



HAL
open science

The circularly permuted globin domain of Androglobin exhibits atypical heme stabilization and nitric oxide interaction

Brandon J Reeder, Giuseppe Deganutti, John Ukeri, Silvia Atanasio, Dimitri A Svistunenko, Christopher Ronchetti, Juan Carlos Mobarec, Elizabeth Welbourn, Jeffrey Asaju, Marten H Vos, et al.

► To cite this version:

Brandon J Reeder, Giuseppe Deganutti, John Ukeri, Silvia Atanasio, Dimitri A Svistunenko, et al.. The circularly permuted globin domain of Androglobin exhibits atypical heme stabilization and nitric oxide interaction. *Chemical Science*, 2024, 15 (18), pp.6738-6751. 10.1039/D4SC00953C . hal-04515181v1

HAL Id: hal-04515181

<https://hal.science/hal-04515181v1>

Submitted on 21 Mar 2024 (v1), last revised 21 May 2024 (v2)

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Chemical Science

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: B. Reeder, G. Deganutti, J. Ukeri, S. Atanasio, D. Svistunenکو, C. Ronchetti, J. C. Mobarec, E. M. Welbourn, M. Vos, J. O. Asaju, M. Wilson and C. A. Reynolds, *Chem. Sci.*, 2024, DOI: 10.1039/D4SC00953C.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.

ARTICLE

The circularly permuted globin domain of Androglobin exhibits atypical heme stabilization and nitric oxide interaction

Brandon J. Reeder^{*a}, Giuseppe Deganutti^{a,b}, John Ukeri^a, Silvia Atanasio^a, Dimitri A. Svistunenko^a, Christopher Ronchetti^a, Juan Carlos Mobarec^{a,b}, Elizabeth Welbourn^a, Jeffrey Asaju^a, Marten H. Vos^c, Michael T. Wilson^a, and Christopher A. Reynolds^{*b}

Received 00th January 20xx,
Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

In the decade since the discovery of androglobin, a multi-domain hemoglobin of metazoans associated with ciliogenesis and spermatogenesis, there has been little advance in the knowledge of the biochemical and structural properties of this unusual member of the hemoglobin superfamily. Using a method for aligning remote homologues, coupled with molecular modelling and molecular dynamics, we have identified a novel structural alignment to other hemoglobins. This has led to the first stable recombinant expression and characterization of the circularly permuted globin domain. Exceptional for eukaryotic globins is that a tyrosine takes the place of the highly conserved phenylalanine in the CD1 position, a critical point in stabilizing the heme. A disulfide bond, similar to that found in neuroglobin, forms a closed loop around the heme pocket, taking the place of androglobin's missing CD loop and further supporting the heme pocket structure. Highly unusual in the globin superfamily is that the heme iron binds nitric oxide as a five-coordinate complex similar to other heme proteins that have nitric oxide storage functions. With rapid autoxidation and high nitrite reductase activity, the globin appears to be more tailored toward nitric oxide homeostasis or buffering. The use of our multi-template profile alignment method to yield the first biochemical characterisation of the circularly permuted globin domain of androglobin expands our knowledge of the fundamental functioning of this elusive protein and provides a pathway to better define the link between the biochemical traits of androglobin with proposed physiological functions.

Introduction

The hemoglobin (Hb) of the erythrocyte is one of the most studied proteins in science. However, a decade after the discovery of androglobin (Adgb), a multi-domain Hb first identified in the testis of metazoans, there is still very little known about this novel member of the Hb superfamily. Adgb is unusual due to its long length of 1667 amino acids (human), with a centrally positioned globin domain.¹ This compares to the 140-190 amino acids of other human globins such as the erythrocyte Hb,² myoglobin (Mb), neuroglobin (Ngb)³ and cytoglobin (Cygb)⁴ and is significantly larger than other multi-domain globins such as flavohemoglobin.⁵ Initial sequence alignment analysis predicted the heme-binding globin domain of Adgb (Adgb-GD) to consist of the eight alpha-helical structure (termed A to H) with the 3-on-3 alpha-helical fold that encloses the heme moiety and is typical of most non-truncated globin architectures.¹ However, highly unusual in the globin family is that Adgb-GD is circularly permuted with a calmodulin binding domain situated between the H and A helices.¹ The N-terminal region of Adgb is reported to contain a

calpain-7-like region and the C-terminal region is reported to contain sequences for a coiled-coil region, a nuclear localization signal (NLS) and an ER membrane endoplasmic reticulum retention signal.¹ Calpain C2-like domains flanking the globin domain are predicted from calpain homologues studies.⁶

The potential interaction of Adgb with nitric oxide (NO) and calcium may underlie its relevance to spermatogenesis, ciliogenesis, as well as to potential diseases. A knockdown study in cells showed enhanced apoptosis and proliferation inhibition in glioma cell lines, relating to changes in the level of several proteins involved in cell proliferation, survival or apoptosis, including STAT3 cleaved caspase-3 and Bcl-2.⁷ Additionally, STXPB5 antisense proficient GR pancreatic cancer cell lines overexpress Adgb through ADGB promoter methylation, leading to drug resistance and inhibition of cell apoptosis. Recently, mRNA-Seq data from mammalian tissue has shown that Adgb is expressed in the female reproductive tract, lungs, and brain. In each case, Adgb is specifically associated with cell types forming motile cilia, with expression linked to transcription factors involved in ciliogenesis.⁸ An absence of Adgb leads to defective sperm head and flagellum formation during spermatogenesis.⁹ There remains a lack of data concerning the structure, properties or functions of this protein. This lack of knowledge on the structural and functional aspects of the protein is due to the difficulty in generating such a large full-length hemoglobin by recombinant techniques and by instabilities of the heme-binding, circularly-permuted globin domain.¹⁰

^a School of Life Sciences, University of Essex, Wivenhoe Park, Colchester, Essex, CO4 3SQ, UK.

^b Centre for Health and Life Sciences (CHLS), Alison Gingell Building, Coventry, CV1 5FB, UK.

^c LOB, CNRS, INSERM, École Polytechnique, Institut Polytechnique de Paris, 91128 Palaiseau, France

* reedb@essex.ac.uk (biochemical), and ad5291@coventry.ac.uk (modelling)
Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here]. See DOI: 10.1039/x0xx00000x



The alignment between Adgb and other proteins with regard to Adgb–GD was reassessed using a novel helix alignment method shown to work in the twilight zone of low sequence similarity.^{11–13} Our unique identification of the globin domain yielded a structure by homology modelling that was stable under initial 500 ns of molecular dynamics simulations, giving support for the potential expression of this protein despite initial negative expression results.¹⁰ Thus, based on our new alternative alignment, we have expressed the circularly-permuted globin domain as a stable recombinant protein. An intramolecular disulfide bond linking the N and C terminal sections of the heme binding globin domain appears to stabilize the ‘CD loop’ heme pocket region and likely the whole globular domain.

With a stable form of the protein expressed, we have characterized the protein using optical, electron paramagnetic resonance (EPR), stopped-flow and femtosecond laser flash photolysis spectroscopy to show that Adgb binds NO as a five-coordinate heme iron. This is unlike other globins, but similar to that observed with other hemoproteins such as guanylate cyclase, cytochrome c prime (cyt c') and dissimilatory nitrate respiration regulator.^{14–16} Furthermore, the protein exhibits high nitrite reductase activity, which is influenced by the redox state of the disulfide bond and exhibits a high autoxidation rate similar to Ngb. Based on our findings we propose that the globin domain may serve as an NO storage or NO synthesis protein under hypoxic conditions. With a binding domain for a calcium-dependent calmodulin and a calcium-dependent calpain-like sequence on the N-terminal domain, Adgb likely plays a role in both NO homeostasis and calcium signaling, with repercussions on spermatogenesis, sperm maturation and motile cilia function.^{17–20}

Results and discussion

The alignment of the androglobin helices to known templates

Fig. 1 (left) shows the scores for various alternative alignments of the six key helices of the Adgb-GD to their counterparts in alpha-Hb, beta-Hb, Cygb, Ngb, and Mb. Helix A (Adgb residues Val939–Glu951) is the first helix in alpha-Hb, beta-Hb, Cygb, Ngb and Mb, but in the naturally circular permuted Adgb it follows helix H (Adgb residues Phe866–Ser877) and the IQ domain (residues Val891–Thr933). The main peak lies at 0 for alpha-Hb, beta-Hb, Cygb, Ngb and Mb, which corresponds to the alignment in Fig. 2; a small alternative peak lies at +4 for each globin bar Mb, which corresponds to shifting the Adgb helix A four residues to the right. For the alpha-Hb–Adgb-GD alignment, there were 17155 pairwise alignments (because there were 235 alpha-Hb sequences and 73 Adgb-GD sequences). When the Adgb sequences were moved between -25 and +25, the 0 alignment obtained 10664 votes and the +4 alignment obtained 3664 votes; when the alpha-Hb sequences were moved between -25 and +25, the zero alignment obtained 10001 votes and the +4 alignment obtained 774 votes. In this case, the mean of 10332 for alignment 0 is considerably higher than the next nearest mean of 2219 (alignment +4). A similar predominant score was obtained for all HA alignments, and so when the scores for all five alignments were multiplied together, Fig. 1 (right), there is an overwhelming preference for alignment 0, despite the low percentage identity for HA of 15.3 %, 15.2 %, 16.6 %, 21.1 %, and 15.1 % for the Adgb-GD alignments to

alpha-Hb, beta-Hb, Cygb, Ngb, and Mb respectively. The results for the other five helices are similar, with percentage identities ranging between 7.8 % (HH of the Adgb-GD–Ngb alignment) and 26 % (HF of the Adgb–Mb alignment), Table S1. Based on these percentages, Adgb-GD is marginally more similar to Mb (mean helical percentage identity 17.8 %) and less similar to beta-Hb (mean helical percentage identity 14.0 %). The full alignment is given in Fig. 2 and the mean percentage identity for the pairwise alignments for each helix-helix profile alignment between androglobin and the other hemoglobins is given in Table S1.

The threefold Modeller structural alignment of both Adgb models (complex 1 and complex 2, see below) with the AlphaFold 2 model shows agreement in the sequence alignment except in two places, namely ⁸²⁸PVPFHDKEL⁸³⁴ (where helix F of the AlphaFold 2 model fills the space normally occupied by the heme) and the distal half of helix H, which terminates prematurely to accommodate the IQ domain (⁸⁷²DLWLLN⁸⁷⁷ is not helical).

