

Intercostal muscle oxygenation and expiratory loaded breathing at rest: Respiratory pattern effect

Quentin Bretonneau, Claire de Bisschop, Valentin Mons, Aurélien Pichon

▶ To cite this version:

Quentin Bretonneau, Claire de Bisschop, Valentin Mons, Aurélien Pichon. Intercostal muscle oxygenation and expiratory loaded breathing at rest: Respiratory pattern effect. Respiratory Physiology & Neurobiology, 2022, 304, pp.103925. 10.1016/j.resp.2022.103925. hal-04563409

HAL Id: hal-04563409 https://hal.science/hal-04563409

Submitted on 22 Jul 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

Version of Record: https://www.sciencedirect.com/science/article/pii/S1569904822000842 Manuscript_1de2d8ac5dff842049b2159a589e9551

1 Intercostal muscle oxygenation and expiratory loaded breathing

2 at rest: respiratory pattern effect

3

4 (Duentin Bretonneau ^{a*} .	Claire de Bisscho	р ^а .	Valentin Mons ^a	Aurélien Pichon
			- T		,

^a Université de Poitiers, Faculté des Sciences du Sport, Laboratoire 'MOVE', EA 6314

6 8, allée Jean Monnet – TSA 31 113 – 86073 Poitiers, Cedex 9 – France

7

- 8 * Corresponding author:
- 9 Quentin Bretonneau
- 10 Université de Poitiers, Faculté des Sciences du Sport, Laboratoire 'MOVE', EA 6314
- 11 8, allée Jean Monnet TSA 31 113 86073 Poitiers, Cedex 9 France
- 12 Tel: +33 5 49 45 40 42 / E-mail: quentin.bretonneau@univ-poitiers.fr
- 13 ORCID id: https://orcid.org/0000-0002-9307-8688
- 14
- 15
- 16

17

19 ABSTRACT

In patients with airway obstruction, an increase in breathing frequency at rest is commonly associated with a dynamic hyperinflation (DH). In such a situation, intercostal muscle oxygenation may be disturbed. This hypothesis was examined in a context of simulated airway obstruction in healthy subjects.

After a control period of 5 minutes, twelve participants (20 ± 2 years) breathed at rest through a 20-cmH₂O expiratory threshold load, either by increasing or reducing their respiratory rate ($_{ETL}F^+$ or $_{ETL}F^-$). Tissue saturation index (TSI) and concentration changes in oxyhaemoglobin (oxy[Hb+Mb]) were measured as well as cardiorespiratory variables.

Inspiratory capacity was decreased in $_{\text{ETL}}\text{F}^+$ (p<0.001) and correlated with dyspnea. An increase in oxy[Hb+Mb] occurred in $_{\text{ETL}}\text{F}^+$ that was higher than in $_{\text{ETL}}\text{F}^-$ (p<0.01). TSI

31 was not different between conditions.

In healthy subjects at rest, an increase in respiratory rate during a simulated obstruction
with an expiratory threshold load resulted in paradoxical response with DH emergence
while intercostal muscle oxygenation was preserved.

35

36

37 **KEYWORDS**

Intercostal muscle oxygenation, NIRS, expiratory threshold load, respiratory pattern,dynamic hyperinflation

40

41

43

44 1. INTRODUCTION

In patients with obstructive lung disease, maximal expiratory flows are reduced due to bronchial obstruction and flows available are usually insufficient to ensure relaxation of respiratory system. In this case, as end expiratory lung volume (EELV) is increased and tidal volume is shifted at higher lung volume, dynamic hyperinflation (DH) can occur. This situation is often observed in obstructive patients, at rest, along with an increase in respiratory rate (Klooster et al., 2015; Lavorini et al., 2013). DH is also usually majored by exercise.

52 Such a context may have deleterious consequences on the perfusion of 53 respiratory muscles because of adverse combined factors. First, ventilatory effort, which is usually modest in healthy subjects, is much more important in patients with 54 55 bronchial obstruction. Thus, maintaining adequate ventilation may require greater contraction of respiratory muscles. This exaggerated effort may affect the capillary 56 perfusion of respiratory muscles while in the same time their oxygen needs are 57 increased (Leenaerts and Decramer, 1990). Furthermore, in a context of DH, intercostal 58 59 muscles are likely to be stretched (Wilson et al., 2001). This change in length may affect 60 the capillaries by reducing their diameter (Poole et al., 1997). Thereby, in context of 61 high expiratory resistance at rest, blood perfusion could be particularly disturbed in intercostal muscles, especially if DH occurs. Muscle oxygen supply could become 62 63 insufficient according to their majored metabolic needs, inducing a metabolic imbalance 64 state.

65 Several studies have already imposed an external expiratory load on healthy subjects in order to induce the ventilatory disorders associated with pulmonary 66 67 pathologies (Athanasopoulos et al., 2010; Barrett et al., 1994; Bretonneau et al., 2020, 2019; Stark-Leyva et al., 2004; Xiao et al., 2012). In our previous study, an expiratory 68 69 threshold load (ETL) had been imposed in healthy subjects at rest to assess ventilatory adaptations (Bretonneau et al., 2019). Ventilatory variables such as tidal volume and 70 71 total respiratory time were increased in response to expiratory loaded breathing while 72 intercostal muscle blood volume and concentration in oxyhaemoglobin were decreased. However, metabolic balance seemed not disturbed in these muscles. A limit of this 73 74 study was nevertheless that DH was probably not induced, as inspiratory capacity was 75 not changed in response to ETL.

In the current study, in order to appreciate the influence of DH on metabolic balance in intercostal muscles, two respiratory patterns were imposed on healthy subjects during simulated obstruction condition. Subjects were asked to breathe either with high respiratory rate or with low breathing frequency against an ETL. High respiratory rate associated with expiratory loaded breathing was hypothesized to induce DH that was expected to disturb importantly intercostal muscle oxygenation.

