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# SCIENTIFIC REPORTS



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## T cell immunity to Zika virus targets immunodominant epitopes that show cross-reactivity with other Flaviviruses

C. J. Reynolds<sup>1</sup>, O. M. Suleyman<sup>1</sup>, A. M. Ortega-Prieto<sup>1</sup>, J. K. Skelton<sup>1</sup>, P. Bonnesoeur<sup>2</sup>, A. Blohm<sup>1</sup>, V. Carregaro<sup>3</sup>, J. S. Silva<sup>3</sup>, E. A. James<sup>4</sup>, B. Maillère<sup>2</sup>, M. Dorner<sup>1</sup>, R. J. Boyton<sup>1</sup> & D. M. Altmann<sup>1</sup>

Zika virus (ZIKV) infection has several outcomes from asymptomatic exposure to rash, conjunctivitis, Guillain-Barré syndrome or congenital Zika syndrome. Analysis of ZIKV immunity is confounded by the fact that several related Flaviviruses infect humans, including Dengue virus 1–4, West Nile virus and Yellow Fever virus. HLA class II restricted T cell cross-reactivity between ZIKV and other Flaviviruses infection(s) or vaccination may contribute to protection or to enhanced immunopathology. We mapped immunodominant, HLA class II restricted, CD4 epitopes from ZIKV Envelope (Env), and Non-structural (NS) NS1, NS3 and NS5 antigens in HLA class II transgenic mice. In several cases, ZIKV primed CD4 cells responded to homologous sequences from other viruses, including DENV1–4, WNV or YFV. However, cross-reactive responses could confer immune deviation - the response to the Env DENV4 p1 epitope in HLA-DR1 resulted in IL-17A immunity, often associated with exacerbated immunopathogenesis. This conservation of recognition across Flaviviruses, may encompass protective and/or pathogenic components and poses challenges to characterization of ZIKV protective immunity.

Zika virus (ZIKV) is a Flavivirus, first discovered in 1947 in Uganda and named after the Zika Forest<sup>1–3</sup>. The current outbreak has been associated also with a large increase in birth defects, ‘congenital syndrome associated with Zika virus infection’, notably microcephaly<sup>1,2</sup>.

There is a pressing need to understand innate and adaptive immune mechanisms protecting against the virus. Human immunity to this virus had not been extensively researched until the current outbreak and analysis has posed important challenges. With several Zika vaccine candidates poised to enter trials, there is an imperative to have agreed measurables to serve as correlates of protection<sup>4</sup>. In ZIKV infection (as indeed for most other Flavivirus infections) we lack the datasets of T cell response patterns that might offer us the immune correlates of differential outcome after exposure; this is equally true whether considering outcomes after natural exposure or in vaccine trials. Neutralizing antibodies, CD4 and CD8 immunity might be envisaged as important. However, knowledge of immunity to other related Flaviviruses such as West Nile virus (WNV), Japanese encephalitis virus (JEV) and Dengue virus (DENV), suggests that it may be predicted that components of anti-viral antibody, CD4 or CD8 responses could themselves be highly pathogenic<sup>5–8</sup>. The pathogenesis of both WNV and JEV infection encompasses T cell mediated pathology<sup>5–7</sup>. Another confounder to simple analysis of Zika virus (ZIKV) immunity is the issue of virus cross-reactivity. In several of the countries in which ZIKV currently poses a threat, there may be as many as 10 different, related Flaviviruses circulating, including DENV serotypes 1–4 and Yellow Fever virus (YFV), as well as YFV vaccination being widespread<sup>9</sup>. This raises important issues. First, that antigenic cross-reactivity has imposed a confounder for design and roll-out of reagents for unequivocal serodiagnostics and, therefore, for seroprevalence reporting<sup>10</sup>. Second, that the impact of anti-viral cross-reactivity has not been taken into account. A simple view would be that a high level of immunity in these populations to some conserved

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HLA transgenic strain	ZIKV antigen, T cell epitopes identified		
	NS1	NS3	NS5
HLA-DR4	p18 [171–190]	p12 [111–130] p28 [271–190] p29 [281–300] p42 [411–430] p51 [501–520]	p22 [211–230] p23 [221–240] p26 [251–270] p28 [271–290] p29 [281–300]
HLA-DR1	p28 [271–290]	p15 [141–160]	—

**Table 1.** CD4 T cell epitopes identified from the Zika virus protein antigens NS1, NS3, and NS5 in HLA-DR1, and -DR4, transgenic lines.

or partially conserved antigen sequences may offer a degree of protection from ZIKV infection. An alternative, non-mutually exclusive hypothesis, is that as with antibody dependent enhancement (ADE) between DENV serotypes, prior immunity to related viruses may potentiate a pathogenic response to subsequent ZIKV exposure or vaccination (or indeed, that ZIKV immunity will alter responses to subsequent challenges with viruses such as DENV or YFV)<sup>4,11</sup>. Such considerations make detailed mapping of interactions between the respective anti-viral repertoires a priority.

Recent studies from mouse models, non-human primates and exposed humans have started to describe features of adaptive immunity to ZIKV<sup>4,11–16</sup>. Furthermore, clues can be extrapolated from previous studies with other Flaviviruses<sup>17</sup>. Attempts to model acute infection in mice have encompassed neonatal or in utero infection of wild-type strains as well as infection of type I interferon receptor knockout strains. These models variably involve central nervous system (CNS) infiltration by CD4 (mainly T helper 1 (Th1)) and CD8 T cells, sometimes with associated neuropathology. In two separate studies, analysis of ZIKV CD8 IFN $\gamma$  production highlighted an immunodominant epitope from envelope, in the region of amino acids 294–302 of the full length viral sequences<sup>13,14</sup>. Work by Elong Ngono and colleagues showed that murine ZIKV acute infection leads to an increase in activated CD8 T cells<sup>14</sup>. This response was implicated in protection since CD8 ablation exacerbated disease and transfer of primed CD8 T cells decreased viral load. Epitope mapping of CD8 responses by screening recall responses to ZIKV peptides, predicted using the [www.iedb.org](http://www.iedb.org) predictive tools, defined epitopes in Env, prM and NS5 proteins, including the aforementioned Env 294–302 epitope. Acute infection of rhesus macaques showed viral persistence at several sites including CNS (with CNS lymphocyte infiltration), CD4 and CD8 cells activation and a neutralizing antibody response targeted largely to envelope and NS1<sup>18</sup>. A study of ZIKV immunity in ZIKV-exposed humans indicated that while antigenic cross-reactivity between DENV and ZIKV was evident in the antibody repertoire, this was less apparent in the T cell response to Env and NS1<sup>12</sup>.