The list of Adgb sequences, greatly expanded since the original sequence analysis in 2012, together with the helix alignment analysis above, permits a re-assessment of the overall sequence identification of the globin domain of Adgb. From the previous assignment of the globin domain structure,¹ helices C to H are the first to be expressed on the N terminal side (His761 onwards). This is followed by C terminal helices A and B from Asp935 onwards. These sections are interconnected by a 33-amino acid section incorporating an IQ calmodulin binding domain (Fig. S1A). This assignment took into account several key amino acids that are highly conserved throughout the hemoglobin superfamily. These comprised of: (i) The proximal His connecting the heme iron to the protein, situated on the F helix (F8); (ii) The E7 distal heme iron ligand, in Adgb this is a Gln instead of the more common His residue;¹ (iii) The CD1 Phe residue functions as anchorage and binding of the heme group within the heme pocket.^{21, 22} While our sequence alignment agrees with most of the original alignment, the identity of the C helix region and the CD1 residue differ. With the Phe770 amino acid assignment as the CD1 residue in human Adgb it becomes clear that in other species ~22% of such alignments places a Cys in the CD1 position (Table S2). Indeed, the previous alignment of Adgb globin domains shows three (9% of sequences) as Cys residues at the CD1 position,¹ but this was interpreted as mismatched alignments. The adjusted sequence alignment, shown in Fig. S1B and C, was used for the recombinant expression.

A Cys residue in the CD1 position is likely to have profound impact on the stabilization of heme binding as the substitution of the CD1 Phe by Cys removes an important contact with heme leaving a gap at the surface of the heme pocket which could result in instability. Mutations in human hemoglobin of CD1 Phe almost invariably result in instabilities in the protein, resulting in Heinz body formation, cyanosis and severe hemolytic anemia. A natural variation with a Cys residue in the CD1 position was found in the beta Hb chain of a Caucasian male infant (Hb Little Venice, β 42[CD1] Phe→Cys).²³ At 2 years of age the infant showed severe chronic hemolytic anaemia, positive Heinz body formation, haptoglobin depletion and required a monthly regular transfusion regime used more commonly for cases of severe forms of thalassemia.²³

We conducted a reassessment of the C to D helix sequence assignment, disregarding the prerequisite of a Phe as the CD1



component of the structure, based on the globin domain helix alignments (Fig. 1 and 2). Our assessment places a different sequence as the C to D helix section thus placing a Tyr residue (Tyr976) in the CD1 position of human Adgb globin domain (Fig. S2, Fig. S1B). Although essentially unique in eukaryotes, the presence of a Tyr in the CD1 position of prokaryotic Hbs is common and does not significantly affect heme binding, but can affect oxygen binding affinity.²⁴ Sequence alignment comparison shows that the frequency of Tyr residues is ~82% with the majority of the remaining sequences being Phe residues (Table S2). Therefore, we propose the assignment of CD1 to Tyr976 instead of Phe770, resulting in a circular permutation where helices D to H are expressed on the N-terminal side, followed by helices A to C following the IQ calmodulin binding domain sequence.

The previous alignment sequence did not allow expression of a stable form of the globin domain to be generated recombinantly,¹⁰ likely as a result of the expression of the incomplete globin domain. However, this new approach to sequence alignment resulted in expression of a stable globin domain (*vide infra*).

The androglobin model structures

Adgb-GD complexes 1 and 2, shown in Fig. 3, align well with the five Hb structural templates. The root mean square deviations (RMSDs) over the common globin domain to Mb are 1.3 Å for complex 1, 1.1 Å for complex 2 and 2.3 Å for the AlphaFold 2 model, over the alpha helices, as determined using SSM.²⁵ The three models differ in the orientation of the IQ (calmodulin-binding) domain, which is not included in the RMSD calculations because it is absent in Mb and the other traditional Hbs. The orientation of the IQ domain differs in all three structures and is connected by a very flexible loop, and this flexibility is probably important for binding calmodulin. When superposed onto the full Adgb AlphaFold structure, all three structures present the IQ domain in an accessible orientation. The AlphaFold 2 and Modeller structures differ in two other aspects. Firstly, helix H is truncated in the AlphaFold 2 model. Secondly, helix E moves slightly into the space occupied by the heme; this structural topology may bear some resemblance to purified Adgb before the heme group is added back (see Methods). The globin domain contains four cysteine residues, none of which form a disulfide bond in the AlphaFold 2 model.

Adgb-GD complexes 1 and 2 were interrogated through molecular (MD) dynamics simulations to assess stability over the time course. Three 500 ns MD replicas were produced for each model in complex with the heme (Fig. 4). The emerging scenario suggests the overall higher stability of complex 1 as indicated by the root mean square fluctuation (RMSF) analysis (Fig. 4A,D) and the RMSD, of the heme (Fig. 4B, D). In both structures, the IQ domain and the loop connecting it to the H helix were the most dynamic parts (Supplementary videos 1 and 2), however, in complex 1 the whole structure underwent high thermal fluctuations. This affected the stability of the heme: it was less steady in complex 1 (Fig. 4B). In both systems, the distal Gln792 remained in the proximity of the heme, while the CD Tyr976 was more flexible during the simulations, especially in complex 2. Models are accessible via <https://zenodo.org/records/10509366>.

Optical properties, heme iron spin state and the intramolecular disulfide bond in the androglobin globin domain

SDS PAGE and western blot analysis show expression and purification of a soluble Adgb-GD protein at ~26 kDa (Fig. S3), close to the predicted 27 kDa weight based on sequence as shown in Fig. S1C. This includes the Histag and linker sequences and the alternative C-terminal C helix identified (*vide supra*). Reverse phase HPLC analysis of Adgb-GD shows the heme content used for extinction coefficient calculations as directly compared to a known concentration of Mb (Fig S4). Size exclusion analysis shows that in a phosphate buffer the protein is primarily monomeric with some dimer (Fig. S5). In 150 mM NaCl buffer, this changed to mostly tetramer, likely as a result of the decrease of charge-repulsion on the protein surface. All subsequent data was collected in the absence of salt unless otherwise stated. The predicted model of this domain (see supporting information videos) results in a large positive surface electrostatic potential for much of the protein. This may explain why SDS facilitates purification, i.e. prevents aggregation with endogenous *E.coli* proteins through blockage of charged sites.

As expressed, the optical properties of the Adgb-GD are shown in Fig. 5 and the calculated extinction coefficient and peak wavelengths are in Table S3. The ferric protein has bands in the visible region at 534 and 566 nm, suggesting a hexacoordinate state of the heme iron like that observed for Cygb and Ngb.^{3, 4} However, a small peak at ~630 nm suggests that the protein also has some pentacoordinate-like properties. The coordination is pH dependent as shown in Fig S6. There is a single transition with pK_a 8.6 that, from the optical spectra, is typical of a transition from penta- to hexa-coordination, with the loss of a 630 nm peak (typical of the water-bound pentacoordinate state) and the evolution of peaks at 566/534 nm.²⁶ Mb and Hb have similar pK_a values (8.93 and 8.0 for Mb and Hb respectively).²⁶ However, the change in coordination is opposite to that of Adgb, with Adgb showing hexa-coordination at more acidic pH values and penta-coordination at alkaline pH. Thus the change in coordination cannot be due to the H₂O/OH⁻ ligands observed in Mb and Hb and is more indicative of the coordination change observed with Cygb (pK 8.2) with His-Fe(III)-X (X=His for Cygb and Gln for Adgb) at acidic pH and His-Fe(III)-H₂O at alkaline pH.²⁷ Reduction to deoxy ferrous iron shows two prominent peaks at 531 and 560 nm indicative of hexacoordinate heme iron configuration. The CO-bound spectrum is typical for most globins, but the NO-bound spectrum exhibits a Soret peak at an unexpectedly hypsochromic (blue) shifted peak at 395 nm (Fig. 5). Typical wavelength maxima for hexacoordinate ferrous-NO bound protein are observed ~420-430 nm in other NO-bound globins such as Mb and Cygb.^{28, 29} This suggests that the NO is bound in an unusual form in Adgb-GD. Optical changes were not observed with addition of NO (20 mM) to ferric Adgb-GD, indicating that NO affinity for the ferric heme iron is low.

The EPR spectra of ferric heme Adgb-GD at a slightly acidic pH (Fig. 6A and 6B, pH 6) measured at 10 K shows a mixture of the high spin (S=5/2, HS) and low spin (S=1/2, LS) signals corresponding to penta- and hexa-coordinated heme iron, respectively. The HS signal with the perpendicular $g_x=g_y=5.95$ and the parallel $g_z=2.00$ components is typical of other globins in a pentacoordinate conformation. The $g=2.95$ and $g=2.26$ EPR signals are the g_x and g_y components of a LS signal, with the third g_z component was not observed as it was too



broad, which is typical for many globins, particularly for the signal-to-noise level. The LS signal with these g -values has been classified as a low spin form in ferric Hb, with one of the axial ligands likely to be a histidine's nitrogen and the other not identified.³⁰ At pH 7, the HS signal at $g=5.95$ is smaller than at pH 6, which is concurrent with a noticeable increase of the LS form (as per both $g=2.95$ and $g=2.26$ signals). This is in agreement with the total ferric heme concentration being conserved in the pH 6 – pH 7 range. The intensity of the HS peak partially returns at pH 8, with a small decrease of the LS form as compared to pH 7. However, this was likely not due to an acid-alkaline transition between the HS and LS ferric heme forms as the pK_a for such a transition is clearly reported in the liquid phase at 8.4 (Fig. S6). The intensity changes are likely an artifact of freezing, similar to that observed with the EPR spectra of Cygb.²⁷ This effect appears largely negated with the disulfide reduced protein where there was no effect of pH on both HS and LS signals (Fig. S7). The reduced cysteine protein shows a change in line shape of the perpendicular components area, at $g\sim 6$. Not only does the $g=6$ signal becomes much wider, the new effective g -values become apparent at $g\sim 6.5$ (or maybe lower/higher). A small amount of 'rhombohedral iron' can be observed as a $g=4.3$ signal in both oxidized and reduced proteins as a typical small component in native and recombinant proteins.³¹

In Ngb, the CD loop contains a disulfide bond between Cys46 (helix C7) and Cys55 (helix D5), known to stabilize the distal histidine ligation and the redox thermodynamics of ferric Ngb.^{34–36} With our reassignment of the C helix sequence, the CD section of Adgb-GD possesses a predicted disulfide bond from residues Cys787 and Cys978 (Fig. S1). Absent in the original sequence alignment, this disulfide bond could stabilize this crucial juncture of the heme pocket, now predicted as the N and C terminal sections of the domain. With four cysteine residues in the globin domain sequence, we determined how many of those are surface exposed and free to bind dithiodipyridine. As shown in Fig. 6C, there were two (1.9 ± 0.3) free sulfhydryl's per heme detected with tris(2-carboxyethyl)phosphine (TCEP) reduced Adgb-GD, meaning that two of the four cysteines are surface exposed. In the protein as expressed in *E.coli* (without reduction by TCEP), no free cysteines were observed. Thus, as expressed, the globin domain possesses a single disulfide bond. From the predicted positions of the cysteines in complex 1 and 2, only two are both surface exposed and close enough to form an intramolecular disulfide, that of Cys787 and 978 in the "CD loop" region of the protein; however, both these two (reduced) cysteine residues and Cys894, Cys970 moved sufficiently close to form a disulfide bond in the MD simulation of the AlphaFold2 structure after 1.4 μ s (Fig. 4C), even though the sulfur atoms were originally 11.6 Å and 17.4 Å apart respectively (Fig. 4D). Further studies will be required to confirm the position and the micro-environmental conditions favoring the formation of the disulfide bond within Adgb-GD.

