

Region specific Raman spectroscopy analysis of the femoral head reveals that trabecular bone is unlikely to contribute to non-traumatic osteonecrosis

Tristan Pascart, Guillaume Falgayrac, Henri Migaud, Jean-François Quinchon, Laurène Norberciak, Jean-François Budzik, Julien Paccou, Anne Cotten, Guillaume Penel, Bernard Cortet

▶ To cite this version:

Tristan Pascart, Guillaume Falgayrac, Henri Migaud, Jean-François Quinchon, Laurène Norberciak, et al.. Region specific Raman spectroscopy analysis of the femoral head reveals that trabecular bone is unlikely to contribute to non-traumatic osteonecrosis. Scientific Reports, 2017, 7 (1), pp.97. 10.1038/s41598-017-00162-3. hal-04445010

HAL Id: hal-04445010 https://hal.science/hal-04445010

Submitted on 4 Apr 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

SCIENTIFIC REPORTS

Received: 4 August 2016 Accepted: 13 February 2017 Published online: 07 March 2017

OPEN Region specific Raman spectroscopy analysis of the femoral head reveals that trabecular bone is unlikely to contribute to non-traumatic osteonecrosis

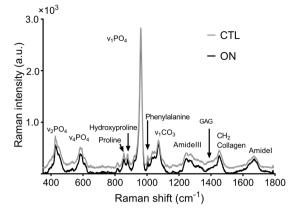
Tristan Pascart^{1,2}, Guillaume Falgayrac¹, Henri Migaud^{1,3}, Jean-François Quinchon⁴, Laurène Norberciak⁵, Jean-François Budzik^{1,6}, Julien Paccou^{1,7}, Anne Cotten^{1,8}, Guillaume Penel¹& Bernard Cortet^{1,7}

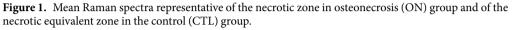
Non-traumatic osteonecrosis (ON) of the femoral head is a common disease affecting a young population as the peak age of diagnosis is in the 40 s. The natural history of non-traumatic ON leads to a collapse of the femoral head requiring prosthetic replacement in a 60% of cases. Although trabecular bone involvement in the collapse is suspected, the underlying modifications induced at a molecular level have not been explored in humans. Here, we examine changes in the molecular composition and structure of bone as evaluated by Raman spectroscopy in human end-stage ON. Comparing samples from femoral heads harvested from 11 patients and 11 cadaveric controls, we show that the mineral and organic chemical composition of trabecular bone in ON is not modified apart from age-related differences. We also show that the molecular composition in the necrotic part of the femoral head is not different from the composition of the remaining 'healthy' trabecular bone of the femoral head. These findings support that quality of trabecular bone is not modified during ON despite extensive bone marrow necrosis and osteocyte death observed even in the 'healthy' zones on histological examination.

Non-traumatic osteonecrosis (ON) of the femoral head is a common disease with an estimated annual incidence of 3/100,000 in Europe¹ and up to 29/100,000 in Asia². The disease affects a young population as the peak age of diagnosis is in the 40 s³. The natural history of ON leads to a collapse of the femoral head requiring prosthetic replacement in a 60% of cases^{3–5}.

Research on the pathophysiology of the disease has not identified a single mechanism and a multiple hit theory has been proposed including vascular occlusion, direct cellular toxicity, altered mesenchymal stem cell differentiation, dysfunctional lipid metabolism and anatomical abnormalities^{4, 6–8}. How the induced infarction eventually leads to the collapse of the femoral head is however not fully understood. Cancellous bone has been suspected to be responsible for the defective mechanical properties of the osteonecrotic femoral head⁹. This suspicion led to explorations of trabecular bone quality during the course of ON using different techniques applied at

¹Lille University, Littoral Côte d'Opale University, EA 4490, PMOI, Physiopathologie des Maladies Osseuses Inflammatoires, F-59000, Lille, France. ²Department of Rheumatology, Saint-Philibert Hospital, Lille University, F-59160, Lomme, France. ³Department of Orthopaedic Surgery, Lille University Hospital, Lille University, F-59000, Lille, France. ⁴Department of Anatomopathology, Saint-Philibert Hospital, Lille University, F-59160, Lomme, France. ⁵Department of biostatistics, Saint-Philibert Hospital, Lille University, F-59160, Lomme, France. ⁶Department of Radiology, Saint-Philibert Hospital, Lille University, F-59160, Lomme, France. ⁷Department of Rheumatology, Lille University Hospital, Lille University, F-59000, Lille, France. ⁸Department of Radiology, Lille University Hospital, Lille University, F-59000, Lille, France. Correspondence and requests for materials should be addressed to T.P. (email: pascart.tristan@qhicl.net)





various scales. The macroscale was investigated with dual-energy X-ray absorptiometry. The bone mineral density (BMD) of osteonecrotic femoral heads was found to be decreased compared with matched controls^{10, 11}. At the mesoscale, histological analyses showed necrosis and decreased osteocyte viability beyond the necrotic zone and in some cases as far as the proximal femur^{12–15}. Micro-computed tomography was applied to study the microscale¹⁶. Wang *et al.* found local alterations of the microarchitecture in 10 necrotic femoral heads. These authors reported cracks and thinning of trabeculae in the necrotic region but a normal microstructure at a distance¹⁷. The molecular scale remains to be explored in depth, particularly looking for local discrepancies, to provide a better understanding of bone quality alterations in affected bone.

