



# Stochastic models help understanding the evolution of partially clonal species and inferring rate of clonality from genetic diversity

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Team DEMEcology: Dynamics, Evolution, Modelling & Ecology

## Clonix consortium:

CNRS, Ifremer, INRAE, IRD, **UACH**, **UC Chile**, University of Kansas (USA),  
UAB (USA), UAY (Mexico), RAS (Russia), CSIRO (Australia)



# Asexuality / Clonality

Reproductive mode in which descendants are genetically identical to their only ancestor at all loci in their genomes, except at sites that have experienced somatic mutations and recombinations.

Richards 2003; DeMeeüs *et al.* 2007

No segregation, No fusion

→  $n$  alleles at each gene are transmitted together to the descendant.

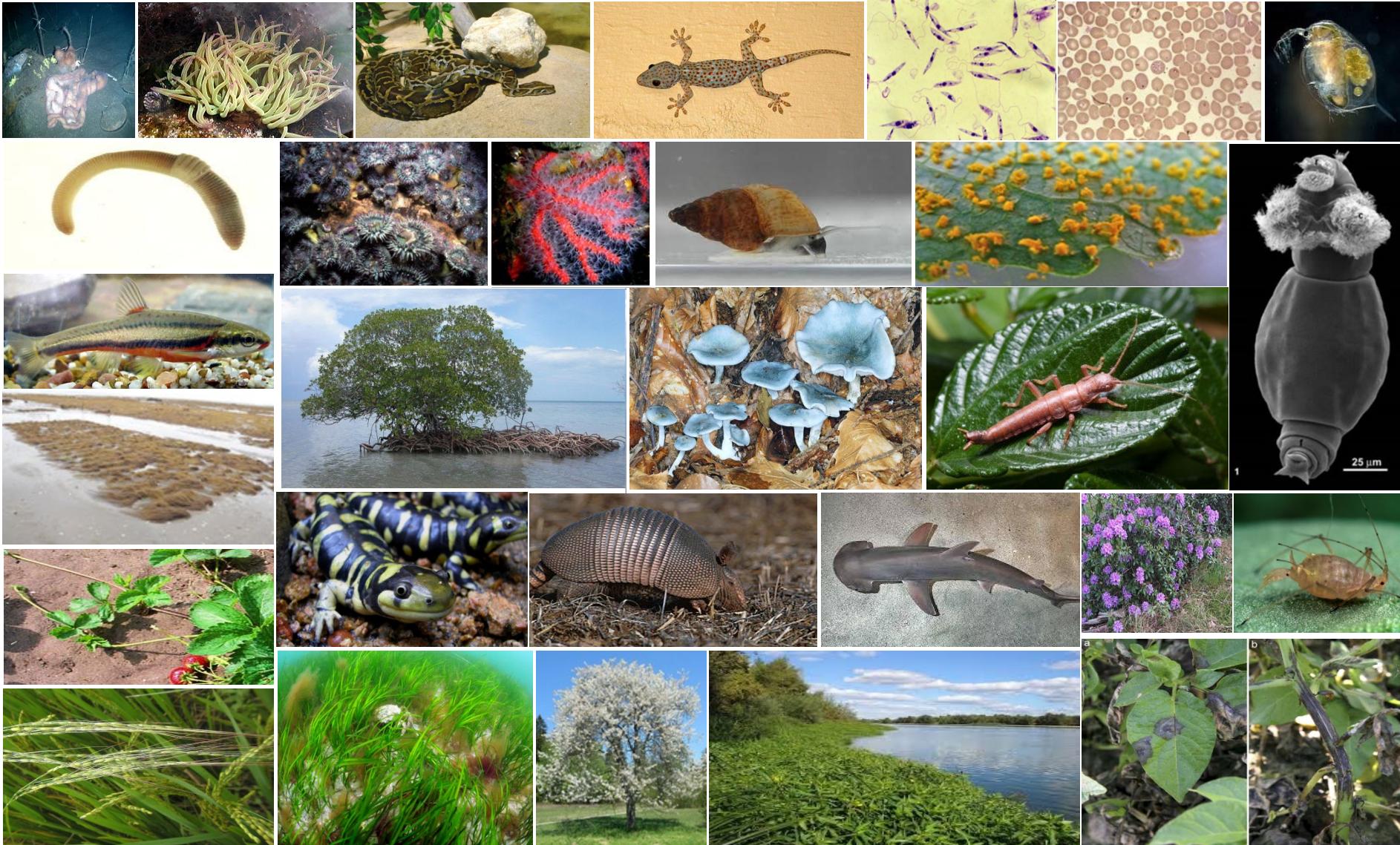


No meiosis (thus no massive recombination)

→ alleles at different genes on a chromatid are transmitted together.

# Why studying clonality?

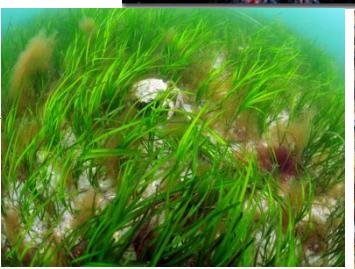
... most Eukaryote clades reproduce using clonality!



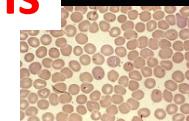
# Species using clonal reproduction: The mainstay of ecosystems and human activities



Engineers



Pathogens



Resources



Invasives



GLOBAL CHANGE, CLONAL GROWTH, AND BIOLOGICAL INVASIONS BY PLANTS

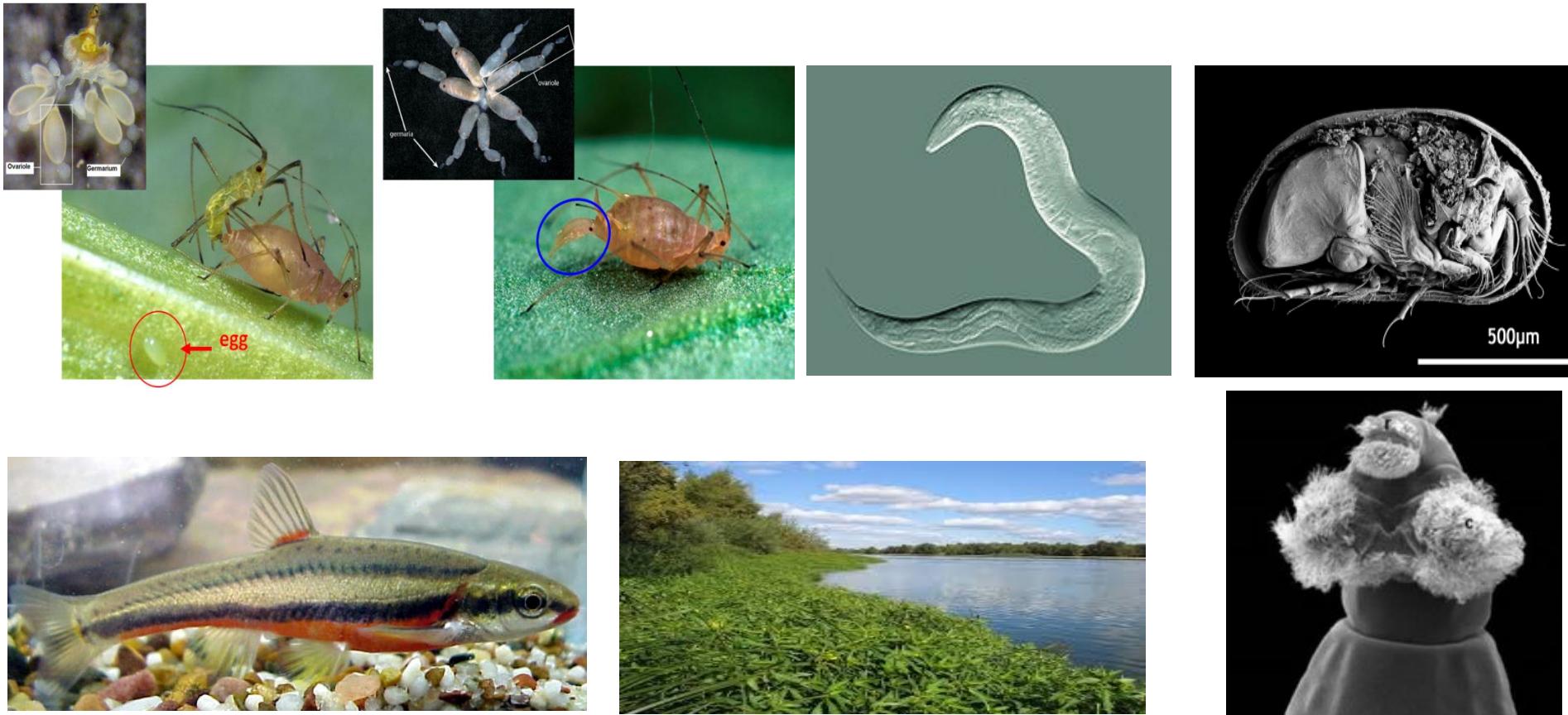
EDITED BY Fei-Hai Yu, Sergio R. Rojas and Peter Alpert  
PUBLISHED IN Frontiers in Plant Science