Ferrous androglobin globin domain binds nitric oxide in a primarily pentacoordinate form

In order to further establish that the heme group is fully incorporated into a heme pocket, as in other heme proteins and not merely adventitiously bound to the surface, we have studied the ligand binding properties of Adgb-GD. By using stopped-flow and femto-second laser spectroscopy, we have shown that the heme behaves as expected for a fully incorporated heme.

Adgb-GD binds NO in the ferrous form to generate an optical spectrum as observed in Fig. 5, suggesting a five-coordinate NO heme iron. This is supported by EPR at 10 K (Fig. 6D). This spectrum showed a characteristic three-line hyperfine signal, essentially identical to that reported for cyt *c'* from *Shewanella frigidimarina* and *Alcaligenes xylosoxidans*.¹⁶ This is again consistent with five-coordinate NO binding to the ferrous heme.^{16, 37}

The optical changes and kinetics of NO binding to deoxyferrous Adgb-GD, followed using stopped-flow spectroscopy are shown in Fig. S8. Increases at 390 nm are concurrent with decreases at 426 nm with a number of isosbestic points (e.g. 407 nm) consistent with a simple $Fe^{2+} + NO \rightarrow Fe^{2+}\text{-NO}$ binding mechanism. However, the time course follows a double exponential function (Fig. S8 C, E and F), suggesting either a heterogeneous population or an intermediate that is distinct from the NO-bound or deoxyferrous species. The former possibility is supported by a global fit to a 3-component parallel or serial mechanism showing essentially identical calculated component spectra; deoxyferrous, NO-bound and a spectrum that is intermediate between the deoxyferrous species and ferrous-NO species with clear isosbestic points (Fig. S8D). This eliminates the possibility that the slow phase results from a displacement of a bound ligand by NO. This suggests a conformational change from a closed to an open form of the protein may be required for NO binding.

The rate constant for NO binding is NO concentration independent for both phases (Fig. S8 E and F), suggesting that there two populations of the heme pocket, both having an occluded (closed) heme where a non-binding ligand sterically hindered the approach of NO. The rate limits observed are thus the rate constants of the conformational change in each population for the closed to open transition. The oxidation state of the cysteines makes no difference to the rate of NO binding, either fast or slow phase. Therefore, any effect of cysteine oxidation state has on heme pocket dynamics does not hinder the entrance, binding and dissociation of NO to the heme iron. An estimate of the maximum rate of binding may be made from Fig S8 E and F, at concentrations of NO less than 12 μ M. The affinity of NO binding is measured to be $\sim 1 \times 10^{-10}$ to 1×10^{-11} M (Table S4) with k_{on} values $\geq 10^7$ and 10^6 $M^{-1}s^{-1}$ based on the lowest concentration of NO used and k_{off} calculated from ligand exchange with oxygen ($1.6 \times 10^{-4} s^{-1}$, Fig. S9). This affinity is consistent with other globins such as Mb, Hb and Ngb where k_{on} is $\sim 10^7$ – 10^8 $M^{-1}s^{-1}$ and k_{off} 10^{-3} to $10^{-5} s^{-1}$.^{38, 39}

CO binding exhibits a similar kinetic pattern to NO binding, albeit at slower rates (Fig. S10). At the concentrations used the CO binding was CO concentration independent, like that of NO. However, unlike NO binding the reduction of the cysteines did show an effect on CO binding, with a decrease in the kinetics of CO binding of ~ 3.5 fold (Fig. 10 E and F). The affinity of the CO binding is $\sim 10^{-7}$ to 10^{-8} M (Table S4) with fast and slow k_{on} values $\geq 3 \times 10^6$ and 3×10^5 $M^{-1}s^{-1}$ respectively based on the lowest concentration of CO used and k_{off} on ligand exchange with NO ($0.31 s^{-1}$, Fig. S11). This affinity is ~ 6000 fold lower



than NO binding, consistent with the 'sliding scale rule' developed by Olson and coworkers⁴⁰ and is consistent with other globins such as Mb and Ngb where k_{on} is $\sim 10^7$ – 10^8 $\text{M}^{-1}\text{s}^{-1}$ and k_{off} 10^{-3} to 10^{-5} s^{-1} .^{40, 41} The five-coordinate NO-bound species found in this work is similar to those reported for cyt *c'*, with the NO on the proximal side of the heme. The kinetics of NO binding to deoxyferrous Adgb-GD (Fig. S8) is consistent with the NO binding mechanism proposed for cyt *c'*, that is a single optical transitions comprising two kinetic phases.¹⁶ NO can only bind to the distal iron location of cyt *c'* following a conformational change involving an occlusion in the heme pocket (by a Phe residue in the case of cyt *c'*). These two conformations of cyt *c'* are optically indistinguishable. Subsequent to proximal histidine displacement, NO binding to the proximal side of the heme and NO dissociation from the distal side are rapid and not observed as separate events, but only as a single observed spectrokinetic event. This model may also be applied to the observations of NO binding to Adgb-GD. However, there is insufficient evidence to assign whether NO binds to the proximal or distal side, although the simple hypothesis is that it binds to the distal side. Furthermore, cyt *c'* shows a mixture of penta- and hexa-coordinate NO bound species as observed by a split Soret peak 395/415 nm.⁴² As a weak band is consistently present in NO binding and NiR experiments at ~ 420 – 430 nm, it is likely that a small fraction of NO-bound Adgb-GD remains a hexa-coordinate species.

In addition to cyt *c'*, Hb has also been reported to bind NO in a pentacoordinate form, but only in a T state at low NO to Hb ratios and only with the alpha chain.^{35, 43, 44}

The transient absorption spectra observed following dissociation of the Adgb-GD:NO complex resulting from a short (femtosecond) light pulse is shown in Fig. 7A. The spectra are characterized by a broad bleaching around 390 nm due to the disappearance of the 5-coordinate NO-bound state and a relatively strong induced absorption centered at 427 nm assigned to the 4-coordinate NO-dissociated state.^{45, 46} This supports the EPR data (Fig. 6D) showing that the NO bound state of the ferrous Adgb is pentacoordinate. Apart from small relaxation signals with a time constant of ~ 1.5 ps, corresponding to a blue shift of the induced absorption band (Fig. 7A inset) and assigned to vibrational cooling, the spectral evolution is characterized by a decay (associated spectra in the inset of Fig. 7A) dominated by a 5.3 ps phase ($1.9 \times 10^{11} \text{ s}^{-1}$) and a minor ($\sim 14\%$) 20 ps phase ($5.0 \times 10^{10} \text{ s}^{-1}$) (Fig. 7B). The remaining spectrum after these phases corresponds to only $\sim 1\%$ of the photo-dissociated NO, meaning that NO rebinding is almost completely geminate. This implies that dissociated NO stays within the confines of the heme pocket and only minor quantities of NO escape the heme pocket. Rebinding of NO to the heme iron from bulk solution outside the heme pocket would be expected to occur on the μs to ms timescale, as observed with Mb and Cygb for NO or other gases such as CO.^{29, 47, 48} High-yield rebinding of NO to heme in a single 5–8 ps phase has been observed upon dissociation in all studied 5-coordinate heme-NO complexes in proteins thus far (Table S5).^{49–51} In Adgb, however a slower, 20 ps phase of NO binding is also present. This finding suggests a relaxation process competing with initial NO rebinding, allowing NO to explore a larger conformational space (rototranslational freedom), and indicating a less constrained heme pocket. Such multiphasic NO rebinding dynamics reflecting a more

accessible heme pocket is also observed in other globins, which all form 6-coordinate NO complexes (Table S5).^{DOI: 10.1039/D4SC00953C}

Androglobin nitrite reductase activity and effect of cysteine redox state on behaviour

Many heme proteins, including Hbs, are noted for their nitrite reductase (NiR) activity, giving potential functions in NO homeostasis depending on the oxygen concentration of their microenvironment.^{29, 52–54} This reaction was assessed for Adgb-GD under anaerobic conditions (Fig. 8). The reaction follows a mechanism in which NO_2^- slowly reacts with ferrous heme to generate NO and ferric heme^{55, 56}. The ferric heme is rapidly reduced by excess dithionite in solution resulting in the formation of the ferrous-NO complex. The optical changes following this reaction are essentially identical to those of NO binding (Fig. 8 A and B; compare with Fig. S8A and B) with a two-phase kinetics (Fig. 8C). The concentration dependence of the fast kinetics of the nitrite reaction (Fig. 8E) exhibits a high error due to the small amplitude of the optical changes observed in the global fit (Fig. 8D) but appears to follow a hyperbolic concentration dependence with an apparent K_D of 2.91 ± 0.57 mM NO_2^- and a maximum rate of $1.54 \pm 0.01 \text{ s}^{-1}$ (Fig. 8E, ●). The slower rate representing the formation of the deoxyferrous-NO bound species also follows a hyperbolic curve as a function of nitrite concentration with an apparent K_D of 8.42 ± 0.46 mM NO_2^- and a maximum rate of $2.54 \times 10^{-1} \text{ s}^{-1} \pm 6.1 \times 10^{-4} \text{ s}^{-1}$ (Fig. 8F, ●). Unlike NO binding, the effect of reduction of the disulfide bond has a significant effect on the rate of NiR activity (Fig. 8E and 8F) with TCEP reduced free sulfhydryl (○) exhibiting significantly decreased kinetics, both for the fast kinetics with an apparent K_D of 9.13 ± 1.38 mM NO_2^- and a maximum rate of $3.23 \times 10^{-1} \text{ s}^{-1} \pm 0.02 \text{ s}^{-1}$ and slower NO binding kinetics of with an apparent K_D of 3.26 ± 1.12 mM NO_2^- and a maximum rate of $3.67 \pm 0.40 \times 10^{-2} \text{ s}^{-1}$. This effect of sulfhydryl reduction on the NiR activity may arise from structural changes in the heme pocket, affecting exogenous ligand affinity, as observed for Ngb,³⁶ or affecting the endogenous distal ligand off-rate, as observed for Ngb⁵⁶ and Cygb.⁴⁷

The oxyform which can only be observed transiently by stopped-flow spectroscopy binds with an association rate constant of $\sim 0.8 \times 10^6 \text{ M}^{-1}\text{s}^{-1}$. This compares to 14, 50, 60 and $250 \times 10^6 \text{ M}^{-1}\text{s}^{-1}$ for Mb, Hb α , Hb β and Ngb respectively.^{57, 58} Autoxidation rates for globins vary greatly, with Mb and Ngb reported at 0.055 and 5.4 h^{-1} .⁵⁷ The measured autoxidation rate for Adgb-GD is $\geq 2.6 \text{ s}^{-1}$ (9360 h^{-1}) being limited by the oxidation of dithionite. This value is three orders of magnitude higher than the corresponding value for Ngb (Fig S12). This remarkable autoxidation rate strongly suggests that O_2 binding is not a physiological function of the protein except for the possibility of oxygen sensing. Similarly, NO dioxygenase activity is unlikely, as this requires a semi-stable ferrous- O_2 complex for the NO to react with. The oxidation state of the iron *in vivo* remains unknown, like that of Ngb and Cygb. However, the oxygen tension in the testes is typically low (~ 10 – 15 mmHg), lower even than venal O_2 levels.^{59–61} Consequently the existence of the ferrous form of the heme iron *in vivo*, at least in the testes, cannot be completely discounted.