Raman spectroscopy enables simultaneous exploration of mineral and organic composition and structure in healthy and pathological bone¹⁸. Physicochemical parameters (PCPs) can be measured to assess relative variations in the composition and structure of a bone sample, providing a reliable grasp of its quality at a molecular level. Raman spectroscopy is increasingly used to understand how changes in bone composition and structure influence tissue-level mechanical properties of bone^{19–22}. Raman spectroscopy can be performed on fresh samples using simple sample preparation to the contrary of other vibrational techniques analyzing bone composition such as Fourier transform infrared (FTIR)^{23, 24}. Raman spectroscopy is thus a promising tool to explore the molecular changes occurring in bone during ON. Aruwajoye *et al.* used Raman spectroscopy to examine early modifications in an animal model of ON of the femoral head and found increased carbonate substitution in the necrotic bone²⁵. Animal models provide insight into early-stage ON, but cannot fully reproduce the features of human disease; such results must be confronted with analyses of human samples²⁶.

So far, the mechanisms leading to the collapse of the femoral head and the anatomical extent of the altered bone remain unclear such that the structural evolution is still unpredictable⁴. The objective of this study was to examine modifications of the molecular composition and structure of bone as evaluated by Raman spectroscopy in human end-stage non-traumatic ON of the feoral head, and to search for relations with histological findings.

Results

Raman Spectroscopy. The Raman spectrum of bone is shown in Fig. 1. Values of each PCP according to the zones of sampling are presented in Table 1.

Inside-group comparisons. The PCPs were not significantly different between the three zones in the control group. The PCPs were not significantly different between the necrotic, sclerotic and distant zones in the ON group.

Between-group comparisons. Uni and bi-variate analysis. No significant differences were found regarding the mineral-to-matrix ratio, carbonate B-substitution, collagen maturity and the hydroxyproline-to-proline ratio for any of the matched zones. Crystallinity was significantly decreased in all zones of the ON group compared with the control group (p = 0.04). The relative proteoglycan content was significantly decreased in the sclerotic zone of ON patients compared with the matched zone of controls (p = 0.03). This decrease was not significant in the necrotic zone (p = 0.28) and tended to be significant in the distant zone (p = 0.07).

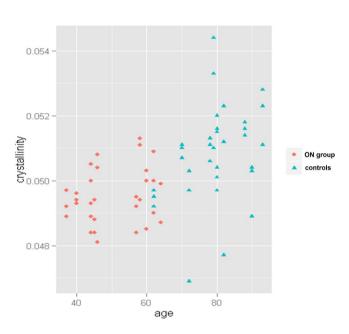
Multivariate analysis. Parameters with significant differences in bivariate analysis were tested for multivariate analysis with adjustment on age and gender using a mixed linear model.

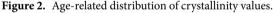
The model regarding crystallinity was found valid after descending selection. Selected variables were age (p = 0.026) and gender (p = 0.056) suggesting that observed differences of crystallinity between groups were age-related (Fig. 2).

The residual normality of the model for the proteoglycan content was negative even after log transformation of the variable. Non-linear models were considered but not performed due to a risk of misinterpreting the data. The decrease of the proteoglycan relative content in bivariate analysis was not explained by multivariate analysis. However, in order to measure a suspected influence of age on the relative proteoglycan content in the