Pests



# In Eukaryotes: Clonality = Partial Clonality

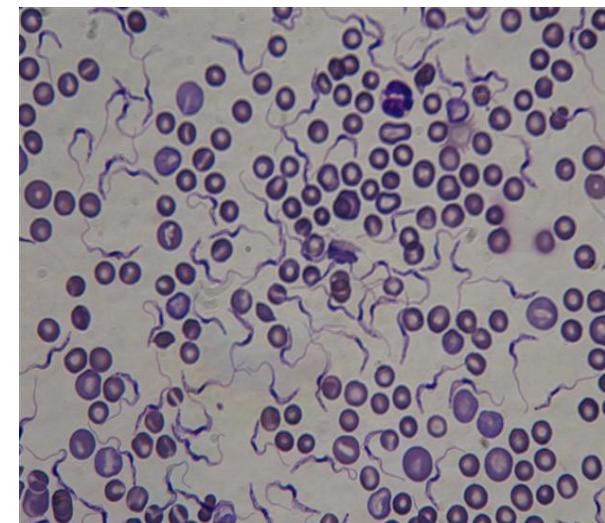
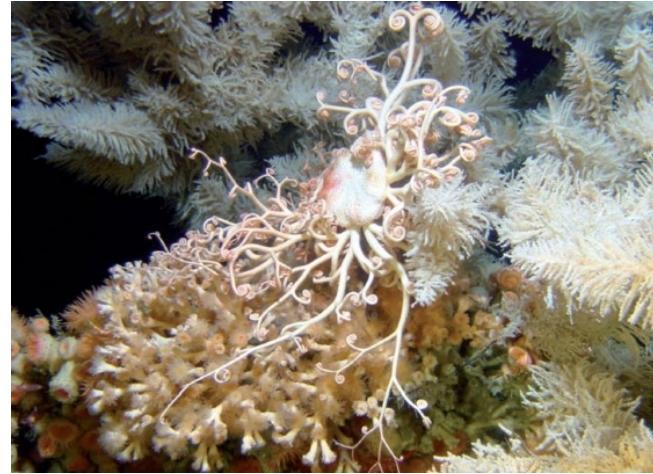


$$\text{Rate of clonality: } c = \frac{\text{nb clonal descendants}}{\text{total nb descendants}}$$

→ How estimating those rates in populations?  
I) Inference

## 1) Identify clonal species, assess their level of clonality

→ troubles: cost of observations, how to deal with cryptic organisms, etc.

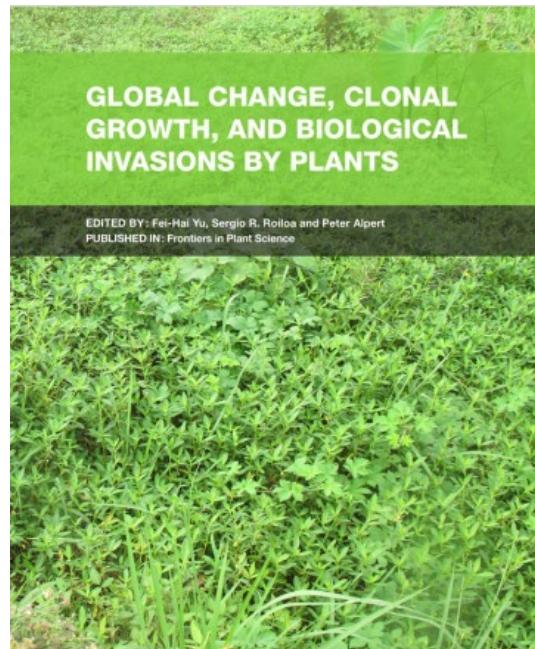


# Understand the effects of partial clonality on ecological and evolutionary dynamics of focal species

- Biotech
- Invasive species under global changes
- Breeding and mass selection

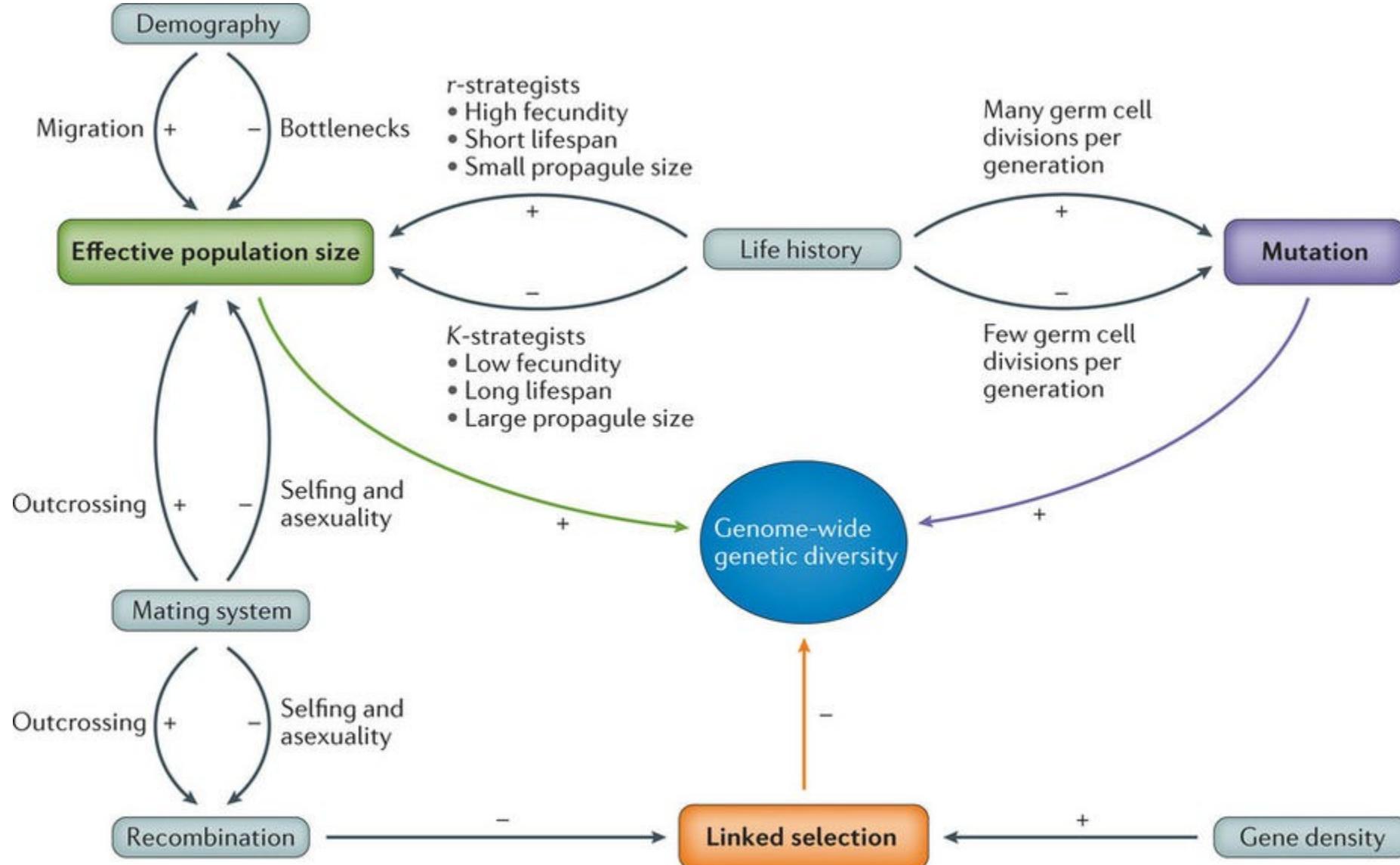
John C. Avise (2015). Clonal reproduction in vertebrate animals. PNAS, 112 (29), p.8867-8873

Baker's conjecture (1955)



→ How evolve genetic diversity and genomes in such species?  
II) Prediction

# DNA = a barcode to record all those effects:



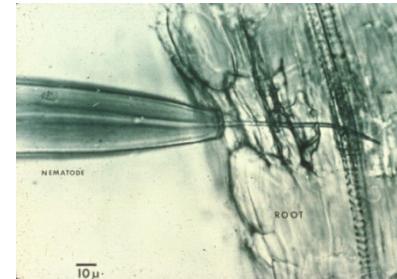
## → What the early models say?

Genetic diversity in partial clonality = sexuality as long as clonality rate is under 90%

$$E[T]_f = 1 + \frac{1}{2\Omega} - \Phi, \text{ where } \Phi = \frac{f(\Omega + f + 1)}{2(2\Omega + 1)}.$$

Marshall & Weir 1979, Orive 1993, Berg & Lascoux 2000, Bengtson 2003, Ceplitis 2003, Balloux et al. 2003, Yonezawa et al. 2004

## → What the data say?



Three main *kuhnian\** anomalies

- 1- over-representation of heterozygotes in partially clonal populations ( $0 < c < 0.9$ )
- 2- Observed genetic variances  $\gg$  predicted variances in highly clonals ( $c > 0.9$ )
- 3- high volatility of genetic indices over time and space  $\neq$  stabilité prédictions

\*Thomas S. Kuhn. (1962). The Structure of Scientific Revolutions. University of Chicago Press.

# Wright-Fisher + clonal rate + genetic drift + reciprocal mutation

Genetic state of a population

State  $t$   
11 AA, 9 Aa, 10 aa

Transition probability

State  $t+1$   
7 AA, 12 Aa, 9 aa

The diagram illustrates the transition from State  $t$  (11 AA, 9 Aa, 10 aa) to State  $t+1$  (7 AA, 12 Aa, 9 aa) through three different reproductive pathways:

- Clonal reproduction:** Represented by the first set of equations under the heading "Ratio Clonality - Sexuality". It shows the probability of each genotype appearing in the next generation given only clonal reproduction.
- Sexual reproduction:** Represented by the second set of equations under the heading "Sexual reproduction". It shows the probability of each genotype appearing in the next generation given only sexual reproduction.
- under infinite pop size:** Represented by the third set of equations at the bottom, which combine the probabilities of clonal and sexual reproduction.

