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Acquiring social safety engages oxytocin neurons in the supraoptic nucleus – role of *Magel2* deficiency

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Abstract

Introduction

Exposure to social trauma may alter engagement with both fear-related and unrelated social stimuli long after. Intriguingly, how simultaneous discrimination of social fear and safety is affected in neurodevelopmental conditions remains underexplored. The role of the neuropeptide oxytocin is established in social behaviors, and yet unexplored during such a challenge post-social trauma.

Methods

Using *Magel2* knockout mice, an animal model of Prader Willi Syndrome (PWS) and Schaaf-Yang Syndrome (SYS), we tested memory of social fear and safety after a modified social fear conditioning task. Additionally, we tracked the activity of oxytocin neurons in the supraoptic (SON) and paraventricular (PVN) nuclei of the hypothalamus by fibre photometry, as animals were simultaneously presented with a choice between a fear and safe social cue during recall.

Results

Male *Magel2* KO mice trained to fear females with electrical footshocks avoided both unfamiliar females and males during recalls, lasting even a week post-conditioning. On the contrary, trained *Magel2* WT avoided only females during recalls, lasting days rather than a week post-conditioning. Inability to overcome social fear and avoidance of social safety in *Magel2* KO mice were associated with reduced engagement of oxytocin neurons in the SON, but not the PVN.

Conclusion

In a preclinical model of PWS/SYS, we demonstrated region-specific deficit in oxytocin neuron activity associated with behavioral generalization of social fear to social safety. Insights from this study add to our understanding of oxytocin action in the brain at the intersection of social trauma and PWS/SYS.

Introduction

Our day-to-day social interactions are complex, requiring discrimination of not only information about the interacting conspecific (age, sex, social status, and so on) but each encounter may also carry a different implicit valence (threat versus mate). Furthermore, several social cues may be present at the same instant (degree of familiarity versus novelty), making such discrimination even more challenging. While neurotypical individuals can appropriately respond to such situations, responses to implicit and explicit social cues are severely compromised in many neurodevelopmental diseases associated with a high penetrance of autism spectrum disorders like the Prader-Willi syndrome (PWS) and Schaaf-Yang syndrome (SYS). PWS has been genetically attributed to a lack of expression of 15q11.2-q13 region containing MAGEL2 gene - due to paternal deletion or imprinting, or maternal uniparental disomy (1). MAGEL2 deficiency is also associated with other disorders like Schaaf-Yang syndrome (SYS), which is linked to >75% risk of developing comorbid autistic spectrum disorders (2). Apart from well-documented developmental deficits, PWS and SYS patients also show a high comorbidity with autism symptoms (3,4) including decreased social responsiveness in non-threatening contexts (5), increased temper outbursts (6–8) and social fear (9). Recent publications have established a link between ASD and PWS/SYS, such that ASDprevalence in MAGEL2-related diseases (25%-84%, varying due to age and ethnic background) sits significantly high above the prevalence in general population (~1.5%) (1,3,4,10). Such a high overlap of ASD related to MAGEL2 deficiency makes its mouse and rat models bridge the gap between ASD and PWS/SYS (11). Remarkably, most of these clinical symptoms are wellreplicated in Magel2 knockout (KO) mice, which report impaired response to social novelty following social habituation (12,13) as well as high social fear that is difficult to extinguish (14).

It is interesting to note that *MAGEL2*-deficient patient-derived cell lines express deficient levels of the social neuropeptide oxytocin (OXT) and vasopressin (AVP) (15). Indeed, clinically supplementing OXT by intranasal delivery significantly improves social trust in PWS subjects (16). In *Magel2* KO mice too, OXT treatment early in life prevents social interaction deficits later (12), while AVP can also correct impaired responses to social-novelty in *Magel2* KO adults (13). Remarkably, recruitment of the specific source of OXT and AVP in the brain may be cued to behavioral experience. For instance, OXT neurons in the supraoptic nucleus (SON), but not in the paraventricular nucleus (PVN), are specifically activated during social fear extinction (14). While we have recently reported that impairment in the SON-lateral septum OXT pathway underlies social fear extinction deficits in *Magel2* KO (14), how the SON neurons *themselves* respond during social fear recall remains unexplored. Intriguingly, common social assays in rodents test response towards a single social cue at one instant. Everyday interactions are complicated by the multiplicity of social cues converging at once, and therefore how *Magel2* KO mice may react to a more ethologically-relevant social challenge remains to be seen.

To address these questions, we designed a novel strategy to condition mice to fear a female, but not a male mouse. We then tested a generalized fear response over days, when unfamiliar males and females were co-presented in a neutral context. Additionally, we tracked the activity of OXT neurons by fiber photometry during these sessions. Our present study was guided by a recent report (14) where we demonstrated that a) OXT neurons in the SON are specifically activated during extinction of social fear, b) *Magel2* KO mice fail to extinguish social fear and c) this failure is coupled with a reduced activation of SON OXT neurons (inferred from Fosmapping). These set of observations gave rise to the following hypotheses and predictions: a)

WT mice capable of social fear extinction, should be able to discriminate between social-safe and social-fear cues after conditioning, b) this should be accompanied by a concurrent change in $OXT^{SON} Ca^{2+}$ dynamics *in vivo*, c) *Magel2* KO mice should show an impairment in correctly interpreting social salience, which would be tracked by d) an insufficient engagement of OXT^{SON} neurons in the mutants. Together, we believe that observations from this study add new insights into the role of OXT in mediating social behavior within the context of PWS.

Methods

Animals. Group-housed animals kept under standard pathogen-free conditions (12/12 light/dark cycle, food and water *ad libitum*) were used in this study. Male *Magel2* KO mice (*Magel2*tm^{1.1Mus}/J) alone, or crossed with *Oxt*-Cre (Oxt^{tm1.1}(CRE)^{Dolsn}/J from Jackson labs and were used in the study. *Magel2* gene is paternally expressed (p) but maternally (m) imprinted, and thus heterozygotes with the paternal null allele (-p) were designated as knockouts (KO). Across experiments, age- and weight-matched heterozygote *Magel2*+m/-p mice were designated as knockouts and *Magel2*+m/+p mice as WT. For stimulus mice, age-matched, non-littermate B6J WT mice were used. *Magel2*KO mice were obtained from F. Muscatelli (INMED, Marseille).

Stereotaxic injection of viral vectors. 6-week old double-transgenic *Magel2* KO or WT mice crossed with *Oxt*-Cre were unilaterally injected with 500 nL AAV2/5 pGC-CAG-FLEX-jGCaMP7s-WPRE::hGH (CERVO, Canada) in either the SON (AP -0.056 cm, ML +/-0.113 cm, DV -0.545 cm) or PVN (AP -0.9cm, ML +/-0.2 cm, DV 0.45 cm). Injections were performed with microinjector-controlled (micro-4, WPI) syringe (nanofil syringe and Nanofil 33G BVLD Needle, WPI). We observed successful recombination leading to expression of GCaMP in 75% OXT neurons (GFP antibody, 1:3000, Abcam, ab13970; NPI-oxytocin antibody, 1:1000, gift from H. Gainer at the National Institute of Health).

Fibre Photometry. Optic fibre was implanted in either the SON (AP -0.056 cm, ML +/-0.113 cm, DV -0.545 cm) or PVN (AP -0.9cm, ML +/-0.2 cm, DV 0.45 cm). Monofibre optic ferrule (MFC 400/430-0.66 5mm MF1.25 FLT, DORIC lenses, Quebec, CA) were fixed with dental cement (Paladur, Henry Schein) and animals were given anti-inflammatory medication (meloxicam, 10 mg/kg) and monitored daily in post-surgery isolation for 3 weeks. Calcium (Ca^{2+}) signals were explored in freely-moving animals by connecting the implanted fiber optic ferrule to a low-autofluorescence fiber optic patchcord connected to a rotary joint. For photostimulation, connecterized LEDs (CLEDs) emitting sinusoidal signals (GCaMP, 465 nm, and isosbestic, 405 nm) were connected to a fluorescence minicube (consisting of integrated photodetector heads) by a fiber optic attenuating patchcord. Fluorescence acquired through the minicube was digitized and converted by the fiber photometry console to voltage signals, and processed with a low-pass filter (12 Hz). Behavioural data was acquired simultaneously using DORIC Neuroscience studio (Doric Lenses, Quebec, Canada). Isosbestic and calcium signals were demodulated in real time with lock-in mode of acquisition. Detection of Ca²⁺ spikes was done using PMATv1-3 setting a threshold of +2.5 SD (17), and custom-written MATLAB code based on (17) were used to calculate positive area under the curve. For this, loss-of-signal artifacts were corrected, followed by smoothing using loess and linearizing using a moving minima algorithm. Next, GCaMP was fit to the isosbestic by using linear regression. The fitted isosbestic was then used to calculate the $\Delta F/F$, by subtracting the fitted isosbestic from the fitted GCaMP, followed by dividing by the fitted isosbestic, and computing a z-score. Behavioral bouts were scored offline during social interactions using BORIS (18).

