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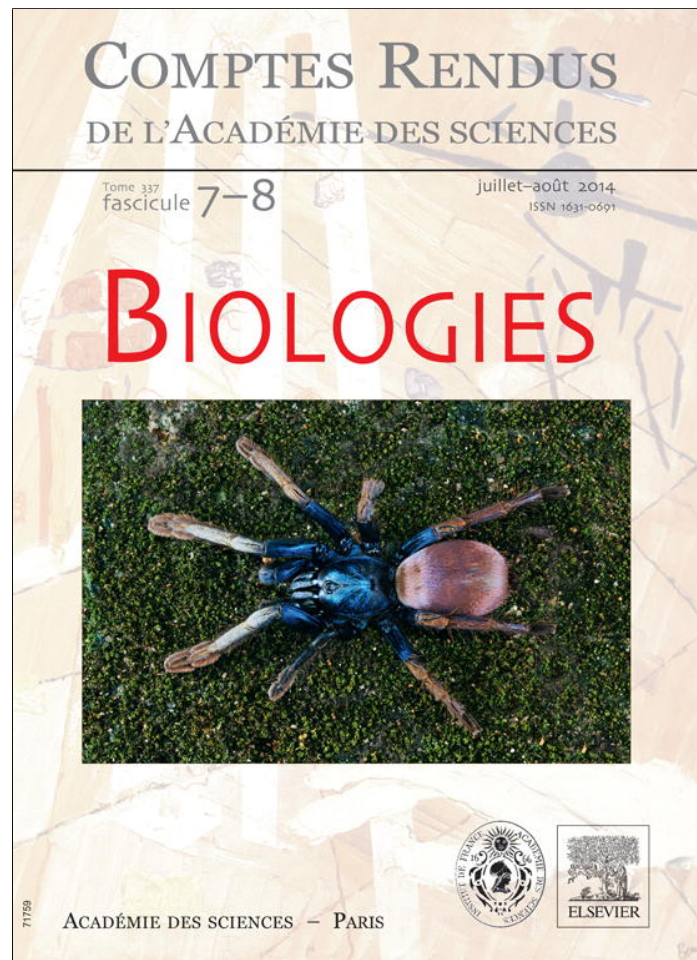
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Review/Revue

Plant-parasitic nematodes associated with olive tree (*Olea europaea* L.) with a focus on the Mediterranean Basin: A review

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ABSTRACT

The olive tree (*Olea europaea* ssp. *europaea*.) is one of the most ancient cultivated trees. It is an emblematic species owing to its ecological, economic and cultural importance, especially in the Mediterranean Basin. Plant-parasitic nematodes are major damaging pests on olive trees, mainly in nurseries. They significantly contribute to economic losses in the top-ten olive-producing countries in the world. However, the damages they induce in orchards and nurseries are specifically documented only in a few countries. This review aims to update knowledge about the olive-nematode pathosystem by: (1) updating the list of plant-parasitic nematodes associated with olive trees; (2) analysing their diversity (taxonomic level, trophic groups, dominance of taxa), which allowed us (i) to assess the richness observed in each country, and (ii) to exhibit and describe the most important taxa able to induce damages on olive trees such as: *Meloidogyne*, *Pratylenchus*, *Helicotylenchus*, *Xiphinema*, *Tylenchulus*, *Rotylenchulus*, *Heterodera* (distribution especially in the Mediterranean Basin, pathogenicity and reactions of olive trees); (3) describing some management strategies focusing on alternative control methods; (4) suggesting new approaches for controlling plant-parasitic nematodes based on the management of the diversity of their communities, which are structured by several environmental factors such as olive diversity (due to domestication of wild olive in the past, and to breeding now), cropping systems (from traditional to high-density orchards), irrigation, and terroirs.

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1. The olive tree: origins, dissemination and cultivation

Six *Olea europaea* subspecies with distinct geographic areas are distinguished [1,2] (Fig. 1):

- subsp. *europaea* present in the Mediterranean Basin, with two botanical varieties: *O. europaea* subsp. *europaea* var. *europaea* for the cultivated forms, and *O. e.* subsp.

e. var. *sylvestris* for wild and spontaneous trees (usually named oleasters) [2];

- subsp. *laperrinei* in the Saharan mountains;
- subsp. *cuspidata* from southern Africa to southern Egypt and from Arabia to China;
- subsp. *guanchica* in the Canary Islands;
- subsp. *maroccana* in southern Morocco;
- subsp. *cerasiformis* in Madeira.

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The Mediterranean olive (*Olea europaea* L. subsp. *europaea*; Oleaceae) is one of the first domesticated tree

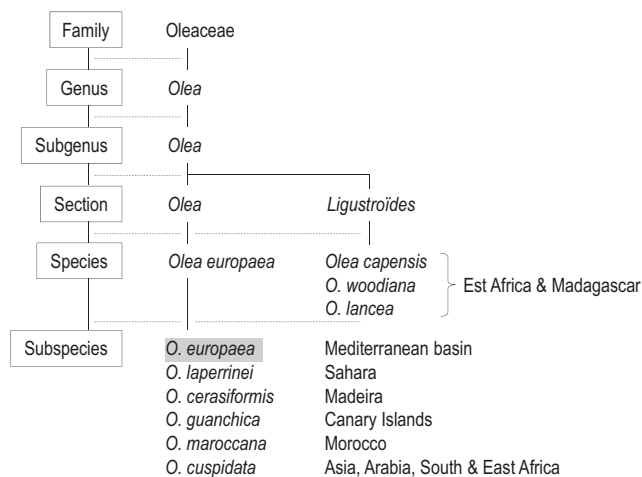


Fig. 1. Simplified diagram of the *Olea* genus (Oleaceae) and biogeography of taxa [11].

species [3,4]. Its origins and domestication history are still highly debated, but archaeological and molecular data sustain that the first cultivars originated from the Levant, very likely 6000 years ago, in a region presently located at the border between southwestern Turkey and northwestern Syria [5,6]. Genetic studies indicated also that olive domestication is a long and on-going process and cultivars have originated from multiple source populations, as for grape and figs [7–9]. It is currently established that western wild olives secondarily contributed to the cultivated gene pool, while most of present cultivars remain mainly related to eastern wild olives [10].

The genetic diversity of oleaster is somewhat greater than that of the cultivated olive [6,10,11]. This diversity is highly structured between the East and the West of the Mediterranean Basin [6,12,13]. The analysis of plastid DNA polymorphisms allowed the identification of 48 distinct profiles delineating three diverging lineages. Lineage E1 is currently distributed over the whole Basin, whereas lineages E2 and E3 exclusively occur in western and central Mediterranean areas [6,14].

The olive tree has long been considered as the most typical Mediterranean tree. It has been widely disseminated through conquests and exchanges that accompanied the expansion of the Mediterranean civilizations (Phoenicians, Greeks, Romans, and Arabs) [15]. Populations of wild olives are restricted to a few isolated areas of native Mediterranean forests, where pollen/stones may be spread by natural factors (wind and birds) [13]. Most other wild-looking forms of olive may include feral forms (either cultivated olive abandoned and becoming wild or olive from stones of cultivated olive spread by birds). The olive dissemination was accompanied by a secondary diversification that has led to a high genetic variability that reflects a wide range of morphological (tree vigour, fruit shape, yield, oil content, etc.) and physiological (adaptation traits to environmental stresses, particularly those related to climate and soil constraints) characteristics that can explain its large distribution in the Mediterranean Basin [16,17]. This diversity can also result from a series of events related to human practices made to

meet microclimate and terroir requirements and to satisfy consumer taste [12].

More than 2000 Mediterranean varieties have been recognised based on fruit, pit, and leaf shapes and colours, tree architecture and phenology (i.e. flowering time) [18–21]. Olive cultivars are propagated vegetatively by cutting or grafting. Most of the modern varieties display a maternal lineage (E1) that is distributed all around the whole Mediterranean area [6,14,22]. However, the genetic diversity of cultivated populations exhibits a complex patchy pattern [23,24].

For cultural (traditions, landscapes) and economic (oil and olives) reasons, this tree is regarded as one of the most cultivated plant in the world, ranking the 24th among 35 species [25]. It is now distributed in the five continents. It grows extensively in Mediterranean climate regions in Australia, South Africa, North and South America and covers about 11 Mha, with 98% located in the Mediterranean Basin [26,27]. Eighty percent of the cultivated olive surface is located in northern Mediterranean countries (Spain, Italy, Greece, Turkey), 17% in the Middle East (Jordan, Syria, Iraq, Iran) and in North Africa (Morocco, Algeria, Tunisia, Egypt), and only 2% is located in North (USA) and South (Mexico, Argentina, Peru) America [28,29] (Fig. 2).

Olive diversity may also reflect a significant diversity of associated pests, some of them being specific. Major olive diseases are due to insects (e.g., the fruit fly *Bactrocera oleae*, the black scale *Saissetia oleae*, the moth *Prays oleae*, the borer *Palpita unionalis*), and to air-borne (e.g., the peacock spot *Spilocaea oleaginea*, the knot *Pseudomonas syringae*) or soil-borne (e.g., verticillium wilt *Verticillium dahliae* and plant-parasitic nematodes) parasites and pathogens.

2. Plant-parasitic nematodes associated with olive trees

2.1. General considerations

Nematodes are roundworms of the phylum Nematoda, Metazoa having unsegmented, cylindrical bodies with a non-ciliated tough outer cuticle. The group includes free-living forms and disease-causing parasites. Plant-parasitic nematodes (PPN) are soil-borne microscopic worms that mainly feed on plant-root cells using a spear-like structure, called a stylet. They introduce it in cells to inject digestion secretions and suck cell contents [30].

Thereby, these parasites cause significant economic damages to many types of crops as cereals, vegetables, tubers, fruit, and floral crops [31]. Annual losses caused by PPN are estimated from 8.8 to 14.6% of the world crop production (100–157 billion USD/year) [32,33]. In Europe, economic losses are estimated to 10% of the grain production and lead to reduce by 20 to 30% Mediterranean citrus production [34].

