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CORRESPONDENCE

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CSF3R T618I mutant chronic myelomonocytic leukemia (CMML) defines a proliferative CMML subtype enriched in ASXL1 mutations with adverse outcomes

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Dear Editor,

Chronic myelomonocytic leukemia (CMML) is a clonal myeloid disorder characterized by peripheral blood (PB) monocytosis (absolute monocyte count (AMC) $\geq 1 \times 10^9/L$ and ≥10% of the total white blood cell count (WBC)) and overlapping features of myelodysplastic syndromes and myeloproliferative neoplasms (MPN)¹. Sustained PB monocytosis is a hallmark of CMML, with a mutational signature consisting of biallelic TET2, or TET2 and SRSF2 co-mutations, biasing hematopoiesis toward monocytosis². Chronic neutrophilic leukemia (CNL) is a rare MPN characterized by sustained mature neutrophilia (WBC≥ 25×10^9 /L, with segmented neutrophils and band forms \geq 80% of the WBC), with an AMC $< 1 \times 10^9/L^3$. CNL commonly occurs due to oncogenic driver mutations involving the colony-stimulating factor receptor, CSF3R (90–100%)^{4,5}. Among CSF3R mutations described in CNL, the T6181 membrane proximal point mutation activating Janus-activated kinase (JAK)/signal transducer and activator of transcription factor signaling is the most common (74%)^{4,6}. These mutations potentially render CNL responsive to therapy with JAK inhibitors, such as ruxolitinib. On the other hand, CSF3R mutations, especially *CSF3R* T618I, are very infrequent in CMML (<5%)^{8,9}. We carried out this study to assess the prevalence, phenotypic features, and outcomes of patients with CMML who harbor the *CSF3R* T618I mutation.

Eight hundred and forty-six patients with CMML, defined per the World Health Organization (WHO) 2016 criteria, from Mayo Clinic, Minnesota and the Groupe Français des Myelodysplasies were included in this study (August 1994 through July 2019). All patients underwent targeted next-generation sequencing for myeloid-relevant genes obtained at diagnosis or at first referral using previously described methods^{2,10}. Categorical variables were compared by Fisher's exact or Pearson's chi square. Continuous variables were compared by Wilcoxon test. Overall survival (OS) was defined as the time from diagnosis to death. Acute myeloid leukemia-free survival (AML-FS) was defined as the time from diagnosis to transformation to AML or death. Patients who underwent hematopoietic cell transplantation prior to AML transformation were censored for AML-FS estimates. Both OS and AML-FS were estimated by the Kaplan-Meier method and compared by log-rank test.

We identified 6 (1%) patients who met the 2016 WHO criteria for CMML and who harbored CSF3R mutations, all of whom had the CSF3R T618I missense alteration; 3 (50%) were males, median age 63 years, with a median AMC of $6\times10^9/L$ (range 2–23). Five (100%) of five evaluable patients had bone marrow (BM) dysplasia, a finding not seen in CNL. As expected, all six CSF3R T618I mutant patients had a proliferative CMML phenotype, with marked neutrophilia in addition to the monocytosis. Four (100%) of the 4 evaluable patients also had circulating immature myeloid cells (myelocytes and metamyelocytes).