We set out to put in place an initial dataset of Human leukocyte antigen (HLA) class II restricted responses to ZIKV epitopes that may facilitate analysis of correlates of protection. We used transgenic mouse models that express human class II with a particular focus on HLA-DRB1\*0101 and HLA-DRB1\*0401 since these alleles are relatively common in ZIKV exposed communities in the Americas. For example, the allele frequencies of HLA-DRB1\*0101 and HLA-DRB1\*0401 in Sao Paulo, Brazil are respectively, 0.100 and 0.119 (n = 800,809) [www.allelefreqencies.net](http://www.allelefreqencies.net). Furthermore, we looked for evidence that ZIKV-primed CD4 T cells may recognize cross-reactive epitopes from the homologous antigens in other Flaviviruses. Using immunization of HLA class II transgenics with ZIKV antigen, and then acute infection of AG129 mice, we describe a large array of ZIKV CD4 T cell epitopes, the most comprehensive thus far. ZIKV primed T cells can in some cases respond to related epitopes from other Flaviviruses. However, this cross-reactive recognition can result in ‘immune deviation’ of the response to an altered cytokine profile of enhanced Interleukin 17A (IL-17A). This argues for the potential of prior ZIKV infection or vaccination to skew subsequent immunopathogenesis on exposure to DENV.

## Results

**Mapping immunodominant CD4 T cell ZIKV epitopes in HLA class II transgenic mice.** As a first step to the definition of HLA-restricted, CD4 T cell epitopes from ZIKV, we immunized transgenic mice that carried human class II molecules (all on an H2A $\beta$  –/– background), with recombinant ZIKV antigens NS1, NS3, NS5 and Env (Table S1) and determined recall T cell responses to the full sequence as peptide 20mers overlapping by 10 amino acids (Table S2). The decision to focus initial efforts on NS1, NS3, NS5 and Env was based on extrapolation from the ranking of immunogenicity of CD4 T cell antigens in WNV<sup>6</sup>. HLA-DR1 and HLA-DR4 transgenics were immunized with ZIKV NS1 and recall T cell responses to the full sequence of peptide 20mers overlapping by 10 amino acids analysed. One CD4 epitope was identified in HLA-DR1 transgenics (p28) and one in HLA-DR4 (p18). Immunization with NS3 demonstrated one epitope presented by HLA-DR1 (p15) and 5 by HLA-DR4 (p12, 28, 29, 42, 51). Immunization with NS5 showed no HLA-DR1 epitopes and 5 HLA-DR4 epitopes (p22, 23, 26, 28, 29) (Table 1 and Fig. S1). We had thus shown, with respect to HLA-DR1 and DR4 restricted epitopes from NS1, 3 and 5, that we could define 13 epitopes, 2 from NS1, 6 from NS3, and 5 from NS5.

T cell epitopes from Env were investigated more broadly, by immunization of mice transgenic for HLA-DR1, DR4, DR1501 or DQ8 (Fig. 1). This enabled us to define a further 29 peptide/HLA combinations. Env was the most CD4 epitope-rich of the antigens. Interestingly, some epitopes were presented by more than one class II heterodimer: for example, Env p1 and p41 were presented by HLA-DR1, DR4, and DR1501, while p1 and p38 were presented by HLA-DR4, HLA-DR1501 and HLA-DQ8. The definition of epitopes that can be presented through both HLA-DR and HLA-DQ peptide binding grooves is interesting in light of the relatively distinct structures and binding pockets of these heterodimers. However, these peptides may be presented in different registers by these



HLA transgenic strain	ZIKV antigen, Epitope richness (no. of epitopes [epitopes per 10 kDa])			
	Env	NS1	NS3	NS5
HLA-DR4	10 [2.03]	1 [0.25]	5 [0.73]	5 [0.49]
HLA-DR1	5 [1.02]	1 [0.25]	1 [0.14]	0 [0]

**Table 2.** Epitope density of ZIKV proteins.

Env peptide	DR1	DR3	DR4	DR7	DR11	DR15	DRB5
p1 [1–20]	638	8	118	86	>2 248	175	474
p14 [131–150]	1	250	0.2	7	0.4	0.2	1
p31 [301–320]	3	>1 129	11	1	50	53	2
p41 [401–420]	900	>1 129	114	>359	4	75	65
p42 [411–430]	319	19	>477	119	5	125	54

**Table 3.** ZIKV Env peptide relative binding affinity to HLA-DR molecules. Data were expressed as relative affinity: ratio of the half maximal inhibitory concentration of the peptide to the half maximal inhibitory concentration of the reference peptide. The peptide concentration preventing 50% binding of labeled peptide (IC<sub>50</sub>) was evaluated and data expressed as relative affinity: the ratio of the IC<sub>50</sub> of test peptide to the IC<sub>50</sub> of reference peptide. Mean ± SEM was calculated from two to three independent experiments. Relative affinities of 10 or less were considered high binders, and relative affinities of 10–100 were moderate binders.

**Cross-reactive T cell epitope recognition between ZIKV and other Flaviviruses.** The T cell epitope sequences identified above were submitted to a BLAST search to identify other viruses containing partially homologous sequences. To analyse potential T cell cross-reactivity between the ZIKV epitopes and epitopes from other Flaviviruses, we aligned the sequences of ZIKV Env p1, p6, p7, p8, p14, p25, p29, p31, p32, p38, p41 and p42 against those of DENV1–4, WNV and YFV. The aligned virus sequences contained partially homologous sequences with an amino acid match at 3 to 17 residues out of 20. We synthesized the variant Env epitopes from the sequences of WNV, YFV and DENV1–4, for p1, p6, p7, p8, p14, p25, p29, p31, p32, p38, p41 and p42 (Table 4).

We screened for potential virus cross-reactive recognition by immunization with recombinant ZIKV Env, then screening T cell responses by IFN $\gamma$  ELISpot for recognition of the selected ZIKV epitopes or the homologous sequences from WNV, YFV or DENV1–4. This was done using mice that were transgenic for HLA-DR1, HLA-DR1501 or HLA-DQ8 (Fig. 2A,B,C).