Olive trees are hosts of these parasites. But very few information is available about PPN communities associated with olive [27]. The first record was from the USA, where root-knot nematodes *Meloidogyne* spp. were detected [35]. Since then, some researches on olive PPN

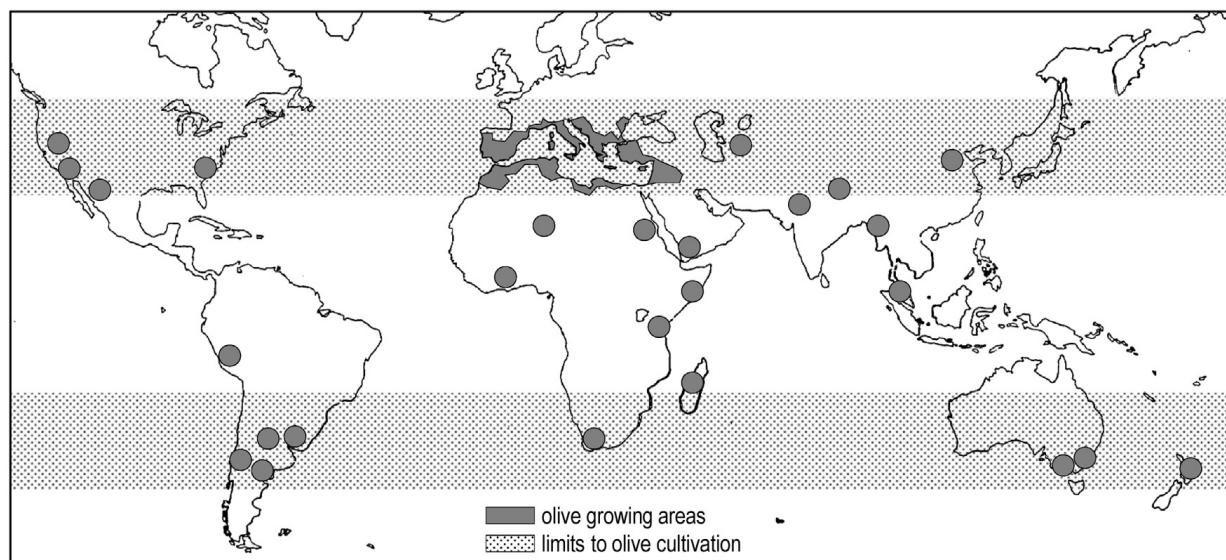


Fig. 2. World distribution of *Olea europaea* L. producing areas [26].

were developed in several parts of the world, mainly in Italy [36–38], Chile [39], Jordan [40], Libya [41], Spain [27,42], Portugal [43], Egypt [44–46], Iran [47,48], Turkey [49,50], Argentina [51], and Syria [52].

Most of these studies have focused on PPN description without studying their potential effects on the crop [53]. However, studies focused on taxa that represent a real threat to olive tree, particularly in the Mediterranean Basin, such as root-knot nematodes *Meloidogyne* [54–57], lesion nematodes *Pratylenchus* [58], spiral nematodes *Helicotylenchus* [59,60], reniform nematodes *Rotylenchulus* [61,62], cyst nematodes *Heterodera* [63,64], and dagger nematodes *Xiphinema* [60].

Other studies focused on assessing the impact of these parasites in nurseries, especially in Spain [27] and Iran [48].

2.2. Diversity of plant-parasitic nematodes associated with olive trees

Previous reviews updated knowledge about nematode diversity associated with olive tree [53,65–68].

Moreover, the analysis of all literature sources let to record 153 species (some as still unidentified) belonging to 56 genera related to the PPN orders Aphelenchida (4 genera and 3 species), Dorylaimida (5 genera and 36 species) and Tylenchida (48 genera and 114 species) (Table 1). Most of the species have been detected on other crops [28]. They belong to three trophic groups: plant feeders, fungal feeders (this group usually feeds on fungi but may alternatively feeds on plants, e.g. Aphelenchida nematodes) and root-hair feeders. Considering their adaptive strategies defined by colonizer/persister values ($=cp$ -values) ranking from 1 (colonizers/ r strategy) to 5 (persisters/ K strategy) [69], most of these species show high reproduction rates and rapid adaptation capacities to environmental changes (cp -3).

While many species such as *Helicotylenchus digonicus*, *Meloidogyne javanica*, *M. incognita*, *Pratylenchus vulnus* are able to parasitize a wide range of cultivated and wild plants,

others, such as *H. oleae*, *H. neopaxilli*, *M. baetica*, *M. lusitanica*, *Rotylenchulus macrosoma*, are specific to olive tree [28].

Considering the species richness encountered in each genus, the genera *Xiphinema*, *Tylenchorhynchus*, *Pratylenchus*, *Helicotylenchus*, *Longidorus*, *Meloidogyne* and *Paratylenchus* are the most diversified with 16, 13, 12, 10, 9, and 8 species respectively, while 41.4% of the genera are monospecific (Fig. 3).

2.3. Distribution of plant-parasitic nematodes

Plant-parasitic nematodes were encountered in most of the olive-producing countries. But 81% of the genera were scattered in less than 25% of the countries (Fig. 4). Twenty-one genera were detected in only one country. The dominant genera (*Criconemoides*, *Helicotylenchus*, *Longidorus*, *Meloidogyne*, *Pratylenchus*, *Rotylenchulus*, *Rotylenchus*, *Tylenchorhynchus*, *Tylenchulus*, *Tylenchus*, and *Xiphinema*) are each present in at least six countries (Fig. 5), especially around the Mediterranean Basin. Root-knot nematodes *Meloidogyne* spp. are distributed in 19 countries. That could explain the economic importance of this group that can represent a real threat for olive.

Modelling the number of genera recorded in each country (Fig. 6A), the highest diversities were recorded in Spain (38 genera), in Jordan (28), in Italy (24), in Greece (20), and in Turkey (16). Ten to twelve genera were detected in Chile, Egypt, Iran, Portugal, and Syria. Only one to five genera were recorded in the other countries. However, these data can in no way reflect the true diversity of the PPN associated with olive trees in each country. Indeed, modelling the research effort by the number of publications as an indicator (Fig. 6B), the diversity found in each country is close to this indicator. Moreover, considering the Mediterranean Basin only, PPN diversity is totally unknown in France, in Morocco, in Adriatic countries, and in most of the Mediterranean islands.

The most common nematode genera detected in Jordan were [40]: *Filenchus*, *Helicotylenchus*, *Merlinius*,

Table 1

Plant-parasitic nematode taxa associated with olive tree.

Order	Family	Species	Country	Reference		
Aphelenchida	Aphelenchidae	<i>Aphelenchus avenae</i> Bastian 1865	Greece	[103]		
			Iran ^a	[48]		
			Jordan	[40]		
		Aphelenchoididae	<i>Aphelenchus</i> sp. Bastian 1865	Spain	[42]	
				Iran	[47]	
			<i>Paraphelenchus pseudoparietinus</i> Micoletzky 1921	Italy	[36]	
				Spain	[42]	
				Egypt	[46]	
				Italy	[36]	
				Jordan	[40]	
	Dorylaimida	Longidoridae	<i>Aprutides guidettii</i> Scognamiglio 1974	Spain	[42]	
				Egypt	[108]	
			<i>Longidorus africanus</i> Merny 1966	Portugal	[79]	
				<i>L. belloi</i> Andres and Arias 1988	Greece	[167]
				<i>L. closelongatus</i> Stoyanov 1964	Greece	[168]
				<i>L. cretensis</i> Tzortzakakis et al., 2001	Egypt	[46]
				<i>L. elongatus</i> De Man 1876	Spain	[42]
				<i>L. macrosoma</i> Hooper 1961	Jordan	[94]
				<i>L. siddiqii</i> Aboul-Eid 1970	Portugal	[79]
				<i>L. vinearum</i> Bravo and Roca 1995	Egypt	[60]
<i>Longidorus</i> sp. Micoletzky 1922	Italy	[36]				
	Spain ^a	[27]				
	Syria	[52]				
	<i>Paralongidorus</i> sp. Siddiqi et al., 1963	Italy		[110]		
	<i>Xiphinema aequum</i> [112]	Italy		[112]		
	<i>X. barensense</i> Lamberti et al., 1986	Italy		[113]		
	<i>X. californicum</i> Lamberti and Bleve-Zacheo 1979	USA		[114]		
	<i>X. diversicaudatum</i> Micoletzky, 1927	Portugal		[115]		
	<i>X. elongatum</i> Schuurmans Stekhoven and Teunissen, 1938	Egypt		[60]		
	<i>X. index</i> Thorne and Allen 1950	Greece		[169]		
<i>X. ingens</i> Luc and Dalmaso 1964	Italy	[121]				
	Jordan	[40]				
	Italy	[116]				
	Jordan	[94]				
	<i>X. italiae</i> Meyl 1953	Italy	[78]			
	<i>X. lusitanicum</i> Sturhan, 1983	Portugal	[79]			
	<i>X. macroacanthum</i> Lamberti et al., 1990	Italy	[117]			
	<i>X. nuragicum</i> Lamberti et al., 1992	Spain	[17]			
	<i>X. pachtaicum</i> Tulaganov 1938	Italy	[68]			
	Jordan	[94]				
<i>X. sahelense</i> Dalmaso, 1969	Portugal	[79]				
	Spain	[17]				
	Spain ^a	[27]				
	Spain	[118]				
	<i>X. turcicum</i> Luc and Dalmaso, 1964	Spain	[42]			
	<i>X. vuittenezi</i> Luc et al., 1964	Spain	[118]			
	<i>Xiphinema</i> sp. Cobb, 1913	Chile	[120]			
	Egypt	[46]				
	Iran	[47]				
	Israel	[170]				
Italy	[36]					
Jordan	[171]					
Spain	[172]					
Trichodoridae	<i>Paratrichodorus alleni</i> Siddiqi, 1974	Italy	[142]			
		<i>P. minor</i> Colbran 1956	Spain ^a	[27]		
		<i>P. teres</i> Hooper 1962	Spain ^a	[27]		
		<i>Paratrichodorus</i> sp. Siddiqi 1974	Jordan	[40]		
		<i>Trichodorus aequalis</i> Allen 1957	Spain	[42]		
		<i>T. andalusicus</i> Decraemer et al., 2012	Spain ^b	[140]		
		<i>T. iliolaensis</i> Decraemer et al., 2012	Spain	[140]		
		<i>T. giennensis</i> Decraemer et al., 1993	Spain ^a	[27]		
		<i>T. lusitanicus</i> Siddiqi, 1974	Portugal	[143]		
		<i>T. parasilvestris</i> Decraemer et al., 2012	Spain	[140]		
		<i>T. primitivus</i> De Man 1880	Portugal	[143]		
		<i>T. silvestris</i> Decraemer et al., 2012	Spain ^b	[140]		
		<i>T. taylori</i> Mancine 1980	Italy	[141]		
		<i>T. variopapillatus</i> Hooper, 1972	Italy	[142]		

