal of pathology* 2013, **182**(6): 2285-2297.
- 531. Ferger B, Leng A, Mura A, Hengerer B, Feldon J. Genetic ablation of tumor necrosis factor-alpha (TNF-alpha) and pharmacological inhibition of TNF-synthesis attenuates MPTP toxicity in mouse striatum. *Journal of neurochemistry* 2004, **89**(4): 822-833.
- Sriram K, Matheson JM, Benkovic SA, Miller DB, Luster MI, O'Callaghan JP. Mice deficient in TNF receptors are protected against dopaminergic neurotoxicity: implications for Parkinson's disease. FASEB journal: official publication of the Federation of American Societies for Experimental Biology 2002, **16**(11): 1474-1476.
- Zhou QH, Sumbria R, Hui EK, Lu JZ, Boado RJ, Pardridge WM. Neuroprotection with a brainpenetrating biologic tumor necrosis factor inhibitor. *The Journal of pharmacology and* experimental therapeutics 2011, **339**(2): 618-623.
- McCoy MK, Martinez TN, Ruhn KA, Szymkowski DE, Smith CG, Botterman BR, et al. Blocking
   soluble tumor necrosis factor signaling with dominant-negative tumor necrosis factor inhibitor

- attenuates loss of dopaminergic neurons in models of Parkinson's disease. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2006, **26**(37): 9365-9375.
- Sriram K, Matheson JM, Benkovic SA, Miller DB, Luster MI, O'Callaghan JP. Deficiency of TNF receptors suppresses microglial activation and alters the susceptibility of brain regions to MPTP-induced neurotoxicity: role of TNF-alpha. FASEB journal: official publication of the Federation of American Societies for Experimental Biology 2006, 20(6): 670-682.

3835

3839

3843

3846

3850

3854

3859

3863

- Dong Y, Fischer R, Naudé PJ, Maier O, Nyakas C, Duffey M, *et al.* Essential protective role of tumor necrosis factor receptor 2 in neurodegeneration. *Proceedings of the National Academy of Sciences of the United States of America* 2016, **113**(43): 12304-12309.
- 537. Shi JQ, Wang BR, Jiang WW, Chen J, Zhu YW, Zhong LL, *et al.* Cognitive improvement with intrathecal administration of infliximab in a woman with Alzheimer's disease. *Journal of the American Geriatrics Society* 2011, **59**(6): 1142-1144.
- Tobinick EL, Gross H. Rapid cognitive improvement in Alzheimer's disease following perispinal etanercept administration. *Journal of neuroinflammation* 2008, **5:** 2.
- 3847 539. Alto LT, Chen X, Ruhn KA, Treviño I, Tansey MG. AAV-dominant negative tumor necrosis factor (DN-TNF) gene transfer to the striatum does not rescue medium spiny neurons in the YAC128 mouse model of Huntington's disease. *PloS one* 2014, **9**(5): e96544.
- 540. Cantarella G, Di Benedetto G, Puzzo D, Privitera L, Loreto C, Saccone S, et al. Neutralization of TNFSF10 ameliorates functional outcome in a murine model of Alzheimer's disease. Brain: a journal of neurology 2015, 138(Pt 1): 203-216.
- 3855 541. Uberti D, Ferrari-Toninelli G, Bonini SA, Sarnico I, Benarese M, Pizzi M, *et al.* Blockade of the tumor necrosis factor-related apoptosis inducing ligand death receptor DR5 prevents beta-amyloid neurotoxicity. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology 2007*, **32**(4): 872-880.
- Takahashi T, Tanaka M, Brannan CI, Jenkins NA, Copeland NG, Suda T, *et al.* Generalized lymphoproliferative disease in mice, caused by a point mutation in the Fas ligand. *Cell* 1994, **76**(6): 969-976.
- Landau AM, Luk KC, Jones ML, Siegrist-Johnstone R, Young YK, Kouassi E, *et al.* Defective Fas expression exacerbates neurotoxicity in a model of Parkinson's disease. *The Journal of experimental medicine* 2005, **202**(5): 575-581.

- Gao L, Brenner D, Llorens-Bobadilla E, Saiz-Castro G, Frank T, Wieghofer P, et al. Infiltration of circulating myeloid cells through CD95L contributes to neurodegeneration in mice. The Journal of experimental medicine 2015, 212(4): 469-480.
- Hayley S, Crocker SJ, Smith PD, Shree T, Jackson-Lewis V, Przedborski S, *et al.* Regulation of dopaminergic loss by Fas in a 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine model of Parkinson's disease. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2004, **24**(8): 2045-2053.

3876 3877

3883

3886

3889

3893

3897

3901

3905

546.

3878 and Parkinson's disease. *Neurobiology of disease* 2008, **31**(3): 309-315.

Betarbet R, Anderson LR, Gearing M, Hodges TR, Fritz JJ, Lah JJ, et al. Fas-associated factor 1

- 3880 547. Sul JW, Park MY, Shin J, Kim YR, Yoo SE, Kong YY, *et al.* Accumulation of the parkin substrate, FAF1, plays a key role in the dopaminergic neurodegeneration. *Human molecular genetics* 2013, **22**(8): 1558-1573.
- Rohn TT, Head E, Nesse WH, Cotman CW, Cribbs DH. Activation of caspase-8 in the Alzheimer's disease brain. *Neurobiology of disease* 2001, **8**(6): 1006-1016.
- Sánchez I, Xu CJ, Juo P, Kakizaka A, Blenis J, Yuan J. Caspase-8 is required for cell death induced by expanded polyglutamine repeats. *Neuron* 1999, **22**(3): 623-633.
- 3890 550. Yang L, Sugama S, Mischak RP, Kiaei M, Bizat N, Brouillet E, *et al.* A novel systemically active caspase inhibitor attenuates the toxicities of MPTP, malonate, and 3NP in vivo. *Neurobiology of disease* 2004, **17**(2): 250-259.
- 551. Viceconte N, Burguillos MA, Herrera AJ, De Pablos RM, Joseph B, Venero JL. Neuromelanin activates proinflammatory microglia through a caspase-8-dependent mechanism. *Journal of neuroinflammation* 2015, **12:** 5.
- Fricker M, Vilalta A, Tolkovsky AM, Brown GC. Caspase inhibitors protect neurons by enabling selective necroptosis of inflamed microglia. *The Journal of biological chemistry* 2013, **288**(13): 9145-9152.
- 3902 553. Burguillos MA, Deierborg T, Kavanagh E, Persson A, Hajji N, Garcia-Quintanilla A, *et al.*3903 Caspase signalling controls microglia activation and neurotoxicity. *Nature* 2011, **472**(7343): 319-324.

- Kavanagh E, Burguillos MA, Carrillo-Jimenez A, Oliva-Martin MJ, Santiago M, Rodhe J, *et al.* Deletion of caspase-8 in mouse myeloid cells blocks microglia pro-inflammatory activation and confers protection in MPTP neurodegeneration model. *Aging* 2015, 7(9): 673-689.
- 3910 555. Xu D, Zhao H, Jin M, Zhu H, Shan B, Geng J, *et al.* Modulating TRADD to restore cellular homeostasis and inhibit apoptosis. *Nature* 2020, **587**(7832): 133-138.

3912

3917

3920

3924

3928

3931

3935

3939

- Hartmann A, Troadec JD, Hunot S, Kikly K, Faucheux BA, Mouatt-Prigent A, et al. Caspase-8 is an effector in apoptotic death of dopaminergic neurons in Parkinson's disease, but pathway inhibition results in neuronal necrosis. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2001, **21**(7): 2247-2255.
- 3918 557. Rehker J, Rodhe J, Nesbitt RR, Boyle EA, Martin BK, Lord J, *et al.* Caspase-8, association with Alzheimer's Disease and functional analysis of rare variants. *PloS one* 2017, **12**(10): e0185777.
- Jasmin M, Ahn EH, Voutilainen MH, Fombonne J, Guix C, Viljakainen T, *et al.* Netrin-1 and its receptor DCC modulate survival and death of dopamine neurons and Parkinson's disease features. *The EMBO journal* 2021, **40**(3): e105537.
- 3925 559. Yu WR, Fehlings MG. Fas/FasL-mediated apoptosis and inflammation are key features of acute human spinal cord injury: implications for translational, clinical application. *Acta neuropathologica* 2011, **122**(6): 747-761.
- 560. Casha S, Yu WR, Fehlings MG. FAS deficiency reduces apoptosis, spares axons and improves function after spinal cord injury. *Experimental neurology* 2005, **196**(2): 390-400.
- 561. Demjen D, Klussmann S, Kleber S, Zuliani C, Stieltjes B, Metzger C, *et al.* Neutralization of CD95 ligand promotes regeneration and functional recovery after spinal cord injury. *Nature medicine* 2004, **10**(4): 389-395.
- 3936 562. Yu WR, Liu T, Fehlings TK, Fehlings MG. Involvement of mitochondrial signaling pathways in the mechanism of Fas-mediated apoptosis after spinal cord injury. *The European journal of neuroscience* 2009, **29**(1): 114-131.
- Letellier E, Kumar S, Sancho-Martinez I, Krauth S, Funke-Kaiser A, Laudenklos S, *et al.* CD95-ligand on peripheral myeloid cells activates Syk kinase to trigger their recruitment to the inflammatory site. *Immunity* 2010, **32**(2): 240-252.

- 564. Ellman DG, Lund MC, Nissen M, Nielsen PS, Sørensen C, Lester EB, *et al.* Conditional Ablation of Myeloid TNF Improves Functional Outcome and Decreases Lesion Size after Spinal Cord Injury in Mice. *Cells* 2020, **9**(11).
- 3948 565. Ziebell JM, Bye N, Semple BD, Kossmann T, Morganti-Kossmann MC. Attenuated neurological deficit, cell death and lesion volume in Fas-mutant mice is associated with altered neuroinflammation following traumatic brain injury. *Brain research* 2011, **1414:** 94-105.
- 3952 566. Yu WR, Liu T, Kiehl TR, Fehlings MG. Human neuropathological and animal model evidence 3953 supporting a role for Fas-mediated apoptosis and inflammation in cervical spondylotic 3954 myelopathy. *Brain : a journal of neurology* 2011, **134**(Pt 5): 1277-1292.
- Yang J, You Z, Kim HH, Hwang SK, Khuman J, Guo S, *et al.* Genetic analysis of the role of tumor necrosis factor receptors in functional outcome after traumatic brain injury in mice. *Journal of neurotrauma* 2010, **27**(6): 1037-1046.
- Bermpohl D, You Z, Lo EH, Kim HH, Whalen MJ. TNF alpha and Fas mediate tissue damage and functional outcome after traumatic brain injury in mice. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* 2007, **27**(11): 1806-1818.
- Longhi L, Perego C, Ortolano F, Aresi S, Fumagalli S, Zanier ER, *et al.* Tumor necrosis factor in traumatic brain injury: effects of genetic deletion of p55 or p75 receptor. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* 2013, **33**(8): 1182-1189.
- Khuman J, Meehan WP, 3rd, Zhu X, Qiu J, Hoffmann U, Zhang J, *et al.* Tumor necrosis factor alpha and Fas receptor contribute to cognitive deficits independent of cell death after concussive traumatic brain injury in mice. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism 2011, 31(2): 778-789.*
- 3975 571. Quintana A, Giralt M, Rojas S, Penkowa M, Campbell IL, Hidalgo J, *et al.* Differential role of tumor necrosis factor receptors in mouse brain inflammatory responses in cryolesion brain injury.

  3977 *Journal of neuroscience research* 2005, **82**(5): 701-716.
- Mironets E, Osei-Owusu P, Bracchi-Ricard V, Fischer R, Owens EA, Ricard J, et al. Soluble
   TNFα Signaling within the Spinal Cord Contributes to the Development of Autonomic
   Dysreflexia and Ensuing Vascular and Immune Dysfunction after Spinal Cord Injury. The
   Journal of neuroscience: the official journal of the Society for Neuroscience 2018, 38(17): 4146-4162.

3951

3955

3959

3964

3969

3974

- Baratz R, Tweedie D, Wang JY, Rubovitch V, Luo W, Hoffer BJ, et al. Transiently lowering tumor necrosis factor-α synthesis ameliorates neuronal cell loss and cognitive impairments induced by minimal traumatic brain injury in mice. Journal of neuroinflammation 2015, 12: 45.
- Chen KB, Uchida K, Nakajima H, Yayama T, Hirai T, Watanabe S, *et al.* Tumor necrosis factor α antagonist reduces apoptosis of neurons and oligodendroglia in rat spinal cord injury. *Spine* 2011, 36(17): 1350-1358.

3992

3997

4001

4005

4010

4015

4020

- 3993 575. O'Reilly ML, Mironets E, Shapiro TM, Crowther K, Collyer E, Bethea JR, *et al.* Pharmacological Inhibition of Soluble Tumor Necrosis Factor-Alpha Two Weeks after High Thoracic Spinal Cord Injury Does Not Affect Sympathetic Hyperreflexia. *Journal of neurotrauma* 2021, **38**(15): 2186-2191.
- 576. Ellman DG, Degn M, Lund MC, Clausen BH, Novrup HG, Flæng SB, et al. Genetic Ablation of
   Soluble TNF Does Not Affect Lesion Size and Functional Recovery after Moderate Spinal Cord
   Injury in Mice. Mediators of inflammation 2016, 2016: 2684098.
- 4002 577. Oshima T, Lee S, Sato A, Oda S, Hirasawa H, Yamashita T. TNF-alpha contributes to axonal sprouting and functional recovery following traumatic brain injury. *Brain research* 2009, **1290**: 102-110.
- Kim GM, Xu J, Xu J, Song SK, Yan P, Ku G, *et al.* Tumor necrosis factor receptor deletion reduces nuclear factor-kappaB activation, cellular inhibitor of apoptosis protein 2 expression, and functional recovery after traumatic spinal cord injury. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2001, **21**(17): 6617-6625.
- 579. Scherbel U, Raghupathi R, Nakamura M, Saatman KE, Trojanowski JQ, Neugebauer E, *et al.*Differential acute and chronic responses of tumor necrosis factor-deficient mice to experimental brain injury. *Proceedings of the National Academy of Sciences of the United States of America* 1999, **96**(15): 8721-8726.
- 4016 580. Cantarella G, Di Benedetto G, Scollo M, Paterniti I, Cuzzocrea S, Bosco P, *et al.* Neutralization of tumor necrosis factor-related apoptosis-inducing ligand reduces spinal cord injury damage in mice. *Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology* 2010, **35**(6): 1302-1314.
- Fang Y, Lu J, Wang X, Wu H, Mei S, Zheng J, et al. HIF-1α Mediates TRAIL-Induced Neuronal
   Apoptosis via Regulating DcR1 Expression Following Traumatic Brain Injury. Frontiers in
   cellular neuroscience 2020, 14: 192.

- 582. Sobrido-Cameán D, Barreiro-Iglesias A. Role of Caspase-8 and Fas in Cell Death After Spinal Cord Injury. Frontiers in molecular neuroscience 2018, 11: 101.
- Sung TC, Chen Z, Thuret S, Vilar M, Gage FH, Riek R, *et al.* P45 forms a complex with FADD and promotes neuronal cell survival following spinal cord injury. *PloS one* 2013, **8**(7): e69286.

4030

4034

4039

4044

4048

4053

4058

- Krajewska M, You Z, Rong J, Kress C, Huang X, Yang J, *et al.* Neuronal deletion of caspase 8 protects against brain injury in mouse models of controlled cortical impact and kainic acid-induced excitotoxicity. *PloS one* 2011, **6**(9): e24341.
- 4035 585. Ugolini G, Raoul C, Ferri A, Haenggeli C, Yamamoto Y, Salaün D, *et al.* Fas/tumor necrosis factor receptor death signaling is required for axotomy-induced death of motoneurons in vivo.

  4037 *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2003, **23**(24): 8526-8531.
- Monnier PP, D'Onofrio PM, Magharious M, Hollander AC, Tassew N, Szydlowska K, *et al.*Involvement of caspase-6 and caspase-8 in neuronal apoptosis and the regenerative failure of injured retinal ganglion cells. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2011, **31**(29): 10494-10505.
- Tezel G, Yang X, Yang J, Wax MB. Role of tumor necrosis factor receptor-1 in the death of retinal ganglion cells following optic nerve crush injury in mice. *Brain research* 2004, **996**(2): 202-212.
- Fontaine V, Mohand-Said S, Hanoteau N, Fuchs C, Pfizenmaier K, Eisel U. Neurodegenerative and neuroprotective effects of tumor Necrosis factor (TNF) in retinal ischemia: opposite roles of TNF receptor 1 and TNF receptor 2. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2002, **22**(7): Rc216.
- Nakazawa T, Nakazawa C, Matsubara A, Noda K, Hisatomi T, She H, *et al.* Tumor necrosis factor-alpha mediates oligodendrocyte death and delayed retinal ganglion cell loss in a mouse model of glaucoma. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2006, **26**(49): 12633-12641.
- Krishnan A, Kocab AJ, Zacks DN, Marshak-Rothstein A, Gregory-Ksander M. A small peptide antagonist of the Fas receptor inhibits neuroinflammation and prevents axon degeneration and retinal ganglion cell death in an inducible mouse model of glaucoma. *Journal of neuroinflammation* 2019, **16**(1): 184.
- Cueva Vargas JL, Osswald IK, Unsain N, Aurousseau MR, Barker PA, Bowie D, *et al.* Soluble
   Tumor Necrosis Factor Alpha Promotes Retinal Ganglion Cell Death in Glaucoma via Calcium-

4066 Permeable AMPA Receptor Activation. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2015, **35**(35): 12088-12102.

4068

4072

4075

4079

4083

4086

4090

4094

4098

- 4069 592. Roh M, Zhang Y, Murakami Y, Thanos A, Lee SC, Vavvas DG, *et al.* Etanercept, a widely used
   4070 inhibitor of tumor necrosis factor-α (TNF-α), prevents retinal ganglion cell loss in a rat model of
   4071 glaucoma. *PloS one* 2012, **7**(7): e40065.
- 4073 593. Yang X, Zeng Q, Tezel G. Regulation of distinct caspase-8 functions in retinal ganglion cells and astroglia in experimental glaucoma. *Neurobiology of disease* 2021, **150:** 105258.
- Tisch N, Freire-Valls A, Yerbes R, Paredes I, La Porta S, Wang X, *et al.* Caspase-8 modulates physiological and pathological angiogenesis during retina development. *The Journal of clinical investigation* 2019, **129**(12): 5092-5107.
- 4080 595. Kang TB, Ben-Moshe T, Varfolomeev EE, Pewzner-Jung Y, Yogev N, Jurewicz A, *et al.*4081 Caspase-8 serves both apoptotic and nonapoptotic roles. *Journal of immunology (Baltimore, Md*4082 : 1950) 2004, **173**(5): 2976-2984.
- Tian Y, Li H, Liu X, Xie L, Huang Z, Li W, *et al.* Pharmacological Inhibition of Caspase-8 Suppresses Inflammation-Induced Angiogenesis in the Cornea. *Biomolecules* 2020, **10**(2).
- 4087 597. Burgaletto C, Platania CBM, Di Benedetto G, Munafò A, Giurdanella G, Federico C, *et al.*4088 Targeting the miRNA-155/TNFSF10 network restrains inflammatory response in the retina in a
  4089 mouse model of Alzheimer's disease. *Cell death & disease* 2021, **12**(10): 905.
- Meng HL, Li XX, Chen YT, Yu LJ, Zhang H, Lao JM, et al. Neuronal Soluble Fas Ligand Drives
   M1-Microglia Polarization after Cerebral Ischemia. CNS neuroscience & therapeutics 2016,
   22(9): 771-781.
- 599. Niu FN, Zhang X, Hu XM, Chen J, Chang LL, Li JW, *et al.* Targeted mutation of Fas ligand gene attenuates brain inflammation in experimental stroke. *Brain, behavior, and immunity* 2012, **26**(1): 61-71.
- 4099 600. Martin-Villalba A, Herr I, Jeremias I, Hahne M, Brandt R, Vogel J, *et al.* CD95 ligand (Fas-4100 L/APO-1L) and tumor necrosis factor-related apoptosis-inducing ligand mediate ischemia-4101 induced apoptosis in neurons. *The Journal of neuroscience : the official journal of the Society for* 4102 *Neuroscience* 1999, **19**(10): 3809-3817.

- Graham EM, Sheldon RA, Flock DL, Ferriero DM, Martin LJ, O'Riordan DP, *et al.* Neonatal mice lacking functional Fas death receptors are resistant to hypoxic-ischemic brain injury.

  Neurobiology of disease 2004, **17**(1): 89-98.
- Dzietko M, Boos V, Sifringer M, Polley O, Gerstner B, Genz K, et al. A critical role for Fas/CD 95 dependent signaling pathways in the pathogenesis of hyperoxia-induced brain injury. Annals
   of neurology 2008, 64(6): 664-673.

4111

4114

4118

4126

4130

4134

4138

- 4112 603. Ullah I, Chung K, Oh J, Beloor J, Bae S, Lee SC, *et al.* Intranasal delivery of a Fas-blocking peptide attenuates Fas-mediated apoptosis in brain ischemia. *Scientific reports* 2018, **8**(1): 15041.
- 4115 604. Martin-Villalba A, Hahne M, Kleber S, Vogel J, Falk W, Schenkel J, *et al.* Therapeutic neutralization of CD95-ligand and TNF attenuates brain damage in stroke. *Cell death and differentiation* 2001, **8**(7): 679-686.
- 4119 605. Xu W, Jin W, Zhang X, Chen J, Ren C. Remote Limb Preconditioning Generates a Neuroprotective Effect by Modulating the Extrinsic Apoptotic Pathway and TRAIL-Receptors Expression. *Cellular and molecular neurobiology* 2017, **37**(1): 169-182.
- 4122
  4123 606. Cui M, Wang L, Liang X, Ma X, Liu Y, Yang M, et al. Blocking TRAIL-DR5 signaling with
  4124 soluble DR5 reduces delayed neuronal damage after transient global cerebral ischemia.
  4125 Neurobiology of disease 2010, 39(2): 138-147.
- Clausen BH, Degn M, Sivasaravanaparan M, Fogtmann T, Andersen MG, Trojanowsky MD, et
   Conditional ablation of myeloid TNF increases lesion volume after experimental stroke in mice, possibly via altered ERK1/2 signaling. Scientific reports 2016, 6: 29291.
- 4131 608. Lambertsen KL, Clausen BH, Babcock AA, Gregersen R, Fenger C, Nielsen HH, *et al.* Microglia protect neurons against ischemia by synthesis of tumor necrosis factor. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2009, **29**(5): 1319-1330.
- 4135 609. Murakami Y, Saito K, Hara A, Zhu Y, Sudo K, Niwa M, *et al.* Increases in tumor necrosis factor-4136 alpha following transient global cerebral ischemia do not contribute to neuron death in mouse 4137 hippocampus. *Journal of neurochemistry* 2005, **93**(6): 1616-1622.
- 4139 610. Bruce AJ, Boling W, Kindy MS, Peschon J, Kraemer PJ, Carpenter MK, *et al.* Altered neuronal and microglial responses to excitotoxic and ischemic brain injury in mice lacking TNF receptors. *Nature medicine* 1996, **2**(7): 788-794.

Lei B, Dawson HN, Roulhac-Wilson B, Wang H, Laskowitz DT, James ML. Tumor necrosis factor α antagonism improves neurological recovery in murine intracerebral hemorrhage. *Journal of neuroinflammation* 2013, **10:** 103.

4146

4147 612. Yli-Karjanmaa M, Clausen BH, Degn M, Novrup HG, Ellman DG, Toft-Jensen P, *et al.* Topical Administration of a Soluble TNF Inhibitor Reduces Infarct Volume After Focal Cerebral Ischemia in Mice. *Frontiers in neuroscience* 2019, **13:** 781.

4150

Madsen PM, Clausen BH, Degn M, Thyssen S, Kristensen LK, Svensson M, et al. Genetic ablation of soluble tumor necrosis factor with preservation of membrane tumor necrosis factor is associated with neuroprotection after focal cerebral ischemia. Journal of cerebral blood flow and metabolism: official journal of the International Society of Cerebral Blood Flow and Metabolism 2016, 36(9): 1553-1569.

4156

Wu MH, Huang CC, Chio CC, Tsai KJ, Chang CP, Lin NK, *et al.* Inhibition of Peripheral TNF α and Downregulation of Microglial Activation by Alpha-Lipoic Acid and Etanercept Protect Rat
 Brain Against Ischemic Stroke. *Molecular neurobiology* 2016, **53**(7): 4961-4971.

4160

4161 615. Clausen BH, Degn M, Martin NA, Couch Y, Karimi L, Ormhøj M, *et al.* Systemically administered anti-TNF therapy ameliorates functional outcomes after focal cerebral ischemia.

4163 *Journal of neuroinflammation* 2014, **11:** 203.

4164

4165 Arango-Dávila CA, Vera A, Londoño AC, Echeverri AF, Cañas F, Cardozo CF, et al. Soluble or soluble/membrane TNF-α inhibitors protect the brain from focal ischemic injury in rats. The
 4167 International journal of neuroscience 2015, 125(12): 936-940.

4168

4169 617. Lu YM, Huang JY, Wang H, Lou XF, Liao MH, Hong LJ, *et al.* Targeted therapy of brain ischaemia using Fas ligand antibody conjugated PEG-lipid nanoparticles. *Biomaterials* 2014, 4171 35(1): 530-537.

4172

4173 618. Nawashiro H, Tasaki K, Ruetzler CA, Hallenbeck JM. TNF-alpha pretreatment induces protective effects against focal cerebral ischemia in mice. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* 1997, **17**(5): 483-490.

4177

4178 619. Kanazawa T, Kurano T, Ibaraki H, Takashima Y, Suzuki T, Seta Y. Therapeutic Effects in a
4179 Transient Middle Cerebral Artery Occlusion Rat Model by Nose-To-Brain Delivery of Anti4180 TNF-Alpha siRNA with Cell-Penetrating Peptide-Modified Polymer Micelles. *Pharmaceutics*4181 2019, **11**(9).

Lin SY, Wang YY, Chang CY, Wu CC, Chen WY, Liao SL, et al. TNF-α Receptor Inhibitor
 Alleviates Metabolic and Inflammatory Changes in a Rat Model of Ischemic Stroke. Antioxidants
 (Basel, Switzerland) 2021, 10(6).

4186

4187 621. Xiaohong W, Jun Z, Hongmei G, Fan Q. CFLAR is a critical regulator of cerebral ischaemiareperfusion injury through regulating inflammation and endoplasmic reticulum (ER) stress. 4189 *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* 2019, **117:** 109155.

4190

4191 622. Taoufik E, Valable S, Müller GJ, Roberts ML, Divoux D, Tinel A, *et al.* FLIP(L) protects neurons against in vivo ischemia and in vitro glucose deprivation-induced cell death. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2007, **27**(25): 6633-6646.

4194

4195 623. Ke DQ, Chen ZY, Li ZL, Huang X, Liang H. Target inhibition of caspase-8 alleviates brain damage after subarachnoid hemorrhage. *Neural regeneration research* 2020, **15**(7): 1283-1289.

4197

4198 624. Shabanzadeh AP, D'Onofrio PM, Monnier PP, Koeberle PD. Targeting caspase-6 and caspase-8 to promote neuronal survival following ischemic stroke. *Cell death & disease* 2015, **6**(11): e1967.

4200

4201 625. Inoue S, Davis DP, Drummond JC, Cole DJ, Patel PM. The combination of isoflurane and caspase 8 inhibition results in sustained neuroprotection in rats subject to focal cerebral ischemia. Anesthesia and analgesia 2006, **102**(5): 1548-1555.

4204

4205 626. Muhammad IF, Borné Y, Melander O, Orho-Melander M, Nilsson J, Söderholm M, *et al.* FADD (Fas-Associated Protein With Death Domain), Caspase-3, and Caspase-8 and Incidence of Ischemic Stroke. *Stroke* 2018, **49**(9): 2224-2226.

4208

4209 627. Rodhe J, Burguillos MA, de Pablos RM, Kavanagh E, Persson A, Englund E, *et al.* Spatiotemporal activation of caspase-8 in myeloid cells upon ischemic stroke. *Acta Neuropathol Commun* 2016, **4**(1): 92.

4212

Taoufik E, Petit E, Divoux D, Tseveleki V, Mengozzi M, Roberts ML, *et al.* TNF receptor I sensitizes neurons to erythropoietin- and VEGF-mediated neuroprotection after ischemic and excitotoxic injury. *Proceedings of the National Academy of Sciences of the United States of America* 2008, **105**(16): 6185-6190.

4217

4218 629. Lu MO, Zhang XM, Mix E, Quezada HC, Jin T, Zhu J, et al. TNF-alpha receptor 1 deficiency 4219 enhances kainic acid-induced hippocampal injury in mice. *Journal of neuroscience research* 4220 2008, **86**(7): 1608-1614.

- 4222 630. Balosso S, Ravizza T, Perego C, Peschon J, Campbell IL, De Simoni MG, *et al.* Tumor necrosis factor-alpha inhibits seizures in mice via p75 receptors. *Annals of neurology* 2005, **57**(6): 804-812.
- 4226 631. Patel DC, Wallis G, Dahle EJ, McElroy PB, Thomson KE, Tesi RJ, et al. Hippocampal TNFα
   4227 Signaling Contributes to Seizure Generation in an Infection-Induced Mouse Model of Limbic
   4228 Epilepsy. eNeuro 2017, 4(2).

4229

4235

4239

4243

4247

4251

4256

- Marchetti L, Klein M, Schlett K, Pfizenmaier K, Eisel UL. Tumor necrosis factor (TNF)mediated neuroprotection against glutamate-induced excitotoxicity is enhanced by N-methyl-Daspartate receptor activation. Essential role of a TNF receptor 2-mediated phosphatidylinositol 3kinase-dependent NF-kappa B pathway. *The Journal of biological chemistry* 2004, **279**(31):
  32869-32881.
- 4236 633. Thompson C, Gary D, Mattson M, Mackenzie A, Robertson GS. Kainic acid-induced naip expression in the hippocampus is blocked in mice lacking TNF receptors. *Brain research Molecular brain research* 2004, **123**(1-2): 126-131.
- 4240 634. Zhang XM, Zheng XY, Sharkawi SS, Ruan Y, Amir N, Azimullah S, *et al.* Possible protecting
   4241 role of TNF-α in kainic acid-induced neurotoxicity via down-regulation of NFκB signaling
   4242 pathway. *Current Alzheimer research* 2013, **10**(6): 660-669.
- Dolga AM, Granic I, Blank T, Knaus HG, Spiess J, Luiten PG, *et al.* TNF-alpha-mediates neuroprotection against glutamate-induced excitotoxicity via NF-kappaB-dependent upregulation of K2.2 channels. *Journal of neurochemistry* 2008, **107**(4): 1158-1167.
- Ettcheto M, Junyent F, de Lemos L, Pallas M, Folch J, Beas-Zarate C, *et al.* Mice Lacking Functional Fas Death Receptors Are Protected from Kainic Acid-Induced Apoptosis in the Hippocampus. *Molecular neurobiology* 2015, **52**(1): 120-129.
- 4252 637. Papazian I, Tsoukala E, Boutou A, Karamita M, Kambas K, Iliopoulou L, *et al.* Fundamentally different roles of neuronal TNF receptors in CNS pathology: TNFR1 and IKKβ promote microglial responses and tissue injury in demyelination while TNFR2 protects against excitotoxicity in mice. *Journal of neuroinflammation* 2021, **18**(1): 222.
- 4257 638. Li T, Lu C, Xia Z, Xiao B, Luo Y. Inhibition of caspase-8 attenuates neuronal death induced by limbic seizures in a cytochrome c-dependent and Smac/DIABLO-independent way. *Brain research* 2006, **1098**(1): 204-211.

Henshall DC, Bonislawski DP, Skradski SL, Lan JQ, Meller R, Simon RP. Cleavage of bid may amplify caspase-8-induced neuronal death following focally evoked limbic seizures.

Neurobiology of disease 2001, **8**(4): 568-580.

4264

Lee P, Sata M, Lefer DJ, Factor SM, Walsh K, Kitsis RN. Fas pathway is a critical mediator of cardiac myocyte death and MI during ischemia-reperfusion in vivo. *American journal of physiology Heart and circulatory physiology* 2003, **284**(2): H456-463.

4268

4269 641. Jeremias I, Kupatt C, Martin-Villalba A, Habazettl H, Schenkel J, Boekstegers P, *et al.*4270 Involvement of CD95/Apo1/Fas in cell death after myocardial ischemia. *Circulation* 2000,
4271 **102**(8): 915-920.

4272

4273 642. Tekin D, Xi L, Kukreja RC. Genetic deletion of fas receptors or Fas ligands does not reduce infarct size after acute global ischemia-reperfusion in isolated mouse heart. *Cell biochemistry and biophysics* 2006, **44**(1): 111-117.

4276

Boisguérin P, Covinhes A, Gallot L, Barrère C, Vincent A, Busson M, et al. A novel therapeutic peptide targeting myocardial reperfusion injury. Cardiovascular research 2020, 116(3): 633-644.

4279

Shiraishi H, Toyozaki T, Tsukamoto Y, Saito T, Masuda Y, Hiroshima K, *et al.* Antibody binding to fas ligand attenuates inflammatory cell infiltration and cytokine secretion, leading to reduction of myocardial infarct areas and reperfusion injury. *Laboratory investigation; a journal of technical methods and pathology* 2002, **82**(9): 1121-1129.

4284

4285 645. Covinhes A, Gallot L, Barrère C, Vincent A, Sportouch C, Piot C, et al. Anti-apoptotic peptide
 4286 for long term cardioprotection in a mouse model of myocardial ischemia-reperfusion injury.
 4287 Scientific reports 2020, 10(1): 18116.

4288

Wang Y, Zhang H, Wang Z, Wei Y, Wang M, Liu M, *et al.* Blocking the death checkpoint protein TRAIL improves cardiac function after myocardial infarction in monkeys, pigs, and rats. *Science translational medicine* 2020, **12**(540).

4292

4293 647. Mattisson IY, Björkbacka H, Wigren M, Edsfeldt A, Melander O, Fredrikson GN, *et al.* Elevated 4294 Markers of Death Receptor-Activated Apoptosis are Associated with Increased Risk for 4295 Development of Diabetes and Cardiovascular Disease. *EBioMedicine* 2017, **26:** 187-197.

4296

4297 648. Stenemo M, Nowak C, Byberg L, Sundström J, Giedraitis V, Lind L, *et al.* Circulating proteins as predictors of incident heart failure in the elderly. *European journal of heart failure* 2018, **20**(1): 55-62.

- 4301 649. Tanner MA, Thomas TP, Grisanti LA. Death receptor 5 contributes to cardiomyocyte hypertrophy through epidermal growth factor receptor transactivation. *Journal of molecular and cellular cardiology* 2019, **136:** 1-14.
- 4305 650. Di Bartolo BA, Cartland SP, Prado-Lourenco L, Griffith TS, Gentile C, Ravindran J, *et al.* Tumor Necrosis Factor-Related Apoptosis-Inducing Ligand (TRAIL) Promotes Angiogenesis and 4307 Ischemia-Induced Neovascularization Via NADPH Oxidase 4 (NOX4) and Nitric Oxide-4308 Dependent Mechanisms. *Journal of the American Heart Association* 2015, **4**(11).

4309

4313

4317

4321

4325

4329

4334

- Hamid T, Gu Y, Ortines RV, Bhattacharya C, Wang G, Xuan YT, *et al.* Divergent tumor necrosis factor receptor-related remodeling responses in heart failure: role of nuclear factor-kappaB and inflammatory activation. *Circulation* 2009, **119**(10): 1386-1397.
- 4314 652. Zhang Y, Zhao J, Lau WB, Jiao LY, Liu B, Yuan Y, *et al.* Tumor necrosis factor-α and lymphotoxin-α mediate myocardial ischemic injury via TNF receptor 1, but are cardioprotective when activating TNF receptor 2. *PloS one* 2013, **8**(5): e60227.
- 4318 653. Kelly ML, Wang M, Crisostomo PR, Abarbanell AM, Herrmann JL, Weil BR, *et al.* TNF receptor 2, not TNF receptor 1, enhances mesenchymal stem cell-mediated cardiac protection following acute ischemia. *Shock (Augusta, Ga)* 2010, **33**(6): 602-607.
- 4322 654. Monden Y, Kubota T, Inoue T, Tsutsumi T, Kawano S, Ide T, *et al.* Tumor necrosis factor-alpha is toxic via receptor 1 and protective via receptor 2 in a murine model of myocardial infarction.

  4324 *American journal of physiology Heart and circulatory physiology* 2007, **293**(1): H743-753.
- 4326 655. Luo D, Luo Y, He Y, Zhang H, Zhang R, Li X, *et al.* Differential functions of tumor necrosis factor receptor 1 and 2 signaling in ischemia-mediated arteriogenesis and angiogenesis. *The American journal of pathology* 2006, **169**(5): 1886-1898.
- Gouweleeuw L, Wajant H, Maier O, Eisel ULM, Blankesteijn WM, Schoemaker RG. Effects of selective TNFR1 inhibition or TNFR2 stimulation, compared to non-selective TNF inhibition, on (neuro)inflammation and behavior after myocardial infarction in male mice. *Brain, behavior, and immunity* 2021, **93:** 156-171.
- 4335 657. Guo X, Yin H, Li L, Chen Y, Li J, Doan J, *et al.* Cardioprotective Role of Tumor Necrosis Factor Receptor-Associated Factor 2 by Suppressing Apoptosis and Necroptosis. *Circulation* 2017, **136**(8): 729-742.
- 4339 658. Higuchi Y, McTiernan CF, Frye CB, McGowan BS, Chan TO, Feldman AM. Tumor necrosis factor receptors 1 and 2 differentially regulate survival, cardiac dysfunction, and remodeling in

transgenic mice with tumor necrosis factor-alpha-induced cardiomyopathy. *Circulation* 2004, **109**(15): 1892-1897.

4343

4344 659. Mann DL, McMurray JJ, Packer M, Swedberg K, Borer JS, Colucci WS, *et al.* Targeted anticytokine therapy in patients with chronic heart failure: results of the Randomized Etanercept Worldwide Evaluation (RENEWAL). *Circulation* 2004, **109**(13): 1594-1602.

4347

d348 660. Deswal A, Bozkurt B, Seta Y, Parilti-Eiswirth S, Hayes FA, Blosch C, *et al.* Safety and efficacy of a soluble P75 tumor necrosis factor receptor (Enbrel, etanercept) in patients with advanced heart failure. *Circulation* 1999, **99**(25): 3224-3226.

4351

Hospital Bozkurt B, Torre-Amione G, Warren MS, Whitmore J, Soran OZ, Feldman AM, *et al.* Results of targeted anti-tumor necrosis factor therapy with etanercept (ENBREL) in patients with advanced heart failure. *Circulation* 2001, **103**(8): 1044-1047.

4355

Generali E, Carrara G, Kallikourdis M, Condorelli G, Bortoluzzi A, Scirè CA, *et al.* Risk of hospitalization for heart failure in rheumatoid arthritis patients treated with etanercept and abatacept. *Rheumatology international* 2019, **39**(2): 239-243.

4359

4360 663. Fan Q, Huang ZM, Boucher M, Shang X, Zuo L, Brinks H, *et al.* Inhibition of Fas-associated death domain-containing protein (FADD) protects against myocardial ischemia/reperfusion injury in a heart failure mouse model. *PloS one* 2013, **8**(9): e73537.

4363

4364 664. Xiao J, Moon M, Yan L, Nian M, Zhang Y, Liu C, *et al.* Cellular FLICE-inhibitory protein protects against cardiac remodelling after myocardial infarction. *Basic research in cardiology* 2012, **107**(1): 239.

4367

4368 665. Liu D, Wu H, Li YZ, Yang J, Yang J, Ding JW, *et al.* Cellular FADD-like IL-1β-converting enzyme-inhibitory protein attenuates myocardial ischemia/reperfusion injury via suppressing apoptosis and autophagy simultaneously. *Nutrition, metabolism, and cardiovascular diseases : NMCD* 2021, **31**(6): 1916-1928.

4372

4373 666. Liang Y, Lin Q, Zhu J, Li X, Fu Y, Zou X, *et al.* The caspase-8 shRNA-modified mesenchymal stem cells improve the function of infarcted heart. *Molecular and cellular biochemistry* 2014, 4375 **397**(1-2): 7-16.

4376

Fauconnier J, Meli AC, Thireau J, Roberge S, Shan J, Sassi Y, *et al.* Ryanodine receptor leak mediated by caspase-8 activation leads to left ventricular injury after myocardial ischemia-reperfusion. *Proceedings of the National Academy of Sciences of the United States of America* 2011, **108**(32): 13258-13263.

4381 4382 668. Scharner D, Rössig L, Carmona G, Chavakis E, Urbich C, Fischer A, et al. Caspase-8 is involved in neovascularization-promoting progenitor cell functions. Arteriosclerosis, thrombosis, and 4383 4384 vascular biology 2009, **29**(4): 571-578. 4385 Koshinuma S, Miyamae M, Kaneda K, Kotani J, Figueredo VM. Combination of necroptosis and 4386 669. apoptosis inhibition enhances cardioprotection against myocardial ischemia-reperfusion injury. 4387 4388 Journal of anesthesia 2014, 28(2): 235-241. 4389 4390 670. Nam SW, Liu H, Wong JZ, Feng AY, Chu G, Merchant N, et al. Cardiomyocyte apoptosis 4391 contributes to pathogenesis of cirrhotic cardiomyopathy in bile duct-ligated mice. Clinical science (London, England: 1979) 2014, 127(8): 519-526. 4392 4393 671. 4394 Toffoli B, Bernardi S, Candido R, Zacchigna S, Fabris B, Secchiero P. TRAIL shows potential cardioprotective activity. *Investigational new drugs* 2012, **30**(3): 1257-1260. 4395 4396 672. Papathanasiou S, Rickelt S, Soriano ME, Schips TG, Maier HJ, Davos CH, et al. Tumor necrosis 4397 4398 factor-α confers cardioprotection through ectopic expression of keratins K8 and K18. Nature medicine 2015, 21(9): 1076-1084. 4399 4400 673. Liu Z, Fitzgerald M, Meisinger T, Batra R, Suh M, Greene H, et al. CD95-ligand contributes to 4401 4402 abdominal aortic aneurysm progression by modulating inflammation. Cardiovascular research 4403 2019, **115**(4): 807-818. 4404 4405 674. Wencker D, Chandra M, Nguyen K, Miao W, Garantziotis S, Factor SM, et al. A mechanistic role for cardiac myocyte apoptosis in heart failure. The Journal of clinical investigation 2003, 4406 4407 **111**(10): 1497-1504. 4408 4409 675. Tanner MA, Grisanti LA. A Dual Role for Death Receptor 5 in Regulating Cardiac Fibroblast Function. Frontiers in cardiovascular medicine 2021, 8: 699102. 4410 4411 4412 676. Pircher J, Merkle M, Wörnle M, Ribeiro A, Czermak T, Stampnik Y, et al. Prothrombotic effects 4413 of tumor necrosis factor alpha in vivo are amplified by the absence of TNF-alpha receptor subtype 1 and require TNF-alpha receptor subtype 2. Arthritis research & therapy 2012, 14(5): R225. 4414 4415

Duerrschmid C, Crawford JR, Reineke E, Taffet GE, Trial J, Entman ML, et al. TNF receptor 1

signaling is critically involved in mediating angiotensin-II-induced cardiac fibrosis. Journal of

molecular and cellular cardiology 2013, **57:** 59-67.

4416

4417

4418

4419

677.

- Woods C, Marques-Lopes J, Contoreggi NH, Milner TA, Pickel VM, Wang G, et al. Tumor
   Necrosis Factor α Receptor Type 1 Activation in the Hypothalamic Paraventricular Nucleus
   Contributes to Glutamate Signaling and Angiotensin II-Dependent Hypertension. The Journal of
   neuroscience: the official journal of the Society for Neuroscience 2021, 41(6): 1349-1362.
- Li H, Tang QZ, Liu C, Moon M, Chen M, Yan L, et al. Cellular FLICE-inhibitory protein protects against cardiac remodeling induced by angiotensin II in mice. Hypertension (Dallas, Tex: 1979) 2010, 56(6): 1109-1117.