Social-fear conditioning. 4 weeks after surgery, all animals were handled for a minimum of 3 days, prior to the beginning of the experiment. A separate cohort of animals were also tested without any surgical implant. For baseline test (day -1, Fig. 1A), the male test subject was put in the experimental arena (22x30x44cm) to interact during 3 minutes with an unfamiliar male and an unfamiliar female, presented together. Social cues were placed in two diametric ends of the arena in chambers with grills that allowed active physical interaction. This would reveal baseline differences in behaviour across genotype, about any preferential bias towards either stimuli-sex, before animals were selectively conditioned to fear female mice (Fig. 1A, day -1). 24 hrs later (Fig. 1A, day 0), the test mouse was put in the conditioning arena (11x15x44cm, cleaned with Anios detergent) to freely interact with a novel male stimulus presented alone (M1), during 5 minutes. Next, they underwent social fear conditioning to an unfamiliar female (F1), such that a scrambled electric footshock (0.5mA, 2 sec) was delivered upon each interaction with the female stimuli (Fig. 1A, day 0). The session was terminated after 5 minutes, or if no further social encounter occurred in the 3 minutes following the last shock. This is to ensure that the conditioned mouse remains at least 3 mins in the fearful context after the last trauma. Finally, we tested the memory of social fear/safety over time wherein new sets of unfamiliar male and female mice (M2 and F2) were simultaneously presented in a neutral context on several days after conditioning (Fig. 1E). Using new stimulus animals would allow us to test a generalized response to unfamiliar social stimulus, which would not be limited to stimulus mice used during conditioning. During recall on days 1, 2 and 7, the test mouse was introduced into a novel arena (22x30x44cm, patterned walls, different light, cleaned with 10% ethanol) for 5 mins, to allow interaction with a novel male (M2) and a novel female (F2) kept at diametrically opposite corners. All arenas were carefully cleaned between successive animals, and recorded videos were analysed offline with BORIS (18). Heatmaps depicting spatial localization of test subjects in the arena were generated using ezTrack (19). Experiments were run by different researchers, in batches of double transgenic animals without surgeries (data in Figure 1) or with surgical implantation of a viral vector and optic fiber (data in Figures 2-5). Analyses were also performed by different researchers to foster replication.

cFos mapping. On day 7, brain samples were collected 60 minutes after recall via transcardiac perfusion of ice-cold 0.9% NaCl and 4% PFA in deeply anesthetized mice (pentobarbital 50 mg/kg). 40 μm coronal sections was collected with a vibratome, and non-specific sites were blocked prior to immunostaining (non-labeled Fab anti mouse IgG (abliance, 1:500) with 3% normal donkey serum in PBS, 0.1% triton X-100). Primary antibodies (c-Fos: Cell Signaling, mAb 2250, NPI-oxytocin, mouse monoclonal antibodies (H. Gainer, NIH)) were incubated on brain sections for 48 hrs at 4°C, and Alexa-Fluor-conjugated secondary antibodies (1:2,000 ThermoFisher Scientific) were incubated for 2 hrs at 25°C. Sections were mounted with fluoromount and stored at 4°C (ThermoFisher Scientific). Stacks were acquired with an epifluorescence microscope (Imager Z1, Carl Zeiss), and the overlap between cFos and OXT cells counted with FIJI by an experimentalist blind to the groups.

Statistics. Prism 9 (GraphPad, La Jolla, CA) was used for statistical analysis. Photometry data were plotted by events. Plotting of fiber photometry data by number of events has been previously used by several studies exploring oxytocin function (20–23). Normally distributed data were compared with two-way ANOVA, mixed-effects analysis, or unpaired, two-tailed t-test, as applicable. Post-hoc Sidak's test was conducted as necessary. Non-normal datasets were compared using Mann Whitney U-test for two-tailed unpaired comparisons. Cohen's d was calculated as an indicator of effect size between groups showing significant difference of the mean according to p values. Therefore, p values describe significant differences between group

comparisons with α =0.05 while *d* values precise the effect size of these comparisons according to the sample size. All data presented are as means ±SEM, N as indicated in figures and statistical tests as indicated in the legends and significance reported as follow: **p*<0.05, ***p*<0.01, ****p*<0.001, ****p*<0.001. Details of statistical tests are enlisted in Supplementary Table 1.

Results

Social fear conditioning with females generalizes to avoidance of males in *Magel2* KO mice Studies on social fear in the past have routinely tested fear memory in response to a single social cue. Here, we designed a new task to integrate social fear and safety association based on stimuli-sex (Fig. 1A). This created an associative memory that was cued to stimuli-sex, wherein unfamiliar females indicated social fear unlike the unfamiliar male mice that indicated social safety. 24 hours before conditioning, both Magel2 KO and Magel2 WT mice explored simultaneously the presented male and female stimuli to a similar extent (two-way ANOVA: Social stimuli sex, $F_{(1,32)} = 0.8080$, p=0.38, Genotype, $F_{(2,32)} = 0.4350$, p=0.98, Interaction, $F_{(2,32)} = 0.4350$, p=0.98, Interaction, $F_{(2,32)} = 0.4350$, p=0.98, $F_{(2,32)} = 0.4350$, $F_{(2,32)} = 0.4350$ $_{32)} = 0.4441$, p=0.65), indicating no baseline behavioural deficits in absence of shock (Fig. 1B). However, both Magel2 WT and KO animals investigated the socially fear conditioned femalestimulus less at conditioning, with a significant effect of genotype (two-way ANOVA: $F_{(2,25)} =$ 53.69, *p*<0.0001), sex of the social stimuli ($F_{(1, 25)} = 16.71$, *p*<0.001) and interaction between the two (Interaction, $F_{(2, 25)} = 21.01$, p < 0.0001) (Fig. 1C), during when they also spent more time in the corner away from the social stimulus (Fig. S1). Cohen's d analysis indicated large effect size of conditioning in both genotypes (conditioned WT: d=3.541, conditioned KO: d=5.397). Additionally, an equivalent number of shocks were delivered to both groups (twosided unpaired t-test, t=0.4605, df=15, p=0.65) (Fig. 1D), suggesting no difference in shocksensitivity due to genotype.

Next, we tested the recall of this memory, such that animals were presented with novel male and female stimuli mice, at the same time, following conditioning. We predicted that wild-type mice should avoid the social-fear cue (female) at early time-points, but should show a gradual extinction and increased social approach over time. Investigation of the male stimulus, which indicates social-safety in comparison, should remain unimpaired. Magel2 WT animals explored the female cue significantly less at days 1 and 2, as compared to unshocked controls (Fig 1E). And, 7 days later, however, female-investigation significantly improved from that at day 1 (Fig. 1E). This is in striking contrast to Magel2 KO animals that robustly avoided the females even at 7 d, suggesting a stronger memory of social fear (Fig. 1E). Comparison across groups showed a significant effect of both conditioning (two-way ANOVA: $F_{(2,26)} = 21.15$, p < 0.0001) and time $(F_{(1,430, 36,47)} = 4.235, p < 0.05)$ that did not interact $(F_{(4,51)} = 1.268, p = 0.29)$. Cohen's d analysis indicated large effect size of a fear memory trace in both genotypes (d1, Con/conditioned WT, d= 2.94, CON/conditioned KO, d= 3.135, conditioned WT/conditioned KO, d= 0.377; d2, Con/conditioned WT, d=1.918, CON/conditioned KO, d=6.273, conditioned WT/conditioned KO, d= 0.553; d7, Con/conditioned WT, d= 1.235, CON/conditioned KO, d= 3.424, conditioned WT/conditioned KO, d = 0.804). On the other hand, while interaction of Magel2 WT animals with the male stimulus was not different from unshocked controls (Fig. 1F), Magel2 KO mice remarkably avoided interaction with the males as well at all timepoints postconditioning (Fig. 1F) (two-way ANOVA: Conditioning, $F_{(2, 26)} = 9.440$, p < 0.001, Time, $F_{(1.859)}$ $_{47.40} = 1.859$, p=0.17, Interaction, $F_{(4, 51)} = 1.182$, p=0.33). Cohen's *d* analysis indicated large effect size of a fear generalization in Magel2 KO only (d1, Con/conditioned WT, d=0.782, CON/conditioned KO, d= 2.842, conditioned WT/conditioned KO, d= 1.174; d2, Con/conditioned WT, d = 0.638, CON/conditioned KO, d = 1.867, conditioned WT/conditioned KO, d= 1.071; d7, Con/conditioned WT, d= 0.302, CON/conditioned KO, d= 1.289,

conditioned WT/conditioned KO, d= 1.141). This indicated that learning to fear females extended to avoidance of males in animals with *Magel2* deficiency.

