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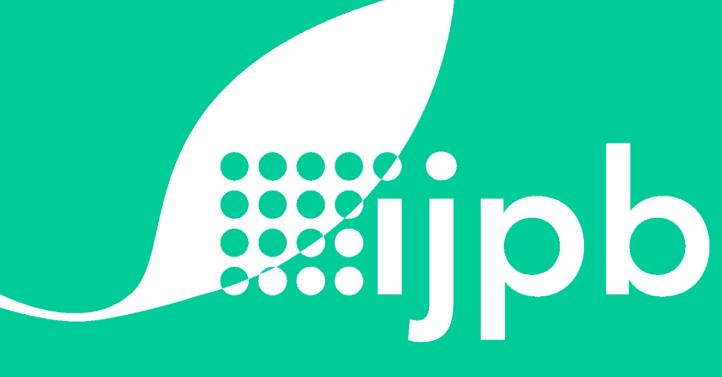
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# **CHARACTERIZATION OF A NITRATE HIGH-AFFINITY TRANSPORT SYSTEM IN BRACHYPODIUM**

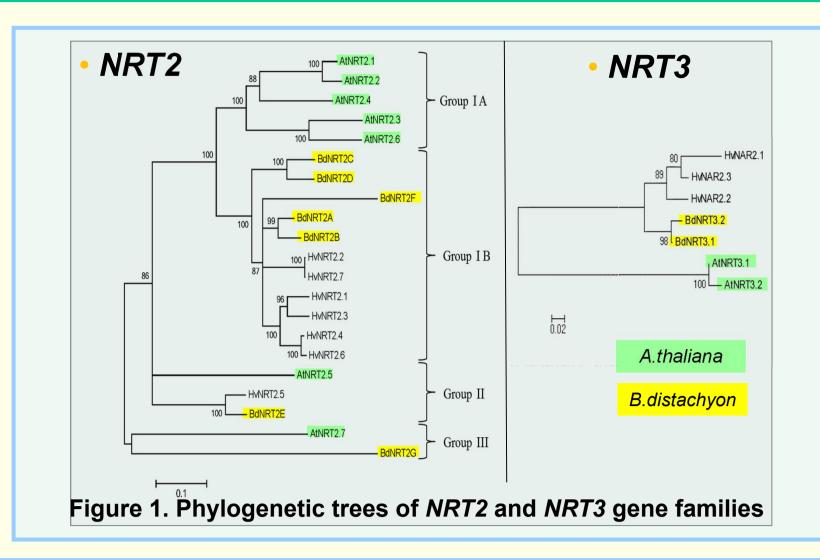
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### Introduction

Nitrogen is an essential element for growth and plant development. Improving the nitrogen use efficiency of crops is a priority for the future in order to limit the fertilizer use and environmental impacts. An efficient nitrate uptake system contributes to improve the nitrogen use efficiency of crops in conditions of low N availability. The High-Affinity nitrate Transport System (HATS) in plants is active in low range of external nitrate and is mediated by a two-component system (the high affinity transporters NRT2 associated to a partner protein NRT3). Using phylogenetic studies, seven NRT2 orthologues and two NRT3 orthologues were found in Brachypodium distachyon (Figure 1) [1].



## **1a)** BdNRT2A/2B and BdNRT3.2 expressions are induced in response to nitrate

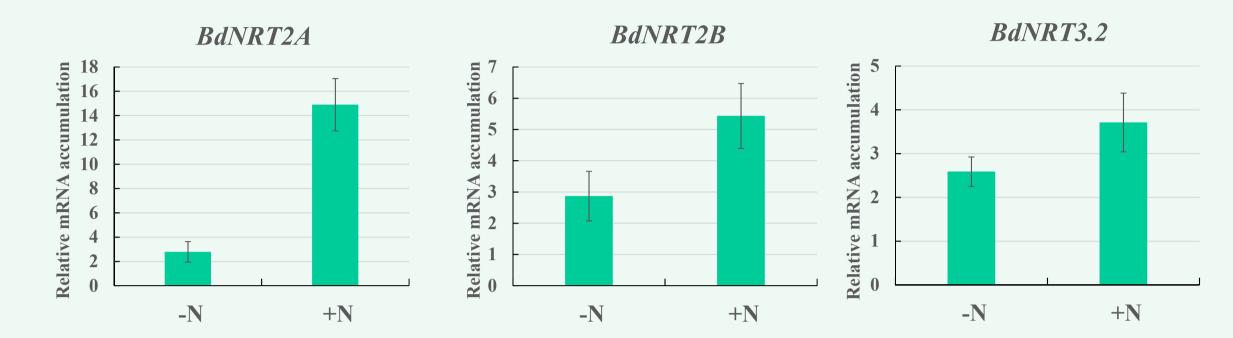


Figure 2. Expression profiles of BdNRT2.A/B and *BdNRT3.2* genes

Real-time RT-PCR performed on roots of 22 day old plants grown in hydroponic on 0.1 mM NO<sub>3</sub><sup>-</sup> for 17 days then on 0 mM  $NO_3^{-1}$  for 4 days

