

Exposure of wild mammals to glyphosate, AMPA, and glufosinate: a case for "emerging organic contaminants"?

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Exposure of wild mammals to glyphosate, AMPA, and glufosinate: a case for "emerging organic 2 contaminants"? 3 4 **Authors and affiliations** Clémentine Fritsch 1*, Brice MR Appenzeller 2, Colette Bertrand 3, Michael Coeurdassier 1, Vincent Driget 1, 5 Emilie M Hardy ², Paul Palazzi ², Charline Schaeffer ², Anne-Claude Goydadin ¹, Sabrina Gaba ^{4,5}, Vincent 6 Bretagnolle 4,5, Céline Pelosi 3,6 7 8 9 ⁽¹⁾ UMR 6249 Chrono-environnement CNRS - Université de Franche-Comté, 16 route de Gray 25030 Besançon 10 Luxembourg Institute of Health, Dpt of Precision Health, 1 A-B rue Thomas Edison, 1445 Strassen, 11 12 (3) UMR 1402 EcoSys, INRAE – AgroParisTech – Université Paris-Saclay, RD 10 Route de St Cyr, 78026 Versailles 13 14 Cedex, France (4) UMR 7372 CEBC, CNRS – La Rochelle Université, USC INRAE, 405 route de Prissé la Charrière, 79360 Villiers-15 en-Bois, France 16 (5) LTSER « Zone Atelier Plaine & Val De Sèvre », 79360 Beauvoir Sur Niort, France 17 18 (6) UMR EMMAH, INRAE - Avignon Université, F-84000, Avignon, France 19 20 21 * Corresponding Author: 22 Dr Clémentine Fritsch 23 UMR 6249 Chrono-environnement 24 CNRS - Université de Franche-Comté 25 16 route de Gray, F-25030 Besançon cedex 26 clementine.fritsch@univ-fcomte.fr 27 clementine.fritsch@cnrs.fr 28 orcid.org/0000-0003-2062-5799 29 30

Abstract

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Glyphosate (GLY) is the most widely used herbicide worldwide, and its use continues to increase. Accumulating evidence shows that GLY and its metabolite aminomethylphosphonic acid (AMPA) are more persistent and toxic than expected, but little is known about the risks to wildlife. Glufosinate (GLUF) was considered an alternative broad-spectrum herbicide, but its field ecotoxicology has rarely been studied. This study aimed to assess the exposure of free-ranging rodents and shrews from treated (cereals under conventional farming) and nontreated habitats (cereals under organic farming and hedgerows) in France to GLY, AMPA, and GLUF through residue analyses in hair. We investigated the patterns of accumulation according to species, habitat, and treatment intensity at the plot, landscape, or township scale. We showed a generalized exposure of small mammals to GLY, AMPA and GLUF, as they were detected in all of the species, and in 64%, 51% and 44% of the hair samples, respectively. The detection and levels of GLY, AMPA and GLUF were greater in herbivorous and omnivorous voles than in insectivorous shrews and omnivorous wild mice. The three compounds showed comparable ranges of concentrations: 0.018-7.74 pg/mg with an outlier of 522 pg/mg for GLY (median = 2.65), 0.240-33.6 pg/mg for AMPA (median = 1.39), and 1.16-25.5 for GLUF (median = 3.51). The frequencies of detection and concentrations did not significantly differ according to the farming practices or proxies of pesticide treatment intensity. The concentrations of GLY were greater in individuals captured in hedgerows than in those captured in cereal fields. On the basis of dose reconstruction approaches and toxicological thresholds from the literature, GLY and GLUF levels may be associated with risk in small mammals and endanger local populations. Our findings raise issues about the omnipresence of GLY, AMPA and GLUF in agricultural landscapes, including in animals from habitats considered refuges, questioning their ecological safety. This work provides new insights into current broad-spectrum herbicide wildlife ecotoxicology that may support decision-making to protect biodiversity.

- **Keywords**: agroecosystem; bioaccumulation; currently used pesticides; landscape ecotoxicology;
- 57 wildlife toxicology

1. Introduction

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Since its registration as a broad-spectrum contact herbicide in 1974, the use of glyphosate (GLY) has continuously increased, making GLY-based pesticides the most widely applied herbicides worldwide (Gandhi et al. 2021; Richmond 2018). Maggi et al. (2020) reported that GLY and its main degradation product aminomethylphosphonic acid (AMPA) may be persistent contaminants in approximately 30% and 93% of global cropland soils, respectively. On the basis of persistence, accumulation and toxicity criteria, they computed that 1% of croplands worldwide (i.e. approximately 385,000 km²) experience mid-high to high hazard conditions. Outside these hotspots, contamination is expected to be low but globally pervasive (Maggi et al. 2020). Initially, GLY was marketed as an "environmentally friendly" herbicide owing to its low expected environmental persistence (easily biodegraded), low predicted toxicity to wildlife and humans (absence of the shikimate pathway in animals, which is found only in plants and microorganisms and is the target pathway of GLY in plants), and lack of potential for bioaccumulation (hydrophilic, low log octanol-water partition coefficient K_{ow} (- 3.12) which is a measure to assess whether a substance is more or less hydrophobic (and/or lipophilic, fat solubility) or hydrophilic (and/or lipophobic, water solubility) and is used as one of the regulatory criteria to classify a compound as bioaccumulative if log K_{ow} > 5, poorly metabolized and rapidly eliminated in mammals) (EFSA 2015a; Gandhi et al. 2021; Kissane and Shephard 2017; Richmond 2018). However, concerns about the environmental safety and health risks associated with GLY have increased within recent decades, especially in the context of its continuously increasing use (Gandhi et al. 2021; Székács and Darvas 2018; Van Bruggen et al. 2018; Vandenberg et al. 2017). GLY and AMPA are more persistent in the environment than initially assessed, as shown by the ubiquitous occurrence of residues in soil, water, and air (Bai and Ogbourne 2016; Gandhi et al. 2021; Richmond 2018; Van Bruggen et al. 2018; Xu et al. 2019). They can accumulate in environmental compartments, crops, and the human food chain (Bai and Ogbourne 2016; Richmond 2018). They are associated with chronic low-dose exposure risks in animals and humans, and can be more harmful to ecosystems than expected (Gandhi et al. 2021; Kissane and Shephard 2017; Ojelade et al. 2022; Peillex and Pelletier 2020; Richmond 2018). Furthermore, GLY and AMPA may be more bioaccumulative substances than predicted from their physico-chemical properties (Pelosi et al. 2022; Richmond 2018; Ruuskanen et al. 2020; Serra et al. 2021). However, the controversial issues regarding GLY-based herbicides have focused mostly on human health (Székács and Darvas 2018; Vandenberg et al. 2017), whereas GLY and AMPA exposure and their impacts on wildlife have been overlooked (Kissane and Shephard 2017). In the USA and EU, registration reviews concerning GLY use as a plant protection product (PPP) identified potential risks to terrestrial and aquatic plants and nontarget terrestrial arthropods and risks to mammals and birds (EFSA 2015b, 2015c, 2015a; US EPA 2020). In the EU assessment for vertebrates, the taxa identified as the most at risk were mammals, particularly herbivorous mammals (EFSA 2015b, 2015a). Glufosinate (GLUF), like GLY, is an organophosphorus compound used as a broad-spectrum nonselective contact herbicide with a systemic action and was considered "one of the very few alternatives to glyphosate" (European Commission 2017). However, risks for herbivorous and insectivorous mammals associated with the herbicidal use of GLUF have been reported in environmental risk assessment (ERA) processes (EFSA 2005, 2012). GLUF was classified as a candidate for substitution in the EU according to Commission Regulation EC No 1107/2009 (PPP Regulation) in 2015, and withdrawal was scheduled at its end of approval in the EU in 2018 (Robin and Marchand, 2023). However, GLUF is still used in many countries worldwide. The singular physicochemical properties of GLY (a particularly hydrophilic polar compound, existing in different ionic states depending on pH, making it difficult to extract) and closely related compounds such as AMPA or GLUF make measurement of their residues challenging and prevent these compounds from being included in multiresidue analyses (Delhomme et al. 2021). Such technical limitations in analytical chemistry may have hampered the development of studies about GLY, AMPA and GLUF exposure and their effects in wildlife and, more generally, knowledge building about the terrestrial ecotoxicology of these compounds. Ultimately, this lack of knowledge could be an impediment for regulatory decisions with respect to the objectives of reducing the risks and impacts

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characterizing the exposure of wildlife to GLY, AMPA and GLUF is therefore a critical issue for environmental protection and biodiversity conservation. The main aim of this study was to investigate the exposure of wildlife in an arable landscape to GLY, AMPA and GLUF and evaluate the risk for toxicological impairment, with a focus on small mammals. This was achieved via a minimally invasive sampling method based on residue analyses in hair and on reverse dosimetry approaches. The compound residues can be detected and quantified in hair, owing to the effectiveness of this matrix for assessing exposure to multiple pesticides in wildlife (Fritsch et al. 2022; Krief et al. 2017), as shown for GLY in bats (Hooper et al. 2022) and both GLY and AMPA in humans (Alvarez et al. 2022). Moreover, GLY concentrations were relative to the intensity of exposure in human samples (i.e. GLY was quantified at lower concentrations in hair samples from nonoccupationally exposed subjects than in hair samples from farmers, who often used GLY) (Alvarez et al. 2022). Differences in exposure between the three compounds were investigated, and the influences of farming practices and herbicide use intensity were studied. We expect exposure to GLY to be greater than that to the other compounds. Indeed, GLY is more commonly used and is still authorized in France, in contrast to GLUF (BNV-D Traçabilité), and AMPA is not a pesticide active substance itself but a metabolite of GLY. Mammals can be exposed to herbicides through diet and overspray (European Commission, 2015; Martinez-Haro et al. 2022; Mayer et al. 2020; Newton et al. 1984). Therefore, small mammals may be exposed to GLY through both trophic exposure and/or overspray, whereas direct exposure during spraying should be negligible for the other two compounds. Higher concentrations in hair were expected in treated areas (i.e. conventional crops) than in nontargeted habitats (i.e. organic farming (OF) fields and hedgerows). Since small mammals are mobile species, the influence of herbicide use intensity was also considered at the landscape scale (i.e. using the proportion of OF fields within the landscape as a proxy) and at the township level (i.e. using sales of active substances as a proxy of usage intensity). To further document the factors that may shape exposure to GLY, AMPA and GLUF, differences between small mammal species

of pesticide use on human health and the environment (e.g. Directive 2009/128/EC). Assessing and

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(rodents and shrews) with various ecological traits were investigated. Herbivorous voles are expected to exhibit greater exposure than are granivorous, omnivorous or insectivorous species according to the higher daily dietary doses estimated during ERA processes (EFSA 2005, 2012; European Commission 2015). Moreover, herbivorous voles are less mobile than omnivorous mice and voles or shrews are (van den Brink et al. 2011; Wijnhoven et al. 2005) and may not avoid treated areas. We expect species that feed on animal matter (i.e. omnivores, carnivores, insectivores or vermivores) to also be exposed because residues of GLY, AMPA and GLUF have been found in earthworms that are the prey of these trophic groups (Pelosi et al. 2022), and GLY residues have been found in small carnivorous and omnivorous mammals in a forest ecosystem (Newton et al. 1984).

2. Material and methods

2.1. Study design

The study site was the Long-Term Socio-Ecological Research (LTSER) "Zone Atelier Plaine & Val de Sèvre" (ZAPVS, https://za-plaineetvaldesevre.com/) in mid-western France (see Bretagnolle et al. (2018) for a general site description). The sampling design was based on 40 sampling squares of 1 km², initially selected on the basis of landscape features (gradient of seminatural habitats such as hedgerows and a gradient of organic farming surfaces), in which sampling by trapping was conducted in an arable cereal field (winter cereals) and a woody patch (hedgerow or woodlot edge) as close as possible to the sampled arable field (cf. Pelosi et al. 2022), for a total of 78 plots investigated (Fig. 1). Cropped cereals and hedgerows are referred to as "habitats" hereafter. Cropped fields in which the sampling was carried out were cultivated either under conventional farming (CF) or organic farming (OF). The farming practices in the organic fields followed the rules of the "AB (Agriculture Biologique) France label" and were under OF for at least three years at the time of sampling. A few fields were in transition from CF to OF and were referred to as "OF/CF". Sampling was conducted in spring 2018 (May-June).

2.2. Sampling process

Small mammals were captured using traplines, which were set in each sampling plot over the sampling squares using lines of nonlethal traps (25 traps, 2 m apart) with dormitory (Le Quilliec and Croci 2006) filled with hay and food (i.e. pieces of carrots and apples, sunflower seeds, cat food and peanut butter). The hay, vegetables, fruits and seeds originated from organic agriculture. The traps were checked every day, rebaited and refilled as necessary, and replaced by a new trap in case of capture. The traplines were set during one to four nights depending on logistics and capture success (i.e. longer duration, if possible, in case of an absence of capture during the first night(s)). Species were determined during the handling of individuals on the basis of morphometrics and confirmed by molecular analyses in case of doubt (see below). A sample of 50-100 mg of hair was taken by shaving

over the posterior part of the back, wrapped in aluminium foil and stored in plastic zip-lock bags at room temperature during the field session and later at -20 °C until analysis. Individuals were released after handling. In case of recapture, the individuals were immediately released without any measurement or sampling. For animals that did not survive capture, the bodies were collected and stored at -20 °C until they were shaved for hair sampling. Because Microtus arvalis and Microtus agrestis are morphologically similar species, the identification of Microtus voles was confirmed by molecular analyses (Sup Info Annex 1. Text A1). The suspected Mus spp individuals were also confirmed genetically. For that, a tiny piece of ear was sampled (using medical clean and disinfected scissors, followed by disinfection of the ear) and stored by freezing (-20 °C) until DNA extraction. The experiments were performed under the authorization of the French National Ethical Committee (Project APAFIS N°5340) by skilled and experienced investigators from the Chrono-environnement research department (EU0592), following directive 2010/63/EU on the protection of animals used for scientific purposes. All precautions to limit stress and deleterious effects on the animals as much as possible were taken. The sampling and handling of small mammals were conducted under supervision and with the participation of people authorized for animal experimentation and experienced in small mammal capture and handling and using appropriate and authorized (EU0592)

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required for animal welfare).

