

## Single cell analysis for intrinsic stemness potential of cancer

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## Single cell analysis for intrinsic stemness potential of cancer

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Stencil.







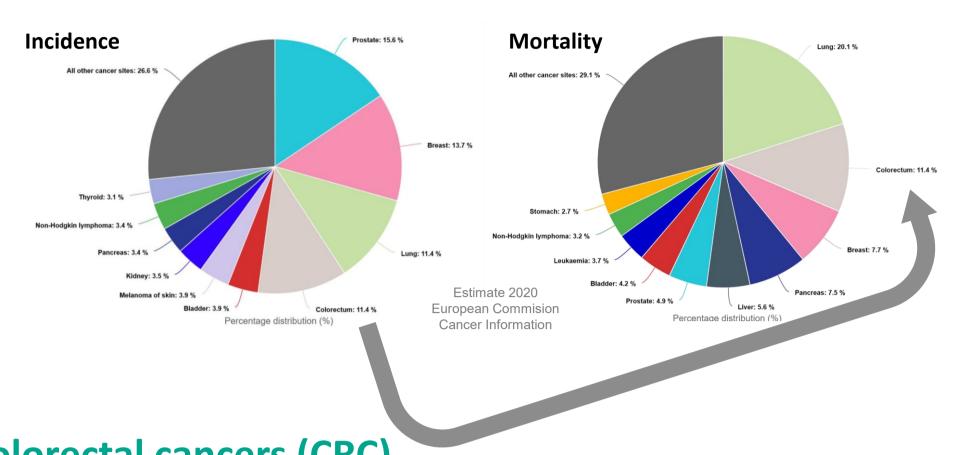




# Canther



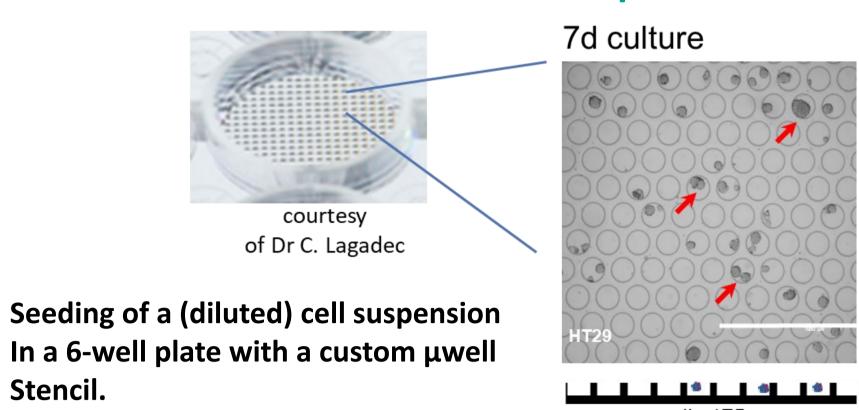
#### **Cancer prevalence in France**



#### **Colorectal cancers (CRC)**

- > Poor survival rates are largely due to resistance to chemotherapeutic treatments and the development of metastatic lesions.
- ➤ More then 15% of patients have metastatic lesions at diagnosis and another 20% develop them. Metastatic CRC has a poor 5-year survival rate (14%)
- > CRC is treated with targeted therapy, immunotherapy and/or surgery combined with chemotherapy (e.g. FOLFIRINOX: 5FU, folic acid, oxaliplatine and irinotécan)

## HT29 cells in custom micro-wells plates

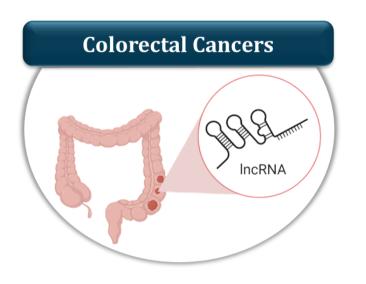


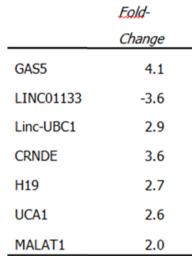
Individual cell ~15 µm Spheres ~ 150 µm

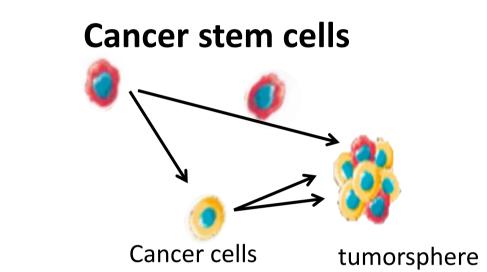
 $\mu$ well = 175  $\mu$ m

- ❖ More then 1 cell / well, no single cell analysis
- Cell distribution sensible to vibrations