Altered calcium dynamics in SON oxytocin neurons of *Magel2* KO during social fear recall

We aimed to track the activity of SON OXT neurons in vivo - that have been previously implicated in social fear (14) - during recall of social fear memory over days while in presence of a socially safe cue. Thus, we stereotaxically targeted the SON in Oxt-Cre mice crossed with Magel2 KO to inject a Cre-dependent virus expressing GCaMP7s (Fig. 2A), causing successful recombination (Fig. 2B). This allowed us to record calcium-dependent changes in fluorescence from OXT neurons during recall (Fig. 2C). As compared to conditioned WT animals, SON OXT neurons in *Magel2* KO showed more Ca^{2+} spikes over time (Fig. 2D) (two-way ANOVA: Genotype, $F_{(1,5)} = 5.065$, p=0.07, Time, $F_{(1.581, 7.907)} = 3.716$, p=0.08, Interaction, $F_{(2,10)} = 1.919$, p=0.19), post-hoc analysis by Sidak's test revealed a significant increase in spike number in mutant mice 7 d later. Cohen's d analysis indicated large effect size of genotype on the number of spikes (WT/KO, d1, d= 1.513, d2, d= 0.467, d7, d= 3.575). Interestingly, a cumulative frequency analysis of spike duration revealed greater proportion of shorter-duration spikes in Magel2 KO at all time-points (Fig. 2E-2G). Yet, Ca²⁺ spike duration significantly decreased in Magel2 KO animals post-conditioning (Fig. 2H) (Kolmogorov-Smirnov test: d1, D=0.26, p<0.0001, d2, D=0.16, p<0.0001, d7, D=0.26, p<0.0001). A decrease in spike duration was coupled with a decrease in inter-spike interval in Magel2 KO (Fig. 2I-K) (Kolmogorov-Smirnov test: d1, D=0.28, p<0.0001, d2, D=0.21, p<0.0001, d7, D=0.22, p<0.0001). While inter-spike interval increased at d 7 in Magel2 WT, it remained low in mutant animals (Fig. 2L). Together, recall of social fear in Magel2 KO mice changes the overall Ca²⁺ dynamics of SON OXT neurons.

Reduced engagement of SON OXT neurons in *Magel2* KO *during* investigation of male and female stimuli during recall

The above analysis revealed a generalized change in activity at recall that prompted us to investigate activity of the SON OXT neurons when given the choice to interact with a male and a female. To address this, we looked at changes in fluorescence as the mice actively investigated either male or female stimulus mice before conditioning, and during the days of recall post-conditioning (Fig. 3A). At baseline, SON OXT neurons activity was not significantly different between sex-stimuli and genotypes (Mann-Whitney test: U=189, p=0.075) (Fig. 3B1). After conditioning, however, a significant reduction in SON OXT activity in *Magel2* KO was observed later at both 1 d (Mann-Whitney test: U=0, p<0.01) (Fig. 3B2) as well as 2 d (Mann-Whitney test: U=0, p<0.01) (Fig. 3B3), and at 7 d the increase was no longer statistically significant (Mann-Whitney test: U=23, p=0.09) (Fig. 3B4). Cohen's d analysis revealed large effect size before as well as days 1 and 2 after conditioning (d-1, d=0.941, d1, d=0.916, d2, d=0.954), and a moderate effect size 7 days later (d=0.72) (24). Overall, we observed a shutdown of SON OXT neuronal activity following social fear conditioning that increased with more exploration of females with time in *Magel2* WT, which is delayed in *Magel2* KO (Fig. 3C).

During investigation of male-stimuli at baseline, genotypic difference in SON OXT activity was not observed (Mann-Whitney test: U=244, p=0.1) (Fig. 3D1). In both groups, social fear conditioning significantly reduced SON OXT activity 1 d (Mann-Whitney test: U=30, p=0.19) (Fig. 3D2) and 2 d later (Fig. 3D3) (Mann-Whitney test: U=107, p=0.98). However, 7 d later, SON OXT activity in WT animals improved, being significantly higher than *Magel2* KO

(Mann-Whitney test: U=56, p<0.01) (Fig. 3D4). Cohen's *d* analysis showed that the effect of genotype is small up to 2 days after recall (d-1, d=0.275, d1, d=0.558, d2, d=0.375) and large at d7 (d=0.944). This suggested a failure to engaged SON OXT neurons in mutant mice a week after conditioning, in contrast to WT animals (Fig. 3E).

PVN OXT neurons in *Magel2* KO show changes in calcium dynamics but not differential engagement with overcoming social fear

Beyond the SON, OXT in the brain can originate from other structures such as the PVN, where social neuropeptides have been implicated across a wide range of behaviors (25–27). We explored their engagement in the present paradigm, by now targeting the PVN with Credependent GCaMP7s in Magel2 KO;Oxt-Cre mice (Fig. 4A), showing efficient recombination in PVN OXT neurons (Fig. 4B). We next analyzed the overall Ca²⁺ spikes during recall (Fig. 4C). We observed a significant decrease in the number of Ca^{2+} spikes during recall (Fig. 4D) that was due to genotype (two-way ANOVA: Genotype, $F_{(1, 19)} = 32.34$, p<0.0001, Time, $F_{(2, 19)} = 32.34$, p<0.0001, Time, $F_{($ $F_{(2, 19)} = 0.8437$, p=0.45, Interaction, $F_{(2, 19)} = 0.41$, p=0.67). This decrease was significant at all recall timepoints, lasting up to 7d post-conditioning (Fig. 4D). Cohen's d analysis indicated large effect size of genotype on the number of spikes (WT/KO, d1, d= 4.234, d2, d= 2.346, d7, d=1.971). On the other hand, the duration of such spikes increased in *Magel2* KO mice (Fig. 4E). A cumulative frequency analysis revealed a significant right-shift in Magel2 KO animals post-conditioning (Fig. S2A-C). Magel2 KO mice also showed increased ISI during recalls (Fig. 4F), with a significant right-shift of cumulative frequency post-conditioning (Fig. S2D-F). We next addressed whether PVN neurons differently engaged *during* investigation with the social stimuli. Interestingly, there was no difference in PVN OXT neuron activity due to genotype during female-investigation on d 1 and d 2 (Fig. 4G, Fig. S2G, H), but only on d 7 (Mann-Whitney test: U=41, **p<0.01; Cohen's d=1.083) (Fig. S2I). On the other hand, Ca²⁺ dynamics in PVN OXT neurons during male investigation was unchanged in Magel2 KO at all time-points post-conditioning (Fig. 4H, Fig. S2J-L). Together, it suggests that social fear conditioning changes the overall Ca²⁺dynamics in *Magel2* KO mice, and its engagement is dampened only 7 days later selectively during exploration of the fear-associated female, but not the male stimulus mouse.