82 2. MATERIAL AND METHODS

83 2.1. Participants

Twelve healthy active men $(20 \pm 2 \text{ years}, 1,7 \pm 0.1 \text{ m}, 70 \pm 9 \text{ kg})$ were included in this study. Anthropometric and pulmonary function data are given in Table 1. All subjects had FEV1 \geq 80% of the predicted value (Quanjer et al., 2012), FEV1/FVC > 70% and skinfold < 20 mm at the seventh left intercostal space (Grassi and Quaresima, 2016).

88 *** Table 1 ***

89 2.2. Ethical approval

90 The protocol was approved by the Ethics Committee in Sport Science Research
91 (CERSTAPS 2018-25-01-22) and was presented to participants prior to the beginning of
92 the study. All subjects gave their written informed consent.

93 2.3. Study protocol

94 2.3.1 Experimental protocol

Two sessions were scheduled. At each session, subjects breathed as naturally as possible for 5 minutes, seated on a chair with a nose clip (control period). Afterwards, participants breathed for 20 minutes through a 20-cmH₂O expiratory threshold load (ETL, Threshold PEP, Respironics Inc., Murrysville, PA, USA). As soon as the ETL was applied at the mouth, participants had to increase their control breathing frequency by 30% or decrease it by 40% ($_{ETL}F^+$ or $_{ETL}F^-$ condition) according to the randomisation (Figure 1). This decrease of 40% was based on our previous study in which subjects were free to adjust their respiratory pattern in response to ETL(Bretonneau et al., 2019). Sound metronome was used to regulate the respiratory rate.

Tidal volume was calculated to obtain minute ventilation close to that experienced
spontaneously during our previous study (Bretonneau et al., 2019). Seven days
separated the two sessions scheduled at the same time of day.

107 2.3.2 Measurements and materials

2.3.2.1 Anthropometry. Skinfold was assessed at the seventh left intercostal space with a
skinfold caliper (Harpenden, British Indicators Ltd., St. Albans, England) and a wholebody bioelectrical impedance technique was used to assess body fat mass (Tanita BC418 MA, Tanita Corporation, Tokyo, Japan).

2.3.2.2 Pulmonary function tests and inspiratory capacity. Slow and forced vital
capacities (SVC and FVC, respectively) were evaluated at each session (Metalyzer-3B,
Cortex Biophysik GmbH, Leipzig, Germany) according to the ATS/ERS Task Force
recommendation (Miller et al., 2005).

Inspiratory capacity (IC) was measured several times during the protocol (Figure 1) to detect ETL-induced DH (Guenette et al., 2013; Yan et al., 1997). During control period (Ctrl), a set of two maximal inspirations were performed and the best maneuver (*i.e.* the one with the highest IC) was chosen as reference IC. During ETL period, two single maximal inspirations were performed. A second trial was realized if the inspiration had been disturbed (*e.g.* saliva swallowing during the maneuver).

2.3.2.3 Maximal respiratory pressures. Maximal expiratory and inspiratory pressures
(PEmax and PImax) were measured before and 30 seconds after ETL period with a non-

invasive manometer (MicroRPM, Micro Medical, Kent, England). Sniff nasal technique
was used to assess PImax while PEmax was measured at the mouth. Three attempts
were performed before the protocol to familiarize with the procedure.

127 2.3.2.4 Intercostal muscle oxygenation. Near-infrared spectroscopy (NIRS) (Oxymon Mk III, Artinis Medical Systems, Zetten, The Netherlands) was used to assess the tissue 128 129 saturation index (TSI) and the concentration changes in oxygenated, deoxygenated and 130 total haemoglobin and myoglobin in the intercostal muscles ($\Delta \text{ oxy}[\text{Hb+Mb}], \Delta$ 131 deoxy[Hb+Mb] and Δ total[Hb+Mb], respectively) (Barstow, 2019; Bretonneau et al., 132 2020, 2019; de Bisschop et al., 2014). While TSI and oxy[Hb+Mb] reflect local tissue oxygenation, deoxy[Hb+Mb] may reflect local oxygen extraction. Furthermore, 133 total[Hb+Mb] (*i.e.* the sum of oxy[Hb+Mb] + deoxy[Hb+Mb]) can be seen as an 134 135 indicator of local blood volume. The concentration changes in response to ETL were calculated from control period. The optode was placed in the seventh intercostal space, 136 137 on the left side of the subject, between the midclavicular and the anterior axillary lines. As maximal penetration depth of the light is approximately half of the distance between 138 transmitter and receiver (Chance et al., 1992), interoptode distance was adjusted 139 140 between 3.3 and 4.8 cm according to adipose tissue thickness. A differential pathlength 141 factor of 4 was chosen (van Beekvelt et al., 2002). Wavelengths of 859 and 763 nm were used. Data were recorded at 10 Hz and exported at 1 Hz after running average 142 143 filtering.

2.3.2.5 Ventilatory variables and gas exchanges. Inspiratory and expiratory times (Ti
and Te, respectively) and tidal volume (Vt) were measured breath-by-breath using the
Metalyzer device (Cortex Biophysik GmbH, Leipzig, Germany). Total respiratory time
(Ttot), inspiratory duty cycle (Ti/Ttot), breathing frequency (BF) and ventilatory flow

148 ($\dot{V}E$) were calculated, as mean inspiratory and expiratory flow rates (Vt/Ti and Vt/Te). 149 Oxygen consumption ($\dot{V}O_2$), carbon dioxide production ($\dot{V}CO_2$) and end-tidal CO₂ 150 pressure (PetCO₂) were likewise recorded. Respiratory exchange ratio (RER) and 151 respiratory equivalents in oxygen ($\dot{V}E/\dot{V}O_2$) and carbon dioxide ($\dot{V}E/\dot{V}CO_2$) were 152 calculated.

2.3.2.6 Dyspnea. Perception of dyspnea was assessed using the modified 0-10 Borgscale, before each IC measurement (Kendrick et al., 2000). Subjects were required to
point on the scale the level of their respiratory discomfort.

2.3.2.7 Pulse oximetry. A pulse oximeter (Wrist Ox2 3150, Nonin Medical Inc.,
Plymouth, MN, USA) was placed at the right index to continuously record pulse
haemoglobin oxygen saturation (SpO₂).