2.3. Proxies related to the intensity of treatments at various spatial scales

First, to investigate the role of agricultural practices in shaping the exposure of small mammals, habitats were classified as cultivated under OF or CF according to the type of agricultural organic or conventional practices in the cultivated fields. To classify hedgerows or woodlots with respect to farming practices, the adjacent plots were considered, and each woody patch was classified as "OF" if adjacent fields were under OF and none of the adjacent fields was under CF, "OF/CF" if at least one of the adjacent fields was under CF, or "CF" when all adjacent fields were under CF.

facilities (a vehicle equipped with a mobile anaesthesia unit and all of the materials and equipment

For the proxies of treatment intensity, at the plot scale (arable field or woody patches where trapping was realized), a general proxy of potential for pesticide treatment was computed. The fields cultivated under OF and hedgerows or woodlots were considered not intentionally targeted by treatments (NT), whereas the arable fields cultivated under CF and transitional fields cultivated from CF to OF were considered possibly targeted (T). A second proxy to characterize the intensity of potential treatments was computed at the landscape scale. The locations and boundaries of cultivated plots in 2018 were obtained from the official French "Registre Parcellaire Graphique" (https://www.data.gouv.fr/fr/datasets/parcelles-en-agriculture-biologique-ab-declarees-a-la-pac). The software Quantum GIS (QGIS 3.16) was used to calculate the proportion of OF plots in the landscape, which was computed in each sampling square of 1 km² as the proportion of the total surface occupied by OF arable fields. The final proxy resulted in a two-level factor, with a category "OF dominant" with proportions varying from 50 to 100% ([49-79], 6 sampling squares), in which treatment intensity at the landscape scale was expected to be lower since arable fields not targeted by pesticide treatments prevailed within the landscape mosaic. The second category, "CF dominant", gathered squares with landscape OF proportions ranging from 0 to 50% ([0-49], 19 sampling squares). Finally, to consider the intensity of potential treatments over the range of several farmlands, a proxy at the township scale was computed. The sales of active substance (a.s., data provided in Kg) were obtained from the "Banque Nationale des ventes de produits phytopharmaceutiques par les Distributeurs agréés (BNV-D Traçabilité)" as released to the public by official agencies at the township level (i.e. according to postal code, https://www.data.gouv.fr/fr). These data were extracted for 2017 and 2018 (the year before and the year when sampling was conducted). The sales of GLY and GLUF were associated with each trapping location on the basis of township positioning. The sales of GLY were highly correlated between years at the township level $(R^2=0.950)$, rendering it impossible to study the influence of potential use intensity separately in the year before or in the year of sampling. The marketing authorisation of pesticides containing GLUF was withdrawn in France by October 2017. The quantities sold in 2017 were very low (i.e. 0.75 kg)

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and sporadic (i.e. only one quantity reported as sold in the townships studied) whereas in 2018, no sales of GLUF were reported in the townships of the study area of concern. The quantities sold in 2017 and 2018 were thus summed for each a.s. and classified into two categories. For GLUF, the categories were "0" and "0.75". For GLY, the median of the summed quantities in the dataset was calculated at 7300, and the categories were set at "lower than 7000" ([4138-7000[, 2 townships) and "higher than 7000" ([7000-9366[, 3 townhips).

2.4. Chemical analyses of glyphosate, AMPA and glufosinate

Full detailed information about chemicals and reagents, sample preparation, methodological approaches used for the chemical analyses and issues related to metrology are provided in Sup Info Annex 1. Text A2.

Briefly, the surface of hair samples were decontaminated with sodium dodecyl sulfate solution and methanol (Duca et al. 2014) After decontamination, the samples were spiked with internal standard solution, and water was added before overnight extraction. The supernatants were evaporated to dryness, and borate buffer and FMOC-Chloride were added before derivatization and injection into the LC-MS/MS Waters system Acquity UPLC H-class system equipped with a Waters Acquity BEH C18 column ($100 \times 2.1 \text{ mm}$, $1.7 \text{ }\mu\text{m}$ particle size) coupled to a Waters Xevo TQ-S triple quadrupole mass spectrometer. With each analytical run, quality controls consisting in hair samples supplemented at several concentrations (0.1 to 100 pg/mg) and a blank sample were included.

The concentrations are expressed as pg/mg hair. The limits of quantification (LOQ) and lowest detected values (Minimum) are presented in Table 1. We did not perform specific experiments and calculations to compute limits of detection (see Sup Info Annex 1. Text A2). Importantly, only one concentration value was reported below the lowest concentration in quality controls (the minimum GLY concentration, see Table 1). All values above the lowest analytical detected value were included as "detects" contrary to "nondetects". All reported concentration values, including values between the LOQ and the lowest detected value, were included as continuous data. The use of concentrations

below the LOQ has been shown to be a suitable method than may be preferred to the absence of data or other established approaches such as imputation-based methods (i.e. efficiency in terms of bias and precision)(Keizer et al. 2015).

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2.5. Dose reconstruction to assess the risk of deleterious effects in small mammals Given the concentrations of GLY, AMPA and GLUF in the keratinized tissues of small mammals, our study highlights the exposure of these animals. However, the toxicological meaning of such hair concentrations is hard to interpret with respect to toxicity benchmarks (which are expressed as daily doses in mg a. s. per kg body weight) because relationships between residues in hair and exposure doses or between residues in hair and in those other organs are not known. To overcome this gap and assess whether exposure levels could reach toxic levels, approaches of reverse dosimetry have been applied to estimate levels of exposure in terms of daily dose intake, to obtain data comparable to toxicological thresholds. Different types of data (i.e. concentrations in mammal body fluids and hair, residues in viscera and body burden in small mammals, toxicokinetics in small mammal blood, pharmacokinetics of fast-elimination chemicals) that may be used for such purposes have been collected from the literature (Sup Info Annex 1. Text A3 for full description), leading to different approaches of back-calculations computed under several assumptions when there was a lack of available data. Five different approaches could be computed for GLY and one for GLUF (Table 2). The objective of our risk assessment was to determine the overall orders of magnitude of daily dose intake relative to the concentrations measured in hair to evaluate whether wild small mammals may experience deleterious toxicological effects. The lack of data and assumptions that had to be applied hamper the accuracy of the calculated dose values. The interpretation of the findings was thus developed to consider ranges and global patterns rather than refined (but biased) toxicological assessments. The doses calculated were compared with toxicity data using the usual and regulatory procedure of toxicity-exposure ratio (TER) calculation, with associated triggers of 5 for chronic toxicity data and of 10 for acute toxicity data. The doses were reconstructed for each species, using four levels of residue concentrations measured in hair: minimum, median, 3rd quartile and maximum values (only one value available in house mice for GLY and in house mice and shrews for GLUF). The dose of GLY was compared to five toxicological thresholds: the short-term dietary no observed effect level (short-term dietary NOEL), the chronic 21 day no observed adverse effect level (chronic NOAEL, which is the same value as the reproductive NOAEL), the long-term NOAEL, the no mortality dose to the Oregon vole, and a "chronic low dose" limit. Details about the values of the thresholds are provided in the Sup Info Annex 1. Text A3. The dose of GLUF was compared to four available toxicological thresholds: the acute oral median lethal dose, the short-term dietary NOEL, the chronic 21 day NOAEL, and the long-term NOAEL.

The process ended with the TER values computed for each approach (i.e. back-calculations on the

basis of concentrations in human urine and hair, body burden in rats, toxicokinetics in rat blood, residues in viscera and body of wild small mammals, prediction of incorporation in hair from pharmacokinetics), under different scenarios of assumption, for the different levels of residue concentrations measured in hair (minimum, median, 3rd quartile and maximum values), for each species, for the different toxicological thresholds (see details in Sup Info Annex 2). Finally, the number of cases where TER exceeded the triggers, which highlights the risk for toxic effects in small mammals, and the number of individuals involved were summarized.

2.6. Data analysis and statistics

Analyses were performed using R (3.3.1), with the additional packages "AICcmodavg", "effects", "EnvStats", "forestmodel", "ggeffects", "ggplot2", "gtsummary", "multcomp", "pgirmess", and "questionr". Since more than 2 individuals were obtained from the same trapline in only 8 out of the 29 where the animals were captured, pseudoreplication was not considered an issue. Moreover, the traplines extended over 50 metres, which should limit the influence of spatial autocorrelation between individuals. The statistical approaches were designed to handle left-censored data within a situation where the smallest observations of data are nondetects, with high proportions of

nondetects, skewed distributions, and small sample sizes, which hampered the use of methods considered "gold standards" to manage left-censored data (ITRC 2013; Shoari and Dubé 2018). Following recommendations to analyse datasets including nondetected and left-censored observations, we used tests dedicated to censored data when possible, computed descriptive statistics separately on detection frequencies and on detected concentrations (for which median, interquartile range, and outer percentiles were reported), performed group comparisons separately on detection frequencies and on detected concentrations, and preferred nonparametric tests to analyse the combined dataset, including uncensored data and nondetects with the latter being substituted by 0 (ITRC 2013; Shoari and Dubé 2018). Compared with other methods (i.e. imputationand nonimputation-based approaches), the traditional substitution approach has been shown to be appropriate for handling nondetects in a study that investigated a dataset of pesticide concentrations in pollen and nectar to assess the exposure of bumble bees (Adjei and Stevens 2022). Substitution with a half limit of detection outperformed the other approaches in the study of Adjei and Stevens (2022), which used parametric tests (nested ANOVA), but we preferred to use 0 because (1) we used all reported values, including those between the LOQ and lowest detected values, instead of data left-censored by the limit of detection (Keizer et al. 2015), (2) the lowest detected values reached low values that would have brought the value of half of the LOD close to 0, (3) this approach allows easy identifying nondetects in the databases and graphics, and (4) nonparametric tests were used to compute statistics on both nondetects and quantified values. Comparisons between the distributions of the compounds were performed using Tarone-Ware two-sample test for censored data (i.e. GLY vs. AMPA, GLY vs. GLUF, and AMPA vs. GLUF), with nondetects set to the corresponding lowest detected value of the corresponding compound as needed in such tests, for all species together (the sample sizes were too small to run the tests by species). Correlations between raw values with nondetects set to 0 of the concentrations of the three

compounds were investigated using Spearman's rank correlation tests. The relationships between

the concentrations of each compound were further studied according to species via generalized

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linear models (Gaussian family) on log-transformed data (log(x+1)) to meet the assumptions of normality and variance homogeneity. The models were checked for homogeneity of variance, normality of error and linearity/additivity and leverage was checked (i.e. standardized residuals vs. leverage). An extreme outlier of GLY concentration was identified (one Microtus vole) as having overinfluence on the estimated parameters (i.e. Cook distance greater than 1). This outlier was removed from the dataset to compute model outputs and graphics in this part of the analyses. Then, statistical analyses were performed on (1) GLY, AMPA or GLUF frequencies of detection and (2) GLY, AMPA or GLUF concentrations quantified above the lowest detected value (i.e. in samples where the compound was detected, without nondetects). According to the preliminary analyses showing differences between species, further analyses were conducted taking species into account in statistics using the factor "species" first in the models with the other factors. Full models including all explanatory factors and first-order interactions could not be used due the large number of NAs. Preliminary tests performed on combinations of factors, including interactions, revealed that none of the interactions were statistically significant and that models without interactions better fit the data (according to corrected Akaike information criterion (AICc) comparisons). The final models used to compute the statistics were therefore run without interactions. We tested whether the detection of GLY, AMPA or GLUF in hair of small mammals was dependent on species, habitat, farming practices and proxies for treatment intensity using binomial GLMs (link = logit). The detection was coded as "0" for nondetected and "1" when detected. The models were built using species alone and species with each of the other variables as explanatory factors. The significance of the factors was checked by the chi-squared test. Odds ratios with confidence intervals were computed to characterize differences between levels. We tested whether the concentrations of GLY, AMPA or GLUF in the hair of small mammals were dependent on species, habitat, farming practices and proxies for treatment intensity using GLMs on log-transformed values above the lowest detected value (i.e. without nondetects) to meet the assumptions of normality and variance homogeneity. The models were built using species alone and species with each of the other variables as explanatory factors. The significance of the

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explanatory factors was checked via an *F* test, and statistical differences between levels were via a *t* test.

To support the results and interpretation, both the detection and concentration data were analyzed via nonparametric tests on all of the data, with nondetects included and set to 0. The differences between species, habitats, farming practices, and the proxies of treatment intensity were tested adequately via the Wilcoxon-Mann-Whitney test or the Kruskal-Wallis test adequately depending on the number of levels of the factors (i.e. two or more). Post hoc multiple comparison tests after Kruskal-Wallis tests were used to investigate pairwise comparisons when relevant. Correlations between concentrations and landscape OF proportions or sales of a.s. used as raw continuous data were tested via Spearman's rank correlation test.