| Group | Zone | Proteoglycan relative content | Collagen maturity | Hydroxyproline- to-proline ratio | Mineral-to- matrix ratio | Carbonate-B- substitution | Crystallinity |
|---------|----------------------|---|---|---|---|-------------------------------|--|
| ON | Necrotic | $\begin{array}{c} 0.064 \pm 0.004 \\ (0.001) \end{array}$ | $\begin{array}{c} 1.476 \pm 0.058 \\ (0.017) \end{array}$ | $0.841 \pm 0.114 \\ (0.034)$ | 8.947±0.688 (0.207) | $0.126 \pm 0.004 \\ (0.001)$ | $\begin{array}{c} 0.05 \pm 0.001 * \\ (0.0003) \end{array}$ |
| ON | Sclerotic | $0.061 \pm 0.004 * \\ (0.001)$ | $\begin{array}{c} 1.464 \pm 0.066 \\ (0.020) \end{array}$ | $0.869 \pm 0.127 \\ (0.038)$ | $9.018 \pm 0.568 \\ (0.171)$ | $0.127 \pm 0.005 \\ (0.002)$ | $\begin{array}{c} 0.049 \pm 0.001 * \\ (0.0003) \end{array}$ |
| ON | Distant | $0.061 \pm 0.004 \\ (0.001)$ | $\begin{array}{c} 1.492 \pm 0.072 \\ (0.022) \end{array}$ | 0.841±0.121 (0.036) | $\begin{array}{c} 8.913 \pm 0.537 \\ (0.162) \end{array}$ | $0.126 \pm 0.004 \\ (0.001)$ | $\begin{array}{c} 0.049 \pm 0.001 * \\ (0.0003) \end{array}$ |
| Control | Necrotic equivalent | $0.075 \pm 0.016 \\ (0.005)$ | $\begin{array}{c} 1.505 \pm 0.069 \\ (0.021) \end{array}$ | $\begin{array}{c} 0.763 \pm 0.152 \\ (0.046) \end{array}$ | 9.624±0.716 (0.216) | $0.126 \pm 0.004 \\ (0.001)$ | $\begin{array}{c} 0.051 \pm 0.001 \\ (0.0003) \end{array}$ |
| Control | Sclerotic equivalent | $0.074 \pm 0.015 \\ (0.005)$ | $\begin{array}{c} 1.453 \pm 0.066 \\ (0.02) \end{array}$ | $0.749 \pm 0.136 \\ (0.041)$ | 9.722±0.640 (0.193) | $0.127 \pm 0.003 \\ (0.0009)$ | $\begin{array}{c} 0.051 \pm 0.002 \\ (0.0006) \end{array}$ |
| Control | Distant equivalent | $\begin{array}{c} 0.075 \pm 0.015 \\ (0.005) \end{array}$ | $\begin{array}{c} 1.465 \pm 0.104 \\ (0.031) \end{array}$ | $\begin{array}{c} 0.752 \pm 0.115 \\ (0.035) \end{array}$ | 9.411±0.353 (0.106) | $0.127 \pm 0.004 \\ (0.001)$ | $\begin{array}{c} 0.051 \pm 0.002 \\ (0.0006) \end{array}$ |

Table 1. Mean values \pm Standard Deviation (Standard Error) for each physicochemical parameter for thenecrotic, sclerotic and distant zones of the non-traumatic osteonecrosis (ON) of the femoral head group andequivalent zones of the control group. None of these differences were significant between zones within groups.*Corrected p-value < 0.05 (univariate between group differences, none of these differences were significant after</td>multivariate analysis).





....

intermediary zone, a Spearman correlation coefficient was calculated. The coefficient was statistically significant (0.57, p = 0.013) showing that the relative proteoglycan content was positively correlated with age.

Histology. Histological analysis confirmed the diagnosis of ON with extended bone marrow necrosis in the necrotic zone. The necrosis was extended to the sclerotic zone in some cases. The bone marrow was normal in the distant zone. Osteocyte lacunae were either empty or filled with dead cells in the necrotic zone. Some dead or intact osteocytes were filling most osteocytes lacunae in the sclerotic zone, with some of them remaining empty. Almost exclusively intact osteocytes were found in the distant zone. Osteocyte lacunae filled with pyknotic cellular structures coexisted with others presenting with nuclei showing spread and faded chromatin. In the sclerotic zone could be found new bone structure due to repairing processes with chondroid and woven bone associated with normal trabeculae (Fig. 3).

Examination of the three zones from the control group showed normal bone structure with intact osteocytes.

Discussion

This study provides evidence that the molecular composition and structure of the trabecular bone is not modified during end-stage non-traumatic ON of the femoral head despite extensive cell death.

The mineral and organic compositions of bone were generally not modified across zones and between groups. These results are in agreement with a previous study of piglets using microscale techniques²⁵. Indeed, using X-ray absorbance near edge structure and Raman spectroscopy, molecular composition was found to be unchanged in the necrotic bone apart from a higher carbonate substitution in the necrotic zone. This increased carbonate

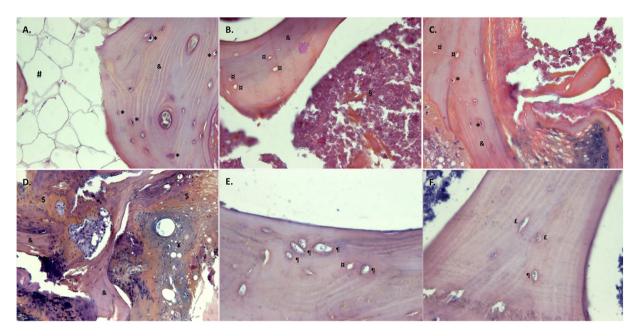


Figure 3. Histological examination using hemtoxylin-eosin-saffron staining. (**A**) Control sclerotic-equivalent zone (x200); (**B**). Necrotic zone (ON group) (x200); (**C**). Sclerotic zone (ON group) (x200); (**D**). Sclerotic zone with coexisting normal trabeculae, woven and chondroid bone (ON group) (x200); (**E**). pyknotic apoptotic osteocytes (ON group) (x400); (**F**). coexistence of pyknotic osteocytes others presenting with nuclei showing spread and faded chromatin (x400). [&]Normal trabecula, ^{\$}woven bone, [¥]chondroid bone, [#]normal medullary space, ^{\$}necrotic medullary space, ^{*}intact osteocyte, [¶]apoptotic osteocyte, [¤]empty osteocyte lacuna. ON: osteonecrosis.

substitution in the necrotic zone, found only in the animal model, could be a result of early stage processes, differences related to animal pathophysiology, but also a result of sample fixation that could have affected carbonate substitution²⁴. Aruwajoye *et al.* found that molecular stability of the necrotic zone was not echoed by structural stability as architectural trabecular modifications were seen in the necrotic zone both with scanning electron microscopy and micro-computed tomography²⁵. Wang *et al.* tested the biomechanical properties of human necrotic femoral heads using nano-indentation. Once again, bone nanoscopic biomechanical properties were similar between necrotic and healthy regions despite clear architectural alterations of the necrotic bone evaluated by micro-computed tomography¹⁷.

