



**HAL**  
open science

# Progesterone in the Brain: Hormone, Neurosteroid and Neuroprotectant

Rachida Guennoun

► **To cite this version:**

Rachida Guennoun. Progesterone in the Brain: Hormone, Neurosteroid and Neuroprotectant. International Journal of Molecular Sciences, 2020, 21 (15), pp.5271. 10.3390/ijms21155271 . hal-04509151

**HAL Id: hal-04509151**

**<https://hal.science/hal-04509151>**

Submitted on 18 Mar 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Review

# Progesterone in the Brain: Hormone, Neurosteroid and Neuroprotectant

Rachida Guennoun 

U 1195 Inserm and University Paris Saclay, University Paris Sud, 94276 Le Kremlin Bicêtre, France;  
rachida.guennoun@inserm.fr

Received: 4 May 2020; Accepted: 22 July 2020; Published: 24 July 2020



**Abstract:** Progesterone has a broad spectrum of actions in the brain. Among these, the neuroprotective effects are well documented. Progesterone neural effects are mediated by multiple signaling pathways involving binding to specific receptors (intracellular progesterone receptors (PR); membrane-associated progesterone receptor membrane component 1 (PGRMC1); and membrane progesterone receptors (mPRs)) and local bioconversion to  $3\alpha,5\alpha$ -tetrahydroprogesterone ( $3\alpha,5\alpha$ -THPROG), which modulates GABA<sub>A</sub> receptors. This brief review aims to give an overview of the synthesis, metabolism, neuroprotective effects, and mechanism of action of progesterone in the rodent and human brain. First, we succinctly describe the biosynthetic pathways and the expression of enzymes and receptors of progesterone; as well as the changes observed after brain injuries and in neurological diseases. Then, we summarize current data on the differential fluctuations in brain levels of progesterone and its neuroactive metabolites according to sex, age, and neuropathological conditions. The third part is devoted to the neuroprotective effects of progesterone and  $3\alpha,5\alpha$ -THPROG in different experimental models, with a focus on traumatic brain injury and stroke. Finally, we highlight the key role of the classical progesterone receptors (PR) in mediating the neuroprotective effects of progesterone after stroke.

**Keywords:** progesterone; PR; allopregnanolone; neuroprotection; neurosteroid; stroke; traumatic brain injury; TBI

## 1. Introduction

Steroid hormones are synthesized by adrenal glands, gonads, and placenta and influence the function of many target tissues including the nervous system. In addition, some steroids are synthesized *de novo* by neurons and glial cells and are called “neurosteroids” to refer to their site of synthesis: the nervous system [1]. Studies from several laboratories have shown the expression in the nervous system of the enzymes involved in steroidogenesis (reviewed in [2–4]). From an evolutionary point of view, data from different species demonstrate that neurosteroidogenesis is a conserved feature across fish, amphibians, birds, and mammals [2,5,6]. In addition to the *de novo* synthesis of neurosteroids from cholesterol, some steroids can be locally converted in the nervous system to neuroactive metabolites, as in the case of progesterone which is metabolized into  $3\alpha,5\alpha$ -tetrahydroprogesterone ( $3\alpha,5\alpha$ -THPROG; allopregnanolone). Neuroactive steroids designate all the steroids that can regulate neural functions including steroid hormones, neurosteroids, and synthetic steroids. Neuroactive steroids exert important functions in both the central and peripheral nervous systems and may represent a promising therapeutic strategy for the treatment of nervous system disorders [4,5,7].

Among neuroactive steroids, progesterone and its metabolite  $3\alpha,5\alpha$ -THPROG have been extensively studied in brain, spinal cord, and sciatic nerve, and have been shown to regulate several functions including myelination, neuroprotection, and neuropathic pain [8–13]. This review aims to

give an overview of the synthesis, metabolism, neuroprotective effects, and mechanism of action of progesterone in the rodent and human brain.

## 2. Progesterone Synthesis, Metabolism and Mechanism of Action in Brain

### 2.1. Progesterone Is an Endogenous Hormone and a Neurosteroid

Progesterone is a steroid hormone synthesized by ovaries and placenta in females and by adrenal glands in males and females. Thanks to its small size and lipid solubility, circulating progesterone easily crosses the blood brain barrier (BBB) by free transmembrane transport and diffuses throughout the nervous tissues [14–17]. However, most steroids bind and are transported by hormone-binding proteins in blood and this binding modulates their availability and their transport via the BBB. Concerning progesterone, this binding has a modest influence on its transport to the brain. Thus, 83% of the  $^3\text{H}$ -labeled progesterone was found in the ipsilateral rat brain hemisphere after 15 s post-injection in Ringer's solution via the common carotid artery. This high transport of  $^3\text{H}$  progesterone via the BBB was not affected when the injection was in 67% rat serum and was slightly reduced by 18% when the bolus injection was in 67% human serum [14].

Progesterone is also a neurosteroid as it can be synthesized locally in the nervous system by glia and neurons [1,2,9,18–21]. Progesterone synthesis requires the conversion of cholesterol to pregnenolone by the cytochrome P450scc enzyme and then the conversion of pregnenolone to progesterone by the  $3\beta$ -hydroxysteroid dehydrogenases ( $3\beta$ -HSD). Furthermore, progesterone can be sequentially converted to its neuroactive  $5\alpha$ -reduced metabolites:  $5\alpha$ -dihydroprogesterone ( $5\alpha$ -DHPROG) by  $5\alpha$ -reductases; and  $3\alpha$ - $5\alpha$ -THPROG by  $3\alpha$ -hydroxysteroid dehydrogenases ( $3\alpha$ -HSD) (Figure 1). Thus, the progesterone pool in the nervous system depends on (1) its peripheral synthesis, brain uptake, and accumulation; (2) its local synthesis; and (3) its metabolism.

### 2.2. Enzymes Involved in the Synthesis of Progesterone and Its Neuroactive Metabolites

The expression in the brain of the enzymes involved in progesterone synthesis and metabolism is well documented (For review see [2–4,9,20,22]).

P450scc expression was first detected by immunohistochemistry in the white matter of rat brain [23]. Subsequent studies detected P450scc expression in Purkinje cells in rat cerebellum [24,25] and pyramidal and granule cells in the hippocampus [26]. Cyp11A1 mRNA encoding P450scc was detected in different brain regions of rats with the highest level in the cerebral cortex [27]. In human brain, P450scc expression was first detected by immunohistochemistry in white matter [28]; and the expression of Cyp11A mRNA was detected in the neocortex, hippocampus, amygdala, caudate nucleus, cerebellum, corpus callosum, and thalamus [29–31]. Activity of the P450scc has been demonstrated in oligodendrocytes from rodents [32] and humans [33].

$3\beta$ -HSD mRNA was detected in several regions of rat brain [22]. Using in situ hybridization, we showed a large distribution of  $3\beta$ -HSD mRNA in particular in the olfactory bulb, striatum, cortex, thalamus, hypothalamus, septum, hippocampus, habenula, and cerebellum [34].  $3\beta$ -HSD mRNA expression was detected in different regions of human brain including the amygdala, caudate nucleus, corpus callosum, cerebellum, hippocampus, thalamus, and pons. The highest levels were measured in the corpus callosum [31,35]. Human oligodendroglial, astroglial, and neuronal cell lines in cultures express  $3\beta$ -HSD [33].  $3\beta$ -HSD activity was demonstrated in rats using homogenates of amygdala and septum [36] and primary cultures of neurons, oligodendrocytes, and astrocytes [37–42].

$5\alpha$ -reductase 1 is the most abundant isoform of  $5\alpha$ -reductases expressed in the brain of mice, rats, and humans [43–49]. Recently, Giatti et al. showed that the expression of  $5\alpha$ -reductase mRNA in the rat cerebellum is higher in males compared to females [50].  $5\alpha$ -reductase 1 mRNA was detected in the mouse brain in neurons of the olfactory bulb, cortex, hippocampus, amygdala, and cerebellum [51]. In humans,  $5\alpha$ -reductase 1 mRNA was detected in the cortex, hippocampus, cerebellum, hypothalamus, and pons [46,47,49]. In rat brain,  $5\alpha$ -reductase 1 was detected by immunohistochemistry in the

hypothalamus, thalamus, hippocampus, cortex, and circumventricular organs [52,53]. At the cellular level, the 5 $\alpha$ -reductase 1 immunoreactivity was described in glial and ependymal cells. 5 $\alpha$ -reductase activity has been demonstrated in rat [54] and human brain [45,46,55]. Using rat neuronal cell cultures, Melcangi et al. demonstrated that neurons have a higher 5 $\alpha$ -reductase activity than astrocytes and oligodendrocytes [56]. Interestingly, the activity of 5 $\alpha$ -reductase in astrocytes is stimulated by co-culture with neurons or simply by addition of neuron-conditioned medium, suggesting stimulating diffusible factors from neurons [57]. One remarkable finding is the dramatic changes in steroidogenic activity observed during oligodendrocyte differentiation [42]. In particular, the activity of the 5 $\alpha$ -reductase was five times higher in mature oligodendrocytes than in their pre-progenitors and their progenitors.

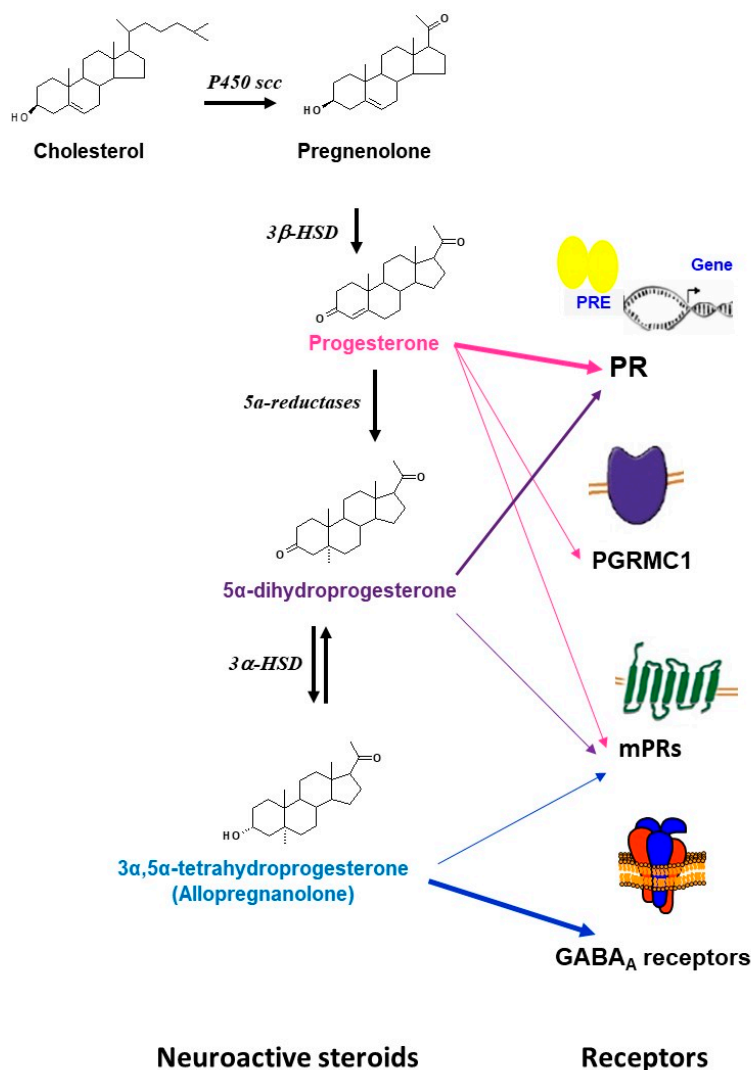
3 $\alpha$ -HSD has been detected in the cortex, hippocampus, olfactory bulb, amygdala, and thalamus of both rat and mouse brain [51,58]. In rat cerebellum, the expression of 3 $\alpha$ -HSD mRNA was higher in females compared to males and decreased after gonadectomy specifically in females [50]. 3 $\alpha$ -HSD activity has been demonstrated in several regions of rat brain [58]. In neuronal cell cultures from rat brain, 3 $\alpha$ -HSD activity was higher in astrocytes than in oligodendrocytes and neurons [56]. The investigation of progesterone metabolism at different stages of the oligodendrocyte lineage differentiation in vitro showed that 3 $\alpha$ -HSD activity was ten times lower in the mature oligodendrocytes than in their pre-progenitors and progenitors [42]. mRNA of 3 $\alpha$ -HSD type 2 and type 3 were detected in several regions of the human brain [22]. 3 $\alpha$ -HSD activity has been demonstrated in cortex and white matter from biopsies from patients suffering from epilepsy [45].

It is important to note that the expression of the enzymes of neurosteroidogenesis has been reported to change under some pathological conditions. For example, there was a decrease in the 3 $\beta$ -HSD mRNA expression in the contusion site within the frontal cortex in both males and pseudopregnant female rats 24 h after traumatic brain injury (TBI) [59]. In a recent study, Leicaj et al. evaluated the expression of different enzymes of steroidogenesis in the brain of mice subjected to demyelination by cuprizone, a model of multiple sclerosis. They showed that the expression of P450scc and 5 $\alpha$ -reductase was decreased in the hippocampus, cortex, and corpus callosum during the demyelination period and was restored to control levels during remyelination [60]. Using another model of multiple sclerosis, Noorbakhsh et al. have reported a significant decrease in 3 $\alpha$ -HSD expression in the brain of autoimmune encephalomyelitis (EAE) mice [61]. Interestingly, studies in the Niemann–Pick type C (NP-C) mice, a model of fatal neurodegenerative human disease, showed a marked reduction in neurosteroidogenesis and suggest that this could contribute to the pathogenesis of the disease [62,63]. In fact, a reduced activity of 3 $\alpha$ -HSD was observed in several brain regions of NP-C mice at birth. A reduced activity of 5 $\alpha$  reductase was observed some weeks later before the onset of symptoms. In adult brains of NP-C mice, the expressions of P450scc, 3 $\beta$ HSD, 5 $\alpha$ - reductase, and 3 $\alpha$ -HSD were decreased in the cortex and cerebellum. In line with this, a dramatic decrease in the brain levels of 3 $\alpha$ ,5 $\alpha$ -THPROG was reported in NP-C mice. Furthermore, symptoms were delayed by the administration of 3 $\alpha$ ,5 $\alpha$ -THPROG [62]. In studies using postmortem human brain tissues, down-regulation of 5 $\alpha$ -reductase type 1 and upregulation of 3 $\alpha$ -HSD type 3 have been reported in substantia nigra of Parkinson's disease patients [64]. In multiple sclerosis lesions, an up-regulation of 3 $\beta$ -HSD mRNA was observed in females but not in males [65]. In another study, a down-regulation of the transcripts of 5 $\alpha$ -reductase type 1 and of 3 $\alpha$ -HSD type 1 has been reported in the brains of multiple sclerosis patients [61].