Inability to overcome social fear is associated with impaired cFos expression in SON of *Magel2* KO mice

Finally, we mapped the OXT activation patterns of SON and PVN 7 d after conditioning (Fig. **5B**), a timepoint where WT animals show a reduction in social fear (Fig. 1E). To that end, we examined the expression of cFos, an immediate early gene produced in response to neuronal activity, in OXT neurons of SON (Fig. **5B**) and PVN (Fig. **5**C) 60 minutes after recall. Interestingly, a significant deficit in activation of SON OXT neurons was seen in fear conditioned *Magel2* KO animals as compared to the conditioned WT mice (Fig. **5D**). A two-way ANOVA showed a significant effect of genotype, with no effect of brain-region ($F_{(1, 24)} = 1.634, p=0.21$) or any interaction between the two (Interaction, $F_{(1, 24)} = 2.808, p=0.17$). Cohen's *d* analysis indicated large effect size of genotype on cFos induction in SON OXT neurons (d=1.813). Such a differential engagement was not observed in the PVN (Fig. **5D**). In non-shocked controls, OXT neurons in both the WT and KO animals showed no divergent activation as well (two-way ANOVA: Genotype, $F_{(1, 14)} = 0.1798, p=0.678$, Brain region, $F_{(1, 14)} = 0.2673, p=0.6132$, Interaction, $F_{(1, 14)} = 0.4131, p=0.5308$) (Fig. **5E**). Thus, activation of OXT neurons that is associated with overcoming social fear in WT is specific to the SON, and such an activation is impaired in the *Magel2* KO model.

Discussion

Using a mouse model of PWS/SYS, the present study examined the activity of OXT neurons as conditioned mice recalled social fear while in the presence of a safe social stimulus. Compared to WT, *Magel2* KO mice had similar response during conditioning and equivalent recall of social fear memory 24 h later, in agreement with our recent observations (14). We add that this memory of social fear is persistent and lasts at least up to 7 days after conditioning. However, while WT animals showed a significant increase in exploring the social fear cue at day 7, *Magel2* KO failed to show such an increase. This is reminiscent of the within-session social extinction deficit seen in *Magel2* KO animals (14). We additionally demonstrate that *Magel2* WT animals remember the association of social safety with a male cue and in remarkable contrast, social fear in *Magel2* KO extends to avoidance of socially safe male stimuli.

It is interesting to note that impaired discrimination of social salience has been seen in PWS/SYS as well. More than 75% of patients with SYS and 25% with PWS have ASD (28,29). These proportions correspond with the frequency of MAGEL2 mutations; causal in the Schaaf-Yang syndrome (2) whereas it is only associated with large chromosomal alterations causing the Prader-Willi disease (30). Despite the genetic heterogeneity between patients, MAGEL2 gene has been a central focus of studies for mechanisms of disease and comorbidities (3). PWS subjects show non-typical responses to identifying facial emotion (31), and have a general inability to correctly interpret social cues (32), including correctly expressing emotion as well (33). Another study assessed both caregiver's response as well as the patients clinically to infer significant impairments in social responsiveness and social functioning in PWS patients (34). Additionally, they also have a significant comorbidity with psychiatric disorders, with one study showing high percentage of disruptive behaviors in males (35). Interestingly, a newly identified genetic variation of Magel2 has showed social fear as well in the studied individual (9). This aligns well with social salience misprocessing seen in ASD - for example, while neurotypical subjects responded faster to a simulated social-hazard, response by autistic subjects was similar towards social and non-social hazards (36). A similar deficit in social salience interpretation may have led to generalization of social fear across stimuli in Magel2deficient animals. Interestingly in humans, presence of another individual improves the learning of social safety (37), while in rats, learning of safety also helps to overcome threat associated with an avoidance task (38). Together, we predict that a) presence of the social safety cue contributes to the delayed response towards overcoming social fear in Magel2 KO mice, such that b) overcoming of social fear is preceded by re-learning of social safety in animals with Magel2 deficiency.

One limitation of the present study was that stimuli-females used were not matched to estrus stage. While our results indicate that the behavioral responses are perhaps generalized to female across estrus, it is interesting to note that female mice only in non-receptive phases of the estrus, but not in the receptive phase, show interest towards a same-sex conspecific (39). In male rats, on the other hand, nasal oxytocin abolishes preference towards virgin estrus female rats by diminishing pERK1/2 levels in the SON (40), suggesting a complex relationship between SON OXT action and estrus-dependent social exploration that remains to be explored further. Additionally, previous mechanistic explorations into social fear have used female mice as test subjects (41). Counterbalancing our present results, therefore, it is necessary to investigate social salience in *Magel2*-deficient females trained to avoid males in a future study.

How is neuronal activity of OXT neurons associated with enhanced social fear in Magel2 KO? To answer this, we recorded activity from OXT neurons in SON and PVN over time in Magel2 KO and WT animals. In Magel2-deficient mice, we observe a shift in Ca²⁺ dynamics leading to an increased number of thinner Ca^{2+} spikes occurring at greater frequency. This is in apparent contrast to previous reports suggesting that reduced OXT signaling contributes to heightened social fear (14,41,42). However, given that OXT receptors in Magel2 KO mice are downregulated (43), it is likely that the increased activity results from homeostatic compensation. A similar compensation is seen in PWS patients - while on one hand they report reduced methylation of OXT-receptor (44) and increased plasma OXT (45), treatment with OXT-receptor agonist improves symptoms of PWS in some clinical studies (46-48) (but not others (49)). Furthermore, spike broadening of OXT neurons as seen here is observed during lactation (50,51), suggesting that reduction of Ca²⁺ spike duration observed in Magel2 KO mice reflects a physiologically relevant deficit. Indeed, when we specifically examined the activity of SON OXT neurons during social investigation, we observed a reduced engagement of these neurons in Magel2 KO mice, aligning well with the behavioral deficits observed in these animals (Fig. 6A). Intriguingly, however, SON OXT neurons in Magel2 KO animals were engaged more during interaction with social cues in the absence of shock (Fig. 6A). Given the positively correlated role of oxytocin in mediating social trust in humans (52), we predict that this aberrant engagement might be necessary to overcome innate threat towards unfamiliar social cues in Magel2 KO, to enable social approach even at baseline. A subsequent exposure to social fear conditioning shuts down the activity of SON OXT neurons in both genotypes (Fig. 6A), until a divergent trajectory emerges over time during recall. As a result, a progressively more engagement of SON OXT neurons in WT was associated with greater exploration of social fear and socially safe stimuli, in contrast to Magel2-deficient animals.

In contrast to SON, cue-specific activity of OXT neurons in the PVN were comparable between WT and Magel2 KO mice at baseline and they shut down after conditioning (Fig. 6B). Exploration of social cues did not significantly engage these neurons at any recall time point (Fig. 6B) but there was an effect of genotype on d 7 during female investigation marking a delay in the Magel2 KO response to the acquisition of social safety. This is in agreement with previous reports that show a selective recruitment of SON but not PVN with social fear extinction (14). Interestingly, a reduction in the number and frequency of Ca^{2+} spikes was observed in PVN OXT neurons of Magel2 KO. This aligns with a previous study reporting decreased firing of PVN OXT neurons in Magel2-deficient animals, due to a selective loss of AMPA-current contributing to increased silent synapses (53). We also observed an increase in Ca^{2+} spike duration at the same time point. Interestingly, magnocellular neurons in the PVN, which release OXT (54), have a greater spike half-width than parvocellular neurons (55). Magnocellular OXT neurons projecting to the central amygdala also modulate expression of non-social fear conditioning (56). While we have previously demonstrated that PVN OXT neurons projecting to the lateral septum are not recruited during social fear extinction (14), exploring the role of magnocellular PVN OXT neurons projecting to other targets like the amygdala in social fear needs to be explored in future.

Consistently, mapping of the immediate early gene cFos confirmed the specific engagement of the SON but not PVN OXT neurons in the present study – suggesting that selective recruitment of neuropeptidergic pathways may facilitate different aspects of social salience. Increased cFos was associated with overcoming of social fear in *Magel2* WT animals 7 days later. It is perhaps intriguing to note that a higher frequency of Ca^{2+} spikes did not lead to more Fos expression in SON OXT neurons of *Magel2* KO. Across brain regions, a complex relationship exists between

neuronal activity and cFos expression. In the granule cells, for example, kindled seizures causing action potentials led to cFos expression, but the same stimulation in the substantia nigra failed to do so (57). In the hippocampal CA2, on the other hand, spiking at 0.1 Hz, but not at 50 Hz, was successful in evoking cFos expression, which indicated participation of the neuron in synchronized synaptic activity (58). Interestingly, cFos expression in magnocellular SON OXT neurons is only seen after synaptic activity induced by an OXT-receptor agonist (59). Investigations in the future should focus on understanding the precise role of synaptic inputs in evoking immediate early gene expression in the SON, its lack in the PVN, and their functional interaction with *Magel2* deficiency.