HLA-DR1 transgenic mice were immunized with ZIKV Env and popliteal lymph node cells analyzed *ex vivo* at d10 by IFN $\gamma$  ELISpot for response to the immunodominant epitopes p1, p14, p31, p41 and p42, or to the variant versions we had identified in the sequences of WNV, YFV and DENV1–4 (Fig. 2A). With respect to p1, we observed significant responses to the cross-reactive epitopes from WNV, YFV and DENV1–4. In the case of YFV, the response was low (though positive by our criteria), showing the lowest sequence identity with the ZIKV p1 epitope, with matches at 12 of 20 residues. Tetramer analysis confirmed the presence of HLA-DRB1\*0101 ZIKV p1 specific T cells and a high degree of cross reactivity with HLA-DRB1\*0101 DENV4 p1 at the clonal level (Fig. S3). We also analyzed other cytokines produced during this response, to look for any indication that the cross-reactive peptides might act as an altered peptide ligand, with a modulated cytokine response relative to the ZIKV epitope. We found that the DENV4 p1 epitope (and to a lesser extent, the DENV3 p1 epitope) triggered an IL-17A response that was not present in the response to ZIKV p1 (Fig. 2D). Thus, stimulation of ZIKV Env primed T cells by the homologous DENV4 sequence shifts the response to a Th17 component. From the tools at [www.iedb.org](http://www.iedb.org), the most likely minimal binding register is FVEGMSGGT (anchors underlined); considering the difference between the DENV4 p1 epitope and either the ZIKV or DENV1–3 epitopes, the variant residue that might influence T cell receptor (TCR) recognition (thereby mediating altered peptide ligand (APL) activity) is replacement of the methionine residue (a TCR contact), with the much smaller valine.

We defined no cross-reactive epitope recognition for the ZIKV Env p14, p31, or p42 epitopes. There was no significant cross-reactive epitope recognition for the ZIKV Env p41. When the same experiment was done immunizing HLA-DR1501 transgenics with ZIKV Env, we observed cross-reactive responses to p1 from WNV, YFV, and DENV1, 2 and 3 (Fig. 2B). There was also cross-reactive epitope recognition for the ZIKV Env immunodominant epitopes p25 and p41, but not for peptides p29, p32, and p42. With respect to p25, we observed significant responses to the cross-reactive epitopes from YFV and DENV4 and for p41 we observed significant responses to the cross-reactive epitopes from DENV1 and DENV3. Similar to HLA-DR1 transgenics, when Env immunized HLA-DQ8 transgenics were analyzed for ELISpot responses to the variant p1 peptides, significant responses were observed to each of the p1 variants, that is, from WNV, YFV, and DENV1–4 (Fig. 2C). There was also cross-reactive epitope recognition for the ZIKV Env immunodominant epitopes p7 (DENV1–4) and p8 (WNV and DENV1–3), but not for peptides p6 and p38. Thus, T cell activation by the cross-reactive variant peptides from other Flaviviruses may be to some extent dependent on the presenting HLA class II allele, and may be expected to differ between individuals.

In summary, these studies show examples where HLA class II transgenic T cells, primed to respond to ZIKV Env, can cross-reactively respond to the partially homologous sequences from other Flaviviruses, though results vary depending on the presenting HLA class II heterodimer. This was seen most extensively for ZIKV Env p1. In the example of p1 presentation by HLA-DR1, the DENV4 epitope functioned as an APL, eliciting an IL-17A

Peptide name	Peptide Sequence	Sequence Identity
<b>Peptide 1 variants</b>		
ZIKV P1	IRCIGVSNRDFVEGMSGGTW	
WNV P1	FNCLGMSNRDFLEGVSGATW	13/20
YFV P1	AHCIGITDRDFIEGVHGGTW	12/20
DENV1 P1	MRCVIGNRDFVEGLSGATW	14/20
DENV2 P1	MRCIGISNRDFVEGVSGGSW	16/20
DENV3 P1	MRCVGVGNRDFVEGLSGATW	15/20
DENV4 P1	MRCVGVGNRDFVEGVSGGAW	15/20
<b>Peptide 6 variants</b>		
ZIKV P6	SNMAEVRSYCYEASISDMAS	
WNV P6	ANLAEVRSYCYLATVSDLST	12/20
YFV P6	DGPAEARKVCYSAVLTNVKI	6/20
DENV1 P6	TNPAVLRKLCIEAKISNTTT	8/20
DENV2 P6	KQPATLRKYCIEAKLTNTTT	6/20
DENV3 P6	TQLATLRKLCIEGKITNITT	5/20
DENV4 P6	KEVALLRTYCIEALISNITT	8/20
<b>Peptide 7 variants</b>		
ZIKV P7	YEASISDMASDRCPTQGEA	
WNV P7	YLATVSDLSTKAACPTMGEA	10/20
YFV P7	YSAVLTNVKINDKCPSTGEA	7/20
DENV1 P7	IEAKISNTTDSRCPTQGEA	14/20
DENV2 P7	IEAKLTNTTESRCPTQGEA	10/20
DENV3 P7	IEGKITNITDSRCPTQGEA	12/20
DENV4 P7	IEALISNITTATRCPTQGEA	11/20
<b>Peptide 8 variants</b>		
ZIKV P8	DSRCPTQGEAYLDKQSDTQY	
WNV P8	KAACPTMGEAHNDKRADPAF	9/20
YFV P8	NDKCPSTGEAHLEENEGDN	6/20
DENV1 P8	DSRCPTQGEATLVEEQDANF	12/20
DENV2 P8	ESRCPTQGEPSLNEEQDKRF	10/20
DENV3 P8	DSRCPTQGEAVLPEEQDQNY	13/20
DENV4 P8	ATRCPTQGEPEYLKEEQDQY	12/20
<b>Peptide 14 variants</b>		
ZIKV P14	QPENLEYRIMLSVHGSQHS	
WNV P14	LKENIKYEVAIFVHGPTTVE	6/20
YFV P14	DQTKIQYVIRAQLHVGAKQE	3/20
DENV1 P14	QYENLKYSVIVTVHTGDQH	7/20
DENV2 P14	QPENLEYTIVITPHSGEEHA	9/20
DENV3 P14	QYENLKYTVIITVHTGDQH	7/20
DENV4 P14	QIENLEYTVVVTVHNGDTHA	8/20
<b>Peptide 25 variants</b>		
ZIKV P25	ALVEFKDAHAKRQTVVVLGS	
WNV P25	TLMEFEPPHATKQSVIALGS	10/20
YFV P25	HLVEFEPPHAATIKVLALGN	9/20
DENV1 P25	LLVTFKTAHAKKQEVVVLGS	15/20
DENV2 P25	TLVTFKNPHAKKQDVVVLGS	14/20
DENV3 P25	LLVTFKNAHAKKQEVVVLGS	15/20
DENV4 P25	RMVTFKVPNAKQDVVVLGS	13/20
<b>Peptide 29 variants</b>		
ZIKV P29	KGRLSSGHLKCRKMDKRL	
WNV P29	TVKLTSGHLKCRVKMEKLQL	13/20
YFV P29	LYKLHGGHVACRVKLSALTL	8/20
DENV1 P29	TTTIFAGHLKCRKMDKLT	13/20
DENV2 P29	GNULLFTGHLKCRKMDKQL	13/20
DENV3 P29	GTSIFAGHLKCRKMDKLEL	13/20
DENV4 P29	GNHMFAGHLKCKVRMEKLRI	7/20
Continued		