4428

4432

4436

4440

4444

4448

4453

- Jobe LJ, Meléndez GC, Levick SP, Du Y, Brower GL, Janicki JS. TNF-alpha inhibition attenuates adverse myocardial remodeling in a rat model of volume overload. *American journal of physiology Heart and circulatory physiology* 2009, 297(4): H1462-1468.
- 4433 681. Sun M, Chen M, Dawood F, Zurawska U, Li JY, Parker T, *et al.* Tumor necrosis factor-alpha mediates cardiac remodeling and ventricular dysfunction after pressure overload state. *Circulation* 2007, **115**(11): 1398-1407.
- 4437 682. Badorff C, Ruetten H, Mueller S, Stahmer M, Gehring D, Jung F, *et al.* Fas receptor signaling inhibits glycogen synthase kinase 3 beta and induces cardiac hypertrophy following pressure overload. *The Journal of clinical investigation* 2002, **109**(3): 373-381.
- Stamm C, Friehs I, Cowan DB, Moran AM, Cao-Danh H, Duebener LF, et al. Inhibition of tumor necrosis factor-alpha improves postischemic recovery of hypertrophied hearts. Circulation 2001, 104(12 Suppl 1): 1350-355.
- Miao K, Zhou L, Ba H, Li C, Gu H, Yin B, *et al.* Transmembrane tumor necrosis factor alpha
   attenuates pressure-overload cardiac hypertrophy via tumor necrosis factor receptor 2. *PLoS biology* 2020, **18**(12): e3000967.
- Mattos BR, Bonacio GF, Vitorino TR, Garcia VT, Amaral JH, Dellalibera-Joviliano R, et al.
   TNF-α inhibition decreases MMP-2 activity, reactive oxygen species formation and improves hypertensive vascular hypertrophy independent of its effects on blood pressure. Biochemical pharmacology 2020, 180: 114121.
- Giampietri C, Petrungaro S, Musumeci M, Coluccia P, Antonangeli F, De Cesaris P, et al. c-Flip overexpression reduces cardiac hypertrophy in response to pressure overload. *Journal of hypertension* 2008, 26(5): 1008-1016.
- Hsu CC, Li Y, Hsu CT, Cheng JT, Lin MH, Cheng KC, *et al.* Etanercept Ameliorates Cardiac Fibrosis in Rats with Diet-Induced Obesity. *Pharmaceuticals (Basel, Switzerland)* 2021, **14**(4).

4461 688. Di Bartolo BA, Cartland SP, Harith HH, Bobryshev YV, Schoppet M, Kavurma MM. TRAIL-4462 deficiency accelerates vascular calcification in atherosclerosis via modulation of RANKL. *PloS* 

*one* 2013, **8**(9): e74211.

4464

4465 689. Di Bartolo BA, Chan J, Bennett MR, Cartland S, Bao S, Tuch BE, *et al.* TNF-related apoptosisinducing ligand (TRAIL) protects against diabetes and atherosclerosis in Apoe <sup>-/-</sup> mice. *Diabetologia* 2011, **54**(12): 3157-3167.

4468

Watt V, Chamberlain J, Steiner T, Francis S, Crossman D. TRAIL attenuates the development of atherosclerosis in apolipoprotein E deficient mice. *Atherosclerosis* 2011, **215**(2): 348-354.

4471

4472 G91. Zadelaar AS, von der Thüsen JH, Boesten LS, Hoeben RC, Kockx MM, Versnel MA, *et al.*4473 Increased vulnerability of pre-existing atherosclerosis in ApoE-deficient mice following
4474 adenovirus-mediated Fas ligand gene transfer. *Atherosclerosis* 2005, **183**(2): 244-250.

4475

4476 692. Yang J, Sato K, Aprahamian T, Brown NJ, Hutcheson J, Bialik A, *et al.* Endothelial overexpression of Fas ligand decreases atherosclerosis in apolipoprotein E-deficient mice.

4478 *Arteriosclerosis, thrombosis, and vascular biology* 2004, **24**(8): 1466-1473.

4479

4480 693. Xanthoulea S, Thelen M, Pöttgens C, Gijbels MJ, Lutgens E, de Winther MP. Absence of p55

TNF receptor reduces atherosclerosis, but has no major effect on angiotensin II induced aneurysms in LDL receptor deficient mice. *PloS one* 2009, **4**(7): e6113.

4483

4484 694. Xanthoulea S, Gijbels MJ, van der Made I, Mujcic H, Thelen M, Vergouwe MN, *et al.* P55 tumour necrosis factor receptor in bone marrow-derived cells promotes atherosclerosis development in low-density lipoprotein receptor knock-out mice. *Cardiovascular research* 2008, **80**(2): 309-318.

4487

Zhang L, Peppel K, Sivashanmugam P, Orman ES, Brian L, Exum ST, et al. Expression of tumor necrosis factor receptor-1 in arterial wall cells promotes atherosclerosis. Arteriosclerosis, thrombosis, and vascular biology 2007, 27(5): 1087-1094.

4491

4492 696. Brånén L, Hovgaard L, Nitulescu M, Bengtsson E, Nilsson J, Jovinge S. Inhibition of tumor necrosis factor-alpha reduces atherosclerosis in apolipoprotein E knockout mice.

4494 Arteriosclerosis, thrombosis, and vascular biology 2004, **24**(11): 2137-2142.

4495

4496 697. Miyata S, Takemura G, Kosai K, Takahashi T, Esaki M, Li L, *et al.* Anti-Fas gene therapy prevents doxorubicin-induced acute cardiotoxicity through mechanisms independent of apoptosis. *The American journal of pathology* 2010, **176**(2): 687-698.

- Niu J, Azfer A, Wang K, Wang X, Kolattukudy PE. Cardiac-targeted expression of soluble fas attenuates doxorubicin-induced cardiotoxicity in mice. *The Journal of pharmacology and experimental therapeutics* 2009, **328**(3): 740-748.
- Clayton ZS, Brunt VE, Hutton DA, Casso AG, Ziemba BP, Melov S, *et al.* Tumor Necrosis Factor Alpha-Mediated Inflammation and Remodeling of the Extracellular Matrix Underlies Aortic Stiffening Induced by the Common Chemotherapeutic Agent Doxorubicin. *Hypertension* (Dallas, Tex: 1979) 2021, **77**(5): 1581-1590.

4508

4512

4516

4520

4525

4531

4535

- 4509 700. Furuichi K, Kokubo S, Hara A, Imamura R, Wang Q, Kitajima S, et al. Fas Ligand Has a Greater
   4510 Impact than TNF-α on Apoptosis and Inflammation in Ischemic Acute Kidney Injury. Nephron
   4511 extra 2012, 2(1): 27-38.
- Ko GJ, Jang HR, Huang Y, Womer KL, Liu M, Higbee E, et al. Blocking Fas ligand on leukocytes
   attenuates kidney ischemia-reperfusion injury. Journal of the American Society of Nephrology:
   JASN 2011, 22(4): 732-742.
- Hamar P, Song E, Kökény G, Chen A, Ouyang N, Lieberman J. Small interfering RNA targeting
   Fas protects mice against renal ischemia-reperfusion injury. *Proceedings of the National Academy of Sciences of the United States of America* 2004, **101**(41): 14883-14888.
- To and the American Society of Transplant Surgeons 2006, **6**(10): 2256-2267.

  Du C, Wang S, Diao H, Guan Q, Zhong R, Jevnikar AM. Increasing resistance of tubular epithelial cells to apoptosis by shRNA therapy ameliorates renal ischemia-reperfusion injury.

  American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons 2006, **6**(10): 2256-2267.
- Hou L, Chen G, Feng B, Zhang XS, Zheng XF, Xiang Y, et al. Small interfering RNA targeting TNF-α gene significantly attenuates renal ischemia-reperfusion injury in mice. Journal of Huazhong University of Science and Technology Medical sciences = Hua zhong ke ji da xue xue bao Yi xue Ying De wen ban = Huazhong keji daxue xuebao Yixue Yingdewen ban 2016, 36(5): 634-638.
- 4532 705. Adachi T, Sugiyama N, Yagita H, Yokoyama T. Renal atrophy after ischemia-reperfusion injury depends on massive tubular apoptosis induced by TNFα in the later phase. *Medical molecular morphology* 2014, **47**(4): 213-223.
- 4536 706. Choi DE, Jeong JY, Lim BJ, Na KR, Shin YT, Lee KW. Pretreatment with the tumor nerosis factor-alpha blocker etanercept attenuated ischemia-reperfusion renal injury. *Transplantation proceedings* 2009, **41**(9): 3590-3596.

4540 707. Adachi T, Sugiyama N, Gondai T, Yagita H, Yokoyama T. Blockade of Death Ligand TRAIL Inhibits Renal Ischemia Reperfusion Injury. *Acta histochemica et cytochemica* 2013, **46**(6): 161-170.

4543

4544 708. Leng X, Zhang Q, Chen Z, Wang D. Blocking TRAIL-DR5 signaling with soluble DR5 alleviates acute kidney injury in a severely burned mouse model. *International journal of clinical and experimental pathology* 2014, **7**(6): 3460-3468.

4547

Ramesh G, Reeves WB. TNF-alpha mediates chemokine and cytokine expression and renal injury in cisplatin nephrotoxicity. *The Journal of clinical investigation* 2002, **110**(6): 835-842.

4550

4551 710. Lin JS, Mamlouk O, Selamet U, Tchakarov A, Glass WF, Sheth RA, *et al.* Infliximab for the treatment of patients with checkpoint inhibitor-associated acute tubular interstitial nephritis.

4553 *Oncoimmunology* 2021, **10**(1): 1877415.

4554

4555 711. Misaki T, Yamamoto T, Suzuki S, Fukasawa H, Togawa A, Ohashi N, *et al.* Decrease in tumor necrosis factor-alpha receptor-associated death domain results from ubiquitin-dependent degradation in obstructive renal injury in rats. *The American journal of pathology* 2009, **175**(1): 74-83.

4559

4560 712. Misseri R, Meldrum DR, Dinarello CA, Dagher P, Hile KL, Rink RC, *et al.* TNF-alpha mediates obstruction-induced renal tubular cell apoptosis and proapoptotic signaling. *American journal of physiology Renal physiology* 2005, **288**(2): F406-411.

4563

4564 713. Morimoto Y, Gai Z, Tanishima H, Kawakatsu M, Itoh S, Hatamura I, *et al.* TNF-alpha deficiency accelerates renal tubular interstitial fibrosis in the late stage of ureteral obstruction. *Experimental and molecular pathology* 2008, **85**(3): 207-213.

4567

4568 714. Guo G, Morrissey J, McCracken R, Tolley T, Klahr S. Role of TNFR1 and TNFR2 receptors in tubulointerstitial fibrosis of obstructive nephropathy. *The American journal of physiology* 1999, 4570 **277**(5): F766-772.

4571

Hughes J, Johnson RJ. Role of Fas (CD95) in tubulointerstitial disease induced by unilateral ureteric obstruction. *The American journal of physiology* 1999, **277**(1): F26-32.

4574

4575 716. Zhang X, Zheng X, Sun H, Feng B, Chen G, Vladau C, *et al.* Prevention of renal ischemic injury by silencing the expression of renal caspase 3 and caspase 8. *Transplantation* 2006, **82**(12): 1728-1732.

Linkermann A, Bräsen JH, Himmerkus N, Liu S, Huber TB, Kunzendorf U, *et al.* Rip1 (receptor-interacting protein kinase 1) mediates necroptosis and contributes to renal ischemia/reperfusion injury. *Kidney international* 2012, **81**(8): 751-761.

4582

4583 718. Linkermann A, Bräsen JH, Darding M, Jin MK, Sanz AB, Heller JO, *et al.* Two independent pathways of regulated necrosis mediate ischemia-reperfusion injury. *Proceedings of the National Academy of Sciences of the United States of America* 2013, **110**(29): 12024-12029.

4586

4587 719. Sung B, Su Y, Jiang J, McLeod P, Liu W, Haig A, *et al.* Loss of receptor interacting protein kinases 3 and caspase-8 augments intrinsic apoptosis in tubular epithelial cell and promote kidney ischaemia-reperfusion injury. *Nephrology (Carlton, Vic)* 2019, **24**(6): 661-669.

4590

4591 720. Awad AS, You H, Gao T, Cooper TK, Nedospasov SA, Vacher J, *et al.* Macrophage-derived tumor necrosis factor-α mediates diabetic renal injury. *Kidney international* 2015, **88**(4): 722-733.

4594

4595 721. Omote K, Gohda T, Murakoshi M, Sasaki Y, Kazuno S, Fujimura T, *et al.* Role of the TNF pathway in the progression of diabetic nephropathy in KK-A(y) mice. *American journal of physiology Renal physiology* 2014, **306**(11): F1335-1347.

4598

4599 722. Moriwaki Y, Inokuchi T, Yamamoto A, Ka T, Tsutsumi Z, Takahashi S, *et al.* Effect of TNF-4600 alpha inhibition on urinary albumin excretion in experimental diabetic rats. *Acta diabetologica* 4601 2007, **44**(4): 215-218.

4602

Cheng D, Liang R, Huang B, Hou J, Yin J, Zhao T, et al. Tumor necrosis factor-α blockade ameliorates diabetic nephropathy in rats. Clinical kidney journal 2021, 14(1): 301-308.

4605

4606 724. Cartland SP, Erlich JH, Kavurma MM. TRAIL deficiency contributes to diabetic nephropathy in fat-fed ApoE-/- mice. *PloS one* 2014, **9**(3): e92952.

4608

4609 725. Lorz C, Benito-Martín A, Boucherot A, Ucero AC, Rastaldi MP, Henger A, *et al.* The death ligand TRAIL in diabetic nephropathy. *Journal of the American Society of Nephrology : JASN* 2008, **19**(5): 904-914.

4612

Toffoli B, Tonon F, Tisato V, Michelli A, Zauli G, Secchiero P, et al. TRAIL treatment prevents renal morphological changes and TGF-β-induced mesenchymal transition associated with diabetic nephropathy. Clinical science (London, England: 1979) 2020, 134(17): 2337-2352.

4616

Roix J, Saha S. TNF-α blockade is ineffective in animal models of established polycystic kidney disease. *BMC nephrology* 2013, **14:** 233.

4620 728. Li X, Magenheimer BS, Xia S, Johnson T, Wallace DP, Calvet JP, *et al.* A tumor necrosis factoralpha-mediated pathway promoting autosomal dominant polycystic kidney disease. *Nature medicine* 2008, **14**(8): 863-868.

4623

Tarzi RM, Sharp PE, McDaid JP, Fossati-Jimack L, Herbert PE, Pusey CD, *et al.* Mice with defective Fas ligand are protected from crescentic glomerulonephritis. *Kidney international* 2012, **81**(2): 170-178.

4627

Khan SB, Cook HT, Bhangal G, Smith J, Tam FW, Pusey CD. Antibody blockade of TNF-alpha reduces inflammation and scarring in experimental crescentic glomerulonephritis. *Kidney international* 2005, **67**(5): 1812-1820.

4631

Zaenker M, Arbach O, Helmchen U, Glorius P, Ludewig S, Braasch E. Crescentic glomerulonephritis associated with myeloperoxidase-antineutrophil-cytoplasmic antibodies: first report on the efficacy of primary anti-TNF-alpha treatment. *International journal of tissue reactions* 2004, **26**(3-4): 85-92.

4636

4637 732. Le Hir M, Haas C, Marino M, Ryffel B. Prevention of crescentic glomerulonephritis induced by anti-glomerular membrane antibody in tumor necrosis factor-deficient mice. *Laboratory investigation; a journal of technical methods and pathology* 1998, **78**(12): 1625-1631.

4640

Wen Y, Rudemiller NP, Zhang J, Robinette T, Lu X, Ren J, et al. TNF-α in T lymphocytes
 attenuates renal injury and fibrosis during nephrotoxic nephritis. American journal of physiology
 Renal physiology 2020, 318(1): F107-f116.

4644

Taubitz A, Schwarz M, Eltrich N, Lindenmeyer MT, Vielhauer V. Distinct contributions of TNF receptor 1 and 2 to TNF-induced glomerular inflammation in mice. *PloS one* 2013, **8**(7): e68167.

4647

Pfeifer E, Polz J, Grieβl S, Mostböck S, Hehlgans T, Männel DN. Mechanisms of immune complex-mediated experimental glomerulonephritis: possible role of the balance between endogenous TNF and soluble TNF receptor type 2. *European cytokine network* 2012, 23(1): 15-20.

4652

Vielhauer V, Stavrakis G, Mayadas TN. Renal cell-expressed TNF receptor 2, not receptor 1, is essential for the development of glomerulonephritis. *The Journal of clinical investigation* 2005, **115**(5): 1199-1209.

4656

4657 737. Ryffel B, Eugster H, Haas C, Le Hir M. Failure to induce anti-glomerular basement membrane glomerulonephritis in TNF alpha/beta deficient mice. *International journal of experimental pathology* 1998, **79**(6): 453-460.

4660
 4661 738. Müller MB, Hoppe JM, Bideak A, Lux M, Lindenmeyer MT, Müller S, et al. Exclusive expression of transmembrane TNF aggravates acute glomerulonephritis despite reduced leukocyte infiltration and inflammation. *Kidney international* 2019, 95(1): 75-93.

4664
 4665 739. Mahmoud MF, El Shazly SM, Barakat W. Inhibition of TNF-α protects against hepatic ischemia-reperfusion injury in rats via NF-κB dependent pathway. *Naunyn-Schmiedeberg's archives of pharmacology* 2012, **385**(5): 465-471.

Hernandez-Alejandro R, Zhang X, Croome KP, Zheng X, Parfitt J, Chen D, et al. Reduction of liver ischemia reperfusion injury by silencing of TNF-α gene with shRNA. The Journal of surgical research 2012, 176(2): 614-620.

4668

4672

4675

4679

4683

4687

4691

- Rüdiger HA, Clavien PA. Tumor necrosis factor alpha, but not Fas, mediates hepatocellular apoptosis in the murine ischemic liver. *Gastroenterology* 2002, **122**(1): 202-210.
- 4676 742. Al-Saeedi M, Steinebrunner N, Kudsi H, Halama N, Mogler C, Büchler MW, *et al.* Neutralization of CD95 ligand protects the liver against ischemia-reperfusion injury and prevents acute liver failure. *Cell death & disease* 2018, **9**(2): 132.
- Nakajima H, Mizuta N, Fujiwara I, Sakaguchi K, Ogata H, Magae J, *et al.* Blockade of the Fas/Fas ligand interaction suppresses hepatocyte apoptosis in ischemia-reperfusion rat liver. *Apoptosis : an international journal on programmed cell death* 2008, **13**(8): 1013-1021.
- Teoh N, Leclercq I, Pena AD, Farrell G. Low-dose TNF-alpha protects against hepatic ischemiareperfusion injury in mice: implications for preconditioning. *Hepatology (Baltimore, Md)* 2003, 37(1): 118-128.
- Fahrner R, Trochsler M, Corazza N, Graubardt N, Keogh A, Candinas D, *et al.* Tumor necrosis factor-related apoptosis-inducing ligand on NK cells protects from hepatic ischemia-reperfusion injury. *Transplantation* 2014, **97**(11): 1102-1109.
- Contreras JL, Vilatoba M, Eckstein C, Bilbao G, Anthony Thompson J, Eckhoff DE. Caspase-8
   and caspase-3 small interfering RNA decreases ischemia/reperfusion injury to the liver in mice.
   Surgery 2004, 136(2): 390-400.
- Kolachala VL, Palle SK, Shen M, Shenoi A, Shayakhmetov DM, Gupta NA. Influence of Fat on Differential Receptor Interacting Serine/Threonine Protein Kinase 1 Activity Leading to Apoptotic Cell Death in Murine Liver Ischemia Reperfusion Injury Through Caspase 8. *Hepatology communications* 2019, **3**(7): 925-942.

Williams CD, McGill MR, Farhood A, Jaeschke H. Fas receptor-deficient lpr mice are protected against acetaminophen hepatotoxicity due to higher glutathione synthesis and enhanced detoxification of oxidant stress. *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association* 2013, 58: 228-235.

749. Chen Q, Yan D, Zhang Q, Zhang G, Xia M, Li J, et al. Treatment of acetaminophen-induced liver failure by blocking the death checkpoint protein TRAIL. Biochimica et biophysica acta Molecular basis of disease 2020, **1866**(1): 165583.

4705

4709

4713

4718

4722

4726

4730

4734

- Tinel M, Berson A, Vadrot N, Descatoire V, Grodet A, Feldmann G, *et al.* Subliminal Fas stimulation increases the hepatotoxicity of acetaminophen and bromobenzene in mice. Hepatology (Baltimore, Md) 2004, **39**(3): 655-666.
- 51. Schattenberg JM, Nagel M, Kim YO, Kohl T, Wörns MA, Zimmermann T, et al. Increased hepatic fibrosis and JNK2-dependent liver injury in mice exhibiting hepatocyte-specific deletion of cFLIP. American journal of physiology Gastrointestinal and liver physiology 2012, 303(4): G498-506.
- Schuchmann M, Varfolomeev EE, Hermann F, Rueckert F, Strand D, Koehler H, et al. Dominant negative MORT1/FADD rescues mice from CD95 and TNF-induced liver failure. Hepatology (Baltimore, Md) 2003, 37(1): 129-135.
- Fastor
   Seino K, Setoguchi Y, Ogino T, Kayagaki N, Akiba H, Nakano H, et al. Protection against Fastor
   mediated and tumor necrosis factor receptor 1-mediated liver injury by blockade of FADD without loss of nuclear factor-kappaB activation. Annals of surgery 2001, 234(5): 681-688.
- Tiedtke C, Bangen JM, Freimuth J, Beraza N, Lambertz D, Cubero FJ, et al. Loss of caspase-8 protects mice against inflammation-related hepatocarcinogenesis but induces non-apoptotic liver injury. Gastroenterology 2011, 141(6): 2176-2187.
- 755. Ni HM, McGill MR, Chao X, Woolbright BL, Jaeschke H, Ding WX. Caspase Inhibition Prevents
   Tumor Necrosis Factor-α-Induced Apoptosis and Promotes Necrotic Cell Death in Mouse
   Hepatocytes in Vivo and in Vitro. *The American journal of pathology* 2016, **186**(10): 2623-2636.
- 4735 756. Wroblewski R, Armaka M, Kondylis V, Pasparakis M, Walczak H, Mittrücker HW, *et al.*4736 Opposing role of tumor necrosis factor receptor 1 signaling in T cell-mediated hepatitis and bacterial infection in mice. *Hepatology (Baltimore, Md)* 2016, **64**(2): 508-521.

757. Zender L, Hutker S, Liedtke C, Tillmann HL, Zender S, Mundt B, et al. Caspase 8 small interfering RNA prevents acute liver failure in mice. Proceedings of the National Academy of Sciences of the United States of America 2003, 100(13): 7797-7802.

4742

58. Schattenberg JM, Zimmermann T, Wörns M, Sprinzl MF, Kreft A, Kohl T, *et al.* Ablation of c-FLIP in hepatocytes enhances death-receptor mediated apoptosis and toxic liver injury in vivo. *Journal of hepatology* 2011, **55**(6): 1272-1280.

4746

4747 759. Lazic M, Eguchi A, Berk MP, Povero D, Papouchado B, Mulya A, *et al.* Differential regulation of inflammation and apoptosis in Fas-resistant hepatocyte-specific Bid-deficient mice. *Journal of hepatology* 2014, **61**(1): 107-115.

4750

Kaufmann T, Tai L, Ekert PG, Huang DC, Norris F, Lindemann RK, *et al.* The BH3-only protein bid is dispensable for DNA damage- and replicative stress-induced apoptosis or cell-cycle arrest.
 *Cell* 2007, **129**(2): 423-433.

4754

Hikita H, Takehara T, Kodama T, Shimizu S, Shigekawa M, Hosui A, *et al.* Delayed-onset caspase-dependent massive hepatocyte apoptosis upon Fas activation in Bak/Bax-deficient mice. *Hepatology (Baltimore, Md)* 2011, **54**(1): 240-251.

4758

4759 762. Rodriguez I, Matsuura K, Khatib K, Reed JC, Nagata S, Vassalli P. A bcl-2 transgene expressed in hepatocytes protects mice from fulminant liver destruction but not from rapid death induced by anti-Fas antibody injection. *The Journal of experimental medicine* 1996, **183**(3): 1031-1036.

4762

4763 763. Lacronique V, Mignon A, Fabre M, Viollet B, Rouquet N, Molina T, *et al.* Bcl-2 protects from lethal hepatic apoptosis induced by an anti-Fas antibody in mice. *Nature medicine* 1996, **2**(1): 80-86.

4766

Tan S, Liu X, Chen L, Wu X, Tao L, Pan X, et al. Fas/FasL mediates NF-κBp65/PUMA-modulated hepatocytes apoptosis via autophagy to drive liver fibrosis. Cell death & disease 2021,
 12(5): 474.

4770

Yan J, Xiang J, Lin Y, Ma J, Zhang J, Zhang H, *et al.* Inactivation of BAD by IKK inhibits TNFα-induced apoptosis independently of NF-κB activation. *Cell* 2013, **152**(1-2): 304-315.

4773

766. Ottina E, Sochalska M, Sgonc R, Villunger A. The BH3-only protein Bad is dispensable for TNF-mediated cell death. *Cell death & disease* 2015, **6**(1): e1611.

Woo M, Hakem A, Elia AJ, Hakem R, Duncan GS, Patterson BJ, *et al.* In vivo evidence that caspase-3 is required for Fas-mediated apoptosis of hepatocytes. *Journal of immunology* (*Baltimore*, *Md* : 1950) 1999, **163**(9): 4909-4916.

4780

4781 768. Bajt ML, Vonderfecht SL, Jaeschke H. Differential protection with inhibitors of caspase-8 and caspase-3 in murine models of tumor necrosis factor and Fas receptor-mediated hepatocellular apoptosis. *Toxicology and applied pharmacology* 2001, **175**(3): 243-252.

4784

Theng TS, Hunot S, Kuida K, Momoi T, Srinivasan A, Nicholson DW, *et al.* Deficiency in caspase-9 or caspase-3 induces compensatory caspase activation. *Nature medicine* 2000, **6**(11): 1241-1247.

4788

4789 770. Sudo K, Yamada Y, Saito K, Shimizu S, Ohashi H, Kato T, *et al.* TNF-alpha and IL-6 signals from the bone marrow derived cells are necessary for normal murine liver regeneration.

4791 *Biochimica et biophysica acta* 2008, **1782**(11): 671-679.

4792

771. Desbarats J, Newell MK. Fas engagement accelerates liver regeneration after partial hepatectomy. *Nature medicine* 2000, **6**(8): 920-923.

4795

Knight B, Yeoh GC. TNF/LTalpha double knockout mice display abnormal inflammatory and regenerative responses to acute and chronic liver injury. *Cell and tissue research* 2005, **319**(1): 61-70.

4799

4800 773. Taira K, Hiroyasu S, Shiraishi M, Muto Y, Koji T. Role of the Fas system in liver regeneration after a partial hepatectomy in rats. European surgical research Europaische chirurgische Forschung Recherches chirurgicales europeennes 2001, **33**(5-6): 334-341.

4803

Schuchmann M, Ruckert F, Garcia-Lazaro JF, Karg A, Burg J, Knorr N, *et al.* MORT1/FADD is involved in liver regeneration. *World journal of gastroenterology* 2005, **11**(46): 7248-7253.

4806

Hepatocyte response to infection and injury in mice. *Hepatology (Baltimore, Md)* 2007, **45**(4): 1014-1024.

4810

Freimuth J, Bangen JM, Lambertz D, Hu W, Nevzorova YA, Sonntag R, *et al.* Loss of caspase-8 in hepatocytes accelerates the onset of liver regeneration in mice through premature nuclear factor kappa B activation. *Hepatology (Baltimore, Md)* 2013, **58**(5): 1779-1789.

4815 777. Isayama F, Moore S, Hines IN, Wheeler MD. Fas Regulates Macrophage Polarization and Fibrogenic Phenotype in a Model of Chronic Ethanol-Induced Hepatocellular Injury. *The American journal of pathology* 2016, **186**(6): 1524-1536.

4818

Verma VK, Li H, Wang R, Hirsova P, Mushref M, Liu Y, *et al.* Alcohol stimulates macrophage activation through caspase-dependent hepatocyte derived release of CD40L containing extracellular vesicles. *Journal of hepatology* 2016, **64**(3): 651-660.

4822

4823 779. Yin M, Wheeler MD, Kono H, Bradford BU, Gallucci RM, Luster MI, *et al.* Essential role of tumor necrosis factor alpha in alcohol-induced liver injury in mice. *Gastroenterology* 1999, **117**(4): 942-952.

4826

4827 780. Mundt B, Wirth T, Zender L, Waltemathe M, Trautwein C, Manns MP, *et al.* Tumour necrosis factor related apoptosis inducing ligand (TRAIL) induces hepatic steatosis in viral hepatitis and after alcohol intake. *Gut* 2005, **54**(11): 1590-1596.

4830

Hao F, Cubero FJ, Ramadori P, Liao L, Haas U, Lambertz D, *et al.* Inhibition of Caspase-8 does not protect from alcohol-induced liver apoptosis but alleviates alcoholic hepatic steatosis in mice. *Cell death & disease* 2017, **8**(10): e3152.

4834

Zhou Z, Sun X, Kang YJ. Ethanol-induced apoptosis in mouse liver: Fas- and cytochrome c-mediated caspase-3 activation pathway. *The American journal of pathology* 2001, **159**(1): 329-338.

4838

4839 783. Item F, Wueest S, Lemos V, Stein S, Lucchini FC, Denzler R, *et al.* Fas cell surface death receptor controls hepatic lipid metabolism by regulating mitochondrial function. *Nature communications* 2017, **8**(1): 480.

4842

Kakino S, Ohki T, Nakayama H, Yuan X, Otabe S, Hashinaga T, et al. Pivotal Role of TNF-α in the Development and Progression of Nonalcoholic Fatty Liver Disease in a Murine Model.
 Hormone and metabolic research = Hormon- und Stoffwechselforschung = Hormones et metabolisme 2018, 50(1): 80-87.

4847

Salles J, Tardif N, Landrier JF, Mothe-Satney I, Guillet C, Boue-Vaysse C, et al. TNFα gene knockout differentially affects lipid deposition in liver and skeletal muscle of high-fat-diet mice.
 The Journal of nutritional biochemistry 2012, 23(12): 1685-1693.

4851

4852 786. Kanuri G, Spruss A, Wagnerberger S, Bischoff SC, Bergheim I. Role of tumor necrosis factor α
 4853 (TNFα) in the onset of fructose-induced nonalcoholic fatty liver disease in mice. *The Journal of nutritional biochemistry* 2011, 22(6): 527-534.

4855 787. Tomita K, Tamiya G, Ando S, Ohsumi K, Chiyo T, Mizutani A, et al. Tumour necrosis factor 4856

alpha signalling through activation of Kupffer cells plays an essential role in liver fibrosis of non-

alcoholic steatohepatitis in mice. *Gut* 2006, **55**(3): 415-424.

4859

4857 4858

4860 788. De Sousa Rodrigues ME, Houser MC, Walker DI, Jones DP, Chang J, Barnum CJ, et al. Targeting soluble tumor necrosis factor as a potential intervention to lower risk for late-onset Alzheimer's 4861 disease associated with obesity, metabolic syndrome, and type 2 diabetes. Alzheimer's research 4862 4863

& therapy 2019, **12**(1): 1.

4864

4865 789. Ilan Y, Ben Ya'acov A, Shabbat Y, Gingis-Velitski S, Almon E, Shaaltiel Y. Oral administration of a non-absorbable plant cell-expressed recombinant anti-TNF fusion protein induces 4866 immunomodulatory effects and alleviates nonalcoholic steatohepatitis. World journal of 4867 gastroenterology 2016, 22(39): 8760-8769. 4868

4869

4870 790. Koca SS, Bahcecioglu IH, Poyrazoglu OK, Ozercan IH, Sahin K, Ustundag B. The treatment 4871 with antibody of TNF-alpha reduces the inflammation, necrosis and fibrosis in the non-alcoholic steatohepatitis induced by methionine- and choline-deficient diet. Inflammation 2008, 31(2): 91-4872

4873 98.

4874

Wandrer F, Liebig S, Marhenke S, Vogel A, John K, Manns MP, et al. TNF-Receptor-1 inhibition 4875 791. 4876 reduces liver steatosis, hepatocellular injury and fibrosis in NAFLD mice. Cell death & disease 4877 2020, **11**(3): 212.

4878

4879 792. Bluemel S, Wang Y, Lee S, Schnabl B. Tumor necrosis factor alpha receptor 1 deficiency in 4880 hepatocytes does not protect from non-alcoholic steatohepatitis, but attenuates insulin resistance in mice. World journal of gastroenterology 2020, **26**(33): 4933-4944. 4881

4882

793. Lambertucci F, Arboatti A, Sedlmeier MG, Motiño O, Alvarez ML, Ceballos MP, et al. 4883 4884 Disruption of tumor necrosis factor alpha receptor 1 signaling accelerates NAFLD progression in mice upon a high-fat diet. The Journal of nutritional biochemistry 2018, 58: 17-27. 4885

4886

4887 794. Bernardi S, Toffoli B, Tisato V, Bossi F, Biffi S, Lorenzon A, et al. TRAIL reduces impaired 4888 glucose tolerance and NAFLD in the high-fat diet fed mouse. Clinical science (London, England : 1979) 2018, **132**(1): 69-83. 4889

4890

Hirsova P, Weng P, Salim W, Bronk SF, Griffith TS, Ibrahim SH, et al. TRAIL Deletion Prevents 4891 795. Liver, but Not Adipose Tissue, Inflammation during Murine Diet-Induced Obesity. Hepatology 4892 4893 communications 2017, **1**(7): 648-662.

4895 796. Cartland SP, Harith HH, Genner SW, Dang L, Cogger VC, Vellozzi M, *et al.* Non-alcoholic fatty liver disease, vascular inflammation and insulin resistance are exacerbated by TRAIL deletion in mice. *Scientific reports* 2017, **7**(1): 1898.

4898

Krishnan A, Katsumi T, Guicciardi ME, Azad AI, Ozturk NB, Trussoni CE, et al. Tumor Necrosis
 Factor-Related Apoptosis-Inducing Ligand Receptor Deficiency Promotes the Ductular Reaction,
 Macrophage Accumulation, and Hepatic Fibrosis in the Abcb4(-/-) Mouse. *The American journal* of pathology 2020, 190(6): 1284-1297.

4903

4904 798. Gujral JS, Liu J, Farhood A, Jaeschke H. Reduced oncotic necrosis in Fas receptor-deficient C57BL/6J-lpr mice after bile duct ligation. *Hepatology (Baltimore, Md)* 2004, **40**(4): 998-1007.

4906

4907 799. Canbay A, Higuchi H, Bronk SF, Taniai M, Sebo TJ, Gores GJ. Fas enhances fibrogenesis in the bile duct ligated mouse: a link between apoptosis and fibrosis. *Gastroenterology* 2002, **123**(4): 1323-1330.

4910

4911 800. Miyoshi H, Rust C, Roberts PJ, Burgart LJ, Gores GJ. Hepatocyte apoptosis after bile duct ligation in the mouse involves Fas. *Gastroenterology* 1999, **117**(3): 669-677.

4913

Osawa Y, Hoshi M, Yasuda I, Saibara T, Moriwaki H, Kozawa O. Tumor necrosis factor-α
 promotes cholestasis-induced liver fibrosis in the mouse through tissue inhibitor of metalloproteinase-1 production in hepatic stellate cells. *PloS one* 2013, 8(6): e65251.

4917

4918 802. Gäbele E, Froh M, Arteel GE, Uesugi T, Hellerbrand C, Schölmerich J, et al. TNFalpha is required for cholestasis-induced liver fibrosis in the mouse. Biochemical and biophysical research communications 2009, **378**(3): 348-353.

4921

4922 803. Takeda K, Kojima Y, Ikejima K, Harada K, Yamashina S, Okumura K, *et al.* Death receptor 5 mediated-apoptosis contributes to cholestatic liver disease. *Proceedings of the National Academy of Sciences of the United States of America* 2008, **105**(31): 10895-10900.

4925

4926 804. Kahraman A, Barreyro FJ, Bronk SF, Werneburg NW, Mott JL, Akazawa Y, et al. TRAIL mediates liver injury by the innate immune system in the bile duct-ligated mouse. *Hepatology* (Baltimore, Md) 2008, **47**(4): 1317-1330.

4929

4930 805. Zhuang H, Wang X, Zha D, Gan Z, Cai F, Du P, *et al.* FADD is a key regulator of lipid metabolism. *EMBO molecular medicine* 2016, **8**(8): 895-918.

- Wang PX, Ji YX, Zhang XJ, Zhao LP, Yan ZZ, Zhang P, *et al.* Targeting CASP8 and FADD-like apoptosis regulator ameliorates nonalcoholic steatohepatitis in mice and nonhuman primates.
- 4935 *Nature medicine* 2017, **23**(4): 439-449.

4937 807. Gehrke N, Nagel M, Straub BK, Wörns MA, Schuchmann M, Galle PR, *et al.* Loss of cellular FLICE-inhibitory protein promotes acute cholestatic liver injury and inflammation from bile duct ligation. *American journal of physiology Gastrointestinal and liver physiology* 2018, **314**(3): G319-g333.

4941

4942 808. Chaudhary K, Liedtke C, Wertenbruch S, Trautwein C, Streetz KL. Caspase 8 differentially controls hepatocytes and non-parenchymal liver cells during chronic cholestatic liver injury in mice. *Journal of hepatology* 2013, **59**(6): 1292-1298.

4945

4946 809. Hatting M, Zhao G, Schumacher F, Sellge G, Al Masaoudi M, Gaβler N, *et al.* Hepatocyte caspase-8 is an essential modulator of steatohepatitis in rodents. *Hepatology (Baltimore, Md)* 2013, **57**(6): 2189-2201.

4949

4950 810. Cubero FJ, Peng J, Liao L, Su H, Zhao G, Zoubek ME, *et al.* Inactivation of caspase 8 in liver parenchymal cells confers protection against murine obstructive cholestasis. *Journal of hepatology* 2018, **69**(6): 1326-1334.

4953

4954 811. Kondylis V, Polykratis A, Ehlken H, Ochoa-Callejero L, Straub BK, Krishna-Subramanian S, *et al.* NEMO Prevents Steatohepatitis and Hepatocellular Carcinoma by Inhibiting RIPK1 Kinase Activity-Mediated Hepatocyte Apoptosis. *Cancer cell* 2015, **28**(5): 582-598.

4957

4958 812. Ehlken H, Krishna-Subramanian S, Ochoa-Callejero L, Kondylis V, Nadi NE, Straub BK, *et al.*4959 Death receptor-independent FADD signalling triggers hepatitis and hepatocellular carcinoma in
4960 mice with liver parenchymal cell-specific NEMO knockout. *Cell death and differentiation* 2014,
4961 **21**(11): 1721-1732.

4962

4963 813. Vogel A, Aslan JE, Willenbring H, Klein C, Finegold M, Mount H, *et al.* Sustained phosphorylation of Bid is a marker for resistance to Fas-induced apoptosis during chronic liver diseases. *Gastroenterology* 2006, **130**(1): 104-119.

4966

4967 814. Straus SE, Jaffe ES, Puck JM, Dale JK, Elkon KB, Rösen-Wolff A, *et al.* The development of lymphomas in families with autoimmune lymphoproliferative syndrome with germline Fas mutations and defective lymphocyte apoptosis. *Blood* 2001, **98**(1): 194-200.

4970

Davidson WF, Giese T, Fredrickson TN. Spontaneous development of plasmacytoid tumors in mice with defective Fas-Fas ligand interactions. *The Journal of experimental medicine* 1998, **187**(11): 1825-1838.

4974
 4975 816. Finnberg N, Klein-Szanto AJ, El-Deiry WS. TRAIL-R deficiency in mice promotes susceptibility to chronic inflammation and tumorigenesis. *The Journal of clinical investigation* 2008, 118(1):

4977 111-123.

4978

4982

4986

4989

4992

4995

4998

5002

5006

- 4979 817. Zerafa N, Westwood JA, Cretney E, Mitchell S, Waring P, Iezzi M, *et al.* Cutting edge: TRAIL deficiency accelerates hematological malignancies. *Journal of immunology (Baltimore, Md:* 1950) 2005, **175**(9): 5586-5590.
- 4983 818. Yue HH, Diehl GE, Winoto A. Loss of TRAIL-R does not affect thymic or intestinal tumor development in p53 and adenomatous polyposis coli mutant mice. *Cell death and differentiation* 2005, **12**(1): 94-97.
- 4987 819. LA OR, Tai L, Lee L, Kruse EA, Grabow S, Fairlie WD, *et al.* Membrane-bound Fas ligand only is essential for Fas-induced apoptosis. *Nature* 2009, **461**(7264): 659-663.
- 4990 820. Guillen-Ahlers H, Suckow MA, Castellino FJ, Ploplis VA. Fas/CD95 deficiency in ApcMin/+ mice increases intestinal tumor burden. *PloS one* 2010, **5**(2): e9070.
- 4993 821. Park SM, Chen L, Zhang M, Ashton-Rickardt P, Turner JR, Peter ME. CD95 is cytoprotective for intestinal epithelial cells in colitis. *Inflammatory bowel diseases* 2010, **16**(6): 1063-1070.
- 4996 822. Fingleton B, Carter KJ, Matrisian LM. Loss of functional Fas ligand enhances intestinal tumorigenesis in the Min mouse model. *Cancer research* 2007, **67**(10): 4800-4806.
- Kim JY, Kim YM, Park JM, Han YM, Lee KC, Hahm KB, *et al.* Cancer preventive effect of recombinant TRAIL by ablation of oncogenic inflammation in colitis-associated cancer rather than anticancer effect. *Oncotarget* 2018, **9**(2): 1705-1716.
- 5003 824. Lopetuso LR, Petito V, Zinicola T, Graziani C, Gerardi V, Arena V, *et al.* Infliximab does not increase colonic cancer risk associated to murine chronic colitis. *World journal of gastroenterology* 2016, **22**(44): 9727-9733.
- 5007 825. Craven B, Zaric V, Martin A, Mureau C, Egan LJ. Effect of genetic deletion or pharmacological antagonism of tumor necrosis factor alpha on colitis-associated carcinogenesis in mice.

  5009 Inflammatory bowel diseases 2015, 21(3): 485-495.
- Nyboe Andersen N, Pasternak B, Basit S, Andersson M, Svanström H, Caspersen S, et al.
   Association between tumor necrosis factor-α antagonists and risk of cancer in patients with inflammatory bowel disease. Jama 2014, 311(23): 2406-2413.

5014
 5015 827. Chang F, Lacey MR, Bouljihad M, Höner Zu Bentrup K, Fortgang IS. Tumor necrosis factor receptor 1 functions as a tumor suppressor. *American journal of physiology Gastrointestinal and liver physiology* 2012, 302(2): G195-206.

5019 828. Ba H, Jiang R, Zhang M, Yin B, Wang J, Li Z, *et al.* Suppression of Transmembrane Tumor 5020 Necrosis Factor Alpha Processing by a Specific Antibody Protects Against Colitis-Associated 5021 Cancer. *Frontiers in immunology* 2021, **12:** 687874.

5018

5022

5026

5030

5035

5039

5043

5047

- 5023 829. Yang Y, Gharaibeh RZ, Newsome RC, Jobin C. Amending microbiota by targeting intestinal inflammation with TNF blockade attenuates development of colorectal cancer. *Nature cancer* 2020, **1**(7): 723-734.
- 5027 830. Kim YJ, Hong KS, Chung JW, Kim JH, Hahm KB. Prevention of colitis-associated carcinogenesis with infliximab. *Cancer prevention research (Philadelphia, Pa)* 2010, **3**(10): 1314-1333.
- 5031 831. Onizawa M, Nagaishi T, Kanai T, Nagano K, Oshima S, Nemoto Y, *et al.* Signaling pathway via TNF-alpha/NF-kappaB in intestinal epithelial cells may be directly involved in colitis-associated carcinogenesis. *American journal of physiology Gastrointestinal and liver physiology* 2009, **296**(4): G850-859.
- 832. Rao VP, Poutahidis T, Ge Z, Nambiar PR, Horwitz BH, Fox JG, *et al.* Proinflammatory CD4+ CD45RB(hi) lymphocytes promote mammary and intestinal carcinogenesis in Apc(Min/+) mice. *Cancer research* 2006, **66**(1): 57-61.
- 5040 833. Oshima H, Ishikawa T, Yoshida GJ, Naoi K, Maeda Y, Naka K, *et al.* TNF-α/TNFR1 signaling promotes gastric tumorigenesis through induction of Noxo1 and Gna14 in tumor cells. *Oncogene* 2014, **33**(29): 3820-3829.
- Popivanova BK, Kitamura K, Wu Y, Kondo T, Kagaya T, Kaneko S, *et al.* Blocking TNF-alpha in mice reduces colorectal carcinogenesis associated with chronic colitis. *The Journal of clinical investigation* 2008, **118**(2): 560-570.
- 5048 835. Castets M, Broutier L, Molin Y, Brevet M, Chazot G, Gadot N, *et al.* DCC constrains tumour progression via its dependence receptor activity. *Nature* 2011, **482**(7386): 534-537.
- 5051 836. Genevois AL, Ichim G, Coissieux MM, Lambert MP, Lavial F, Goldschneider D, et al.
  5052 Dependence receptor TrkC is a putative colon cancer tumor suppressor. Proceedings of the
  5053 National Academy of Sciences of the United States of America 2013, 110(8): 3017-3022.