What could be the targets downstream of SON that mediates these effects? One key functional projection output of SON OXT neurons is the lateral septum (LS) (14,41). Genetic tracing tools have previously revealed functional projections of oxytocin (OXT) neurons between the supraoptic nucleus (SON) and the lateral septum (LS) of female mice (41). We recently extended this observation in males, and also that *Magel2*KO mice lack SON-derived OXT support in the LS which prevents the extinction of social fear conditioning (14). We predict that a similar pathway is involved in social safety discrimination in the present study. SON OXT neurons may also project to different amygdalar nuclei. Activation of SON OXT neurons by secretin, for example, facilitates social recognition by their dense projections to the medial amygdala (60). Hypothalamic OXT neurons also project to the central amygdala, and opto-activation of this pathway attenuates non-social fear (56).

Additionally, SON neurons also project to the pituitary, particularly to its posterior division (61). In the pituitary, the molecule CD38 plays a critical role in OXT function, such that both CD38 knockout mice as well as pharmalogically blocking CD38 in the pituitary impeded release of OXT (62). Association between CD38 polymorphisms and autism spectrum disorders (ASD) has been reported as well, with a subset of such individuals showing lower levels of plasma OXT (63). Additionally, the social buffering effect of OXT on the stress response is mediated by the blockade of ACTH release from the pituitary and its downstream effector cortisol secretion from adrenal glands (64). Prader-Willi patients suffer from central adrenal insufficiency under stressful conditions (65), delayed peak response of cortisol to insulin challenge (66), and small-sized adrenal glands in autopsies (67). But how does cortisol and OXT function interact in PWS/SYS patients? Do PWS/SYS patients show CD38 polymorphisms as well? And in *Magel2*KO animals, is OXT function in the pituitary impaired too? These questions are yet to be explored.

Finally, it is interesting to note that somatodendritic release of OXT creates a long-lasting action of these neuropeptides in relevant circuits. Dendritic release of OXT has been seen in the magnocellular OXT neurons in the SON (68). Would a similar mechanism been utilised in the present study? Seems possible, as calcium dependent, somatodendritic release of OXT has been implicated in regulating social memory (69). This is also dependent on CD38, and associated with ASD (62,69,70). Its precise involvement in overcoming social fear needs future investigation.

Apart from central OXT action, OXT release in the periphery has been well demonstrated in the past. Beyond physiologically-relevant challenges like change in osmotic balance and suckling, stimulation of the SON also causes increase in peripheral OXT levels (71). However, response to emotionally stressful states differ. For instance, rats subjected to social defeat show a sharp increase in OXT levels in the SON, while peripheral levels remain unchanged (72).

Chronic homotypic stress also selectively affects central OXT, sparing peripheral OXT levels (73). Therefore, we predict that OXT neurons in the SON would engage central OXT response to social stress in *Magel2* WT, and less so in the *Magel2*KO model, while peripheral responses have been overlooked in this study.

Finally, based on our observations, we propose a model of divergent OXT activation that maps with social fear/safety deficits in *Magel2* KO mice (Fig. 6C-D). In *Magel2* WT animals, exposure to social fear shuts down OXT activity 24 h later in both the SON and PVN (Fig. 6C). However, overcoming social fear to transition towards social safety selectively engages SON OXT neurons (Fig. 6C). In *Magel2* KO, OXT neurons activity shuts down in both the SON and PVN during the initial phase of social fear recall, similar to WT animals (Fig. 6D). However, a failure to engage SON OXT neurons during the behavioural bout accompanies the long-lasting deficit in miscomprehending social safety and thus, the inability to overcome social fear (Fig. 6D). In sum, our present demonstration of how OXT neuron activity associates with behavioural approach towards socially salient stimuli provides a fundamental insight into the disease pathology of *Magel2* deficiency. We believe that this set of observations can guide future explorations into treatment strategies in PWS, and other psychiatric disorders with impaired interpretation of social salience.

Statements

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Statement of Ethics

Experiments were performed according to the Directive by the Council of the European Communities (86/609/EEC) following guidelines from the French Ministry of research and ethics committee for the care and use of laboratory animals (approved protocols APAFIS-5133, 8940, 11468). All efforts were made to minimize animal suffering and to reduce the number of mice utilized in each series of experiments. All experimental procedures in this study were performed between 8:00 and 15:00-hr according to the ARRIVE guidelines.

Conflict of Interest Statement

The authors declare no conflict of interest.

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Author Contributions

Conceptualization: PC, FJ; Methodology: PC, FJ, Investigations: PC, HL, EA; Photometry analysis code: PF, Funding acquisition: FJ; Project administration and supervision: FJ; Writing–original draft: PC, FJ.

Data Availability Statement

Data will be posted on public repository before publication.

References

- 1. Dykens EM, Roof E, Hunt-Hawkins H, Dankner N, Lee EB, Shivers CM, et al. Diagnoses and characteristics of autism spectrum disorders in children with Prader-Willi syndrome. J Neurodevelop Disord. 2017 Dec;9(1):18.
- 2. Schaaf CP, Gonzalez-Garay ML, Xia F, Potocki L, Gripp KW, Zhang B, et al. Truncating mutations of MAGEL2 cause Prader-Willi phenotypes and autism. Nat Genet. 2013 Nov;45(11):1405–8.
- 3. Fountain M, Schaaf C. Prader-Willi Syndrome and Schaaf-Yang Syndrome: Neurodevelopmental Diseases Intersecting at the MAGEL2 Gene. Diseases. 2016 Jan 13;4(1):2.
- 4. Kong X, Zhu J, Tian R, Liu S, Sherman HT, Zhang X, et al. Early Screening and Risk Factors of Autism Spectrum Disorder in a Large Cohort of Chinese Patients With Prader-Willi Syndrome. Front Psychiatry. 2020 Nov 26;11:594934.
- Dimitropoulos A, Ho A, Feldman B. Social Responsiveness and Competence in Prader-Willi Syndrome: Direct Comparison to Autism Spectrum Disorder. J Autism Dev Disord. 2013 Jan;43(1):103–13.
- 6. Rice LJ, Gray KM, Howlin P, Taffe J, Tonge BJ, Einfeld SL. The developmental trajectory of disruptive behavior in Down syndrome, fragile X syndrome, Prader–Willi syndrome and Williams syndrome. Am J Med Genet. 2015 Jun;169(2):182–7.
- Singh D, Wakimoto Y, Filangieri C, Pinkhasov A, Angulo M. Guanfacine Extended Release for the Reduction of Aggression, Attention-Deficit/Hyperactivity Disorder Symptoms, and Self-Injurious Behavior in Prader-Willi Syndrome—A Retrospective Cohort Study. Journal of Child and Adolescent Psychopharmacology. 2019 May;29(4):313–7.
- 8. Tunnicliffe P, Woodcock K, Bull L, Oliver C, Penhallow J. Temper outbursts in Prader-Willi syndrome: causes, behavioural and emotional sequence and responses by carers: Temper outbursts in Prader-Willi syndrome. J Intellect Disabil Res. 2014 Feb;58(2):134–50.
- 9. Duan Y, Liu L, Zhang X, Jiang X, Xu J, Guan Q. Phenotypic spectrum and mechanism analysis of Schaff Yang syndrome: A case report on new mutation of MAGEL2 gene. Medicine. 2021 Jun 18;100(24):e26309.
- 10. Bennett JA, Germani T, Haqq AM, Zwaigenbaum L. Autism spectrum disorder in Prader-Willi syndrome: A systematic review. Am J Med Genet. 2015 Dec;167(12):2936–44.
- 11. Reznik DL, Yang MV, Albelda De La Haza P, Jain A, Spanjaard M, Theiss S, et al. *Magel2* truncation alters select behavioral and physiological outcomes in a rat model of Schaaf-Yang syndrome. Disease Models & Mechanisms. 2023 Feb 1;16(2):dmm049829.