Below these, a general formula for the transition probability is provided, involving factorials and multinomial coefficients.

$$P(s_{aa}, s_{Aa}, s_{AA} / r_{aa}, r_{Aa}, r_{AA}) = \frac{N!}{s_{aa}! s_{Aa}! s_{AA}!} (\pi_{aa}^{n+1})^{s_{aa}} (\pi_{Aa}^{n+1})^{s_{Aa}} (\pi_{AA}^{n+1})^{s_{AA}}$$

Transition probabilities under finite size  $N$

# Wright-Fisher + clonal rate + genetic drift + reciprocal mutation

$\left\{ \begin{array}{l} \text{Taux de reproduction asexué} = c \\ \text{Taux de reproduction sexué} = 1 - c \end{array} \right.$

$$\left\{ \begin{array}{l} P(\text{observer } aa | \text{repro. asexuée}) = \frac{r_{aa}}{N} = p_{aa} \\ P(\text{observer } aA | \text{repro. asexuée}) = \frac{r_{aA}}{N} = p_{aA} \\ P(\text{observer } AA | \text{repro. asexuée}) = \frac{r_{AA}}{N} = p_{AA} \end{array} \right.$$

Let  $u$  being a symetrical mutation rate

$$\left\{ \begin{array}{l} p_{aa}^{(n+1)} = (1-u)^2 \cdot p_{aa}^{(n)} + u \cdot (1-u) \cdot p_{aA}^{(n)} + u^2 \cdot p_{AA}^{(n)} \\ p_{aA}^{(n+1)} = 2 \cdot u \cdot (1-u) \cdot p_{aa}^{(n)} + [(1-u)^2 + u^2] \cdot p_{aA}^{(n)} + 2 \cdot u \cdot (1-u) \cdot p_{AA}^{(n)} \\ p_{AA}^{(n+1)} = u^2 \cdot p_{aa}^{(n)} + u \cdot (1-u) \cdot p_{aA}^{(n)} + (1-u)^2 \cdot p_{AA}^{(n)} \end{array} \right.$$

$$\left\{ \begin{array}{l} P(\text{observer } aa | \text{repro. sexuée}) = q_{aa} = \left( \frac{2r_{aa} + r_{aA} + r_{AA}}{2N} \right) \\ P(\text{observer } aA | \text{repro. sexuée}) = q_{aA} = 2 \cdot \left( \frac{r_{aa} + r_{aA}}{2N} \right) \\ P(\text{observer } AA | \text{repro. sexuée}) = q_{AA} = \left( \frac{r_{AA}}{2N} \right) \end{array} \right.$$

$$\left\{ \begin{array}{l} q_{aa}^{n+1} = \left[ (1-\mu) \cdot p_{aa}^n + \mu \cdot p_{AA}^n + \frac{1}{2} p_{Aa}^n \right]^2 \\ q_{Aa}^{n+1} = 2 \cdot \left[ (1-\mu) \cdot p_{aa}^n + \frac{1}{2} \cdot p_{Aa}^n + \mu \cdot p_{AA}^n \right] \cdot \left[ (1-\mu) \cdot p_{AA}^n + \frac{1}{2} \cdot p_{Aa}^n + \mu \cdot p_{aa}^n \right] \\ q_{AA}^{n+1} = \left[ (1-\mu) \cdot p_{AA}^n + \mu \cdot p_{aa}^n + \frac{1}{2} p_{Aa}^n \right]^2 \end{array} \right.$$

$$\left\{ \begin{array}{l} \pi_{aa}^{n+1} = p_{aa} \cdot c + q_{aa} \cdot (1-c) \\ \pi_{Aa}^{n+1} = p_{aA} \cdot c + q_{aA} \cdot (1-c) \\ \pi_{AA}^{n+1} = p_{AA} \cdot c + q_{AA} \cdot (1-c) \end{array} \right.$$

$$P(S_{aa}, S_{Aa}, S_{AA} / r_{aa}, r_{aA}, r_{AA}) = \frac{N!}{S_{aa}! S_{Aa}! S_{AA}!} (\pi_{aa}^{n+1})^{S_{aa}} (\pi_{Aa}^{n+1})^{S_{Aa}} (\pi_{AA}^{n+1})^{S_{AA}}$$

Computational/calculus complexity:

$$\Upsilon = \frac{(N + \frac{(n_a+pl-1)!}{pl! \cdot (n_a-1)!} - 1)!}{(N! \frac{(n_a+pl-1)!}{pl! \cdot (n_a-1)!})}$$



$N$	$\mathcal{P}$	$\mathcal{L}$	$\mathcal{A}$	$g$	$ S $	memory uptake
20	2	1	2	3	231	420 KB
100	2	1	2	3	5 151	205 MB
500	2	1	2	3	125 751	120 GB
1000	2	1	2	3	501 501	2 TB
20	4	1	2	5	10 626	865 MB
20	2	2	2	9	3 108 105	75 TB
20	2	1	4	10	10 015 005	730 TB
20	2	2	4	100	$9.8 \times 10^{20}$	$6.5 \times 10^{21}$ YB

Table 1: Examples of matrix size based on the Stoeckel-Masson model. Memory sizes are approximate and assume 64-bit accuracy.

## Forward $\hat{I}$ to diffusive SDE approximation

Assuming that  $1-c$  and  $u$  converge to 0 at the speed of  $1/N$

$$dY_t = b(Y_t)dt + \sigma(Y_t)dW_t$$

$$u = \frac{\alpha}{N} \quad c = 1 - \frac{\beta}{N}$$

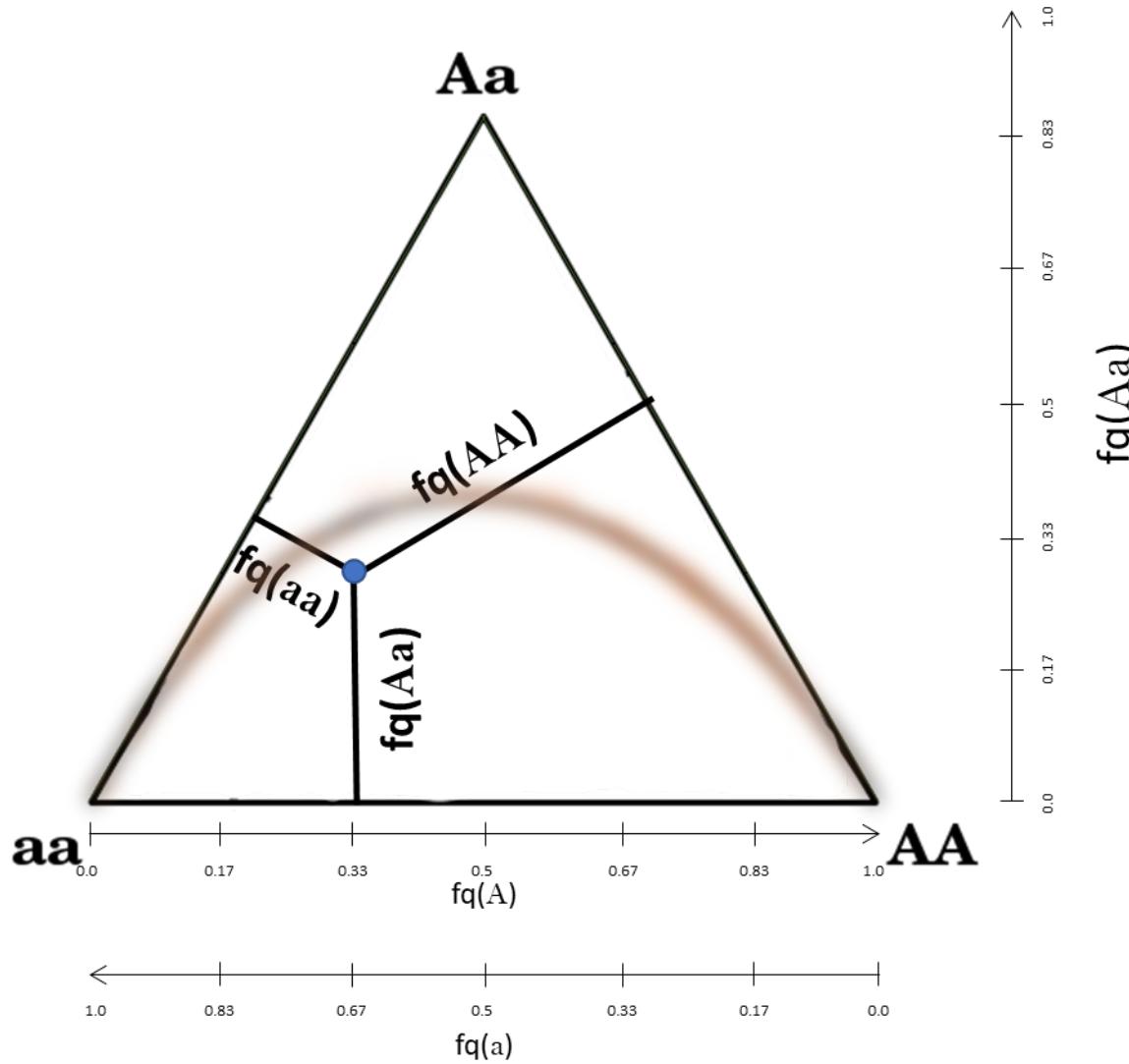
$$Y = (\pi_{aa}, \pi_{Aa}, \pi_{AA}) = (p, q, r)$$

↓

Homoz	$dp = \left[ (-2p+q)\alpha + \left( (p+q/2)^2 - p \right) \beta \right] dt + (1-p)\sqrt{p}dB^p - p\sqrt{q}dB^q - p\sqrt{r}dB^r$
Heteroz	$dq = \left[ 2(p-q+r)\alpha + \left( 2(p+q/2)(r+q/2) - q \right) \beta \right] dt - q\sqrt{p}dB^p + (1-q)\sqrt{q}dB^q - q\sqrt{r}dB^r$

