

Bisphosphonates as Radiopharmaceuticals: Spotlight on the Development and Clinical Use of DOTAZOL in Diagnostics and Palliative Radionuclide Therapy

Céleste Souche, Juliette Fouillet, Léa Rubira, Charlotte Donzé, Charlotte

Donzé, Emmanuel Deshayes, Cyril Fersing

▶ To cite this version:

Céleste Souche, Juliette Fouillet, Léa Rubira, Charlotte Donzé, Charlotte Donzé, et al.. Bisphosphonates as Radiopharmaceuticals: Spotlight on the Development and Clinical Use of DOTAZOL in Diagnostics and Palliative Radionuclide Therapy. International Journal of Molecular Sciences, 2023, 25 (1), pp.462. 10.3390/ijms25010462. hal-04544442

HAL Id: hal-04544442 https://hal.science/hal-04544442

Submitted on 12 Apr 2024 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License





Bisphosphonates as Radiopharmaceuticals: Spotlight on the Development and Clinical Use of DOTAZOL in Diagnostics and Palliative Radionuclide Therapy

Céleste Souche¹, Juliette Fouillet¹, Léa Rubira¹, Charlotte Donzé¹, Emmanuel Deshayes^{1,2} and Cyril Fersing^{1,3,*}

- ¹ Nuclear Medicine Department, Institut Régional du Cancer de Montpellier (ICM), University Montpellier, 34298 Montpellier, France
- ² Institut de Recherche en Cancérologie de Montpellier (IRCM), INSERM U1194, University Montpellier, Institut Régional du Cancer de Montpellier (ICM), 34298 Montpellier, France
- ³ IBMM, University Montpellier, CNRS, ENSCM, 34293 Montpellier, France
- * Correspondence: cyril.fersing@icm.unicancer.fr; Tel.: +33-4-67-61-24-78

Abstract: Bisphosphonates are therapeutic agents that have been used for almost five decades in the treatment of various bone diseases, such as osteoporosis, Paget disease and prevention of osseous complications in cancer patients. In nuclear medicine, simple bisphosphonates such as ^{99m}Tc-radiolabelled oxidronate and medronate remain first-line bone scintigraphic imaging agents for both oncology and non-oncology indications. In line with the growing interest in theranostic molecules, bifunctional bisphosphonates bearing a chelating moiety capable of complexing a variety of radiometals were designed. Among them, DOTA-conjugated zoledronate (DOTAZOL) emerged as an ideal derivative for both PET imaging (when radiolabeled with ⁶⁸Ga) and management of bone metastases from various types of cancer (when radiolabeled with ¹⁷⁷Lu). In this context, this report provides an overview of the main medicinal chemistry aspects concerning bisphosphonates, discussing their roles in molecular oncology imaging and targeted radionuclide therapy with a particular focus on bifunctional bisphosphonates. Particular attention is also paid to the development of DOTAZOL, with emphasis on the radiochemistry and quality control aspects of its preparation, before outlining the preclinical and clinical data obtained so far with this radiopharmaceutical candidate.

Keywords: DOTAZOL; bisphosphonates; bone metastases; theranostics; nuclear medicine; ⁶⁸Ga; ¹⁷⁷Lu

1. Introduction

Breast, prostate and lung are the solid cancers with the highest incidence of bone metastases [1], which can lead to complications such as severe bone pain, pathological fractures, nerve compression, hypercalcemia and overall morbidity [2]. Since the 1970s, nuclear medicine has played a key role in the diagnosis of bone metastases with high-sensitivity molecular imaging approaches such as whole-body bone scintigraphy [3–6]. Furthermore, the discipline is well-suited to the concept of theranostics, which refers to the convergence of diagnostic and therapeutic procedures by using the same molecular platform combined with either a photon-emitting radioelement (diagnostic) or a particle-emitting radioelement (therapeutic) [7]. In this context, the most appropriate molecules for bone matrix targeting tend to be bisphosphonates; as a matter of fact, the simplest ^{99m}Tc-labeled derivatives of this chemical family, i.e., medronate and oxidronate, have been extensively used as bone single-photon emission computed tomography (SPECT) imaging agents for over 40 years [8]. Therapeutic counterparts to these SPECT tracers have been designed and studied in humans, such as [¹⁸⁸Re]Re-etidronate, which showed some potential in terms of efficacy [9–21] and improvement of patients' quality of life [22]. However,



Citation: Souche, C.; Fouillet, J.; Rubira, L.; Donzé, C.; Deshayes, E.; Fersing, C. Bisphosphonates as Radiopharmaceuticals: Spotlight on the Development and Clinical Use of DOTAZOL in Diagnostics and Palliative Radionuclide Therapy. *Int. J. Mol. Sci.* **2024**, *25*, 462. https:// doi.org/10.3390/ijms25010462

Academic Editor: Beatriz Salinas Rodríguez

Received: 4 December 2023 Revised: 25 December 2023 Accepted: 27 December 2023 Published: 29 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). rhenium's complex redox chemistry hampered the expansion of this therapeutic option, as it is challenging to implement in a conventional radiopharmaceutical environment [23]. Likewise, rhenium-188 can be obtained from a $^{188}W/^{188}Re$ radioelement generator, but no fully pharmaceutical-grade generator is currently available [24], causing preference to be given to "simpler-to-use" radioelements.

In connection with the ever-growing development of a wide variety of radiotracer candidates for PET imaging, the use of ⁶⁸Ga has recently become increasingly popular, owing to its availability via GMP-grade ⁶⁸Ge/⁶⁸Ga generators [25] and to its rich yet sufficiently straightforward radiochemistry [26]. With regard to therapy, ¹⁷⁷Lu has lately emerged as a key radioelement, with a 6.7 day half-life (allowing the easier logistics of production and supply of either the radioisotope alone or radiolabeled molecules); low-energy beta particles (E_{β} (max) = 497 keV [78.6%], 384 keV [9.1%] and 176 keV [12.2%], theoretically allowing the irradiation of small lesions and decreased damages to non-target tissues because of the short beta particles range); and radiochemical properties that are now largely documented and mastered [27]. To date, the ⁶⁸Ga/¹⁷⁷Lu theranostic pair has demonstrated its clinical reliability with FDA-approved radiopharmaceuticals in the management of neuroendocrine tumors (DOTATOC and DOTATATE vector molecules) [28,29] and metastatic prostate cancer (PSMA ligands vector molecules) [30,31]. This ⁶⁸Ga/¹⁷⁷Lu approach was then transposed to bone targeting with the use of chelator-bearing bisphosphonates, either originally conceived as MRI contrast agents or directly designed as nuclear medicine theranostic agents. Depending on the nature of the chelating agent functionalizing the bisphosphonate, several radioelements could be complexed by these bifunctional molecules, either for diagnostic or therapeutic purposes (Figure 1) [32]. Among these chelator-bearing bisphosphonate, DOTAZOL is one of the most recent derivatives to have been used on humans, for both PET imaging with [68Ga]Ga-DOTAZOL and targeted radionuclide therapy (TRT) with [¹⁷⁷Lu]Lu-DOTAZOL.



Figure 1. General principle of a theranostic approach in nuclear medicine, applied to bifunctional bisphosphonate derivatives.

After an initial overview of the main chemistry and biology aspects of bisphosphonates and their use as radiopharmaceutical agents, this review will provide a summary of the preclinical development of several chelate-conjugated bisphosphonates to finally focus on DOTAZOL, with a particular emphasis on the chemical and radiochemical aspects. DOTAZOL's evaluation and use on humans, for both diagnostic and therapeutic indications, will be discussed with a view to defining the current and possible future roles of this radiopharmaceutical in PET diagnostics and the TRT of bone metastases.

2. Bisphosphonates: From Conventional Drugs to Radiopharmaceuticals

2.1. Development and Pharmacology Basics of Bisphosphonates

Before their use as medicinal products for treating bone or calcium metabolism disorders, bisphosphonates were originally designed as softeners and complexing agents for textile, food and oil industries [33] dating from their first synthesis in 1865 [34]. Initial biological applications of bisphosphonates were related to their structural analogy with inorganic pyrophosphate, which confer in vitro and in vivo inhibitory properties on bone resorption, combined with resistance to acid- or phosphatase-catalyzed hydrolysis [35,36]. These effects on bone were mainly mediated by the inhibition of the hydroxyapatite crystal dissolution but were associated, for the earliest bisphosphonates derivatives, to the inhibitory properties of skeletal mineralization [37]. This was therefore one of the characteristics to be controlled as successive bisphosphonate analogs were developed.

The chemical structure of bisphosphonates is centered on the P-C-P pharmacophore, which is responsible for inhibiting bone resorption, whereas several analogs such as ethilenediphosphonates (P-C-C-P) or iminodiphosphonates (P-N-P) are inactive. This chemical motif is responsible for high affinity and strong binding to hydroxyapatite crystals in the bone mineral matrix; in addition, the central sp³ carbon offers numerous pharmacomodulation possibilities by introducing a variety of substituents in this position. The natures of the residues on this central carbon (referred as R_1 and R_2) allow the identification of several groups of molecules, the first being the non-hydroxy- and non-nitrogen-containing bisphosphonates groups, originally headed by clodronate [38]. Since the late 1960s, it has been demonstrated that derivatives containing a R_1 hydroxy group on the central carbon exhibited improved hydroxyapatite binding, with better blockage of hydroxyapatite crystal dissolution and growth, combined with a greater affinity for calcium (particularly in the bone mineral matrix) [39]. The first hydroxylated bisphosphonate to be used on humans was etidronate, which was originally synthesized in the late 19th century [40] and first used as a calcification inhibitor in the mid-1970s to treat patients with fibrodysplasia ossificans progressiva [41]. In addition to their binding to the mineral components of the bone, an intracellular action of bisphosphonates related to etidronate and clodronate has been identified, based on their structural homology with inorganic pyrophosphate. Such molecules could be incorporated in nucleotide precursors to form non-hydrolysable analogs [42–45], which could then accumulate in osteoclasts and lead to cell death (Figure 2) [46–48]. It was then evidenced that, with R_2 being a basic primary alkylamine as in pamidronate and alendronate [49,50], antiresorptive effects were 10 to 100 times higher than with the non-nitrogen-containing derivatives. This suggested alternative specific intracellular effects associated with the R₂ nitrogen chemical moiety [51]. Notably, N-containing bisphosphonates displayed inhibitory properties against several enzymes involved in the mevalonate biosynthesis pathway, contributing to sterol production, especially farnesyl-pyrophosphate synthase [52,53]. Thus, because of the critical role of the mevalonate pathway for osteoclastic functions, this mechanistic aspect of nitrogen-containing bisphosphonates is of major relevance for this cell type (Figure 2) [54-58]. The importance of the nitrogen atom in the R_2 side chain appeared to be even greater as a tertiary amine motif further increases potency (e.g., in ibandronate) as well as the inclusion of the nitrogen in an aromatic ring (e.g., a pyridine in risedronate or an imidazole in zoledronate) [59]. In addition to their effect on osteoclasts, some bisphosphonate derivatives appear to have a beneficial effect on osteoblasts, enhancing their inhibitory properties on osteoclastogenesis [60,61] and inducing proliferative [62–65] or anti-apoptotic effects [66].



Figure 2. Extracellular and molecular mechanisms of the action of bisphosphonates, exemplified with zoledronate for nitrogen-containing BPs. Molecular interaction representations between zoledronate and farnesyl-PP synthase were adapted from schematic 2-D features of protein-ligand complexes from the Protein Data Bank (PDB) [67]. BP: bisphosphonate; HA: hydroxyapatite; PP: pyrophosphate.

Overall, thorough investigations of bisphosphonates structure-activity relationships, with the significant contribution of crystallographic studies [68], have led to a better understanding of the main mechanistic aspects of these compounds [69] and has opened the way to potential novel utilizations [70–72]. Figure 3 summarizes the key medicinal chemistry elements outlined in bisphosphonate series. To date, depending on the country, around 10 bisphosphonates for pharmacological uses have been approved for clinical indications in humans (almost all being hydroxy- and nitrogen-containing compounds). Their main applications are the treatment of post-menopausal osteoporosis, the management of osteolysis and hypercalcemia of malignant origin, the prevention of bone complications (fractures and spinal cord compression) in cancer patients and the treatment of Paget's disease. The safety profile of bisphosphonates is well-defined and the most serious adverse affects are either rare or predictable, including gastrointestinal (nausea, vomiting, epigastric pain, esophagitis, gastric ulcer and dyspepsia), kidney (renal failure if given in IV), musculoskeletal (pain, atypical femoral fractures and osteonecrosis of the jaw) or cutaneous side effects. Some of the most recent drugs, such as zoledronate, tend to be positioned as first- or second-line therapies, given their efficacy and convenient treatment regimens (injectable drugs are characterized by 12- or 18-month dosing intervals, which ensures compliance). Their use as adjuvant therapy in selected malignancies such as breast cancer [73–76] shows positive effects, although small, on bone recurrence, fracture rates, cancer mortality and overall survival [77], possibly via their ancillary effects on cancer cells including immunomodulation and synergy with anticancer agents [78]. Moreover, beyond their pharmacological utility, selected bisphosphonates are currently used or investigated as bone molecular targeting agents for nuclear medicine applications.



Figure 3. Main structure-activity relationships among bisphosphonates and the relevant chemical structures of bisphosphonates used for clinical applications. FPPS: farnesyl-pyrophosphate synthase; HA: hydroxyapatite; Ppi: inorganic pyrophosphate.

2.2. Bone Metastases Molecular Targeting and Early Bisphosphonate-Based Radiopharmaceuticals

Bone microenvironments provide a privileged metastatic niche, especially when tumor cells display a particular affinity for bone tissue [79]. In several diseases such as lung cancer, bone metastases are associated with predominant osteoclast-mediated bone resorption lesions which are described as lytic. Conversely, prostate cancer patients with bone metastases often exhibit dense osteosclerotic lesions with increased osteoblast activity [80]. Mixed lesions are prevalent across various tumor types but are especially frequent among patients with metastatic breast cancer [81]. In nuclear medicine, several bone-addressed radiopharmaceuticals have therefore been developed for diagnostic or therapeutic purposes, exploiting two main principles for targeting bone metastases (Figure 4):



Figure 4. Overview of radiopharmaceuticals for the diagnosis and treatment of bone metastases, depending on their targeting pattern.