Negulescu AM, Mehlen P. Dependence receptors - the dark side awakens. *The FEBS journal* 2018, 285(21): 3909-3924.
Grandin M, Meier M, Delcros JG, Nikodemus D, Reuten R, Patel TR, *et al.* Structural Decoding of the Netrin-1/UNC5 Interaction and its Therapeutical Implications in Cancers. *Cancer cell* 2016, 29(2): 173-185.

5062 839. Chen L, Park SM, Tumanov AV, Hau A, Sawada K, Feig C, *et al.* CD95 promotes tumour growth. *Nature* 2010, **465**(7297): 492-496.

5061

5064

5067

5070

5074

5078

5082

5086

5090

Pikarsky E, Porat RM, Stein I, Abramovitch R, Amit S, Kasem S, *et al.* NF-kappaB functions as a tumour promoter in inflammation-associated cancer. *Nature* 2004, **431**(7007): 461-466.

5068 841. Schneider AT, Gautheron J, Feoktistova M, Roderburg C, Loosen SH, Roy S, *et al.* RIPK1 Suppresses a TRAF2-Dependent Pathway to Liver Cancer. *Cancer cell* 2017, **31**(1): 94-109.

- 5071 842. Vucur M, Reisinger F, Gautheron J, Janssen J, Roderburg C, Cardenas DV, *et al.* RIP3 inhibits inflammatory hepatocarcinogenesis but promotes cholestasis by controlling caspase-8- and JNK-dependent compensatory cell proliferation. *Cell reports* 2013, **4**(4): 776-790.
- 5075 843. Vredevoogd DW, Kuilman T, Ligtenberg MA, Boshuizen J, Stecker KE, de Bruijn B, *et al.*5076 Augmenting Immunotherapy Impact by Lowering Tumor TNF Cytotoxicity Threshold. *Cell*5077 2019, **178**(3): 585-599.e515.
- Boege Y, Malehmir M, Healy ME, Bettermann K, Lorentzen A, Vucur M, *et al.* A Dual Role of Caspase-8 in Triggering and Sensing Proliferation-Associated DNA Damage, a Key Determinant of Liver Cancer Development. *Cancer cell* 2017, 32(3): 342-359.e310.
- 5083 845. Liccardi G, Ramos Garcia L, Tenev T, Annibaldi A, Legrand AJ, Robertson D, *et al.* RIPK1 and Caspase-8 Ensure Chromosome Stability Independently of Their Role in Cell Death and Inflammation. *Molecular cell* 2019, **73**(3): 413-428.e417.
- Hakem A, El Ghamrasni S, Maire G, Lemmers B, Karaskova J, Jurisicova A, *et al.* Caspase-8 is essential for maintaining chromosomal stability and suppressing B-cell lymphomagenesis. *Blood* 2012, **119**(15): 3495-3502.
- Krelin Y, Zhang L, Kang TB, Appel E, Kovalenko A, Wallach D. Caspase-8 deficiency facilitates cellular transformation in vitro. *Cell death and differentiation* 2008, **15**(9): 1350-1355.

5093 5094 848. Rodriguez YI, Campos LE, Castro MG, Bannoud N, Blidner AG, Filippa VP, et al. Tumor Necrosis Factor Receptor-1 (p55) Deficiency Attenuates Tumor Growth and Intratumoral 5095 5096 Angiogenesis and Stimulates CD8(+) T Cell Function in Melanoma. *Cells* 2020, **9**(11). 5097 5098 849. Schioppa T, Moore R, Thompson RG, Rosser EC, Kulbe H, Nedospasov S, et al. B regulatory cells and the tumor-promoting actions of TNF-\alpha during squamous carcinogenesis. *Proceedings* 5099 of the National Academy of Sciences of the United States of America 2011, 108(26): 10662-5100 5101 10667. 5102 5103 850. Arnott CH, Scott KA, Moore RJ, Robinson SC, Thompson RG, Balkwill FR. Expression of both TNF-alpha receptor subtypes is essential for optimal skin tumour development. Oncogene 2004, 5104 **23**(10): 1902-1910. 5105 5106 5107 851. Scott KA, Moore RJ, Arnott CH, East N, Thompson RG, Scallon BJ, et al. An anti-tumor necrosis 5108 factor-alpha antibody inhibits the development of experimental skin tumors. Molecular cancer 5109 therapeutics 2003, **2**(5): 445-451. 5110 Suganuma M, Okabe S, Marino MW, Sakai A, Sueoka E, Fujiki H. Essential role of tumor 852. 5111 necrosis factor alpha (TNF-alpha) in tumor promotion as revealed by TNF-alpha-deficient mice. 5112 Cancer research 1999, **59**(18): 4516-4518. 5113 5114 853. Moore RJ, Owens DM, Stamp G, Arnott C, Burke F, East N, et al. Mice deficient in tumor 5115 necrosis factor-alpha are resistant to skin carcinogenesis. *Nature medicine* 1999, **5**(7): 828-831. 5116 5117 854. 5118 Calıskan E, Gamsızkan M, Yurekli A, Botsali A, Kabalar ME, Demiriz M, et al. Anti-TNF agent etanercept augments UV-induced skin cancer development in SKH-1 mice. The Journal of 5119 dermatological treatment 2021, **32**(7): 812-818. 5120 5121 Singh A, Singh A, Bauer SJ, Wheeler DL, Havighurst TC, Kim K, et al. Genetic deletion of TNFα 5122 855. 5123 inhibits ultraviolet radiation-induced development of cutaneous squamous cell carcinomas in PKCe transgenic mice via inhibition of cell survival signals. *Carcinogenesis* 2016, **37**(1): 72-80. 5124 5125 5126 856. Lind MH, Rozell B, Wallin RP, van Hogerlinden M, Ljunggren HG, Toftgård R, et al. Tumor necrosis factor receptor 1-mediated signaling is required for skin cancer development induced by 5127 5128 NF-kappaB inhibition. Proceedings of the National Academy of Sciences of the United States of

5131 857. Galheigo MR, Cruz AR, Cabral Á S, Faria PR, Cordeiro RS, Silva MJ, et al. Role of the TNF-α receptor type 1 on prostate carcinogenesis in knockout mice. The Prostate 2016, 76(10): 917-926.

America 2004, **101**(14): 4972-4977.

5129

5134 858. 5135 Sobo-Vujanovic A, Vujanovic L, DeLeo AB, Concha-Benavente F, Ferris RL, Lin Y, et al. Inhibition of Soluble Tumor Necrosis Factor Prevents Chemically Induced Carcinogenesis in 5136 5137 Mice. *Cancer immunology research* 2016, **4**(5): 441-451. 5138 He L, Bhat K, Duhacheck-Muggy S, Ioannidis A, Zhang L, Nguyen NT, et al. Tumor necrosis 5139 859. factor receptor signaling modulates carcinogenesis in a mouse model of breast cancer. Neoplasia 5140 (New York, NY) 2021, 23(2): 197-209. 5141 5142 5143 860. Sangaletti S, Tripodo C, Ratti C, Piconese S, Porcasi R, Salcedo R, et al. Oncogene-driven intrinsic inflammation induces leukocyte production of tumor necrosis factor that critically 5144 contributes to mammary carcinogenesis. Cancer research 2010, **70**(20): 7764-7775. 5145 5146 Chadwick JW, Macdonald R, Ali AA, Glogauer M, Magalhaes MA. TNFa Signaling Is Increased 5147 861. in Progressing Oral Potentially Malignant Disorders and Regulates Malignant Transformation in 5148 5149 an Oral Carcinogenesis Model. Frontiers in oncology 2021, 11: 741013. 5150 5151 862. Karabela SP, Kairi CA, Magkouta S, Psallidas I, Moschos C, Stathopoulos I, et al. Neutralization of tumor necrosis factor bioactivity ameliorates urethane-induced pulmonary oncogenesis in 5152 mice. Neoplasia (New York, NY) 2011, 13(12): 1143-1151. 5153 5154 5155 Gong L, da Silva Caetano M, Cumpian AM, Daliri S, Garza Flores A, Chang SH, et al. Tumor 863. necrosis factor links chronic obstructive pulmonary disease and K-ras mutant lung cancer through 5156 induction of an immunosuppressive pro-tumor microenvironment. Oncoimmunology 2016, 5157 5158 **5**(10): e1229724. 5159 5160 864. Kedinger V, Muller S, Gronemeyer H. Targeted expression of tumor necrosis factor-related apoptosis-inducing ligand TRAIL in skin protects mice against chemical carcinogenesis. 5161 5162 *Molecular cancer* 2011, **10:** 34. 5163 5164 865. Chio, II, Sasaki M, Ghazarian D, Moreno J, Done S, Ueda T, et al. TRADD contributes to tumour suppression by regulating ULF-dependent p19Arf ubiquitylation. Nature cell biology 2012, 5165 **14**(6): 625-633. 5166 5167 Grosse-Wilde A, Voloshanenko O, Bailey SL, Longton GM, Schaefer U, Csernok AI, et al. 5168 866. 5169 TRAIL-R deficiency in mice enhances lymph node metastasis without affecting primary tumor development. The Journal of clinical investigation 2008, 118(1): 100-110. 5170 5171 Takeda K, Smyth MJ, Cretney E, Hayakawa Y, Kayagaki N, Yagita H, et al. Critical role for

tumor necrosis factor-related apoptosis-inducing ligand in immune surveillance against tumor

development. The Journal of experimental medicine 2002, 195(2): 161-169.

5172

5173 5174 867.

5175
5176
868. Cretney E, Takeda K, Yagita H, Glaccum M, Peschon JJ, Smyth MJ. Increased susceptibility to tumor initiation and metastasis in TNF-related apoptosis-inducing ligand-deficient mice. *Journal of immunology (Baltimore, Md : 1950)* 2002, **168**(3): 1356-1361.

5179
5180
869. Montinaro A, Areso Zubiaur I, Saggau J, Kretz AL, Ferreira RMM, Hassan O. *et al.* Potent pro-

Montinaro A, Areso Zubiaur I, Saggau J, Kretz AL, Ferreira RMM, Hassan O, *et al.* Potent proapoptotic combination therapy is highly effective in a broad range of cancers. *Cell death and* differentiation 2022, **29**(3): 492-503.

5184 870. von Karstedt S, Conti A, Nobis M, Montinaro A, Hartwig T, Lemke J, *et al.* Cancer cellautonomous TRAIL-R signaling promotes KRAS-driven cancer progression, invasion, and metastasis. *Cancer cell* 2015, **27**(4): 561-573.

5183

5187

5191

5195

5199

5203

5207

- Hartwig T, Montinaro A, von Karstedt S, Sevko A, Surinova S, Chakravarthy A, *et al.* The TRAIL-Induced Cancer Secretome Promotes a Tumor-Supportive Immune Microenvironment via CCR2. *Molecular cell* 2017, **65**(4): 730-742.e735.
- 5192 872. Hoogwater FJ, Nijkamp MW, Smakman N, Steller EJ, Emmink BL, Westendorp BF, *et al.*5193 Oncogenic K-Ras turns death receptors into metastasis-promoting receptors in human and mouse colorectal cancer cells. *Gastroenterology* 2010, **138**(7): 2357-2367.
- 5196 873. Rieux-Laucat F, Le Deist F, Hivroz C, Roberts IA, Debatin KM, Fischer A, *et al.* Mutations in Fas associated with human lymphoproliferative syndrome and autoimmunity. *Science (New York, NY)* 1995, **268**(5215): 1347-1349.
- Watanabe-Fukunaga R, Brannan CI, Copeland NG, Jenkins NA, Nagata S. Lymphoproliferation disorder in mice explained by defects in Fas antigen that mediates apoptosis. *Nature* 1992, **356**(6367): 314-317.
- 5204 875. Alvarez-Diaz S, Dillon CP, Lalaoui N, Tanzer MC, Rodriguez DA, Lin A, *et al.* The Pseudokinase MLKL and the Kinase RIPK3 Have Distinct Roles in Autoimmune Disease Caused by Loss of Death-Receptor-Induced Apoptosis. *Immunity* 2016, **45**(3): 513-526.
- 5208 876. Chyuan IT, Tsai HF, Wu CS, Sung CC, Hsu PN. TRAIL-Mediated Suppression of T Cell Faceptor Signaling Inhibits T Cell Activation and Inflammation in Experimental Autoimmune Encephalomyelitis. *Frontiers in immunology* 2018, **9:** 15.
- 5212 877. Ikeda T, Hirata S, Fukushima S, Matsunaga Y, Ito T, Uchino M, *et al.* Dual effects of TRAIL in suppression of autoimmunity: the inhibition of Th1 cells and the promotion of regulatory T cells.

  5214 *Journal of immunology (Baltimore, Md : 1950)* 2010, **185**(9): 5259-5267.

5216 878. Cretney E, McQualter JL, Kayagaki N, Yagita H, Bernard CC, Grewal IS, *et al.* TNF-related apoptosis-inducing ligand (TRAIL)/Apo2L suppresses experimental autoimmune encephalomyelitis in mice. *Immunology and cell biology* 2005, **83**(5): 511-519.

5219

5220 879. Razmara M, Hilliard B, Ziarani AK, Murali R, Yellayi S, Ghazanfar M, *et al.* Fn14-TRAIL, a chimeric intercellular signal exchanger, attenuates experimental autoimmune encephalomyelitis.

5222 *The American journal of pathology* 2009, **174**(2): 460-474.

5223

5224 880. Aktas O, Smorodchenko A, Brocke S, Infante-Duarte C, Schulze Topphoff U, Vogt J, et al.
5225 Neuronal damage in autoimmune neuroinflammation mediated by the death ligand TRAIL.
5226 Neuron 2005, **46**(3): 421-432.

5227 5228

5228 881. Hilliard B, Wilmen A, Seidel C, Liu TS, Göke R, Chen Y. Roles of TNF-related apoptosis-5229 inducing ligand in experimental autoimmune encephalomyelitis. *Journal of immunology* 5230 (*Baltimore, Md*: 1950) 2001, **166**(2): 1314-1319.

5231

Lamhamedi-Cherradi SE, Zheng SJ, Maguschak KA, Peschon J, Chen YH. Defective thymocyte apoptosis and accelerated autoimmune diseases in TRAIL-/- mice. *Nature immunology* 2003, 4(3): 255-260.

5235

5236 883. Song K, Chen Y, Göke R, Wilmen A, Seidel C, Göke A, *et al.* Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) is an inhibitor of autoimmune inflammation and cell cycle progression. *The Journal of experimental medicine* 2000, **191**(7): 1095-1104.

5239

5240 884. Park JS, Oh Y, Park O, Foss CA, Lim SM, Jo DG, et al. PEGylated TRAIL ameliorates 5241 experimental inflammatory arthritis by regulation of Th17 cells and regulatory T cells. *Journal* 5242 of controlled release: official journal of the Controlled Release Society 2017, **267**: 163-171.

5243

5244 885. Chyuan IT, Tsai HF, Liao HJ, Wu CS, Hsu PN. An apoptosis-independent role of TRAIL in suppressing joint inflammation and inhibiting T-cell activation in inflammatory arthritis. *Cellular & molecular immunology* 2018, **15**(9): 846-857.

5247

5248 886. Jin CH, Chae SY, Kim TH, Yang HK, Lee EY, Song YW, *et al.* Effect of tumor necrosis factorrelated apoptosis-inducing ligand on the reduction of joint inflammation in experimental rheumatoid arthritis. *The Journal of pharmacology and experimental therapeutics* 2010, **332**(3): 858-865.

5252

5253 887. Kang S, Park EJ, Joe Y, Seo E, Park MK, Seo SY, *et al.* Systemic delivery of TNF-related apoptosis-inducing ligand (TRAIL) elevates levels of tissue inhibitor of metalloproteinase-1

5255 (TIMP-1) and prevents type 1 diabetes in nonobese diabetic mice. *Endocrinology* 2010, **151**(12): 5638-5646.

5257

5258 888. Mi QS, Ly D, Lamhamedi-Cherradi SE, Salojin KV, Zhou L, Grattan M, *et al.* Blockade of tumor 5259 necrosis factor-related apoptosis-inducing ligand exacerbates type 1 diabetes in NOD mice. 5260 *Diabetes* 2003, **52**(8): 1967-1975.

5261

5262 889. Bossi F, Bernardi S, Zauli G, Secchiero P, Fabris B. TRAIL modulates the immune system and protects against the development of diabetes. *Journal of immunology research* 2015, **2015**: 680749.

5265

5266 890. Lamhamedi-Cherradi SE, Zheng S, Tisch RM, Chen YH. Critical roles of tumor necrosis factorrelated apoptosis-inducing ligand in type 1 diabetes. *Diabetes* 2003, **52**(9): 2274-2278.

5268

Bachmann R, Eugster HP, Frei K, Fontana A, Lassmann H. Impairment of TNF-receptor-1 signaling but not fas signaling diminishes T-cell apoptosis in myelin oligodendrocyte glycoprotein peptide-induced chronic demyelinating autoimmune encephalomyelitis in mice. *The American journal of pathology* 1999, **154**(5): 1417-1422.

5273

Malipiero U, Frei K, Spanaus KS, Agresti C, Lassmann H, Hahne M, *et al.* Myelin oligodendrocyte glycoprotein-induced autoimmune encephalomyelitis is chronic/relapsing in perforin knockout mice, but monophasic in Fas- and Fas ligand-deficient lpr and gld mice. *European journal of immunology* 1997, **27**(12): 3151-3160.

5278

5279 893. Waldner H, Sobel RA, Howard E, Kuchroo VK. Fas- and FasL-deficient mice are resistant to induction of autoimmune encephalomyelitis. *Journal of immunology (Baltimore, Md : 1950)* 1997, **159**(7): 3100-3103.

5282

5283 894. Sabelko KA, Kelly KA, Nahm MH, Cross AH, Russell JH. Fas and Fas ligand enhance the pathogenesis of experimental allergic encephalomyelitis, but are not essential for immune privilege in the central nervous system. *Journal of immunology (Baltimore, Md : 1950)* 1997, 5286 159(7): 3096-3099.

5287

Wolf Y, Shemer A, Polonsky M, Gross M, Mildner A, Yona S, *et al.* Autonomous TNF is critical for in vivo monocyte survival in steady state and inflammation. *The Journal of experimental medicine* 2017, **214**(4): 905-917.

5291

Williams SK, Fairless R, Maier O, Liermann PC, Pichi K, Fischer R, *et al.* Anti-TNFR1 targeting in humanized mice ameliorates disease in a model of multiple sclerosis. *Scientific reports* 2018, **8**(1): 13628.

Williams SK, Maier O, Fischer R, Fairless R, Hochmeister S, Stojic A, *et al.* Antibody-mediated inhibition of TNFR1 attenuates disease in a mouse model of multiple sclerosis. *PloS one* 2014, 9(2): e90117.

Nomura T, Abe Y, Kamada H, Shibata H, Kayamuro H, Inoue M, *et al.* Therapeutic effect of PEGylated TNFR1-selective antagonistic mutant TNF in experimental autoimmune encephalomyelitis mice. *Journal of controlled release : official journal of the Controlled Release Society* 2011, **149**(1): 8-14.

5299

5304

5308

5313

5318

5323

5327

5331

5335

5305 Steeland S, Van Ryckeghem S, Van Imschoot G, De Rycke R, Toussaint W, Vanhoutte L, *et al.*5306 TNFR1 inhibition with a Nanobody protects against EAE development in mice. *Scientific reports*5307 2017, **7**(1): 13646.

5309 900. Brambilla R, Ashbaugh JJ, Magliozzi R, Dellarole A, Karmally S, Szymkowski DE, *et al.*15310 Inhibition of soluble tumour necrosis factor is therapeutic in experimental autoimmune encephalomyelitis and promotes axon preservation and remyelination. *Brain : a journal of neurology* 2011, **134**(Pt 9): 2736-2754.

Körner H, Lemckert FA, Chaudhri G, Etteldorf S, Sedgwick JD. Tumor necrosis factor blockade in actively induced experimental autoimmune encephalomyelitis prevents clinical disease despite activated T cell infiltration to the central nervous system. *European journal of immunology* 1997, 27(8): 1973-1981.

5319 902. Körner H, Goodsall AL, Lemckert FA, Scallon BJ, Ghrayeb J, Ford AL, *et al.* Unimpaired autoreactive T-cell traffic within the central nervous system during tumor necrosis factor receptor-mediated inhibition of experimental autoimmune encephalomyelitis. *Proceedings of the National Academy of Sciences of the United States of America* 1995, **92**(24): 11066-11070.

5324 903. Richter F, Williams SK, John K, Huber C, Vaslin C, Zanker H, *et al.* The TNFR1 Antagonist Atrosimab Is Therapeutic in Mouse Models of Acute and Chronic Inflammation. *Frontiers in immunology* 2021, **12:** 705485.

5328 904. Dittel BN, Merchant RM, Janeway CA, Jr. Evidence for Fas-dependent and Fas-independent mechanisms in the pathogenesis of experimental autoimmune encephalomyelitis. *Journal of immunology (Baltimore, Md : 1950)* 1999, **162**(11): 6392-6400.

5332 905. Suvannavejh GC, Dal Canto MC, Matis LA, Miller SD. Fas-mediated apoptosis in clinical remissions of relapsing experimental autoimmune encephalomyelitis. *The Journal of clinical investigation* 2000, **105**(2): 223-231.

- 5336 906. Wang X, Haroon F, Karray S, Martina D, Schlüter D. Astrocytic Fas ligand expression is required to induce T-cell apoptosis and recovery from experimental autoimmune encephalomyelitis. *European journal of immunology* 2013, **43**(1): 115-124.
- 5340 907. Sabelko-Downes KA, Cross AH, Russell JH. Dual role for Fas ligand in the initiation of and recovery from experimental allergic encephalomyelitis. *The Journal of experimental medicine* 1999, **189**(8): 1195-1205.

5343

5348

5352

5356

5360

5364

5368

- Batoulis H, Recks MS, Holland FO, Thomalla F, Williams RO, Kuerten S. Blockade of tumour necrosis factor-α in experimental autoimmune encephalomyelitis reveals differential effects on the antigen-specific immune response and central nervous system histopathology. *Clinical and experimental immunology* 2014, **175**(1): 41-48.
- 5349 909. Liu J, Marino MW, Wong G, Grail D, Dunn A, Bettadapura J, *et al.* TNF is a potent antiinflammatory cytokine in autoimmune-mediated demyelination. *Nature medicine* 1998, **4**(1): 78-83.
- 5353 910. Tu-Rapp H, Hammermüller A, Mix E, Kreutzer HJ, Goerlich R, Köhler H, *et al.* A proinflammatory role for Fas in joints of mice with collagen-induced arthritis. *Arthritis research* & *therapy* 2004, **6**(5): R404-414.
- Shen F, Verma AH, Volk A, Jones B, Coleman BM, Loza MJ, et al. Combined Blockade of TNF α and IL-17A Alleviates Progression of Collagen-Induced Arthritis without Causing Serious
   Infections in Mice. Journal of immunology (Baltimore, Md: 1950) 2019, 202(7): 2017-2026.
- Moore AR, Allden S, Bourne T, Denis MC, Kranidioti K, Okoye R, et al. Collagen II antibody induced arthritis in Tg1278TNFko mice: optimization of a novel model to assess treatments
   targeting human TNFα in rheumatoid arthritis. Journal of translational medicine 2014, 12: 285.
- 5365 913. Zalevsky J, Secher T, Ezhevsky SA, Janot L, Steed PM, O'Brien C, *et al.* Dominant-negative inhibitors of soluble TNF attenuate experimental arthritis without suppressing innate immunity to infection. *Journal of immunology (Baltimore, Md : 1950)* 2007, **179**(3): 1872-1883.
- 5369 914. Zhao Y, Yang X, Li S, Zhang B, Li S, Wang X, et al. sTNFRII-Fc modification protects human
   5370 UC-MSCs against apoptosis/autophagy induced by TNF-α and enhances their efficacy in
   5371 alleviating inflammatory arthritis. Stem cell research & therapy 2021, 12(1): 535.
- Huang QQ, Birkett R, Koessler RE, Cuda CM, Haines GK, 3rd, Jin JP, *et al.* Fas signaling in macrophages promotes chronicity in K/BxN serum-induced arthritis. *Arthritis & rheumatology* (*Hoboken, NJ*) 2014, **66**(1): 68-77.

916. 5377 Kang SE, Park JK, Yoo HJ, Kang HS, Park YW, Park BC, et al. Efficacy of novel bispecific antibody targeting TNF-α/CXCL10 in the treatment of experimental arthritis. Translational 5378 5379 research: the journal of laboratory and clinical medicine 2021, 232: 75-87.

5380

Itoh N, Imagawa A, Hanafusa T, Waguri M, Yamamoto K, Iwahashi H, et al. Requirement of 5381 917. Fas for the development of autoimmune diabetes in nonobese diabetic mice. The Journal of 5382 experimental medicine 1997, **186**(4): 613-618. 5383

5384

5385 918. Su X, Hu Q, Kristan JM, Costa C, Shen Y, Gero D, et al. Significant role for Fas in the 5386 pathogenesis of autoimmune diabetes. Journal of immunology (Baltimore, Md: 1950) 2000, **164**(5): 2523-2532. 5387

5391

5388

919. 5389 Chervonsky AV, Wang Y, Wong FS, Visintin I, Flavell RA, Janeway CA, Jr., et al. The role of Fas in autoimmune diabetes. *Cell* 1997, **89**(1): 17-24. 5390

920. Vence L, Benoist C, Mathis D. Fas deficiency prevents type 1 diabetes by inducing 5392 hyporesponsiveness in islet beta-cell-reactive T-cells. *Diabetes* 2004, **53**(11): 2797-2803. 5393

5394

5395 921. Mohamood AS, Guler ML, Xiao Z, Zheng D, Hess A, Wang Y, et al. Protection from autoimmune diabetes and T-cell lymphoproliferation induced by FasL mutation are differentially 5396 regulated and can be uncoupled pharmacologically. The American journal of pathology 2007, 5397 **171**(1): 97-106. 5398

5399

922. Jeong JH, Kim SH, Lee M, Kim WJ, Park TG, Ko KS, et al. Non-viral systemic delivery of Fas 5400 5401 siRNA suppresses cyclophosphamide-induced diabetes in NOD mice. Journal of controlled release: official journal of the Controlled Release Society 2010, 143(1): 88-94. 5402

5403

5404 923. Trivedi PM, Fynch S, Kennedy LM, Chee J, Krishnamurthy B, O'Reilly LA, et al. Soluble FAS ligand is not required for pancreatic islet inflammation or beta-cell destruction in non-obese 5405 5406 diabetic mice. Cell death discovery 2019, 5: 136.

5407

5408 924. Choi D, Radziszewska A, Schroer SA, Liadis N, Liu Y, Zhang Y, et al. Deletion of Fas in the 5409 pancreatic beta-cells leads to enhanced insulin secretion. American journal of physiology *Endocrinology and metabolism* 2009, **297**(6): E1304-1312. 5410

5411

5412 925. Thomas HE, Darwiche R, Corbett JA, Kay TW. Evidence that beta cell death in the nonobese 5413 diabetic mouse is Fas independent. Journal of immunology (Baltimore, Md: 1950) 1999, 163(3): 1562-1569. 5414

5416 926. Biemans VBC, Sleutjes JAM, de Vries AC, Bodelier AGL, Dijkstra G, Oldenburg B, *et al.*Tofacitinib for ulcerative colitis: results of the prospective Dutch Initiative on Crohn and Colitis
(ICC) registry. *Alimentary pharmacology & therapeutics* 2020, **51**(9): 880-888.

5419

5420 927. Almon E, Shaaltiel Y, Sbeit W, Fich A, Schwartz D, Waterman M, *et al.* Novel Orally 5421 Administered Recombinant Anti-TNF Alpha Fusion Protein for the Treatment of Ulcerative 5422 Colitis: Results From a Phase 2a Clinical Trial. *Journal of clinical gastroenterology* 2021, **55**(2): 5423 134-140.

5424

5425 928. Liu CY, Tam SS, Huang Y, Dubé PE, Alhosh R, Girish N, *et al.* TNF Receptor 1 Promotes Early-5426 Life Immunity and Protects against Colitis in Mice. *Cell reports* 2020, **33**(3): 108275.

5427

5428 929. Lin DP, Jin YL, Hu DY, Ying SJ, Jiang Y. Influence of TRAIL Deficiency on Th17 Cells and Colonic Microbiota in Experimental Colitis Mouse Model. *The American journal of the medical sciences* 2021, **362**(2): 188-197.

5431

5432 930. Chyuan IT, Tsai HF, Wu CS, Hsu PN. TRAIL suppresses gut inflammation and inhibits colitogeic T-cell activation in experimental colitis via an apoptosis-independent pathway. *Mucosal immunology* 2019, **12**(4): 980-989.

5435

5436 931. Pinhu L, Qin Y, Xiong B, You Y, Li J, Sooranna SR. Overexpression of Fas and FasL is associated with infectious complications and severity of experimental severe acute pancreatitis by promoting apoptosis of lymphocytes. *Inflammation* 2014, **37**(4): 1202-1212.

5439

Fig. 1840 Page 1841 Randhi R, Damon M, Dixon KJ. Selective inhibition of soluble TNF using XPro1595 relieves pain and attenuates cerulein-induced pathology in mice. *BMC gastroenterology* 2021, **21**(1): 243.

5442

5443 933. Mao XJ, Zhang XM, Zhang HL, Quezada HC, Mix E, Yang X, *et al.* TNF-alpha receptor 1 deficiency reduces antigen-presenting capacity of Schwann cells and ameliorates experimental autoimmune neuritis in mice. *Neuroscience letters* 2010, **470**(1): 19-23.

5446

5447 934. Taylor JM, Pollard JD. Soluble TNFR1 inhibits the development of experimental autoimmune 5448 neuritis by modulating blood-nerve-barrier permeability and inflammation. *Journal of* 5449 *neuroimmunology* 2007, **183**(1-2): 118-124.

5450

5451 935. Bao L, Lindgren JU, Zhu Y, Ljunggren HG, Zhu J. Exogenous soluble tumor necrosis factor receptor type I ameliorates murine experimental autoimmune neuritis. *Neurobiology of disease* 2003, **12**(1): 73-81.

5455 936. Lu MO, Duan RS, Quezada HC, Chen ZG, Mix E, Jin T, *et al.* Aggravation of experimental autoimmune neuritis in TNF-alpha receptor 1 deficient mice. *Journal of neuroimmunology* 2007, **186**(1-2): 19-26.

5458

5459 937. Kaaij MH, Rip J, Jeucken KCM, Kan YY, van Rooijen CCN, Saris J, *et al.* Overexpression of Transmembrane TNF Drives Development of Ectopic Lymphoid Structures in the Bone Marrow and B Cell Lineage Alterations in Experimental Spondyloarthritis. *Journal of immunology* (*Baltimore*, *Md* : 1950) 2021, **207**(9): 2337-2346.

5463

5464 938. Chen S, Lin Z, Xi L, Zheng Y, Zhou Q, Chen X. Differential role of TNFR1 and TNFR2 in the development of imiquimod-induced mouse psoriasis. *Journal of leukocyte biology* 2021, **110**(6): 1047-1055.

5467 5468

5468 939. Yu X, Li L, Li Q, Zang X, Liu Z. TRAIL and DR5 promote thyroid follicular cell apoptosis in iodine excess-induced experimental autoimmune thyroiditis in NOD mice. *Biological trace element research* 2011, **143**(2): 1064-1076.

5471

Fang Y, Sharp GC, Yagita H, Braley-Mullen H. A critical role for TRAIL in resolution of granulomatous experimental autoimmune thyroiditis. *The Journal of pathology* 2008, **216**(4): 505-513.

5475

5476 941. Wei Y, Chen K, Sharp GC, Braley-Mullen H. Fas ligand is required for resolution of granulomatous experimental autoimmune thyroiditis. *Journal of immunology (Baltimore, Md:* 1950) 2004, **173**(12): 7615-7621.

5479

5480 942. Wang SH, Chen GH, Fan Y, Van Antwerp M, Baker JR, Jr. Tumor necrosis factor-related apoptosis-inducing ligand inhibits experimental autoimmune thyroiditis by the expansion of CD4+CD25+ regulatory T cells. *Endocrinology* 2009, **150**(4): 2000-2007.

5483

5484 943. Wang SH, Cao Z, Wolf JM, Van Antwerp M, Baker JR, Jr. Death ligand tumor necrosis factorrelated apoptosis-inducing ligand inhibits experimental autoimmune thyroiditis. *Endocrinology* 2005, **146**(11): 4721-4726.

5487

5488 944. Patankar JV, Müller TM, Kantham S, Acera MG, Mascia F, Scheibe K, *et al.* E-type prostanoid receptor 4 drives resolution of intestinal inflammation by blocking epithelial necroptosis. *Nature cell biology* 2021, **23**(7): 796-807.

5491

5492 945. Kang TB, Jeong JS, Yang SH, Kovalenko A, Wallach D. Caspase-8 deficiency in mouse embryos 5493 triggers chronic RIPK1-dependent activation of inflammatory genes, independently of RIPK3. 5494 *Cell death and differentiation* 2018, **25**(6): 1107-1117. 5495 5496 946. Rajput A, Kovalenko A, Bogdanov K, Yang SH, Kang TB, Kim JC, et al. RIG-I RNA helicase activation of IRF3 transcription factor is negatively regulated by caspase-8-mediated cleavage of 5497 5498 the RIP1 protein. *Immunity* 2011, **34**(3): 340-351. 5499 Laurien L, Nagata M, Schünke H, Delanghe T, Wiederstein JL, Kumari S, et al. 5500 947. Autophosphorylation at serine 166 regulates RIP kinase 1-mediated cell death and inflammation. 5501 *Nature communications* 2020, **11**(1): 1747. 5502 5503 948. 5504 Rickard JA, Anderton H, Etemadi N, Nachbur U, Darding M, Peltzer N, et al. TNFR1-dependent cell death drives inflammation in Sharpin-deficient mice. eLife 2014, 3. 5505 5506 5507 949. Kumari S, Redouane Y, Lopez-Mosqueda J, Shiraishi R, Romanowska M, Lutzmayer S, et al. Sharpin prevents skin inflammation by inhibiting TNFR1-induced keratinocyte apoptosis. eLife 5508 2014, 3. 5509 5510 950. Berger SB, Kasparcova V, Hoffman S, Swift B, Dare L, Schaeffer M, et al. Cutting Edge: RIP1 5511 5512 kinase activity is dispensable for normal development but is a key regulator of inflammation in 5513 SHARPIN-deficient mice. Journal of immunology (Baltimore, Md: 1950) 2014, 192(12): 5476-5514 5480. 5515

5516 951. Taraborrelli L, Peltzer N, Montinaro A, Kupka S, Rieser E, Hartwig T, *et al.* LUBAC prevents lethal dermatitis by inhibiting cell death induced by TNF, TRAIL and CD95L. *Nature communications* 2018, **9**(1): 3910.

5520 952. Mc Guire C, Volckaert T, Wolke U, Sze M, de Rycke R, Waisman A, *et al.* Oligodendrocyte-5521 specific FADD deletion protects mice from autoimmune-mediated demyelination. *Journal of immunology (Baltimore, Md : 1950)* 2010, **185**(12): 7646-7653.

5519

5523

5527

5531

5524 953. Sun J, Hilliard B, Xu L, Chen YH. Essential roles of the Fas-associated death domain in autoimmune encephalomyelitis. *Journal of immunology (Baltimore, Md : 1950)* 2005, **175**(7): 4783-4788.

5528 954. Newton K, Harris AW, Bath ML, Smith KG, Strasser A. A dominant interfering mutant of FADD/MORT1 enhances deletion of autoreactive thymocytes and inhibits proliferation of mature T lymphocytes. *The EMBO journal* 1998, **17**(3): 706-718.

5532 955. Zhang CJ, Jiang M, Zhou H, Liu W, Wang C, Kang Z, *et al.* TLR-stimulated IRAKM activates caspase-8 inflammasome in microglia and promotes neuroinflammation. *The Journal of clinical investigation* 2018, **128**(12): 5399-5412.

5535
5536
956. Allison J, Thomas HE, Catterall T, Kay TW, Strasser A. Transgenic expression of dominantnegative Fas-associated death domain protein in beta cells protects against Fas ligand-induced
apoptosis and reduces spontaneous diabetes in nonobese diabetic mice. *Journal of immunology*(Baltimore, Md: 1950) 2005, **175**(1): 293-301.

Mollah ZU, Wali J, McKenzie MD, Krishnamurthy B, Graham KL, Fynch S, *et al.* The proapoptotic BH3-only protein Bid is dispensable for development of insulitis and diabetes in the non-obese diabetic mouse. *Apoptosis : an international journal on programmed cell death* 2011, **16**(8): 822-830.

5540

5545

5549

5553

5557

5561

5564

5568

5572

Huang QQ, Birkett R, Doyle RE, Haines GK, Perlman H, Shi B, et al. Association of Increased
 F4/80(high) Macrophages With Suppression of Serum-Transfer Arthritis in Mice With Reduced
 FLIP in Myeloid Cells. Arthritis & rheumatology (Hoboken, NJ) 2017, 69(9): 1762-1771.

- 5550 959. Dominguez S, Montgomery AB, Haines GK, 3rd, Bloomfield CL, Cuda CM. The caspase-8/RIPK3 signaling axis in antigen presenting cells controls the inflammatory arthritic response. 5552 Arthritis research & therapy 2017, **19**(1): 224.
- Pearson JS, Giogha C, Ong SY, Kennedy CL, Kelly M, Robinson KS, *et al.* A type III effector antagonizes death receptor signalling during bacterial gut infection. *Nature* 2013, **501**(7466): 247-251.
- Uchiyama R, Yonehara S, Taniguchi S, Ishido S, Ishii KJ, Tsutsui H. Inflammasome and Fas Mediated IL-1β Contributes to Th17/Th1 Cell Induction in Pathogenic Bacterial Infection In
   Vivo. Journal of immunology (Baltimore, Md: 1950) 2017, 199(3): 1122-1130.
- 5562 962. Maudet C, Kheloufi M, Levallois S, Gaillard J, Huang L, Gaultier C, *et al.* Bacterial inhibition of Fas-mediated killing promotes neuroinvasion and persistence. *Nature* 2022, **603**(7903): 900-906.
- 5565 963. Krzyzowska M, Baska P, Orlowski P, Zdanowski R, Winnicka A, Eriksson K, *et al.* HSV-2 regulates monocyte inflammatory response via the Fas/FasL pathway. *PloS one* 2013, **8**(7): e70308.
- 5569 964. O'Donnell JA, Kennedy CL, Pellegrini M, Nowell CJ, Zhang JG, O'Reilly LA, *et al.* Fas regulates 5570 neutrophil lifespan during viral and bacterial infection. *Journal of leukocyte biology* 2015, **97**(2): 5571 321-326.
- 5573 965. Peterson LW, Philip NH, DeLaney A, Wynosky-Dolfi MA, Asklof K, Gray F, *et al.* RIPK1-5574 dependent apoptosis bypasses pathogen blockade of innate signaling to promote immune defense. 5575 *The Journal of experimental medicine* 2017, **214**(11): 3171-3182.

5576 5577 966. Weng D, Marty-Roix R, Ganesan S, Proulx MK, Vladimer GI, Kaiser WJ, et al. Caspase-8 and RIP kinases regulate bacteria-induced innate immune responses and cell death. *Proceedings of* 5578 5579 the National Academy of Sciences of the United States of America 2014, 111(20): 7391-7396. 5580 5581 967. DeLaney AA, Berry CT, Christian DA, Hart A, Bjanes E, Wynosky-Dolfi MA, et al. Caspase-8 promotes c-Rel-dependent inflammatory cytokine expression and resistance against Toxoplasma 5582 gondii. Proceedings of the National Academy of Sciences of the United States of America 2019, 5583 **116**(24): 11926-11935. 5584 5585

5586 968. Kuriakose T, Man SM, Malireddi RK, Karki R, Kesavardhana S, Place DE, *et al.* ZBP1/DAI is an innate sensor of influenza virus triggering the NLRP3 inflammasome and programmed cell death pathways. *Sci Immunol* 2016, **1**(2).

Thapa RJ, Ingram JP, Ragan KB, Nogusa S, Boyd DF, Benitez AA, et al. DAI Senses Influenza
 A Virus Genomic RNA and Activates RIPK3-Dependent Cell Death. Cell host & microbe 2016,
 20(5): 674-681.

5589

5593

5597

5601

5605

5609

5613

5594 970. Place DE, Christgen S, Tuladhar S, Vogel P, Malireddi RKS, Kanneganti TD. Hierarchical Cell Death Program Disrupts the Intracellular Niche Required for Burkholderia thailandensis Pathogenesis. *mBio* 2021, **12**(3): e0105921.

- 5598 971. Alikhani M, Alikhani Z, He H, Liu R, Popek BI, Graves DT. Lipopolysaccharides indirectly stimulate apoptosis and global induction of apoptotic genes in fibroblasts. *The Journal of biological chemistry* 2003, **278**(52): 52901-52908.
- 5602 972. Sarid R, Ben-Moshe T, Kazimirsky G, Weisberg S, Appel E, Kobiler D, *et al.* vFLIP protects 5603 PC-12 cells from apoptosis induced by Sindbis virus: implications for the role of TNF-alpha. *Cell death and differentiation* 2001, **8**(12): 1224-1231.
- 973. Qian Z, Shuying W, Ranran D. Inhibitory effects of JQ1 on listeria monocytogenes-induced acute liver injury by blocking BRD4/RIPK1 axis. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* 2020, **125:** 109818.
- 5610 974. Kim H, Rhee SH, Pothoulakis C, Lamont JT. Inflammation and apoptosis in Clostridium difficile enteritis is mediated by PGE2 up-regulation of Fas ligand. *Gastroenterology* 2007, **133**(3): 875-886.
- 5614 975. He BL, Yuan JM, Yang LY, Xie JF, Weng SP, Yu XQ, *et al.* The viral TRAF protein (ORF111L) from infectious spleen and kidney necrosis virus interacts with TRADD and induces caspase 8-mediated apoptosis. *PloS one* 2012, **7**(5): e37001.

5617 976. 5618 Mandal P, Feng Y, Lyons JD, Berger SB, Otani S, DeLaney A, et al. Caspase-8 Collaborates with Caspase-11 to Drive Tissue Damage and Execution of Endotoxic Shock. *Immunity* 2018, **49**(1): 5619 5620 42-55.e46. 5621 Karki R, Sharma BR, Tuladhar S, Williams EP, Zalduondo L, Samir P, et al. Synergism of TNF-5622 977. α and IFN-γ Triggers Inflammatory Cell Death, Tissue Damage, and Mortality in SARS-CoV-2 5623 Infection and Cytokine Shock Syndromes. Cell 2021, 184(1): 149-168.e117. 5624

5626 978. Langen RC, Van Der Velden JL, Schols AM, Kelders MC, Wouters EF, Janssen-Heininger YM.
Tumor necrosis factor-alpha inhibits myogenic differentiation through MyoD protein
destabilization. FASEB journal: official publication of the Federation of American Societies for
Experimental Biology 2004, **18**(2): 227-237.

5625

5630

5633

5637

5640

5644

5648

5652

5656

5631 979. Kim D, Singh N, Waldemer-Streyer RJ, Yoon MS, Chen J. Muscle-derived TRAIL negatively regulates myogenic differentiation. *Experimental cell research* 2020, **394**(1): 112165.

5634 980. Zhang R, Wang L, He L, Yang B, Yao C, Du P, *et al.* Fas-Associated Protein with Death Domain Regulates Notch Signaling during Muscle Regeneration. *Cells, tissues, organs* 2014, **200**(3-4): 253-264.

5638 981. Chen SE, Jin B, Li YP. TNF-alpha regulates myogenesis and muscle regeneration by activating p38 MAPK. *American journal of physiology Cell physiology* 2007, **292**(5): C1660-1671.

5641 982. Chen SE, Gerken E, Zhang Y, Zhan M, Mohan RK, Li AS, *et al.* Role of TNF-{alpha} signaling in regeneration of cardiotoxin-injured muscle. *American journal of physiology Cell physiology* 2005, **289**(5): C1179-1187.

Dufresne SS, Boulanger-Piette A, Bossé S, Argaw A, Hamoudi D, Marcadet L, *et al.* Genetic deletion of muscle RANK or selective inhibition of RANKL is not as effective as full-length OPG-fc in mitigating muscular dystrophy. *Acta Neuropathol Commun* 2018, **6**(1): 31.

Alger HM, Raben N, Pistilli E, Francia DL, Rawat R, Getnet D, *et al.* The role of TRAIL in mediating autophagy in myositis skeletal muscle: a potential nonimmune mechanism of muscle damage. *Arthritis and rheumatism* 2011, **63**(11): 3448-3457.