• and then 0 mM (-N)

• or  $1 \text{ mM NO}_3^-$  for 3 h (+N).

### **1b)** BdNRT2A/2B and BdNRT3.2 expressions in response to nitrate availability fits with the characteristics of a HATS activity [2]

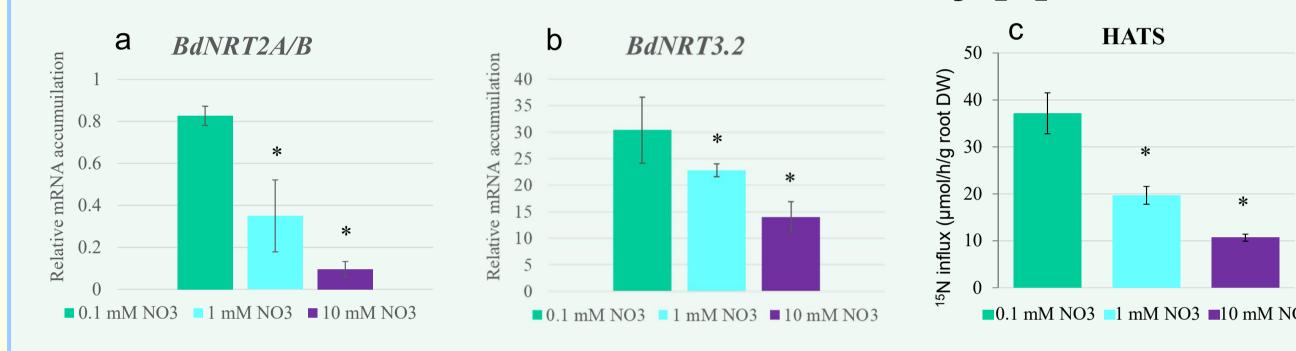


Figure 3. Expression profiles of BdNRT2A/B and *BdNRT3.2* genes and HATS activity

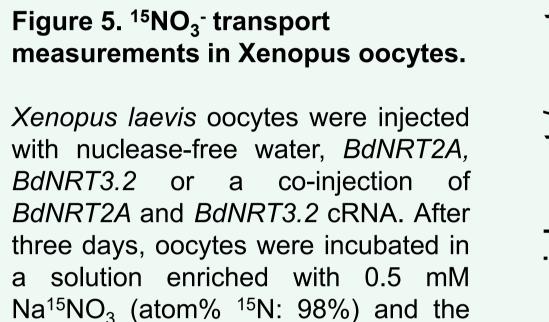
• Real-time RT-PCR performed on roots of 17 day old plants grown in hydroponic on 0.1; 1 or  $10 \text{ mM NO}_3^-$  (a,b).

<sup>15</sup>NO<sub>3</sub><sup>-</sup> influx measurements into roots (0.2mM  $^{15}NO_{3}^{-}$  for 5 min ) (HATS), (c).

**3)** Co-expression of *BdNRT2A* and *BdNRT3.2* is required for an effective nitrate transport in heterologous expression the system Xenopus oocytes

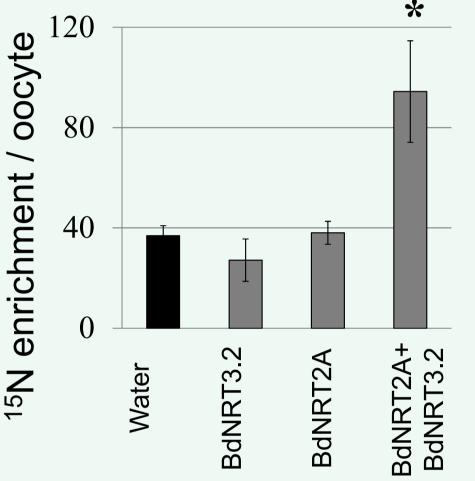
Functional interaction between BdNRT2A and BdNRT3.2 was shown by  $NO_3^-$  uptake measurements in a heterologous expression system.