- Specific bone targeting based on the electronic properties of a free radioactive atom: two isotopes of the same atom do not differ in their electronic structure and therefore have identical chemical properties, which allows skeletal targeting by the isotopes of atoms with natural affinities for bone. This applies to $[^{18}F]$ fluoride ions, which replace hydroxide anions in hydroxyapatite crystals to form fluoroapatite in the bone mineral [82]. The [¹⁸F]NaF PET imaging agent tends to be more accurate than bone scintigraphy for the detection of skeletal metastatic lesions in several types of cancer [83–86]. In therapy, [³²P]orthophosphate ($t_{1/2}$ = 14.3 days; $E_{\beta-max}$ = 1.71 MeV) is also incorporated into hydroxyapatite crystals [87] and was historically used to treat painful osteoblastic metastases [88]. This treatment showed moderate efficacy with the disappearance of pain symptoms in almost half of patients [89] but caused common bone marrow toxicity, especially in patients with renal impairment. Likewise, the chemical elements in the same group/column of the periodic table of elements are characterized by the same number of valence electrons, and therefore, by usually comparable chemical reactivity. Part of the same group as calcium, ⁸⁹Sr ($t_{1/2}$ = 50.5 days; $E_{\beta-max}$ = 1.49 MeV) is an alkaline earth metal that accumulates in lesions with high osteoblastic activity and has been used under its dichloride salt form in the palliative treatment of pain associated with bone metastases [90], especially in prostate cancer. The initial clinical trials with this radiopharmaceutical in monotherapy showed modest effects on pain control in bone metastases associated with substantial bone marrow toxicity [91-93], while its use in patients treated with doxorubicin [94] or docetaxel [95] tended to improve both the symptoms associated with bone metastases and survival. Similarly, alpha-emitting radionuclide ²²³Ra ($t_{1/2}$ = 11.4 days; E α = 5.0 to 7.5 MeV [95.3%]; E $_{\beta-max}$ = 1.37 MeV and 1.42 MeV [3.6%]) showed an overall survival benefit in patients with metastatic prostate cancer, with a significant 9-month delay in bone-related events when associated with a bone-protecting agent (e.g., denosumab) [96]. To date, radium-223 dichloride (Xofigo[®], Bayer, Leverkusen, Germany) is the only FDA- and EMA-approved targeted alpha therapy available.
- Specific targeting based on a vector molecule with bone tropism: in scintigraphic imaging, medronate (MDP) [97] and oxidronate (HMDP) [98,99] were among the first bisphosphonates to be used as bone scintigraphy imaging vectors on humans after the pioneering application of [99mTc]Tc-etidronate [100-103] (Figure 5). These two compounds are characterized by their simple chemical structures, which do not contain dedicated chelation sites. Although their formulation in single-vial cold kits for radiopharmaceutical preparation makes ^{99m}Tc radiolabeling simple and ensures high radiochemical purity levels, [99mTc]Tc-MDP and [99mTc]Tc-HMDP complexes do not form a single defined chemical entity but are rather structured into a mixture of monomers, oxo-bridged dimers and oligomeric clusters of varying sizes, featuring diverse technetium-oxo core arrangements, oxidation states and ligand coordination numbers [104] with a composition that varies according to pH, technetium concentration and oxygen amount [105]. Lastly, the phosphonate groups of MDP and HMDP (as well as the hydroxyl group of HMDP) serve both as coordination sites with ^{99m}Tc and as recognition sites for the bone mineral matrix. Consequently, the bone affinity of the corresponding ^{99m}Tc complexes is intrinsically reduced [106]. Even so, these radiopharmaceuticals remain reference bone scintigraphy agents, either in oncology for cancer staging [107,108] and therapeutic response evaluation [109–111] or in benign bone disorders such as Paget disease [112,113] or primary hyperparathyroidism [114,115]. Interestingly, ^{99m}Tc-radiolabeled butedronate (2,3-dicarboxypropane-1,1-diphosphonate, DPD, Figure 5) [116] is another SPECT imaging agent with the same indications as [99mTc]Tc-MDP and [99mTc]Tc-HMDP but also has a particular role in the detection of cardiac amyloidosis [117–119]. Concerning therapy, a bisphosphonate-related derivative with an ethylenediamine tetraphosphonate structure (EDTMP, also named lexidronam) radiolabeled with 153 Sm ($t_{1/2}$ = 1.9 days; $E_{\beta-max}$ = 0.81 MeV) has also been used since the late 1980s [120,121] for the relief of pain resulting from bone metastases;



Figure 5. Chemical structure of monofunctional bisphosphonates and related agents for ^{99m}Tc (MDP, HMDP, HEDP, DPD), ^{188/186}Re (HEDP) or ¹⁵³Sm (EDTMP) radiolabeling.

2.3. From Standard Bisphosphonates to Bifunctional Derivatives Optimized for Nuclear Medicine

Beyond the simple, monofunctional bisphosphonate vector molecules mentioned above, other radiopharmaceutical candidates have been designed on the concept of bifunctional agents, dissociating the radiophore part of the molecule that binds to the radioactive atom from the vector part responsible for tropism specificities [106] (Figure 6). In particular, acyclic complexing moieties were used to enable undemanding radiolabeling conditions. Thus, numerous agents for ^{99m}Tc radiolabeling were designed, with a chemical variety based on the selected chelating agent. Notably, aminoalkyl derivatives such as pamidronate and alendronate are particularly well-suited to conjugation with a bifunctional chelator because the alkyl chain serves as a spacer and the primary amine serves as an anchoring site for the chelating group. Bisphosphonates bearing an ethylenedicysteine [124], a mercaptoacetyltriglycine (MAG₃) [125,126], a hydrazinonicotinamide (HYNIC) [125,127], an imidazole- [128], a pyrazole- [129,130] or a pyridine-containing chelator [131] were reported in the literature but were not studied in humans. More recently, a pamidronate derivative with a tris(3,4-hydroxypyridinone) (THP) ligand was reported to allow quantitative ⁶⁸Ga radiolabeling at room temperature and to promise in vivo properties (high bone/muscle and bone/blood ratios and fast blood clearance) comparable to [18F]NaF [132]. Lastly, a non-hydroxylated bisphosphonate derivative called P15-041, conjugated to the linear chelator HBED-CC, displayed excellent ⁶⁸Ga labeling properties (>95% radiochemical yield (RCY) at room temperature in 5 min, with a single-vial cold kit formulation developed later [133]), good stability and high bone uptake in normal mice [134]. It was evaluated on humans in early-phase clinical trials for imaging bone metastases in prostate cancer, showing high accumulation in bone lesions and rapid clearance from the blood, soft tissues and normal bone, with tumor-to-normal bone ratios up to 8 at 90 min and reasonable dosimetry estimates [135,136]. A larger clinical study on 51 cancer patients with bone metastases from various types of primary cancers showed the superiority of [68Ga]Ga-P15-041 over [^{99m}Tc]Tc-MDP in terms of sensitivity, positive predictive value, negative predictive value and accuracy, making P15-041 a valuable PET radiotracer candidate for a wider use in the detection of skeletal metastases [137]. Nevertheless, a direct comparison with another bone-targeting PET radiotracer would be necessary to confirm its efficiency.

To access other radiometals and to form complexes with high thermodynamic stability, the functionalization of the bisphosphonate derivatives with cyclic chelators was also studied. More specifically, compounds containing a nine-membered cyclic chelator related to NOTA or NODA were designed for easy radiolabeling with ⁶⁸Ga and were used as PET imaging agents (Figure 7). These include NODAGA-Alendronate [138], NODAGA-Pamidronate [139] and NO2AP-Bisphosphonate [140], which has demonstrated high diagnostic efficiency in humans, with bone metastasis detection properties comparable to [¹⁸F]NaF [141]. Another compound, NODAGA-Zoledronate, also achieved quantitative yields when radiolabeled with ⁶⁸Ga [142] and demonstrated its clinical value in the detection of bone metastases in prostate cancer patients who experienced PSA progression on PSMA-based radioligand therapy or in the setting of restaging [143,144]. NODAGA-Zoledronate also displayed an important uptake in atherosclerotic plaques, which could make it suitable for risk assessment by PET/CT imaging in patients with atherosclerotic cardiovascular disease [145,146]. Lastly, from a radiochemical point of view, it is interesting to note that derivatives with a nine-membered cyclic chelator can be labeled with ¹⁸F via a non-covalent radiofluorination approach that relies on the formation of [¹⁸F]aluminum fluoride [147], as exemplified with [¹⁸F]AlF-NOTA-Pamidronate [148]. Aside from NOTA and NODA derivatives, mesocyclic chelators are a recent group of complexing moieties suitable for ⁶⁸Ga labeling [149]. Derived from 6-amino-1,4-diazepine-triacetic acid, these AAZTA-derived chelators allow fast ⁶⁸Ga radiolabeling under mild conditions. Such a chelator was used to functionalize several bisphosphonate derivatives which could offer promising diagnostic potential, although they have only been evaluated in mice up until now [150].



Figure 6. Chemical structure of several bifunctional bisphosphonates conjugated to an acyclic chelator moiety for ^{99m}Tc (**left**) or ⁶⁸Ga (**right**) radiolabeling. The oxidation states of ^{99m}Tc in these complexes are not shown. The name of each chelating motif is indicated in color.



Figure 7. Chemical structure of bifunctional bisphosphonates bearing a nine-membered cyclic chelator, which is particularly suitable for ⁶⁸Ga complexation.

The functionalization of bisphosphonates with a DOTA chelator has also been investigated, with a view to developing theranostic approaches based on a unique vector molecule for both diagnostic purposes and therapy (Figure 8). DOTA is a 12-membered macrocyclic chelator characterized by its ability to form thermodynamically and kinetically

stable complexes with a variety of radiometals including ¹¹¹In and ⁶⁸Ga for imaging purposes, or ⁹⁰Y and ¹⁷⁷Lu for TRT applications [32]. One of the simplest DOTA-containing bisphosphonates is the bisphosphonate monoamide analogue of DOTA, BPAMD, which can be prepared via a rather convenient synthesis route [151] and radiolabeled with 68 Ga in fairly straightforward conditions. It initially demonstrated promising potential for high bone accumulation in mice PET imaging studies [152]. Subsequently, it showed positive results in its first-in-human application, with significantly high target-to-soft tissue ratios and rapid renal clearance [153]. Its standardized uptake values (SUVs) were comparable to [¹⁸F]NaF, and some metastases even displayed higher bisphosphonate accumulations. These encouraging diagnostic outcomes paved the way for the first therapeutic applications of BPAMD, replacing 68 Ga with β -emitter 177 Lu. In this way, the in-house preparation protocols of [¹⁷⁷Lu]Lu-BPAMD were developed for routine production to treat patients with skeletal metastases [154,155]. After conclusive preclinical evaluation against other simple DOTA-bisphosphonate derivatives (including original dimeric molecules bearing bisphosphonate groups on two of the four acetate arms of the DOTA core) [156], [¹⁷⁷Lu]Lu-BPAMD was successfully administered to a number of patients, showing a biodistribution similar to its ⁶⁸Ga-labeled counterpart [157,158]. The prolonged contact time of [¹⁷⁷Lu]Lu-BPAMD with bone metastases resulted in high tumor doses, leading to a significant reduction in osteoblastic activity in bone lesions. Moreover, the therapy did not cause any significant adverse events. In comparison with [177Lu]Lu-EDTMP, [177Lu]Lu-BPAMD demonstrated higher bone uptake and a higher target-to-background ratio; however, this could be due to the much higher amount of ligand used in the synthesis of [¹⁷⁷Lu]Lu-EDTMP and its potential target blocking [159]. Although BPAMD displayed good results in both diagnostic and therapeutic applications in humans, the further chemical optimization (potentially requiring a more demanding synthesis route) of such a bisphosphonate-based probe could still be considered. For example, a BPAMD analog bearing a reversible albumin-binding domain was designed to delay body elimination and enhance target accumulation for TRT purposes [160]. More practically, a hydroxy derivative could be used to improve its affinity for the bone matrix, or a nitrogen-containing substituent might be introduced as a spacer, because close proximity of the chelating group to the vector moiety could impair target recognition [161]. The very recent DOTA-ibandronate (DOTA-IBA) was designed on these criteria and displayed both good in vitro characteristics (stability > 91% over 4h after ⁶⁸Ga radiolabeling, high hydrophilicity and 80% plasma protein binding) and in vivo properties (no tissue or organ toxicity in mice, rapid blood clearance with high bone uptake and accurate bone PET/CT images in mice and rabbits); a first-in-patient study of [68Ga]Ga-DOTA-IBA also showed comparable results to [99mTc]Tc-MDP in the identification of skeletal lesions, with a higher sensitivity for showing small lesions and a higher target-to-non-target ratio for [⁶⁸Ga]Ga-DOTA-IBA compared to [^{99m}Tc]Tc-MDP [162]. In addition, a dosimetry study confirmed its high activity uptake and long retention in bone, with absorbed doses to the whole body and critical organs (red marrow and drug-excretion organs such as kidneys and bladder) within the safety limit [163]. The therapeutic pendant [¹⁷⁷Lu]Lu-DOTA-IBA was logically developed and could be efficiently radiolabeled after 15 min at 95 °C in a 0.25 M sodium acetate buffer pH 10.4 [164]. The early clinical evaluation of [177Lu]Lu-DOTA-IBA (370-1110 MBq) in five patients with bone metastases of malignant diseases showed fast and long-lasting bone pain palliation in three patients, with no detectable hematological or organ toxicity. Subsequently, a phase 0/1 trial on 18 patients with bone metastases first provided dosimetry data consistent with those of [⁶⁸Ga]Ga-DOTA-IBA [165]. Next, the single administration of 891.5 \pm 301.3 MBq of [¹⁷⁷Lu]Lu-DOTA-IBA caused bone pain palliation in 82% of patients and was associated with partial responses in three patients, disease progression in one patient, and stable disease in 14 patients according to the [⁶⁸Ga]Ga-DOTA-IBA PET/CT follow-up at 8 weeks. The highly promising effects of this DOTA-bisphosphonate, radiolabeled with either ⁶⁸Ga for PET imaging or ¹⁷⁷Lu for TRT, should be confirmed in a larger population but already suggest the theranostic potential



of this molecule. Several case reports support these data [166], some even reporting the efficacy of DOTA-IBA radiolabeled with alpha emitter actinium-225 [167].

Figure 8. Chemical structure of bifunctional DOTA-bisphosphonates for Me(III) (e.g., ⁶⁸Ga or ¹⁷⁷Lu) radiolabeling.

Overall, although the newly developed candidate DOTA-IBA offers undeniable advantages, its use in humans is still scarcely reported. To date, other molecules have been developed on the same model such as DOTA-Pamidronate [168] and DOTA-Alendronate [169–172]; however, the optimized theranostic DOTA-bisphosphonate that is most widely used in humans remains DOTAZOL.

3. DOTAZOL: Chemistry and Radiochemistry Considerations

DOTAZOL is based on the structure of third-generation hydroxylated amino-bisphosphonate zoledronate, functionalized at position four of the imidazole ring by an ethyl-monoamide-DOTA moiety [173].

3.1. Chemical Synthesis of DOTAZOL

The first synthesis of DOTAZOL was reported by Meckel et al. in 2017, who presented this macrocyclic bisphosphonate as a promising potential theranostic tool in the management of skeletal metastases [168]. The first step of the synthesis sequence (Scheme 1) consisted of the functionalization of ω -N-acetylhistamine with an acetic arm on the imidazole nitrogen. The use of benzyl bromoacetate with cesium carbonate in DMF formed the desired compound in the average yield via a N-alkylation reaction, followed by a deprotection step catalyzed by palladium on carbon in methanol under hydrogen atmosphere. The hydroxybisphosphonate core was then obtained in a modest yield by reacting a mixture of phosphorus trichloride and phosphorus acid (probably to increase reaction yield compared to PCl₃ alone [174]) in methanesulfonic acid with the latter 1H-imidazole-1-acetic acid derivative. Interestingly, cleavage of the acetamide to form the primary amine was achieved during the same step, based on the strong acidic condition after the hydrolysis of phosphorus trichloride. Finally, functionalization with the chelating moiety was achieved through the nucleophilic substitution of activated DOTA-NHS-ester in water in the presence of sodium carbonate [169].

The overall yield of this synthesis sequence is 2.5%, the most limiting steps being the formation of the bisphosphonate motif (28%) and, most importantly, the anchoring of the DOTA group (16%). Regarding this final step, because unprotected bisphosphonates have poor solubility in organic solvents and dissolve only in aqueous buffers, an amide bond formation is not an appropriate strategy because most active esters tend to hydrolyze in aqueous buffers. Thus, the solution proposed here is to use DOTA-NHS-ester; however, it was reported that this compound led to an increased amount of free DOTA as a hydrolysis product of the NHS ester, resulting in successive, laborious final purification cycles. In 2022, Greifenstein et al. proposed a practical and efficient alternative coupling method between

the 12-membered macrocycle and the bisphosphonate moiety. This ligation method, which relies on the pH-sensitive reactivity of diethyl squarate, is progressively gaining prominence in the development of novel radiopharmaceuticals [175]. Although the compound that was ultimately obtained was not DOTAZOL (the squaric acid replacing the imidazole ring), the stability of squaric acid esters in aqueous buffers would provide a much more straightforward coupling reaction method (Scheme 2), with no significant formation of side products.



Scheme 1. Original synthesis sequence of DOTAZOL as described by Meckel et al. [168].



Scheme 2. Synthesis sequence of DOTAGA.SA.PAM, a DOTAZOL-related bisphosphonate, as described by Greifenstein et al. [176].