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Table 1

Patient	Patient Age, years Gender CMM subty	's Gende	r CMI.	ML type	CBC						ВМ	Karyotype		CSF3R VAF %	CSF3R Co-mutations Treatment Outcomes VAF % (VAF %)	Treatment	Outcomes
			FAB	FAB WHO	Hb, g/dL	Plts, ×10 ⁹ /L	WBC, ×10 ⁹ /L	ANC, ×10 ⁹ / L (%)	AMC, ×10 ⁹ / L (%)	PB blasts (%)	Blasts (%) Dysplasia	plasia					
<u></u>	62	ш	۵	2	13	50	40	21 (53)	6 (15)	12	16	46, X, [14]/4t	46, X, i(Xq13) [14]/46, XX [6]	10	SRSF2 (49)	HU/ induction	AML transformation in 1 month and death 9 months later
2	39	ш	۵	2	9	50	134	114 (85)	19 (14)	9	15 +	46, X, .	46, X, i(Xq13) [7]/ 63 46, XX [4]	63	ASXL1 (35)/ STEBP1 (51)	H	No follow-up
m	48	≥	Д	0	_	168	180	151 (84)	23 (13)	0	+	46, XY [20]		28	ASXL1 (25)	HMA	Refractory disease and died in 20 months
4	99	Σ	Д	0	∞	09	31	25 (81)	4 (13)	0	+	46, XY [20]		51	ASXL1 (45)	H	Alive at 6 months
2	69	ட	۵	_	∞	173	35	23 (66)	5 (14)	2	+	46, X, del() 46, XX [8]	46, X, del(X) [12]/ 12 46, XX [8]	12	ASXL1 (46)	₽ H	No follow-up
9	64	≥	۵	0	=	302	21	18 (86)	2 (11)	0	3 UK	46, XY [UK]		UK	ASXL1 (49)/ UA2F1 (44)	HO	Died in 17 months

Individual characteristics and outcomes of CMML patients with CSF3R T618I mutation. Characteristics are based on diagnosis or first referral.

M male, F female, WHO World Health Organization, FAB French-American-British, CBC complete blood count, Hb hemoglobin, WBC white blood count, AMC absolute neutrophils count, AMC absolute monocyte count, Plt platelets, PB peripheral blood, BM bone marrow, VAF variant allele frequency, HU hydroxyurea, induction cytarabine-based intense chemotherapy regimens, RBC red blood cells, AML acute myeloid leukemia, UK unknown.

The *CSF3R* T618I variant allele frequency (VAF) burdens were >40% in 3 (60%) of 5 evaluable cases (range 10-63%). While the *CSF3R* mutational VAF in three patients was in the heterozygous range, all three had adult-onset leukocytosis/monocytosis, making the likelihood that these variants were germline, quite low^{11} . We, however, acknowledge that we did not have germline tissue for further confirmation. Three patients had a normal karyotype, while the remaining three had X chromosome abnormalities (Table 1).

In comparison to CSF3R wild-type (WT) CMML patients, those with the CSF3R T618I mutation were younger (median 63 vs. 72 years; p = 0.0055), had lower hemoglobin values (Hb, median 8 vs. 11 g/dl; p = 0.0186), higher WBC (median 38 vs. $13 \times 10^9 / \text{L}$; p = 0.0013), higher percentage of neutrophils (median 83 vs. 52%; p =0.001), higher AMC (median 6 vs. $2 \times 10^9 / \text{L}$; p = 0.024), lower percentage of monocytes (median 13 vs. 23%; p =0.0069), higher percentage of PB blasts (median 1 vs. 0%; p = 0.0472), and were more likely to be classified as proliferative CMML (100 vs. 48%; p = 0.012). There were no TET2 mutations in the CSF3R T618I mutant CMML group, vs. 59% in the WT group (p = 0.005), while ASXL1 mutations were more frequent in the CSF3R T618I mutant group (83 vs. 41%; p = 0.045) in comparison to the WT group (Supplementary Table 1). While not statistically significant, 5 of the 6 (83%) CSF3R T618I mutant CMML patients did not have concomitant SRSF2 mutations, with 4 of the 6 (66%) not having any splicing mutation at all.

At last follow-up, 3 (50%) and 400 (48%) deaths and 1 (17%) and 122 (15%) leukemic transformations were documented in the *CSF3R* T618I mutant and WT CMML groups, respectively. The median OS was significantly shorter in CMML patients with the *CSF3R* T618I mutation (median 1.4 vs. 3.0 years; p=0.049), in comparison to the WT patients, with no difference in the AML-FS (1.4 vs. 3.2 years; p=0.17) (Fig. 1). None of our patients received ruxolitinib or underwent allogeneic stem cell transplantation. Five of the six patients were treated with hydroxyurea and one patient was treated with hypomethylating agent therapy, with no response.