159 2.3.2.8 Cardiovascular parameters. A sphygmomanometer (Omron M3, Health Care 160 CO. Ltd, Kyoto, Japan) was used to measure systolic and diastolic blood pressure (SBP 161 and DBP) before each IC measurement. Mean arterial pressure (MAP) was calculated as 162 follows: MAP = DBP + 1/3 (SBP - DBP). A PhysioFlow device was used to measure 163 stroke volume (SV), heart rate (HR) and cardiac output (Qc) non-invasively and beat-164 by-beat (PF-05, Manatec Medical, Poissy, France). This recording was triggered 165 synchronously with the recordings of the other variables measured continuously.

166 2.4. Statistical analysis

Statistical analysis was performed with Statistica (StatSoft Inc., Tulsa, OK, USA). For
variables recorded continuously, mean values were calculated from the last 30 seconds
preceding IC measurements. Following the verification of parametric conditions, a first

170 two-way repeated measures ANOVA was performed to evaluate the responses to ETL (control period vs. ETL10') according to the respiratory patterns (ETLF⁺ vs. ETLF⁻). A 171 172 second ANOVA was conducted to test the effect of ETL duration (ETL10' vs. ETL20') according to the respiratory patterns. A two-way repeated measures ANOVA was also 173 174 conducted to compare maximal respiratory pressures before and after ETL period. A 175 Tukey post-hoc analysis was performed when ANOVA was significant. Furthermore, in order to detect DH induced by ETL, IC measured at ETL10' and ETL20' were compared 176 with the corresponding one measured during control period. Hedges' g was calculated 177 178 to define effect size (0.2: small; 0.5: medium; 0.8: large; 1.2: very large; 2.0: huge). Correlation coefficients were assessed using the Pearson or Spearman test, according to 179 180 Gaussian distribution. Values were expressed as mean ± standard deviation (SD) and results were considered as significant for a p-value < 0.05. 181

182 **3. RESULTS**

183 *3.1. Ventilation*

184 VE was not different between the two experimental conditions and Vt was higher in 185 $_{\text{ETL}}\text{F}^-$ as compared to $_{\text{ETL}}\text{F}^+$ (p < 0.001). Ti/Ttot, Vt/Ti and Vt/Te were not different 186 between conditions (Table 2 and Figure 2).

- 187 In $_{\text{ETL}}\text{F}^+$, IC was decreased from Ctrl (2.8 L at 20' $_{\text{ETL}}\text{F}^+$ vs. 3.5 L during Ctrl; 188 p < 0.001, Hedges' g = 0.8) (Figure 3).
- 189 Dyspnea was not different between conditions and increased significantly during $_{\text{ETL}}\text{F}^+$ 190 (from 2.5/10 on Borg scale at 10' to 4.1/10 at 20') (Figure 3).

191 *** Table 2 ***

192 3.2. Intercostal muscle oxygenation

NIRS values are shown in Figure 4. In response to ETL, oxy[Hb+Mb] was significantly 193 increased in $ETLF^+$ but not in $ETLF^-$ (at 20': +4.40 μ M vs. +0.44 μ M; p < 0.01, 194 Hedges' g = 0.9). These changes were significantly different between breathing 195 patterns. Total[Hb+Mb] was increased in ETLF⁺ while it was decreased after a few 196 minutes in $_{ETL}F^-$ (at 20': +2.3 μ M vs. -1.5 μ M; p < 0.05). These changes were 197 significantly different between breathing patterns. Deoxy[Hb+Mb] decreased identically 198 199 in both conditions after a few minutes of loaded breathing. TSI was not different between conditions and did not vary from control period, neither in ETF^+ nor in ETF^- . 200

201 3.3. Maximal respiratory pressures

PEmax and PImax were not different between conditions and did not change after ETL
period (from 150 cmH₂O to 147 cmH₂O for PEmax and from 114 cmH₂O to 110
cmH₂O for PImax).

205 3.4. Gas exchanges

- As shown in Table 2, $\dot{V}E/\dot{V}O_2$ and $\dot{V}E/\dot{V}CO_2$ were increased from Ctrl while PetCO₂
- 207 was decreased. RER, $\dot{V}E/\dot{V}O_2$ and $\dot{V}E/\dot{V}CO_2$ were lower in $ETLF^+$ than in $ETLF^-$.
- 208 Conversely, PetCO₂ was higher in $ETLF^+$. $\dot{V}O_2$, $\dot{V}CO_2$ and SpO₂ were not different
- between conditions. At 20', SpO₂ was 98% in the two conditions.

210 3.5. Cardiovascular variables

- 211 Neither cardiac parameters (HR, SV and Qc; Table 2) nor arterial pressures (SBP, DBP,
- and MAP) were significantly different between conditions. At 20', MAP was 99 mmHg
- 213 in $ETLF^+$ and 100 mmHg in $ETLF^-$.

214 3.6. Correlations

- In $ETLF^+$, changes in IC ($\Delta = ETL20'$ control period) were correlated with changes in
- 216 dyspnea (r = -0.67, p = 0.02).

217 **4. DISCUSSION**

218 This study aimed to investigate intercostal muscle oxygenation in a context of 219 high expiratory resistance that frequently induces DH. In such a situation in which 220 intercostal muscles are likely to be stretched, blood perfusion could be impaired and 221 oxygenation disturbed. In order to appreciate the influence of DH on metabolic balance 222 in intercostal muscles, two respiratory patterns were imposed on healthy subjects by 223 simulating obstruction with an ETL. We found that at rest, increasing respiratory rate 224 resulted in paradoxical response with DH and dyspnea onset while blood perfusion was enhanced and oxygenation of intercostal muscles preserved. 225

226 4.1. Methodological aspects – Breathing frequency

In $_{\text{ETL}}\text{F}^+$ condition, control breathing frequency was increased by 30% in order to induce DH (*i.e.* to increase end-expiratory lung volume) (Gelb et al., 2004). Initially, a rise of 40% was planned but during our pre-experimentations, several participants expressed difficulties to maintain the rhythm during 20 minutes.