3.2. DOTAZOL Radiolabeling with ⁶⁸Ga or ¹⁷⁷Lu

The initial ⁶⁸Ga radiolabeling of DOTAZOL was performed manually, involving sodium acetate buffer 0.5 M pH 4 and post-processed ⁶⁸Ga in acetone/HCl and 25 nmol ligand; the mixture was heated at 98 °C for 15 min. After terminal purification on a weak anion exchange cartridge, the radiochemical purity (RCP) of [68Ga]Ga-DOTAZOL was >98% with the RCY between 80 and 95% [168]. The same method was reported for the preparation of ⁶⁸Ga-labeled DOTAZOL for clinical use [177]. Meisenheimer et al. proposed an alternate ⁶⁸Ga radiolabeling protocol using ammonium acetate 0.08 M with ethanol as a cosolvent, concentrated ⁶⁸Ga in NaCl 5 M pH 1, and heated at 95 °C for 10 min [178]. A thorough study of the reaction conditions was carried out simultaneously and showed that, interestingly, the nature of the reaction vial had a significant influence on the overall reaction outcome. Several polyvalent cations present in the glass could interact with the bisphosphonate molecules, leading to their precipitation and/or adsorption on the reaction vessel. This issue has apparently not been reported in the literature for other DOTA-bisphosphonates; however, it should be considered in the context of unsatisfactory radiolabeling results involving the ionic molecules likely to form this type of interaction [179]. The terminal purification step via solid phase extraction (SPE) was also identified as critical in the [⁶⁸Ga]Ga-DOTAZOL preparation process. The use of a weak anion exchange cartridge to

trap the product and then elute it with phosphate buffer saline could be considered [168]; however, the screening of dozens of SPE cartridges did not identify any fully suitable model as the artridges showed either no retention, the retention of all chemical species, or e thpartial/imperfect retention of one or other species [178]. This parameter therefore requires in-depth study and adaptation to each DOTA-Bisphosphonate used. Indeed, similar issues have been reported with other compounds such as BPAMD, although apparently reliable solutions have been found for this derivative [152,180]. Finally, the automation of the ⁶⁸Ga radiolabeling of DOTAZOL was reported to be quite challenging (notably due to the influence of the reaction vial and to the SPE purification step) [178], which could complicate its use in humans on a larger scale. Thus, the optimized conditions for the automated preparation of [⁶⁸Ga]Ga-DOTAZOL have yet to be identified and adapted to the various GMP-compliant synthesis modules available at present [181]. The ideal scenario for successful and lasting integration into routine clinical practice would probably be the industrial development of DOTAZOL, formulated in a single-vial cold kit for ⁶⁸Ga radiolabeling, which would guarantee the easy preparation of the radiopharmaceutical.

The original ¹⁷⁷Lu radiolabeling protocol was in line with the usual conditions for this type of reaction [27], involving sodium acetate buffer 0.25 M, [¹⁷⁷Lu]LuCl₃ (1 GBq) in 0.04 M HCl and only 20 nmol (~14 µg) DOTAZOL, which were heated at 98 °C for 30 min [168]. In comparison, the ¹⁷⁷Lu radiolabeling of DOTA-IBA employed the exact same reaction conditions, with a heating time of 15 min. The quantitative complexation yields were reached with this method, exempting it from a final purification step. Khawar et al. later described a modified radiolabeling protocol replacing sodium acetate with ascorbate buffer 1.3 M plus gentisic acid 0.3 M as an anti-radiolysis compound [182]. Although the ¹⁷⁷Lu activity used was higher (~6 GBq was added to the reaction medium and heated at 95 °C for 30 min), excellent RCP and RCY values were reached (\geq 98% and \geq 95%, respectively); the radiopharmaceutical thus prepared was subsequently administered to humans. This protocol was also applied by both Yadav et al. [183] and Kreppel et al. [184], who gave details of the quantities of the vector used for ¹⁷⁷Lu radiolabeling (60 μ g), which was logically higher than that used for ⁶⁸Ga radiolabeling. Regardless of the radiolabeling conditions, the pH of the reaction medium should be carefully controlled to be around 5, which is ideal for lutetium-177 complexation [27]. It is worth mentioning that these preparations were carried out manually, again raising the issue of the need for pharmacotechnical works aimed at developing efficient and reliable automated synthesis routes in accordance with GMP guidelines. Additionally, although all reported clinical uses of [¹⁷⁷Lu]Lu-DOTAZOL mention in-house preparation, the industrialized production of this radiopharmaceutical would be a further step toward wider clinical applications.

3.3. Quality Controls of Radiolabeled DOTAZOL: A Critical Step

As well as several parameters involved in the radiolabeling of DOTAZOL, the quality control methods for determining th RCP of this ⁶⁸Ga/¹⁷⁷Lu-labeled bisphosphonate proved to be critical points in the preparation process of such investigational radiopharmaceuticals.

The initial radio-TLC conditions for controlling the RCP of [⁶⁸Ga]Ga-DOTAZOL used an acetone/acetylacetone/HCl mixture (10/10/1) as the mobile phase and silica 60 F254 TLC plates as the stationary phase [168]. With this eluent composition, the unbound ⁶⁸Ga is chelated by acetylacetone to form gallium(III) tris(acetylacetonate), migrating to the solvent front. Colloidal gallium-68 should also convert to ionic [⁶⁸Ga]Ga³⁺ due to its extremely low pH, facilitating its subsequent chelation by acetylacetone and its migration to the solvent front. Notably, slightly adapted analytical conditions (e.g., using acetonitrile instead of acetone) have been reported for other ⁶⁸Ga-labeled bifunctional bisphosphonates [146]. To guarantee the reliability of the results obtained by this analysis and to verify that part of the free gallium identified in radio-TLC does not come from the dissociation of the DOTA-complex [185,186], a threeplate TLC system was later proposed in order to more formally identify all the chemical species potentially present [178]. Table 1 summarizes the migration profiles expected under the different analytical conditions. Interestingly, these conditions were also used to control the RCP of the $[^{68}Ga]Ga$ -DOTAZOL used in humans, but only partially, with the single acetylacetone/acetone (1:1) system [177].

Table 1. Radio-TLC conditions proposed by Meisenheimer et al. for [⁶⁸Ga]Ga-DOTAZOL analysis [178]. The green-colored cells indicate the chemical species identified in each condition, potentially by cross-referencing with other radio-TLC analyses.

Mobile Phase	R _f [⁶⁸ Ga]Ga-DOTAZOL	R _f [⁶⁸ Ga]Ga ³⁺	R _f ⁶⁸ Ga-Colloids
TBAP 60 mM/MeOH (9:1)	0.7–0.8	0.1-0.3	0.1-0.2
Citrate buffer pH 4	0-0.1	0.7–1	0.1-0.2
Acetylacetone/acetone (1:1)	0-0.1	0.7–0.8	0-0.1/0.5-0.9
TBAP: tetrabutylammonium phosph	nate.		

I BAP: tetrabutylammonium phosphate.

Radio-TLC analytical conditions for [177 Lu]Lu-DOTAZOL tended to be much more straightforward and relied on iTLC plates with the citrate buffer 0.1 M pH 4 [168,183], although a three-plate protocol similar to the one for [68 Ga]Ga-DOTAZOL was proposed [182]. Overall, in view of the difficulty of discriminating by TLC between two highly hydrophilic chemical entities such as free 68 Ga $^{3+}/{}^{177}$ Lu $^{3+}$ and radiolabeled bisphosphonates, particular attention should be paid to the rigorous validation of the RCP control conditions for each of these different radiopharmaceuticals. In addition, special consideration should be given to the redaction of the experimental protocols provided in scientific publications to ensure maximum reproducibility of such analytical processes.

The HPLC methods for the RCP determination of radiolabeled DOTA-bisphosphonates are considered difficult to implement [176]. For some derivatives, the use of an anionexchange stationary phase is preferred in order to properly discriminate free radiometal from radiolabeled bisphosphonate [152,180]. Nevertheless, in order to take advantage of the reverse-phase radio-HPLC configuration usually employed for radiopharmaceutical quality controls, several protocols relying on reverse-phase columns were proposed. The original HPLC analysis method of [68Ga]Ga-DOTAZOL used a water/acetonitrile gradient and required sample incubation in 0.25 M desferoxamine for 1 min to complex free ⁶⁸Ga³⁺ and discriminate it from the radiolabeled bisphosphonate [168]. However, these conditions made [68Ga]Ga-DOTAZOL pass through the column without retention and were therefore unsuitable for the formal identification of the radiolabeling product. An alternative method using an isocratic mixture of tetrabutylammonium phosphate 59 mM and methanol (9:1) as the mobile phase allowed the slight retention of the [⁶⁸Ga]Ga-DOTAZOL complex on the reverse stationary phase [178]. This method was also used for quality control of the ⁶⁸Ga-labeled DOTAZOL evaluated in humans [182]. Finally, an HPLC analysis method for [¹⁷⁷Lu]Lu-DOTAZOL that is transposable to other phosphonatecontaining radiopharmaceuticals was developed by Eryilmaz et al., based on a disodium hydrogen phosphate mobile phase adjusted at pH 1.5-1.8 with orthophosphoric acid and containing N,N-dimethyltetradecylamine [187]. This latter reagent can form ion pairs with phosphonate groups and increase the retention of bisphosphonates via the lipophilic moiety of the alkylamine. In summary, similar to radio-TLC methods, particular attention should be given to the validation of a reliable and efficient radio-HPLC method for the RCP determination of radiolabeled bisphosphonates to guarantee the accurate analysis of the radiopharmaceutical preparation and to avoid either false positive or false negative results.

4. Preclinical Investigations on DOTAZOL

4.1. [⁶⁸Ga]Ga-DOTAZOL

Only limited data on the non-clinical evaluation of DOTAZOL are available in the literature, reflecting its rapid transfer to clinical use. Initial works on DOTAZOL relate its extremely high in vitro adsorption on hydroxyapatite when radiolabeled with ⁶⁸Ga, which is slightly better than [⁶⁸Ga]Ga-DOTA-Pamidronate and its dehydroxylated analog [⁶⁸Ga]Ga-BPAPD (92.7 \pm 1.3% vs. 91.2 \pm 2.7% and 83.0 \pm 0.8%, respectively) [168]. An ex vivo organ biodistribution study on healthy rats showed high accumulation in bone and

low uptake in soft tissues, with 40 to 50% of the injected doses detected in the skeletons at 60 min post-injection (p.i.). Notably, [⁶⁸Ga]DOTAZOL displayed the highest bone-to-blood ratio in comparison with [⁶⁸Ga]Ga-DOTA-Pamidronate and [⁶⁸Ga]Ga-BPAPD (11.5 vs. 7.6 and 3.7, respectively). The in vivo biodistribution experiments were in line with the above results, showing the rapid bone accumulation of [⁶⁸Ga]Ga-DOTAZOL with a plateau level at 45 min p.i. and a fast blood clearance. Interestingly, t hepreclinical evaluation of [⁶⁸Ga]Ga-NODAGAZOL highlighted, in comparison with [⁶⁸Ga]Ga-DOTAZOL, its higher femur accumulation (SUV = 4.53 ± 0.28 vs. 3.67 ± 0.37 at 60 min p.i., respectively). [⁶⁸Ga]Ga-NODAGAZOL also displayed higher tumor-to-background ratios than [⁶⁸Ga]Ga-DOTAZOL (femur-to-blood = 57.5 ± 5 vs. 8.4 ± 1.4 at 60 min p.i. respectively), making it even more suitable than DOTAZOL for diagnostic use. However, this molecule cannot be used for therapy applications, as the NODAGA chelator is not suitable for large radiometals such as ¹⁷⁷Lu [142].

4.2. [¹⁷⁷Lu]Lu-DOTAZOL

Similar to its ⁶⁸Ga-labeled counterpart, [¹⁷⁷Lu]Lu-DOTAZOL displayed fast and high uptake in bones and low accumulation in soft tissues, according to the ex vivo biodistribution studies in male Wistar rats [168]. Importantly, the ¹⁷⁷Lu complex showed longer renal clearance and higher accumulation in the kidneys than [⁶⁸Ga]Ga-DOTAZOL ($1.70 \pm 0.13\%$ ID/g vs. $0.53 \pm 0.04\%$ ID/g at 60 min p.i., respectively), suggesting the importance of close monitoring of the renal function before and during treatment (particularly in the case of high and/or repeated doses). The in vivo evaluation of [¹⁷⁷Lu]Lu-DOTAZOL was consistent with ex vivo experiments, positioning this investigational radiopharmaceutical as a good candidate for TRT approach in view of its high skeletal accumulation and good target-to-soft tissues ratio.

In the perspective of a potential ⁶⁸Ga/¹⁷⁷Lu/²²⁵Ac theranostic triplet, the early preclinical evaluation of DOTAZOL labeled with actinium-225 evidenced high bone uptake in healthy Wistar rats, which was comparable to the ⁶⁸Ga and ¹⁷⁷Lu-containing derivatives [188]. No clinical signs of toxicity were evidenced over a 2-month observation period; however, the microscopic histopathological analysis of the kidneys 3 months after a single dose of ~400 kBq [²²⁵Ac]Ac-DOTAZOL revealed significant tubular damage in most of the seven animals studied. As suspected with the ¹⁷⁷Lu-radiolabeled derivative, DOTAZOL complexed with radiometals for TRT purposes may present a renal toxicity that should be considered and may require concomitant strategies to reduce these adverse effects [189].

5. Clinical Uses of DOTAZOL

5.1. PET Imaging with [68Ga]Ga-DOTAZOL

An early experience with [⁶⁸Ga]Ga-DOTAZOL imaging in humans was reported by Zhang et al. [190], particularly in prostate cancer patients for whom DOTAZOL showed an approximately threefold higher uptake in skeletal lesions than [⁶⁸Ga]Ga-PSMA-11. The introduction of this second-generation bifunctional bisphosphonate follows from the initial use of [⁶⁸Ga]Ga-BPAMD in the same indications and was associated with a perfectible radiosynthesis process, suboptimal accumulation in bone metastases (possibly due to its non-hydroxylated chemical structure) and significant uptake in healthy tissues [191].

The initial observation of [⁶⁸Ga]Ga-DOTAZOL biodistribution in a single patient with prostate cancer evidenced, in comparison with [⁶⁸Ga]Ga-PSMA-11, a more intense uptake in the bone metastases associated with lower activity in background and in non-target organs (Figure 9) [158]. In this context, in order to better characterize the fate of [⁶⁸Ga]Ga-DOTAZOL in the human body, a larger biodistribution study was carried out on five patients with metastatic breast, bronchial or prostate carcinoma [177]. After the injection of 2.15 ± 0.17 MBq/kg of [⁶⁸Ga]Ga-DOTAZOL, the initial rapid uptake of the tracer in bone could be observed until 30 min p.i., followed by a further gradual rise. The comparison of PET/CT images in the same bronchial carcinoma patient showed a 2.56-fold higher uptake for [⁶⁸Ga]Ga-DOTAZOL than [¹⁸F]FDG. In addition, a greater number of apparent lesions

was found in each patient with the ⁶⁸Ga-labeled bisphosphonate than with the [¹⁸F]FDG in bronchial carcinoma, the [⁶⁸Ga]Ga-PSMA-617 in prostate cancer and th [^{99m}Tc]Tc-MDP in breast cancer, respectively. Dosimetric analysis showed that [⁶⁸Ga]Ga-DOTAZOL, like other bone-seeking agents, delivered the highest radiation-absorbed dose to the bladder (0.368 mSv/MBq), followed by the osteogenic cells (0.040 mSv/MBq), the kidneys (0.031 mSv/MBq) and red marrow (0.027 mSv/MBq). In a theranostic perspective, these high bladder- and kidney-absorbed doses have to be considered but should be easily reduced by proper hydration and rapid diuresis [189].