5653 985. Kondo M, Murakawa Y, Harashima N, Kobayashi S, Yamaguchi S, Harada M. Roles of proinflammatory cytokines and the Fas/Fas ligand interaction in the pathogenesis of inflammatory myopathies. *Immunology* 2009, **128**(1 Suppl): e589-599.

- 5657 986. Del Sorbo L, Costamagna A, Muraca G, Rotondo G, Civiletti F, Vizio B, *et al.* Intratracheal Administration of Small Interfering RNA Targeting Fas Reduces Lung Ischemia-Reperfusion Injury. *Critical care medicine* 2016, **44**(8): e604-613.
- 5661 987. An S, Hishikawa Y, Liu J, Koji T. Lung injury after ischemia-reperfusion of small intestine in rats involves apoptosis of type II alveolar epithelial cells mediated by TNF-alpha and activation of Bid pathway. *Apoptosis : an international journal on programmed cell death* 2007, **12**(11): 1989-2001.

5665

5669

5673

5678

5681

5685

- Patel BV, Wilson MR, O'Dea KP, Takata M. TNF-induced death signaling triggers alveolar epithelial dysfunction in acute lung injury. *Journal of immunology (Baltimore, Md : 1950)* 2013, **190**(8): 4274-4282.
- 5670 989. Wilson MR, Wakabayashi K, Bertok S, Oakley CM, Patel BV, O'Dea KP, *et al.* Inhibition of TNF Receptor p55 By a Domain Antibody Attenuates the Initial Phase of Acid-Induced Lung Injury in Mice. *Frontiers in immunology* 2017, **8:** 128.
- Bohr A, Tsapis N, Foged C, Andreana I, Yang M, Fattal E. Treatment of acute lung inflammation
   by pulmonary delivery of anti-TNF-α siRNA with PAMAM dendrimers in a murine model.
   European journal of pharmaceutics and biopharmaceutics: official journal of
   Arbeitsgemeinschaft fur Pharmazeutische Verfahrenstechnik eV 2020, 156: 114-120.
- 5679 991. Lai WY, Wang JW, Huang BT, Lin EP, Yang PC. A Novel TNF-α-Targeting Aptamer for TNF 5680 α-Mediated Acute Lung Injury and Acute Liver Failure. *Theranostics* 2019, 9(6): 1741-1751.
- 5682 992. Proudfoot A, Bayliffe A, O'Kane CM, Wright T, Serone A, Bareille PJ, *et al.* Novel anti-tumour necrosis factor receptor-1 (TNFR1) domain antibody prevents pulmonary inflammation in experimental acute lung injury. *Thorax* 2018, **73**(8): 723-730.
- Bohr A, Tsapis N, Andreana I, Chamarat A, Foged C, Delomenie C, et al. Anti-Inflammatory
   Effect of Anti-TNF-α SiRNA Cationic Phosphorus Dendrimer Nanocomplexes Administered
   Intranasally in a Murine Acute Lung Injury Model. Biomacromolecules 2017, 18(8): 2379-2388.
- Weifeng Y, Li L, Yujie H, Weifeng L, Zhenhui G, Wenjie H. Inhibition of Acute Lung Injury by
   TNFR-Fc through Regulation of an Inflammation-Oxidative Stress Pathway. *PloS one* 2016,
   11(3): e0151672.
- 5693
  5694
  995. Cakarova L, Marsh LM, Wilhelm J, Mayer K, Grimminger F, Seeger W, *et al.* Macrophage tumor
  5695 necrosis factor-alpha induces epithelial expression of granulocyte-macrophage colony5696 stimulating factor: impact on alveolar epithelial repair. *American journal of respiratory and*5697 *critical care medicine* 2009, **180**(6): 521-532.

5698
 5699
 5690
 Matute-Bello G, Winn RK, Martin TR, Liles WC. Sustained lipopolysaccharide-induced lung inflammation in mice is attenuated by functional deficiency of the Fas/Fas ligand system. *Clinical and diagnostic laboratory immunology* 2004, 11(2): 358-361.

5703 997. Janssen WJ, Barthel L, Muldrow A, Oberley-Deegan RE, Kearns MT, Jakubzick C, *et al.* Fas determines differential fates of resident and recruited macrophages during resolution of acute lung injury. *American journal of respiratory and critical care medicine* 2011, **184**(5): 547-560.

5702

5706

5710

5715

5719

5723

5728

5733

- 998. Qian L, Yin X, Ji J, Chen Z, Fang H, Li H, *et al.* Tumor necrosis factor-α small interfering RNA alveolar epithelial cell-targeting nanoparticles reduce lung injury in C57BL/6J mice with sepsis.
   709 *The Journal of international medical research* 2021, 49(1): 300060520984652.
- Weckbach S, Hohmann C, Denk S, Kellermann P, Huber-Lang MS, Baumann B, *et al.* Apoptotic and inflammatory signaling via Fas and tumor necrosis factor receptor I contribute to the development of chest trauma-induced septic acute lung injury. *The journal of trauma and acute care surgery* 2013, **74**(3): 792-800.
- 5716 1000. Thakkar RK, Chung CS, Chen Y, Monaghan SF, Lomas-Neira J, Heffernan DS, *et al.* Local tissue expression of the cell death ligand, fas ligand, plays a central role in the development of extrapulmonary acute lung injury. *Shock (Augusta, Ga)* 2011, **36**(2): 138-143.
- 5720 1001. Perl M, Chung CS, Perl U, Lomas-Neira J, de Paepe M, Cioffi WG, *et al.* Fas-induced pulmonary apoptosis and inflammation during indirect acute lung injury. *American journal of respiratory and critical care medicine* 2007, **176**(6): 591-601.
- 5724 1002. Perl M, Chung CS, Lomas-Neira J, Rachel TM, Biffl WL, Cioffi WG, *et al.* Silencing of Fas, but not caspase-8, in lung epithelial cells ameliorates pulmonary apoptosis, inflammation, and neutrophil influx after hemorrhagic shock and sepsis. *The American journal of pathology* 2005, **167**(6): 1545-1559.
- 5729 1003. Messer MP, Kellermann P, Weber SJ, Hohmann C, Denk S, Klohs B, *et al.* Silencing of fas, fasassociated via death domain, or caspase 3 differentially affects lung inflammation, apoptosis, and development of trauma-induced septic acute lung injury. *Shock (Augusta, Ga)* 2013, **39**(1): 19-27.
- 5734 1004. Matsuda N, Yamamoto S, Takano K, Kageyama S, Kurobe Y, Yoshihara Y, *et al.* Silencing of fas-associated death domain protects mice from septic lung inflammation and apoptosis.

  5736 *American journal of respiratory and critical care medicine* 2009, **179**(9): 806-815.

- 5738 1005. Ehrhardt H, Pritzke T, Oak P, Kossert M, Biebach L, Förster K, *et al.* Absence of TNF-α enhances inflammatory response in the newborn lung undergoing mechanical ventilation. *American journal of physiology Lung cellular and molecular physiology* 2016, **310**(10): L909-918.
- 5742 1006. Mao Q, Gundavarapu S, Patel C, Tsai A, Luks FI, De Paepe ME. The Fas system confers 5743 protection against alveolar disruption in hyperoxia-exposed newborn mice. *American journal of* 5744 respiratory cell and molecular biology 2008, **39**(6): 717-729.

5745

5748

5752

5756

5760

5764

5768

5772

- 5746 Guthmann F, Wissel H, Rüstow B. Early subcutaneous administration of etanercept (Enbrel) prevents from hyperoxia-induced lung injury. *Experimental lung research* 2009, **35**(9): 770-780.
- 5749 1008. Kaya G, Saldir M, Polat A, Fidanci MK, Erdem A, Erdem G, et al. Evaluation of Etanercept 5750 Treatment in Newborn Rat Model with Hyperoxic Lung Injury. Fetal and pediatric pathology 5751 2016, **35**(5): 327-338.
- 5753 1009. Wolthuis EK, Vlaar AP, Choi G, Roelofs JJ, Haitsma JJ, van der Poll T, *et al.* Recombinant human soluble tumor necrosis factor-alpha receptor fusion protein partly attenuates ventilator-induced lung injury. *Shock (Augusta, Ga)* 2009, **31**(3): 262-266.
- 5757 1010. Pryhuber GS, O'Brien DP, Baggs R, Phipps R, Huyck H, Sanz I, *et al.* Ablation of tumor necrosis 5758 factor receptor type I (p55) alters oxygen-induced lung injury. *American journal of physiology* 5759 *Lung cellular and molecular physiology* 2000, **278**(5): L1082-1090.
- 5761 1011. Redente EF, Chakraborty S, Sajuthi S, Black BP, Edelman BL, Seibold MA, *et al.* Loss of Fas signaling in fibroblasts impairs homeostatic fibrosis resolution and promotes persistent pulmonary fibrosis. *JCI insight* 2020, **6**(1).
- 5765 1012. Hao Z, Hampel B, Yagita H, Rajewsky K. T cell-specific ablation of Fas leads to Fas ligand-5766 mediated lymphocyte depletion and inflammatory pulmonary fibrosis. *The Journal of* 5767 *experimental medicine* 2004, **199**(10): 1355-1365.
- 5769 1013. Aoshiba K, Yasui S, Tamaoki J, Nagai A. The Fas/Fas-ligand system is not required for bleomycin-induced pulmonary fibrosis in mice. *American journal of respiratory and critical care medicine* 2000, **162**(2 Pt 1): 695-700.
- 5773 1014. Kuwano K, Hagimoto N, Kawasaki M, Yatomi T, Nakamura N, Nagata S, *et al.* Essential roles 5774 of the Fas-Fas ligand pathway in the development of pulmonary fibrosis. *The Journal of clinical* 5775 *investigation* 1999, **104**(1): 13-19.

1015. Redente EF, Keith RC, Janssen W, Henson PM, Ortiz LA, Downey GP, et al. Tumor necrosis factor-α accelerates the resolution of established pulmonary fibrosis in mice by targeting profibrotic lung macrophages. American journal of respiratory cell and molecular biology 2014, (4): 825-837.

1016. Oikonomou N, Harokopos V, Zalevsky J, Valavanis C, Kotanidou A, Szymkowski DE, et al. Soluble TNF mediates the transition from pulmonary inflammation to fibrosis. *PloS one* 2006, (1): e108. 

1017. Kuroki M, Noguchi Y, Shimono M, Tomono K, Tashiro T, Obata Y, et al. Repression of bleomycin-induced pneumopathy by TNF. Journal of immunology (Baltimore, Md: 1950) 2003, (1): 567-574. 

1018. Collison AM, Li J, de Siqueira AP, Lv X, Toop HD, Morris JC, et al. TRAIL signals through the ubiquitin ligase MID1 to promote pulmonary fibrosis. BMC pulmonary medicine 2019, 19(1): 31.

1019. McGrath EE, Lawrie A, Marriott HM, Mercer P, Cross SS, Arnold N, et al. Deficiency of tumour necrosis factor-related apoptosis-inducing ligand exacerbates lung injury and fibrosis. Thorax 2012, **67**(9): 796-803. 

1020. Malaviya R, Sunil VR, Venosa A, Verissimo VL, Cervelli JA, Vayas KN, et al. Attenuation of Nitrogen Mustard-Induced Pulmonary Injury and Fibrosis by Anti-Tumor Necrosis Factor-α Antibody. Toxicological sciences: an official journal of the Society of Toxicology 2015, 148(1): 71-88.

1021. Tan J, Ni X. TNF-α antagonist may not be suitable for severe rituximab-induced interstitial lung disease. Journal of clinical pharmacy and therapeutics 2015, 40(3): 249-250.

1022. Santos LD, Antunes KH, Muraro SP, de Souza GF, da Silva AG, Felipe JS, et al. TNF-mediated alveolar macrophage necroptosis drives disease pathogenesis during respiratory syncytial virus infection. The European respiratory journal 2021, 57(6). 

1023. Morris DR, Ansar M, Ivanciuc T, Qu Y, Casola A, Garofalo RP. Selective Blockade of TNFR1 Improves Clinical Disease and Bronchoconstriction in Experimental RSV Infection. Viruses 2020, **12**(10). 

1024. Nguyen TH, Maltby S, Simpson JL, Eyers F, Baines KJ, Gibson PG, et al. TNF-α and Macrophages Are Critical for Respiratory Syncytial Virus-Induced Exacerbations in a Mouse Model of Allergic Airways Disease. Journal of immunology (Baltimore, Md: 1950) 2016, (9): 3547-3558.

5818 1025. van den Berg E, van Woensel JB, Bos AP, Bem RA, Altemeier WA, Gill SE, et al. Role of the 5819 Fas/FasL system in a model of RSV infection in mechanically ventilated mice. American journal 5820 5821 of physiology Lung cellular and molecular physiology 2011, **301**(4): L451-460. 5822 1026. Lopez AD, Avasarala S, Grewal S, Murali AK, London L. Differential role of the Fas/Fas ligand 5823 apoptotic pathway in inflammation and lung fibrosis associated with reovirus 1/L-induced 5824 bronchiolitis obliterans organizing pneumonia and acute respiratory distress syndrome. Journal 5825 of immunology (Baltimore, Md: 1950) 2009, **183**(12): 8244-8257. 5826 5827 1027. Bem RA, Bos AP, Wösten-van Asperen RM, Bruijn M, Lutter R, Sprick MR, et al. Potential role 5828 of soluble TRAIL in epithelial injury in children with severe RSV infection. American journal of 5829 respiratory cell and molecular biology 2010, 42(6): 697-705. 5830 5831 5832 1028. Neuzil KM, Tang YW, Graham BS. Protective Role of TNF-alpha in respiratory syncytial virus 5833 infection in vitro and in vivo. The American journal of the medical sciences 1996, 311(5): 201-5834 204. 5835 1029. Pant K, Chandrasekaran A, Chang CJ, Vageesh A, Popkov AJ, Weinberg JB. Effects of tumor 5836 necrosis factor on viral replication and pulmonary inflammation during acute mouse adenovirus 5837 type 1 respiratory infection. Virology 2020, 547: 12-19. 5838 5839 5840 1030. Adkins LJ, Molloy CT, Weinberg JB. Fas activity mediates airway inflammation during mouse adenovirus type 1 respiratory infection. Virology 2018, **521**: 129-137. 5841 5842 5843 1031. Li XM, Chen X, Gu W, Guo YJ, Cheng Y, Peng J, et al. Impaired TNF/TNFR2 signaling 5844 enhances Th2 and Th17 polarization and aggravates allergic airway inflammation. American 5845 journal of physiology Lung cellular and molecular physiology 2017, 313(3): L592-1601. 5846 5847 1032. Starkhammar M, Kumlien Georén S, Dahlén SE, Cardell LO, Adner M. TNFα-blockade stabilizes local airway hyperresponsiveness during TLR-induced exacerbations in murine model 5848 of asthma. Respiratory research 2015, 16: 129. 5849 5850 1033. Faustino L, Fonseca DM, Florsheim EB, Resende RR, Lepique AP, Faquim-Mauro E, et al. 5851 Tumor necrosis factor-related apoptosis-inducing ligand mediates the resolution of allergic 5852 5853 airway inflammation induced by chronic allergen inhalation. Mucosal immunology 2014, 7(5): 1199-1208. 5854 5855 5856 1034. Yilmaz O, Karaman M, Bagriyanik HA, Firinci F, Kiray M, Turkeli A, et al. Comparison of TNF

antagonism by etanercept and dexamethasone on airway epithelium and remodeling in an

experimental model of asthma. *International immunopharmacology* 2013, **17**(3): 768-773.

5859
5860
1035. Sharma SK, Almeida FA, Kierstein S, Hortobagyi L, Lin T, Larkin A, *et al.* Systemic FasL neutralization increases eosinophilic inflammation in a mouse model of asthma. *Allergy* 2012, 67(3): 328-335.

5863
5864
1036. Hwang SJ, Kim HS, Chung DH. Fas/Fas ligand-mediated apoptosis promotes hypersensitivity

5864 1036. Hwang SJ, Kim HS, Chung DH. Fas/Fas figand-mediated apoptosis promotes hypersensitivity pneumonitis in mice by enhancing maturation of dendritic cells. *American journal of respiratory and critical care medicine* 2010, **181**(11): 1250-1261.

5868 1037. Weckmann M, Collison A, Simpson JL, Kopp MV, Wark PA, Smyth MJ, *et al.* Critical link 5869 between TRAIL and CCL20 for the activation of TH2 cells and the expression of allergic airway 5870 disease. *Nature medicine* 2007, **13**(11): 1308-1315.

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5872 1038. Chuang YH, Suen JL, Chiang BL. Fas-ligand-expressing adenovirus-transfected dendritic cells decrease allergen-specific T cells and airway inflammation in a murine model of asthma. *Journal of molecular medicine (Berlin, Germany)* 2006, **84**(7): 595-603.

5876 1039. Broide DH, Stachnick G, Castaneda D, Nayar J, Sriramarao P. Inhibition of eosinophilic inflammation in allergen-challenged TNF receptor p55/p75--and TNF receptor p55-deficient mice. *American journal of respiratory cell and molecular biology* 2001, **24**(3): 304-311.

5880 1040. Whitehead GS, Thomas SY, Shalaby KH, Nakano K, Moran TP, Ward JM, *et al.* TNF is required for TLR ligand-mediated but not protease-mediated allergic airway inflammation. *The Journal of clinical investigation* 2017, **127**(9): 3313-3326.

5884 1041. Maillet I, Schnyder-Candrian S, Couillin I, Quesniaux VF, Erard F, Moser R, *et al.* Allergic lung inflammation is mediated by soluble tumor necrosis factor (TNF) and attenuated by dominant-negative TNF biologics. *American journal of respiratory cell and molecular biology* 2011, **45**(4): 731-739.

5889 1042. Choi IW, Sun K, Kim YS, Ko HM, Im SY, Kim JH, *et al.* TNF-alpha induces the late-phase airway hyperresponsiveness and airway inflammation through cytosolic phospholipase A(2) activation. *The Journal of allergy and clinical immunology* 2005, **116**(3): 537-543.

5893 1043. Hildebrandt GC, Olkiewicz KM, Corrion L, Clouthier SG, Pierce EM, Liu C, *et al.* A role for TNF receptor type II in leukocyte infiltration into the lung during experimental idiopathic pneumonia syndrome. *Biology of blood and marrow transplantation : journal of the American Society for Blood and Marrow Transplantation* 2008, **14**(4): 385-396.

5898 5899 5900	1044.	Wu Y, Shen Y, Zhang J, Wan C, Wang T, Xu D, <i>et al.</i> Increased serum TRAIL and DR5 levels correlated with lung function and inflammation in stable COPD patients. <i>International journal of chronic obstructive pulmonary disease</i> 2015, <b>10:</b> 2405-2412.
5901 5902 5903 5904	1045.	Haw TJ, Starkey MR, Nair PM, Pavlidis S, Liu G, Nguyen DH, <i>et al.</i> A pathogenic role for tumor necrosis factor-related apoptosis-inducing ligand in chronic obstructive pulmonary disease. <i>Mucosal immunology</i> 2016, <b>9</b> (4): 859-872.
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### **Legends to Figures**

Figure 1. Principal causes of the therapeutic failure of intrinsic or extrinsic apoptosis inhibitors.

The clinical development and success of agents inhibiting apoptosis is limited by multiple contributory causes, including potential non-apoptotic, accessory or even protective roles of the targeted proteins (exemplified by the involvement of certain BCL2 family members, caspases and death receptors in processes as diverse as inflammation, cell differentiation, cell proliferation and cell survival), the high interconnectivity between RCD pathway (potentially leading to the activation of compensatory RCD variants in response to the inhibition of a specific RCD type), the low specificity and selectivity of the inhibitors developed so far (exemplified by the broad-spectrum caspase inhibitors) and the difficulty to precisely determine and quantify cell death *in vivo*. RCD, regulated cell death.

Figure 2. Molecular machinery of the intrinsic apoptosis. Intrinsic apoptosis can be activated by a range of extracellular or intracellular stimuli, including, but not limited to, DNA damage, endoplasmic reticulum (ER) or oxidative stress, growth factor withdrawal or microtubular alterations. The critical step of the intrinsic apoptosis is the activation of the pro-apoptotic effectors of the BCL2 family, BAX, BAK and possibly BOK, which drives the outer membrane permeabilization (MOMP) and commits cells to death. MOMP results in the release from the mitochondrial intermembrane space into the cytosol of proapoptotic proteins, including CYCS and SMAC. CYCS assembles with APAF1, dATP and pro-CASP9 into the apoptosome, leading to the activation of CASP9, which in turn promotes the activation of the executioner caspases CASP3 and CASP7. The activation of the executioner caspases is facilitated by SMAC, which sequesters and/or degrades members of IAP family that inhibit apoptosis.

Figure 3. Impact of intrinsic apoptosis players on neurological disorders. Intrinsic apoptosis is directly or indirectly involved in the pathogenesis of multiple neurological disorders, including neurodegenerative diseases, such as AD and PD, in brain damage caused by traumatic injury or

5931 neurotoxicity as well as in neuromuscular and retinal disorders. Pro- and anti-apoptotic members of the BCL2 family are depicted, respectively, in blue and green, while caspases are illustrated in pale violet. 5932 5933 Figure 4. Molecular machinery of the extrinsic apoptosis pathway. Extrinsic apoptosis is initiated by 5934 the binding of FASL to FAS or TRAIL to TRAIL-R1 or TRAIL-R2, which promotes the assembly, on 5935 the cytoplasmic tail of these death receptors, of a platform known as the DISC. Extrinsic apoptosis is 5936 also triggered by the binding of TNF to TNF-R1, which promotes the assembly of the Complex II. The 5937 DISC comprises FADD, c-FLIPs and pro-CASP8. Complex II is a platform consisting of FADD and 5938 pro-CASP8 in association with either TRADD (complex IIa) or RIPK1 (complex IIb). The assembly of these complexes promotes the activation of CASP8, which 5939 mediates CASP3 and CASP7 activation either directly, by catalyzing the proteolytic activation of CASP3 5940 5941 and CASP7 (in type I cells) or indirectly, via the proteolytic activation of the BH3-only protein BID and 5942 the outer membrane permeabilization (MOMP) (in type II cells). Extrinsic apoptosis can also be induced 5943 by dependence receptors like DCC, NTRK3, PTCH1, or UNC5A-D, which are activated by decreased 5944 concentration of the related ligand, as illustrated in the figure. However, the role of this pathway in normal physiology and disease is not yet established. 5945 5946 Figure 5. Impact of extrinsic apoptosis players on neurological disorders. Death receptor-induced 5947

apoptosis is directly or indirectly involved in the pathogenesis of multiple neurological disorders, including neurodegenerative diseases, such as AD and PD, in brain damage due to traumatic injury or neurotoxicity as well as in neuromuscular and retinal disorders.

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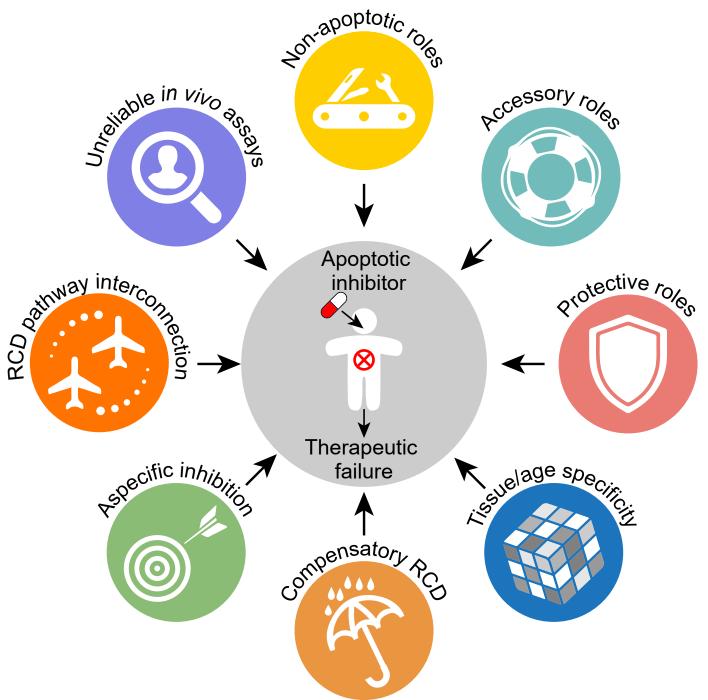


Figure 1

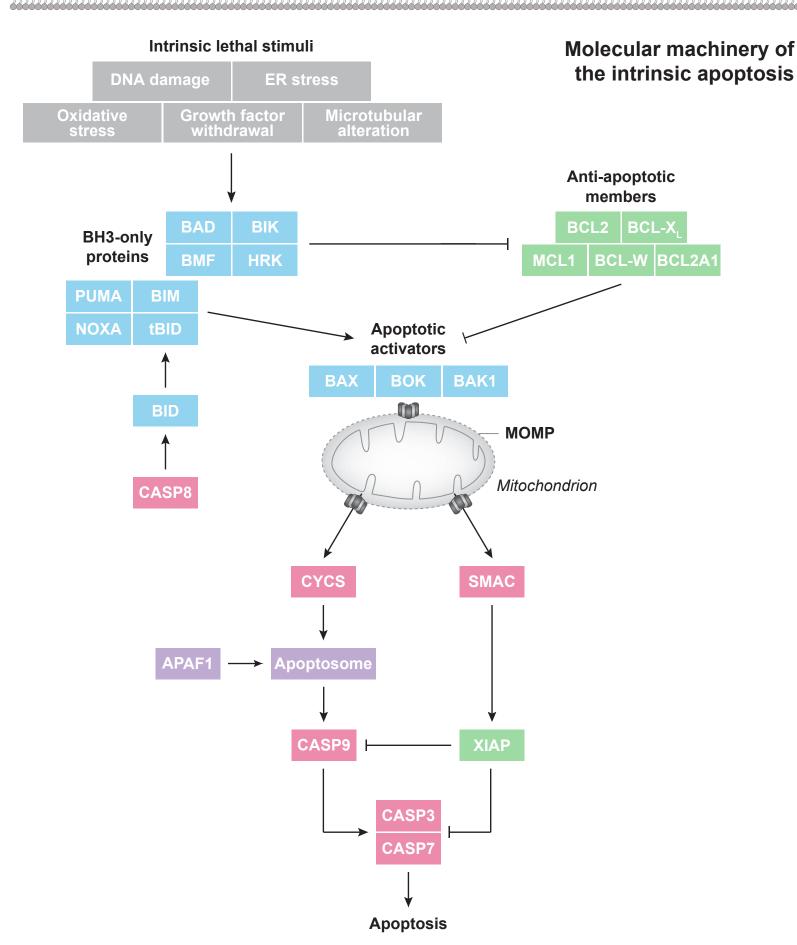


Figure 2

# **Neurodegenerative disorders**

**PUMA** 

**BAX** 

CASP3

#### **Neuromuscular disorders**

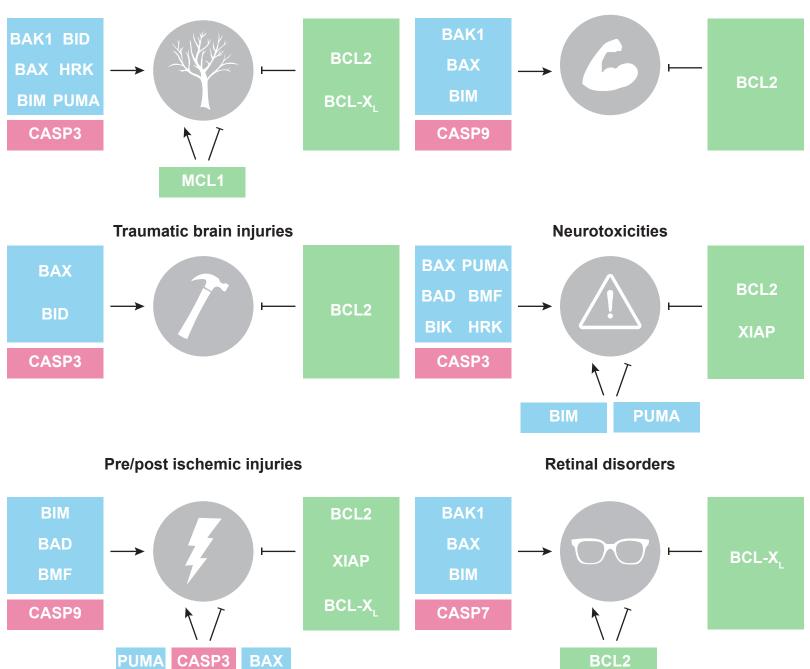


Figure 3

# Molecular machinery of the extrinsic apoptosis

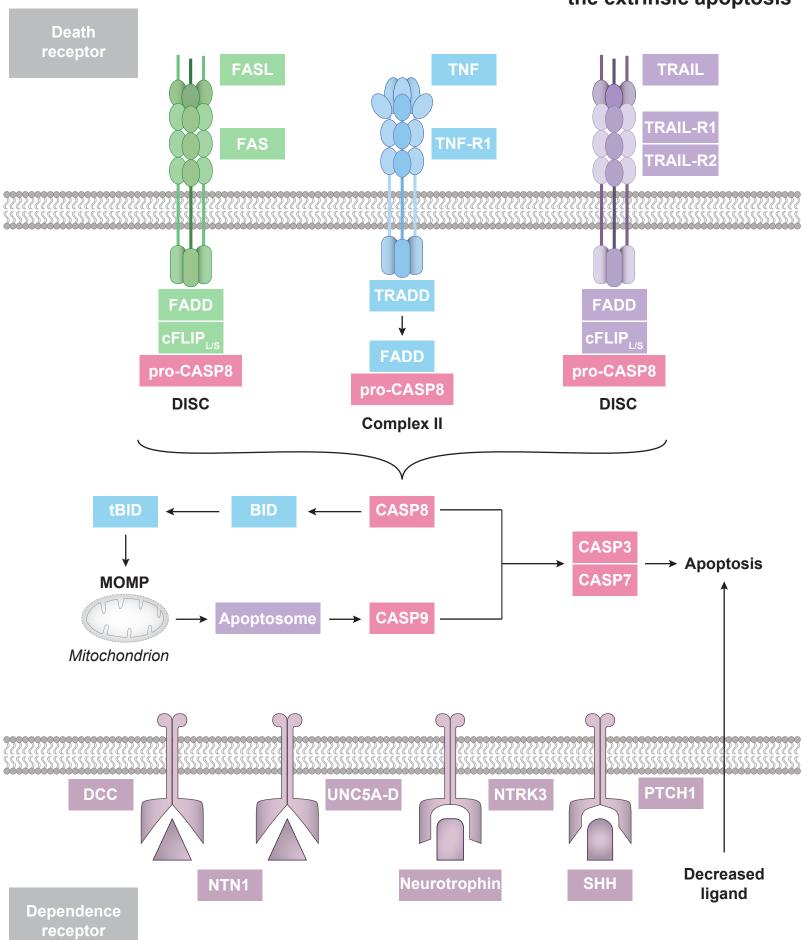


Figure 4

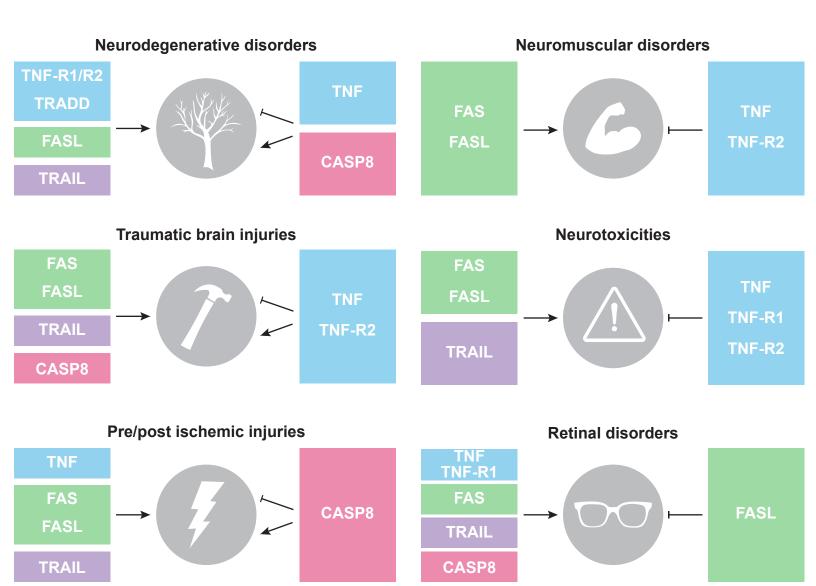


Figure 5

# **Box 1.** Principle of intrinsic apoptosis.

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Intrinsic apoptosis is a type of regulated cell death (RCD) initiated by perturbations of the extracellular 2 3 or intracellular microenvironment including (but not limited to) DNA damage, endoplasmic reticulum or oxidative stress, growth factor withdrawal, microtubular alteration. The critical step is mitochondrial 4 outer membrane permeabilization (MOMP) <sup>1,2,3,4</sup>. MOMP - which involves constitutive outer membrane 5 6 proteins, such as the voltage-dependent anion channel (VDAC), is modulated by the activity of multiple pro-apoptotic and anti-apoptotic members of the BCL2, apoptosis regulator (BCL2) protein family <sup>5, 6, 7,</sup> 7 <sup>8,9</sup>. In response to apoptotic stimuli, MOMP leads to the sequential activation of the initiator caspase 9 8 (CASP9) and then the executioner caspases CASP3 and CASP7 10, 11, 12, 13, 14. Two functionally distinct 9 classes of pro-apoptotic BCL2 proteins have been identified. The first class encompasses the apoptotic 10 activators BCL2 associated X, apoptosis regulator (BAX), BCL2 antagonist/killer 1 (BAK1), and BCL2 11 family apoptosis regulator (BOK) <sup>15</sup>. Once activated by apoptotic stimuli, BAX, BAK1 and BOK induce 12 MOMP by generating pores across the outer mitochondrial membrane (OMM) <sup>16, 17, 18, 19, 20</sup>. These pro-13 apoptotic factors promote the release into the cytosol of several apoptogenic factors, including 14 15 cytochrome c, somatic (CYCS) and diablo IAP-binding mitochondrial protein (DIABLO; also known as second mitochondrial activator of caspases, SMAC) <sup>21</sup>. CYCS exerts apoptogenic activity by associating 16 17 with apoptotic peptidase activating factor 1 (APAF1) and pro-CASP9 to generate a complex known as the apoptosome, leading to sequential activation of CASP9 and the executioner caspases CASP3 and 18 CASP7<sup>22</sup>. DIABLO/SMAC contributes to CASP3 and CASP7 activation by associating with and 19 20 inhibiting X-linked inhibitor of apoptosis (XIAP) and other members of the inhibitor of apoptosis (IAP) protein family which restrain caspase activation <sup>23</sup>. 21 The second class of pro-apoptotic BCL2 proteins (known as BH3-only proteins <sup>24</sup>) include BCL2 22

associated agonist of cell death (BAD), BCL2 binding component 3 (BBC3; best known as p53-

upregulated modulator of apoptosis, PUMA), BCL2 interacting killer (BIK), BCL2 like 11 (BCL2L11; best known as BCL2-interacting mediator of cell death, BIM), Bcl2 modifying factor (BMF), BH3 interacting domain death agonist (BID), BCL2 interacting protein harakiri (HRK, also known as DP5), and phorbol-12-myristate-13-acetate-induced protein 1 (PMAIP1; best known as NOXA <sup>25, 26</sup>). Of these, caspase-cleaved BID (tBID), BIM, PUMA, and NOXA have been reported to also be able to promote BAX and BAK1 activation through a direct interaction with these proteins at the mitochondria <sup>27, 28, 29, 30</sup>, <sup>31, 32, 33</sup>. All BH3-only proteins, including BAD, BIK, BMF and HRK activate BAX and BAK1 indirectly by associating with anti-apoptotic BCL2 family members, thereby blocking the inhibitory binding of the latter to BAX and BAK1 5, 9, 31, 33, 34, 35. Some BH3-only proteins, particularly BIM, PUMA and tBID, can potently bind and inhibit all anti-apoptotic BCL-2 proteins whereas others bind only some (e.g., NOXA only binds MCL1 and A1) <sup>31, 33, 36, 37</sup>. It is noteworthy that BAX and BAK1 can induce apoptosis in the absence of all BH3-only proteins when the anti-apoptotic BCL2 proteins are genetically removed or inhibited by BH3 mimetic drugs <sup>33, 34</sup>. However, BAX and BAK1 activation in the absence of BH3only proteins occurs at slower kinetics compared to that in the presence of BH3-only proteins <sup>33</sup>. These findings support the existence of both BH3-dependent and BH3-independent activation of BAX and BAK1 where BH3-only proteins function as catalysts for BAX and BAK activation <sup>33,38</sup>.. In this context, BAX and BAK are also reported to be activated by the tumor protein p53 (TP53; best known as p53) in a fashion independent of BH3-only proteins <sup>39, 40</sup>. The anti-apoptotic members of the BCL2 family encompass BCL2, apoptosis regulator (BCL2), BCL2 like 1 (BCL2L1; best known as BCL-X<sub>L</sub>), MCL1, BCL2 family apoptosis regulator (MCL1), BCL2 like 2 (BCL2L2; best known as BCL-W), and BCL2 related protein A1 (BCL2A1; best known as A1) <sup>5, 6, 7, 8</sup>. The anti-apoptotic activity of these BCL2 proteins mainly involves MOMP maintenance, although, a non-canonical, cellular redox-dependent mechanism of cytoprotection has been ascribed in cancer cells at least for BCL2 41, 42, 43, 44

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# Box 2. Impact of pro-apoptotic BCL2 proteins on health.

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Deletion of BCL2-associated X protein (Bax), BCL2-antagonist/killer 1 (Bak1) or BCL2-related ovarian killer (Bok) does not significantly affect mouse development 45, 46, 47, with the exception of a mild lymphocyte and neuron accumulation in Bax-/- mice which also exhibit male infertility due to seminiferous tubule malformation 45, 48. Of note, a recent study has demonstrated that such defects in germ cells occur in the fetal period <sup>49</sup>, supporting the requirement for intrinsic apoptosis in testicular development <sup>50, 51</sup>. Subsequent studies confirmed the role of BAX in neurogenesis, in particular the development of hippocampal and cerebellar neurons, cortical interneurons and astrocytes <sup>52, 53, 54, 55, 56, 57</sup>. Accordingly, Bax<sup>-/-</sup> mice exhibit impaired neurological functions manifesting with increased anxiety, depression-like traits, compromised social and sexual behavior, and impaired spatial representation and olfactory system function <sup>58, 59, 60</sup>. These mice also show accelerated medulloblastoma formation <sup>61</sup>, which is in line with the oncosuppressive activity of apoptotic (and non-apoptotic) regulated cell death (RCD) 62. Ablation of Bok does not compromise the relatively normal development of BAK1- or BAX-deficient mice, although  $Bax^{-/-}Bok^{-/-}$  mice exhibit an increased number of mature oocytes <sup>63</sup>. In contrast, co-deletion of Bax and Bak1 causes perinatal death in the vast majority (more than 90%) of mice, mainly due to multiple developmental abnormalities and feeding difficulties 46, 64. Importantly, the developmental defects of Bax--Bak1-- mice are exacerbated by additional deletion of Bok, underscoring not only some functional redundancy between BAX, BAK1 and BOK, but also a crucial role of pro-apoptotic BCL2 family members in the development of the central nervous system (CNS) and hematopoietic compartment <sup>64</sup>. However, since some Bax<sup>-/-</sup>Bak1<sup>-/-</sup> and Bax<sup>-/-</sup>Bak1<sup>-/-</sup>Bok<sup>-/-</sup> mice can reach adulthood <sup>46</sup>, <sup>64</sup>, additional systems must be at play to compensate for defects in apoptosis in other organs. In is worth noting that the developmental defects of Bax--Bak1-- mice can be further aggravated by deletion of autophagy related 5 (*Atg5*) <sup>65</sup>, which is involved in autophagy as well as in non-canonical vesicular pathways like LC3-associated phagocytosis <sup>66, 67</sup>. However, whether autophagy-dependent cell death compensates for the apoptotic defects of *Bax-/-Bak1-/-* mice remains to be formally determined <sup>68, 69</sup>.