# Parenthesis: Visualizing equation results using DeFinetti Diagram

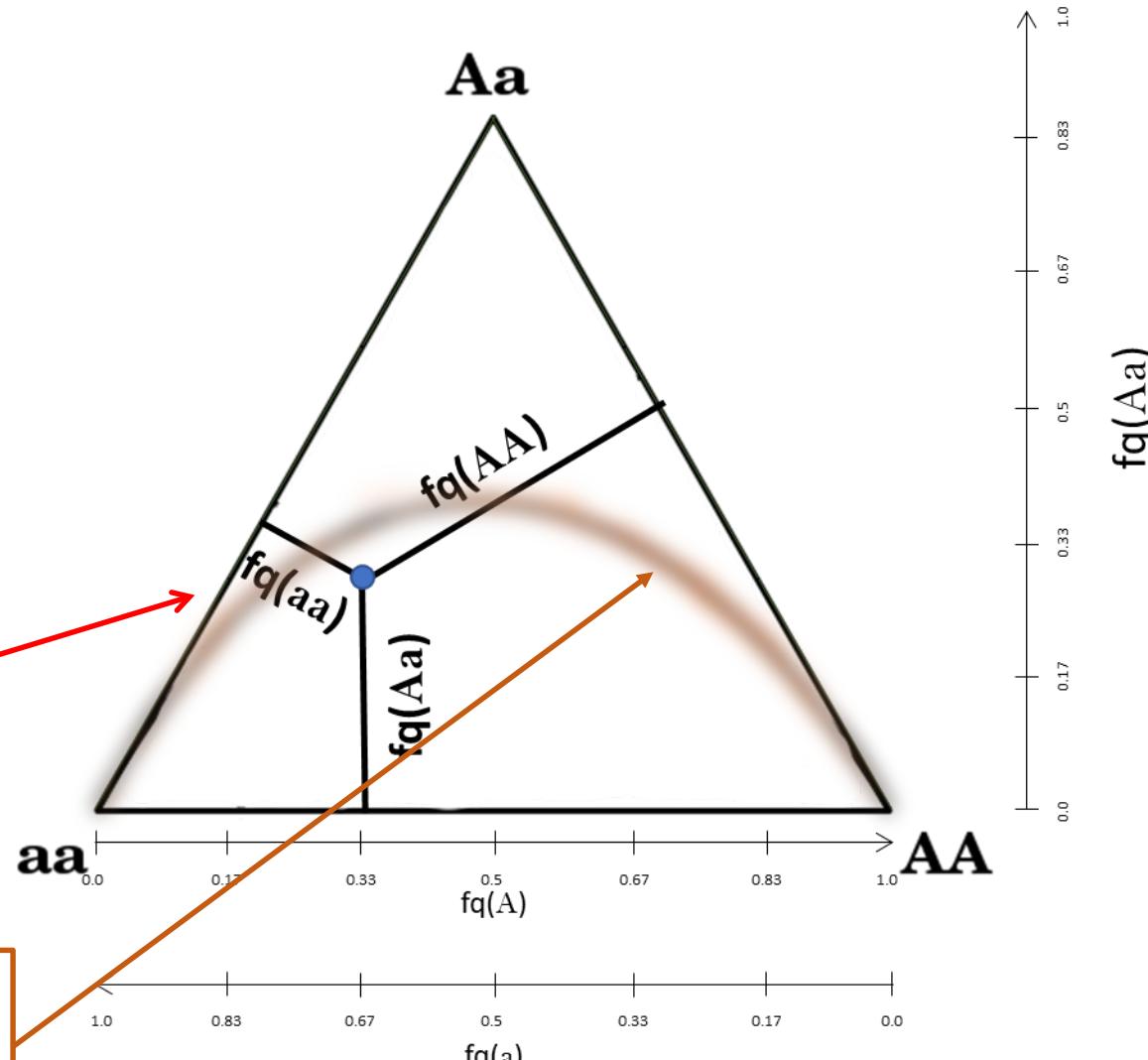
1 locus, 2 alleles (A & a)



# DeFinetti Diagram

1 locus, 2 alleles (A&a)

N=6



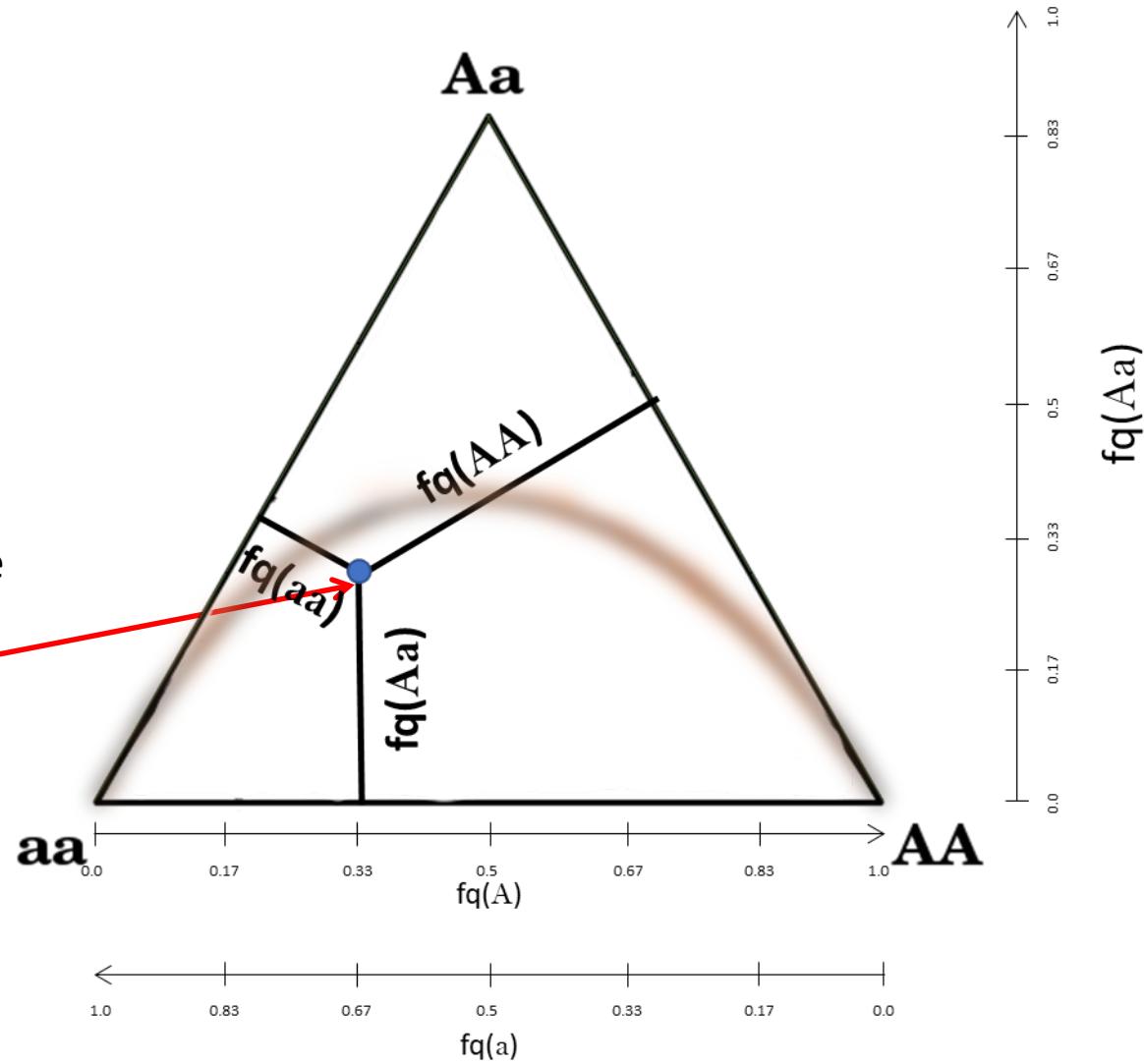
# DeFinetti Diagram

1 locus, 2 alleles (A&a)

N=6

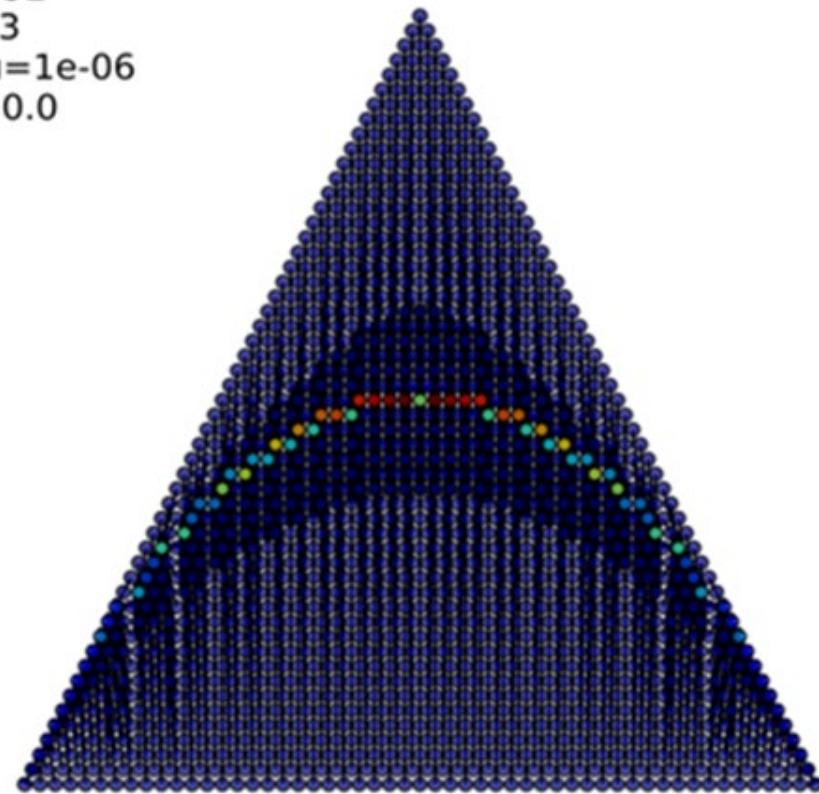
One population genetic state

(aa=1,Aa=2,AA=3)