Figure 9. Whole-body PET/CT images acquired 60 min p.i. 170 MBq [68 Ga]Ga-PSMA-11 and 155 MBq [68 Ga]Ga-DOTAZOL, respectively, in a single prostate cancer patient (71-years-old, Gleason 4 + 4), as reported by Pfannkuchen et al. [158].

Aside from this specific study, the clinical use of [⁶⁸Ga]Ga-DOTAZOL is also reported for the selection and follow-up of patients considered for TRT with [¹⁷⁷Lu]Lu-DOTAZOL [182–184].

5.2. Targeted Radionuclide Therapy with [¹⁷⁷Lu]Lu-DOTAZOL

The initial assessment of biodistribution and normal organ-absorbed doses after therapeutic doses of [¹⁷⁷Lu]Lu-DOTAZOL was performed by Khawar et al. in patients with either metastatic prostate cancer (n = 2) or bronchial carcinoma (n = 2) (Figure 10) [182]. After a single mean injected dose of 5968 \pm 64 MBq, qualitative SPECT analysis evidenced early and high uptake in the bladder, followed by the kidneys and soft tissues. Consistent with preclinical data, the kidneys showed a rapid decrease in activity at 3 h p.i., concomitantly with an intense and continuous skeletal uptake. Interestingly, despite the small sample size, the skeletal-to-soft tissue contrast values appeared better in bronchial carcinoma patients than in prostate cancer patients. The mean organ-absorbed doses were the highest for the osteogenic cells (3.33 \pm 0.35 mSv/MBq), followed by the kidneys (0.49 \pm 0.16 mSv/MBq), red marrow (0.46 \pm 0.06 mSv/MBq) and the urinary bladder wall $(0.32 \pm 0.02 \text{ mSv/MBq})$. As a comparison, the dose delivered to the kidneys by [¹⁷⁷Lu]Lu-DOTAZOL ranged from 1.2 to 1.88 times lower than that of [¹⁷⁷Lu]Lu-PSMA-617 [192,193].



Figure 10. Planar scintigraphy (anterior and posterior views) after the infusion of $[^{177}Lu]Lu$ -DOTAZOL at (**A**) 20 min, (**B**) 3 h, (**C**) 24 h and (**D**) 168 h, and (**E**) PET/CT after injection of $[^{68}Ga]Ga$ -DOTAZOL in a patient with bone metastases secondary to bronchial carcinoma, as reported by Khawar et al. [182].

The early safety data on the clinical use of [¹⁷⁷Lu]Lu-DOTAZOL were collected by Fernández et al. [194]. After a single mean injected dose of 5780 ± 329 MBq in bone metastatic prostate cancer patients (n = 9), the normalized absorbed doses ranged from 0.206 to 0.564 Gy/GBq for red marrow, from 0.053 to 0.691 Gy/GBq for the kidneys and from 0.635 to 1.980 Gy/GBq for bone surfaces. Assuming a maximum tolerated dose of 2, 23 and 10 Gy for red marrow, kidneys and bone surfaces, red marrow was identified as the dose-limiting organ for all patients, with maximum safely injectable activity ranging from 3.5 to 9.7 GBq (median 6.0 GBq). Regarding the hematological and biochemical parameters, a significant, transitory reduction was observed for leukocytes at 4 and 10 weeks p.i. and for platelets at 4 weeks p.i., with recovery from weeks 4 to 10. Conversely, the treatment had no significant effect on lactate dehydrogenase, alkaline phosphatase, creatinine, hemoglobin or hematocrit levels. No other relevant adverse events such as xerostomia, fatigue, nausea, loss of appetite, nephrotoxicity or hepatotoxicity was experienced by any patient.

The therapeutic safety and efficacy of [177Lu]Lu-DOTAZOL was further documented by Yadav et al., who conducted a prospective, single-arm study in 40 patients experiencing bone pain due to skeletal metastases of either prostate (n = 11), breast (n = 23) or lung cancer (n = 6) [183]. Notably, a fixed activity of 1295 MBq (i.e., 2.7 times lower than the most conservative activity of 3.5 GBq recommended by Fernández et al. [194]) was administered as a single-dose treatment in 15 patients, whereas 25 patients received two cycles with a 3-month interval. Overall, 27.5% patients experienced a >70% reduction in the visual analogue scale (VAS) for pain intensity measurement, 50% had a 40-70% decrease in VAS, 12.5% had a 20–40% decrease in VAS and 10% had a <20% decrease in VAS or an increase in pain. In addition, significant improvement was identified in the Karnofski performance scale (pre-therapy = 60 vs. post-therapy = 80) and in the ECOG performance status (pretherapy = 3 vs. post-therapy = 2). The median time for the initiation of pain relief was \leq 7 days and the median duration of sustained response after the last treatment cycle was 3 months. As patients were not treated by any other anticancer treatment during [¹⁷⁷Lu]Lu-DOTAZOL evaluation (except for eight patients with prostate cancer having hormonal therapy), the median survival from TRT was 13 months (95% CI 10–14 months) and the 1-year survival probability was 55.4%. Regarding toxicity, none of the patients experienced hematologically serious adverse events and only two patients showed grade II transient anemia after [177Lu]Lu-DOTAZOL therapy. No adverse renal or biochemical side

effects were recorded. In summary, although this cohort study was not randomized and had no comparative arm, it provided more detailed information on [¹⁷⁷Lu]Lu-DOTAZOL safety and efficacy in the management of metastatic bone pain in several types of cancers. In future similar studies, it would be of interest to investigate other key parameters, such as metastatic burden reduction or the possibility of more retreatment cycles.

Aside from the previously presented studies involving [¹⁷⁷Lu]Lu-DOTAZOL, a few case reports also exemplify the use of this radioligand therapy in single patients. For instance, a 50-year-old woman with breast cancer previously treated with chemotherapy, radiotherapy and antihormone therapy and displaying extensive bone metastases, was first treated with five cycles of [177Lu]Lu-BPAMD (~3.2 GBq/cycle) that led to a partial remission of the disease [190]. Approximately 15 months later, the patient was treated with three cycles of [177Lu]Lu-DOTAZOL (~11.5 GBq/cycle with ~5 months between each cycle) that achieved a partial remission of the disease after treatment. Comparably, a 64-year-old man with a poorly differentiated, osseous metastatic adenocarcinoma of the lung who had been previously treated with radiation therapy of bone metastases, systemic chemotherapy and immunotherapy (checkpoint inhibitor nivolumab) received two cycles of ~5.5 GBq [¹⁷⁷Lu]Lu-DOTAZOL [184]. As expected, the radiopharmaceutical showed intense accumulation in the bone metastases and caused a decrease in osteoblastic activity with no apparent side effects. These two case reports combined with the previously discussed studies suggest that, similarl to the radium-223 that is available for the treatment of prostate cancer bone metastases, [¹⁷⁷Lu]Lu-DOTAZOL could be an acceptable alternative for the management of skeletal metastases from other types of cancer, e.g., breast or bronchial carcinoma.

6. Conclusions

The early diagnosis and management of skeletal metastases can significantly improve patients quality of life as the disease is considered a common cause of morbidity and mortality with a high socio-economic impact. Among the radionuclide therapy alternatives available in nuclear medicine, several options are reserved for the management of prostate cancer bone metastases, as is the case for ²²³Ra and, non-specific for bone metastases, [¹⁷⁷Lu]Lu-PSMA-617. Combining diagnostic and therapeutic dimensions, the ^{99m}Tc-labeled bisphosphonate and the [¹⁵³Sm]Sm-lexidronam pair are currently available. However, the samarium-tetramethylenephosphonate complex presents significant drawbacks, including a moderate clinical impact and the instability of the coordination compound, associated with delivery and storage constraints (freezing between -10 °C and -20 °C). In this context, second-generation bifunctional bisphosphonates such as DOTAZOL offer promising prospects for the diagnosis and treatment—at present, as a last-line, palliative approach—of skeletal lesions in a variety of cancer diseases. Functionalized by a DOTA-chelating group with versatile features, DOTAZOL even offers compatibility with the theranostic triplet ⁶⁸Ga/¹⁷⁷Lu/²²⁵Ac, similar to somatostatin analogs [195]. To date, the few results available on the use of DOTAZOL in humans, particularly in relation to its therapeutic aspects, are encouraging regarding both efficacy (especially in the reduction of pain associated with bone metastases) and safety. Other promising DOTA-bisphosphonates such as DOTA-IBA seem to follow a similar trend. Nevertheless, the ⁶⁸Ga/¹⁷⁷Lu-labeled DOTAZOL theranostic combination will have to find its place among the therapeutic alternatives available to date. In addition, further prospective patient studies would be needed to reliably validate the optimal dosing and frequency of cures and assess the potential of DOTAZOL for the diagnosis and treatment of bone metastases in selected types of cancer.

Author Contributions: Writing—original draft preparation, C.S. and C.F.; writing—review and editing, L.R., C.D., and J.F.; visualization, E.D.; supervision, C.F. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Ryan, C.; Stoltzfus, K.C.; Horn, S.; Chen, H.; Louie, A.V.; Lehrer, E.J.; Trifiletti, D.M.; Fox, E.J.; Abraham, J.A.; Zaorsky, N.G. Epidemiology of Bone Metastases. *Bone* 2022, 158, 115783. [CrossRef] [PubMed]
- Coleman, R.E.; Croucher, P.I.; Padhani, A.R.; Clézardin, P.; Chow, E.; Fallon, M.; Guise, T.; Colangeli, S.; Capanna, R.; Costa, L. Bone Metastases. *Nat. Rev. Dis. Primers.* 2020, *6*, 83. [CrossRef]
- 3. O'Mara, R.E. Skeletal Scanning in Neoplastic Disease. *Cancer* **1976**, 37, 480–486. [CrossRef] [PubMed]
- 4. Hortobagyi, G.N.; Libshitz, H.I.; Seabold, J.E. Osseous Metastases of Breast Cancer. Clinical, Biochemical, Radiographic, and Scintigraphic Evaluation of Response to Therapy. *Cancer* **1984**, *53*, 577–582. [CrossRef] [PubMed]
- 5. Vinholes, J.; Coleman, R.; Eastell, R. Effects of Bone Metastases on Bone Metabolism: Implications for Diagnosis, Imaging and Assessment of Response to Cancer Treatment. *Cancer Treat. Rev.* **1996**, *22*, 289–331. [CrossRef]
- 6. Rybak, L.D.; Rosenthal, D.I. Radiological Imaging for the Diagnosis of Bone Metastases. Q. J. Nucl. Med. 2001, 45, 53–64.
- Filippi, L.; Chiaravalloti, A.; Schillaci, O.; Cianni, R.; Bagni, O. Theranostic Approaches in Nuclear Medicine: Current Status and Future Prospects. *Expert. Rev. Med. Devices* 2020, *17*, 331–343. [CrossRef]
- Ogawa, K.; Saji, H. Advances in Drug Design of Radiometal-Based Imaging Agents for Bone Disorders. Int. J. Mol. Imaging. 2011, 2011, 537687. [CrossRef]
- 9. de Klerk, J.M.; van Dijk, A.; van het Schip, A.D.; Zonnenberg, B.A.; van Rijk, P.P. Pharmacokinetics of Rhenium-186 after Administration of Rhenium-186-HEDP to Patients with Bone Metastases. *J. Nucl. Med.* **1992**, *33*, 646–651.
- 10. Maxon, H.R.; Schroder, L.E.; Washburn, L.C.; Thomas, S.R.; Samaratunga, R.C.; Biniakiewicz, D.; Moulton, J.S.; Cummings, D.; Ehrhardt, G.J.; Morris, V. Rhenium-188(Sn)HEDP for Treatment of Osseous Metastases. *J. Nucl. Med.* **1998**, *39*, 659–663.
- Palmedo, H.; Guhlke, S.; Bender, H.; Sartor, J.; Schoeneich, G.; Risse, J.; Grünwald, F.; Knapp, F.F.; Biersack, H.J. Dose Escalation Study with Rhenium-188 Hydroxyethylidene Diphosphonate in Prostate Cancer Patients with Osseous Metastases. *Eur. J. Nucl. Med.* 2000, 27, 123–130. [CrossRef] [PubMed]
- 12. Li, S.; Liu, J.; Zhang, H.; Tian, M.; Wang, J.; Zheng, X. Rhenium-188 HEDP To Treat Painful Bone Metastases. *Clin. Nucl. Med.* **2001**, *26*, 919–922. [CrossRef] [PubMed]
- Han, S.H.; de Klerk, J.M.H.; Tan, S.; van het Schip, A.D.; Derksen, B.H.; van Dijk, A.; Kruitwagen, C.L.J.J.; Blijham, G.H.; van Rijk, P.P.; Zonnenberg, B.A. The Placorhen Study: A Double-Blind, Placebo-Controlled, Randomized Radionuclide Study with 186Re-Etidronate in Hormone-Resistant Prostate Cancer Patients with Painful Bone Metastases. J. Nucl. Med. 2002, 43, 1150–1156. [PubMed]
- 14. Liepe, K.; Kropp, J.; Runge, R.; Kotzerke, J. Therapeutic Efficiency of Rhenium-188-HEDP in Human Prostate Cancer Skeletal Metastases. *Br. J. Cancer.* 2003, *89*, 625–629. [CrossRef] [PubMed]
- 15. Zhang, H.; Tian, M.; Li, S.; Liu, J.; Tanada, S.; Endo, K. Rhenium-188-HEDP Therapy for the Palliation of Pain Due to Osseous Metastases in Lung Cancer Patients. *Cancer. Biother. Radiopharm.* **2003**, *18*, 719–726. [CrossRef] [PubMed]
- 16. Liepe, K.; Hliscs, R.; Kropp, J.; Runge, R.; Knapp, F.F.; Franke, W.-G. Dosimetry of 188Re-Hydroxyethylidene Diphosphonate in Human Prostate Cancer Skeletal Metastases. *J. Nucl. Med.* **2003**, *44*, 953–960.
- 17. Liepe, K.; Runge, R.; Kotzerke, J. The Benefit of Bone-Seeking Radiopharmaceuticals in the Treatment of Metastatic Bone Pain. J. Cancer. Res. Clin. Oncol. 2005, 131, 60–66. [CrossRef]
- Liepe, K.; Kotzerke, J. A Comparative Study of ¹⁸⁸Re-HEDP, ¹⁸⁶Re-HEDP, ¹⁵³Sm-EDTMP and ⁸⁹Sr in the Treatment of Painful Skeletal Metastases. *Nucl. Med. Commun.* 2007, 28, 623–630. [CrossRef]
- Cheng, A.; Chen, S.; Zhang, Y.; Yin, D.; Dong, M. The Tolerance and Therapeutic Efficacy of Rhenium-188 Hydroxyethylidene Diphosphonate in Advanced Cancer Patients with Painful Osseous Metastases. *Cancer. Biother. Radiopharm.* 2011, 26, 237–244. [CrossRef]
- Shinto, A.; Mallia, M.; Kameswaran, M.; Kamaleshwaran, K.; Joseph, J.; Radhakrishnan, E.; Upadhyay, I.; Subramaniam, R.; Sairam, M.; Banerjee, S.; et al. Clinical Utility of ¹⁸⁸Rhenium-Hydroxyethylidene-1,1-Diphosphonate as a Bone Pain Palliative in Multiple Malignancies. *World J. Nucl. Med.* 2018, 17, 228–235. [CrossRef]
- Chen, P.; Li, J.; Gui, J.; Liu, C.; Wang, Y.; Zhang, G.; Kuai, D.; Wu, Y.; Liu, Z.; Zuo, C.; et al. Efficacy and Safety of 188Re-HEDP in Lung Cancer Patients with Bone Metastases: A Randomized, Multicenter, Multiple-Dose Phase IIa Study. *Int. J. Clin. Oncol.* 2021, 26, 1212–1220. [CrossRef] [PubMed]
- Lange, R.; Overbeek, F.; de Klerk, J.; Pasker-de Jong, P.; van den Berk, A.; ter Heine, R.; Rodenburg, C.; Kooistra, A.; Hendrikse, N.H.; Bloemendal, H. Treatment of painful bone metastases in prostate and breast cancer patients with the therapeutic radiopharmaceutical rhenium-188-HEDP: Clinical benefit in a real-world study. *Nuklearmedizin* 2016, 55, 188–195. [CrossRef] [PubMed]
- Kleynhans, J.; Duatti, A.; Bolzati, C. Fundamentals of Rhenium-188 Radiopharmaceutical Chemistry. *Molecules* 2023, 28, 1487. [CrossRef] [PubMed]