The decrease of the relative proteoglycan content in the sclerotic region is probably age-related and linked to increased bone remodelling in this zone. Proteoglycan content was previously shown to increase with age²⁷ as further confirmed by a significant correlation between the two variables in our study. In addition, it is also known that proteoglycans act as a scaffold of bone mineralisation and down-regulate the bone mineral apposition rate to obtain a well-organized bone^{27, 28}. Histological examination showed the presence of woven bone that corresponds to rapid, disorganised and unregulated new bone apposition expected to have low proteoglycan content.

The absence of modification of bone composition despite extensive osteocyte death suggests that osteocytes may not play a central role in ON. Previous studies found an increased cell apoptosis rate in the osteocyte/oste-oblast lineage and hypothesized that osteocyte apoptosis in the cancellous bone was a key mechanism in the pathogenesis of osteonecrosis^{12, 15, 29-31}. Osteocytes are known to act on the mineral homeostasis of bone³². There is however no evidence from our study that the mineral composition is modified, advocating that the observed osteocyte death did not impact on the trabecular bone quality. This would imply that osteocyte death may simply be a marker of trabecular bone stress rather than an active participant to its collapse. Consistently with this hypothesis, Weinstein *et al.* have shown that this cell death is predominant near the fractured subchondral region, suggesting that trabecular bone suffers from the nearby defective subchondral bone²⁹. Mutijima *et al.* also showed an increase of apoptotic osteocytes in the sclerotic region¹⁵ that is also in contact with the compromised necrotic zone¹⁷.

Our study supports the hypothesis that the quality of trabecular bone may not be affected directly by ON. Several studies looked into potential modifications of the trabecular architecture of human or animal femoral heads undergoing or having undergone osteonecrosis. Two studies compared the remaining "healthy" trabecular bone using micro-computed tomography of human osteonecrotic femoral heads versus osteoarthritic controls and none of them found differences^{33, 34}. The same results were found in a model of corticosteroid-induced ON of the femoral head in bipedal emus³⁵. The total preservation of the trabecular microarchitecture, apart from the necrotic zone, is intriguing. Indeed, if trabecular microarchitectural modifications were a pathophysiological feature of the disease, it can be hypothesized that they would not be cut off-limited to the collapsing necrotic zone but would also be present to some extent in its periphery. The microarchitectural modifications observed only in the necrotic zone could rather be the consequence of excessive mechanical stress and aggravated by increased

| Patient number | Age (y.o) | Gender | Side | Bilateral osteonecrosis | Disease duration (months) | Cause | FICAT stage |
|-------------------|--------------|--------|-------|-------------------------|---------------------------------|---------------------|----------------|
| 1 | 58 | male | Left | Unknown | 36 | Alcohol abuse | 4 |
| 2 | 45 | male | Left | Yes | 6 | Alcohol abuse | 3 |
| 3 | 46 | male | Right | Yes | 24 | Alcohol abuse | 3 |
| 4 | 64 | male | Right | Yes | 12 | Glucocorticoids use | 4 |
| 5 | 57 | male | Right | Yes | unknown | Alcohol abuse | 3 |
| 6 | 44 | male | Left | Yes | unknown | Alcohol abuse | 4 |
| 7 | 40 | male | Right | Yes | 36 | Alcohol abuse | 4 |
| 8 | 62 | male | Left | Yes | unknown | Alcohol abuse | 4 |
| 9 | 44 | male | Right | Unknown | unknown | Alcohol abuse | 4 |
| 10 | 60 | male | Right | Yes | unknown | Alcohol abuse | 4 |
| 11 | 37 | male | Right | Yes | 18 | Alcohol abuse | 3 |

Table 2. Disease history and patients' characteristics.

bone resorption³⁶. The study of osteoclast activity and growth factor expression in human ON showed a particular increase in the sclerotic trabecular region and also nearby subchondral fractures suggesting that these mechanisms react to mechanically compromised zones³⁷.

We acknowledge that this study has limitations. The sample size of the study is modest but keeping in the context of non-traumatic ON, contrary to many studies^{17, 25, 36, 37}, a control group was provided and the population size of the control group was larger than observed in most similar studies^{15, 33, 35}. Age-related increase of crystallinity (and to some extent the decrease of the proteoglycan content) was an expected between-group difference³⁸ and was confirmed by multivariate analysis. Age differences between groups did not seem to have induced significant differences regarding the other parameters according to univariate analysis. The histological analyses that were performed with hematoxylin-erythrosin-saffron (HES) staining are qualitative only and no TUNEL assay was performed, therefore these results must be interpreted with caution. Nonetheless, given the specific morphological aspects of the various cell deaths³⁹, these first observations of both necrotic and apoptotic osteocytes are convincing.