In $_{\text{ETL}}\text{F}^-$ condition, control breathing frequency was decreased by 40% as soon as the ETL was applied. This drop was imposed in order to reproduce the spontaneous change previously observed in healthy subjects in similar context (Bretonneau et al., 2019). Furthermore, it was imposed with the aim of maintaining (or decreasing) the end-expiratory lung volume with the ETL (Bretonneau et al., 2019). Thus, in the current study, $_{\text{ETL}}\text{F}^-$ condition can be seen as a control condition.

237 4.2. Respiratory patterns, hyperinflation and intercostal muscle oxygenation

238 In obstructive lung diseases, worsening of DH is usually demonstrated by a decrease in IC from steady-state period (Guenette et al., 2013; Yan et al., 1997). In 239 $_{\rm ETI} F^{-}$ condition, IC did not vary from control period. In contrast, in $_{\rm ETI} F^{+}$ condition, IC 240 decreased to a significant and large extent according to the Hedges' g (-19%, reduction 241 242 > 150 ml in 9 participants). These results confirm that in a context of high expiratory 243 resistance, DH is favoured by rapid and shallow breathing. Our results agree with those reported by Gelb et al. (2004) as in their study performed in obstructive patients, a 18% 244 245 decreased in inspiratory capacity was induced by a two-fold increase in respiratory rate.

When an external expiratory resistance is applied at the mouth, healthy subjects 246 usually increase spontaneously their tidal volume (Barrett et al., 1994; Bretonneau et al., 247 2019; Suzuki et al., 1991; Xiao et al., 2012), probably to minimize unconsciously the 248 work of expiratory muscles. Indeed, by increasing tidal volume, a greater elastic recoil 249 of lung may facilitate expiration. In $_{ETL}F^+$ condition, as tidal volume is decreased, only 250 a shift in tidal breathing to higher lung volumes seems able to increase the elastic recoil. 251 These suggestions could explain the occurrence of DH with a rapid and shallow 252 breathing but not with a deep and slow ventilation in healthy population with ETL. 253

During expiratory loaded period, total[Hb+Mb] and oxy[Hb+Mb] were not different from baseline in $_{ETL}F^-$ while it increased in $_{ETL}F^+$. In parallel, change in deoxy[Hb+Mb] was not different between conditions. These results suggest that oxygen extraction was likely similar between conditions despite a higher blood volume and oxyhaemoglobin concentration in $_{ETL}F^+$. These "over-oxygenation" could be explained by a pump effect of respiratory muscles resulting from the increase in respiratory frequency.

261 During ETL period, TSI was not different between conditions and was not modified from control period, neither in $ETLF^+$ nor in $ETLF^-$, which is in agreement with 262 our previous results (Bretonneau et al., 2019). As intercostal muscles were probably 263 264 recruited during ETL period (de Bisschop et al., 2017), these results suggest that higher 265 metabolic needs of intercostal muscles were adequately satisfied in the two conditions 266 (Ferrari et al., 2011). Maximal respiratory pressures being not changed after expiratory 267 loaded breathing, the preservation of metabolic balance in respiratory muscles could explain the lack of respiratory muscle fatigue at the end of the two conditions (Verges et 268 269 al., 2010).

270 According to our present results, obtained at rest, DH did not seem to have an effect on the oxygen saturation of intercostal muscles but this might be somewhat 271 different in exercise conditions. Indeed, Vogiatzis et al. (2010, 2011) recorded a 272 273 decrease in intercostal muscle blood flow and oxygenation in obstructive patients when their end-expiratory volume increased in response to exercise, while conversely a better 274 275 tissue oxygenation was observed when end-expiratory volume normalized (Vogiatzis et 276 al., 2010, 2011). In agreement, in a previous study performed at exercise, our team had observed an increase in oxy[Hb+Mb] and deoxy[Hb+Mb] that stopped when DH 277 278 occurred (Bretonneau et al., 2020). Besides, in the study from Louvaris et al. (2018), in 279 which obstructive patients were asked to reproduce different levels of exercise-induced hyperpnea, both intercostal muscles blood flow and DH leveled off from an hyperpnea 280 281 associated with 75% of maximal work rate. Finally, the causal links between DH and 282 respiratory muscle oxygenation are still to be clarified.

283 **4.3.** Dyspnea

Dyspnea was not different between conditions at $_{ETL}10'$ and increased thereafter only in $_{ETL}F^+$ condition (up until 4 on the modified Borg scale, dyspnea somewhat severe). Furthermore, a negative correlation between IC and dyspnea was observed in this condition. In agreement, Lavorini et al. (2013) showed that dyspnea was more intense for asthmatic patients who increased their breathing frequency in response to induced-bronchoconstriction, as compared to those who decreased it. Likewise, these patients were those who showed the greatest DH.

291 Paradoxical effects on respiratory mechanics appears associated to DH. On one 292 side, DH increases inspiratory muscle efferent due to higher work of breathing and on 293 the other side, DH reduces the ability of inspiratory muscles to produce strength by 294 disrupting their tension-length relationship (Braun et al., 1982). Hence, due to the shift 295 in tidal breathing to higher lung volumes, mismatch between inspiratory muscle efferent 296 and mechanical/muscular response (*i.e.* neuromuscular dissociation) may occur, which 297 could worsen dyspnea (Lougheed et al., 1993; O'Donnell et al., 1997). The correlation between IC and dyspnea highlighted in the current study, is consistent with the 298 hypothesis that subject who reported the higher dyspnea in $ETLF^+$ could be those who 299 300 experienced the higher neuromuscular dissociation.

It could be also hypothesized that dyspnea is linked to oxygenation state of the respiratory muscles. Indeed, a correlation between dyspnea and deoxygenation of the serratus muscle has been observed in patients with heart failure (Mancini et al., 1992). Moreover, in obstructive patients, a reduction of dyspnea has been observed during exercise while oxygen supply to the intercostal muscles was improved by oxygen

supplementation (Louvaris et al., 2016). Unfortunately, in our study, we did not findany association between dyspnea and intercostal muscle oxygenation.