- 24. Lepareur, N.; Lacœuille, F.; Bouvry, C.; Hindré, F.; Garcion, E.; Chérel, M.; Noiret, N.; Garin, E.; Knapp, F.F.R. Rhenium-188 Labeled Radiopharmaceuticals: Current Clinical Applications in Oncology and Promising Perspectives. *Front. Med.* **2019**, *6*, 132. [CrossRef]
- 25. Lepareur, N. Cold Kit Labeling: The Future of 68Ga Radiopharmaceuticals? Front. Med. 2022, 9, 812050. [CrossRef] [PubMed]
- 26. Meisenheimer, M.; Saenko, Y.; Eppard, E. Gallium-68: Radiolabeling of Radiopharmaceuticals for PET Imaging—A Lot to Consider. In *Medical Isotopes*; Ali Raza Naqvi, S., Babar Imrani, M., Eds.; IntechOpen: London, UK, 2021, ISBN 978-1-83880-627-9.
- 27. Banerjee, S.; Pillai, M.R.A.; Knapp, F.F. (Russ) Lutetium-177 Therapeutic Radiopharmaceuticals: Linking Chemistry, Radiochemistry, and Practical Applications. *Chem. Rev.* 2015, 115, 2934–2974. [CrossRef]
- 28. Hennrich, U.; Benešová, M. [68Ga]Ga-DOTA-TOC: The First FDA-Approved 68Ga-Radiopharmaceutical for PET Imaging. *Pharmaceuticals* **2020**, *13*, 38. [CrossRef]
- 29. Hennrich, U.; Kopka, K. Lutathera[®]: The First FDA- and EMA-Approved Radiopharmaceutical for Peptide Receptor Radionuclide Therapy. *Pharmaceuticals* **2019**, *12*, 114. [CrossRef]
- Hennrich, U.; Eder, M. [⁶⁸Ga]Ga-PSMA-11: The First FDA-Approved 68Ga-Radiopharmaceutical for PET Imaging of Prostate Cancer. *Pharmaceuticals* 2021, 14, 713. [CrossRef]
- Hennrich, U.; Eder, M. [¹⁷⁷Lu]Lu-PSMA-617 (PluvictoTM): The First FDA-Approved Radiotherapeutical for Treatment of Prostate Cancer. *Pharmaceuticals* 2022, 15, 1292. [CrossRef]
- Price, E.W.; Orvig, C. Matching Chelators to Radiometals for Radiopharmaceuticals. Chem. Soc. Rev. 2014, 43, 260–290. [CrossRef] [PubMed]
- 33. Blomen, L.J.M.J. Discovery and History of the Non-Medical Uses of Bisphosphonates. In *Bisphosphonates on Bone*; Chapter 7; Bijvoet, O., Fleisch, H.A., Canfield, R.E., Russell, R.G.G., Eds.; Elsevier Science: Amsterdam, The Netherlands, 1995; pp. 111–124.
- 34. Menschutkin, N. Ueber die Einwirkung des Chloracetyls auf phosphorige Säure. *Ann. Chem. Pharm.* **1865**, 133, 317–320. [CrossRef]
- 35. Francis, M.D.; Graham, R.; Russell, G.; Fleisch, H. Diphosphonates Inhibit Formation of Calcium Phosphate Crystals in Vitro and Pathological Calcification in Vivo. *Science* **1969**, *165*, 1264–1266. [CrossRef] [PubMed]
- 36. Fleisch, H.; Russell, R.G.; Francis, M.D. Diphosphonates Inhibit Hydroxyapatite Dissolution in Vitro and Bone Resorption in Tissue Culture and in Vivo. *Science* **1969**, *165*, 1262–1264. [CrossRef] [PubMed]
- Russell, R.G.; Mühlbauer, R.C.; Bisaz, S.; Williams, D.A.; Fleisch, H. The Influence of Pyrophosphate, Condensed Phosphates, Phosphonates and Other Phosphate Compounds on the Dissolution of Hydroxyapatite in Vitro and on Bone Resorption Induced by Parathyroid Hormone in Tissue Culture and in Thyroparathyroidectomised Rats. *Calcif. Tissue. Res.* 1970, *6*, 183–196. [CrossRef]
- 38. Quimby, O.T.; Prentice, J.B.; Nicholson, D.A. Tetrasodium Carbonyldiphosphonate. Synthesis, Reactions, and Spectral Properties. J. Org. Chem. 1967, 32, 4111–4114. [CrossRef]
- 39. Ebrahimpour, A.; Francis, M.D. Bisphosphonate Therapy in Acute and Chronic Bone Loss: Physical Chemical Considerations in Bisphosphonate-Related Therapies. In *Bisphosphonates on Bone*; Bijvoet, O., Fleisch, H.A., Canfield, R.E., Russell, R.G.G., Eds.; Elsevier Science: Amsterdam, The Netherlands, 1995; pp. 125–136.
- 40. Baeyer, H.V.; Hofmann, K.A. Acetodiphosphorige Säure. Berichte. Dtsch. Chem. Ges. 1897, 30, 1973–1978. [CrossRef]
- Bijvoet, O.L.M.; Nollen, A.J.G.; Slooff, T.J.J.H.; Feith, R. Effect of a Diphosphonate on Para-Articular Ossification after Total Hip Replacement. Acta Orthop. Scand. 1974, 45, 926–934. [CrossRef]
- 42. Pelorgeas, S.; Martin, J.-B.; Satre, M. Cytotoxicity of Dichloromethane Diphosphonate and of 1-Hydroxyethane-1,1-Diphosphonate in the Amoebae of the Slime Mould Dictyostelium Discoideum. *Biochem. Pharmacol.* **1992**, *44*, 2157–2163. [CrossRef]
- Frith, J.C.; Mönkkönen, J.; Blackburn, G.M.; Russell, R.G.G.; Rogers, M.J. Clodronate and Liposome-Encapsulated Clodronate Are Metabolized to a Toxic ATP Analog, Adenosine 5'-(β,γ-Dichloromethylene) Triphosphate, by Mammalian Cells In Vitro. *J. Bone Min. Res.* 1997, *12*, 1358–1367. [CrossRef]
- Rogers, M.J.; Brown, R.J.; Hodkin, V.; Blackburn, G.M.; Russell, R.G.G.; Watts, D.J. Bisphosphonates Are Incorporated into Adenine Nucleotides by Human Aminoacyl-tRNA Synthetase Enzymes. *Biochem. Biophys. Res. Commun.* 1996, 224, 863–869. [CrossRef] [PubMed]
- Auriola, S.; Frith, J.; Rogers, M.J.; Koivuniemi, A.; Mönkkönen, J. Identification of Adenine Nucleotide-Containing Metabolites of Bisphosphonate Drugs Using Ion-Pair Liquid Chromatography–Electrospray Mass Spectrometry. J. Chromatogr. B Biomed. Sci. Appl. 1997, 704, 187–195. [CrossRef] [PubMed]
- Rogers, M.J.; Xiong, X.; Ji, X.; Mönkkönen, J.; Russell, R.G.G.; Williamson, M.P.; Ebetino, F.H.; Watts, D.J. Inhibition of Growth of Dictyostelium Discoideum Amoebae by Bisphosphonate Drugs Is Dependent on Cellular Uptake. *Pharm. Res.* 1997, 14, 625–630. [CrossRef] [PubMed]
- Frith, J.C.; Mönkkönen, J.; Auriola, S.; Mönkkönen, H.; Rogers, M.J. The Molecular Mechanism of Action of the Antiresorptive and Antiinflammatory Drug Clodronate: Evidence for the Formation in Vivo of a Metabolite That Inhibits Bone Resorption and Causes Osteoclast and Macrophage Apoptosis. *Arthritis. Rheum.* 2001, 44, 2201–2210. [CrossRef] [PubMed]
- Lehenkari, P.P.; Kellinsalmi, M.; Näpänkangas, J.P.; Ylitalo, K.V.; Mönkkönen, J.; Rogers, M.J.; Azhayev, A.; Väänänen, H.K.; Hassinen, I.E. Further Insight into Mechanism of Action of Clodronate: Inhibition of Mitochondrial ADP/ATP Translocase by a Nonhydrolyzable, Adenine-Containing Metabolite. *Mol. Pharmacol.* 2002, *61*, 1255–1262. [CrossRef] [PubMed]

- 49. Shinoda, H.; Adamek, G.; Felix, R.; Fleisch, H.; Schenk, R.; Hagan, P. Structure-Activity Relationships of Various Bisphosphonates. *Calcif. Tissue. Int.* **1983**, *35*, 87–99. [CrossRef]
- Schenk, R.; Eggli, P.; Fleisch, H.; Rosini, S. Quantitative Morphometric Evaluation of the Inhibitory Activity of New Aminobisphosphonates on Bone Resorption in the Rat. *Calcif. Tissue. Int.* 1986, *38*, 342–349. [CrossRef]
- 51. Russell, R.G.G.; Watts, N.B.; Ebetino, F.H.; Rogers, M.J. Mechanisms of Action of Bisphosphonates: Similarities and Differences and Their Potential Influence on Clinical Efficacy. *Osteoporos. Int.* **2008**, *19*, 733–759. [CrossRef]
- Amin, D.; Cornell, S.A.; Gustafson, S.K.; Needle, S.J.; Ullrich, J.W.; Bilder, G.E.; Perrone, M.H. Bisphosphonates Used for the Treatment of Bone Disorders Inhibit Squalene Synthase and Cholesterol Biosynthesis. J. Lipid. Res. 1992, 33, 1657–1663. [CrossRef]
- Amin, D.; Cornell, S.A.; Perrone, M.H.; Bilder, G.E. 1-Hydroxy-3-(Methylpentylamino)-Propylidene-1,1-Bisphosphonic Acid as a Potent Inhibitor of Squalene Synthase. *Arzneimittelforschung* 1996, 46, 759–762.
- Luckman, S.P.; Hughes, D.E.; Coxon, F.P.; Russell, R.G.G.; Rogers, M.J. Nitrogen-Containing Bisphosphonates Inhibit the Mevalonate Pathway and Prevent Post-Translational Prenylation of GTP-Binding Proteins, Including Ras. *J. Bone Min. Res.* 1998, 13, 581–589. [CrossRef] [PubMed]
- 55. Fisher, J.E.; Rogers, M.J.; Halasy, J.M.; Luckman, S.P.; Hughes, D.E.; Masarachia, P.J.; Wesolowski, G.; Russell, R.G.; Rodan, G.A.; Reszka, A.A. Alendronate Mechanism of Action: Geranylgeraniol, an Intermediate in the Mevalonate Pathway, Prevents Inhibition of Osteoclast Formation, Bone Resorption, and Kinase Activation in Vitro. *Proc. Natl. Acad. Sci. USA* 1999, *96*, 133–138. [CrossRef] [PubMed]
- Van Beek, E.; Pieterman, E.; Cohen, L.; Löwik, C.; Papapoulos, S. Nitrogen-Containing Bisphosphonates Inhibit Isopentenyl Pyrophosphate Isomerase/Farnesyl Pyrophosphate Synthase Activity with Relative Potencies Corresponding to Their Antiresorptive Potencies in Vitroandin Vivo. *Biochem. Biophys. Res. Commun.* 1999, 255, 491–494. [CrossRef] [PubMed]
- 57. Dunford, J.E.; Thompson, K.; Coxon, F.P.; Luckman, S.P.; Hahn, F.M.; Poulter, C.D.; Ebetino, F.H.; Rogers, M.J. Structure-Activity Relationships for Inhibition of Farnesyl Diphosphate Synthase in Vitro and Inhibition of Bone Resorption in Vivo by Nitrogen-Containing Bisphosphonates. *J. Pharmacol. Exp. Ther.* **2001**, *296*, 235–242.
- Kavanagh, K.L.; Guo, K.; Dunford, J.E.; Wu, X.; Knapp, S.; Ebetino, F.H.; Rogers, M.J.; Russell, R.G.G.; Oppermann, U. The Molecular Mechanism of Nitrogen-Containing Bisphosphonates as Antiosteoporosis Drugs. *Proc. Natl. Acad. Sci. USA* 2006, 103, 7829–7834. [CrossRef]
- Luckman, S.P.; Coxon, F.P.; Ebetino, F.H.; Russell, R.G.G.; Rogers, M.J. Heterocycle-Containing Bisphosphonates Cause Apoptosis and Inhibit Bone Resorption by Preventing Protein Prenylation: Evidence from Structure-Activity Relationships in J774 Macrophages. J. Bone. Min. Res. 1998, 13, 1668–1678. [CrossRef]
- 60. Sahni, M.; Guenther, H.L.; Fleisch, H.; Collin, P.; Martin, T.J. Bisphosphonates Act on Rat Bone Resorption through the Mediation of Osteoblasts. *J. Clin. Invest.* **1993**, *91*, 2004–2011. [CrossRef]
- Nishikawa, M.; Akatsu, T.; Katayama, Y.; Yasutomo, Y.; Kado, S.; Kugai, N.; Yamamoto, M.; Nagata, N. Bisphosphonates Act on Osteoblastic Cells and Inhibit Osteoclast Formation in Mouse Marrow Cultures. *Bone* 1996, 18, 9–14. [CrossRef]
- 62. Giuliani, N.; Pedrazzoni, M.; Passeri, G.; Girasole, G. Pass Bisphosphonates Inhibit IL-6 Production by Human Osteoblast-like Cells. *Scand. J. Rheumatol.* **1998**, *27*, 38–41. [CrossRef]
- 63. D'Aoust, P.; McCulloch, C.A.G.; Tenenbaum, H.C.; Lekic, P.C. Etidronate (HEBP) Promotes Osteoblast Differentiation and Wound Closure in Rat Calvaria. *Cell Tissue Res.* **2000**, *302*, 353–363. [CrossRef]
- 64. Itoh, F.; Aoyagi, S.; Furihata-Komatsu, H.; Aoki, M.; Kusama, H.; Kojima, M.; Kogo, H. Clodronate Stimulates Osteoblast Differentiation in ST2 and MC3T3-E1 Cells and Rat Organ Cultures. *Eur. J. Pharmacol.* **2003**, 477, 9–16. [CrossRef] [PubMed]
- 65. Im, G.-I.; Qureshi, S.A.; Kenney, J.; Rubash, H.E.; Shanbhag, A.S. Osteoblast Proliferation and Maturation by Bisphosphonates. *Biomaterials* **2004**, *25*, 4105–4115. [CrossRef] [PubMed]
- Bellido, T.; Plotkin, L.I. Novel Actions of Bisphosphonates in Bone: Preservation of Osteoblast and Osteocyte Viability. *Bone* 2011, 49, 50–55. [CrossRef] [PubMed]
- 67. Ohno, K.; Mori, K.; Orita, M.; Takeuchi, M. Computational Insights into Binding of Bisphosphates to Farnesyl Pyrophosphate Synthase. *CMC* 2011, *18*, 220–233. [CrossRef]
- Rogers, M.J.; Mönkkönen, J.; Munoz, M.A. Molecular Mechanisms of Action of Bisphosphonates and New Insights into Their Effects Outside the Skeleton. *Bone* 2020, 139, 115493. [CrossRef]
- 69. Ebetino, F.H.; Sun, S.; Cherian, P.; Roshandel, S.; Neighbors, J.D.; Hu, E.; Dunford, J.E.; Sedghizadeh, P.P.; McKenna, C.E.; Srinivasan, V.; et al. Bisphosphonates: The Role of Chemistry in Understanding Their Biological Actions and Structure-Activity Relationships, and New Directions for Their Therapeutic Use. *Bone* **2022**, *156*, 116289. [CrossRef]
- Varela, I.; Pereira, S.; Ugalde, A.P.; Navarro, C.L.; Suárez, M.F.; Cau, P.; Cadiñanos, J.; Osorio, F.G.; Foray, N.; Cobo, J.; et al. Combined Treatment with Statins and Aminobisphosphonates Extends Longevity in a Mouse Model of Human Premature Aging. *Nat. Med.* 2008, 14, 767–772. [CrossRef]
- 71. Misra, J.; Mohanty, S.T.; Madan, S.; Fernandes, J.A.; Hal Ebetino, F.; Russell, R.G.G.; Bellantuono, I. Zoledronate Attenuates Accumulation of DNA Damage in Mesenchymal Stem Cells and Protects Their Function. *Stem. Cells* 2016, 34, 756–767. [CrossRef]
- Gordon, L.B.; Kleinman, M.E.; Massaro, J.; D'Agostino, R.B.; Shappell, H.; Gerhard-Herman, M.; Smoot, L.B.; Gordon, C.M.; Cleveland, R.H.; Nazarian, A.; et al. Clinical Trial of the Protein Farnesylation Inhibitors Lonafarnib, Pravastatin, and Zoledronic Acid in Children With Hutchinson-Gilford Progeria Syndrome. *Circulation* 2016, 134, 114–125. [CrossRef]