360 Figure 1. Maps of the study design (A) and location of the traplines where hair samples were collected (B and C) across the study area. Legend 361 Land use Forest, woodlot & hedgerow 362 Cropped field Grassland Vineyard Roads, urban & non-agricultural areas 363 Water Organic Farming 364 365 Sampling plots Landscape squares (1 km2) 366 B) Traplines hair sampling 367 5 km 5 km 368 C 369 C) List of species by traplines 370 Bank vole ▲ Common vole Common vole & Bank vole 371 Common vole & House mouse △ House mouse ■ Shrew Shrew & Bank vole 372 Shrew & Common vole ▲ Wood mouse Wood mouse & Bank vole 373 Wood mouse & Shrew

Wood mouse & Shrew & Common voleWood mouse and Common vole

3. Results & Discussion

3.1. Captures of small mammals

The sample size resulted in hair samples from 61 individuals captured in 29 traplines distributed over 25 squares of 1 km² out of the sampling plots initially surveyed (Fig. 1, Table 1; further details about the sample sizes are provided in Sup Info Annex 1. Table A1). Six individuals were recaptured. The number of recaptures was quite low since most of the traplines were set for only one night (1 night: n = 63, 2 nights: n = 12, 4 nights: n = 3). Individual samples of 18 wood mice (*Apodemus sylvaticus*), 14 greater white-toothed shrews (*Crocidura russula*), 16 common voles (*Microtus arvalis*), 11 bank voles (*Myodes glareolus*), and two house mice (*Mus musculus*) were analysed. A total of 41 samples were collected from individuals captured in habitats under CF farming, 10 from OF/CF category, and 10 from the OF category. A total of 39 samples were collected from hedgerows or woodlots, and 22 were collected from cereal fields.

3.2. Detection frequency of GLY, AMPA and GLUF in the hair of small mammals

Considering all of the species together, GLY, AMPA and GLUF were detected in 64%, 51%, and 44% of the individuals, respectively (Table 1). Compared with the quantities sold in the townships studied, the quantities of GLUF were 2484 to 4844 times lower than those of GLY in 2018 and 1653 to 4548 times lower in 2017. Indeed, after its ban in 2017, GLUF could only have been used until stock depletion or usage derogation. Focusing on the part of the region where the study was carried out (i.e. "Département des Deux-Sèvres"), the purchases of GLUF rapidly decreased after 2015 and reached 0 in 2019 (BNV-D"). Despite its low intensity of application compared with that of GLY, GLUF was detected in approximately half of the small mammal individuals. Within the "PING" research programme, the part of the study conducted over the same year over the same sampling sites as the present one detected GLY in 88% of the soil samples and 74% of the earthworm samples, AMPA in 58% of the soil samples and 38% of the earthworms, and GLUF in 35% of the soil samples and 12% of the earthworms (Pelosi et al. 2022). These detections in soil, earthworms and small mammals

collected from arable fields and hedgerows, both in conventional and organic farming fields, revealed a generalized occurrence of GLY, AMPA and GLUF in the agricultural environment and fauna across this typical French agricultural area. In comparison, in three county-level hair pools of carcasses of big brown bats (*Eptesicus fuscus*; an insectivorous species weighing 15 to 26 g) in Missouri (USA), GLY was detected in all three analysed sample pools (100% detection frequency) (Hooper et al. 2022). The frequency of exposure to GLY in a farmland herbivorous mammal, the Iberian hare (*Lepus granatensis*) via analyses of residues in samples of gastric content, reached 45% of individuals found dead due to roadkill or unknown causes (Martinez-Haro et al. 2022). In hunted hares from pesticide-treated areas the frequency of detection in gastric content ranged between 9% and 22%. These values look lower than the frequencies found in rodents in our study, possibly because residues in gastric content represent a snapshot of recent exposure, whereas measurements in hair represent a temporally integrative exposure assessment during the time of hair growth. Issues related to local contamination and/or different analytical sensitivities may further explain the lower detection frequency in hares.

Descriptive statistics on concentrations were computed for samples where the given compound was detected, i.e. nondetects (samples whose concentrations were below the lowest detected value (Minimum)) were excluded. Home range size were obtained from Chambers et al. (2000) and Wijnhoven et al. (2005). The bank vole is often considered a mostly granivorous species but recent studies have shown omnivory in this taxon. Abbreviations: OF=Organic farming, CF=Conventional farming, NT=Not targeted by treatments, T=Targeted by treatments, LOQ= Limit of quantification.

	All species	Wood mouse Apodemus sylvaticus	Greater white-toothed shrew Crocidura russula	Common vole Microtus arvalis	House mouse Mus musculus	Bank vole Myodes glareolus
Diet	All species	Omnivore	Insectivore	Herbivore	Omnivore	Omnivore
Mean home-range size (m ²)		230–12200	50–200	161–1350	2710	260-4100
NUMBER OF INDIVIDUALS						
Habitat						
Cereal	22	9	2	9	2	
Hedgerow or woodlot	39	9	12	7		11
Farming						
CF	41	12	9	9	1	10
OF/CF	10	4	3	2		1
OF	10	2	2	5	1	
Not treated or treated						
« NT »	41	9	12	8	1	11
«T»	20	9	2	8	1	
Landscape OF proportion						
0-50%	40	11	9	8	1	11
50-100%	21	7	5	8	1	
Sales GLY						
< 7000	25	6	4	11	2	2
> 7000	36	12	10	5		9
Sales GLUF						
0	44	15	11	7	_	11
0.75	17	3	3	9	2	
Total	61	18	14	16	2	11
GLYPHOSATE						
Detections			_			
Number of detection	39	6	7	15	1	10
Frequency (%) Concentrations (pg/mg)	64	33	50	94	50	91
LOQ	0.018	0.600	0.600	0.010		1 42
Minimum Q1	0.018 1.040	0.600 0.795	0.600 0.600	0.018 2.23		1.43 2.60
Median	2.65	1.77	0.800	4.42		3.65
Q3	5.77	4.83	0.950	8.13		5.63
Maximum	522	5.92	1.22	522	3.79	7.74
AMPA						
Detections						
Number of detection	31	4	2	14	1	10
Frequency (%)	51	22	14	88	50	91
Concentrations (pg/mg)						
LOQ	1	0.422	0.455	0.5.5		0
Minimum	0.240	0.400	0.400	0.240		0.515
Q1	1.025	0.519	0.617	1.08		1.19
Median Q3	1.39 6.45	1.29 2.03	0.833 1.05	2.51 7.65		1.61 6.51
Maximum	33.6	2.07	1.27	19.4	1.37	33.6
GLUFOSINATE						
Detections						
Number of detection	27	2	1	13	1	10
Frequency (%)	44	11	7	81	50	91
Concentrations (pg/mg)						
LOQ	1	2.04		1.30		1.10
Minimum	1.16	2.04		1.29		1.16
Q1 Median	2.01	7.88 13.8		2.39		1.59 3.01
Median Q3	3.51 7.25	13.8 19.7		4.05 5.80		3.01 4.87
Maximum	25.5	25.5	1.48	21.8	9.75	4.87 12.6
iviaxiiIIuIII	23.3	دی.ی	1.40	21.0	3.13	14.0

3.3. Concentrations of GLY, AMPA and GLUF in the hair of small mammals

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With respect to residue concentrations above the lowest detected value, we found that the three compounds presented overall similar ranges of concentrations, and the highest concentrations were reached for GLY, with values varying from 0.018 to 522 pg/mg (median = 2.65), whereas the concentrations of AMPA ranged from 0.240 to 33.6 pg/mg (median = 1.39), and those of GLUF ranged from 1.16 to 25.5 pg/mg (median = 3.51) (Table 1). The concentrations of GLY did not differ significantly from those of AMPA or GLUF, and the concentrations of AMPA did not differ from those of GLUF (Tarone-Ware tests, p-values > 0.05, Sup Info Annex 1. Text A4). In county-level hair pools from big brown bats in Missouri, Hooper et al. (2022) reported concentrations of GLY varying from 5.7 to 4505.2 pg/mg. These maximum concentrations are much higher than those measured here in shrews (median = 0.8 pg/mg) and almost ten times higher than the maximum concentration measured in our study in a vole (i.e. 522 pg/mg). Several nonexclusive hypotheses may be proposed to explain such high levels found in bats: analytical issues, physiological and ecological traits of the species enhancing exposure and/or bioaccumulation (Fritsch et al. 2024), and issues related to the amount of GLY used and the timing of exposure of the animals after GLY application. Newton et al. (1984) studied the fate and behaviour of GLY and AMPA in forest brush field ecosystems in the Oregon Coast Range (USA) following aerial application of GLY at doses comparable to the recommended GLY application rate in the EU (i.e. 3.3 kg a.s./ha there, maximum field application of 4.32 kg a.s./ha in the EU (European Commission 2015)). They measured the residues of GLY and AMPA in the viscera and body of free-ranging small mammals captured at the sites before treatment and over the 55 days posttreatment. As in the present work, GLY was detected in all of the trophic groups (LOD of 0.10 mg/kg = 100 pg/mg). However, AMPA was detected in two voles only at 130 and <160 pg/mg (LOD of 100 pg/mg) (Newton et al. 1984), which is far lower than in our study. This may be because the lowest detected value in our work was much lower (i.e.

0.240 pg/mg) and because different tissues were used to measure residues.

Interestingly, considering these previous findings of Newton et al. (1984) in forest small mammal tissues, the concentrations varied over the same order of magnitude as the maximum concentrations measured in our hair samples from agricultural landscapes. They reported concentrations in viscera ranging from < 100 pg/mg (LOD) to 1690 pg/mg in shrews, 5080 pg/mg in deermice and 1700 pg/mg in voles, and concentrations in body tissues ranging from < 100 to 410 pg/mg in shrews, 400 pg/mg in deermice and 250 pg/mg in voles. The concentrations were the highest during the first days following herbicide treatment both in the viscera and body tissues. Body concentrations drecreased under the LOD by 2 weeks after treatment while they were still detectable in the viscera, and concentrations in the viscera decreased to nondetectable concentrations or near the LOD by day 55 after treatment (Newton et al. 1984). Together with differences in the matrices analysed, this (rapid) concentration decrease over time following a single GLY application may explain the overall lower GLY levels in our study and suggests that the highest concentrations found here may correspond to animals exposed to GLY soon after its application. Our samples were collected during springtime, from May-June, whereas glyphosate is usually applied to winter cereals during late summer or fall (often in October). The seasonal moult of small mammals is expected to be completed mostly by May-June, when they were captured, and the sampled fur might have partly or even totally grown within the period of February-May (Abad 1991; López-Fuster et al. 1986; Sealander 1972). However, the postwinter moult may be incomplete, depending on the age of individuals and the species of concern, allowing the possibility for hair grown during autumn to be included in the analysed fur sample (Beltran et al. 2018).

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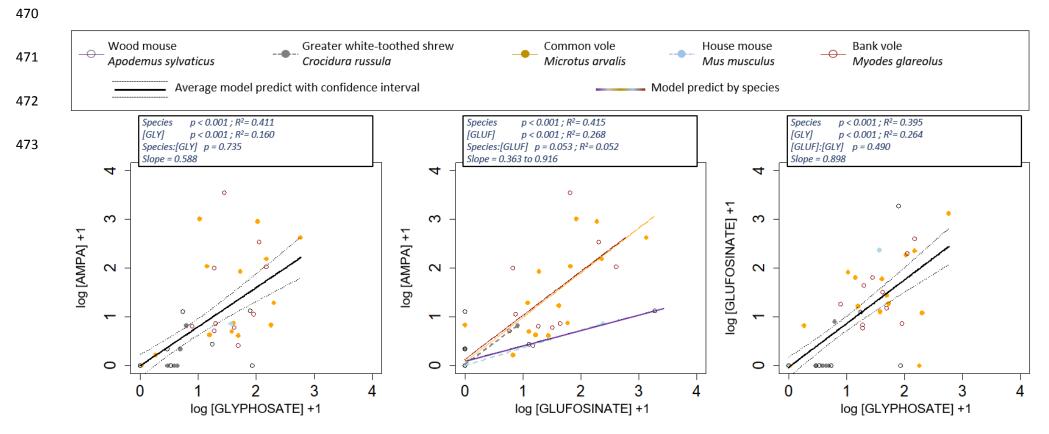
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Figure 2. Correlations between the concentrations of glyphosate, AMPA and glufosinate in the hair of small mammals.

The concentrations are expressed in pg/mg. The statistical outputs of general linear models are provided on graphs. Predictions are plotted according to species when the interaction between the factors is significant, and average predictions are provided when the interaction is not significant.



The concentrations of the three compounds in the hair of small mammals were significantly related (Fig. 2, Annex Fig. A1). Positive correlations were observed between AMPA and GLY (correlation coefficient Spearman's rho=0.84), between GLUF and GLY (rho=0.79), and between AMPA and GLUF (rho=0.89) (Annex Fig. A1). The variance explained (cf. determination coefficient R²) was greater for the factor species (0.41 to 0.42) than for compounds (0.16 to 0.27) (Fig. 2). The correlation between AMPA and GLUF differed only according to species, with greater slopes for shrews and voles than for the two species of wild mice (Fig. 2). The correlations between AMPA and GLY and between GLUF and GLY were similar between species (Fig. 2). The strength of the association between AMPA and GLUF was greater than that between AMPA and GLY (i.e. greater rho values, greater R² values; Fig. 2, Sup Info Annex 1. Fig. A1), and the relationship presented greater slope coefficients. This may be seen as unexpected since AMPA is a GLY transformation product. Such patterns of correlation between the compounds may reflect different exposure pathway contributions between GLY versus AMPA and GLUF. As GLY is poorly metabolized in mammals (EFSA 2015a), the origin of AMPA should be external (environmental) rather than internal (degradation of GLY in the body), as suggested in humans (Alvarez et al., 2022). Studying GLY and AMPA residues in human hair, Alvarez et al. (2022) reported different origins of GLY and AMPA because AMPA was rarely detected in comparison with GLY in both farmers and nonoccupationally exposed subjects, and the ratio of GLY to AMPA greatly differed between the subjects. The correlation between GLY and AMPA or between GLY and GLUF likely originates from a spatial relationship related to the geographic occurrence of the herbicidal treatments, which increases the probability of co-occurrence of parent and transformation products in herbicide-treated areas. As hypothesized, small mammals may be exposed to GLY via the dietary route but also via direct spraying if they are present in the plots during the treatments and orally after spraying treatments through grooming activities. Conversely, dietary exposure, related to the accumulation of the compounds in the soil, biota and food webs, may be the predominant exposure pathway to AMPA and to GLUF (whose use as a PPP is no longer approved) in small mammals.