Recent studies have shown many globins appear to have properties relating to NO homeostasis, although the exact nature of this biochemistry *in vivo* is still under debate. Our results in Fig. 8 confirm that Adgb-GD reacts with nitrite to generate NO under



hypoxic/anoxic conditions and subsequently binds NO as a pentacoordinate species. The NiR rate constants of other globins are typically linear as a function of nitrite concentration.²⁹ Assuming that the rate of NO formation relates to the rate of ferrous-NO generation (Fig. 8F), the initial slope of $\sim 25 \text{ M}^{-1} \text{ s}^{-1}$ at low nitrite concentrations is higher than that for Ngb, Mb or Hb, and similar to that reported for globin X⁵² and Cygb (Table S6).^{29, 62}

NO has important roles in the testes and hence NO homeostasis is also important. Four Nitric Oxide Synthases (NOS) are present in the testes, endothelial NOS (eNOS), inducible NOS (iNOS), and neuronal NOS (nNOS) and a testis-specific NOS (TnNOS).¹⁷ NO has been proposed to play a unique role in modulating germ cell viability and development,²⁰ with high NO concentrations exhibiting a deleterious role in the mobility of spermatozoa⁶³ and thus acting on some aspects of male infertility.¹⁷ NO also has vital roles in steroidogenesis, gametogenesis and the regulation of germ-cell apoptosis with excess NO linked to induction of germ cell apoptosis and oxidative damage.⁶⁴ Many globins have proven or proposed functions in NO regulation, either in primary or secondary roles. Thus, there is a distinct possibility that Adgb could also play a role in NO homeostasis and hence germ cell viability. Our data showing an unusual NO binding and high NiR activity provides proof-of-principle that Adgb could be involved in this process, but requires further studies to verify.

The presence of a calpain-like motif on the N terminal side of the globin domain and a calmodulin binding domain directly dissecting the globin domain strongly suggest a role for calcium in the functional mechanism of this protein. The links between NO and calcium are well established.⁶⁵ Calcium channel blockers decrease intracellular calcium levels and increase the vasodilator efficacy of NO in smooth muscle.⁶⁶ Conversely, an increase in NO by vascular endothelial cells in the liver enhances calcium signaling in surrounding hepatocytes.⁶⁷ In the testes, calcium channel blockers to relieve hypertension causes reversible male infertility in mice.⁶⁸ Hence it is not implausible that calcium and NO binding are linked in Adgb function.

Recently, the establishment of a role for Adgb in ciliogenesis has heightened the importance of Adgb. Overexpression studies show an Adgb-dependent increase in ciliated cells.⁸ Expression of Adgb is in turn linked to Forkhead Box J1 (FOXJ1), a transcription factor involved in ciliogenesis, as overexpression of FOXJ1 directly led to increased Adgb mRNA levels through binding to the ADGB promotor.⁸ Furthermore, Adgb has been shown to directly interact with the cytoskeletal protein Sept10.⁹ Adgb knockout in mice leads to mislocalisation of Sept10 in sperm, resulting in malformations of the flagella and head. The mechanism of Adgb action appears to be via proteolytic cleavage of Sept10, with the proteolytic activity controlled via calmodulin-dependent binding to the IQ motif of the circularly permuted globin domain of Adgb.⁹

Conclusions

In summary, the helical alignment for Adgb-GD from our method, designed to work in the twilight zone, yielded an alternate helix alignment around the C and D helical region. This alignment is consistent with that obtained from the AlphaFold-2 model. Identification of a Tyr in the CD1 position (Tyr977 in

the human Adgb) is to our knowledge unique for eukaryotic globins, but is common in prokaryotes. DOI: 10.1039/D4SC00953C

Validation of our proposed helix alignment lies, at least in part, in the agreement with the AlphaFold-2 model and the generation of a stable recombinant form of the protein using the alternative globin domain sequence. This is supported by heme insertion and tight heme binding to generate optical and EPR spectra, typical of pure authentic heme proteins. The femtosecond laser flash data indicates a true heme pocket from which ligand (NO) cannot escape and kinetics measured by stopped-flow that conform in general to known heme proteins. The presence of an intramolecular disulfide bond goes some way to explain the stability of the recombinant protein, given that the N and C terminal regions of this circularly permuted globin domain are in the critical area of the heme-binding pocket.

The biochemical evaluation presented here illustrates the characteristics of the globin domain of Adgb, showing that it can potentially participate in NO sensing or regulation. Our results, together with the known calcium-linked structural aspects of calpain and calmodulin binding properties of Adgb, lay the foundation for further investigation into the functional role of Adgb, given its potential medical significance.

Author Contributions

BJR and CAR conceived the study, BJR, JU and MTW performed the biochemical analysis of the protein and wrote the paper. CAR, CR, JCM, SA, and GD conducted the sequence, helix alignments and molecular dynamics studies, analysed the data and wrote the paper. CAR and CR conducted the helix alignments. EW and JA performed supplementary biochemical data collection and analysis for size exclusion (EW) and SDS PAGE / western blot (JA). DAS, BJR and JA performed and analysed the EPR study. MHV performed and analysed the ultra-fast laser flash study.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

BJR, DAS, MTW and CAR thank the Biotechnology and Biological Sciences Research Council (UK) (BB/T01802X/1) for financial support.

Notes and references

1. D. Hoogewijs, B. Ebner, F. Germani, F. G. Hoffmann, A. Fabrizio, L. Moens, T. Burmester, S. Dewilde, J. F. Storz, S. N. Vinogradov and T. Hankeln, Androglobin: a chimeric globin in metazoans that is preferentially expressed in Mammalian testes, *Mol Biol Evol*, 2012, **29**, 1105-1114.
2. M. F. Perutz, Structure of hemoglobin, *Brookhaven Symp Biol*, 1960, **13**, 165-183.



