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## A non-invasive method to appraise time-dependent effects of venom toxin-based therapeutics on the mouse neuromuscular excitability *in vivo*

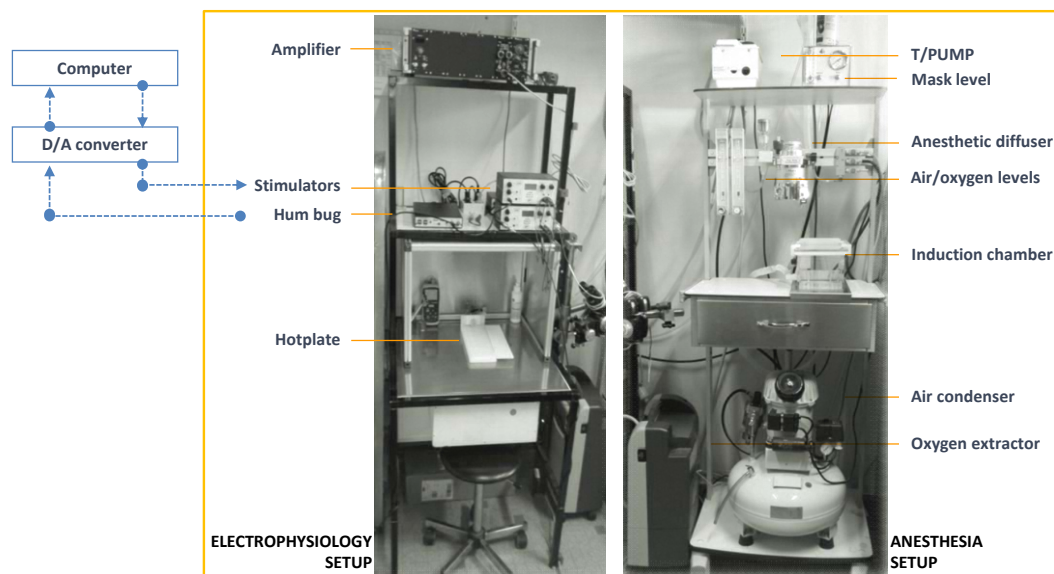
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The conventional methods, usually performed *in vivo*, provide limited information regarding sensory and neuromuscular excitability properties. In the present work, we review the new minimally invasive methods that have been improved to supplement the conventional electrophysiological methods. In particular, the automated sequence of multiple excitability tests developed by Professor Bostock (Qtrac®, Institute of Neurology, London, UK) to assess sensory and neuromuscular excitability in clinical neurophysiology, has been adapted to animal models *in vivo*, initially in rats and, more recently, in mice. This method is particularly appropriate for venom toxin injections and repeated recordings from the same animal cohort, making it a promising tool to appraise the progression of toxin effects, and that of a given treatment, on the sensory and neuromuscular excitability of animal models *in vivo*. The *in vivo* efficiency in mice and long-term follow-up of the effects of sub-lethal doses of spider and *conus* venom toxins will be emphasised. It is indubitable that this method brings specific information regarding changes in ion transfer and membrane properties induced by a given venom toxin which, thereby, will help to a better characterization of toxin action on the sensory and neuromuscular excitability properties, *in vivo*.



Recordings are performed by means of a minimally-invasive electrophysiological method, under isoflurane anesthesia (inhalation). The principle is to electrically stimulate a nerve trunk, by means of surface electrodes (cathode and anode), using two stimulators. In return, the compound action potential, which propagates in either the stimulated nerve or muscle, is collected using very fine needle electrodes.

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