308 4.4. Limits of the study

309 Some points which can be considered as limits are presented below. First, 310 respiratory muscle oxygenation was investigated in intercostal muscles by NIRS. 311 However, the exact depth of the intercostal muscles under the skin was unknown, which 312 represents a limit for NIRS investigations. The depth of measurement was chosen 313 according to a previous estimation of external muscle thickness by De Troyer et al. 314 (2003) at 7-10 mm, which seems reasonable.

Secondly, in the present study, the activity of the intercostal muscles was not measured electromyographically. However, the activation of intercostal muscles has previously been demonstrated under similar conditions in humans (de Bisschop et al., 2017; Sears and Newsom Davis, 1968). In addition, a two-fold increase in Vt/Te from baseline was observed in the two conditions, which indicates the recruitment of intercostal muscles.

Thirdly, cardiac values were obtained with a Physioflow device, whose accuracy is still debated in obstructive patients, especially during exercise (Bougault et al., 2005; Louvaris et al., 2019). In our study however, cardiac parameters were measured only at rest, and in view of carrying out a general survey of cardiac adjustments alongside DH and respiratory muscle activity.

Finally, the study was undertaken on healthy subjects using simulation conditions. Thus, the physiological responses described in this study may differ from the real-condition responses that occur in patients with obstructive pulmonary diseases.

329 4.5. Strength, clinical implication and perspective of the study

In the present study, we demonstrated that DH was induced by breathing rapidly against an expiratory load and that the shift in tidal breathing to higher lung volumes was associated with dyspnea. Thus, our observations carried out on healthy subjects suggest to advise slow and deep breathing to patients during exacerbation of bronchial obstruction in order to limit DH and dyspnea. Nevertheless, these recommendations should be confirmed on patients to be clinically relevant.

In addition, this study confirms that the use of a PEP can be an excellent way to increase EELV in healthy subjects. This model seems particularly interesting for studying the isolated effects of DH without including the others potential effects associated with obstructive pulmonary diseases (Erram et al., 2021).

340 **5. CONCLUSION**

In conclusion, this study performed in healthy subjects demonstrated that during expiratory loaded breathing at rest, increased respiratory rate results in paradoxical adaptations as DH and dyspnea happen without negative change neither in blood perfusion nor in oxygenation variables in intercostal muscles. Thus, shift in tidal breathing to higher lung volume at rest probably does not induce mismatch between oxygen need and supply in intercostal muscles.

347 ACKNOWLEDGEMENTS

348 This study was supported in part by the European Union and the New Aquitaine region

through the Habisan program (CPER-FEDER).

350

351 DECLARATION OF INTEREST

352 None.

353

354 AUTHORS' CONTRIBUTIONS

All authors have made substantial contributions to all of the following: (1) the conception and design of the study, or acquisition of data, or analysis and interpretation of data, (2) drafting the article or revising it critically for important intellectual content, (3) final approval of the version to be submitted.

359

360 FUNDING

361 This research did not receive any specific grant from funding agencies in the public,

362 commercial, or not-for-profit sectors.

REFERENCES

364	Athanasopoulos, D., Louvaris, Z., Cherouveim, E., Andrianopoulos, V., Roussos, C.,
365	Zakynthinos, S., Vogiatzis, I., 2010. Expiratory muscle loading increases
366	intercostal muscle blood flow during leg exercise in healthy humans. J. Appl.
367	Physiol. 109, 388-395. https://doi.org/10.1152/japplphysiol.01290.2009
368	Barrett, J., Cerny, F., Hirsch, J.A., Bishop, B., 1994. Control of breathing patterns and
369	abdominal muscles during graded loads and tilt. J. Appl. Physiol. 76, 2473–
370	2480. https://doi.org/10.1152/jappl.1994.76.6.2473
371	Barstow, T.J., 2019. Understanding near infrared spectroscopy and its application to
372	skeletal muscle research. J. Appl. Physiol. 126, 1360–1376.
373	https://doi.org/10.1152/japplphysiol.00166.2018
374	Bougault, V., Lonsdorfer-Wolf, E., Charloux, A., Richard, R., Geny, B., Oswald-
375	Mammosser, M., 2005. Does thoracic bioimpedance accurately determine
376	cardiac output in COPD patients during maximal or intermittent exercise? Chest
377	127, 1122–1131. https://doi.org/10.1378/chest.127.4.1122
378	Braun, N.M., Arora, N.S., Rochester, D.F., 1982. Force-length relationship of the
379	normal human diaphragm. J. Appl. Physiol. 53, 405–412.
380	https://doi.org/10.1152/jappl.1982.53.2.405
381	Bretonneau, Q., Pichon, A., de Bisschop, C., 2020. Effect of expiratory loaded
382	breathing during moderate exercise on intercostal muscle oxygenation.
383	Multidiscip. Respir. Med. 15. https://doi.org/10.4081/mrm.2020.702
384	Bretonneau, Q., Pichon, A., de Bisschop, C., 2019. Intercostal muscle oxygenation
385	during expiratory load breathing at rest. Respir. Physiol. Neurobiol. 261, 24–30.
386	https://doi.org/10.1016/j.resp.2018.12.007
387	Chance, B., Dait, M.T., Zhang, C., Hamaoka, T., Hagerman, F., 1992. Recovery from
388	exercise-induced desaturation in the quadriceps muscles of elite competitive
389	rowers. Am. J. PhysiolCell Physiol. 262, 766–775.
390	https://doi.org/10.1152/ajpcell.1992.262.3.C766
391	de Bisschop, C., Beloka, S., Groepenhoff, H., van der Plas, M.N., Overbeek, M.J.,
392	Naeije, R., Guenard, H., 2014. Is there a competition for oxygen availability
393	between respiratory and limb muscles? Respir. Physiol. Neurobiol. 196, 8–16.
394	https://doi.org/10.1016/j.resp.2014.02.011
395	de Bisschop, C., Montaudon, M., Glénet, S., Guénard, H., 2017. Feasibility of
396	intercostal blood flow measurement by echo-Doppler technique in healthy
397	subjects. Clin. Physiol. Funct. Imaging 37, 282–287.
398	https://doi.org/10.1111/cpf.12298
399	De Troyer, A., Gorman, R.B., Gandevia, S.C., 2003. Distribution of inspiratory drive to
400	the external intercostal muscles in humans. J. Physiol. 546, 943–954.
401	https://doi.org/10.1113/jphysiol.2002.028696
402	Erram, J., Bari, M., Domingo, A., Cannon, D.T., 2021. Pulmonary function with
403	expiratory resistive loading in healthy volunteers. PLOS ONE 16, e0252916.
404	https://doi.org/10.1371/journal.pone.0252916
405	Ferrari, M., Muthalib, M., Quaresima, V., 2011. The use of near-infrared spectroscopy
406	in understanding skeletal muscle physiology: recent developments. Philos.
407	Trans. R. Soc. Math. Phys. Eng. Sci. 369, 4577–4590.
408	https://doi.org/10.1098/rsta.2011.0230