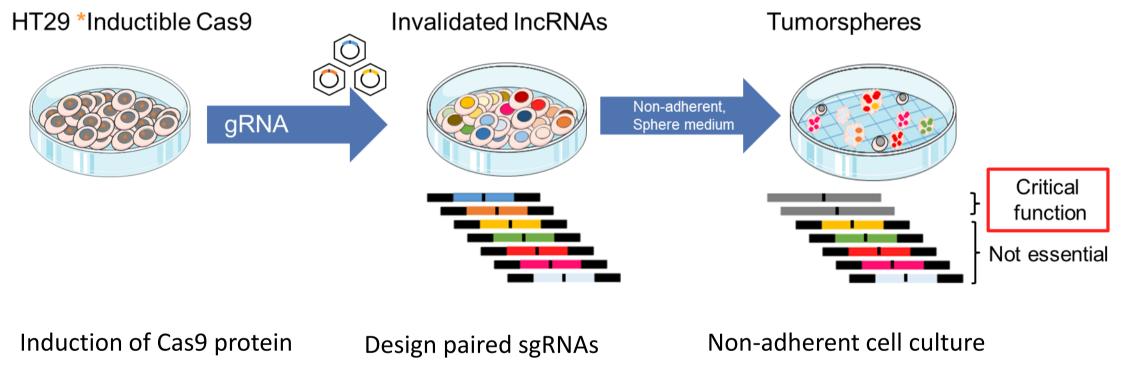
Role of IncRNAs in cancer persistent cells

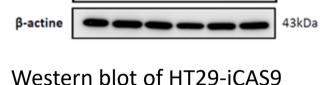




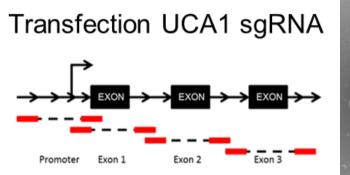


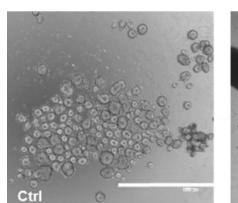
## **Lentiviral CRISPR-Cas9 library screen**

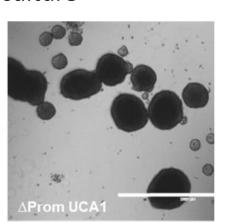




cells with 2 μg/mL Doxicycline (TET-on promoter)



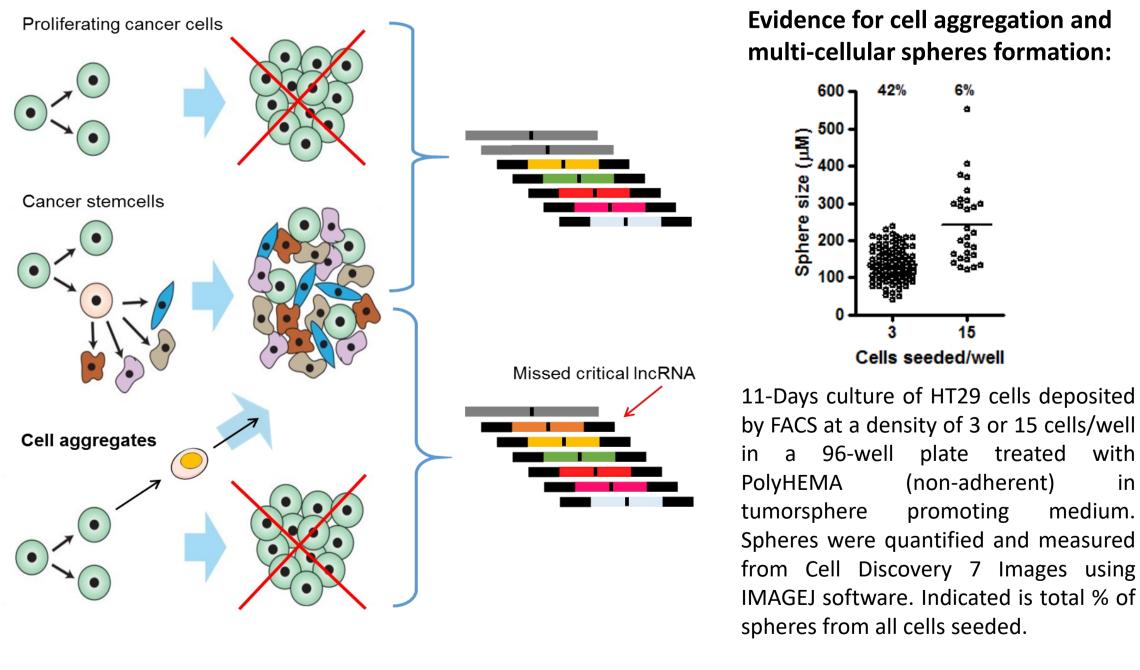




Ctrl/HT29iCAS9-dUCA1 cells cultured for 7 days in tumorsphere promoting conditions