Peptide name	Peptide Sequence	Sequence Identity
<b>Peptide 31 variants</b>		
ZIKV P31	KGVSYSLCTAAFTFTKIPAE	
WNV P31	KGTTYGVCSKAFKFLGTPAD	9/20
YFV P31	KGTSYKMCTDKMSFVKNPTD	9/20
DENV1 P31	KGMSYVMCTGSFKLEKEVAE	10/20
DENV2 P31	KGMSYSMCTGKFKVVKEIAE	11/20
DENV3 P31	KGMSYAMCTNTFVLKKEVSE	9/20
DENV4 P31	KGMSYTMCSGKFSIDKEMAE	9/20
<b>Peptide 32 variants</b>		
ZIKV P32	AFTFTKIPAETLHGTVTVEV	
WNV P32	AFKFLGTPADTGHGTVVLEL	11/20
YFV P32	KMSFVKNPTDTGHGTAVMQV	8/20
DENV1 P32	SFKLEKEVAETQHGTVLVQV	11/20
DENV2 P32	KFKVVKEIAETQHGTIVIRV	9/20
DENV3 P32	TFVLKKEVSETQHGTILIKV	8/20
DENV4 P32	KFSIDKEMAETQHGTVVVKV	10/20
<b>Peptide 38 variants</b>		
ZIKV P38	NSKMMLDPPFGDSYIVIG	
WNV P38	NAKVLIELEPPFGDSYIVVG	14/20
YFV P38	DDEVLIENPPFGDSYIIVG	11/20
DENV1 P38	EKPVNIETPPFGESYIVVG	10/20
DENV2 P38	DSPVNIEAEPFGDSYIIVG	11/20
DENV3 P38	DEPVNIEAEPFGESNIVIG	10/20
DENV4 P38	NSVTNIELEPPFGDSYIVIG	15/20
<b>Peptide 41 variants</b>		
ZIKV P41	HRSGSTIGKAFEATVRGAKR	
WNV P41	HKSGSSIGKAFTTTLKGAQR	13/20
YFV P41	HKEGSSIGKLFQTMMKAER	11/20
DENV1 P41	FKKGSSIGKMFEATARGARR	13/20
DENV2 P41	FKKGSSIGQMFETMRGAKR	12/20
DENV3 P41	YKKGSSIGKMFEATARGARR	13/20
DENV4 P41	FRKGSSIGKMFESTYRGAKR	14/20
<b>Peptide 42 variants</b>		
ZIKV P42	FEATVRGAKRMAVLGDTAWD	
WNV P42	FTTTLKGAQRLAALGDTAWD	13/20
YFV P42	FTQTMKAERLAVMGDAAWD	12/20
DENV1 P42	FEATARGARRMAILGDTAWD	17/20
DENV2 P42	FETMRGAKRMAILGDTAWD	17/20
DENV3 P42	FEATARGARRMAILGDTAWD	17/20
DENV4 P42	FESTYRGAKRMAILGETAWD	16/20

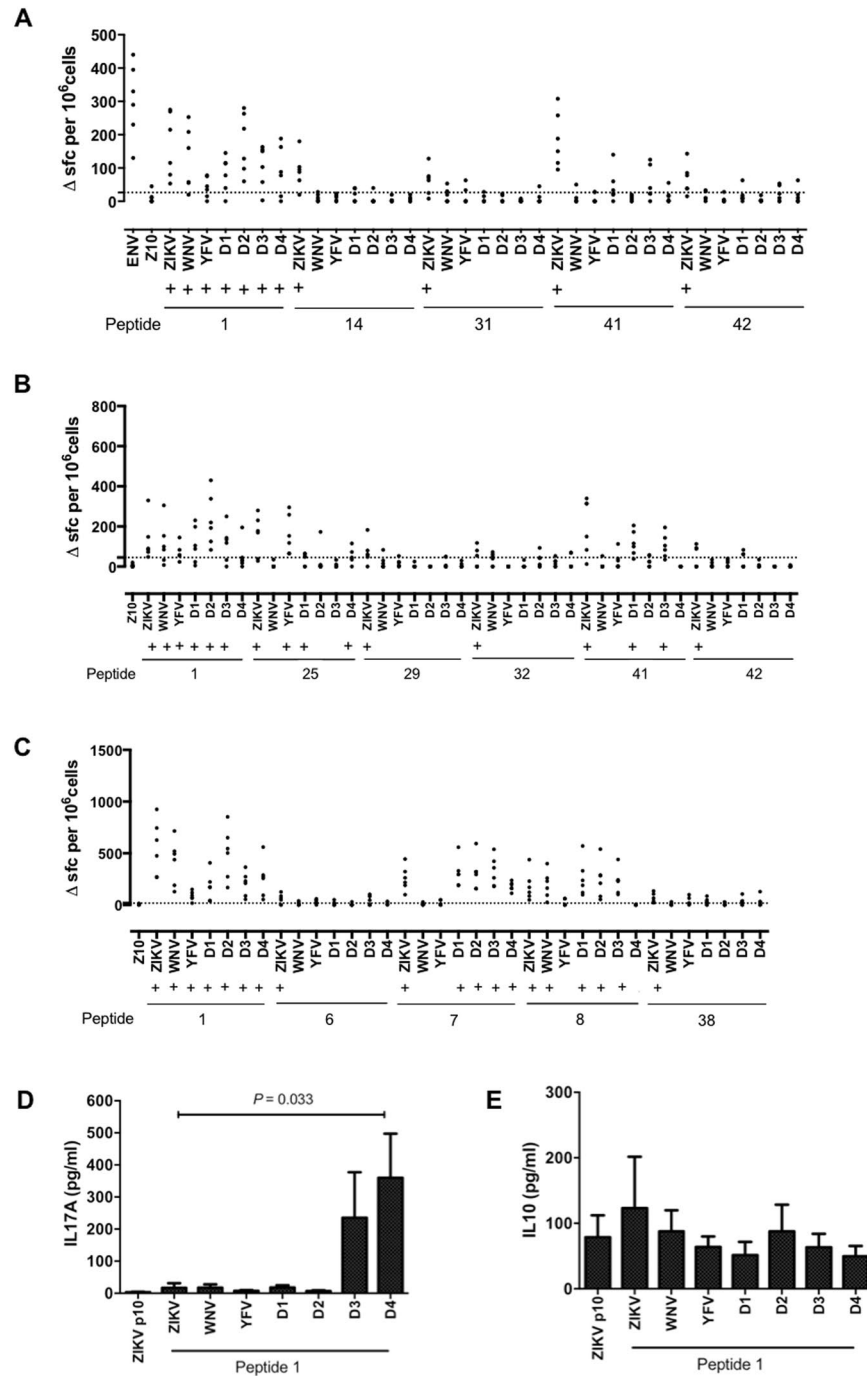
**Table 4.** Aligned the sequences of ZIKV Env p1, p6, p7, p8, p14, p29, p31, p32, p38, p41 and p42 against those of DENV 1–4, WNV and YFV.

response not seen with the ZIKV epitope. Tetramer analysis confirmed a high degree of cross reactivity at the clonal level.