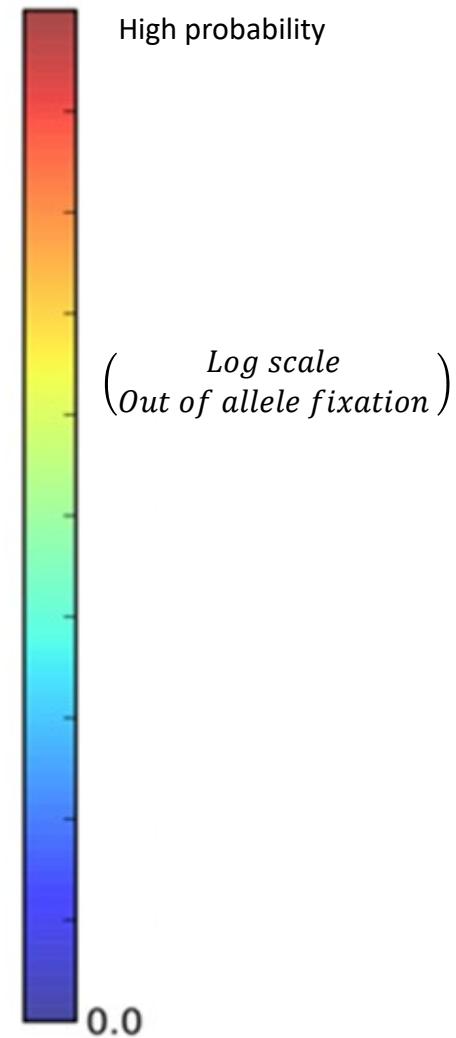
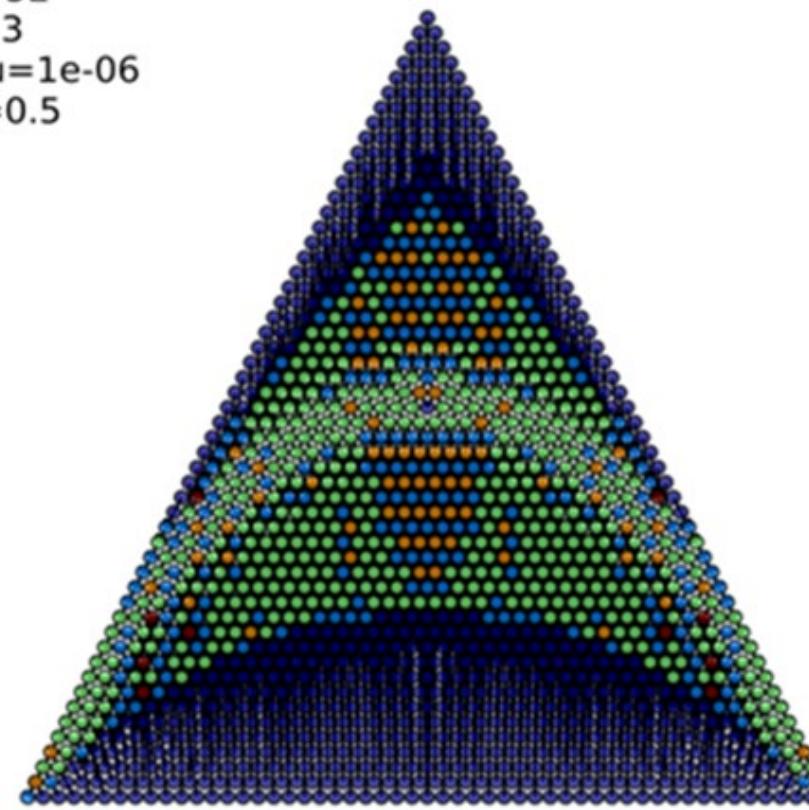


# results

N=52  
n=3  
mu=1e-06  
C=0.0



N=52  
n=3  
mu=1e-06  
C=0.5



- 1/ a few clonality change the probabilities of population genetic states at stationarity (equilibrium)
- 2/ Transition probabilities are identifiable considering rates of clonality

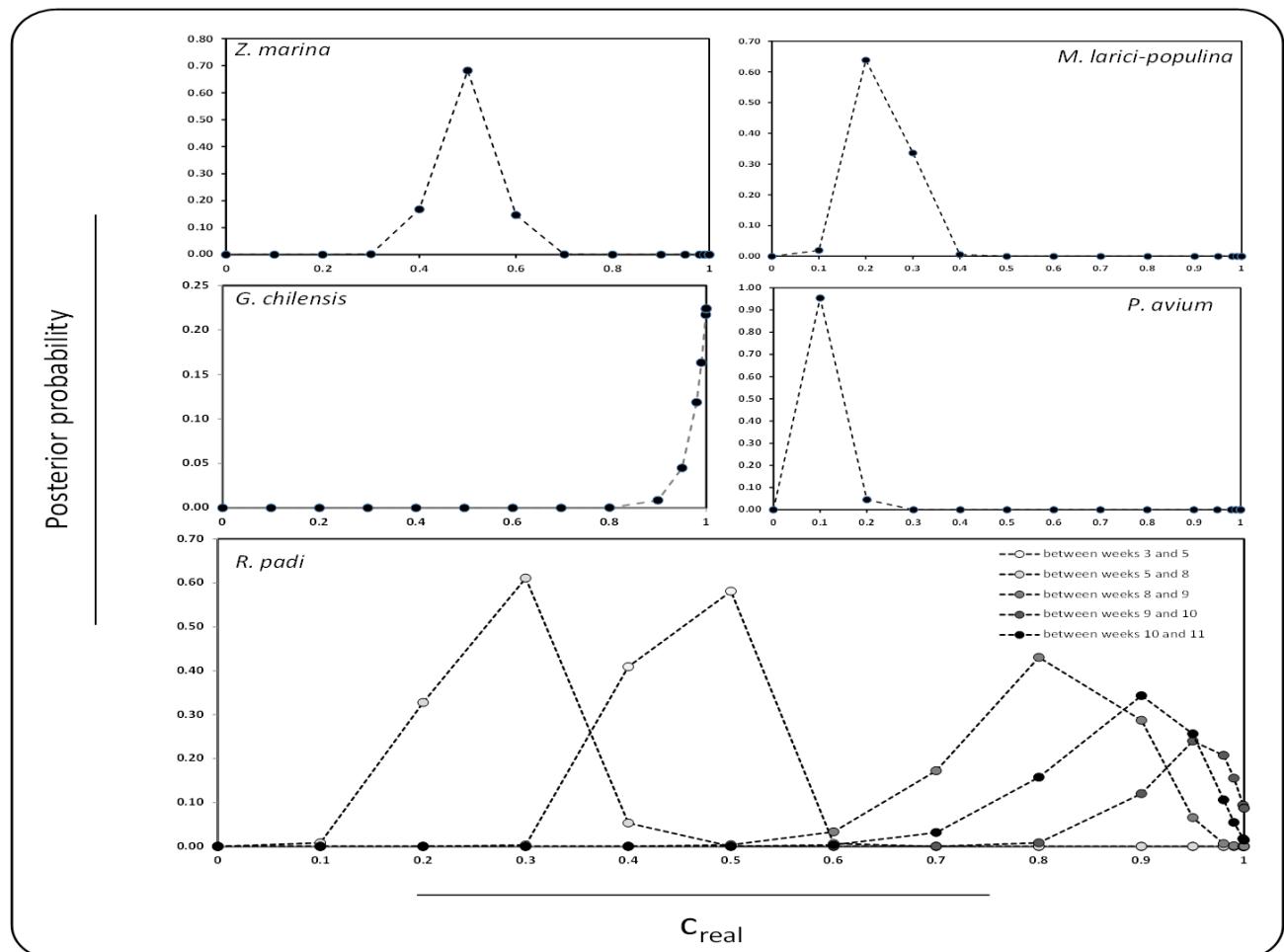
# ClonEstiMate: a Bayesian method for quantifying rates of clonality of populations genotyped at two-time steps

$$L(\tilde{X}_t, \tilde{X}_{t+1}) = \frac{N_\sigma!}{\prod_{i=1}^n (q_{A_i A_i})! \prod_{i=1, j \neq i}^n (q_{A_i A_j})!} \cdot \prod_{i=1}^n (p_{A_i A_i}^*)^{q_{A_i A_i}} \cdot \prod_{i=1, j \neq i}^n (p_{A_i A_j}^*)^{q_{A_i A_j}}$$

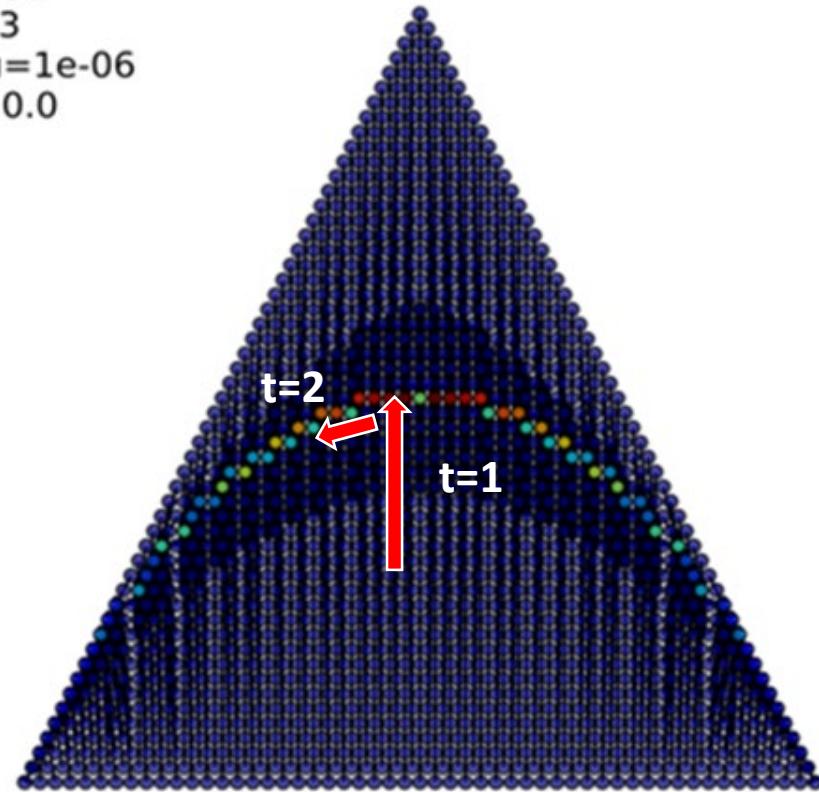
with  $N_\sigma = \sum_i q_{A_i A_i} + \sum_{i,j} q_{A_i A_j}$  and  $\forall i, j \in \{1, \dots, n\}$