3. J. T. Trent, 3rd, R. A. Watts and M. S. Hargrove, Human neuroglobin, a hexacoordinate hemoglobin that reversibly binds oxygen, *J Biol Chem*, 2001, **276**, 30106-30110.
4. T. Burmester, B. Ebner, B. Weich and T. Hankeln, Cytoglobin: a novel globin type ubiquitously expressed in vertebrate tissues, *Mol Biol Evol*, 2002, **19**, 416-421.
5. H. Zhu and A. F. Riggs, Yeast flavohemoglobin is an ancient protein related to globins and a reductase family, *Proc Natl Acad Sci U S A*, 1992, **89**, 5015-5019.
6. N. D. Rawlings, Bacterial calpains and the evolution of the calpain (C2) family of peptidases, *Biol Direct*, 2015, **10**, 66.
7. B. Huang, Y. S. Lu, X. Li, Z. C. Zhu, K. Li, J. W. Liu, J. Zheng and Z. L. Hu, Androglobin knockdown inhibits growth of glioma cell lines, *Int J Clin Exp Pathol*, 2014, **7**, 2179-2184.
8. T. W. Koay, C. Osterhof, I. M. C. Orlando, A. Keppner, D. Andre, S. Yousefian, M. Suarez Alonso, M. Correia, R. Markworth, J. Schodel, T. Hankeln and D. Hoogewijs, Androglobin gene expression patterns and FOXJ1-dependent regulation indicate its functional association with ciliogenesis, *J Biol Chem*, 2021, **296**, 100291.
9. A. Keppner, M. Correia, S. Santambrogio, T. W. Koay, D. Maric, C. Osterhof, D. V. Winter, A. Clerc, M. Stumpe, F. Chalmel, S. Dewilde, A. Odermatt, D. Kressler, T. Hankeln, R. H. Wenger and D. Hoogewijs, Androglobin, a chimeric mammalian globin, is required for male fertility, *Elife*, 2022, **11**.
10. A. Bracke, D. Hoogewijs and S. Dewilde, Exploring three different expression systems for recombinant expression of globins: *Escherichia coli*, *Pichia pastoris* and *Spodoptera frugiperda*, *Anal Biochem*, 2018, **543**, 62-70.
11. A. Lock, R. Forfar, C. Weston, L. Bowsher, G. J. Upton, C. A. Reynolds, G. Ladds and A. M. Dixon, One motif to bind them: A small-XXX-small motif affects transmembrane domain 1 oligomerization, function, localization, and cross-talk between two yeast GPCRs, *Biochim Biophys Acta*, 2014, **1838**, 3036-3051.
12. B. Taddese, G. J. Upton, G. R. Bailey, S. R. Jordan, N. Y. Abdulla, P. J. Reeves and C. A. Reynolds, Do plants contain g protein-coupled receptors?, *Plant Physiol*, 2014, **164**, 287-307.
13. H. A. Watkins, M. Chakravarthy, R. S. Abhayawardana, J. J. Gingell, M. Garelja, M. Pardamwar, J. M. McElhinney, A. Lathbridge, A. Constantine, P. W. Harris, T. Y. Yuen, M. A. Brimble, J. Barwell, D. R. Poyner, M. J. Woolley, A. C. Conner, A. A. Pioszak, C. A. Reynolds and D. L. Hay, Receptor Activity-modifying Proteins 2 and 3 Generate Adrenomedullin Receptor Subtypes with Distinct Molecular Properties, *J Biol Chem*, 2016, **291**, 11657-11675.
14. F. Cutruzzola, A. Arcovito, G. Giardina, S. della Longa, P. D'Angelo and S. Rinaldo, Distal-proximal crosstalk in the heme binding pocket of the NO sensor DNR, *Biometals*, 2014, **27**, 763-773.
15. D. M. Lawson, C. E. Stevenson, C. R. Andrew and R. R. Eady, Unprecedented proximal binding of nitric oxide to heme: implications for guanylate cyclase, *EMBO J*, 2000, **19**, 5661-5671.
16. A. Manole, D. Kekilli, D. A. Svistunenko, M. T. Wilson, P. S. Dobbin and M. A. Hough, Conformational control of the binding of diatomic gases to cytochrome c', *J Biol Inorg Chem*, 2015, **20**, 675-686.
17. N. P. Lee and C. Y. Cheng, Nitric oxide and cyclic nucleotides: their roles in junction dynamics and spermatogenesis, *Adv Exp Med Biol*, 2008, **636**, 172-185.
18. M. V. Sataric, S. Zdravkovic, T. Nemes and B. M. Sataric, Calcium signaling modulates the dynamics of cilia and flagella, *Eur Biophys J*, 2020, **49**, 619-631.
19. H. C. Saternos and W. A. AbouAlaiwi, Signaling interplay between primary cilia and nitric oxide: A mini review, *Nitric Oxide*, 2018, **80**, 108-112.
20. A. Zini, M. K. O'Bryan, M. S. Magid and P. N. Schlegel, Immunohistochemical localization of endothelial nitric oxide synthase in human testis, epididymis, and vas deferens suggests a possible role for nitric oxide in spermatogenesis, sperm maturation, and programmed cell death, *Biol Reprod*, 1996, **55**, 935-941.
21. M. S. Hargrove, S. Krzywdka, A. J. Wilkinson, Y. Dou, M. Ikeda-Saito and J. S. Olson, Stability of myoglobin: a model for the folding of heme proteins, *Biochemistry*, 1994, **33**, 11767-11775.
22. A. Roesner, C. Fuchs, T. Hankeln and T. Burmester, A globin gene of ancient evolutionary origin in lower vertebrates: evidence for two distinct globin families in animals, *Mol Biol Evol*, 2005, **22**, 12-20.
23. S. J. Henderson, A. T. Timbs, J. McCarthy, A. E. Gallienne, M. Proven, M. J. Rugless, H. Lopez, J. Eglinton, D. Dziedzic, M. Beardsall, M. S. Khalil and J. M. Old, Ten Years of Routine alpha- and beta-Globin Gene Sequencing in UK Hemoglobinopathy Referrals Reveals 60 Novel Mutations, *Hemoglobin*, 2016, **40**, 75-84.
24. H. Ouellet, M. Milani, M. LaBarre, M. Bolognesi, M. Couture and M. Guertin, The roles of Tyr(CD1) and Trp(G8) in *Mycobacterium tuberculosis* truncated hemoglobin O in ligand binding and on the heme distal site architecture, *Biochemistry*, 2007, **46**, 11440-11450.
25. E. M. Mitchell, P. J. Artymiuk, D. W. Rice and P. Willett, Use of Techniques Derived from Graph-Theory to Compare Secondary Structure Motifs in Proteins, *Journal of Molecular Biology*, 1990, **212**, 151-166.
26. E. Antonini and M. Brunori, in *Frontiers in Biology*, eds. A. Neuberger and E. L. Tatum, North-Holland publishing company, Amsterdam-London, 1971, vol. 21, pp. 13-52.
27. B. J. Reeder, D. A. Svistunenko and M. T. Wilson, Lipid binding to cytoglobin leads to a change in haem coordination: a role for cytoglobin in lipid signalling of oxidative stress, *Biochem J*, 2011, **434**, 483-492.
28. H. Li, C. Hemann, T. M. Abdelghany, M. A. El-Mahdy and J. L. Zweier, Characterization of the mechanism and magnitude of cytoglobin-mediated nitrite reduction and nitric oxide generation under anaerobic conditions, *J Biol Chem*, 2012, **287**, 36623-36633.
29. B. J. Reeder and J. Ukeri, Strong modulation of nitrite reductase activity of cytoglobin by disulfide bond oxidation: Implications for nitric oxide homeostasis, *Nitric Oxide*, 2018, **72**, 16-23.
30. W. E. Blumberg and J. Peisach, in *Probes of structure and function of macromolecules and membranes*, eds. B. Chance, T. Yonetani and A. S. Mildvan, Academic Press, New York, 1971, vol. 2, pp. 215-229.
31. D. A. Svistunenko, B. J. Reeder, M. M. Wankasi, R. L. Silaghi-Dumitrescu, C. E. Cooper, S. Rinaldo, F. Cutruzzola and M. T. Wilson, Reaction of *Aplysia limacina* metmyoglobin with



- hydrogen peroxide, *Dalton Trans*, 2007, DOI: 10.1039/b615770j, 840-850.
32. E. Droghetti, B. D. Howes, A. Feis, P. Dominici, M. Fittipaldi and G. Smulevich, The quantum mechanically mixed-spin state in a non-symbiotic plant hemoglobin: the effect of distal mutation on AHb1 from *Arabidopsis thaliana*, *J Inorg Biochem*, 2007, **101**, 1812-1819.
33. M. M. Maltempo, T. H. Moss and M. A. Cusanovich, Magnetic studies on the changes in the iron environment in Chromatium ferricytochrome *c*', *Biochim Biophys Acta*, 1974, **342**, 290-305.
34. M. Bellei, C. A. Bortolotti, G. Di Rocco, M. Borsari, L. Lancellotti, A. Ranieri, M. Sola and G. Battistuzzi, The influence of the Cys46/Cys55 disulfide bond on the redox and spectroscopic properties of human neuroglobin, *J Inorg Biochem*, 2018, **178**, 70-86.
35. D. Hamdane, L. Kiger, S. Dewilde, B. N. Green, A. Pesce, J. Uzan, T. Burmester, T. Hankeln, M. Bolognesi, L. Moens and M. C. Marden, Coupling of the heme and an internal disulfide bond in human neuroglobin, *Micron*, 2004, **35**, 59-62.
36. E. Vinck, S. Van Doorslaer, S. Dewilde and L. Moens, Structural change of the heme pocket due to disulfide bridge formation is significantly larger for neuroglobin than for cytoglobin, *J Am Chem Soc*, 2004, **126**, 4516-4517.
37. Y. Zhao, C. Hoganson, G. T. Babcock and M. A. Marletta, Structural changes in the heme proximal pocket induced by nitric oxide binding to soluble guanylate cyclase, *Biochemistry*, 1998, **37**, 12458-12464.
38. M. Brunori, A. Giuffrè, K. Nienhaus, G. U. Nienhaus, F. M. Scandurra and B. Vallone, Neuroglobin, nitric oxide, and oxygen: functional pathways and conformational changes, *Proc Natl Acad Sci U S A*, 2005, **102**, 8483-8488.
39. C. E. Cooper, Nitric oxide and iron proteins, *Biochim Biophys Acta*, 1999, **1411**, 290-309.
40. A. L. Tsai, V. Berka, E. Martin and J. S. Olson, A "sliding scale rule" for selectivity among NO, CO, and O(2) by heme protein sensors, *Biochemistry*, 2012, **51**, 172-186.
41. J. Uzan, S. Dewilde, T. Burmester, T. Hankeln, L. Moens, D. Hamdane, M. C. Marden and L. Kiger, Neuroglobin and other hexacoordinated hemoglobins show a weak temperature dependence of oxygen binding, *Biophys J*, 2004, **87**, 1196-1204.
42. H. R. Adams, D. A. Svistunenko, M. T. Wilson, S. Fujii, R. W. Strange, Z. A. Hardy, P. A. Vazquez, T. Dabritz, G. J. Streblov, C. R. Andrew and M. A. Hough, A heme pocket aromatic quadrupole modulates gas binding to cytochrome *c*'-beta: Implications for NO sensors, *J Biol Chem*, 2023, **299**, 104742.
43. A. Fago, A. L. Crumbliss, J. Peterson, L. L. Pearce and C. Bonaventura, The case of the missing NO-hemoglobin: spectral changes suggestive of heme redox reactions reflect changes in NO-heme geometry, *Proc Natl Acad Sci U S A*, 2003, **100**, 12087-12092.
44. T. Yonetani, A. Tsuneshige, Y. Zhou and X. Chen, Electron paramagnetic resonance and oxygen binding studies of alpha-Nitrosyl hemoglobin. A novel oxygen carrier having no-assisted allosteric functions, *J Biol Chem*, 1998, **273**, 20323-20333.
45. M. Negrerie, L. Bouzahir, J. L. Martin and U. Liebl, Control of nitric oxide dynamics by guanylate cyclase in its activated state, *J Biol Chem*, 2001, **276**, 46815-46821.
46. G. Silkstone, S. M. Kapetanaki, I. Husu, M. H. Vos and M. T. Wilson, Nitric oxide binds to the proximal heme coordination site of the ferrocyclochrome *c*/cardiolipin complex: formation mechanism and dynamics, *J Biol Chem*, 2010, **285**, 19785-19792.
47. P. Beckerson, B. J. Reeder and M. T. Wilson, Coupling of disulfide bond and distal histidine dissociation in human ferrous cytoglobin regulates ligand binding, *FEBS Lett*, 2015, **589**, 507-512.
48. R. W. Romberg and R. J. Kassner, Nitric oxide and carbon monoxide equilibria of horse myoglobin and (N-methylimidazole)protoheme. Evidence for steric interaction with the distal residues, *Biochemistry*, 1979, **18**, 5387-5392.
49. S. G. Kruglik, J. C. Lambry, S. Cianetti, J. L. Martin, R. R. Eady, C. R. Andrew and M. Negrerie, Molecular basis for nitric oxide dynamics and affinity with *Alcaligenes xylosoxidans* cytochrome *c*, *J Biol Chem*, 2007, **282**, 5053-5062.
50. U. Liebl, J. C. Lambry and M. H. Vos, Primary processes in heme-based sensor proteins, *Biochim Biophys Acta*, 2013, **1834**, 1684-1692.
51. L. Lobato, L. Bouzahir-Sima, T. Yamashita, M. T. Wilson, M. H. Vos and U. Liebl, Dynamics of the heme-binding bacterial gas-sensing dissimilative nitrate respiration regulator (DNR) and activation barriers for ligand binding and escape, *J Biol Chem*, 2014, **289**, 26514-26524.
52. P. Corti, J. Xue, J. Tejero, N. Wajih, M. Sun, D. B. Stolz, M. Tsang, D. B. Kim-Shapiro and M. T. Gladwin, Globin X is a six-coordinate globin that reduces nitrite to nitric oxide in fish red blood cells, *Proc Natl Acad Sci U S A*, 2016, **113**, 8538-8543.
53. M. T. Gladwin and D. B. Kim-Shapiro, The functional nitrite reductase activity of the heme-globins, *Blood*, 2008, **112**, 2636-2647.
54. F. B. Jensen, The role of nitrite in nitric oxide homeostasis: a comparative perspective, *Biochim Biophys Acta*, 2009, **1787**, 841-848.
55. S. Shiva, Z. Huang, R. Grubina, J. Sun, L. A. Ringwood, P. H. MacArthur, X. Xu, E. Murphy, V. M. Darley-Usmar and M. T. Gladwin, Deoxymyoglobin is a nitrite reductase that generates nitric oxide and regulates mitochondrial respiration, *Circ Res*, 2007, **100**, 654-661.
56. M. Tiso, J. Tejero, S. Basu, I. Azarov, X. Wang, V. Simplaceanu, S. Frizzell, T. Jayaraman, L. Geary, C. Shapiro, C. Ho, S. Shiva, D. B. Kim-Shapiro and M. T. Gladwin, Human neuroglobin functions as a redox-regulated nitrite reductase, *J Biol Chem*, 2011, **286**, 18277-18289.
57. S. Dewilde, L. Kiger, T. Burmester, T. Hankeln, V. Baudin-Creuz, T. Aerts, M. C. Marden, R. Caubergs and L. Moens, Biochemical characterization and ligand binding properties of neuroglobin, a novel member of the globin family, *J Biol Chem*, 2001, **276**, 38949-38955.
58. R. J. Rohlfs, J. S. Olson and Q. H. Gibson, A comparison of the geminate recombination kinetics of several monomeric heme proteins, *J Biol Chem*, 1988, **263**, 1803-1813.
59. M. J. Free, G. A. Schluntz and R. A. Jaffe, Respiratory gas tensions in tissues and fluids of the male rat reproductive tract, *Biol Reprod*, 1976, **14**, 481-488.
60. E. D. Massie, W. R. Gomes and N. L. VanDemark, Oxygen tension in testes of normal and cryptorchid rats, *J Reprod Fertil*, 1969, **19**, 559-561.