Together with biomechanical and microarchitectural studies, our study advocates for a relative innocence of cancellous bone in both the disease pathogenesis and the collapse of the osteonecrotic femoral head. In the meantime, evidence is being gathered against suspect number two: subchondral bone. Osteoclast activity and growth factors tend to be concentrated in the subchondral region^{17, 37}. Microarchitectural modifications of the subchondral bone and especially thinning and increased porosity of the subchondral plate have been observed in animal models^{35, 40}. In addition, clinical outcomes of hip resurfacing preserving the trabecular bone but replacing the subchondral bone are very promising even in case of extensive osteonecrosis^{41–44}. Further controlled studies on human ON exploring modifications of the subchondral bone involving its architecture, composition and structure within the necrotic zone but also in more distant regions are required to provide a better understanding of the pathophysiology and course of ON.

Methods

Samples. Between April 2010 and March 2014, samples from femoral heads were harvested from eleven patients and eleven cadaveric controls. The patients gave informed consent to participate and the study was approved by the ethical review board of the *Direction Générale de la recherche et de l'innovation* (French Ministry for Research) (DC-2008-642). The study was carried out in accordance with the World Medical Association Declaration of Helsinki. Patients undergoing total hip arthroplasty with the diagnosis of magnetic resonance imaging (MRI)-proven non-traumatic ON of the femoral head were included in the study. All patients were male (mean age (\pm SD) 50.6 (\pm 9.6) years). Demographic and clinical characteristics of patients are detailed in Table 2. The ON group was composed of biopsies of the femoral heads collected in the operating room just after the resection of the femoral head. Biopsies were performed using a 10 mm diameter and 25 mm hole saw in three zones pre-determined by a systematically performed pre-operative MRI examination and confirmed by visual macroscopic observation: the necrotic zone, the sclerotic zone and in the distant zone (far off the necrotic zone) (Fig. 4).

The control group was composed of bone biopsies from cadaveric donors provided by the Lille University anatomy department. The donors had informed of their will to donate their body to science after their death. The donors had no recorded history of bone disease. Femoral heads were harvested immediately upon arrival of the corpses at the anatomy department before any embalmment procedure was performed²⁴. Samples were harvested from the corpses of seven male and four female subjects who died at a mean age of 79.4 (\pm 9.1) years. The femur was sectioned at the base of the neck to harvest the entire femoral head from each subject. Three separate biopsies were sampled in equivalent zones comparatively to the ON group: below the subchondral region, base of the femoral neck, and in between the two.

Sample preparation. Samples were stored at -80 °C until analysis. An 8 mm height section of each biopsy was cut 8 mm below the cartilage surface using a speed diamond blade saw. The section was fixed in 70% ethanol solution for 48 H to avoid sample degradation during the Raman analysis, then polished using abrasive papers

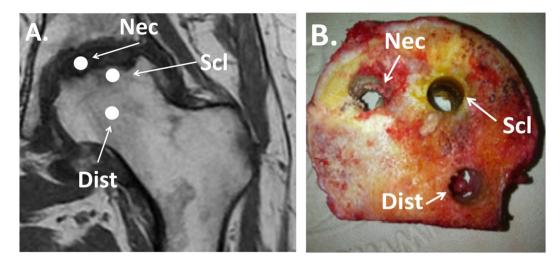


Figure 4. Magnetic resonance imaging (**A**) and macroscopic (**B**) localizations of the three sampled zones. Nec = necrotic zone, Scl = sclerotic zone, Dist = distant zone.

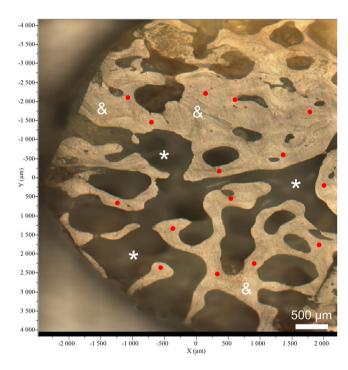


Figure 5. Partial overview of a single sample in confocal microscopy (X10) with each of the 15 (/30 of the entire sample) selected spots selected for Raman spectral acquisition (red dots). *Marrow regions, [&]mineral trabeculae.

with decreasing grain size (30, 3 and $0.3 \,\mu$ m) and set on a microscope slide. A contiguous sample of the same size was used for histology.

Histology. The samples were decalcified with a solution of formaldehyde, methanol and formic acid. Sections of paraffin-embedded sample blocks were stained with HES. Optical microscopic examination was performed. The extents of necrosis and osteocyte viability were assessed by qualitative histological examination across the three zones.