### 2.3. Progesterone Receptors' Expression in the Brain

Progesterone has multiple receptors and may activate several signaling pathways [9,20,21,66–69]. The first characterized are the intracellular receptors (PR). PR-A and PR-B are transcribed from two promoters of the same gene [70,71]. PRs act as nuclear transcription factors [9,70]. However, they have been also shown to interact with membrane-associated kinases and activate extranuclear signaling pathways [72]. In addition to PR, progesterone binds to the putative membrane-associated progesterone receptor component 1 (PGRMC1) that activates Jak/STAT, Src pathways, and protein kinase G [73–76].

Progesterone may also act after binding to the membrane progesterone receptors (mPR) that activate G-proteins and MAPK cascades. Five isoforms of mPR (mPR $\alpha$ , mPR $\beta$ , mPR $\gamma$ , mPR $\delta$ , mPR $\epsilon$ ) encoded by distinct genes were cloned and characterized by Thomas and colleagues [77–79]. In addition, progesterone can be locally bio-converted to 3 $\alpha$ ,5 $\alpha$ -THP (allopregnanolone) that modulates GABA $_A$  receptors [9,80] (Figure 1). All these progesterone receptors are expressed and widely distributed in the brain and can account for the progesterone actions (for review see [9,20,66,68,69,73]).



**Figure 1.** Schematic representation of the synthesis, metabolism, and main receptors that are involved in the actions of progesterone and its metabolites in brain. The precursor cholesterol is first converted into pregnenolone by the side chain cleavage cytochrome (P450scc) enzyme, pregnenolone is then converted to progesterone by the 3 $\beta$ -hydroxysteroid dehydrogenases (3 $\beta$ -HSD). Progesterone can further be bio-converted to 5 $\alpha$ -dihydroprogesterone by 5 $\alpha$ -reductases. 5 $\alpha$ -dihydroprogesterone is converted to 3 $\alpha$ ,5 $\alpha$ -tetrahydroprogesterone (allopregnanolone) by the 3 $\alpha$ -hydroxysteroid oxidoreductase (3 $\alpha$ -HSD) enzyme. Progesterone can bind to multiple receptors including the classical intracellular receptors (PR), the membrane receptors (mPRs: mPR  $\alpha$ , mPR $\beta$ , mPR $\gamma$ , mPR $\delta$ , mPR $\epsilon$ ), and the membrane-binding sites (PGRMC1). Some effects of progesterone may be mediated by its neuroactive metabolites. 5 $\alpha$ -dihydroprogesterone binds to the classical receptors PR and has relatively high binding affinity for mPR $\alpha$ . 3 $\alpha$ ,5 $\alpha$ -tetrahydroprogesterone (allopregnanolone) has no affinity for the intracellular PR, but is a potent allosteric modulator of GABA $_A$  receptors. Some neuroprotective effects of allopregnanolone may be mediated by the membrane progesterone receptor mPR $\delta$ .

It is important to note that after brain injury, changes in progesterone receptors' expression have been observed. For example, we have shown that PGRMC1 was expressed in regions involved in cerebrospinal fluid production and in osmoregulation, and that its expression was increased in neurons and induced in astrocytes after traumatic brain injury (TBI) in both male and pseudopregnant female rats [81]. These observations suggest that PGRMC1 may be involved in the maintenance of water homeostasis and in decreasing edema formation after TBI. We have also shown that mPR $\alpha$  is expressed only by neurons in the absence of injury and that its expression is induced in microglia, astrocytes, and oligodendrocytes after TBI, suggesting its potential role in mediating the progesterone effects in inflammation, edema formation, and myelin repair after TBI [82]. After a transient cerebral ischemia, no significant changes in PR mRNA were noticed in the cortical penumbra obtained from cerebral cortex of male rats at 24h after ischemia [83]. However, another study showed an up-regulation of PR protein in the cytosol fraction prepared from the cortical penumbra at 24 and 48h post-ischemia [84]. After a permanent cerebral ischemia in rats, Stanojlovic et al. showed that levels of PR were decreased in both the cytosolic and nuclear fractions from the prefrontal cortex at 7 days post-injury and that progesterone treatment restored the PR levels to those observed in rats subjected to sham-operation [85]. Interestingly, a recent study in mice showed a decrease in PGRMC1 protein level in the cortical penumbra at day 7 following cerebral ischemia [86]. In studies using postmortem brain tissues, an up-regulation of PR expression was observed in multiple sclerosis lesions from female patients but not in male patients [65]. This difference may contribute to the observed gender differences in multiple sclerosis.

These observations provide examples indicating that progesterone signaling may be adjusted in response to injuries and thus it may differ according to physiological conditions and after brain injuries.

### 3. Brain Levels of Progesterone and Its Metabolites

#### 3.1. Sex Differences and Decline during Aging

Effects of cycle, sex, and age on brain levels of progesterone and its metabolites are summarized in Table 1.

In mice, significant levels of progesterone, 5 $\alpha$ -DHPROG, and 3 $\alpha$ -5 $\alpha$ -THPROG were measured in brains of both young adult males and females [87–90]. In female mice, we have shown that brain levels of progesterone and 5 $\alpha$ -DHPROG changed significantly according to estrus cycle [87]. Thus, brain levels of progesterone were in the same range or much higher in the brain of females compared to males depending on the phase of the cycle. In contrast, brain levels of 3 $\alpha$ -5 $\alpha$ -THPROG were not significantly different between males and females [87]. In rats, significant levels of progesterone, 5 $\alpha$ -DHPROG, and 3 $\alpha$ -5 $\alpha$ -THPROG were measured in both young adult males and females [91,92]. Increased brain levels of progesterone, 5 $\alpha$ -DHPROG, and 3 $\alpha$ -5 $\alpha$ -THPROG have been reported in pseudopregnant rats [91]. Evaluation of neurosteroid levels in different brain regions of male and female (at diestrus) rats showed subtle sex differences [92]. For instance, levels of progesterone in the cerebral cortex and the cerebellum were higher in males compared to females. In contrast, levels of 5 $\alpha$ -DHPROG were higher in the hippocampus, cerebral cortex, and cerebellum in females compared to males. Levels of 3 $\alpha$ -5 $\alpha$ -THPROG were higher in the hippocampus and cerebral cortex in females compared to males [92]. Bixo and colleagues have measured the levels of 5 $\alpha$ -DHPROG, and 3 $\alpha$ -5 $\alpha$ -THPROG in different regions of postmortem brain tissues from young and postmenopausal women. The highest levels of progesterone were measured in the hypothalamus, amygdala, and cerebellum; while the highest levels of 5 $\alpha$ -DHPROG, and 3 $\alpha$ -5 $\alpha$ -THPROG were measured in the substantia nigra and hypothalamus [93].

Aging is associated with a decrease in steroid production. We have evaluated the profile of progesterone and its metabolites in the brain of young (3-month-old) and aged (20-month-old) male and female mice. Our data showed a strong decrease in brain levels of progesterone, 5 $\alpha$ -DHPROG, and 3 $\alpha$ -5 $\alpha$ -THPROG in aged mice of both sexes. Interestingly, the sex differences observed in young mice were no longer observed in aged mice [87]. In rats, comparison of profiles in the limbic region



of 7- and 24-month-old males showed decreased levels of progesterone and 3 $\alpha$ -5 $\alpha$ -THPROG but surprisingly increased levels of 5 $\alpha$ -DHPROG [94]. Interestingly, analysis of postmortem human brain tissues showed that brain levels of progesterone, 5 $\alpha$ -DHPROG, and 3 $\alpha$ -5 $\alpha$ -THPROG were higher in young women at the luteal phase of the cycle than in postmenopausal women suggesting a dependence of their brain levels on the synthesis in the periphery and a decline during aging [93].

**Table 1.** Brain levels of progesterone (PROG), 5 $\alpha$ -dihydroprogesterone (5 $\alpha$ -DHPROG), and 3 $\alpha$ ,5 $\alpha$ -tetrahydroprogesterone (3 $\alpha$ ,5 $\alpha$ -THPROG): effect of cycle, sex, and age.

Steroid	Cycle Effect	Sex Effect	Age Effect	Species/Structures	References
PROG	D > P > E	F (D) > M	Decrease		
5 $\alpha$ -DHPROG	D > P > E	NS	Decrease	Mice/Cerebral hemisphere	[87]
3 $\alpha$ ,5 $\alpha$ -THPROG	NS	NS	NS		
		NS		Rats/Hippocampus	
PROG		M > F (D)		Rats/Cortex, Cerebellum	[92]
5 $\alpha$ -DHPROG		F (D) > M		Rats/Hippocampus, Cortex, Cerebellum	
3 $\alpha$ ,5 $\alpha$ -THPROG		F (D) > M		Rats/Hippocampus, Cortex	
PROG			Decrease		
5 $\alpha$ -DHPROG			Increase	Rats/Limbic regions	[94]
3 $\alpha$ ,5 $\alpha$ -THPROG			Decrease		
PROG			Decrease		
5 $\alpha$ -DHPROG			Decrease	Human/Cortex, Amygdala, Hippocampus, Striatum, Thalamus	[93]
3 $\alpha$ ,5 $\alpha$ -THPROG			Decrease		

Sex effect: 3-month-old male mice vs. 3-month-old female mice in diestrus stage. Age effect: 20-month-old mice vs. 3-month-old mice of both sexes; 24-month-old male rats vs. 7-month-old male rats; postmenopausal (59–75 years) women vs. fertile women (18–38 years) at the luteal phase. Stages of the estrus cycle: proestrus (P); estrus (E); diestrus (D). F: females; M: males; NS: not significant.

### 3.2. Levels in Neurodegenerative Conditions and in Response to Brain Injuries

Changes in brain levels of progesterone and its neuroactive metabolites were observed under neurodegenerative conditions and in response to brain injuries (Table 2).

Increased levels of 5 $\alpha$ -DHPROG were measured in the limbic region of the brain of Alzheimer's disease mice model 3xTg-AD mice compared to wild type littermates [94]. In contrast, decreased levels of 5 $\alpha$ -DHPROG were measured in the striatum of male rats treated with 6-hydroxydopamine (an experimental model of Parkinson's disease) [95], and in the cerebral spinal fluid of Parkinson's disease patients [96]. Analysis of steroid levels in an experimental autoimmune encephalomyelitis (EAE) rat model, showed sex- and region-specific changes at the acute phase (14 days after immunization). For instance, in the cerebellum there was a decrease in progesterone in males and a decrease in 3 $\alpha$ ,5 $\alpha$ -THPROG in females. In the cerebral cortex, there was a decrease in progesterone and 5 $\alpha$ -DHPROG levels and an increase in 3 $\alpha$ ,5 $\alpha$ -THPROG levels in males; while in females no changes were observed [97]. The same group has also measured the levels of steroids at the chronic phase of EAE (40 days post-immunization). They showed a decrease in progesterone, 5 $\alpha$ -DHPROG, and 3 $\alpha$ ,5 $\alpha$ -THPROG in the cerebral cortex of males and in the cerebellum of females [98]. Decreased levels of 3 $\alpha$ ,5 $\alpha$ -THPROG were also reported in frontal lobe normal white matter near lesions obtained from male multiple sclerosis patients [61]. Levels of progesterone and its reduced metabolites were also altered after brain injuries. Thus, we have shown a transient increase in brain levels of progesterone and 5 $\alpha$ -DHPROG, 6 h post-injury in male rats subjected to bilateral cortex contusion, a model of TBI. A transient increase of 5 $\alpha$ -DHPROG level at 6 h post-injury, was also observed in pseudopregnant rat females that already had very high brain levels of progesterone and 5 $\alpha$ -DHPROG at the time of injury. At 24 h post-injury, no significant differences were observed in both males and pseudopregnant females [91]. More recent studies investigated the potential dysregulation of steroids at 24 h, 72 h, and 2 weeks post-injury using the weight drop model of TBI in young male and female mice. In male

mice, no differences were observed in brain levels at the three end points analyzed [89]. Surprisingly, decreased brain levels of progesterone, 5 $\alpha$ -DHPROG, and 3 $\alpha$ ,5 $\alpha$ -THPROG were measured in brain of female mice at the three times studied [88].

**Table 2.** Levels of progesterone (PROG), 5 $\alpha$ -dihydroprogesterone (5 $\alpha$ -DHPROG), and 3 $\alpha$ ,5 $\alpha$ -tetrahydroprogesterone (3 $\alpha$ ,5 $\alpha$ -THPROG) under neurodegenerative conditions and in response to brain injuries.