Table 1 (Continued)

Order	Family	Species	Country	Reference
Tylenchida	Anguinidae	<i>Trichodorus</i> sp. Cobb 1913	Chile	[120]
			Italy	[36]
		<i>Ditylenchus anchiliposomus</i> Tarjan 1958	Spain	[42]
		<i>D. dipsaci</i> Kuhn 1857	Jordan	[40]
			Spain	[173]
		<i>D. virtudesae</i> Tobar-Jimenez 1964	Spain	[172]
		<i>Ditylenchus</i> sp. Filipjev 1936	Italy	[36]
			Jordan	[171]
			Spain	[42]
			Spain	[42]
	Criconematidae	<i>Criconema annuliferum</i> De Man 1921	Spain	[42]
		<i>C. princeps</i> Andr�assy 1962	Portugal	[68]
		<i>Criconema</i> sp. Hofm�anner and Menzel 1914	Greece	[169]
		<i>Criconemella informis</i> Micoletzky 1922	Spain	[42]
		<i>C. sphaerocephala</i> Taylor 1936	Spain	[42]
		<i>C. xenoplax</i> Raski 1952	Jordan	[40]
			Spain	[27]
		<i>Criconemella</i> sp. De Grisse and Loof 1965	Spain	[42]
		<i>Criconemoides informis</i> Micoletzky 1922	Jordan	[94]
			Spain ^a	[27]
		<i>Criconemoides</i> sp. Taylor 1936	Chile	[120]
			Egypt	[60]
			Greece	[103]
			Italy	[36]
			Jordan	[171]
			Zimbabwe	[174]
		<i>Crossonema multisquamatum</i> Kirjanova 1948	Turkey	[49]
		<i>Hemicriconemoides gaddi</i> Loos 1949	Spain	[68]
		<i>Mesocriconema curvatum</i> Raski 1952	Brazil	[175]
		<i>M. onoense</i> Luc 1959	Italy	[176]
		<i>M. sicula</i> Loof and De Grisse 1989	Spain	[68]
		<i>M. sphaerocephalum</i> Taylor 1936	Spain ^a	[27]
		<i>M. xenoplax</i> Raski 1952	Jordan	[177]
		<i>Neolobocriconema olearum</i> Hashim 1984	Portugal	[142]
		<i>Nothocriconema princeps</i> De Grisse and Loof 1965	Zimbabwe	[174]
		<i>Ogma civellae</i> Steiner 1949	Spain	[42]
		<i>O. rhombosquamatum</i> Mehta and Raski 1971	Jordan	[40]
		<i>O. seymouri</i> Wu 1965	Italy	[148]
			Portugal	[146]
			Spain	[68]
		Italy	[110]	
	Dolichodoridae	<i>Ogma</i> sp. Mehta and Raski 1971	Italy	[178]
		<i>Dolichodorus heterocephalus</i> Cobb 1911	Italy	[110]
		<i>Dolichodorus</i> sp. Cobb 1915	Italy	[36]
	Hemicycliophoridae	<i>Trophurus</i> sp. Loof 1956	Turkey	[49]
		<i>Hemicycliophora sturhani</i> Loof 1984	Chile	[120]
	Heteroderidae	<i>Hemicycliophora</i> sp. De Man 1921	Spain ^a	[27]
<i>Heterodera avenae</i> Filipjev 1934		Spain ^a	[27]	
Hoplolaimidae	<i>H. mediterranea</i> Vovlas et al., 1981	Italy	[63]	
		Spain	[64]	
	<i>Heterodera</i> sp. Filipjev and Schuurmans Stekhoven 1941	Italy	[36]	
	Heteroderinae Filipjev and Schuurmans Stekhoven 1941	Spain	[42]	
	<i>Aorolaimus perscitus</i> Doucet 1980	Spain	[147]	
	<i>Helicotylenchus digonicus</i> Perry 1959	Cyprus	[179]	
		Greece	[103]	
		Jordan	[180]	
		Spain	[17]	
		Spain ^a	[27]	
		Turkey	[49]	
	<i>H. dihystra</i> Cobb 1893	Brazil	[181]	
		Cyprus	[179]	
		Egypt	[108]	
	Italy	[111]		
	Jordan	[180]		
	Spain ^a	[27]		
	Spain	[182]		
	Zimbabwe	[109]		
<i>H. erythrinae</i> Zimmermann 1904	Italy	[59]		
<i>H. minzi</i> Sher 1966	Jordan	[40]		
<i>H. neopaxili</i> Inserra et al., 1979	Italy	[70]		
<i>H. oleae</i> Inserra et al., 1979	Italy	[70]		

Table 1 (Continued)

Order	Family	Species	Country	Reference
		<i>H. pseudorobustus</i> Steiner 1914	Greece Iran ^a Italy Jordan Spain ^a Spain	[103] [48] [68] [94] [27] [27]
		<i>H. tunisiensis</i> Siddiqi 1963	Israel Jordan Portugal Spain Turkey	[109] [94] [146] [173] [49]
		<i>H. vulgaris</i> Yuen 1964	Jordan Portugal Spain ^a Spain Turkey	[40] [146] [27] [173] [49]
		<i>Helicotylenchus</i> sp. Steiner 1945	Algeria Chile Egypt Greece Iran Italy Jordan Syria	[104] [120] [46] [103] [47] [36] [171] [52]
		<i>Hoplolaimus aorolaimoides</i> Siddiqi 1972	Portugal	[146]
		<i>H. galeatus</i> Cobb 1913	Turkey	[49]
		<i>Hoplolaimus</i> sp. Daday 1905	Egypt Syria	[60] [52]
		<i>Plesiorotylenchus striaticeps</i> Vovlas et al., 1993	Turkey	[49]
		<i>Rotylenchus buxophilus</i> Golden 1956	Turkey	[49]
		<i>R. cypriensis</i> Antoniou 1980	Jordan Turkey	[40] [49]
		<i>R. robustus</i> De Man 1876	Portugal	[142]
		<i>Rotylenchus</i> sp. Filipjev 1936	Cyprus Greece Italy Spain ^a Spain	[179] [103] [36] [27] [42]
	Meloidogynidae	Hoplolaiminae Filipjev 1934	Spain	[89]
		<i>Meloidogyne acrita</i> Chitwood 1949	China	[89]
		<i>M. arenaria</i> Neal 1889	Chile China Spain ^a Spain ^b	[39] [89] [27] [90]
		<i>M. baetica</i> Castillo et al. 2003	Chile	[39]
		<i>M. hapla</i> Chitwood 1949	Israel Portugal	[88] [96]
		<i>M. incognita</i> Kofoid and White 1919	Argentina Brazil Chile China Egypt India Israel Italy Jordan Lebanon Lybia Portugal Spain ^a	[68] [181] [39] [89] [45] [183] [88] [184] [185] [186] [41] [187] [27]

Table 1 (Continued)

Order	Family	Species	Country	Reference
		<i>M. javanica</i> Treub 1885	Argentina	[51]
			Chile	[39]
			China	[89]
			Egypt	[60]
			Greece	[103]
			Iran ^a	[48]
			Israel	[87]
			Italy	[184]
			Jordan	[94]
			Lybia	[41]
			Pakistan	[68]
			Portugal	[68]
			Spain ^a	[27]
		<i>M. lusitanica</i> Abrantes and Santos 1991	Portugal	[43]
			Spain ^a	[27]
		<i>Meloidogyne</i> sp. Goeldi 1887	Chile	[120]
			Cyprus	[179]
			Iran	[47]
			Israel	[87]
			Italy	[59]
			Jordan	[171]
			Portugal	[188]
			Syria	[52]
			USA	[35]
	Paratylenchidae	<i>Gracilacus peratica</i> Raski 1962	Italy	[36]
			Portugal	[146]
			Spain	[68]
		<i>G. teres</i> Raski 1976	Spain	[147]
		<i>Gracilacus</i> sp. Raski 1962	Italy	[37]
			Spain	[42]
		<i>Paratylenchus arcuatus</i> Luc and de Guiran 1962	Spain ^a	[27]
			Turkey	[49]
		<i>P. baldacii</i> Raski 1975	Spain	[42]
		<i>P. ciccaronei</i> Raski 1975	Spain	[17]
		<i>P. microdorus</i> Andrassy 1959	Spain	[173]
		<i>P. nanus</i> Cobb 1923	Spain	[173]
		<i>P. sheri</i> Raski 1973	Spain	[173]
		<i>P. vandenbrandei</i> De Grisse 1962	Italy	[95]
		<i>Paratylenchus</i> sp. Filipjev 1936	Italy	[36]
			Jordan	[171]
			Syria	[52]
	Pratylenchidae	<i>Pratylenchoides erzurumensis</i> Yuksel 1977	Turkey	[49]
		<i>P. ritteri</i> Sher 1970	Spain ^a	[27]
			Iran	[190]
		<i>Pratylenchoides</i> sp. Winslow 1958	Jordan	[171]
		<i>Pratylenchus coffeae</i> Zimmerman 1898	Australia	[101]
			Jordan	[40]
		<i>P. crenatus</i> Loof 1960	Italy	[190]
		<i>P. fallax</i> Seinhorst 1968	Spain ^a	[27]
		<i>P. mediterraneus</i> Corbett 1983	Turkey	[49]
		<i>P. musicola</i> Cobb 1919	USA	[102]
		<i>P. neglectus</i> Rensch 1924	Greece	[103]
			Italy	[95]
			Iran	[189]
			Jordan	[40]
			Spain	[173]
			Spain ^a	[27]
		<i>P. penetrans</i> Cobb 1917	Australia	[191]
			Italy	[95]
			Jordan	[40]
			Spain	[42]
			Turkey	[68]
		<i>P. thornei</i> Sher and Allen 1953	Iran ^a	[48]
			Jordan	[94]
			Spain ^a	[27]
		<i>P. vulnus</i> Allen and Johnson 1951	Algeria	[104]
			Australia	[191]
			Italy	[58]
			Spain ^a	[27]
			USA	[102]
		<i>P. zaeae</i> Graham 1951	Turkey	[49]