61. J. G. Reyes, J. G. Farias, S. Henriquez-Olavarrieta, E. Madrid, M. Parraga, A. B. Zepeda and R. D. Moreno, The hypoxic testicle: physiology and pathophysiology, *Oxid Med Cell Longev*, 2012, **2012**, 929285.
62. P. Corti, M. Ieraci and J. Tejero, Characterization of zebrafish neuroglobin and cytoglobins 1 and 2: Zebrafish cytoglobins provide insights into the transition from six-coordinate to five-coordinate globins, *Nitric Oxide*, 2016, **53**, 22-34.
63. G. Balercia, S. Moretti, A. Vignini, M. Magagnini, F. Mantero, M. Boscaro, G. Ricciardo-Lamonica and L. Mazzanti, Role of nitric oxide concentrations on human sperm motility, *J Androl*, 2004, **25**, 245-249.
64. S. Dutta and P. Sengupta, The Role of Nitric Oxide on Male and Female Reproduction, *Malays J Med Sci*, 2022, **29**, 18-30.
65. U. Muller and G. Bicker, Calcium-activated release of nitric oxide and cellular distribution of nitric oxide-synthesizing neurons in the nervous system of the locust, *J Neurosci*, 1994, **14**, 7521-7528. DOI: 10.1039/D4SC00953C
66. C. E. Van Hove, C. Van der Donckt, A. G. Herman, H. Bult and P. Franssen, Vasodilator efficacy of nitric oxide depends on mechanisms of intracellular calcium mobilization in mouse aortic smooth muscle cells, *Br J Pharmacol*, 2009, **158**, 920-930.
67. A. Charles, Nitric oxide pumps up calcium signalling, *Nat Cell Biol*, 1999, **1**, E193-195.
68. J. H. Lee, H. Kim, D. H. Kim and M. C. Gye, Effects of calcium channel blockers on the spermatogenesis and gene expression in peripubertal mouse testis, *Arch Androl*, 2006, **52**, 311-318.



ARTICLE

Figures

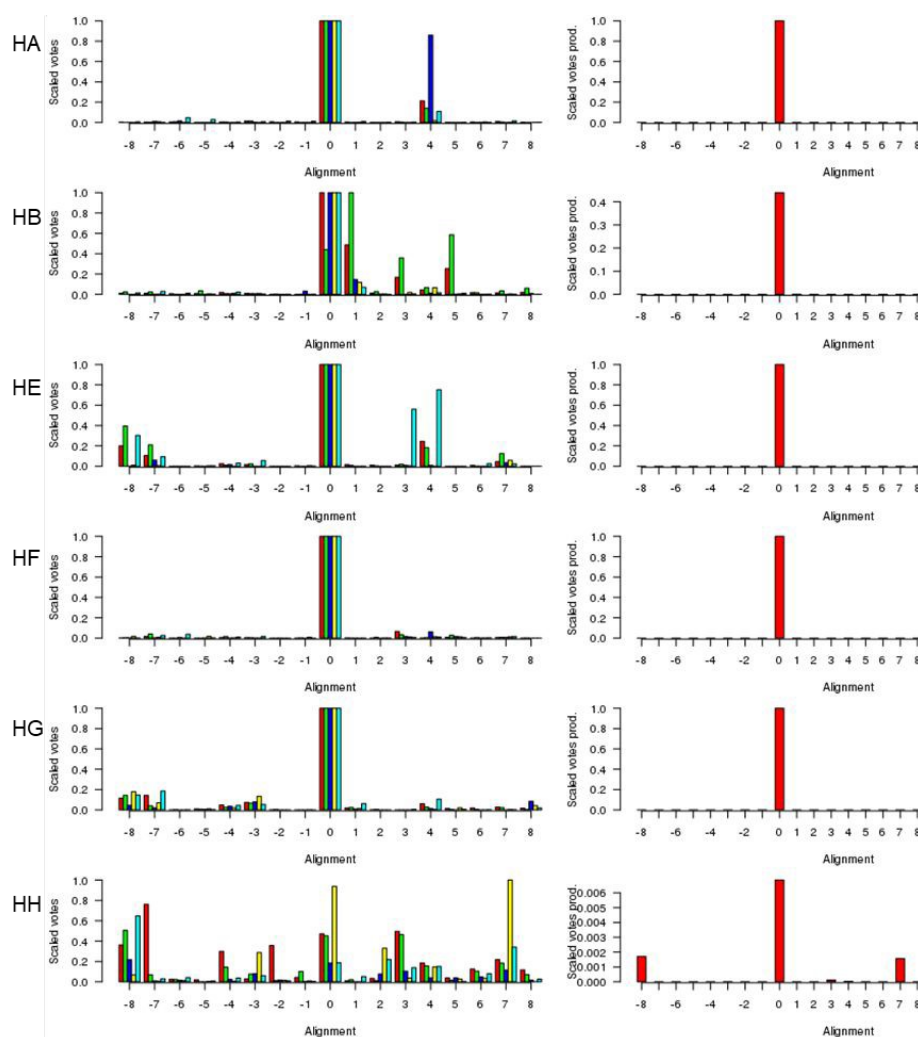


Figure 1. The alignment of the androglobin heme-binding globin domain helices. The left-hand column gives the individual alignments for helices A-B and helices E-H, one per row (denoted HA, HB etc.). The alpha-Hb (red), beta-Hb (green), Cygb (blue), Mb (yellow), and Ngb (cyan) alignments to Adgb are denoted by the bars representing the number of votes (scaled between 0 and 1). The right-hand column gives the consensus alignment where the votes for the individual alignments are multiplied together. The results points overwhelmingly to the 0 alignment given in Figure 2. An alignment of +1 would correspond to movement of the Adgb helix one residue to the right in Figure 2; an alignment of say -3 would correspond to movement of the Adgb helix three residues to the left in Figure 2.



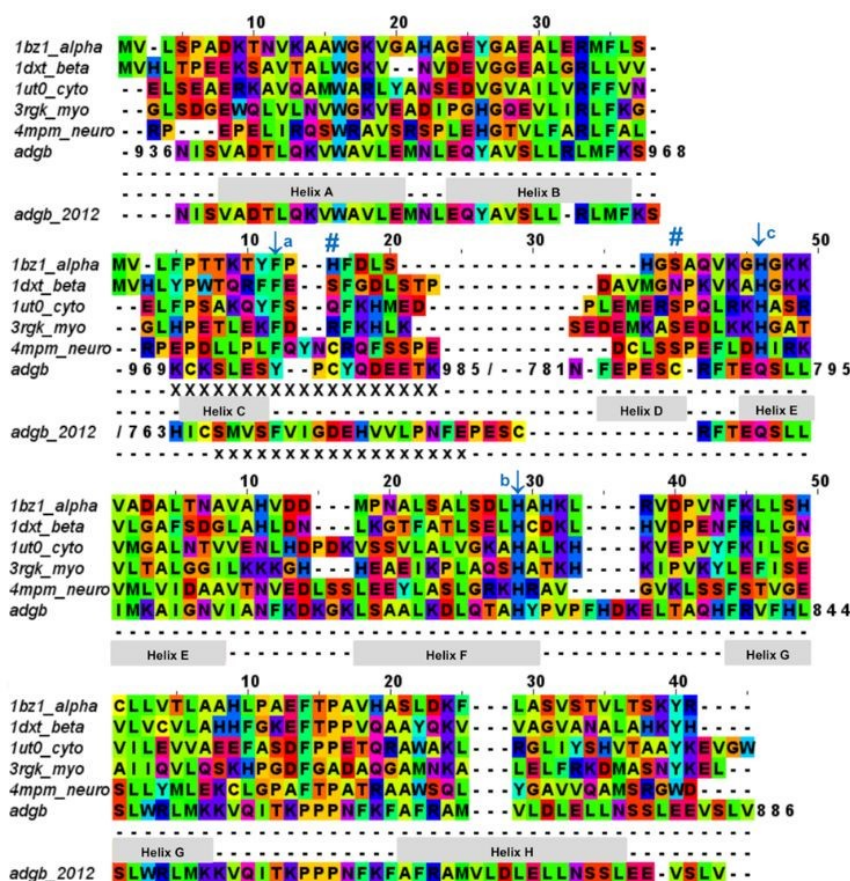


Figure 2. Androglobin heme-binding globin domain alignment. The alignment of the traditional globins was generated by structural alignment; the alignment of Adgb was determined using the in-house multi-template approach. The alignment denoted Adgb_2012, previously reported by Hoogewijs et al. 1, is shown where it differs from our alternative proposed alignment. The six alignment zones coincide (bar helices C and D) with globin helices denoted 'Helix A', 'Helix B' etc. as determined by inspection from the X-ray crystal structures. '/' denotes the chain break in the discontinuous Adgb sequence, 'XXX'... denotes the extra residues used in the Adgb construct (this work) while 'xxx...' denotes the extra residues in the globin domain of Adgb reported by Hoogewijs et al., 2012 and which here are identified by us as part of the preceding calpain C2-like domain (unpublished work). Key amino acids are denoted by labelled arrows, 'a' for the conserved CD1 loop aromatic residue, 'b' for proximal histidine and 'c' for the distal histidine/glutamine; the disulfide bond is denoted by '#'.
 Open Access Article. Published on 21 March 2024. Downloaded on 3/21/2024 12:33:10 PM.
 This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