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3.4. Differences between species in detection frequency and concentration of the compounds The frequency of detection differed among species for the three compounds (Fig. 3, Sup Info Annex 1. Table A2). In all cases, the odds ratios were significantly different between the voles and the reference (i.e. wood mice) (Sup Info Annex 1. Table A2). The probabilities of detection of GLY, AMPA and GLUF were significantly greater in common and bank voles than in shrews and wood mice (Fig. 3). The results were contrasted for the house mouse, likely because of the small sample size (Fig. 3). When analyses were run on the subset of data with quantified concentrations in small mammals (i.e. without nondetects), the concentrations of GLY, AMPA and GLUF did not significantly differ among species (Sup Info Annex 1. Table A3). However, the trends considering GLY and AMPA were comparable overall, since estimates for concentrations were found to be the lowest in shrews and wood mice for GLY and the highest in voles for AMPA (Sup Info Annex 1. Table A3). Considering GLUF, the highest estimates were observed for wood mice and house mice (Sup Info Annex 1. Table A3). When the tests included nondetects set to 0, the results revealed higher levels of GLY, AMPA, and GLUF in common voles and bank voles than in wood mice and shrews (Fig. 4). The concentrations in house mice did not significantly differ from those in other species, with intermediate values (note that the sample size is small) (Fig. 4).

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Figure 3. Probabilities of the detection of glyphosate, AMPA and glufosinate in the hair of small mammals according to species.

The predicted probabilities of detection were obtained from the statistical binomial models "Detection ~ Species". Statistical outputs of comparisons between levels are provided, and the species sharing similar letters did not differ statistically. Abbreviations: GWT shrew stands for greater white-toothed shrew. The numbers of individuals in each category are provided in Table 1.

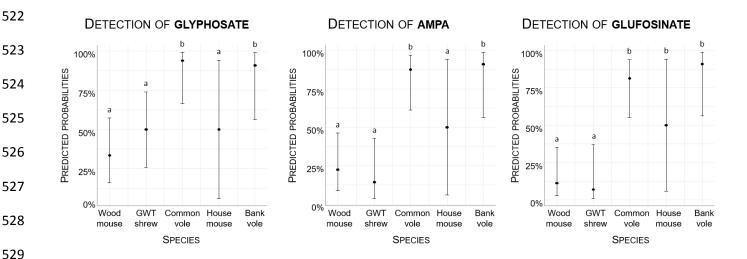
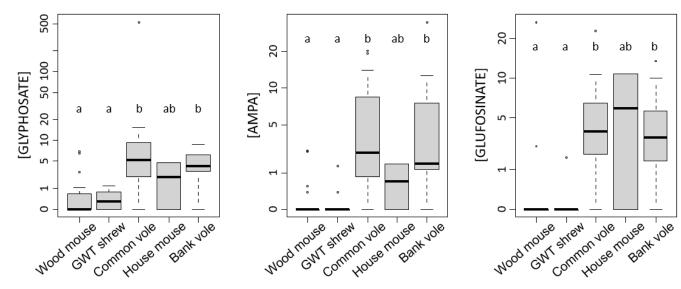


Figure 4. Concentrations of glyphosate, AMPA, and glufosinate in the hair of small mammals according to species.

The concentrations are expressed in pg/mg. Nondetects are set to 0. Statistical significance is indicated with letters, and different letters between levels indicate significant differences. Statistics were performed via the Kruskal-Wallis test followed by posthoc multiple comparison tests to investigate pairwise differences. Abbreviations: GWT shrew stands for greater white-toothed shrew. The sample sizes are provided in Table 1.



In their forest study where residues of GLY and AMPA were measured in several free-ranging mammalian herbivores, carnivores, and omnivores, Newton et al. (1984) reported that GLY was present in all trophic groups, whereas AMPA was present in only a few herbivorous voles. Our results are in line with these findings, with confirmed exposure to GLY (and AMPA) in both herbivorous and granivorous or omnivorous rodents and insectivorous shrews and greater levels of AMPA in herbivorous mammals (note that the bank vole was often considered a granivore, but recent literature has shown omnivory (Balčiauskas et al., 2022)). Newton et al. (1984) emphasized that exposure to and accumulation of GLY in mammals varied according to diet preference, since omnivores (deermice) had the highest concentrations of GLY in the viscera during the 2 weeks posttreatment, suggesting a higher initial intake of GLY in omnivores, but such visceral levels decreased the fastest in comparison to carnivores (shrews and weasel) and herbivores (squirrel, vole, chipmunk). The detailed data revealed greater body concentrations in shrews and deermice (0.35 to 0.41 mg/kg) than in squirrels and voles (0.13 to 0.25 mg/kg) within the first days posttreatment. Our results similarly revealed differences in the accumulation or uptake of GLY and AMPA between trophic groups or species, but various trends were identified according to trophic preferences. Overall, we found lower detection frequencies and concentrations in insectivores, whereas the highest detection frequencies and concentrations were observed in herbivorous and omnivorous voles. Both the detection probability and the concentrations were greater for the two species of voles than for the omnivorous wood mice and house mice in our study. The differences in the matrices analysed may explain the discrepancies between the findings of Newton et al. (1984) and our findings regarding the influence of diet on accumulation, in addition to differences in the identities of the species (i.e. European vs. American species) and ecosystems (i.e. agrosystem vs. forest). The physiological and metabolic traits affecting uptake and accumulation in body tissues/fluids and the ecological traits affecting exposure may shape the differences found here between species or trophic groups, but their role is difficult to disentangle. The species exhibiting the lowest detection frequency differ in terms of phylogeny, since both Apodemus and Mus mice of the

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subfamily Murinae as well as Crocidurinae Crocidura shrews were represented (Sup Info Annex 1. Fig. A4). They also differ in their ecological traits since they are omnivorous and insectivorous. The most herbivorous and, to a lesser extent, the least mobile species, which are the two species of voles studied here, presented the highest detection frequencies and greatest concentrations. Microtus and Myodes voles are classified into two different genera, but both belong to the same subfamily, Arvicolinae (Sup Info Annex 1. Fig. A2). The correlation between the levels of GLUF and AMPA varied under species-specific patterns, as detailed above. The wood mouse and the house mouse, which are both omnivorous Murinae species, presented a slope different from that of the herbivorous common vole, the omnivorous bank vole and the insectivorous greater white-toothed shrew which presented a steeper slope. The bank vole is usually classified as showing intermediate features of traits between the wood mouse and the common vole in terms of mobility and homerange size, habitat selectivity, and diet breadth (van den Brink et al. 2011). According to ecotoxicological studies where the dietary route is the predominant exposure pathway, its responses are expected to be intermediate or closer to the trends exhibited by the wood mouse than by the common vole (Baudrot et al. 2018; van den Brink et al. 2011). Although the importance of toxicokinetics on accumulation in hair cannot be ignored (Faÿs et al. 2023), our results may again suggest the involvement of different origins of the compounds and of a different contribution of the various exposure routes in shaping interspecies accumulation dissimilarities. The differences found here between species and between animals captured in different habitats within the landscape highlights the need to include both ecological and physiological features in the wildlife risk assessment process and postregistration surveys of the unintentional effects of PPP (Morrissey et al. 2023). Importantly, these results highlight the need for additional research to gain knowledge about the traits that shape the exposure and uptake of PPP in wildlife and about all the interacting mechanisms involved.

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3.5. Influence of habitat, farming practices, and proxies of treatment intensity on the detection frequency of the compounds The frequency of detection did not differ according to habitat or farming practices (Sup Info Annex 1. Table A2). Accordingly, the study of Pelosi et al. (2022), which was done within the same research project as the present study (same location of sampled plots and sampling in 2018) did not show differences in GLY detection frequency in soils sampled from cereal fields or hedgerows. However, in earthworms, GLY and AMPA were detected more frequently in cereal fields than in hedgerows. The frequency of detection of GLY, AMPA and GLUF was not significantly dependent on the proxies of treatment intensity at the plot, landscape and township scales, except in the case of GLUF sales (Sup Info Annex 1. Table A2, Fig. A3, Fig. A4). However, such an effect of a.s. sales on GLUF detection probabilities was negative, i.e. the detection probabilities were lower in townships where GLUF sales were the highest, and the differences between categories were not significant (Sup Info Annex 1. Table A2, Fig. A4). To investigate this paradoxical result further and avoid any interpretation bias, a model with GLUF sales as the only explanatory factor (i.e. without the factor "species" first in the model formula) was run, revealing an absence of significant differences (Sup Info Annex 1. Table A2, Fig. A4). Hedgerows, woodlots and fields cultivated under OF are not supposed to be targeted by pesticide treatments, but they may be unintentionally contaminated due to drift or run-off (Gandhi et al. 2021), and the home range of small mammals can overlap wooded patches, nontarget crops and surrounding cropped habitats. In agricultural topsoil samples from conventional and organic farms across Europe screened for chemicals, GLY and AMPA were among the compounds with the highest frequency of detection and the greatest concentrations. (Geissen et al. 2021). Moreover, AMPA was the residue most often detected in both conventional and organic fields, with 96% occurrence in conventional systems and 83% in organic systems (Geissen et al. 2021). AMPA and GLY were also the most commonly detected pesticides in soils under various land uses sampled over mainland France, in which 111 pesticide residues were screened (Froger et al. 2023). The detection frequency was 83% for AMPA and 70% for GLY, and the two compounds were found both in treated cultivated fields and nontargeted habitats, as they occurred both in conventional and organic farming

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plots, and in different types of land uses: arable lands, orchards, forests, grasslands, and brownfields (Froger et al. 2023). Such a global occurrence of these two compounds, including in organic fields and grasslands or forests, echoes the pervasive exposure found here in small mammals. Martinez-Haro et al. (2022) reported results contrasted with the present findings about the influence of farming practices on exposure to GLY: the prevalence was 9-22% in the gastric content of hunted Iberian hares and 45% in hares found dead in pesticide-treated areas, whereas no residues were detected in hares from organic crops. This pattern may be due to the differences in the matrices analysed, with an assessment of recent short-term exposure in the case of gastric content versus past integrated measures of exposure in the case of hair. Furthermore, the Iberian hares from conventional and organic crops were collected by hunting from October to January, while our sampling session was conducted during springtime. The authors indicated that the frequency of detection in hares progressively increased over the hunting season, being 0% and 11% in October and November and 50% in December and January (Martinez-Haro et al. 2022). The hares found dead were collected in March-April (45% positive for GLY). Finally, the intensity and extent of the GLY treatments may have been greater in our study region, and the OF and CF plots may have been geographically closer in our case. In addition to the study of GLY levels in environmental matrices such as soil or biota, more research is needed to characterize the patterns of exposure of wildlife to GLY, AMPA and GLUF and quantify the underlying processes and spatiotemporal dynamics.

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3.6. Influence of habitat, farming practices, and proxies of treatment intensity on the concentrations of the compounds

In terms of the concentrations of the compounds when quantified, the GLY concentrations differed among the small mammals captured in the different habitats: they were greater in individuals from hedgerows than in those from cereal fields (Fig. 5, Sup Info Annex 1. Table A3, Fig. A3). Considering both nondetects set to 0 and quantified concentrations, the trends were similar but not significant (Fig. 6). Such a pattern was not observed in the cases of AMPA and GLUF (Fig. 6, Sup Info Annex 1. Table A3).

This may be a confirmation of the different contributions of exposure pathways for GLY in comparison to AMPA and GLUF. This pattern may indeed be related to a contribution of overspray in addition to dietary exposure because of direct spraying and/or elevated GLY deposits on hedgerow vegetation (1) during spraying due to drift and/or (2) due to (wet) deposition (Gandhi et al. 2021). GLY is known to have a low volatility but is found in air and rainfall in intensively treated areas, at relatively high frequencies and concentrations, and is subjected to wet deposition (Chang et al. 2011). Compared with those in open habitats, contaminant deposits, including PPP, on vegetation in hedgerows are known to be enhanced by larger surfaces of interception and by "edge effect" (Davis et al. 1994; Fowler et al. 2004; Lazzaro et al. 2008; Ould-Dada et al. 2002). Mayer et al. (2020) reported for European hares (Lepus europaeus) that pesticide uptake by foraging could be 7-fold lower than uptake via overspray/oral grooming. Moreover, GLY has been found to persist in vegetation for up to one year following treatments in perennial forested areas, representing a potent dietary source of exposure for wildlife (Edge et al. 2021). In a previous study addressing the exposure of small mammals to banned and currently used PPP by residue screening in hair, which was based on animals sampled from the same study area and another region in France, we reported greater contamination (in terms of the number of compounds and concentrations) in individuals captured in hedgerows than in those captured in grasslands; however, the contamination did not differ between animals captured in hedgerows and those sampled from cereal crops (Fritsch et al. 2022). In Pelosi et al. (2022), GLY and AMPA concentrations in both soils and earthworms were, however, higher in samples from cereal fields than in those from grasslands and hedgerows. This may be an argument in favour of the contribution of processes related to overspraying in determining small mammal exposure to GLY. The role of external exposure to herbicides in small mammals and the importance of oral exposure by grooming remain to be investigated. The contribution of nondietary routes to wildlife exposure is still an overlooked field of research that needs to be addressed.