formation

## **→** Need for single cell analysis



## ➤ Need for accessible high-throughput readout assay



- FACS sorting 1 cell / well

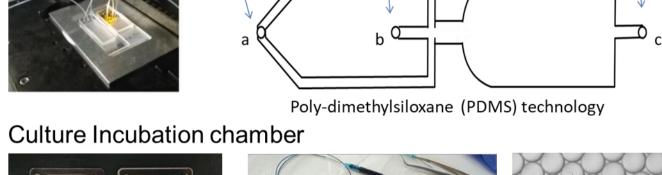
- One sgRNA-pair / cell => low MOI, more cells to infect

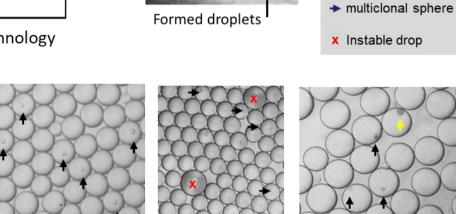
- Negative screen: each sgRNA 1000x represented in library

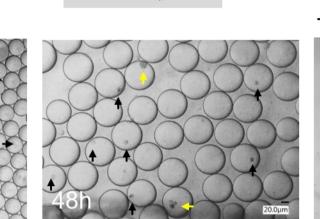
- RNA analysis of all spheres: 96-well format not practical