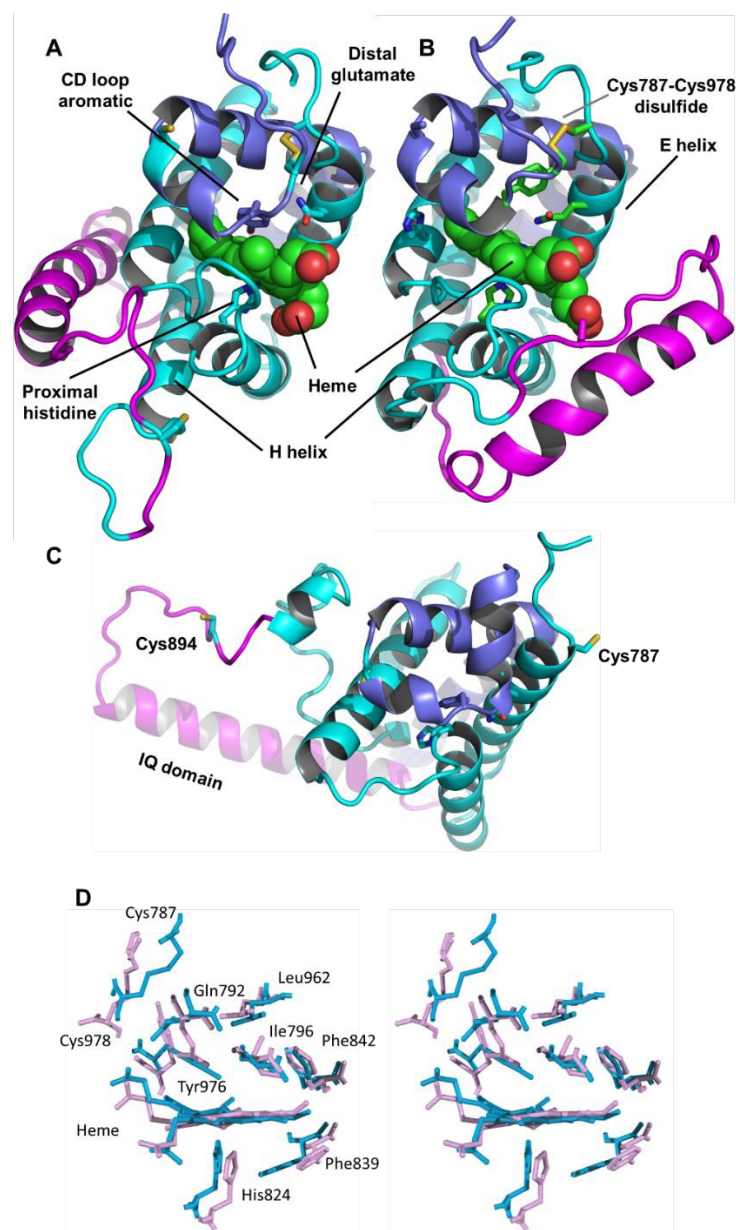
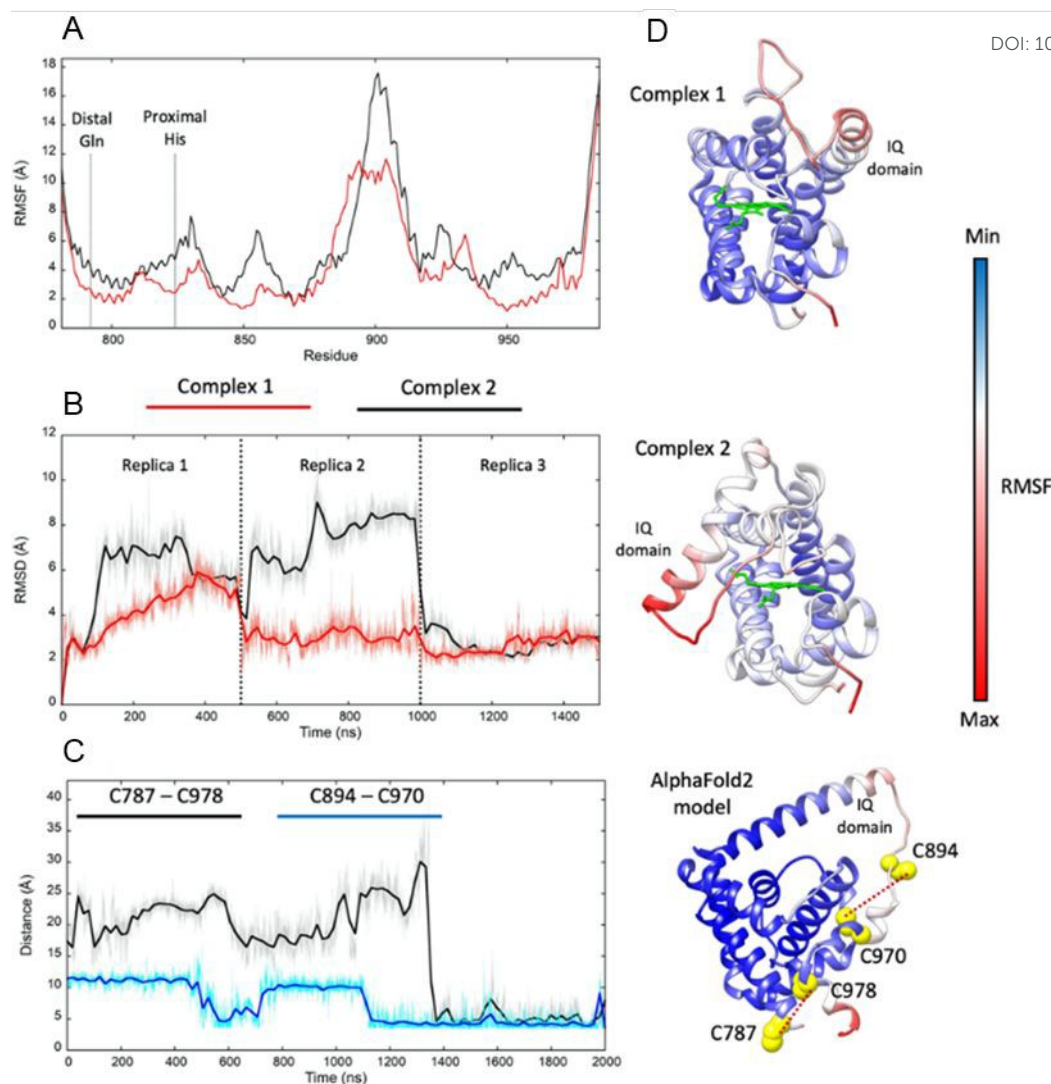


Figure 3. Structures of the heme-binding globin domain of androglobin. (A) The complex 1 structure determined using Modeller, (B) the complex 2 structure determined using Modeller and (C) The AlphaFold 2 structure. The structure is coloured blue for the N-terminus (helices D-H), magenta for the IQ domain insertion and cyan for the C terminus (helices A - C). (D) Stereoview of predicted heme environment from complex1 (blue) and complex 2 (purple).





View Article Online

DOI: 10.1039/D4SC00953C

Figure 4. Molecular dynamics simulations of Adgb-GD complex 1, complex 2, and AlphaFold2 model. (A) RMSF comparison between complex 1 (red) and 2 (black); (B) heme RMSD within complex 1 and 2 during the MD replicas; (C) distances between pairs of cysteine residues during the MD simulation of the AlphaFold2 model (C787-C978 in black, C894-C970 in blue); (D) RMSF plotted on complex 1, complex 2, and AlphaFold2 model (represented as ribbon, heme is represented as green stick); red ribbon colour indicates high structural flexibility; the four cysteine of the domain and their distance are shown on the AlphaFold2 model.



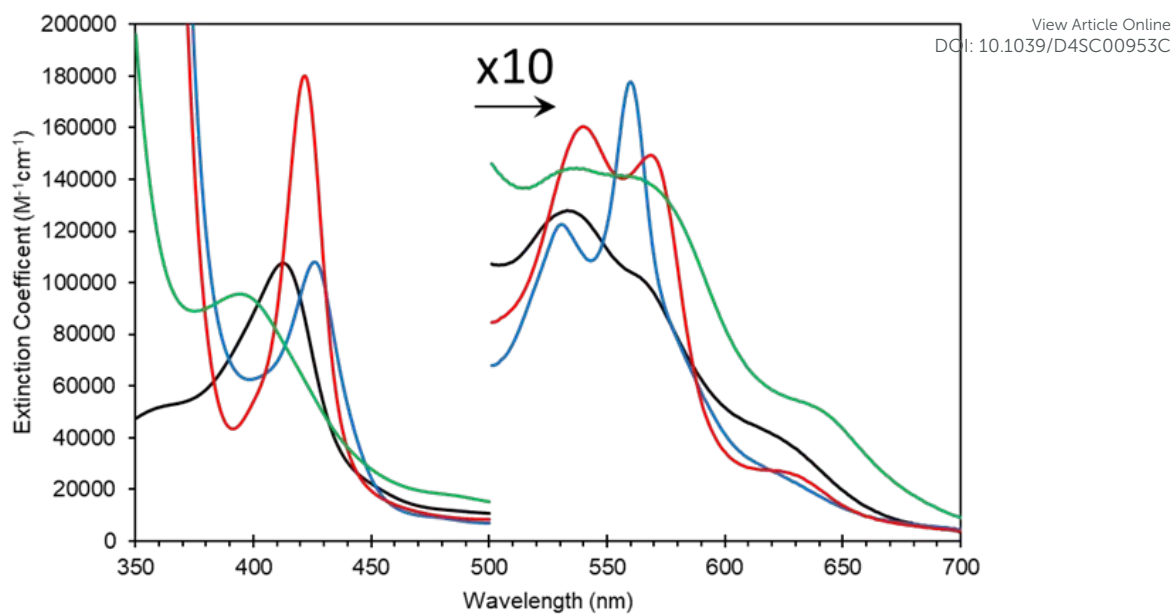


Figure 5. Optical characteristics of androglobin globin domain. Protein was expressed based on revised amino acid assignment shown in Figure 2 and S2. Ferric (black line), dithionite reduced deoxy ferrous (blue line), ferrous-CO bound (red line) and ferrous-NO bound (green line).