## Efficient inference of clonal rate recipe

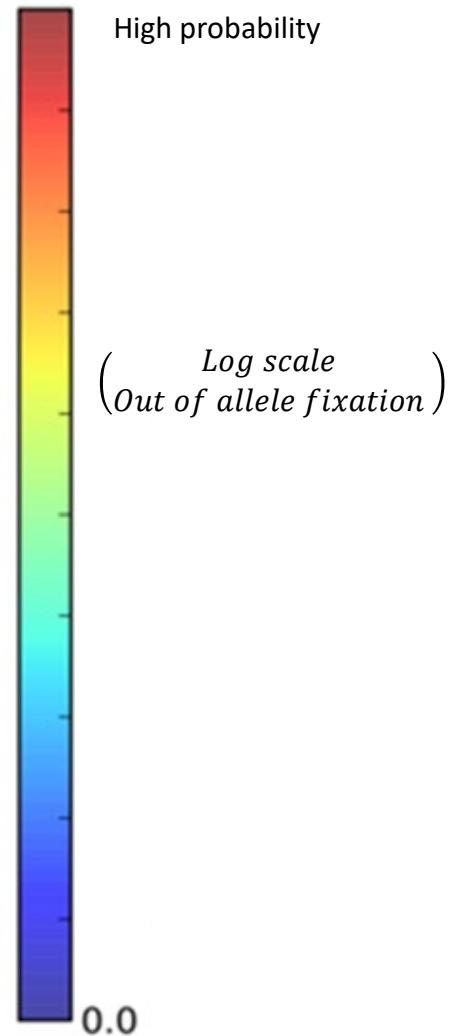
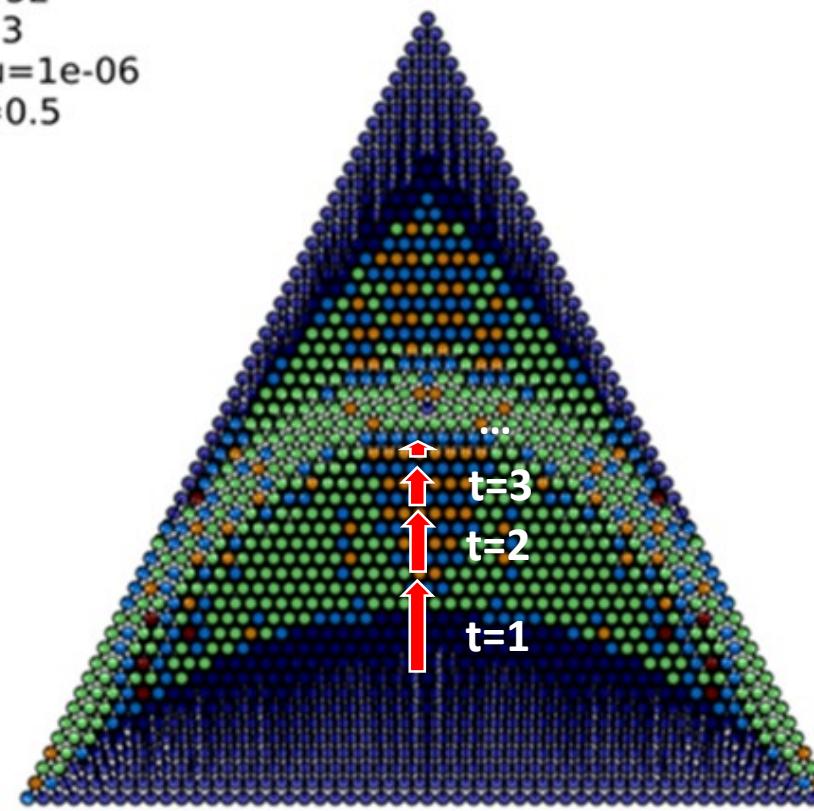
- 30 genotyped individuals
- 10 loci with 2 to 4 alleles (SNP)
- Two-time step sampling



$N=52$   
 $n=3$   
 $\mu=1e-06$   
 $C=0.0$

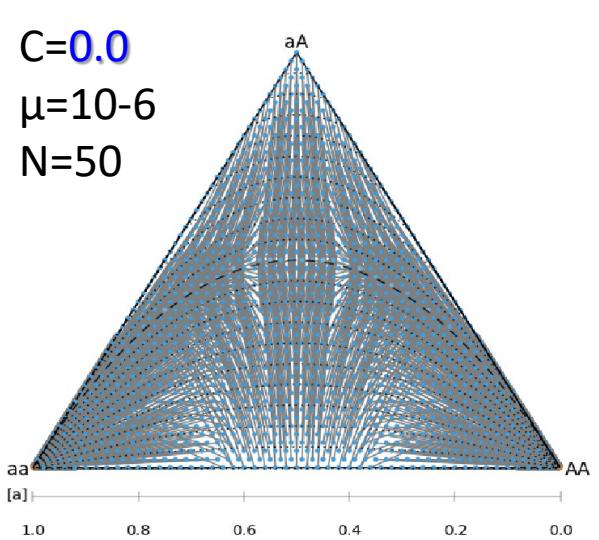


$N=52$   
 $n=3$   
 $\mu=1e-06$   
 $C=0.5$

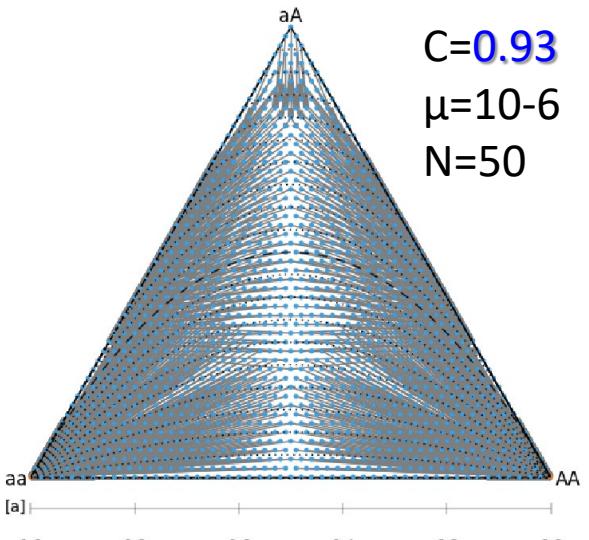


From low rate of clonality, getting back to genotype frequencies expected under Hardy-Weinberg hypotheses takes more and more generations as we are approaching them!