- 12. Meziane H, Schaller F, Bauer S, Villard C, Matarazzo V, Riet F, et al. An Early Postnatal Oxytocin Treatment Prevents Social and Learning Deficits in Adult Mice Deficient for Magel2, a Gene Involved in Prader-Willi Syndrome and Autism. Biological Psychiatry. 2015 Jul;78(2):85–94.
- 13. Borie AM, Dromard Y, Guillon G, Olma A, Manning M, Muscatelli F, et al. Correction of vasopressin deficit in the lateral septum ameliorates social deficits of mouse autism model. Journal of Clinical Investigation. 2021 Jan 19;131(2):e144450.
- 14. Dromard Y, Borie AM, Chakraborty P, Muscatelli F, Guillon G, Desarménien MG, et al. Disengagement of somatostatin neurons from lateral septum circuitry by oxytocin and vasopressin restores social-fear extinction and suppresses aggression outbursts in Prader-Willi syndrome model. Biological Psychiatry [Internet]. 2023; Available from: https://www.sciencedirect.com/science/article/pii/S000632232301661X
- 15. Chen H, Victor AK, Klein J, Tacer KF, Tai DJC, De Esch C, et al. Loss of MAGEL2 in Prader-Willi syndrome leads to decreased secretory granule and neuropeptide production. JCI Insight. 2020 Sep 3;5(17):e138576.
- 16. Oztan O, Zyga O, Stafford DEJ, Parker KJ. Linking oxytocin and arginine vasopressin signaling abnormalities to social behavior impairments in Prader-Willi syndrome. Neuroscience & Biobehavioral Reviews. 2022 Nov;142:104870.
- 17. Bruno CA, O'Brien C, Bryant S, Mejaes JI, Estrin DJ, Pizzano C, et al. pMAT: An open-source software suite for the analysis of fiber photometry data. Pharmacology Biochemistry and Behavior. 2021 Feb;201:173093.
- 18. Friard O, Gamba M. BORIS : a free, versatile open-source event-logging software for video/audio coding and live observations. Fitzjohn R, editor. Methods Ecol Evol. 2016 Nov;7(11):1325–30.
- 19. Pennington ZT, Dong Z, Feng Y, Vetere LM, Page-Harley L, Shuman T, et al. ezTrack: An opensource video analysis pipeline for the investigation of animal behavior. Sci Rep. 2019 Dec 27;9(1):19979.
- 20. Tang Y, Benusiglio D, Lefevre A, Hilfiger L, Althammer F, Bludau A, et al. Social touch promotes interfemale communication via activation of parvocellular oxytocin neurons. Nat Neurosci. 2020 Sep;23(9):1125–37.
- 21. Yukinaga H, Hagihara M, Tsujimoto K, Chiang HL, Kato S, Kobayashi K, et al. Recording and manipulation of the maternal oxytocin neural activities in mice. Current Biology. 2022 Sep;32(17):3821-3829.e6.
- 22. Rychlik JJ. CHARACTERIZATION OF ASTROCYTE AND OXYTOCIN NEURON ACTIVITY DYNAMICS IN THE PARAVENTRICULAR NUCLEUS OF THE HYPOTHALAMUS.
- 23. Ino D, Tanaka Y, Hibino H, Nishiyama M. A fluorescent sensor for real-time measurement of extracellular oxytocin dynamics in the brain. Nat Methods. 2022 Oct;19(10):1286–94.
- 24. Lakens D. Calculating and reporting effect sizes to facilitate cumulative science: a practical primer for t-tests and ANOVAs. Front Psychol [Internet]. 2013 [cited 2024 Feb 29];4. Available from: http://journal.frontiersin.org/article/10.3389/fpsyg.2013.00863/abstract

- 25. Hirota Y, Arai A, Young LJ, Osako Y, Yuri K, Mitsui S. Oxytocin receptor antagonist reverses the blunting effect of pair bonding on fear learning in monogamous prairie voles. Hormones and Behavior. 2020 Apr 1;120:104685.
- 26. Carcea I, Caraballo NL, Marlin BJ, Ooyama R, Riceberg JS, Mendoza Navarro JM, et al. Oxytocin neurons enable social transmission of maternal behaviour. Nature. 2021 Aug 1;596(7873):553–7.
- 27. Smith AS, Tabbaa M, Lei K, Eastham P, Butler MJ, Linton L, et al. Local oxytocin tempers anxiety by activating GABAA receptors in the hypothalamic paraventricular nucleus. Psychoneuroendocrinology. 2016 Jan;63:50–8.
- 28. Dykens EM, Lee E, Roof E. Prader–Willi syndrome and autism spectrum disorders: an evolving story. Journal of Neurodevelopmental Disorders. 2011 Sep 1;3(3):225–37.
- 29. McCarthy J, Lupo PJ, Kovar E, Rech M, Bostwick B, Scott D, et al. Schaaf-Yang syndrome overview: Report of 78 individuals. American Journal of Medical Genetics Part A. 2018 Dec 1;176(12):2564–74.
- 30. Chung MS, Langouët M, Chamberlain SJ, Carmichael GG. Prader-Willi syndrome: reflections on seminal studies and future therapies. Open Biology. 2020 Sep 23;10(9):200195.
- 31. Debladis J, Valette M, Strenilkov K, Mantoulan C, Thuilleaux D, Laurier V, et al. Face processing and exploration of social signals in Prader-Willi syndrome: a genetic signature. Orphanet J Rare Dis. 2019 Dec;14(1):262.
- 32. Dykens EM, Roof E, Hunt-Hawkins H, Daniell C, Jurgensmeyer S. Profiles and trajectories of impaired social cognition in people with Prader-Willi syndrome. Senju A, editor. PLoS ONE. 2019 Oct 17;14(10):e0223162.
- 33. Famelart N, Diene G, Çabal-Berthoumieu S, Glattard M, Molinas C, Guidetti M, et al. Equivocal expression of emotions in children with Prader-Willi syndrome: what are the consequences for emotional abilities and social adjustment? Orphanet J Rare Dis. 2020 Dec;15(1):55.
- 34. Fernández-Lafitte M, Cobo J, Coronas R, Parra I, Oliva JC, Àlvarez A, et al. Social Responsiveness and Psychosocial Functioning in Adults with Prader–Willi Syndrome. JCM. 2022 Mar 5;11(5):1433.
- 35. Shriki-Tal L, Avrahamy H, Pollak Y, Gross-Tsur V, Genstil L, Hirsch HJ, et al. Psychiatric disorders in a cohort of individuals with Prader–Willi syndrome. Eur psychiatr. 2017 Jul;44:47–52.
- 36. Bishop HJ, Biasini FJ, Stavrinos D. Social and Non-social Hazard Response in Drivers with Autism Spectrum Disorder. J Autism Dev Disord. 2017 Apr;47(4):905–17.
- 37. Pan Y, Olsson A, Golkar A. Social safety learning: Shared safety abolishes the recovery of learned threat. Behaviour Research and Therapy. 2020 Dec;135:103733.
- 38. Micale V, Stepan J, Jurik A, Pamplona FA, Marsch R, Drago F, et al. Extinction of avoidance behavior by safety learning depends on endocannabinoid signaling in the hippocampus. Journal of Psychiatric Research. 2017 Jul;90:46–59.