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Further corroborating the relevance of intrinsic apoptosis for proper development, the few surviving Bax /-Bak1-/- mice and Bax-/-Bak1-/- mice display phenotypes related to defective programmed cell death (PCD), including webbed feet (due to the incomplete removal of interdigital webs), imperforate vagina and midline fusion defects including facial cleft 46,64. CNS issues exhibited by these animals include a striking expansion of the tissue regions that harbor the neural stem cell pool 46, 64 as well as impaired function of the motor <sup>70</sup> and visual <sup>71, 72</sup> systems. Although the number of apoptotic cells were reduced to the limit of detection in embryos lacking BAX, BAK1 and BOK <sup>64</sup>, anomalies in the urinary tract were conspicuously absent in these animals <sup>64</sup>. This sparked a study examining if BID, in addition to linking the death receptor (DR) pathway and the intrinsic apoptotic pathway (Box 5), could act in a way similar to BAX and BAK1. Indeed, while loss of BID alone did not lead to anomalies during embryonic and fetal development, additional deletion of *Bid* in  $Bax^{-/-}Bak1^{-/-}Bok^{-/-}$ mice mice revealed a redundant requirement for BID in urogenital tract development <sup>73</sup>. In its previously recognized role, BID in the form of tBID activates BAX and BAK1, which would not have caused additional anomalies in the absence of BAX and BAK1. Therefore, these results indicate that BID can act in parallel with BAX, BAK1 and BOK. Congruently, full-length BID <sup>73</sup> or tBID <sup>74</sup> can mediate mitochondrial permeabilization and cause cytochrome c, somatic (CYCS) release. In this context it is worth considering that BID has been reported to be structurally similar to the multi-BH domain BCL2 family proteins, such as BAX and BCL- $X_L$   $^{9,75,}$ 76, 77

Tissue-specific ablation of Bax and Bak1, confirmed the crucial role of these proteins in the hematopoietic system, and specifically in the homeostasis and functionality of B cells <sup>78</sup>, T cells <sup>79</sup>, megakaryocytes <sup>80</sup>

and platelets 81. Mice reconstituted with fetal liver cells from Bax--Bak1-- mice display massive lymphadenopathy and defective T cell proliferation, and the severity of these defects is even more pronounced when Bak1<sup>-/-</sup>Bax<sup>-/-</sup>Bok<sup>-/-</sup> fetal liver cells are used for reconstitution, an experimental setting that also reveals signs of autoimmunity 82, 83, 84. Similarly, mice reconstituted with a Bak1<sup>-/-</sup>Bax<sup>-/-</sup> hematopoietic compartment develop a fatal systemic lupus erythematosus (SLE)-like autoimmune disease 85. Moreover, the inducible co-deletion of Bax and Bak1 in lymphocytes of adult mice results in the development of severe autoimmune glomerulonephritis <sup>78</sup>. Finally, conditional knockout mouse models reveal a crucial contribution of BAX and BAK1 to endothelial cell homeostasis 86,87, but little impact on cardiac and intestinal functions, as shown by the absence of hyperplasia <sup>88, 89</sup>. These results demonstrate that the multi-BH domain pro-apoptotic BCL2 proteins play critical roles for the normal development of multiple tissues but that, surprisingly, a few mice can reach weaning or even adulthood when all of these effectors of apoptosis are removed <sup>64</sup>. Amongst the BH3-only proteins, BCL2 like 11 (BCL2L11, best known as BIM) appears the most critical for embryonic development and tissue homeostasis, as shown by the fact that approximately 30% of BIM-deficient mice die during embryogenesis <sup>90</sup>. Surviving BIM-deficient mice display severe defects in the hematopoietic system including lymphoid hyperplasia and marked splenomegaly, and on a mixed C57BL/6 x 129SV background many of these mice spontaneously develop systemic autoimmunity often resulting in fatal kidney disease 90, a condition that can be accelerated by depletion of immunosuppressive CD4+CD25+FOXP3+ regulatory T (T<sub>REG</sub>) cells <sup>91</sup>. Cells from BIM-deficient mice are profoundly resistant to growth factor deprivation, glucocorticoids, deregulated calcium flux and ER stress 90, 92. Accordingly, BIM-deficient mice also display dysregulated T cell development and homeostasis 93, 94, 95, <sup>96, 97</sup> and hence exhibit defective cellular <sup>98, 99, 100</sup> and humoral <sup>101, 102, 103</sup> immune responses. *Bcl2l11* deletion (loss of BIM) has also been shown to extend the survival of granulocytes 104 and to perturb the development of mammary glands <sup>105, 106</sup>, gastric epithelium <sup>107</sup> and the retina <sup>108</sup>. Moreover, aged BIM

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deficient mice show reduced adiposity <sup>109</sup>. Of note, systemic deletion of Bax or Bak1 exacerbates the hematopoietic dysregulation of BIM-deficient mice <sup>110</sup>. Conditional knockout systems confirmed a key role for BIM in the hematopoietic system homeostasis 111, 112, 113, 114, and revealed a role for BIM in the survival and differentiation of hippocampal neurons 115. Of note, myeloid cell-specific deletion of Bcl2111 induces a systemic lupus erythematosus (SLE)-like disease that resembles the pathology developing in mice that lack BIM in all cells <sup>116</sup>. Mice lacking BH3 interacting domain death agonist (BID), phorbol-12-myristate-13-acetate-induced protein 1 (PMAIP1, best known as NOXA) or BCL2 binding component 3 (BBC3, best known as PUMA) display normal embryonic development <sup>117, 118, 119, 120</sup>. In these studies on BID-deficient mice, substantial reduction in FAS ligand-induced apoptosis was seen in hepatocytes <sup>117, 121</sup>, pancreatic cells <sup>117, 122, 123</sup> and possibly neurons <sup>124, 125</sup>. Moreover, *Bid*<sup>-/-</sup> mice display a dysregulated myeloid compartment resulting in an increased likelihood of leukemogenesis <sup>125</sup>, as well as cardiac dysfunction <sup>126</sup>. Conditional gene deletion studies confirmed the relevance of BID in the homeostasis and functionality of hepatocytes and T cells 127, 128, 129. PUMA contributes to normal ovarian development, as shown by the evidence that two-thirds of the germ cells produced during embryonic development undergo PUMA-mediated cell death shortly after formation <sup>130</sup>. Moreover, cells from PUMA-deficient mice are profoundly resistant to p53-induced apoptosis triggered by genotoxic drugs and lymphoid cells are also resistant to glucocorticoids, phorbol ester and growth factor deprivation <sup>119, 120, 131, 132, 133</sup>. Cells from NOXA-deficient mice also showed resistance to DNA damage-inducing drugs, although to a lesser extent compared to cells lacking PUMA <sup>119, 134</sup>. Moreover, *Pmaip1*<sup>-/-</sup> mice (lacking NOXA) show limited stress-induced erythropoiesis <sup>135</sup>. Germline deletion of the gene encoding PUMA or NOXA also affects humoral immune responses <sup>136, 137</sup> and increases the abundance of multiple cell types in the retina 138. Interestingly, the loss of PUMA

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greatly impairs radiation induced thymic lymphoma development and the formation of liver cancer <sup>139</sup>, 142 <sup>140, 141, 142</sup> (see main text), potentially reflecting the ability of apoptotic cells to secrete mitogenic and 143 immunosuppressive molecules such as prostaglandin E2 (PGE<sub>2</sub>) <sup>143, 144</sup>. PUMA was also shown to play 144 a role in radiation-induced intestinal damage <sup>145</sup>. 145 Co-deletion of two or more genes coding for BH3-only proteins confirmed the pronounced relevance of 146 BIM for development and underscored some degree of functional redundancy in the system. On the one 147 hand, mice lacking both PUMA and NOXA develop normally but their cells are profoundly resistant to 148 genotoxic agents, as resistant as cells lacking p53 <sup>146</sup>. Concomitant loss of PUMA but not the additional 149 loss of NOXA, BAD, BID or BIK increases the severity of hematopoietic defects imposed by the lack of 150 BIM <sup>147, 148, 149, 150</sup>. On the other hand, *Bcl2l11*<sup>-/-</sup>*Bbc3*<sup>-/-</sup>*Bid*<sup>-/-</sup>and *Bcl2l11*<sup>-/-</sup>*Bbc3*<sup>-/-</sup>*Bid*<sup>-/-</sup> mice 151 displayed perinatal embryonic lethality and increased incidence of developmental defects, including 152 webbed feet, imperforate vagina, and supernumerary neurons similar in extent to those seen in Bax<sup>-/-</sup> 153 Bak1<sup>-/-</sup> mice <sup>33, 151</sup>. Of note, triple deficiency of BID, BIM, and PUMA completely abrogates BAX/BAK1 154 dependent apoptosis in cerebellar granule neurons and T lymphocytes <sup>151</sup>, providing in vivo evidence 155 supporting direct activation of BAX and BAK1 by the BH3-only proteins. 156 Mice lacking BCL2-associated agonist of cell death (Bad), BCL2 interacting killer (Bik), BCL2 157 modifying factor (Bmf) and harakiri, BCL2 interacting protein (contains only BH3 domain) (Hrk) are 158 viable and develop normally <sup>152, 153, 154, 155</sup>. That said, BAD-deficient mice display a prolonged platelet 159 lifespan <sup>156</sup>, while *Bmf*<sup>-/-</sup> mice are characterized by mild lymphadenopathy, vaginal atresia <sup>154, 157</sup> as well 160 as minor defects in mammary gland development and oogenesis  $^{106, 158}$ . Interestingly, female  $Bmf^{-/-}$  mice 161 had significantly more primordial follicles than wild-type control animals associated with an extended 162 fertile life span 159, while Bmf<sup>-/-</sup> mice developed an accelerated gamma irradiation-induced thymic 163 lymphoma <sup>154</sup>. Combined deletion of some of the above listed BH3-only protein-coding genes does not 164

cause significant embryonic lethality or developmental abnormalities. Moreover, the combined absence of BIK and NOXA did not accelerate c-MYC-driven lymphoma development <sup>160</sup>, while increased spontaneous tumorigenesis has been documented in *Bad*<sup>-/-</sup>*Bmf*<sup>-/-</sup> mice <sup>161</sup>. Conversely, the absence of some of these BH3-only proteins aggravates the defects caused by the loss of *Bcl2l11* (the gene encoding BIM). This applies to: (1) *Bad* co-deletion with *Bcl2l11*, which enhances lymphocyte accumulation <sup>156</sup>, (2) *Bik* co-deletion with *Bcl2l11*, which causes male infertility due to defective spermatogenesis <sup>162</sup>, a phenotype resembling that of BAX-deficient mice, and (3) *Bmf* co-deletion with *Bcl2l11*, which considerably increases the incidence of developmental defects, vaginal atresia, lymphadenopathy, autoimmune glomerulonephritis, and spontaneous development of hematological malignancies <sup>157, 163, 164</sup>.

## Box 3. Impact of anti-apoptotic BCL2 proteins on health.

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While myeloid cell leukemia sequence 1 (Mcl1) deletion in mice induces embryonic lethality at the blastocyst (embryonic E3) stage prior to implantation <sup>165, 166</sup>, embryos lacking BCL2-like 1 (BCL2L1, best known as BCL-X<sub>L</sub>) die around embryonic day 13.5) with substantial cell depletion in the developing central nervous system (CNS) and erythroid progenitors <sup>167</sup>. Concomitant deletion of BCL2-associated X protein (Bax) or caspase 9 (Casp9) considerably limited neuronal cell death genotype caused by the absence of BCL-X<sub>L</sub> <sup>168, 169</sup>. Concomitant deletion of BCL2 like 11 (*Bcl2l11*, the gene encoding BIM) rescues the erythroid progenitors (but not the neuronal) cells from death in BCL-X<sub>L</sub>-deficient mice <sup>170</sup>. Bcl2<sup>-/-</sup> mice are born but exhibit severe defects in their kidneys, alterations of the CNS, lymphoid cell depletion as well as premature graying of their hair and they succumb to polycystic kidney disease at a young age <sup>171, 172, 173, 174, 175, 176, 177</sup>. These defects can all be rescued by concomitant deletion of the gene encoding BIM, and, remarkably, in the case of some defects the loss of even a single allele of Bim is sufficient <sup>171</sup>. Mice with deletion of B cell leukemia/lymphoma 2 related protein A1a (Bcl2a1a, one of three isoforms of BCL2A1 in mice) or loss of all isoforms of BCL2A1 (best known as A1) show no developmental defects but display minor defects in the hematopoietic compartment <sup>178, 179, 180, 181</sup>. The absence of BCL-W results in male infertility due to defective spermatogenesis <sup>182, 183, 184</sup>. As opposed to homozygous deletion, haploinsufficiency for genes encoding MCL1 or BCL- $X_L$  did not result in defects in normal development <sup>165, 167</sup>. However, *Mcl1*<sup>+/-</sup> mice display significant, albeit minor decreases in certain hematopoietic cell types <sup>185, 186</sup>, and poor hematopoietic recovery from stress, such as gamma-radiation or treatment with 5-FU, which can be rescued by deletion of BCL2 binding component 3 (*Bbc3*; the gene encoding PUMA) <sup>186</sup>. Moreover, the loss of one *Bcl2l1* allele (encoding BCL-X<sub>L</sub>) limits male fertility due to defects in germ cell development <sup>187</sup> and shortens platelet lifespan <sup>188</sup>. Of note, while combined haploinsufficiency for *Mcl1* and *Bcl2*, for *Mcl1* and *Bcl2a1a* or for *Bcl2l1* 

and Bcl2 does not markedly affect embryonic development in mice 189, 190, 191, Mcl1+/-Bcl2l1+/- double heterozygote mice display severe developmental defects and die during embryogenesis or early postnatally <sup>190</sup>.Remarkably this defect that can be rescued by concomitant deletion of a single allele of the gene encoding BIM. These observations suggest that embryonic development is safeguarded by a delicate balance between pro- and anti-apoptotic BCL2 proteins. Conditional knockout studies confirmed the importance of the different pro-survival BCL2 family members in specific tissues at precise developmental stages. These studies showed that MCL1 is critical for the development and/or maintenance of most (but not all) hematopoietic cell populations including stem and progenitor cells <sup>192</sup>, immature as well as mature B and T lymphocytes <sup>193, 194, 195, 196</sup> Jain, 2017, 28972012;<sup>197</sup>, natural killer (NK) cells <sup>198</sup>, neutrophils <sup>199, 200</sup>, mast cells and basophils <sup>201</sup>, as well as Ig secreting plasma cells <sup>202, 203</sup>. Accumulating evidence suggests that the survival of some hematopoietic cell subsets is safeguarded by the combined activity of two or even more anti-apoptotic BCL2 family members <sup>204</sup>. Conditional deletion of *Bcl2l1* alone (leading to lack of BCL-X<sub>L</sub>) or in combination with loss of Mcl1 demonstrated functional redundancy between BCL-X<sub>L</sub> and MCL1 in developing lymphocytes <sup>205, 206</sup> and megakaryocytes <sup>188, 207, 208, 209</sup>. Conversely, BCL2 and A1 appear to have overlapping actions in the survival of B cells and neutrophils <sup>189, 210, 211</sup> but not megakaryocytes and platelets <sup>212</sup>. Data from hematopoietic chimeric mice confirm the role of these proteins in hematopoiesis <sup>104, 167, 213, 214</sup>. BCL2 is reported to contribute to the development and homeostasis of the mouse epidermis <sup>215</sup>. Along similar lines, MCL1 and BCL- X<sub>L</sub> play roles in the development and homeostasis of several tissues including the myocardium <sup>216, 217</sup>, the CNS <sup>218, 219, 220, 221, 222, 223, 224, 225, 226</sup>, the hepatic parenchyma <sup>227, 228, 229, 230, 231</sup>, vascular endothelium <sup>232</sup>, thymic epithelium <sup>233</sup>, as well as the intestinal <sup>234</sup>, mammary <sup>235, 236</sup>, lung <sup>237</sup> and renal <sup>238</sup> epithelium.

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There are substantial differences in the severity of the defects caused by the conditional deletion of different pro-survival BCL2 family genes and between distinct tissues. For instance, conditional deletion of Mcl1 in mouse hematopoietic stem/progenitor cells <sup>192</sup>, erythroid cells <sup>239</sup> or T<sub>REG</sub> cells <sup>240</sup> is lethal. In the latter case, lethality is ascribed to multiorgan autoimmunity caused by the depletion of the pool of T<sub>REG</sub> cells <sup>240</sup>. Similarly, the megakaryocyte-specific combined deletion of the genes encoding MCL1 and BCL-X<sub>L</sub> provokes embryonic or perinatal lethality <sup>207</sup>, which can be rescued by the absence of BAK1 <sup>80</sup>. Similar findings have been obtained upon the ablation of *Mcl1* from the CNS or the myocardium, or the specific removal of the gene encoding BCL-X<sub>L</sub> from the respiratory epithelium, although these experiments did not include rescue approaches <sup>217, 218, 219, 237</sup>. The functional overlap between MCL1 and BCL-X<sub>L</sub> appears to be particularly relevant in the CNS and liver <sup>225, 228</sup>. Of note, the requirement of MCL1 and BCL-X<sub>L</sub> for neurogenesis appears to fluctuate between different stages of differentiation. The neurodevelopmental defects imposed by the deletion of Mcl1 or Bcl2l1 can be rescued by the absence of BAX <sup>169, 225</sup>. The detrimental effects of the hepatocyte-specific ablation of *Bcl2l1* or *Mcl1* can be rescued by deletion of Bax and Bak1 as well as by that of Bcl2l11 (encoding BIM) and/or BH3 interacting domain death agonist (Bid) 241, 242. These observations demonstrate that organogenesis and adult tissue homeostasis depend on the balance between both anti-apoptotic and pro-apoptotic members of the BCL2 family. Further substantiating this notion, deletion of the gene encoding BCL-X<sub>L</sub> from keratinocyte precursors limits skin cancer development driven by ultraviolet B (UVB) rays and chemical carcinogens  $^{243}$ . Conversely, the hepatocyte-specific deletion of *Mcl1* promotes hepatic carcinogenesis  $^{244}$ , as does the deletion of *Mcl1* in intestinal epithelial cells <sup>234</sup>. These latter findings may appear counterintuitive, as pre-malignant cells are expected to be more susceptible to succumb to environmental stress in the absence of MCL1 or BCL-X<sub>L</sub>. However, both hepatic and intestinal carcinogenesis involve a robust inflammatory component that is exacerbated by tissue damage and cell death <sup>245</sup>. Moreover, MCL1-deficient tissues show an increased cell turnover, which results in elevated level of replicative stress and genetic

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instability, potentially promoting carcinogenesis <sup>231, 234</sup>. Also, when many cells die, progenitors get mobilized and must divide extensively. This increases the risk of such cells acquiring mutations that may drive neoplastic transformation, as firstly shown in a murine model of radiation induced thymic T cell lymphoma development <sup>139, 140</sup>.

## Box 4. Impact of the apoptosome and apoptotic caspases on health

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The whole-body deletion of apoptotic peptidase activating factor 1 (Apaf1) or caspase 9 (Casp9) is associated with fetal lethality around E14.5 to E16.5 <sup>246, 247, 248</sup>. Severe abnormalities in APAF1-deficient fetuses include webbed feet, craniofacial malformations, incomplete neural tube closure and/or excessive brain growth and exencephaly resulting in alteration of the central nervous system (CNS) including in the visual, olfactory, and auditory systems <sup>246, 248, 249, 250, 251, 252</sup>. Similar defects in the developing brain result from Casp9 deletion <sup>166, 248, 253</sup>, a phenotype that was not exacerbated by Casp2 co-deletion <sup>254</sup>. The absence of CASP9 did not rescue neuronal defects due p53 hyperactivation in neural crest cells <sup>255</sup>. Of note, evidence linking mutations in APAF1, CASP9 and CASP3 to neural tube defects in humans has been reported <sup>256, 257</sup>. Mice lacking cytochrome c, somatic (CYCS) die in midgestation <sup>258</sup>, while the deletion of cytochrome c, testis (Cyct), which is specifically expressed in male gonads is associated with normal development but male infertility <sup>259</sup>. The neuron-specific ablation of *Cycs* results in postnatal cell death <sup>260</sup>. Confirming that the detrimental effects of Cycs deletion result from impaired apoptosis, mice expressing a mutant CYCS that retains the ability to shuttle electrons as a component of the mitochondrial respiratory chain but is unable to assemble the apoptosome exhibit perinatal lethality and developmental brain defects similar to APAF1- and CASP9-deficient mice <sup>261</sup>. Importantly, the genetic background of the mouse strains appears to significantly influence the impact of the absence of core components of the apoptotic machinery on embryonic development. Thus, while genetic deletion of Casp3 in 129S1/SvImJ mice results in embryonic or early postnatal lethality due to the severe defects in brain development that are only partially rescued by concomitant deletion of the gene encoding BCL-X<sub>L</sub>, on a C57BL/6 background Casp3<sup>-/-</sup> mice develop normally and survive into adulthood <sup>262, 263, 264, 265</sup>. A similar impact of genetic background on phenotype has also been observed for *Apaf1*<sup>-/-</sup> and *Casp9*<sup>-/-</sup> mice <sup>266, 267</sup>. Although *Casp3*<sup>-/-</sup> mice reach adulthood on a C57BL/6 background, they exhibit defects in complex brain functions including attention and (in males) social behavior <sup>268, 269</sup>, as well as ear and vestibular dysfunction including hearing loss <sup>270, 271, 272, 273, 274</sup>, Abnormalities were also seen in the kidney and spleen of aged *Casp3*<sup>-/-</sup> mice <sup>275</sup>. Survival of *Casp3*<sup>-/-</sup> mice to adulthood in C57BL/6 mice was ascribed to the compensatory activation of CASP7 <sup>276</sup>. The combined ablation of *Casp3* and *Casp7* causes embryonic lethality on the C57BL/6 background, although death is caused by severe cardiac rather than brain defects <sup>277</sup>. Such phenotypic differences may originate from some degree of substrate selectivity exhibited by CASP3 *vs.* CASP7 <sup>278, 279, 280, 281, 282</sup>. Moreover, a recent study performed in *Casp7*<sup>-/-</sup> mice indicates that CASP7 acts as a facilitator of the variants of RCD occurring in the context of pore-driven lysis rather than an apoptotic executioner <sup>283</sup>.

Approximately 5% of APAF1-deficient mice develop normally and survive into adulthood, although males are often sterile due to defective spermatogenesis <sup>247</sup>; their phenotype is reminiscent of the phenotype of mice deficient for BAX, BAK1 and BOK (*i.e.*, *Bak1*<sup>-/-</sup>*Bax*<sup>-/-</sup>*Bok*<sup>-/-</sup> mice) <sup>64</sup>. Of note, rare adult *Apaf1*<sup>-/-</sup> male mice that retain fertility display expansion of the lateral brain ventricles coupled with behavioral abnormalities and growth retardation <sup>267</sup>. Conversely, the rare mice expressing a CYCS variant specifically deficient in apoptotic functions that survive into adulthood exhibit impaired lymphocyte homeostasis <sup>261</sup>. Whole-body deletion of diablo, IAP-binding mitochondrial protein (*Diablo*, coding for a pro-apoptotic factor also known as SMAC) alone or along with HtrA serine peptidase 2 (*Htra2*) does not result in developmental defects in mice <sup>284, 285</sup>, while the *Diablo*<sup>-/-</sup>*Casp3*<sup>-/-</sup> genotype accrues the perinatal lethality observed in *Casp3*<sup>-/-</sup> mice <sup>286</sup>. Mice lacking the X-linked inhibitor of apoptosis (XIAP, the main target of the pro-apoptotic activity of SMAC and HTRA2) are also viable and develop normally, possibly due to functional compensation by other members of the inhibitor of apoptosis protein (IAP) family <sup>287, 288</sup>, but they exhibit mild defects in late pregnancy that do not compromise lactation <sup>287</sup>. Consistent with this SMAC-mimetic drugs that were designed to induce

apoptosis by antagonizing IAPs are quite well tolerated  $^{289}$ .  $Xiap^{-/-}$  mice also show dysregulated innate immune responses  $^{290}$ , most likely linked to the modulatory role of XIAP in inflammation and necroptosis  $^{291, 292, 293}$ , or to the inability of these animals to resolve infections  $^{294}$ . Accordingly, loss-of-function mutations in XIAP are associated with X-linked lymphoproliferative syndrome type 2 in humans  $^{291, 295, 296, 297}$ 

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The myocardium-specific deletion of Casp3 and Casp7 impairs heart development in mice resulting in myocyte hypertrophy <sup>298</sup>. The role of APAF1, CASP9 and CASP3 in hematopoiesis remains debated. Specific ablation of *Apaf1* or *Casp9* from the hematopoietic system using lethally irradiated wild-type mice reconstituted with hematopoietic stem/progenitor cells deficient for these factors did not expand the lymphoid or myeloid cell compartments <sup>299</sup>. Likewise, no hematopoietic defects emerge from the whole-body deletion of Casp3 <sup>277</sup>. Moreover, mice lacking Casp9 in the hematopoietic system display a proper generation and functionality of megakaryocytes and platelets 300. Moreover, the clearance of Casp9<sup>-/-</sup> thymocytes seems to occur in a caspase-independent fashion <sup>301</sup>. In the same line, although apoptosis is widely believed to be crucial for epithelial cell death and shedding in the intestine, during steady state, executioner CASP3 and CASP7 are dispensable for intestinal epithelial cell turnover at the top of intestinal villi, intestinal tissue dynamics, microbiome, and immune cell composition, suggesting high redundancy in non-challenged conditions <sup>302</sup>. Apparently at odds with these observations, Casp3<sup>-/-</sup> mice were reported to have abnormally increased numbers of splenic B cells manifesting increased proliferative capacity 303, as well as a dysregulated activity in bone marrow stromal stem cells that attenuated osteogenic differentiation <sup>304</sup>. A similar debate revolves around the requirement for APAF1 and caspase activity in thymocyte selection and/or T cell responses <sup>299, 305, 306, 307, 308, 309</sup>. Mouse bone marrow chimeras deficient for APAF1 or CASP9 in their hematopoietic cells displayed a defect in hematopoietic stem/progenitor cells that is caused by the aberrant type 1 interferon production caused by the fact that hematopoietic cells undergoing normal programmed cell death do not die in a "neat" noninflammatory manner <sup>310, 311</sup>. Taken together, these findings suggest that BAX/BAK1 dependent death of hematopoietic cells does not require caspases but that caspases are needed to prevent an inflammation causing form of cell demolition <sup>312, 313, 314, 315</sup>. However, neither the degree of functional redundancy exhibited by CASP3, CASP6 and CASP7, nor the potential for APAF1-independent CASP3 activation has been formally excluded in these studies, most of which involved single genetic alterations.

# **Box 5.** Principles of extrinsic apoptosis.

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Extrinsic apoptosis is a regulated cell death (RCD) process frequently triggered by immune effector cells expressing TNF superfamily death ligands binding the death receptor (DRs) upon binding of a cognate ligand <sup>316, 317, 318</sup>. The principal DRs which will be discussed in the review are the Fas cell surface death receptor (FAS; also known as CD95 or APO-1), the TNF receptor superfamily member 1A (TNFRSF1A; best known as TNF-R1), the TNF receptor superfamily member 10a (TNFRSF10A; best known as TRAIL-R1 or DR4) and the TNF receptor superfamily member 10b (TNFRSF10B; best known as TRAIL-R2 or DR5). FAS is activated by the binding of FAS ligand (FASLG; also known as CD95L or APO-1L; FASL in mice), which is primarily expressed by effector immune cells <sup>318</sup>. TNF-R1 is activated by tumor necrosis factor (TNF), a functionally pleiotropic cytokine expressed in cells in the spleen, thymus and certain other adult tissues <sup>316</sup>. Of note, while the soluble form of TNF preferentially binds to TNF-R1, the membrane-anchored form mainly interacts with the TNF receptor superfamily member 1B (TNFRSF1B, best known as TNF-R2), which does not have death domain and therefore is not a DR <sup>319</sup>. Finally, TRAIL-R1 and TRAIL-R2 are specifically activated by the binding of TNF superfamily member 10 (TNFSF10; best known as TRAIL), which is expressed by a variety of cell subtypes of the innate as well as adaptive system, including monocytes, macrophages and effector T cells, as either a soluble or membrane-bound version <sup>320</sup>. Of note, mice express only one TRAIL receptor (TRAIL-R2, referred in this article as mTRAIL-R) which is equally homologous to human TRAIL-R1 and TRAIL-R2. Upon ligand binding and trimerization and in certain instances formation of higher order complexes, the engagement of DRs promotes the assembly of multi-protein complexes, such as the death-inducing signaling complex (DISC) and complex II, resulting in the activation of caspase 8 (CASP8) and apoptosis 321, 322, 323, 324. The DISC, which is assembled on the cytoplasmic tail of ligated FAS, TNF-R1, TRAIL-R1 or TRAIL-R2, is comprised of the molecular adaptor Fas-associated death domain protein (FADD),

Fas (TNFRSF6)-associated via death domain (FADD), CASP8, and (FADD-like IL-1β-converting enzyme)-inhibitory protein distinct isoforms of CASP8 and FADD like apoptosis regulator (CFLAR; best known as c-FLIP), including the alternative splicing variants, the long form c-FLIP<sub>L</sub> and the short form c-FLIPs and (in human) c-FLIP<sub>R</sub> <sup>325, 326, 327, 328, 329, 330</sup>. Of note, c-FLIPs are catalytically inactive CASP8-like molecules acting as a modulator of CASP8 activation. Unlike FAS- and TRAIL-Rassociated DISCs, complex II is a cytosolic complex assembled secondarily upon TNF-R1 ligation, in conditions of reduced pro-survival signaling and protein synthesis as for instance upon administration of inhibitors of inhibitor of apoptosis proteins (IAPs) and cycloheximide <sup>331</sup>. Complex II consists of FADD and CASP8 in association with either TNF-R1-associated death domain protein (TRADD) (complex IIa) or receptor interacting serine/threonine kinase 1 (RIPK1) (complex IIb), which is involved in the modulation of apoptosis and necroptosis <sup>332</sup>. Upon the recruitment to the DISC (complex I), CASP8 is activated by a process involving CASP8 oligomerization and autoproteolysis. CASP8 then acts as the executor of extrinsic apoptosis by favoring the proteolytic activation of the effector caspases CASP3 and CASP7 <sup>333</sup>. This direct pathway is sufficient for FAS ligand induced killing thymocytes and mature lymphocytes (so-called type 1 cells), but the efficient killing of hepatocytes, pancreatic  $\beta$  cells, and most cancer cells (so-called type 2 cells) requires pathway amplification through caspase-8 mediated proteolytic activation of the BH3-only protein BID, leading to engagement of the intrinsic apoptotic pathway 117, 334, 335, 336, 337, 338, 339 PMID: 9501089 PMID: 9501089 PMID: 9501089 PMID: 9501089. Of note, the absence of XIAP converts type 2 cells into type 1 cells <sup>123</sup>, indicating that a limited amount of caspase activity is needed for cell killing. Once activated, CASP8 also cleaves RIPK1 leading to the inhibition of necroptosis and the maintenance of inflammatory homeostasis <sup>340</sup>. As a further layer of complication, the engagement of DRs by their

respective ligands does not necessarily culminate in the activation of the extrinsic apoptosis signaling

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pathway. Indeed, the engagement of FAS, TRAIL-Rs and TNF-R1 can also result in the activation of pro-survival pathways which is often but not always dependent on NF-κB signaling 320, 341, or, alternatively, in the initiation of inflammatory responses, the promotion of processes including cell differentiation/activation (as is the case of lymphocytes), and the activation or inhibition of other RCD variants, particularly necroptosis and pyroptosis <sup>342</sup>. The induction of inflammatory chemokines and cytokines downstream of the activation of FAS and TRAIL-Rs is mediated by FADD and CASP8 by a mechanism that can be independent of apoptosis induction <sup>343, 344</sup>. Extrinsic apoptosis can be activated by another class of cell surface receptors known as dependence receptor. In this case, cell death is ignited by the decrease in the availability of a specific ligand on which these receptors depend, while the latter through the binding of a cognate ligand <sup>345, 346</sup>. The dependence receptors include (but are not limited to) the DCC netrin 1 receptor (DCC) and distinct types of unc-5 netrin receptors (UNC5A, UNC5B, UNC5C, and UNC5D), all of which are bound by netrin 1 (NTN1), and the neurotrophic receptor tyrosine kinase 3 (NTRK3) and patched 1 (PTCH1), which are, respectively, ligated by neurotrophin and sonic hedgehog (SHH). The activation of dependence receptors stimulates hitherto poorly characterized signaling cascade often dependent on caspase activation, leading

to the induction of cell death <sup>347, 348</sup>. It is noteworthy that the relevance of the dependence receptor-

induced apoptosis for normal physiology and disease is not established.

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# **Box 6.** Impact of death receptors on health.

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A large body of data demonstrates that death receptor (DR) signaling is crucial for the maintenance of adult tissue homeostasis but nor for embryonic development as demonstrated by the normal appearance of mice double knockout for caspase 8 and mixed lineage kinase domain like pseudokinase (Casp8<sup>-/-</sup> *Mlkl*<sup>-/-</sup> mice) or CASP8 and receptor-interacting serine-threonine kinase 3 (*Casp*8<sup>-/-</sup>*Ripk*3<sup>-/-</sup> mice) (before they develop lymphadenopathy and splenomegaly) <sup>349, 350, 351, 352, 353, 354</sup>. Mouse strains with spontaneous mutations in TNF receptor superfamily member 6 (Fas) - the so-called lpr/lpr mice - or Fas ligand (TNF superfamily, member 6) (Fasl) - the so-called gld/gld mice - are viable but develop progressive lymphoproliferative and systemic lupus erythematosus (SLE)-like disorders 355, 356, 357, 358, 359. The severity of these pathologies is greatly influenced by genetic background: fairly mild on a C57BL/6 background but very severe on the MRL or NOD backgrounds. Mice with heterozygous Fas or Fasl mutations are normal <sup>359</sup>. These lymphoproliferative and autoimmune disorders are not accompanied by impaired thymocyte development 360. Transgenic overexpression of BCL2 335 or MCL1 361 in the lymphocyte compartment of lpr/lpr mice or the absence of BIM 362 massively exacerbate lymphadenopathy. This is consistent with the notion that intrinsic apoptosis and DR-induced apoptosis are distinct in lymphoid cells and act additively. FAS or FASL deficiency also perturbs the homeostasis or function of other mouse tissues, including (but not limited to) the liver <sup>360</sup>, kidney <sup>363</sup>, retina <sup>364</sup>, pancreas <sup>365</sup> and intestinal epithelium <sup>366</sup>, but these effects may all be a consequence of the deregulation of the lymphoid system in these mice, for example causing excess production of certain cytokines and chemokines. Conditional deletion of Fas and Fasl in specific immune cell subsets as well as transgenic expression of FAS in lymphocytes confirms the crucial role of FASL-FAS signaling in the homeostasis of lymphocytes and dendritic cells (DCs) <sup>367, 368, 369, 370, 371</sup>. In this context, experiments in *lpr/lpr* mice deleted of BH3-

only protein BCL2 like 11 (Bcl2111, the gene encoding BIM) demonstrate some degree of cooperation between FAS and BIM in preserving the functionality of the immune system <sup>362</sup>. However, abrogating FAS-FASL signaling ultimately has heterogeneous organismal consequences. The lymphoproliferative disorder caused by Fas or Fasl deletion confers protection from autoimmune diabetes <sup>372</sup>. This may be explained by the fact that the distortion of the T cell repertoire caused by the lymphadenopathy in the lpr/lpr and gld/gld mice prevents the development of diabetogenic T cells. Finally, FAS appears to exert tumor suppressive effects in lymphoid cells. Indeed, both gld/gld mice as well as lpr/lpr mice lacking the T cell compartment have increased incidence of B cell lymphoma <sup>373, 374, 375</sup>. Loss of FAS also predisposes humans to B lymphoma (see below). As for the other DRs, mice lacking TNF receptor superfamily member 10b (TNFRSF10B, best known as TRAIL-R2 or mTRAIL-R) or its ligand TNF superfamily member 10 (TNFSF10, beast known as TRAIL) are viable, fertile, and do not spontaneously develop autoimmune diseases <sup>376, 377, 378, 379</sup>. Moreover, these mice exhibit normal immune system development and function <sup>380, 381, 382, 383</sup>. Along similar lines, the whole-body deletion of the DR ligand tumor necrosis factor (Tnf) does not affect mouse development and fertility 384, 385. However, Tnf<sup>-/-</sup> mice often show early hearing loss and, despite presenting with an overtly functional immune system, these mice exhibit abnormally increased susceptibility to spontaneous bacterial infection, which has been ascribed to multiple defects including defective lymphoid organ architecture as well as deficient granuloma and germinal center formation <sup>384</sup>,  $^{385,386,387,388}$ . Impaired responses to pathogens have been documented in  $Tnf^{+/-}$  mice  $^{384}$  as well as in mice lacking TNF receptor superfamily member 1A (TNFRSF1A, best known as TNF-R1) 386, 389, 390. Conversely, mice overexpressing TNF in cardiomyocytes suffer from lethal dilated cardiomyopathy, demonstrating that balanced TNF signaling is essential for the homeostasis of the cardiac tissue <sup>391, 392,</sup> <sup>393</sup>. Of note, while the lack of TRAIL enhances the severity of lymphoproliferative and autoimmune disorders in gld/gld mice <sup>394</sup>, the lack of TNF attenuates the lymphoproliferative phenotype, extending

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the survival of *gld/gld* mice <sup>395</sup>. The latter is probably due to the reduction in TNF-mediated inflammation attenuating lymphadenopathy caused by the absence of FAS ligand. These findings confirm the pleiotropy and redundancy of DR signaling, encompassing not only apoptotic and non-apoptotic regulated cell death (RCD)-related effects, but also various pro-survival and pro-inflammatory modules. Multiple clinical observations support the role of FAS ligand/FAS signaling in human hematopoiesis <sup>396</sup>. Most human patients with autoimmune lymphoproliferative syndrome (ALPS) - a primary immunodeficiency manifesting with lymphadenopathy, splenomegaly as well as abnormal numbers, development and function of lymphocytes carry loss-of-function mutations in *FAS* or *FASLG* <sup>398</sup>, <sup>399</sup>, <sup>400</sup>, <sup>401</sup>, <sup>402</sup>, <sup>403</sup>, <sup>404</sup>. ALPS patients also display an increased incidence of non-Hodgkin and Hodgkin lymphoma <sup>405</sup>. While no mutations in the genes encoding TRAIL, TRAIL-R1 and TRAIL-R2 have so far been linked to human autoimmune diseases, autosomal dominant mutations in *TNFRSF1A* (leading to lack of TNF-R1) have been identified in patients affected by TNF receptor-associated periodic syndrome (TRAPS), characterized by severe abdominal pain, arthralgias, and myalgias <sup>406</sup>, <sup>407</sup>, <sup>408</sup>.

# **Box 7.** Impact of extrinsic apoptosis complexes and caspases on health.

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Several signal transducers in the death receptor (DRs) pathway are essential for embryonic development in mice. Thus, deletion of Fas (TNFRSF6)-associated via death domain (Fadd), caspase 8 (Casp8) or CASP8 and FADD-like apoptosis regulator (*Cflar*) is embryonic lethal at mid-gestation as a consequence of severe vascular as well as cardiac defects associated with spontaneous intra-abdominal hemorrhage 409, 410, 411, 412, 413, 414. Of note, CASP8-deficient mice also exhibit neural tube defects 413. A similar embryonic lethality has also been documented in mice expressing a mutant form of FADD deficient in its death domain <sup>410</sup>. The absence of other components of DR-associated signaling complexes, such as TNFRSF1A associated via death domain (TRADD) and receptor-interacting serine/threonine kinase 1 (RIPK1), causes different abnormalities. Thus, while *Tradd*<sup>-/-</sup> mice develop normally and do not display major hematopoietic defects 415, 416, 417, Ripk1-- mice die early after birth due to severe multiorgan inflammation <sup>418, 419</sup>. These findings are attributed to the pleiotropic contribution of RIPK1 and TRADD to a variety of processes beyond apoptosis, most notably necroptotic regulated cell death (RCD) and inflammation. This is exemplified by the observation that the embryonic lethality caused by the absence of CASP8 or FADD can be rescued by the concomitant loss of MLKL or RIPK3 (see text). Mice lacking baculoviral IAP repeat-containing 3 (BIRC3; best known as IAP1) and X-linked inhibitor of apoptosis (XIAP) or IAP1 and BIRC2 (best known as IAP2) but not mice lacking IAP2 and XIAP display embryonic lethality 420. These findings indicate specific functional redundancies among the inhibitor of apoptosis protein family. IAP1/IAP2-deficient mice display mid-gestation lethality, which was rescued to birth by the deletion of TNF receptor superfamily member 1A (Tnfrsfla, encoding TNF-R1) but not that of TNF receptor superfamily member 1B (*Tnfrsf1b*, best known as TNF-R2) <sup>420</sup>. Loss of one allele of Ripk1 or loss of Ripk3 prolonged embryonic survival of these mice 420. It is noteworthy, that, as discussed above, genetic background effects might contribute to the phenotype, as mice with concomitant

knockout of the genes encoding IAP1 and IAP2 using mutant alleles generated in C57BL/6 embryonic 473 stem cells die in midgestation <sup>420</sup>, whereas *Iap1*<sup>-/-</sup> and *Xiap*<sup>-/-</sup> double mutants generated using 129Sv 474 embryonic stem cells are viable <sup>421</sup>. 475 It was demonstrated that embryonic lethality in Casp8-/- and Fadd-/- mice is due to excessive necroptosis, 476 reflecting the ability of CASP8 to limit necroptosis downstream of DR activation 349, 350, 422, 423. 477 Accordingly, deletion of genes encoding key components of the necroptotic machinery such as RIPK3 478 or MLKL prevents all developmental defects and embryonic lethality in FADD- or CASP8-deficient 479 embryos <sup>349, 350, 351, 352, 354, 423, 424</sup>. Of note, *Casp8-/-Ripk3-/-* and *Casp8-/-Mlkl-/-* mice develop progressive 480 lymphoproliferative disorders that resemble those caused by the absence of FAS or FASL 350, 351, 423. 481 Moreover, embryonic lethality around E10.5 in mice lacking c-FLIP and the perinatal lethality of Ripk1 482 <sup>/-</sup> mice depend on aberrant activation of both DR-induced apoptosis and DR-induced necroptosis. Indeed, 483 the lethality of these animals can be rescued by concomitant deletion of Fadd and Ripk3, Casp8 and 484 Ripk3, or Fadd and Mlkl <sup>349, 350, 351, 352, 353, 354</sup>. Of note, mice with loss of Ripk1 that prevents its CASP8-485 mediated cleavage die around E10.5 of embryonic development and this can be prevented by the 486 combined absence of RIPK3 and CASP8 340, 425, 426. In a heterozygous state these mutations in the gene 487 encoding RIPK1 cause severe auto-inflammation. As an additional layer of complexity, although the 488 deletion of Tradd rescues Ripk1--Ripk3-- embryos from perinatal lethality, triple knockout mice die 489 postnatally <sup>427, 428</sup>. Moreover, TRADD deficiency does not prevent the embryonic lethality caused by the 490 loss of FADD <sup>428</sup>. Additional studies confirm the importance of the inter-connectivity between multiple 491 regulated cell death (RCD) pathways. Mice with a mutation that prevents auto-proteolytic activation of 492 CASP8 develop normally <sup>429</sup>, but akin to complete loss of *CASP8*, mutations in the CASP8 catalytic site 493 result in embryonic lethality around E10.5 due to aberrant necroptosis 425, 430, while the genetic ablation 494 of Mlkl or Mlkl plus Fadd prevent E10.5 embryonic lethality in these mice, the compound mutant mice 495

die soon after birth, likely due to aberrant inflammation and pyroptosis <sup>431, 432</sup>. These observations point to the central role for CASP8 in the regulation of multiple RCD variants and inflammatory processes <sup>433</sup>. The tissue-specific deletion of Fadd or Casp8 in mouse endothelial cells results in an embryonic lethal phenotype that resembles that of germline Fadd or Casp8 deletion <sup>434, 435</sup>. Conversely, the absence of FADD in cardiomyocytes or cardiac progenitor cells appears to have no impact on embryonic development <sup>434</sup>. Again, abrogation of necroptosis rescued the lethal phenotype of endothelial cell specific Fadd or Casp8 deletion <sup>434</sup>, lending additional support to inhibitory role of FADD and CASP8 in necroptotic RCD. FADD, CASP and CFLAR (best known as c-FLIP) have also been implicated in hematopoietic system homeostasis. However, the absence of FADD in specific immune cell subsets in mice via distinct experimental approaches, such as conditional gene deletion, injection of Fadd-/embryonic stem cells into Rag 1<sup>-/-</sup> blastocysts or transgenic expression of a dominant-negative variant of FADD does not drive lymphoproliferative disorders. Instead, FADD appears to be critical for the proliferation and/or development of T lymphocytes 436, 437, 438, 439, 440, 441, 442, 443, 444, 445, 446 and B cells 447, most likely by preventing necroptosis through activation of CASP8 which then prevents RIPK1/RIPK3 mediated activation of MLKL. Similar conclusions were derived from the analysis of mice with lymphocyte-specific ablation of Casp8 or Cflar 448, 449, 450, 451, 452, 453. A role for CASP8 in T cell proliferation has also emerged from the realization of the anti-proliferative effects of caspase inhibitors <sup>454</sup>. The T cell-specific deletion of *Casp8* attenuates autoimmunity and improved the survival of mice lacking the BH3-only protein BCL2 like 11 (BCL2L11, best known as BIM) by limiting T cell proliferation and survival <sup>455</sup>. Apparently at odds with these findings, the conditional deletion of *Casp8* in T cells has also been associated with an age-dependent, lymphoproliferative immune disorder resembling the condition of patients with CASP8 mutations 456. Whether mouse genetic background or other contextual variables (e.g., the mouse microbiota) underlie such apparent discrepancies remains to be elucidated.

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The conditional loss of the functions of FADD or CASP8 also revealed a role for these proteins in early hematopoiesis, which may relate to their ability to promote the proliferation and differentiation of hematopoietic stem and progenitor cells by preventing necroptosis 435, 457, 458. Conditional deletion of Fadd in myeloid cells resulted in increased myeloid and B cell populations coupled to activation of inflammatory responses <sup>459</sup>. Along similar lines, the macrophage-restricted deletion of *Casp8* induced a mild systemic inflammatory disease potentially linked to altered macrophage polarization 460, 461, while the DC-specific deletion of the genes encoding c-FLIP or CASP8 elicited splenomegaly, inflammatory responses and autoimmune disorders 462, 463, 464. These effects all seem to be unrelated to the pro-apoptotic functions of FADD and CASP8 but reflect their ability to prevent necroptosis 350, 423, 443, 459, 460, 465, 466, 467. Corroborating these findings, loss-of-function mutations in FADD 468, 469, 470, 471, CASP8 or CASP10 472, <sup>473, 474</sup> and *TRADD* <sup>475</sup> have been associated with ALPS-like syndromes and certain hematological diseases in humans. Of note, patients with ALPS bearing mutations in FADD or CASP8 but not ALPS patients with mutations in FAS or FASLG also exhibit immunodeficiency coupled with lymphocytic infiltrations in multiple organs, granulomas and/or inflammatory bowel disease 468, 472, 476, 477, 478. Tissue-specific deletion of Fadd, Casp8 and Cflar has also revealed a role for these proteins in the homeostasis of the liver, skin and intestine, although severity of the phenotype varies quite considerably, ranging from mild inflammatory response to embryonic or early postnatal lethality, again likely due to unleashed necroptosis. Conditional deletion of *Cflar* (resulting in lack of c-FLIP) in intestinal epithelial cells, hepatocytes or keratinocytes resulted in embryonic or perinatal lethality due to aberrant activation of cell death <sup>479, 480, 481, 482</sup>. The inducible deletion of *Cflar* from the intestinal epithelium of adult mice caused severe inflammation that was often fatal 482. These findings are in line with the crucial role of c-FLIP as an inhibitor of necroptosis <sup>349, 483</sup>. Along similar lines, *Fadd* deletion in epidermal keratinocytes or intestinal epithelial cells causes severe chronic inflammation due to the induction of aberrant necroptosis 484, 485, 486, 487, 488, 489, 490. Accordingly, the removal of FADD (or CASP8) in intestinal

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epithelial cells resulted in chronic inflammatory colitis and ileitis, which was prevented by concomitant deletion of Ripk3 or Mklk 430, 432, 485, 487, 490, 491. In one of these studies, acute deletion of Casp8 in the gut of adult mice resulted in enterocyte death, leading to disruption of tissue homeostasis, sepsis and death <sup>490</sup>. In this context, CASP8-deficient enterocytes displayed decreased *in vivo* survival and migration potential <sup>492</sup>. Specific deletion of *Casp8* in endothelial cells results in small intestinal hemorrhage and bowel inflammation, suggesting a key role of CASP8 in vascular homeostasis in the small intestine <sup>493</sup>. Expression of a catalytically inactive variant of CASP8 resulted in embryonic lethality similar to Casp8 <sup>/-</sup> mice, which was rescued by concomitant deletion of *Mlkl* <sup>430</sup>. However, unexpectedly, catalytically inactive CASP8 mutant mice also deficient for MLKL died perinatally. Loss of CASP8 catalytic activity specifically in intestinal epithelial cells induced intestinal inflammation similar to absence of CASP8 in the intestinal epithelium. This intestinal phenotype was aggravated by Mlkl deletion, resulting in premature death dependent on the induction of inflammatory responses and pyroptosis <sup>430</sup>. As an added layer of complexity, deletion of tumor necrosis factor (Tnf) or Tnfrsfla (encoding TNF-R1) attenuated colitis, but not ileitis, in mice with an intestinal epithelial cell-specific deletion of *Fadd* or *Casp*8 <sup>482, 485</sup>. A recent study indicated that this effect may also involve the aberrant activation of pyroptosis. Indeed, the CASP8-dependent activation of gasdermin D (GSDMD) appears to promote ileitis in mice with FADD-deficient intestinal epithelial cells <sup>494</sup>. These results are in line with the crucial involvement of CASP8 and FADD in the activation of inflammation 495, 496 and indicate that the FADD-CASP8 axis regulates tissue homeostasis by balancing apoptosis, necroptosis, pyroptosis and inflammation.