**Antigen recognition, immunodominant epitopes and cross-reactivity in the CD4 T cell response to acute ZIKV infection.** We next investigated ZIKV antigen and epitope recognition in the context of acute ZIKV infection of adult AG129 mice. Mice were infected intraperitoneally with  $10^5$  focus forming unit (FFU) Zika virus (PF13/251013-18) and were culled at day 7, having reached their clinical endpoint, including rapid weight loss and clinical signs of disease. Infected mice exhibited splenomegaly as well as marked liver abnormalities. ZIKV infection was confirmed by ZIKV RNA quantification (Fig. 3A,B).

Splenic T cells were screened by IFN $\gamma$  ELISpot for their response to whole ZIKV Env, NS1, NS3 and NS5 proteins. A strong response was observed to Env but not to any of the non-structural proteins (Fig. 3C).

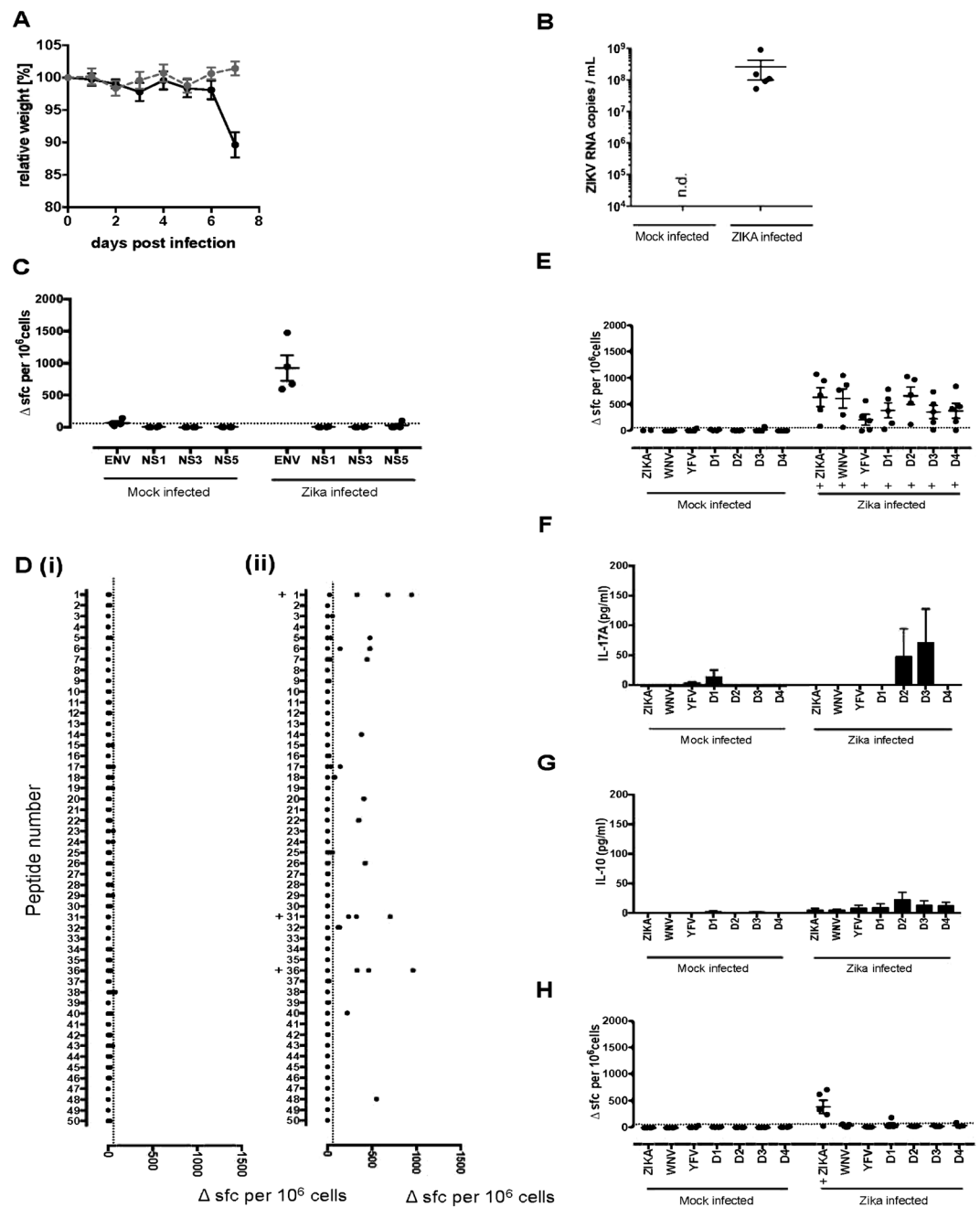
We then mapped the epitope specificity of AG129 splenic cells from d7 ZIKV infected mice. Strong Env peptide specific responses were seen against p1, p31 and p36 (Fig. 3D). We next determined whether priming in the context of acute infection stimulated a T cell response that recognized the cross-reactive p1 epitopes from WNV, YFV, and DENV1–4 (Fig. 3E). Strong responses were observed to each of these epitopes, the response to the WNV and DENV2 epitopes being of similar magnitude to the ZIKV p1 response. In this context, the response to



**Figure 2.** T cells responding to ZIKV envelope protein peptide 1 are crossreactive with variant peptides from other flavivirus species. Mice transgenic for (A) HLA-DR1 (DRB1\*0101)  $n = 6$ , (B) HLA-DR15:01 (DRB1\*1501)  $n = 6$  and (C) HLA-DQ8 (DQB1\*0301)  $n = 6$  were primed with  $25\mu\text{g}$  of recombinant ZIKV envelope protein (Env). 10 days after immunisation, DLNs were assayed by IFN $\gamma$  ELISpot for recall responses to the Env protein, Env peptide 10, Env peptides as shown and the corresponding peptide variants of west nile virus (WNV), yellow fever virus (YFV), dengue virus 1 (D1), dengue virus 2 (D2), dengue virus 3 (D3) and dengue virus 4 (D4). Data are plotted as number of spot forming cells (SFC) per  $10^6$  cells for individual mice. Responses were considered positive (+) if the response was greater than 2 SD above the mean of the response in the absence of any antigen (shown as horizontal dotted line). HLA-DR1 ELISpot supernatants were collected prior to assay development and levels of (D) IL-17A and (E) IL-10 measured by ELISA. Data shown represent mean  $\pm$  SEM. Statistical significance was determined using an unpaired t-test.