- Chari T, Griswold S, Andrews NA, Fagiolini M. The Stage of the Estrus Cycle Is Critical for Interpretation of Female Mouse Social Interaction Behavior. Front Behav Neurosci. 2020 Jun 30;14:113.
- 40. Liu XY, Cui D, Li D, Jiao R, Wang X, Jia S, et al. Oxytocin Removes Estrous Female vs. Male Preference of Virgin Male Rats: Mediation of the Supraoptic Nucleus Via Olfactory Bulbs. Front Cell Neurosci. 2017 Oct 23;11:327.
- 41. Menon R, Grund T, Zoicas I, Althammer F, Fiedler D, Biermeier V, et al. Oxytocin Signaling in the Lateral Septum Prevents Social Fear during Lactation. Current Biology. 2018 Apr;28(7):1066-1078.e6.
- 42. Zoicas I, Slattery DA, Neumann ID. Brain Oxytocin in Social Fear Conditioning and Its Extinction: Involvement of the Lateral Septum. Neuropsychopharmacol. 2014 Dec;39(13):3027–35.
- 43. Gigliucci V, Busnelli M, Santini F, Paolini C, Bertoni A, Schaller F, et al. Oxytocin receptors in the Magel2 mouse model of autism: Specific region, age, sex and oxytocin treatment effects. Front Neurosci. 2023 Mar 14;17:1026939.
- 44. Heseding HM, Jahn K, Eberlein CK, Wieting J, Maier HB, Proskynitopoulos PJ, et al. Distinct promoter regions of the oxytocin receptor gene are hypomethylated in Prader-Willi syndrome and in Prader-Willi syndrome associated psychosis. Transl Psychiatry. 2022 Jun 10;12(1):246.
- 45. Johnson L, Manzardo AM, Miller JL, Driscoll DJ, Butler MG. Elevated plasma oxytocin levels in children with Prader–Willi syndrome compared with healthy unrelated siblings. American J of Med Genetics Pt A. 2016 Mar;170(3):594–601.
- 46. Tauber M, Boulanouar K, Diene G, Çabal-Berthoumieu S, Ehlinger V, Fichaux-Bourin P, et al. The Use of Oxytocin to Improve Feeding and Social Skills in Infants With Prader–Willi Syndrome. Pediatrics. 2017 Feb 1;139(2):e20162976.
- 47. Tauber M, Mantoulan C, Copet P, Jauregui J, Demeer G, Diene G, et al. Oxytocin may be useful to increase trust in others and decrease disruptive behaviours in patients with Prader-Willi syndrome: a randomised placebo-controlled trial in 24 patients. Orphanet J Rare Dis. 2011;6(1):47.
- 48. Dykens EM, Miller J, Angulo M, Roof E, Reidy M, Hatoum HT, et al. Intranasal carbetocin reduces hyperphagia in individuals with Prader-Willi syndrome. JCI Insight. 2018 Jun 21;3(12):e98333.
- 49. Althammer F, Muscatelli F, Grinevich V, Schaaf CP. Oxytocin-based therapies for treatment of Prader-Willi and Schaaf-Yang syndromes: evidence, disappointments, and future research strategies. Transl Psychiatry. 2022 Aug 8;12(1):318.
- 50. Armstrong WE, Stern JE, Teruyama R. Plasticity in the electrophysiological properties of oxytocin neurons. Microscopy Res & Technique. 2002 Jan 15;56(2):73–80.
- 51. Stern JE, Armstrong WE. Changes in the Electrical Properties of Supraoptic Nucleus Oxytocin and Vasopressin Neurons during Lactation. J Neurosci. 1996 Aug 15;16(16):4861–71.
- 52. Tops M, Huffmeijer R, Linting M, Grewen KM, Light KC, Koole SL, et al. The role of oxytocin in familiarization-habituation responses to social novelty. Front Psychol [Internet]. 2013 [cited 2023]

Dec 10];4. Available from: http://journal.frontiersin.org/article/10.3389/fpsyg.2013.00761/abstract

- 53. Ates T, Oncul M, Dilsiz P, Topcu IC, Civas CC, Alp MI, et al. Inactivation of Magel2 suppresses oxytocin neurons through synaptic excitation-inhibition imbalance. Neurobiology of Disease. 2019 Jan;121:58–64.
- 54. Liu Y, Rao B, Li S, Zheng N, Wang J, Bi L, et al. Distinct Hypothalamic Paraventricular Nucleus Inputs to the Cingulate Cortex and Paraventricular Thalamic Nucleus Modulate Anxiety and Arousal. Front Pharmacol. 2022 Jan 28;13:814623.
- 55. Chen S, Xu H, Dong S, Xiao L. Morpho-Electric Properties and Diversity of Oxytocin Neurons in Paraventricular Nucleus of Hypothalamus in Female and Male Mice. J Neurosci. 2022 Apr 6;42(14):2885–904.
- Knobloch HS, Charlet A, Hoffmann LC, Eliava M, Khrulev S, Cetin AH, et al. Evoked Axonal Oxytocin Release in the Central Amygdala Attenuates Fear Response. Neuron. 2012 Feb;73(3):553–66.
- 57. Labiner D, Butler L, Cao Z, Hosford D, Shin C, McNamara J. Induction of c-fos mRNA by kindled seizures: complex relationship with neuronal burst firing. J Neurosci. 1993 Feb 1;13(2):744–51.
- 58. Anisimova M, Lamothe-Molina PJ, Franzelin A, Aberra AS, Hoppa MB, Gee CE, et al. Neuronal FOS reports synchronized activity of presynaptic neurons [Internet]. Neuroscience; 2023 Sep [cited 2023 Dec 10]. Available from: http://biorxiv.org/lookup/doi/10.1101/2023.09.04.556168
- 59. Luckman S, Dyball R, Leng G. Induction of c-fos expression in hypothalamic magnocellular neurons requires synaptic activation and not simply increased spike activity. J Neurosci. 1994 Aug 1;14(8):4825–30.
- Takayanagi Y, Yoshida M, Takashima A, Takanami K, Yoshida S, Nishimori K, et al. Activation of Supraoptic Oxytocin Neurons by Secretin Facilitates Social Recognition. Biological Psychiatry. 2017 Feb;81(3):243–51.
- 61. Hashimoto H, Matsuura T, Ueta Y. Fluorescent visualization of oxytocin in the hypothalamoneurohypophysial system. Front Neurosci [Internet]. 2014 Jul 23 [cited 2024 Feb 16];8. Available from: http://journal.frontiersin.org/article/10.3389/fnins.2014.00213/abstract
- 62. Jin D, Liu HX, Hirai H, Torashima T, Nagai T, Lopatina O, et al. CD38 is critical for social behaviour by regulating oxytocin secretion. Nature. 2007 Mar;446(7131):41–5.
- 63. Munesue T, Yokoyama S, Nakamura K, Anitha A, Yamada K, Hayashi K, et al. Two genetic variants of CD38 in subjects with autism spectrum disorder and controls. Neuroscience Research. 2010 Jun;67(2):181–91.
- 64. Heinrichs M, Baumgartner T, Kirschbaum C, Ehlert U. Social support and oxytocin interact to suppress cortisol and subjective responses to psychosocial stress. Biological Psychiatry. 2003 Dec 15;54(12):1389–98.
- 65. de Lind van Wijngaarden RFA, Otten BJ, Festen DAM, Joosten KFM, de Jong FH, Sweep FCGJ, et al. High Prevalence of Central Adrenal Insufficiency in Patients with Prader-Willi Syndrome. The Journal of Clinical Endocrinology & Metabolism. 2008 May 1;93(5):1649–54.

- 66. Oto Y, Matsubara K, Ayabe T, Shiraishi M, Murakami N, Ihara H, et al. Delayed peak response of cortisol to insulin tolerance test in patients with Prader–Willi syndrome. American Journal of Medical Genetics Part A. 2018 Jun 1;176(6):1369–74.
- 67. Stevenson DA, Anaya TM, Clayton-Smith J, Hall BD, Van Allen MI, Zori RT, et al. Unexpected death and critical illness in Prader–Willi syndrome: Report of ten individuals. American Journal of Medical Genetics Part A. 2004 Jan 15;124A(2):158–64.
- 68. Ludwig M, Leng G. Dendritic peptide release and peptide-dependent behaviours. Nat Rev Neurosci. 2006 Feb;7(2):126–36.
- 69. Higashida H. Somato-axodendritic release of oxytocin into the brain due to calcium amplification is essential for social memory. J Physiol Sci. 2016 Jul;66(4):275–82.
- 70. Higashida H, Yokoyama S, Munesue T, Kikuchi M, Minabe Y, Lopatina O. Translational Research in Neurodevelopmental Disorders: Development of Etiology-Based Animal Models. 2011;34(9).
- Neumann I, Ludwig M, Engelmann M, Pittman QJ, Landgraf R. Simultaneous Microdialysis in Blood and Brain: Oxytocin and Vasopressin Release in Response to Central and Peripheral Osmotic Stimulation and Suckling in the Rat. Neuroendocrinology. 2008 Apr 8;58(6):637–45.
- Engelmann, Ebner, Landgraf, Holsboer, Wotjak. Emotional Stress Triggers Intrahypothalamic But Not Peripheral Release of Oxytocin in Male Rats. J Neuroendocrinology. 1999 Nov;11(11):867– 72.
- 73. Babygirija R, Bülbül M, Yoshimoto S, Ludwig K, Takahashi T. Central and peripheral release of oxytocin following chronic homotypic stress in rats. Autonomic Neuroscience: Basic and Clinical. 2012 Apr 3;167(1):56–60.