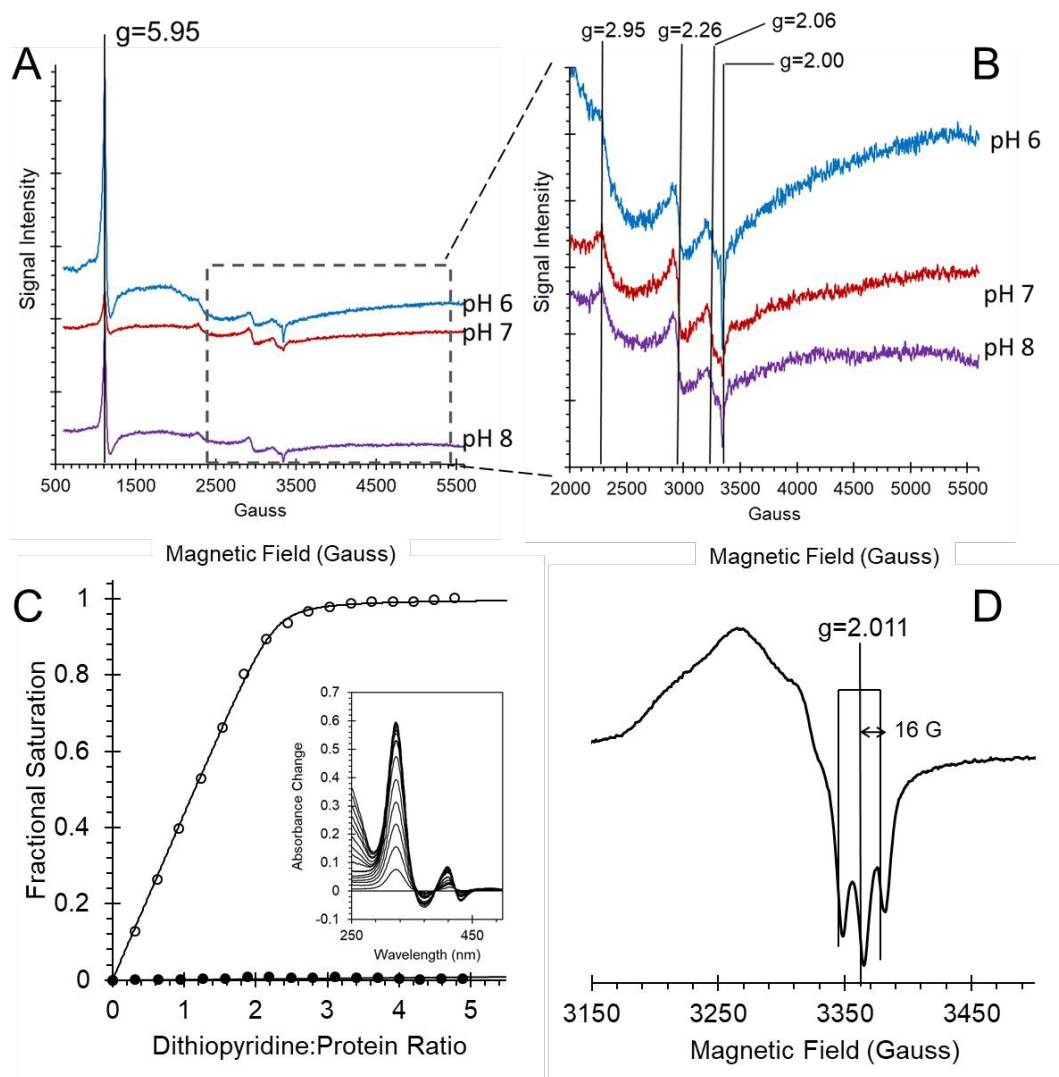


Figure 6. Androglobin heme iron ligation and cysteine oxidation states. (A), EPR spectra of 80 μM ferric androglobin globin domain showing both high spin and low spin Fe^{3+} EPR signals at various pH values. The spectra were recorded at 10 K, 3.16 mW microwave power and 5 G modulation amplitude. (B), Expanded view of (A) highlighting the low spin signals. (C), Measurement of surface exposed disulfide number and oxidation state. Protein (6.4 μM) was titrated with 4,4' dithiopyridine and the optical changes followed at 324 nm. The fractional saturation of dithiopyridine binding to free sulfhydryl as a function of dithiopyridine concentration for TCEP reduced (\circ) and purified form (no disulfide reduction, \bullet) shows that one surface-exposed disulfide bond is present in the globin domain when expressed. Inset shows optical changes of titration of TCEP reduced Adgb-GD with dithiopyridine. (D), An EPR spectrum of ferrous NO bound Adgb-GD, exhibiting three lines, separated by 16 G, around $g=2.011$, typical of other EPR spectra of five coordinate NO binding such as cyt c' . The spectrum was recorded at 10 K, 3.16 mW microwave power and 3 G modulation amplitude.



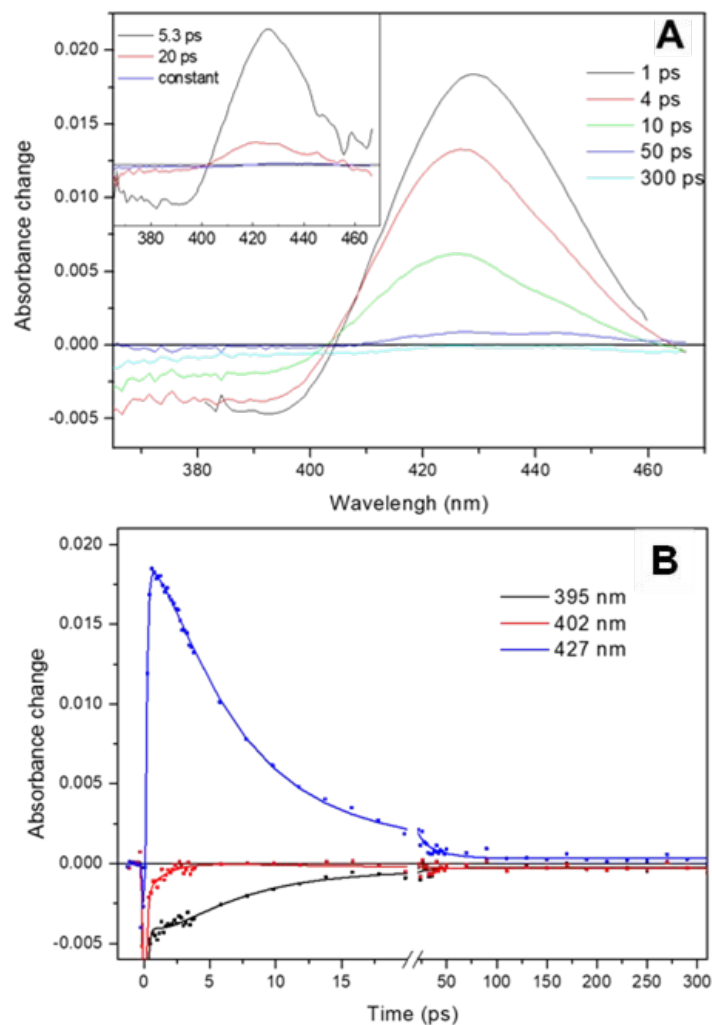


Figure 7. Ultrafast photo-dissociation and rebinding of NO from the ferrous Adgb-GD five coordinate NO complex. (A) Transient absorption spectra after different delay times upon excitation at 570 nm. Inset: Decay Associated Spectra corresponding to the NO geminate rebinding phases obtained from a global analysis (B) Dual-timescale kinetics and fits at selected wavelengths.



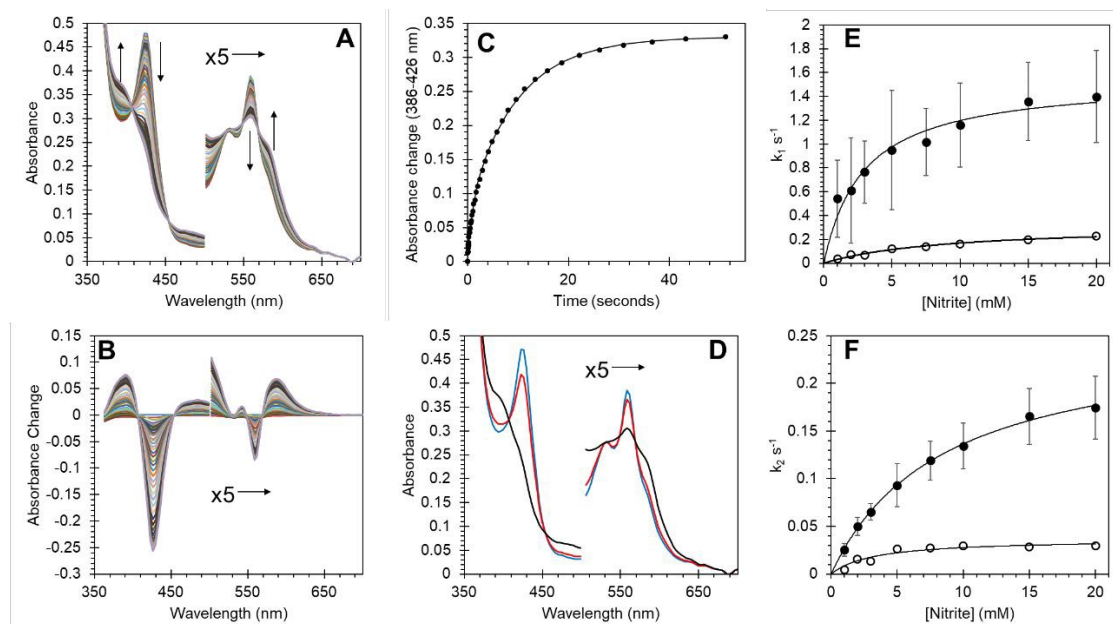


Figure 8. Nitrite reductase activity of Adgb-GD and effect of disulfide redox state on reductase activity. (A), Optical spectra of deoxyferrous Adgb-GD (5 μM) with sodium nitrite (5 mM). (B), Difference spectra with initial ferrous protein set to zero. (C), Time course of optical changes fitted to a double exponential function ($k_1 = 9.25 \times 10^{-1} \text{ s}^{-1}$, $k_2 = 1.02 \times 10^{-1} \text{ s}^{-1}$). (D), Global fit of initial deoxyferrous protein (blue), intermediate (red) and final ferrous-NO bound spectrum (black). (E) and (F), The dependence of the rate constants for Adgb nitrite reductase activity. The observed rate constants of the fast (E) and slow (F) phase on the reaction in the presence (●) or absence (by reduction using TCEP, ○) of the CD disulfide bond.