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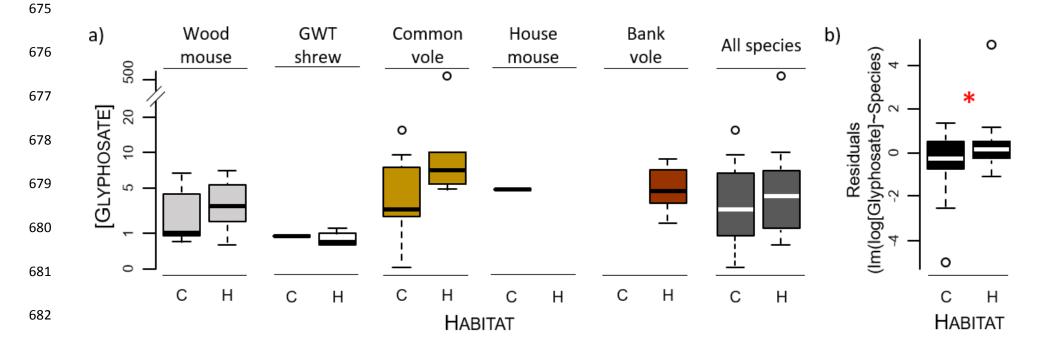
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Figure 5. Concentrations of glyphosate in the hair of small mammals without nondetects (a) according to species and habitat and (b) according to habitat taking species into account statistically in modelling.

The concentrations are expressed in pg/mg in hair. The statistical significance is depicted by a red asterisk. The habitats are represented as "C" for cereals and "H" for hedgerows. The GWT shrew stands for the greater white-toothed shrew. The sample sizes are provided in Table 1 and Table A1.



No differences in the quantified concentrations of GLY, AMPA or GLUF in the hair of small mammals were detected according to farming practices or proxies of pesticide treatment intensity at the different scales (Sup Info Annex 1. Table A3). Running the analyses on the basis of concentrations and including nondetects as 0 provided similar results (Fig. 6, Sup Info Annex 1. Fig. A3). Previously, we also highlighted the absence of differences in contamination by currently used pesticides other than GLY between small mammals captured in conventional or organic farming plots (neither the number of compounds nor their concentrations in individuals) (Fritsch et al. 2022).

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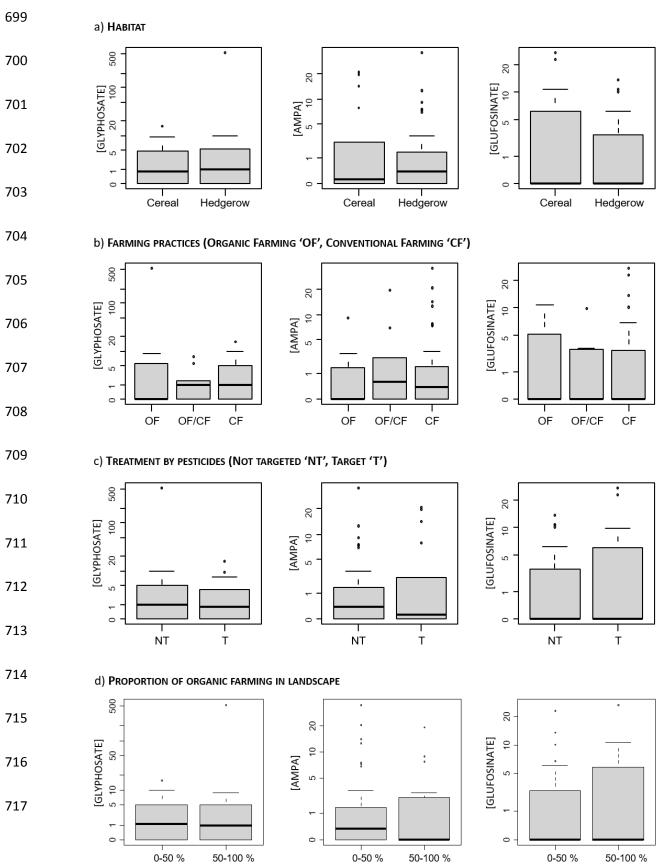
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The concentrations are expressed in pg/mg. Statistics were perfomed via the Wilcoxon-Mann-Whitney test or the Kruskal-Wallis test depending on the number of levels of the factor of concern. Posthoc multiple comparison tests after Kruskal-Wallis tests were used to investigate pairwise comparisons when relevant. No significant differences were found.



0-50 %

50-100 %

In soils and earthworms sampled within the ZAPVS, Pelosi et al. (2022) also reported no significant differences in GLY, AMPA or GLUF levels according to pesticide use (i.e. comparing treated conventional fields to nontreated habitats such as grasslands, hedgerows and organic fields) or according to farming systems in cropped fields (i.e. fields cropped under conventional or organic farming). The ubiquity of exposure to GLY, AMPA and GLUF in arable landscapes, both in crops and agroecological infrastructure, was therefore shown in recent studies for soils and for both terrestrial invertebrates and vertebrates belonging to various taxa and of different trophic levels. The transfer of GLY, AMPA and GLUF in food webs is thus questioned and requires further attention. This widespread occurrence of GLY, AMPA and GLUF in the environment and biota, regardless of the farming practices and intensity of use at the landscape scale, indicates pervasive contamination that may reflect the magnitude of repeated GLY-based (and formerly GLUF-based) herbicidal use over large surfaces in French agrosystems. On the basis of our results and recent literature, the beneficial role of agroecological infrastructures (e.g. hedgerows) and nontreated crops as refuges or recovery areas towards pesticide treatments in agroecosystems under the current agricultural model is questioned. Our findings highlight that the current surfaces of nontreated habitats within the agricultural landscapes studied are insufficient to mitigate the exposure of wildlife to GLY, AMPA and GLUF. We reached similar conclusions when we studied the exposure of small mammals to 73 residues of fungicides, herbicides (excluding GLY, AMPA and GLUF), insecticides, and some of their metabolites in Fritsch et al. (2022). With respect to the mitigation of unintentional effects of PPP on wildlife, these results call for a reassessment of the relevance and efficiency of the current procedures and a strengthening of national and regional initiatives for agri-environmental schemes. Further investigations into the pathways of wildlife exposure to PPP and responses at the individual and population levels are needed to understand and predict the role of agroecological infrastructures in shaping the direct and indirect effects of PPP. The beneficial role of hedgerows in limiting inputs of PPP on non-target plots around treated fields can be leveraged in agri-environmental schemes, but this desirable use of agroecological infrastructures to

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protect the environment and biodiversity may turn seminatural habitats into "ecotoxicological traps". These issues deserve attention and should be addressed in both future research studies and monitoring schemes.

3.7. Dose reconstruction for risk assessment

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To evaluate the risk of unintentional impacts in small mammals by comparing exposure dose values estimated from back-calculation using the concentrations measured in hair to toxicological data, we found that toxic thresholds could be exceeded for both GLY and GLUF. In the case of GLY, at least one of the five considered toxicological thresholds was exceeded according to the trigger of concern for at least one species in all of the approaches (Table 2). The potential for toxicological impairment was thus observed in all of the approaches developed. This finding suggests a risk for small mammals, especially rodents, and particularly for herbivorous voles (Table 2). Often, only the maximum value measured in Microtus voles reached toxic levels (10 times out of 16 scenarios). In five of the 16 scenarios computed via the different approaches, all of the species were found to be at risk, with maximum but also median and minimum concentrations measured being involved. At the population level, in the "best-case" scenario, toxicological effects related to GLY exposure impaired at least 6% of individuals in the common voles studied here, which represents a risk of effects in 2.1% of the rodents and in 1.6% of the small mammal community studied, including insectivorous species (Table 2). In the worst-case scenario, toxicological impairment occurred in 87.5% of the common voles, 88.9% of the voles (bank voles and common voles), 66% of all of the rodent species, 50% of the shrews and 62.3% of the individuals within the small mammal community (Table 2). Up to 93.8% of the common voles and 92.6% of the individuals of all vole species may be impacted if individuals close to the trigger value of 5 are included in the calculation (Table 2). For GLUF, the four toxicological thresholds considered were exceeded in the two scenarios for all

rodent species, including the maximum concentrations measured in our samples but also the third

quartile, median and even minimum values (Table 2). This would translate, on the basis of our

captured individuals, into a possible impairment related to GLUF exposure in 31.3 to 75% of the common voles, 25.9 to 70.4% of the vole individuals, 19.1 to 46.8% of individuals within the rodent population and 14.8 to 36.1% of individuals within the small mammal community (Table 2). This result suggests that many of the free-ranging nontargeted small mammals could be at risk for toxicological effects related to exposure to GLUF, especially herbivorous and granivorous or omnivorous species, at the time of sampling (i.e. 2018), although the use of this compound in France has largely decreased since 2015.

Thus, our findings are consistent with the first conclusions of the risk assessments performed during the registration processes, which identified potential threats for both GLY and GLUF to herbivorous, granivorous and insectivorous terrestrial vertebrates, especially herbivorous voles (EFSA 2005, 2012, 2015a). Note that the regulatory assessments were based on dietary exposure, but wildlife may be exposed to PPP - and this is especially the case for GLY when used through spraying - via additional routes such as oral uptake by grooming, inhalation, dermal contact, and/or ocular contact, as mentioned for human populations (Gandhi et al. 2021; Mayer et al. 2020; Mineau 2011).

Table 2. Summary of risk assessment for glyphosate and glufosinate based on reverse dosimetry approaches.

 Details are provided in Sup Info Annex 1 and Annex 2. Abbreviations: Nb number; Ind individuals; L-T NOAEL long-term no observed adverse effect level; S-T NOEL short-term dietary no observed effect level; Ac NOAEL acute NOAEL; Ch NOAEL chronic NOAEL; CLD chronic low dose threshold; Apsy Apodemus sylvaticus (wood mouse); Crru Crocidura russula (greater white-toothed shrew); Miar Microtus arvalis (common vole); Mumu Mus musculus (house mouse); Mygl Myodes glareolus (bank vole); Max maximum; Q3 3rd quartile; Med median; Min minimum; PK pharmacokinetics.

Compounds Approach Scenarios		Cases when Toxicity/exposure ratio exceed trigger value (of 10 for Ac toxicity data or of 5 for Ch toxicity data)													
	Type of data / Specific data / Assumptions	Nb	Threshold of concern	Species of concern	Level [C] _{hair}	Nb ind	Nb rodents	Nb shrews	% All species	% Rodents	% Voles	% Microtus	% Shrews		
Scenarios	, 100 a.m. p 110 m.	cases			or concern			(includi	ng individuals	close to th	e trigge	r*)			
GLYPHOSATE								,							
Approach 1	[GLY] in human urine and hair														
Scenario 1.1	Head hair														
Scenario 1.1.a	Urinary excretion 10%	3	L-T NOAEL, S-T NOEL, CLD	Miar	Max	1	1		2	2	4	6			
Scenario 1.1.b	Urinary excretion 20%	2	L-T NOAEL, CLD	Miar	Max	1	1		2	2	4	6			
Scenario 1.1.c	Urinary excretion 1%	16	L-T NOAEL, S-T NOEL, Ac NOAEL, Ch NOAEL, CLD	Miar, Apsy, Mumu, Mygl, Crru	Max, Q3, Med	29	25	4	48	53	74	69	29		
			,	707		(33)	(26)	(7)	(54)	(55)	(78)	(75)	(50)		
Scenario 1.2	Pubic hair														
Scenario 1.2.a	Urinary excretion 10%	12	L-T NOAEL, S-T NOEL, Ac NOAEL, Ch NOAEL, CLD	Miar, Apsy, Mygl	Max, Q3, Med	16	16		26	34	48	50			
			,			(18)	(17)	(1)	(30)	(36)	(48)	(50)	(7)		
Scenario 1.2.b	Urinary excretion 20%	4	L-T NOAEL, S-T NOEL, Ac NOAEL, CLD	Miar	Max	1	1		2	2		6			
Scenario 1.2.c	Urinary excretion 1%	31	L-T NOAEL, S-T NOEL, Ac NOAEL, Ch NOAEL, CLD	Miar, Apsy, Mumu, Mygl, Crru	Max, Q3, Med, Min	37	30	7	61	79	85	81	50		
Approach 2	Body burden in small mammals 7 days after dose														
Approach 2			L-T NOAEL, S-T NOEL, Ac												
Scenario 2.1	Proportions hair/body burden	4	NOAEL, CLD	Miar	Max	1	1		2	2		6			
Scenario 2.2	$[C]_{\text{whole body}} = [C]_{\text{hair}}$	4	L-T NOAEL, S-T NOEL, Ac NOAEL, CLD	Miar	Max	1	1		2	2		6			