DENV3 p1 (and to a lesser extent, DENV2 p1) yielded an APL IL-17A response not seen in response to ZIKV p1 (Fig. 3F) although this result did not achieve statistical significance. No APL IL-10 response to ZIKV p1 variant peptides was seen (Fig. 3G). No cross-reactive responses were identified to ZIKV Env p31 variants (Fig. 3H).





**Figure 3.** ZIKV virus infection in AG129 mice is associated with a strong T-cell response to ZIKV Env peptide 1 which is crossreactive with variant peptides from other flavivirus species. AG129 mice were (i) mock infected ( $n = 4$ ) or (ii) infected intraperitoneally with  $10^5$  FFU of ZIKV (PF13/251013-18) ( $n = 5$ ). (A) Mock (grey circles) and ZIKV infected (black circles) mice were monitored daily for signs of weight loss and culled at 7 days post infection. (B) ZIKV RNA load was quantified by real time PCR. Splenocytes from both groups of mice were assayed by IFN $\gamma$  ELISpot for recall responses to (C) ZIKV proteins Env, NS1, NS3 and NS5, (D) an overlapping panel of 50 Env peptides and flavivirus variant peptides from west Nile virus (WNV), yellow fever virus (YFV), dengue virus 1 (D1), dengue virus 2 (D2), dengue virus 3 (D3) and dengue virus 4 (D4) for peptides 1 (E) and 31 (H). Data are plotted as number of spot forming cells (SFC) per  $10^6$  cells for individual mice. Responses were considered positive (+) if the response was greater than 2 SD above the mean of the response in the absence of any antigen (shown as horizontal dotted line). ELISpot supernatants from peptide 1 variants were also collected prior to assay development and levels of IL-17A (F) and IL-10 (G) measured by ELISA. Data shown represent mean  $\pm$  SEM.

## Discussion

While the incidence of new Zika cases in Central and South America during the 2017 rainy season has, thus far, been lighter than 2016, Zika infection and the associated impacts with respect to congenital Zika syndrome

continue to be a source of public health concern in more than 80 countries. Zika virus disease is notifiable in the USA, with more than 5000 cases notified, mainly of travelers returning from affected areas, with some hundreds resulting from local mosquito-borne transmission<sup>19</sup>. A very large number of candidate Zika vaccines, protective in animal models and based on diverse vaccination platforms, currently await trial<sup>4</sup>. While a number have already been demonstrated to confer protection from acute infection in non-human primate (NHP) challenge models, roll-out of Flavivirus vaccination programs can be non-trivial. Among the many issues is the fact that in many of the affected countries, ZIKV is one of several circulating Flaviviruses<sup>9</sup>. In some cases there is antigen cross-reactivity between the viruses. Whether such cross-reactivity, either ensuing from natural infection or from vaccination, might confer cross-protection and/or immune potentiation of enhanced disease is uncharted. Furthermore, the immunocompetent animal challenge models do not, in most cases, encompass the range of clinical phenotypes seen in human infection, so that protection from these remains hard to evaluate. Knowledge of protective adaptive immune mechanisms is, thus far, based on protection from acute experimental challenge, whereas understanding the consequences of exposure will require understanding of the control of viral persistence in diverse tissues<sup>18</sup>. At least as much as for any other virus genus, outcome after infection by Flaviviruses is driven by a complex interplay between viral pathogenicity and the multiple, protective or pathogenic components of host immunity. In any case, studies of adaptive immunity to ZIKV are in their infancy and there is a lack of any clear consensus of the correlates of protection.

Basic features of ZIKV protective immunity can be inferred from protective transfer studies in mice and from NHP vaccine studies. The data have recently been reviewed by Barouch and colleagues<sup>4</sup>. From the mouse studies, acute infection causes systemic activation of Natural killer (NK), CD4 and CD8 cells as well as the expected antibody response<sup>15</sup>. The ZIKV neutralizing antibody response has been documented and encompasses specificities that are cross-reactive with other Flaviviruses. Passive transfer of antibody can confer protection from acute challenge in the absence of a CD4 or CD8 response<sup>4</sup>. In support of the functional role of the CD8 response, CD8 ablated mice show enhanced susceptibility, while transfer of primed CD8 cells lowers viral load<sup>14</sup>. The T cell cytokine profile appears IFN $\gamma$  rather than IL-17A focused, and an immunodominant epitope in Env has been described. A number of studies have described initial ZIKV T cell epitopes by screening responses of infected humans or mice to candidate epitopes predicted by the predictive tools in [www.iedb.org](http://www.iedb.org). These have included CD4 and CD8 epitopes in Cap, Env, NS1, 2, 3, and 5<sup>12–14</sup>.

In the present study we started by immunizing mice that were endogenous class II-null but HLA transgenic for HLA-DR1 and DR4 with the ZIKV antigens Env, NS1, 3, or 5. Mapping the T cell epitopes within these antigens, we found a hierarchy of epitope density across the transgenic lines of Env > NS5 > NS3 > NS1. This is in contrast to epitope density in YFV vaccinees, where C > Env > NS2A = NS3 > NS1<sup>20</sup>, or DENV infection, where C > NS3 > NS5/NS2A > Env<sup>21</sup>. Overall, we have here been able to identify forty two ZIKV T cell epitopes. However, we note the caveat that CD4 epitopes predicted by HLA class II transgenic studies are not always found identically in humans of the same HLA type. At the initiation of this program, we had based our expectations to some extent on what was already known about human CD4 T cell epitopes from WNV, which showed precisely the same hierarchy<sup>6</sup>. The analogy with WNV T cell immunity also serves to remind that the anti-viral T cell response may encompass both protective and pathogenic phenotypes. In the WNV example, patients with neuroinvasive disease have raised numbers of specific T cells. Furthermore, these WNV responses included a CXCR3 + CCR4 + CCR6- T cell population, secreting IFN $\gamma$  and IL-4, highly correlated with neuroinvasive disease<sup>6</sup>. On the other hand, neuroinflammation in JEV infection may depend on a pathogenic Th17 response under poor Treg control<sup>7</sup>. In terms of antibody responses to Flavivirus epitopes, anti-DENV antibodies can confer potentiation of virus uptake and disease exacerbation – ‘antibody dependent enhancement’ (ADE) – with respect either to subsequent ZIKV or DENV infection<sup>8</sup>.