Raman Acquisitions. Once the sample is grinded to obtain an even surface, mineralized trabeculae are distinguished from bone marrow regions, and zones of the trabeculae were pre-selected for Raman spectral acquisitions (Fig. 5). Spectra were acquired with a Raman microspectrometer LabRAM HR800 (HORIBA, Jobin-Yvon, France) provided with DuoScan technology. The instrument is equipped with a XYZ motorized stage and a diode laser at 785 nm. DuoScan technology provides an average spectrum representative of a rastered area giving the opportunity to explore more rapidly broader samples⁴⁵. The DuoScan averaging mode was used with an $\times 50$ objective and a rastered area of $30 \times 30 \,\mu$ m. Thirty spectra were acquired for each sample, with an integration time of 60 seconds and 5 accumulations. The spectral range was 300 to $1700 \,\text{cm}^{-1}$ with a resolution of $4 \,\text{cm}^{-1}$.

All Raman spectra were processed using Labspec software (HORIBA, Jobin-Yvon, France). A Savitzky-Golay smoothing filter (filter width: 3; polynomial order: 2) and polynomial baseline correction (degree 4) were applied to the data set prior to evaluation of physicochemical parameters.

Bone physicochemical parameters. The physicochemical parameters characterizing bone composition and structure have been described previously¹⁸. Mineral-to-matrix ratio was calculated from the ratio of the phosphate symmetric stretch band intensity (960 cm⁻¹) to the intensity of vibrations resulting primarily from the CH_2 side-chains of collagen molecules (1450 cm⁻¹). This parameter measures relative bone mineral-to-organic content that provides an assessment of mineralization. Crystallinity was calculated as the inverse of the full width at half maximum (FWHM) of the phosphate symmetric stretch band. Crystallinity gives an assessment of crystal size and perfection. Type-B carbonatation was calculated as the ratio between the respective intensity of the type-B carbonate symmetric stretch band (at 1073 cm⁻¹) and the phosphate symmetric stretch band. This ratio represents the level of type-B carbonate substitution in the apatite crystal. The relative proteoglycan content was calculated from the ratio of integrated areas of the glycosaminoglycan (GAG)/CH₃ band (1365-1390 cm⁻¹) to the amide III band (1243–1269 cm⁻¹). The hydroxyproline-to-proline ratio was calculated as the intensity ratio of hydroxyproline (877 cm^{-1}) and proline (853 cm^{-1}). The study of the hydroxyproline and proline Raman bands was shown to give a reliable evaluation of post-translational modifications of collagen in bone^{22, 46, 47}. This ratio provides an assessment of the collagen secondary structure with implications for bone mineralization capability⁴⁸. Collagen maturity was calculated as the intensity ratio of two sub-component bands in the amide I band commonly described approximately at 1660 cm⁻¹ for the amide I peak and 1690 cm⁻¹ for the subsequent shoulder²².

Statistical analysis. Statistical analysis was performed using the R software. The value of each physicochemical parameter of each biopsy sample was the average of the 30 spectra collected. Results are presented as mean (\pm SD (SE)). Comparisons between the three zones inside groups were performed using the non-parametric Quade test as repeated measures within the same subjects were considered. The Mann-Whitney-Wilcoxon test was used to compare matched zones of the ON and control groups. P-values were corrected for each value using Holm's technique to avoid the inflation of the α risk due to repeated comparisons of each zone. Multivariate analysis was performed for parameters with significant p-value in bi-variate analysis using mixed linear models given the repeated measurements (3 zones) performed on each subject. Age, sex, group, and zone were integrated as fixed effects in the models. The small sample size required the application of a variable selection procedure. The step by step, backward method, based on the Akaïke criterion was chosen. The model validation was assessed graphically using the residuals (normality and homoscedasticity). Statistical significance was assigned to p < 0.05.