Given the higher prevalence of *ASXL1* mutations in the *CSF3R* T618I mutated cohort, and the known detrimental prognostic impact of *ASXL1* mutations in CMML, we performed a subgroup analysis comparing *CSF3R* T618I mutant CMML with *ASXL1* mutant CMML patients (Supplementary Table 1)¹⁰. In this comparison, once again, *CSF3R* mutant CMML patients were more likely to be younger in age (p = 0.009), have lower Hb levels (p = 0.005), have higher WBC (p = 0.009), have lower AMC (p = 0.012), and were less likely to have *TET2* mutations (p = 0.029). There was no OS difference between the two groups (Supplementary Fig. 1).

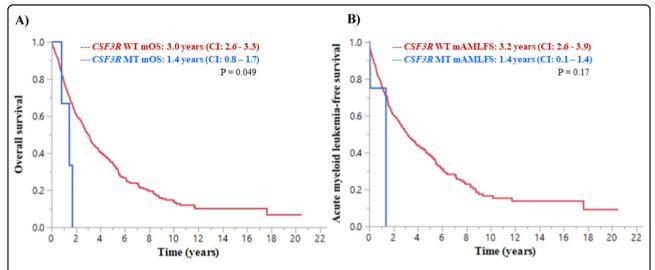


Fig. 1 Kaplan–Meier survival analysis of CMML patients stratified by the presence or absence of the CSF3RT618I mutation. A Inferior overall survival of CSF3RT618I mutant CMML in comparison to CSF3R wildtype patients. B Inferior acute myeloid leukemia-free survival of CSF3RT618I mutant CMML in comparison to CSF3R wildtype patients.

In summary, we present a cohort of patients who met 2016 WHO criteria for CMML and who harbored an oncogenic driver mutation in CSF3R T618I. While this mutation is fairly specific to CNL, infrequent occurrences in other myeloid neoplasms have been documented^{8,9}. In our opinion, in the context of accurate histopathological analyses, the true frequency of these mutations in CMML is <1%. In addition, the CSF3R T618I mutation when present defines a unique proliferative CMML subtype characterized by pronounced leukocytosis/neutrophilia/monocytosis, with a unique molecular signature (ASXL1mt/TET2wt), with infrequent splicing mutations. The high frequency of X chromosome abnormalities remains to be defined. We have in the past demonstrated the negative prognostic impact of the ASXL1mt/TET2wt genotype in CMML, a finding once again validated by the poor OS of this group in the current study (median OS 1.4 years)^{2,12}. In fact, the molecular landscape of CSF3R T618I mutant CMML is more akin to BCR-ABL1-negative atypical chronic myeloid leukemia (CML), where ASXL1 mutations are frequent (80%) and splicing mutations are less common (30%)¹³. However, atypical CML is not associated with monocytosis, and given the fact that in CMML biallelic TET2 or TET2/SRSF2 comutations skew hematopoiesis toward monocytosis, the etiology of monocytosis in these patients remains to be elucidated. While the presence of BM dysplasia along with concomitant monocytosis/neutrophilia morphologically distinguishes CSF3R T618I mutant CMML from CNL, further work is needed to see whether CSF3R T618I mutant CMML is truly a CMML subtype, or if akin to PDGFRA/B and FGFR1 rearranged myeloid neoplasms, it deserves its own classification schema as a CSF3R T618I mutant myeloid neoplasm with monocytosis.

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Author contributions

E.D.B. collected and interpreted the data, performed statistical analysis, and wrote the manuscript; T.L.L. interpreted the data and reviewed the manuscript; C.M.F. collected the data; A.N.S., M.A.E., A.D.P., N.G., A.A.M., R.P.K., A.T., and E.S. reviewed the manuscript; and M.M.P. concepted the study, interpreted the data, and revised the manuscript.

Conflict of interest

The authors declare no competing interests.

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