Table 1 (Continued)

Order	Family	Species	Country	Reference
		<i>Pratylenchus</i> sp. Thorne 1949	Algeria	[104]
			Chile	[120]
			Egypt	[46]
			Greece	[103]
			Iran	[47]
			Italy	[36]
			Jordan	[171]
			Spain	[42]
			Syria	[52]
		<i>Radopholus</i> sp. Thorne 1949	Greece	[103]
		<i>Zygotylenchus guevarai</i> Tobar-Jimenez 1963	Spain	[42]
			Spain ^a	[27]
			Turkey	[49]
	Psilenchidae	<i>Psilenchus hilarulus</i> De Man 1921	Iran ^a	[48]
		<i>P. iranicus</i> Kheiri 1970	Jordan	[40]
			Spain	[42]
		<i>Psilenchus</i> sp. De Man 1921	Greece	[169]
			Iran	[47]
	Rotylenchulidae	<i>Rotylenchulus macrorodatus</i> Dasgupta et al., 1968	Greece	[133]
			Italy	[131]
		<i>R. macrosoma</i> Dasgupta et al., 1968	Israel	[134]
			Jordan	[40]
			Spain ^b	[62]
			Turkey	[49]
		<i>R. reniformis</i> Linford and Oliveira 1940	Egypt	[192]
			Greece	[103]
		<i>Rotylenchulus</i> sp. Linford and Oliveira 1940	Algeria	[104]
			Syria	[52]
	Telotylenchidae	<i>Amplimerlinius amplus</i> Siddiqi 1976	Portugal	[193]
		<i>A. dubius</i> Steiner 1914	Turkey	[49]
		<i>A. macrurus</i> Goodey 1932	Jordan	[94]
		<i>A. paraglobigerus</i> Castillo et al., 1990	Spain ^a	[27]
		<i>Amplimerlinius</i> sp. Siddiqi 1976	Spain	[42]
		<i>Bitylenchus goffarti</i> Sturhan 1966	Turkey	[49]
		<i>Merlinius brevidens</i> Allen 1955	Cyprus	[179]
			Greece	[103]
			Iran ^a	[48]
			Jordan	[94]
			Spain	[42]
		<i>M. microdorus</i> Geraert 1966	Jordan	[40]
			Spain	[173]
		<i>M. nothus</i> Allen 1955	Jordan	[40]
		<i>Quinisulcius acutus</i> Allen 1955	Turkey	[49]
		<i>Paratrophurus loofi</i> Arias 1970	Spain	[42]
		<i>Scutylenchus lenorus</i> Brown 1956	Turkey	[49]
		<i>Telotylenchus</i> sp. Siddiqi 1960	Greece	[103]
		<i>Trophurus</i> sp.	Italy	[36]
		<i>Tylenchorhynchus aduncus</i> De Guiran 1967	Spain ^a	[27]
		<i>T. clarus</i> Allen 1955	Jordan	[94]
			Spain ^a	[27]
		<i>T. claytoni</i> Steiner 1937	Cyprus	[68]
			Turkey	[49]
		<i>T. cylindricus</i> Cobb 1913	Turkey	[49]
		<i>T. dubius</i> Buetschli 1873	Greece	[103]
			Spain	[42]
			Spain ^a	[27]
		<i>T. goffarti</i> Sturhan 1966	Jordan	[94]
		<i>T. huesingi</i> Paetzold 1958	Spain ^a	[27]
		<i>T. mamillatus</i> Tobar-Jimenez 1966	Spain ^a	[27]
		<i>T. penniseti</i> Gupta and Uma 1980	Turkey	[49]
		<i>T. striatus</i> Allen 1955	Greece	[103]
		<i>T. tenuis</i> Micoletzky 1922	Jordan	[194]
		<i>T. tritici</i> Golden et al., 1987	Turkey	[49]

Table 1 (Continued)

Order	Family	Species	Country	Reference
		<i>Tylenchorhynchus</i> sp. Cobb 1913	Algeria Chile Egypt Greece Iran Italy Jordan Spain ^a	[104] [120] [45] [103] [47] [36] [171] [27]
	Tylenchidae	<i>Aglenchus agricola</i> De Man 1884 <i>Basiria duplexa</i> Hagemeyer and Allen 1952 <i>Basiria</i> sp. Siddiqi 1959 <i>Boleodorus thylactus</i> Thorne 1941 <i>Boleodorus</i> sp. Thorne 1941	Syria Spain Jordan Iran ^a Iran Jordan Spain	[52] [42] [42] [40] [48] [47] [40] [42]
		<i>Coslenchus cancellatus</i> Cobb 1925 <i>C. costatus</i> De Man 1921 <i>C. lateralis</i> Andrassy 1982 <i>Coslenchus</i> sp. Siddiqi 1978 <i>Discotylenchus discretus</i> Siddiqi 1980 <i>Filenchus discretus</i> Siddiqi 1979 <i>F. filiformis</i> Buetschli 1873 <i>F. sandneri</i> Wasilewska 1965 <i>Filenchus</i> sp. Andrassy 1954	Spain Greece Spain Spain Jordan Spain Greece Spain Jordan Spain Iran ^a Spain Spain Spain Chile Greece Iran Italy Jordan Spain	[42] [103] [42] [42] [40] [42] [103] [17] [40] [17] [48] [42] [42] [17] [120] [103] [47] [36] [94] [17]
	Tylenchulidae	<i>Trophotylenchulus saltensis</i> Hashim 1983 <i>Tylenchulus semipenetrans</i> Cobb 1913	Jordan Australia Chile Egypt Greece Italy USA Chile Italy Syria	[40] [101] [68] [60] [169] [38] [195] [120] [110] [52]
		<i>Tylenchulus</i> sp. Cobb 1913	Chile Italy Syria	[120] [110] [52]

^a In nurseries.^b On wild olive.

Pratylenchus, and *Xiphinema*. A more recent study conducted in Iran revealed other frequent genera, such as *Aphelenchus*, *Boleodorus*, *Meloidogyne*, *Psilenchus*, *Tylenchus*, and *Tylenchorhynchus* [47].

The spatial distribution of PPN species in different fields sampled in Spain is homogeneous [42]. However, this author noticed seasonal fluctuations according to local climate. High-level PPN populations were observed in autumn, low levels in summer and intermediate levels in winter and spring. He hypothesized that microclimatic soil variations could modify root tissue plasticity, which consequently could change the aptitude of nematodes to penetrate roots [70].

Nematode occurrence usually differs in nurseries: the most common species detected in southern Spain are *M. incognita*, *M. javanica*, *M. arenaria*, *Mesocriconema xenoplax*, *Pratylenchus penetrans*, and *P. vulnus* [27]. In

contrast, the most common nematodes detected in nurseries in northern Iran are [48]: *Aphelenchus avenae*, *M. javanica*, *Irantylenchus* sp. (a new genus reported for the first time as associated with olive), *Pratylenchus thornei*, *Helicotylenchus pseudorobustus*, *Boleodorus thylactus*, *Psilenchus hilarulus*, and *Merlinius brevidens*. But in nurseries, the occurrence of PPN is directly related to the origin of the cutting substrates that are often not sterilized, and to the susceptibility of olive cultivars (e.g., *P. vulnus* is more frequent on cv. Manzanilla than on other cultivars [27]).