Several of the defined epitopes were in regions of sequence showing relatively high sequence conservation with other Flaviviruses, including WNV, DENV1–4 and YFV. We therefore took the opportunity to investigate cross-reactive recognition by ZIKV-primed T cells of the variant sequences from other viruses. This was done for the variant Env sequences from WNV, YFV and DENV1–4 for p1, p14, p31, p41 and p42 first identified in HLA-DR1 transgenics. Cross-reactive responses were observed with each of these specificities, most commonly with the WNV sequence. Peptide p1 (AA 1–20) from ZIKV Env was noteworthy since it has now been defined to contain an immunodominant epitope that can be presented both through class I and class II for CD4 and CD8 responses (as we found in our Env primed mice). This Env 1–20 peptide (or core epitopes within this peptide) have been previously described in a number of other T cell studies, including humans infected with DENV1<sup>22</sup>, DENV3 immunized HLA-DR3 transgenic mice<sup>23</sup> as well as CD4 responses of DENV2 Env-immunized mice<sup>24</sup>. The variant Env p1–20 sequences in WNV, YFV and DENV1–4 (as well as other closely related viruses such as Spondweni virus) each retain the residues xxCxGxxxRDFxEGxxGxxW. It is well established in other settings of T cell immunity, notably autoimmune or antiviral responses, that variant stimulatory peptides may interact differentially with TCR contact residues, thus acting as altered peptide ligands (APL) to deviate the phenotype of the effector response<sup>25–28</sup>. This was indeed the case for DENV variants of p1, showing an ability not seen with the wild-type ZIKV sequence, to elicit an IL-17A response. This APL effect was observed both after priming with Env protein and after acute ZIKV infection. Indeed, this peptide encompasses an epitope that has previously been described as a DENV1 T cell epitope that is cross-reactively recognized with DENV2<sup>22</sup>. It remains as yet unclear whether Th17 immunity may be neuropathogenic in ZIKV infection, as for JEV<sup>7</sup>. It might be predicted that there would be an increased risk of immunopathological effects if DENV infection (or vaccination) follows ZIKV infection. Th17 responses have been implicated in the immunopathogenesis of Dengue hemorrhagic fever<sup>29</sup>. Clearly, DENV1–4 and ZIKV show a highly overlapping distribution in Central and South America. Furthermore, there is potential overlap between ZIKV and WNV exposures in North America and in Asia. Possible impacts of T cell cross-reactivity between ZIKV and YFV merits some mention; it is unclear whether any such interaction could

underpin the fact that the peak of ZIKV cases in Brazil during 2016 has been followed in 2017 by the sharpest peak for decades of severe YFV cases<sup>30</sup>.

Our findings serve to underline the point that many of the countries that have been facing a healthcare crisis of ZIKV infections are also at risk from several other, related Flaviviruses. This not only poses challenges to implementation of specific serodiagnosistics due to antigenic cross-reactivity, it also poses currently unanswered questions about the wider implications of cross-reactive immunity. At the most simplistic level, some of the cross-reactive overlaps may serve to confer a degree of cross-protection between infections. It could be speculated that such effects may underpin the observation that in Brazil, which has very high DENV seroprevalence, the vast majority of ZIKV exposures during the past seasons have been asymptomatic. However, the examples of ADE, both within DENV exposures and, potentially, between DENV and ZIKV, along with the possibility that cross-reactive stimulation may deviate T cell phenotypes to a more pathogenic profile, prompts a need for greater efforts to characterize these relationships, as well as greater vigilance and precision in our vaccination programs and understanding of their true correlates of protection.

## Materials and Methods

**Ethics statement.** Mouse experiments were performed within U.K Home Office legislation under the terms of Home Office Project Licenses granted for this work under the “Animals (Scientific Procedures) Act 1986”. Local ethical review and formal approval was also obtained through the Imperial College Ethical Review Process Committee.

**Recombinant proteins and peptides.** Sequences of the envelope protein (Env) and non-structural proteins NS1, NS3 and NS5 of the ZIKV were taken from a recent Brazilian isolate (GenBank accession no. AMH87239.1). Following codon optimisation, recombinant proteins were expressed in *Escherichia coli* and purified by His-Tag. The hydrophobic transmembrane domain (456–504) of Env was not included to improve protein expression, solubility and stability. (Biomatik, Cambridge, ON, Canada) (Supplementary Table 1). Synthetic peptides of 20 aa in length and overlapping by 10 aa were generated for each of the recombinant proteins (GL Biochem, Shanghai, China) (Supplementary Table 2). Flavivirus variants of Env peptides 1, 6, 7, 8, 14, 29, 31, 32, 38, 41 and 42 for WNV (accession no. AFJ05105.1), YFV (accession no. AIZ07887.1), DENV1 (accession no. AKQ00039.1), DENV2 (accession no. AKQ00040.1), DENV3 (accession no. ACO06174.1) and DENV4 (accession no. AKQ00037.1) were also synthesised (Table 4).

**HLA-peptide binding assays.** HLA-DR heterodimers were purified from B cell lines by affinity purification on L243. Peptide binding was evaluated with competitive ELISA using an automated workstation. HLA heterodimers were incubated with biotinylated indicator peptide and serial dilutions of competitor peptide. As reference peptides, unlabelled forms of the biotinylated indicator peptide were used to assess validity in each experiment. The following reference sequences were used as labelled indicator peptides, and their IC<sub>50</sub> values are indicated in parentheses: HA 306–318 (PKYVKQNTLKLAT) for DRB1\*01:01 (3 nM), DRB1\*04:01 (45 nM), DRB1\*07:01 (28 nM) DRB1\*1101 (21 nM) and DRB5\*01:01 (10 nM), MT2-16 (AAKTIAYDEEARRGLE) for DRB1\*03:01 (50 nM) and A3 152–166 (EAEQLRAYLDGTGVE) for DRB1\*15:01 (40 nM). After 24–72 h incubation (37 °C), samples were neutralised with 50 µl 450 mM Tris-HCl (pH 7.5), 0.3% BSA, 1 mM DM buffer and applied to 96-well MaxiSorp ELISA plates (Nunc) coated with 10 µg/ml L243. Bound biotinylated peptide was detected by successive addition of streptavidin-alkaline phosphatase conjugate (GE Healthcare, Saclay, France) and 4-methylumbelliferyl phosphate substrate (Sigma, France). Emitted fluorescence was measured at 450 nm upon excitation at 365 nm. The peptide concentration preventing 50% binding of labelled peptide (IC<sub>50</sub>) was evaluated and data expressed as relative affinity: the ratio of the IC<sub>50</sub> of test peptide to the IC<sub>50</sub> of reference peptide. Mean ± SEM was calculated from two to three independent experiments. Relative affinities of 10 or less were considered high binders, and relative affinities of 10–100 were moderate binders.