Only Xenopus oocytes co-injected with BdNRT2A and BdNRT3.2 were able to transport  $NO_3^{-1}$ .



Na<sup>15</sup>N enrichment of individual

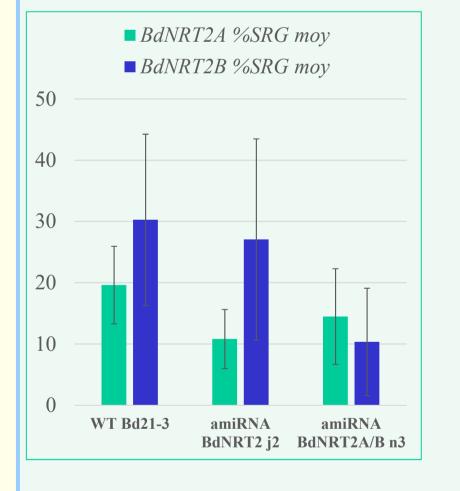
oocytes was measured after 16h.



BdNRT2A/2B and BdNRT3.2 gene expressions were enhanced by low nitrate supply in root in correlation with the HATS activity, and thus are good candidates for HATS activity in Brachypodium.

**2)** BdNRT2A has a major role in  $NO_3^-$  uptake in low N availability (HATS). Growth and HATS activity are lowered in Brachypodium 4) Species specificity of BdNRT2A/BdNRT3 interaction : BdNRT2A is targeted to plasma membrane in presence of BdNRT3.2 but not AtNRT3.1

### mutants defective in *BdNRT2A*



### Brachypodium mutants defective in BdNRT2A or BdNRT2A and BdNRT2B.

- amiRj2 : mutant with an amiRNA construct targeted to **BdNRT2A and BdNRT2B** (55% and 90% of the expression of each gene respectively)
- amiRn3 : mutant with an amiRNA construct targeted to **BdNRT2A, 2B**, (74%, 34%), and <u>2C, 2D, 2F</u>
- 7.1 (and 9.2) : tilling mutant with a STOP codon (W248) in **BdNRT2A W248/STOP**
- az1 and az2 : two azygote lines of the BdNRT2A tilling mutant