- 73. Eidtmann, H.; De Boer, R.; Bundred, N.; Llombart-Cussac, A.; Davidson, N.; Neven, P.; Von Minckwitz, G.; Miller, J.; Schenk, N.; Coleman, R. Efficacy of Zoledronic Acid in Postmenopausal Women with Early Breast Cancer Receiving Adjuvant Letrozole: 36-Month Results of the ZO-FAST Study. Ann. Oncol. 2010, 21, 2188–2194. [CrossRef]
- 74. Gnant, M.; Mlineritsch, B.; Stoeger, H.; Luschin-Ebengreuth, G.; Heck, D.; Menzel, C.; Jakesz, R.; Seifert, M.; Hubalek, M.; Pristauz, G.; et al. Adjuvant Endocrine Therapy plus Zoledronic Acid in Premenopausal Women with Early-Stage Breast Cancer: 62-Month Follow-up from the ABCSG-12 Randomised Trial. *Lancet Oncol.* 2011, *12*, 631–641. [CrossRef] [PubMed]
- 75. Brufsky, A.M.; Harker, W.G.; Beck, J.T.; Bosserman, L.; Vogel, C.; Seidler, C.; Jin, L.; Warsi, G.; Argonza-Aviles, E.; Hohneker, J.; et al. Final 5-Year Results of Z-FAST Trial: Adjuvant Zoledronic Acid Maintains Bone Mass in Postmenopausal Breast Cancer Patients Receiving Letrozole. *Cancer* 2012, 118, 1192–1201. [CrossRef] [PubMed]
- 76. Friedl, T.W.P.; Fehm, T.; Müller, V.; Lichtenegger, W.; Blohmer, J.; Lorenz, R.; Forstbauer, H.; Fink, V.; Bekes, I.; Huober, J.; et al. Prognosis of Patients With Early Breast Cancer Receiving 5 Years vs 2 Years of Adjuvant Bisphosphonate Treatment: A Phase 3 Randomized Clinical Trial. *JAMA Oncol.* 2021, 7, 1149. [CrossRef]
- 77. Early Breast Cancer Trialists' Collaborative Group. Adjuvant Bisphosphonate Treatment in Early Breast Cancer: Meta-Analyses of Individual Patient Data from Randomised Trials. *Lancet* 2015, *386*, 1353–1361. [CrossRef] [PubMed]
- Coleman, R.E. Adjuvant Bone-Targeted Therapy to Prevent Metastasis: Lessons from the AZURE Study. Curr. Opin. Support. Palliat. Care 2012, 6, 322–329. [CrossRef] [PubMed]
- 79. Coleman, R.E.; Brown, J.; Holen, I. Bone Metastases. In *Abeloff's Clinical. Oncology*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 809–830.e3. ISBN 978-0-323-47674-4.
- 80. Macedo, F.; Ladeira, K.; Pinho, F.; Saraiva, N.; Bonito, N.; Pinto, L.; Gonçalves, F. Bone Metastases: An Overview. *Oncol. Rev.* 2017, 11, 321. [CrossRef]
- 81. Coleman, R.E. Skeletal Complications of Malignancy. Cancer 1997, 80, 1588–1594. [CrossRef]
- Czernin, J.; Satyamurthy, N.; Schiepers, C. Molecular Mechanisms of Bone ¹⁸ F-NaF Deposition. J. Nucl. Med. 2010, 51, 1826–1829. [CrossRef]
- Even-Sapir, E.; Metser, U.; Flusser, G.; Zuriel, L.; Kollender, Y.; Lerman, H.; Lievshitz, G.; Ron, I.; Mishani, E. Assessment of Malignant Skeletal Disease: Initial Experience with 18F-Fluoride PET/CT and Comparison between 18F-Fluoride PET and 18F-Fluoride PET/CT. J. Nucl. Med. 2004, 45, 272–278.
- Withofs, N.; Grayet, B.; Tancredi, T.; Rorive, A.; Mella, C.; Giacomelli, F.; Mievis, F.; Aerts, J.; Waltregny, D.; Jerusalem, G.; et al. 18F-Fluoride PET/CT for Assessing Bone Involvement in Prostate and Breast Cancers. *Nucl. Med. Commun.* 2011, 32, 168–176. [CrossRef]
- Damle, N.A.; Bal, C.; Bandopadhyaya, G.P.; Kumar, L.; Kumar, P.; Malhotra, A.; Lata, S. The Role of 18F-Fluoride PET-CT in the Detection of Bone Metastases in Patients with Breast, Lung and Prostate Carcinoma: A Comparison with FDG PET/CT and 99mTc-MDP Bone Scan. *Jpn. J. Radiol.* 2013, *31*, 262–269. [CrossRef] [PubMed]
- Apolo, A.B.; Lindenberg, L.; Shih, J.H.; Mena, E.; Kim, J.W.; Park, J.C.; Alikhani, A.; McKinney, Y.Y.; Weaver, J.; Turkbey, B.; et al. Prospective Study Evaluating Na ¹⁸F PET/CT in Predicting Clinical Outcomes and Survival in Advanced Prostate Cancer. *J. Nucl. Med.* 2016, 57, 886–892. [CrossRef] [PubMed]
- 87. Silberstein, E.B. Teletherapy and Radiopharmaceutical Therapy of Painful Bone Metastases. *Semin. Nucl. Med.* **2005**, *35*, 152–158. [CrossRef] [PubMed]
- Silberstein, E.B. The Treatment of Painful Osseous Metastases with Phosphorus-32-Labeled Phosphates. Semin. Oncol. 1993, 20, 10–21. [PubMed]
- 89. Silberstein, E.B.; Elgazzar, A.H.; Kapilivsky, A. Phosphorus-32 Radiopharmaceuticals for the Treatment of Painful Osseous Metastases. *Semin. Nucl. Med.* **1992**, 22, 17–27. [CrossRef] [PubMed]
- 90. Silberstein, E.B.; Williams, C. Strontium-89 Therapy for the Pain of Osseous Metastases. J. Nucl. Med. 1985, 26, 345–348. [PubMed]
- Porter, A.T.; McEwan, A.J.B.; Powe, J.E.; Reid, R.; McGowan, D.G.; Lukka, H.; Sathyanarayana, J.R.; Yakemchuk, V.N.; Thomas, G.M.; Erlich, L.E.; et al. Results of a Randomized Phase-III Trial to Evaluate the Efficacy of Strontium-89 Adjuvant to Local Field External Beam Irradiation in the Management of Endocrine Resistant Metastatic Prostate Cancer. *Int. J. Radiat. Oncol. Biol. Phys.* 1993, 25, 805–813. [CrossRef]
- Smeland, S.; Erikstein, B.; Aas, M.; Skovlund, E.; Hess, S.L.; Fosså, S.D. Role of Strontium-89 as Adjuvant to Palliative External Beam Radiotherapy Is Questionable: Results of a Double-Blind Randomized Study. *Int. J. Radiat. Oncol. Biol. Phys.* 2003, 56, 1397–1404. [CrossRef]
- Oosterhof, G.O.N.; Roberts, J.T.; De Reijke, T.M.; Engelholm, S.A.; Horenblas, S.; Von Der Maase, H.; Neymark, N.; Debois, M.; Collette, L. Strontium-89 Chloride versus Palliative Local Field Radiotherapy in Patients with Hormonal Escaped Prostate Cancer: A Phase III Study of the European Organisation for Research and Treatment of Cancer Genitourinary Group. *Eur. Urol.* 2003, 44, 519–526. [CrossRef]
- 94. Tu, S.-M.; Millikan, R.E.; Mengistu, B.; Delpassand, E.S.; Amato, R.J.; Pagliaro, L.C.; Daliani, D.; Papandreou, C.N.; Smith, T.L.; Kim, J.; et al. Bone-Targeted Therapy for Advanced Androgen-Independent Carcinoma of the Prostate: A Randomised Phase II Trial. *Lancet* 2001, 357, 336–341. [CrossRef]

- 95. James, N.D.; Pirrie, S.J.; Pope, A.M.; Barton, D.; Andronis, L.; Goranitis, I.; Collins, S.; Daunton, A.; McLaren, D.; O'Sullivan, J.; et al. Clinical Outcomes and Survival Following Treatment of Metastatic Castrate-Refractory Prostate Cancer with Docetaxel Alone or With Strontium-89, Zoledronic Acid, or Both: The TRAPEZE Randomized Clinical Trial. *JAMA Oncol.* 2016, 2, 493. [CrossRef] [PubMed]
- Parker, C.; Nilsson, S.; Heinrich, D.; Helle, S.I.; O'Sullivan, J.M.; Fosså, S.D.; Chodacki, A.; Wiechno, P.; Logue, J.; Seke, M.; et al. Alpha Emitter Radium-223 and Survival in Metastatic Prostate Cancer. N. Engl. J. Med. 2013, 369, 213–223. [CrossRef] [PubMed]
- 97. Subramanian, G.; McAfee, J.G.; Blair, R.J.; Kallfelz, F.A.; Thomas, F.D. Technetium-99m-Methylene Diphosphonate--a Superior Agent for Skeletal Imaging: Comparison with Other Technetium Complexes. J. Nucl. Med. 1975, 16, 744–755.
- Bevan, J.A.; Tofe, A.J.; Benedict, J.J.; Francis, M.D.; Barnett, B.L. Tc-99m HMDP (Hydroxymethylene Diphosphonate): A Radiopharmaceutical for Skeletal and Acute Myocardial Infarct Imaging. I. Synthesis and Distribution in Animals. *J. Nucl. Med.* 1980, 21, 961–966. [PubMed]
- 99. Domstad, P.A.; Coupal, J.J.; Kim, E.E.; Blake, J.S.; DeLand, F.H. 99mTc-Hydroxymethane Diphosphonate: A New Bone Imaging Agent with a Low Tin Content. *Radiology* **1980**, *136*, 209–211. [CrossRef] [PubMed]
- Castronovo, F.P.; Callahan, R.J. New Bone Scanning Agent: 99mTc-Labeled 1-Hydroxy-Ethylidene-1, 1-Disodium Phosphonate. J. Nucl. Med. 1972, 13, 823–827. [PubMed]
- 101. Subramanian, G.; McAfee, J.G.; Blair, R.J.; Mehter, A.; Connor, T. 99m Tc-EHDP: A Potential Radiopharmaceutical for Skeletal Imaging. *J. Nucl. Med.* **1972**, *13*, 947–950.
- 102. Yano, Y.; McRae, J.; Van Dyke, D.C.; Anger, H.O. Technetium-99m-Labeled Stannous Ethane-1-Hydroxy-1 1-Diphosphonate: A New Bone Scanning Agent. J. Nucl. Med. 1973, 14, 73–78.
- Pendergrass, H.P.; Potsaid, M.S.; Castronovo, F.P. The Clinical Use of 99m Tc-Diphosphonate (HEDSPA). A New Agent for Skeletal Imaging. *Radiology* 1973, 107, 557–562. [CrossRef]
- 104. Wilson, G.M.; Pinkerton, T.C. Determination of Charge and Size of Technetium Diphosphonate Complexes by Anion-Exchange Liquid Chromatography. *Anal. Chem.* **1985**, *57*, 246–253. [CrossRef]
- 105. Tanabe, S.; Zodda, J.P.; Deutsch, E.; Heineman, W.R. Effect of pH on the Formation of Tc(NaBH4)-MDP Radiopharmaceutical Analogues. *Int. J. Appl. Radiat. Isot.* **1983**, *34*, 1577–1584. [CrossRef] [PubMed]
- Libson, K.; Deutsch, E.; Barnett, B.L. Structural Characterization of a Technetium-99-Diphosphonate Complex. Implications for the Chemistry of Technetium-99m Skeletal Imaging Agents. J. Am. Chem. Soc. 1980, 102, 2476–2478. [CrossRef]
- 107. Sharma, P.; Kumar, R.; Singh, H.; Bal, C.; Julka, P.K.; Thulkar, S.; Malhotra, A. Indeterminate Lesions on Planar Bone Scintigraphy in Lung Cancer Patients: SPECT, CT or SPECT-CT? *Skelet. Radiol.* 2012, 41, 843–850. [CrossRef] [PubMed]
- Chang, M.-C.; Chen, J.-H.; Liang, J.-A.; Lin, C.-C.; Yang, K.-T.; Cheng, K.-Y.; Yeh, J.-J.; Kao, C.-H. Meta-Analysis: Comparison of F-18 Fluorodeoxyglucose-Positron Emission Tomography and Bone Scintigraphy in the Detection of Bone Metastasis in Patients with Lung Cancer. Acad. Radiol. 2012, 19, 349–357. [CrossRef] [PubMed]
- Kubo, T.; Furuta, T.; Sakuda, T.; Ochi, M.; Adachi, N. Conventional 99mTc-(Hydroxy) Methylene Diphosphate Remains Useful to Predict Osteosarcoma Response to Neoadjuvant Chemotherapy: Individual Patient Data and Aggregate Data Meta-Analyses. *Medicine* 2018, 97, e13308. [CrossRef] [PubMed]
- 110. Lee, I.; Byun, B.H.; Lim, I.; Kim, B.I.; Kong, C.-B.; Song, W.S.; Cho, W.H.; Koh, J.-S.; Lim, S.M. Comparison of 99mTc-Methyl Diphosphonate Bone Scintigraphy and 18F-Fluorodeoxyglucose Positron Emission Tomography/Computed Tomography to Predict Histologic Response to Neoadjuvant Chemotherapy in Patients with Osteosarcoma. *Medicine* 2018, 97, e12318. [CrossRef] [PubMed]
- 111. Dennis, E.R.; Jia, X.; Mezheritskiy, I.S.; Stephenson, R.D.; Schoder, H.; Fox, J.J.; Heller, G.; Scher, H.I.; Larson, S.M.; Morris, M.J. Bone Scan Index: A Quantitative Treatment Response Biomarker for Castration-Resistant Metastatic Prostate Cancer. J. Clin. Oncol. 2012, 30, 519–524. [CrossRef]
- 112. Rubini, G.; Lauriero, F.; Rubini, D.; D'Addabbo, A. 99Tcm-MDP Global Skeletal Uptake and Markers of Bone Metabolism in Patients with Bone Diseases. *Nucl. Med. Commun.* **1993**, *14*, 567–572. [CrossRef]
- 113. Griffith, K.; Pearson, D.; Parker, C.; Thorpe, S.; Vincent, R.M.; Hosking, D.J. The Use of a Whole Body Index with Bone Scintigraphy to Monitor the Response to Therapy in Paget's Disease. *Nucl. Med. Commun.* **2001**, *22*, 1069–1075. [CrossRef]
- 114. Israel, O.; Front, D.; Hardoff, R.; Ish-Shalom, S.; Jerushalmi, J.; Kolodny, G.M. In Vivo SPECT Quantitation of Bone Metabolism in Hyperparathyroidism and Thyrotoxicosis. *J. Nucl. Med.* **1991**, *32*, 1157–1161.
- 115. Brenner, A.I.; Koshy, J.; Morey, J.; Lin, C.; DiPoce, J. The Bone Scan. Semin. Nucl. Med. 2012, 42, 11–26. [CrossRef] [PubMed]
- 116. Schwarz, A.; Kloss, G. Technetium-99m DPD-A New Skeletal Imaging Agent. J. Nucl. Med. 1981, 22, P77.
- 117. Perugini, E.; Guidalotti, P.L.; Salvi, F.; Cooke, R.M.T.; Pettinato, C.; Riva, L.; Leone, O.; Farsad, M.; Ciliberti, P.; Bacchi-Reggiani, L.; et al. Noninvasive Etiologic Diagnosis of Cardiac Amyloidosis Using 99mTc-3,3-Diphosphono-1,2-Propanodicarboxylic Acid Scintigraphy. J. Am. Coll. Cardiol. 2005, 46, 1076–1084. [CrossRef] [PubMed]
- 118. Rapezzi, C.; Quarta, C.C.; Guidalotti, P.L.; Longhi, S.; Pettinato, C.; Leone, O.; Ferlini, A.; Salvi, F.; Gallo, P.; Gagliardi, C.; et al. Usefulness and Limitations of ^{99m}Tc-3,3-Diphosphono-1,2-Propanodicarboxylic Acid Scintigraphy in the Aetiological Diagnosis of Amyloidotic Cardiomyopathy. *Eur. J. Nucl. Med. Mol. Imaging* **2011**, *38*, 470–478. [CrossRef] [PubMed]
- Quarta, C.C.; Zheng, J.; Hutt, D.; Grigore, S.F.; Manwani, R.; Sachchithanantham, S.; Mahmood, S.A.; Whelan, C.J.; Fontana, M.; Martinez-Naharro, A.; et al. ^{99m}Tc-DPD Scintigraphy in Immunoglobulin Light Chain (AL) Cardiac Amyloidosis. *Eur. Heart J. Cardiovasc. Imaging* 2021, 22, 1304–1311. [CrossRef]