# HT29 cells in microfluidic droplets



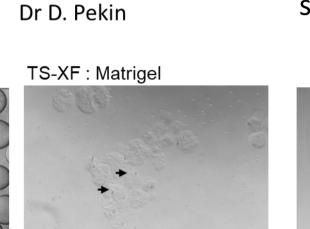


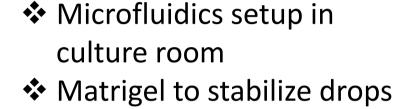




➤ single cell

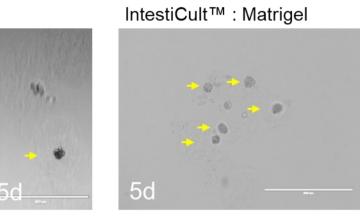
monoclonal sphere



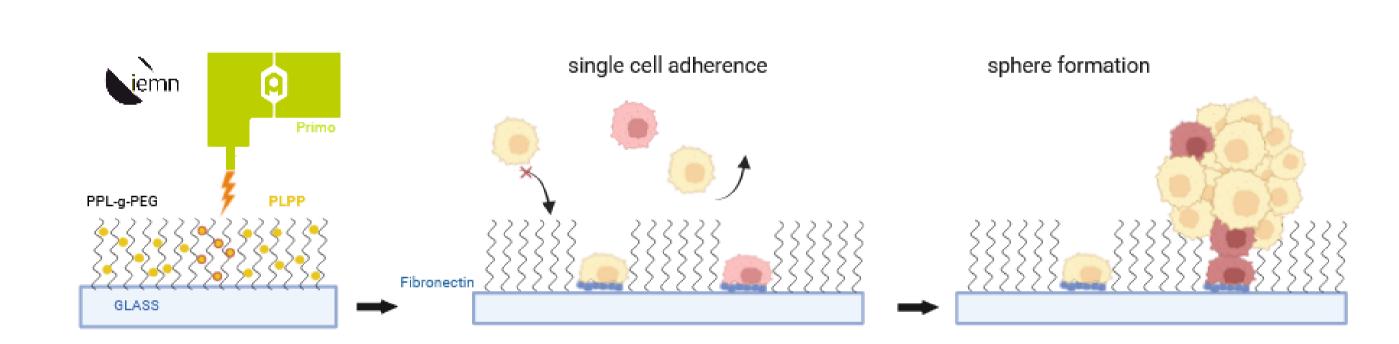


Each experiment needed fine-tuning of flux, difficult to

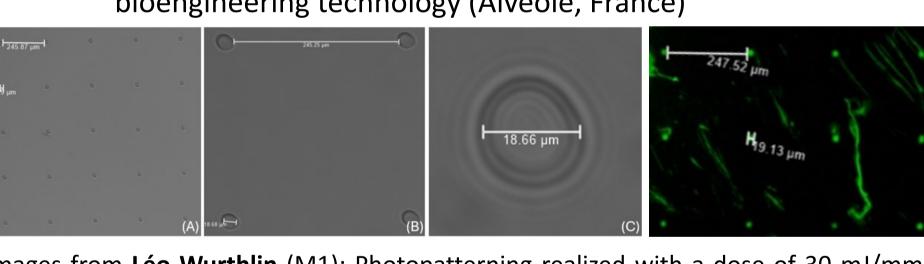
standardize using matrigel



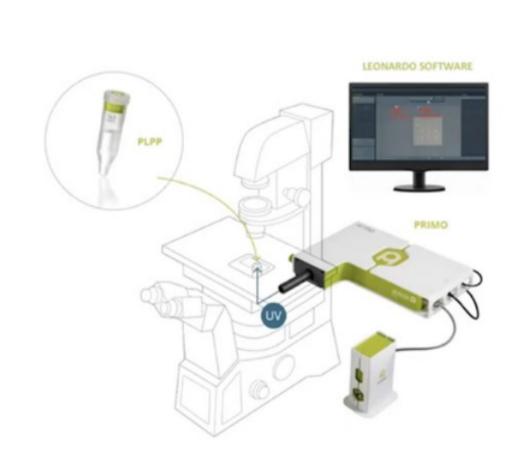
## HT29 cells in single cell adherence plates followed by tumorsphere culture



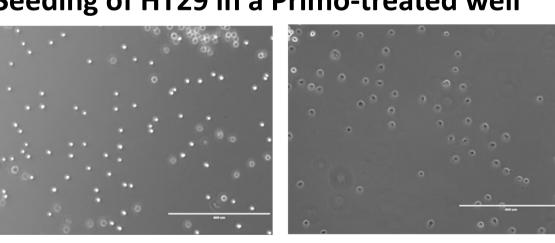
#### **Photopatterning** Primo bioengineering technology (Alvéole, France)



Images from Léo Wurthlin (M1): Photopatterning realized with a dose of 30 mJ/mm<sup>2</sup> to obtain a matrix of 50 x 50 circles with a diametre of 18 μm separated by 250 μm. (A-C) Images with different magnification acquired by a DMi8 inverted microscope, after the photopattern was visualized by DWS LAB Vitra<sup>TM</sup> 429 ink. (D) DMi8-acquired image of a green-fluorescent Alexa Fluor® 488 fibrinogen conjugate deposited in a 6-well glass bottom plate after photopatterning.

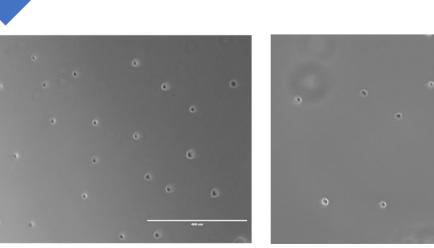


## Seeding of HT29 in a Primo-treated well



HT29 cells incubated at 37C in standard culture medium for at least 2h. O/N culture results in inert cells

Washes with PBS to remove cells not

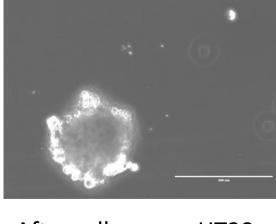


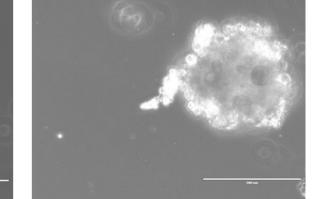
in adherence spots

HT29 cells seeded at 200K cells/well adhere in an individual cell pattern. Primo-treated plates are best used within 1-2 days for best results.



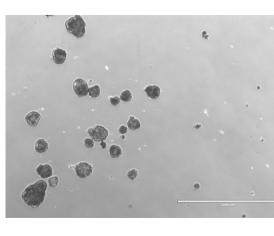
> Seeding of a cell suspension, but individual cell culture





After adherence HT29 cells were cultured in 3D Tumorsphere Medium XF (Promocell). Only stem-like cells will proliferate and develop tumorspheres

Swirling the plate will release the tumorspheres in suspension (not the adherent cancer cells)



- > This method is suitable as high-throughput readout assay for library screens
- > Single-cell originated tumorspheres can easily be recolted for RNAseq analyses assessing sgRNA expression and transcriptomes