794 Table 2. continued

			Cases when To	oxicity/exposure ratio	exceed trigge	r value	(of 10 for A	Ac toxicity	data or of 5 f	or Ch toxici	ity data)		
Compounds Approach Scenarii	Type of data / Specific data / Assumptions	Nb cases	Threshold of concern	Species of concern	Level [C] _{hair}	Nb ind	Nb rodents	Nb shrews	% All species	% Rodents	% Voles	% Microtus	% Shrews
	•	cases			or concern			(including individuals close to the trigger*)					
GLYPHOSATE													
Approach 3	Toxicokinetics in small mammal blood												
Scenario 3.1	Anadon et al. (2009)												
Scenario 3.1.a	GLY residues	4 L-T NOAEL, S-T NOEL, AC NOAEL, CLD		Miar	Max	1	1		2	2		6	
Scenario 3.1.b	AMPA residues	16	L-T NOAEL, S-T NOEL, CLD	Miar, Apsy, Mumu, Mygl, Crru	Max, Q3, Med	24	23	1	39	49	74	63	7
Scenario 3.2	Kim et al. (2023) GLY residues	1	CLD	Miar	Max	1	1		2	2		6	
Approach 4	Residues in viscera and body of small mammals												
Scenario 4.1	Min ratio viscera/body residues	1	CLD	Miar	Max	1	1		2	2		6	
Scenario 4.2	Average ratio viscera/body residues	1	CLD	Miar	Max	1	1		2	2		6	
						(2)	(2)		(3)	(4)	(7)	(13)	
Scenario 4.3	Max ratio viscera/body residues	2	L-T NOAEL, CLD	Miar	Max	1	1		2	2		6	
Approach 5	Prediction of incorporation in hair from PK												
Scenario 5.1	Glyphosate residues	20	L-T NOAEL, S-T NOEL, Ac	Miar, Apsy, Mumu,	Max, Q3,	38	31	7	62	66	89	88	50
			NOAEL, Ch NOAEL, CLD	Mygl, Crru	Med, Min								
				N.4: A N.4	Ma.: 03	(39)	(32)	(7)	(64)	(68)	(93)	(94)	(50)
Scenario 5.2	AMPA residues	20	L-T NOAEL, S-T NOEL, CLD	Miar, Apsy, Mumu, Mygl, Crru	Max, Q3, Med, Min	30	28	2	49	60	85	81	14
				iviygi, cira	ivica, iviiii	(31)	(29)	(2)	(51)	(62)	(89)	(88)	(14)
GLUFOSINATE													
Approach 1	Toxicokinetics in small mammal blood												
Scenario 1.1	values in males	18	L-T NOAEL, S-T NOEL, Acute LD50, Chronic NOAEL	Miar, Apsy, Mumu, Mygl	Max, Q3, Med, Min	22	22		36	47	70	75	
						(23)	(22)	(1)	(38)	(47)	(70)	(75)	(7)
			L-T NOAEL, S-T NOEL,	Miar, Apsy, Mumu,	Max, Q3,								
Scenario 1.2	values in females	10	Acute LD50, Chronic	Mygl	Med	9	9		15	19	26	31	
			NOAEL			(10)	(10)		(16)	(21)	(30)	(31)	
						. ,					• •		

^{*}Values considered close to the trigger: less than 1 of difference, e.g. value of 5.2 for a trigger of 5

Importantly, in the study on small mammals carried out in the same area and mentioned above (Fritsch et al. 2022), 112 different residues of banned and currently used PPP (parent chemicals and metabolites) were detected in hair samples from wood mice and greater white-toothed shrews, with 32 to 65 residues detected per individual. This means that "cocktail effects" are likely to occur, which can ultimately increase the risk of GLY- or GLUF-induced toxicity assessed here owing to mechanisms of additivity and synergy (Martin et al. 2021). In their review, Martin et al. (2021) reported that additivity was the interaction that most often occurred in chemical mixtures, followed by synergistic interactions. For example, Brodeur et al. (2014) reported synergic toxic impacts in the common South American toad (Rhinella arenarum) of binary mixtures of GLY-and cypermethrin-based commercial PPP products; two compounds that have been detected in more than half of the small mammal individuals in our studies. They repoted that the degree of synergy ranged from 2 to 9 times depending on the mixture tested (Brodeur et al. 2014). From terrestrial wildlife and biodiversity conservation perspectives, our findings may be worrisome. As mentioned earlier, GLY concentrations were found at higher levels in bats than in this study (Hooper et al. 2022). The authors emphasized that GLY reached the highest concentrations measured in their study, where 8 compounds, including neonicotinoids and several systemic or selective herbicides, were analysed. These authors suggested that extremely high environmental inputs of GLY may counteract the role of its short half-life in mitigating risks to wildlife (Hooper et al. 2022). A growing body of evidence raises concerns about the lethal and sublethal toxic effects of GLY in biota and highlights endocrine disruption and reproductive impairment in vertebrates chronically exposed to environmental concentrations (Fritsch et al., 2024). Recent studies have shown the toxic effects of GLUF at environmentally relevant concentrations and suggest that it has deleterious consequences on biota health under chronic exposure scenarios (Santovito et al., 2024). For example, genomic instability and cellular damage in vitro in cultured human lymphocytes and in vivo in Lymnaea stagnalis haemocytes following low-dose exposure to glufosinate-ammonium have been reported, as have negative impacts on reproductive health in L. stagnalis (Santovito et al., 2024). Furthermore,

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some wild taxa may be highly sensitive to GLY, AMPA or GLUF, more than expected from laboratory studies on models such as rats and mice, and commercial formulations can be more toxic than a.s. alone (Fritsch et al., 2024). For example, the mortality rate of postmetamorphosis juveniles of the species Rana sylvatica, Bufo woodhousii fowleri, and Hyla versicolor reached 68-86% when they were exposed via spraying to a GLY-based commercial herbicide at the recommended dose (Relyea 2005). The exposure of spined toads (Bufo spinosus) to AMPA at environmental concentrations affects hatchling and tadpole morphology and decreases embryonic and tadpole survival (Cheron and Brischoux, 2020, 2023). Risks of indirect effects of GLY on wildlife via disturbances in natural habitats and dietary resources due to effects on nontarget plants and arthropods [from offsite spray drift] have been noted in risk assessments (EFSA 2015c; US EPA 2020). Following GLY treatments, altered habitat and/or food resources have been associated with reduced (breeding) populations and negative effects on reproduction in birds (Lazaran et al. 2013; Ojelade et al. 2022), ra eduction in the frequentation of elk populations (Milner et al. 2013), and a reduced abundance of small mammals (especially insectivores Soricidae and herbivores Microtinae) (Santillo et al. 1989). Our results show a pervasive exposure, and they question the magnitude of positive compensatory effects (i.e. refuges and connectivity recolonization) expected from untreated areas within the agricultural landscape mosaic (see, for instance, Dalkvist et al. (2013) about the role of landscape features in the impacts of pesticides on vole population dynamics). Moreover, recurring treatments over seasons and years and use over large surfaces may lead to pseudopersistence within the environment (Hvězdová et al. 2018) and thus likely limit the possibility for recovery. Indirect effects may thus occur in addition to direct toxic effects. Our findings warrant further research to confirm their interpretation and gather knowledge about the ecological impacts of the use of GLY and GLUF as PPP. There is a need to characterize the toxicokinetics and toxicodynamics of these compounds in wild species better and to provide data to

relate concentrations in hair with exposure dose, accumulation in other tissues, and toxicological

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responses. Furthermore, more research is needed to assess whether the potential risks computed here actually translate into deleterious effects in free-ranging populations, which would require field surveys and monitoring of wildlife exposure and health. The dynamics of GLY, AMPA and GLUF transfer and their impacts on food webs crucially need to be investigated. Here, we provide here lines of evidence to support the identification of the organophosphate chemicals GLY and GLUF as 'emerging organic contaminants', as emphasized by Kissane and Shephard (2017). Given the pervasive exposure of wildlife to GLY, AMPA and GLUF and suggesting chronic exposure at sublethal levels on the basis of nondestructive sampling in biosentinel species, our study highlights the need for research to fill gaps in knowledge about early warning signals of impacts as emphasized by Kissane and Shephard (2017).

5. Conclusion

Our results provide new insights into the ecological risks of GLY- and GLUF-based herbicides, showing that in addition to the evidence related to environmental and human health issues, these PPP are of concern for wildlife exposure and may be of concern for impacts and further trophic cascading effects. The exposure of wild small mammals to GLY, AMPA and GLUF was shown to be widespread in all trophic groups or taxa and in treated and nontreated habitats within the studied agricultural area and was unrelated to the intensity of the treatments at the plot or landscape scale. These results support the most recent findings concerning pervasive GLY environmental contamination, occurrence in food webs, and associated ecological risks. Our results warrant further research on the ecological safety of the use of GLY and GLUF as PPP and provide support for assessing their potential as 'emerging organic contaminants'.

Our study relied on measurements of residues in hair samples, a method that is developing in human epidemiology, and looks a promising nondestructive approach to study the field ecotoxicology of PPP and monitor wildlife exposure or detect early signals in wild species. Although this approach should be strengthened by further assessment of exposure doses and measurements of effects in animals,

such surveys may represent valuable opportunities for the assessment of regulatory measures and postregistration monitoring and provide support for phytopharmacovigilance and toxicovigilance schemes (Vijver et al., 2017). For example, the presence of pesticides in the hair of small mammals reported in Fritsch et al. (2022) was reported as an exposure warning for nontarget species by the French department of the agency ANSES in charge of phytopharmacovigilance.

This study highlights the need for research on the pathways of wildlife exposure to PPP and the toxicokinetics and toxicodynamics of GLY, AMPA and GLUF and, more broadly, of PPP initially assessed as not persistent and not bioaccumulative on the basis of criteria established on legacy compounds. Our findings strongly suggest that current agri-environmental schemes do achieve the objectives of ecological risk mitigation. They call for additional research and monitoring of the role of agroecological infrastructure in shaping PPP impacts on biodiversity and provide support for an overall reflection, if not an overhaul, of agroenvironmental policies.

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898 899	Clémentine Fritsch: Conceptualization, Funding acquisition, Project administration, Supervision, Investigation, Data curation, Formal analysis, Validation, Visualization, Writing - original draft
900 901	Brice Appenzeller: Project administration, Investigation, Methodology; Supervision, Validation, Writing - review & editing
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917	Data accessibility
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919	Supplementary information
920	List of information in the appendix document:
921	Annex 1.
922	Text A1: Molecular identification of small mammal species
923 924	Text A2: Reagents and procedures used for the chemical analyses of glyphosate, AMPA and glufosinate
925 926	Text A3: Approaches of reverse dosimetry to assess the risk for deleterious effects in small mammals
927	Text A4: Statistical outputs of the Tarone-Ware tests

928 929	Figure A1. Correlations between the concentrations of glyphosate, AMPA, and glufosinate in small mammal hair.
930	Figure A2. Simplified phylogenetic tree of the rodents and shrews included in the study.
931 932 933 934	Figure A3. Concentrations of glyphosate, AMPA, and glufosinate in the hair of small mammals with nondetects set to 0 according to the proportion of organic farming in the landscape as a continuous factor (a) and to the proxy of treatment intensity at township scale as a continuous factor (b) and as a categorical factor (c).
935 936	Figure A4. Probabilities of detection of glufosinate in hair of small mammals according to the sales of the active substance at the township scale.
937	Table A.1. Overview of sample size.
938 939 940	Table A.2. Statistical outputs of binomial GLMs used to investigate the influence of explanatory factors on the detection frequencies of glyphosate, AMPA, and glufosinate in the hair of small mammals.
941 942 943	Table A.3. Statistical outputs of the GLMs used to investigate the influence of explanatory factors on the concentrations of glyphosate, AMPA, and glufosinate in the hair of small mammals.
944	Annex 2.
945	Computation of reverse dosimetry approaches (calculation spreadsheets).
946	computation of reverse dosiment, approaches (calculation spreadsheets).
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Appendix - Annex 1

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Text A1: Molecular identification of small mammal species

Total genomic DNA was extracted from 10-25 mg of ear tissue using the DNeasy Blood and Tissue kit (Qiagen, Courtaboeuf, France) according to the manufacturer's recommendations (pp 28-30, version 07/2006). Each sample was processed independently in an automated manner using the QIAcube robot (Qiagen). The DNA concentration was then measured using a NanoVue Plus spectrophotometer (Biochrom). The DNA extracts were stored at -20 °C until DNA amplification.

All of the DNA extracts were then amplified into an approximately 900 bp-long fragment (excluding primers) of the *cytochrome* b gene. We used the primers CytB Uni fw 5' – TCATCMTGATGAAAYTTYGG – 3' and CytB Uni rev 5' – ACTGGYTGDCCBCCRATTCA – 3' published in Schlegel et al. (2011). Amplifications were performed in 50 μ L reactions containing $1\times$ HotStart Taq Master Mix (Qiagen), 0.32 μ M of each primer, 0.5 ng/μ L of bovine serum albumin, 0.75 mM of MgCl₂, and 5 μ L of extracted DNA (10-70 ng/μ L). The PCR program consisted of an activation step of 15 mn at 95 °C, followed by 40 cycles of denaturation at 94 °C during 30 s, annealing at 53 °C during 45 s, and extension at 72 °C during 1 min. A final extension was performed at 72 °C for 10 min. Amplification reactions were performed using an Eppendorf Mastercycler DNA Engine.

The PCR products were separated and visualized using the QIAxcel device and a QIAxcel DNA Screening kit (Qiagen). The amplified products were purified using the Illustra GFX PCR DNA and Gel Band Purification Kit (GE Healthcare) according to the manufacturer's instructions. Direct sequencing of the PCR products was performed with an automated sequencer (Applied Biosystems Seqstudio Genetic Analyzer). All of the samples were sequenced with the primers employed for the PCR reactions. The obtained DNA sequences were submitted to GenBank via the BLAST algorithm (http://blast.ncbi.nlm.nih.gov/Blast.cgi). A query sequence was then assigned to a given species when identity and coverage matched the database sequence with a threshold of at least 99% for only one species.