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#### References

- Tait SW, Green DR. Mitochondria and cell death: outer membrane permeabilization and beyond.

  Nature reviews Molecular cell biology 2010, **11**(9): 621-632.
- 571 2. Galluzzi L, Kepp O, Kroemer G. Mitochondrial regulation of cell death: a phylogenetically conserved control. *Microbial cell (Graz, Austria)* 2016, **3**(3): 101-108.
- 574 3. Dadsena S, Zollo C, García-Sáez AJ. Mechanisms of mitochondrial cell death. *Biochemical Society transactions* 2021, **49**(2): 663-674.
- 577 4. Bock FJ, Tait SWG. Mitochondria as multifaceted regulators of cell death. *Nature reviews Molecular cell biology* 2020, **21**(2): 85-100.
- 580 5. Czabotar PE, Lessene G, Strasser A, Adams JM. Control of apoptosis by the BCL-2 protein family: implications for physiology and therapy. *Nature reviews Molecular cell biology* 2014, **15**(1): 49-63.
- 584 6. Shamas-Din A, Kale J, Leber B, Andrews DW. Mechanisms of action of Bcl-2 family proteins. *Cold Spring Harbor perspectives in biology* 2013, **5**(4): a008714.
- 587 7. Kalkavan H, Green DR. MOMP, cell suicide as a BCL-2 family business. *Cell death and differentiation* 2018, **25**(1): 46-55.
- Birkinshaw RW, Czabotar PE. The BCL-2 family of proteins and mitochondrial outer membrane permeabilisation. *Seminars in cell & developmental biology* 2017, **72:** 152-162.
- 593 9. Youle RJ, Strasser A. The BCL-2 protein family: opposing activities that mediate cell death.

  Nature reviews Molecular cell biology 2008, **9**(1): 47-59.
- 596 10. Julien O, Wells JA. Caspases and their substrates. *Cell death and differentiation* 2017, **24**(8): 1380-1389.
- 599 11. Shalini S, Dorstyn L, Dawar S, Kumar S. Old, new and emerging functions of caspases. *Cell death and differentiation* 2015, **22**(4): 526-539.

- 602 12. Green DR. Caspases and Their Substrates. *Cold Spring Harbor perspectives in biology* 2022, 603 **14**(3).
- Kumar S, Dorstyn L, Lim Y. The role of caspases as executioners of apoptosis. *Biochemical Society transactions* 2022, **50**(1): 33-45.

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- 608 14. Kesavardhana S, Malireddi RKS, Kanneganti TD. Caspases in Cell Death, Inflammation, and Pyroptosis. *Annual review of immunology* 2020, **38:** 567-595.
- Moldoveanu T, Czabotar PE. BAX, BAK, and BOK: A Coming of Age for the BCL-2 Family Effector Proteins. *Cold Spring Harbor perspectives in biology* 2020, **12**(4).
- Llambi F, Wang YM, Victor B, Yang M, Schneider DM, Gingras S, *et al.* BOK Is a Noncanonical BCL-2 Family Effector of Apoptosis Regulated by ER-Associated Degradation. *Cell* 2016, **165**(2): 421-433.
- Bleicken S, Landeta O, Landajuela A, Basañez G, García-Sáez AJ. Proapoptotic Bax and Bak proteins form stable protein-permeable pores of tunable size. *The Journal of biological chemistry* 2013, **288**(46): 33241-33252.
- Bleicken S, Wagner C, García-Sáez AJ. Mechanistic differences in the membrane activity of Bax and Bcl-xL correlate with their opposing roles in apoptosis. *Biophysical journal* 2013, **104**(2): 421-431.
- Dewson G, Kratina T, Czabotar P, Day CL, Adams JM, Kluck RM. Bak activation for apoptosis involves oligomerization of dimers via their alpha6 helices. *Molecular cell* 2009, **36**(4): 696-703.
- Dewson G, Kluck RM. Mechanisms by which Bak and Bax permeabilise mitochondria during apoptosis. *Journal of cell science* 2009, **122**(Pt 16): 2801-2808.
- Verhagen AM, Ekert PG, Pakusch M, Silke J, Connolly LM, Reid GE, *et al.* Identification of DIABLO, a mammalian protein that promotes apoptosis by binding to and antagonizing IAP proteins. *Cell* 2000, **102**(1): 43-53.
- Dorstyn L, Akey CW, Kumar S. New insights into apoptosome structure and function. *Cell death and differentiation* 2018, **25**(7): 1194-1208.
- Shiozaki EN, Shi Y. Caspases, IAPs and Smac/DIABLO: mechanisms from structural biology.
   Trends in biochemical sciences 2004, 29(9): 486-494.

641 24. 642 Huang DC, Strasser A. BH3-Only proteins-essential initiators of apoptotic cell death. *Cell* 2000, **103**(6): 839-842. 643 644 25. Kale J, Osterlund EJ, Andrews DW. BCL-2 family proteins: changing partners in the dance 645 towards death. Cell death and differentiation 2018, 25(1): 65-80. 646 647 648 26. Giam M, Huang DC, Bouillet P. BH3-only proteins and their roles in programmed cell death. Oncogene 2008, **27 Suppl 1:** S128-136. 649 650 651 27. Gavathiotis E, Suzuki M, Davis ML, Pitter K, Bird GH, Katz SG, et al. BAX activation is initiated at a novel interaction site. *Nature* 2008, **455**(7216): 1076-1081. 652 653 28. Gavathiotis E, Reyna DE, Davis ML, Bird GH, Walensky LD. BH3-triggered structural 654 reorganization drives the activation of proapoptotic BAX. *Molecular cell* 2010, **40**(3): 481-492. 655 656 657 29. Kim H, Tu HC, Ren D, Takeuchi O, Jeffers JR, Zambetti GP, et al. Stepwise activation of BAX and BAK by tBID, BIM, and PUMA initiates mitochondrial apoptosis. Molecular cell 2009, 658 **36**(3): 487-499. 659 660 30. 661 Wei MC, Zong WX, Cheng EH, Lindsten T, Panoutsakopoulou V, Ross AJ, et al. Proapoptotic BAX and BAK: a requisite gateway to mitochondrial dysfunction and death. Science (New York, 662 *NY*) 2001, **292**(5517): 727-730. 663 664 31. Kim H, Rafiuddin-Shah M, Tu HC, Jeffers JR, Zambetti GP, Hsieh JJ, et al. Hierarchical 665 regulation of mitochondrion-dependent apoptosis by BCL-2 subfamilies. Nature cell biology 666 2006, **8**(12): 1348-1358. 667 668 32. Dai H, Smith A, Meng XW, Schneider PA, Pang YP, Kaufmann SH. Transient binding of an 669 activator BH3 domain to the Bak BH3-binding groove initiates Bak oligomerization. The Journal 670 of cell biology 2011, **194**(1): 39-48. 671 672 33. Chen HC, Kanai M, Inoue-Yamauchi A, Tu HC, Huang Y, Ren D, et al. An interconnected 673 hierarchical model of cell death regulation by the BCL-2 family. Nature cell biology 2015, 674 675 **17**(10): 1270-1281. 676 677 34. O'Neill KL, Huang K, Zhang J, Chen Y, Luo X. Inactivation of prosurvival Bcl-2 proteins

activates Bax/Bak through the outer mitochondrial membrane. Genes & development 2016,

678

679

**30**(8): 973-988.

680
681 35. Letai A, Bassik MC, Walensky LD, Sorcinelli MD, Weiler S, Korsmeyer SJ. Distinct BH3
682 domains either sensitize or activate mitochondrial apoptosis, serving as prototype cancer
683 therapeutics. *Cancer cell* 2002, **2**(3): 183-192.

684

Kuwana T, Bouchier-Hayes L, Chipuk JE, Bonzon C, Sullivan BA, Green DR, *et al.* BH3 domains of BH3-only proteins differentially regulate Bax-mediated mitochondrial membrane permeabilization both directly and indirectly. *Molecular cell* 2005, **17**(4): 525-535.

688

689 37. Chen L, Willis SN, Wei A, Smith BJ, Fletcher JI, Hinds MG, *et al.* Differential targeting of prosurvival Bcl-2 proteins by their BH3-only ligands allows complementary apoptotic function.
691 *Molecular cell* 2005, **17**(3): 393-403.

692

Jeng PS, Inoue-Yamauchi A, Hsieh JJ, Cheng EH. BH3-Dependent and Independent Activation of BAX and BAK in Mitochondrial Apoptosis. *Curr Opin Physiol* 2018, **3:** 71-81.

695

Vaseva AV, Moll UM. The mitochondrial p53 pathway. *Biochimica et biophysica acta* 2009, **1787**(5): 414-420.

698

Mihara M, Erster S, Zaika A, Petrenko O, Chittenden T, Pancoska P, *et al.* p53 has a direct apoptogenic role at the mitochondria. *Molecular cell* 2003, **11**(3): 577-590.

701

702 41. Chen ZX, Pervaiz S. Involvement of cytochrome c oxidase subunits Va and Vb in the regulation of cancer cell metabolism by Bcl-2. *Cell death and differentiation* 2010, **17**(3): 408-420.

704

705 42. Chong SJF, Iskandar K, Lai JXH, Qu J, Raman D, Valentin R, *et al.* Serine-70 phosphorylated Bcl-2 prevents oxidative stress-induced DNA damage by modulating the mitochondrial redox metabolism. *Nucleic acids research* 2020, **48**(22): 12727-12745.

708

709 43. Clément MV, Hirpara JL, Pervaiz S. Decrease in intracellular superoxide sensitizes Bcl-2-710 overexpressing tumor cells to receptor and drug-induced apoptosis independent of the 711 mitochondria. *Cell death and differentiation* 2003, **10**(11): 1273-1285.

712

Low IC, Loh T, Huang Y, Virshup DM, Pervaiz S. Ser70 phosphorylation of Bcl-2 by selective tyrosine nitration of PP2A-B56δ stabilizes its antiapoptotic activity. *Blood* 2014, **124**(14): 2223-2234.

716

717 45. Knudson CM, Tung KS, Tourtellotte WG, Brown GA, Korsmeyer SJ. Bax-deficient mice with lymphoid hyperplasia and male germ cell death. *Science (New York, NY)* 1995, **270**(5233): 96-719 99.

Lindsten T, Ross AJ, King A, Zong WX, Rathmell JC, Shiels HA, *et al.* The combined functions of proapoptotic Bcl-2 family members bak and bax are essential for normal development of multiple tissues. *Molecular cell* 2000, 6(6): 1389-1399.

725 47. Ke F, Voss A, Kerr JB, O'Reilly LA, Tai L, Echeverry N, et al. BCL-2 family member BOK is 726 widely expressed but its loss has only minimal impact in mice. *Cell death and differentiation* 727 2012, **19**(6): 915-925.

724

728

732

735

739

743

747

751

- 729 48. Deckwerth TL, Elliott JL, Knudson CM, Johnson EM, Jr., Snider WD, Korsmeyer SJ. BAX is 730 required for neuronal death after trophic factor deprivation and during development. *Neuron* 731 1996, **17**(3): 401-411.
- 733 49. Nguyen DH, Soygur B, Peng SP, Malki S, Hu G, Laird DJ. Apoptosis in the fetal testis eliminates developmentally defective germ cell clones. *Nature cell biology* 2020, **22**(12): 1423-1435.
- 736 50. Russell LD, Chiarini-Garcia H, Korsmeyer SJ, Knudson CM. Bax-dependent spermatogonia apoptosis is required for testicular development and spermatogenesis. *Biology of reproduction* 2002, **66**(4): 950-958.
- 740 51. Rodriguez I, Ody C, Araki K, Garcia I, Vassalli P. An early and massive wave of germinal cell 741 apoptosis is required for the development of functional spermatogenesis. *The EMBO journal* 742 1997, **16**(9): 2262-2270.
- White FA, Keller-Peck CR, Knudson CM, Korsmeyer SJ, Snider WD. Widespread elimination of naturally occurring neuronal death in Bax-deficient mice. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 1998, **18**(4): 1428-1439.
- Fan H, Favero M, Vogel MW. Elimination of Bax expression in mice increases cerebellar purkinje cell numbers but not the number of granule cells. *The Journal of comparative neurology* 2001, **436**(1): 82-91.
- Jung AR, Kim TW, Rhyu IJ, Kim H, Lee YD, Vinsant S, *et al.* Misplacement of Purkinje cells during postnatal development in Bax knock-out mice: a novel role for programmed cell death in the nervous system? *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2008, **28**(11): 2941-2948.
- 55. Sun W, Winseck A, Vinsant S, Park OH, Kim H, Oppenheim RW. Programmed cell death of adult-generated hippocampal neurons is mediated by the proapoptotic gene Bax. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2004, **24**(49): 11205-11213.

760
 761 56. Chang MY, Sun W, Ochiai W, Nakashima K, Kim SY, Park CH, et al. Bcl-XL/Bax proteins direct the fate of embryonic cortical precursor cells. Molecular and cellular biology 2007, 27(12): 4293-4305.

57. Southwell DG, Paredes MF, Galvao RP, Jones DL, Froemke RC, Sebe JY, *et al.* Intrinsically determined cell death of developing cortical interneurons. *Nature* 2012, **491**(7422): 109-113.

764

767

771

774

777

781

784

787

791

795

- Jyotika J, McCutcheon J, Laroche J, Blaustein JD, Forger NG. Deletion of the Bax gene disrupts
   sexual behavior and modestly impairs motor function in mice. *Developmental neurobiology* 2007, 67(11): 1511-1519.
- 59. Luedke AC, Boucher PO, Niel L, Holmes MM. Altered anxiety and defensive behaviors in Bax knockout mice. *Behavioural brain research* 2013, 239: 115-120.
- Krahe TE, Medina AE, Lantz CL, Filgueiras CC. Hyperactivity and depression-like traits in Bax
   KO mice. *Brain research* 2015, **1625**: 246-254.
- Garcia I, Crowther AJ, Gama V, Miller CR, Deshmukh M, Gershon TR. Bax deficiency prolongs cerebellar neurogenesis, accelerates medulloblastoma formation and paradoxically increases both malignancy and differentiation. *Oncogene* 2013, **32**(18): 2304-2314.
- 782 62. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011, **144**(5): 646-783 674.
- 785 63. Ke F, Bouillet P, Kaufmann T, Strasser A, Kerr J, Voss AK. Consequences of the combined loss of BOK and BAK or BOK and BAX. *Cell death & disease* 2013, **4**(6): e650.
- Ke FFS, Vanyai HK, Cowan AD, Delbridge ARD, Whitehead L, Grabow S, *et al.* Embryogenesis and Adult Life in the Absence of Intrinsic Apoptosis Effectors BAX, BAK, and BOK. *Cell* 2018, 173(5): 1217-1230.e1217.
- 792 65. Arakawa S, Tsujioka M, Yoshida T, Tajima-Sakurai H, Nishida Y, Matsuoka Y, *et al.* Role of Atg5-dependent cell death in the embryonic development of Bax/Bak double-knockout mice. *Cell death and differentiation* 2017, **24**(9): 1598-1608.
- Rybstein MD, Bravo-San Pedro JM, Kroemer G, Galluzzi L. The autophagic network and cancer.
   Nature cell biology 2018, 20(3): 243-251.

- Galluzzi L, Green DR. Autophagy-Independent Functions of the Autophagy Machinery. *Cell* 2019, **177**(7): 1682-1699.
- Miller DR, Cramer SD, Thorburn A. The interplay of autophagy and non-apoptotic cell death pathways. *International review of cell and molecular biology* 2020, **352:** 159-187.

804

807

811

815

819

824

828

831

- Fairlie WD, Tran S, Lee EF. Crosstalk between apoptosis and autophagy signaling pathways. *International review of cell and molecular biology* 2020, **352:** 115-158.
- Gu Z, Serradj N, Ueno M, Liang M, Li J, Baccei ML, *et al.* Skilled Movements Require Non-apoptotic Bax/Bak Pathway-Mediated Corticospinal Circuit Reorganization. *Neuron* 2017, **94**(3): 626-641.e624.
- Hahn P, Lindsten T, Ying GS, Bennett J, Milam AH, Thompson CB, *et al.* Proapoptotic bcl-2 family members, Bax and Bak, are essential for developmental photoreceptor apoptosis. *Investigative ophthalmology & visual science* 2003, **44**(8): 3598-3605.
- Hahn P, Lindsten T, Tolentino M, Thompson CB, Bennett J, Dunaief JL. Persistent fetal ocular vasculature in mice deficient in bax and bak. *Archives of ophthalmology (Chicago, Ill: 1960)* 2005, **123**(6): 797-802.
- Ke FS, Holloway S, Uren RT, Wong AW, Little MH, Kluck RM, *et al.* The BCL-2 family member BID plays a role during embryonic development in addition to its BH3-only protein function by acting in parallel to BAX, BAK and BOK. *The EMBO journal* 2022, **41**(15): e110300.
- Flores-Romero H, Hohorst L, John M, Albert MC, King LE, Beckmann L, *et al.* BCL-2-family protein tBID can act as a BAX-like effector of apoptosis. *The EMBO journal* 2022, **41**(2): e108690.
- Suzuki M, Youle RJ, Tjandra N. Structure of Bax: coregulation of dimer formation and intracellular localization. *Cell* 2000, **103**(4): 645-654.
- McDonnell JM, Fushman D, Milliman CL, Korsmeyer SJ, Cowburn D. Solution structure of the proapoptotic molecule BID: a structural basis for apoptotic agonists and antagonists. *Cell* 1999, **96**(5): 625-634.
- Chou JJ, Li H, Salvesen GS, Yuan J, Wagner G. Solution structure of BID, an intracellular amplifier of apoptotic signaling. *Cell* 1999, **96**(5): 615-624.

Takeuchi O, Fisher J, Suh H, Harada H, Malynn BA, Korsmeyer SJ. Essential role of BAX,BAK
in B cell homeostasis and prevention of autoimmune disease. *Proceedings of the National Academy of Sciences of the United States of America* 2005, **102**(32): 11272-11277.

Biswas S, Shi Q, Matise L, Cleveland S, Dave U, Zinkel S. A role for proapoptotic Bax and Bak in T-cell differentiation and transformation. *Blood* 2010, **116**(24): 5237-5246.

842

845

849

853

857

860

864

868

872

- 80. Kodama T, Hikita H, Kawaguchi T, Shigekawa M, Shimizu S, Hayashi Y, *et al.* Mcl-1 and BclxL regulate Bak/Bax-dependent apoptosis of the megakaryocytic lineage at multistages. *Cell death and differentiation* 2012, **19**(11): 1856-1869.
- 81. Pleines I, Lebois M, Gangatirkar P, Au AE, Lane RM, Henley KJ, *et al.* Intrinsic apoptosis circumvents the functional decline of circulating platelets but does not cause the storage lesion. *Blood* 2018, **132**(2): 197-209.
- 82. Ke F, Grabow S, Kelly GL, Lin A, O'Reilly LA, Strasser A. Impact of the combined loss of BOK, BAX and BAK on the hematopoietic system is slightly more severe than compound loss of BAX and BAK. *Cell death & disease* 2015, **6**(10): e1938.
- 858 83. Rathmell JC, Lindsten T, Zong WX, Cinalli RM, Thompson CB. Deficiency in Bak and Bax perturbs thymic selection and lymphoid homeostasis. *Nature immunology* 2002, **3**(10): 932-939.
- 84. Jones RG, Bui T, White C, Madesh M, Krawczyk CM, Lindsten T, *et al.* The proapoptotic factors
  Bax and Bak regulate T Cell proliferation through control of endoplasmic reticulum Ca(2+)
  homeostasis. *Immunity* 2007, **27**(2): 268-280.
- 85. Mason KD, Lin A, Robb L, Josefsson EC, Henley KJ, Gray DH, *et al.* Proapoptotic Bak and Bax guard against fatal systemic and organ-specific autoimmune disease. *Proceedings of the National Academy of Sciences of the United States of America* 2013, **110**(7): 2599-2604.
- 86. Watson EC, Koenig MN, Grant ZL, Whitehead L, Trounson E, Dewson G, *et al.* Apoptosis regulates endothelial cell number and capillary vessel diameter but not vessel regression during retinal angiogenesis. *Development (Cambridge, England)* 2016, **143**(16): 2973-2982.
- 873 87. Grant ZL, Whitehead L, Wong VH, He Z, Yan RY, Miles AR, *et al.* Blocking endothelial apoptosis revascularizes the retina in a model of ischemic retinopathy. *The Journal of clinical investigation* 2020, **130**(8): 4235-4251.

- 88. Whelan RS, Konstantinidis K, Wei AC, Chen Y, Reyna DE, Jha S, *et al.* Bax regulates primary necrosis through mitochondrial dynamics. *Proceedings of the National Academy of Sciences of the United States of America* 2012, **109**(17): 6566-6571.
- 881 89. Kirsch DG, Santiago PM, di Tomaso E, Sullivan JM, Hou WS, Dayton T, *et al.* p53 controls radiation-induced gastrointestinal syndrome in mice independent of apoptosis. *Science (New York, NY)* 2010, **327**(5965): 593-596.

884

888

892

895

898

902

906

910

- 885 90. Bouillet P, Metcalf D, Huang DC, Tarlinton DM, Kay TW, Köntgen F, *et al.* Proapoptotic Bcl-2 relative Bim required for certain apoptotic responses, leukocyte homeostasis, and to preclude autoimmunity. *Science (New York, NY)* 1999, **286**(5445): 1735-1738.
- Wang YM, Zhang GY, Wang Y, Hu M, Zhou JJ, Sawyer A, *et al.* Exacerbation of spontaneous autoimmune nephritis following regulatory T cell depletion in B cell lymphoma 2-interacting mediator knock-out mice. *Clinical and experimental immunology* 2017, **188**(2): 195-207.
- Puthalakath H, O'Reilly LA, Gunn P, Lee L, Kelly PN, Huntington ND, *et al.* ER stress triggers apoptosis by activating BH3-only protein Bim. *Cell* 2007, **129**(7): 1337-1349.
- Hutcheson J, Perlman H. Loss of Bim results in abnormal accumulation of mature CD4-CD8-CD44-CD25- thymocytes. *Immunobiology* 2007, **212**(8): 629-636.
- Chougnet CA, Tripathi P, Lages CS, Raynor J, Sholl A, Fink P, *et al.* A major role for Bim in regulatory T cell homeostasis. *Journal of immunology (Baltimore, Md : 1950)* 2011, **186**(1): 156-163.
- 903 95. Bouillet P, Purton JF, Godfrey DI, Zhang LC, Coultas L, Puthalakath H, *et al.* BH3-only Bcl-2 family member Bim is required for apoptosis of autoreactive thymocytes. *Nature* 2002, **415**(6874): 922-926.
- 907 96. Enders A, Bouillet P, Puthalakath H, Xu Y, Tarlinton DM, Strasser A. Loss of the pro-apoptotic BH3-only Bcl-2 family member Bim inhibits BCR stimulation-induced apoptosis and deletion of autoreactive B cells. *The Journal of experimental medicine* 2003, **198**(7): 1119-1126.
- 91. Zhan Y, Zhang Y, Gray D, Carrington EM, Bouillet P, Ko HJ, et al. Defects in the Bcl-2-regulated
   91. apoptotic pathway lead to preferential increase of CD25 low Foxp3+ anergic CD4+ T cells.
   91. Journal of immunology (Baltimore, Md: 1950) 2011, 187(4): 1566-1577.
- 915 98. Pellegrini M, Belz G, Bouillet P, Strasser A. Shutdown of an acute T cell immune response to viral infection is mediated by the proapoptotic Bcl-2 homology 3-only protein Bim. *Proceedings*

- of the National Academy of Sciences of the United States of America 2003, **100**(24): 14175-14180.
- 99. Hildeman DA, Zhu Y, Mitchell TC, Bouillet P, Strasser A, Kappler J, *et al.* Activated T cell death
   in vivo mediated by proapoptotic bcl-2 family member bim. *Immunity* 2002, **16**(6): 759-767.

922

926

930

934

938

942

946

950

- 923 100. Pellegrini M, Bouillet P, Robati M, Belz GT, Davey GM, Strasser A. Loss of Bim increases T cell production and function in interleukin 7 receptor-deficient mice. *The Journal of experimental medicine* 2004, **200**(9): 1189-1195.
- 927 101. Fischer SF, Bouillet P, O'Donnell K, Light A, Tarlinton DM, Strasser A. Proapoptotic BH3-only 928 protein Bim is essential for developmentally programmed death of germinal center-derived 929 memory B cells and antibody-forming cells. *Blood* 2007, **110**(12): 3978-3984.
- 931 102. Sugimoto-Ishige A, Harada M, Tanaka M, Terooatea T, Adachi Y, Takahashi Y, *et al.* Bim establishes the B-cell repertoire from early to late in the immune response. *International immunology* 2021, **33**(2): 79-90.
- 935 103. Oliver PM, Wang M, Zhu Y, White J, Kappler J, Marrack P. Loss of Bim allows precursor B cell survival but not precursor B cell differentiation in the absence of interleukin 7. *The Journal of experimental medicine* 2004, **200**(9): 1179-1187.
- Villunger A, Scott C, Bouillet P, Strasser A. Essential role for the BH3-only protein Bim but redundant roles for Bax, Bcl-2, and Bcl-w in the control of granulocyte survival. *Blood* 2003, **101**(6): 2393-2400.
- 943 105. Mailleux AA, Overholtzer M, Schmelzle T, Bouillet P, Strasser A, Brugge JS. BIM regulates 944 apoptosis during mammary ductal morphogenesis, and its absence reveals alternative cell death 945 mechanisms. *Developmental cell* 2007, **12**(2): 221-234.
- 947 106. Schuler F, Baumgartner F, Klepsch V, Chamson M, Müller-Holzner E, Watson CJ, *et al.* The 948 BH3-only protein BIM contributes to late-stage involution in the mouse mammary gland. *Cell* 949 *death and differentiation* 2016, **23**(1): 41-51.
- 951 107. Ohgushi M, Kuroki S, Fukamachi H, O'Reilly LA, Kuida K, Strasser A, *et al.* Transforming growth factor beta-dependent sequential activation of Smad, Bim, and caspase-9 mediates physiological apoptosis in gastric epithelial cells. *Molecular and cellular biology* 2005, **25**(22): 10017-10028.

- Doonan F, Donovan M, Gomez-Vicente V, Bouillet P, Cotter TG. Bim expression indicates the pathway to retinal cell death in development and degeneration. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2007, **27**(40): 10887-10894.
- 960 109. Wali JA, Galic S, Tan CY, Gurzov EN, Frazier AE, Connor T, *et al.* Loss of BIM increases mitochondrial oxygen consumption and lipid oxidation, reduces adiposity and improves insulin sensitivity in mice. *Cell death and differentiation* 2018, **25**(1): 217-225.

963

967

971

975

979

983

987

991

- Hutcheson J, Scatizzi JC, Bickel E, Brown NJ, Bouillet P, Strasser A, *et al.* Combined loss of proapoptotic genes Bak or Bax with Bim synergizes to cause defects in hematopoiesis and in thymocyte apoptosis. *The Journal of experimental medicine* 2005, **201**(12): 1949-1960.
- Liu R, King A, Bouillet P, Tarlinton DM, Strasser A, Heierhorst J. Proapoptotic BIM Impacts B
   Lymphoid Homeostasis by Limiting the Survival of Mature B Cells in a Cell-Autonomous
   Manner. Frontiers in immunology 2018, 9: 592.
- 972 112. Herold MJ, Stuchbery R, Mérino D, Willson T, Strasser A, Hildeman D, *et al.* Impact of conditional deletion of the pro-apoptotic BCL-2 family member BIM in mice. *Cell death & disease* 2014, **5**(10): e1446.
- Huntington ND, Labi V, Cumano A, Vieira P, Strasser A, Villunger A, et al. Loss of the pro apoptotic BH3-only Bcl-2 family member Bim sustains B lymphopoiesis in the absence of IL-7.
   *International immunology* 2009, 21(6): 715-725.
- Ludwig LM, Roach LE, Katz SG, LaBelle JL. Loss of BIM in T cells results in BCL-2 family
   BH3-member compensation but incomplete cell death sensitivity normalization. *Apoptosis : an international journal on programmed cell death* 2020, 25(3-4): 247-260.
- 984 115. Bunk EC, König HG, Bernas T, Engel T, Henshall DC, Kirby BP, *et al.* BH3-only proteins BIM and PUMA in the regulation of survival and neuronal differentiation of newly generated cells in the adult mouse hippocampus. *Cell death & disease* 2010, **1**(1): e15.
- 988 116. Tsai F, Homan PJ, Agrawal H, Misharin AV, Abdala-Valencia H, Haines GK, 3rd, *et al.* Bim suppresses the development of SLE by limiting myeloid inflammatory responses. *The Journal of experimental medicine* 2017, **214**(12): 3753-3773.
- Yin XM, Wang K, Gross A, Zhao Y, Zinkel S, Klocke B, et al. Bid-deficient mice are resistant to Fas-induced hepatocellular apoptosis. Nature 1999, 400(6747): 886-891.

- Leonard JR, D'Sa C, Cahn BR, Korsmeyer SJ, Roth KA. Bid regulation of neuronal apoptosis.
   *Brain research Developmental brain research* 2001, 128(2): 187-190.
- 998 119. Villunger A, Michalak EM, Coultas L, Müllauer F, Böck G, Ausserlechner MJ, *et al.* p53- and drug-induced apoptotic responses mediated by BH3-only proteins puma and noxa. *Science (New York, NY)* 2003, **302**(5647): 1036-1038.

1001

1004

1008

1012

1015

1019

1023

1027

- 120. Jeffers JR, Parganas E, Lee Y, Yang C, Wang J, Brennan J, et al. Puma is an essential mediator of p53-dependent and -independent apoptotic pathways. *Cancer cell* 2003, **4**(4): 321-328.
- 121. Kaufmann T, Tai L, Ekert PG, Huang DC, Norris F, Lindemann RK, *et al.* The BH3-only protein bid is dispensable for DNA damage- and replicative stress-induced apoptosis or cell-cycle arrest. *Cell* 2007, **129**(2): 423-433.
- 1009 122. McKenzie MD, Carrington EM, Kaufmann T, Strasser A, Huang DC, Kay TW, *et al.*1010 Proapoptotic BH3-only protein Bid is essential for death receptor-induced apoptosis of pancreatic
  1011 beta-cells. *Diabetes* 2008, **57**(5): 1284-1292.
- 1013 123. Jost PJ, Grabow S, Gray D, McKenzie MD, Nachbur U, Huang DC, *et al.* XIAP discriminates between type I and type II FAS-induced apoptosis. *Nature* 2009, **460**(7258): 1035-1039.
- 1016 124. Engel T, Caballero-Caballero A, Schindler CK, Plesnila N, Strasser A, Prehn JH, *et al.* BH3-only protein Bid is dispensable for seizure-induced neuronal death and the associated nuclear accumulation of apoptosis-inducing factor. *Journal of neurochemistry* 2010, **115**(1): 92-101.
- 125. Zinkel SS, Ong CC, Ferguson DO, Iwasaki H, Akashi K, Bronson RT, *et al.* Proapoptotic BID is required for myeloid homeostasis and tumor suppression. *Genes & development* 2003, **17**(2): 229-239.
- 126. Salisbury-Ruf CT, Bertram CC, Vergeade A, Lark DS, Shi Q, Heberling ML, *et al.* Bid maintains mitochondrial cristae structure and function and protects against cardiac disease in an integrative genomics study. *eLife* 2018, **7**.
- Wree A, Johnson CD, Font-Burgada J, Eguchi A, Povero D, Karin M, *et al.* Hepatocyte-specific Bid depletion reduces tumor development by suppressing inflammation-related compensatory proliferation. *Cell death and differentiation* 2015, **22**(12): 1985-1994.
- 1032 128. Tischner D, Gaggl I, Peschel I, Kaufmann M, Tuzlak S, Drach M, *et al.* Defective cell death signalling along the Bcl-2 regulated apoptosis pathway compromises Treg cell development and limits their functionality in mice. *Journal of autoimmunity* 2012, **38**(1): 59-69.

Lazic M, Eguchi A, Berk MP, Povero D, Papouchado B, Mulya A, et al. Differential regulation of inflammation and apoptosis in Fas-resistant hepatocyte-specific Bid-deficient mice. *Journal of hepatology* 2014, 61(1): 107-115.

130. Myers M, Morgan FH, Liew SH, Zerafa N, Gamage TU, Sarraj M, *et al.* PUMA regulates germ cell loss and primordial follicle endowment in mice. *Reproduction (Cambridge, England)* 2014, **148**(2): 211-219.

1039

1043

1047

1050

1054

1058

1062

1065

1069

- 131. Erlacher M, Michalak EM, Kelly PN, Labi V, Niederegger H, Coultas L, *et al.* BH3-only proteins Puma and Bim are rate-limiting for gamma-radiation- and glucocorticoid-induced apoptosis of lymphoid cells in vivo. *Blood* 2005, **106**(13): 4131-4138.
- Wang J, Thomas HR, Li Z, Yeo NCF, Scott HE, Dang N, *et al.* Puma, noxa, p53, and p63 differentially mediate stress pathway induced apoptosis. *Cell death & disease* 2021, **12**(7): 659.
- 1051 133. Kerr JB, Hutt KJ, Michalak EM, Cook M, Vandenberg CJ, Liew SH, *et al.* DNA damage-induced primordial follicle oocyte apoptosis and loss of fertility require TAp63-mediated induction of Puma and Noxa. *Molecular cell* 2012, **48**(3): 343-352.
- 134. Naik E, Michalak EM, Villunger A, Adams JM, Strasser A. Ultraviolet radiation triggers apoptosis of fibroblasts and skin keratinocytes mainly via the BH3-only protein Noxa. *The Journal of cell biology* 2007, **176**(4): 415-424.
- 1059 135. Wensveen FM, Geest CR, Libregts S, Derks IAM, Ekert PG, Labi V, et al. BH3-only protein 1060 Noxa contributes to apoptotic control of stress-erythropoiesis. Apoptosis: an international 1061 journal on programmed cell death 2013, **18**(11): 1306-1318.
- 136. Clybouw C, Fischer S, Auffredou MT, Hugues P, Alexia C, Bouillet P, *et al.* Regulation of memory B-cell survival by the BH3-only protein Puma. *Blood* 2011, **118**(15): 4120-4128.
- 137. Wensveen FM, Derks IA, van Gisbergen KP, de Bruin AM, Meijers JC, Yigittop H, *et al.* BH3-1067 only protein Noxa regulates apoptosis in activated B cells and controls high-affinity antibody 1068 formation. *Blood* 2012, **119**(6): 1440-1449.
- 138. Harder JM, Libby RT. BBC3 (PUMA) regulates developmental apoptosis but not axonal injury induced death in the retina. *Molecular neurodegeneration* 2011, **6:** 50.

- 139. Labi V, Erlacher M, Krumschnabel G, Manzl C, Tzankov A, Pinon J, et al. Apoptosis of leukocytes triggered by acute DNA damage promotes lymphoma formation. Genes & development 2010, 24(15): 1602-1607.
- 140. Michalak EM, Vandenberg CJ, Delbridge AR, Wu L, Scott CL, Adams JM, *et al.* Apoptosis-1078 promoted tumorigenesis: gamma-irradiation-induced thymic lymphomagenesis requires Puma-1079 driven leukocyte death. *Genes & development* 2010, **24**(15): 1608-1613.

1080

1084

1087

1090

1094

1098

1102

1106

- 1081 141. Michalak EM, Jansen ES, Happo L, Cragg MS, Tai L, Smyth GK, *et al.* Puma and to a lesser extent Noxa are suppressors of Myc-induced lymphomagenesis. *Cell death and differentiation* 2009, **16**(5): 684-696.
- 1085 142. Qiu W, Wang X, Leibowitz B, Yang W, Zhang L, Yu J. PUMA-mediated apoptosis drives chemical hepatocarcinogenesis in mice. *Hepatology (Baltimore, Md)* 2011, **54**(4): 1249-1258.
- Huang Q, Li F, Liu X, Li W, Shi W, Liu FF, *et al.* Caspase 3-mediated stimulation of tumor cell repopulation during cancer radiotherapy. *Nature medicine* 2011, **17**(7): 860-866.
- 1091 144. Böttcher JP, Bonavita E, Chakravarty P, Blees H, Cabeza-Cabrerizo M, Sammicheli S, *et al.* NK Cells Stimulate Recruitment of cDC1 into the Tumor Microenvironment Promoting Cancer Immune Control. *Cell* 2018, **172**(5): 1022-1037.e1014.
- 1095 145. Qiu W, Carson-Walter EB, Liu H, Epperly M, Greenberger JS, Zambetti GP, *et al.* PUMA regulates intestinal progenitor cell radiosensitivity and gastrointestinal syndrome. *Cell stem cell* 2008, **2**(6): 576-583.
- 1099 146. Michalak EM, Villunger A, Adams JM, Strasser A. In several cell types tumour suppressor p53
   1100 induces apoptosis largely via Puma but Noxa can contribute. *Cell death and differentiation* 2008,
   1101 15(6): 1019-1029.
- 1103 147. Erlacher M, Labi V, Manzl C, Böck G, Tzankov A, Häcker G, *et al.* Puma cooperates with Bim, the rate-limiting BH3-only protein in cell death during lymphocyte development, in apoptosis induction. *The Journal of experimental medicine* 2006, **203**(13): 2939-2951.
- 148. Gray DH, Kupresanin F, Berzins SP, Herold MJ, O'Reilly LA, Bouillet P, *et al.* The BH3-only proteins Bim and Puma cooperate to impose deletional tolerance of organ-specific antigens. *Immunity* 2012, **37**(3): 451-462.

Happo L, Cragg MS, Phipson B, Haga JM, Jansen ES, Herold MJ, *et al.* Maximal killing of lymphoma cells by DNA damage-inducing therapy requires not only the p53 targets Puma and Noxa, but also Bim. *Blood* 2010, **116**(24): 5256-5267.

1114

1115 150. Kaufmann T, Jost PJ, Pellegrini M, Puthalakath H, Gugasyan R, Gerondakis S, *et al.* Fatal hepatitis mediated by tumor necrosis factor TNFalpha requires caspase-8 and involves the BH3-1117 only proteins Bid and Bim. *Immunity* 2009, **30**(1): 56-66.

1118

1119 151. Ren D, Tu HC, Kim H, Wang GX, Bean GR, Takeuchi O, *et al.* BID, BIM, and PUMA are essential for activation of the BAX- and BAK-dependent cell death program. *Science (New York, NY)* 2010, **330**(6009): 1390-1393.

1122

1123 152. Ranger AM, Zha J, Harada H, Datta SR, Danial NN, Gilmore AP, *et al.* Bad-deficient mice develop diffuse large B cell lymphoma. *Proceedings of the National Academy of Sciences of the United States of America* 2003, **100**(16): 9324-9329.

1126

1127 153. Imaizumi K, Benito A, Kiryu-Seo S, Gonzalez V, Inohara N, Lieberman AP, *et al.* Critical role for DP5/Harakiri, a Bcl-2 homology domain 3-only Bcl-2 family member, in axotomy-induced neuronal cell death. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2004, **24**(15): 3721-3725.

1131

1132 Labi V, Erlacher M, Kiessling S, Manzl C, Frenzel A, O'Reilly L, *et al.* Loss of the BH3-only protein Bmf impairs B cell homeostasis and accelerates gamma irradiation-induced thymic lymphoma development. *The Journal of experimental medicine* 2008, **205**(3): 641-655.

1135

136 155. Coultas L, Bouillet P, Stanley EG, Brodnicki TC, Adams JM, Strasser A. Proapoptotic BH3-only Bcl-2 family member Bik/Blk/Nbk is expressed in hemopoietic and endothelial cells but is redundant for their programmed death. *Molecular and cellular biology* 2004, **24**(4): 1570-1581.

1139

156. Kelly PN, White MJ, Goschnick MW, Fairfax KA, Tarlinton DM, Kinkel SA, *et al.* Individual and overlapping roles of BH3-only proteins Bim and Bad in apoptosis of lymphocytes and platelets and in suppression of thymic lymphoma development. *Cell death and differentiation* 2010, **17**(10): 1655-1664.

1144

Hübner A, Cavanagh-Kyros J, Rincon M, Flavell RA, Davis RJ. Functional cooperation of the proapoptotic Bcl2 family proteins Bmf and Bim in vivo. *Molecular and cellular biology* 2010, 30(1): 98-105.

1148

1149 158. Vaithiyanathan K, Liew SH, Zerafa N, Gamage T, Cook M, O'Reilly LA, *et al.* BCL2-modifying factor promotes germ cell loss during murine oogenesis. *Reproduction (Cambridge, England)* 2016, **151**(5): 553-562.

1152 159. 1153 Liew SH, Vaithiyanathan K, Cook M, Bouillet P, Scott CL, Kerr JB, et al. Loss of the proapoptotic BH3-only protein BCL-2 modifying factor prolongs the fertile life span in female 1154

1155 mice. Biology of reproduction 2014, **90**(4): 77.

1160

1164

1168

1172

1176

1179

1182

1186

1190

1156 Happo L, Phipson B, Smyth GK, Strasser A, Scott CL. Neither loss of Bik alone, nor combined 1157 160. loss of Bik and Noxa, accelerate murine lymphoma development or render lymphoma cells 1158

resistant to DNA damaging drugs. *Cell death & disease* 2012, **3**(5): e306. 1159

Baumgartner F, Woess C, Pedit V, Tzankov A, Labi V, Villunger A. Minor cell-death defects but 1161 161. 1162 reduced tumor latency in mice lacking the BH3-only proteins Bad and Bmf. Oncogene 2013, **32**(5): 621-630. 1163

1165 162. Coultas L, Bouillet P, Loveland KL, Meachem S, Perlman H, Adams JM, et al. Concomitant loss of proapoptotic BH3-only Bcl-2 antagonists Bik and Bim arrests spermatogenesis. The EMBO 1166 1167 journal 2005, **24**(22): 3963-3973.

1169 163. Labi V, Woess C, Tuzlak S, Erlacher M, Bouillet P, Strasser A, et al. Deregulated cell death and lymphocyte homeostasis cause premature lethality in mice lacking the BH3-only proteins Bim 1170 and Bmf. Blood 2014, 123(17): 2652-2662. 1171

164. Woess C, Tuzlak S, Labi V, Drach M, Bertele D, Schneider P, et al. Combined loss of the BH3-1173 only proteins Bim and Bmf restores B-cell development and function in TACI-Ig transgenic mice. 1174 *Cell death and differentiation* 2015, **22**(9): 1477-1488. 1175

1177 165. Rinkenberger JL, Horning S, Klocke B, Roth K, Korsmeyer SJ. Mcl-1 deficiency results in periimplantation embryonic lethality. Genes & development 2000, **14**(1): 23-27. 1178

166. Kuida K, Haydar TF, Kuan CY, Gu Y, Taya C, Karasuyama H, et al. Reduced apoptosis and 1180 cytochrome c-mediated caspase activation in mice lacking caspase 9. Cell 1998, **94**(3): 325-337. 1181

167. Motoyama N, Wang F, Roth KA, Sawa H, Nakayama K, Nakayama K, et al. Massive cell death 1183 of immature hematopoietic cells and neurons in Bcl-x-deficient mice. Science (New York, NY) 1184 1995, **267**(5203): 1506-1510. 1185

Zaidi AU, D'Sa-Eipper C, Brenner J, Kuida K, Zheng TS, Flavell RA, et al. Bcl-X(L)-caspase-9 1187 168. 1188 interactions in the developing nervous system: evidence for multiple death pathways. The Journal 1189 of neuroscience: the official journal of the Society for Neuroscience 2001, **21**(1): 169-175.

1191 169. Shindler KS, Latham CB, Roth KA. Bax deficiency prevents the increased cell death of immature neurons in bcl-x-deficient mice. *The Journal of neuroscience : the official journal of the Society* 1193 for Neuroscience 1997, **17**(9): 3112-3119.

1194

170. Akhtar RS, Klocke BJ, Strasser A, Roth KA. Loss of BH3-only protein Bim inhibits apoptosis of hemopoietic cells in the fetal liver and male germ cells but not neuronal cells in bcl-x-deficient mice. *The journal of histochemistry and cytochemistry : official journal of the Histochemistry Society* 2008, **56**(10): 921-927.