2.4. Impacts of plant-parasitic nematodes on olive trees

Host–parasite interactions between PPN and olive trees have been scarcely studied. However, controlled experiments showed that root-knot nematodes *Meloidogyne* (e.g., *M. arenaria*, *M. incognita*, and *M. javanica*) significantly

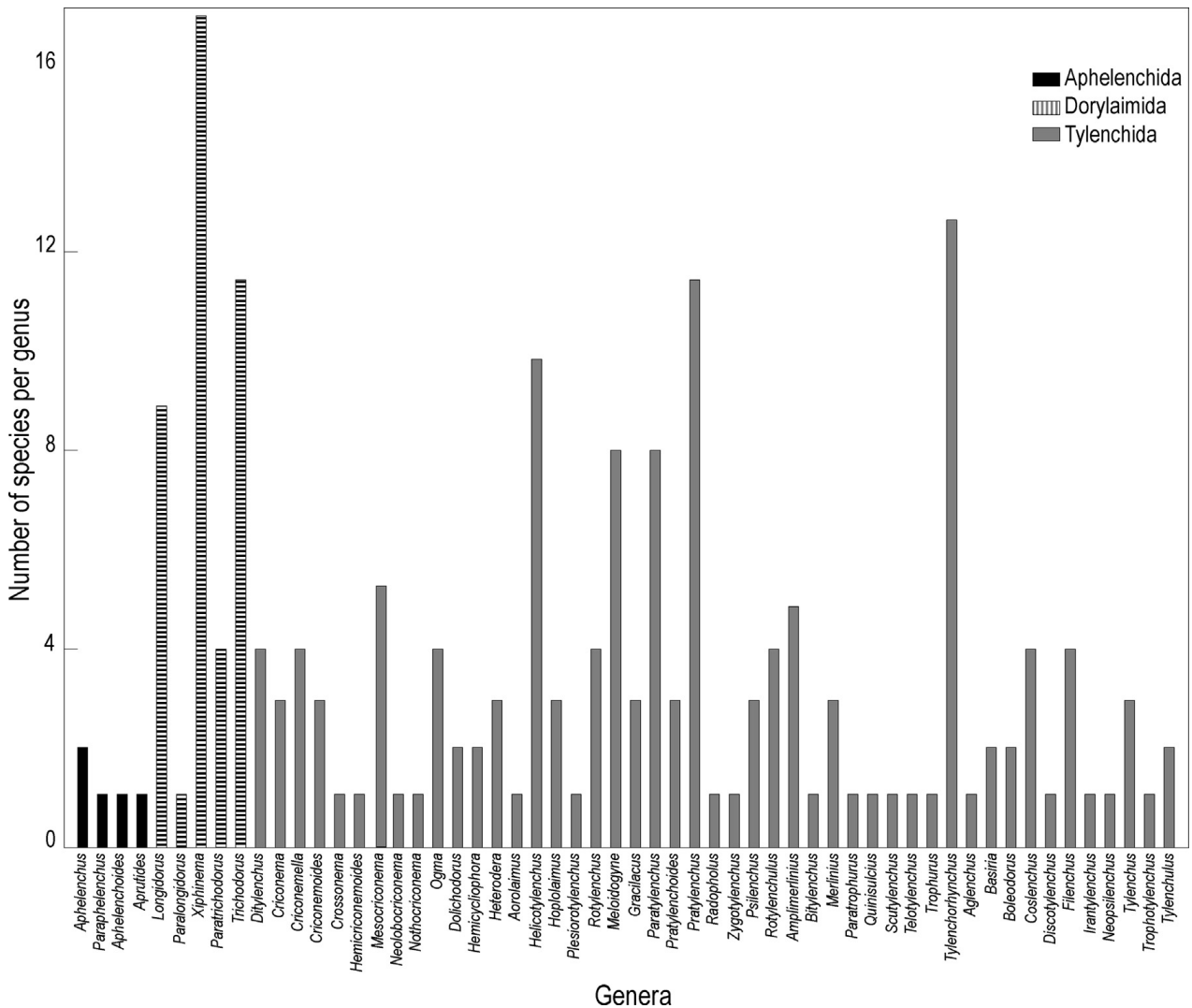


Fig. 3. Species diversity in each plant-parasitic nematode genus.

reduce tree growth [55–57,71] and may be responsible for 5 to 10% crop losses [33], while these damages are often difficult to assign to PPN [28]. Indeed, the symptoms due to PPN parasitism are not clear, because

- the hardness of this plant is able to hide them;
 - population levels are usually low in the rhizosphere [68].
- In any case, this is commonly observed in traditional olive cropping systems.

Three species of fruit crop infecting nematodes that could damage olive (*Longidorus diadecturus*, *Xiphinema americanum*, *X. californicum*) are absent in Europe [68]. These species have been declared as quarantine pests in Europe (EPPO/OEPP). *Heterodera mediterranea* and *X. italiae* are also quarantine pests in Chile.

The nematode parasitism impacts directly the plant growth and yield. Basically, root functions are disturbed by histological alterations of root tissues. Physiological disorders limit water and mineral absorption, leading to major deficits [66]. These phenomena are less prominent on wild olive trees that seems more tolerant to PPN infection [28].

The impact of nematodes is much more significant in nurseries where irrigation conditions favour the development of roots and therefore PPN multiplication [27,28,68]. Humidification of the substrates moderates the temperature, two factors that support the development of nematodes [72]. Moreover, roots of olive seedlings are more susceptible to PPN than older plants [27]. Thus, infestation levels with *Meloidogyne arenaria*, *M. incognita*, *M. javanica*, *Mesocriconema xenoplax*, *Pratylenchus penetrans* and *P. vulnus* are significantly higher in nurseries than in orchards [27,48]. But although the same genera are known for their harmfulness on other fruit trees such as apple and pear, the damaging threshold of PPN is poorly evaluated on olive [55,58,73].

Indirect root damages are also related to nematode infection, whose penetration opens pathways to other soil-borne pathogens (bacteria, fungi) [67,74]. The best example is the association of PPN, such as *M. incognita* and *P. vulnus*, with the fungal pathogen *Verticillium dahliae* (*Verticillium wilt*). The presence of nematodes enhances the symptoms induced by the fungus [75–77]. Experiments have demonstrated a positive correlation between the number of root galls induced by *M. incognita* and the

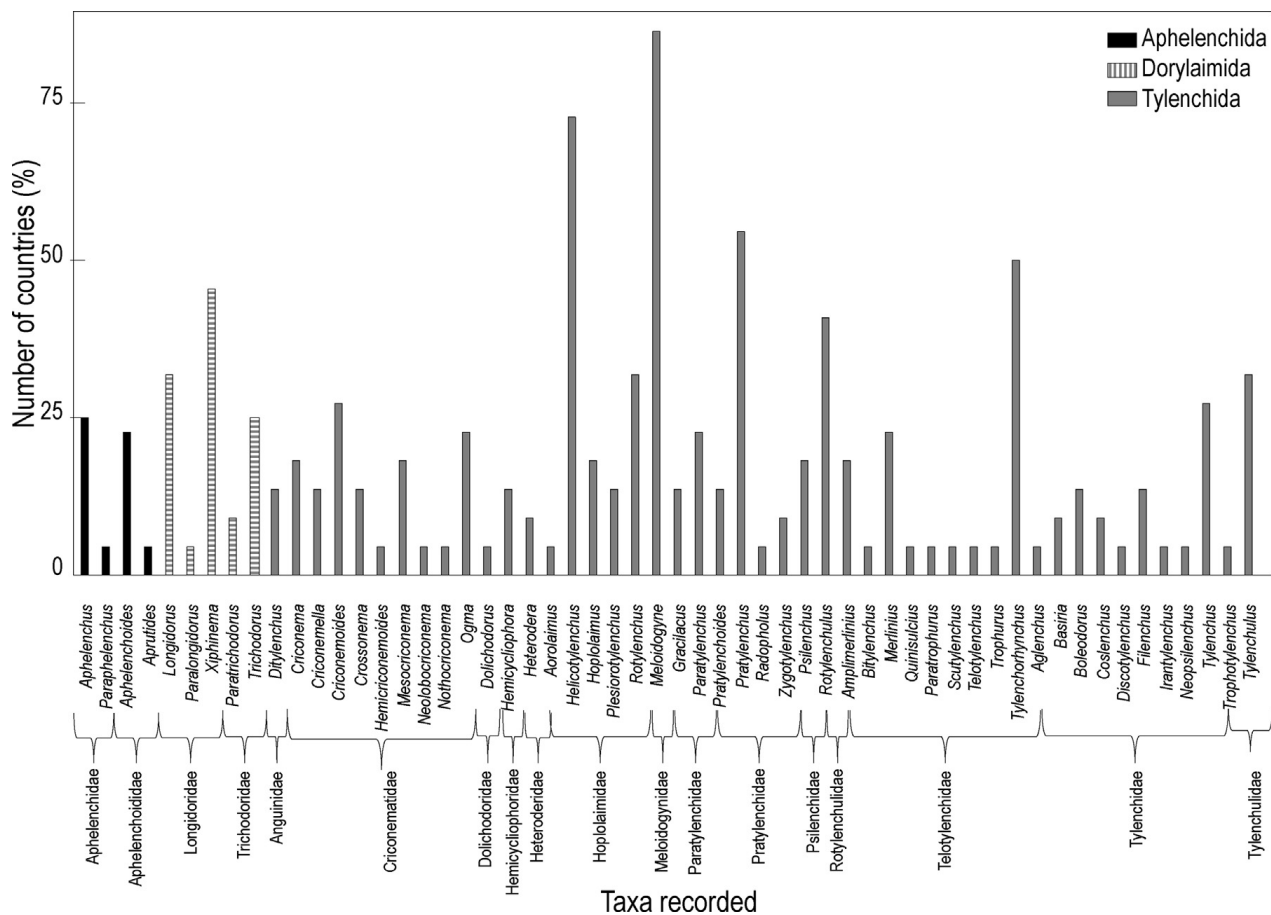


Fig. 4. Frequency of the plant-parasitic nematode genera detected (% of countries).