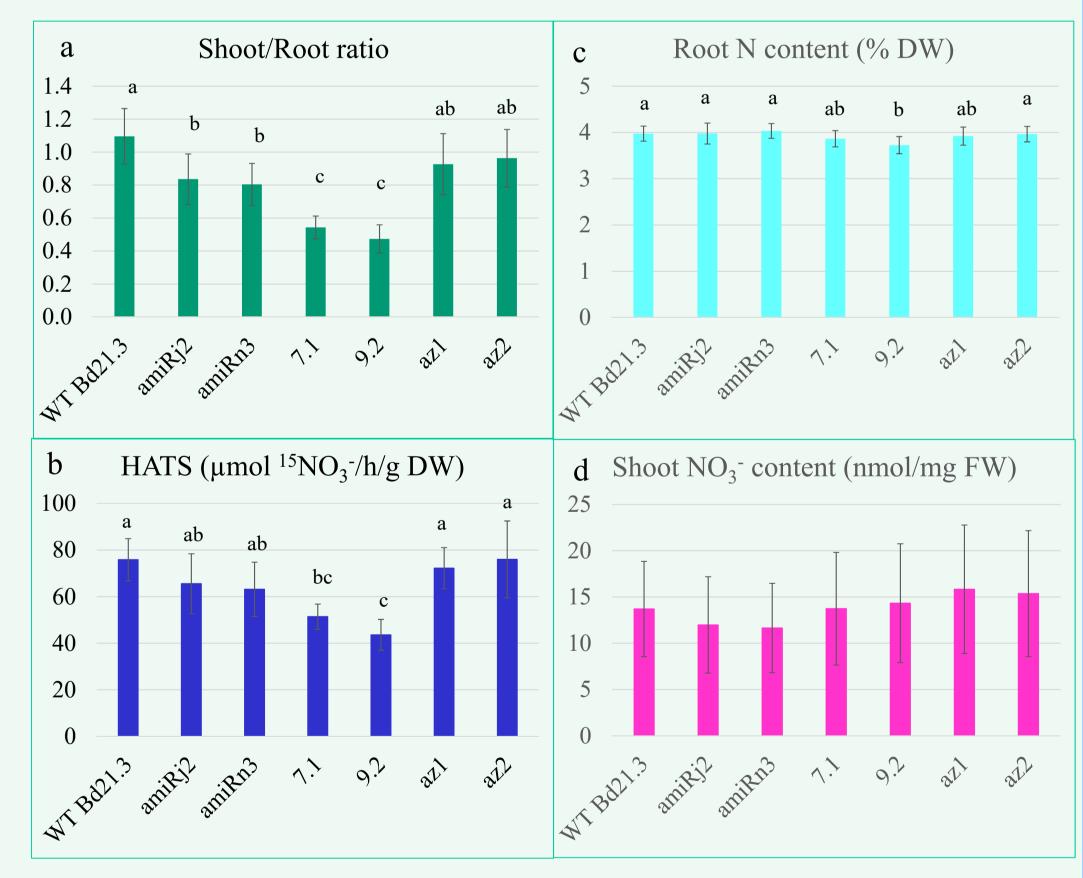


Figure 4. Phenotype of Brachypodium mutants defective in BdNRT2A.

(a) Shoot/root biomass ratio of 3 week-old plants of Brachypodium grown in hydroponics with  $0.2 \text{ mM NO}_{3}$ -. (b)  $15NO_3^-$  (0.2 mM) influx measured during 5 min uptake (HATS). (c) N content in roots (d) Nitrate content in shoots

NRT2 targeting to plasma membrane requires a protein-protein interaction with NRT3. Transient expression expreriments in *atnrt2.1-2* protoplasts showed that BdNRT2A interact with BdNRT3.2 but not with AtNRT3.1.

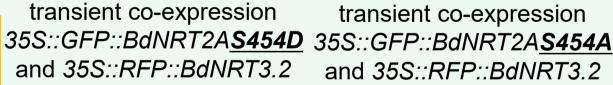
A Ser specifically conserved in the C-terminal extension of NRT2.1 in monocotyledons has been identified as responsible of the NRT2/NRT3 interaction in Barley [3]. Site directed mutagenesis of BdNRT2ASer454 mimicking (or not) a constitutive phosphorylation status of this Ser (S454D or S454A respectively) was used to study the species specificity of the interaction.

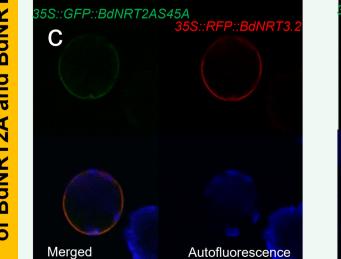
The results showed that Ser454 is not sufficient to explain the species specificity of the interaction.

### atnrt2.1-2 protoplasts transient co-expression of transient expression of 35S::GFP::BdNRT2A 35S::GFP::BdNRT2A and 35S::RFP::BdNRT3.2 b

### atnrt2.1-2 x atnrt3.1 protoplasts

lirected mutagenesis on *BdNRT2ASer454* 





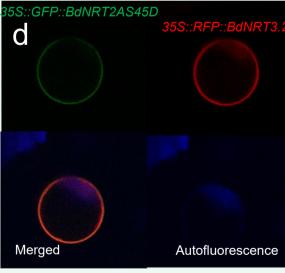


Figure 6. Subcellar localisation of BdNRT2A in Arabidopsis protoplasts observed with confocal imaging.

### Conclusion

Using various approaches, these results show that BdNRT2A and BdNRT3.2 are the main component of the HATS activity of NO<sub>3<sup>-</sup></sub> (up to 30-40%) in Brachypodium and allow an optimal growth in low N conditions. The functional interaction between BdNRT2A and BdNRT3.2 seem species specific.

Cytosol of

### References

[1] Girin T, David LC, Chardin C, Sibout R, Krapp A, Ferrario-Méry S, Daniel-Vedele F. 2014 Brachypodium: a promising hub between model species and cereals. J. Exp. Bot 65: (19) 5683-5696.

[2] David LC, Girin T, Fleurisson E, Phommabouth E, Mahfoudhi A, Citerne S, Berquin P, Daniel-Vedele F, Krapp A, Ferrario-Méry S. 2019 Developmental and physiological responses of Brachypodium distachyon to fluctuating nitrogen availability. Sci Rep. 9(1):3824 [3] Ishikawa S, Yuka Ito, Sato Y, Fukaya Y, Takahashi M, Morikawa H, Ohtake N, Ohyama T, Sueyoshi K. 2009 Two-component highaffinity nitrate transport system in barley: Membrane localization, protein expression in roots and a direct protein-protein interaction. Plant Biotechnology 26, 197–205.

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