**HLA transgenic mouse studies.** This study used HLA class II transgenic mouse lines for the alleles HLA-DR1 (DRB1\*0101), HLA-DR4 (DRB1\*0401), HLA-DR1501 (DRB1\*1501) and HLA-DQ8 (DQB1\*0302), which were all maintained in the context of a homozygous knockout for murine H2-Aβ, as described previously<sup>31–36</sup>. Mice were maintained in individually ventilated cages and used in experiments as young adults. For T cell epitope mapping studies, mice were primed sub-cutaneously in one hind footpad with 25 µg recombinant protein emulsified in Hunters Titermax Gold adjuvant (Sigma-Aldrich). Ten days after immunisation the draining popliteal lymph nodes were removed and disaggregated into a single-cell suspension for ELISpot assays. The frequency of cells producing IFNγ in response to antigen was quantified by ELISpot using a murine IFN gamma ELISpot set kit (862.031.020, Diaclone; 2B Scientific, Oxon, U.K)<sup>31–36</sup>. Briefly, ELISpot plates (MSIPN4550, Merck Millipore, Massachusetts, USA) were coated with anti-mouse IFNγ capture antibody overnight before blocking with skimmed milk. Two × 10<sup>5</sup> cells, from either lymph node or spleen, plus 25 µg/ml protein or peptide were added to wells in HL-1 serum free medium (Lonza, Slough, U.K), supplemented with L-glutamine and penicillin-streptomycin (ThermoFisher Scientific, U.K). Plates were incubated for 72 h, 37 °C with 5% CO<sub>2</sub>. Following assay development, spots were counted on an automated ELISpot reader (Autoimmun Diagnostika, Strasbourg, France). Response frequencies were expressed as Δ spot forming cells (SFC) per 10<sup>6</sup> cells, with an epitope confirmed when the majority of immunised mice responded with a magnitude greater than the mean SFCs in the absence of any antigen +2 SD. Mean + 2 SD background SFC for each ELISpot is indicated by a dotted line. For some experiments, supernatants from ELISpot plates were collected prior to assay development and levels of IL-17A and IL-10 were quantified by ELISA using paired antibodies (DY421, R&D systems, Abingdon, UK) and (3432-1 H, Mabtech, Nacka Strand, Sweden) for IL-17A and IL-10 respectively.

**Flow Cytometry.** Draining lymph node cells from HLA-DQ8 (DQB1\*0302) mice primed 11 days previously with 25 µg/ml ZIKV Env protein were stimulated *in vitro* with 25 µg/ml of the relevant peptide, in the presence of 3 µg/ml Brefeldin A (Affymetrix eBioscience, USA) or with Brefeldin A alone. Cells were stimulated for 7 hours in RPMI medium supplemented with L-glutamine and Penicillin-streptomycin at 37 °C with 5% CO<sub>2</sub>. Stimulated cells were stained with the cell surface antibodies PE-anti mouse CD4 (clone GK1.5 from BD Pharmingen, USA), PE-Cy5 anti-mouse CD8a (clone 53-6.7 from Affymetrix eBioscience, USA), FITC anti-mouse CD3e (clone 145-2C11 from BD Pharmingen, USA) in PBS containing 10% fetal calf serum before fixation, permeabilisation and intracellular antibody staining with Alexa Fluor<sup>®</sup> 700 anti-mouse IFN $\gamma$  $\gamma$  (clone XMG1.2 from BD Pharmingen, USA). R-phycoerythrin (R-PE) and Allophycocyanin (APC) labelled HLA-DRB1\*0101 tetramers loaded with either the ZIKV Env p1 or the DENV4 Env p1 variant respectively were constructed as previously described<sup>6,37</sup>. Labelled tetramers containing no peptide were generated for use as controls<sup>38</sup>. 1 × 10<sup>6</sup> DLN cells from ZIKV ENV primed HLA-DR1 transgenic mice were stained with 10 µg/ml of tetramer in 100 µl DMEM-10 media for 2.75 hours at 37 °C. Cells were then incubated with Fc block (BD Pharmingen, USA) for 15 minutes on ice followed by addition of Fluorescein conjugated anti-mouse CD4 (clone:GKL5, catalogue number:100406, Biologend, USA) for a further 15 minutes. Data was collected using an Attune NxT flow cytometer (ThermoFisher, UK) and analysed using FlowJo software (FlowJo, LLC, USA).

**Zika virus infection in AG129.** AG129 mice were obtained from Marshall BioResources (Hull, UK). Animals were supplied with food and water *ad libitum* and monitored daily for signs of illness. Zika virus (PF13/251013-18, provided by European Commission Seventh Framework Program [FP7/2007-2013] for the DENFREE project under Grant Agreement n<sup>o</sup>282 378) was grown in C6/36 cells, and viral titers were determined by focus formation assay. Mice were infected intraperitoneally with 10<sup>5</sup> FFU in a volume of 100 µL. They were weighed and clinically scored daily to monitor disease severity. After 7 d, mice were euthanized and blood and spleen was collected for further analysis.

**Zika virus RNA quantification.** Zika virus RNA was quantified as previously described<sup>39</sup>. Briefly, RNA was isolated from plasma using a viral RNA extraction kit (Qiagen, UK) according to the manufacturer's instructions and RNA was quantified using a Viia 7 real-time PCR system (Thermo Fisher Scientific, UK) using the TaqMan RNA-to-Ct 1-step kit (Thermo Fisher Scientific, UK). Plasmid pMA-T containing the complete 5'UTR of Zika virus was used for T7 polymerase *in vitro* transcription using the T7 Ribomax Express Large Scale RNA Production System (Promega, UK) and used as RNA standard.

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## Author Contributions

C.J.R. developed and performed experiments, analysed and interpreted data and helped prepare the manuscript. O.M.S., A.B., and V.C. performed experiments and analysed data. A.M.O., J.K.S. and M.D. developed and carried out live ZIKV infection studies. P.B. and B.M. developed, carried out and analysed HLA class II binding studies. E.J. made the HLA class II tetramers and contributed to study design, data analysis and interpretation. J.S.S. contributed to study design and data interpretation. R.J.B. and D.M.A. conceived and designed the study, designed experiments, analysed and interpreted data, and wrote the manuscript. R.J.B. and D.M.A. supervised the research, are co-senior authors and contributed equally to the study. All authors discussed the results and commented on the manuscript.

## Additional Information

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**Competing Interests:** The authors declare that they have no competing interests.

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