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Text A2: Reagents and procedures used for the chemical analyses of glyphosate, AMPA and glufosinate

Chemicals and Reagents

Methanol Pesti-S grade, acetonitrile, ammonium acetate and water ULC-MS grade were purchased from Biosolve (Dieuze, France). Sodium dodecyl sulfate, FMOC chloride, sodium tetraborate decahydrate and glyphosate, glufosinate ammonium, aminomethylphosphonic acid and glyphosate-1,2-13C2,15N standards were purchased from Sigma Aldrich (Diegem, Belgium).

Sample preparation

To differentiate compounds biologically incorporated into hair from chemicals externally deposited on the hair surface due to contaminated air or dust, the samples were decontaminated with sodium dodecyl sulfate (SDS) solution and methanol (Duca et al. 2014). This procedure was developed to be as "universal" as possible, and was tested on a large panel of chemicals with different physicochemical properties including rather hydrophilic chemicals. The two steps (SDS and methanol) were found to be the best compromise to remove both lipophilic and hydrophilic chemicals from hair surface. Importantly, SDS solution being an aqueous solution (5% SDS / 95% water), it is therefore well adapted to remove hydrophilic compounds such as GLU, GLY and AMPA.

Note that in other studies where hair samples were washed prior to analyses of GLY, Hooper et al. (2022) decontaminated hair of bats using ultrapure (MilliQ) water and subsequently isopropanol, and Alvarez et al. (2022) used dichloromethane twice to wash human hair samples.

Analysis of glyphosate, glufosinate and AMPA in hair

After decontamination, the samples (50 mg) were spiked with 5 μ L of internal standard solution at 1 mg/L, and 1 mL of water was added before overnight extraction at 40 °C with 350 rpm agitation. The supernatants were evaporated to dryness at 50 °C under N₂ flow, and 100 μ L of borate buffer and FMOC-Cl were added before derivatization and injection into the LC-MS/MS Waters system Acquity UPLC H-class system equipped with a Waters Acquity BEH C18 column (100 × 2.1 mm, 1.7 μ m particle

size) and with a binary mobile phase consisting of ammonium acetate 10mM in water (A) and acetonitrile (B). The flow rate was initially set at 0.4mL/min with 98% A and 2% B and reached 2% A and 98 B after 6 min. From 6.0 to 9min the column was re-equilibrated to 98% A and 2% B. The UPLC system was coupled to Waters Xevo TQ-S triple quadrupole mass spectrometer. With each analytical run, quality controls consisting of hair samples supplemented at several concentrations (0.1, 0.2, 0.5, 1, 2, 5, 10, 20, 50, and 100 pg/mg) were used to control for possible sensitivity drift, and a blank sample (sample without the target chemicals) was used to verify possible cross-contamination. Since field samples were not available in sufficient amount to conduct method validation, it has to be conducted on blank samples from laboratory animals supplemented with standards. The slopes of the calibration curves were 13.56, 6.31 and 10.68 for GLU, GLY and AMPA respectively. The corresponding R^2 values were all > 0.998. The concentrations are expressed as pg/mg hair. The limits of quantification (LOQ) and lowest detected values (Minimum) are presented in Table 1. The LOQ was determined as the lowest concentration level presenting variability and accuracy < 25%. The estimation of the limit of detection (LOD, the lowest concentration of a substance that can be reliably detected by the analytical method) can be done through various experimental and computational approaches, and choices must be made according to the type and the objective of the analytical method (Evard et al. 2016b, 2016a). We did not apply specific experiments and calculations to compute the LOD in this study owing to several reasons: (1) the purpose of the study was not focused on an analytical method development and validation; (2) high performance and resolution of current instrumentation and modes hamper the use of methods based on signal-to-noise to estimate the LOD; (3) the analyses were not run within a framework requiring a specific guideline to be followed; and (4) no critical decision has to be based on the LOD. As described above, the analytics were based on up-todate protocols and materials and followed the general principles of metrology (such as inclusion of blank samples and over 5 calibration or quality control levels, among others). The accuracy and sensivity of the method were not fully computed but the inclusion of blanks and pretty low

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concentrations in standards (i.e. 0.1 and 0.2 pg/mg) limits the potential for bias towards false positive. All values above the lowest analytical detected value were included as "detects" contrary to "nondetects". Importantly, only one concentration was reported below the lowest concentration in quality controls (the minimum GLY concentration, see Table 1). All reported concentration values, involving values between the LOQ and the lowest detected value, were included as continuous data. The use of concentrations below the LOQ has been shown to be a suitable method preferable to the absence of data, which shows better efficiency in terms of bias and precision than several other established methods (Keizer et al. 2015).

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Text A3: Approaches of reverse dosimetry to assess the risk for deleterious effects in small mammals

In this part of the work, reverse dosimetry approaches have been applied to estimate the daily dose intake of small mammals from the concentrations of glyphosate (GLY), AMPA or glufosinate (GLUF) measured in hair, and several types of data that may be useful for applying dose reconstruction processes have been used. Concentrations in mammal body fluids and hair, residues in viscera and body burden in small mammals, and toxicokinetics in small mammal blood have been collected from the literature. Several approaches of back-calculations were thus applied depending on the type of input data and the different approaches were computed under several assumptions in cases with a lack of available data. For GLY, four different approaches have been applied on the basis of data about (1) concentrations in human urine and hair (Alvarez et al. 2022), (2) body burden in rats 7 days after dose administration (Brewster, 1991), (3) toxicokinetics in rat blood (Anadón et al. 2009; Kim et al. 2023), (4) residues of GLY in the viscera and body of wild small mammals in a field study (Newton et al. 1984), and (5) equations for fast-elimination chemicals that relate concentrations in hair in rats and the level of exposure, taking pharmacokinetics into account (Faÿs et al. 2023). For GLUF, one approach could be proposed on the basis of toxicokinetics in rat blood (Joint FAO/WHO Meeting on Pesticide Residues (JMPR) et al. 2012). The body weights of small mammals were obtained from actual morphometrics measured in our experiment by weighing individuals in which residues of GLY, AMPA and GLUF were measured (Sup Info Annex 2). Data concerning urinary excretion percentage of GLY were obtained in documents released by the EFSA and recent reviews (Connolly et al. 2020; EFSA 2015a; Peillex and Pelletier 2020). The concentrations of GLY in the viscera and body reported in Newton et al. (1984) for shrews (carnivores), deermice (omnivores) and voles (herbivores) were used separately according

to diet preference to compute reverse dosimetry in shrews (carnivores), wood mice, house mice and

bank voles (omnivores), and common voles (herbivores), respectively. The food intake of the different species was computed using the mean food intake of the different items provided by Crocker et al. (2002). The mean food intake of arthropods and cereal seeds provided for the wood mouse has been used for the wood mouse and the house mouse; the mean food intake of arthropods and earthworms provided for the common shrew has been used for the greater whitetoothed shrew; and the mean food intake of grasses and forbs provided for the field vole has been used for the common vole (Crocker et al. 2002). The assumptions that had to be made to allow the calculations were that the distribution was equivalent between body parts and hair, that the concentration in hair was equivalent to the concentration in plasma/blood and proportional to the administered dose, and that the concentration in viscera reflected the concentration in the diet. These assumptions, if not verified, might, however, be considered acceptable according to current knowledge about GLY pharmacokinetics. It is known that "absorbed GLY is poorly metabolised, widely distributed in the body, does not undergo enterohepatic circulation and is rapidly eliminated" (EFSA 2015a). Like GLY, GLUF is distributed ubiquitously in the body rapidly after administration and is mostly excreted in faeces and urine (Joint FAO/WHO Meeting on Pesticide Residues (JMPR) et al. 2012). Chemicals are mostly incorporated into living hair bulb cells from the bloodstream, and residue concentrations in hair are representative of the internal dose during the time of hair sample growth (Appenzeller et al. 2017; Faÿs et al. 2023). In a field study where GLY was aerially applied at 3.3 kg a.s./ha in a forest, the concentrations measured in the remainder of small mammal bodies (without viscera) were of the same order of magnitude as the maximum concentrations measured here in hair; owing that in the EU the maximum field application of GLY in cropped fields is comparable (4.32 g a.s./ha) (Newton et al. 1984). Newton et al. (1984) considered concentrations in viscera as reflecting intake and reported that "GLY in the viscera of herbivores was roughly equal to or somewhat below concentrations found in litter and ground cover". The dose of GLY was compared with five toxicological thresholds. Three of these thresholds were

regulatory thresholds: the short-term dietary NOEL (no observed effect level, 150 mg active

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substance/kg body weight), the chronic 21 day NOAEL, which has the same value as the reproductive NOAEL (no observed adverse effect level, 351 mg a.s./kg bw/d), and the long-term NOAEL (maternal and developmental NOAEL, which is the same than long-term NOAEL for terrestrial vertebrates, 50 mg a.s./kg bw/d) (EFSA, 2015a; PPDB, http://sitem.herts.ac.uk/). The No Mortality Dose to the Oregon vole (NOAEL Acute toxicity (Intraperitoneal), 450 mg a.s./kg bw) was also used to consider a wild species (McComb et al. 2008). Some more recent publications highlighted that the harmful effects of GLY were detected below regulatory limits (e.g. within the spectrum of acceptable daily intake) (Cai et al. 2017; Clair et al. 2012; Mesnage et al. 2015). We therefore added a "chronic low-dose" limit, with a value of 5 mg a.s./kg bw/d obtained from studies showing effects on reproductive development, endocrine disruption, maternal behaviour, neuroplasticity and the gut microbiota in rats (Dechartres et al. 2019; Romano et al. 2010).

The dose of GLUF was compared to four available toxicological thresholds: the acute oral LD50 (median lethal dose; 416 mg/kg bw), the short-term dietary NOEL (64 mg a.s./kg bw), the chronic 21 days NOAEL (6.3 mg a.s./kg bw/d) and the long-term NOAEL (2 mg a.s./kg bw/d) (Joint FAO/WHO Meeting on Pesticide Residues (JMPR) et al. 2012; PPDB, http://sitem.herts.ac.uk/).

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1390 *Text A4: Statistical outputs of the Tarone-Ware tests*

```
1391
          twoSampleLinearRankTestCensored(x
                                             =
                                                  TAB$GLY,
                                                              x.censored
1392
      TAB$GLY.censored,
1393
                                                   y = TAB\$AMPA, y.censored =
1394
      TAB$AMPA.censored,
1395
                                     test = "tarone-ware")
1396
      Results of Hypothesis Test
1397
      Based on Censored Data
1398
      _____
1399
      Null Hypothesis:
                                     Fy(t) = Fx(t)
      Alternative Hypothesis:
1400
                                     Fy(t) != Fx(t) for at least one t
1401
      Test Name:
                                     Two-Sample Linear Rank Test:
1402
                                     Tarone-Ware Test
1403
                                     with Hypergeometric Variance
1404
      Censoring Side:
                                     left
1405
      Censoring Level(s):
                                    x = 0.018
                                    y = 0.24
1406
1407
      Data:
                                     x = TAB\$GLY
1408
                                    y = TAB$AMPA
1409
                                    x = TAB\$GLY.censored
      Censoring Variable:
1410
                                    y = TAB\$AMPA.censored
1411
                                    nx = 61
     Sample Sizes:
1412
                                    ny = 61
1413
     Percent Censored:
                                    x = 36.1%
1414
                                    y = 49.2%
1415
     Test Statistics:
                                    nu = 58.055344
1416
                                    var.nu = 1504.925791
1417
                                               1.496527
                                    z =
1418
                                     0.1345163
     P-value:
1419
1420
1421
1422
          twoSampleLinearRankTestCensored(x = TAB$GLY, x.censored =
1423
     TAB$GLY.censored,
1424
                                                   y = TAB$GLUF, y.censored =
1425
     TAB$GLUF.censored,
1426
                                     test = "tarone-ware")
1427
     Results of Hypothesis Test
1428
     Based on Censored Data
1429
      _____
1430
     Null Hypothesis:
                                    Fy(t) = Fx(t)
                                Fy(t) != Fx(t) for at least one t
Two-Sample Linear Rank Test:
1431
     Alternative Hypothesis:
1432
      Test Name:
1433
                                    Tarone-Ware Test
1434
                                     with Hypergeometric Variance
1435
                                    left
     Censoring Side:
                                    x = 0.018
1436
      Censoring Level(s):
                                     y = 1.161
1437
                                     x = TAB\$GLY
1438
     Data:
                                    y = TAB$GLUF
x = TAB$GLY.censored
1439
1440
      Censoring Variable:
                                    y = TAB$GLUF.censored
nx = 61
1441
1442
      Sample Sizes:
1443
                                     ny = 61
1444
                                     x = 36.1%
      Percent Censored:
                                    y = 55.7%
1445
                                     nu = 13.9088437
1446
      Test Statistics:
1447
                                     var.nu = 1304.2735588
```

```
z = 0.3851294
1448
1449
     P-value:
                                     0.7001415
1450
1451
1452
      > twoSampleLinearRankTestCensored(x = TAB$AMPA, x.censored
1453
      TAB$AMPA.censored,
1454
                                                  y = TAB$GLUF, y.censored =
1455
      TAB$GLUF.censored,
1456
                                    test = "tarone-ware")
1457
1458
      Results of Hypothesis Test
1459
      Based on Censored Data
1460
     _____
1461
     Null Hypothesis:
                                    Fy(t) = Fx(t)
1462
                                    Fy(t) != Fx(t) for at least one t
      Alternative Hypothesis:
1463
     Test Name:
                                    Two-Sample Linear Rank Test:
1464
                                    Tarone-Ware Test
1465
                                    with Hypergeometric Variance
1466
      Censoring Side:
                                    left
1467
                                    x = 0.24
      Censoring Level(s):
                                    y = 1.161
1468
1469
                                    x = TAB\$AMPA
     Data:
                                    y = TAB$GLUF
1470
                                   x = TAB\$AMPA.censored
1471
     Censoring Variable:
                                    y = TAB$GLUF.censored
1472
1473
     Sample Sizes:
                                    nx = 61
                                    ny = 61
1474
                                    x = 49.2\%
1475
     Percent Censored:
                                    y = 55.7%
1476
                                    nu = -33.2689131
1477
     Test Statistics:
1478
                                    var.nu = 1197.6199535
1479
                                    z = -0.9613446
1480
                                    0.3363789
     P-value:
1481
1482
```

Figure A1. Correlations between the concentrations of glyphosate, AMPA, and glufosinate in small mammal hair.