- 120. Turner, J.H.; Martindale, A.A.; Sorby, P.; Hetherington, E.L.; Fleay, R.F.; Hoffman, R.F.; Claringbold, P.G. Samarium-153 EDTMP Therapy of Disseminated Skeletal Metastasis. *Eur. J. Nucl. Med.* **1989**, *15*, 784–795. [CrossRef]
- 121. Turner, J.H.; Claringbold, P.G.; Hetherington, E.L.; Sorby, P.; Martindale, A.A. A Phase I Study of Samarium-153 Ethylenediaminetetramethylene Phosphonate Therapy for Disseminated Skeletal Metastases. J. Clin. Oncol. 1989, 7, 1926–1931. [CrossRef]
- 122. Serafini, A.N.; Houston, S.J.; Resche, I.; Quick, D.P.; Grund, F.M.; Ell, P.J.; Bertrand, A.; Ahmann, F.R.; Orihuela, E.; Reid, R.H.; et al. Palliation of Pain Associated with Metastatic Bone Cancer Using Samarium-153 Lexidronam: A Double-Blind Placebo-Controlled Clinical Trial. JCO 1998, 16, 1574–1581. [CrossRef]
- 123. Sartor, O.; Reid, R.H.; Hoskin, P.J.; Quick, D.P.; Ell, P.J.; Coleman, R.E.; Kotler, J.A.; Freeman, L.M.; Olivier, P. Samarium-153-Lexidronam Complex for Treatment of Painful Bone Metastases in Hormone-Refractory Prostate Cancer. Urology 2004, 63, 940–945. [CrossRef]
- 124. Verbeke, K.; Rozenski, J.; Cleynhens, B.; Vanbilloen, H.; De Groot, T.; Weyns, N.; Bormans, G.; Verbruggen, A. Development of a Conjugate of ^{99m} Tc-EC with Aminomethylenediphosphonate in the Search for a Bone Tracer with Fast Clearance from Soft Tissue. *Bioconjug. Chem.* 2002, 13, 16–22. [CrossRef]
- 125. Ogawa, K.; Mukai, T.; Inoue, Y.; Ono, M.; Saji, H. Development of a Novel 99mTc-Chelate-Conjugated Bisphosphonate with High Affinity for Bone as a Bone Scintigraphic Agent. *J. Nucl. Med.* **2006**, *47*, 2042–2047. [PubMed]
- 126. Ogawa, K.; Mukai, T.; Kawai, K.; Takamura, N.; Hanaoka, H.; Hashimoto, K.; Shiba, K.; Mori, H.; Saji, H. Usefulness of Competitive Inhibitors of Protein Binding for Improving the Pharmacokinetics of ¹⁸⁶Re-MAG3-Conjugated Bisphosphonate (¹⁸⁶Re-MAG3-HBP), an Agent for Treatment of Painful Bone Metastases. *Eur. J. Nucl. Med. Mol. Imaging* **2009**, *36*, 115–121. [CrossRef] [PubMed]
- 127. Liu, L.; Zhong, G.; Wei, Y.; Zhang, M.; Wang, X. Synthesis and Biological Evaluation of a Novel 99mTc Complex of HYNIC-Conjugated Aminomethylenediphosphonate as a Potential Bone Imaging Agent. J. Radioanal. Nucl. Chem. 2011, 288, 467–473. [CrossRef]
- 128. Yazdani, A.; Bilton, H.; Vito, A.; Genady, A.R.; Rathmann, S.M.; Ahmad, Z.; Janzen, N.; Czorny, S.; Zeglis, B.M.; Francesconi, L.C.; et al. A Bone-Seeking Trans-Cyclooctene for Pretargeting and Bioorthogonal Chemistry: A Proof of Concept Study Using 99mTcand 177Lu-Labeled Tetrazines. J. Med. Chem. 2016, 59, 9381–9389. [CrossRef]
- Palma, E.; Oliveira, B.L.; Correia, J.D.G.; Gano, L.; Maria, L.; Santos, I.C.; Santos, I. A New Bisphosphonate-Containing (99m)Tc(I) Tricarbonyl Complex Potentially Useful as Bone-Seeking Agent: Synthesis and Biological Evaluation. *J. Biol. Inorg. Chem.* 2007, 12, 667–679. [CrossRef]
- Palma, E.; Correia, J.D.G.; Oliveira, B.L.; Gano, L.; Santos, I.C.; Santos, I. ^{99m}Tc(CO)3-Labeled Pamidronate and Alendronate for Bone Imaging. *Dalton Trans.* 2011, 40, 2787–2796. [CrossRef]
- 131. Torres Martin de Rosales, R.; Finucane, C.; Foster, J.; Mather, S.J.; Blower, P.J. ¹⁸⁸Re(CO)3-Dipicolylamine-Alendronate: A New Bisphosphonate Conjugate for the Radiotherapy of Bone Metastases. *Bioconjug. Chem.* **2010**, *21*, 811–815. [CrossRef]
- 132. Keeling, G.P.; Sherin, B.; Kim, J.; San Juan, B.; Grus, T.; Eykyn, T.R.; Rösch, F.; Smith, G.E.; Blower, P.J.; Terry, S.Y.A.; et al. [⁶⁸Ga]Ga-THP-Pam: A Bisphosphonate PET Tracer with Facile Radiolabeling and Broad Calcium Mineral Affinity. *Bioconjug. Chem.* 2021, 32, 1276–1289. [CrossRef]
- 133. Hong, H.; Ploessl, K.; Zha, Z.; Wang, H.; Guo, R.; Xie, Q.; Zhu, H.; Yang, Z.; Zhu, L.; Kung, H.F. Development and Validation of a Kit Formulation of [⁶⁸Ga]Ga-P15-041 as a Bone Imaging Agent. *Appl. Radiat. Isot.* 2021, 169, 109485. [CrossRef]
- 134. Zha, Z.; Wu, Z.; Choi, S.R.; Ploessl, K.; Smith, M.; Alexoff, D.; Zhu, L.; Kung, H.F. A New [⁶⁸Ga]Ga-HBED-CC-Bisphosphonate as a Bone Imaging Agent. *Mol. Pharm.* **2020**, *17*, 1674–1684. [CrossRef]
- 135. Alexoff, D.; Doot, R.; Pryma, D.; Schubert, E.; Wu, Z.; Zha, Z.; Choi, S.; Ploessl, K.; Lee, S.; Kao, C.; et al. Preliminary Kinetic Analysis of [⁶⁸Ga]P15-041, a Novel ⁶⁸Ga Labeled Bisphosphonate, from First-in-Human Studies. J. Nucl. Med. 2017, 58, 388.
- 136. Doot, R.K.; Young, A.J.; Daube-Witherspoon, M.E.; Alexoff, D.; Labban, K.J.; Lee, H.; Wu, Z.; Zha, Z.; Choi, S.R.; Ploessl, K.H.; et al. Biodistribution, Dosimetry, and Temporal Signal-to-Noise Ratio Analyses of Normal and Cancer Uptake of [⁶⁸Ga]Ga-P15-041, a Gallium-68 Labeled Bisphosphonate, from First-in-Human Studies. *Nucl. Med. Biol.* **2020**, *86*, 1–8. [CrossRef]
- 137. Guo, R.; Meng, X.; Wang, F.; Yu, J.; Xie, Q.; Zhao, W.; Zhu, L.; Kung, H.F.; Yang, Z.; Li, N. ⁶⁸Ga-P15-041, A Novel Bone Imaging Agent for Diagnosis of Bone Metastases. *Front. Oncol.* **2021**, *11*, 766851. [CrossRef] [PubMed]
- Suzuki, K.; Satake, M.; Suwada, J.; Oshikiri, S.; Ashino, H.; Dozono, H.; Hino, A.; Kasahara, H.; Minamizawa, T. Synthesis and Evaluation of a Novel ⁶⁸Ga-Chelate-Conjugated Bisphosphonate as a Bone-Seeking Agent for PET Imaging. *Nucl. Med. Biol.* 2011, 38, 1011–1018. [CrossRef] [PubMed]
- 139. Ashhar, Z.; Yusof, N.A.; Ahmad Saad, F.F.; Mohd Nor, S.M.; Mohammad, F.; Bahrin Wan Kamal, W.H.; Hassan, M.H.; Ahmad Hassali, H.; Al-Lohedan, H.A. Preparation, Characterization, and Radiolabeling of [⁶⁸Ga]Ga-NODAGA-Pamidronic Acid: A Potential PET Bone Imaging Agent. *Molecules* 2020, 25, 2668. [CrossRef] [PubMed]
- Holub, J.; Meckel, M.; Kubíček, V.; Rösch, F.; Hermann, P. Gallium(III) Complexes of NOTA-Bis (Phosphonate) Conjugates as PET Radiotracers for Bone Imaging. *Contrast Media. Mol. Imaging* 2015, 10, 122–134. [CrossRef] [PubMed]
- 141. Passah, A.; Tripathi, M.; Ballal, S.; Yadav, M.P.; Kumar, R.; Roesch, F.; Meckel, M.; Sarathi Chakraborty, P.; Bal, C. Evaluation of Bone-Seeking Novel Radiotracer ⁶⁸Ga-NO2AP-Bisphosphonate for the Detection of Skeletal Metastases in Carcinoma Breast. *Eur. J. Nucl. Med. Mol. Imaging* **2017**, *44*, 41–49. [CrossRef] [PubMed]
- 142. Pfannkuchen, N.; Bergmann, R.; Pietzsch, J.; Bachmann, M.; Roesch, F. DOTAZOL and NODAGAZOL for Theranostics of Bone Metastases. J. Nucl. Med. 2017, 58, 324.