409	Gelb, A.F., Gutierrez, C.A., Weisman, I.M., Newsom, R., Taylor, C.F., Zamel, N.,
410	2004. Simplified detection of dynamic hyperinflation. Chest 126, 1855–1860.
411	https://doi.org/10.1378/chest.126.6.1855
412	Grassi, B., Quaresima, V., 2016. Near-infrared spectroscopy and skeletal muscle
413	oxidative function in vivo in health and disease: a review from an exercise
414	physiology perspective. J. Biomed. Opt. 21.
415	https://doi.org/10.1117/1.JBO.21.9.091313
416	Guenette, J.A., Chin, R.C., Cory, J.M., Webb, K.A., O'Donnell, D.E., 2013. Inspiratory
417	capacity during exercise: measurement, analysis, and interpretation. Pulm. Med.
418	2013, 1–13. https://doi.org/10.1155/2013/956081
419	Kendrick, K.R., Baxi, S.C., Smith, R.M., 2000. Usefulness of the modified 0-10 Borg
420	scale in assessing the degree of dyspnea in patients with COPD and asthma. J.
421	Emerg. Nurs. 26, 216–222. https://doi.org/10.1016/S0099-1767(00)90093-X
422	Klooster, K., ten Hacken, N.H.T., Hartman, J.E., Sciurba, F.C., Kerstjens, H.A.M.,
423	Slebos, DJ., 2015. Determining the role of dynamic hyperinflation in patients
424	with severe chronic obstructive pulmonary disease. Respiration 90, 306–313.
425	https://doi.org/10.1159/000439056
426	Lavorini, F., Magni, C., Chellini, E., Camiciottoli, G., Pistolesi, M., Fontana, G.A.,
427	2013. Different respiratory behaviors disclosed by induced bronchoconstriction
428	in mild asthma patients. Respir. Physiol. Neurobiol. 189, 521–529.
429	https://doi.org/10.1016/j.resp.2013.08.013
430	Leenaerts, P., Decramer, M., 1990. Respiratory changes in parasternal intercostal
431	intramuscular pressure. J. Appl. Physiol. 68, 868–875.
432	https://doi.org/10.1152/jappl.1990.68.3.868
433	Lougheed, M.D., Lam, M., Forkert, L., Webb, K.A., O'Donnell, D.E., 1993.
434	Breathlessness during acute bronchoconstriction in asthma: pathophysiologic
435	mechanisms. Am. Rev. Respir. Dis. 148, 1452–1459.
436	https://doi.org/10.1164/ajrccm/148.6_Pt_1.1452
437	Louvaris, Z., Habazettl, H., Wagner, H., Wagner, P., Zakynthinos, S., Vogiatzis, I.,
438	2016. Improvement in respiratory muscle O2 delivery by heliox or pure O2
439	breathing is associated with reduced dyspnea sensations during exercise in
440	COPD, in: 4.1 Clinical Physiology, Exercise and Functional Imaging. Presented
441	at the ERS International Congress 2016 abstracts, European Respiratory Society,
442	p. OA2012. https://doi.org/10.1183/13993003.congress-2016.OA2012
443	Louvaris, Z., Habazettl, H., Wagner, H., Zakynthinos, S., Wagner, P., Vogiatzis, I.,
444	2018. Near-infrared spectroscopy using indocyanine green dye for minimally
445	invasive measurement of respiratory and leg muscle blood flow in patients with
446	COPD. J. Appl. Physiol. 125, 947–959.
447	https://doi.org/10.1152/japplphysiol.00959.2017
448	Louvaris, Z., Spetsioti, S., Andrianopoulos, V., Chynkiamis, N., Habazettl, H., Wagner,
449	H., Zakynthinos, S., Wagner, P.D., Vogiatzis, I., 2019. Cardiac output
450	measurement during exercise in COPD: A comparison of dye dilution and
451	impedance cardiography. Clin. Respir. J. 13, 222–231.
452	https://doi.org/10.1111/crj.13002
453	Mancini, D.M., Henson, D., LaManca, J., Levine, S., 1992. Respiratory muscle function
454	and dyspnea in patients with chronic congestive heart failure. Circulation 86,
455	909–918. https://doi.org/10.1161/01.CIR.86.3.909