Pathology/Experimental Model	Sex/Species	Steroids/Change	Structures	References
AD/3xTg-AD mice	Male mice	5 $\alpha$ -DHPROG $\uparrow$	Limbic region	[94]
P/ 6-OHDA	Male rats	5 $\alpha$ -DHPROG $\downarrow$	Cortex, Striatum	[95]
PD	Men	5 $\alpha$ -DHPROG $\downarrow$	CSF	[96]
EAE acute phase	Male rats	PROG $\downarrow$	Cerebellum	[97]
		5 $\alpha$ -DHPROG $\downarrow$ 3 $\alpha$ ,5 $\alpha$ -THPROG $\uparrow$		
EAE chronic phase	Female rats	3 $\alpha$ ,5 $\alpha$ -THPROG $\downarrow$	Cerebellum	[98]
	Male and female rats	PROG $\downarrow$ 5 $\alpha$ -DHPROG $\downarrow$ 3 $\alpha$ ,5 $\alpha$ -THPROG $\downarrow$	Cortex of males Cerebellum of females	
MS	Men	3 $\alpha$ ,5 $\alpha$ -THPROG $\downarrow$	Frontal lobe	[61]
TBI/Bilateral cortex contusion	Male rats	PROG $\uparrow$ 5 $\alpha$ -DHPROG $\uparrow$ (6 h post-TBI)	Brain	[91]
	Pseudopregnant Female rats	5 $\alpha$ -DHPROG $\uparrow$ (6 h post-TBI)		
	Male mice	PROG = 5 $\alpha$ -DHPROG = 3 $\alpha$ ,5 $\alpha$ -THPROG = (24 h, 72 h, 2 weeks post-TBI)		
TBI/weight drop model	Female mice	PROG $\downarrow$ 5 $\alpha$ -DHPROG $\downarrow$ 3 $\alpha$ ,5 $\alpha$ -THPROG $\downarrow$ (24 h, 72 h, 2 weeks post-TBI)	Brain	[88]
Severe TBI	Men	PROG $\uparrow$ (Day 0 post-TBI)	Plasma	[99]
	Men and Women	PROG $\downarrow$ (Days 1 to 6 post-TBI)		
Stroke MCAO/R	Male mice	PROG $\uparrow$ (4 h, 6 h post-MCAO) 5 $\alpha$ -DHPROG $\uparrow$ 3 $\alpha$ ,5 $\alpha$ -THPROG = (1,2,4,6, 24 h post-MCAO)	Cerebral hemisphere	[90]
	Female mice	PROG = 5 $\alpha$ -DHPROG = 3 $\alpha$ ,5 $\alpha$ -THPROG = (1,2,4,6, 24 h post-MCAO)		

6-OHDA: 6-hydroxydopamine; AD: Alzheimer's disease; CSF: cerebrospinal fluid; EAE: Experimental autoimmune encephalomyelitis; MCAO/R: Middle cerebral artery occlusion/reperfusion; MS: multiple sclerosis; PD: Parkinson's disease; TBI: traumatic brain injury.  $\uparrow$  Increase;  $\downarrow$  Decrease; = No change.

Unfortunately, there are no studies using post-mortem brain tissue from patients that died after TBI. However, a study using plasma from patients suffering from severe TBI showed a transient increase of progesterone levels specifically in men at day 0 post-injury, then a dramatic decrease from day 1 to day 6 post-injury in both men and women [99].

Changes in brain progesterone and 5 $\alpha$ -DHPROG levels were also found in experimental stroke. For instance, using an experimental model of stroke by occlusion of the middle cerebral artery followed by reperfusion (MCAO/R) we showed an up-regulation of progesterone and 5 $\alpha$ -DHPROG in both the contralateral and ipsilateral hemisphere of male mice at 6 h post-MCAO/R [100]. In a subsequent study, we performed a steroid profiling at different end points: 1, 2, 4, 6, and 24 h post-MCAO/R in both male and female mice [90]. Interestingly, in males we confirmed the increase at 6 h and showed



that progesterone levels increased as early as 4 h, reaching a maximum at 6 h, then decreased to levels similar to controls at 24 h. Surprisingly, no significant variations were observed in these steroid levels in brains of female mice [90].

These observations provide examples indicating that brain levels of progesterone and its metabolites are dysregulated under pathological conditions and may be adjusted to respond to neurodegeneration and injuries. It is important to keep in mind that these regulations may be age-, sex-, and time-dependent. The decreases may reflect dysregulations associated with the pathologies and in this case normalization to control levels may have beneficial effects. On the other hand, increases may be part of endogenous neuroprotective and rescue processes and stimulation of the endogenous synthesis or exogenous administration of these steroids may optimize these natural responses.

#### 4. Progesterone and Its Metabolites for Brain Neuroprotection

Progesterone has a broad spectrum of actions in the brain including reproduction and sexual behavior [101,102], CNS development and differentiation [103,104], myelination [105], and neuroprotection. In this review, we focus on the neuroprotective effects.

##### 4.1. Neuroprotective Effects in Neurodegenerative Diseases Models

The neuroprotective effects of progesterone and its derivative  $3\alpha,5\alpha$ -THPROG were demonstrated in different experimental models of neurodegenerative disease. For example, treatment of ovariectomized female 3xTg-AD mice (a model of Alzheimer's disease) with progesterone alone or in combination with estradiol for 3 months specifically attenuated the hyperphosphorylation of Tau [106]. Extensive work by Brinton's team focuses on the development of  $3\alpha,5\alpha$ -THPROG as therapeutic for the treatment of Alzheimer's disease. They showed in particular that  $3\alpha,5\alpha$ -THPROG increased neurogenesis, improved cognitive function, and memory; decreased neuroinflammation and beta-amyloid accumulation; and restored the deficits of bioenergetics in the 3xTgAD mice [107–110]. Alzheimer's disease patients at the early phase of the disease are now recruited for clinical trial-phase 1, to identify the effective dose of  $3\alpha,5\alpha$ -THPROG (4–18 mg intramuscular weekly injections for 12 weeks) as a potential regenerative therapeutic for Alzheimer's disease (Allopregnanolone regenerative therapeutic for early Alzheimer's disease: intramuscular study (Allo-IM); Clinical Trials. Gov Identifier: NCT03748303).

In the case of Parkinson's disease (PD), no definitive conclusion could be drawn from studies that investigated the therapeutic potential of progesterone using experimental models. Indeed, the results depend on the dose of progesterone, the treatment regimen, the sex, and the model used [111–117]. Interestingly, treatment by  $3\alpha,5\alpha$ -THPROG improved cognitive and motor functions in Parkinson's disease experimental models [118,119].

Importantly, progesterone showed efficient remyelinating and anti-inflammatory effects in different experimental models of multiple sclerosis and demyelination. For example, progesterone was shown to reduce the degeneration of oligodendrocytes, to increase the number of their progenitors, and to reduce astrogliosis, demyelination, and neurological deficits induced by cuprizone in male mice [120]. In female mice, progesterone and its synthetic derivative Nestorone, the high affinity selective ligand of PR, promoted myelin repair in cerebral cortex and corpus callosum with chronic demyelinating lesions induced by Cuprizone. In particular, progesterone increased the density of mature oligodendrocytes and their progenitors and decreased the density of activated astrocytes and microglia [121]. More recently, the beneficial effects of progesterone in this model were confirmed and extended with a focus on the anti-inflammatory effects [122]. This study showed that progesterone treatment decreased neurological deficits and demyelination; induced a switch in microglia phenotype from pro-inflammatory M1 to anti-inflammatory M2 type as shown by the expression of different specific markers; and decreased the NOD-like receptor pyrin domain containing 3 (NLRP3) inflammasome and the pro-inflammatory interferon-gamma inducing factor 18 (Il-18). In addition to the cuprizone model, the effectiveness of progesterone and  $3\alpha,5\alpha$ -THPROG has been reported in the experimental

autoimmune encephalomyelitis (EAE) model. In this model, progesterone showed promyelinating, neuroprotective, and anti-inflammatory effects, as recently reviewed [123]. In addition, the selective progesterone agonist Nestorone improved functional outcomes, increased neurogenesis, and decreased inflammation in the brains of female EAE mice [124]. In EAE mice, 3 $\alpha$ ,5 $\alpha$ -THPROG limited axonal injury, demyelination, neuroinflammation, and functional deficits [61]. For more information regarding the effects of 3 $\alpha$ ,5 $\alpha$ -THPROG on inflammation see [125,126]. For readers interested in the role of sex and sex hormones and their neuromodulatory effects and underlying mechanisms in the context of neurodegenerative diseases, we recommend two excellent recent reviews [127,128].

#### 4.2. Neuroprotective Effects in TBI

There is an extensive literature reporting the neuroprotective effects of progesterone in experimental models of TBI. Briefly, progesterone treatment decreased edema, inflammation, BBB dysfunction, and promoted survival of newborn neurons and functional recovery (for review see [8,9,20,129–133]). Interestingly, 3 $\alpha$ ,5 $\alpha$ -THPROG was also shown to be neuroprotective in experimental TBI. In particular, it decreased cell death, cognitive deficits, mitochondrial dysfunction, and inflammation [134–137]. Recruitment of patients will soon start for a phase II clinical trial (Allopregnanolone in chronic complex traumatic injury; Clinical Trials. Gov Identifier: NCT04003285). The trial aims to evaluate the potential of 3 $\alpha$ ,5 $\alpha$ -THPROG (allopregnanolone) to improve depression, pain symptoms, and functional outcomes at several time points post-administration in patients with a mild traumatic injury.

Despite the multifactorial benefits of progesterone obtained in the experimental models of TBI and the promising results of two Phase II clinical trials [138,139], two Phase III clinical trials failed to show benefits of progesterone [140,141]. The possible reasons for this failure were recently discussed [142–144]. Among the concerns that have been raised we can cite: the high heterogeneity of the enrolled patients concerning sex, age, and severity of TBI; the high dose of progesterone used; the lack of stratification of patients; the subjective outcome measures; and the lack of follow up for a longer period. Several meta-analyses of the clinical trials are available [145–148]. Ma et al. included five randomized clinical trials. The results of the meta-analysis did not find evidence that progesterone could reduce mortality or disability. However, authors pointed to some inconsistencies and discussed potential bias that might reduce confidence in the conclusions. They recommend stratifying TBI patients and optimizing the dosage of progesterone for future trials [146]. The meta-analysis by Lu et al. (2016) included 8 clinical trials and did not reveal a significant effect of progesterone treatment in mortality or in neurological function in patients with acute TBI, and raised the same concerns [147]. The meta-analysis by Pan et al. included 8 clinical trials and showed that progesterone improved clinical outcomes in severe TBI at 3 months but not at 6 months [148].

In clinical trials, levels of progesterone in plasma of patients treated with progesterone were 100 times higher than the levels in patients treated with placebo; they reached a concentration of

1.1  $\mu$ M [140,141,149], that is 10 to 1000 times higher than the Kd of progesterone receptors (PR: Kd = 1nM; mPRs: Kd = 10 nM; PGRMC1: Kd = 100 nM) [21]. The efficient cerebroprotective dose of progesterone (8 mg/kg) used in experimental rodent models of both TBI (Guennoun et al., unpublished results) and stroke [150] results in plasmatic levels of 150 nM, which is 10 times lower than the levels reached in patients treated with progesterone in the clinical trials (1.1 $\mu$ M). In the experimental models of stroke, progesterone brain levels reached 100 nM at 2 h after the last administration of progesterone [150]; these levels are compatible with the activation of PR (Kd = 1 nM); mPRs (Kd = 10 nM) and PGRMC1 (Kd = 100 nM). Depending on its brain concentration, progesterone may differentially activate its different receptors: higher doses of progesterone may saturate nuclear receptors (PR) while still activating membrane receptors mPR and PGRMC1. Higher doses might also induce the desensitization of receptors or decrease their expression, the saturation of pathways leading to neuroactive metabolites or the induction of inactivating metabolic pathways.

In summary, progesterone could be repurposed and repositioned in the treatment of TBI patients [151]. However, more pre-clinical studies with a better optimization of the formulation,

dosing, and route of administration are needed [144]. It was also recommended to create a collaborative research network, to pool and more rigorously report preclinical studies, and to do more exploratory phase 2 trials [152]. Better definition of TBI and stratification according to sex, age, and severity, as well as development of biomarkers and better outcome measures are essential for future successful trials [133,142,143,147].

#### 4.3. Neuroprotective Effects in Stroke Models

There is strong evidence that progesterone reduces the infarct volume and improves functional recovery in experimental models of stroke (see systematic reviews [153,154]). However, the meta-analysis by Wong et al. revealed that most studies used young male animals and none used gonadally intact young adult females. Different studies provided important information. For example, progesterone treatment decreased BBB disruption [150,155], hemorrhagic transformation [156], inflammatory response [84,157–160], mitochondrial dysfunction and oxidative damage [161–165], and apoptosis [166]. Moreover, progesterone has been reported to increase neurogenesis [167] and the survival of newborn neurons [168]. For further details see review [169]. Dose–response studies showed that the optimal neuroprotective dose is 8 mg/kg [170–173]. Of note, progesterone provides neuroprotection even when administered as late as 6 h after ischemia [171,174]. The observed transient increase of endogenous cerebral levels of progesterone after stroke [90,100] may provide an early endogenous neuroprotection that may contribute to this large therapeutic window.

Studies using  $3\alpha,5\alpha$ -THPROG in ischemic stroke showed that it is efficient in reducing infarct size, BBB dysfunction, inflammation response, and behavioral deficits [155,163,175]. These findings led the authors to propose that the neuroprotective effects of progesterone may be mediated by its neuroactive metabolite  $3\alpha,5\alpha$ -THPROG. However, our study using PR knock-out mice showed that although both treatments with progesterone or  $3\alpha,5\alpha$ -THPROG are neuroprotective, PR play a key role in the mediation of the effect of progesterone [100].