Figure Legends



Figure 1 - Learning to fear females (social fear cue) generalizes to avoidance of males (social safety cue) in *Magel2* KO mice. (A) Experimental design and timeline. One day before conditioning, test animals interacted with simultaneously presented male and female mice (M0 and F0) in the absence of any shock. During conditioning, *Magel2* KO and WT mice first explored a male stimulus mouse (M1) alone without any shock. This was followed by a session where shock was delivered upon every interaction with a female mouse (F1) (social fear conditioning). 24 h, 48 h and 7 d later, unfamiliar male and female stimulus mice (M2 and F2)

were presented at the same time in a novel arena. (B) At baseline before conditioning, all groups interact with M0 and F0 comparably. (Control WT, N = 12, Conditioned *Magel2* WT = 16, Conditioned *Magel2* KO = 7) (C) During social fear conditioning, conditioned *Magel2* WT and KO mice show significantly less interaction with females than males. (Control WT, N = 6, Conditioned *Magel2* WT = 16, Conditioned *Magel2* KO = 7). (E) Following conditioning, *Magel2* KO mice show impaired investigation of F2 over time. Conditioned *Magel2* WT animals show a significant increase at day 7, as compared to day 1. (F) Conditioned *Magel2* KO mice show a robust avoidance of M2 as well, 7 d later. *p<0.05, **p<0.01, ***p<0.001, ****p<0.001 in post-hoc Sidak's test. Coloured * indicates comparison to respective group (black, Control, *Magel2* WT, red, Conditioned, *Magel2* WT, blue, Conditioned, *Magel2* KO).



Figure 2 - Learning to fear females increases the number, frequency and decreases the duration of Ca²⁺ spikes in SON OXT neurons of *Magel2* KO. (A) Experimental timeline. (B) Representative image showing GCaMP7s expression in SON OXT neurons. Scale bar 50 μ m. (C) Representative traces showing Ca²⁺ activity in SON OXT neurons of conditioned Magel2 WT;Oxt-Cre and Magel2 KO;Oxt-Cre mice during recall, 7 d later. Dotted lines indicate threshold for spike determination at 2.5 SD. (D) Social fear conditioning increases the overall number of Ca2+ spikes in SON OXT of Magel2 KO mice. Conditioned Magel2 WT;Oxt-Cre N=4, Conditioned Magel2 KO;Oxt-Cre N=3. * p<0.05, post-hoc Sidak's test. (E-G) Cumulative frequency of Ca^{2+} spike duration shows a significant left shift post-conditioning. (Conditioned Magel2 WT; Oxt-Cre, d1=338 events, d2=327 events, d7=187 events, 4 animals, Conditioned Magel2 KO;Oxt-Cre, d1=349 events, d2=303 events, d7=292 events, 4 animals) (H) Radar plot showing decrease in spike duration in Magel2 KO;OXT-Cre mice. (I-K) Cumulative frequency of interval between subsequent Ca²⁺ spikes shows a significant left shift post-conditioning. (Conditioned Magel2 WT; Oxt-Cre, d1, n=334 events, d2, n=323 events, d 7, n=183 events, 4 animals, Conditioned Magel2 KO;Oxt-Cre, d1, n=346 events, d2, n=369 events, d 7, n=300 events, 3 animals). (L) Radar plot showing decrease in inter-spike-interval,

indicating increase spike frequency, in *Magel2* KO;*Oxt*-Cre mice. **p<0.01, ***p<0.001, ****p<0.001 in KS test.



Figure 3 - Learning to fear females reduces engagement of SON OXT neurons in Magel2 KO. (A) Experimental timeline. (B1) Magel2 KO mice show a non-significant increase in activity during increase of novel female before conditioning. (B2) Post-conditioning, a significant decrease in Ca²⁺ activity is seen during social investigation of novel female mice (social fear cue) at day 1 and (B3) day 2 after conditioning. (B4) Engagement of SON OXT neurons are still low at day 7, although not statistically different (Conditioned Magel2 WT; Oxt-Cre, d 0, n=37, d 1, n=10, d 2, n=20, d 7, n=39, 4 animals, Conditioned Magel2 KO;Oxt-Cre, d 0, n=15, d 1, n=3, d 2, n=3, d 7, n=3, 3 animals). (C) Radar plot depicts trajectory of SON OXT engagement during investigation of female stimulus mice, before and after social fear conditioning. (D1-3) During exploration of an unfamiliar male social-stimuli, difference in SON OXT engagement seen at baseline or at days 1 and 2 after conditioning. (D4) Significant decrease in Ca²⁺ engagement is seen during social investigation of novel male mice at day 7 (Conditioned Magel2 WT; Oxt-Cre, d 0, n=29, d 1, n=26, d 2, n=36, d 7, n=23, 4 animals, Conditioned Magel2 KO;Oxt-Cre, d0, n=4, d 1, n=3, d 2, n=6, d 7, n=12, 3 animals). (E) Radar plot depicts trajectory of SON OXT engagement during investigation of male stimulus mice, before and after social fear conditioning. **p<0.01, in Mann-Whitney U test.



Figure 4 - Learning to fear females changes calcium dynamics in PVN OXT neurons in Magel2 KO, but does not affect their engagement during social investigation. (A) Experimental timeline. (B) Representative image showing GCaMP7s expression in PVN OXT neurons. Scale bar 50 µm. (C) Representative traces showing Ca²⁺ activity in PVN OXT neurons of conditioned Magel2 WT; Oxt-Cre and Magel2 KO; Oxt-Cre mice during recall, 7 days later. Dotted lines indicate threshold for spike determination at +2.5 standard deviation from the mean (SD). (D) Social fear conditioning decreases the overall number of Ca^{2+} spikes in PVN OXT of Magel2 KO mice. Conditioned Magel2 WT; Oxt-Cre N=5-6, Conditioned *Magel2* KO; *Oxt*-Cre N=3. ** p<0.01, post-hoc Sidak's test, #p<0.05, main effect of genotype. (E) Post-conditioning, both Ca²⁺ spike duration as well as (F) inter-spike interval increases in Magel2 KO; Oxt-Cre, at all timepoints. (G) Engagement of PVN OXT neurons at days 1, 2 and 7 after conditioning depicted during exploration of female stimulus (Conditioned Magel2 WT; Oxt-Cre, d 1, n=9, d 2, n=8, d 7, n=14, 5-6 animals, Conditioned Magel2 KO; Oxt-Cre, d 1, n=12, d 2, n=14, d 7, n=15, 3 animals) (F2) and (H) male stimulus (M2) (Conditioned Magel2 WT; Oxt-Cre, d 1, n=45, d 2, n=110, d 7, n=59, 5-6 animals, Conditioned Magel2 KO; Oxt-Cre, d 1, n=16, d 2, n=16, d 7, n=28, 3 animals).



Figure 5 – Impaired cFos expression in SON OXT neurons of *Magel2* KO mice, but not PVN, 7 days after conditioning. (A) Experimental timeline. (B) Representative images showing cFos expressing OXT neurons in SON and (C) PVN. Scale bar= 50 μ m. (D) *Magel2* KO mice show a significant deficit in cFos expression in SON OXT during recall, 7 d after conditioning, but not in PVN OXT neurons. (*Magel2* WT SON, n=7 roi, N=4 animals, *Magel2* KO SON, n=5 roi, N=3 animals, *Magel2* WT PVN, n=8 roi, N=4 animals) (E) No significant change in cFos expression seen in the absence of shock in either group (*Magel2* WT SON, n=5 roi, N=3 animals), *Magel2* WT SON, n=6 roi, N=3 animals). **p<0.01, post-hoc Tukey's test.



Figure 6 – Overcoming social fear and expression of social safety engages SON OXT neurons, and is impaired in *Magel2* deficiency. (A) *Magel2* KO animals show increased engagement of SON OXT neurons during social cue exploration prior to shock-exposure. Social fear conditioning robustly shuts down such engagement in both KO and WT animals. While *Magel2* WT mice can re-engage these neurons over time, *Magel2* KO fails to do so. ***p<0.001, post-hoc Sidak's test, ###p<0.001, main effect of interaction (genotype x time).(C) No genotypic difference observed in engagement of PVN OXT neurons before or after social fear conditioning. (C) In *Magel2* WT, social fear shuts down OXT neurons in both PVN and SON. Overcoming social fear and expression of social safety selectively reengages SON OXT neurons. (D) In *Magel2* KO mice, social fear also shuts down OXT neuron activity in both SON and PVN, which fail to reengage post-conditioning.