References

- 1. Cooper, C., Steinbuch, M., Stevenson, R., Miday, R. & Watts, N. B. The epidemiology of osteonecrosis: findings from the GPRD and THIN databases in the UK. *Osteoporos. Int.* **21**, 569–577 (2010).
- Kang, J. S. *et al.* Prevalence of osteonecrosis of the femoral head: a nationwide epidemiologic analysis in Korea. J. Arthroplasty 24, 1178–1183 (2009).
- 3. Fukushima, W. *et al.* Nationwide epidemiologic survey of idiopathic osteonecrosis of the femoral head. *Clin. Orthop.* **468**, 2715–2724 (2010).
- Mont, M. A., Cherian, J. J., Sierra, R. J., Jones, L. C. & Lieberman, J. R. Nontraumatic Osteonecrosis of the Femoral Head: Where Do We Stand Today? A Ten-Year Update. J. Bone Joint Surg. Am. 97, 1604–1627 (2015).
- Mont, M. A., Zywiel, M. G., Marker, D. R., McGrath, M. S. & Delanois, R. E. The natural history of untreated asymptomatic osteonecrosis of the femoral head: a systematic literature review. J. Bone Joint Surg. Am. 92, 2165–2170 (2010).
- Moya-Angeler, J., Gianakos, A. L., Villa, J. C., Ni, A. & Lane, J. M. Current concepts on osteonecrosis of the femoral head. World J. Orthop. 6, 590–601 (2015).
- Penel, G. *et al.* Bone and marrow fat interactions in osteonecrosis of the femoral head (ONFH). *Osteoporos. Int.* 24, S458-+ (2013).
 Ollivier, M., Lunebourg, A., Abdel, M. P., Parratte, S. & Argenson, J.-N. Anatomical Findings in Patients Undergoing Total Hip
- Arthroplasty for Idiopathic Femoral Head Osteonecrosis. J. Bone Joint Surg. Am. 98, 672–676 (2016).
 Brown, T. D., Baker, K. J. & Brand, R. A. Structural consequences of subchondral bone involvement in segmental osteonecrosis of the femoral head. J. Orthop. Res. 10, 79–87 (1992).
- 10. Laroche, M. et al. Dual-energy X-ray absorptiometry in osteonecrosis of the femoral head. Rev. Rhum. Engl. Ed 65, 393-396 (1998).
- Craiovan, B. *et al.* Decreased femoral periprosthetic bone mineral density: a comparative study using DXA in patients after cementless total hip arthroplasty with osteonecrosis of the femoral head versus primary osteoarthritis. *Arch. Orthop. Trauma Surg.* 136, 709–713 (2016).
- 12. Calder, J. D., Pearse, M. F. & Revell, P. A. The extent of osteocyte death in the proximal femur of patients with osteonecrosis of the femoral head. *J. Bone Joint Surg. Br.* 83, 419–422 (2001).
- Calder, J. D., Hine, A. L., Pearse, M. F. & Revell, P. A. The relationship between osteonecrosis of the proximal femur identified by MRI and lesions proven by histological examination. J. Bone Joint Surg. Br. 90, 154–158 (2008).
- 14. Kim, Y.-H. & Kim, J.-S. Histologic analysis of acetabular and proximal femoral bone in patients with osteonecrosis of the femoral head. *J. Bone Joint Surg. Am.* 86–A, 2471–2474 (2004).
- Mutijima, E., De Maertelaer, V., Deprez, M., Malaise, M. & Hauzeur, J.-P. The apoptosis of osteoblasts and osteocytes in femoral head osteonecrosis: its specificity and its distribution. *Clin. Rheumatol.* 33, 1791–1795 (2014).
- Tingart, M. *et al.* Influence of factors regulating bone formation and remodeling on bone quality in osteonecrosis of the femoral head. *Calcif. Tissue Int.* 82, 300–308 (2008).
- 17. Wang, C. et al. Bone microstructure and regional distribution of osteoblast and osteoclast activity in the osteonecrotic femoral head. PloS One 9, e96361 (2014).
- 18. Morris, M. D. & Mandair, G. S. Raman assessment of bone quality. Clin. Orthop. 469, 2160-2169 (2011).