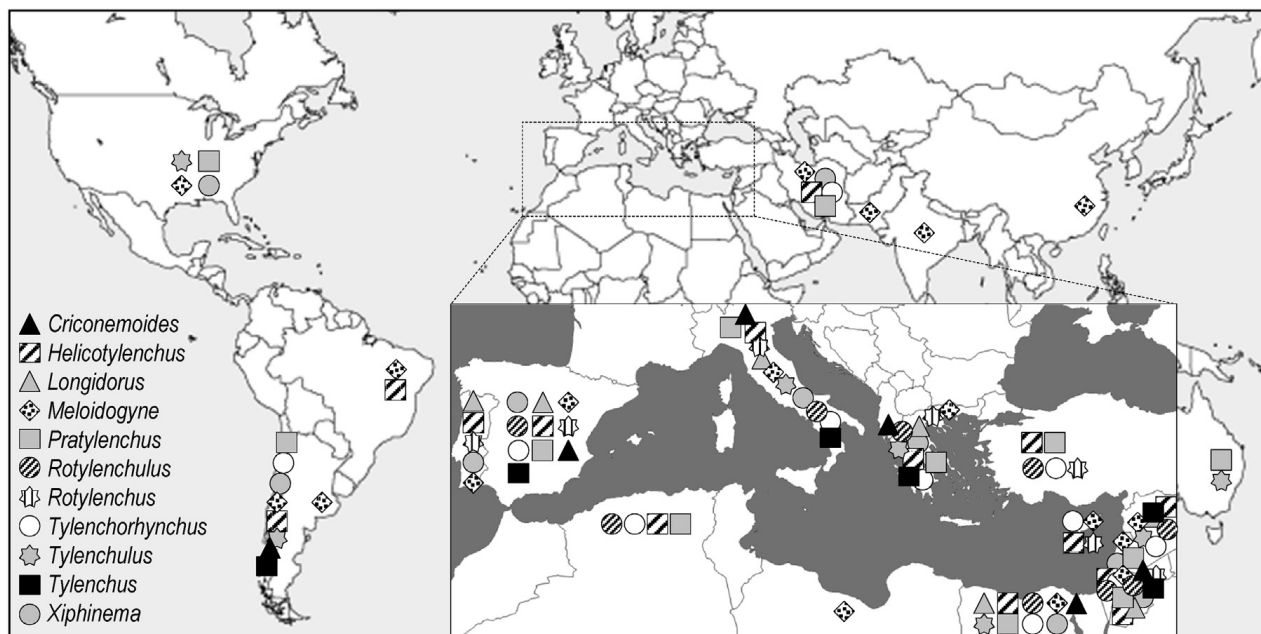


Fig. 5. World and Mediterranean distribution of the most frequent plant-parasitic nematode genera associated with olive tree (i.e. present in more than 25% of countries).

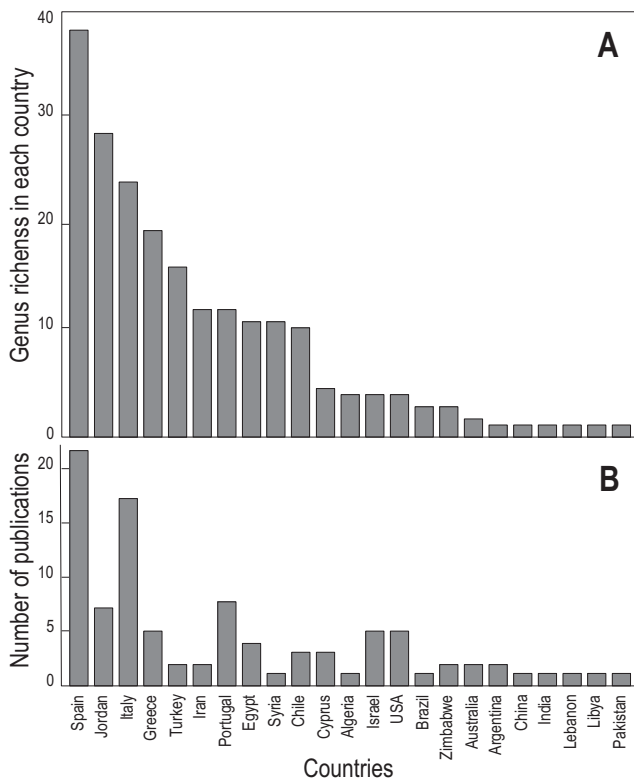


Fig. 6. Genus richness of plant-parasitic nematode recorded in each Mediterranean country (A) and associated publications (B).

extent of root rots and vascular discoloration, and the *M. incognita*/*V. dahliae* combination reduced rhizogenesis [76]. When the three parasites *M. incognita*/*P. vulnus*/*V. dahliae* were associated, the reproduction of each of two nematode species was affected by the presence of the other species and/or by the fungus.

Similarly, damages induced by *Heterodera mediterranea* could be intensified by the presence of *V. dahliae* [64]. The

M. javanica/*Fusarium* sp. complex would also be responsible for the “Seca” disease that induces drying of young olive trees in nurseries [51].

Virus transmission by some PPN species is also a major risk to the olive tree. Thus, the olive tree is a natural host to Nepoviruses such as the “Arabis Mosaic Nepovirus” (ArMV), the “Strawberry Latent Ringspot Sadwavirus” (SLRSV) and the “Cherry Leaf Roll Virus” (CLRV) transmitted by *Xiphinema diversicaudatum* and *X. vuittenzei* [78,79]. These viruses do not produce symptoms on olive [80], except SLRSV that is responsible for leaf and fruit distortion on the variety Ascolana tenera [81].

Biochemical reactions in olive tree roots are diverse, depending on the nematode species that induce specific physiological mechanisms, as for lesion nematodes as *P. vulnus* [58,82], for root-knot nematodes as *M. javanica* [55,56], or for dagger nematodes as *Xiphinema index* [83]. The secondary metabolism would be involved in plant reactions, which would confer resistance (i.e. lower infestation and reproduction) on some varieties [84].

3. The dominant plant-parasitic nematodes associated with olive trees

3.1. *Meloidogyne* spp. (root-knot nematodes)

Root-knot nematodes (RKN) are sedentary endoparasites. Ninety-seven species have been described [85]. They have developed reproduction and parasitism strategies that allow them to infect thousands of plant species. Most RKN species induce galls that can invade the whole root system. They are widespread all over the world [66,74,86].

These nematodes have been firstly reported on olive trees in the United States [35]. Thereafter, several *Meloidogyne* species were detected: *M. javanica* [87], *M. incognita*, and *M. hapla* [88], *M. arenaria* [89], *M. lusitanica* [43], and

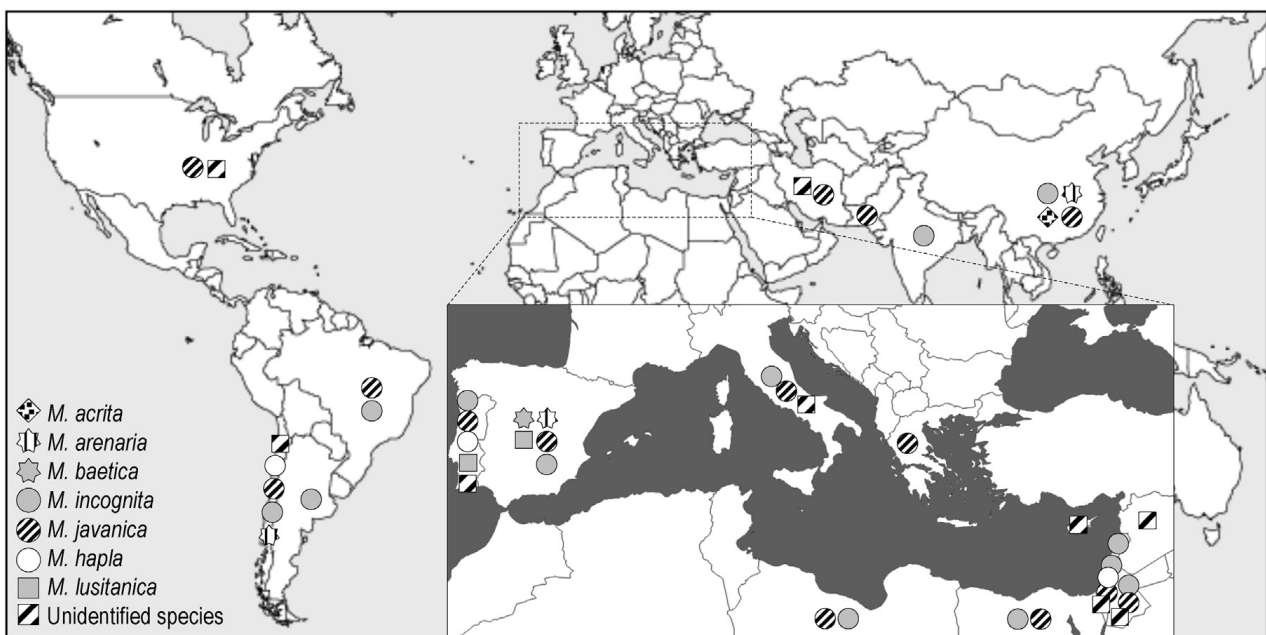


Fig. 7. World and Mediterranean distribution of root-knot nematode species (*Meloidogyne* spp.) associated with olive tree.

M. baetica [90] recently. These nematodes occur on wild and cultivated olive and also in nurseries (except for *M. baetica* [27] (Table 1). Among these species, *M. baetica* was detected on wild olive in southern Spain, and could infect olive orchards. Experiments have shown that this species does not reproduce on tomato (on which it induces small galls without any egg mass) or on legumes. That would confirm its specificity to olive, which led to call it “Mediterranean olive root-knot nematode” [90]. This species was also detected on *Pistacia lentiscus* and *Aristolochia baetica* [91].

RKN are very well adapted to a wide range of temperatures, which explains their worldwide distribution (Table 1 and Fig. 7). However, *M. incognita* and *M. javanica* are the most widespread species in the world [53,92] especially in the Mediterranean Basin [93].

But RKN are at least the less frequent species in olive orchards: they were detected in less than 4% of the samples in Jordan, mainly in irrigated areas and nurseries [40,94], in 2% of the samples in southern Italy (the main production area of the country) and in less than 1% in the other producing areas of the country [95]. That supports other observations made elsewhere in the world [54,55,60,92].