456	Miller, M.R., Hankinson, J., Brusasco, V., Burgos, F., Casaburi, R., Coates, A., Crapo,
457	R., Enright, P., van der Grinten, C.P.M., Gustafsson, P., Jensen, R., Johnson,
458	D.C., MacIntyre, N., McKay, R., Navajas, D., Pedersen, O.F., Pellegrino, R.,
459	Viegi, G., Wanger, J., ATS/ERS Task Force, 2005. Standardisation of
460	spirometry. Eur. Respir. J. 26, 319–338.
461	https://doi.org/10.1183/09031936.05.00034805
462	O'Donnell, D.E., Bertley, J.C., Chau, L.K., Webb, K.A., 1997. Qualitative aspects of
463	exertional breathlessness in chronic airflow limitation: pathophysiologic
464	mechanisms. Am. J. Respir. Crit. Care Med. 155, 109–115.
465	https://doi.org/10.1164/ajrccm.155.1.9001298
466	Poole, D.C., Musch, T.I., Kindig, C.A., 1997. In vivo microvascular structural and
467	functional consequences of muscle length changes. Am. J. PhysiolHeart Circ.
468	Physiol. 272, 2107–2114. https://doi.org/10.1152/ajpheart.1997.272.5.H2107
469	Quanjer, P.H., Stanojevic, S., Cole, T.J., Baur, X., Hall, G.L., Culver, B.H., Enright,
470	P.L., Hankinson, J.L., Ip, M.S.M., Zheng, J., Stocks, J., the ERS Global Lung
471	Function Initiative, 2012. Multi-ethnic reference values for spirometry for the 3–
472	95-yr age range: the global lung function 2012 equations. Eur. Respir. J. 40,
473	1324–1343. https://doi.org/10.1183/09031936.00080312
474	Sears, T., Newsom Davis, J., 1968. The control of respiratory muscles during voluntary
475	breathing. Ann. N. Y. Acad. Sci. 155, 183–190. https://doi.org/10.1111/j.1749-
476	6632.1968.tb56762.x
477	Stark-Leyva, K.N., Beck, K.C., Johnson, B.D., 2004. Influence of expiratory loading
478	and hyperinflation on cardiac output during exercise. J. Appl. Physiol. 96, 1920-
479	1927. https://doi.org/10.1152/japplphysiol.00756.2003
480	Suzuki, S., Suzuki, J., Okubo, T., 1991. Expiratory muscle fatigue in normal subjects. J.
481	Appl. Physiol. 70, 2632–2639. https://doi.org/10.1152/jappl.1991.70.6.2632
482	van Beekvelt, M.C.P., van Engelen, B.G.M., Wevers, R.A., Colier, W.N.J.M., 2002. In
483	vivo quantitative near-infrared spectroscopy in skeletal muscle during
484	incremental isometric handgrip exercise. Clin. Physiol. Funct. Imaging 22, 210-
485	217. https://doi.org/10.1046/j.1475-097X.2002.00420.x
486	Verges, S., Bachasson, D., Wuyam, B., 2010. Effect of acute hypoxia on respiratory
487	muscle fatigue in healthy humans. Respir. Res. 11. https://doi.org/10.1186/1465-
488	9921-11-109
489	Vogiatzis, I., Athanasopoulos, D., Habazettl, H., Aliverti, A., Louvaris, Z., Cherouveim,
490	E., Wagner, H., Roussos, C., Wagner, P.D., Zakynthinos, S., 2010. Intercostal
491	muscle blood flow limitation during exercise in chronic obstructive pulmonary
492	disease. Am. J. Respir. Crit. Care Med. 182, 1105–1113.
493	https://doi.org/10.1164/rccm.201002-0172OC
494	Vogiatzis, I., Habazettl, H., Aliverti, A., Athanasopoulos, D., Louvaris, Z., LoMauro,
495	A., Wagner, H., Roussos, C., Wagner, P.D., Zakynthinos, S., 2011. Effect of
496	helium breathing on intercostal and quadriceps muscle blood flow during
497	exercise in COPD patients. Am. J. PhysiolRegul. Integr. Comp. Physiol. 300,
498	1549–1559. https://doi.org/10.1152/ajpregu.00671.2010
499	Wilson, T.A., Legrand, A., Gevenois, PA., De Troyer, A., 2001. Respiratory effects of
500	the external and internal intercostal muscles in humans. J. Physiol. 530, 319-
501	330. https://doi.org/10.1111/j.1469-7793.2001.0319l.x

502	Xiao, SC., Lu, YR., Guo, HX., Qiu, ZH., Luo, YM., 2012. Effect of expiratory
503	load on neural inspiratory drive. Chin. Med. J. (Engl.) 125, 3629-3634.
504	https://doi.org/10.3760/cma.j.issn.0366-6999.2012.20.006
505	Yan, S., Kaminski, D., Sliwinski, P., 1997. Reliability of inspiratory capacity for
506	estimating end-expiratory lung volume changes during exercise in patients with
507	chronic obstructive pulmonary disease. Am. J. Respir. Crit. Care Med. 156, 55-
508	59. https://doi.org/10.1164/ajrccm.156.1.9608113

510 FIGURE LEGENDS

Figure 1. Study design and time measurements. With AP, arterial pressure; Ctrl, control;
ETL, expiratory threshold load; IC inspiratory capacity; PEmax, maximal expiratory
pressure and PImax, maximal inspiratory pressure.

Figure 2. Respiratory times and ventilatory flows during control (Ctrl) and expiratory threshold load (ETL) periods. In $_{ETL}F^-$ condition, the breathing frequency should be decreased as soon as the ETL was applied (panel A). In $_{ETL}F^+$ condition, the breathing frequency should be increased (panel B). During ETL periods, data were analysed at the 10th and the 20th minute ($_{ETL}10'$ and $_{ETL}20'$). Values are mean ± SD. With Te, expiratory time; Ti, inspiratory time; Ttot, total respiratory time; Vt/Te, spontaneous expiratory flow and Vt/Ti, spontaneous inspiratory flow.