- 143. Lawal, I.; Ebenhan, T.; Mahapane, J.; Vorster, M.; Mokoala, K.; Meckel, M.; Rosch, F.; Sathekge, M. Assessment of Skeletal Metastasis in Prostate Cancer Staging: An Intra-Individual Comparison of ⁶⁸Ga-PSMA PET/CT, ⁶⁸Ga-NODAGA-Zoledronate PET/CT, and ^{99m}Tc-MDP Bone Scan. J. Nucl. Med. **2020**, *61*, 1256.
- 144. Lawal, I.O.; Mokoala, K.M.G.; Mahapane, J.; Kleyhans, J.; Meckel, M.; Vorster, M.; Ebenhan, T.; Rösch, F.; Sathekge, M.M. A Prospective Intra-Individual Comparison of [⁶⁸Ga]Ga-PSMA-11 PET/CT, [⁶⁸Ga]Ga-NODAGAZOL PET/CT, and [^{99m}Tc]Tc-MDP Bone Scintigraphy for Radionuclide Imaging of Prostate Cancer Skeletal Metastases. *Eur. J. Nucl. Med. Mol. Imaging* 2021, 48, 134–142. [CrossRef]
- 145. Ndlovu, H.; Lawal, I.; Popoola, G.; Brits, B.; Mokoala, K.; Mahapane, J.; Davis, C.; Sathekge, M. PET Imaging of Atherosclerotic Plaque Calcification with [⁶⁸Ga]Ga-NODAGAZOL: Correlation of Uptake with Cardiovascular Risk Profile of Patients. *J. Nucl. Med.* **2022**, *63*, 2222.
- 146. Ndlovu, H.; Lawal, I.O.; Popoola, G.O.; Brits, B.; Mokoala, K.M.G.; Maserumule, L.C.; Hlongwa, K.N.; Mahapane, J.; Davis, C.; Sathekge, M.M. [⁶⁸Ga]Ga-NODAGAZOL Uptake in Atherosclerotic Plaques Correlates with the Cardiovascular Risk Profile of Patients. Ann. Nucl. Med. 2022, 36, 684–692. [CrossRef] [PubMed]
- 147. Fersing, C.; Bouhlel, A.; Cantelli, C.; Garrigue, P.; Lisowski, V.; Guillet, B. A Comprehensive Review of Non-Covalent Radiofluorination Approaches Using Aluminum [¹⁸F]Fluoride: Will [¹⁸F]AIF Replace 68Ga for Metal Chelate Labeling? *Molecules* 2019, 24, 2866. [CrossRef] [PubMed]
- 148. Hassan, H.; Othman, M.F.; Abdul Razak, H.R.; Zakaria, Z.A.; Ahmad Saad, F.F.; Osman, M.A.; Yi, L.H.; Ashhar, Z.; Idris, J.; Abdul Hamid, M.H.N.; et al. Preparation, Optimisation, and In Vitro Evaluation of [¹⁸F]AlF-NOTA-Pamidronic Acid for Bone Imaging PET. *Molecules* 2022, 27, 7969. [CrossRef] [PubMed]
- 149. Fersing, C.; Masurier, N.; Rubira, L.; Deshayes, E.; Lisowski, V. AAZTA-Derived Chelators for the Design of Innovative Radiopharmaceuticals with Theranostic Applications. *Pharmaceuticals* **2022**, *15*, 234. [CrossRef]
- 150. Wu, Z.; Zha, Z.; Choi, S.R.; Plössl, K.; Zhu, L.; Kung, H.F. New 68Ga-PhenA Bisphosphonates as Potential Bone Imaging Agents. *Nucl. Med. Biol.* **2016**, *43*, 360–371. [CrossRef]
- 151. Kubícek, V.; Rudovský, J.; Kotek, J.; Hermann, P.; Vander Elst, L.; Muller, R.N.; Kolar, Z.I.; Wolterbeek, H.T.; Peters, J.A.; Lukes, I. A Bisphosphonate Monoamide Analogue of DOTA: A Potential Agent for Bone Targeting. J. Am. Chem. Soc. 2005, 127, 16477–16485. [CrossRef]
- Fellner, M.; Biesalski, B.; Bausbacher, N.; Kubícek, V.; Hermann, P.; Rösch, F.; Thews, O. ⁶⁸Ga-BPAMD: PET-Imaging of Bone Metastases with a Generator Based Positron Emitter. *Nucl. Med. Biol.* 2012, *39*, 993–999. [CrossRef]
- 153. Fellner, M.; Baum, R.P.; Kubícek, V.; Hermann, P.; Lukes, I.; Prasad, V.; Rösch, F. PET/CT Imaging of Osteoblastic Bone Metastases with ⁶⁸Ga-Bisphosphonates: First Human Study. *Eur. J. Nucl. Med. Mol. Imaging* **2010**, *37*, 834. [CrossRef]
- 154. Mueller, D.; Klette, I.; Baum, R. Clinical Routine Production of ¹⁷⁷Lu-BPAMD. J. Nucl. Med. 2013, 54, 1191.
- 155. Meckel, M.; Nauth, A.; Timpe, J.; Zhernosekov, K.; Puranik, A.D.; Baum, R.P.; Rösch, F. Development of a [177Lu]BPAMD Labeling Kit and an Automated Synthesis Module for Routine Bone Targeted Endoradiotherapy. *Cancer Biother. Radiopharm.* 2015, 30, 94–99. [CrossRef] [PubMed]
- 156. Bergmann, R.; Meckel, M.; Kubíček, V.; Pietzsch, J.; Steinbach, J.; Hermann, P.; Rösch, F. ¹⁷⁷Lu-Labelled Macrocyclic Bisphosphonates for Targeting Bone Metastasis in Cancer Treatment. *EJNMMI Res.* **2016**, *6*, 5. [CrossRef] [PubMed]
- 157. Fellner, M.; Baum, R.; Kubicek, V.; Hermann, P.; Roesch, F. ¹⁷⁷Lu-BPAMD—From Bone Imaging to Therapy with a Macrocycle-Bisphosphonate Ligand. *J. Nucl. Med.* **2010**, *51*, 1164.
- 158. Pfannkuchen, N.; Meckel, M.; Bergmann, R.; Bachmann, M.; Bal, C.; Sathekge, M.; Mohnike, W.; Baum, R.P.; Rösch, F. Novel Radiolabeled Bisphosphonates for PET Diagnosis and Endoradiotherapy of Bone Metastases. *Pharmaceuticals* 2017, 10, 45. [CrossRef] [PubMed]
- Yousefnia, H.; Zolghadri, S.; Sadeghi, H.R.; Naderi, M.; Jalilian, A.R.; Shanehsazzadeh, S. Preparation and Biological Assessment of 177Lu-BPAMD as a High Potential Agent for Bone Pain Palliation Therapy: Comparison with ¹⁷⁷Lu-EDTMP. *J. Radioanal. Nucl. Chem.* 2016, 307, 1243–1251. [CrossRef]
- 160. Meckel, M.; Kubíček, V.; Hermann, P.; Miederer, M.; Rösch, F. A DOTA Based Bisphosphonate with an Albumin Binding Moiety for Delayed Body Clearance for Bone Targeting. *Nucl. Med. Biol.* **2016**, *43*, 670–678. [CrossRef] [PubMed]
- 161. Velikyan, I. Prospective of ⁶⁸Ga-Radiopharmaceutical Development. Theranostics 2013, 4, 47–80. [CrossRef]
- 162. Wang, Y.; Wang, Q.; Chen, Z.; Yang, J.; Liu, H.; Peng, D.; Lei, L.; Liu, L.; Wang, L.; Xing, N.; et al. Preparation, Biological Characterization and Preliminary Human Imaging Studies of ⁶⁸Ga-DOTA-IBA. *Front. Oncol.* **2022**, *12*, 1027792. [CrossRef]
- 163. Yang, J.; Deng, J.; Fan, D.; Chen, G.; Lu, Z.; Liu, H.; Mok, G.S.P.; Chen, Y. Biodistribution and Internal Dosimetry of 68Ga-DOTA-IBA PET Imaging for Patients with Bone Metastases. *Clin. Nucl. Med.* **2023**, *48*, 847–852. [CrossRef]
- Wang, Q.; Yang, J.; Wang, Y.; Liu, H.; Feng, Y.; Qiu, L.; Chen, Y. Lutetium177-Labeled DOTA-Ibandronate: A Novel Radiopharmaceutical for Targeted Treatment of Bone Metastases. *Mol. Pharm.* 2023, 20, 1788–1795. [CrossRef]
- 165. Qiu, L.; Wang, Y.; Liu, H.; Wang, Q.; Chen, L.; Liu, L.; Wang, L.; Feng, Y.; Chen, Y. Safety and Efficacy of 68Ga- or ¹⁷⁷Lu-Labeled DOTA-IBA as a Novel Theranostic Radiopharmaceutical for Bone Metastases: A Phase 0/I Study. *Clin. Nucl. Med.* 2023, 48, 489. [CrossRef] [PubMed]
- 166. Li, H.; Xu, T.; Hua, Q.; Wang, L.; Chen, Y. 177Lu-DOTA-IBA Therapy in Prostate Cancer With Bone Metastases. *Clin. Nucl. Med.* 2023, 48, 740. [CrossRef] [PubMed]

- Xu, T.; Qu, G.; Liu, G.; Wang, L.; Chen, Y. A New Radiopharmaceutical ²²⁵Ac-DOTA-IBA in the Treatment of a Case of Bone Metastases. *Clin. Nucl. Med.* 2023, 48, 650. [CrossRef] [PubMed]
- 168. Meckel, M.; Bergmann, R.; Miederer, M.; Roesch, F. Bone Targeting Compounds for Radiotherapy and Imaging: *Me(III)-DOTA Conjugates of Bisphosphonic Acid, Pamidronic Acid and Zoledronic Acid. *EJNMMI Radiopharm. Chem.* 2017, 1, 14. [CrossRef] [PubMed]
- Ogawa, K.; Kawashima, H.; Shiba, K.; Washiyama, K.; Yoshimoto, M.; Kiyono, Y.; Ueda, M.; Mori, H.; Saji, H. Development of [(90)Y]DOTA-Conjugated Bisphosphonate for Treatment of Painful Bone Metastases. *Nucl. Med. Biol.* 2009, 36, 129–135. [CrossRef]
- 170. Ogawa, K.; Takai, K.; Kanbara, H.; Kiwada, T.; Kitamura, Y.; Shiba, K.; Odani, A. Preparation and Evaluation of a Radiogallium Complex-Conjugated Bisphosphonate as a Bone Scintigraphy Agent. *Nucl. Med. Biol.* **2011**, *38*, 631–636. [CrossRef] [PubMed]
- 171. Fakhari, A.; Jalilian, A.R.; Johari-Daha, F.; Shafiee-Ardestani, M.; Khalaj, A. Preparation and Biological Study of ⁶⁸Ga-DOTA-Alendronate. *Asia. Ocean J. Nucl. Med. Biol.* **2016**, *4*, 98–105. [CrossRef] [PubMed]
- 172. Ahrens, B.J.; Li, L.; Ciminera, A.K.; Chea, J.; Poku, E.; Bading, J.R.; Weist, M.R.; Miller, M.M.; Colcher, D.M.; Shively, J.E. Diagnostic PET Imaging of Mammary Microcalcifications Using ⁶⁴ Cu-DOTA-Alendronate in a Rat Model of Breast Cancer. *J. Nucl. Med.* 2017, 58, 1373–1379. [CrossRef]
- Ballinger, J.R. 68Ga-DOTA-Zoledronate. In PET Radiopharmaceuticals; Springer International Publishing: Cham, Switzerland, 2022; pp. 62–63. ISBN 978-3-031-10270-7.
- 174. Grun, A.; Kovacs, R.; Nagy, D.I.; Garadnay, S.; Greiner, I.; Keglevich, G. Efficient Synthesis of Benzidronate Applying of Phosphorus Trichloride and Phosphorous Acid. *Lett. Drug. Des. Discov.* **2015**, *12*, 78–84. [CrossRef]
- 175. Grus, T.; Lahnif, H.; Klasen, B.; Moon, E.-S.; Greifenstein, L.; Roesch, F. Squaric Acid-Based Radiopharmaceuticals for Tumor Imaging and Therapy. *Bioconjug. Chem.* 2021, *32*, 1223–1231. [CrossRef]
- 176. Greifenstein, L.; Engelbogen, N.; Máthé, D.; Grus, T.; Rösch, F.; Bergmann, R. Squaric Acid Bisphposphonates for Theranostics of Bone Metastasis—The Easy DOTA-Zoledronate. *Front. Nucl. Med.* **2022**, *2*, 870910. [CrossRef]
- 177. Khawar, A.; Eppard, E.; Roesch, F.; Ahmadzadehfar, H.; Kürpig, S.; Meisenheimer, M.; Gaertner, F.C.; Essler, M.; Bundschuh, R.A. Preliminary Results of Biodistribution and Dosimetric Analysis of [⁶⁸Ga]Ga-DOTAZOL: A New Zoledronate-Based Bisphosphonate for PET/CT Diagnosis of Bone Diseases. *Ann. Nucl. Med.* 2019, *33*, 404–413. [CrossRef]
- 178. Meisenheimer, M.; Kürpig, S.; Essler, M.; Eppard, E. DOTA-ZOL: A Promising Tool in Diagnosis and Palliative Therapy of Bone Metastasis-Challenges and Critical Points in Implementation into Clinical Routine. *Molecules* 2020, 25, 2988. [CrossRef] [PubMed]
- 179. Pillai, S.A.; Chobisa, D.; Urimi, D.; Ravindra, N. Pharmaceutical Glass Interactions: A Review of Possibilities. J. Pharm. Sci. 2016, 8, 103–111.
- Meckel, M.; Fellner, M.; Thieme, N.; Bergmann, R.; Kubicek, V.; Rösch, F. In Vivo Comparison of DOTA Based ⁶⁸Ga-Labelled Bisphosphonates for Bone Imaging in Non-Tumour Models. *Nucl. Med. Biol.* 2013, 40, 823–830. [CrossRef] [PubMed]
- 181. Eppard, E.; Meisenheimer, M.; Fuente, A.D.L.; Kurpig, S.; Essler, M.; Roesch, F. Radiolabelling of DOTAMZOL with ⁶⁸Ga and ⁴⁴Sc for Clinical Application. *EJEA* 2016, 47, OC34. [CrossRef]
- 182. Khawar, A.; Eppard, E.; Roesch, F.; Ahmadzadehfar, H.; Kürpig, S.; Meisenheimer, M.; Gaertner, F.C.; Essler, M.; Bundschuh, R.A. Biodistribution and Post-Therapy Dosimetric Analysis of [¹⁷⁷Lu]Lu-DOTAZOL in Patients with Osteoblastic Metastases: First Results. *EJNMMI Res.* 2019, 9, 102. [CrossRef]
- 183. Yadav, M.P.; Ballal, S.; Meckel, M.; Roesch, F.; Bal, C. [¹⁷⁷Lu]Lu-DOTA-ZOL Bone Pain Palliation in Patients with Skeletal Metastases from Various Cancers: Efficacy and Safety Results. *EJNMMI Res.* **2020**, *10*, 130. [CrossRef]
- 184. Kreppel, B.; Gaertner, F.C.; Ahmadzadehfar, H.; Khawar, A.; Roesch, F.; Kürpig, S.; Meisenheimer, M.; Essler, M.; Bundschuh, R.A. [177Lu]Lu-DOTA-Zoledronate Therapy—First Application in a Patient with Primary Osseous Metastatic Bronchial Carcinoma. *Nuklearmedizin* 2020, 59, 281–283. [CrossRef]
- 185. Clarke, E.T.; Martell, A.E. Stabilities of Trivalent Metal Ion Complexes of the Tetraacetate Derivatives of 12-, 13- and 14-Membered Tetraazamacrocycles. *Inorganica Chim. Acta.* **1991**, 190, 37–46. [CrossRef]
- 186. Kubíček, V.; Havlíčková, J.; Kotek, J.; Tircsó, G.; Hermann, P.; Tóth, É.; Lukeš, I. Gallium(III) Complexes of DOTA and DOTA–Monoamide: Kinetic and Thermodynamic Studies. *Inorg. Chem.* **2010**, *49*, 10960–10969. [CrossRef] [PubMed]
- 187. Eryilmaz, K.; Bakar, H.E.; Kilbas, B. Novel Developed HPLC Analyses of [⁶⁸Ga]Ga/[¹⁷⁷Lu]Lu-EDTMP and [⁶⁸Ga]Ga/[¹⁷⁷Lu]Lu-DOTA-Zoledronate. J. Label Comp. Radiopharm. 2022, 65, 178–186. [CrossRef] [PubMed]
- 188. Pfannkuchen, N.; Bausbacher, N.; Pektor, S.; Miederer, M.; Rosch, F. In Vivo Evaluation of [²²⁵Ac]Ac-DOTAZOL for α-Therapy of Bone Metastases. *Curr. Radiopharm.* 2018, 11, 223–230. [CrossRef]
- Park, E.A.; Graves, S.A.; Menda, Y. The Impact of Radiopharmaceutical Therapy on Renal Function. Semin. Nucl. Med. 2022, 52, 467–474. [CrossRef] [PubMed]
- 190. Zhang, J.; Singh, A.; Kulkarni, H.R.; Schuchardt, C.; Müller, D.; Wester, H.-J.; Maina, T.; Rösch, F.; van der Meulen, N.P.; Müller, C.; et al. From Bench to Bedside—The Bad Berka Experience With First-in-Human Studies. *Semin. Nucl. Med.* 2019, 49, 422–437. [CrossRef] [PubMed]
- 191. International Atomic Energy Agency. Pain Palliation of Bone Metastases: Production, Quality Control and Dosimetry of Radiopharmaceuticals, IAEA Radioisotopes and Radiopharmaceuticals Series No.9 [IAEA Preprint]. 2021. Available online: https://Preprint.Iaea.Org/Search.Aspx?orig_q=RN:54079457 (accessed on 3 December 2023).

- 192. Kabasakal, L.; AbuQbeitah, M.; Aygün, A.; Yeyin, N.; Ocak, M.; Demirci, E.; Toklu, T. Pre-Therapeutic Dosimetry of Normal Organs and Tissues of 177Lu-PSMA-617 Prostate-Specific Membrane Antigen (PSMA) Inhibitor in Patients with Castration-Resistant Prostate Cancer. *Eur. J. Nucl. Med. Mol. Imaging* **2015**, *42*, 1976–1983. [CrossRef]
- 193. Scarpa, L.; Buxbaum, S.; Kendler, D.; Fink, K.; Bektic, J.; Gruber, L.; Decristoforo, C.; Uprimny, C.; Lukas, P.; Horninger, W.; et al. The ⁶⁸Ga/¹⁷⁷Lu Theragnostic Concept in PSMA Targeting of Castration-Resistant Prostate Cancer: Correlation of SUVmax Values and Absorbed Dose Estimates. *Eur. J. Nucl. Med. Mol. Imaging* **2017**, *44*, 788–800. [CrossRef]
- 194. Fernández, R.; Eppard, E.; Lehnert, W.; Jiménez-Franco, L.D.; Soza-Ried, C.; Ceballos, M.; Ribbeck, J.; Kluge, A.; Rösch, F.; Meckel, M.; et al. Evaluation of Safety and Dosimetry of ¹⁷⁷Lu-DOTA-ZOL for Therapy of Bone Metastases. *J. Nucl. Med.* **2021**, *62*, 1126–1132. [CrossRef]
- 195. Rubira, L.; Deshayes, E.; Santoro, L.; Kotzki, P.O.; Fersing, C. 225Ac-Labeled Somatostatin Analogs in the Management of Neuroendocrine Tumors: From Radiochemistry to Clinic. *Pharmaceutics* **2023**, *15*, 1051. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.