### 5. Mechanism of Action of Progesterone after Stroke: A Key Role of PR

Our recent studies have revealed a key role of PR in neuroprotection after stroke. Thus, we have shown that at 6 h and 24 h post-MCAO, PR-dependent signaling of endogenous brain progesterone limited the extent of infarct size and the impairment of motor functions. Additional treatment with exogenous progesterone is required for optimal longer-term neuroprotection and is also PR-dependent [90,100,176]. Using total PR KO male mice, we first showed that PR expression is a limiting factor. Thus PR<sup>-/-</sup> and even the heterozygous PR<sup>+/-</sup> mice showed larger infarcts compared to wild type PR<sup>+/+</sup> mice at 6 and 24 h post-MCAO [100]. We also showed the importance of PR after treatment with progesterone, as progesterone decreased infarct volume and motor deficits in wild type PR<sup>+/+</sup> mice but not in PR<sup>-/-</sup> mice [100]. Our findings, also demonstrated that the bio-conversion of progesterone to  $3\alpha,5\alpha$ -THPROG is not the mechanism through which progesterone provides neuroprotection, otherwise progesterone treatment would have been neuroprotective in PR knockout (PR<sup>-/-</sup>) mice [100]. Another study confirms the importance of the level of expression of PR [177]. Indeed, progesterone decreased infarct size in wild type PR<sup>+/+</sup> mice but did not in heterozygous mice PR<sup>+/-</sup>. To further clarify the role of PR specifically expressed in brain, we then generated a new transgenic mice line PR<sup>NesCre</sup> mice in which PR expression was selectively invalidated in neural cells using the Cre-Lox strategy [90]. Invalidation of PR in neural cells resulted in increased brain tissue damage and neurological deficits in both male and female, young and aging mice at the early phase after stroke [90]. Interestingly, a greater effect was observed in young males compared to young females suggesting that females may have additional neuroprotective agents independent of PR. Using PR<sup>NesCre</sup> mice and their control littermates PR<sup>loxP/loxP</sup>, we have also shown a key role of PR after progesterone treatment [176]. Thus, progesterone treatment improved motor coordination and reduced neurological deficits and infarct size in PR<sup>loxP/loxP</sup> mice that express normal brain levels of PR but not in PR<sup>NesCre</sup>. At the cellular level, progesterone treatment increased the density of neurons,

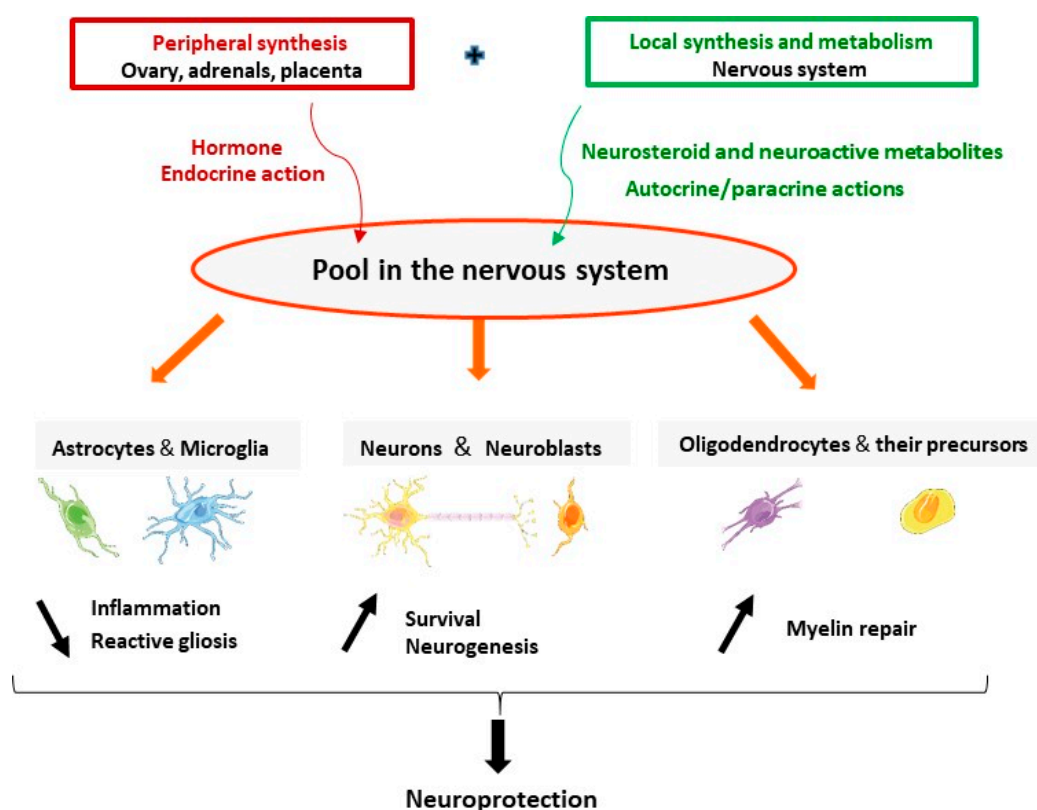
cells of the oligodendroglial lineage, and oligodendrocyte progenitors. Progesterone decreased the density of activated astrocytes and reactive microglia. These effects were observed in PR<sup>loxP/loxP</sup> mice but not in PR<sup>NesCre</sup> mice [176]. The key role of PR in neuroprotection was also shown by using Nestorone, the potent and selective PR agonist which does not interact with other receptors [178] and which is not bio-converted to GABA<sub>A</sub>-active metabolites [179]. We showed that, in male mice, Nestorone at very low dose decreased infarct volume and deficits in motor coordination [100]. Recently, a study by Tanaka et al. showed that the administration of Nestorone to male rats at 6 h post-MCAO reduced the infarct size at 48h post-MCAO and improved functional outcomes at long term (28 days post-MCAO) [180].

Although, there is strong evidence that PR is a major mediator of the neuroprotective effects of progesterone after cerebral ischemia, this does not exclude the involvement of additional mediators and cross talk with specific signaling pathways. The activation of these mechanisms may depend on the dose and timing of administration, on the time of analysis, and the outcomes measured. For example, Cai et al. showed that the activation of Src-ERK1/2 cascade via PR mediated the neuroprotective effects of progesterone observed at 48 h post-MCAO [181]. In addition, inhibiting the phosphoinositide 3-kinase/protein kinase B (PI3K/Akt) pathway, which regulates inflammation and cell survival, decreased the effects of progesterone observed on edema, infarct size, and VEGF levels at 24 h post-MCAO [166].

miRNAs (small noncoding RNAs that regulate gene expression) may also contribute to the neuroprotective effects of progesterone by acting post-transcriptionally to regulate the level of the mRNAs of the progesterone-related enzymes and receptors; progesterone may in turn regulate some miRNAs that are involved in cell death and inflammatory responses [68,182–185]. In the particular context of cerebral ischemia, a recent study in male mice showed a decrease in PGRMC1 protein level in the cortical penumbra at day 7 post-MCAO and that inhibition of let-7i miRNA alleviated the ischemia-induced decrease of PGMRC1 expression and increased the beneficial effects of progesterone [86]. Recently, Herzog et al. investigated the role of estradiol and progesterone on the regulation of selected miRNA and the genes they regulate. They showed a time-dependent increase in the levels of specific miRNAs in the peri-infarct; and that progesterone like estradiol decreased the levels of miR-223 and mi-214 and increased the level of mi-375. Furthermore, progesterone regulated the related target genes of miR-223 and miR-375 in the peri-infarct at 24 h post-MCAO. Thus, this indirect control of the genes involved in apoptosis and inflammation may contribute to the neuroprotective effects of progesterone after stroke [186].

## 6. Summary and Concluding Remarks

The brain is a site of synthesis, metabolism, and action of progesterone. Progesterone and its neuroactive metabolites have pleiotropic protective effects in neurons and glial cells including reduction of inflammation and reactive gliosis; and promoting neuroprotection and neurogenesis and myelin repair (Figure 2). Synthesis and actions of progesterone may be adjusted on demand, to respond to brain injuries and may be affected by neurodegenerative diseases. The stimulation of neurosteroidogenesis may be part of the endogenous neuroprotective processes. One therapeutic strategy is to reinforce these intrinsic natural mechanisms by pharmacological treatments. The data reviewed here give a strong hint that treatment by progesterone has a great therapeutic potential as a protective agent to reduce brain damage. The failure of the clinical trials using progesterone for TBI patients pointed to many aspects that should be considered and revised. Progress in understanding the mechanisms of action of progesterone in specific pathological conditions will help to better design future clinical trials. In the particular case of stroke, the identification of PR as a major mediator of neuroprotection indicates that PR agonists may also represent a potential efficient treatment.



**Figure 2.** The brain is a site of synthesis, metabolism, and action of progesterone. Progesterone (either synthesized in the endocrine glands or locally in the nervous system) and its neuroactive metabolites have pleiotropic neuroprotective effects. They act in astrocytes and microglia to reduce inflammation and reactive gliosis; promote survival of neurons and neurogenesis; and increase myelination by acting in oligodendrocytes and their precursors.

**Author Contributions:** R.G. designed and wrote the manuscript. The author has read and agreed to the published version of the manuscript.

**Funding:** No funding was received for the preparation of the review. The work of the team, summarized in this review, was supported by Inserm, Paris Sud/Paris Saclay University and the Mattern Foundation.

**Acknowledgments:** I would like to thank the former and present members of my team and the colleagues who contributed to the work summarized in this review.

**Conflicts of Interest:** The author declares no conflict of interest.

## Abbreviations

3 $\alpha$ -HSD	3 $\alpha$ -hydroxysteroid dehydrogenase
3 $\beta$ -HSD	3 $\beta$ -hydroxysteroid dehydrogenase
5 $\alpha$ -DHPROG	5 $\alpha$ -dihydroprogesterone
3 $\alpha$ ,5 $\alpha$ -THPROG	3 $\alpha$ ,5 $\alpha$ -tetrahydroprogesterone (allopregnanolone)
6-OHDA	6-hydroxydopamine
AD	Alzheimer's disease
BBB	Blood brain barrier
CNS	Central nervous system
CSF	Cerebrospinal fluid
EAE	Experimental autoimmune encephalomyelitis
GABA	$\gamma$ -aminobutyric acid
Il-18	Interferon-gamma inducing factor 18



MCAO/R	Middle cerebral artery occlusion/reperfusion
mPR	Membrane progesterone receptors
MS	Multiple sclerosis
NLRP3	NOD-like receptor pyrin domain containing 3
NP-C	Niemann–Pick type C
P450scc	Cholesterol side-chain cleavage
PD	Parkinson’s disease
PGRMC1	Membrane-associated progesterone receptor membrane component 1
PR	Progesterone receptors
PROG	Progesterone
TBI	Traumatic brain injury

## References

- Baulieu, E.E.; Robel, P. Neurosteroids: A new brain function? *J. Steroid Biochem. Mol. Biol.* **1990**, *37*, 395–403. [[CrossRef](#)]
- Mensah-Nyagan, A.G.; Do-Rego, J.L.; Beaujean, D.; Luu-The, V.; Pelletier, G.; Vaudry, H. Neurosteroids: Expression of steroidogenic enzymes and regulation of steroid biosynthesis in the central nervous system. *Pharmacol. Rev.* **1999**, *51*, 63–81.
- Compagnone, N.A.; Mellon, S.H. Neurosteroids: Biosynthesis and function of these novel neuromodulators. *Front. Neuroendocrinol.* **2000**, *21*, 1–56. [[CrossRef](#)] [[PubMed](#)]
- Mellon, S.H.; Griffin, L.D. Neurosteroids: Biochemistry and clinical significance. *Trends Endocrinol. Metab.* **2002**, *13*, 35–43. [[CrossRef](#)]
- Melcangi, R.C.; Garcia-Segura, L.M.; Mensah-Nyagan, A.G. Neuroactive steroids: State of the art and new perspectives. *Cell Mol. Life Sci.* **2008**, *65*, 777–797. [[CrossRef](#)] [[PubMed](#)]
- Diotel, N.; Charlier, T.D.; Lefebvre d’Hellencourt, C.; Couret, D.; Trudeau, V.L.; Nicolau, J.C.; Meilhac, O.; Kah, O.; Pellegrini, E. Steroid Transport, Local Synthesis, and Signaling within the Brain: Roles in Neurogenesis, Neuroprotection, and Sexual Behaviors. *Front. Neurosci.* **2018**, *12*, 84. [[CrossRef](#)] [[PubMed](#)]
- Mendell, A.L.; MacLusky, N.J. Neurosteroid Metabolites of Gonadal Steroid Hormones in Neuroprotection: Implications for Sex Differences in Neurodegenerative Disease. *Front. Mol. Neurosci.* **2018**, *11*, 359. [[CrossRef](#)]
- De Nicola, A.F.; Labombarda, F.; Gonzalez Deniselle, M.C.; Gonzalez, S.L.; Garay, L.; Meyer, M.; Gargiulo, G.; Guennoun, R.; Schumacher, M. Progesterone neuroprotection in traumatic CNS injury and motoneuron degeneration. *Front. Neuroendocrinol.* **2009**, *30*, 173–187. [[CrossRef](#)]
- Schumacher, M.; Mattern, C.; Ghomari, A.; Oudinet, J.P.; Liere, P.; Labombarda, F.; Sitruk-Ware, R.; De Nicola, A.F.; Guennoun, R. Revisiting the roles of progesterone and allopregnanolone in the nervous system: Resurgence of the progesterone receptors. *Prog. Neurobiol.* **2014**, *113*, 6–39. [[CrossRef](#)]
- Melcangi, R.C.; Giatti, S.; Calabrese, D.; Pesaresi, M.; Cermenati, G.; Mitro, N.; Viviani, B.; Garcia-Segura, L.M.; Caruso, D. Levels and actions of progesterone and its metabolites in the nervous system during physiological and pathological conditions. *Prog. Neurobiol.* **2014**, *113*, 56–69. [[CrossRef](#)]
- Melcangi, R.C.; Panzica, G.C. Allopregnanolone: State of the art. *Prog. Neurobiol.* **2014**, *113*, 1–5. [[CrossRef](#)]
- Patte-Mensah, C.; Meyer, L.; Taleb, O.; Mensah-Nyagan, A.G. Potential role of allopregnanolone for a safe and effective therapy of neuropathic pain. *Prog. Neurobiol.* **2014**, *113*, 70–78. [[CrossRef](#)]
- Meyer, L.; Taleb, O.; Patte-Mensah, C.; Mensah-Nyagan, A.G. Neurosteroids and neuropathic pain management: Basic evidence and therapeutic perspectives. *Front. Neuroendocrinol.* **2019**, *55*, 100795. [[CrossRef](#)]
- Pardridge, W.M.; Mietus, L.J. Transport of steroid hormones through the rat blood-brain barrier. Primary role of albumin-bound hormone. *J. Clin. Investig.* **1979**, *64*, 145–154. [[CrossRef](#)]
- Pardridge, W.M. Drug transport across the blood-brain barrier. *J. Cereb. Blood Flow Metab.* **2012**, *32*, 1959–1972. [[CrossRef](#)]
- Witt, K.A.; Sandoval, K.E. Steroids and the blood-brain barrier: Therapeutic implications. *Adv. Pharmacol.* **2014**, *71*, 361–390.
- Banks, W.A. Brain meets body: The blood-brain barrier as an endocrine interface. *Endocrinology* **2012**, *153*, 4111–4119. [[CrossRef](#)]