Figure 3. Inspiratory capacity (IC, panel A) and dyspnea (panel B) during control (Ctrl) 521 and expiratory threshold load (ETL) periods. In ETLF condition, the breathing 522 frequency should be decreased as soon as the ETL was applied. In $ETLF^+$ condition, the 523 breathing frequency should be increased. During ETL periods, data were analysed at the 524 10^{th} and the 20^{th} minute (ETL10' and ETL20'). Values are mean ± SD. ^{\$}, difference 525 between Ctrl and ETL10' regardless of condition or [#], for the corresponding condition (p 526 < 0.05). [†], difference between $_{ETL}10'$ and $_{ETL}20'$ for the corresponding condition (p < 527 0.05).^{*}, difference between conditions during ETL period (p < 0.05). 528

Figure 4. Intercostal muscle oxygenation during control (Ctrl) and expiratory threshold load (ETL) periods. In $_{ETL}F^-$ condition, the breathing frequency should be decreased as soon as the ETL was applied. In $_{ETL}F^+$ condition, the breathing frequency should be

increased. During ETL periods, data were analysed at the 10th and the 20th minute 532 (ETL10' and ETL20'). Values are mean \pm SD. Tissue saturation index (TSI, panel A). 533 oxyhaemoglobin (Δ 534 Concentration changes in oxy[Hb+Mb], panel B), deoxyhaemoglobin (Δ deoxy[Hb+Mb], panel C), and total haemoglobin (Δ 535 total[Hb+Mb], panel D).[#], difference between Ctrl and _{ETL}10' for the corresponding 536 condition (p < 0.05). \ddagger , difference between _{ETL}10' and _{ETL}20' (p < 0.05). \ast , difference 537 between conditions during ETL period (p < 0.05). 538

539 TABLES

_	Absolute values			Predicted values (%	Predicted values (%)			
BMI (kg/m²)	23	±	2	-				
Body fat mass (%)	14	±	4	-				
Left 7 th IS ATT (mm)	4.7	±	2.0	-				
ERV (L)	1.5	±	0.4	-				
IRV (L)	3.1	±	0.4	-				
IC (L)	3.8	±	0.3	-				
SVC (L)	5.3	±	0.4	99 ±	7			
FEV1 (L)	4.6	±	0.5	112 ±	5			
FVC (L)	5.3	±	0.5	113 ±	6			
FEV1/FVC (%)	86	±	4	99 ±	5			
MMEF 25-75% (L/sec)	5.0	±	0.9	109 ±	15			

540 **Table 1.** Anthropometric and pulmonary function characteristics.

Values are mean ± SD. With BMI, body mass index; ERV, expiratory reserve volume;
FEV1, forced expiratory volume during the first second; FVC, forced vital capacity; IC,
inspiratory capacity; IRV, inspiratory reserve volume; IS ATT, intercostal space
adipose tissue thickness; MMEF, mean median expiratory flow between 25% and 75%
of FVC and SVC, slow vital capacity.

Table 2. Respiratory, metabolic and cardiac variables during control period (Ctrl) and with expiratory threshold load when the breathing frequency was decreased ($_{ETL}F^{-}$) or increased ($_{ETL}F^{+}$).

	$_{ m ETL}{f F}^-$				$_{ m ETL}{f F}$ +		Difference between	
	Ctrl	ETL 10'	ETL 20'	Ctrl -	ETL 10'	ETL 20'	Ctrl and _{ETL} 10'	ETL 10' and ETL 20'
Ventilatory variables								
Vt (L)	0.7 (0.1)	1.7 (0.3)	1.8 (0.3)	0.7 (0.1)	0.8 (0.1)	0.8 (0.1)*	#	
ΫΕ (L/min)	11.8 (2.4)	17.4 (2.6)	18.8 (2.4)	11.8 (2.0)	17.6 (2.4)	17.9 (2.5)	\$	
Ti/Ttot (%)	41.6 (4.1)	50.0 (7.3)	48.9 (6.8)	41.8 (4.3)	51.6 (4.7)	51.3 (2.5)	\$	
Metabolic variables								
^{VO} ₂ (ml/min/kg)	0.38 (0.06)	0.35 (0.06)	0.38 (0.05)	0.36 (0.05)	0.39 (0.04)	0.38 (0.04)		
VCO ₂ (ml/min/kg)	0.32 (0.05)	0.36 (0.06)	0.38 (0.05)	0.31 (0.03)	0.35 (0.04)	0.35 (0.03)		
Gas exchanges								
RER	0.84 (0.05)	1.04 (0.08)	0.99 (0.06)	0.86 (0.07)	0.91 (0.07)	0.91 (0.07)*	#	
V̈Ε/V̈O ₂	19.5 (2.2)	43.2 (8.8)	43.0 (6.0)	20.3 (4.4)	30.6 (4.8)	31.5 (5.4)*	\$	
[.] VE/VCO ₂	23.2 (2.7)	41.4 (6.1)	43.4 (5.2)	23.5 (3.4)	33.7 (4.1)	34.5 (4.5)*	\$	
PetCO ₂ (mmHg)	39.6 (2.4)	26.5 (2.9)	24.7 (2.2)	39.1 (3.5)	31.7 (3.3)	30.7 (3.8)*	\$	‡
Cardiac variables								
HR (bpm)	74.0 (10.5)	83.4 (8.7)	80.2 (9.0)	73.6 (10.9)	83.0 (8.6)	80.7 (7.8)	\$	‡
SV (ml)	75.0 (17.5)	68.6 (14.9)	69.9 (15.0)	83.5 (8.7)	78.2 (11.3)	72.9 (12.5)	\$	
Qc (L/min)	5.5 (1.1)	5.7 (1.2)	5.6 (1.4)	6.1 (0.6)	6.5 (1.0)	5.9 (1.1)		

Values are mean (SD). During ETL periods, data were analysed at the 10th and the 20th minute ($_{ETL}10'$ and $_{ETL}20'$). With HR, heart rate; PetCO₂, end-tidal CO₂ pressure; RER, respiratory exchange ratio; SV, stroke volume; Ti/Ttot, inspiratory duty cycle; Vt, tidal volume; Qc, cardiac output; VCO₂, carbon dioxide exhalation per unit of time; VE, ventilatory flow; VO₂, oxygen consumption per unit of time; VE/VCO₂, ventilatory equivalent for CO₂ and VE/VO₂, ventilatory equivalent for O₂. \$, difference between Ctrl and $_{ETL}10'$ regardless of condition (p < 0.05). #, difference between Ctrl and $_{ETL}20'$ regardless of condition (p < 0.05). *, difference between conditions during ETL period, regardless of time (p < 0.05).



Continuous measurement:

- Intercostal muscle oxygenation
- Ventilatory, metabolic and cardiac parameters
- Mean of continuous measurement (30 sec)