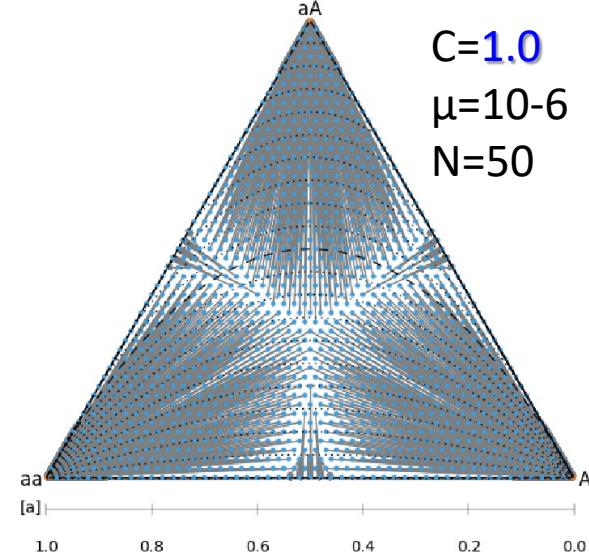
$C=0.0$   
 $\mu=10^{-6}$   
 $N=50$



$C=0.93$   
 $\mu=10^{-6}$   
 $N=50$

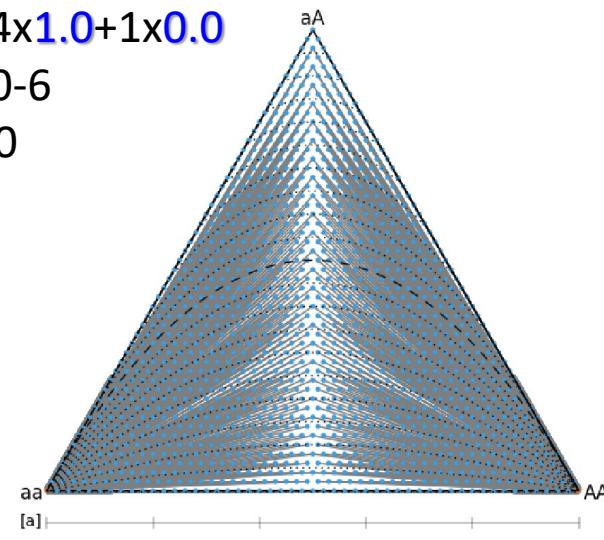


$C=1.0$   
 $\mu=10^{-6}$   
 $N=50$



### Non cyclical partial clonality

$C=14 \times 1.0 + 1 \times 0.0$   
 $\mu=10^{-6}$   
 $N=50$

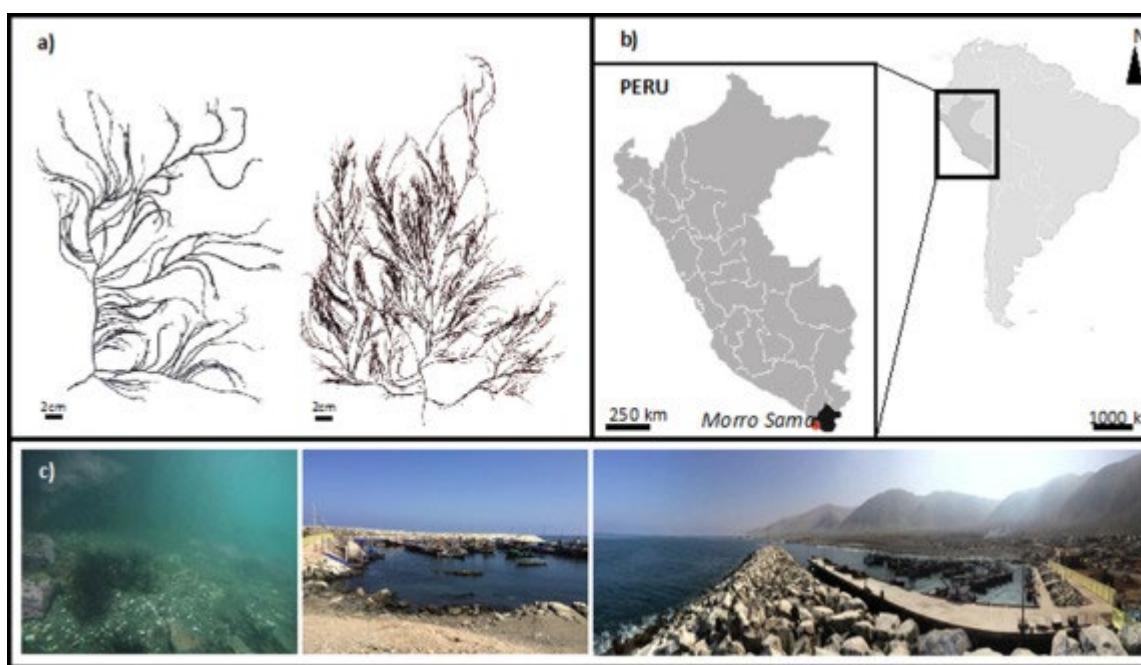


### Cyclical parthenogenesis

(Fungi, Insects, Crustaceans, Rotifers,...)

$C=14 \times 1.0 + 1 \times 0.0$   
 $\mu=10^{-6}$   
 $N=50$   
+ selection  
AA 1.0  
Aa 0.95  
Aa 0.9





## Collaborations with

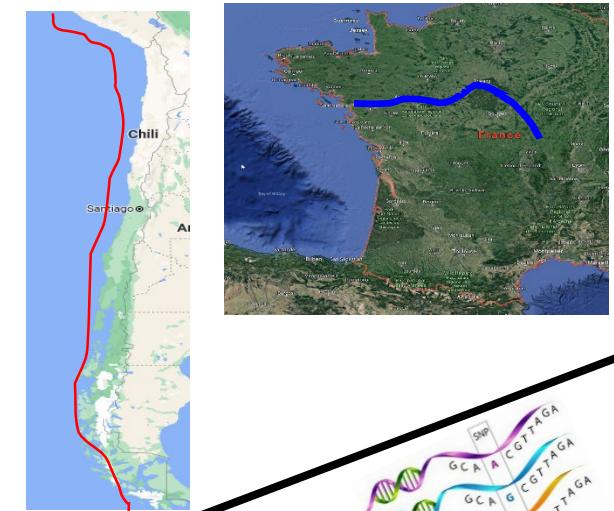
Marie-Laure Guillemin,

Ciencias Ambientales y Evolutivas, Universidad Austral de Chile

Sylvain Faugeron,

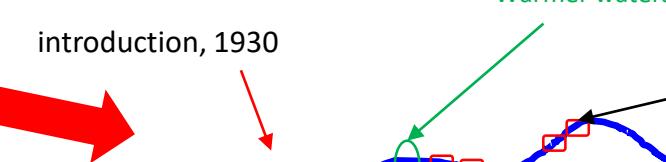
Ecología, Pontificia Universidad Católica de Chile

# Simplified biologic system & environment



Hiplex  
Genotyping populations

introduction, 1930



Warmer waters

First recorded sexual reproduction 1993

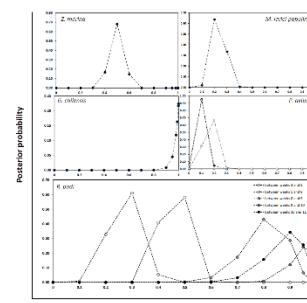
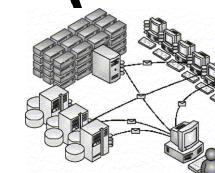
Population adventice isolée pendant 30 ans

$$\begin{cases} dp = [(-2p+q)\alpha + ((p+q/2)^2 - p)\beta]dt + (1-p)\sqrt{pdB^p} - p\sqrt{qdB^q} - p\sqrt{rdB^r} \\ dq = [2(p-q+r)\alpha + (2(p+q/2)(r+q/2) - q)\beta]dt - q\sqrt{pdB^p} + (1-q)\sqrt{qdB^q} - q\sqrt{rdB^r} \\ dr = [(-2r+q)\alpha + ((r+q/2)^2 - r)\beta]dt - r\sqrt{pdB^p} - r\sqrt{qdB^q} + (1-r)\sqrt{rdB^r} \end{cases}$$

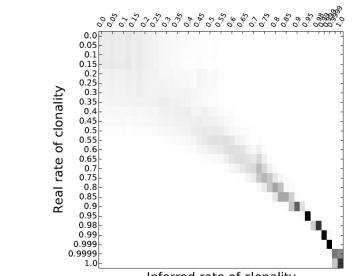
## Stochastic mathematical models

```
## reproduction
for p in xrange(nbpop):
    #####
    # SEXUAL #
    #####
    NbDesX = (1-o)*(nbbind+0.0000000000001)+restesex
    restesex = NbDesX - int(NbDesX)
    NbDesX = int(NbDesX)
    EtatX = EtatX + NbDesX
    EtatX = EtatX % nbbind
    EtatX = EtatX + 1
    #####
    # ASEXUAL #
    #####
    NbDesA = nbbind - NbDesX
    pl=numpy.random.randint(nbbind, size=NbDesA)
```

## Simulations



## Quantitative statistical inferences



## Signal identifiability Model analyses

## Clonix2D

### The genetic consequences of partial clonality in populations colonizing new areas

#### Task1:

Theoretical dynamics  
of genetic diversity in  
populations colonizing  
new areas at  
increasing rates of  
clonality

Resp. S.Stoeckel

#### Task2:

Inference of clonal  
dispersal from various  
datasets

Resp. S.Arnaud-Haond

#### Task3:

Inference of selective  
events when partially  
clonal populations  
colonize new areas

Resp. F.Halkett

# Clonix2D

## The genetic consequences of partial clonality in populations colonizing new areas

**Task1:**  
Theoretical dynamics  
of genetic diversity in  
populations colonizing  
new areas at  
increasing rates of  
clonality

**Task2:**  
Inference of clonal  
dispersal from various

**Task3:**  
Inference of selective  
events when partially  
clonal populations  
colonize new areas

**WG1: Data acquisitions  
and managements**  
SM & MLG

**WG2: Handling and  
production of genomic  
data**  
MV & DB

**WG5: Sharing of  
progresses**  
SAH & SKH

**WG4: Statistical  
inferences**  
SdM & NP

**WG3: Mathematical  
models, numerical  
computations**  
MEO, FM & SS



Institut de Recherche  
pour le Développement  
FRANCE



# INRAe



# UQÀM



# ClonX consortium

A comprehensive theoretical framework to interpret molecular data  
in the framework of population genetics

- WHO?

- INRAE: aphids, fungi, trees, plants, daphnia
- Ifremer: Seagrasses and corals
- CNRS: Algae
- IRD: human pathogens

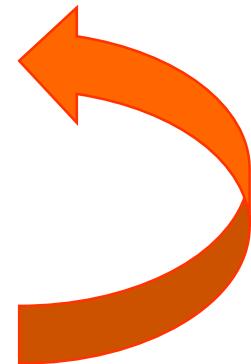
- OBJECTIVES:

through mathematics and simulations:

- assess the effect of increasing rate of clonality on the genotypic and genetic composition of populations
- In turn develop estimates of rates of clonality
- Improve dispersal estimates



How does clonality affect the genomic and genetic architecture of populations?



How do genetic and genotypic parameters relate to the extent of clonality and can they lead to its estimation?