1199

1200 171. Bouillet P, Cory S, Zhang LC, Strasser A, Adams JM. Degenerative disorders caused by Bcl-2 1201 deficiency prevented by loss of its BH3-only antagonist Bim. *Developmental cell* 2001, **1**(5): 1202 645-653.

1203

1204 172. Veis DJ, Sorenson CM, Shutter JR, Korsmeyer SJ. Bcl-2-deficient mice demonstrate fulminant lymphoid apoptosis, polycystic kidneys, and hypopigmented hair. *Cell* 1993, **75**(2): 229-240.

1206

1207 173. Nakayama K, Nakayama K, Negishi I, Kuida K, Shinkai Y, Louie MC, *et al.* Disappearance of the lymphoid system in Bcl-2 homozygous mutant chimeric mice. *Science (New York, NY)* 1993, 261(5128): 1584-1588.

1210

174. Kamada S, Shimono A, Shinto Y, Tsujimura T, Takahashi T, Noda T, *et al.* bcl-2 deficiency in mice leads to pleiotropic abnormalities: accelerated lymphoid cell death in thymus and spleen, polycystic kidney, hair hypopigmentation, and distorted small intestine. *Cancer research* 1995, 55(2): 354-359.

1215

175. Michaelidis TM, Sendtner M, Cooper JD, Airaksinen MS, Holtmann B, Meyer M, *et al.*1217 Inactivation of bcl-2 results in progressive degeneration of motoneurons, sympathetic and sensory
1218 neurons during early postnatal development. *Neuron* 1996, **17**(1): 75-89.

1219

1220 176. Manzl C, Baumgartner F, Peintner L, Schuler F, Villunger A. Possible pitfalls investigating cell death responses in genetically engineered mouse models and derived cell lines. *Methods (San Diego, Calif)* 2013, **61**(2): 130-137.

1223

1224 177. Carpinelli MR, Wise AK, Arhatari BD, Bouillet P, Manji SS, Manning MG, *et al.* Anti-apoptotic gene Bcl2 is required for stapes development and hearing. *Cell death & disease* 2012, **3**(8): e362.

1226

1227 178. Hamasaki A, Sendo F, Nakayama K, Ishida N, Negishi I, Nakayama K, *et al.* Accelerated neutrophil apoptosis in mice lacking A1-a, a subtype of the bcl-2-related A1 gene. *The Journal of experimental medicine* 1998, **188**(11): 1985-1992.

- 1231 179. Xiang Z, Ahmed AA, Möller C, Nakayama K, Hatakeyama S, Nilsson G. Essential role of the prosurvival bcl-2 homologue A1 in mast cell survival after allergic activation. *The Journal of experimental medicine* 2001, **194**(11): 1561-1569.
- 180. Schenk RL, Tuzlak S, Carrington EM, Zhan Y, Heinzel S, Teh CE, *et al.* Characterisation of mice lacking all functional isoforms of the pro-survival BCL-2 family member A1 reveals minor defects in the haematopoietic compartment. *Cell death and differentiation* 2017, **24**(3): 534-545.

1238

1242

1246

1249

1252

1256

1260

1264

- 1239 181. Tuzlak S, Schenk RL, Vasanthakumar A, Preston SP, Haschka MD, Zotos D, *et al.* The BCL-2 pro-survival protein A1 is dispensable for T cell homeostasis on viral infection. *Cell death and differentiation* 2017, **24**(3): 523-533.
- 1243 182. Print CG, Loveland KL, Gibson L, Meehan T, Stylianou A, Wreford N, et al. Apoptosis regulator 1244 bcl-w is essential for spermatogenesis but appears otherwise redundant. Proceedings of the 1245 National Academy of Sciences of the United States of America 1998, **95**(21): 12424-12431.
- 1247 183. Ross AJ, Waymire KG, Moss JE, Parlow AF, Skinner MK, Russell LD, *et al.* Testicular degeneration in Bclw-deficient mice. *Nature genetics* 1998, **18**(3): 251-256.
- 1250 184. Russell LD, Warren J, Debeljuk L, Richardson LL, Mahar PL, Waymire KG, *et al.*1251 Spermatogenesis in Bclw-deficient mice. *Biology of reproduction* 2001, **65**(1): 318-332.
- 1253 185. Brinkmann K, Grabow S, Hyland CD, Teh CE, Alexander WS, Herold MJ, et al. The combination of reduced MCL-1 and standard chemotherapeutics is tolerable in mice. Cell death and differentiation 2017, **24**(12): 2032-2043.
- 1257 186. Delbridge AR, Opferman JT, Grabow S, Strasser A. Antagonism between MCL-1 and PUMA governs stem/progenitor cell survival during hematopoietic recovery from stress. *Blood* 2015, 125(21): 3273-3280.
- 1261 187. Kasai S, Chuma S, Motoyama N, Nakatsuji N. Haploinsufficiency of Bcl-x leads to male-specific defects in fetal germ cells: differential regulation of germ cell apoptosis between the sexes.

  1263 Developmental biology 2003, 264(1): 202-216.
- 188. Mason KD, Carpinelli MR, Fletcher JI, Collinge JE, Hilton AA, Ellis S, *et al.* Programmed anuclear cell death delimits platelet life span. *Cell* 2007, **128**(6): 1173-1186.
- 1268 189. Schenk RL, Gangoda L, Lawlor KE, O'Reilly LA, Strasser A, Herold MJ. The pro-survival Bcl-1269 2 family member A1 delays spontaneous and FAS ligand-induced apoptosis of activated 1270 neutrophils. *Cell death & disease* 2020, **11**(6): 474.

1271 190. 1272 Grabow S, Kueh AJ, Ke F, Vanyai HK, Sheikh BN, Dengler MA, et al. Subtle Changes in the Levels of BCL-2 Proteins Cause Severe Craniofacial Abnormalities. Cell reports 2018, 24(12): 1273 1274 3285-3295.e3284. 1275 191. 1276 Ke F, Lancaster GI, Grabow S, Murphy AJ, Strasser A. Combined reduction in the expression of MCL-1 and BCL-2 reduces organismal size in mice. Cell death & disease 2020, 11(3): 185. 1277 1278 1279 192. Opferman JT, Iwasaki H, Ong CC, Suh H, Mizuno S, Akashi K, et al. Obligate role of antiapoptotic MCL-1 in the survival of hematopoietic stem cells. Science (New York, NY) 2005, 1280 1281 **307**(5712)**:** 1101-1104. 1282 1283 193. Opferman JT, Letai A, Beard C, Sorcinelli MD, Ong CC, Korsmeyer SJ. Development and maintenance of B and T lymphocytes requires antiapoptotic MCL-1. Nature 2003, 426(6967): 1284 671-676. 1285 1286 194. Pierson W, Cauwe B, Policheni A, Schlenner SM, Franckaert D, Berges J, et al. Antiapoptotic 1287 1288 Mcl-1 is critical for the survival and niche-filling capacity of Foxp3+ regulatory T cells. *Nature* immunology 2013, **14**(9): 959-965. 1289 1290 195. Vikstrom I, Carotta S, Lüthje K, Peperzak V, Jost PJ, Glaser S, et al. Mcl-1 is essential for 1291 germinal center formation and B cell memory. Science (New York, NY) 2010, 330(6007): 1095-1292 1293 1099. 1294 1295 196. Tripathi P, Koss B, Opferman JT, Hildeman DA. Mcl-1 antagonizes Bax/Bak to promote effector 1296 CD4(+) and CD8(+) T-cell responses. *Cell death and differentiation* 2013, **20**(8): 998-1007. 1297 1298 197. Dunkle A, Dzhagalov I, He YW. Mcl-1 promotes survival of thymocytes by inhibition of Bak in a pathway separate from Bcl-2. Cell death and differentiation 2010, 17(6): 994-1002. 1299 1300 198. 1301 Sathe P, Delconte RB, Souza-Fonseca-Guimaraes F, Seillet C, Chopin M, Vandenberg CJ, et al. Innate immunodeficiency following genetic ablation of Mcl1 in natural killer cells. Nature 1302 communications 2014, **5:** 4539. 1303 1304 1305 199. Dzhagalov I, St John A, He YW. The antiapoptotic protein Mcl-1 is essential for the survival of neutrophils but not macrophages. *Blood* 2007, **109**(4): 1620-1626. 1306 1307 Steimer DA, Boyd K, Takeuchi O, Fisher JK, Zambetti GP, Opferman JT. Selective roles for 1308 200.

antiapoptotic MCL-1 during granulocyte development and macrophage effector function. Blood

1309 1310

2009, **113**(12): 2805-2815.

1311 1312 201. Lilla JN, Chen CC, Mukai K, BenBarak MJ, Franco CB, Kalesnikoff J, et al. Reduced mast cell and basophil numbers and function in Cpa3-Cre; Mcl-1fl/fl mice. Blood 2011, 118(26): 6930-1313 1314 6938. 1315 1316 202. Slomp A, Peperzak V. Role and Regulation of Pro-survival BCL-2 Proteins in Multiple Myeloma. Frontiers in oncology 2018, 8: 533. 1317 1318 1319 203. Peperzak V, Vikström I, Walker J, Glaser SP, LePage M, Coquery CM, et al. Mcl-1 is essential for the survival of plasma cells. *Nature immunology* 2013, **14**(3): 290-297. 1320 1321 Carrington EM, Zhan Y, Brady JL, Zhang JG, Sutherland RM, Anstee NS, et al. Anti-apoptotic 204. 1322 1323 proteins BCL-2, MCL-1 and A1 summate collectively to maintain survival of immune cell populations both in vitro and in vivo. Cell death and differentiation 2017, 24(5): 878-888. 1324 1325 1326 205. Dzhagalov I, Dunkle A, He YW. The anti-apoptotic Bcl-2 family member Mcl-1 promotes T lymphocyte survival at multiple stages. Journal of immunology (Baltimore, Md: 1950) 2008, 1327 1328 **181**(1): 521-528. 1329 1330 206. Malin S, McManus S, Cobaleda C, Novatchkova M, Delogu A, Bouillet P, et al. Role of STAT5 in controlling cell survival and immunoglobulin gene recombination during pro-B cell 1331 development. *Nature immunology* 2010, **11**(2): 171-179. 1332 1333 1334 207. Debrincat MA, Josefsson EC, James C, Henley KJ, Ellis S, Lebois M, et al. Mcl-1 and Bcl-x(L) coordinately regulate megakaryocyte survival. *Blood* 2012, **119**(24): 5850-5858. 1335 1336 208. Josefsson EC, James C, Henley KJ, Debrincat MA, Rogers KL, Dowling MR, et al. 1337 1338 Megakaryocytes possess a functional intrinsic apoptosis pathway that must be restrained to survive and produce platelets. The Journal of experimental medicine 2011, 208(10): 2017-2031. 1339 1340 209. Wagner KU, Claudio E, Rucker EB, 3rd, Riedlinger G, Broussard C, Schwartzberg PL, et al. 1341 Conditional deletion of the Bcl-x gene from erythroid cells results in hemolytic anemia and 1342 1343 profound splenomegaly. Development (Cambridge, England) 2000, 127(22): 4949-4958. 1344 1345 210. Vikström IB, Slomp A, Carrington EM, Moesbergen LM, Chang C, Kelly GL, et al. MCL-1 is required throughout B-cell development and its loss sensitizes specific B-cell subsets to inhibition 1346 1347 of BCL-2 or BCL-XL. *Cell death & disease* 2016, **7**(8): e2345.

- Sochalska M, Ottina E, Tuzlak S, Herzog S, Herold M, Villunger A. Conditional knockdown of
   BCL2A1 reveals rate-limiting roles in BCR-dependent B-cell survival. *Cell death and differentiation* 2016, 23(4): 628-639.
- Debrincat MA, Pleines I, Lebois M, Lane RM, Holmes ML, Corbin J, *et al.* BCL-2 is dispensable for thrombopoiesis and platelet survival. *Cell death & disease* 2015, **6**(4): e1721.

1355

1359

1362

1365

1368

1372

1376

- Ma A, Pena JC, Chang B, Margosian E, Davidson L, Alt FW, et al. Bclx regulates the survival of double-positive thymocytes. Proceedings of the National Academy of Sciences of the United States of America 1995, 92(11): 4763-4767.
- Matsuzaki Y, Nakayama K, Nakayama K, Tomita T, Isoda M, Loh DY, *et al.* Role of bcl-2 in the development of lymphoid cells from the hematopoietic stem cell. *Blood* 1997, **89**(3): 853-862.
- 1363 215. Geueke A, Mantellato G, Kuester F, Schettina P, Nelles M, Seeger JM, *et al.* The anti-apoptotic Bcl-2 protein regulates hair follicle stem cell function. *EMBO reports* 2021, **22**(10): e52301.
- Thomas RL, Gustafsson AB. MCL1 is critical for mitochondrial function and autophagy in the heart. *Autophagy* 2013, **9**(11): 1902-1903.
- Wang X, Bathina M, Lynch J, Koss B, Calabrese C, Frase S, *et al.* Deletion of MCL-1 causes lethal cardiac failure and mitochondrial dysfunction. *Genes & development* 2013, **27**(12): 1351-1364.
- 1373 218. Arbour N, Vanderluit JL, Le Grand JN, Jahani-Asl A, Ruzhynsky VA, Cheung EC, *et al.* Mcl-1 is a key regulator of apoptosis during CNS development and after DNA damage. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2008, **28**(24): 6068-6078.
- 1377 219. Germain M, Nguyen AP, Le Grand JN, Arbour N, Vanderluit JL, Park DS, *et al.* MCL-1 is a stress sensor that regulates autophagy in a developmentally regulated manner. *The EMBO journal* 2011, **30**(2): 395-407.
- 1381 220. Malone CD, Hasan SM, Roome RB, Xiong J, Furlong M, Opferman JT, *et al.* Mcl-1 regulates the survival of adult neural precursor cells. *Molecular and cellular neurosciences* 2012, **49**(4): 439-447.
- Harder JM, Ding Q, Fernandes KA, Cherry JD, Gan L, Libby RT. BCL2L1 (BCL-X) promotes survival of adult and developing retinal ganglion cells. *Molecular and cellular neurosciences* 2012, 51(1-2): 53-59.

1388
1389
222. Nakamura A, Swahari V, Plestant C, Smith I, McCoy E, Smith S, et al. Bcl-xL Is Essential for the Survival and Function of Differentiated Neurons in the Cortex That Control Complex Behaviors. The Journal of neuroscience : the official journal of the Society for Neuroscience 2016, 36(20): 5448-5461.

Savitt JM, Jang SS, Mu W, Dawson VL, Dawson TM. Bcl-x is required for proper development of the mouse substantia nigra. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2005, **25**(29): 6721-6728.

1393

1397

1401

1405

1410

1414

1418

1422

1426

Fogarty LC, Song B, Suppiah Y, Hasan SMM, Martin HC, Hogan SE, *et al.* Bcl-xL dependency coincides with the onset of neurogenesis in the developing mammalian spinal cord. *Molecular and cellular neurosciences* 2016, **77:** 34-46.

1402 225. Fogarty LC, Flemmer RT, Geizer BA, Licursi M, Karunanithy A, Opferman JT, *et al.* Mcl-1 and Bcl-xL are essential for survival of the developing nervous system. *Cell death and differentiation* 2019, **26**(8): 1501-1515.

Veleta KA, Cleveland AH, Babcock BR, He YW, Hwang D, Sokolsky-Papkov M, *et al.*Antiapoptotic Bcl-2 family proteins BCL-xL and MCL-1 integrate neural progenitor survival and proliferation during postnatal cerebellar neurogenesis. *Cell death and differentiation* 2021, **28**(5): 1579-1592.

Weng SY, Yang CY, Li CC, Sun TP, Tung SY, Yen JJ, *et al.* Synergism between p53 and Mcl-1 in protecting from hepatic injury, fibrosis and cancer. *Journal of hepatology* 2011, **54**(4): 685-694.

Hikita H, Takehara T, Shimizu S, Kodama T, Li W, Miyagi T, *et al.* Mcl-1 and Bcl-xL cooperatively maintain integrity of hepatocytes in developing and adult murine liver. *Hepatology* (*Baltimore, Md*) 2009, **50**(4): 1217-1226.

Takehara T, Tatsumi T, Suzuki T, Rucker EB, 3rd, Hennighausen L, Jinushi M, *et al.* Hepatocytespecific disruption of Bcl-xL leads to continuous hepatocyte apoptosis and liver fibrotic responses. *Gastroenterology* 2004, **127**(4): 1189-1197.

1423 230. Vick B, Weber A, Urbanik T, Maass T, Teufel A, Krammer PH, *et al.* Knockout of myeloid cell leukemia-1 induces liver damage and increases apoptosis susceptibility of murine hepatocytes.
 1424 Hepatology (Baltimore, Md) 2009, 49(2): 627-636.

- Boege Y, Malehmir M, Healy ME, Bettermann K, Lorentzen A, Vucur M, *et al.* A Dual Role of Caspase-8 in Triggering and Sensing Proliferation-Associated DNA Damage, a Key Determinant of Liver Cancer Development. *Cancer cell* 2017, 32(3): 342-359.e310.
- Watson EC, Whitehead L, Adams RH, Dewson G, Coultas L. Endothelial cell survival during angiogenesis requires the pro-survival protein MCL1. *Cell death and differentiation* 2016, 23(8): 1371-1379.

1434

1438

1442

1447

1451

1455

1459

1462

- Jain R, Sheridan JM, Policheni A, Heinlein M, Gandolfo LC, Dewson G, *et al.* A critical epithelial survival axis regulated by MCL-1 maintains thymic function in mice. *Blood* 2017, **130**(23): 2504-2515.
- 1439 234. Healy ME, Boege Y, Hodder MC, Böhm F, Malehmir M, Scherr AL, et al. MCL1 Is Required
   1440 for Maintenance of Intestinal Homeostasis and Prevention of Carcinogenesis in Mice.
   1441 Gastroenterology 2020, 159(1): 183-199.
- Walton KD, Wagner KU, Rucker EB, 3rd, Shillingford JM, Miyoshi K, Hennighausen L. Conditional deletion of the bcl-x gene from mouse mammary epithelium results in accelerated apoptosis during involution but does not compromise cell function during lactation. *Mechanisms of development* 2001, **109**(2): 281-293.
- Fu NY, Rios AC, Pal B, Soetanto R, Lun AT, Liu K, *et al.* EGF-mediated induction of Mcl-1 at the switch to lactation is essential for alveolar cell survival. *Nature cell biology* 2015, **17**(4): 365-375.
- Staversky RJ, Vitiello PF, Yee M, Callahan LM, Dean DA, O'Reilly MA. Epithelial ablation of
   Bcl-XL increases sensitivity to oxygen without disrupting lung development. *American journal* of respiratory cell and molecular biology 2010, 43(3): 376-385.
- 238. Brinkmann K, Waring P, Glaser SP, Wimmer V, Cottle DL, Tham MS, *et al.* BCL-XL exerts a protective role against anemia caused by radiation-induced kidney damage. *The EMBO journal* 2020, 39(24): e105561.
- Turnis ME, Kaminska E, Smith KH, Kartchner BJ, Vogel P, Laxton JD, *et al.* Requirement for antiapoptotic MCL-1 during early erythropoiesis. *Blood* 2021, **137**(14): 1945-1958.
- Teh CE, Robbins AK, Henstridge DC, Dewson G, Diepstraten ST, Kelly G, et al. MCL-1 is essential for survival but dispensable for metabolic fitness of FOXP3(+) regulatory T cells. Cell death and differentiation 2020, 27(12): 3374-3385.

- Hikita H, Takehara T, Kodama T, Shimizu S, Hosui A, Miyagi T, *et al.* BH3-only protein bid participates in the Bcl-2 network in healthy liver cells. *Hepatology (Baltimore, Md)* 2009, **50**(6): 1972-1980.
- 1470
  1471 242. Kodama T, Hikita H, Kawaguchi T, Saito Y, Tanaka S, Shigekawa M, et al. The Bcl-2 homology domain 3 (BH3)-only proteins Bim and bid are functionally active and restrained by anti-apoptotic Bcl-2 family proteins in healthy liver. The Journal of biological chemistry 2013, 288(42): 30009-30018.
- 1476 243. Kim DJ, Kataoka K, Sano S, Connolly K, Kiguchi K, DiGiovanni J. Targeted disruption of Bcl 1477 xL in mouse keratinocytes inhibits both UVB- and chemically induced skin carcinogenesis.
   1478 *Molecular carcinogenesis* 2009, 48(10): 873-885.

1479

1483

1486

1489

1492

1495

1499

- Weber A, Boger R, Vick B, Urbanik T, Haybaeck J, Zoller S, *et al.* Hepatocyte-specific deletion of the antiapoptotic protein myeloid cell leukemia-1 triggers proliferation and hepatocarcinogenesis in mice. *Hepatology (Baltimore, Md)* 2010, **51**(4): 1226-1236.
- 1484 245. Coussens LM, Zitvogel L, Palucka AK. Neutralizing tumor-promoting chronic inflammation: a magic bullet? *Science (New York, NY)* 2013, **339**(6117): 286-291.
- 1487 246. Cecconi F, Alvarez-Bolado G, Meyer BI, Roth KA, Gruss P. Apaf1 (CED-4 homolog) regulates programmed cell death in mammalian development. *Cell* 1998, **94**(6): 727-737.
- Honarpour N, Du C, Richardson JA, Hammer RE, Wang X, Herz J. Adult Apaf-1-deficient mice exhibit male infertility. *Developmental biology* 2000, 218(2): 248-258.
- 1493 248. Yoshida H, Kong YY, Yoshida R, Elia AJ, Hakem A, Hakem R, *et al.* Apaf1 is required for mitochondrial pathways of apoptosis and brain development. *Cell* 1998, **94**(6): 739-750.
- 1496 249. Cecconi F, Roth KA, Dolgov O, Munarriz E, Anokhin K, Gruss P, et al. Apaf1-dependent 1497 programmed cell death is required for inner ear morphogenesis and growth. *Development* 1498 (*Cambridge, England*) 2004, **131**(9): 2125-2135.
- 1500 250. Nonomura K, Yamaguchi Y, Hamachi M, Koike M, Uchiyama Y, Nakazato K, *et al.* Local apoptosis modulates early mammalian brain development through the elimination of morphogen-producing cells. *Developmental cell* 2013, **27**(6): 621-634.
- Long AB, Kaiser WJ, Mocarski ES, Caspary T. Apaf1 apoptotic function critically limits Sonic hedgehog signaling during craniofacial development. *Cell death and differentiation* 2013, **20**(11): 1510-1520.

1507 252. 1508 Ohsawa S, Hamada S, Kuida K, Yoshida H, Igaki T, Miura M. Maturation of the olfactory sensory neurons by Apaf-1/caspase-9-mediated caspase activity. Proceedings of the National Academy of 1509 1510 *Sciences of the United States of America* 2010, **107**(30): 13366-13371. 1511 Hakem R, Hakem A, Duncan GS, Henderson JT, Woo M, Soengas MS, et al. Differential 1512 253. requirement for caspase 9 in apoptotic pathways in vivo. *Cell* 1998, **94**(3): 339-352. 1513 1514 254. 1515 Marsden VS, Ekert PG, Van Delft M, Vaux DL, Adams JM, Strasser A. Bcl-2-regulated apoptosis and cytochrome c release can occur independently of both caspase-2 and caspase-9. The Journal 1516 of cell biology 2004, **165**(6): 775-780. 1517 1518 1519 255. Bowen ME, Mulligan AS, Sorayya A, Attardi LD. Puma- and Caspase9-mediated apoptosis is dispensable for p53-driven neural crest-based developmental defects. Cell death and 1520 differentiation 2021, 28(7): 2083-2094. 1521 1522 256. Spellicy CJ, Norris J, Bend R, Bupp C, Mester P, Reynolds T, et al. Key apoptotic genes APAF1 1523 1524 and CASP9 implicated in recurrent folate-resistant neural tube defects. European journal of human genetics: EJHG 2018, **26**(3): 420-427. 1525 1526 257. Zhou X, Zeng W, Li H, Chen H, Wei G, Yang X, et al. Rare mutations in apoptosis related genes 1527 APAF1, CASP9, and CASP3 contribute to human neural tube defects. Cell death & disease 2018, 1528 **9**(2): 43. 1529 1530 258. Li K, Li Y, Shelton JM, Richardson JA, Spencer E, Chen ZJ, et al. Cytochrome c deficiency 1531 causes embryonic lethality and attenuates stress-induced apoptosis. *Cell* 2000, **101**(4): 389-399. 1532 1533 1534 259. Narisawa S, Hecht NB, Goldberg E, Boatright KM, Reed JC, Millán JL. Testis-specific cytochrome c-null mice produce functional sperm but undergo early testicular atrophy. *Molecular* 1535 and cellular biology 2002, **22**(15): 5554-5562. 1536 1537 260. Pinto M, Vempati UD, Diaz F, Peralta S, Moraes CT. Ablation of Cytochrome c in Adult 1538 1539 Forebrain Neurons Impairs Oxidative Phosphorylation Without Detectable Apoptosis. *Molecular* neurobiology 2019, **56**(5): 3722-3735. 1540 1541

Hao Z, Duncan GS, Chang CC, Elia A, Fang M, Wakeham A, et al. Specific ablation of the

apoptotic functions of cytochrome C reveals a differential requirement for cytochrome C and

Apaf-1 in apoptosis. *Cell* 2005, **121**(4): 579-591.

1542

1543

1544

1545

261.

- Woo M, Hakem R, Soengas MS, Duncan GS, Shahinian A, Kägi D, *et al.* Essential contribution of caspase 3/CPP32 to apoptosis and its associated nuclear changes. *Genes & development* 1998, **12**(6): 806-819.
- 1550 263. Kuida K, Zheng TS, Na S, Kuan C, Yang D, Karasuyama H, *et al.* Decreased apoptosis in the brain and premature lethality in CPP32-deficient mice. *Nature* 1996, **384**(6607): 368-372.

1552

1556

1561

1565

1568

1571

1575

1579

- Leonard JR, Klocke BJ, D'Sa C, Flavell RA, Roth KA. Strain-dependent neurodevelopmental abnormalities in caspase-3-deficient mice. *Journal of neuropathology and experimental neurology* 2002, **61**(8): 673-677.
- Roth KA, Kuan C, Haydar TF, D'Sa-Eipper C, Shindler KS, Zheng TS, *et al.* Epistatic and independent functions of caspase-3 and Bcl-X(L) in developmental programmed cell death. *Proceedings of the National Academy of Sciences of the United States of America* 2000, **97**(1): 466-471.
- 1562 266. Matsumoto Y, Yamaguchi Y, Hamachi M, Nonomura K, Muramatsu Y, Yoshida H, *et al.*1563 Apoptosis is involved in maintaining the character of the midbrain and the diencephalon roof
  1564 plate after neural tube closure. *Developmental biology* 2020, **468**(1-2): 101-109.
- Okamoto H, Shiraishi H, Yoshida H. Histological analyses of normally grown, fertile Apaf1-deficient mice. *Cell death and differentiation* 2006, **13**(4): 668-671.
- 1569 268. Lo SC, Scearce-Levie K, Sheng M. Characterization of social behaviors in caspase-3 deficient mice. *Scientific reports* 2016, **6:** 18335.
- Lo SC, Wang Y, Weber M, Larson JL, Scearce-Levie K, Sheng M. Caspase-3 deficiency results in disrupted synaptic homeostasis and impaired attention control. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2015, **35**(5): 2118-2132.
- Takahashi K, Kamiya K, Urase K, Suga M, Takizawa T, Mori H, *et al.* Caspase-3-deficiency induces hyperplasia of supporting cells and degeneration of sensory cells resulting in the hearing loss. *Brain research* 2001, **894**(2): 359-367.
- 1580 271. Morishita H, Makishima T, Kaneko C, Lee YS, Segil N, Takahashi K, *et al.* Deafness due to degeneration of cochlear neurons in caspase-3-deficient mice. *Biochemical and biophysical research communications* 2001, **284**(1): 142-149.
- Parker A, Hardisty-Hughes RE, Wisby L, Joyce S, Brown SD. Melody, an ENU mutation in Caspase 3, alters the catalytic cysteine residue and causes sensorineural hearing loss in mice.

- Mammalian genome : official journal of the International Mammalian Genome Society 2010, **21**(11-12): 565-576.
- 1588
   1589 273. Armstrong PA, Wood SJ, Shimizu N, Kuster K, Perachio A, Makishima T. Preserved otolith organ function in caspase-3-deficient mice with impaired horizontal semicircular canal function.
   1591 Experimental brain research 2015, 233(6): 1825-1835.
- Makishima T, Hochman L, Armstrong P, Rosenberger E, Ridley R, Woo M, *et al.* Inner ear dysfunction in caspase-3 deficient mice. *BMC neuroscience* 2011, **12:** 102.

1595

1598

1603

1607

1611

1615

1619

- Suzuki T, Ichii O, Nakamura T, Horino T, Elewa YHA, Kon Y. Immune-associated renal disease found in caspase 3-deficient mice. *Cell and tissue research* 2020, **379**(2): 323-335.
- Houde C, Banks KG, Coulombe N, Rasper D, Grimm E, Roy S, *et al.* Caspase-7 expanded function and intrinsic expression level underlies strain-specific brain phenotype of caspase-3-null mice. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2004, **24**(44): 9977-9984.
- Lakhani SA, Masud A, Kuida K, Porter GA, Jr., Booth CJ, Mehal WZ, *et al.* Caspases 3 and 7: key mediators of mitochondrial events of apoptosis. *Science (New York, NY)* 2006, **311**(5762): 847-851.
- McComb S, Chan PK, Guinot A, Hartmannsdottir H, Jenni S, Dobay MP, *et al.* Efficient apoptosis requires feedback amplification of upstream apoptotic signals by effector caspase-3 or -7. *Science advances* 2019, **5**(7): eaau9433.
- Lamkanfi M, Moreira LO, Makena P, Spierings DC, Boyd K, Murray PJ, *et al.* Caspase-7 deficiency protects from endotoxin-induced lymphocyte apoptosis and improves survival. *Blood* 2009, **113**(12): 2742-2745.
- Walsh JG, Cullen SP, Sheridan C, Lüthi AU, Gerner C, Martin SJ. Executioner caspase-3 and caspase-7 are functionally distinct proteases. *Proceedings of the National Academy of Sciences of the United States of America* 2008, **105**(35): 12815-12819.
- Yoshida A, Kawata D, Shinotsuka N, Yoshida M, Yamaguchi Y, Miura M. Evidence for the involvement of caspases in establishing proper cerebrospinal fluid hydrodynamics. *Neuroscience research* 2021, **170**: 145-153.

- Demon D, Van Damme P, Vanden Berghe T, Deceuninck A, Van Durme J, Verspurten J, *et al.*Proteome-wide substrate analysis indicates substrate exclusion as a mechanism to generate caspase-7 versus caspase-3 specificity. *Mol Cell Proteomics* 2009, **8**(12): 2700-2714.
- 1628 283. Nozaki K, Maltez VI, Rayamajhi M, Tubbs AL, Mitchell JE, Lacey CA, *et al.* Caspase-7 activates ASM to repair gasdermin and perforin pores. *Nature* 2022, **606**(7916): 960-967.

1630

1633

1637

1641

1645

1648

1651

1655

1659

- Okada H, Suh WK, Jin J, Woo M, Du C, Elia A, *et al.* Generation and characterization of Smac/DIABLO-deficient mice. *Molecular and cellular biology* 2002, **22**(10): 3509-3517.
- 1634 285. Martins LM, Morrison A, Klupsch K, Fedele V, Moisoi N, Teismann P, *et al.* Neuroprotective role of the Reaper-related serine protease HtrA2/Omi revealed by targeted deletion in mice.

  1636 *Molecular and cellular biology* 2004, **24**(22): 9848-9862.
- Hui KK, Kanungo AK, Elia AJ, Henderson JT. Caspase-3 deficiency reveals a physiologic role for Smac/DIABLO in regulating programmed cell death. *Cell death and differentiation* 2011, 18(11): 1780-1790.
- Olayioye MA, Kaufmann H, Pakusch M, Vaux DL, Lindeman GJ, Visvader JE. XIAP-deficiency leads to delayed lobuloalveolar development in the mammary gland. *Cell death and differentiation* 2005, **12**(1): 87-90.
- Harlin H, Reffey SB, Duckett CS, Lindsten T, Thompson CB. Characterization of XIAP-deficient mice. *Molecular and cellular biology* 2001, 21(10): 3604-3608.
- 1649 289. Morrish E, Brumatti G, Silke J. Future Therapeutic Directions for Smac-Mimetics. *Cells* 2020, **9**(2).
- Prakash H, Albrecht M, Becker D, Kuhlmann T, Rudel T. Deficiency of XIAP leads to sensitization for Chlamydophila pneumoniae pulmonary infection and dysregulation of innate immune response in mice. *The Journal of biological chemistry* 2010, **285**(26): 20291-20302.
- Damgaard RB, Nachbur U, Yabal M, Wong WW, Fiil BK, Kastirr M, *et al.* The ubiquitin ligase XIAP recruits LUBAC for NOD2 signaling in inflammation and innate immunity. *Molecular cell* 2012, **46**(6): 746-758.
- Yabal M, Müller N, Adler H, Knies N, Groß CJ, Damgaard RB, *et al.* XIAP restricts TNF- and RIP3-dependent cell death and inflammasome activation. *Cell reports* 2014, **7**(6): 1796-1808.

- Wahida A, Muller M, Hiergeist A, Popper B, Steiger K, Branca C, *et al.* XIAP restrains TNF-driven intestinal inflammation and dysbiosis by promoting innate immune responses of Paneth and dendritic cells. *Sci Immunol* 2021, **6**(65): eabf7235.
- Hsieh WC, Chuang YT, Chiang IH, Hsu SC, Miaw SC, Lai MZ. Inability to resolve specific infection generates innate immunodeficiency syndrome in Xiap-/- mice. *Blood* 2014, **124**(18): 2847-2857.

1670

1674

1678

1682

1686

1690

1694

- Salzer U, Hagena T, Webster DB, Grimbacher B. Sequence analysis of BIRC4/XIAP in male patients with common variable immunodeficiency. *International archives of allergy and immunology* 2008, **147**(2): 147-151.
- Yang X, Kanegane H, Nishida N, Imamura T, Hamamoto K, Miyashita R, et al. Clinical and genetic characteristics of XIAP deficiency in Japan. *Journal of clinical immunology* 2012, 32(3): 411-420.
- Damgaard RB, Fiil BK, Speckmann C, Yabal M, zur Stadt U, Bekker-Jensen S, *et al.* Disease-causing mutations in the XIAP BIR2 domain impair NOD2-dependent immune signalling. *EMBO molecular medicine* 2013, **5**(8): 1278-1295.
- 298. Cardona M, López JA, Serafín A, Rongvaux A, Inserte J, García-Dorado D, *et al.* Executioner
   Caspase-3 and 7 Deficiency Reduces Myocyte Number in the Developing Mouse Heart. *PloS one* 2015, 10(6): e0131411.
- Marsden VS, O'Connor L, O'Reilly LA, Silke J, Metcalf D, Ekert PG, *et al.* Apoptosis initiated by Bcl-2-regulated caspase activation independently of the cytochrome c/Apaf-1/caspase-9 apoptosome. *Nature* 2002, **419**(6907): 634-637.
- 1691 300. White MJ, Schoenwaelder SM, Josefsson EC, Jarman KE, Henley KJ, James C, *et al.* Caspase-9 mediates the apoptotic death of megakaryocytes and platelets, but is dispensable for their generation and function. *Blood* 2012, **119**(18): 4283-4290.
- van Delft MF, Smith DP, Lahoud MH, Huang DC, Adams JM. Apoptosis and non-inflammatory phagocytosis can be induced by mitochondrial damage without caspases. *Cell death and differentiation* 2010, **17**(5): 821-832.
- Ghazavi F, Huysentruyt J, De Coninck J, Kourula S, Martens S, Hassannia B, *et al.* Executioner caspases 3 and 7 are dispensable for intestinal epithelium turnover and homeostasis at steady state. *Proceedings of the National Academy of Sciences of the United States of America* 2022, **119**(6).

1703 303. 1704 Woo M, Hakem R, Furlonger C, Hakem A, Duncan GS, Sasaki T, et al. Caspase-3 regulates cell cycle in B cells: a consequence of substrate specificity. *Nature immunology* 2003, **4**(10): 1016-1705 1706 1022. 1707 Miura M, Chen XD, Allen MR, Bi Y, Gronthos S, Seo BM, et al. A crucial role of caspase-3 in 1708 304. osteogenic differentiation of bone marrow stromal stem cells. The Journal of clinical 1709 investigation 2004, **114**(12): 1704-1713. 1710 1711 Tong H, Miyake Y, Mi-Ichi F, Iwakura Y, Hara H, Yoshida H. Apaf1 plays a negative regulatory 1712 305. role in T cell responses by suppressing activation of antigen-stimulated T cells. PloS one 2018, 1713 **13**(3): e0195119. 1714 1715 Hara H, Takeda A, Takeuchi M, Wakeham AC, Itié A, Sasaki M, et al. The apoptotic protease-1716 306. activating factor 1-mediated pathway of apoptosis is dispensable for negative selection of 1717 1718 thymocytes. *Journal of immunology (Baltimore, Md: 1950)* 2002, **168**(5): 2288-2295. 1719 1720 307. Nagasaka A, Kawane K, Yoshida H, Nagata S. Apaf-1-independent programmed cell death in mouse development. *Cell death and differentiation* 2010, **17**(6): 931-941. 1721 1722 1723 308. Doerfler P, Forbush KA, Perlmutter RM. Caspase enzyme activity is not essential for apoptosis during thymocyte development. Journal of immunology (Baltimore, Md: 1950) 2000, 164(8): 1724 1725 4071-4079. 1726 1727 309. Izquierdo M, Grandien A, Criado LM, Robles S, Leonardo E, Albar JP, et al. Blocked negative 1728 selection of developing T cells in mice expressing the baculovirus p35 caspase inhibitor. The EMBO journal 1999, 18(1): 156-166. 1729 1730 1731 310. Lu EP, McLellan M, Ding L, Fulton R, Mardis ER, Wilson RK, et al. Caspase-9 is required for normal hematopoietic development and protection from alkylator-induced DNA damage in mice. 1732 Blood 2014, 124(26): 3887-3895. 1733 1734 1735 311. White MJ, McArthur K, Metcalf D, Lane RM, Cambier JC, Herold MJ, et al. Apoptotic caspases suppress mtDNA-induced STING-mediated type I IFN production. Cell 2014, 159(7): 1549-1736 1737 1562. 1738 1739 312. Oppenheim RW, Blomgren K, Ethell DW, Koike M, Komatsu M, Prevette D, et al. Developing 1740 postmitotic mammalian neurons in vivo lacking Apaf-1 undergo programmed cell death by a

caspase-independent, nonapoptotic pathway involving autophagy. The Journal of neuroscience:

the official journal of the Society for Neuroscience 2008, **28**(6): 1490-1497.

1741

1743
 1744 313. Oppenheim RW, Flavell RA, Vinsant S, Prevette D, Kuan CY, Rakic P. Programmed cell death of developing mammalian neurons after genetic deletion of caspases. *The Journal of neuroscience* : the official journal of the Society for Neuroscience 2001, 21(13): 4752-4760.

1748 314. Yaginuma H, Shiraiwa N, Shimada T, Nishiyama K, Hong J, Wang S, *et al.* Caspase activity is involved in, but is dispensable for, early motoneuron death in the chick embryo cervical spinal cord. *Molecular and cellular neurosciences* 2001, **18**(2): 168-182.

1747

1751

1755

1758

1761

1764

1767

1770

1774

1778

1781

Honarpour N, Tabuchi K, Stark JM, Hammer RE, Südhof TC, Parada LF, *et al.* Embryonic neuronal death due to neurotrophin and neurotransmitter deprivation occurs independent of Apaf-1. *Neuroscience* 2001, **106**(2): 263-274.

1756 316. Aggarwal BB, Gupta SC, Kim JH. Historical perspectives on tumor necrosis factor and its superfamily: 25 years later, a golden journey. *Blood* 2012, **119**(3): 651-665.

1759 317. Strasser A, Jost PJ, Nagata S. The many roles of FAS receptor signaling in the immune system. *Immunity* 2009, **30**(2): 180-192.

1762 318. Wajant H. The Fas signaling pathway: more than a paradigm. *Science (New York, NY)* 2002, **296**(5573): 1635-1636.

Wallach D. The Tumor Necrosis Factor Family: Family Conventions and Private Idiosyncrasies.
 Cold Spring Harbor perspectives in biology 2018, 10(10).

von Karstedt S, Montinaro A, Walczak H. Exploring the TRAILs less travelled: TRAIL in cancer biology and therapy. *Nature reviews Cancer* 2017, **17**(6): 352-366.

1771 321. Kischkel FC, Hellbardt S, Behrmann I, Germer M, Pawlita M, Krammer PH, *et al.* Cytotoxicity-1772 dependent APO-1 (Fas/CD95)-associated proteins form a death-inducing signaling complex 1773 (DISC) with the receptor. *The EMBO journal* 1995, **14**(22): 5579-5588.

Boldin MP, Goncharov TM, Goltsev YV, Wallach D. Involvement of MACH, a novel MORT1/FADD-interacting protease, in Fas/APO-1- and TNF receptor-induced cell death. *Cell* 1996, **85**(6): 803-815.

Dickens LS, Powley IR, Hughes MA, MacFarlane M. The 'complexities' of life and death: death receptor signalling platforms. *Experimental cell research* 2012, **318**(11): 1269-1277.

Muzio M, Chinnaiyan AM, Kischkel FC, O'Rourke K, Shevchenko A, Ni J, *et al.* FLICE, a novel FADD-homologous ICE/CED-3-like protease, is recruited to the CD95 (Fas/APO-1) death-inducing signaling complex. *Cell* 1996, **85**(6): 817-827.

1785

Boldin MP, Varfolomeev EE, Pancer Z, Mett IL, Camonis JH, Wallach D. A novel protein that interacts with the death domain of Fas/APO1 contains a sequence motif related to the death domain. *The Journal of biological chemistry* 1995, **270**(14): 7795-7798.

1789

1790 326. Chinnaiyan AM, O'Rourke K, Tewari M, Dixit VM. FADD, a novel death domain-containing protein, interacts with the death domain of Fas and initiates apoptosis. *Cell* 1995, **81**(4): 505-512.

1792

1793 327. Kischkel FC, Lawrence DA, Chuntharapai A, Schow P, Kim KJ, Ashkenazi A. Apo2L/TRAIL-1794 dependent recruitment of endogenous FADD and caspase-8 to death receptors 4 and 5. *Immunity* 1795 2000, **12**(6): 611-620.

1796

328. Scott FL, Stec B, Pop C, Dobaczewska MK, Lee JJ, Monosov E, *et al.* The Fas-FADD death domain complex structure unravels signalling by receptor clustering. *Nature* 2009, **457**(7232): 1019-1022.

1800

1801 329. Chan FK, Chun HJ, Zheng L, Siegel RM, Bui KL, Lenardo MJ. A domain in TNF receptors that mediates ligand-independent receptor assembly and signaling. *Science (New York, NY)* 2000, **288**(5475): 2351-2354.

1804

Fu Q, Fu TM, Cruz AC, Sengupta P, Thomas SK, Wang S, et al. Structural Basis and Functional Role of Intramembrane Trimerization of the Fas/CD95 Death Receptor. Molecular cell 2016, 61(4): 602-613.

1808

1809 331. Brenner D, Blaser H, Mak TW. Regulation of tumour necrosis factor signalling: live or let die. *Nature reviews Immunology* 2015, **15**(6): 362-374.

1811

1812 332. Galluzzi L, Kepp O, Chan FK, Kroemer G. Necroptosis: Mechanisms and Relevance to Disease.
 1813 Annual review of pathology 2017, 12: 103-130.

1814

1815 333. Tummers B, Green DR. Caspase-8: regulating life and death. *Immunological reviews* 2017, 1816 277(1): 76-89.

1817

1818 334. Barnhart BC, Alappat EC, Peter ME. The CD95 type I/type II model. *Seminars in immunology* 2003, **15**(3): 185-193.

- 1821 335. Strasser A, Harris AW, Huang DC, Krammer PH, Cory S. Bcl-2 and Fas/APO-1 regulate distinct pathways to lymphocyte apoptosis. *The EMBO journal* 1995, **14**(24): 6136-6147.
- 1823
  1824 336. Li H, Zhu H, Xu CJ, Yuan J. Cleavage of BID by caspase 8 mediates the mitochondrial damage
  1825 in the Fas pathway of apoptosis. *Cell* 1998, **94**(4): 491-501.
- 1826
  1827 337. Luo X, Budihardjo I, Zou H, Slaughter C, Wang X. Bid, a Bcl2 interacting protein, mediates cytochrome c release from mitochondria in response to activation of cell surface death receptors.
  1829 Cell 1998, 94(4): 481-490.
- 338. Gross A, Yin XM, Wang K, Wei MC, Jockel J, Milliman C, *et al.* Caspase cleaved BID targets mitochondria and is required for cytochrome c release, while BCL-XL prevents this release but not tumor necrosis factor-R1/Fas death. *The Journal of biological chemistry* 1999, **274**(2): 1156-1163.