In spite of their scarce distribution, RKN are considered as major pests on olive [53]. Thus, *M. javanica* is the main cause of “Seca” syndrome in new olive orchards in Argentina [51]. This species is responsible for olive decay in nurseries [40]. RKN reduce plant growth [27,43,55,56]. Pathogenicity tests showed that *M. arenaria*, *M. incognita* and *M. javanica* grow as well on rootstocks [56,57] and on plants cv. Picual and cv. Arbequina, which are widely distributed in the Mediterranean Basin [71]. Controlled experiments showed that six months after inoculation with 5000 or 10,000 *M. javanica* juveniles per plant, plant growth was significantly reduced: 28% [60] and 45–52%, according to the cultivar [55], respectively. The stem diameter is the most sensitive parameter to infestation [57,71]. Olive trees infected by *Meloidogyne* spp. show yellowing of the highest branches, and then defoliation [68]. These symptoms are observed everywhere, as well as in China [89], Portugal [96], and Chile [39].

Histological observations of roots infected with *M. incognita* [97], with *M. baetica* [90,91], or with *M. javanica* and *M. lusitanica* [92] indicate different sizes and different localisations of the galls they induce. *M. javanica* galls are usually located on the root tip, but some galls are also distributed along roots. Clusters of several galls were also observed and no secondary roots seen near them [92]. *M. lusitanica* galls are usually distributed as individual galls along roots. *M. baetica* induces small galls, frequently located along the taproot and very rarely on secondary roots, without the development of egg masses [90].

3.2. *Pratylenchus* spp. (lesion nematodes)

Lesion nematodes (LN) are migratory endoparasites. They are worldwide distributed, and are able to infect a wide range of plants. They are involved with other soil-borne pathogens (e.g., fungi) in parasitic complexes. LN are the most damaging nematodes with RKN [98–100].

Several *Pratylenchus* species have been detected on olive (Table 1 and Fig. 5): *P. coffeae* [101], *P. crenatus* [95], *P. mediterraneus* and *P. musicola* and *P. vulnus* [102], *P. neglectus* [103], *P. penetrans* [95], *P. thornei* [104] and *P. zeae* [49]. Some of them (*P. neglectus*, *P. penetrans*, *P. thornei*, *P. vulnus* and *P. fallax*) have been also observed in nurseries [27,48].

Among these species, *P. vulnus* and *P. penetrans* are considered the most frequent and pathogenic on olive [58,71].

Infection with LN induces large necrotic areas on roots, which may cover all the root system. This may rot the cortical parenchyma [66,102] and reduce both number and size of infected roots [105]. Other symptoms such as leaf chlorosis, internode shortening and defoliation can be observed [106].

Controlled inoculations revealed that most of the LN species do not reproduce on olive, leading one to minimize the impact of these nematodes on this crop [71]. Field observations pointed that LN reproduce primarily in weeds without injuring olive trees [107]. Nevertheless, *P. vulnus* is the most pathogenic species [58]. It is responsible for tree decline [53,106]. Significant damages attributed to this species have been recorded in several olive orchards in Italy [38,95,106], in the USA [102], and in Algeria [104]. Inoculations with *P. vulnus* performed on several olive cultivars (Arbequina, Ascolana, Manzanilla, Picual) confirmed its strong pathogenic potential [58,71]. Competitions between RKN and LN have been recorded on olive. Reproduction of *P. vulnus* populations was reduced with root-knot nematode co-infection [76].

3.3. *Helicotylenchus* spp. (spiral nematodes)

Spiral nematodes (SN) are migratory ectoparasites. They are also widely distributed in the world.

Several *Helicotylenchus* species were detected in the olive rhizosphere such as *H. digonicus* [103], *H. erythrinae* [59], *H. dihystra* [108], *H. minzi* [40], *H. neopaxili* and *H. oleae* [70], *H. pseudorobustus* [103], *H. tunisiensis* [109], and *H. vulgaris* [40]. Some species, such as *H. digonicus*, *H. pseudorobustus* and *H. vulgaris* were also detected in nurseries [27,48]. These species have been observed in different olive-producing areas in the world (Table 1 and Fig. 5).

H. digonicus is dominant in many olive orchards [17,42]. For example, it occurs in more than 45% of the orchards in Jordan [40]. Nevertheless, *H. oleae* is widely distributed in Italy [70], where *H. digonicus* was not detected [42]. Other studies also demonstrate the dominance of other spiral nematodes, as *Rotylenchus* spp. in Italy [37,110,111].

Pathogenicity processes caused by SN are still unclear [53]. However, some few studies indicate that *H. dihystra*, *H. erythrinae* or *H. oleae* induces brown lesions on roots and chlorosis of leaf tips, leading to root-mass and plant-growth reduction [59,60,66,70]. Growth delay was observed on plants infected by *H. digonicus* [93]. A 78% plant-growth reduction was measured on plants inoculated with 1000 *H. dihystra* six months before [60]. Some species (e.g., *H. digonicus* and *H. oleae*) have a semi-endoparasitic behaviour (i.e. introduction of the anterior body part into roots) [40,70]. *H. oleae* penetrates up to the root epidermis

and to the cortical parenchyma. It induces cell wall necroses, feed from place to place, leaving cavities in the cortex.

3.4. *Xiphinema* spp. (dagger nematodes)

Dagger nematodes (DN) are ectoparasites, frequently found in the olive rhizosphere [66]. Different species have been recorded (Table 1 and Fig. 5): *Xiphinema aequum* [112], *X. barensense* [113], *X. californicum* (*X. americanum*) [114], *X. diversicaudatum* [115], *X. elongatum* [60], *X. index* [40], *X. ingens* [116], *X. italiae* [78], *X. lusitanicum* [79], *X. macroacanthum* [117], *X. nuragicum* [17] and *X. pachtaicum* [94], *X. sahelense* and *X. vuittenezi* [118], and *X. turcicum* [42]. Despite the low population densities, *X. pachtaicum* is the most common DN species on olive [40,94].

X. diversicaudatum and *X. pachtaicum* show different distribution areas [115]. *X. diversicaudatum* could be considered as an Atlantic species, distributed in both cultivated and uncultivated habitats, but is more abundant in uncultivated ones with weak environmental stresses. *X. pachtaicum* is rather a Mediterranean species, associated also with other fruit trees (apple and peach). This would explain why *X. diversicaudatum* has been reported on olive in central and northern Spain [115], but not in the South [119].

These nematodes are considered as major pathogens on olive trees in Chile [120]. In the USA, they are responsible for 5 to 10% of the damages, which corresponds to an estimated \$ 38.9 million loss [33]. *X. pachtaicum* could be a real nuisance in olive orchards [42]. DNs affect plant growth initiated by root necrosis [60,121].

A 65% plant-growth reduction was observed on plants inoculated with 500 *X. elongatum* six months before [60]. However, significant development of *X. californicum* populations did not affect plant growth [114]. In fact, DNs are usually difficult to rear in controlled conditions because of their susceptibility to moisture and temperature variations [122]. That could explain failures in pathogenicity experiments [53].

3.5. *Tylenchulus* spp. (citrus nematodes)

Usually known as citrus parasites, *T. semipenetrans* was reported on olive in California, Chile and Italy [123] (Table 1 and Fig. 5). Studying several life history traits associated with infection and reproduction on specific host plants, six biotypes were identified: four in California [124], one in Florida [125], and one in Italy [126]. This diversity was revised and reduced to four biotypes [127]:

- the “Poncirus biotype” that reproduces on *Poncirus trifoliata* and grapevine but not on olive (= California biotype C3);
- the “Citrus biotype” that reproduces on *Citrus* spp., grapevine and olive (= California biotype C1)—both previous C2 and C4 California biotypes were included in this biotype group, since they cannot reproduce on *P. trifoliata*; but they were detected on olives in Italy;
- the “Mediterranean biotype” is very close to the “Citrus biotype” for its host range except olive—this biotype has

been found in all olive-producing Mediterranean countries, in South Africa and possibly in India;

- the “Grass biotype” was reported in Florida only, where it reproduces only on *Andropogon rhizomatus*, but this biotype corresponds to the new species *T. graminis* that only parasitizes monocots [128].

The limits of these biotypes are rather unclear. C1 and C2 reproduce on grapevine and olive, whereas C3 affects grapevine but not olive, and C4 reproduces only on olive [124]. Furthermore, C1 does not reproduce on *P. trifoliata* and olive [126], while other studies showed that it is the only biotype able to reproduce on olive [127]. Moreover, *T. semipenetrans* was found in olive roots in northern Italy where citrus is not cultivated [127]. C1 has not been reported in Spain [129] either in orchards or in nurseries [27].

Histological disturbances induced by these nematodes in olive roots are similar to those observed on citrus [127]. However, life history traits vary from one variety to another. C1 is more aggressive and reproduces faster on cvs. Manzanilla and Ascolana than on other cultivars, and much more than on *Citrus sinensis*, delaying plant growth [130]. Similarly, C1 populations significantly reduced cv. Frangivento root growth, even no reproduction was observed one year after infection [126].

3.6. *Rotylenchulus* spp. (reniform nematodes)

Reniform nematodes (RN) are semi-endoparasites that infect herbaceous and woody plants, especially in tropical and subtropical regions. Three RN species were detected on olive (Table 1 and Fig. 5): *R. macrodoratus* [131], *R. macrosoma* [62,132] and *R. reniformis* [103,133]. These species were registered in Algeria [104], Greece [103], Israel [134], and Italy [95,131,135], but their pathogenicity on olive remains poorly documented [53].

Non-mature females penetrate the roots and remain in the epidermis on feeding sites [61,136,137]. Mature female produce a gelatinous matrix, and lays eggs within it [62].

Distribution of *R. macrodoratus* is restricted to Mediterranean countries, especially to France, Greece, Italy, and Malta. It is the most common RN species in Italy, where it has been detected in about 20% of the samples in the South and 22% somewhere else [38,95].