18. Baulieu, E.E.; Robel, P.; Schumacher, M. Neurosteroids: Beginning of the story. *Int. Rev. Neurobiol.* **2001**, *46*, 1–32.
19. Mellon, S.H.; Vaudry, H. Biosynthesis of neurosteroids and regulation of their synthesis. *Int. Rev. Neurobiol.* **2001**, *46*, 33–78.
20. Guennoun, R.; Labombarda, F.; Gonzalez Deniselle, M.C.; Liere, P.; De Nicola, A.F.; Schumacher, M. Progesterone and allopregnanolone in the central nervous system: Response to injury and implication for neuroprotection. *J. Steroid Biochem. Mol. Biol.* **2015**, *146*, 48–61. [[CrossRef](#)]
21. Schumacher, M.; Guennoun, R.; Ghomari, A.; Massaad, C.; Robert, F.; El-Etr, M.; Akwa, Y.; Rajkowski, K.; Baulieu, E.E. Novel perspectives for progesterone in hormone replacement therapy, with special reference to the nervous system. *Endocr. Rev.* **2007**, *28*, 387–439. [[CrossRef](#)] [[PubMed](#)]
22. Pelletier, G. Steroidogenic enzymes in the brain: Morphological aspects. *Prog. Brain Res.* **2010**, *181*, 193–207. [[PubMed](#)]
23. Le Goascogne, C.; Robel, P.; Gouezou, M.; Sananes, N.; Baulieu, E.E.; Waterman, M. Neurosteroids: Cytochrome P-450scc in rat brain. *Science* **1987**, *237*, 1212–1215. [[CrossRef](#)] [[PubMed](#)]
24. Tsutsui, K.; Ukena, K. Neurosteroids in the cerebellar Purkinje neuron and their actions (review). *Int. J. Mol. Med.* **1999**, *4*, 49–56. [[CrossRef](#)] [[PubMed](#)]
25. Ukena, K.; Usui, M.; Kohchi, C.; Tsutsui, K. Cytochrome P450 side-chain cleavage enzyme in the cerebellar Purkinje neuron and its neonatal change in rats. *Endocrinology* **1998**, *139*, 137–147. [[CrossRef](#)] [[PubMed](#)]
26. Kimoto, T.; Tsurugizawa, T.; Ohta, Y.; Makino, J.; Tamura, H.; Hojo, Y.; Takata, N.; Kawato, S. Neurosteroid synthesis by cytochrome p450-containing systems localized in the rat brain hippocampal neurons: N-methyl-D-aspartate and calcium-dependent synthesis. *Endocrinology* **2001**, *142*, 3578–3589. [[CrossRef](#)]
27. Mellon, S.H.; Deschepper, C.F. Neurosteroid biosynthesis: Genes for adrenal steroidogenic enzymes are expressed in the brain. *Brain Res.* **1993**, *629*, 283–292. [[CrossRef](#)]
28. Goascogne, C.; Gouezou, M.; Robel, P.; Defaye, G.; Chambaz, E.; Waterman, M.R.; Baulieu, E.E. The cholesterol side-chain cleavage complex in human brain white matter. *J. Neuroendocrinol.* **1989**, *1*, 153–156. [[CrossRef](#)]
29. Beyenburg, S.; Stoffel-Wagner, B.; Watzka, M.; Blumcke, I.; Bauer, J.; Schramm, J.; Bidlingmaier, F.; Elger, C.E. Expression of cytochrome P450scc mRNA in the hippocampus of patients with temporal lobe epilepsy. *Neuroreport* **1999**, *10*, 3067–3070. [[CrossRef](#)]
30. Watzka, M.; Bidlingmaier, F.; Schramm, J.; Klingmuller, D.; Stoffel-Wagner, B. Sex- and age-specific differences in human brain CYP11A1 mRNA expression. *J. Neuroendocrinol.* **1999**, *11*, 901–905. [[CrossRef](#)]
31. Yu, L.; Romero, D.G.; Gomez-Sanchez, C.E.; Gomez-Sanchez, E.P. Steroidogenic enzyme gene expression in the human brain. *Mol. Cell Endocrinol.* **2002**, *190*, 9–17. [[CrossRef](#)]
32. Hu, Z.Y.; Bourreau, E.; Jung-Testas, I.; Robel, P.; Baulieu, E.E. Neurosteroids: Oligodendrocyte mitochondria convert cholesterol to pregnenolone. *Proc. Natl. Acad. Sci. USA* **1987**, *84*, 8215–8219. [[CrossRef](#)] [[PubMed](#)]
33. Brown, R.C.; Cascio, C.; Papadopoulos, V. Pathways of neurosteroid biosynthesis in cell lines from human brain: Regulation of dehydroepiandrosterone formation by oxidative stress and beta-amyloid peptide. *J. Neurochem.* **2000**, *74*, 847–859. [[CrossRef](#)] [[PubMed](#)]
34. Guennoun, R.; Fiddes, R.J.; Gouezou, M.; Lombes, M.; Baulieu, E.E. A key enzyme in the biosynthesis of neurosteroids, 3 beta-hydroxysteroid dehydrogenase/delta 5-delta 4-isomerase (3 beta-HSD), is expressed in rat brain. *Mol. Brain Res.* **1995**, *30*, 287–300. [[CrossRef](#)]
35. Inoue, T.; Akahira, J.; Suzuki, T.; Darnel, A.D.; Kaneko, C.; Takahashi, K.; Hatori, M.; Shirane, R.; Kumabe, T.; Kurokawa, Y.; et al. Progesterone production and actions in the human central nervous system and neurogenic tumors. *J. Clin. Endocrinol. Metab.* **2002**, *87*, 5325–5331. [[CrossRef](#)]
36. Weidenfeld, J.; Siegel, R.A.; Chowers, I. In vitro conversion of pregnenolone to progesterone by discrete brain areas of the male rat. *J. Steroid Biochem.* **1980**, *13*, 961–963. [[CrossRef](#)]
37. Bauer, H.C.; Bauer, H. Micromethod for the determination of 3-beta-HSD activity in cultured cells. *J. Steroid Biochem.* **1989**, *33*, 643–646. [[CrossRef](#)]
38. Jung-Testas, I.; Hu, Z.Y.; Baulieu, E.E.; Robel, P. Neurosteroids: Biosynthesis of pregnenolone and progesterone in primary cultures of rat glial cells. *Endocrinology* **1989**, *125*, 2083–2091. [[CrossRef](#)]
39. Kabbadj, K.; El-Etr, M.; Baulieu, E.E.; Robel, P. Pregnenolone metabolism in rodent embryonic neurons and astrocytes. *Glia* **1993**, *7*, 170–175. [[CrossRef](#)]

40. Akwa, Y.; Sananes, N.; Guezou, M.; Robel, P.; Baulieu, E.E.; Le Goascogne, C. Astrocytes and neurosteroids: Metabolism of pregnenolone and dehydroepiandrosterone. Regulation by cell density. *J. Cell Biol.* **1993**, *121*, 135–143. [[CrossRef](#)]
41. Zwain, I.H.; Yen, S.S. Neurosteroidogenesis in astrocytes, oligodendrocytes, and neurons of cerebral cortex of rat brain. *Endocrinology* **1999**, *140*, 3843–3852. [[CrossRef](#)] [[PubMed](#)]
42. Gago, N.; Akwa, Y.; Sananes, N.; Guennoun, R.; Baulieu, E.E.; El-Etr, M.; Schumacher, M. Progesterone and the oligodendroglial lineage: Stage-dependent biosynthesis and metabolism. *Glia* **2001**, *36*, 295–308. [[CrossRef](#)]
43. Melcangi, R.C.; Poletti, A.; Cavarretta, I.; Celotti, F.; Colciago, A.; Magnaghi, V.; Motta, M.; Negri-Cesi, P.; Martini, L. The 5 $\alpha$ -reductase in the central nervous system: Expression and modes of control. *J. Steroid Biochem. Mol. Biol.* **1998**, *65*, 295–299. [[CrossRef](#)]
44. Russell, D.W.; Wilson, J.D. Steroid 5  $\alpha$ -reductase: Two genes/two enzymes. *Ann. Rev. Biochem.* **1994**, *63*, 25–61. [[CrossRef](#)] [[PubMed](#)]
45. Steckelbroeck, S.; Watzka, M.; Reichelt, R.; Hans, V.H.; Stoffel-Wagner, B.; Heidrich, D.D.; Schramm, J.; Bidlingmaier, F.; Klingmuller, D. Characterization of the 5 $\alpha$ -reductase-3 $\alpha$ -hydroxysteroid dehydrogenase complex in the human brain. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 1324–1331.
46. Stoffel-Wagner, B. Neurosteroid biosynthesis in the human brain and its clinical implications. *Ann. N. Y. Acad. Sci.* **2003**, *1007*, 64–78. [[CrossRef](#)]
47. Thigpen, A.E.; Silver, R.I.; Guileyardo, J.M.; Casey, M.L.; McConnell, J.D.; Russell, D.W. Tissue distribution and ontogeny of steroid 5  $\alpha$ -reductase isozyme expression. *J. Clin. Investig.* **1993**, *92*, 903–910. [[CrossRef](#)]
48. Torres, J.M.; Ortega, E. Differential regulation of steroid 5 $\alpha$ -reductase isozymes expression by androgens in the adult rat brain. *FASEB J.* **2003**, *17*, 1428–1433. [[CrossRef](#)]
49. Luchetti, S.; Huitinga, I.; Swaab, D.F. Neurosteroid and GABA-A receptor alterations in Alzheimer's disease, Parkinson's disease and multiple sclerosis. *Neuroscience* **2011**, *191*, 6–21. [[CrossRef](#)]
50. Giatti, S.; Diviccaro, S.; Garcia-Segura, L.M.; Melcangi, R.C. Sex differences in the brain expression of steroidogenic molecules under basal conditions and after gonadectomy. *J. Neuroendocrinol.* **2019**, *31*, e12736. [[CrossRef](#)]
51. Agis-Balboa, R.C.; Pinna, G.; Zhubi, A.; Maloku, E.; Veldic, M.; Costa, E.; Guidotti, A. Characterization of brain neurons that express enzymes mediating neurosteroid biosynthesis. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 14602–14607. [[CrossRef](#)] [[PubMed](#)]
52. Pelletier, G.; Luu-The, V.; Labrie, F. Immunocytochemical localization of 5  $\alpha$ -reductase in rat brain. *Mol. Cell Neurosci.* **1994**, *5*, 394–399. [[CrossRef](#)] [[PubMed](#)]
53. Tsuruo, Y.; Miyamoto, T.; Yokoi, H.; Kitagawa, K.; Futaki, S.; Ishimura, K. Immunohistochemical presence of 5  $\alpha$ -reductase rat type 1-containing cells in the rat brain. *Brain Res.* **1996**, *722*, 207–211. [[CrossRef](#)]
54. Celotti, F.; Melcangi, R.C.; Martini, L. The 5  $\alpha$ -reductase in the brain: Molecular aspects and relation to brain function. *Front. Neuroendocrinol.* **1992**, *13*, 163–215. [[PubMed](#)]
55. Saitoh, H.; Hirato, K.; Yanaihara, T.; Nakayama, T. A study of 5  $\alpha$ -reductase in human fetal brain. *Endocrinol. Jpn.* **1982**, *29*, 461–467. [[CrossRef](#)] [[PubMed](#)]
56. Melcangi, R.C.; Celotti, F.; Martini, L. Progesterone 5- $\alpha$ -reduction in neuronal and in different types of glial cell cultures: Type 1 and 2 astrocytes and oligodendrocytes. *Brain Res.* **1994**, *639*, 202–206. [[CrossRef](#)]
57. Melcangi, R.C.; Ballabio, M.; Magnaghi, V.; Celotti, F. Metabolism of steroids in pure cultures of neurons and glial cells: Role of intracellular signalling. *J. Steroid Biochem. Mol. Biol.* **1995**, *53*, 331–336. [[CrossRef](#)]
58. Khanna, M.; Qin, K.N.; Cheng, K.C. Distribution of 3  $\alpha$ -hydroxysteroid dehydrogenase in rat brain and molecular cloning of multiple cDNAs encoding structurally related proteins in humans. *J. Steroid Biochem. Mol. Biol.* **1995**, *53*, 41–46. [[CrossRef](#)]
59. Meffre, D.; Delespierre, B.; Guezou, M.; Schumacher, M.; Stein, D.G.; Guennoun, R. 3 $\beta$ -Hydroxysteroid dehydrogenase/5-ene-4-ene isomerase mRNA expression in rat brain: Effect of pseudopregnancy and traumatic brain injury. *J. Steroid Biochem. Mol. Biol.* **2007**, *104*, 293–300. [[CrossRef](#)]
60. Leicaj, M.L.; Pasquini, L.A.; Lima, A.; Gonzalez Deniselle, M.C.; Pasquini, J.M.; De Nicola, A.F.; Garay, L.I. Changes in neurosteroidogenesis during demyelination and remyelination in cuprizone-treated mice. *J. Neuroendocrinol.* **2018**, *30*, e12649. [[CrossRef](#)]
61. Noorbakhsh, F.; Ellestad, K.K.; Maingat, F.; Warren, K.G.; Han, M.H.; Steinman, L.; Baker, G.B.; Power, C. Impaired neurosteroid synthesis in multiple sclerosis. *Brain* **2011**, *134*, 2703–2721. [[CrossRef](#)] [[PubMed](#)]