1835

1839

1842

1845

1848

1852

1856

- 1836 339. Huang K, Zhang J, O'Neill KL, Gurumurthy CB, Quadros RM, Tu Y, et al. Cleavage by Caspase
   1837 8 and Mitochondrial Membrane Association Activate the BH3-only Protein Bid during TRAIL-induced Apoptosis. The Journal of biological chemistry 2016, 291(22): 11843-11851.
- 1840 340. Lalaoui N, Boyden SE, Oda H, Wood GM, Stone DL, Chau D, *et al.* Mutations that prevent caspase cleavage of RIPK1 cause autoinflammatory disease. *Nature* 2020, **577**(7788): 103-108.
- 1843 341. Hayden MS, Ghosh S. Regulation of NF-κB by TNF family cytokines. *Seminars in immunology* 2014, **26**(3): 253-266.
- Bertheloot D, Latz E, Franklin BS. Necroptosis, pyroptosis and apoptosis: an intricate game of cell death. *Cellular & molecular immunology* 2021, **18**(5): 1106-1121.
- Hartwig T, Montinaro A, von Karstedt S, Sevko A, Surinova S, Chakravarthy A, et al. The
   TRAIL-Induced Cancer Secretome Promotes a Tumor-Supportive Immune Microenvironment
   via CCR2. Molecular cell 2017, 65(4): 730-742.e735.
- 344. Henry CM, Martin SJ. Caspase-8 Acts in a Non-enzymatic Role as a Scaffold for Assembly of a
   Pro-inflammatory "FADDosome" Complex upon TRAIL Stimulation. *Molecular cell* 2017,
   65(4): 715-729.e715.
- 345. Gibert B, Mehlen P. Dependence Receptors and Cancer: Addiction to Trophic Ligands. *Cancer research* 2015, **75**(24): 5171-5175.

- 1860 346. Mehlen P, Bredesen DE. Dependence receptors: from basic research to drug development. 1861 Science signaling 2011, **4**(157): mr2.
- 1862
   1863 347. Brisset M, Grandin M, Bernet A, Mehlen P, Hollande F. Dependence receptors: new targets for cancer therapy. *EMBO molecular medicine* 2021, 13(11): e14495.
- 1866 348. Negulescu AM, Mehlen P. Dependence receptors the dark side awakens. *The FEBS journal* 2018, **285**(21): 3909-3924.

1868

1871

1874

1878

1881

1886

1890

1894

- 349. Dillon CP, Oberst A, Weinlich R, Janke LJ, Kang TB, Ben-Moshe T, et al. Survival function of the FADD-CASPASE-8-cFLIP(L) complex. Cell reports 2012, 1(5): 401-407.
- 1872 350. Kaiser WJ, Upton JW, Long AB, Livingston-Rosanoff D, Daley-Bauer LP, Hakem R, *et al.* RIP3 mediates the embryonic lethality of caspase-8-deficient mice. *Nature* 2011, **471**(7338): 368-372.
- 1875 351. Alvarez-Diaz S, Dillon CP, Lalaoui N, Tanzer MC, Rodriguez DA, Lin A, *et al.* The Pseudokinase MLKL and the Kinase RIPK3 Have Distinct Roles in Autoimmune Disease Caused by Loss of Death-Receptor-Induced Apoptosis. *Immunity* 2016, **45**(3): 513-526.
- 1879 352. Dillon CP, Weinlich R, Rodriguez DA, Cripps JG, Quarato G, Gurung P, *et al.* RIPK1 blocks early postnatal lethality mediated by caspase-8 and RIPK3. *Cell* 2014, **157**(5): 1189-1202.
- 1882 353. Kaiser WJ, Daley-Bauer LP, Thapa RJ, Mandal P, Berger SB, Huang C, et al. RIP1 suppresses
   1883 innate immune necrotic as well as apoptotic cell death during mammalian parturition.
   1884 Proceedings of the National Academy of Sciences of the United States of America 2014, 111(21):
   1885 7753-7758.
- 1887 354. Rickard JA, O'Donnell JA, Evans JM, Lalaoui N, Poh AR, Rogers T, *et al.* RIPK1 regulates RIPK3-MLKL-driven systemic inflammation and emergency hematopoiesis. *Cell* 2014, **157**(5): 1175-1188.
- Takahashi T, Tanaka M, Brannan CI, Jenkins NA, Copeland NG, Suda T, *et al.* Generalized lymphoproliferative disease in mice, caused by a point mutation in the Fas ligand. *Cell* 1994, **76**(6): 969-976.
- 1895 356. Watanabe-Fukunaga R, Brannan CI, Copeland NG, Jenkins NA, Nagata S. Lymphoproliferation disorder in mice explained by defects in Fas antigen that mediates apoptosis. *Nature* 1992, **356**(6367): 314-317.

- 1899 357. Lynch DH, Watson ML, Alderson MR, Baum PR, Miller RE, Tough T, *et al.* The mouse Fas-1900 ligand gene is mutated in gld mice and is part of a TNF family gene cluster. *Immunity* 1994, **1**(2): 131-136.
- 1903 358. Roths JB, Murphy ED, Eicher EM. A new mutation, gld, that produces lymphoproliferation and autoimmunity in C3H/HeJ mice. *The Journal of experimental medicine* 1984, **159**(1): 1-20.

1905

1909

1913

1917

1921

1925

1928

1932

- 1906 359. Matsuzawa A, Moriyama T, Kaneko T, Tanaka M, Kimura M, Ikeda H, *et al.* A new allele of the lpr locus, lprcg, that complements the gld gene in induction of lymphadenopathy in the mouse.

  1908 *The Journal of experimental medicine* 1990, **171**(2): 519-531.
- 1910 360. Adachi M, Suematsu S, Kondo T, Ogasawara J, Tanaka T, Yoshida N, *et al.* Targeted mutation in the Fas gene causes hyperplasia in peripheral lymphoid organs and liver. *Nature genetics* 1995, 1912 11(3): 294-300.
- 1914 361. Anstee NS, Vandenberg CJ, Campbell KJ, Hughes PD, O'Reilly LA, Cory S. Overexpression of Mcl-1 exacerbates lymphocyte accumulation and autoimmune kidney disease in lpr mice. *Cell death and differentiation* 2017, **24**(3): 397-408.
- Hughes PD, Belz GT, Fortner KA, Budd RC, Strasser A, Bouillet P. Apoptosis regulators Fas and Bim cooperate in shutdown of chronic immune responses and prevention of autoimmunity. *Immunity* 2008, **28**(2): 197-205.
- 1922 363. Karray S, Kress C, Cuvellier S, Hue-Beauvais C, Damotte D, Babinet C, *et al.* Complete loss of Fas ligand gene causes massive lymphoproliferation and early death, indicating a residual activity of gld allele. *Journal of immunology (Baltimore, Md : 1950)* 2004, **172**(4): 2118-2125.
- Davies MH, Eubanks JP, Powers MR. Increased retinal neovascularization in Fas ligand-deficient mice. *Investigative ophthalmology & visual science* 2003, **44**(7): 3202-3210.
- 1929 365. Schumann DM, Maedler K, Franklin I, Konrad D, Størling J, Böni-Schnetzler M, *et al.* The Fas pathway is involved in pancreatic beta cell secretory function. *Proceedings of the National* 1931 *Academy of Sciences of the United States of America* 2007, **104**(8): 2861-2866.
- 1933 366. Trumpi K, Steller EJ, de Leng WW, Raats DA, Nijman IJ, Morsink FH, *et al.* Mice lacking functional CD95-ligand display reduced proliferation of the intestinal epithelium without gross homeostatic alterations. *Medical molecular morphology* 2016, **49**(2): 110-118.

- 1937 367. Hao Z, Hampel B, Yagita H, Rajewsky K. T cell-specific ablation of Fas leads to Fas ligand-1938 mediated lymphocyte depletion and inflammatory pulmonary fibrosis. *The Journal of* 1939 *experimental medicine* 2004, **199**(10): 1355-1365.
- 1941 368. Fukuyama H, Adachi M, Suematsu S, Miwa K, Suda T, Yoshida N, *et al.* Transgenic expression of Fas in T cells blocks lymphoproliferation but not autoimmune disease in MRL-lpr mice.

  1943 *Journal of immunology (Baltimore, Md : 1950)* 1998, **160**(8): 3805-3811.

1944

1948

1952

1956

1961

1965

1968

- 1945 369. Komano H, Ikegami Y, Yokoyama M, Suzuki R, Yonehara S, Yamasaki Y, *et al.* Severe impairment of B cell function in lpr/lpr mice expressing transgenic Fas selectively on B cells.

  1947 *International immunology* 1999, **11**(7): 1035-1042.
- 1949 370. Stranges PB, Watson J, Cooper CJ, Choisy-Rossi CM, Stonebraker AC, Beighton RA, *et al.*1950 Elimination of antigen-presenting cells and autoreactive T cells by Fas contributes to prevention
  1951 of autoimmunity. *Immunity* 2007, **26**(5): 629-641.
- 1953 371. Rathmell JC, Cooke MP, Ho WY, Grein J, Townsend SE, Davis MM, *et al.* CD95 (Fas)1954 dependent elimination of self-reactive B cells upon interaction with CD4+ T cells. *Nature* 1995,
  1955 376(6536): 181-184.
- Mohamood AS, Guler ML, Xiao Z, Zheng D, Hess A, Wang Y, *et al.* Protection from autoimmune diabetes and T-cell lymphoproliferation induced by FasL mutation are differentially regulated and can be uncoupled pharmacologically. *The American journal of pathology* 2007, **171**(1): 97-106.
- Thang JQ, Okumura C, McCarty T, Shin MS, Mukhopadhyay P, Hori M, *et al.* Evidence for selective transformation of autoreactive immature plasma cells in mice deficient in Fasl. *The Journal of experimental medicine* 2004, **200**(11): 1467-1478.
- Peng SL, Robert ME, Hayday AC, Craft J. A tumor-suppressor function for Fas (CD95) revealed in T cell-deficient mice. *The Journal of experimental medicine* 1996, **184**(3): 1149-1154.
- Davidson WF, Giese T, Fredrickson TN. Spontaneous development of plasmacytoid tumors in mice with defective Fas-Fas ligand interactions. *The Journal of experimental medicine* 1998, 187(11): 1825-1838.
- 1973 376. Sedger LM, Glaccum MB, Schuh JC, Kanaly ST, Williamson E, Kayagaki N, *et al.*1974 Characterization of the in vivo function of TNF-alpha-related apoptosis-inducing ligand,
  1975 TRAIL/Apo2L, using TRAIL/Apo2L gene-deficient mice. *European journal of immunology*1976 2002, **32**(8): 2246-2254.

1977
1978
377. Diehl GE, Yue HH, Hsieh K, Kuang AA, Ho M, Morici LA, *et al.* TRAIL-R as a negative regulator of innate immune cell responses. *Immunity* 2004, **21**(6): 877-889.

1981 378. Finnberg N, Gruber JJ, Fei P, Rudolph D, Bric A, Kim SH, *et al.* DR5 knockout mice are compromised in radiation-induced apoptosis. *Molecular and cellular biology* 2005, **25**(5): 2000-2013.

1980

1984

1988

1992

1996

2000

2004

2008

2013

1985 379. Lamhamedi-Cherradi SE, Zheng SJ, Maguschak KA, Peschon J, Chen YH. Defective thymocyte apoptosis and accelerated autoimmune diseases in TRAIL-/- mice. *Nature immunology* 2003, 4(3): 255-260.

1989 380. Lehnert C, Weiswange M, Jeremias I, Bayer C, Grunert M, Debatin KM, *et al.* TRAIL-receptor costimulation inhibits proximal TCR signaling and suppresses human T cell activation and proliferation. *Journal of immunology (Baltimore, Md : 1950)* 2014, **193**(8): 4021-4031.

1993 381. McGrath EE, Marriott HM, Lawrie A, Francis SE, Sabroe I, Renshaw SA, *et al.* TNF-related apoptosis-inducing ligand (TRAIL) regulates inflammatory neutrophil apoptosis and enhances resolution of inflammation. *Journal of leukocyte biology* 2011, **90**(5): 855-865.

Sacks JA, Bevan MJ. TRAIL deficiency does not rescue impaired CD8+ T cell memory generated in the absence of CD4+ T cell help. *Journal of immunology (Baltimore, Md: 1950)* 2008, **180**(7): 4570-4576.

2001 383. Cretney E, Uldrich AP, Berzins SP, Strasser A, Godfrey DI, Smyth MJ. Normal thymocyte negative selection in TRAIL-deficient mice. *The Journal of experimental medicine* 2003, **198**(3): 491-496.

Marino MW, Dunn A, Grail D, Inglese M, Noguchi Y, Richards E, et al. Characterization of tumor necrosis factor-deficient mice. Proceedings of the National Academy of Sciences of the United States of America 1997, **94**(15): 8093-8098.

Pasparakis M, Alexopoulou L, Episkopou V, Kollias G. Immune and inflammatory responses in TNF alpha-deficient mice: a critical requirement for TNF alpha in the formation of primary B cell follicles, follicular dendritic cell networks and germinal centers, and in the maturation of the humoral immune response. *The Journal of experimental medicine* 1996, **184**(4): 1397-1411.

2014 386. Pasparakis M, Alexopoulou L, Grell M, Pfizenmaier K, Bluethmann H, Kollias G. Peyer's patch organogenesis is intact yet formation of B lymphocyte follicles is defective in peripheral lymphoid organs of mice deficient for tumor necrosis factor and its 55-kDa receptor. *Proceedings of the National Academy of Sciences of the United States of America* 1997, **94**(12): 6319-6323.

2018
 2019 387. Körner H, Cook M, Riminton DS, Lemckert FA, Hoek RM, Ledermann B, *et al.* Distinct roles for lymphotoxin-alpha and tumor necrosis factor in organogenesis and spatial organization of lymphoid tissue. *European journal of immunology* 1997, 27(10): 2600-2609.

Oishi N, Chen J, Zheng HW, Hill K, Schacht J, Sha SH. Tumor necrosis factor-alpha-mutant mice exhibit high frequency hearing loss. *Journal of the Association for Research in Otolaryngology: JARO* 2013, **14**(6): 801-811.

2022

2026

2030

2034

2038

2042

2047

2051

- Pfeffer K, Matsuyama T, Kündig TM, Wakeham A, Kishihara K, Shahinian A, *et al.* Mice deficient for the 55 kd tumor necrosis factor receptor are resistant to endotoxic shock, yet succumb to L. monocytogenes infection. *Cell* 1993, **73**(3): 457-467.
- 2031 390. Rothe J, Lesslauer W, Lötscher H, Lang Y, Koebel P, Köntgen F, *et al.* Mice lacking the tumour necrosis factor receptor 1 are resistant to TNF-mediated toxicity but highly susceptible to infection by Listeria monocytogenes. *Nature* 1993, **364**(6440): 798-802.
- 2035 391. Kubota T, McTiernan CF, Frye CS, Demetris AJ, Feldman AM. Cardiac-specific overexpression of tumor necrosis factor-alpha causes lethal myocarditis in transgenic mice. *Journal of cardiac failure* 1997, **3**(2): 117-124.
- 2039 392. Kubota T, McTiernan CF, Frye CS, Slawson SE, Lemster BH, Koretsky AP, *et al.* Dilated cardiomyopathy in transgenic mice with cardiac-specific overexpression of tumor necrosis factor-alpha. *Circulation research* 1997, **81**(4): 627-635.
- 2043 393. Lacey D, Hickey P, Arhatari BD, O'Reilly LA, Rohrbeck L, Kiriazis H, *et al.* Spontaneous retrotransposon insertion into TNF 3'UTR causes heart valve disease and chronic polyarthritis.

  2045 *Proceedings of the National Academy of Sciences of the United States of America* 2015, **112**(31): 9698-9703.
- 2048 394. Sedger LM, Katewa A, Pettersen AK, Osvath SR, Farrell GC, Stewart GJ, *et al.* Extreme lymphoproliferative disease and fatal autoimmune thrombocytopenia in FasL and TRAIL double-deficient mice. *Blood* 2010, **115**(16): 3258-3268.
- 2052 395. Körner H, Cretney E, Wilhelm P, Kelly JM, Röllinghoff M, Sedgwick JD, *et al.* Tumor necrosis factor sustains the generalized lymphoproliferative disorder (gld) phenotype. *The Journal of experimental medicine* 2000, **191**(1): 89-96.
- 2056 396. Meynier S, Rieux-Laucat F. FAS and RAS related Apoptosis defects: From autoimmunity to leukemia. *Immunological reviews* 2019, **287**(1): 50-61.

2058
 2059 397. Rieux-Laucat F, Magérus-Chatinet A, Neven B. The Autoimmune Lymphoproliferative
 2060 Syndrome with Defective FAS or FAS-Ligand Functions. *Journal of clinical immunology* 2018,
 2061 38(5): 558-568.

2063 398. Del-Rey M, Ruiz-Contreras J, Bosque A, Calleja S, Gomez-Rial J, Roldan E, *et al.* A homozygous Fas ligand gene mutation in a patient causes a new type of autoimmune lymphoproliferative syndrome. *Blood* 2006, **108**(4): 1306-1312.

2062

2066

2070

2074

2079

2083

2087

2091

2095

Fisher GH, Rosenberg FJ, Straus SE, Dale JK, Middleton LA, Lin AY, *et al.* Dominant interfering Fas gene mutations impair apoptosis in a human autoimmune lymphoproliferative syndrome. *Cell* 1995, **81**(6): 935-946.

- 2071 400. Rieux-Laucat F, Le Deist F, Hivroz C, Roberts IA, Debatin KM, Fischer A, *et al.* Mutations in Fas associated with human lymphoproliferative syndrome and autoimmunity. *Science (New York, NY)* 1995, **268**(5215): 1347-1349.
- Magerus-Chatinet A, Stolzenberg MC, Loffredo MS, Neven B, Schaffner C, Ducrot N, *et al.*FAS-L, IL-10, and double-negative CD4- CD8- TCR alpha/beta+ T cells are reliable markers of autoimmune lymphoproliferative syndrome (ALPS) associated with FAS loss of function. *Blood* 2009, **113**(13): 3027-3030.
- 2080 402. Rensing-Ehl A, Völkl S, Speckmann C, Lorenz MR, Ritter J, Janda A, *et al.* Abnormally differentiated CD4+ or CD8+ T cells with phenotypic and genetic features of double negative T cells in human Fas deficiency. *Blood* 2014, **124**(6): 851-860.
- 2084 403. Price S, Shaw PA, Seitz A, Joshi G, Davis J, Niemela JE, *et al.* Natural history of autoimmune lymphoproliferative syndrome associated with FAS gene mutations. *Blood* 2014, **123**(13): 1989-1999.
- 2088 404. Bi LL, Pan G, Atkinson TP, Zheng L, Dale JK, Makris C, *et al.* Dominant inhibition of Fas ligand-2089 mediated apoptosis due to a heterozygous mutation associated with autoimmune 2090 lymphoproliferative syndrome (ALPS) Type Ib. *BMC medical genetics* 2007, **8:** 41.
- Venkataraman G, McClain KL, Pittaluga S, Rao VK, Jaffe ES. Development of disseminated histocytic sarcoma in a patient with autoimmune lymphoproliferative syndrome and associated Rosai-Dorfman disease. *The American journal of surgical pathology* 2010, **34**(4): 589-594.
- Haas SL, Lohse P, Schmitt WH, Hildenbrand R, Karaorman M, Singer MV, *et al.* Severe TNF receptor-associated periodic syndrome due to 2 TNFRSF1A mutations including a new F60V substitution. *Gastroenterology* 2006, **130**(1): 172-178.

Tsuji S, Matsuzaki H, Iseki M, Nagasu A, Hirano H, Ishihara K, *et al.* Functional analysis of a novel G87V TNFRSF1A mutation in patients with TNF receptor-associated periodic syndrome.
 Clinical and experimental immunology 2019, 198(3): 416-429.

2104 408. McDermott MF, Aksentijevich I, Galon J, McDermott EM, Ogunkolade BW, Centola M, *et al.*2105 Germline mutations in the extracellular domains of the 55 kDa TNF receptor, TNFR1, define a
2106 family of dominantly inherited autoinflammatory syndromes. *Cell* 1999, **97**(1): 133-144.

2103

2107

2111

2115

2118

2122

2126

2130

2134

2138

Yeh WC, de la Pompa JL, McCurrach ME, Shu HB, Elia AJ, Shahinian A, et al. FADD: essential for embryo development and signaling from some, but not all, inducers of apoptosis. Science (New York, NY) 1998, 279(5358): 1954-1958.

- Imtiyaz HZ, Zhou X, Zhang H, Chen D, Hu T, Zhang J. The death domain of FADD is essential for embryogenesis, lymphocyte development, and proliferation. *The Journal of biological chemistry* 2009, **284**(15): 9917-9926.
- Zhang H, Zhou X, McQuade T, Li J, Chan FK, Zhang J. Functional complementation between
   FADD and RIP1 in embryos and lymphocytes. *Nature* 2011, 471(7338): 373-376.
- Varfolomeev EE, Schuchmann M, Luria V, Chiannilkulchai N, Beckmann JS, Mett IL, *et al.*Targeted disruption of the mouse Caspase 8 gene ablates cell death induction by the TNF receptors, Fas/Apo1, and DR3 and is lethal prenatally. *Immunity* 1998, **9**(2): 267-276.
- Sakamaki K, Inoue T, Asano M, Sudo K, Kazama H, Sakagami J, *et al.* Ex vivo whole-embryo culture of caspase-8-deficient embryos normalize their aberrant phenotypes in the developing neural tube and heart. *Cell death and differentiation* 2002, **9**(11): 1196-1206.
- Yeh WC, Itie A, Elia AJ, Ng M, Shu HB, Wakeham A, *et al.* Requirement for Casper (c-FLIP) in regulation of death receptor-induced apoptosis and embryonic development. *Immunity* 2000, **12**(6): 633-642.
- 2131 415. Chen NJ, Chio, II, Lin WJ, Duncan G, Chau H, Katz D, et al. Beyond tumor necrosis factor receptor: TRADD signaling in toll-like receptors. *Proceedings of the National Academy of Sciences of the United States of America* 2008, **105**(34): 12429-12434.
- 2135 416. Ermolaeva MA, Michallet MC, Papadopoulou N, Utermöhlen O, Kranidioti K, Kollias G, *et al.*2136 Function of TRADD in tumor necrosis factor receptor 1 signaling and in TRIF-dependent
  2137 inflammatory responses. *Nature immunology* 2008, **9**(9): 1037-1046.

- 2139 417. Pobezinskaya YL, Kim YS, Choksi S, Morgan MJ, Li T, Liu C, *et al.* The function of TRADD in signaling through tumor necrosis factor receptor 1 and TRIF-dependent Toll-like receptors.
- 2141 *Nature immunology* 2008, **9**(9): 1047-1054.

418. Kelliher MA, Grimm S, Ishida Y, Kuo F, Stanger BZ, Leder P. The death domain kinase RIP mediates the TNF-induced NF-kappaB signal. *Immunity* 1998, **8**(3): 297-303.

2145

2146 419. Roderick JE, Hermance N, Zelic M, Simmons MJ, Polykratis A, Pasparakis M, *et al.*2147 Hematopoietic RIPK1 deficiency results in bone marrow failure caused by apoptosis and RIPK32148 mediated necroptosis. *Proceedings of the National Academy of Sciences of the United States of*2149 *America* 2014, **111**(40): 14436-14441.

2150

Moulin M, Anderton H, Voss AK, Thomas T, Wong WW, Bankovacki A, et al. IAPs limit activation of RIP kinases by TNF receptor 1 during development. The EMBO journal 2012, 31(7): 1679-1691.

2154

Heard KN, Bertrand MJ, Barker PA. cIAP2 supports viability of mice lacking cIAP1 and XIAP. The EMBO journal 2015, **34**(19): 2393-2395.

2157

O'Donnell MA, Perez-Jimenez E, Oberst A, Ng A, Massoumi R, Xavier R, *et al.* Caspase 8 inhibits programmed necrosis by processing CYLD. *Nature cell biology* 2011, **13**(12): 1437-1442.

2161

2162 423. Oberst A, Dillon CP, Weinlich R, McCormick LL, Fitzgerald P, Pop C, *et al.* Catalytic activity of the caspase-8-FLIP(L) complex inhibits RIPK3-dependent necrosis. *Nature* 2011, **471**(7338): 363-367.

2165

Zhao Q, Yu X, Zhang H, Liu Y, Zhang X, Wu X, et al. RIPK3 Mediates Necroptosis during
 Embryonic Development and Postnatal Inflammation in Fadd-Deficient Mice. Cell reports 2017,
 19(4): 798-808.

2169

2170 425. Newton K, Wickliffe KE, Dugger DL, Maltzman A, Roose-Girma M, Dohse M, *et al.* Cleavage of RIPK1 by caspase-8 is crucial for limiting apoptosis and necroptosis. *Nature* 2019, **574**(7778): 428-431.

2173

2174 426. Zhang X, Dowling JP, Zhang J. RIPK1 can mediate apoptosis in addition to necroptosis during embryonic development. *Cell death & disease* 2019, **10**(3): 245.

- Anderton H, Bandala-Sanchez E, Simpson DS, Rickard JA, Ng AP, Di Rago L, et al. RIPK1
   prevents TRADD-driven, but TNFR1 independent, apoptosis during development. Cell death and
   differentiation 2019, 26(5): 877-889.
- Dowling JP, Alsabbagh M, Del Casale C, Liu ZG, Zhang J. TRADD regulates perinatal development and adulthood survival in mice lacking RIPK1 and RIPK3. *Nature communications* 2019, **10**(1): 705.

2184

2189

2193

2197

2200

2203

2206

2210

- 2185 429. Kang TB, Oh GS, Scandella E, Bolinger B, Ludewig B, Kovalenko A, *et al.* Mutation of a self-2186 processing site in caspase-8 compromises its apoptotic but not its nonapoptotic functions in 2187 bacterial artificial chromosome-transgenic mice. *Journal of immunology (Baltimore, Md : 1950)* 2188 2008, **181**(4): 2522-2532.
- 2190 430. Fritsch M, Günther SD, Schwarzer R, Albert MC, Schorn F, Werthenbach JP, *et al.* Caspase-8 is 2191 the molecular switch for apoptosis, necroptosis and pyroptosis. *Nature* 2019, **575**(7784): 683-2192 687.
- Tummers B, Mari L, Guy CS, Heckmann BL, Rodriguez DA, Rühl S, et al. Caspase-8-Dependent
   Inflammatory Responses Are Controlled by Its Adaptor, FADD, and Necroptosis. Immunity
   2020, 52(6): 994-1006.e1008.
- Newton K, Wickliffe KE, Maltzman A, Dugger DL, Reja R, Zhang Y, *et al.* Activity of caspase-8 determines plasticity between cell death pathways. *Nature* 2019, **575**(7784): 679-682.
- 2201 433. Bedoui S, Herold MJ, Strasser A. Emerging connectivity of programmed cell death pathways and its physiological implications. *Nature reviews Molecular cell biology* 2020, **21**(11): 678-695.
- Fan C, Pu W, Wu X, Zhang X, He L, Zhou B, *et al.* Lack of FADD in Tie-2 expressing cells causes RIPK3-mediated embryonic lethality. *Cell death & disease* 2016, **7**(9): e2351.
- 2207 435. Kang TB, Ben-Moshe T, Varfolomeev EE, Pewzner-Jung Y, Yogev N, Jurewicz A, et al.
  2208 Caspase-8 serves both apoptotic and nonapoptotic roles. Journal of immunology (Baltimore, Md
  2209 : 1950) 2004, 173(5): 2976-2984.
- 2211 436. Newton K, Harris AW, Bath ML, Smith KG, Strasser A. A dominant interfering mutant of FADD/MORT1 enhances deletion of autoreactive thymocytes and inhibits proliferation of mature T lymphocytes. *The EMBO journal* 1998, **17**(3): 706-718.
- 2215 437. Zörnig M, Hueber AO, Evan G. p53-dependent impairment of T-cell proliferation in FADD dominant-negative transgenic mice. *Current biology: CB* 1998, **8**(8): 467-470.

Zhang J, Cado D, Chen A, Kabra NH, Winoto A. Fas-mediated apoptosis and activation-induced
 T-cell proliferation are defective in mice lacking FADD/Mort1. *Nature* 1998, 392(6673): 296-300.

Zhang X, Dong X, Wang H, Li J, Yang B, Zhang J, et al. FADD regulates thymocyte development at the β-selection checkpoint by modulating Notch signaling. *Cell death & disease* 2014, 5(6): e1273.

2221

2225

2228

2232

2237

2241

2245

2248

2251

2255

Newton K, Harris AW, Strasser A. FADD/MORT1 regulates the pre-TCR checkpoint and can function as a tumour suppressor. *The EMBO journal* 2000, **19**(5): 931-941.

- Kabra NH, Kang C, Hsing LC, Zhang J, Winoto A. T cell-specific FADD-deficient mice: FADD is required for early T cell development. *Proceedings of the National Academy of Sciences of the United States of America* 2001, **98**(11): 6307-6312.
- 2233 442. Zhang Y, Rosenberg S, Wang H, Imtiyaz HZ, Hou YJ, Zhang J. Conditional Fas-associated death domain protein (FADD): GFP knockout mice reveal FADD is dispensable in thymic development but essential in peripheral T cell homeostasis. *Journal of immunology (Baltimore, Md : 1950)* 2005, **175**(5): 3033-3044.
- 2238 443. Osborn SL, Diehl G, Han SJ, Xue L, Kurd N, Hsieh K, *et al.* Fas-associated death domain (FADD) is a negative regulator of T-cell receptor-mediated necroptosis. *Proceedings of the National Academy of Sciences of the United States of America* 2010, **107**(29): 13034-13039.
- 2242 444. Zhang XY, Yang BY, Wang JY, Mo X, Zhang J, Hua ZC. FADD is essential for glucose uptake 2243 and survival of thymocytes. *Biochemical and biophysical research communications* 2014, 2244 451(2): 202-207.
- 2246 445. Walsh CM, Wen BG, Chinnaiyan AM, O'Rourke K, Dixit VM, Hedrick SM. A role for FADD in T cell activation and development. *Immunity* 1998, **8**(4): 439-449.
- Newton K, Kurts C, Harris AW, Strasser A. Effects of a dominant interfering mutant of FADD on signal transduction in activated T cells. *Current biology : CB* 2001, **11**(4): 273-276.
- Imtiyaz HZ, Rosenberg S, Zhang Y, Rahman ZS, Hou YJ, Manser T, et al. The Fas-associated death domain protein is required in apoptosis and TLR-induced proliferative responses in B cells.
   Journal of immunology (Baltimore, Md: 1950) 2006, 176(11): 6852-6861.

- 2256 448. Salmena L, Lemmers B, Hakem A, Matysiak-Zablocki E, Murakami K, Au PY, *et al.* Essential role for caspase 8 in T-cell homeostasis and T-cell-mediated immunity. *Genes & development* 2003, **17**(7): 883-895.
- 2260 449. Beisner DR, Ch'en IL, Kolla RV, Hoffmann A, Hedrick SM. Cutting edge: innate immunity conferred by B cells is regulated by caspase-8. *Journal of immunology (Baltimore, Md : 1950)* 2005, **175**(6): 3469-3473.

2263

2267

2270

2274

2278

2281

2285

2288

- Lemmers B, Salmena L, Bidère N, Su H, Matysiak-Zablocki E, Murakami K, et al. Essential role for caspase-8 in Toll-like receptors and NFkappaB signaling. The Journal of biological chemistry 2007, 282(10): 7416-7423.
- 2268 451. Zhang N, He YW. An essential role for c-FLIP in the efficient development of mature T lymphocytes. *The Journal of experimental medicine* 2005, **202**(3): 395-404.
- 2271 452. Chau H, Wong V, Chen NJ, Huang HL, Lin WJ, Mirtsos C, et al. Cellular FLICE-inhibitory protein is required for T cell survival and cycling. The Journal of experimental medicine 2005, 202(3): 405-413.
- Zhang H, Rosenberg S, Coffey FJ, He YW, Manser T, Hardy RR, et al. A role for cFLIP in B cell proliferation and stress MAPK regulation. Journal of immunology (Baltimore, Md: 1950) 2009, 182(1): 207-215.
- 2279 454. Kennedy NJ, Kataoka T, Tschopp J, Budd RC. Caspase activation is required for T cell proliferation. *The Journal of experimental medicine* 1999, **190**(12): 1891-1896.
- 2282 455. Bohgaki T, Mozo J, Salmena L, Matysiak-Zablocki E, Bohgaki M, Sanchez O, *et al.* Caspase-8 inactivation in T cells increases necroptosis and suppresses autoimmunity in Bim-/- mice. *The Journal of cell biology* 2011, **195**(2): 277-291.
- Salmena L, Hakem R. Caspase-8 deficiency in T cells leads to a lethal lymphoinfiltrative immune disorder. *The Journal of experimental medicine* 2005, **202**(6): 727-732.
- 2289 457. Rosenberg S, Zhang H, Zhang J. FADD deficiency impairs early hematopoiesis in the bone marrow. *Journal of immunology (Baltimore, Md : 1950)* 2011, **186**(1): 203-213.
- Pellegrini M, Bath S, Marsden VS, Huang DC, Metcalf D, Harris AW, *et al.* FADD and caspase-8 are required for cytokine-induced proliferation of hemopoietic progenitor cells. *Blood* 2005, **106**(5): 1581-1589.

2295 459. Schock SN, Young JA, He TH, Sun Y, Winoto A. Deletion of FADD in macrophages and 2296 granulocytes results in RIP3- and MyD88-dependent systemic inflammation. PloS one 2015, 2297 2298 **10**(4): e0124391. 2299 2300 460. Cuda CM, Misharin AV, Khare S, Saber R, Tsai F, Archer AM, et al. Conditional deletion of caspase-8 in macrophages alters macrophage activation in a RIPK-dependent manner. Arthritis 2301 research & therapy 2015, 17: 291. 2302

Vitale I, Manic G, Coussens LM, Kroemer G, Galluzzi L. Macrophages and Metabolism in the Tumor Microenvironment. *Cell metabolism* 2019, **30**(1): 36-50.

2303

2306

2310

2314

2318

2321

2325

2333

Cuda CM, Misharin AV, Gierut AK, Saber R, Haines GK, 3rd, Hutcheson J, et al. Caspase-8 acts as a molecular rheostat to limit RIPK1- and MyD88-mediated dendritic cell activation. Journal of immunology (Baltimore, Md: 1950) 2014, 192(12): 5548-5560.

Huang QQ, Perlman H, Birkett R, Doyle R, Fang D, Haines GK, *et al.* CD11c-mediated deletion of Flip promotes autoreactivity and inflammatory arthritis. *Nature communications* 2015, **6:** 7086.

2315 464. Wu YJ, Wu YH, Mo ST, Hsiao HW, He YW, Lai MZ. Cellular FLIP Inhibits Myeloid Cell Activation by Suppressing Selective Innate Signaling. *Journal of immunology (Baltimore, Md: 1950)* 2015, **195**(6): 2612-2623.

2319 465. Ch'en IL, Tsau JS, Molkentin JD, Komatsu M, Hedrick SM. Mechanisms of necroptosis in T cells. *The Journal of experimental medicine* 2011, **208**(4): 633-641.

Bell BD, Leverrier S, Weist BM, Newton RH, Arechiga AF, Luhrs KA, et al. FADD and caspase 8 control the outcome of autophagic signaling in proliferating T cells. Proceedings of the National
 Academy of Sciences of the United States of America 2008, 105(43): 16677-16682.

2326 467. Ch'en IL, Beisner DR, Degterev A, Lynch C, Yuan J, Hoffmann A, *et al.* Antigen-mediated T cell expansion regulated by parallel pathways of death. *Proceedings of the National Academy of Sciences of the United States of America* 2008, **105**(45): 17463-17468.

Bolze A, Byun M, McDonald D, Morgan NV, Abhyankar A, Premkumar L, et al. Whole-exomesequencing-based discovery of human FADD deficiency. *American journal of human genetics* 2332 2010, 87(6): 873-881.

- 2334 469. Kuehn HS, Caminha I, Niemela JE, Rao VK, Davis J, Fleisher TA, et al. FAS haploinsufficiency is a common disease mechanism in the human autoimmune lymphoproliferative syndrome. 2335
- *Journal of immunology (Baltimore, Md: 1950)* 2011, **186**(10): 6035-6043. 2336

2341

2345

2349

2353

2358

2362

2366

- 470. Kohn LA, Long JD, Trope EC, Kuo CY. Novel Compound Heterozygote Variations in FADD 2338 Identified to Cause FAS-Associated Protein with Death Domain Deficiency. Journal of clinical 2339 immunology 2020, **40**(4): 658-661. 2340
- 2342 471. Savic S, Parry D, Carter C, Johnson C, Logan C, Gutierrez BM, et al. A new case of Fasassociated death domain protein deficiency and update on treatment outcomes. The Journal of 2343 allergy and clinical immunology 2015, **136**(2): 502-505.e504. 2344
- 2346 472. Chun HJ, Zheng L, Ahmad M, Wang J, Speirs CK, Siegel RM, et al. Pleiotropic defects in lymphocyte activation caused by caspase-8 mutations lead to human immunodeficiency. *Nature* 2347 2002, **419**(6905): 395-399. 2348
- Wang J, Zheng L, Lobito A, Chan FK, Dale J, Sneller M, et al. Inherited human Caspase 10 2350 473. 2351 mutations underlie defective lymphocyte and dendritic cell apoptosis in autoimmune lymphoproliferative syndrome type II. Cell 1999, **98**(1): 47-58. 2352
- 474. Martínez-Feito A, Melero J, Mora-Díaz S, Rodríguez-Vigil C, Elduayen R, González-Granado 2354 LI, et al. Autoimmune lymphoproliferative syndrome due to somatic FAS mutation (ALPS-2355 sFAS) combined with a germline caspase-10 (CASP10) variation. *Immunobiology* 2016, **221**(1): 2356 40-47. 2357
- Dechant MJ, Scheuerpflug CG, Pauly E, van der Werff Ten Bosch J, Debatin KM, Fellenberg J. 2359 475. 2360 Screening, identification, and functional analysis of three novel missense mutations in the TRADD gene in children with ALL and ALPS. *Pediatric blood & cancer* 2008, **51**(5): 616-620. 2361
- 2363 476. Lehle AS, Farin HF, Marquardt B, Michels BE, Magg T, Li Y, et al. Intestinal Inflammation and Dysregulated Immunity in Patients With Inherited Caspase-8 Deficiency. Gastroenterology 2364 2019, **156**(1): 275-278. 2365
- 477. Niemela J, Kuehn HS, Kelly C, Zhang M, Davies J, Melendez J, et al. Caspase-8 Deficiency 2367 Presenting as Late-Onset Multi-Organ Lymphocytic Infiltration with Granulomas in two Adult 2368 2369 Siblings. *Journal of clinical immunology* 2015, **35**(4): 348-355.
- 2371 478. Kanderova V, Grombirikova H, Zentsova I, Reblova K, Klocperk A, Fejtkova M, et al. Lymphoproliferation, immunodeficiency and early-onset inflammatory bowel disease associated 2372 with a novel mutation in Caspase 8. *Haematologica* 2019, **104**(1): e32-e34. 2373

2374 2375 479. Piao X, Komazawa-Sakon S, Nishina T, Koike M, Piao JH, Ehlken H, et al. c-FLIP maintains tissue homeostasis by preventing apoptosis and programmed necrosis. Science signaling 2012, 2376 2377 **5**(255): ra93. 2378 2379 480. Panayotova-Dimitrova D, Feoktistova M, Ploesser M, Kellert B, Hupe M, Horn S, et al. cFLIP regulates skin homeostasis and protects against TNF-induced keratinocyte apoptosis. Cell reports 2380 2013, **5**(2): 397-408. 2381 2382

Feoktistova M, Makarov R, Leverkus M, Yazdi AS, Panayotova-Dimitrova D. TNF Is Partially Required for Cell-Death-Triggered Skin Inflammation upon Acute Loss of cFLIP. *International journal of molecular sciences* 2020, **21**(22).

Wittkopf N, Günther C, Martini E, He G, Amann K, He YW, *et al.* Cellular FLICE-like inhibitory protein secures intestinal epithelial cell survival and immune homeostasis by regulating caspase-8. *Gastroenterology* 2013, **145**(6): 1369-1379.

2386

2390

2394

2398

2402

2406

2410

2391 483. Gehrke N, Garcia-Bardon D, Mann A, Schad A, Alt Y, Wörns MA, *et al.* Acute organ failure following the loss of anti-apoptotic cellular FLICE-inhibitory protein involves activation of innate immune receptors. *Cell death and differentiation* 2015, **22**(5): 826-837.

2395 484. Bonnet MC, Preukschat D, Welz PS, van Loo G, Ermolaeva MA, Bloch W, *et al.* The adaptor protein FADD protects epidermal keratinocytes from necroptosis in vivo and prevents skin inflammation. *Immunity* 2011, **35**(4): 572-582.

Welz PS, Wullaert A, Vlantis K, Kondylis V, Fernández-Majada V, Ermolaeva M, et al. FADD prevents RIP3-mediated epithelial cell necrosis and chronic intestinal inflammation. *Nature* 2011, 477(7364): 330-334.

486. Kovalenko A, Kim JC, Kang TB, Rajput A, Bogdanov K, Dittrich-Breiholz O, *et al.* Caspase-8 deficiency in epidermal keratinocytes triggers an inflammatory skin disease. *The Journal of experimental medicine* 2009, **206**(10): 2161-2177.

487. Günther C, Martini E, Wittkopf N, Amann K, Weigmann B, Neumann H, *et al.* Caspase-8 regulates TNF-α-induced epithelial necroptosis and terminal ileitis. *Nature* 2011, **477**(7364): 335-339.

2411 488. Li C, Lasse S, Lee P, Nakasaki M, Chen SW, Yamasaki K, et al. Development of atopic dermatitis-like skin disease from the chronic loss of epidermal caspase-8. Proceedings of the National Academy of Sciences of the United States of America 2010, 107(51): 22249-22254.

2414 489. Kaden-Volynets V, Günther C, Zimmermann J, Beisner J, Becker C, Bischoff SC. Deletion of 2415 the Casp8 gene in mice results in ileocolitis, gut barrier dysfunction, and malassimilation, which 2416 2417 can be partially attenuated by inulin or sodium butyrate. American journal of physiology *Gastrointestinal and liver physiology* 2019, **317**(4): G493-g507. 2418 2419 490. 2420 Weinlich R, Oberst A, Dillon CP, Janke LJ, Milasta S, Lukens JR, et al. Protective roles for caspase-8 and cFLIP in adult homeostasis. *Cell reports* 2013, **5**(2): 340-348. 2421 2422 2423 491. Stolzer I, Kaden-Volynets V, Ruder B, Letizia M, Bittel M, Rausch P, et al. Environmental 2424 Microbial Factors Determine the Pattern of Inflammatory Lesions in a Murine Model of Crohn's Disease-Like Inflammation. *Inflammatory bowel diseases* 2020, **26**(1): 66-79. 2425 2426 2427 492. Kaemmerer E, Kuhn P, Schneider U, Jeon MK, Klaus C, Schiffer M, et al. Intestinal genetic inactivation of caspase-8 diminishes migration of enterocytes. World journal of gastroenterology 2428 2429 2015, **21**(15): 4499-4508. 2430 2431 493. Tisch N, Mogler C, Stojanovic A, Luck R, Korhonen EA, Ellerkmann A, et al. Caspase-8 in endothelial cells maintains gut homeostasis and prevents small bowel inflammation in mice. 2432 *EMBO molecular medicine* 2022, **14**(6): e14121. 2433 2434 494. Schwarzer R, Jiao H, Wachsmuth L, Tresch A, Pasparakis M. FADD and Caspase-8 Regulate 2435 Gut Homeostasis and Inflammation by Controlling MLKL- and GSDMD-Mediated Death of 2436 Intestinal Epithelial Cells. *Immunity* 2020, **52**(6): 978-993.e976. 2437 2438 2439 495. Galluzzi L, López-Soto A, Kumar S, Kroemer G. Caspases Connect Cell-Death Signaling to 2440 Organismal Homeostasis. *Immunity* 2016, **44**(2): 221-231. 2441 496. 2442 Karki R, Kanneganti TD. Diverging inflammasome signals in tumorigenesis and potential targeting. *Nature reviews Cancer* 2019, **19**(4): 197-214. 2443

2444