- Donnelly, E., Chen, D. X., Boskey, A. L., Baker, S. P. & van der Meulen, M. C. H. Contribution of mineral to bone structural behavior and tissue mechanical properties. *Calcif. Tissue Int.* 87, 450–460 (2010).
- Yerramshetty, J. S. & Akkus, O. The associations between mineral crystallinity and the mechanical properties of human cortical bone. Bone 42, 476–482 (2008).
- 21. Misof, B. M. et al. Bone material properties in premenopausal women with idiopathic osteoporosis. J. Bone Miner. Res. 27, 2551–2561 (2012).
- 22. Mandair, G. S. & Morris, M. D. Contributions of Raman spectroscopy to the understanding of bone strength. *BoneKEy Rep.* 4, 620 (2015).
- 23. Turunen, M. J. *et al.* Comparison between infrared and Raman spectroscopic analysis of maturing rabbit cortical bone. *Appl. Spectrosc.* **65**, 595–603 (2011).
- Pascart, T. et al. Bone Samples Extracted from Embalmed Subjects Are Not Appropriate for the Assessment of Bone Quality at the Molecular Level Using Raman Spectroscopy. Anal. Chem. 88, 2777–2783 (2016).
- Aruwajoye, O. O., Kim, H. K. W. & Aswath, P. B. Bone apatite composition of necrotic trabecular bone in the femoral head of immature piglets. *Calcif. Tissue Int.* 96, 324–334 (2015).
- 26. Fan, M., Peng, J., Qin, L. & Lu, S. Experimental animal models of osteonecrosis. Rheumatol. Int. 31, 983-994 (2011).
- Gamsjaeger, S., Brozek, W., Recker, R., Klaushofer, K. & Paschalis, E. P. Transmenopausal changes in trabecular bone quality. J. Bone Miner. Res. 29, 608–617 (2014).
- 28. Gualeni, B. et al. Alteration of proteoglycan sulfation affects bone growth and remodeling. Bone 54, 83–91 (2013).
- Weinstein, R. S., Nicholas, R. W. & Manolagas, S. C. Apoptosis of osteocytes in glucocorticoid-induced osteonecrosis of the hip. J. Clin. Endocrinol. Metab. 85, 2907–2912 (2000).
- Calder, J. D. F., Buttery, L., Revell, P. A., Pearse, M. & Polak, J. M. Apoptosis-a significant cause of bone cell death in osteonecrosis of the femoral head. J. Bone Joint Surg. Br. 86, 1209–1213 (2004).
- 31. Wang, J. et al. iNOS expression and osteocyte apoptosis in idiopathic, non-traumatic osteonecrosis. Acta Orthop. 86, 134–141 (2015).
- 32. Atkins, G. J. & Findlay, D. M. Osteocyte regulation of bone mineral: a little give and take. Osteoporos. Int. 23, 2067-2079 (2012).
- Beckmann, J. et al. A muCT analysis of the femoral bone stock in osteonecrosis of the femoral head compared to osteoarthrosis. Arch. Orthop. Trauma Surg. 129, 501–505 (2009).
- 34. Tingart, M. *et al.* Analysis of bone matrix composition and trabecular microarchitecture of the femoral metaphysis in patients with osteonecrosis of the femoral head. *J. Orthop. Res.* 27, 1175–1181 (2009).
- 35. Zheng, L.-Z. et al. Steroid-associated hip joint collapse in bipedal emus. PloS One 8, e76797 (2013).
- 36. Plenk, H. et al. Magnetic resonance imaging and histology of repair in femoral head osteonecrosis. Clin. Orthop. 42-53 (2001).
- Li, W. et al. Distribution of TRAP-positive cells and expression of HIF-1alpha, VEGF, and FGF-2 in the reparative reaction in patients with osteonecrosis of the femoral head. J. Orthop. Res. 27, 694–700 (2009).
- Paschalis, E. P. et al. Aging Versus Postmenopausal Osteoporosis: Bone Composition and Maturation Kinetics at Actively-Forming Trabecular Surfaces of Female Subjects Aged 1 to 84 Years. J. Bone Miner. Res. 31, 347–357 (2016).
- 39. Lockshin, R. A. & Zakeri, Z. Apoptosis, autophagy, and more. Int. J. Biochem. Cell Biol. 36, 2405-2419 (2004).
- Hofstaetter, J. G., Wang, J., Yan, J. & Glimcher, M. J. Changes in bone microarchitecture and bone mineral density following experimental osteonecrosis of the hip in rabbits. *Cells Tissues Organs* 184, 138–147 (2006).
- Mont, M. A., Seyler, T. M., Marker, D. R., Marulanda, G. A. & Delanois, R. E. Use of metal-on-metal total hip resurfacing for the treatment of osteonecrosis of the femoral head. J. Bone Joint Surg. Am. 88 (Suppl 3), 90–97 (2006).
- 42. Nakasone, S., Takao, M., Sakai, T., Nishii, T. & Sugano, N. Does the extent of osteonecrosis affect the survival of hip resurfacing? *Clin. Orthop.* **471**, 1926–1934 (2013).
- 43. Amstutz, H. C. & Le Duff, M. J. Hip resurfacing results for osteonecrosis are as good as for other etiologies at 2 to 12 years. *Clin. Orthop.* **468**, 375–381 (2010).
- Pyda, M. et al. Hip resurfacing arthroplasty in treatment of avascular necrosis of the femoral head. Med. Sci. Monit. Int. Med. J. Exp. Clin. Res. 21, 304–309 (2015).
- 45. Falgayrac, G. *et al.* Comparison of two-dimensional fast raman imaging versus point-by-point acquisition mode for human bone characterization. *Anal. Chem.* 84, 9116–9123 (2012).
- 46. Karampas, I. A., Orkoula, M. G. & Kontoyannis, C. G. A quantitative bioapatite/collagen calibration method using Raman spectroscopy of bone. *J. Biophotonics* **6**, 573–586 (2013).
- Olejnik, C., Falgayrac, G., During, A., Cortet, B. & Penel, G. Doses effects of zoledronic acid on mineral apatite and collagen quality of newly-formed bone in the rat's calvaria defect. *Bone* 89, 32–39 (2016).
- Aparicio, S. et al. Optimal methods for processing mineralized tissues for Fourier transform infrared microspectroscopy. Calcif. Tissue Int. 70, 422–429 (2002).

Acknowledgements

The authors particularly acknowledge Professor Pierre Gosset for his insightful comments on the histological analyses. The authors also thank Professor Christian Fontaine and M. Maurice Demeulaere from the Lille University Laboratory of Anatomy for their participation in sample collection. We thank M. Patrick Gelé and M. Bertrand Accart for the management and storage of samples.

Author Contributions

T.P. design, data acquisition, analysis, interpretation of data, manuscript drafting. G.F. design, data acquisition, analysis, interpretation of data, manuscript drafting. J.F.Q. design, histological analyses, interpretation of data, drafting and critical revision of the manuscript. L.N. design, statistical analyses, drafting and critical revision of the manuscript. J.P. interpretation of the data and critical revision of the manuscript. B.C. Design, interpretation of data and critical revision of the manuscript. A.C. Design, MRI analyses, critical revision of the manuscript. J.F.B. Design, MRI analyses, critical revision of the manuscript. H.M. Design, Patient recruitment and sampling, critical revision of the manuscript. G.P. Design, interpretation of the data and critical revision of the manuscript.

Additional Information

Competing Interests: The authors declare that they have no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/

© The Author(s) 2017