62. Griffin, L.D.; Gong, W.; Verot, L.; Mellon, S.H. Niemann-Pick type C disease involves disrupted neurosteroidogenesis and responds to allopregnanolone. *Nat. Med.* **2004**, *10*, 704–711. [[CrossRef](#)] [[PubMed](#)]
63. Mellon, S.H.; Gong, W.; Schonemann, M.D. Endogenous and synthetic neurosteroids in treatment of Niemann-Pick Type C disease. *Brain Res. Rev.* **2008**, *57*, 410–420. [[CrossRef](#)] [[PubMed](#)]
64. Luchetti, S.; Bossers, K.; Frajese, G.V.; Swaab, D.F. Neurosteroid biosynthetic pathway changes in substantia nigra and caudate nucleus in Parkinson's disease. *Brain Pathol.* **2010**, *20*, 945–951. [[CrossRef](#)] [[PubMed](#)]
65. Luchetti, S.; Van Eden, C.G.; Schuurman, K.; van Strien, M.E.; Swaab, D.F.; Huitinga, I. Gender differences in multiple sclerosis: Induction of estrogen signaling in male and progesterone signaling in female lesions. *J. Neuropathol. Exp. Neurol.* **2014**, *73*, 123–135. [[CrossRef](#)]
66. Brinton, R.D.; Thompson, R.F.; Foy, M.R.; Baudry, M.; Wang, J.; Finch, C.E.; Morgan, T.E.; Pike, C.J.; Mack, W.J.; Stanczyk, F.Z.; et al. Progesterone receptors: Form and function in brain. *Front. Neuroendocrinol.* **2008**, *29*, 313–339. [[CrossRef](#)]
67. Singh, M.; Su, C.; Ng, S. Non-genomic mechanisms of progesterone action in the brain. *Front. Neurosci.* **2013**, *7*, 159. [[CrossRef](#)]
68. Gonzalez, S.L.; Coronel, M.F.; Raggio, M.C.; Labombarda, F. Progesterone receptor-mediated actions and the treatment of central nervous system disorders: An up-date of the known and the challenge of the unknown. *Steroids* **2020**, *153*, 108525. [[CrossRef](#)]
69. Gonzalez, S.L. Progesterone for the treatment of central nervous system disorders: The many signaling roads for a single molecule. *Neural Regen. Res.* **2020**, *15*, 1846–1847. [[CrossRef](#)]
70. Kastner, P.; Krust, A.; Turcotte, B.; Stropp, U.; Tora, L.; Gronemeyer, H.; Chambon, P. Two distinct estrogen-regulated promoters generate transcripts encoding the two functionally different human progesterone receptor forms A and B. *EMBO J.* **1990**, *9*, 1603–1614. [[CrossRef](#)]
71. Kraus, W.L.; Montano, M.M.; Katzenellenbogen, B.S. Cloning of the rat progesterone receptor gene 5'-region and identification of two functionally distinct promoters. *Mol. Endocrinol.* **1993**, *7*, 1603–1616.
72. Hagan, C.R.; Daniel, A.R.; Dressing, G.E.; Lange, C.A. Role of phosphorylation in progesterone receptor signaling and specificity. *Mol. Cell Endocrinol.* **2012**, *357*, 43–49. [[CrossRef](#)] [[PubMed](#)]
73. Guennoun, R.; Meffre, D.; Labombarda, F.; Gonzalez, S.L.; Gonzalez Deniselle, M.C.; Stein, D.G.; De Nicola, A.F.; Schumacher, M. The membrane-associated progesterone-binding protein 25-Dx: Expression, cellular localization and up-regulation after brain and spinal cord injuries. *Brain Res. Rev.* **2008**, *57*, 493–505. [[CrossRef](#)] [[PubMed](#)]
74. Losel, R.M.; Besong, D.; Peluso, J.J.; Wehling, M. Progesterone receptor membrane component 1—many tasks for a versatile protein. *Steroids* **2008**, *73*, 929–934. [[CrossRef](#)] [[PubMed](#)]
75. Rohe, H.J.; Ahmed, I.S.; Twist, K.E.; Craven, R.J. PGRMC1 (progesterone receptor membrane component 1): A targetable protein with multiple functions in steroid signaling, P450 activation and drug binding. *Pharmacol. Ther.* **2009**, *121*, 14–19. [[CrossRef](#)]
76. Thomas, P.; Pang, Y.; Dong, J. Enhancement of cell surface expression and receptor functions of membrane progesterone receptor alpha (mPRalpha) by progesterone receptor membrane component 1 (PGRMC1): Evidence for a role of PGRMC1 as an adaptor protein for steroid receptors. *Endocrinology* **2014**, *155*, 1107–1119. [[CrossRef](#)] [[PubMed](#)]
77. Zhu, Y.; Bond, J.; Thomas, P. Identification, classification, and partial characterization of genes in humans and other vertebrates homologous to a fish membrane progesterone receptor. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 2237–2242. [[CrossRef](#)]
78. Zhu, Y.; Rice, C.D.; Pang, Y.; Pace, M.; Thomas, P. Cloning, expression, and characterization of a membrane progesterone receptor and evidence it is an intermediary in meiotic maturation of fish oocytes. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 2231–2236. [[CrossRef](#)]
79. Pang, Y.; Dong, J.; Thomas, P. Characterization, neurosteroid binding and brain distribution of human membrane progesterone receptors delta and {epsilon} (mPRdelta and mPR{epsilon}) and mPRdelta involvement in neurosteroid inhibition of apoptosis. *Endocrinology* **2013**, *154*, 283–295. [[CrossRef](#)]
80. Hosie, A.M.; Wilkins, M.E.; da Silva, H.M.; Smart, T.G. Endogenous neurosteroids regulate GABAA receptors through two discrete transmembrane sites. *Nature* **2006**, *444*, 486–489. [[CrossRef](#)]
81. Meffre, D.; Delespierre, B.; Gouezou, M.; Leclerc, P.; Vinson, G.P.; Schumacher, M.; Stein, D.G.; Guennoun, R. The membrane-associated progesterone-binding protein 25-Dx is expressed in brain regions involved in

- water homeostasis and is up-regulated after traumatic brain injury. *J. Neurochem.* **2005**, *93*, 1314–1326. [[CrossRef](#)] [[PubMed](#)]
82. Meffre, D.; Labombarda, F.; Delespierre, B.; Chastre, A.; De Nicola, A.F.; Stein, D.G.; Schumacher, M.; Guennoun, R. Distribution of membrane progesterone receptor alpha in the male mouse and rat brain and its regulation after traumatic brain injury. *Neuroscience* **2013**, *231*, 111–124. [[CrossRef](#)] [[PubMed](#)]
83. Dang, J.; Mitkari, B.; Kipp, M.; Beyer, C. Gonadal steroids prevent cell damage and stimulate behavioral recovery after transient middle cerebral artery occlusion in male and female rats. *Brain Behav. Immun.* **2011**, *25*, 715–726. [[CrossRef](#)] [[PubMed](#)]
84. Allen, R.S.; Sayeed, I.; Oumarbaeva, Y.; Morrison, K.C.; Choi, P.H.; Pardue, M.T.; Stein, D.G. Progesterone treatment shows greater protection in brain vs. retina in a rat model of middle cerebral artery occlusion: Progesterone receptor levels may play an important role. *Restor. Neurol. Neurosci.* **2016**, *34*, 947–963. [[CrossRef](#)] [[PubMed](#)]
85. Stanojlovic, M.; Gusevac Stojanovic, I.; Zaric, M.; Martinovic, J.; Mitrovic, N.; Grkovic, I.; Drakulic, D. Progesterone Protects Prefrontal Cortex in Rat Model of Permanent Bilateral Common Carotid Occlusion via Progesterone Receptors and Akt/Erk/eNOS. *Cell Mol. Neurobiol.* **2019**, *40*, 829–843. [[CrossRef](#)] [[PubMed](#)]
86. Nguyen, T.; Su, C.; Singh, M. Let-7i inhibition enhances progesterone-induced functional recovery in a mouse model of ischemia. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E9668–E9677. [[CrossRef](#)]
87. Gaignard, P.; Savouroux, S.; Liere, P.; Pianos, A.; Therond, P.; Schumacher, M.; Slama, A.; Guennoun, R. Effect of Sex Differences on Brain Mitochondrial Function and Its Suppression by Ovariectomy and in Aged Mice. *Endocrinology* **2015**, *156*, 2893–2904. [[CrossRef](#)]
88. Lopez-Rodriguez, A.B.; Acaz-Fonseca, E.; Giatti, S.; Caruso, D.; Viveros, M.P.; Melcangi, R.C.; Garcia-Segura, L.M. Correlation of brain levels of progesterone and dehydroepiandrosterone with neurological recovery after traumatic brain injury in female mice. *Psychoneuroendocrinology* **2015**, *56*, 1–11. [[CrossRef](#)]
89. Lopez-Rodriguez, A.B.; Acaz-Fonseca, E.; Spezzano, R.; Giatti, S.; Caruso, D.; Viveros, M.P.; Melcangi, R.C.; Garcia-Segura, L.M. Profiling Neuroactive Steroid Levels After Traumatic Brain Injury in Male Mice. *Endocrinology* **2016**, *157*, 3983–3993. [[CrossRef](#)]
90. Zhu, X.; Frechou, M.; Liere, P.; Zhang, S.; Pianos, A.; Fernandez, N.; Denier, C.; Mattern, C.; Schumacher, M.; Guennoun, R. A Role of Endogenous Progesterone in Stroke Cerebroprotection Revealed by the Neural-Specific Deletion of Its Intracellular Receptors. *J. Neurosci.* **2017**, *37*, 10998–11020. [[CrossRef](#)]
91. Meffre, D.; Pianos, A.; Liere, P.; Eychenne, B.; Cambourg, A.; Schumacher, M.; Stein, D.G.; Guennoun, R. Steroid profiling in brain and plasma of male and pseudopregnant female rats after traumatic brain injury: Analysis by gas chromatography/mass spectrometry. *Endocrinology* **2007**, *148*, 2505–2517. [[CrossRef](#)] [[PubMed](#)]
92. Caruso, D.; Pesaresi, M.; Abbiati, F.; Calabrese, D.; Giatti, S.; Garcia-Segura, L.M.; Melcangi, R.C. Comparison of plasma and cerebrospinal fluid levels of neuroactive steroids with their brain, spinal cord and peripheral nerve levels in male and female rats. *Psychoneuroendocrinology* **2013**, *38*, 2278–2290. [[CrossRef](#)] [[PubMed](#)]
93. Bixo, M.; Andersson, A.; Winblad, B.; Purdy, R.H.; Backstrom, T. Progesterone, 5alpha-pregnane-3,20-dione and 3alpha-hydroxy-5alpha-pregnane-20-one in specific regions of the human female brain in different endocrine states. *Brain Res.* **1997**, *764*, 173–178. [[CrossRef](#)]
94. Caruso, D.; Barron, A.M.; Brown, M.A.; Abbiati, F.; Carrero, P.; Pike, C.J.; Garcia-Segura, L.M.; Melcangi, R.C. Age-related changes in neuroactive steroid levels in 3xTg-AD mice. *Neurobiol. Aging* **2013**, *34*, 1080–1089. [[CrossRef](#)] [[PubMed](#)]
95. Melcangi, R.C.; Caruso, D.; Levandis, G.; Abbiati, F.; Armentero, M.T.; Blandini, F. Modifications of neuroactive steroid levels in an experimental model of nigrostriatal degeneration: Potential relevance to the pathophysiology of Parkinson's disease. *J. Mol. Neurosci.* **2012**, *46*, 177–183. [[CrossRef](#)] [[PubMed](#)]
96. Di Michele, F.; Longone, P.; Romeo, E.; Lucchetti, S.; Brusa, L.; Pierantozzi, M.; Bassi, A.; Bernardi, G.; Stanzione, P. Decreased plasma and cerebrospinal fluid content of neuroactive steroids in Parkinson's disease. *Neurol. Sci.* **2003**, *24*, 172–173. [[CrossRef](#)]
97. Giatti, S.; D'Intino, G.; Maschi, O.; Pesaresi, M.; Garcia-Segura, L.M.; Calza, L.; Caruso, D.; Melcangi, R.C. Acute experimental autoimmune encephalomyelitis induces sex dimorphic changes in neuroactive steroid levels. *Neurochem. Int.* **2010**, *56*, 118–127. [[CrossRef](#)]