R. macrosoma is more abundant on wild olive in southern Spain [62], but it has not been detected in nurseries in the same region [27]. It is able to reproduce on common olive cultivars (Picual, Arbequina). Potential damages may occur in irrigated sandy soils for 0.5–1 nematode/cm³ population levels, but infected plants show no symptoms [62].

Feeder cell disturbances are different according to RN species: *R. macrodoratus* and *R. reniformis* induce extended mononucleated cells [131], whereas *R. macrosoma* induces syncytia [62]. High infection with *R. reniformis* leads to weak tree growth [103].

3.7. *Heterodera* spp. (cyst nematodes)

Cyst nematodes (CN) are sedentary endoparasites. Females lay eggs inside them which remain as a persistence structure, and/or in a yellow gelatinous matrix.

When they die, their cuticle hardens, forming a cyst that may persist in the soil for several years in the absence of host plant [66], and hatch when soil conditions are favourable. CN are widespread and economically important nematodes. Very few species infect woody plants [138]. They were not recorded as olive parasites until *H. mediterranea* was described on cultivated olive (cv Manzanilla) in Spain [64]. More recently, *H. avenae* was detected in olive nurseries on grasses associated with substrates [27].

H. mediterranea differs from other CN species by its semi-endoparasitic behaviour which would be explained by the woody structure of olive roots [64]. Identical observations have been reported on pistachio tree [63].

Infected olive trees do not exhibit any symptoms on leaves, even in heavily infested soils. However, necrosis and root distortion were observed near root penetration sites [64], suggesting that *H. mediterranea* could contribute to plant decline in young orchards and nurseries [64,66,139], where high densities occur [66].

3.8. Other plant-parasitic nematodes

Trichodoridae nematodes are also frequently detected on olive trees. They are polyphagous root ectoparasites and are distributed worldwide. They cause damage to a wide range of crops and natural vegetation by direct feeding on roots, causing stunted roots. Some species (13%) are known as Tobravirus vectors, resulting in important yield losses [140]. Three species of *Paratrichodorus* and 10 species of *Trichodorus* were recorded on olive trees, especially in Italy [141,142], Portugal [143], and Spain [27,42,140] (Table 1).

Nematodes belonging to the Telotylenchidae family are not major crop parasites, although they are mostly associated with several crops. The genus *Tylenchorhynchus* seems to be the most diverse, with 13 species recorded in different olive-producing areas, mainly in the Mediterranean Basin (Table 1 and Fig. 5).

Gracilacus peratica (family Paratylenchidae) is an ectoparasite detected on olives in Italy [36,95,144,145], Spain [42], and Portugal [146] (Table 1). Juveniles, young females and males were detected only in soil, while mature females were observed in roots. Cell walls become thick and lignified [145]. Pathogenicity processes on olive are not known [53]. *G. teres* was also detected on olive in Spain [147].

Ogma spp. nematodes are sedentary ectoparasites. *O. rhombosquamatum* has been reported in different orchards in Spain [42], Italy [38,148], Jordan [40], and Portugal [146] (Table 1). Nematode infection induces cortical cell wall thickening and cortex and epidermis necrosis.

4. Management of plant-parasitic nematodes on olive

4.1. Exclusion of PPN infection

Given that PPN are parasites living in the soil and/or in roots, olive tree protection relies first on the use of healthy plant material transplanted in a soil free of these parasites. That is why the first step in avoiding PPN on olive starts in nurseries where the best conditions for PPN multiplication are met, and from where they could be widely disseminated in orchards [53]. In that way, most

infestations by RKN and LN nematodes in olive orchards originate from unsanitized plant material produced in uncertified nurseries [149]. Chemical disinfection of substrates is a suitable approach for production of nematode-free nursery stock [68]. Although laws have been established by the European Union for healthy olive seedlings certification [150], their application is unfortunately still limited [68]. Cultural practices such as fertilization, weed control with herbicides, could lead to increased populations of NPP [17]. On the other hand, intensive production systems (high-yield varieties, high-density plantations, permanent grassing, irrigation, fertilization, etc.) induce environments suitable for the development of soil-borne diseases caused directly or indirectly (e.g., verticillium wilt) by PPN [68]. These cropping systems could lead to PPN communities dominated by the most damaging species to olive tree.

4.2. Plant resistance

The use of resistant cultivars is an effective alternative to chemical control of PPN [151], but their productivity in olive fruits and oil can be a limiting factor to use them [53]. Some cultivars are resistant or tolerant to some species of *Meloidogyne*, *Pratylenchus*, and *Tylenchulus*.

Olive varieties can be distinguished as susceptible (nematode reproduction and plant disturbance), tolerant (nematode reproduction without plant disturbance) and resistant (no nematode reproduction) to RKN. Cvs. Manzanillo and Ascolano are very susceptible to *M. javanica* [54]. But they are resistant to *M. arenaria* and *M. hapla* [55]. In nurseries, Arbequina, Cornicabra, Hojiblanca, Manzanilla and Picual are the most susceptible cultivars to *M. arenaria*, *M. incognita*, and *M. javanica* [27].

Olive cultivars show also various reactions towards *P. vulnus*. This species does not reproduce on cv. Verdalion [82] while cvs. Ascolana and Manzanilla are very susceptible [55]. However, cv. Ascolana is more tolerant than cv. Manzanilla [53]. The host–parasite interaction would also depend on the diversity of both *P. vulnus* geographical strains and host plants on which they were detected [152].

Several olive cultivars are susceptible to *X. index* except cvs. FS17 and DA12I that exhibited resistance [121].

Comparing CN reproduction traits, such as soil (juveniles and cysts) and root (juveniles) infestation levels and reproduction rates on different olive cultivars, cv. Arbequina appeared as the most susceptible cultivar to *H. mediterranea* [68].

These resistant varieties can be also used as stocks for susceptible cultivars that have yield interests.

In that way, resistance screenings were developed, and *in vitro* tests have proved to be valid alternatives to traditional resistance screenings trials [151].

4.3. Soil solarisation

This method was successfully used to control soil-borne parasites: fungi [153,154], bacteria [155], and PPN [156]. It can be an efficient method especially in the Mediterranean Basin, where climatic conditions are optimal with high

summer temperatures. In nurseries, soil solarisation can therefore be used for controlling RKN in substrates [157], affecting 95% of *M. incognita* eggs up to a 40-cm depth after a 10-day application. However, the gelatinous egg mass matrix protects them against several environmental stresses. Then, *Meloidogyne* spp. is sometimes heat-tolerant and difficult to control by soil solarisation [158]. This method may also be useful to control other PPN, e.g., *P. vulnus* and *P. penetrans*, and other soil-borne pathogens as *Verticillium dahliae* [157].

4.4. Biological control

4.4.1. Organic residues

The addition of forest residues provides organic matter that could contribute to PPN suppression, by increasing soil microflora [159]. Composted dry cork, especially in nurseries, may be effective for *M. incognita* suppression, due to the toxicity of released products (e.g., ammonia, phenolic compounds) [160].

4.4.2. Biofumigation

Some plants (e.g., *Brassica oleracea*, *B. nigra*, *B. juncea*, *Raphanus sativum*, *Crambe abyssinica*, *Sorghum* spp.) are able to release volatile biocides after ploughing in soil that could have toxic effects on PPN. Such soil biofumigations are ecological alternatives to chemical fumigation against PPN and other soil-borne pathogens [161]. Trials to assess the effect of *Sorghum sudanense* on the suppression of *M. incognita* showed a significant reduction in final nematode populations 30 to 60 days after treatment [68]. Brassicaceae species showed suppressive effects on *M. incognita* parasitizing tomato plants by reducing root galls and nematode reproduction [162]. The association of these plants with olive trees could be a way to reduce PPN populations.

4.4.3. Microbial control

Soil inoculation with mycorrhizal fungi can be a useful practice to reduce RKN populations. An early inoculation of olive stocks cvs. Arbequina and Picual with *Glomus intraradices*, *G. mosseae* and *G. viscosum* enhanced plant-growth and controlled *M. incognita* race 1 and *M. javanica* populations by inhibiting nematode reproduction and reducing root galling [163]. Other microbial antagonists to PPN (such as nematophagous fungi and *Pasteuria penetrans*) can be also used for nematode suppression.

5. Prospects: better knowledge on PPN biodiversity for better management

This review clearly shows that the diversity of the PPN communities strongly differ across countries. Considering olive protection, PPN control strategies appear to be developed in each Mediterranean producing country according to its own ecological and agronomical constraints, and the PPN diversity encountered.

As seen above, most of the researches emphasized studies targeting some PPN species. Because resistance and biocontrol are specific to emblematic species and because PPN are everywhere found in communities (i.e. species

mixtures), these control strategies may induce changes in nematode communities, but do not necessarily decrease their overall pathogenicity. Consequently, such biological strategies may be unsustainable from an ecological point of view [164], and they must focus on community rearrangements instead of reducing the most pathogenic species.

Considering olive tree history and diversity of associated PPN around the Mediterranean Basin, local PPN communities could be originating from historical mixtures set up with native (before olive introduction) and invasive (with rootstocks from oleasters) communities. Olive domestication in the past, and breeding and cropping now could explain the present distribution of PPN. Consequently, management strategies must take into account the spatial distribution of the PPN diversity all around the Mediterranean Basin (regional scale) and in each olive-producing country (local scale). A special focus on PPN communities associated with wild olive would give information about trade-offs between PPN communities and olive trees in non-disturbed situations. Moreover, investigation about co-adaptation between PPN and olive tree (wild and varieties) would provide new insights into assembling mechanisms between PPN and olive trees. In this way, a large survey with high sampling efforts is initiated [165] in order to evaluate as precisely as possible the diversity of PPN communities parasitizing wild and cultivated olive, and to target the historical (evolution) and environmental (cropping systems, olive varieties, soil factors) filters involved in community patterns. As a confirmation, interesting effects of olive genotypes on PPN community patterns were observed [17]. Such observations could lead to the development of new management strategies in a diversity-conservation approach.

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