Sophie Arnaud-Haond

# ClonX



Katja Reichel



Ronan Becheler



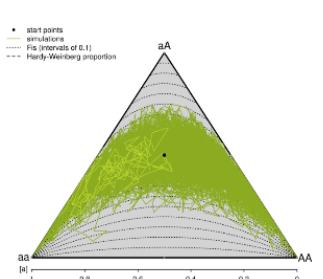
Diane Bailleul



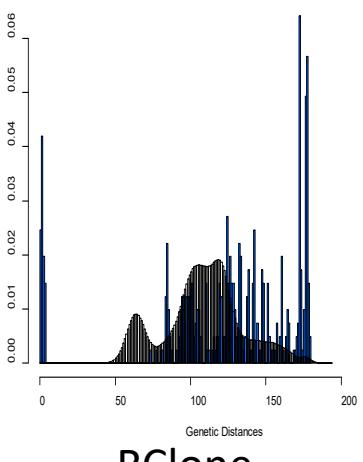
Romuald Rouger



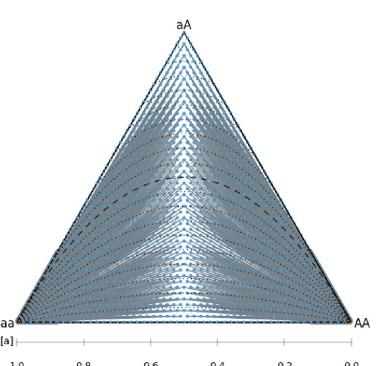
Barbara Porro



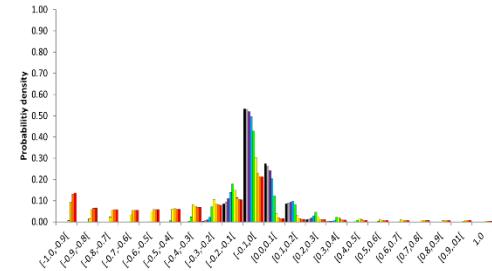
ClonEstiMate



RClone



Cyclical partheno



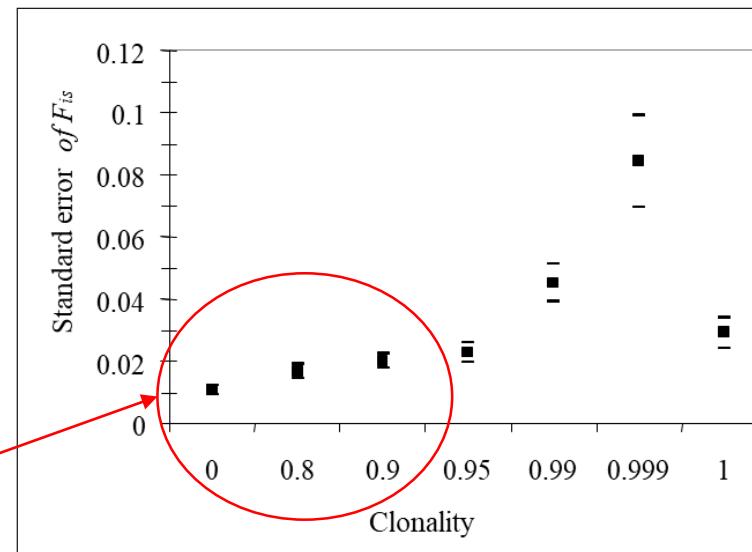
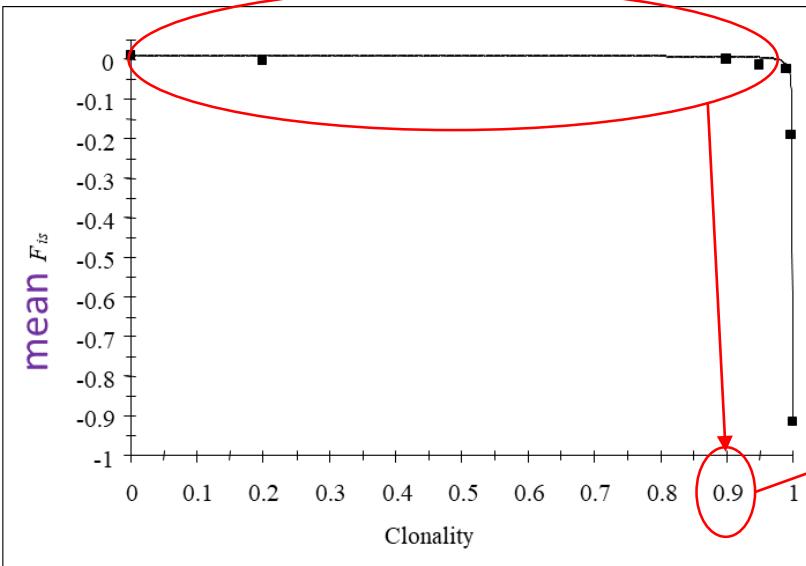
Fabien Halkett

# What the early models say?

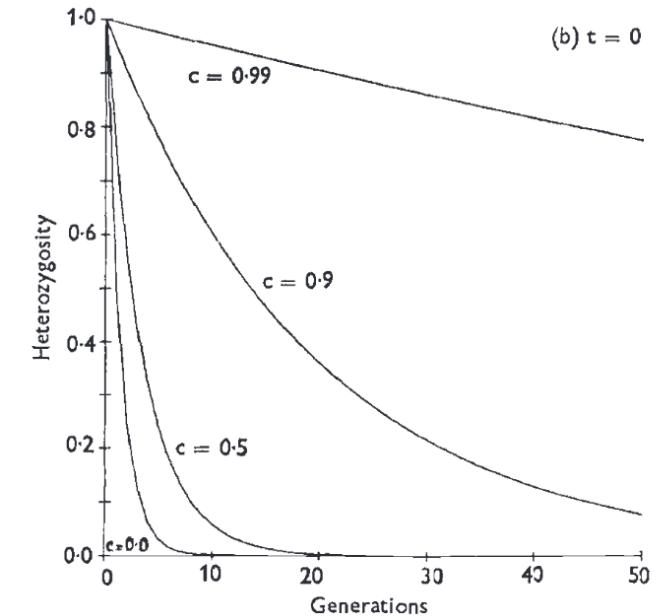
Hypothesis: Clonality impacts genomes as its rate increases (Lewis 1987)?

Marshall & Weir 1979, Orive 1993, Berg & Lascoux 2000, Bengtson 2003, Ceplitis 2003, Balloux et al. 2003, Yonezawa et al. 2004

$$E[T]_f = 1 + \frac{1}{2\Omega} - \Phi, \text{ where } \Phi = \frac{f(\Omega + f + 1)}{2(2\Omega + 1)}.$$



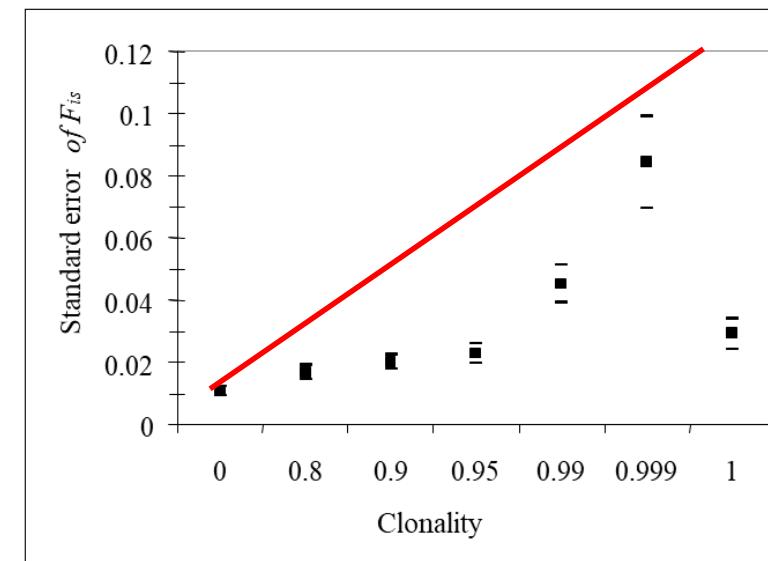
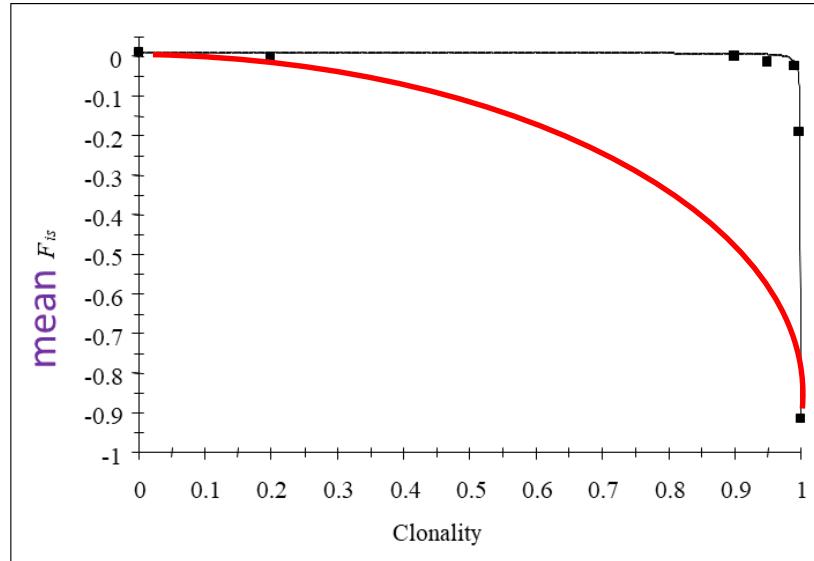
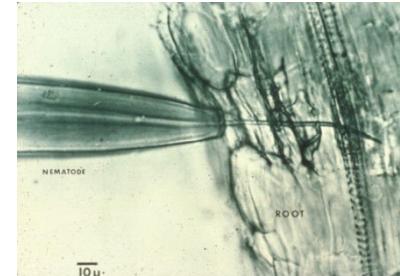
Balloux et al. 2003 Genetics



Marshall & Weir 1979

→ No, as long as rates of clonality are not very high ( $c > 0.9$ ), genetic diversity in partial clonality = sexuality.

# What the data say?



Balloux et al. 2003 Genetics

Three main *kuhnian\** anomalies

- 1- over-representation of heterozygotes in partially clonal populations ( $0 < c < 0.9$ )
- 2- Observed genetic variances  $>>$  predicted variances in highly clonal ( $c > 0.9$ )
- 3- high volatility of genetic indices over time and space  $\neq$  stabilité prédictions

\*Thomas S. Kuhn. (1962). The Structure of Scientific Revolutions. University of Chicago Press.

# Reconciling theory with observations

I) Inference: How estimating those rates in field populations?

II) Prediction: How evolve genetic diversity and genomes in such species?

→ Models allowing tracking trajectories, putting probabilities on observations rather than average values of indices  
(empirically-inspired models rather than theoretically-driven models or data-driven models)

→ Adapted methodologies

→ Adapted datasets



**consortium**

<https://wwz.ifremer.fr/clonix/>

[https://www6.rennes.inra.fr/igepp\\_eng/Productions/Software](https://www6.rennes.inra.fr/igepp_eng/Productions/Software)