98. Caruso, D.; D'Intino, G.; Giatti, S.; Maschi, O.; Pesaresi, M.; Calabrese, D.; Garcia-Segura, L.M.; Calza, L.; Melcangi, R.C. Sex-dimorphic changes in neuroactive steroid levels after chronic experimental autoimmune encephalomyelitis. *J. Neurochem.* **2010**, *114*, 921–932. [[CrossRef](#)]
99. Wagner, A.K.; McCullough, E.H.; Niyonkuru, C.; Ozawa, H.; Loucks, T.L.; Dobos, J.A.; Brett, C.A.; Santarsieri, M.; Dixon, C.E.; Berga, S.L.; et al. Acute serum hormone levels: Characterization and prognosis after severe traumatic brain injury. *J. Neurotrauma* **2011**, *28*, 871–888. [[CrossRef](#)]
100. Liu, A.; Margai, I.; Zhang, S.; Labombarda, F.; Coqueran, B.; Delespierre, B.; Liere, P.; Marchand-Leroux, C.; O'Malley, B.W.; Lydon, J.P.; et al. Progesterone receptors: A key for neuroprotection in experimental stroke. *Endocrinology* **2012**, *153*, 3747–3757. [[CrossRef](#)]
101. Mani, S.K.; Oyola, M.G. Progesterone signaling mechanisms in brain and behavior. *Front. Endocrinol.* **2012**, *3*, 7. [[CrossRef](#)] [[PubMed](#)]
102. Graham, J.D.; Clarke, C.L. Physiological action of progesterone in target tissues. *Endocr. Rev.* **1997**, *18*, 502–519. [[PubMed](#)]
103. Tsutsui, K. Progesterone biosynthesis and action in the developing neuron. *Endocrinology* **2008**, *149*, 2757–2761. [[CrossRef](#)]
104. Gonzalez-Orozco, J.C.; Camacho-Arroyo, I. Progesterone Actions During Central Nervous System Development. *Front. Neurosci.* **2019**, *13*, 503. [[CrossRef](#)] [[PubMed](#)]
105. Schumacher, M.; Hussain, R.; Gago, N.; Oudinet, J.P.; Mattern, C.; Ghomari, A.M. Progesterone synthesis in the nervous system: Implications for myelination and myelin repair. *Front. Neurosci.* **2012**, *6*, 10. [[CrossRef](#)]
106. Carroll, J.C.; Rosario, E.R.; Chang, L.; Stanczyk, F.Z.; Oddo, S.; LaFerla, F.M.; Pike, C.J. Progesterone and estrogen regulate Alzheimer-like neuropathology in female 3xTg-AD mice. *J. Neurosci.* **2007**, *27*, 13357–13365. [[CrossRef](#)]
107. Chen, S.; Wang, J.M.; Irwin, R.W.; Yao, J.; Liu, L.; Brinton, R.D. Allopregnanolone promotes regeneration and reduces beta-amyloid burden in a preclinical model of Alzheimer's disease. *PLoS ONE* **2011**, *6*, e24293.
108. Irwin, R.W.; Wang, J.M.; Chen, S.; Brinton, R.D. Neuroregenerative mechanisms of allopregnanolone in Alzheimer's disease. *Front. Endocrinol.* **2011**, *2*, 117. [[CrossRef](#)]
109. Irwin, R.W.; Brinton, R.D. Allopregnanolone as regenerative therapeutic for Alzheimer's disease: Translational development and clinical promise. *Prog. Neurobiol.* **2014**, *113*, 40–55. [[CrossRef](#)]
110. Wang, T.; Yao, J.; Chen, S.; Mao, Z.; Brinton, R.D. Allopregnanolone Reverses Bioenergetic Deficits in Female Triple Transgenic Alzheimer's Mouse Model. *Neurotherapeutics* **2020**, *17*, 178–188. [[CrossRef](#)]
111. Yu, L.; Kuo, Y.; Cherng, C.G.; Chen, H.H.; Hsu, C.H. Ovarian hormones do not attenuate methamphetamine-induced dopaminergic neurotoxicity in mice gonadectomized at 4 weeks postpartum. *Neuroendocrinology* **2002**, *75*, 282–287. [[CrossRef](#)] [[PubMed](#)]
112. Sanchez, M.G.; Bourque, M.; Morissette, M.; Di Paolo, T. Steroids-dopamine interactions in the pathophysiology and treatment of CNS disorders. *CNS Neurosci. Ther.* **2010**, *16*, e43–e71. [[CrossRef](#)] [[PubMed](#)]
113. Casas, S.; Garcia, S.; Cabrera, R.; Nanfaro, F.; Escudero, C.; Yunes, R. Progesterone prevents depression-like behavior in a model of Parkinson's disease induced by 6-hydroxydopamine in male rats. *Pharmacol. Biochem. Behav.* **2011**, *99*, 614–618. [[CrossRef](#)] [[PubMed](#)]
114. Litim, N.; Morissette, M.; Di Paolo, T. Effects of progesterone administered after MPTP on dopaminergic neurons of male mice. *Neuropharmacology* **2017**, *117*, 209–218. [[CrossRef](#)] [[PubMed](#)]
115. Bourque, M.; Dluzen, D.E.; Di Paolo, T. Neuroprotective actions of sex steroids in Parkinson's disease. *Front. Neuroendocrinol.* **2009**, *30*, 142–157. [[CrossRef](#)] [[PubMed](#)]
116. Bourque, M.; Morissette, M.; Al Sweidi, S.; Caruso, D.; Melcangi, R.C.; Di Paolo, T. Neuroprotective Effect of Progesterone in MPTP-Treated Male Mice. *Neuroendocrinology* **2016**, *103*, 300–314. [[CrossRef](#)]
117. Bourque, M.; Morissette, M.; Di Paolo, T. Repurposing sex steroids and related drugs as potential treatment for Parkinson's disease. *Neuropharmacology* **2019**, *147*, 37–54. [[CrossRef](#)]
118. Adeosun, S.O.; Hou, X.; Jiao, Y.; Zheng, B.; Henry, S.; Hill, R.; He, Z.; Pani, A.; Kyle, P.; Ou, X.; et al. Allopregnanolone reinstates tyrosine hydroxylase immunoreactive neurons and motor performance in an MPTP-lesioned mouse model of Parkinson's disease. *PLoS ONE* **2012**, *7*, e50040. [[CrossRef](#)]
119. Nezhadi, A.; Sheibani, V.; Esmaeilpour, K.; Shabani, M.; Esmaeili-Mahani, S. Neurosteroid allopregnanolone attenuates cognitive dysfunctions in 6-OHDA-induced rat model of Parkinson's disease. *Behav. Brain Res.* **2016**, *305*, 258–264. [[CrossRef](#)]

120. Ye, J.N.; Chen, X.S.; Su, L.; Liu, Y.L.; Cai, Q.Y.; Zhan, X.L.; Xu, Y.; Zhao, S.F.; Yao, Z.X. Progesterone alleviates neural behavioral deficits and demyelination with reduced degeneration of oligodendroglial cells in cuprizone-induced mice. *PLoS ONE* **2013**, *8*, e54590. [[CrossRef](#)]
121. El-Etr, M.; Rame, M.; Boucher, C.; Ghomari, A.M.; Kumar, N.; Liere, P.; Pianos, A.; Schumacher, M.; Sitruk-Ware, R. Progesterone and nestorone promote myelin regeneration in chronic demyelinating lesions of corpus callosum and cerebral cortex. *Glia* **2015**, *63*, 104–117. [[CrossRef](#)] [[PubMed](#)]
122. Aryanpour, R.; Pasbakhsh, P.; Zibara, K.; Namjoo, Z.; Beigi Boroujeni, F.; Shahbeigi, S.; Kashani, I.R.; Beyer, C.; Zendehdel, A. Progesterone therapy induces an M1 to M2 switch in microglia phenotype and suppresses NLRP3 inflammasome in a cuprizone-induced demyelination mouse model. *Int. Immunopharmacol.* **2017**, *51*, 131–139. [[CrossRef](#)]
123. De Nicola, A.F.; Garay, L.I.; Meyer, M.; Guennoun, R.; Sitruk-Ware, R.; Schumacher, M.; Gonzalez Deniselle, M.C. Neurosteroidogenesis and progesterone anti-inflammatory/neuroprotective effects. *J. Neuroendocrinol.* **2018**, *30*, e12502. [[CrossRef](#)]
124. Garay, L.; Gonzalez Deniselle, M.C.; Sitruk-Ware, R.; Guennoun, R.; Schumacher, M.; De Nicola, A.F. Efficacy of the selective progesterone receptor agonist Nestorone for chronic experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* **2014**, *276*, 89–97. [[CrossRef](#)] [[PubMed](#)]
125. Noorbakhsh, F.; Baker, G.B.; Power, C. Allopregnanolone and neuroinflammation: A focus on multiple sclerosis. *Front. Cell Neurosci.* **2014**, *8*, 134. [[CrossRef](#)]
126. Giatti, S.; Boraso, M.; Melcangi, R.C.; Viviani, B. Neuroactive steroids, their metabolites, and neuroinflammation. *J. Mol. Endocrinol.* **2012**, *49*, R125–R134. [[CrossRef](#)]
127. Vegeto, E.; Villa, A.; Della Torre, S.; Crippa, V.; Rusmini, P.; Cristofani, R.; Galbiati, M.; Maggi, A.; Poletti, A. The Role of Sex and Sex Hormones in Neurodegenerative Diseases. *Endocr. Rev.* **2020**, *41*, 273–319. [[CrossRef](#)]
128. Yilmaz, C.; Karali, K.; Fodelianaki, G.; Gravanis, A.; Chavakis, T.; Charalampopoulos, I.; Alexaki, V.I. Neurosteroids as regulators of neuroinflammation. *Front. Neuroendocrinol.* **2019**, *55*, 100788. [[CrossRef](#)]
129. Gibson, C.L.; Gray, L.J.; Bath, P.M.; Murphy, S.P. Progesterone for the treatment of experimental brain injury; a systematic review. *Brain* **2008**, *131*, 318–328. [[CrossRef](#)]
130. Sayeed, I.; Stein, D.G. Progesterone as a neuroprotective factor in traumatic and ischemic brain injury. *Prog. Brain Res.* **2009**, *175*, 219–237.
131. Stein, D.G. Is progesterone a worthy candidate as a novel therapy for traumatic brain injury? *Dialogues Clin. Neurosci.* **2011**, *13*, 352–359. [[PubMed](#)]
132. Brotfain, E.; Gruenbaum, S.E.; Boyko, M.; Kutz, R.; Zlotnik, A.; Klein, M. Neuroprotection by Estrogen and Progesterone in Traumatic Brain Injury and Spinal Cord Injury. *Curr. Neuropharmacol.* **2016**, *14*, 641–653. [[CrossRef](#)] [[PubMed](#)]
133. Spani, C.B.; Braun, D.J.; Van Eldik, L.J. Sex-related responses after traumatic brain injury: Considerations for preclinical modeling. *Front. Neuroendocrinol.* **2018**, *50*, 52–66. [[CrossRef](#)] [[PubMed](#)]
134. Djebaili, M.; Hoffman, S.W.; Stein, D.G. Allopregnanolone and progesterone decrease cell death and cognitive deficits after a contusion of the rat pre-frontal cortex. *Neuroscience* **2004**, *123*, 349–359. [[CrossRef](#)]
135. Djebaili, M.; Guo, Q.; Pettus, E.H.; Hoffman, S.W.; Stein, D.G. The neurosteroids progesterone and allopregnanolone reduce cell death, gliosis, and functional deficits after traumatic brain injury in rats. *J. Neurotrauma* **2005**, *22*, 106–118. [[CrossRef](#)]
136. He, J.; Hoffman, S.W.; Stein, D.G. Allopregnanolone, a progesterone metabolite, enhances behavioral recovery and decreases neuronal loss after traumatic brain injury. *Restor. Neurol. Neurosci.* **2004**, *22*, 19–31.
137. Van Lindingham, J.W.; Cekic, M.; Cutler, S.; Hoffman, S.W.; Stein, D.G. Neurosteroids reduce inflammation after TBI through CD55 induction. *Neurosci. Lett.* **2007**, *425*, 94–98. [[CrossRef](#)]
138. Wright, D.W.; Kellermann, A.L.; Hertzberg, V.S.; Clark, P.L.; Frankel, M.; Goldstein, F.C.; Salomone, J.P.; Dent, L.L.; Harris, O.A.; Ander, D.S.; et al. ProTECT: A randomized clinical trial of progesterone for acute traumatic brain injury. *Ann. Emerg. Med.* **2007**, *49*, 391–402. [[CrossRef](#)]
139. Xiao, G.; Wei, J.; Yan, W.; Wang, W.; Lu, Z. Improved outcomes from the administration of progesterone for patients with acute severe traumatic brain injury: A randomized controlled trial. *Crit. Care* **2008**, *12*, R61. [[CrossRef](#)]
140. Wright, D.W.; Yeatts, S.D.; Silbergleit, R.; Palesch, Y.Y.; Hertzberg, V.S.; Frankel, M.; Goldstein, F.C.; Caveney, A.F.; Howlett-Smith, H.; Bengelink, E.M.; et al. Very early administration of progesterone for acute traumatic brain injury. *N. Engl. J. Med.* **2014**, *371*, 2457–2466. [[CrossRef](#)]