The concentrations are expressed in pg/mg. Nondetects are set to 0.



1483

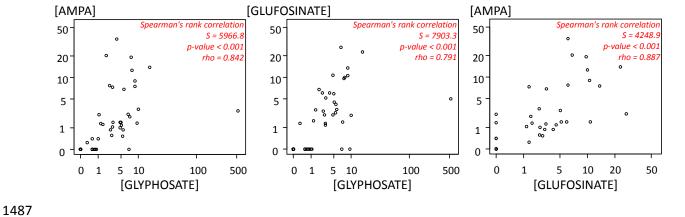
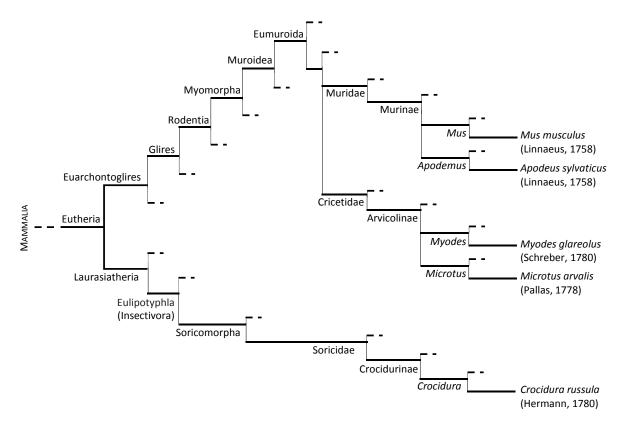


Figure A2. Simplified phylogenetic tree of the rodents and shrews included in the study.

Adapted from (Brace et al. 2016; Springer 2004; Swanson et al. 2019).



References

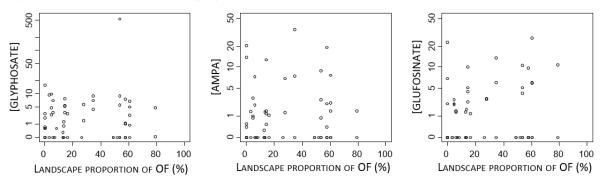
Brace S, Thomas JA, Dalén L, Burger J, MacPhee RDE, Barnes I, et al. 2016. Evolutionary History of the Nesophontidae, the Last Unplaced Recent Mammal Family. Mol Biol Evol 33:3095–3103; doi:10.1093/molbev/msw186.

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Swanson MT, Oliveros CH, Esselstyn JA. 2019. A phylogenomic rodent tree reveals the repeated evolution of masseter architectures. Proc R Soc B 286:20190672; doi:10.1098/rspb.2019.0672.

The concentrations are expressed in pg/mg. Nondetects are set to 0. Statistics were perfomed via the Wilcoxon-Mann-Whitney test or the Kruskal-Wallis test depending on the number of levels of the factor of concern and Spearman's rank correlation test for continuous factors. No significant differences were found.

a) PROPORTION OF ORGANIC FARMING (OF) IN LANDSCAPE as a continuous factor



b) SALES OF ACTIVE SUBSTANCE IN THE TOWNSHIP as a continuous factor and by category

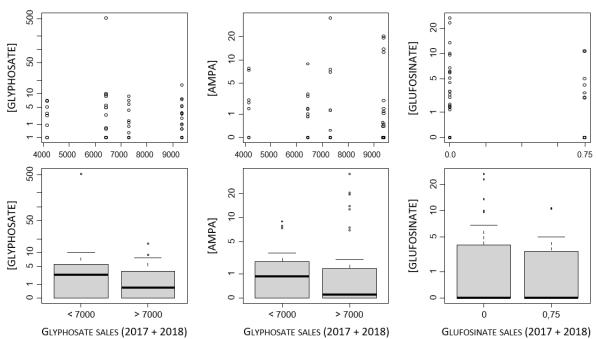


Figure A4. Probabilities of detection of glufosinate in the hair of small mammals according to the sales of the active substance at the township scale.

The predicted probabilities of detection were obtained from the statistical binomial models "Detection ~ species + sales" (left panel) and "Detection ~ sales" (right panel).

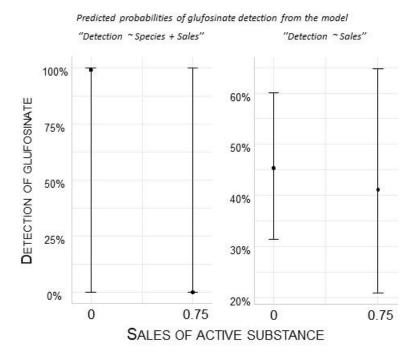


Table A1. Overview of sample size.

Abbreviations: OF=Organic farming, CF=Conventional farming, NT=Not targeted by treatments, T=Targeted by treatments, AS=Active substance.

Species Habitat Farming	Wood mouse Apodemus sylvaticus	Greater white- toothed shrew Crocidura russula	Common vole Microtus arvalis	Bank vole Myodes glareolus	House mouse Mus musculus	Total
Cereal						
CF	5	2	7		1	15
OF/CF	4		1			5
OF			1		1	2
Total	9	2	9		2	22
Hedgerow or woodlot						
CF	7	7	2	10		26
OF/CF		3	1	1		5
OF	2	2	4			8
Total	9	12	7	11		39
Total CF	12	9	9	10	1	41
Total OF/CF	4	3	2	1		10
Total OF	2	2	5		1	10
Total « NT »	9	12	8	11	1	41
Total « T »	9	2	8		1	20
Landscape OF pro	nortion					
0-50%	11	9	8	11	1	40
50-100%	7	5	8		1	21
AS sales GLY						
< 7000	6	4	11	2	2	25
> 7000	12	10	5	9		36
AS sales GLUF						
0	15	11	7	11		44
0.75	3	3	9		2	17
Total	18	14	16	11	2	61

Table A2. Statistical outputs of binomial GLMs used to investigate the influence of explanatory factors on the detection frequencies of glyphosate, AMPA, and glufosinate in the hair of small mammals.

The binomial models were computed as "Detection ~ Species" and "Detection ~ Species + Habitat or Farming or Target treatment or Landscape OF proportion or AS sales". * alternative model with the factor "glufosinate sales" alone without the factor "species". Abbreviations: OF=Organic Farming, CF=Conventional farming, Apsy=Apodemus sylvaticus (Wood mouse), Crru=Crocidura russula (Greater white-toothed shrew), Miar=Microtus arvalis (Common vole), Mumu=Mus musculus (House mouse), Mygl=Myodes glareolus (Bank vole), NT=Not targeted, T=Targeted, AS=Active substance, p=p-value. Significant p-values are indicated in bold.

			GLYPHOSATE							AMPA							(GLUFOSINATE		
			Factor			Levels			Factor			Levels			Factor			Levels		
Factor	Levels	n	Deviance	Df	р	Odds Ratio	95% CI (lower , upper)	р	Deviance	Df	р	Odds Ratio	95% CI (lower , upper)	р	Deviance	Df	р	Odds Ratio	95% CI (lower , upper)	р
Species			20.484	56	< 0.001				32.464	56	< 0.001				39.079	56	<0.001			
	Apsy	18				Reference						Reference						Reference		
	Crru	14				2.00	(0.48, 8.76)	0.344				0.58	(0.07, 3.55)	0.571				0.62	(0.03, 7.14)	0.700
	Miar	16				30.0	(4.48,612)	0.003				24.5	(4.56, 208)	<0.001				34.7	(6.05, 321)	<0.001
	Mumu	2				2.00	(0.07 , 56.9)	0.644				3.50	(0.12 , 103)	0.411				8.00	(0.25 , 273)	0.200
	Mygl	11				20.0	(2.86 , 414)	0.010				35.0	(4.73 , 753)	0.003				80.0	(9.16 , 1999)	<0.001
Habitat			0.191	55					0.018	55	0.892				0.182	55	0.669			
	Cereal	22			0.662	Reference						Reference						Reference		
	Hedgerows	39				0.71	(0.14, 3.31)	0.663				1.12	(0.22 , 6.59)	0.893				1.50	(0.24 , 12.3)	0.673
Farming			3.273	54	0.195				3.364	54	0.186				1.496	54	0.473			
	OF	10				Reference						Reference						Reference		
	OF/CF	10				6.98	(0.57 , 217)	0.169				12.7	(0.79 , 4745)	0.102				5.83	(0.35 , 133)	0.200
	CF	41				6.96	(0.84 , 174)	0.119				6.35	(0.65 , 172)	0.161				2.49	(0.30, 27.1	0.400
Treatment	t		0.000	55	0.999				0.030	55	0.863				0.016	55	0.900			
	NT	41				Reference						Reference						Reference		
	T	20				1.00	(0.22 , 4.47)	0.999				1.15	(0.23, 5.63)	0.863				0.90	(0.14, 5.12)	0.900
Landscape	e OF		0.331	55	0.565				0.031	55	0.860				0.013	55	0.908			
	0-50%	40				Reference						Reference						Reference		
	50-100%	21				0.67	(0.16, 2.60)	0.568				1.14	(0.24, 5.18)	0.860				0.91	(0.15 , 4.88)	0.908
AS sales			0.009	55	0.925				1.257	55	0.262				5.266 (0.091)*	55 (59)*	0.022 (0.763)*			
L	Low category	25				Reference						Reference						Reference		
Н	ligh category	36				1.07	(0.26 , 4.58)	0.925				2.66	(0.51 , 21.9)	0.289				0.00 (0.84)*	NA (0.26 , 2.59)*	> 0.900 (0.800)*

Table A3. Statistical outputs of the GLMs used to investigate the influence of explanatory factors on the concentrations of glyphosate, AMPA, and glufosinate in the hair of small mammals.

The models were computed as "log(concentration) ~ Species" and "log(concentration) ~ Species + Habitat or Farming or Target treatment or Landscape OF proportion or Al sales". Abbreviations: OF=Organic Farming, CF=Conventional farming, Apsy=Apodemus sylvaticus (Wood mouse), Crru=Crocidura russula (Greater white-toothed shrew),

Miar=Microtus arvalis (Common vole), Mumu=Mus musculus (House mouse), Mygl=Myodes glareolus (Bank vole), NT=Not targeted, T=Targeted, AS=Active substance, p=p-value. Significant p-values are indicated in bold.

			GLYPHOSATE									АМРА							GLUFOSINATE							
			Factor				Mode I	Levels			Factor				Model	Levels			Factor				Model	Levels		
Factor	Levels	n	F	Df	р	R ²	R ² R ² Adj	Estimate	SE	р	F	Df	р	R ²	R ² R ² Adj	Estimate	SE	р	F	Df	р	R ²	R ² R ² Adj	Estimate	SE	р
Species			1.643	4	0.186	0.162	0.162 0.063				1.110	4	0.373	0.146	0.146 0.014				1.008	4	0.425	0.155	0.155 0.002			
	Apsy Crru Miar Mumu Mygl	6 7 15 1						Reference -0.840 0.723 0.729 0.701	0.815 0.707 1.58 0.756	0.310 0.314 0.648 0.360						Reference -0.322 1.08 0.333 1.06	1.09 0.715 1.41 0.746	0.770 0.142 0.815 0.169						Reference -1.58 -0.546 0.310 -0.819	1.06 0.657 1.06 0.670	0.150 0.415 0.773 0.235
Habitat			4.37	1	0.044	0.098	0.260 0.148				0.750	1	0.395	0.025	0.171 0.005				2.759	1	0.112	0.098	0.253 0.075			
	Cereal Hedger ows	14 25						Reference 1.20	0.572	0.044						Reference -0.521	0.602	0.395						Reference -0.716	0.431	0.112
Farming			0.017	2	0.983	0.001	0.162 0.006				0.725	2	0.495	0.049	0.195 0.000				0.087	2	0.917	0.007	0.162 0.000			
	OF OF/CF CF	5 6 28						Reference -0.184 -0.146	1.05 0.890	0.861 0.871						Reference 0.800 0.022	0.988 0.838	0.426 0.979						Reference -0.194 -0.252	0.750 0.606	0.799 0.682
Treatme	Treatment		0.232	1	0.633	0.006	0.168 0.042				0.750	1	0.395	0.025	0.171 0.005				2.759	1	0.112	0.098	0.253 0.075			
	NT T	27 12						Reference -0.289	0.600	0.633						Reference 0.521	0.602	0.395						Reference 0.716		0.112
Landscap	oe OF		0.151	1	0.700	0.004	0.166 0.039				0.754	1	0.393	0.025	0.171 0.005				3.454	1	0.077	0.119	0.274 0.101			
	0-50% 50-	27 12						Reference								Reference								Reference		
	100%	12						0.227	0.582	0.700						0.496	0.570	0.393						0.790	0.425	0.077
AS sales			0.104	1	0.749	0.003	0.165 0.038				0.595	1	0.448	0.020	0.166 0.000				0.884	1	0.358	0.034	0.189 0.000			
	category category							Reference -0.172	0.535	0.749						Reference 0.416	 0.539	0.448						Reference -0.454	0.483	0.358