141. Skolnick, B.E.; Maas, A.I.; Narayan, R.K.; van der Hoop, R.G.; MacAllister, T.; Ward, J.D.; Nelson, N.R.; Stocchetti, N.; Investigators, S.T. A clinical trial of progesterone for severe traumatic brain injury. *N. Engl. J. Med.* **2014**, *371*, 2467–2476. [[CrossRef](#)] [[PubMed](#)]
142. Stein, D.G. Embracing failure: What the Phase III progesterone studies can teach about TBI clinical trials. *Brain Inj.* **2015**, *29*, 1259–1272. [[CrossRef](#)] [[PubMed](#)]
143. Schumacher, M.; Denier, C.; Oudinet, J.P.; Adams, D.; Guennoun, R. Progesterone neuroprotection: The background of clinical trial failure. *J. Steroid Biochem. Mol. Biol.* **2016**, *160*, 53–66. [[CrossRef](#)] [[PubMed](#)]
144. Guennoun, R.; Frechou, M.; Gaignard, P.; Liere, P.; Slama, A.; Schumacher, M.; Denier, C.; Mattern, C. Intranasal administration of progesterone: A potential efficient route of delivery for cerebroprotection after acute brain injuries. *Neuropharmacology* **2019**, *145*, 283–291. [[CrossRef](#)]
145. Lin, C.; He, H.; Li, Z.; Liu, Y.; Chao, H.; Ji, J.; Liu, N. Efficacy of progesterone for moderate to severe traumatic brain injury: A meta-analysis of randomized clinical trials. *Sci. Rep.* **2015**, *5*, 13442. [[CrossRef](#)]
146. Ma, J.; Huang, S.; Qin, S.; You, C.; Zeng, Y. Progesterone for acute traumatic brain injury. *Cochrane Database Syst. Rev.* **2016**, *12*, CD008409. [[CrossRef](#)]
147. Lu, X.Y.; Sun, H.; Li, Q.Y.; Lu, P.S. Progesterone for Traumatic Brain Injury: A Meta-Analysis Review of Randomized Controlled Trials. *World Neurosurg.* **2016**, *90*, 199–210. [[CrossRef](#)]
148. Pan, Z.Y.; Zhao, Y.H.; Huang, W.H.; Xiao, Z.Z.; Li, Z.Q. Effect of progesterone administration on the prognosis of patients with severe traumatic brain injury: A meta-analysis of randomized clinical trials. *Drug Des. Dev. Ther.* **2019**, *13*, 265–273. [[CrossRef](#)]
149. Wright, D.W.; Ritchie, J.C.; Mullins, R.E.; Kellermann, A.L.; Denson, D.D. Steady-state serum concentrations of progesterone following continuous intravenous infusion in patients with acute moderate to severe traumatic brain injury. *J. Clin. Pharmacol.* **2005**, *45*, 640–648. [[CrossRef](#)]
150. Frechou, M.; Zhang, S.; Liere, P.; Deslespierre, B.; Soyed, N.; Pianos, A.; Schumacher, M.; Mattern, C.; Guennoun, R. Intranasal delivery of progesterone after transient ischemic stroke decreases mortality and provides neuroprotection. *Neuropharmacology* **2015**, *97*, 394–403. [[CrossRef](#)]
151. Stein, D.G.; Sayeed, I. Repurposing and repositioning neurosteroids in the treatment of traumatic brain injury: A report from the trenches. *Neuropharmacology* **2019**, *147*, 66–73. [[CrossRef](#)] [[PubMed](#)]
152. Schwamm, L.H. Progesterone for traumatic brain injury-resisting the sirens' song. *N. Engl. J. Med.* **2014**, *371*, 2522–2523. [[CrossRef](#)] [[PubMed](#)]
153. Gibson, C.L.; Coomber, B.; Rathbone, J. Is progesterone a candidate neuroprotective factor for treatment following ischemic stroke? *Neuroscientist* **2009**, *15*, 324–332. [[CrossRef](#)]
154. Wong, R.; Renton, C.; Gibson, C.L.; Murphy, S.J.; Kendall, D.A.; Bath, P.M. Progesterone Pre-Clinical Stroke Pooling Project, C., Progesterone treatment for experimental stroke: An individual animal meta-analysis. *J. Cereb. Blood Flow Metab.* **2013**, *33*, 1362–1372. [[CrossRef](#)]
155. Ishrat, T.; Sayeed, I.; Atif, F.; Hua, F.; Stein, D.G. Progesterone and allopregnanolone attenuate blood-brain barrier dysfunction following permanent focal ischemia by regulating the expression of matrix metalloproteinases. *Exp. Neurol.* **2010**, *226*, 183–190. [[CrossRef](#)]
156. Won, S.; Lee, J.H.; Wali, B.; Stein, D.G.; Sayeed, I. Progesterone attenuates hemorrhagic transformation after delayed tPA treatment in an experimental model of stroke in rats: Involvement of the VEGF-MMP pathway. *J. Cereb. Blood Flow Metab.* **2014**, *34*, 72–80. [[CrossRef](#)]
157. Grossman, K.J.; Goss, C.W.; Stein, D.G. Effects of progesterone on the inflammatory response to brain injury in the rat. *Brain Res.* **2004**, *1008*, 29–39. [[CrossRef](#)]
158. Won, S.; Lee, J.K.; Stein, D.G. Recombinant tissue plasminogen activator promotes, and progesterone attenuates, microglia/macrophage M1 polarization and recruitment of microglia after MCAO stroke in rats. *Brain Behav. Immun.* **2015**, *49*, 267–279. [[CrossRef](#)]
159. Lammerding, L.; Slowik, A.; Johann, S.; Beyer, C.; Zendedel, A. Poststroke Inflammation Expression and Regulation in the Peri-Infarct Area by Gonadal Steroids after Transient Focal Ischemia in the Rat Brain. *Neuroendocrinology* **2016**, *103*, 460–475. [[CrossRef](#)]
160. Yousuf, S.; Atif, F.; Sayeed, I.; Wang, J.; Stein, D.G. Neuroprotection by progesterone after transient cerebral ischemia in stroke-prone spontaneously hypertensive rats. *Horm. Behav.* **2016**, *84*, 29–40. [[CrossRef](#)]

161. Aggarwal, R.; Medhi, B.; Pathak, A.; Dhawan, V.; Chakrabarti, A. Neuroprotective effect of progesterone on acute phase changes induced by partial global cerebral ischaemia in mice. *J. Pharm. Pharmacol.* **2008**, *60*, 731–737. [[CrossRef](#)] [[PubMed](#)]
162. Ozacmak, V.H.; Sayan, H. The effects of 17beta estradiol, 17alpha estradiol and progesterone on oxidative stress biomarkers in ovariectomized female rat brain subjected to global cerebral ischemia. *Physiol. Res.* **2009**, *58*, 909–912. [[PubMed](#)]
163. Sayeed, I.; Parvez, S.; Wali, B.; Siemen, D.; Stein, D.G. Direct inhibition of the mitochondrial permeability transition pore: A possible mechanism for better neuroprotective effects of allopregnanolone over progesterone. *Brain Res.* **2009**, *1263*, 165–173. [[CrossRef](#)] [[PubMed](#)]
164. Gaignard, P.; Frechou, M.; Schumacher, M.; Therond, P.; Mattern, C.; Slama, A.; Guennoun, R. Progesterone reduces brain mitochondrial dysfunction after transient focal ischemia in male and female mice. *J. Cereb. Blood Flow Metab.* **2016**, *36*, 562–568. [[CrossRef](#)] [[PubMed](#)]
165. Andrabi, S.S.; Parvez, S.; Tabassum, H. Progesterone induces neuroprotection following reperfusion-promoted mitochondrial dysfunction after focal cerebral ischemia in rats. *Dis. Models Mech.* **2017**, *10*, 787–796. [[CrossRef](#)]
166. Ishrat, T.; Sayeed, I.; Atif, F.; Hua, F.; Stein, D.G. Progesterone is neuroprotective against ischemic brain injury through its effects on the phosphoinositide 3-kinase/protein kinase B signaling pathway. *Neuroscience* **2012**, *210*, 442–450. [[CrossRef](#)]
167. Jiang, C.; Zuo, F.; Wang, Y.; Lu, H.; Yang, Q.; Wang, J. Progesterone Changes VEGF and BDNF Expression and Promotes Neurogenesis After Ischemic Stroke. *Mol. Neurobiol.* **2016**, *54*, 571–581. [[CrossRef](#)]
168. Zhang, Z.; Yang, R.; Cai, W.; Bai, Y.; Sokabe, M.; Chen, L. Treatment with progesterone after focal cerebral ischemia suppresses proliferation of progenitor cells but enhances survival of newborn neurons in adult male mice. *Neuropharmacology* **2010**, *58*, 930–939. [[CrossRef](#)]
169. Guennoun, R.; Zhu, X.; Frechou, M.; Gaignard, P.; Slama, A.; Liere, P.; Schumacher, M. Steroids in Stroke with Special Reference to Progesterone. *Cell Mol. Neurobiol.* **2019**, *39*, 551–568. [[CrossRef](#)]
170. Chen, J.; Chopp, M.; Li, Y. Neuroprotective effects of progesterone after transient middle cerebral artery occlusion in rat. *J. Neurol. Sci.* **1999**, *171*, 24–30. [[CrossRef](#)]
171. Wali, B.; Ishrat, T.; Won, S.; Stein, D.G.; Sayeed, I. Progesterone in experimental permanent stroke: A dose-response and therapeutic time-window study. *Brain* **2014**, *137*, 486–502. [[CrossRef](#)]
172. Yousuf, S.; Atif, F.; Sayeed, I.; Tang, H.; Stein, D.G. Progesterone in transient ischemic stroke: A dose-response study. *Psychopharmacology* **2014**, *231*, 3313–3323. [[CrossRef](#)] [[PubMed](#)]
173. Frechou, M.; Zhu, X.; Liere, P.; Pianos, A.; Schumacher, M.; Mattern, C.; Guennoun, R. Dose-dependent and long-term cerebroprotective effects of intranasal delivery of progesterone after ischemic stroke in male mice. *Neuropharmacology* **2020**, *170*, 108038. [[CrossRef](#)]
174. Yousuf, S.; Sayeed, I.; Atif, F.; Tang, H.; Wang, J.; Stein, D.G. Delayed progesterone treatment reduces brain infarction and improves functional outcomes after ischemic stroke: A time-window study in middle-aged rats. *J. Cereb. Blood Flow Metab.* **2014**, *34*, 297–306. [[CrossRef](#)]
175. Sayeed, I.; Guo, Q.; Hoffman, S.W.; Stein, D.G. Allopregnanolone, a progesterone metabolite, is more effective than progesterone in reducing cortical infarct volume after transient middle cerebral artery occlusion. *Ann. Emerg. Med.* **2006**, *47*, 381–389. [[CrossRef](#)]
176. Zhu, X.; Frechou, M.; Schumacher, M.; Guennoun, R. Cerebroprotection by progesterone following ischemic stroke: Multiple effects and role of the neural progesterone receptors. *J. Steroid Biochem. Mol. Biol.* **2019**, *185*, 90–102. [[CrossRef](#)] [[PubMed](#)]
177. Lee, R.J.; Kim, J.K.; Chao, D.; Kuo, L.; Mally, A.; McClean, M.E.; Pemberton, H.E.; Wilmington, A.R.; Wong, J.; Murphy, S.P. Progesterone and allopregnanolone improves stroke outcome in male mice via distinct mechanisms but neither promotes neurogenesis. *J. Neurochem.* **2015**, *132*, 32–37. [[CrossRef](#)] [[PubMed](#)]
178. Kumar, N.; Koide, S.S.; Tsong, Y.; Sundaram, K. Nestorone: A progestin with a unique pharmacological profile. *Steroids* **2000**, *65*, 629–636. [[CrossRef](#)]
179. Kumar, N.; Fagart, J.; Liere, P.; Mitchell, S.J.; Knibb, A.R.; Petit-Topin, I.; Rame, M.; El-Etr, M.; Schumacher, M.; Lambert, J.J.; et al. Nestorone(R) as a Novel Progestin for Nonoral Contraception: Structure-Activity Relationships and Brain Metabolism Studies. *Endocrinology* **2017**, *158*, 170–182. [[CrossRef](#)]
180. Tanaka, M.; Ogaeri, T.; Samsonov, M.; Sokabe, M. Nestorone exerts long-term neuroprotective effects against transient focal cerebral ischemia in adult male rats. *Brain Res.* **2019**, *1719*, 288–296. [[CrossRef](#)]

181. Cai, W.; Zhu, Y.; Furuya, K.; Li, Z.; Sokabe, M.; Chen, L. Two different molecular mechanisms underlying progesterone neuroprotection against ischemic brain damage. *Neuropharmacology* **2008**, *55*, 127–138. [[CrossRef](#)] [[PubMed](#)]
182. Cochrane, D.R.; Spoelstra, N.S.; Richer, J.K. The role of miRNAs in progesterone action. *Mol. Cell Endocrinol.* **2012**, *357*, 50–59. [[CrossRef](#)] [[PubMed](#)]
183. Theis, V.; Theiss, C. Progesterone: A universal stimulus for neuronal cells? *Neural Regen. Res.* **2015**, *10*, 547–549. [[CrossRef](#)] [[PubMed](#)]
184. Theis, V.; Theiss, C. Progesterone Effects in the Nervous System. *Anat. Rec.* **2019**, *302*, 1276–1286. [[CrossRef](#)] [[PubMed](#)]
185. Pieczora, L.; Stracke, L.; Vorgerd, M.; Hahn, S.; Theiss, C.; Theis, V. Unveiling of miRNA Expression Patterns in Purkinje Cells During Development. *Cerebellum* **2017**, *16*, 376–387. [[CrossRef](#)] [[PubMed](#)]
186. Herzog, R.; Zendedel, A.; Lammerding, L.; Beyer, C.; Slowik, A. Impact of 17beta-estradiol and progesterone on inflammatory and apoptotic microRNA expression after ischemia in a rat model. *J. Steroid Biochem. Mol. Biol.* **2017**, *167*, 126–134. [[CrossRef](#)] [[PubMed](#)]



© 2020 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).